Supporting Information for

Chirality induction in metal-induced achiral polythiophene aggregates assisted by optically active amines and polythiophene[†]

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1. Experimental Procedures

1-1. Instruments

The melting points were measured using a Yanaco MP-500D melting point apparatus (Kyoto, Japan) and were uncorrected. The IR spectra were recorded using a JASCO Fourier Transform IR-680 spectrophotometer (Hachioji, Japan). The NMR spectra were obtained using a Varian UNITY INOVA 500AS spectrometer operating at 500 MHz for ¹H and 125 MHz for ¹³C using tetramethylsilane (TMS) or the solvent residual peaks as the internal standards. The DOSY experiments were carried out by using pulsed field gradient with the BPPSTE pulse sequence.¹ The hydrodynamic diameters (d_h) were calculated from the diffusion constants (D) obtained by DOSY measurements according to the Einstein-Stokes equation, $d_{\rm b} = k_{\rm B}T/3\pi\eta_0 D$ ($k_{\rm B}$: Boltzmann constant; T: absolute temperature; η_0 : solvent viscosity; D: diffusion constant). The electron spray ionization mass spectra (ESI-MS) were recorded on a JEOL JMS-T100CS spectrometer (Akishima, Japan). The absorption and CD spectra were measured in a 2- or 5-mm quartz cell on a JASCO V-570 spectrophotometer and a JASCO J-820 spectropolarimeter, respectively. The temperature was controlled by a JASCO PTC-423L apparatus (-5 to 55 °C). The elemental analyses were performed by the Analytical Laboratory in the Graduate School of Bioagricultural Sciences, Nagoya University. The size exclusion chromatography (SEC) was performed with a JASCO PU-980 liquid chromatograph equipped with a JASCO DG-980-50 degasser and a UV-visible detector (254 nm; JASCO UV-970) at 40 °C. An SEC column (Tosoh TSK-GEL Multipore H_{xL}-M: 30 x 0.78 cm (i.d.): pore size, 0.01–0.6 μ m; bead size, 5 μ m) was connected, and CHCl₃ was used as eluent at a flow rate of 1.0 mL/min. The molecular weight calibration curve was obtained with standard polystyrenes (Tosoh).

1-2. Materials

All starting materials and dehydrated solvents were purchased from Sigma-Aldrich (St. Louis, Missouri, USA), Wako Pure Chemical Industries, Ltd. (Osaka, Japan), and Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). Triethylamine was distilled over CaH₂ under Ar after being stirred with KOH pellets overnight. The deuterium solvents were purchased from Merck (Darmstadt, Germany) or Cambridge Isotope Laboratories (Andover, Massachusetts, USA) and degassed with Ar and used throughout all the experiments. Silica gel (SiO₂) for the flash chromatography was purchased from Merck. An optically active regioregular polythiophene, poly[3-[4-((*R*)-4-ethyl-2-oxazolin-2-yl)phenyl]thiophene] (PEOPT) ($M_n = 5.3 \times 10^3$, DP_n = 21, $M_w/M_n = 1.25$),² and achiral regioregular poly(3-hexylthiophene) (PHT) ($M_n = 8.2 \times 10^3$, DP_n = 49, $M_w/M_n = 1.80$)³ were synthesized according to the previously reported methods.

1-3. Synthetic procedures

The polythiophene derivative bearing an achiral oxazoline group (P1) was synthesized according to Scheme S1.² The reaction progresses were monitored by TLC or ESI-MS.





^aReagents and conditions: (a) 2-amino-2-methyl-1-propanol, Me₃Al, toluene, reflux, 2 days; (b) 4-toluenesulfonyl chloride, Et₃N, CH₂Cl₂, rt, 6 days; (c) 3-thiopheneboronic acid, Pd(PPh₃)₄, K₃PO₄, toluene, reflux, 2 days; (d) NBS, DMF, rt, 3 days, (e) (i) LDA, THF, -98 °C; (ii) MgBr₂•Et₂O, -98 °C, 10 min; (iii) Ni(dppp)Cl₂, -98 °C (15 min), -60 °C (1 h), rt (2.5 h), and then reflux (20 h).

2-[*N*-(**4-Iodobenzoyl)amino**]-**2-methyl-1-propanol.** To a toluene (100 mL) solution of 2-amino-2-methyl-1-propanol (11.0 mL, 115 mmol) was added dropwise trimethylaluminium in *n*-hexane (1.56 M) (100 mL, 156 mmol) at 0 °C via a cannula within 40 min under nitrogen. The reaction mixture was then allowed to warm to room temperature and further stirred for 1 h. A toluene (150 mL) solution of ethyl 4-iodobenzoate (25.0 g, 90.6 mmol) was added dropwise to the mixture via a dropping funnel within 50 min at that temperature, and the mixture was then refluxed for 2 days. After cooling to room temperature, 20% aqueous solution of Rochelle salt (100 mL) was added. After filtration, the filtrate was dissolved in ethanol and the solvent was removed by evaporation. The crude product was purified by recrystallization with ethanol (10 mL) and *n*-hexane (20 mL) to give the desired product as a slightly yellow needle crystal (24.6 g, 77.1 mmol, 85.1% yield). IR (KBr, cm⁻¹): 3375, 3301, 3081, 2968, 2873, 1639, 1587, 1543, 1331, 1049, 877, 715. ¹H NMR (CDCl₃): δ 1.00 (s, CH₃, 6H), 2.33 (s, OH, 1H), 3.70 (s, OCH₂, 2H), 6.22 (s, NH, 1H), 7.48 (d, Ph-H₂, *J* = 8.4 Hz, 2H), 7.77 (d, Ph-H₃, *J* = 8.4 Hz, 2H).

2-(4-Iodophenyl)-4,4-dimethyl-2-oxazoline. To solution of а 2-[N-(4-iodobenzoyl)amino]-2-methyl-1-propanol (24.5 g, 76.8 mmol) prepared above in CH₂Cl₂ (200 mL) and triethylamine (60 mL, 430 mmol) was added dropwise a CH₂Cl₂ (20 mL) solution of 4-toluenesulfonyl chloride (22.0 g, 116 mmol) at 0 °C under nitrogen. The reaction mixture was allowed to warm to room temperature and further stirred for 6 days. The mixture was then poured into saturated aqueous NH₄Cl (100 mL) and extracted with CHCl₃. The extract was washed with water and brine and dried over anhydrous Na₂SO₄. After evaporation, the crude product was purified by SiO₂ chromatography with *n*-hexane/ethyl acetate (9/1, vol/vol) as the eluent to give the desired product as a colorless needle crystal (21.8 g, 72.4 mmol, 94.3% yield). IR (KBr, cm⁻¹): 2956, 2930, 2872, 1645, 1588, 1483, 1460, 1393, 1348, 1332, 1071, 1007, 897, 829, 728. ¹H NMR (CDCl₃): δ 0.99 (s, CH₃, 6H), 4.06 (s, OCH₂, 2H), 7.67 (d, Ph-H₂, *J* = 6.8 Hz, 2H), 7.76 (d, Ph-H₃, *J* = 6.8 Hz, 2H).

3-[4-(4,4-Dimethyl-2-oxazolin-2-yl)phenyl]thiophene То mixture of (1). a 3-thiopheneboronic acid (10.3 g, 80.5 mmol), tetrakis(triphenylphosphine)palladium(0) (2.53 g, 2.19 mmol), and potassium phosphate (25.4 g, 120 mmol) in toluene (285 mL) was added 2-(4-iodophenyl)-4,4-dimethyl-2-oxazoline (21.3 g, 70.7 mmol) at room temperature under nitrogen. The mixture was refluxed under nitrogen for 2 days. After cooling to room temperature, the mixture was poured into saturated aqueous NH₄Cl (200 mL) and extracted with CHCl₃. The extract was washed with water and brine and dried over anhydrous Na₂SO₄. After evaporation, the crude product was purified by SiO₂ chromatography with *n*-hexane/ethyl acetate (9/1, vol/vol) as the eluent to give **1** as a white plate crystal (16.7 g, 64.9 mmol, 91.8% yield). Mp: 123–125 °C. IR (KBr, cm⁻¹): 3102, 2969, 2926, 2896, 1643, 1608, 1357, 1320, 1304, 1185, 1068, 963, 849, 784, 740, 631. ¹H NMR (CDCl₃): δ 1.39 (s, CH₃, 6H), 4.11 (s, OCH₂, 2H), 7.39 (dd, Th-H₄, *J* = 5.0, 3.0 Hz, 2H), 7.42 (dd, Th-H₅, *J* = 5.0, 1.5 Hz, 2H), 7.52 (dd, Th-H₂, J = 3.0, 1.5 Hz, 2H), 7.62 (d, Ph-H₂, J = 8.5 Hz, 2H), 7.96 (d, Ph-H₃, J = 8.5 Hz, 2H). ¹³C NMR (CDCl₃): δ 28.40, 67.56, 79.05, 121.24, 126.09, 126.13, 126.46, 126.62, 128.73, 138.30, 141.41, 161.79. ESI-MS (CH₃OH/CH₂Cl₂ (80/20, vol/vol), positive): Calcd for $C_{15}H_{16}NOS \ [1+H]^+: m/z = 258.10$. Found: m/z = 258.08. Elemental Anal. Calcd for C₁₅H₁₅NOS: C, 70.01; H, 5.87; N, 5.44. Found: C, 69.97; H, 5.92; N, 5.29.

2-Bromo-3-[4-(4,4-dimethyl-2-oxazolin-2-yl)phenyl]thiophene

N-Bromosuccinimide (11.7 g, 65.7 mmol) was added to a solution of **1** (16.7 g, 64.9 mmol) in DMF (100 mL) at room temperature, and the mixture was stirred for 3 days under shielded light. After the solvent was evaporated under reduced pressure, the mixture was poured into saturated aqueous NH₄Cl (100 mL) and extracted with diethyl ether. The extract was washed with water and brine and dried over anhydrous Na₂SO₄. The solvent was removed by evaporation, and the crude product was purified by SiO₂ chromatography with *n*-hexane/ethyl

(**1Br**).

acetate (10/1, vol/vol) as the eluent to give **1Br** as a slightly yellow solid (19.9 g, 59.2 mmol, 91.2% yield). Mp: 56–57 °C. IR (KBr, cm⁻¹): 3106, 2966, 2926, 2891, 1648, 1612, 1352, 1315, 1301, 1184, 1061, 1018, 988, 967, 876, 849, 717, 688. ¹H NMR (CDCl₃): δ 1.40 (s, CH₃, 6H), 4.12 (s, OCH₂, 2H), 7.04 (d, Th-H₄, *J* = 5.5 Hz, 2H), 7.31 (d, Ph-H₅, *J* = 5.5 Hz, 2H), 7.59 (d, Ph-H₂, *J* = 8.5 Hz, 2H), 8.00 (d, Ph-H₃, *J* = 8.5 Hz, 2H). ¹³C NMR (CDCl₃): δ 28.38, 67.60, 79.14, 109.27, 126.13, 127.15, 128.25, 128.48, 128.88, 137.73, 140.36, 161.76. ESI-MS (CH₃OH/CH₂Cl₂ (80/20, vol/vol), positive): Calcd for C₁₅H₁₅BrNOS [**1Br**+H]⁺: *m/z* = 338.00. Found: *m/z* = 337.99. Elemental Anal. Calcd for C₁₅H₁₄BrNOS: C, 53.58; H, 4.20; N, 4.17. Found: C, 53.64; H, 4.29; N, 4.04.

Poly[3-[4-(4,4-dimethyl-2-oxazolin-2-yl)phenyl]thiophene] (P1). Polymerization of 1Br was carried out according to the reported method.^{2,3} 1Br (2.00 g, 5.95 mmol) and MgBr₂•Et₂O (1.84 g, 7.14 mmol) weighed under nitrogen were placed in a two-necked flask equipped with a magnetic stirrer bar and a three-way stopcock under dry nitrogen, and THF (38 mL) was then added by a syringe. The mixture was cooled to -98 °C, and to this suspension solution was added lithium diisopropylamide solution (6.8 mmol based on *n*-BuLi) in THF (12 mL) via a cannula under dry nitrogen -98 °C. After stirring for 10 min at that temperature, [1,3-bis(diphenylphosphino)propane]nickel(II) chloride (Ni(dppp)Cl₂) (32 mg, 68 µmol) was added, and the mixture was allowed to warm to -60 °C over 15 min. After the further addition of Ni(dppp)Cl₂ (32 mg, 68 μ mol), the mixture was gradually allowed to warm to room temperature, whereupon the solution became clear within 1 h. Ni(dppp)Cl₂ (32 mg, 68 μ mol) was added to the mixture at room temperature. After stirring for 2.5 h, the solution was refluxed under stirring for 20 h, giving the purple suspension. After cooling to room temperature, the solvent was removed by evaporation and the dark purple suspension was poured into a large amount of CH₃OH. The purple precipitate was collected by centrifugation, washed with CH₃OH and THF, and dried in vacuo at room temperature overnight. The polymer was then dissolved in CHCl₃ and the solution was washed with saturated aqueous EDTA and dried over anhydrous Na₂SO₄. After the solvent was evaporated, the polymer was again dissolved in CHCl₃ and the solution was poured in a large amount of n-hexane. The precipitate was collected by centrifugation and dried in vacuo at room temperature overnight to afford **P1** as a dark purple powder (0.37 g, 1.46 mmol, 24.5% yield). $M_{\rm n} = 5.8 \times 10^3$, $DP_{\rm n} = 23$, $M_{\rm w}/M_{\rm n} = 1.27$. IR (KBr, cm⁻¹): 3063, 2964, 2927, 2893, 1650, 1609, 1352, 1314, 1300, 1189, 1063, 1018, 967, 830, 689. ¹H NMR (CDCl₃): δ 1.32–1.44 (br, CH₃, 6H), 4.04–4.18 (br, OCH₂, 2H), 6.85–6.93 (br, Th-H₄, 1H), 7.31–7.57 (d, Ph-H₂, 2H), 7.86–8.01 (d, Ph-H₃, 2H). ¹³C NMR (CDCl₃): δ 28.39, 67.59, 79.10, 127.37, 128.42, 129.15, 129.89, 131.55, 134.08, 138.21, 138.60, 161.77. Elemental Anal. Calcd for (C₁₅H₁₃NOS•0.3H₂O)_n: C, 69.10; H, 5.26; N, 5.37. Found: C, 69.00; H, 5.23; N, 5.12.

1-4. Measurements

Absorption and CD measurements of P1 in the presence of various chiral amines and metal salts. A typical experimental procedure is described below. Stock solutions of P1 (2.6 mg/5 mL, 2 mM) and (1*R*,2*S*)-1-amino-2-indanol [(1*R*,2*S*)-2] (3.0 mg/5 mL, 4 mM) in CHCl₃ and silver trifluoromethanesulfonate (AgOTf) (10.3 mg/1 mL, 40 mM) in CH₃CN were prepared in a dry box (NX1-M00320, Vacuum Atmosphere Company, Hawthorne, USA) under an argon atmosphere. The stock solutions of P1 (400 μ L) and (1*R*,2*S*)-2 (200 μ L) were transferred to a 0.2-cm quartz cell with a stopcock using Hamilton microsyringes. To this was added CHCl₃ (180 μ L) and the stock solution of AgOTf (20 μ L) subsequently. After the mixture was allowed to stand for 30 min, the absorption and CD spectra were recorded. In a similar manner, the absorption and CD spectra of P1 in the presence of other chiral amines and metal salts were performed.

¹H NMR and DOSY measurements of P1 in the presence of (1R,2S)-2 and AgOTf. Stock solutions of P1 (3.4 mg/3 mL, 4.4 mM) and (1R,2S)-2 (2.0 mg/3 mL, 4.4 mM) in CDCl₃, and AgOTf (10.3 mg/1 mL, 40 mM) in CD₃CN were prepared in a dry box under an argon atmosphere. The stock solutions of P1 (270 μ L), (1R,2S)-2 (270 μ L), and AgOTf (30 μ L) were transferred to a 5-mm NMR tube using microsyringes. After the mixture was allowed to stand for 30 min, the ¹H NMR and DOSY measurements were performed.

Absorption and CD titrations of P1 with AgOTf-(1*R*,2*S*)-2 mixture. Stock solutions solution of P1 (2.7 mg/10 mL, 1.05 mM) and (1*R*,2*S*)-2 (11.9 mg/1 mL, 80 mM) in CHCl₃ and AgOTf (20.6 mg/1 mL, 80 mM) in CH₃CN were prepared in a dry box under an argon atmosphere. The stock solutions of (1*R*,2*S*)-2 (500 μ L) and AgOTf (500 μ L) were mixed in a vessel equipped with a stopcock using microsyringes to prepare a 1:1 mixture solution of AgOTf and (1*R*,2*S*)-2. The stock solution of P1 (760 μ L) was transferred to a 0.2-mm quartz cell, and its initial absorption spectrum was recorded. To this was added increasing volumes of the AgOTf-(1*R*,2*S*)-2 solution (0–40 μ L, 0–2 equiv.), and their absorption and CD spectra were recorded for each addition after the mixtures had been allowed to stand for 30 min.

Absorption and CD measurements of P1–PEOPT mixtures in the presence of metal salts. A typical experimental procedure is described below. Stock solutions of P1 (2.6 mg/50 mL, 0.20 mM) and PEOPT (2.6 mg/50 mL, 0.20 mM) in CHCl₃ and copper(II) trifluoromethanesulfonate (Cu(OTf)₂) (14.5 mg/2 mL, 20 mM) in CH₃CN were prepared in a dry box under an argon atmosphere. The stock solutions of P1 (0–2 mL) and PEOPT (0–2 mL) were transferred to a 0.5-cm quartz cell with a stopcock using Hamilton microsyringes, keeping the total volume to be 2 mL. To this was added the stock solution of Cu(OTf)₂ (20 μ L). After the mixtures were allowed to stand for 3 h, the absorption and CD spectra were

recorded (Fig. 2a). In a similar manner, the absorption and CD spectra of **P1**–PEOPT mixtures in the presence of other metal salts were performed.

Absorption and CD measurements of PHT–PEOPT mixtures in poor solvents. A typical experimental procedure is described below. Stock solutions of PHT (1.7 mg/10 mL, 1.0 mM) and PEOPT (2.6 mg/10 mL, 1.0 mM) in CHCl₃ were prepared. The stock solutions of PHT (0–0.4 mL) and PEOPT (0–0.4 mL) were transferred to a 0.5-cm quartz cell with a stopcock using Hamilton microsyringes, keeping the total volume to be 0.4 mL. To this was added CH₃CN (1.6 mL). After the mixtures were allowed to stand for 15 min, the absorption and CD spectra were recorded (Fig. S8). In a similar manner, the absorption and CD spectra of PHT–PEOPT mixtures in CHCl₃/CH₃OH (20/80, vol/vol) were performed.

Molecular Modeling and Calculations. Molecular modeling and molecular mechanics calculations were performed using the Dreiding Force Field as implemented in the Materials Studio software (version 3.0; Accerlys Inc.). The polymer model (20 repeating monomer units) of **P1** was constructed with Polymer Builder in the Materials Studio. Charges on the atoms of **P1** were calculated by using the charge equilibration (QEq) in the Materials Studio; the total charge of the molecule was set to zero. The starting main-chain conformation of a polymer model was defined as a rotational conformation of a single bond between neighboring thiophene rings. The initial dihedral angles of a single bond from planarity were set to 120 (*s*-*trans*) and 20° (*s*-*cis*) to give polymer models with a twisted helical conformation. The energy minimization was accomplished by the conjugate gradient method and further continued until the root-mean-square (rms) value became less than 0.1 kcal mol⁻¹ Å⁻¹.

2. Chiral aggregate formation of P1 with chiral metal complexes2-1. Absorption titration of P1 with Ag(I)



Fig. S1. Absorption spectral changes of **P1** in CHCl₃ upon the addition of AgOTf in CH₃CN (CHCl₃/CH₃CN = 100/0 to 97.5/2.5, vol/vol) at 25 °C; [**P1**] = 1 mM unit⁻¹, [AgOTf]/[**P1**] = 0-1. The spectra were measured after the mixtures had been allowed to stand for 30 min at rt.

2-2. Time dependent CD intensity changes of P1 with chiral Ag(I) complex



Fig. S2. Time dependent changes of the molar circular dichroism ($\Delta \varepsilon_{435}$) and molar absorptivity (ε_{435}) at 435 nm of **P1** upon the addition of AgOTf-(1*R*,2*S*)-**2** complex in CHCl₃/CH₃CN (97.5/2.5, vol/vol) at 25 °C; [**P1**] = 1 mM unit⁻¹, [**P1**]:[AgOTf]:[(1*R*,2*S*)-**2**] = 1:1:1.



2-3. CD and absorption titrations of P1 with chiral Ag(I) complex

Fig. S3. CD and absorption spectra of **P1** in the presence of AgOTf–(1R,2S)-2 complex in CHCl₃/CH₃CN (97.5/2.5, vol/vol) at 25 °C; [**P1**] = 1 mM unit⁻¹, [AgOTf–(1R,2S)-2]/[**P1**] = 0–0.5 (a), 0.5–1 (b), and 1–2 (c). These spectra were measured after the mixtures had been allowed to stand for 30 min at rt. (d) Plots of the molar circular dichroism at 435 nm ($\Delta \varepsilon_{435}$) and the molar absorptivities at 320 nm (ε_{320}) of **P1** upon the addition of AgOTf–(1R,2S)-2 complex.

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2-4. ¹H NMR spectra of P1 with chiral Ag(I) complexes



Fig. S4. ¹H NMR spectra of **P1**, (1*R*,2*S*)-**2**, and their complexes in CDCl₃/CD₃CN (10/1, vol/vol) at 25 °C; [**P1**] = 2 mM unit⁻¹.

2-5. DOSY measurements

			$D / 10^{-10} \text{ m}^2 \text{ s}^{-1}$	
sample	conc. / mM	P1	(1 <i>R</i> ,2 <i>S</i>)- 2	TMS ^b
P1	2	5.25		21.5
(1 <i>R</i> ,2 <i>S</i>)- 2	2		13.8	20.2
P1/Ag ^c	2/1	2.09		19.3
Ag/(1 <i>R</i> ,2 <i>S</i>)- 2	1/2		9.09	21.0
P1 /Ag/(1 <i>R</i> ,2 <i>S</i>)- 2	2/2/2	2.50	7.43	19.4

Table S1. Diffusion coefficients (D) of P1, (1R,2S)-2, and their AgOTf complexes^a

^aMeasured in CDCl₃/CD₃CN (10/1, vol/vol) at 25 °C. ^bTMS was used as the internal standard. ^cAfter the measurement (*ca*. 4 h), a small amount of an orange-red precipitate derived from the **P1**–AgOTf complex was observed in the NMR tube.



2-6. Temperature effect

Fig. S5. (a) Temperature dependent absorption and CD spectral changes of P1–AgOTf–(1*R*,2*S*)-2 complex. (b) Plots of the molar circular dichroism at 430 nm ($\Delta \varepsilon_{430}$) and molar absorptivity at 400 nm (ε_{400}) of P1–AgOTf–(1*R*,2*S*)-2 complex in CHCl₃/CH₃CN (97.5/2.5, vol/vol); [P1] = 1 mM unit⁻¹, [P1]:[AgOTf]:[(1*R*,2*S*)-2] = 1:1:1. These spectra were measured after the mixtures had been allowed to stand for 30 min at rt.

2-7. Effect of chiral amines

		CD spectra (2nd Cotton)		absorption spectra		
			$\Delta arepsilon$	λ	ε x 10 ⁻³	λ
amine	configuration ^b	sign	$/ \text{ cm}^{-1} \text{ M}^{-1} \text{ unit}^{-1}$	/ nm	$/ \text{ cm}^{-1} \text{ M}^{-1} \text{ unit}^{-1}$	/ nm
none					6.98	449
(<u>1S</u> ,2R)- 2	S	+	2.66	438	6.98	432
(<u>1R</u> ,2S)- 2	R	_	2.71	434	6.83	430
(1 <i>R</i> , <u>2<i>S</i></u>)- 3	S	+	2.28	441	7.09	441
(1 <i>R</i> , <u>2<i>S</i></u>)- 4	S	+	1.94	444	6.90	443
(<i>S</i>)- 5	S	+	0.72	453	7.19	436
(<i>S</i>)- 6	S	_	0.91	438	7.07	441
(1 <i>R</i> , <u>2</u> <i>S</i>)-7	S	+	0.62	435	6.92	448

Table S2. Molar circular dichroisms ($\Delta \varepsilon_{max}$) and molar absorptivities (ε_{max}) of **P1** in the presence of AgOTf–chiral amine complexes^a

^aSolvent: CHCl₃:CH₃CN = 97.5:2.5; temp: 25 °C; cell length: 2 mm; [**P1**] = 1 mM unit⁻¹ [**P1**]:[AgOTf]:[chiral amine] = 1:1:1. The spectra were measured after the mixtures had been allowed to stand for 30 min at rt. ^bAbsolute configuration of the amino group-bound asymmetric carbon atom.

2-8. Effect of metal salts



Fig. S6. CD and absorption spectra of **P1** in the presence of metal salt–(1R,2S)-**2** (a) and –(1R,2S)-**3** (b) complexes in CHCl₃/CH₃CN (97.5/2.5, vol/vol) at 25 °C; [**P1**] = 1 mM unit⁻¹, [**P1**]:[metal salt]:[(1R,2S)-**2** or (1R,2S)-**3**] = 1:1:1. These spectra were measured after the mixtures had been allowed to stand for 30 min at rt.

	CD spectra			absorption spectra		
		$\Delta arepsilon$	λ	$\varepsilon \ge 10^{-3}$	λ	
metal salt	sign	$/ \text{ cm}^{-1} \text{ M}^{-1} \text{ unit}^{-1}$	/ nm	$/ \text{ cm}^{-1} \text{ M}^{-1} \text{ unit}^{-1}$	/ nm	
$Fe(ClO_4)_2$		n.d. ^b		7.78	438	
$Co(ClO_4)_2$		n.d. ^b		7.15	446	
$Ni(ClO_4)_2$		n.d. ^b		6.68	448	
$Cu(ClO_4)_2$	+	1.06	466	6.37	428	
Cu(OTf) ₂	+	2.89	471	6.52	439	
Zn(OTf) ₂		n.d. ^b		6.62	422	
CuOTf		n.d. ^b		6.54	447	
AgOTf	_	2.71	434	6.83	430	

Table S3. Molar circular dichroisms ($\Delta \varepsilon_{max}$) and molar absorptivities (ε_{max}) of **P1** in the presence of metal salt–(1*R*,2*S*)-2 complexes^a

^aSolvent: CHCl₃:CH₃CN = 97.5:2.5; temp: 25 °C; cell length: 2 mm; [P1] = 1 mM unit⁻¹, [P1]:[metal salt]:[(1*R*,2*S*)-2] = 1:1:1. The spectra were measured after the mixtures had been allowed to stand for 30 min at rt. ^bNot detected.



2-9. Chiral Co-aggregate Formation of P1–PEOPT Mixture with Cu(II) and Ag(I)

Fig. S7. (a, d) CD and absorption spectra of **P1**–PEOPT mixtures with Cu(OTf)₂ (a) and AgOTf (d) in CHCl₃/CH₃CN (99/1, vol/vol) at 25 °C; [**P1**]+[PEOPT] = 0.2 mM unit⁻¹, ([**P1**]+[PEOPT]):[Cu(OTf)₂ or AgOTf] = 1:1. These spectra were measured after the mixtures had been allowed to stand for 3 h at rt. (b, c, e) Time-dependent CD intensity changes ($\Delta \varepsilon_{2nd}$) of **P1**–PEOPT mixtures with Cu(OTf)₂ (b, c) and AgOTf (e) in CHCl₃/CH₃CN (99/1, vol/vol); [PEOPT]/([**P1**]+[PEOPT]) = 0–0.55 (b), 0.55–1 (c), and 0–1 (e).



2-10. Chiral Co-aggregate Formation of PHT-PEOPT Mixture in Poor Solvent

Fig. S8. (a, b) CD and absorption spectra of PHT–PEOPT mixtures in CHCl₃/CH₃OH (20/80, vol/vol) (a) and CHCl₃/CH₃CN (20/80, vol/vol) (b) at 25 °C; [PHT]+[PEOPT] = 0.2 mM unit⁻¹. These spectra were measured after the mixtures had been allowed to stand for 15 min at rt. (c) Plots of the molar circular dichroism (1st Cotton effect) ($\Delta \varepsilon_{1st}$) of PHT–PEOPT mixtures in CHCl₃/CH₃OH (20/80, vol/vol) (blue) and CHCl₃/CH₃CN (20/80, vol/vol) (red) versus PEOPT content; [**P1**]+[PEOPT] = 0.2 mM unit⁻¹. These spectra were measured after the mixtures had been allowed to stand for 15 min at rt.

3. Supporting references

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