

Electronic Supplementary Materials

**Practical Biocatalytic Desymmetrization of Meso *N*-Heterocyclic
Dicarboxamides and Their Application in the Construction of
Aza-sugar Containing Nucleoside Analogs**

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Table of Contents

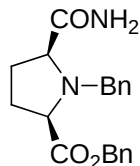
1. General information
2. General procedure for biocatalytic desymmetrization
3. Synthesis of aza-sugar containing nucleoside analogs
4. HPLC analysis of products
5. ¹H and ¹³C NMR spectra of all products

1. General Information

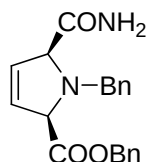
¹H and ¹³C NMR spectra were recorded on 300MHz and 400MHz spectrometers. Chemical shifts are reported in ppm versus tetramethylsilane or the residual solvent resonance used as an internal standard. Melting points are uncorrected. Enantiomeric excess (ee) values of all compounds were obtained from HPLC analyses using chiral stationary phases.

2. General Procedure for Biocatalytic Desymmetrization

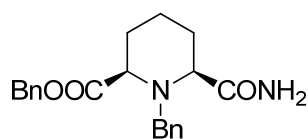
To an Erlenmeyer flask (150 mL) with a screw cap were added aqueous potassium phosphate buffer (0.1 M, pH 7.0, 250 mL) and *Rhodococcus erythropolis* AJ270 cells (2 g wet weight). The resting cells were activated at 30 °C for 0.5 h with orbital shaking. Dicarboxamide substrates **1** (1 mmol) was added in one portion to the flask, and the resulting mixture was incubated at 30 °C using an orbital shaker (200 rpm). The reaction, monitored TLC, was quenched after a period of time (see Table 1 and Scheme 1) by removing the biomass by filtration through a Celite pad, and the filtrate was freeze-dried. The residue was mixed with DMF (2 mL), BnBr (1 mL) and K₂CO₃ (1.38g, 10 mmol). After stirring for 24 h, water (10 mL) was added and the resulting mixture was extracted with ethyl acetate (3 × 100 mL), washed with brine (3 × 2 mL) and dried with anhydrous Na₂SO₄. After removal of solvent, the residue was chromatographed using a silica gel column with ethyl acetate as eluent to give pure products **2**.



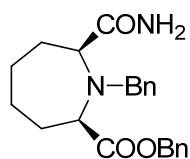
Benzyl N-benzyl-5-carbamoylpyrrolidine-2-carboxylate (2a): white solid; mp 58 °C; $[\alpha]_D^{25} +19.2^\circ$ (c 1, CH₂Cl₂); ee >99.5% (HPLC analysis using a Diacel-ADH column); IR (KBr) ν 3422, 3369, 1740, 1663 cm⁻¹; ¹H NMR (300 MHz/CDCl₃) δ 7.89 (br, s, 1H), 7.37-7.22 (m, 10H), 5.90 (br, s, 1H), 4.98 (s, 2H), 3.88 (d, $J = 13.2$ Hz, 1H), 3.79 (d, $J = 13.2$ Hz, 1H), 3.63 (t, $J = 7.4$ Hz, 1H), 3.54-3.50 (m, 1H), 2.19-1.88 (m, 4H); ¹³C NMR (75MHz/CDCl₃) δ 177.1, 174.1, 136.4, 134.7, 128.5, 127.8, 127.6, 127.56, 127.3, 126.8, 67.1, 65.9, 65.5, 58.2, 29.8, 29.5; MS (ESI) m/z 339.1 (M+1, 86), 361.2 (M+23, 100%). Anal. Calcd. for C₂₀H₂₂N₂O₃: C, 70.99; H, 6.55; N, 8.28. Found: C, 70.65; H, 6.71; N, 8.53.



Benzyl N-benzyl-5-carbamoyl-2,5-dihydro-1H-pyrrole-2-carboxylate (2b): mp 127-128 °C; $[\alpha]_D^{25}$ -36.4° (c 1, CH₂Cl₂); ee >99.5% (HPLC analysis using a Diacel-ADH column); IR (KBr) ν 3361, 3344, 3223, 2527, 1707, 1682, 1627, 1177 cm⁻¹; ¹H NMR (300 MHz/Acetone-*d*₆) δ 7.88 (br, s, 1H), 7.41-7.26 (m, 10H), 6.43 (br, s, 1H), 5.95-5.91 (m, 1H), 5.84-5.82 (m, 1H), 5.07 (d, *J* = 12.3 Hz, 1H), 5.04 (d, *J* = 12.3 Hz, 1H), 4.63-4.61 (m, 1H), 4.27-4.25 (m, 1H), 4.05 (d, *J* = 13.5 Hz, 1H), 3.97 (d, *J* = 13.5 Hz, 1H); ¹³C NMR (75 MHz/Acetone-*d*₆) δ 173.9, 172.7, 137.9, 136.0, 130.9, 129.2, 128.5, 128.3, 128.1, 128.0, 127.3, 126.6, 76.2, 73.6, 66.4, 58.4; MS (ESI) *m/z* 338.3 (M+D, 90), 360.3 (M(D)+23, 100%). Anal. Calcd. for C₂₀H₂₀N₂O₃: C, 71.41; H, 5.99; N, 8.33. Found: C, 71.15; H, 5.98; N, 8.17.



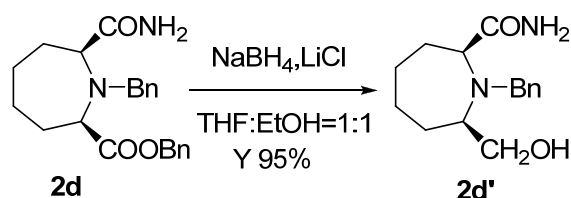
Benzyl N-benzyl-6-carbamoylpiperidine-2-carboxylate (2c): mp 105 °C; $[\alpha]_D^{25}$ -25.6° (c 1, CH₂Cl₂); ee >99.5% (HPLC analysis using a Diacel-ADH column); IR (KBr) ν 3420, 1744, 1632 cm⁻¹; ¹H NMR (300 MHz/CDCl₃) δ 7.36-7.17 (m, 11 H), 5.71 (br, s, 0.5 H), 5.64 (br, s, 0.5H), 5.07 (s, 2 H), 3.77 (s, 2 H), 3.23 (dd, 1H, *J* = 10.3, 3.7 Hz), 3.09 (dd, 1H, *J* = 9.4, 3.7 Hz), 2.00-1.84 (m, 2H), 1.73-1.63 (m, 3H), 1.26-1.17 (m, 2H); ¹³C NMR (75 MHz/CDCl₃) δ 177.3, 174.0, 135.6, 134.9, 130.3, 128.6, 128.5, 128.4, 128.2, 127.6, 66.7, 63.3, 61.6, 58.3, 28.2, 28.1, 20.7; MS (ESI) *m/z* 353.2 (M+1, 100%), 375.2 (M+23, 17%). Anal. Calcd. for C₂₁H₂₄N₂O₃: C, 71.57; H, 6.86; N, 7.95. Found: C, 71.58; H, 6.84; N, 7.96.



Benzyl 1-benzyl-7-carbamoylazepane-2-carboxylate (2d): oil; $[\alpha]_D^{25}$ -25.6° (c 1,

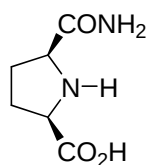
CH₂Cl₂); IR (KBr) ν 3410, 1737, 1670 cm⁻¹; ¹H NMR (300 MHz/CDCl₃) δ 8.42 (br, s, 1 H), 7.38-7.23 (m, 10 H), 5.88 (br, s, 1 H), 5.15 (d, 1 H, J = 12.1 Hz), 5.07 (d, 1 H, J = 12.1 Hz), 3.75 (d, 1 H, J = 8.7 Hz), 3.72 (d, 1 H, J = 8.7 Hz), 3.62 (dd, 1 H, J = 8.7, 1.8 Hz), 3.42 (d, 1 H, J = 8.1 Hz), 2.02-1.46 (m, 8 H); ¹³C NMR (75 MHz/CDCl₃) δ 178.4, 176.3, 138.0, 135.5, 129.0, 128.7, 128.54, 128.50, 128.3, 127.6, 67.0, 66.3, 60.6, 29.1, 28.2, 28.1, 26.8; MS (ESI) m/z 367.2 (M+1, 20%), 389.3 (M+23,100). Anal. Calcd. for C₂₂H₂₆N₂O₃: C, 72.11; H, 7.15; N, 7.64. Found: C, 72.07; H, 7.10; N, 7.79.

To determine the ee value of the product, **2d** was converted into **2d'**.



Preparation of 2d'. To a solution of **2d** (70 mg, 0.2 mmol) in a mixture of EtOH (5 mL) and THF (5 mL) was added consecutively NaBH₄ (15.2 mg, 0.4 mmol) and LiCl (15.2 mg, 0.4 mmol). After the resulting mixture was stirred for 3 h at ambient temperature, water (20 mL) was added and the resulting mixture was extracted with ethyl acetate (3 × 15 mL). Combined organic phase was washed with brine (3 × 5 mL) and dried with anhydrous Na₂SO₄. After removal of solvent, the residue was chromatographed using a silica gel column with ethyl acetate as an eluent to give pure products **2d'** (50 mg, 95%): mp 146 °C; [α]_D²⁵ +22.9° (c 0.35, CH₂Cl₂); ee 63% (HPLC analysis using a Diacel-OJH column); IR (KBr) ν 3364, 3274, 2943, 2919, 1664 cm⁻¹; ¹H NMR (300 MHz/ DMSO-*d*₆) δ 7.86 (br, s, 0.5 H), 7.85 (br, s, 0.5 H), 7.38-7.21 (m, 5 H), 7.02 (br, s, 1 H), 7.02 (br, s, 1 H), 4.98 (t, 1 H, J = 4.8 Hz), 3.81 (d, 1 H, J = 14.4 Hz), 3.67 (d, 1 H, J = 14.4 Hz), 3.53-3.47 (m, 2H), 3.17 (dd, 1 H, J = 15.3, 2.4 Hz), 2.67 (t, 1 H, J = 4.2 Hz), 1.98-1.91 (m, 1 H), 1.72-1.46 (m, 6 H), 1.34-1.17 (m, 1 H); ¹³C NMR (75 MHz/ DMSO-*d*₆) δ 176.4, 140.5, 128.4, 128.1, 126.8, 66.6, 65.0, 63.2, 58.0, 29.2, 28.0, 26.3, 25.2; MS (ESI) m/z 263.4 (M+1, 75%), 285.3(M+23,100). Anal. Calcd. for C₁₅H₂₂N₂O₂: C, 68.67; H, 8.45; N, 10.68. Found: C, 68.60; H, 8.38; N, 10.72.

3. Procedure for large scale biocatalytic desymmetric hydrolysis of 1a. To an Erlenmeyer flask (500 mL) with a screw cap *Rhodococcus erythropolis* AJ270 cells (2 g wet weight) were suspended in aqueous potassium phosphate buffer (0.1 M, pH 7.0, 250 mL), and activated at 30 °C for 0.5 hour with orbital shaking. Dicarboxamide **1a** (20 g) was added in one portion to the flask, and the resulting mixture was incubated at 30 °C using an orbital shaker (200 rpm). The reaction, monitored by HPLC, was quenched after 12 h by removing the biomass by filtration through a Celite pad. The filtrate was concentrated to 100 mL using a rotary evaporator under reduced pressure, and product **3** (8.0 g) precipitated from solution. The mother liquid was subjected to an ion-exchange column and eluted with aqueous ammonia solution (1%) to give another portion of product **3** (11.0 g).

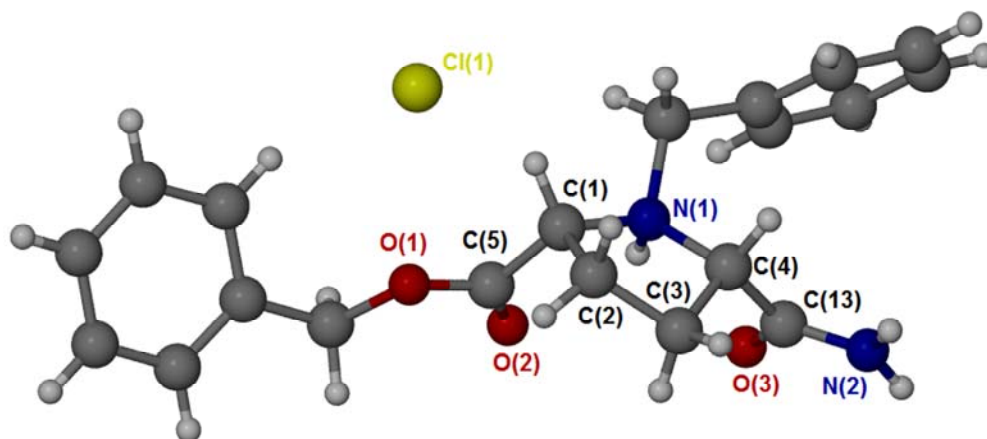


Product **3** (19.0 g, 94%): mp 234-233 °C; $[\alpha]_D^{25}$ -9.6° (c 1, H₂O); IR(KBr) ν 3308, 3153, 2361, 1696, 1642, 1576 cm⁻¹; ¹H NMR (D₂O/300MHz) 4.34 (t, *J* = 7.2 Hz, 1 H), 4.11 (t, *J* = 6.3 Hz, 1 H), 2.39-2.25 (m, 2 H), 2.05-1.91 (m, 2 H); ¹³C NMR (75MHz /D₂O) 173.8, 171.4, 61.8, 60.31, 29.8, 29.2; MS (ESI) *m/z* 159 (*M*+1, 7), 181 (*M*+23, 100%). Anal. Calcd. for C₆H₁₀N₂O₃: C, 45.57; H, 6.37; N, 17.71. Found: C, 45.54; H, 6.53; N, 17.78.

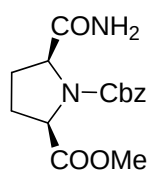
To determine the enantiomeric excess (ee) value and the absolute configuration of product **3**, transformation of **3** into **2a** was conducted. Thus, a mixture of **3** (158 mg), K₂CO₃ (1.38 g), benzyl bromide (1 mL) in dry DMF (2 mL) was stirred at ambient temperature for 24 h. Water (10 mL) was then added, and the mixture was extracted with ethyl acetate (3×10 mL). The combined organic layer was washed with brine (3×10 mL), and dried with anhydrous Na₂SO₄. After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with ethyl acetate to give **2a** (320 mg, 95%). Slow vapor diffusion of diethyl ether into a mixture of **2a** (15.7

mg), concentrated hydrochloric acid (15 μ L) and ethanol (0.5 mL) at 5 $^{\circ}$ C for 2 days led to the formation of a single crystal of **2a**·HCl.

X-ray structure:

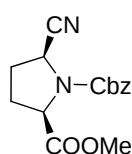


4. Synthesis of Aza sugar containing nucleoside Analogs



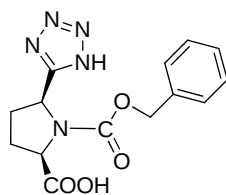
Preparation of 4. To a mixture of **3** (4.0 g, 25 mmol) in ethanol (20 mL) and saturated NaHCO_3 aqueous solution (20 mL) was added drop-wise CbzCl (6 mL) at room temperature while stirring. The resulting mixture was kept stirring overnight. The reaction was quenched by adding hydrochloric acid (2 N) until the pH was adjusted to 7. Under reduced pressure, the solvent was completely removed. The residue was mixed with methanol (100 mL) and SOCl_2 (4 mL) at ambient temperature. After stirring for another 4 h at ambient temperature, saturated NaHCO_3 aqueous solution (200 mL) was added. The mixture was extracted with ethyl acetate (3×100 mL). The combined organic layer was washed with brine (3×20 mL) and dried with anhydrous Na_2SO_4 . After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with ethyl acetate to give product **4** (6.9 g, 91%): mp 104 $^{\circ}$ C; $[\alpha]_D^{25} +46.6^{\circ}$ (c 0.6, CHCl_3); IR (KBr) ν 3435, 3401, 1734,

1700, 1683 cm^{-1} ; ^1H NMR (300 MHz/DMSO- d_6 , 375 K) δ 7.35-7.25 (m, 5H), 6.97 (br, s, 2H), 5.09 (d, J = 12.6 Hz, 1H), 5.05 (d, J = 12.9 Hz, 1H), 4.42 (t, J = 7.2 Hz, 1H), 4.20-4.16 (m, 1 H), 3.66 (s, 3 H), 2.31-2.17 (m, 2 H), 1.98-1.87 (m, 2 H); ^{13}C NMR (75 MHz/DMSO- d_6) δ 175.0, 174.8, 174.3, 174.0, 154.4, 154.1, 136.9, 136.7, 128.8, 128.4, 128.3, 127.7, 127.6, 67.2, 67.1, 62.5, 62.1, 60.5, 60.1, 53.1, 53.0, 30.6, 29.7, 29.6, 28.8; MS (ESI) m/z 307 ($M+1$, 20), 329 ($M+23$, 100%). Anal. Calcd. for $\text{C}_{15}\text{H}_{18}\text{N}_2\text{O}_5$: C, 58.82; H, 5.92; N, 9.15. Found: C, 58.87; H, 5.99; N, 9.10.

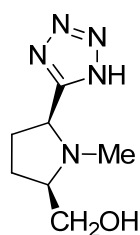


Synthesis of 5.

To a solution of **4** (3.06 g, 10 mmol) in dry DMF (4 mL) was added SOCl_2 (1 mL) at 0 °C. After stirring for another 1 h at room temperature, the reaction was quenched by the addition of saturated NaHCO_3 aqueous solution (10 mL). The mixture was extracted with ethyl acetate (3×20 mL). The combined organic layer was washed with brine (3×4mL) and dried with anhydrous Na_2SO_4 . After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with a mixture of ethyl acetate and petroleum ether (1:2) to give compound **5** (2.6 g, 90%): oil; $[\alpha]_D^{25}$ -14° (c 1, CHCl_3); IR (KBr) ν 1746, 1715 cm^{-1} ; ^1H NMR (300MHz/ CDCl_3) δ 7.40-7.33 (m, 5H), 5.28-5.09 (m, 2H), 4.72-4.63 (m, 1H), 4.47-4.37 (m, 1H), 3.79 (s, 1.5H), 3.64 (s, 1.5H), 2.44-2.21 (m, 4H); ^1H NMR (300 MHz/DMSO- d_6 , 375K) δ 7.39-7.28 (m, 5H), 5.18-5.06 (m, 2H), 4.84-4.80 (m, 1H), 4.42-4.37 (m, 1H), 3.64 (s, 3H), 2.42-2.30 (m, 2H), 2.24-2.12 (m, 1H), 2.10-2.02 (m, 1H); ^{13}C NMR (75 MHz/DMSO- d_6) δ 171.6, 171.4, 153.3, 135.6, 128.6, 128.4, 128.2, 128.1, 128.0, 127.8, 118.0, 117.8, 68.3, 68.0, 67.5, 59.6, 59.3, 52.7, 52.5, 48.1, 47.5, 30.6, 29.8, 29.7, 28.7; MS (ESI) m/z 311 ($M+23$, 100%). Anal. Calcd. for $\text{C}_{15}\text{H}_{16}\text{N}_2\text{O}_4$: C, 62.49; H, 5.59; N, 9.72. Found: C, 62.70; H, 5.62; N, 9.57.

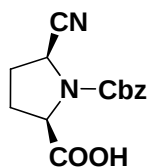


Synthesis of 6. A mixture of **5** (576 mg, 2 mmol), NaN₃ (260 mg, 4 mmol), ZnBr₂ (225 mg, 1 mmol) in water (6 mL) and 2-propanol (3 mL) was refluxed for 16 h. After cooling down to room temperature, hydrochloric acid (1 N, 10 mL) was added. The mixture was extracted with ethyl acetate (3×20 mL). The combined organic layer was washed with brine (3×4 mL) and dried with anhydrous Na₂SO₄. After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with a mixture of ethyl acetate and methanol (20:1) to give compound **6** (367 mg, 58%): mp 183-185 °C; oil; [α]_D²⁵ -22.0° (c 0.5, CHCl₃); IR (KBr) ν 2960, 1707, 1451, 1422 cm⁻¹; ¹H NMR (300 MHz/DMSO-*d*₆, 373K) δ 7.31-7.23 (m, 5H), 5.33-5.31 (m, 1H), 5.05 (s, 2H), 4.41 (t, 1H, *J* = 8.0 Hz), 2.46-2.35 (m, 2H), 2.19-2.05 (m, 2H); ¹³C NMR (75 MHz/ DMSO-*d*₆) δ 177.53, 177.05, 157.7, 157.3, 152.8, 134.4, 134.1, 127.6, 127.4, 126.9, 67.2, 67.1, 58.97, 58.5, 52.5, 52.1, 31.6, 30.5, 29.9, 28.7, 27.8; MS (ESI) *m/z* 340 (M⁺+23, 100%). Anal. Calcd. for C₁₄H₁₆N₅O₄ [M+H]: 318.1202. Found: 318.1192.



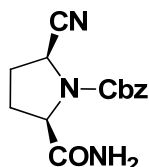
Synthesis of 7. Under protection of argon, a mixture of **6** (317 mg, 1 mmol) and LiAlH₄ (380 mg, 10 mmol) in dry THF (10 mL) was refluxed for 24 h. While the mixture was kept at -20°C, NaOH aqueous solution (2 N, 1 mL) was slowly injected through a syringe. The resulting mixture was filtrate through a Celite pad and washed thoroughly with a mixture of CH₂Cl₂ and MeOH (20:1). After removal of solvent, the

residue was mixed with hydrochloric acid (1 N) with pH value of the mixture being adjusted to pH 2. The resulting mixture was subjected to an ion-exchange column (nk 001×7) and then eluted with pure water. After the pH of the fraction was around 7, the column was eluted with aqueous ammonia solution (1%). After removal of solvent, product **7** was obtained. **7** (152 mg, 83%): mp 185 °C; $[\alpha]_D^{25} +10.3^\circ$ (c 0.78, H₂O); IR (KBr) ν 3447, 2959, 2922, 1578, 1411, 1075, 1030 cm⁻¹; ¹H NMR (300 MHz/D₂O) δ 4.53 (t, *J* = 8.7 Hz, 1H), 3.77-3.64 (m, 2H), 3.54-3.45 (m, 1H), 2.72 (s, 3H), 2.38-2.26 (m, 2H), 2.23-2.14 (m, 1H), 1.94-1.83 (m, 1H); ¹³C NMR (75MHz/D₂O) δ 157.1, 70.3, 64.2, 59.6, 39.1, 27.7, 24.9; MS (ESI) *m/z* 184.1 (M+1, 15%), 206.1 (M+23, 100%). Anal. Calcd. for C₇H₁₄N₅O [M+H]: 184.1198. Found: 184.1193.

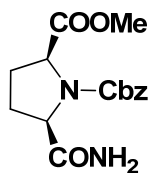


Preparation of 8. A mixture of **5** (100 mg, 0.33 mmol) and LiOH·H₂O (23mg, 0.55 mmol) in THF (3 mL) and water (3 mL) was stirring at ambient temperature for 2 h. The mixture was then treated with hydrochloric acid (1 N) until the pH of the mixture was adjusted to 2, and extracted with ethyl acetate (3×10 mL). The combined organic layer was washed with brine (3×3 mL) and dried with anhydrous Na₂SO₄. After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with a mixture of petroleum ether, ethyl acetate and acetic acid (200:100:0.5). The collected fractions were concentrated to about 10 mL at 30-40 °C under reduced pressure using a rotary evaporator. The resulting solution was mixed with ethyl acetate (50 mL) and washed with brine (3×10 mL). After drying with anhydrous Na₂SO₄ and removing solvent, pure **8** was obtained. **8** (90 mg, 95%): oil; $[\alpha]_D^{25} -24.6^\circ$ (c 4.6, CHCl₃); IR (KBr) ν 3065, 3038, 2960, 1715, 1414, 1358 cm⁻¹; ¹H NMR (300MHz/DMSO-*d*₆, 375K) δ 7.36-7.28 (m, 5H), 5.14 (s, 2H), 4.82-4.78 (m, 1H), 4.34-4.30 (m, 1H), 2.42-2.29 (m, 2H), 2.24-2.11 (m, 1H), 2.09-2.00 (m, 1H); ¹³C

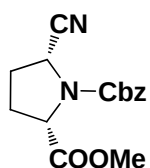
NMR (75 MHz/DMSO-*d*₆) δ 173.3, 173.0, 153.7, 153.4, 136.7, 128.8, 128.4, 128.3, 127.9, 127.7, 119.7, 119.4, 67.4, 67.3, 60.1, 59.6, 48.4, 47.8, 30.7, 29.9, 29.7, 28.9; MS (ESI) *m/z* 273.1 (M-1, 100%). Anal. Calcd. for C₁₄H₁₅N₂O₄ [M+H]: 275.1032. Found: 275.1022.



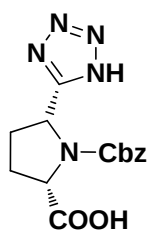
9: To a solution of **8** (66.1mg, 0.24 mmol) in CH₂Cl₂ (4 mL) was added consecutively 2-chloro-4,6-dimethoxy-1,3,5-triazine (CDMT, 50.5 mg, 0.29 mmol) and *N*-methylmorpholine (NMM, 83 μ L, 0.72 mmol). After stirring at room temperature for 2.5 h, an ammonia gas balloon (250 mL) was connected, and the reaction mixture was stirred for 1 h. After removal of the ammonia gas balloon, the mixture was kept stirring for another 1 h. The reaction was quenched by adding hydrochloric acid (1 N, 20 mL). After extraction with ethyl acetate (3 $\times$ 10 mL), the combined organic layer was dried with anhydrous Na₂SO₄. After removal of organic solvent, the residue was chromatographed on a silica gel column eluted with a mixture of petroleum ether and ethyl acetate (1:2) to afford **9** (44 mg, 67%): mp 44 °C; [α]_D²⁵ -7.0° (*c* 0.58, CHCl₃); IR (KBr) ν 3428, 3352, 3200, 1708, 1412, 1356, 1119 cm⁻¹; ¹H NMR (300 MHz/DMSO-*d*₆, 375K) δ 7.38-7.29 (m, 5H), 6.87 (s, br, 1H), 5.13 (d, *J* = 12.9 Hz, 1H), 5.11 (d, *J* = 12.9 Hz, 1H), 4.76 (dd, *J* = 7.4, 4.8 Hz, 1H), 4.29 (dd, *J* = 7.4, 5.1 Hz, 1H), 2.32-2.14 (m, 3H), 2.02-1.96 (s, br, 1H); ¹³C NMR (75MHz/DMSO-*d*₆) δ 173.0, 172.8, 153.9 153.5, 136.8, 128.8, 128.4, 128.2, 127.9, 127.6, 119.8, 119.5, 67.2, 67.1, 60.7, 60.2, 48.6, 48.0, 30.6, 29.7, 29.5; MS (ESI) *m/z* 296.2 (M+23, 100%). Anal. Calcd. for C₁₄H₁₆N₃O₃ [M+H]: 274.1192. Found: 274.1186. Anal. Calcd. for C₁₄H₁₅N₃NaO₃ [M+Na]: 296.1011. Found: 296.1004.



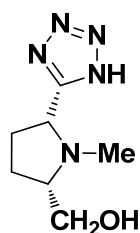
Synthesis of ent-4. To a solution of **9** (155mg, 0.57 mmol) in MeOH (4 mL) at -15 °C was bubbled with dry HCl (g) for 1 h. After the resulting solution was kept at -20 °C for 12 h, hydrochloric acid (6 N, 10 mL) was added. After extraction with ethyl acetate (3×100mL) and washing with brine (3×10 mL), the combined organic phase was dried with anhydrous Na₂SO₄. The solvent was removed and the residue was chromatographed on a silica gel column eluted with ethyl acetate to give ent-**4** (133 mg, 76%): $[\alpha]_D^{25}$ -47.3° (c 1.7, CHCl₃). Identical IR and ¹H NMR spectra as that of **4** were obtained.



Ent-**5** was synthesized from ent-**4** following the same procedure for the conversion of **4** to **5**. Ent-**5**: $[\alpha]_D^{25}$ +14.7° (c 3.0, CHCl₃). Identical IR and ¹H NMR spectra as that of **5** were obtained.



Ent-**6** was synthesized from ent-**5** following the same procedure for the transformation of **5** into **6**. Ent-**6**: $[\alpha]_D^{25}$ +24.0° (c 1.0, MeOH). Identical IR and ¹H NMR spectra as that of **6** were obtained.



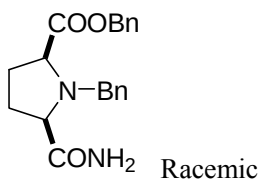
Ent-7 was synthesized from ent-6 following the same procedure for the transformation from 6 into 7. Ent-7: mp 187 °C; $[\alpha]_D^{25}$ -11.0° (c 2.0, H₂O). Following identical IR, ¹H and ¹³C NMR spectra as that of 7 were obtained. IR (KBr) ν 3447, 2958, 2921, 1577, 1409, 1077, 1025 cm⁻¹; ¹H NMR (300 MHz/D₂O) δ 4.53 (t, *J* = 8.7 Hz, 1H), 3.77-3.64 (m, 2H), 3.54-3.45 (m, 1H), 2.72 (s, 3H), 2.38-2.26 (m, 2H), 2.23-2.14 (m, 1H), 1.94-1.83 (m, 1H); ¹³C NMR (75 MHz/D₂O) δ 157.1, 70.3, 64.2, 59.6, 39.1, 27.7, 24.9; MS (ESI) *m/z* 184.1 (M+1, 100%). Anal. Calcd. for C₇H₁₄N₅O [M+H]: 184.1198. Found: 184.1190.

4. HPLC analysis of products.

A Shimadzu LC-10AVP HPLC system was used to analyze enantiomeric excess values of all products.

Table S1: HPLC analysis of amides 2a-d

Compound	Hexane 2-propanol	/ Temperature (°C)	Rate (mL/min)	<i>t</i> ₁ (min)	<i>t</i> ₂ (min)	Column
2a	9/1	15	0.5	41.9	49.1	Chiralcel ADH
2b	9/1	15	0.5	51.4	57.2	Chiralcel ADH
2c	9/1	15	0.5	26.4	27.9	Chiralcel ADH
2d	9/1	20	0.5	25.2	28.0	Chiralcel OJH



cShimadzu CLASS-VP V6.13 SP2

Area % Report

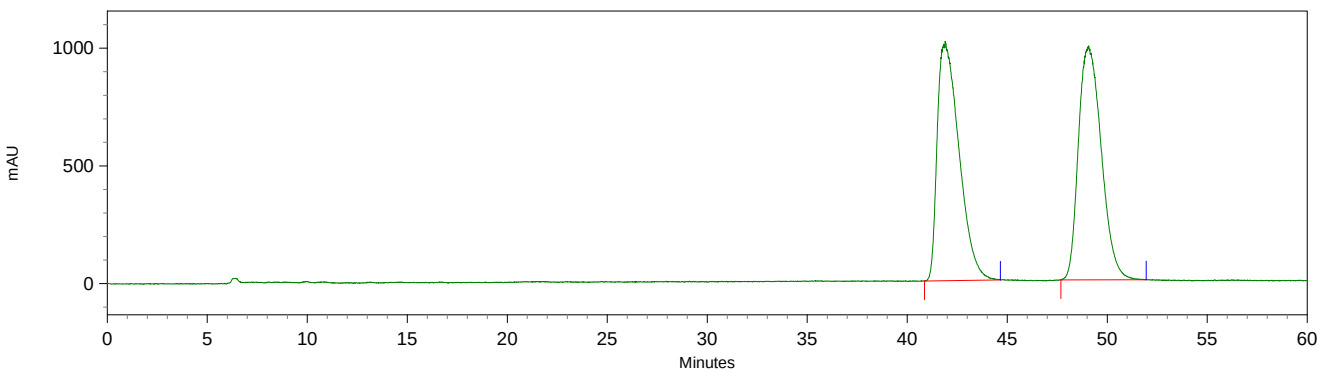
Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\2011-9-10\211b\rac-211b001

User: System

Acquired: 2011-9-10 18:16:06

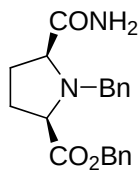
Printed: 2011-9-19 18:38:04



Detector A-213

nm

Pk #	Retention Time	Area	Area %
1	41.899	76472298	49.2
2	49.067	78818336	50.8
Totals		155290634	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

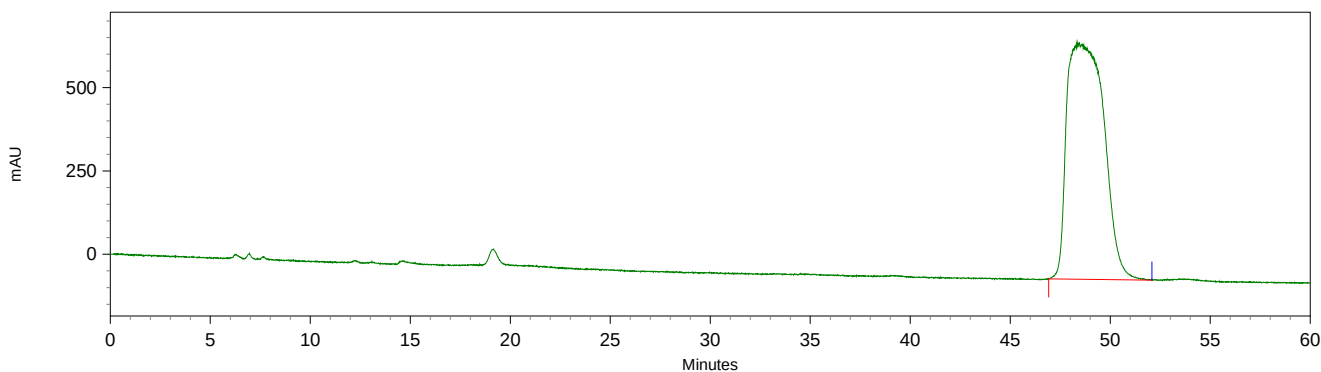
Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\2011-9-10 211b\211b-16h001

User: System

Acquired: 2011-9-11 13:16:57

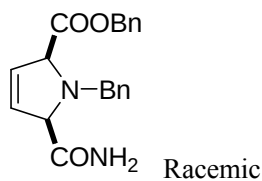
Printed: 2011-9-19 18:38:57



Detector A-213

nm

Pk #	Retention Time	Area	Area %
1	48.459	91678707	100.0
Totals		91678707	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

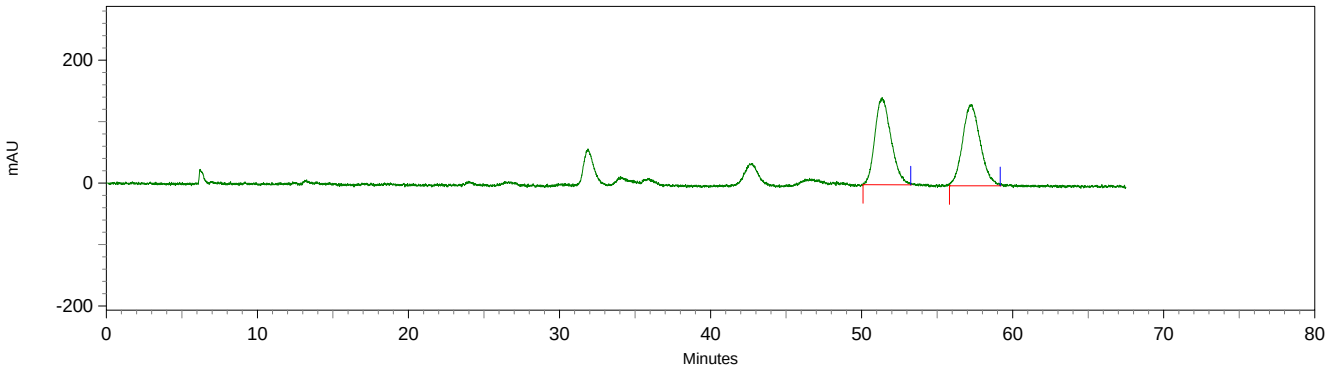
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Gaoming212nm.met

Data Name: E:\chenpeng\rac-d5-15c-0001.5-adh

User: System

Acquired: 2009-12-19 19:12:10

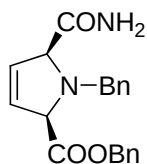
Printed: 2010-4-1 20:53:21



Detector

A-207 nm

Pk #	Retention Time	Area	Area %
1	51.360	10488937	49.3
2	57.205	10770562	50.7
Totals		21259499	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

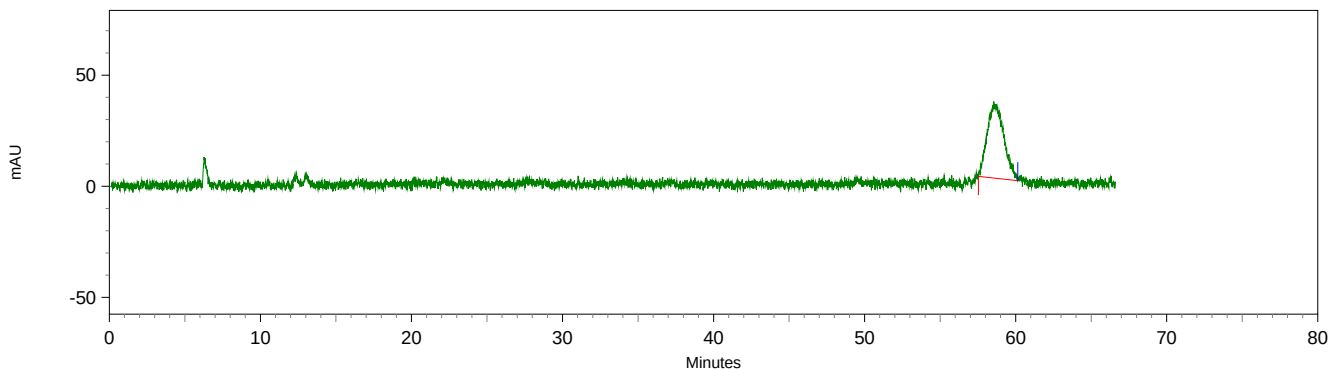
Method Name: D:\HPLC\Yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\2009\12\31\2009-12-31\12-31-04\12-31-04-0001.5-adh

User: System

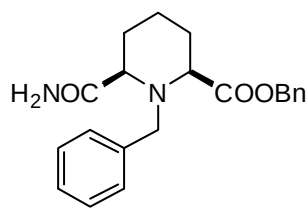
Acquired: 2009-12-31 12:38:04

Printed: 2010-4-1 20:39:37



Detector
A-207 nm

Pk #	Name	Retention Time	Area	Area %
1		58.549	2586761	100.0
Totals			2586761	100.0



Racemic

Shimadzu CLASS-VP V6.13 SP2

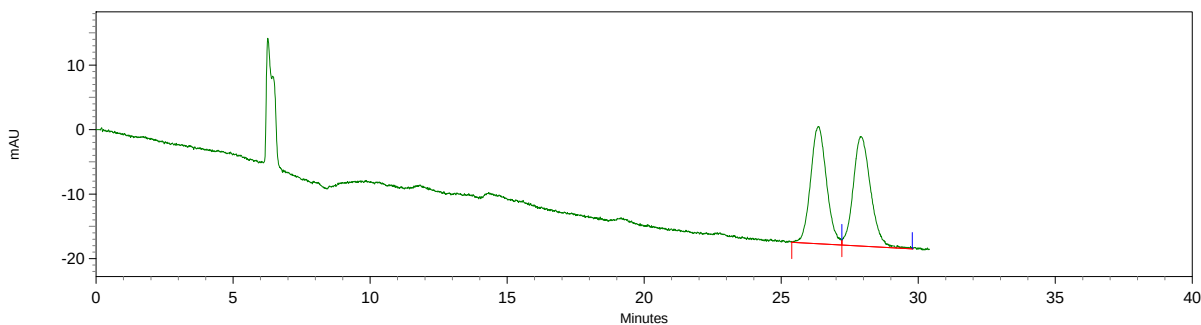
Area % Report

Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\hexdiamide\rac-h-3-15c-0.5-adh'

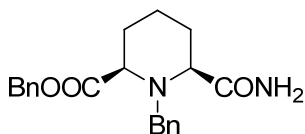
User: System

Acquired: 2008-7-9 20:33:29



1: 254 nm, 8
nm

Pk #	Retention Time	Area	Area %
1	26.368	735902	49.5
2	27.915	750097	50.5
Totals		1485999	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

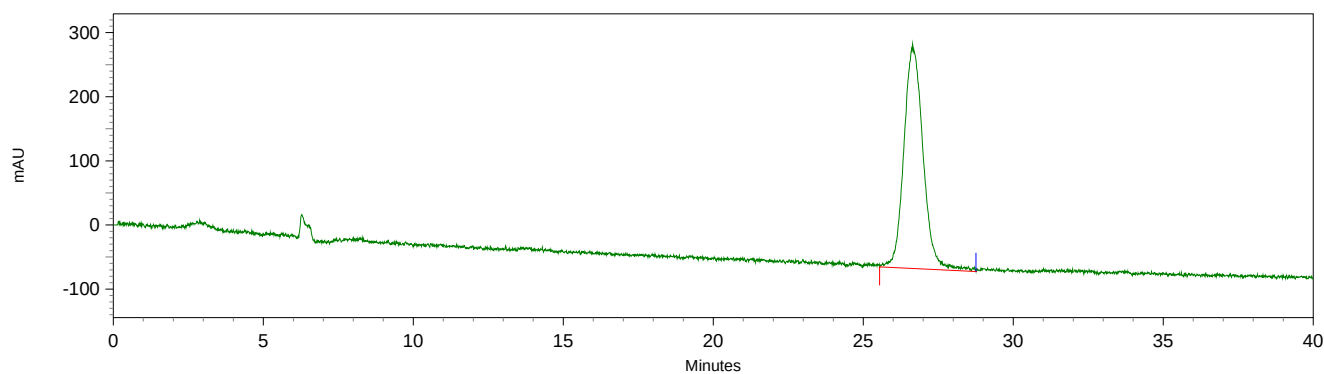
Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

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User: System

Acquired: 2010-6-25 17:25:06

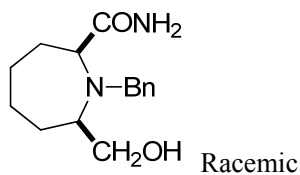
Printed: 2011-9-13 16:29:39



Detector A-216

nm

Pk #	Retention Time	Area	Area %
1	26.645	15439731	100.0
Totals		15439731	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

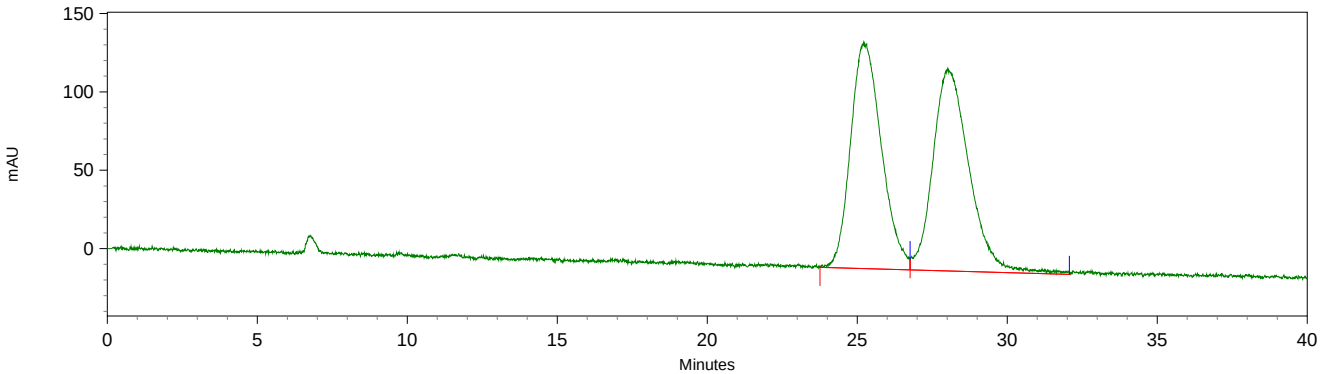
Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\q\RAC-Q5-0001.5-20C-OJH'

User: System

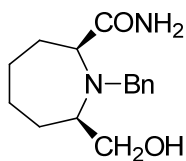
Acquired: 2009-7-18 16:18:44

Printed: 2011-9-13 16:55:06



Detector A-215
nm

Pk #	Retention Time	Area	Area %
1	25.216	10316827	48.8
2	28.021	10827528	51.2
Totals		21144355	100.0



cShimadzu CLASS-VP V6.13 SP2

Area % Report

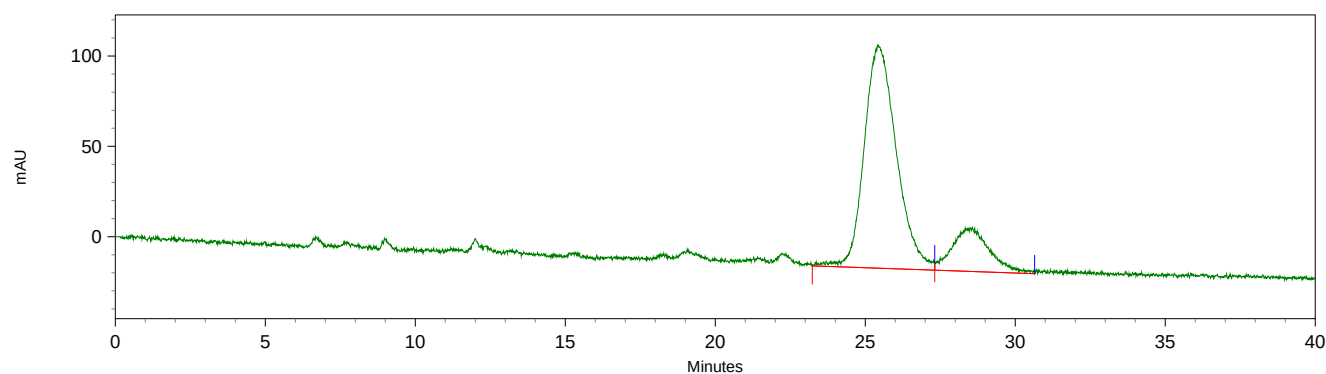
Method Name: D:\HPLC\yangluo\2006\method\14p
Gaoming212nm.met

Data Name: E:\chenpeng\q\bio-q5-0002.5-20c-ojh

User: System

Acquired: 2009-7-18 9:50:01

Printed: 2011-9-13 16:58:38

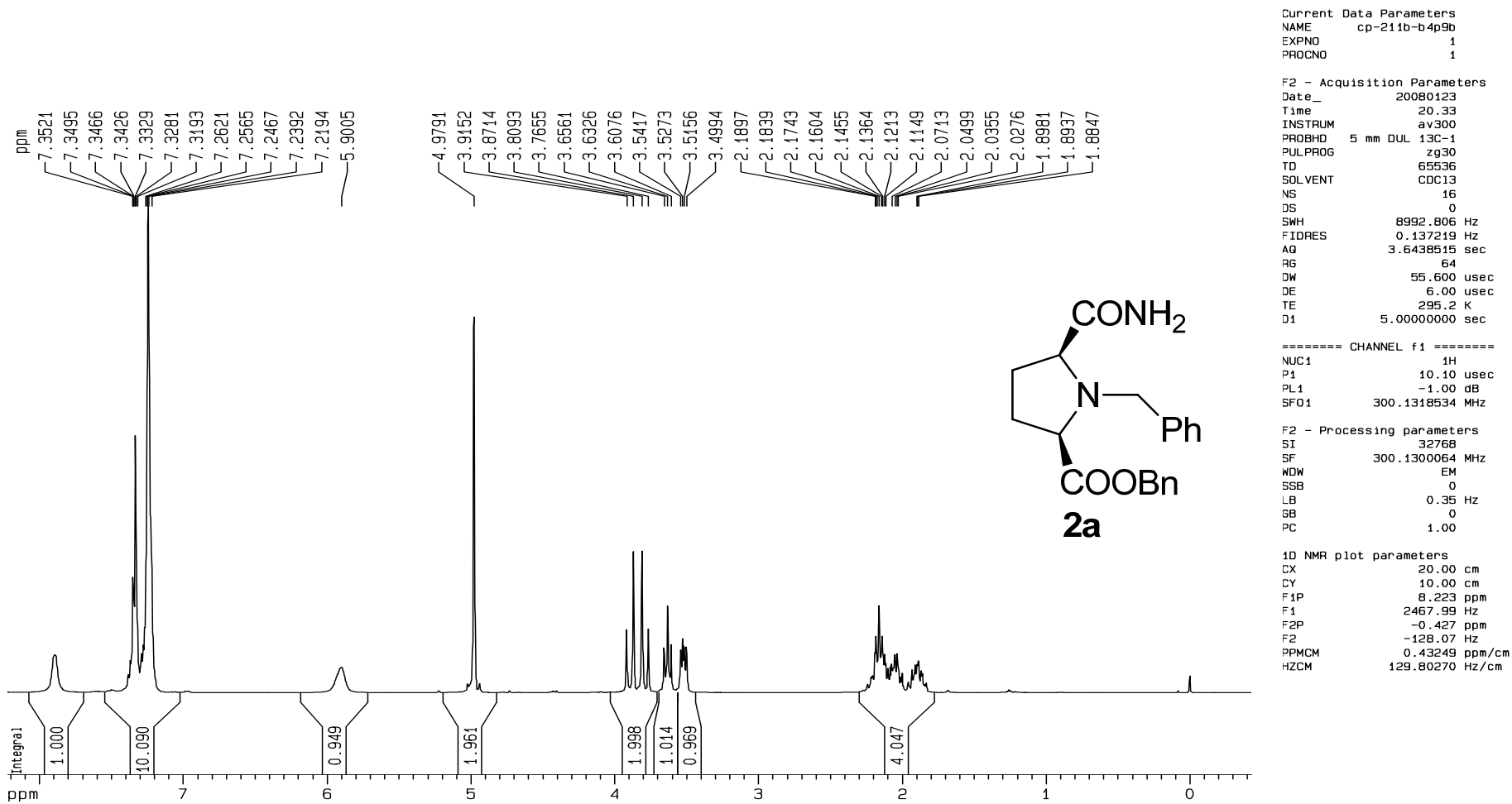


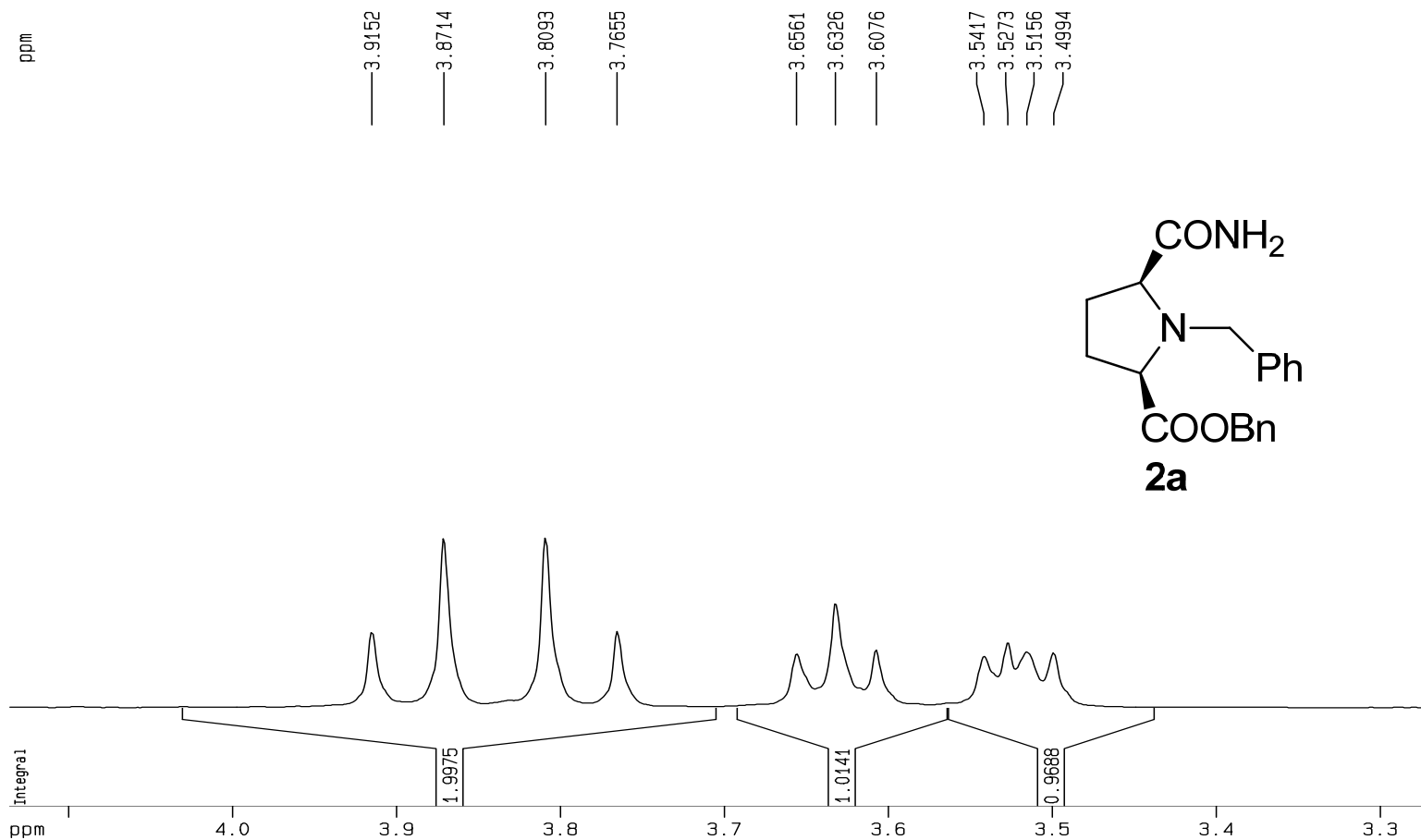
Detector A-215

nm

Pk #	Retention Time	Area	Area %
1	25.419	9222160	81.7
2	28.512	2072085	18.3
Totals		11294245	100.0

5. ^1H and ^{13}C NMR spectra of all products





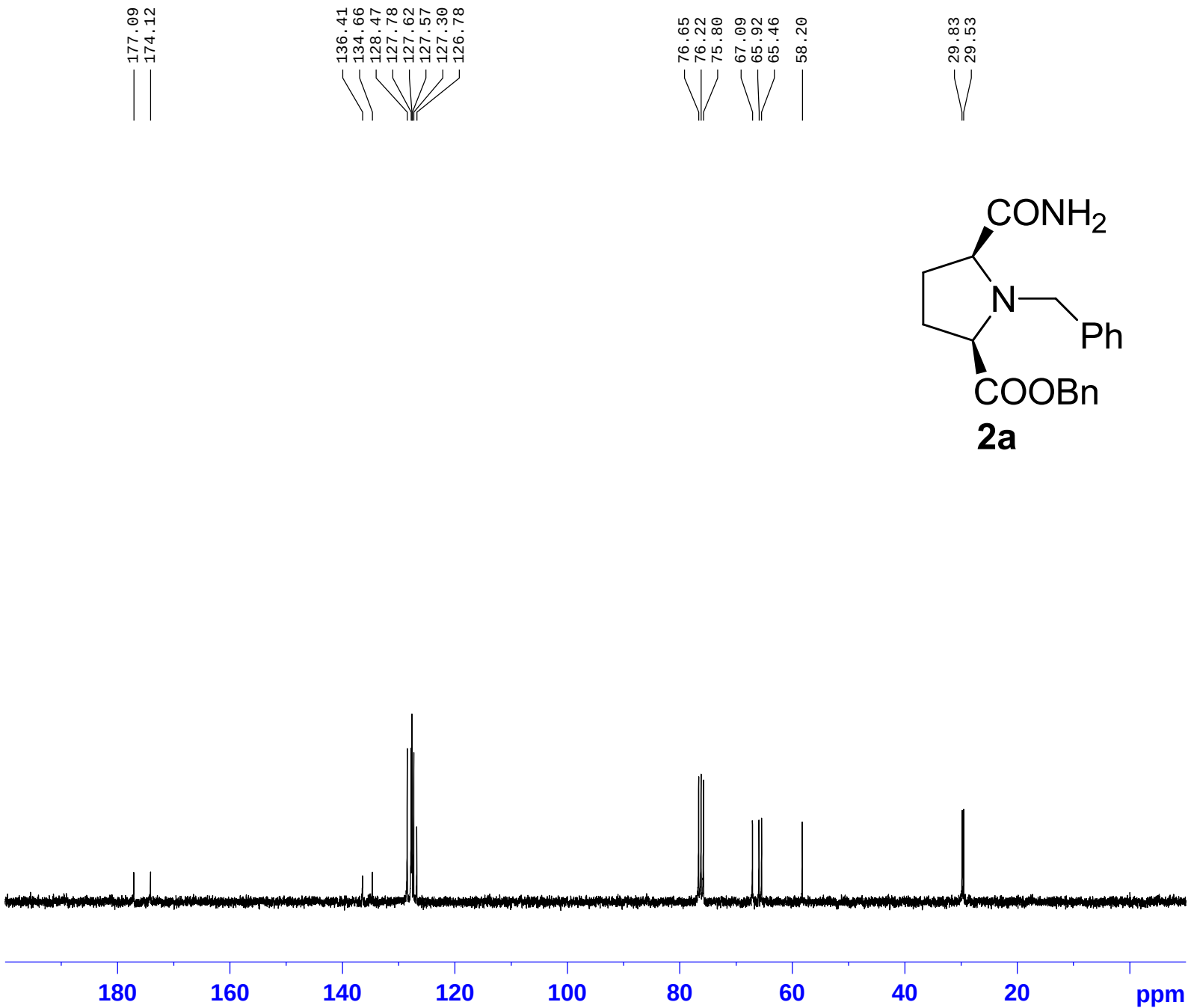
Current Data Parameters
NAME cp-211b-b4p9b
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080123
Time 20.33
INSTRUM av300
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 64
DW 55.600 usec
DE 6.00 usec
TE 295.2 K
D1 5.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.10 usec
PL1 -1.00 dB
SF01 300.1318534 MHz

F2 - Processing parameters
SI 32768
SF 300.1300064 MHz
WDW EM
SSB 0
LB 0.35 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 10.00 cm
F1P 4.136 ppm
F1 1241.40 Hz
F2P 3.274 ppm
F2 982.60 Hz
PPMCM 0.04311 ppm/cm
HZCM 12.93984 Hz/cm



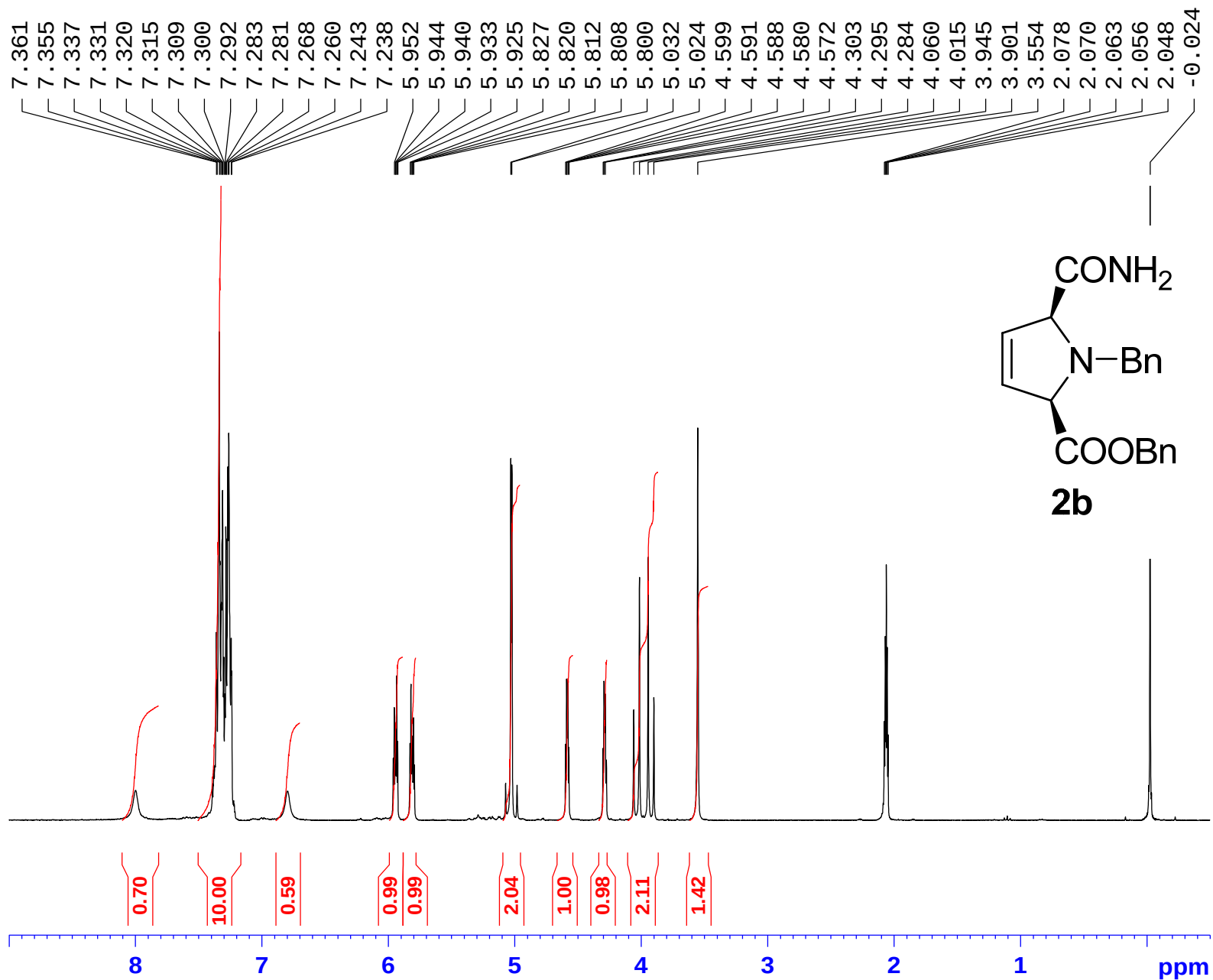
Current Data Parameters
NAME cp-2d
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090821
Time 9.41
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PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 92
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 1625.5
DW 27.800 usec
DE 6.00 usec
TE 301.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 20.16 dB
PL13 16.98 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4678131 MHz
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SSB 0
LB 1.00 Hz
GB 0
PC 1.40

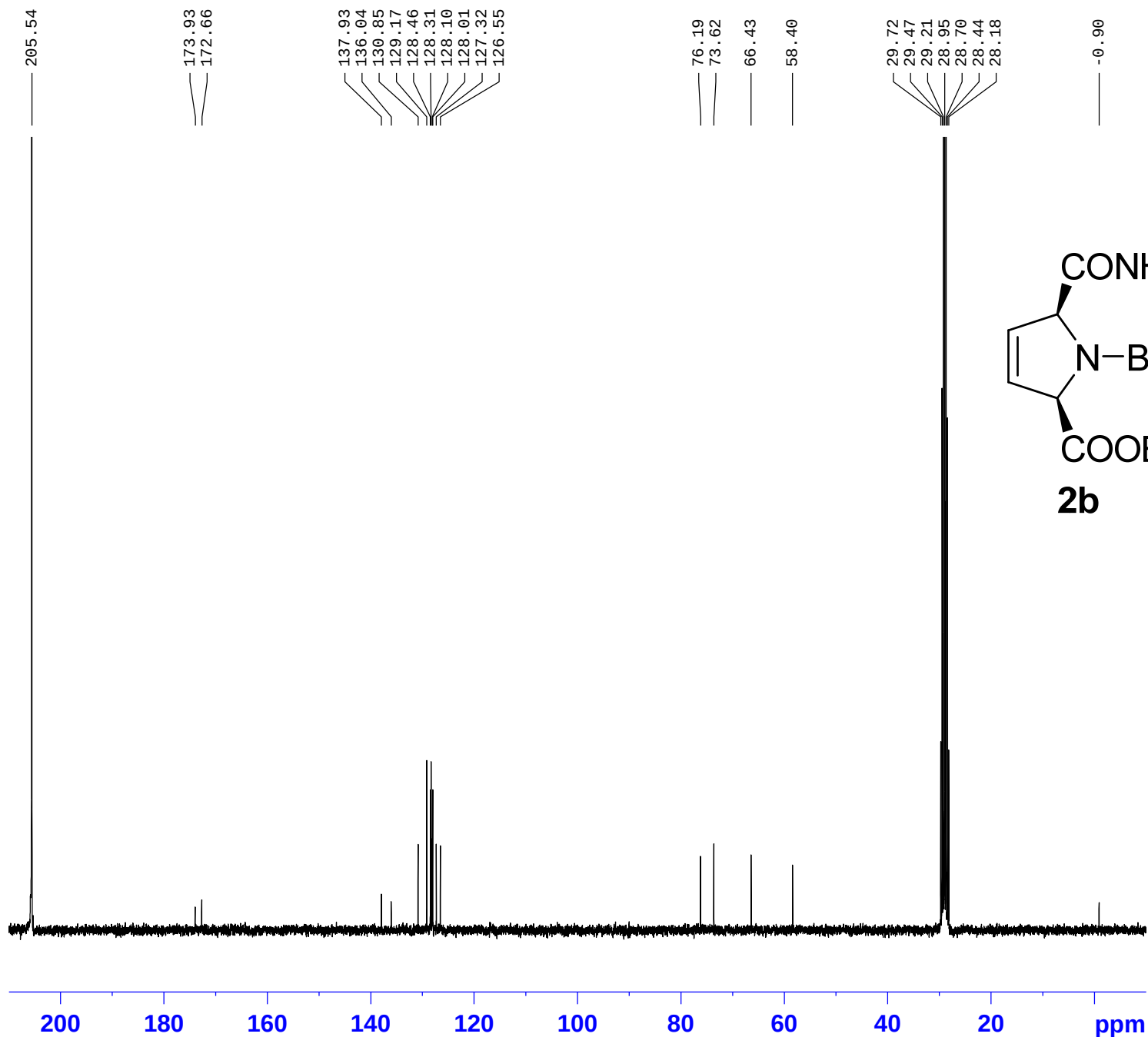


Current Data Parameters
NAME cp-d5'
EXPNO 46
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111006
Time 18.53
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT Acetone/D2O
NS 12
DS 0
SWH 8992.806 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 256
DW 55.600 usec
DE 8.00 usec
TE 299.2 K
D1 2.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1318008 MHz

F2 - Processing parameters
SI 32768
SF 300.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME cp-d5'
EXPNO 44
PROCNO 1

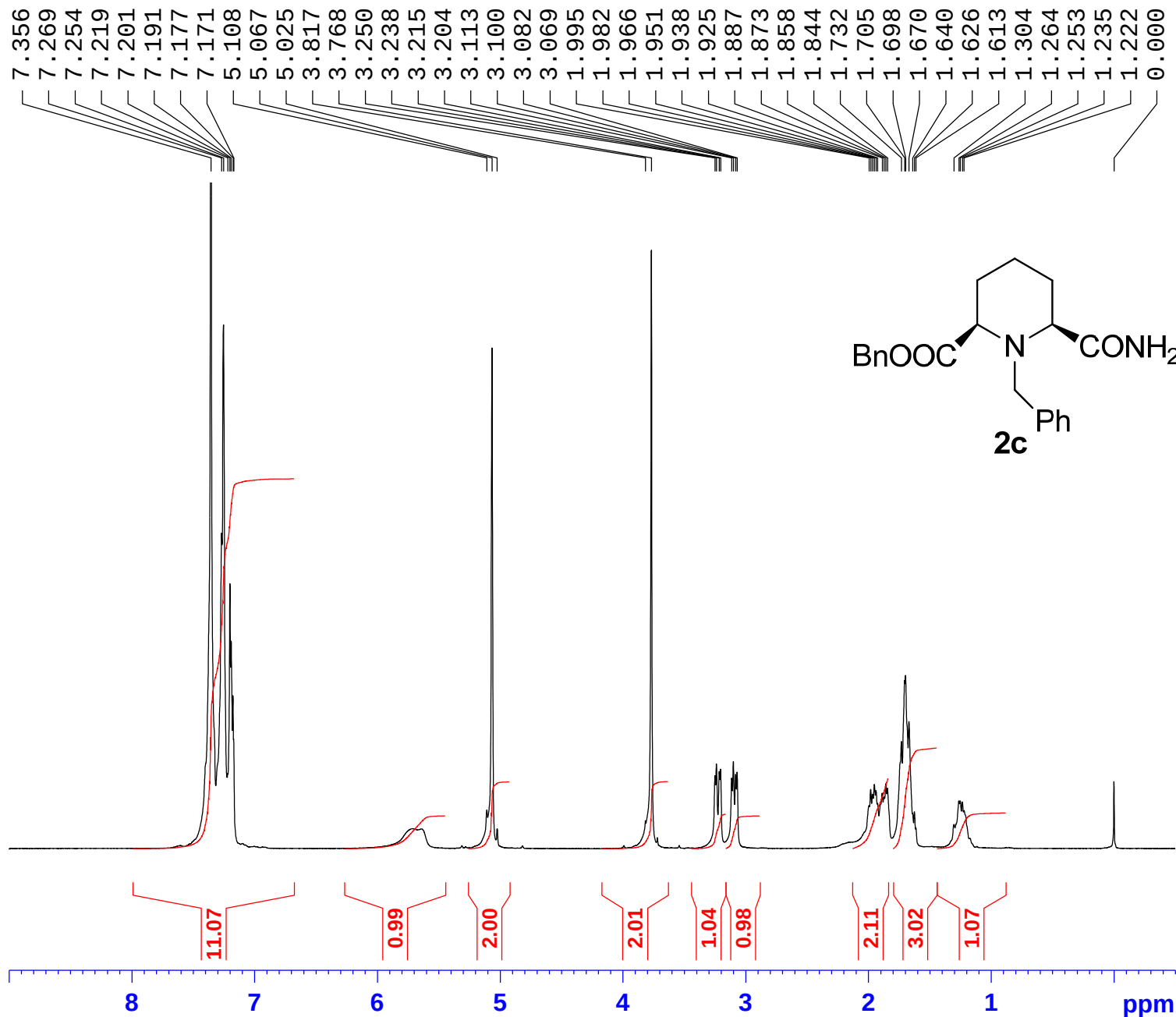
F2 - Acquisition Parameters

Date_ 20111006
Time 17.53
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgig30
TD 32768
SOLVENT Acetone
NS 500
DS 0
SWH 18832.393 Hz
FIDRES 0.574719 Hz
AQ 0.8700404 sec
RG 256
DW 26.550 usec
DE 8.00 usec
TE 299.7 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
SF02 300.1312005 MHz

F2 - Processing parameters
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SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

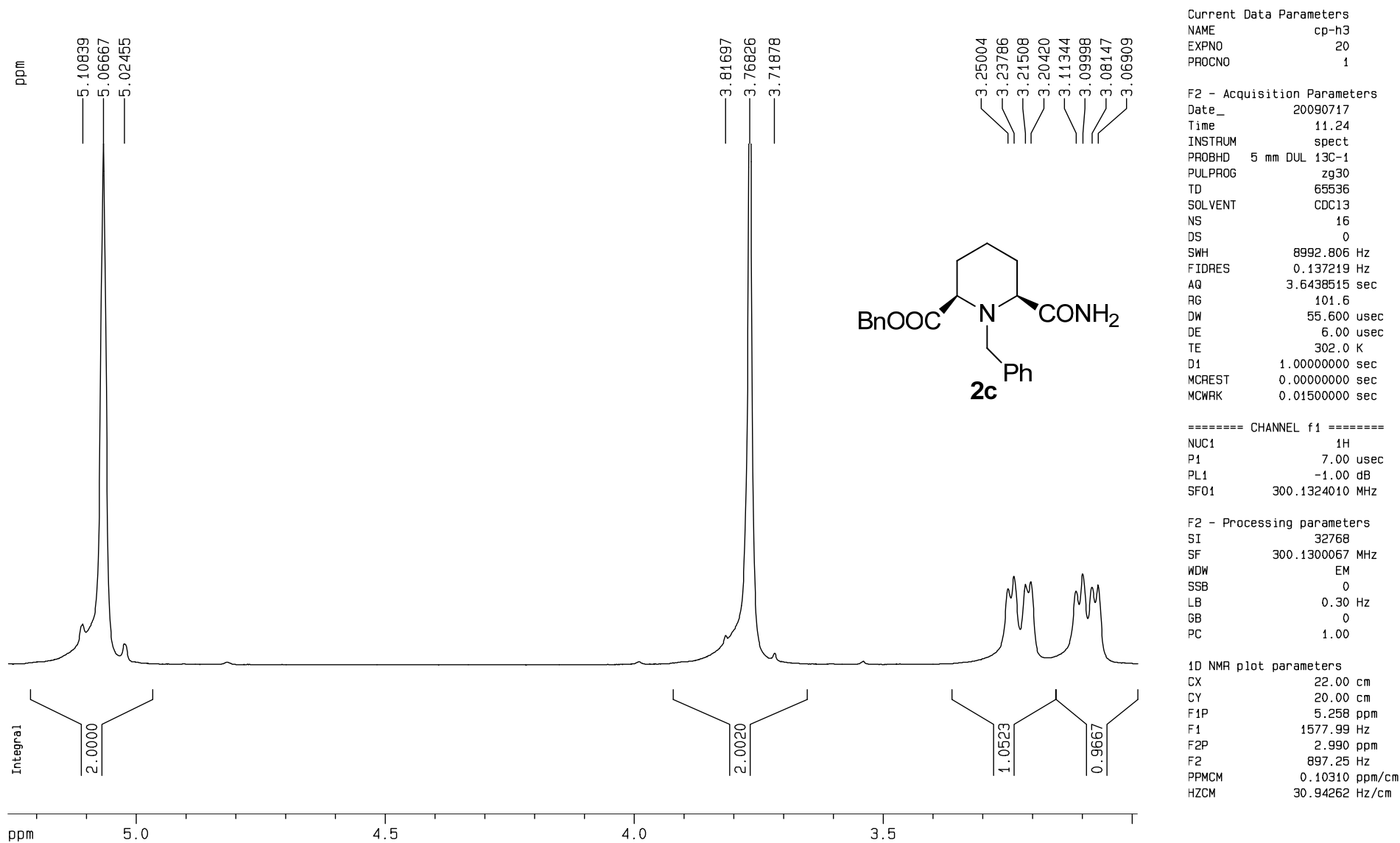


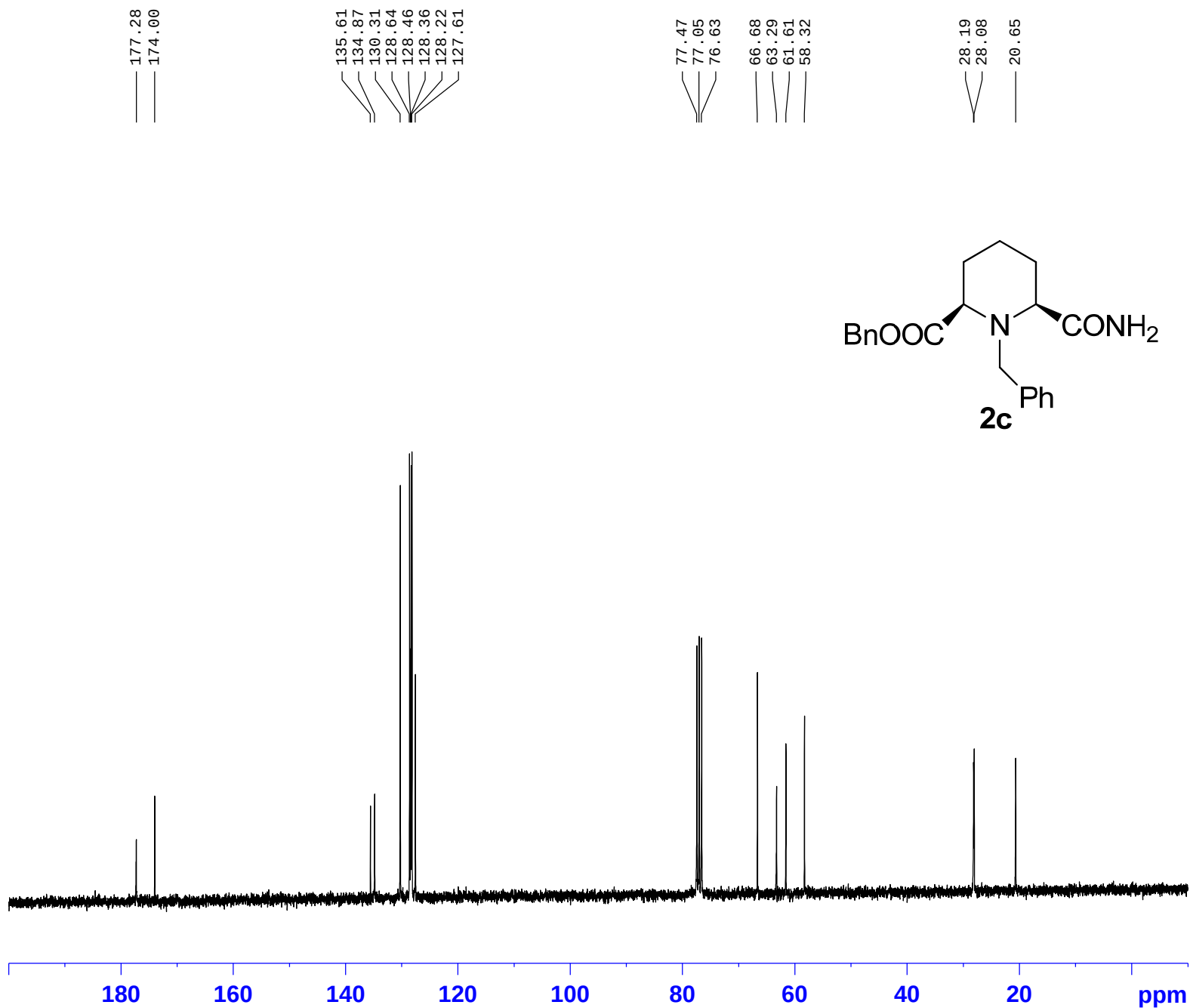
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EXPNO 20
PROCNO 1

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Time 11.24
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PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 101.6
DW 55.600 usec
DE 6.00 usec
TE 302.0 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 -1.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300067 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





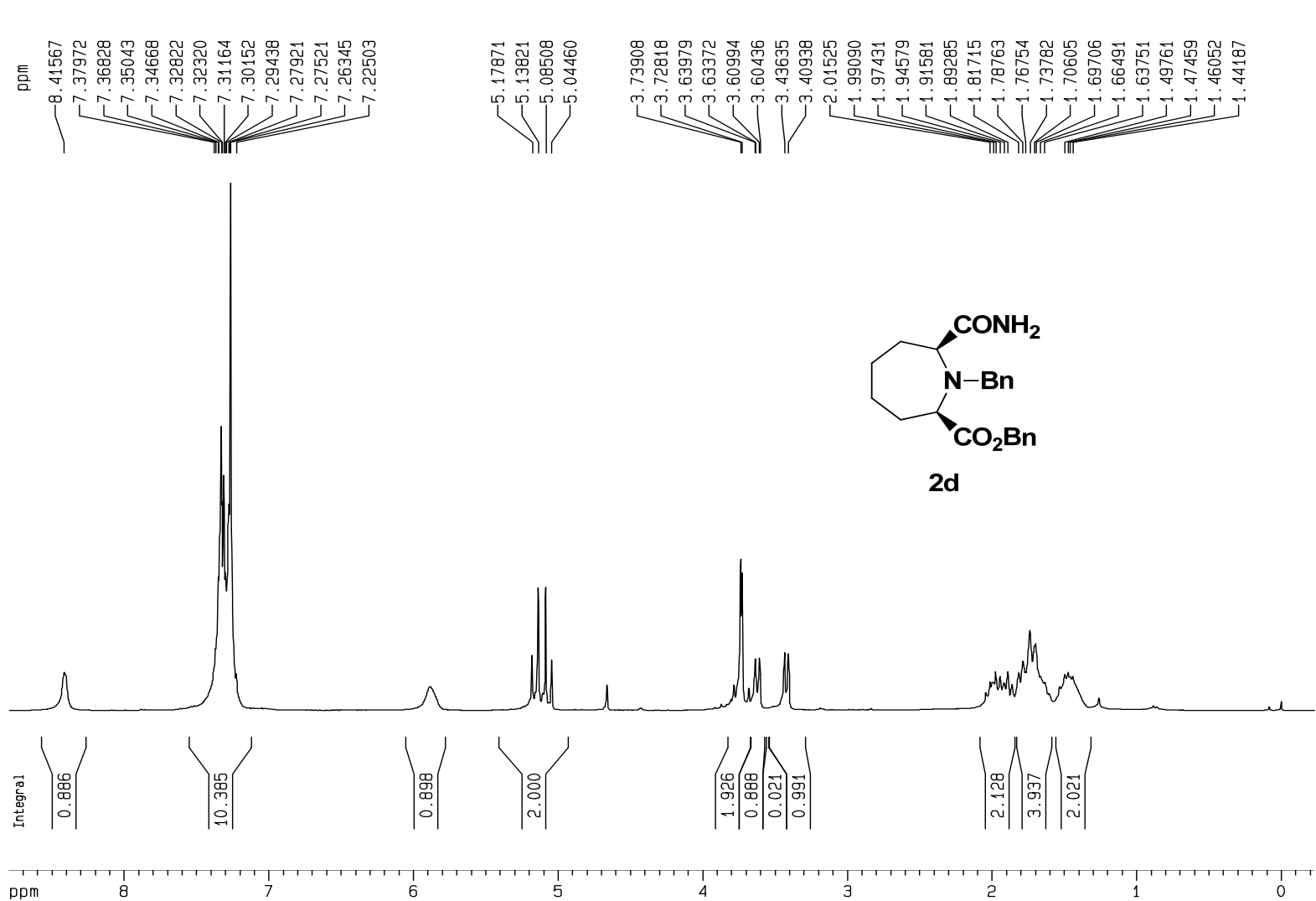
Current Data Parameters
NAME cp-h3
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090717
Time 11.38
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 261
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 1149.4
DW 27.800 usec
DE 6.00 usec
TE 302.1 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 20.16 dB
PL13 16.98 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677487 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



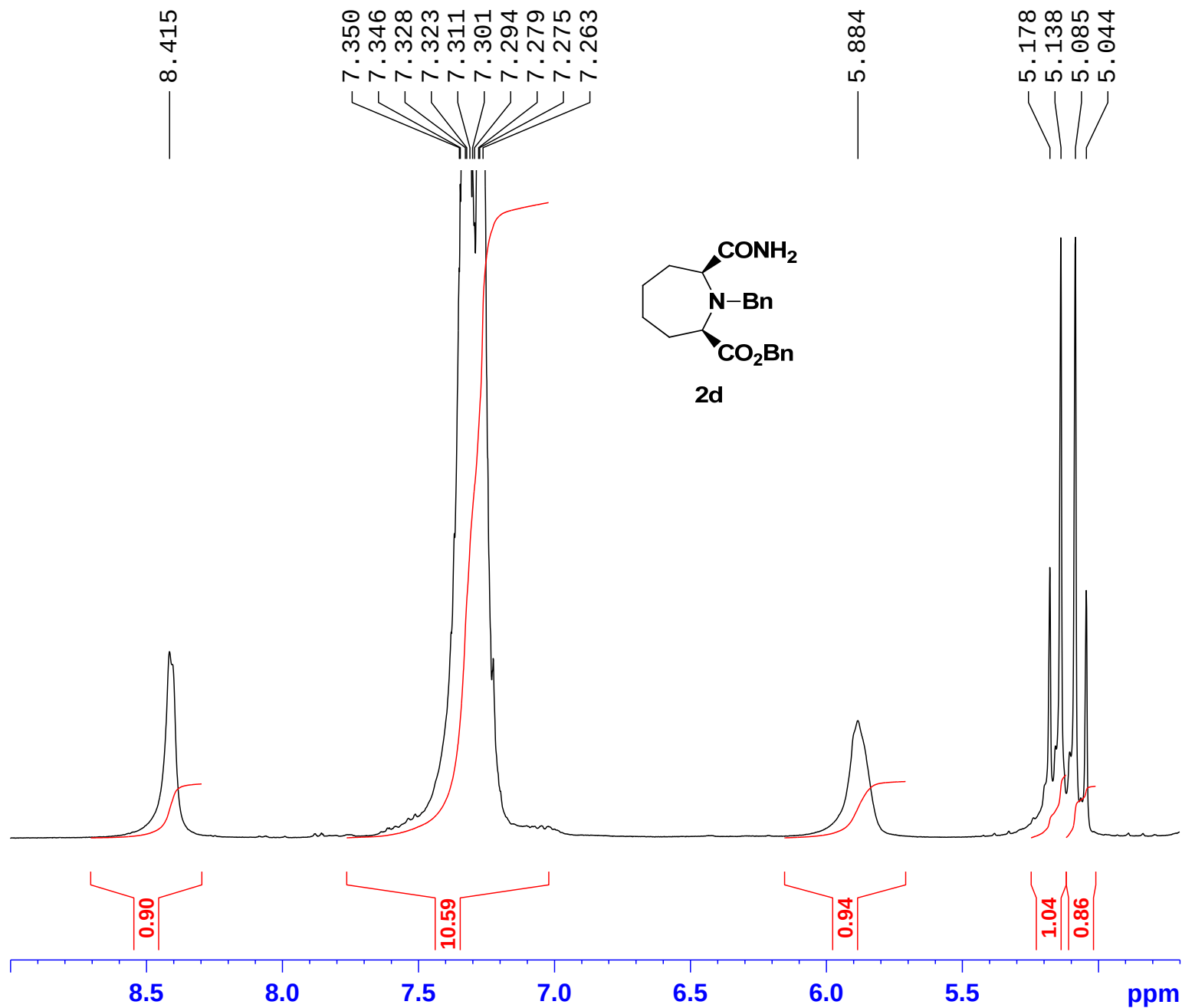
Current Data Parameters
NAME cp-q4
EXPNO 60
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090630
Time 14.12
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 57
DW 55.600 usec
DE 6.00 usec
TE 299.7 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 -1.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300086 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 22.00 cm
CY 10.00 cm
F1P 8.796 ppm
F1 2639.94 Hz
F2P -0.227 ppm
F2 -68.17 Hz
PPMCM 0.41014 ppm/cm
HZCM 123.09608 Hz/cm



Current Data Parameters
NAME cp-q4
EXPNO 60
PROCNO 1

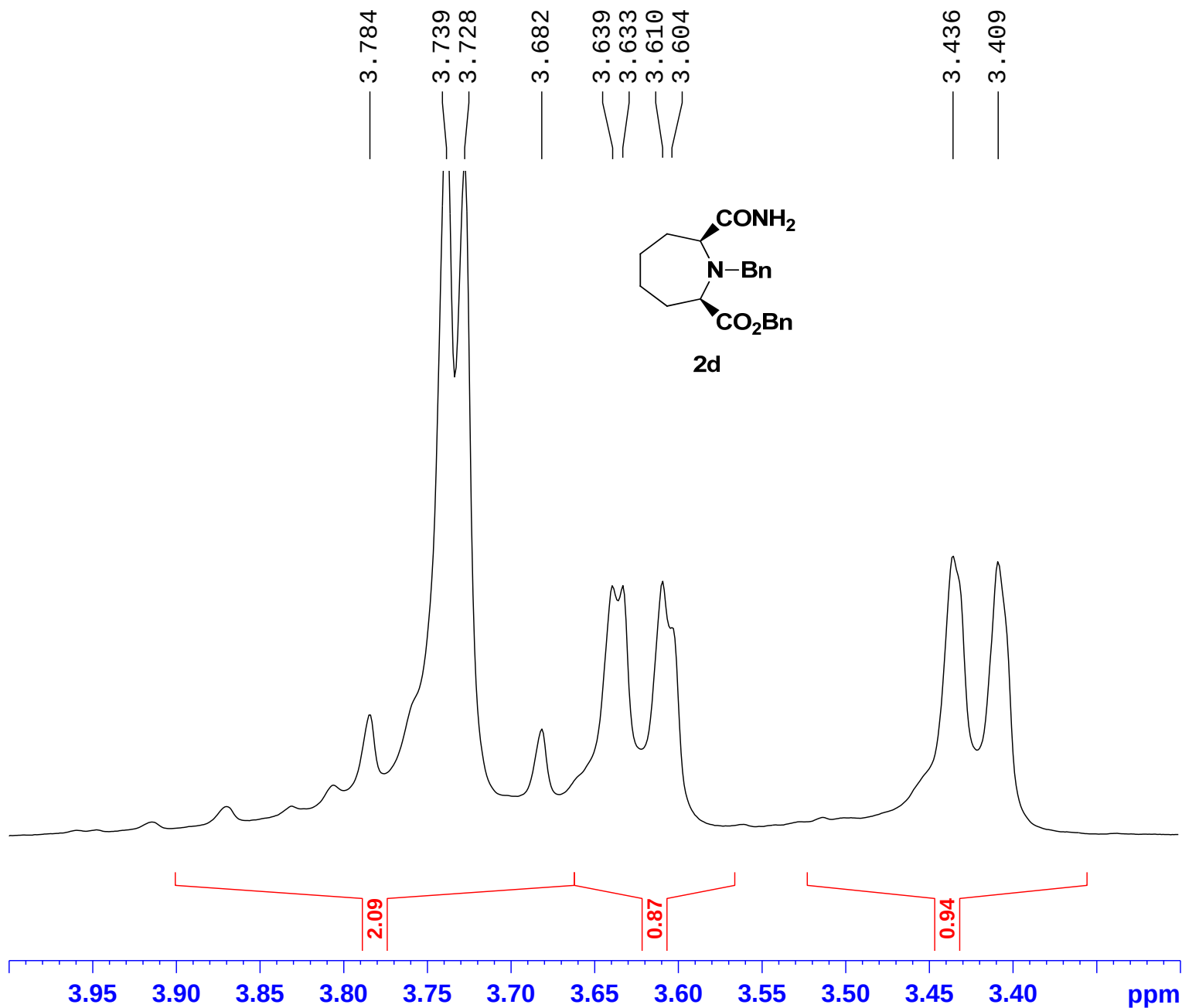
F2 - Acquisition Parameters

Date_ 20090630
Time 14.12
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 57
DW 55.600 usec
DE 6.00 usec
TE 299.7 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 -1.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters

SI 32768
SF 300.1300086 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME cp-q4
EXPNO 60
PROCNO 1

F2 - Acquisition Parameters

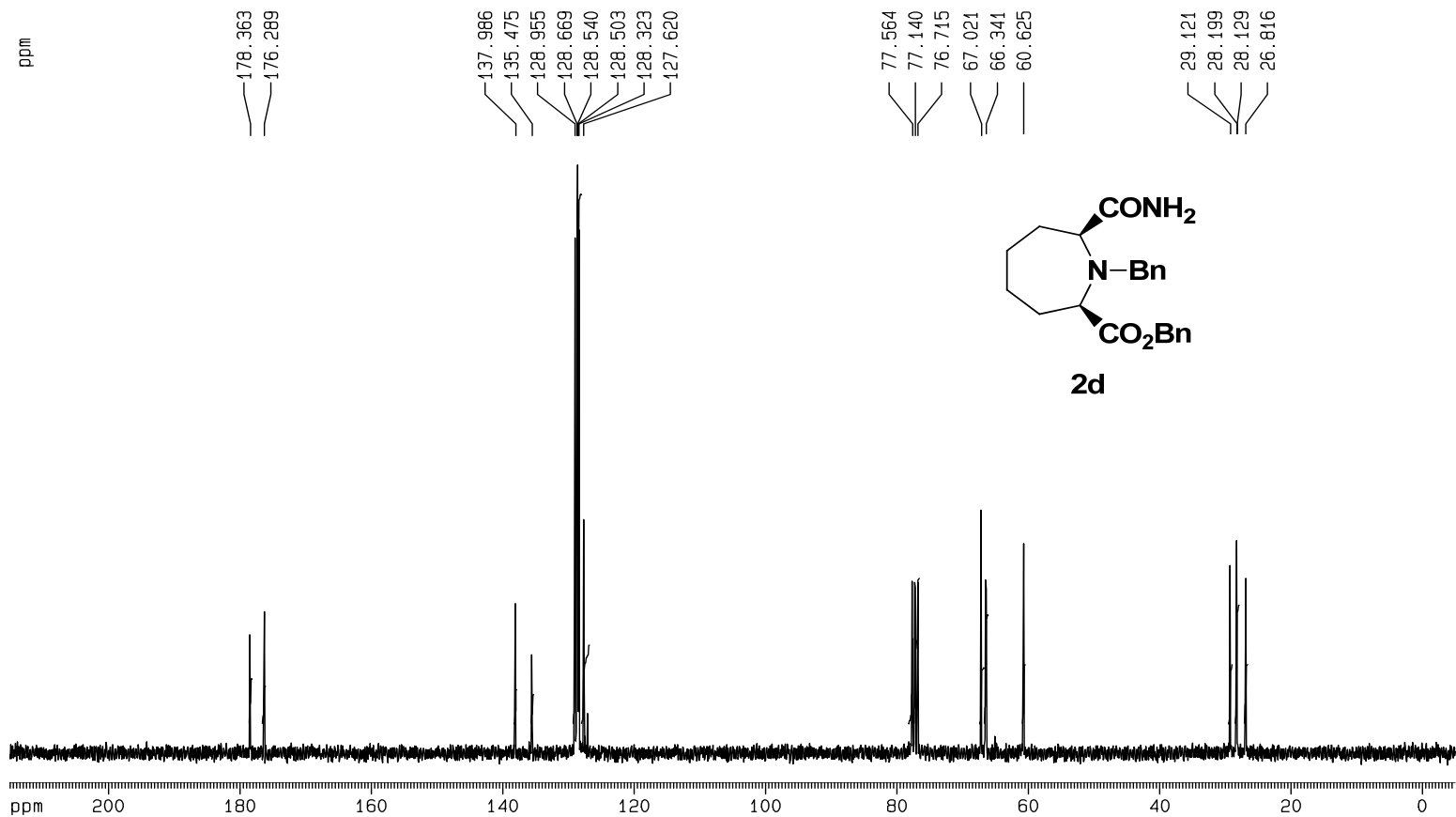
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Time 14.12
INSTRUM spect
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PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 57
DW 55.600 usec
DE 6.00 usec
TE 299.7 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====

NUC1 1H
P1 7.00 usec
PL1 -1.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters

SI 32768
SF 300.1300086 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME cp-q4
EXPNO 61
PROCNO 1

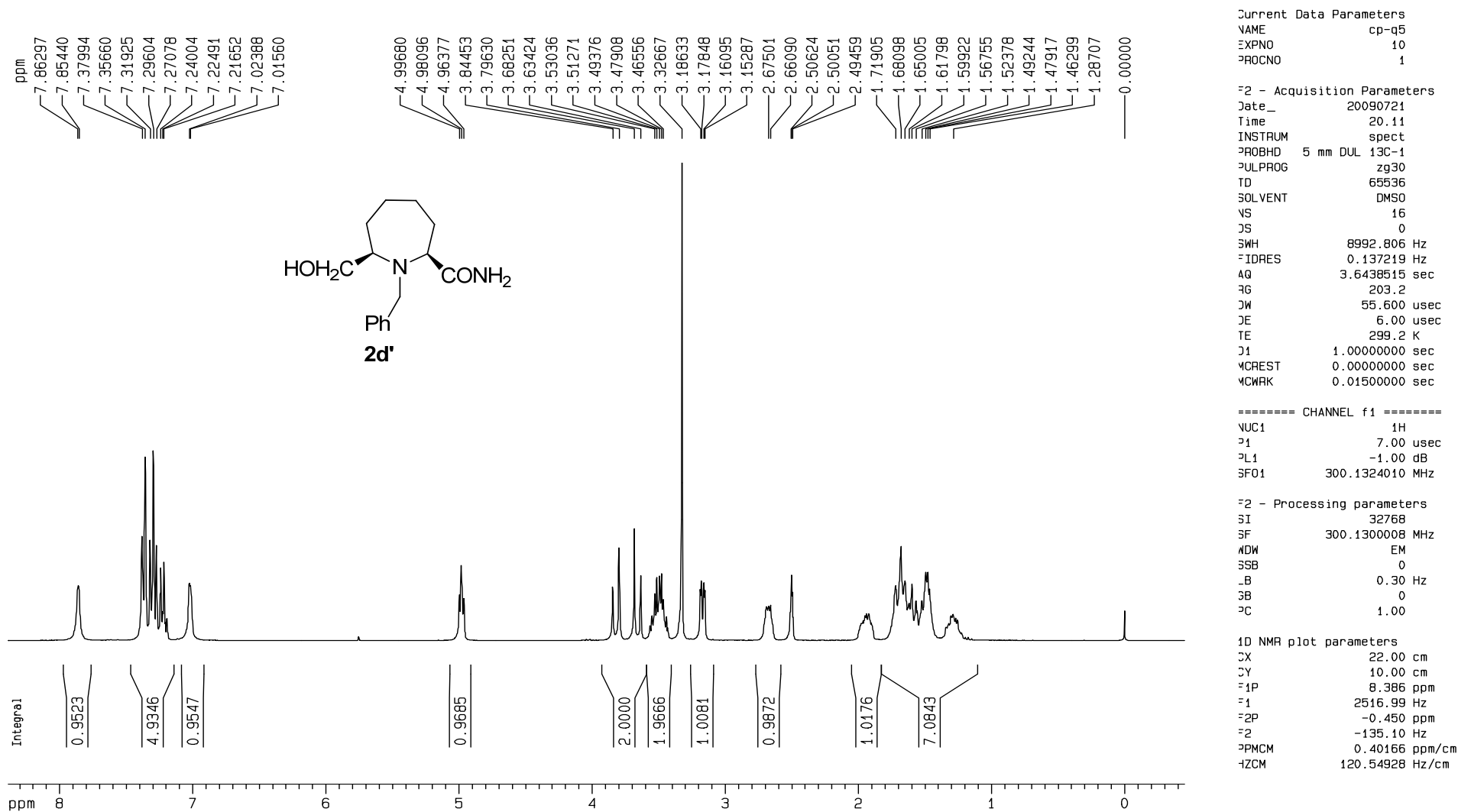
F2 - Acquisition Parameters
Date_ 20090530
Time_ 14.20
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
VS 87
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 8192
RW 27.800 usec
DE 6.00 usec
TE 301.1 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
VCREST 0.00000000 sec
VCMRK 0.01500000 sec

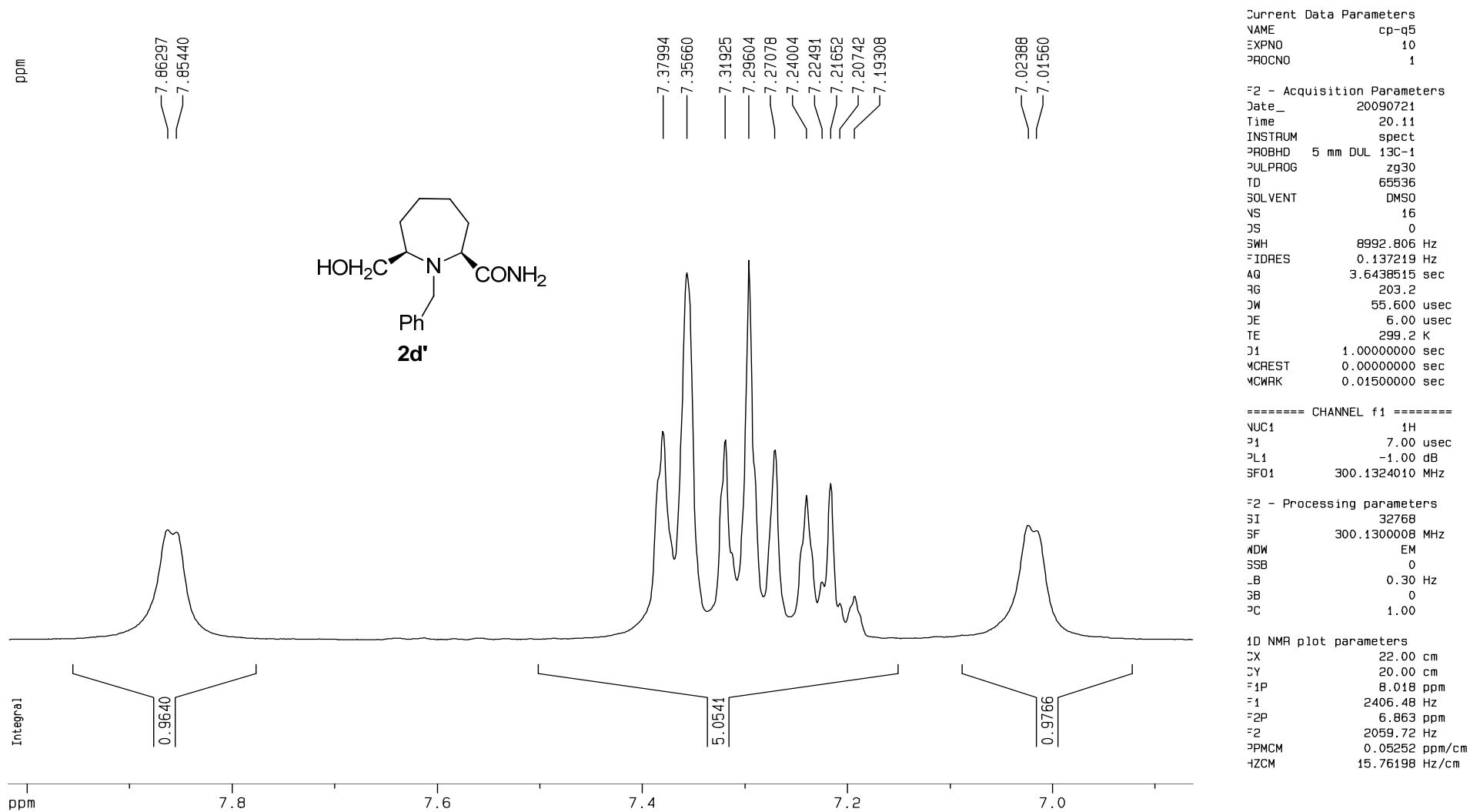
===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
RF01 75.4752953 MHz

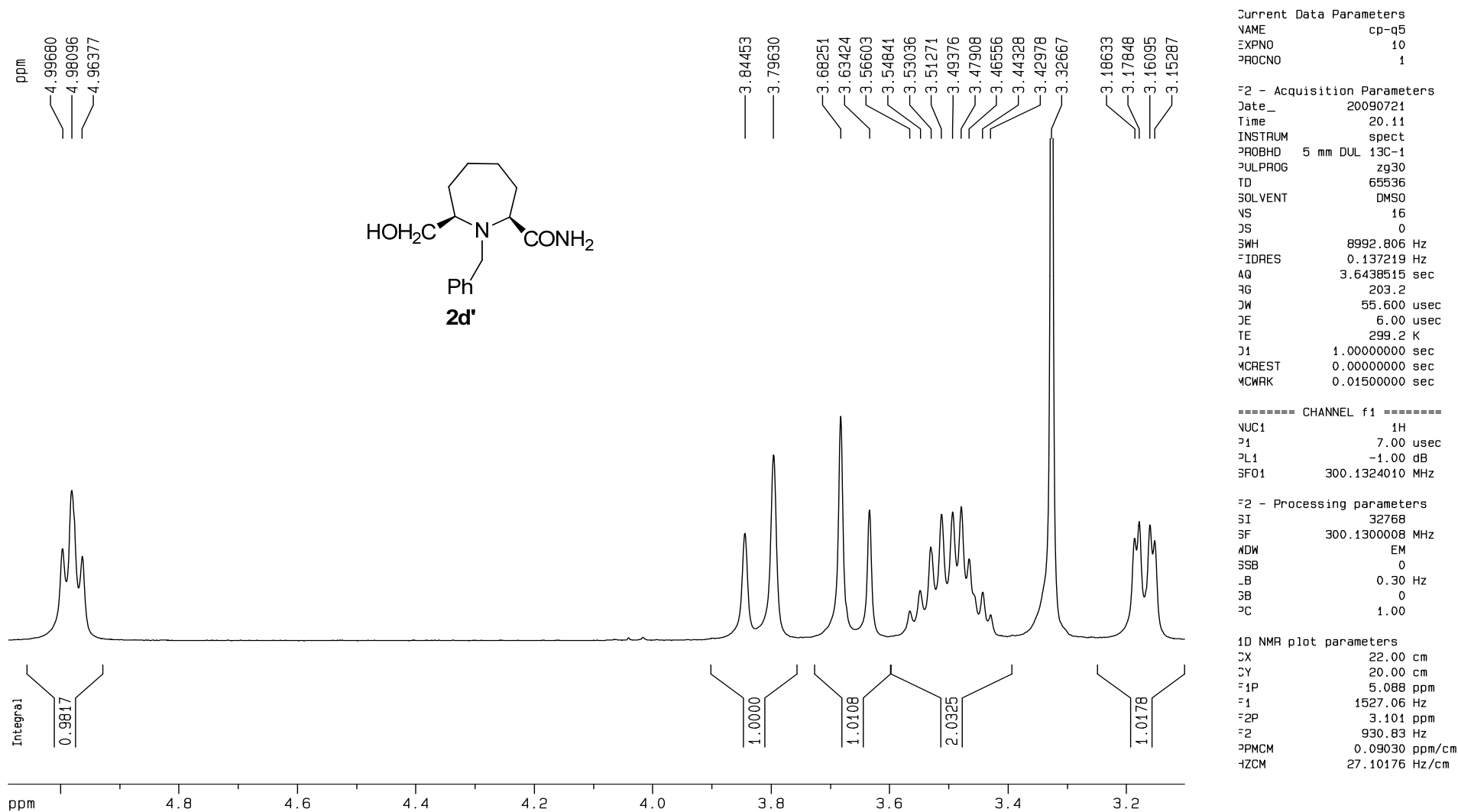
===== CHANNEL f2 =====
PDPG62 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 20.16 dB
PL13 16.98 dB
RF02 300.1312005 MHz

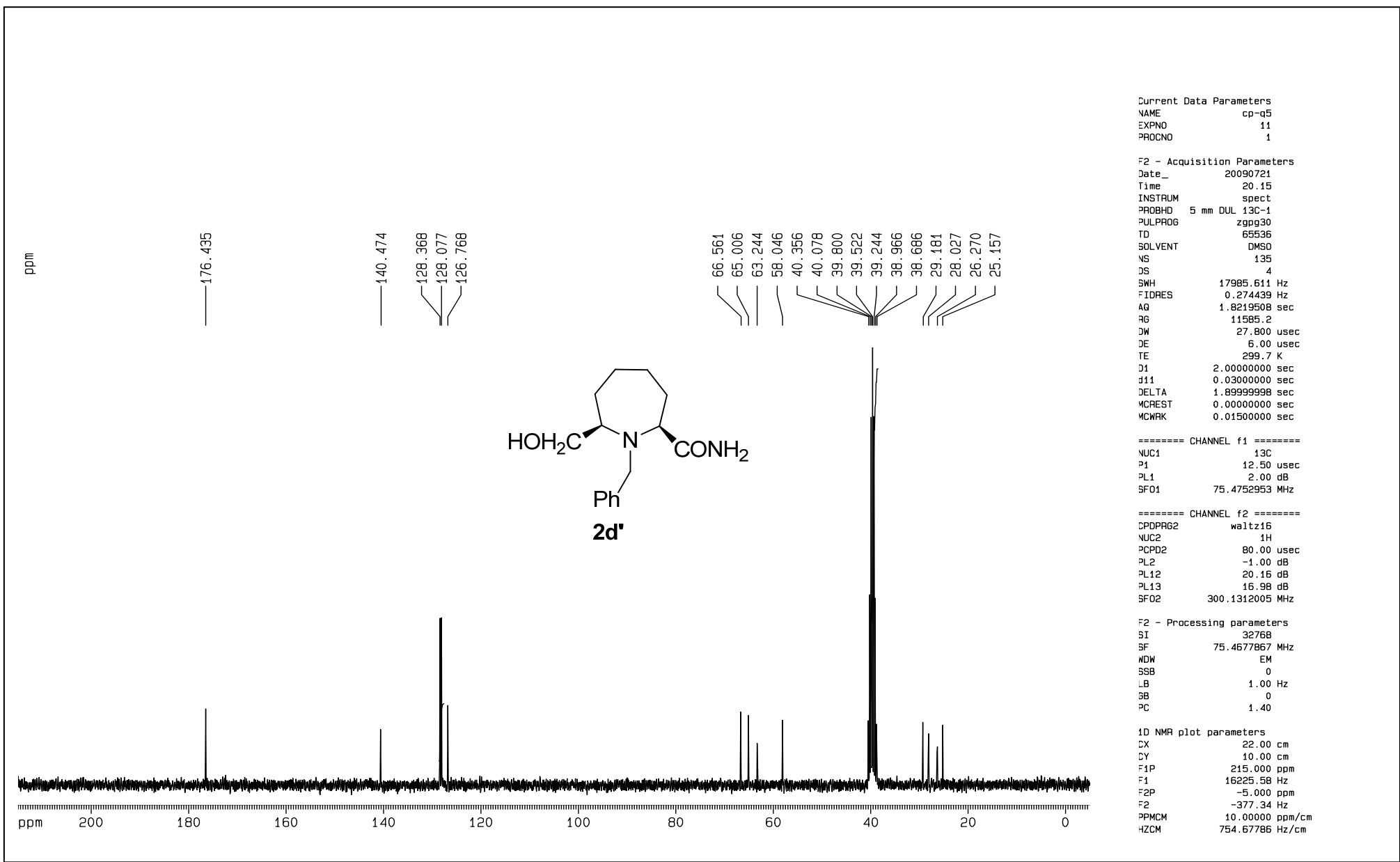
F2 - Processing parameters
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

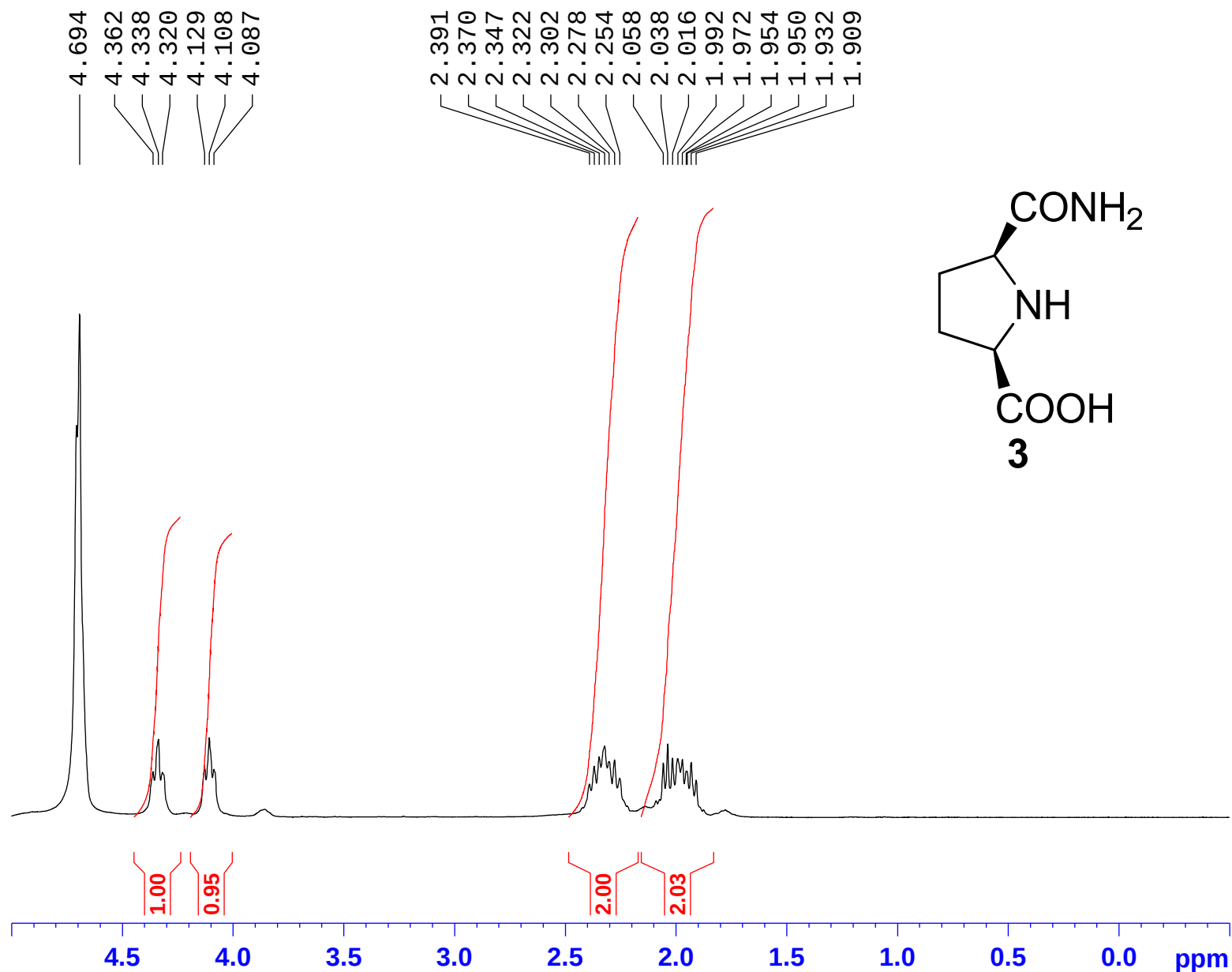
1D NMR plot parameters
CX 22.00 cm
CY 10.00 cm
F1P 215.000 ppm
F1 16225.57 Hz
F2P -5.000 ppm
F2 -377.34 Hz
PMCM 10.00000 ppm/cm
HZCM 754.67749 Hz/cm





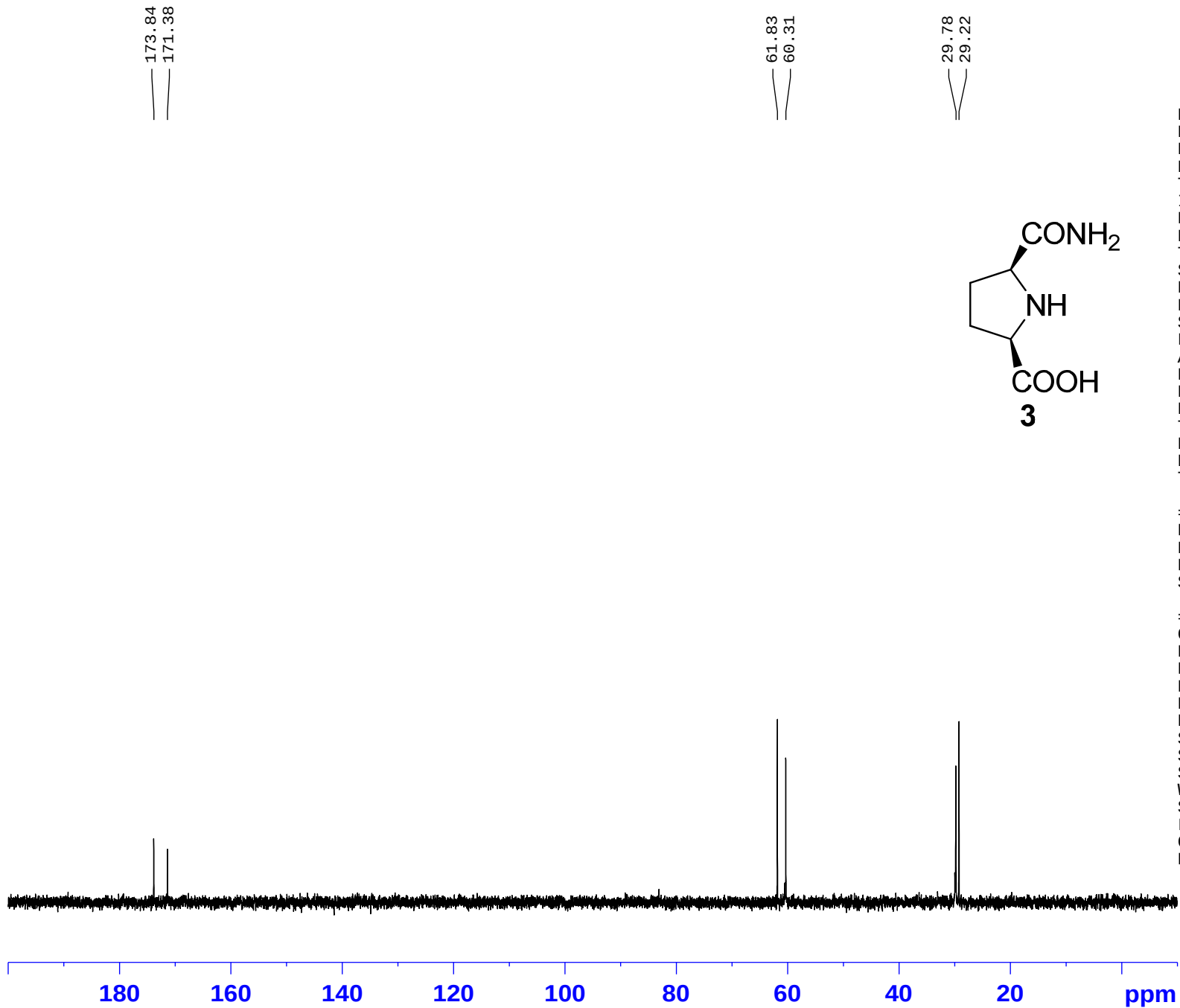






NAME cp-p0
EXPNO 20
PROCNO 1
Date_ 20101130
Time 20.09
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT D2O
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 128
DW 55.600 usec
DE 8.00 usec
TE 296.0 K
D1 1.00000000 sec
TD0 1

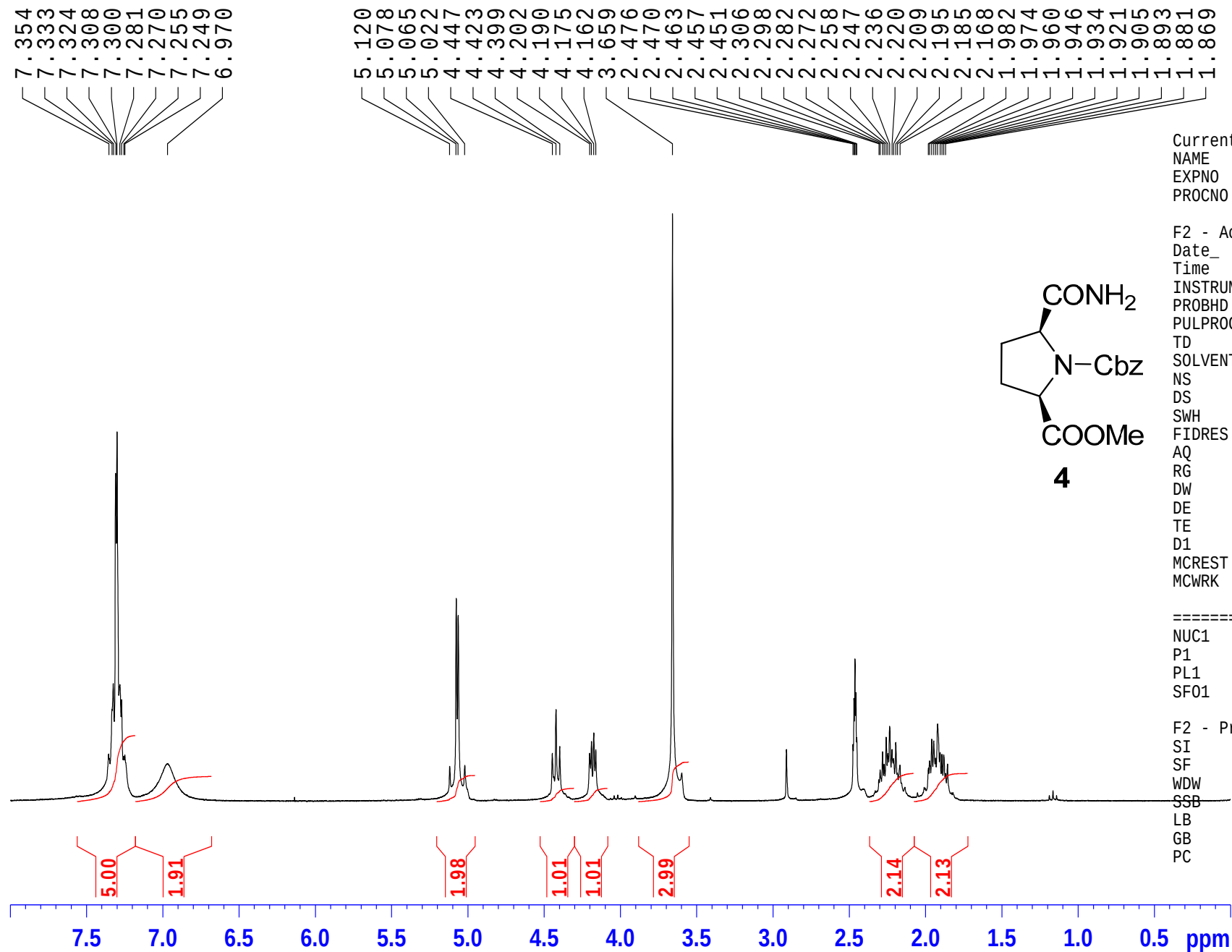
===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz
SI 32768
SF 300.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME cp-p0
EXPNO 21
PROCNO 1
Date_ 20101130
Time 20.14
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT D2O
NS 262
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 14596.5
DW 27.800 usec
DE 8.00 usec
TE 296.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

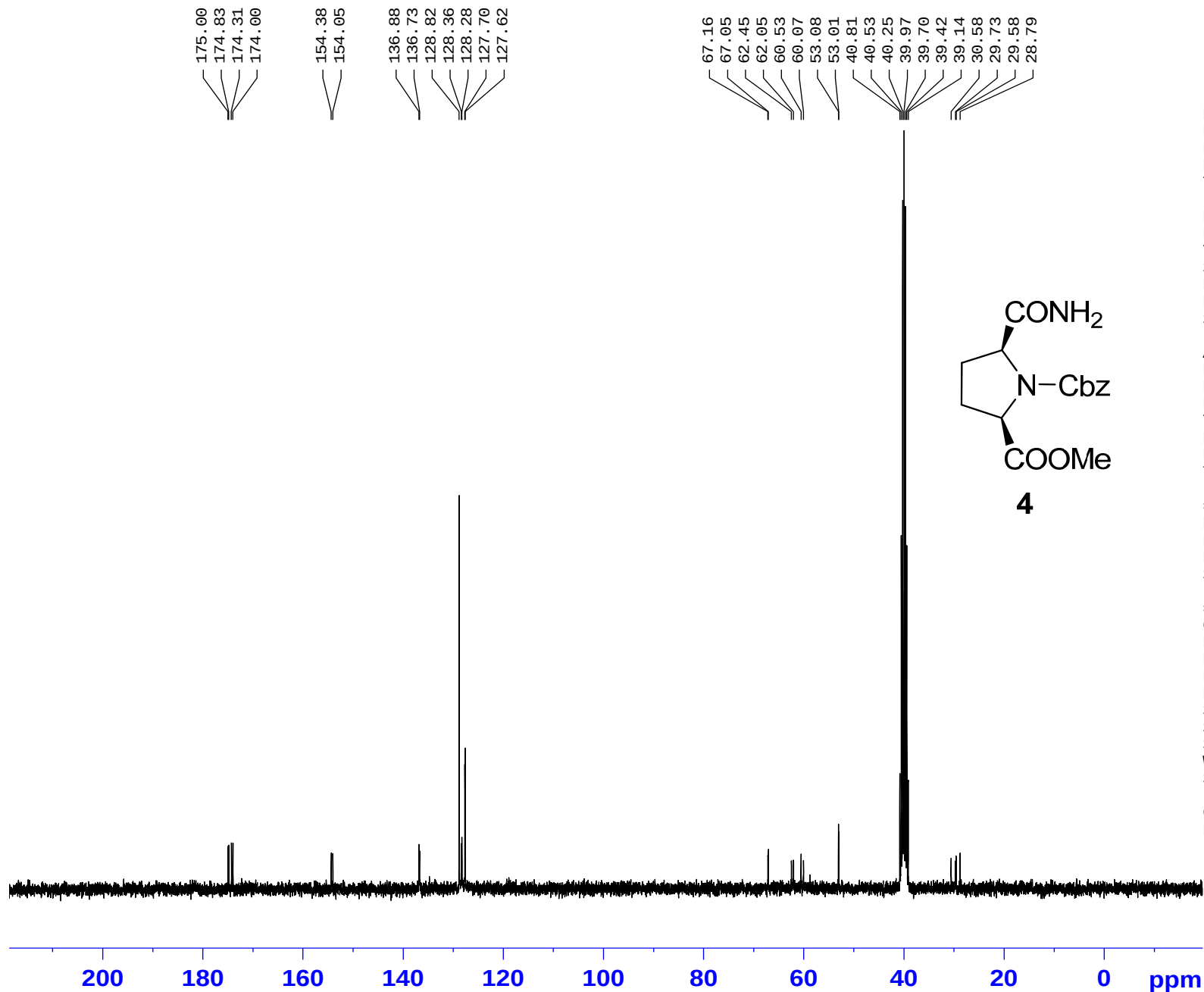


Current Data Parameters
NAME cp-p13 375K
EXPNO 397
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110326
Time 18.48
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 0
SWH 6188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953587 sec
RG 8192
DW 80.800 usec
DE 6.00 usec
TE 375.1 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 13.80 usec
PL1 3.00 dB
SF01 300.1318534 MHz

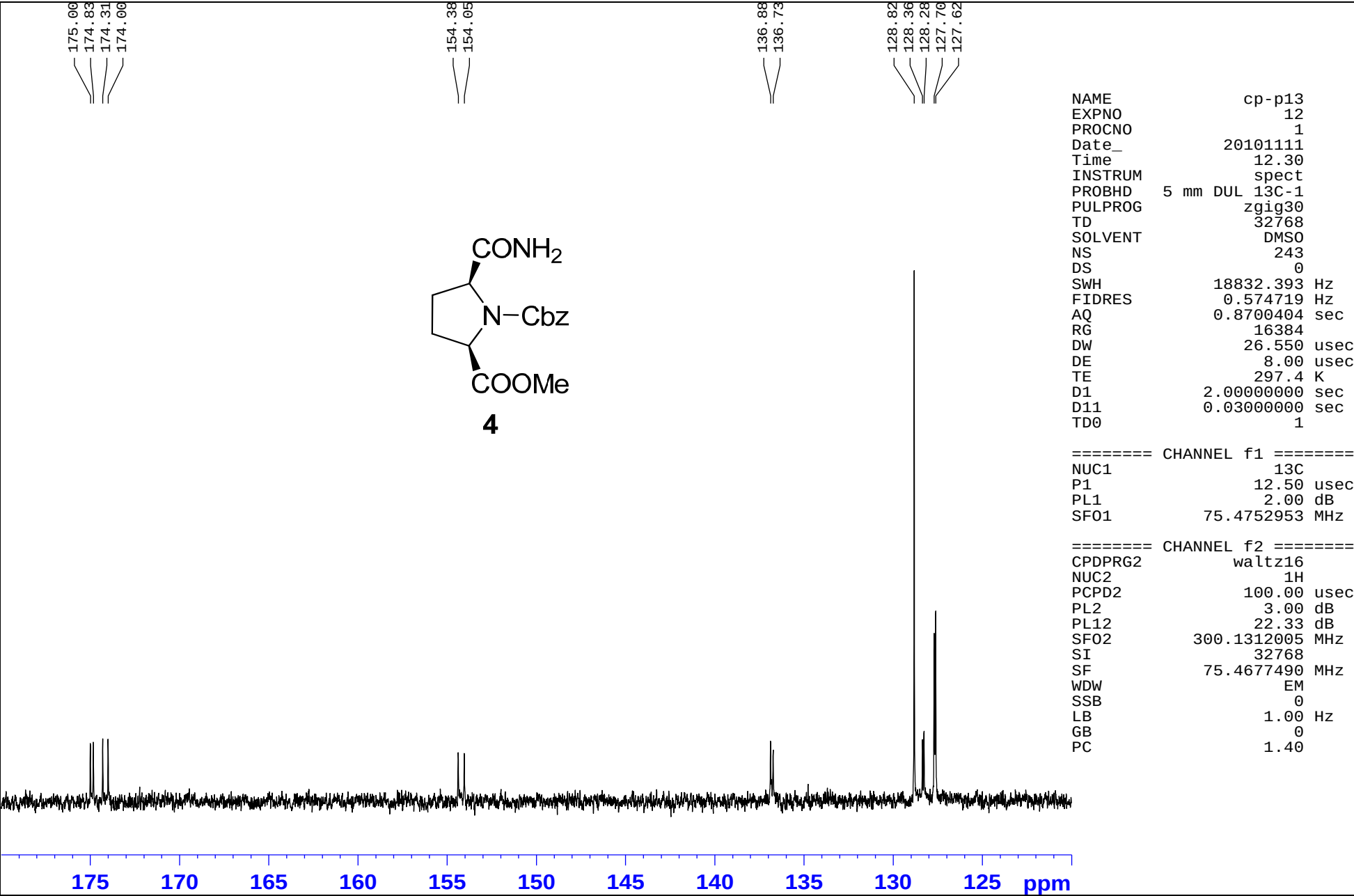
F2 - Processing parameters
SI 32768
SF 300.1300114 MHz
WDW EM
SSB 0
LB 0.20 Hz
GB 0
PC 1.00



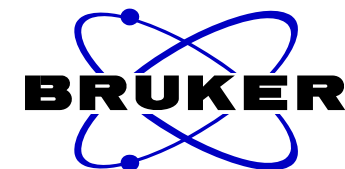
NAME cp-p13
EXPNO 12
PROCNO 1
Date_ 20101111
Time 12.30
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgig30
TD 32768
SOLVENT DMSO
NS 243
DS 0
SWH 18832.393 Hz
FIDRES 0.574719 Hz
AQ 0.8700404 sec
RG 16384
DW 26.550 usec
DE 8.00 usec
TE 297.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



cp-p35 375K

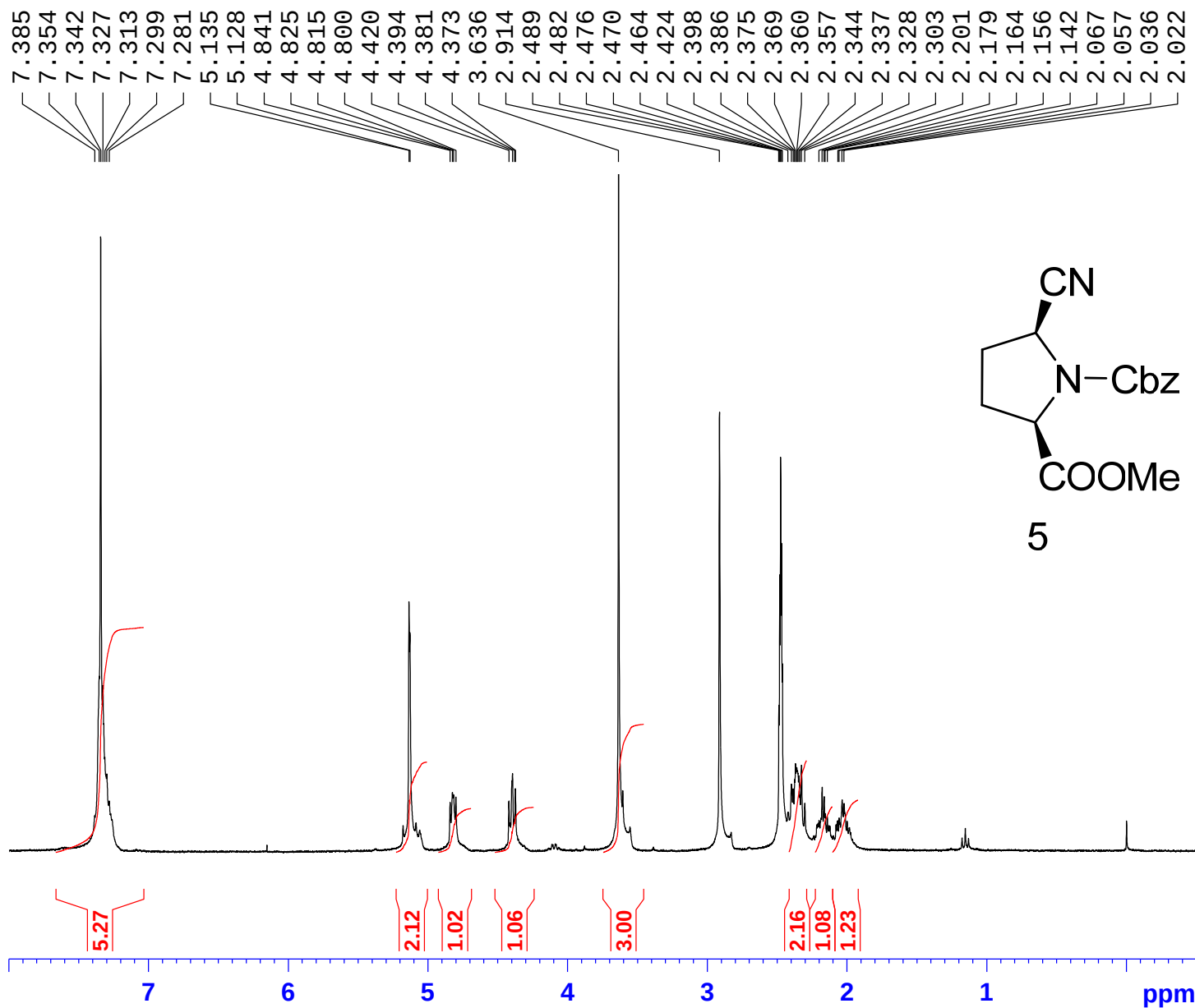


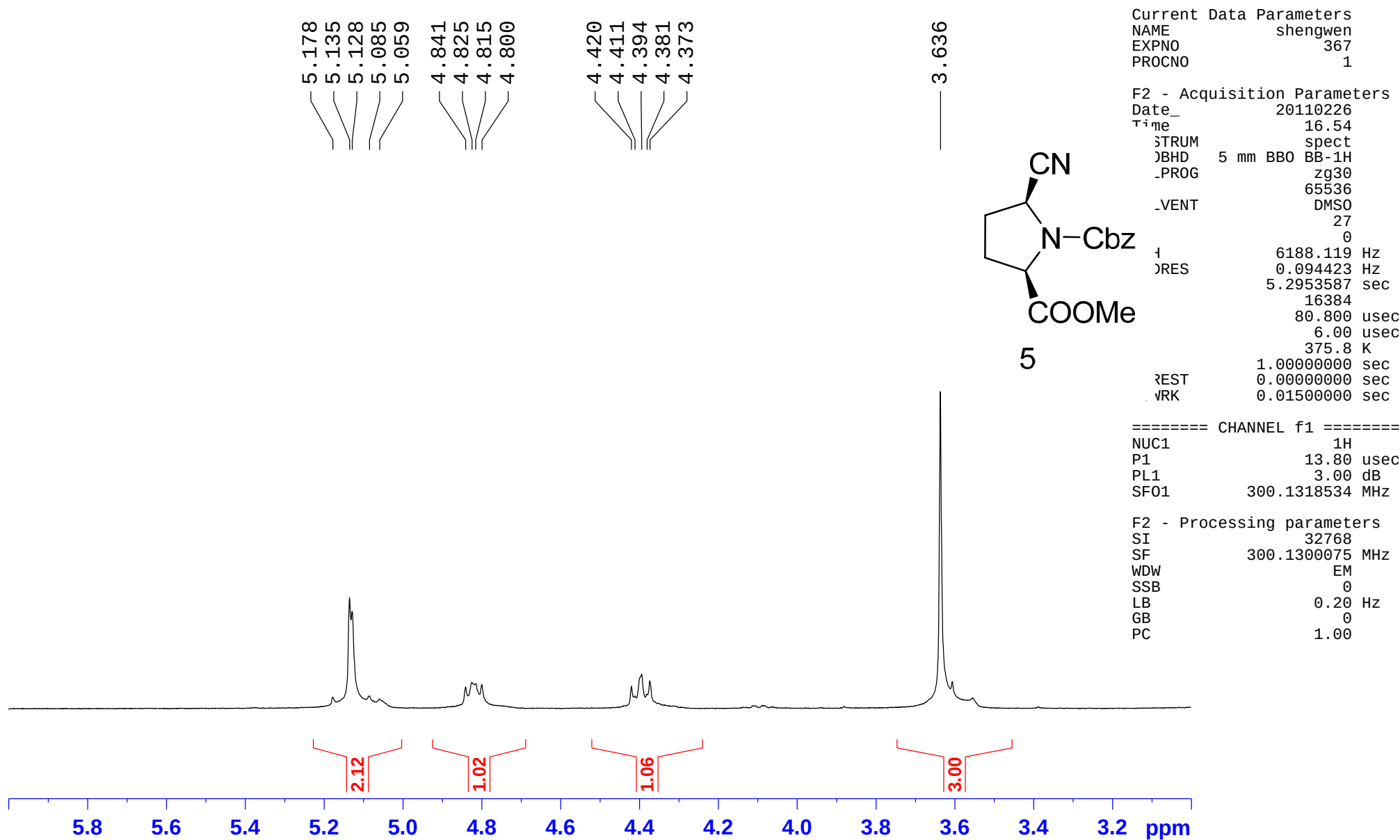
Current Data Parameters
NAME shengwen
EXPNO 367
PROCNO 1

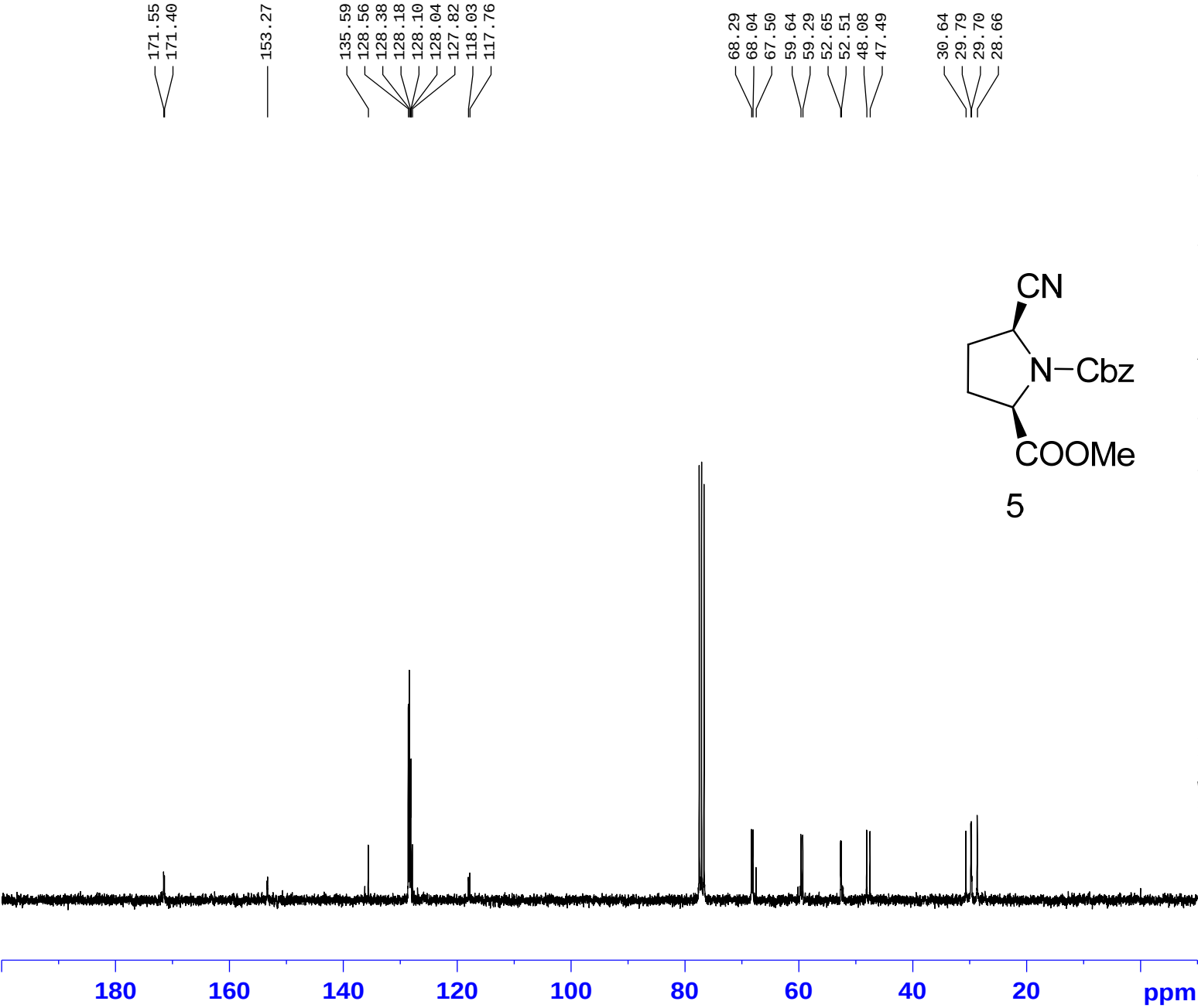
F2 - Acquisition Parameters
Date_ 20110226
Time 16.54
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 27
DS 0
SWH 6188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953587 sec
RG 16384
DW 80.800 usec
DE 6.00 usec
TE 375.8 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 13.80 usec
PL1 3.00 dB
SF01 300.1318534 MHz

F2 - Processing parameters
SI 32768
SF 300.1300075 MHz
WDW EM
SSB 0
LB 0.20 Hz
GB 0
PC 1.00



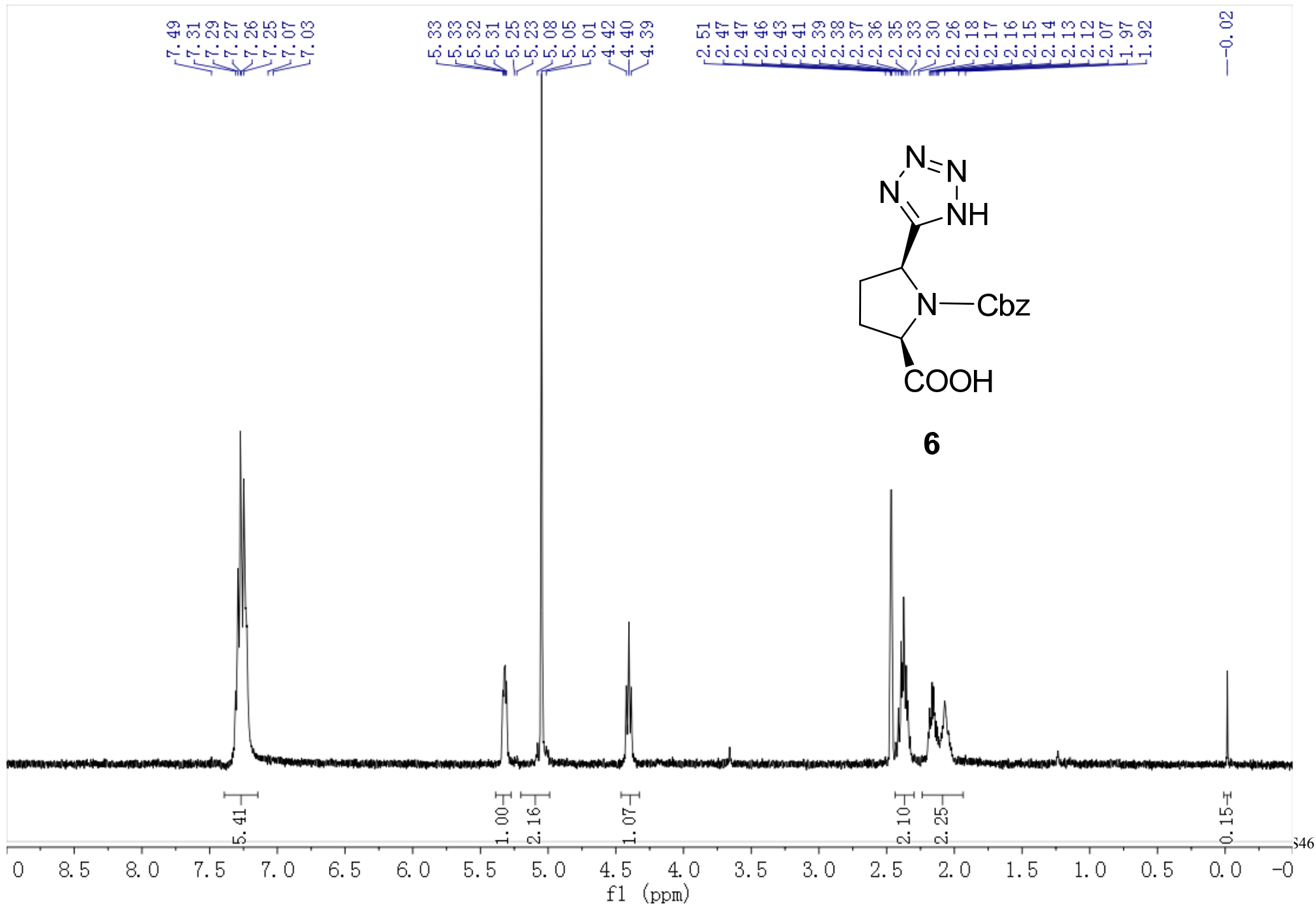


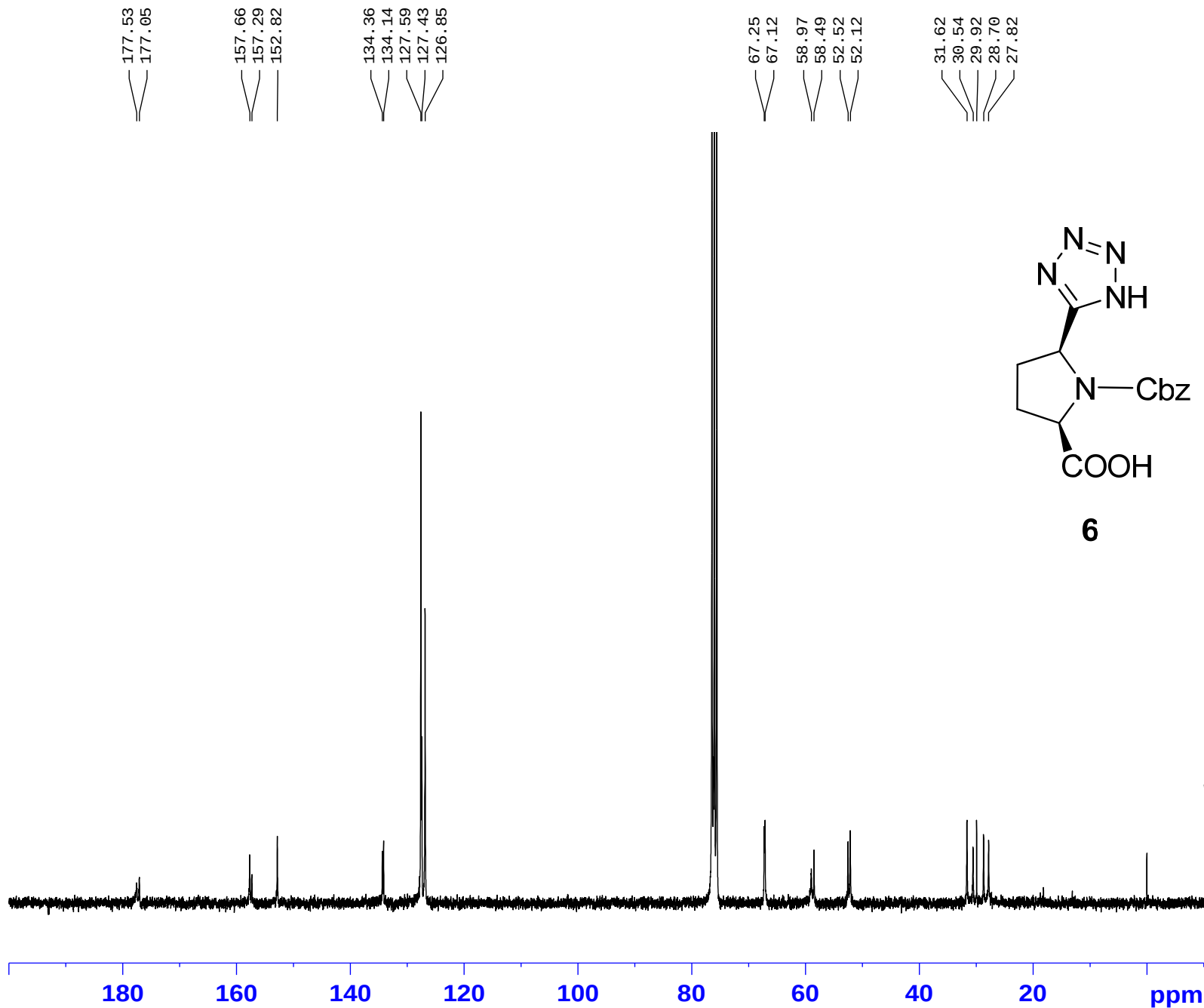


NAME cp-p35
EXPNO 11
PROCNO 1
Date_ 20100818
Time 20.20
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 398
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 16384
DW 27.800 usec
DE 8.00 usec
TE 301.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

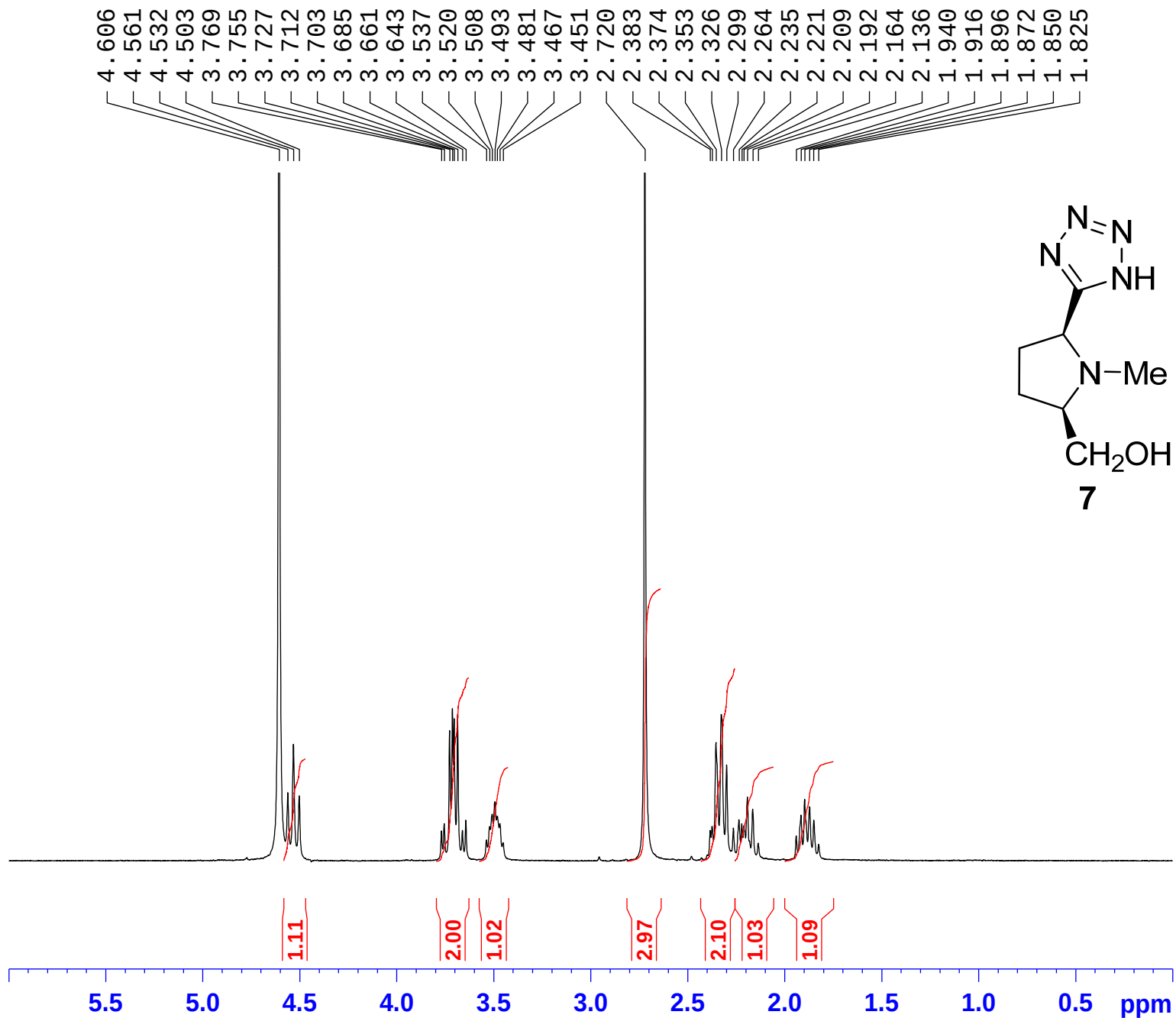




NAME cp-p803
EXPNO 20
PROCNO 1
Date_ 20101220
Time 8.20
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
IS 9544
JS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 9195.2
W 27.800 usec
DE 8.00 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
D0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4678257 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



Current Data Parameters
NAME cp-p82
EXPNO 80
PROCNO 1

F2 - Acquisition Parameters

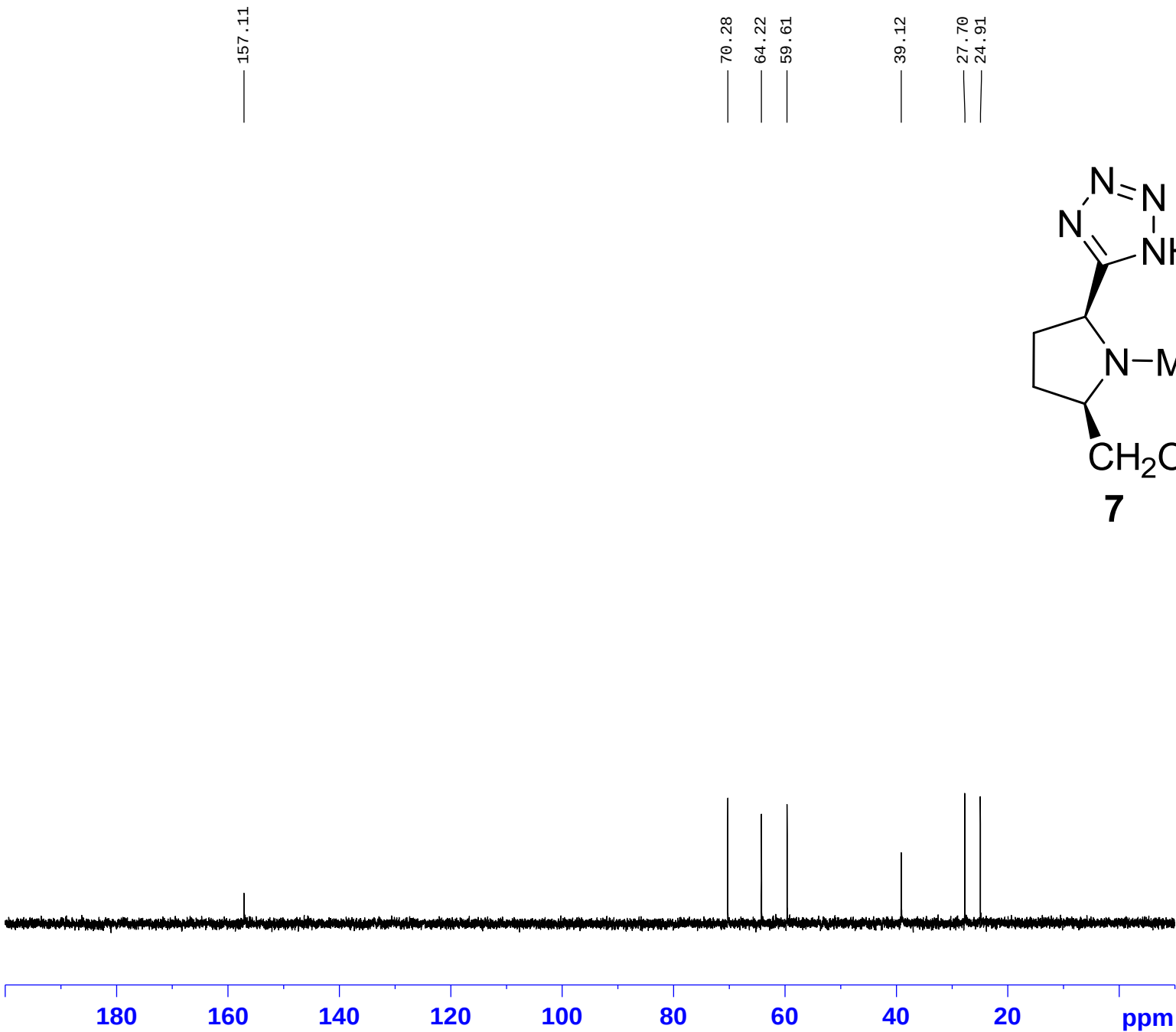
Date_ 20111105
Time 0.35
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT D2O
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 256
DW 55.600 usec
DE 8.00 usec
TE 296.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====

NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters

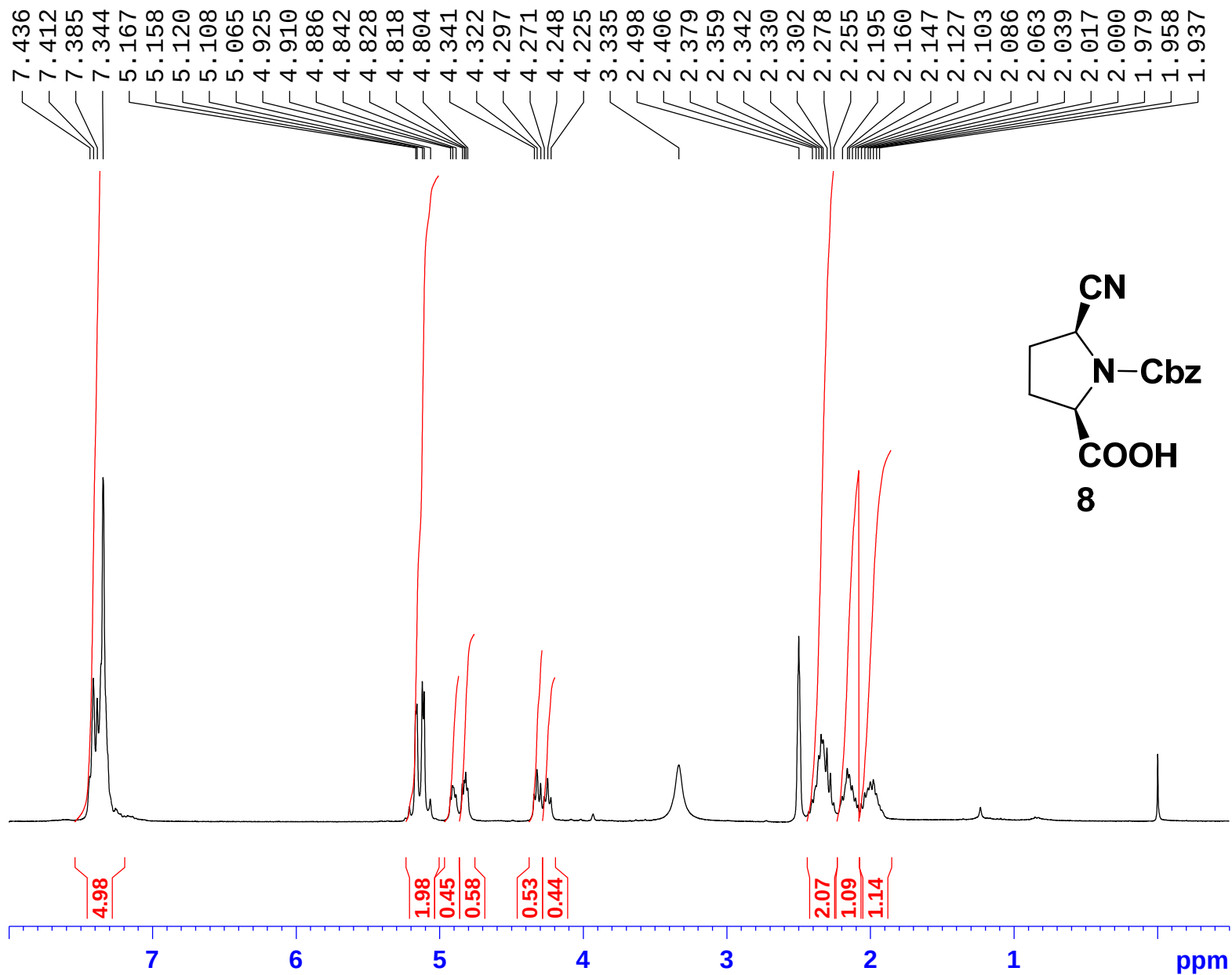
SI 32768
SF 300.1300268 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME cp-p82
EXPNO 11
PROCNO 1
Date_ 20111104
Time 10.54
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
PD 65536
SOLVENT D2O
DS 26
OS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 11585.2
AW 27.800 usec
DE 6.50 usec
TE 298.0 K
SI 2.00000000 sec
SI1 0.03000000 sec
SI0 1

===== CHANNEL f1 =====
JUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

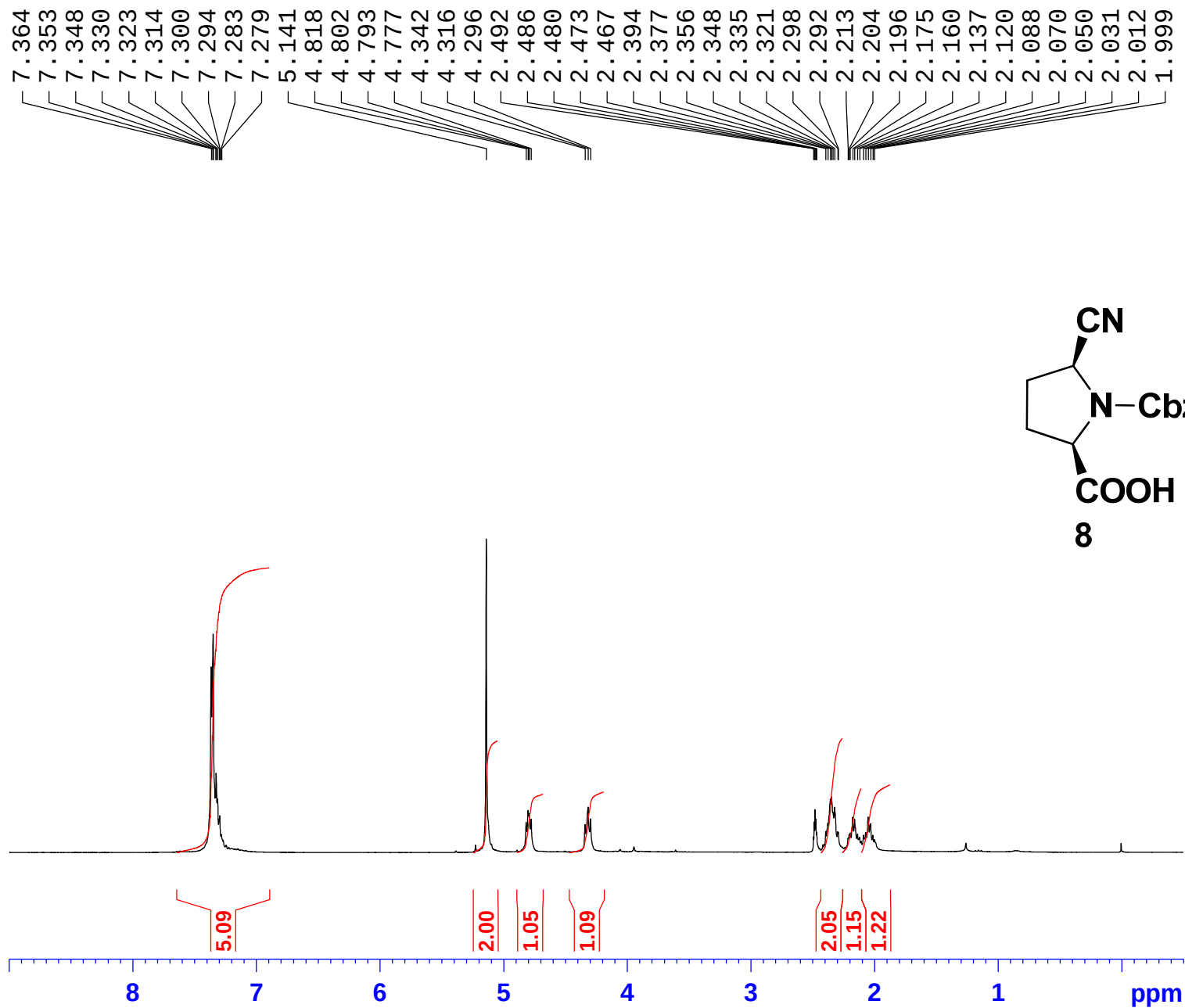


Current Data Parameters
NAME cp-p90
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111022
Time 10.55
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 256
DW 55.600 usec
DE 8.00 usec
TE 297.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300012 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

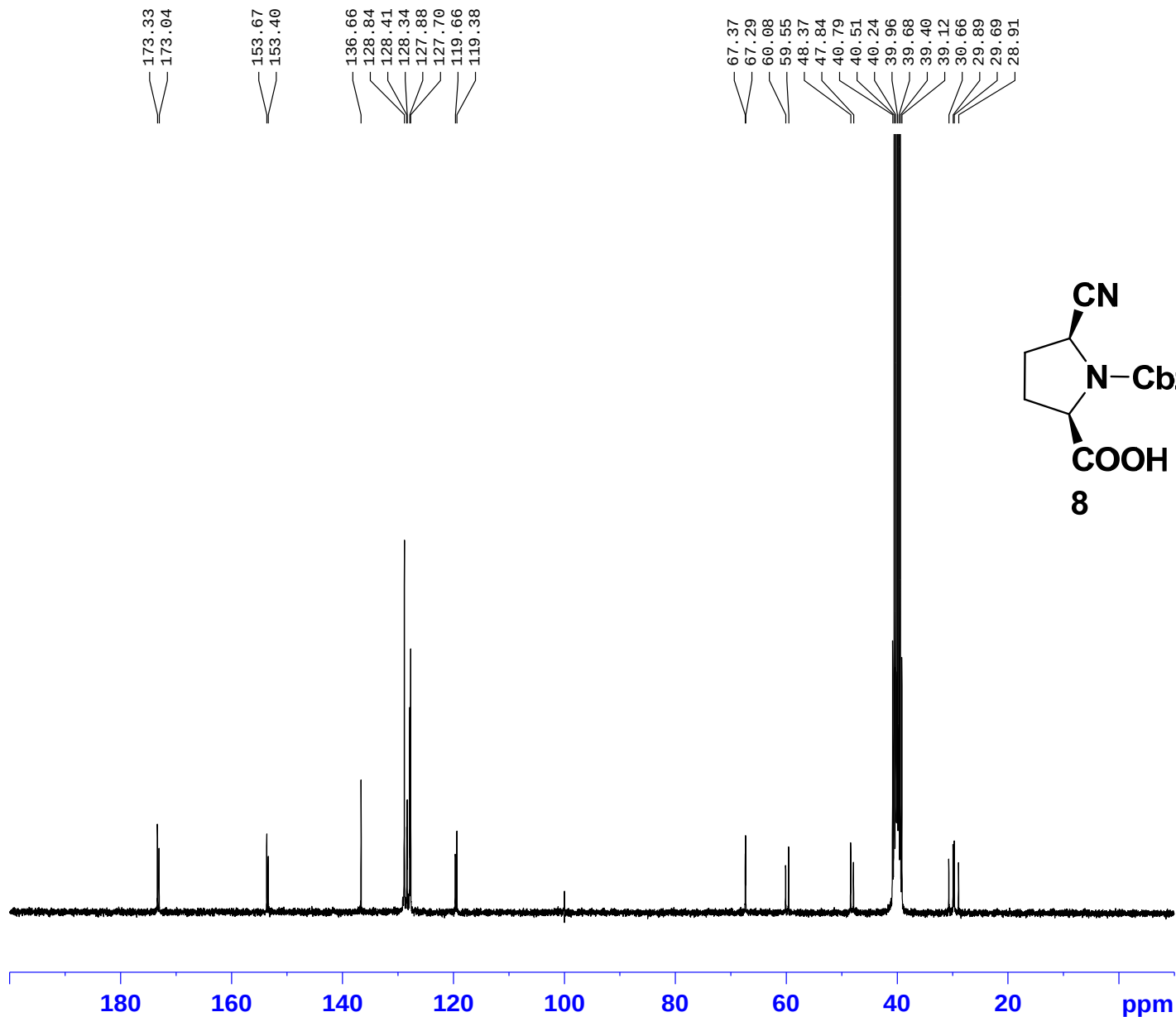


Current Data Parameters
NAME p90 375K
EXPNO 415
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111031
Time 17.02
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 2048
DW 55.600 usec
DE 6.00 usec
TE 375.1 K
D1 2.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.80 usec
PL1 3.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters
SI 32768
SF 300.1300071 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



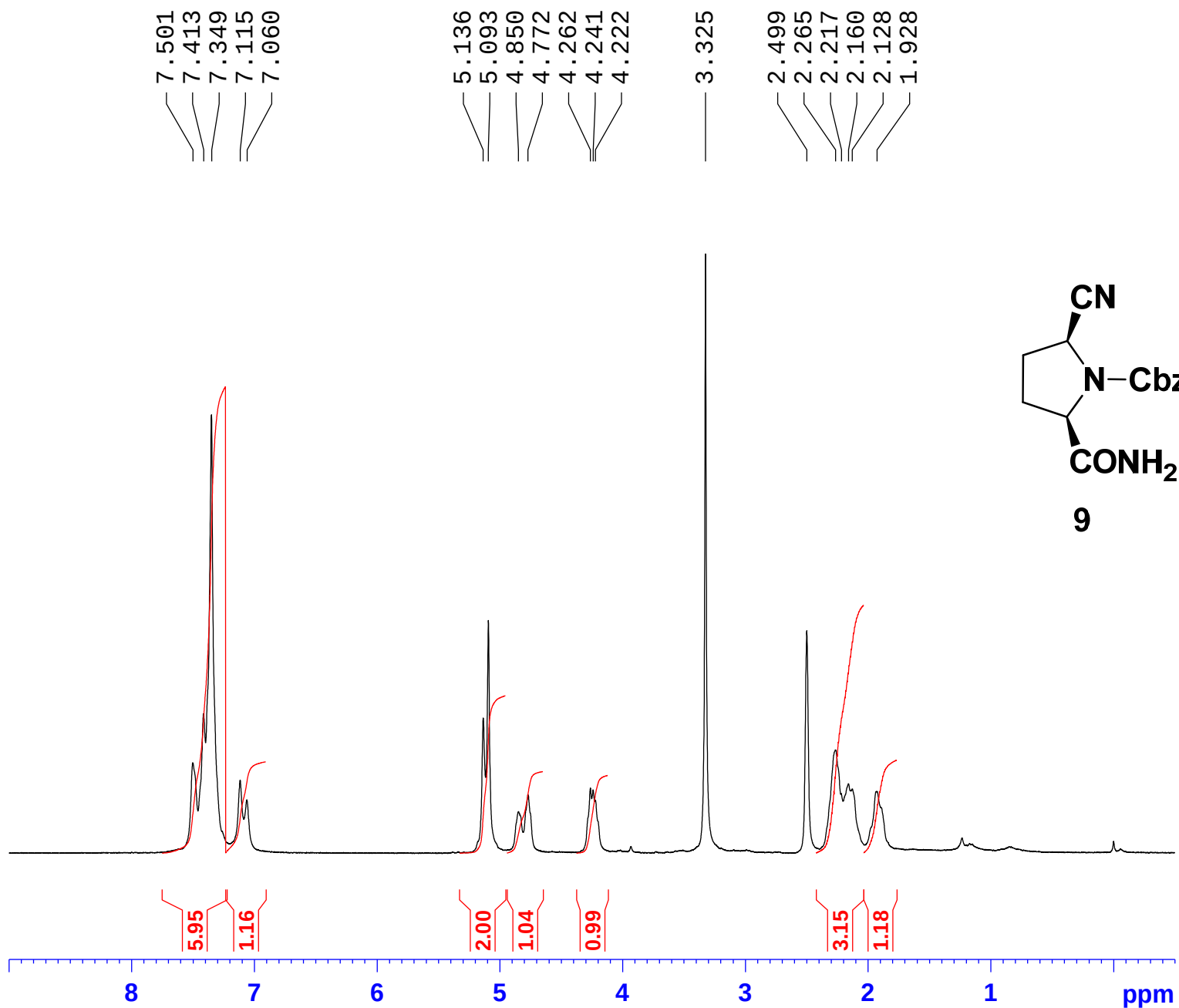
Current Data Parameters
NAME cp-p90
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111022
Time 11.25
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgig30
TD 32768
SOLVENT DMSO
NS 3216
DS 0
SWH 18832.393 Hz
FIDRES 0.574719 Hz
AQ 0.8700404 sec
RG 11585.2
DW 26.550 usec
DE 8.00 usec
TE 297.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

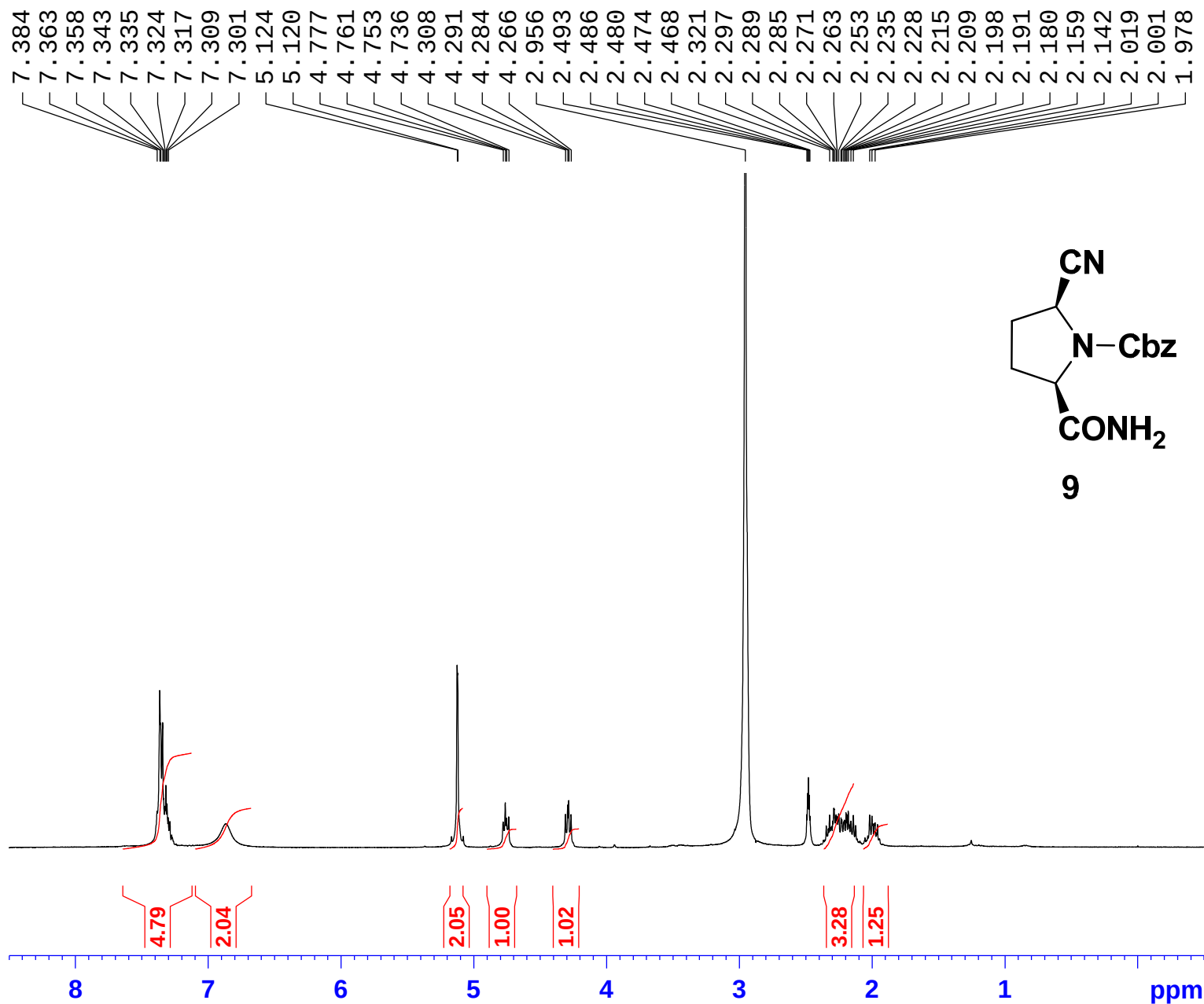
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



NAME cp-p91
EXPNO 20
PROCNO 1
Date_ 20111103
Time 19.57
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 256
DW 55.600 usec
DE 8.00 usec
TE 298.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz
SI 32768
SF 300.1300009 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME shengwen
EXPNO 424
PROCNO 1

F2 - Acquisition Parameters

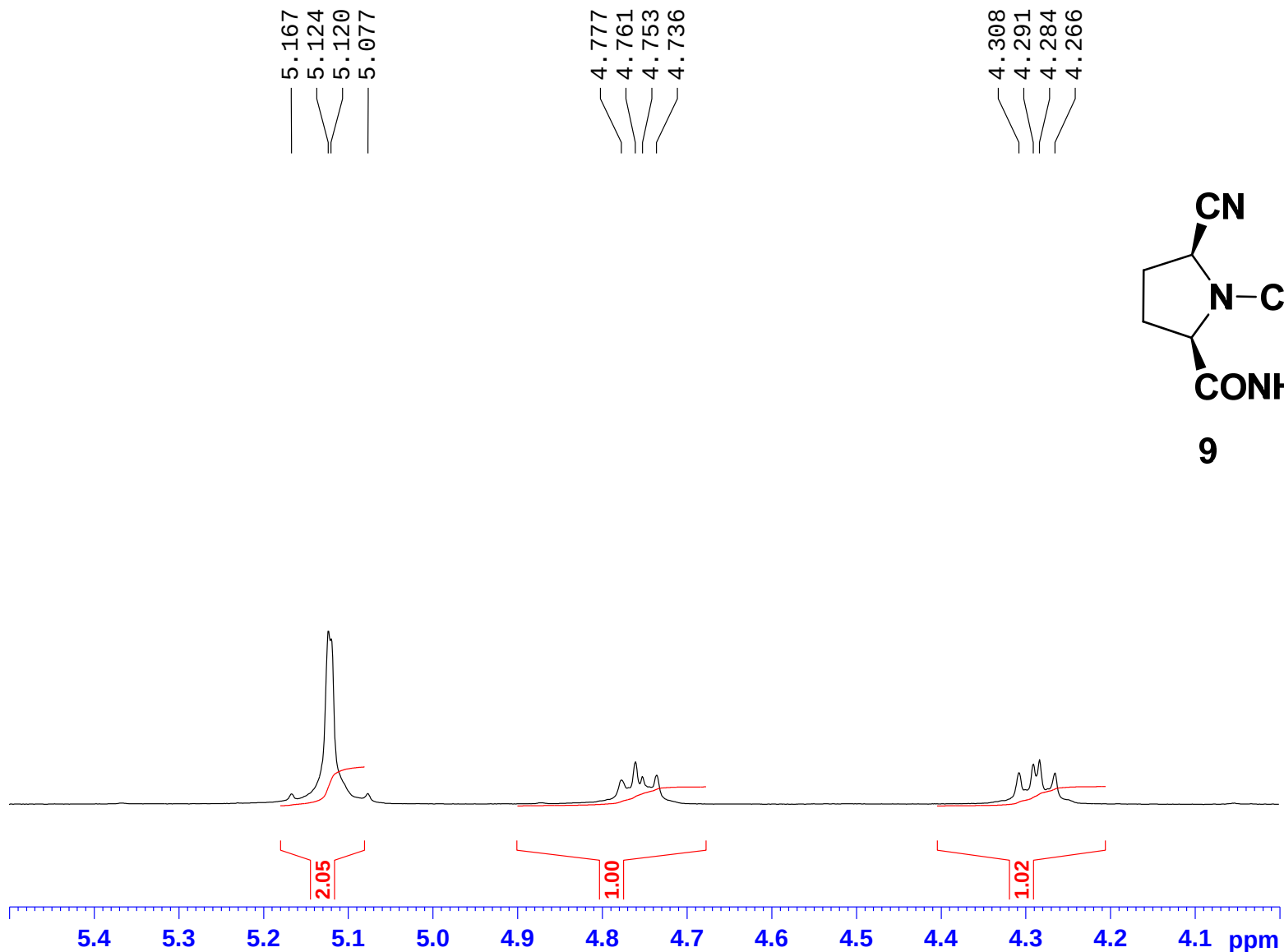
Date_ 20111110
Time 8.35
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 2048
DW 55.600 usec
DE 6.00 usec
TE 372.9 K
D1 2.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====

NUC1 1H
P1 13.80 usec
PL1 3.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters

SI 32768
SF 300.1300071 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0

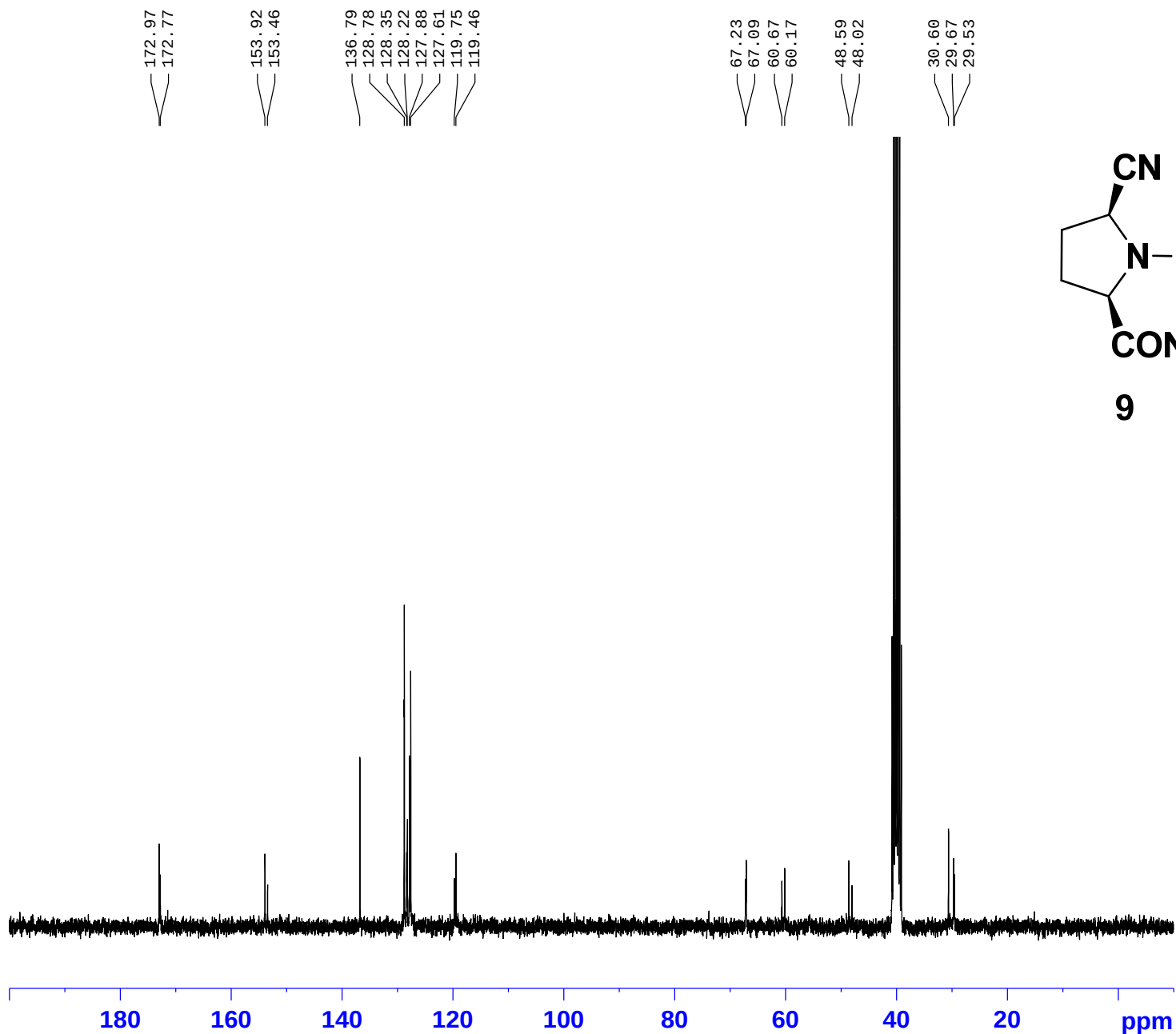


Current Data Parameters
NAME shengwen
EXPNO 424
PROCNO 1

F2 - Acquisition Parameters
:e_ 20111110
ie 8.35
;TRUM spect
;BHD 5 mm BBO BB-1H
;PROG zg30
32768
;VENT DMSO
16
0
f 8992.806 Hz
RES 0.274439 Hz
1.8219508 sec
2048
55.600 usec
6.00 usec
372.9 K
D1 2.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 13.80 usec
PL1 3.00 dB
SF01 300.1315007 MHz

F2 - Processing parameters
SI 32768
SF 300.1300071 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0



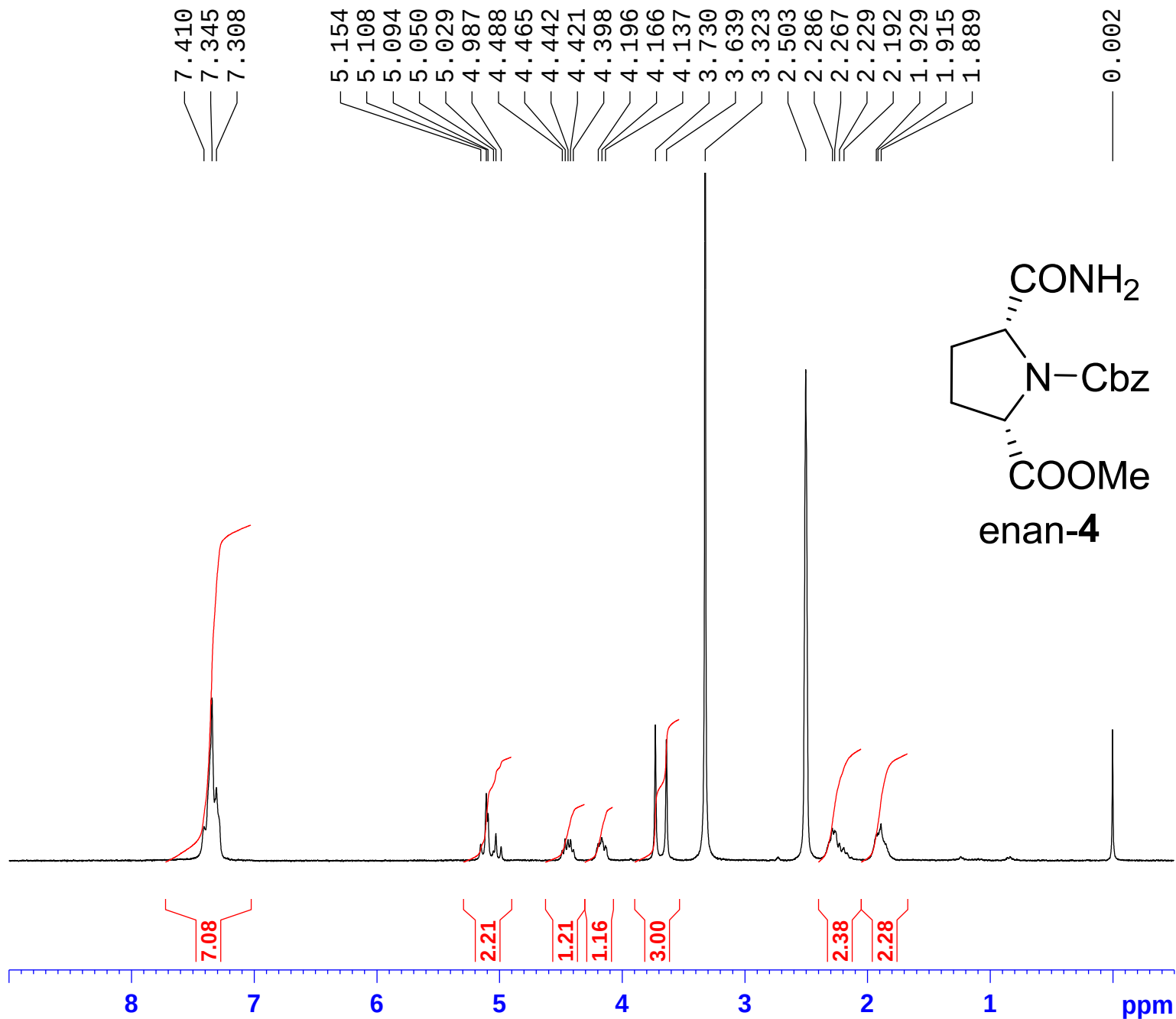
Current Data Parameters
NAME cp-p91
EXPNO 13
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111103
Time 11.51
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgig30
TD 32768
SOLVENT DMSO
NS 1119
DS 0
SWH 18832.393 Hz
FIDRES 0.574719 Hz
AQ 0.8700404 sec
RG 256
DW 26.550 usec
DE 8.00 usec
TE 297.8 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
SF02 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

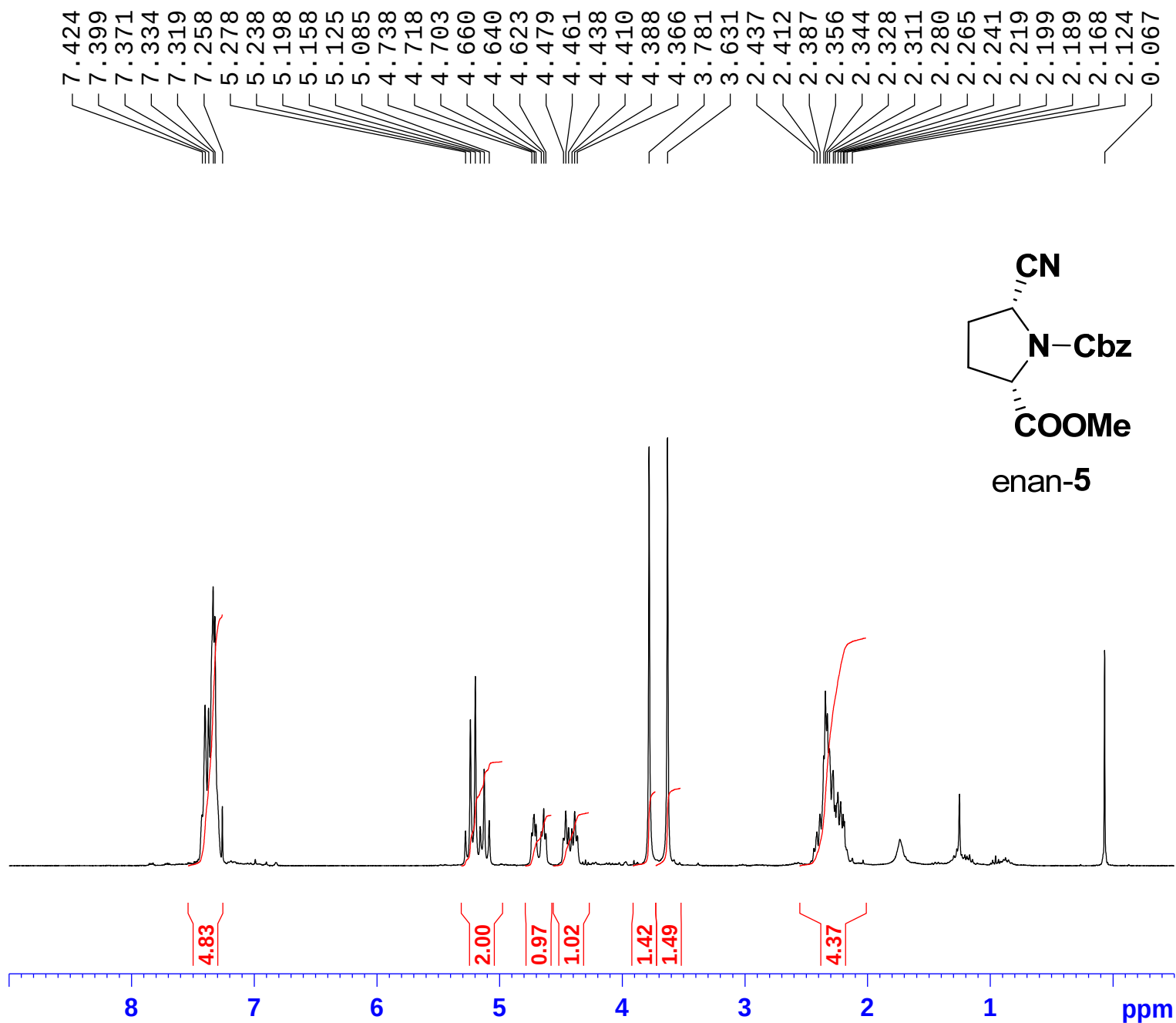


Current Data Parameters
NAME cp-p13e
EXPNO 190
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111024
Time 15.49
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 512
DW 55.600 usec
DE 8.00 usec
TE 297.8 K
D1 2.00000000 sec
TD0 1

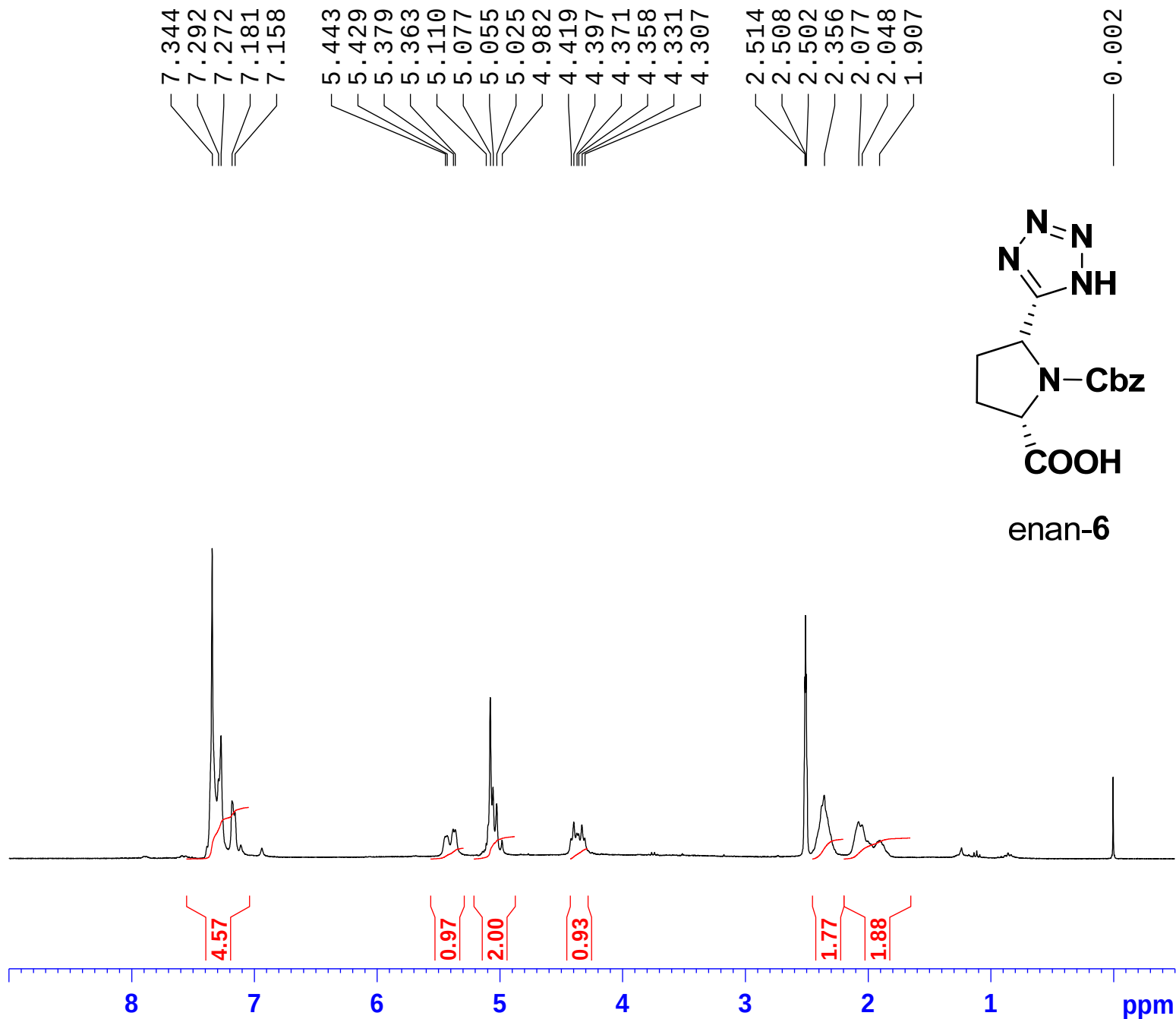
===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1318008 MHz

F2 - Processing parameters
SI 32768
SF 300.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME cp-p35e
EXPNO 10
PROCNO 1
Date_ 20111104
Time 19.21
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 181
DW 55.600 usec
DE 8.00 usec
TE 297.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz
SI 32768
SF 300.1300266 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

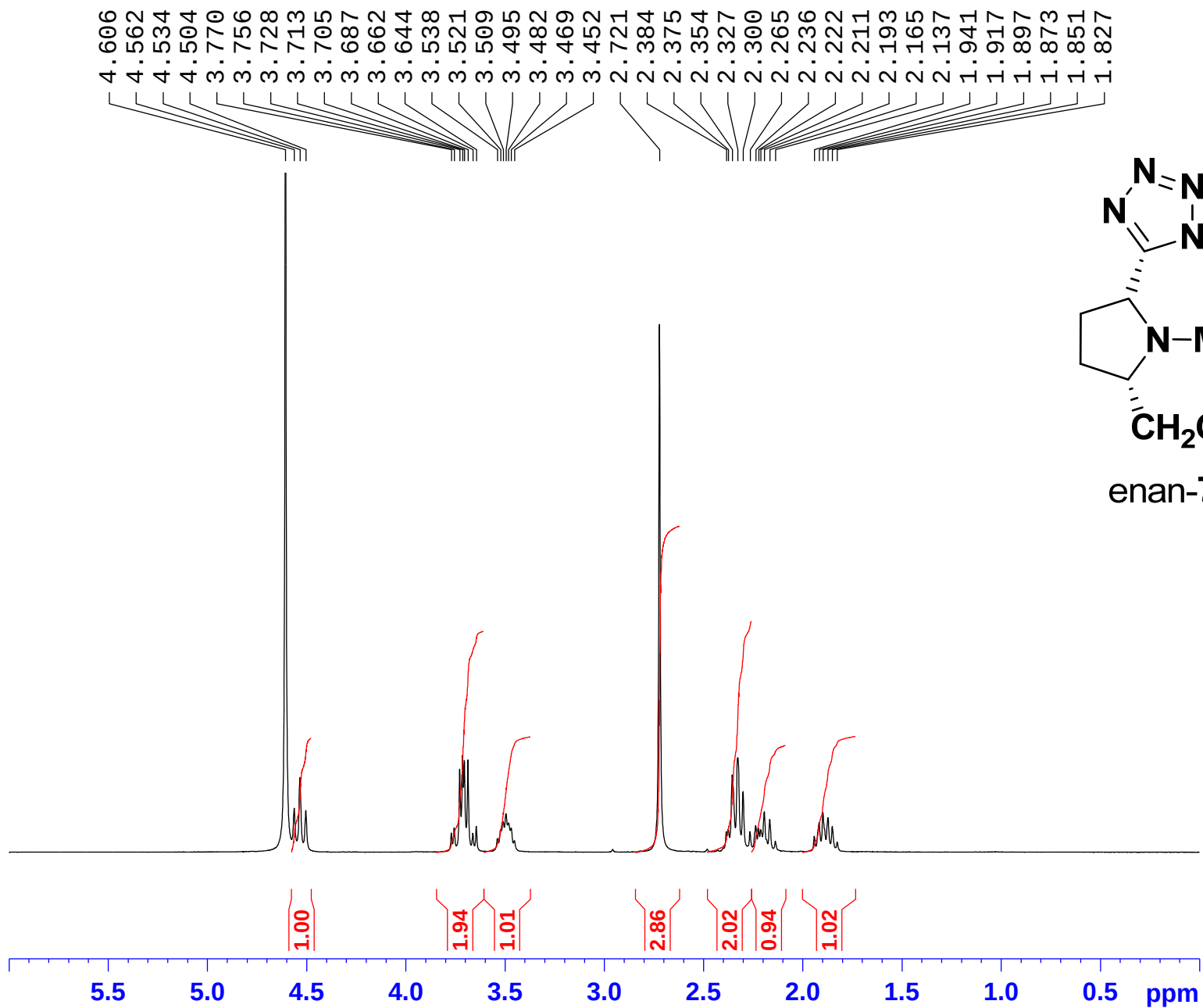


Current Data Parameters
NAME cp-p80-3e
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111105
Time 1.57
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 256
DW 55.600 usec
DE 8.00 usec
TE 296.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1299984 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME cp-p82e
EXPNO 14
PROCNO 1

F2 - Acquisition Parameters

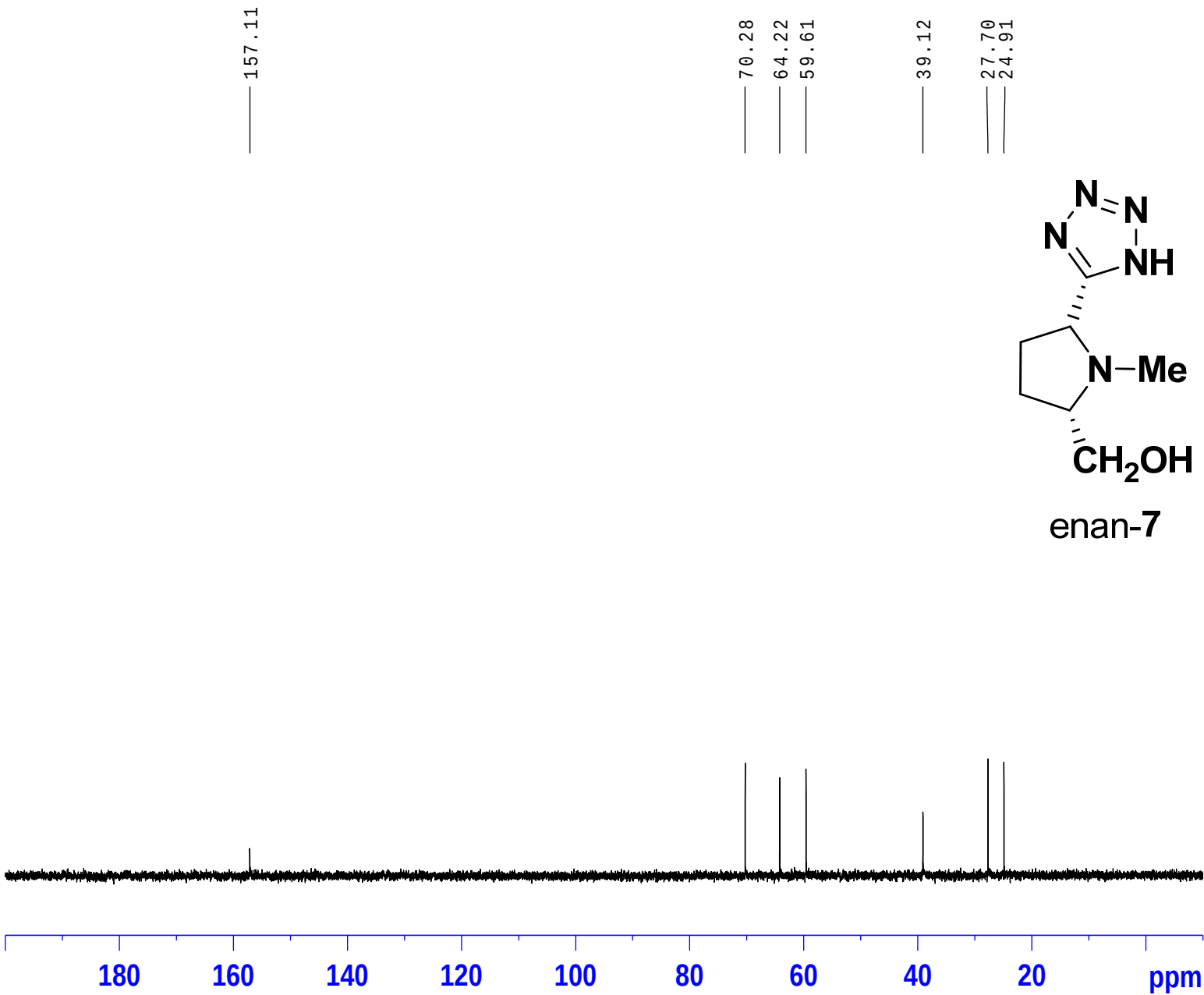
te_ 20111105
me 0.32
STRUM spect
OBHD 5 mm DUL 13C-1
LPROG zg30
65536
LVENT D20
16
0
H 8992.806 Hz
DRES 0.137219 Hz
3.6438515 sec
256
55.600 usec
8.00 usec
296.9 K
1.00000000 sec
1

==== CHANNEL f1 =====

C1 1H
10.80 usec
PL1 3.00 dB
SF01 300.1324010 MHz

F2 - Processing parameters

SI 32768
SF 300.1300268 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME cp-p82e
EXPNO 11
PROCNO 1
Date_ 20111103
Time 10.54
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT D2O
NS 26
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 11585.2
DW 27.800 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SF01 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SF02 300.1312005 MHz
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40