Supporting information for

The role of aristolochene synthase in diphosphate activation

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Table of contents

| Site directed mutagenesis of recombinant PR-AS cDNA | S2 |
|--|--------|
| Expression of wild type PR-AS and mutants | S3 |
| GC/MS analysis | S3 |
| Steady-state kinetic assays | S3 |
| Kinetic parameters and product distributions for PR-AS and mutants | S4 |
| Amino acid sequence alignment of fungal and bacterial sesquiterpene synthase | sS5 |
| Representative GC/MS spectra | S6-S9 |
| Representative Lineweaver-Burk plots | S9-S10 |
| Representative Michaelis-Menten plots | |
| References | S12 |

Site directed mutagenesis of recombinant PR-AS cDNA

The Quickchange site-directed mutagenesis kit (Stratagene) was used to introduce the desired mutations according to the manufacturer's instructions. The primers used for mutagenesis were as follows:

5'-GGGCATTATCTCGAGTACAAAGAGAAGGATGTAGGCAAG-3' and 5'-CTTGCCTACATCCTTCTCTTTGTACTCGAGATAATGCCC-3' for R200K; 5'-GGCATTATCTCGAGTACGAAGAGAAGGATGTAGGCAAG-3' and 5'-CTTGCCTACATCCTTCTCTTCGTACTCGAGATAATGCC-3' for R200E; 5'-GCATTATCTCGAGTACCAGGAGAAGGATGTAGGCAAG-3' and 5'-CTTGCCTACATCCTTCTCCTGGTACTCGAGATAATGC-3' for R200Q; 5'-TCGAGTACCGTGAGAAGCTGGTAGGCAAGGCGTTGC-3' and 5'-GCAACGCCTTGCCTACCAGCTTCTCACGGTACTCGA-3' for D203L; 5'-CATATACAGTTATGACCAGGAAGAGGAAGCGTC-3' and 5'-GACGCTTCCTCTTCCTGGTCATAACTGTATATG-3' for K251Q; 5'-GACATATACAGTTATGACCGTGAAGAGGAAGCGTCTCG-3' and 5'-CGAGACGCTTCCTCTTCACGGTCATAACTGTATATGTC-3' for K251R 5'-GGAGCAAGACCACGCGTAAATACAACTAAGGATCCTC-3' and 5'-GAGGATCCTTAGTTGTATTTACGCGTGGTCTTGCTCC-3' for R340K; 5'-GAGCAAGACCACGCGTATGTACAACTAAGGATCCTC-3' and 5'- GAGGATCCTTAGTTGTACATACGCGTGGTCTTGCTC-3' for R340M; 5'-CAAGACCACGCGTCGCTTTAACTAAGGATCCTCTAG-3' and 5'-CTAGAGGATCCTTAGTTAAAGCGACGCGTGGTCTTG-3' for Y341F;

Plasmids were purified from overnight cultures (10 mL LB medium containing ampicillin 50 μ M/mL) using the QIAGEN miniprep kit as described by the manufacturer. Mutations were confirmed by DNA sequence analysis using Eurofins MWG.

Expression of wild type PR-AS and mutants

PR-AS and mutants were overproduced in *E. coli* BL21(DE3) cells and purified as previously described.¹

GC/MS analysis

Proteins (50 μ M) were incubated with FPP (1 mM) in incubation buffer (25 mM Tris, pH 7.5, 5 mM beta-mercaptoethanol, 15% glycerol and 5 mM MgCl₂) in a total volume of 250 μ L. The reaction was overlaid with pentane and left overnight. Products were extracted with 2 portions of 500 μ L pentane and the combined organic layers passed through a short column of aluminium oxide. GC-MS analysis of incubation products was performed using a HP 6890 GC-MS system fitted with J & W Scientific HP-5MS column (30 m length and internal diameter of 0.25 mm) and a Micromass GCT Premiere mass spectrometer for detection. The program uses an initial oven temperature of 50 °C with a gradient of 4 °C min⁻¹ over 25 min followed by a gradient of 20 °C min⁻¹ over 5 min to a final temperature of 250 °C.

Steady-state kinetic assays

Kinetic assays were carried out as previously described.²⁻⁴ Steady-state parameters were obtained by direct fitting of the data to the Michaelis-Menten equation by nonlinear least squares regression using the graphical procedures developed by Lineweaver-Burk⁵ within the SigmaPlot package (Systat Software).

| | 4 | 2 | 5 | K _M | $k_{\rm cat} \mathrm{x10^{-3}}$ | k _{cat} /K _M |
|-------|------|------|-----|----------------|----------------------------------|----------------------------------|
| | (%) | (%) | (%) | (µM) | (s^{-1}) | $(M^{-1}s^{-1})$ |
| WT | 91.5 | 7.5 | 1 | 0.53 | 84 | 158490 |
| R200K | 19.2 | 78.9 | 1.8 | 1.04 | 0.6 | 620 |
| R200Q | | | | Inactive | | |
| R200E | | | | Inactive | | |
| D203L | 17.3 | 79.7 | 3 | 0.74 | 0.75 | 1013 |
| K251R | 90.2 | 6 | 3.8 | 0.14 | 3.7 | 26428 |
| K251Q | 84.6 | 9.8 | 5.6 | 0.24 | 2 | 8500 |
| R340K | 54.4 | 42.8 | 2.8 | 1.59 | 0.8 | 510 |
| R340M | | | | Inactive | | |
| Y341F | 93.3 | 3.8 | 2.9 | 0.18 | 2.5 | 14000 |

Table 1: Kinetic parameters and product distributions for PR-AS and mutants^a

^aReported values are the average of 3 measurements; all values were within 5% of the average.

| AT-AS | ASSSLEPPPSTFQPLCHPLVEEVSKE 35 | |
|---------|--|-----|
| PR-AS | MATSTETISSLAQPFVHLENPINSPLVKETIRPRNDTTITPPPTQWSYLCHPRVKEVQDE 60 | |
| SC-EIZS | PSLRLPVIEAAFPROLHPYWPKLOET 44 | |
| SP-PS | LPGROSPDHARAEAE 25 | |
| FS-TS | MENFPTEYFLNTTVRLLEYIRYRDSNYTREERIENLHYAYNKAAHHF 47 | |
| AT-AS | VDGYFLQHWNFPNEKARKKFVAAGFSRVTCLYFPKALDDRIHFACRLLTVLFLIDDLLEY 95 | |
| PR-AS | VDGYFLENWKFPSFKAVRTFLDAKFSEVTCLYFPLALDDRIHFACRLLTVLFLIDDVLEH 120 | 0 |
| SC-EIZS | TRTWLLEKRLMPADKVEEYADGLCYTDLMAGYYLGAPDEVLOAIADYSAWFFVWDDRHDR 104 | 4 |
| SP-PS | OLAWPRSLGLIRSDAAAERHLRGGYADLASRFYPHATGADLDLGVDLMSWFFLFDDLFDG 85 | |
| FS-TS | AOPROOOLLKVDPKRLOASLOTIVGMVVYSWAKVSKECMADLSIHYTYTLVLDDSKD- 104 | 4 |
| | | |
| | | |
| AT-AS | MSFEEGSAYNEKLIPISRGDVLPDRSIPVEYIIYDLWESMRAHDR 14 | 40 |
| PR-AS | MSFADGEAYNNRLIPISRGDVLPDRTKPEEFILYDLWESMRAHDA 10 | 65 |
| SC-EIZS | DIVHGRAGAWRRLRGLLHTALDSPGDHLHHEDTLVAGFADSVRRLYAFLPATWNA 1 | 59 |
| SP-PS | PRGEN-PEDTKQLTDQVAAALDGPLPDTAPPIAHGFADIWRRTCEGMTPAWCA 1 | 37 |
| FS-TS | DPYPTMVNYFDDLQAGREQAHPWWALVNEHFPNVLRHFGPFCSLNLIRSTLDFFEGCWIE 1 | 64 |
| | * : : : * | |
| | | 0.5 |
| AI-AS | EMADELLEPVFLFMRAQIDRIR-ARPMGLGGI-LEIRERDVGRELLAALMRFS-MGLK 1 | 20 |
| PR-AS | ELANEVLEPTFVFMRAQIDRAR-LSTHELGHT-LETRERDVGRALLSALMRFSMGLR 2. | 20 |
| SC-EIZS | RFARHFHTVIEAYDREFHNRTR-GIVPGVEEYLELRRLTFAHWIWTDLLEPSSGCE 2. | 14 |
| SP-PS | RSARHWRNYFDGYVDEAESRFWNAPCDSAAQY - LAMRRHTIGVQPTVDLAERA - GRFE 1 | 93 |
| FS-TS | QYNFGGFPGSHDYPQFLRRMNGLGHCVGASLWPKEQFNERSLFLEITSAIAQMENWMVWV 2 | 24 |
| | . : : : . | |
| AT-AS | LSPSELORVREIDANCSKHLSVVNDIYSYEKELYTSKTAHSEGGILCTSVOILAOEA- 2 | 52 |
| PR-AS | LSADELÕDMKALEANCAKOLSVVNDIYSYDKEEEASRTGHKEGAFLCSAVKVLAEES- 2 | 77 |
| SC-EIZS | IPDAVRKHPAYRRAALLSOEFAAWYNDLCSLPKEIAGDEVHNLGISLITHHSLTLEEAIG 2 | 74 |
| SP-PS | VPHRVEDSAVMSAMLOIAVDVNIJINDIASLEKEEARGEONNMVMILRREHGWSKSESVS 2 | 53 |
| FS-TS | NDLMSEYKEEDDERDOISLVKNYVVSDEISLNEALEKLTODTLHSSKOMVAVESDKDPOV 2 | 84 |
| 10 10 | · · · * * : | 01 |
| | | |
| AT-AS | DVTAEAAKRVLFVMCREWELRHQLLVAR-LSAEGLETPGLAAYVEGLEYQMSGNELWS 3 | 09 |
| PR-AS | KLGIPATKRVLWSMTREWETVHDEIVAEKIASPDGCSEAAKAYMKGLEYQMSGNEQWS 3 | 35 |
| SC-EIZS | EVRRRVEECITEFLAVERDALRFADELADGTVRGKE-LSGAVRANVGNMRNWFSSVYWFH 3 | 33 |
| SP-PS | HMQNEVRARLEQYLLLESCLPKVGEIYQLDTAEREALERYRTDAVRTVIRGSYDWH 3 | 09 |
| FS-TS | MDTIECFMHGYVTWHLCDRRYRLSEIYEKVKEEKTEDAQKFCKFYEQAANVGAVSPSEWA 34 | 44 |
| | : : : | |
| ΔT-ΔS | ОТТІ. <mark>РУ</mark> SVVVD 320 | |
| DR-JC | XTTRVV | |
| SC-FIZS | HESCEVINDSWIDDRSTDDVVNINFAACEK 361 | |
| | DCC(DVD) EEVI'' V WREWERE CCCVT = - 201 | |
| | | |
| 1.9-19 | TELAVÖNUN VENÄKLIPPIENNE 214 | |

Figure S1. Amino acid sequence alignment of fungal/bacterial sesquiterpene synthases

ClustalW2.⁶ AT-AS and PR-AS, *A. terreus* and *P. roqueforti* aristolochene synthases; EIZS, *S. coelicolor* epi-isozizane synthase; PS, *Streptomyces sp.* pentalenene synthase; TS, *Fusarium sporotrichioides* trichodiene synthase.





Figure S2. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS: aristolochene (retention time, 24.60 min), valencene (24.93 min) and germacrene A (25.22 min).



Figure S3. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-R200K: aristolochene (retention time, 24.60 min), valencene (24.93 min) and germacrene A (25.22 min).



Figure S4. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-D203L: aristolochene (retention time, 25.18 min), valencene (25.48 min) and germacrene A (25.82 min).



Figure S5. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-K251Q: aristolochene (retention time, 25.60 min), valencene (25.88 min) and germacrene A (26.23 min).



Figure S6. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-K251R: aristolochene (retention time, 25.60 min), valencene (25.88 min) and germacrene A (26.23 min).



Figure S7. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-R340K: aristolochene (retention time, 24.55 min), valencene (24.93 min) and germacrene A (25.22 min).



Figure S8. Representative total ion chromatogram of the pentane extractable products formed from incubation of FDP and PR-AS-Y341F: aristolochene (retention time, 25.60 min), valencene (25.88 min) and germacrene A (26.23 min).



Lineweaver-Burk plots

Figure S9. Representative Lineweaver-Burk plot for PR-AS-R200K.



Figure S10. Representative Lineweaver-Burk plot for PR-AS-D203L.



Figure S11. Representative Lineweaver-Burk plot for PR-AS-R340K.

Michaelis-Menten plots



Figure S12. Representative Michaelis-Menten plot for PR-AS-K251R.



Figure S13. Representative Michaelis-Menten plot for PR-AS-K251Q.



Figure S14. Representative Michaelis-Menten plot for PR-AS-Y341F.

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