

## **Electronic Supplementary Information**

### **Modular synthesis of optically active lactones by Ru-catalyzed asymmetric allylic carboxylation and ring-closing metathesis reaction**

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## Experimental Procedures

### General.

All reactions were carried out under argon atmosphere using standard Schlenk techniques, and the workup was performed in air.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded on Varian Mercury 300, JEOL JNM-GSX 400, JEOL JNM-ECS 400 and JEOL JNM-ECA 500 spectrometers. Enantiomeric excess was determined by HPLC analysis using Hitachi L-2130/L-2455 and Shimadzu LC -10/SPD-10AV equipped with DAICEL Chiralcel OJ-H, OD-H, OB-H and Chiralpak AS-H columns. Optical rotation was measured on JASCO DIP-100. HRMS measurements were carried out on Thermo Fisher Scientific LTQ-Orbitrap XL.

### Materials.

All solvents used for reactions were passed through purification columns just before use. Planar-chiral Cp'Ru complex **1a-1c** were synthesized as reported previously.<sup>1,2</sup> Cinnamyl chloride **2a** was purchased from TCI. Allylic chlorides **2b-2d**, **2f** and **2g** were prepared by Corey-Kim chlorination of the corresponding allylic alcohols,<sup>3</sup> whereas **2e** was prepared by the method according to that for analogous bromide.<sup>4</sup> All allylic chlorides were purified by distillation (**2a**, **2c**, **2f** and **2g**) or recrystallization from *n*-hexane (**2b**, **2d** and **2e**) prior to use. Unsaturated carboxylic acids were available from commercial source and used without any purification. Sodium carboxylate was synthesized by the reaction of the corresponding unsaturated carboxylic acid with  $\text{Na}_2\text{CO}_3$  in acetone/water (*ca.* v/v = 1/1). 2nd generation Grubbs' catalyst (**G-II**) was purchased from Sigma-Aldrich.

### Standard Procedure for Asymmetric Allylic Carboxylation.

#### **Method A.**

To a THF solution (1 mL) of (*R*)-**1c** (5  $\mu\text{mol}$ , 1 mol%), allylic chloride (1.0 mmol) and  $\text{Na}_2\text{CO}_3$  (1.5 mmol) was added a THF solution (1 mL) of unsaturated carboxylic acid (0.5 mmol), and the resulting mixture was stirred for 4 h at 25 °C. After dilution with *n*-hexane, the insoluble parts were filtered off through Celite and the filtrate was concentrated under reduced pressure. The residue was purified by  $\text{SiO}_2$  column chromatography using a mixture of *n*-hexane and  $\text{Et}_2\text{O}$  (v/v = 20/1) as the eluent. Evaporation of the solvent gave branched allylic ester as colorless oil.

#### **Method B.**

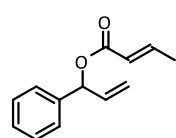
To a THF solution (1 mL) of (*R*)-**1c** (5  $\mu\text{mol}$ , 1 mol%), allylic chloride (1.0 mmol) and  $\text{Na}_2\text{CO}_3$  (1.5 mmol) was added a THF solution (1 mL) of unsaturated carboxylic acid (0.5 mmol), and the resulting mixture was stirred for 4 h at 25 °C. After dilution with *n*-hexane, the insoluble parts were filtered off through Celite and the filtrate was concentrated under reduced pressure. Acetone (5 mL) and diethylamine (5 mL) were added, and the mixture was stirred for 6 h at room temperature. After removal of the solvent under reduced pressure, the resulting crude material was purified by  $\text{SiO}_2$  column chromatography to give branched allylic ester as colorless oil.

## Method C.

To a THF solution (2 mL) of (*R*)-**1c** (15  $\mu$ mol, 3 mol%) and allylic chloride (2.5 mmol) was added sodium carboxylate (0.5 mmol) portionwise, and the resulting mixture was stirred for 4 h at 25 °C. Workup was performed by the same method as **Method A**.

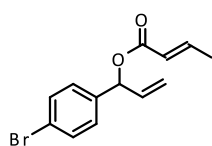
## Characterization of Allylic Esters.

### (*E*)-1-phenylallyl but-2-enoate (**4ab**, Method A)



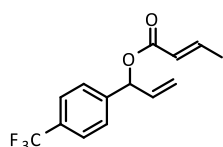
$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz):  $\delta$  7.40–7.26 (m, 5H, Ph), 7.02 (dq, 1H,  $J = 15.5$ , 7.0 Hz,  $\text{CHCH}_3$ ), 6.32 (ddd, 1H,  $J = 5.8$ , 1.4, 1.4 Hz,  $\text{PhCH}$ ), 6.03 (ddd, 1H,  $J = 17.0$ , 10.3, 5.8 Hz,  $\text{CH=}$ ), 5.91 (dq, 1H,  $J = 15.5$ , 1.7 Hz,  $\text{COCH=}$ ), 5.30 (ddd, 1H,  $J = 17.0$ , 1.4, 1.4 Hz,  $=\text{CH}_2$ ), 5.24 (ddd, 1H,  $J = 10.3$ , 1.4, 1.4 Hz,  $=\text{CH}_2$ ), 1.88 (dd, 3H,  $J = 7.0$ , 1.7 Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz):  $\delta$  165.3, 145.1, 139.1, 136.4, 128.5, 128.0, 127.1, 122.7, 116.7, 75.8, 17.9. HPLC analysis: Chiralcel OJ-H column, *n*-hexane/*i*PrOH = 98/2 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 20.1$  min, minor enantiomer  $t = 16.9$  min, 97% ee.  $[\alpha]_{\text{D}}^{28} = -38.2$  ( $c$  0.27,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{13}\text{H}_{14}\text{O}_2\text{Na}$   $[\text{M}+\text{Na}^+]$ : 225.0886, found:  $m/z = 225.0889$ .

### (*E*)-1-(4-bromophenyl)allyl but-2-enoate (**4bb**, Method A)



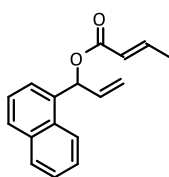
$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  7.50–7.43 (m, 2H, Ar), 7.26–7.21 (m, 2H, Ar), 7.02 (dq,  $J = 15.7$ , 6.9 Hz,  $=\text{CHCH}_3$ ), 6.28–6.23 (m, 1H,  $\text{ArCH}$ ), 5.98 (ddd, 1H,  $J = 17.7$ , 10.4, 5.8 Hz,  $\text{CH=}$ ), 5.90 (dq, 1H,  $J = 15.7$ , 1.7 Hz,  $\text{COCH=}$ ), 5.33–5.22 (m, 2H,  $=\text{CH}_2$ ), 1.89 (dd, 3H,  $J = 6.9$ , 1.7 Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  147.6, 131.8, 125.8, 124.0, 120.6, 118.4, 113.3, 113.0, 109.1, 75.4, 29.7. HPLC analysis: Chiralcel OD-H column, *n*-hexane/*i*PrOH = 100/1 (v/v), 0.5 mL/min, 220 nm; major enantiomer  $t = 13.2$  min, minor enantiomer  $t = 12.2$  min, 96% ee.  $[\alpha]_{\text{D}}^{29} = -21.3$  ( $c$  0.19,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{13}\text{H}_{13}\text{O}_2\text{BrNa}$   $[\text{M}+\text{Na}^+]$ : 302.9991, found:  $m/z = 302.9996$ .

### (*E*)-1-(4-trifluoromethylphenyl)allyl but-2-enoate (**4cb**, Method A)



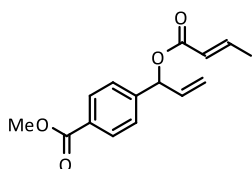
$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  7.61 (d, 2H,  $J = 8.3$  Hz, Ar), 7.47 (d, 2H,  $J = 8.3$  Hz, Ar), 7.04 (dq,  $J = 15.5$ , 6.9 Hz,  $=\text{CHCH}_3$ ), 6.36–6.31 (m, 1H,  $\text{ArCH}$ ), 6.00 (ddd, 1H,  $J = 17.3$ , 10.4, 5.9 Hz,  $\text{CH=}$ ), 5.92 (dq, 1H,  $J = 15.5$ , 1.7 Hz,  $\text{COCH=}$ ), 5.36–5.25 (m, 2H,  $=\text{CH}_2$ ), 1.90 (dd, 3H,  $J = 6.9$ , 1.7 Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz):  $\delta$  166.2, 145.8, 143.1 (q,  $J = 1.3$  Hz), 135.7, 130.2 (q,  $J = 32.5$  Hz), 127.3, 125.5 (q,  $J = 3.8$  Hz), 124.0 (q,  $J = 272.1$  Hz), 122.3, 117.6, 75.2, 17.9. HPLC analysis: Chiralcel OD-H column, *n*-hexane/*i*PrOH = 200/1 (v/v), 0.5 mL/min, 220 nm; major enantiomer  $t = 15.4$  min, minor enantiomer  $t = 13.9$  min, 98% ee.  $[\alpha]_{\text{D}}^{30} = -11.9$  ( $c$  0.28,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{14}\text{H}_{13}\text{O}_2\text{F}_3\text{Na}$   $[\text{M}+\text{Na}^+]$ : 293.0765, found:  $m/z = 293.0760$ .

**(E)-1-(naphthalene-1-yl)allyl but-2-enoate (4db, Method B)**



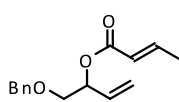
$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  8.16–8.11 (m, 1H, Ar), 7.89–7.79 (m, 2H, Ar), 7.63–7.57 (m, 1H, Ar), 7.56–7.42 (m, 3H, Ar), 7.10–6.98 (m, 2H, ArCH and  $=\text{CHCH}_3$ ), 6.21 (ddd, 1H,  $J = 17.0, 9.8, 5.2$  Hz, CH=), 5.93 (dq, 1H,  $J = 15.6, 1.7$  Hz, COCH=), 5.36–5.25 (m, 2H,  $=\text{CH}_2$ ), 1.88 (dd, 3H,  $J = 6.9, 1.7$  Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  147.7, 131.6, 124.2, 123.1, 122.5, 112.0, 118.5, 118.4, 116.3, 115.9, 115.6, 115.5, 114.4, 113.4, 109.0, 74.0, 29.7. HPLC analysis: Chiralcel OD-H column, *n*-hexane/*i*PrOH = 200/1 (v/v), 0.5 mL/min, 220 nm; major enantiomer  $t = 30.7$  min, minor enantiomer  $t = 28.9$  min, 96% ee.  $[\alpha]_{\text{D}}^{29} = -24.4$  ( $c$  0.27,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{17}\text{H}_{16}\text{O}_2\text{Na}$  [ $\text{M}+\text{Na}^+$ ]: 275.1043, found:  $m/z = 275.1047$ .

**(E)-methyl 4-(1-(but-2-enyloxy)allyl)benzoate (4eb, Method B)**



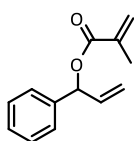
$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz):  $\delta$  8.04–7.99 (m, 2H, Ar), 7.45–7.39 (m, 2H, Ar), 7.04 (dq,  $J = 15.7, 6.9$  Hz,  $=\text{CHCH}_3$ ), 6.37–6.31 (m, 1H, ArCH), 6.00 (ddd, 1H,  $J = 17.7, 10.4, 5.8$  Hz, CH=), 5.92 (dq, 1H,  $J = 15.7, 1.7$  Hz, COCH), 5.36–5.23 (m, 2H,  $=\text{CH}_2$ ), 3.91 (s, 3H,  $\text{OCH}_3$ ), 1.90 (dd, 3H,  $J = 6.9, 1.7$  Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  148.7, 147.5, 131.9, 130.6, 124.0, 119.2, 116.9, 113.3, 109.4, 75.6, 57.0, 29.8, 16.2. HPLC analysis: Chiralcel OJ-H column, *n*-hexane/*i*PrOH = 98/2 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 39.8$  min, minor enantiomer  $t = 22.1$  min. 95% ee.  $[\alpha]_{\text{D}}^{29} = -18.4$  ( $c$  0.28,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{15}\text{H}_{16}\text{O}_4\text{Na}$  [ $\text{M}+\text{Na}^+$ ]: 283.0941, found:  $m/z = 283.0945$ .

**(E)-1-(benzyloxy)but-3-en-2-yl but-2-enoate (4fb, Method A)**



$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  7.37–7.25 (m, 5H, Ph), 7.01 (dq, 1H,  $J = 15.7, 6.9$  Hz,  $=\text{CHCH}_3$ ), 5.93–5.91 (m, 2H, COCH= and CH=), 5.57–5.50 (m, 1H, CHCH=), 5.32 (ddd, 1H,  $J = 17.3, 1.5, 1.4$  Hz,  $=\text{CH}_2$ ), 5.23 (ddd, 1H,  $J = 10.7, 1.4, 1.2$  Hz,  $=\text{CH}_2$ ), 4.59 (d, 1H,  $J = 12.2$  Hz,  $\text{PhCH}_2$ ), 4.54 (d, 1H,  $J = 12.2$  Hz,  $\text{PhCH}_2$ ), 3.61 (dd, 1H,  $J = 10.7, 4.0$  Hz,  $\text{CH}_2\text{CH}$ ), 3.59 (dd, 1H,  $J = 10.7, 2.8$  Hz,  $\text{CH}_2\text{CH}$ ), 1.89 (dd, 3H,  $J = 6.9, 1.7$  Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz):  $\delta$  165.5, 144.9, 137.9, 133.5, 128.3, 127.5, 122.6, 117.6, 73.1, 72.8, 71.3, 17.9. One carbon signal could not be detected probably due to overlapping. HPLC analysis: Chiralcel OJ-H column, 2% *i*PrOH, 1.0 mL/min, 220 nm; major enantiomer  $t = 18.3$  min, minor enantiomer  $t = 15.1$  min. 90% ee.  $[\alpha]_{\text{D}}^{28} = -11.0$  ( $c$  0.22,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{15}\text{H}_{18}\text{O}_3\text{Na}$  [ $\text{M}+\text{Na}^+$ ]: 269.1148, found:  $m/z = 269.1152$ .

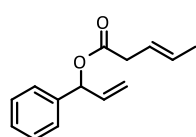
**(E)-1-phenylallyl methacrylate (4ac, Method A)**



$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz):  $\delta$  7.41–7.27 (m, 5H, Ph), 6.32 (ddd, 1H,  $J = 5.8, 1.7, 1.5$  Hz, PhCH), 6.22–6.17 (m, 1H,  $=\text{CH}_2$ ), 6.04 (ddd, 1H,  $J = 17.1, 10.5, 5.8$  Hz, CH=), 5.59 (dq, 1H,  $J = 1.6, 1.6$  Hz,  $\text{CCH}_3=\text{CH}_2$ ), 5.32 (ddd, 1H,  $J = 17.1, 1.7, 1.4$  Hz, CH= $\text{CH}_2$ ), 5.25 (ddd, 1H,  $J = 10.5, 1.5, 1.4$  Hz, CH= $\text{CH}_2$ ), 1.98 (dd, 3H,  $J = 1.6, 0.9$  Hz,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  166.3, 139.1, 136.5, 136.4, 128.6, 128.1, 127.0, 125.7, 116.8, 76.3, 18.3. HPLC analysis: Chiralcel OJ-H column, *n*-hexane/*i*PrOH = 100/2 (v/v), 0.5 mL/min, 220 nm; major enantiomer  $t = 15.6$

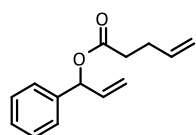
min, minor enantiomer  $t = 12.8$  min, 94% ee.  $[\alpha]_D^{29} = -25.4$  ( $c$  0.23,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{14}\text{H}_{16}\text{O}_2\text{Na}$   $[\text{M}+\text{Na}^+]$ : 225.0886, found:  $m/z = 225.0889$ .

**(E)-1-phenylallyl pent-3-enoate (4ad, Method C)**



$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  7.40–7.26 (m, 5H, Ph), 6.26 (ddd, 1H,  $J = 6.0, 1.7, 1.5$  Hz, PhCH), 6.00 (ddd, 1H,  $J = 17.1, 10.4, 6.0$  Hz, CH=), 5.64–5.49 (m, 2H, CH=CH), 5.28 (ddd, 1H,  $J = 17.1, 1.7, 1.3$  Hz,  $=\text{CH}_2$ ), 5.24 (ddd, 1H,  $J = 10.4, 1.5, 1.3$  Hz,  $=\text{CH}_2$ ), 3.10–3.05 (m, 2H,  $\text{CH}_2$ ), 1.71–1.66 (m, 3H,  $\text{CH}_3$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz):  $\delta$  171.1, 138.9, 136.3, 129.6, 128.5, 128.1, 127.1, 122.5, 116.9, 76.2, 38.2, 17.9. HPLC analysis: Chiralcel OJ-H column,  $n$ -hexane/ $i$ PrOH = 98/2 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 9.7$  min, minor enantiomer  $t = 7.5$  min, 97% ee.  $[\alpha]_D^{29} = -27.2$  ( $c$  0.23,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{14}\text{H}_{16}\text{O}_2\text{Na}$   $[\text{M}+\text{Na}^+]$ : 239.1043, found:  $m/z = 239.1047$ .

**1-phenylallyl pent-4-enoate (4ae, Method C)**



$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz):  $\delta$  7.40–7.26 (m, 5H, Ph), 6.32 (ddd, 1H,  $J = 5.9, 1.6, 1.4$  Hz, PhCH), 6.00 (ddd, 1H,  $J = 17.1, 10.4, 5.9$  Hz, CH=), 5.86–5.76 (m, 1H,  $\text{CH}_2\text{CH=}$ ), 5.30 (ddd, 1H,  $J = 17.1, 1.6, 1.3$  Hz,  $=\text{CH}_2$ ), 5.23 (ddd, 1H,  $J = 10.4, 1.4, 1.3$  Hz,  $=\text{CH}_2$ ), 5.09–4.90 (m, 2H,  $\text{CH}_2\text{CH=CH}_2$ ), 2.51–2.35 (m, 4H,  $\text{CH}_2\text{CH}_2\text{CH=}$ ).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  152.9, 126.5, 124.6, 124.4, 118.2, 117.8, 117.0, 108.9, 107.8, 76.3, 42.4, 38.4. HPLC analysis: Chiralcel OJ-H column,  $n$ -hexane/ $i$ PrOH = 98/2 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 8.7$  min, minor enantiomer  $t = 7.9$  min, 95% ee.  $[\alpha]_D^{29} = -34.7$  ( $c$  0.25,  $\text{CHCl}_3$ ). HRMS (ESI): Calcd for  $\text{C}_{14}\text{H}_{16}\text{O}_2\text{Na}$   $[\text{M}+\text{Na}^+]$ : 239.1043, found:  $m/z = 239.1046$ .

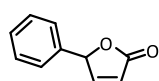
**Standard Procedure for Ring-Closing Metathesis (RCM) of Branched Allylic Ester.**

To a  $\text{CH}_2\text{Cl}_2$  solution (5 mL) of **G-II** (6  $\mu\text{mol}$ , 2 mol%) was added a  $\text{CH}_2\text{Cl}_2$  solution (1 mL) of branched allylic ester (0.3 mmol) and the mixture was stirred for 16 h at 25  $^\circ\text{C}$ . After  $\text{CH}_2\text{Cl}_2$  was removed under reduced pressure, the residue was purified by  $\text{SiO}_2$  column chromatography using a mixture of  $n$ -hexane and AcOEt (v/v = 7/3) as the eluent. Concentration of the resulting solution gave optically active lactones.

For the syntheses of **6db**, **6fb**, **6ac**, and **7**, the reactions were performed under reflux, whereas **8** was prepared by the reaction at 30  $^\circ\text{C}$ .

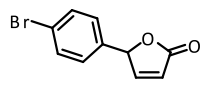
**Characterization of Lactones.**

**5-phenylfuran-2(5H)-one (6ab).<sup>5</sup>**

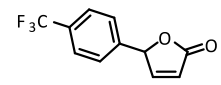


$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta$  7.52 (dd, 1H,  $J = 5.5, 2.0$  Hz, CH=), 7.43–7.32 (m, 3H, Ph), 7.29–7.24 (m, 2H, Ph), 6.23 (dd, 1H,  $J = 5.5, 1.8$  Hz, PhCH), 6.01 (dd, 1H,  $J = 2.0, 1.8$  Hz, COCH=).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 76 MHz):  $\delta$  173.0, 155.8, 134.3, 129.3, 129.0, 126.5, 121.0, 84.3. HPLC analysis: Chiralcel OJ-H column,  $n$ -hexane/ $i$ PrOH = 9/1 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 31.1$  min, minor enantiomer  $t = 34.5$  min, 97% ee.

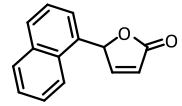
**5-(4-bromophenyl)furan-2(5H)-one (6bb).<sup>5</sup>**

  
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ 7.54 (d, 2H, *J* = 8.4 Hz, Ar), 7.50 (dd, 1H, *J* = 5.6, 1.8 Hz, CH=), 7.15 (d, 2H, *J* = 8.4 Hz, Ar), 6.24 (dd, 1H, *J* = 5.6, 2.2 Hz, ArCH), 5.97 (dd, 1H, *J* = 2.2, 1.8 Hz, COCH=). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 172.6, 155.2, 133.3, 132.3, 128.1, 123.4, 121.3, 83.5. HPLC analysis: Chiralpak AS-H column, *n*-hexane/*i*PrOH = 4/1 (v/v), 0.4 mL/min, 220 nm; major enantiomer *t* = 62.7 min, minor enantiomer *t* = 60.9 min, 98% ee. [α]<sub>D</sub><sup>27</sup> = −31.2 (*c* 0.28, CHCl<sub>3</sub>).

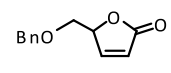
**5-(4-trifluoromethylphenyl)furan-2(5H)-one (6cb).**

  
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.67 (d, 2H, *J* = 8.2 Hz, Ar), 7.54 (dd, 1H, *J* = 5.6, 1.7 Hz, CH=), 7.42 (d, 2H, *J* = 8.2 Hz, Ar), 6.27 (dd, 1H, *J* = 5.6, 2.2 Hz, ArCH), 6.07 (br, 1H, COCH=). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 172.6, 155.2, 138.3 (q, *J* = 1.4 Hz), 131.5 (q, *J* = 32.8 Hz), 122.6, 126.0, (q, *J* = 3.8 Hz), 123.7 (q, *J* = 272.3 Hz), 121.3, 88.3. HPLC analysis: Chiralpak AS-H column, *n*-hexane/*i*PrOH = 9/1 (v/v), 0.3 mL/min, 210 nm; major enantiomer *t* = 77.3 min, minor enantiomer *t* = 74.0 min, 95% ee. [α]<sub>D</sub><sup>29</sup> = −140.6 (*c* 0.12, CHCl<sub>3</sub>). HRMS (ESI): Calcd for C<sub>11</sub>H<sub>7</sub>F<sub>3</sub>O<sub>2</sub>Na [M+Na<sup>+</sup>]: 251.0290, found: *m/z* = 251.0293.

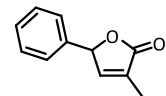
**5-(naphthalene-1-yl)furan-2(5H)-one (6db)**

  
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.91–7.81 (m, 3H, Ar), 7.79 (br, 1H Ar), 7.60 (dd, 1H, *J* = 5.6, 1.6 Hz, CH=), 7.57–7.48 (m, 2H, Ar), 7.31 (dd, 1H, *J* = 8.5, 1.7 Hz, Ar), 6.28 (br, 1H, *J* = 5.6, 2.0 Hz, COCH=), 6.18 (br, 1H, ArCH). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 173.1, 155.8, 133.6, 133.2, 131.6, 129.1, 128.0, 127.8, 126.9, 126.8, 126.2, 123.3, 121.2, 84.5. HPLC analysis: Chiralpak AS-H column, *n*-hexane/*i*PrOH = 4/1 (v/v), 1.0 mL/min, 220 nm; major enantiomer *t* = 29.4 min, minor enantiomer *t* = 24.1 min, 99% ee. [α]<sub>D</sub><sup>30</sup> = −196.6 (*c* 0.14, CHCl<sub>3</sub>). HRMS (ESI): Calcd for C<sub>14</sub>H<sub>10</sub>O<sub>2</sub>Na [M+Na<sup>+</sup>]: 233.0573, found: *m/z* = 233.0577.

**5-(benzyloxymethyl)furan-2(5H)-one (6fb).<sup>6</sup>**

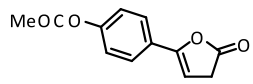
  
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.50 (dd, 1H, *J* = 5.8, 1.6 Hz, CH=), 7.41–7.27 (m, 5H, Ph), 6.17 (dd, 1H, *J* = 5.8, 2.1 Hz, CH=), 5.17 (tdd, 1H, *J* = 5.2, 2.1, 1.6 Hz, CHCH=), 4.57 (s, 2H, PhCH<sub>2</sub>), 3.75 (dd, 1H, *J* = 10.4, 5.2, OCH<sub>2</sub>CH), 3.67 (dd, 1H, *J* = 10.4, 5.2 Hz, OCH<sub>2</sub>CH). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 172.7, 153.8, 137.3, 128.5, 128.0, 127.7, 122.6, 82.1, 73.8, 69.5. HPLC analysis: Chiralpak AS-H column, *n*-hexane/*i*PrOH = 4/1 (v/v), 1.0 mL/min, 220 nm; major enantiomer *t* = 24.1 min, minor enantiomer *t* = 20.4 min, 95% ee.

**3-methyl-5-phenylfuran-2(5H)-one (6ac).<sup>7</sup>**

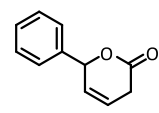
  
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ 7.43–7.32 (m, 3H, Ph), 7.28–7.23 (m, 2H, Ph), 7.12 (dq, 1H, *J* = 1.7, 1.7 Hz, CH=), 5.88–5.83 (m, 1H, PhCH), 1.99 (dd, 3H, *J* = 1.7, 1.7 Hz, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 174.3, 148.4, 135.4, 129.5, 129.0, 128.9, 126.4, 82.1, 10.6. HPLC analysis: Chiralpak AS-H column, *n*-hexane/*i*PrOH = 9/1 (v/v), 1.0 mL/min, 220 nm;

major enantiomer  $t = 21.7$  min, minor enantiomer  $t = 17.1$  min, 97% ee.

**Methyl 4-(5-oxo-4,5-dihydrofuran-2-yl)benzoate (7).**

  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 500 Hz):  $\delta$  8.08 (d, 2H,  $J = 8.7$  Hz, Ar), 7.67 (d, 2H,  $J = 8.7$  Hz, Ar), 5.93 (t, 1H,  $J = 2.8$  Hz, =CH), 3.94 (s, 3H, Me), 3.46 (d, 2H,  $J = 2.8$  Hz,  $\text{CH}_2$ ).  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 75 MHz):  $\delta$  175.2, 166.4, 153.2, 132.3, 130.9, 130.0, 124.6, 100.2, 52.2, 34.7. HRMS (ESI) Calcd for  $\text{C}_{12}\text{H}_{10}\text{O}_4\text{Na}$  [ $\text{M}+\text{Na}^+$ ]: 241.0471, found:  $m/z = 241.0475$ .

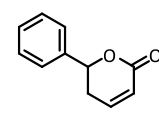
**6-phenyl-3,6-dihydro-2H-pyran-2-one (8).<sup>8</sup>**

  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 500 MHz):  $\delta$  7.43–7.31 (m, 5H, Ph), 6.07–5.97 (m, 3H, PhCH and CH=CH), 3.19–3.15 (m, 2H,  $\text{CH}_2$ ).  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  168.23, 138.20, 128.89, 126.83, 126.18, 121.80, 81.28, 29.91. One carbon signal could not be detected probably due to overlapping. HPLC analysis: Chiralpak AS-H column,  $n$ -hexane/ $i$ PrOH = 4/1 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 27.6$  min, minor enantiomer  $t = 25.1$  min, 93% ee.  $[\alpha]_{\text{D}}^{28} = -125.0$  (c 0.12,  $\text{CHCl}_3$ ).

**Procedure for Tandem RCM—Olefin Isomerization.**

To a 1,2-dichloroethane solution (5 mL) of **G-II** (6  $\mu\text{mol}$ , 2 mol%) was added a 1,2-dichloroethane solution (1 mL) of **4ad** (0.3 mmol), and the mixture was stirred for 16 h at 50 °C. After addition of 2-propanol (0.2 mL), the mixture was refluxed for 48 h. After cooling to room temperature, the solvent was removed under reduced pressure and the residue was purified by  $\text{SiO}_2$  column chromatography using a mixture of  $n$ -hexane and AcOEt (v/v = 7/3) as the eluent. Evaporation of the solvent under reduced pressure gave **9** as white solid.

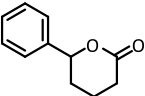
**6-phenyl-5,6-dihydro-2H-pyran-2-one (9).<sup>9</sup>**

  $^1\text{H NMR}$  ( $\text{CDCl}_3$ , 300 MHz):  $\delta$  7.46–7.30 (m, 5H, Ph), 6.97 (ddd, 1H,  $J = 9.7, 5.5, 3.0$  Hz,  $\text{CH}_2\text{CH}=\text{CH}$ ), 6.15 (ddd, 1H,  $J = 9.7, 2.4, 1.3$  Hz, COCH=), 5.46 (dd, 1H,  $J = 10.5, 5.5$  Hz, 1H, PhCH), 2.76–2.54 (m, 2H,  $\text{CH}_2$ ).  $^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 126 MHz):  $\delta$  164.0, 144.8, 138.5, 128.7, 128.6, 126.0, 121.7, 79.2, 31.6. HPLC analysis: Chiralpak AS-H column,  $n$ -hexane/ $i$ PrOH = 4/1 (v/v), 1.0 mL/min, 220 nm; major enantiomer  $t = 31.1$  min, minor enantiomer  $t = 20.4$  min, 94% ee.

**Procedure for Tandem RCM—Hydrogenation.**

To a  $\text{CH}_2\text{Cl}_2$  solution (5 mL) of **G-II** (6  $\mu\text{mol}$ , 2 mol%) was added a  $\text{CH}_2\text{Cl}_2$  solution (1 mL) of **4ad** (0.3 mmol). After stirring for 12 h at 30 °C, the mixture was transferred to autoclave through cannula, rinsing with  $\text{CH}_2\text{Cl}_2$  (3  $\times$  2 mL). The mixture was stirred for 24 h at 70 °C under  $\text{H}_2$  pressure (1 MPa). After cooling to room temperature,  $\text{CH}_2\text{Cl}_2$  was removed under reduced pressure, and the residue was purified by  $\text{SiO}_2$  column chromatography using a mixture of  $n$ -hexane and AcOEt (v/v = 7/3) as the eluent. Recrystallization from  $n$ -hexane gave **10** as colorless needle.

### 6-phenyltetrahydro-2H-pyran-2-one (**10**).<sup>10</sup>

 <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz): δ 7.41–7.27 (m, 5H, Ph), 5.36 (dd, 1H, *J* = 10.1, 3.4 Hz, PhCH), 2.79–2.49 (m, 2H, COCH<sub>2</sub>), 2.25–2.11 (m, 1H, CHCH<sub>2</sub>), 2.07–1.80 (m, 3H, CHCH<sub>2</sub> and CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz): δ 171.2, 139.7, 128.6, 128.2, 125.7, 81.5, 30.5, 29.5, 18.6. HPLC analysis: Chiralcel OB-H column, hexane/*i*PrOH = 4/1 (v/v), 1.0 mL/min, 254 nm; major enantiomer *t* = 20.3 min, minor enantiomer *t* = 24.7 min, >99% ee.

### Metathesis Reaction of **4ae**.

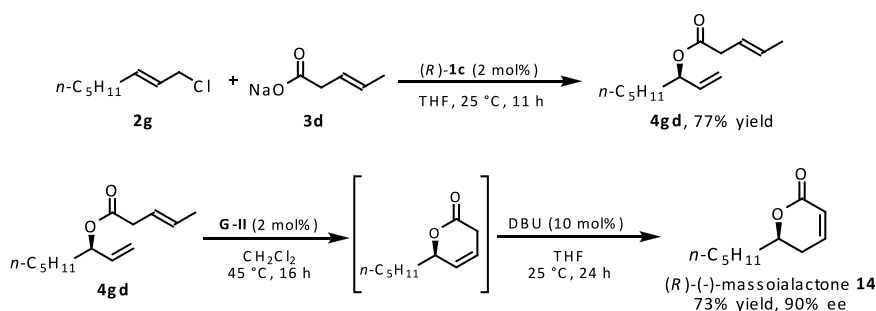
The reaction was performed according to the standard procedure. The resulting brown oil was analyzed by mass spectrometry.

ESI MS: *m/z* = 189.2(10.5), 399.3(100), 400.3(24.0), 427.3(15.2), 587.2(10.1), 775.3(9.6). Calcd for C<sub>12</sub>H<sub>12</sub>O<sub>2</sub> [**11**+H]<sup>+</sup>, 189.1; C<sub>24</sub>H<sub>24</sub>O<sub>4</sub>Na [**12a**+Na]<sup>+</sup>, 399.2; C<sub>24</sub>H<sub>24</sub>O<sub>4</sub>Na [**13**+Na]<sup>+</sup>, 427.2; C<sub>36</sub>H<sub>36</sub>O<sub>6</sub>Na [**12b**+Na]<sup>+</sup>, 587.2; C<sub>48</sub>H<sub>48</sub>O<sub>8</sub>Na [**12c**+Na]<sup>+</sup>, 775.3.

### One-Pot Sequential Asymmetric Allylic Carboxylation—Ring-Closing Metathesis.

To a THF solution (1 mL) of (*R*)-**1c** (5 μmol, 1 mol%), allylic chloride (1.0 mmol) and Na<sub>2</sub>CO<sub>3</sub> (1.5 mmol) was added a 1 mL of THF solution of crotonic acid **3b** (0.5 mmol), and the resulting mixture was stirred for 4 h at 25 °C. **G-II** (10 μmol, 2 mol%) and CH<sub>2</sub>Cl<sub>2</sub> (8 mL) were added, and the mixture was stirred for 16 h. After removal of the solvent under reduced pressure, the residue was purified by SiO<sub>2</sub> column chromatography using a mixture of *n*-hexane and AcOEt (v/v = 7/3) as the eluent.

### Enantioselective Synthesis of (*R*)-(-)-Massoialactone (**14**).

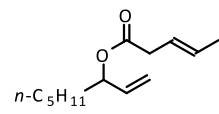


To a THF solution (2 mL) of (*R*)-**1c** (10 μmol, 2 mol%) and allylic chloride **2g** (1.0 mmol) was added sodium pent-3-enoate **3d** (0.5 mmol), and the resulting mixture was stirred for 11 h at 25 °C. After dilution with hexane, insoluble parts were removed by filtration. The filtrate was concentrated under reduced pressure, and the residue was purified by flash column chromatography on SiO<sub>2</sub> using the mixture of *n*-hexane and Et<sub>2</sub>O (v/v = 20/1) as the eluent. Evaporation of the solvent gave the ester **4gd** as colorless oil (81 mg, 77% yield).

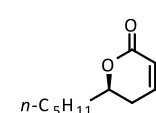
**G-II** (6 μmol, 2 mol%) was added to a CH<sub>2</sub>Cl<sub>2</sub> solution (6 mL) of **4gd** (63 mg, 0.3 mmol), and the mixture was stirred for 16 h at 45 °C. After removal of the solvent under reduced pressure, a THF

solution (2 mL) of 1,8-diazabicyclo-[5,4,0]-undec-7-ene (30  $\mu$ mol, 10 mol%) was added and the mixture was stirred for 24 h at 25 °C. The resulting mixture was poured into a mixture of Et<sub>2</sub>O (20 mL) and NH<sub>4</sub>Cl aq., and the organic layer was washed with NH<sub>4</sub>Cl aq., water and brine successively. The solution was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by SiO<sub>2</sub> column chromatography using a mixture of *n*-hexane and AcOEt (v/v = 7/3) as the eluent to give **14** as colorless oil (36.8 mg, 73% yield).

**(E)-oct-1-en-3-yl pent-3-enoate (4gd).**

 <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  5.77 (ddd, 1H, *J* = 17.0, 10.6, 6.4 Hz, CHCH=), 5.65–5.50 (m, 2H, COCH<sub>2</sub>CH=CH), 5.27–5.19 (m, 2H, OCH and =CH<sub>2</sub>), 5.15 (ddd, 1H, *J* = 10.6, 1.3, 1.3 Hz, =CH<sub>2</sub>), 3.04–2.99 (m, 2H, COCH<sub>2</sub>), 1.69 (d, 3H, *J* = 4.9 Hz, =CHCH<sub>3</sub>), 1.67–1.54 (m, 1H, CHCH<sub>2</sub>CH<sub>2</sub>), 1.38–1.21 (m, 7H, CH<sub>2</sub>CH<sub>2</sub>), 0.88 (t, 3H, *J* = 6.9 Hz, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz):  $\delta$  171.4, 136.7, 129.3, 122.8, 116.5, 74.9, 38.3, 34.1, 31.5, 24.7, 22.5, 17.9, 13.9.  $[\alpha]_D^{26} = +4.1$  (*c* 0.07, CHCl<sub>3</sub>). HRMS (ESI): Calcd for C<sub>13</sub>H<sub>22</sub>O<sub>2</sub>Na [M+Na<sup>+</sup>]: 233.1512, found: *m/z* = 233.1516.

**(R)-(-)-6-pentyl-5,6-dihydro-2H-pyran-2-one, (R)-(-)-massoialactone (14).<sup>11</sup>**

 <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  6.89–6.83 (m, 1H, CH<sub>2</sub>CH=), 6.01 (ddd, 1H, *J* = 9.7, 2.4, 1.4 Hz, COCH=), 4.45–4.38 (m, 1H, OCH), 2.39–2.28 (m, 2H, CH<sub>2</sub>CH=), 1.86–1.74 (m, 1H, CH<sub>2</sub>CH), 1.69–1.59 (m, 1H, CH<sub>2</sub>CH), 1.54–1.47 (m, 1H, CHCH<sub>2</sub>CH<sub>2</sub>), 1.46–1.25 (m, 5H, CH<sub>2</sub>CH<sub>2</sub>), 0.90 (t, 3H, *J* = 7.0 Hz, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz):  $\delta$  164.5, 144.9, 121.5, 78.0, 34.8, 31.5, 29.4, 24.5, 22.4, 13.9. HPLC analysis: Chiralpak AS-H column, hexane/*i*PrOH = 9/1 (v/v), 1.0 mL/min, 210 nm; major enantiomer *t* = 18.8 min, minor enantiomer *t* = 22.4 min. 90% ee.  $[\alpha]_D^{27} = -103.7$  (*c* 0.20, CHCl<sub>3</sub>).

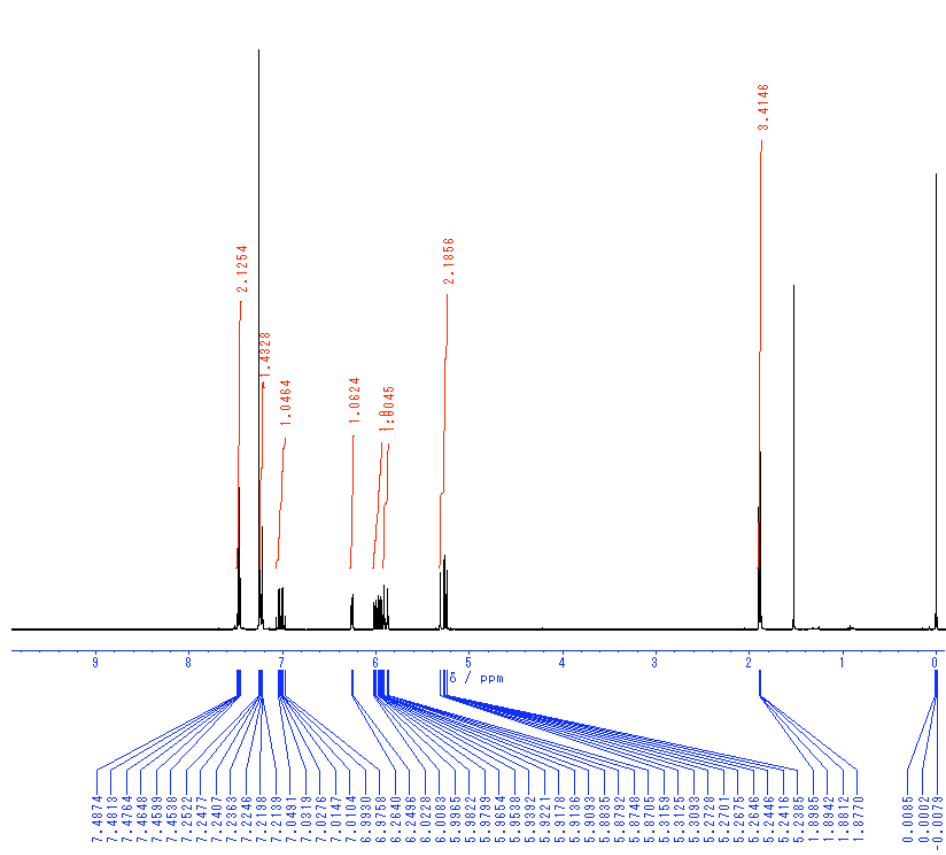
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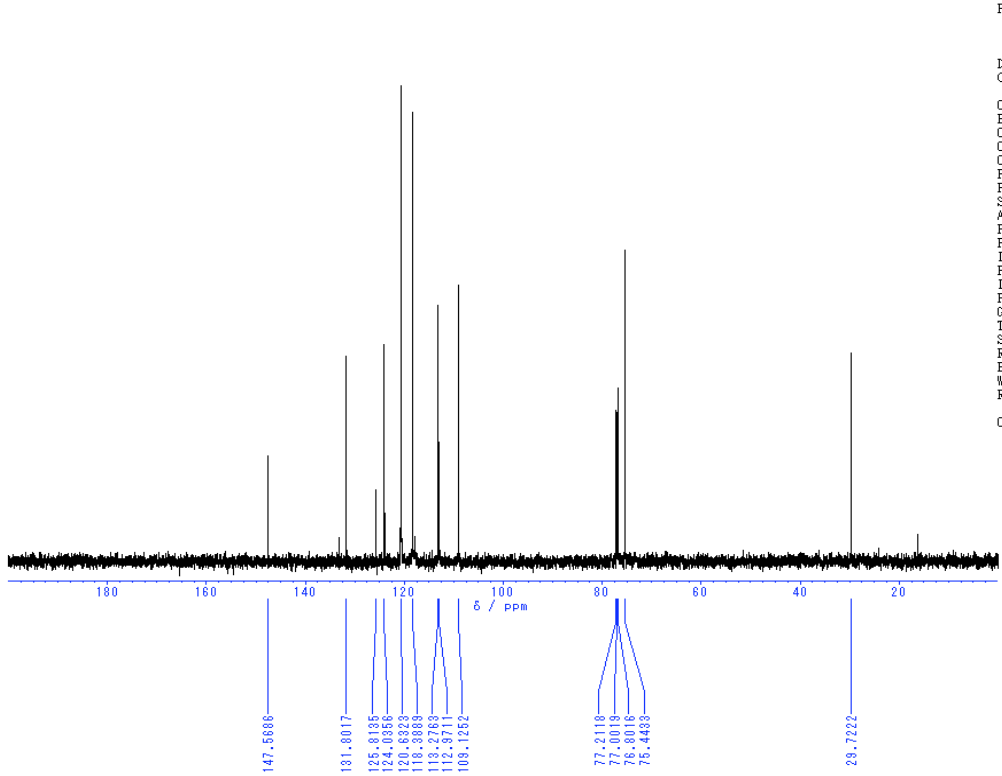
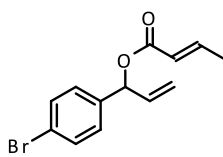
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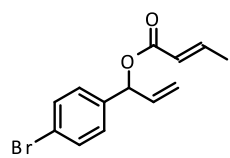
**(E)-1-(4-bromophenyl)allyl but-2-enoate (4bb).**



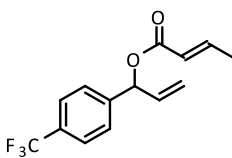
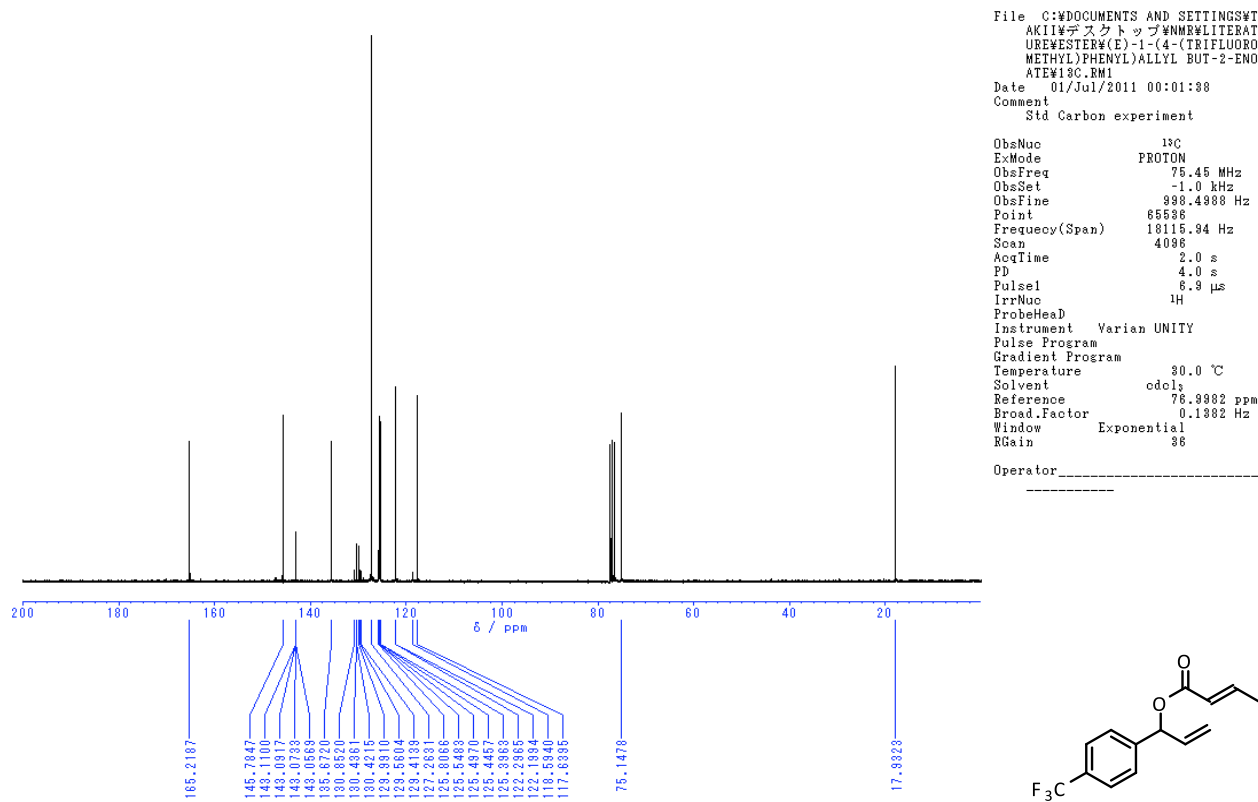
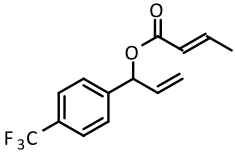
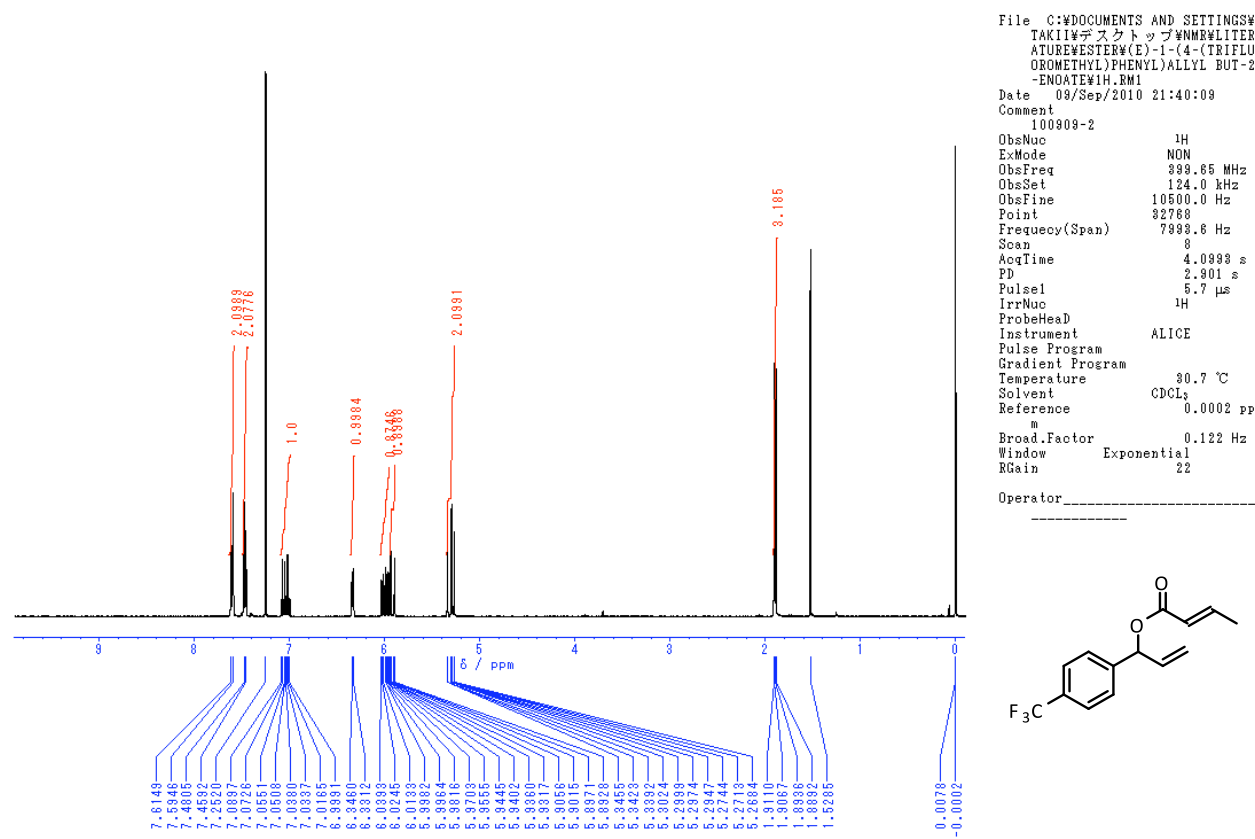
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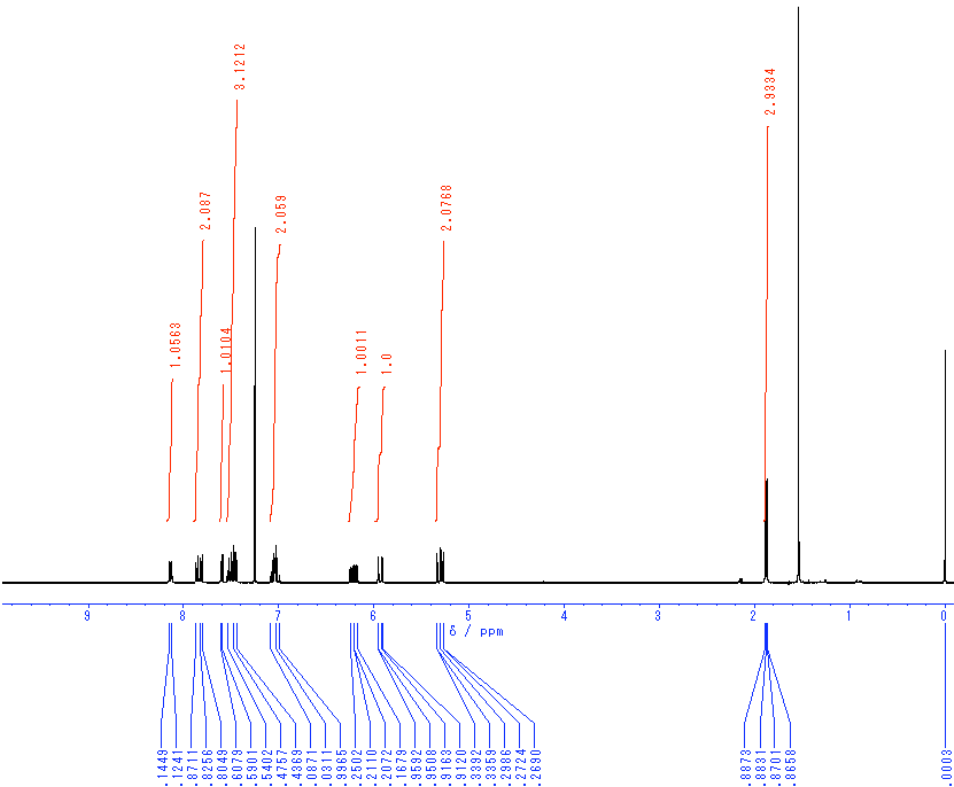
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PD 2.0 s  
Pulsel 5.0 µs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFO  
Reference 76.9981 ppm  
Broad.Factor 0.9586 Hz  
Window Exponential  
RGain 56  
Operator \_\_\_\_\_



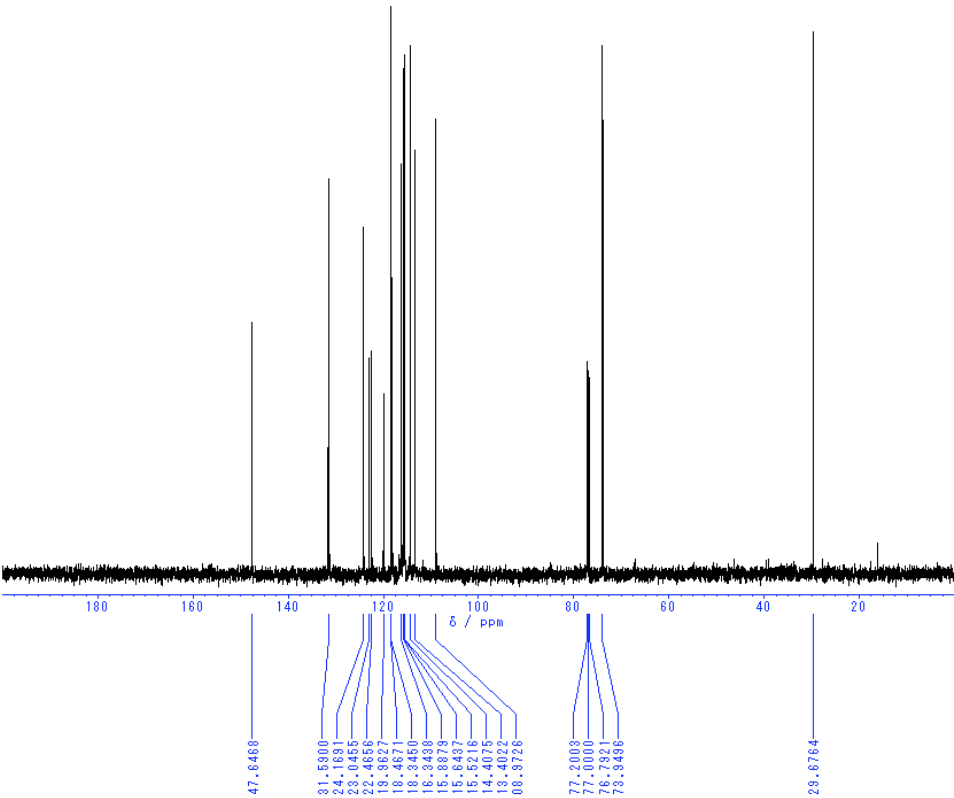
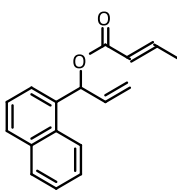
**(E)-1-(4-trifluoromethylphenyl)allyl but-2-enoate (4cb).**



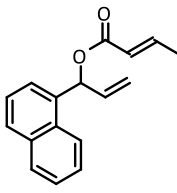
**(E)-1-(naphthalene-1-yl)allyl but-2-enoate (4db).**



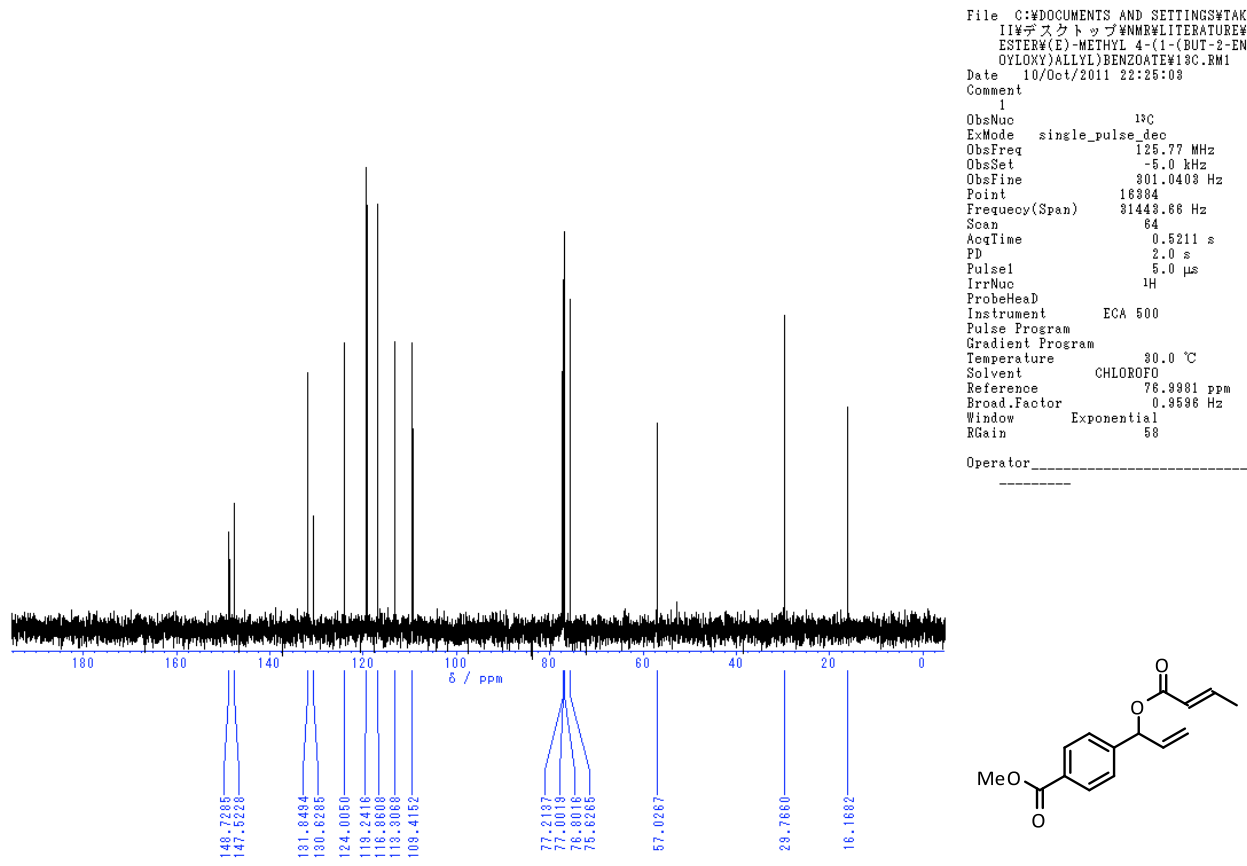
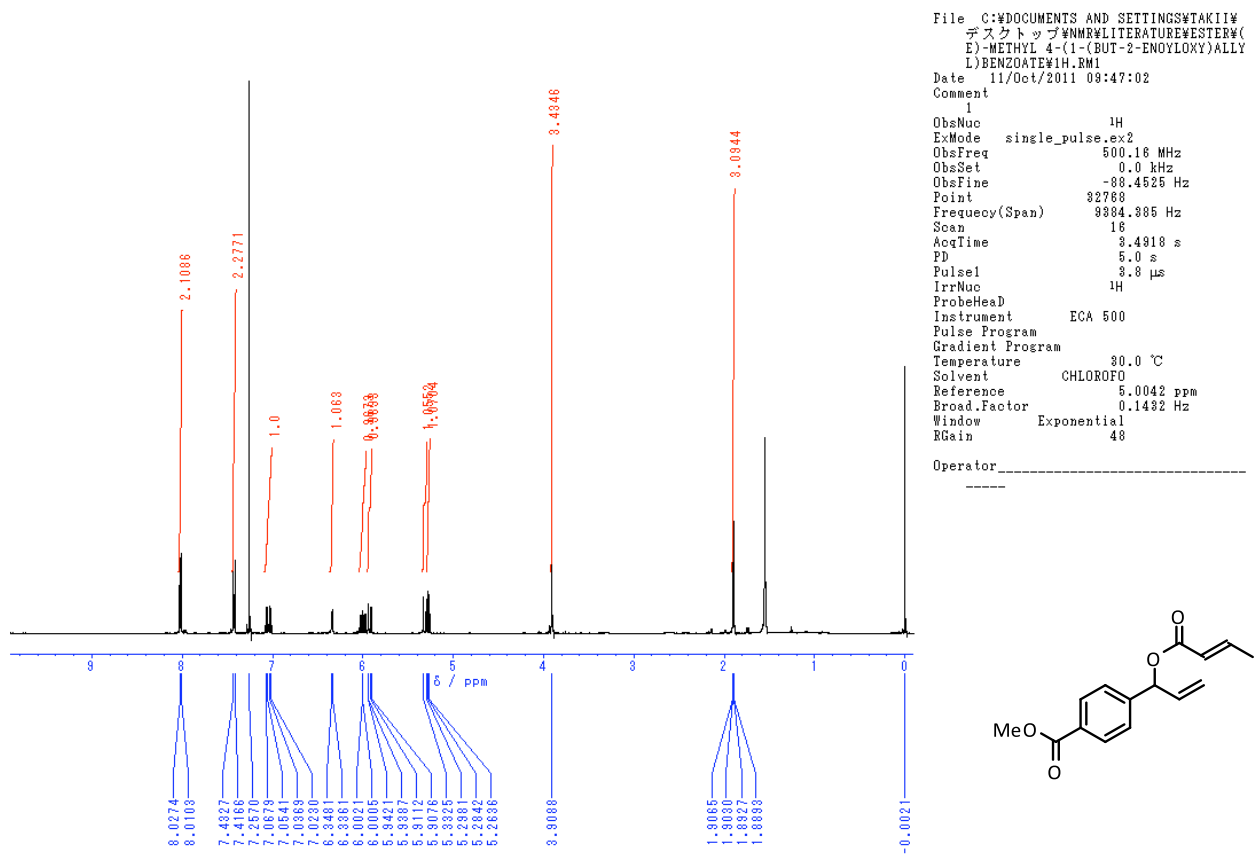
File C:\DOCUMENTS AND SETTINGS\TAK  
II\デスクトップ\NMR\LITERATURE\ESTER(E)-1-(NAPHTHALEN-1-YL)ALLYL BUT-2-ENOATE\1H.RM1  
Date 01/Oct/2010 18:15:32  
Comment 101001-1  
ObsNuc 1H  
ExMode NON  
ObsFreq 399.65 MHz  
ObsSet 124.0 kHz  
ObsFine 10500.0 Hz  
Point 32768  
Frequency(Span) 7993.6 Hz  
Scan 4  
AcqTime 4.0998 s  
PD 2.901 s  
Pulse 6.7 µs  
IrrNuc 1H  
ProbeHead  
Instrument ALICE  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CDCL<sub>3</sub>  
Reference 0.0003 ppm  
Broad.Factor 0.122 Hz  
Window Exponential  
RGain 23  
Operator



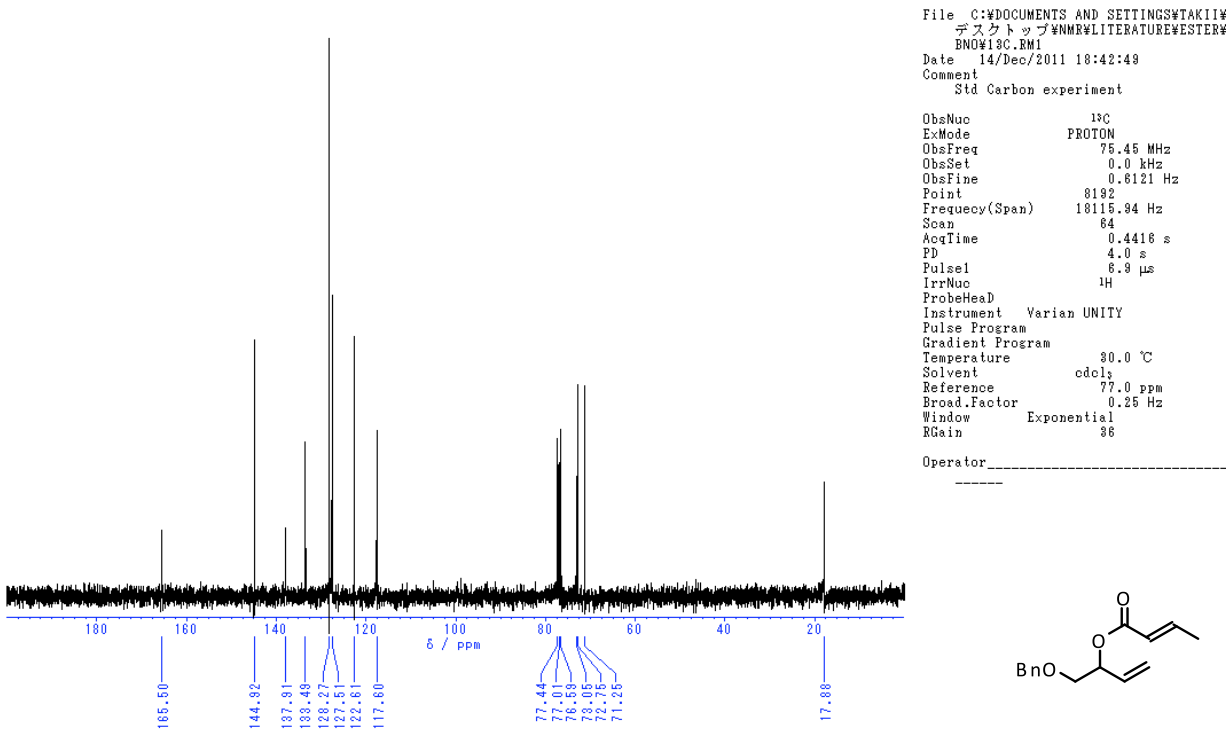
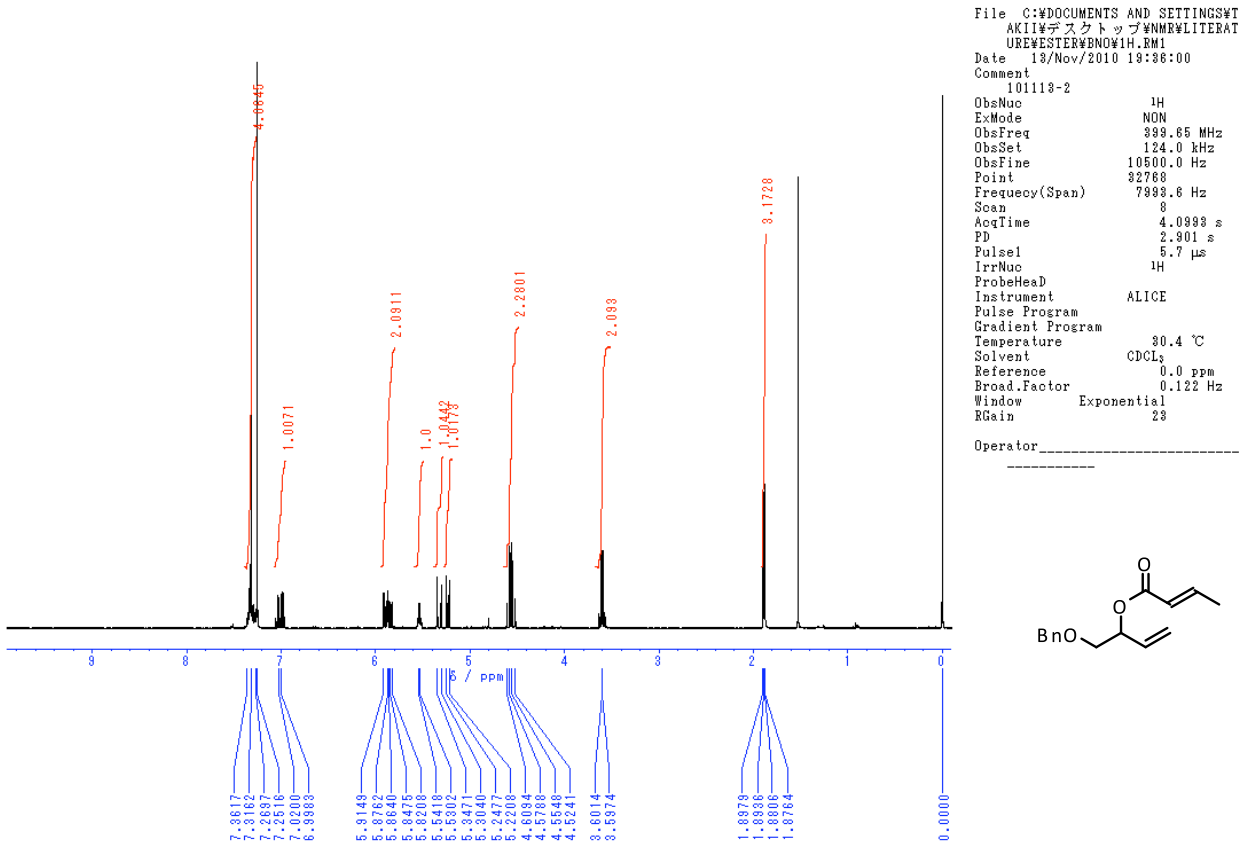
File C:\DOCUMENTS AND SETTINGS\TAK  
II\デスクトップ\NMR\LITERATURE\ESTER(E)-1-(NAPHTHALEN-1-YL)ALLYL BUT-2-ENOATE\13C.RM1  
Date 10/Oct/2011 22:11:07  
Comment 1  
ObsNuc 13C  
ExMode single\_pulse\_dec  
ObsFreq 125.77 MHz  
ObsSet -5.0 kHz  
ObsFine 301.0403 Hz  
Point 16384  
Frequency(Span) 31443.66 Hz  
Scan 128  
AcqTime 0.5211 s  
PD 2.0 s  
Pulse 5.0 µs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFORM-D  
Reference 77.0 ppm  
Broad.Factor 0.25 Hz  
Window Exponential  
RGain 58  
Operator



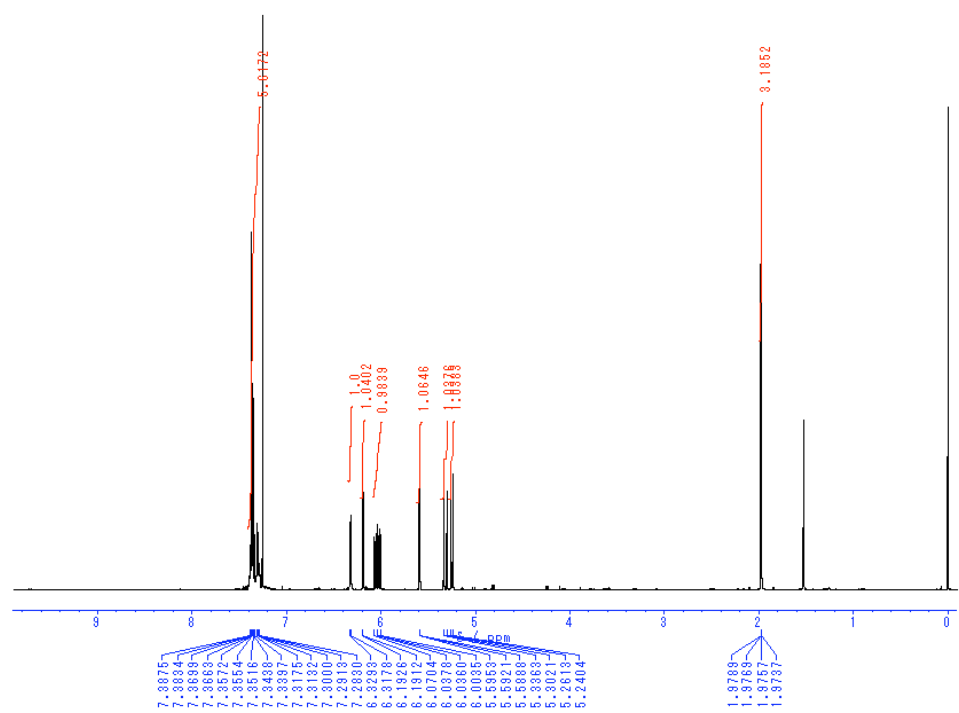
(E)-methyl 4-(1-(but-2-enyloxy)allyl)benzoate (4eb)



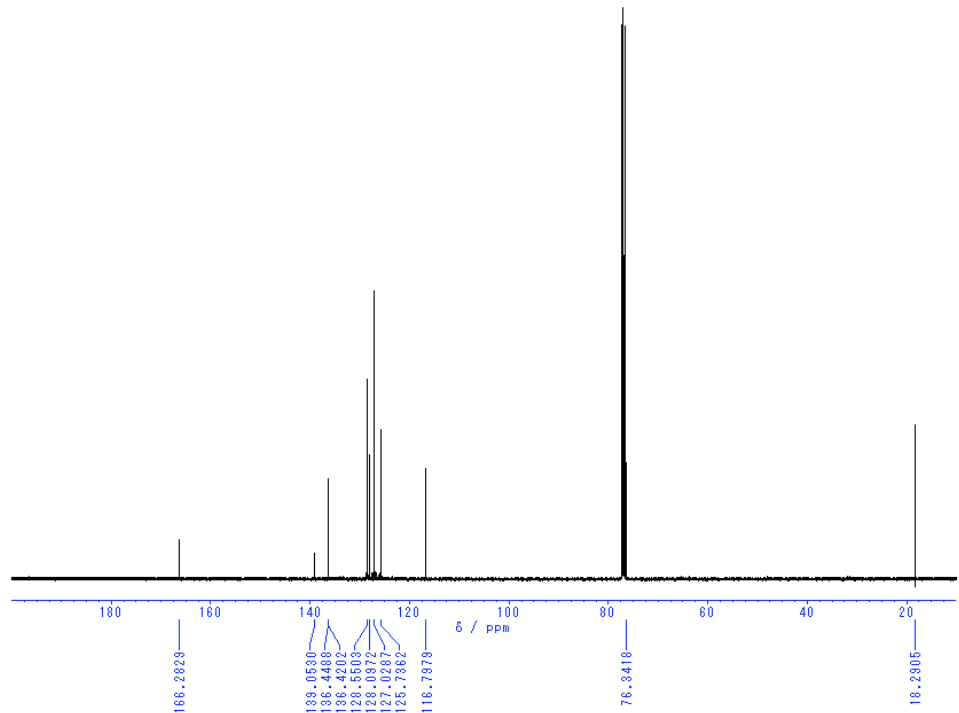
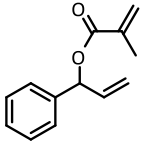
(E)-1-(benzyloxy)but-3-en-2-yl but-2-enoate (4fb).



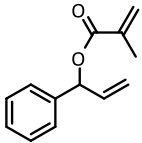
(E)-1-phenylallyl methacrylate (4ac).



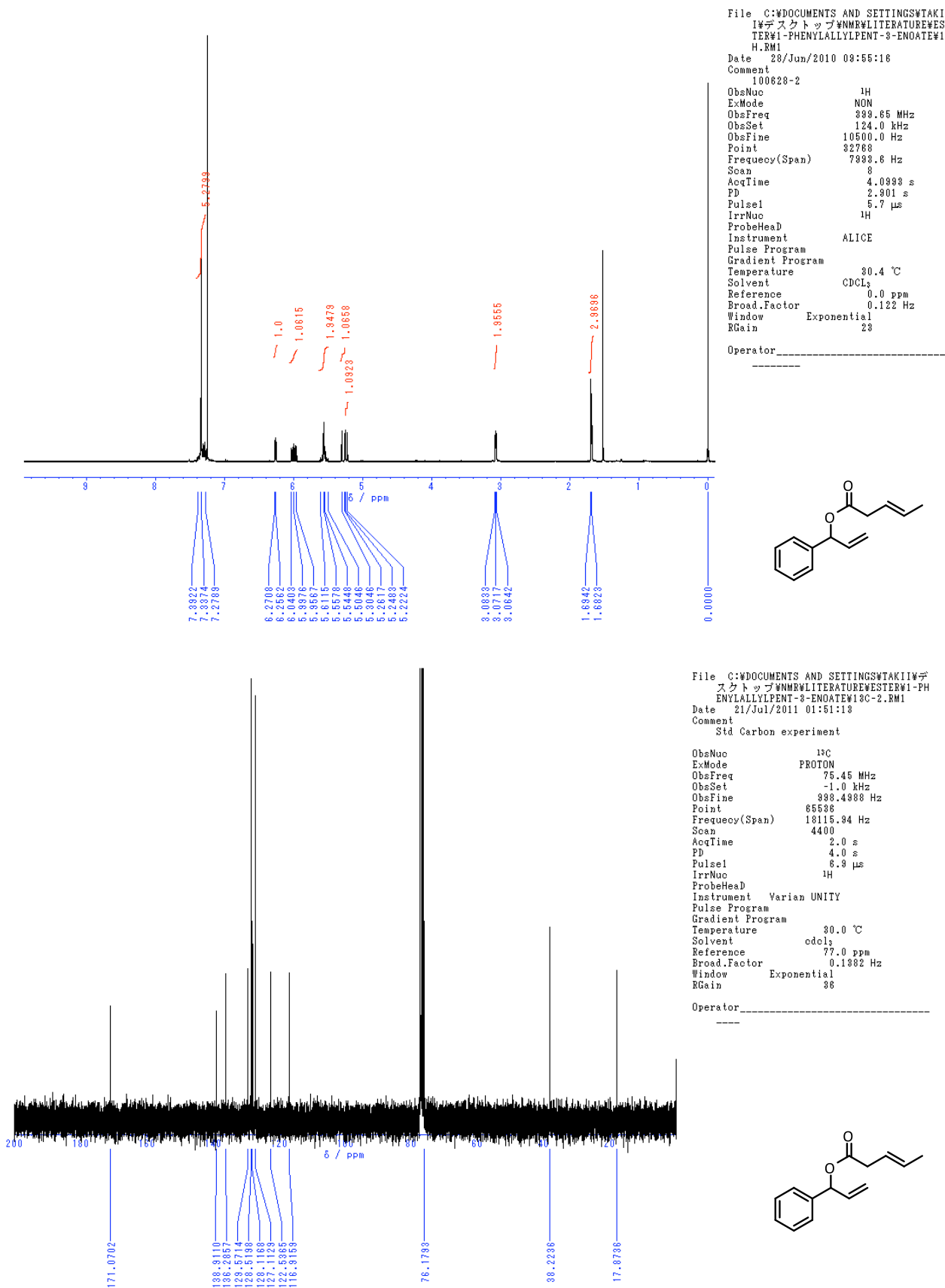
File C:\DOCUMENTS AND SETTINGS\WTAKI\デスクトップ\NNMR\LITERATURE\URE\ESTER\1-PHENYLALLYL METHACRYLATE\1H.RM1  
Date 26/Apr/2011 23:54:39  
Comment 1  
ObsNuc 1H  
ExMode single\_pulse.ex2  
ObsFreq 500.16 MHz  
ObsSet 0.0 kHz  
ObsTime -99.4525 Hz  
Point 85538  
Frequency(Span) 3384.385 Hz  
Scan 8  
AcqTime 6.9885 s  
PD 5.0 s  
Pulse1 6.25 µs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFO  
Reference -0.0001 ppm  
Broad.Factor 0.0716 Hz  
Window Exponential  
RGain 48  
Operator \_\_\_\_\_



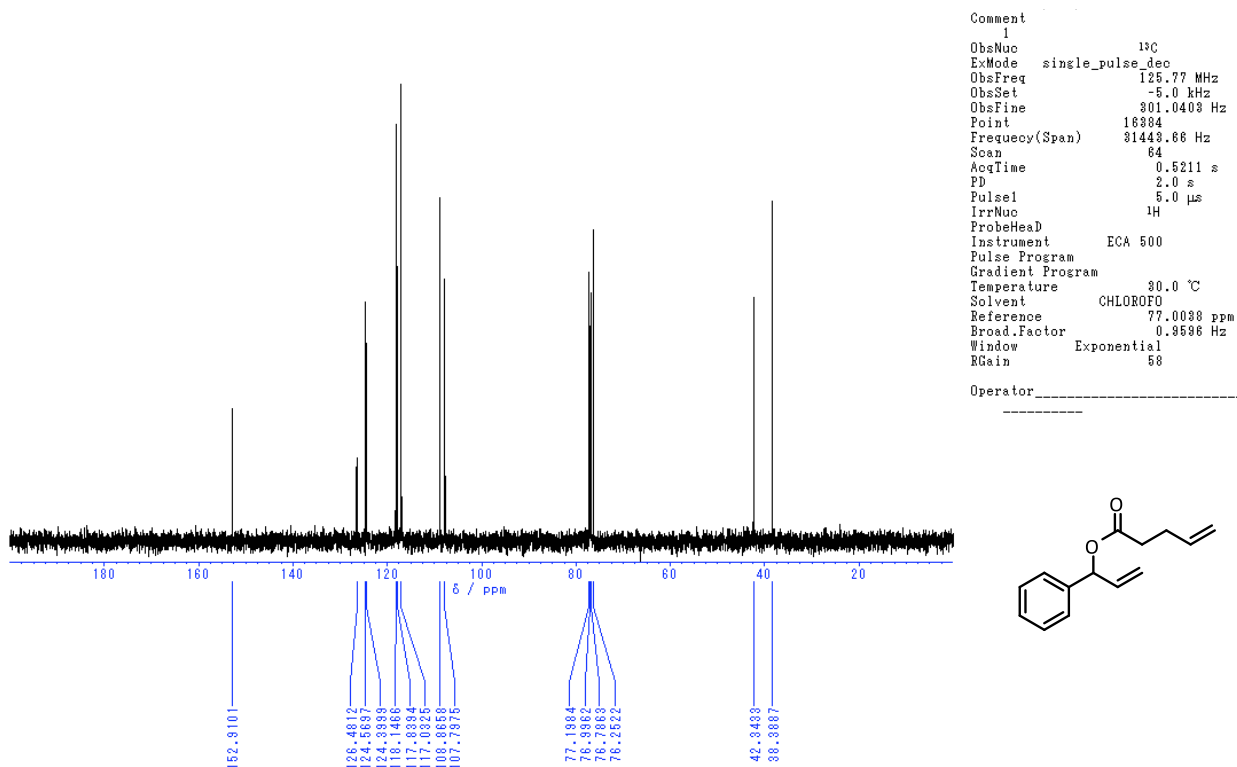
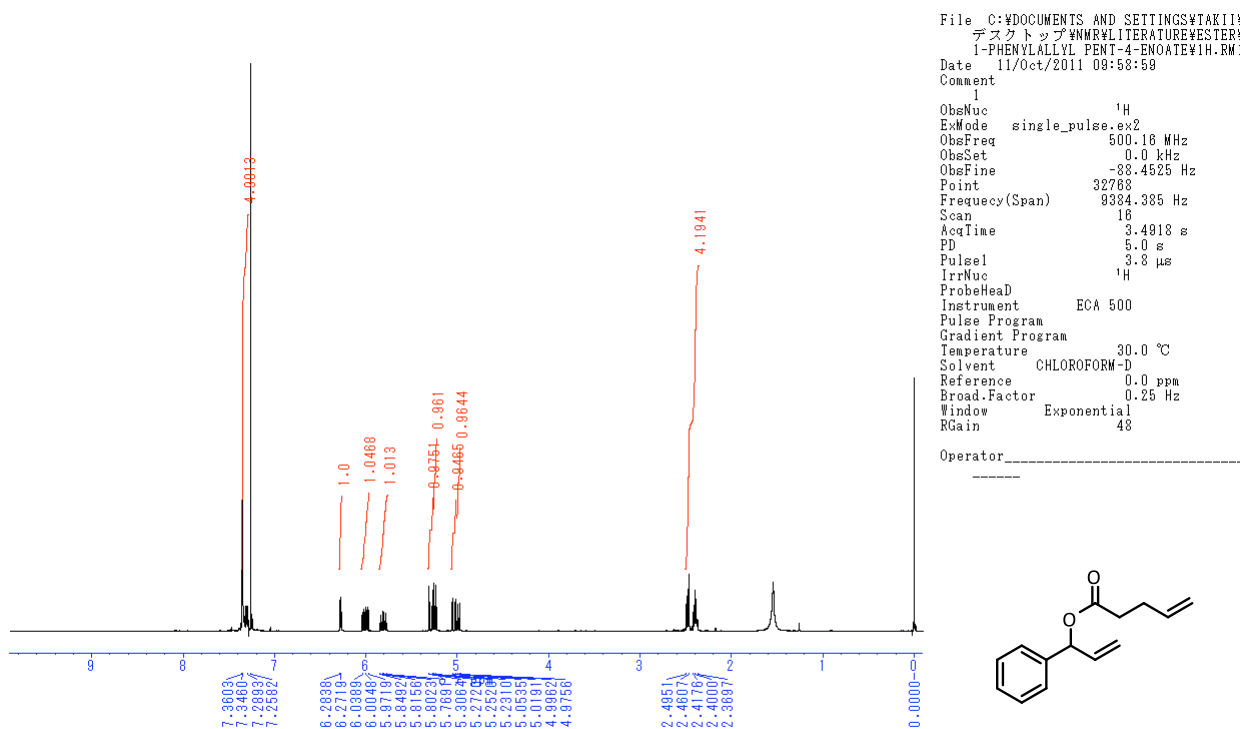
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Date 27/Apr/2011 08:08:00  
Comment 1  
ObsNuc 13C  
ExMode single\_pulse\_dec  
ObsFreq 125.77 MHz  
ObsSet -5.0 kHz  
ObsTime 201.0403 Hz  
Point 85538  
Frequency(Span) 33309.18 Hz  
Scan 7987  
AcqTime 1.6672 s  
PD 2.0 s  
Pulse1 4.55 µs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFO  
Reference 77.0 ppm  
Broad.Factor 0.2999 Hz  
Window Exponential  
RGain 58  
Operator \_\_\_\_\_



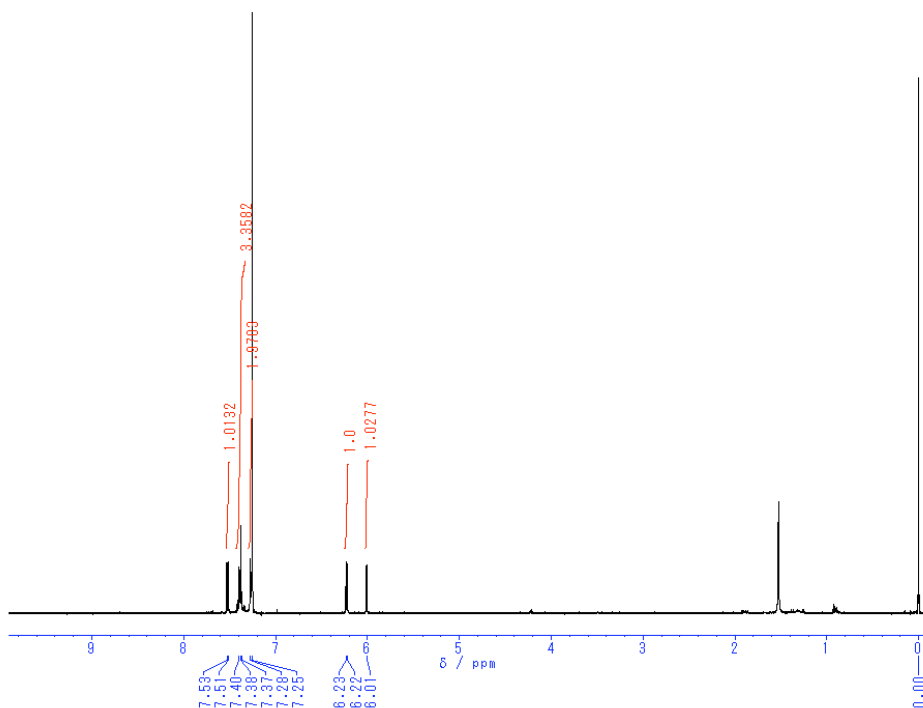
**(E)-1-phenylallyl pent-3-enoate (4ad).**



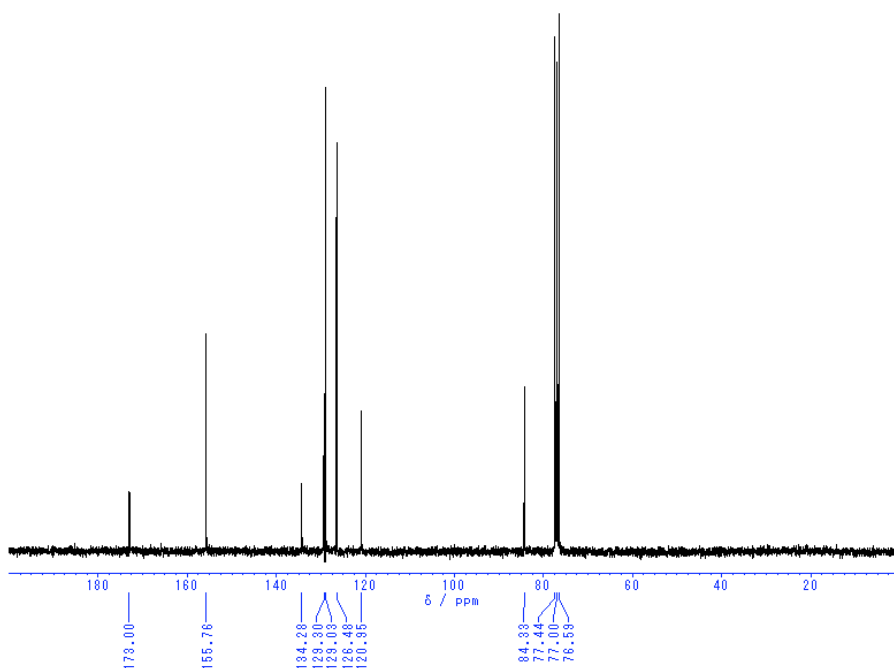
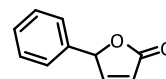
**1-phenylallyl pent-4-enoate (4ae).**



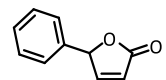
5-phenylfuran-2(5H)-one (6ab).



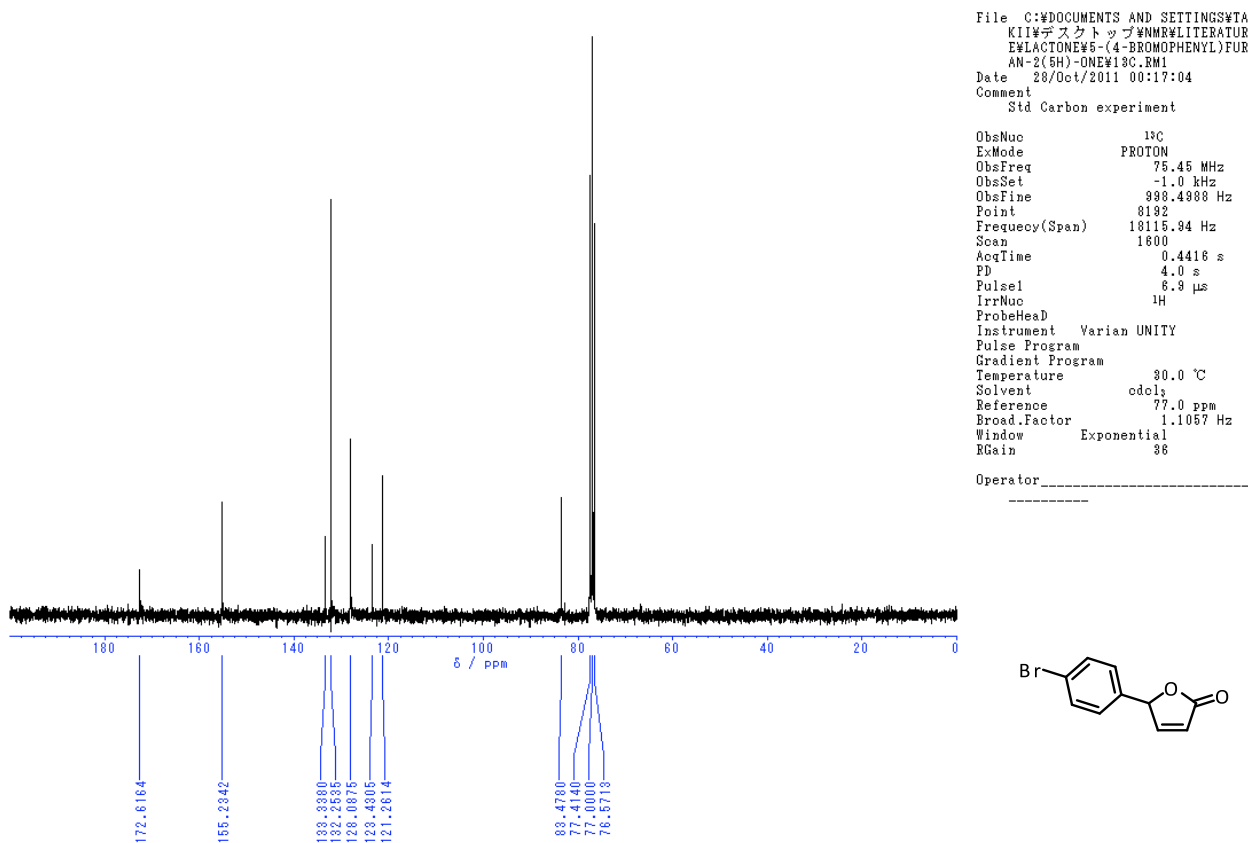
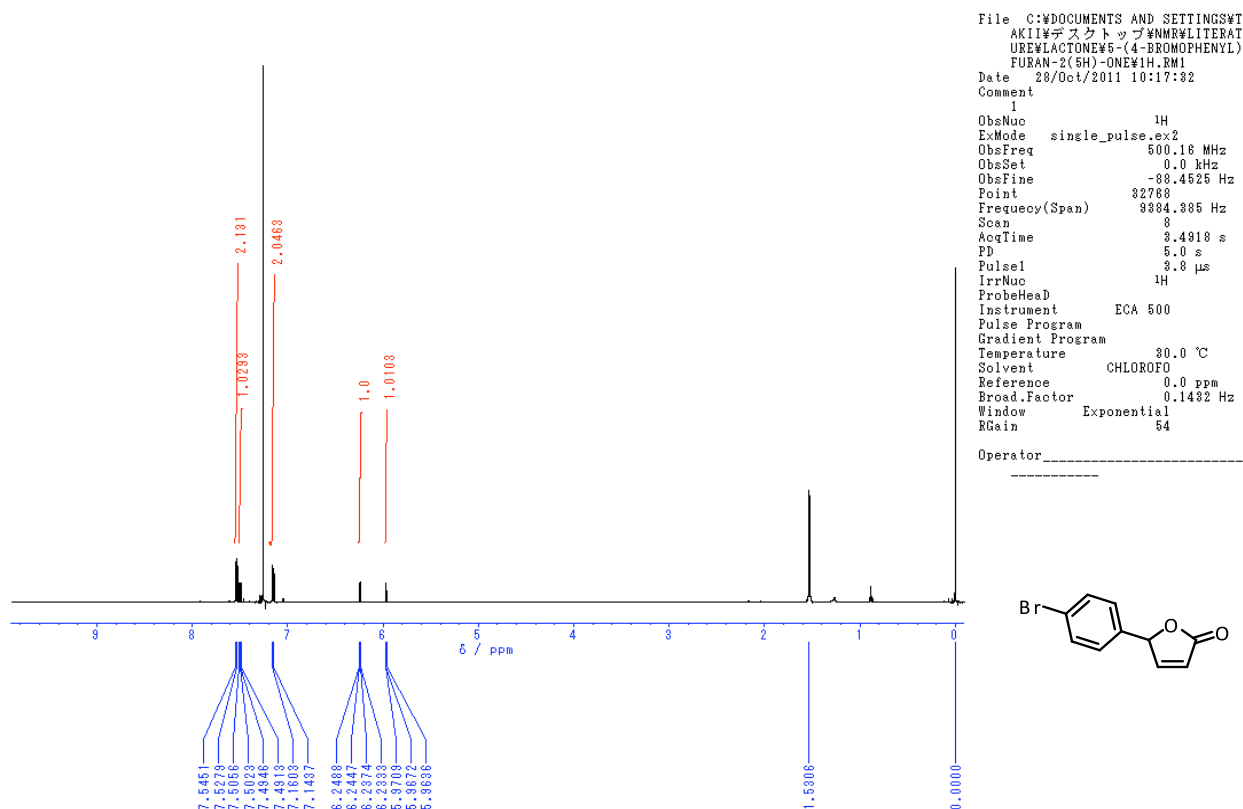
File C:\DOCUMENTS AND SETTINGS\TAKI  
I\デスクトップ\NMR\LITERATURE\LA  
CTONE\5-PHENYLFURAN-2(5H)-ONE\1H  
.RM1  
Date 05/Jan/2011 13:41:19  
Comment  
110105-2  
ObsNuc <sup>1</sup>H  
ExMode NON  
ObsFreq 399.85 MHz  
ObsSet 124.0 kHz  
ObsFine 10500.0 Hz  
Point 32768  
Frequency(Span) 7993.6 Hz  
Scan 4  
AcqTime 4.0893 s  
PD 2.901 s  
Pulse 5.7 μs  
IrrNuc <sup>1</sup>H  
ProbeHead ALICE  
Instrument  
Pulse Program  
Gradient Program  
Temperature 31.2 °C  
Solvent CDCl<sub>3</sub>  
Reference 0.0 ppm  
Broad.Factor 0.122 Hz  
Window Exponential  
RGain 25  
Operator \_\_\_\_\_



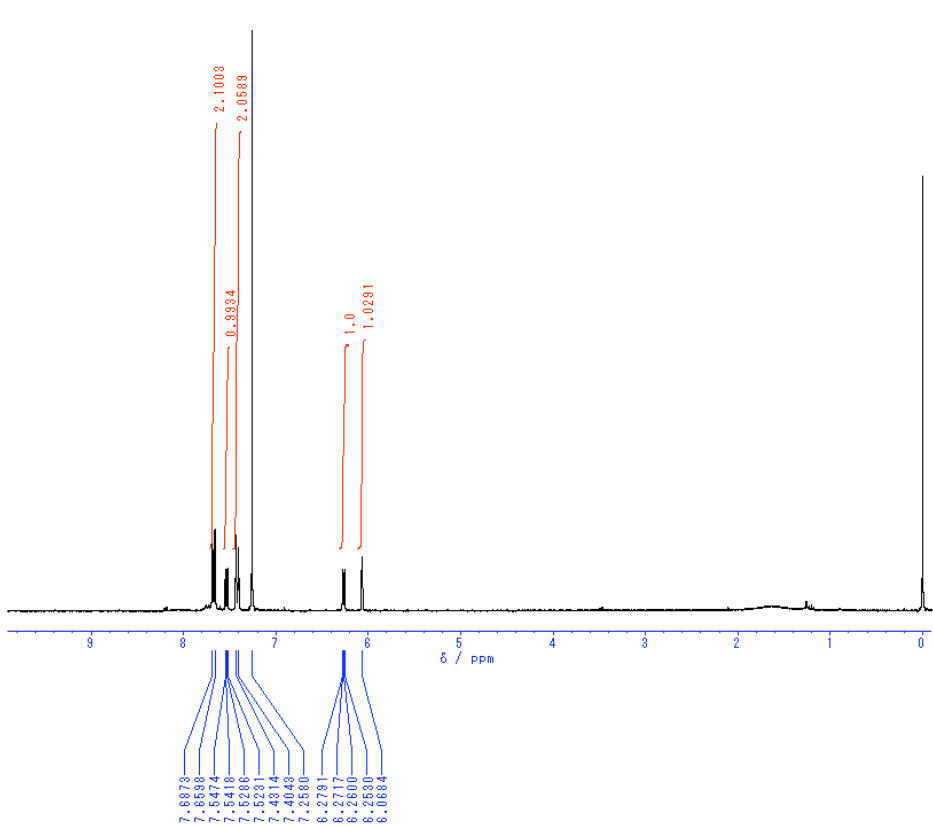
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デスクトップ\NMR\LITERATURE\LACTON  
E\5-PHENYLFURAN-2(5H)-ONE\13C.RM1  
Date 22/Dec/2011 01:43:07  
Comment  
Std Carbon experiment  
ObsNuc <sup>13</sup>C  
ExMode PROTON  
ObsFreq 75.45 MHz  
ObsSet 0.0 kHz  
ObsFine 0.6121 Hz  
Point 8192  
Frequency(Span) 18115.94 Hz  
Scan 2280  
AcqTime 0.4416 s  
PD 4.0 s  
Pulse 6.9 μs  
IrrNuc <sup>1</sup>H  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl<sub>3</sub>  
Reference 77.0 ppm  
Broad.Factor 0.25 Hz  
Window Exponential  
RGain 36  
Operator \_\_\_\_\_



**5-(4-bromophenyl)furan-2(5H)-one (6bb).**

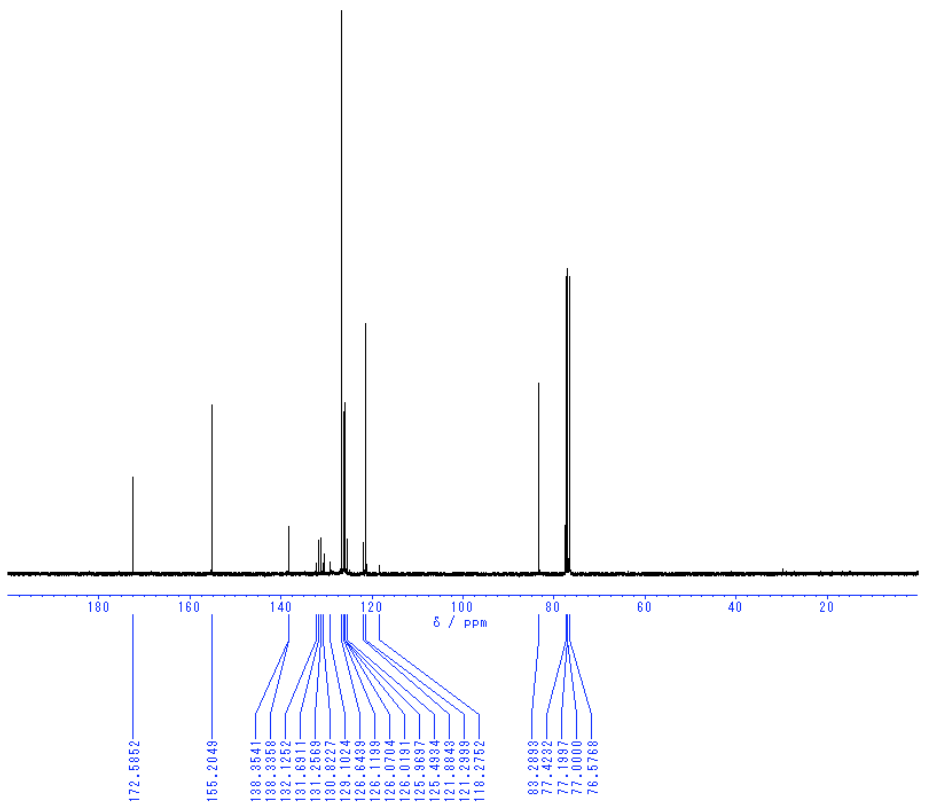
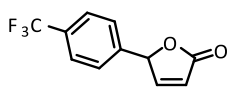


5-(4-trifluoromethylphenyl)furan-2(5H)-one (6cb).



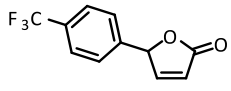
File C:\DOCUMENTS AND SETTINGS\TAKI\14デスクトップ\NMR\LITERATURE\LACTONE\5-(4-TRIFLUOROMETHYLPHENYL)FURAN-2(5H)-ONE\1H.RM1  
Date 05/Jul/2011 17:40:49  
Comment  
Std Proton parameters

ObsNuc <sup>1</sup>H  
ExMode NON  
ObsFreq 300.04 MHz  
ObsSet -1.0 kHz  
ObsFine 994.528 Hz  
Point 16884  
Frequency(Span) 4808.074 Hz  
Scan 8  
AcqTime 3.4095 s  
PD 6.5872 s  
Pulse 4.7 μs  
IrrNuc <sup>13</sup>C  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl<sub>3</sub>  
Reference 0.0004 ppm  
Broad.Factor 0.1466 Hz  
Window Exponential  
RGain 36  
Operator \_\_\_\_\_

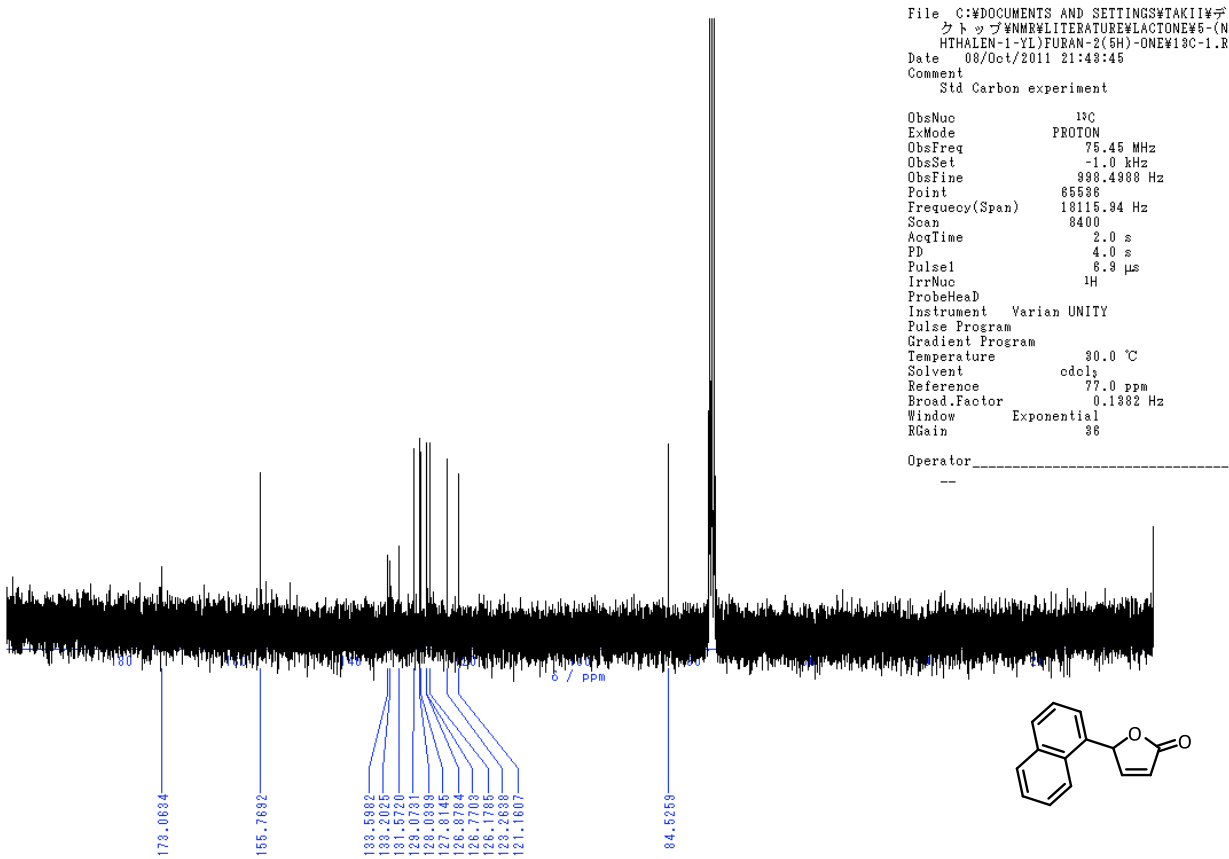
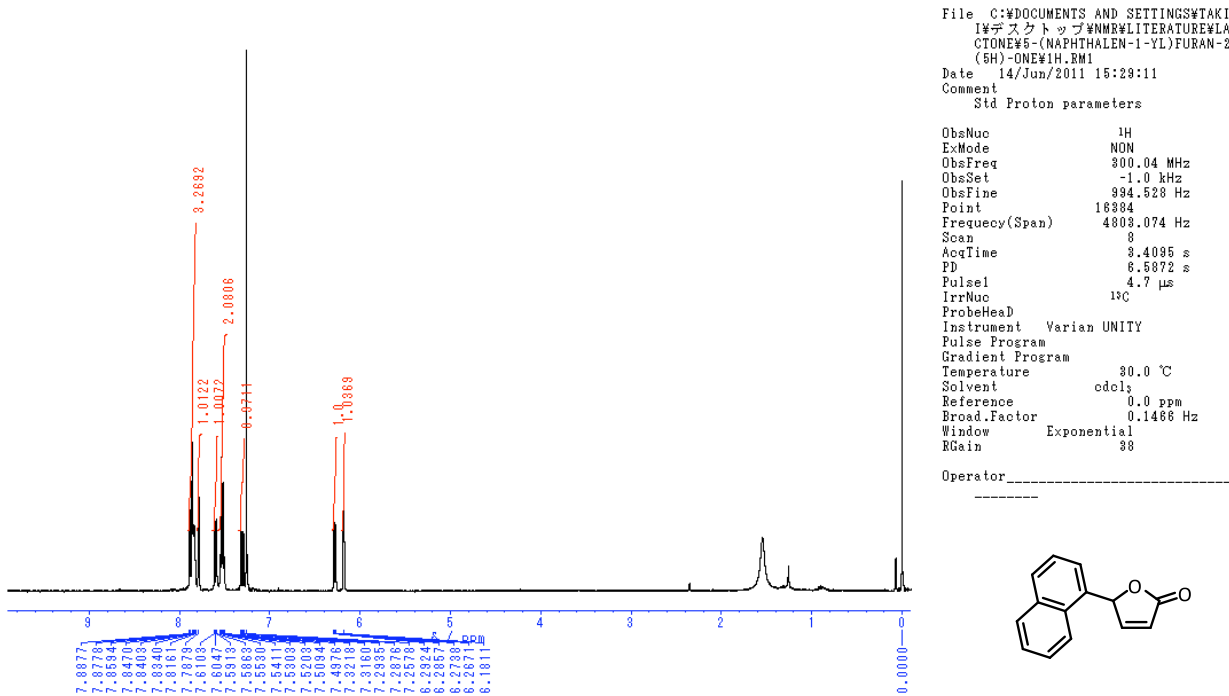


File C:\DOCUMENTS AND SETTINGS\TAKI\14デスクトップ\NMR\LITERATURE\LACTONE\5-(4-TRIFLUOROMETHYLPHENYL)FURAN-2(5H)-ONE\13C.RM1  
Date 06/Jul/2011 22:28:05  
Comment  
Std Carbon experiment

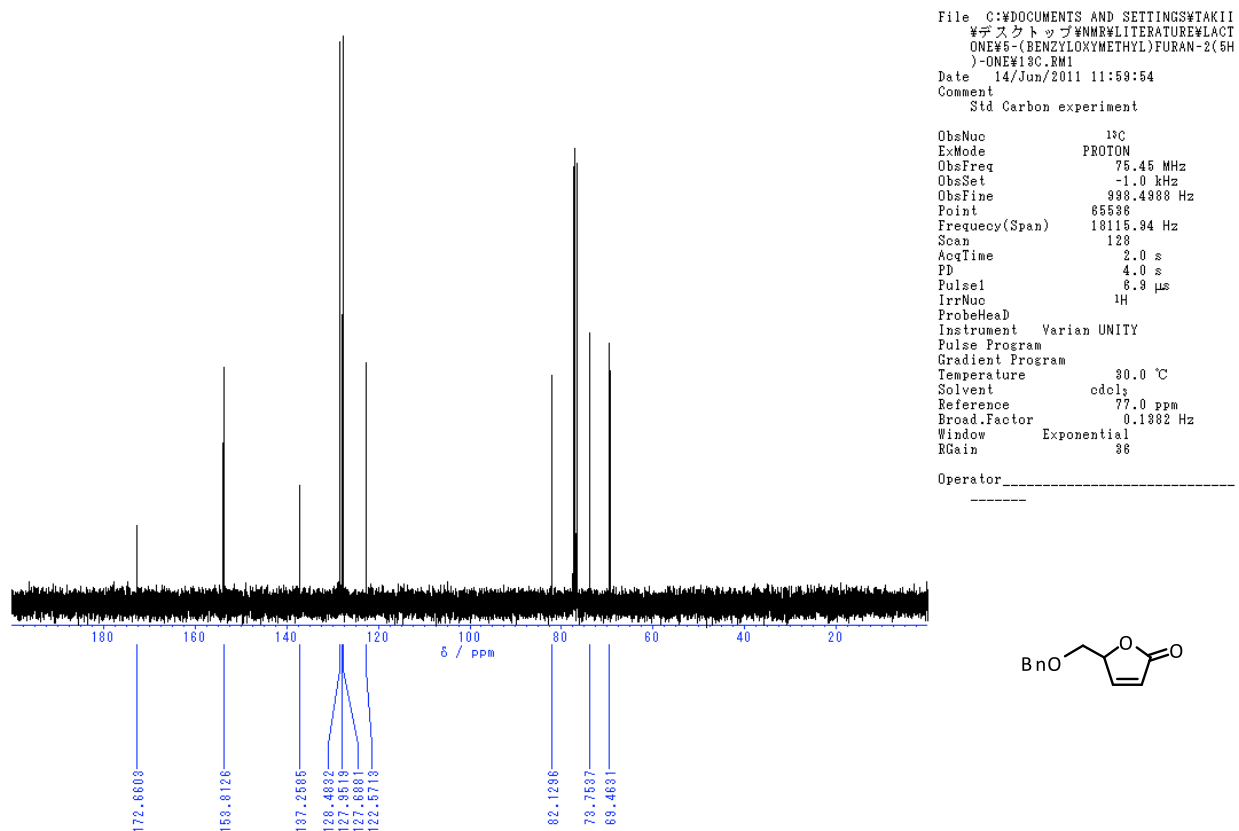
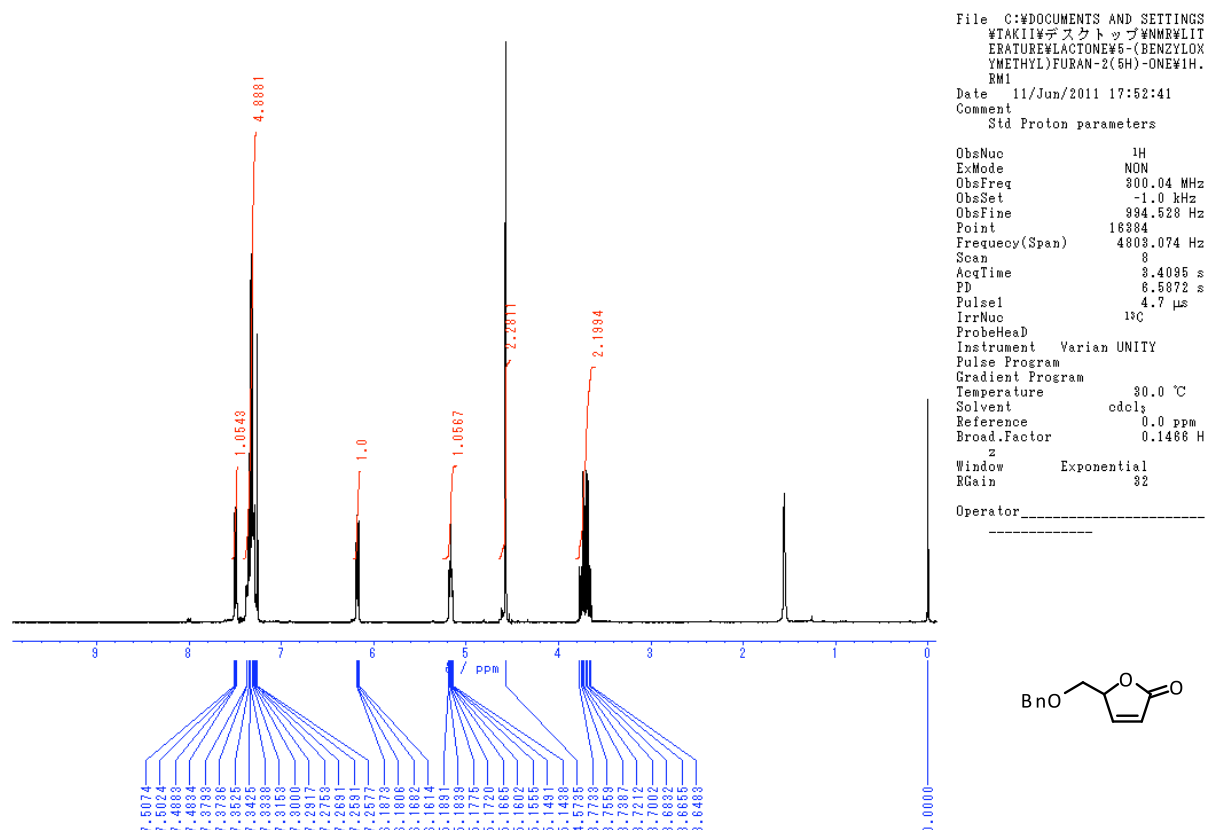
ObsNuc <sup>13</sup>C  
ExMode PROTON  
ObsFreq 75.45 MHz  
ObsSet -1.0 kHz  
ObsFine 998.4988 Hz  
Point 65536  
Frequency(Span) 18115.94 Hz  
Scan 6400  
AcqTime 2.0 s  
PD 4.0 s  
Pulse 6.9 μs  
IrrNuc <sup>1</sup>H  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl<sub>3</sub>  
Reference 77.0 ppm  
Broad.Factor 0.1382 Hz  
Window Exponential  
RGain 36  
Operator \_\_\_\_\_



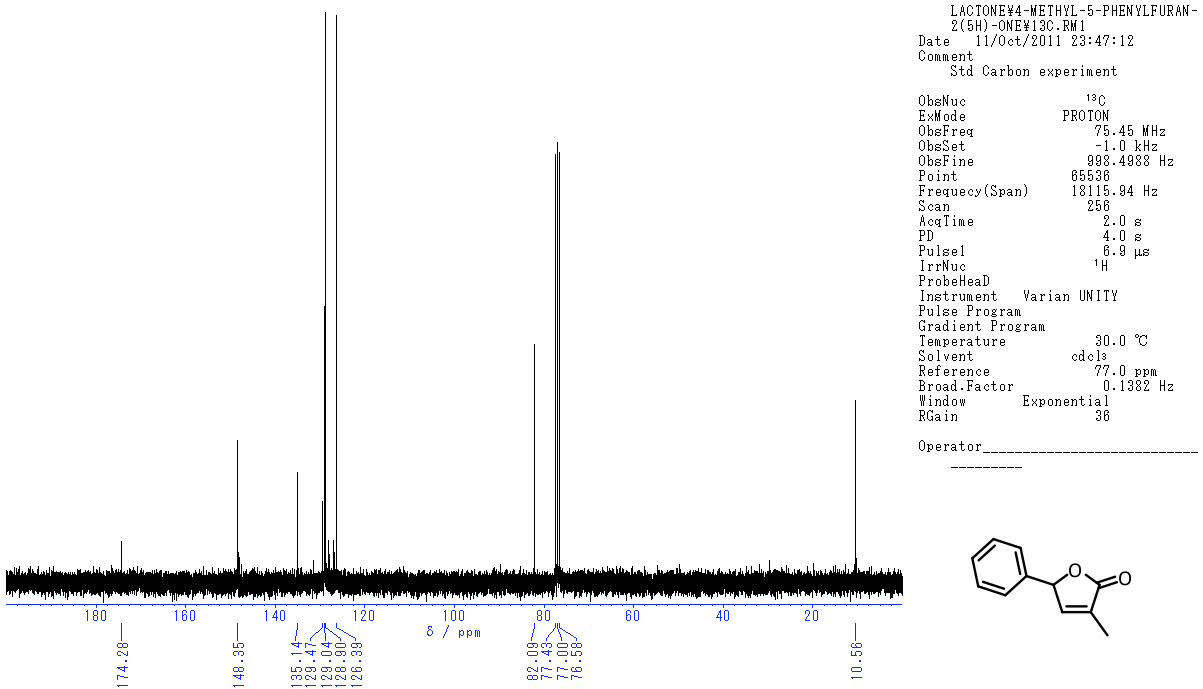
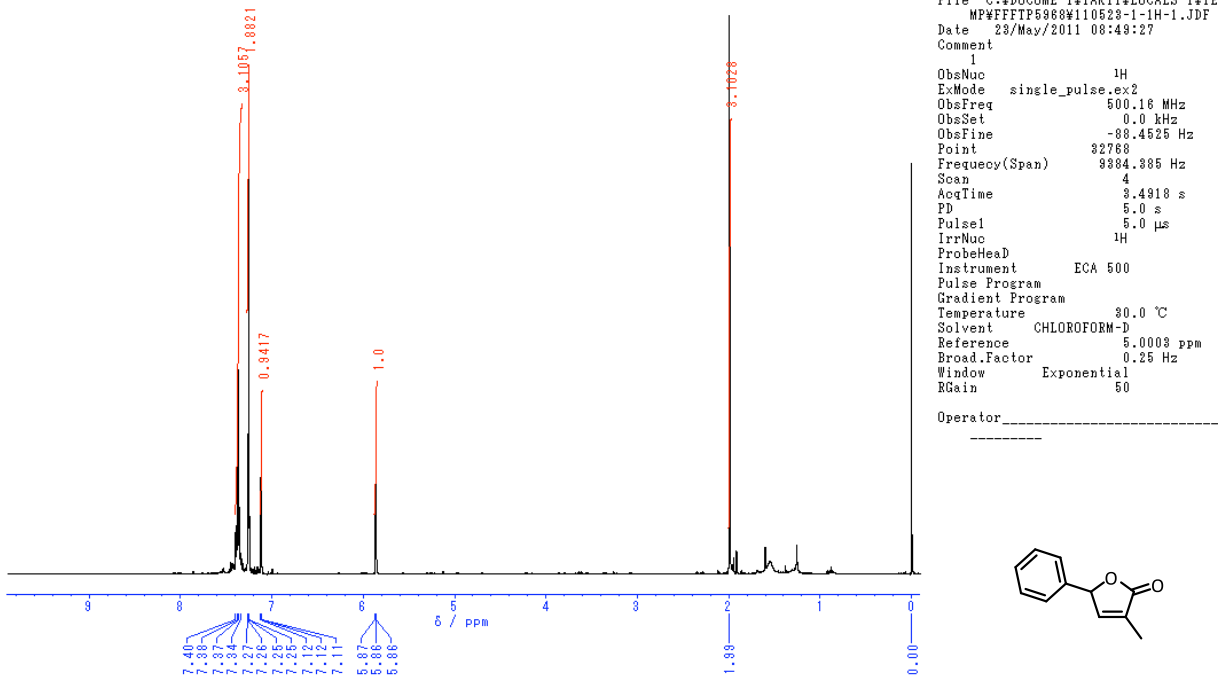
5-(naphthalene-1-yl)furan-2(5H)-one (6db).



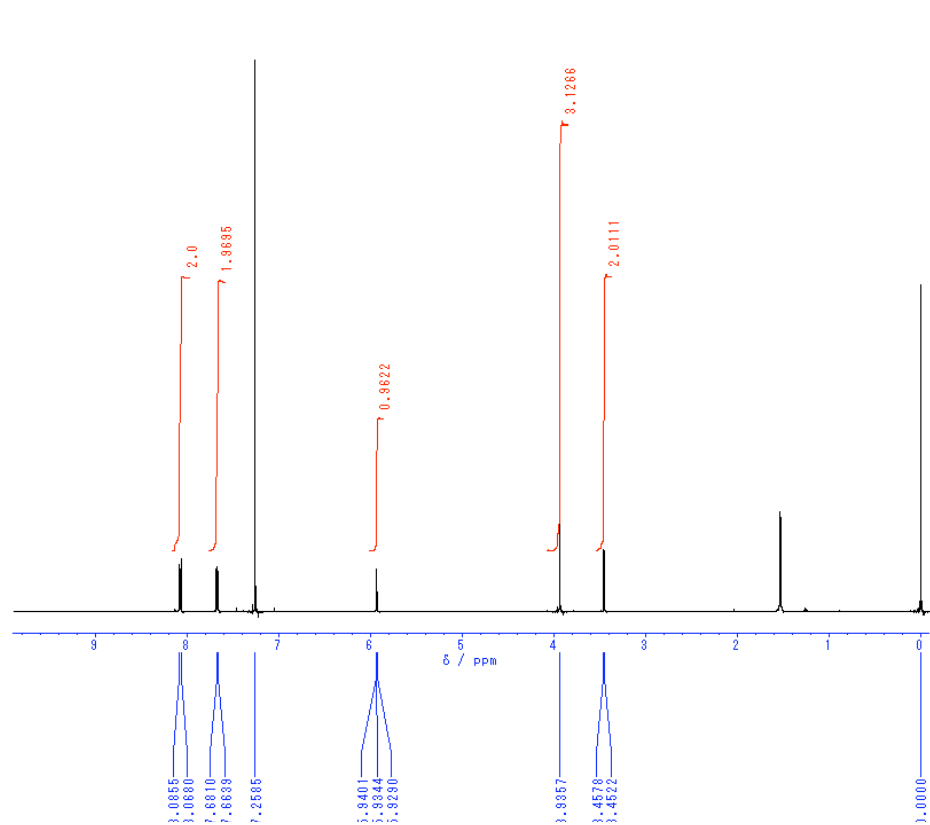
5-(benzyloxymethyl)furan-2(5H)-one (6fb).



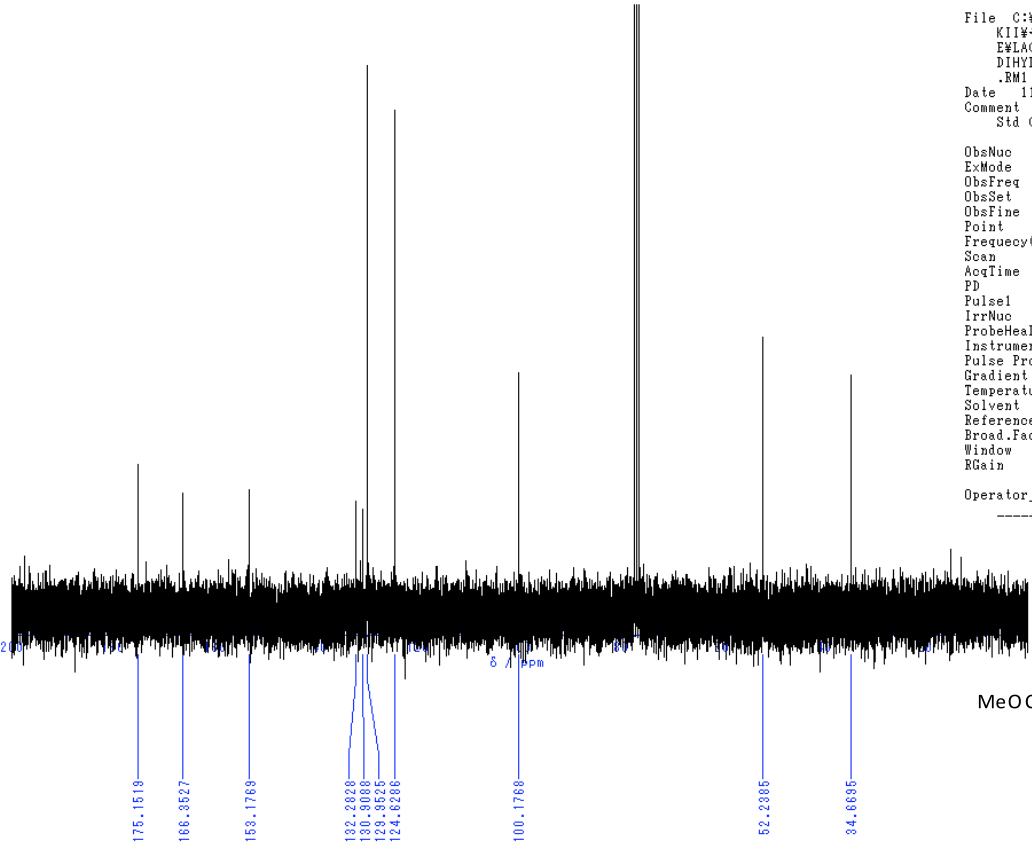
3-methyl-5-phenylfuran-2(5H)-one (6ac).



Methyl 4-(5-oxo-4,5-dihydrofuran-2-yl)benzoate (7).

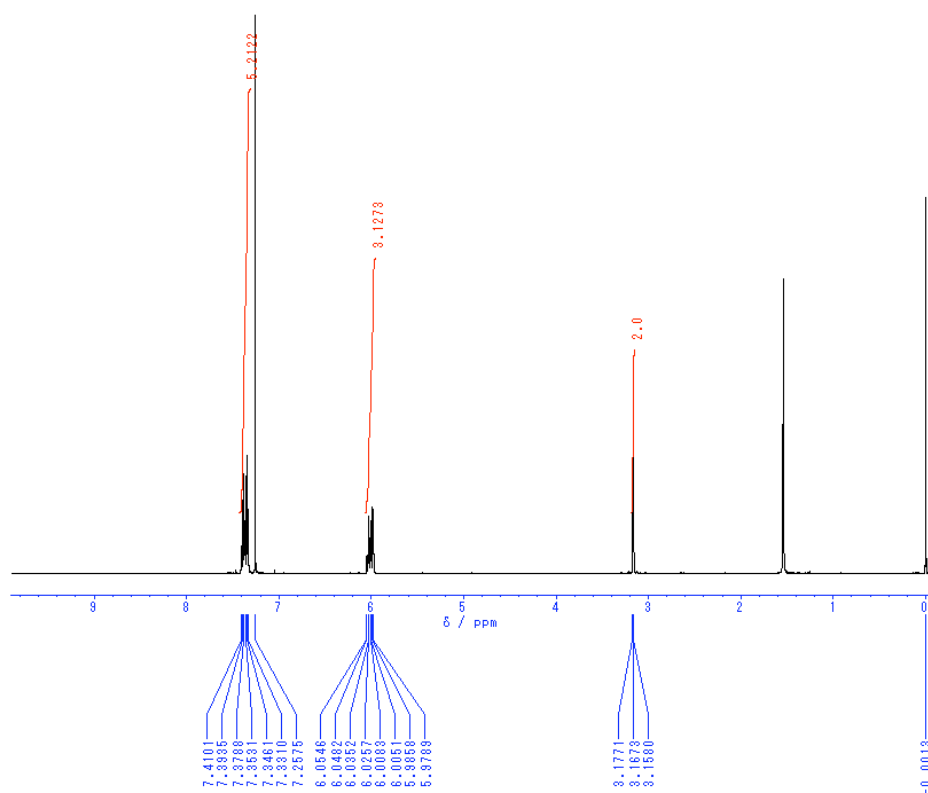


File C:\DOCUMENTS AND SETTINGS\TAKI  
I\デスクトップ\NMR\LITERATURE\LA  
CTONE\METHYL 4-(5-OXO-4,5-DIHYDR  
OFURAN-2-YL)BENZOATE\1H-1.RM1  
Date 28/Oct/2011 10:28:06  
Comment  
1  
ObsNuc 1H  
ExMode single\_pulse.ex2  
ObsFreq 500.16 MHz  
ObsSet 0.0 kHz  
ObsFine -88.4525 Hz  
Point 32768  
Frequency(Span) 9984.385 Hz  
Scan 16  
AcqTime 3.4918 s  
PD 5.0 s  
Pulse1 3.8 µs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.1 °C  
Solvent CHLOROFO  
Reference 0.0 ppm  
Broad.Factor 0.1432 Hz  
Window Exponential  
RGain 50  
Operator\_\_\_\_\_

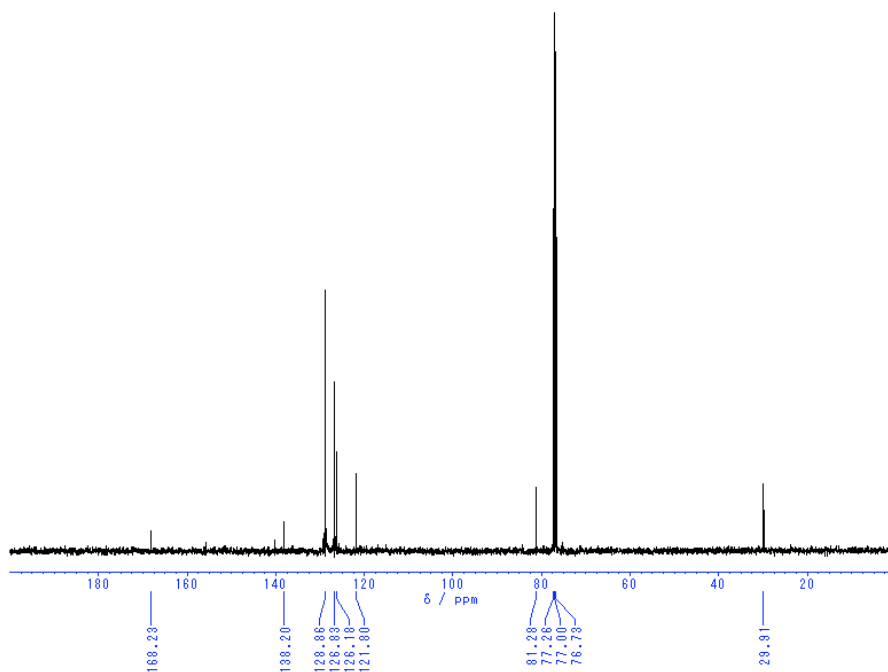
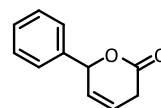


File C:\DOCUMENTS AND SETTINGS\TAKI  
KII\デスクトップ\NMR\LITERATUR  
E\LACTONE\METHYL 4-(5-OXO-4,5-  
DIHYDROFURAN-2-YL)BENZOATE\13C  
.RM1  
Date 11/Oct/2011 23:32:04  
Comment  
Std Carbon experiment  
ObsNuc 13C  
ExMode PROTON  
ObsFreq 75.45 MHz  
ObsSet -1.0 kHz  
ObsFine 998.4988 Hz  
Point 65536  
Frequency(Span) 18115.94 Hz  
Scan 64  
AcqTime 2.0 s  
PD 4.0 s  
Pulse1 5.3 µs  
IrrNuc 1H  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl3  
Reference 77.0 ppm  
Broad.Factor 0.1382 Hz  
Window Exponential  
RGain 36  
Operator\_\_\_\_\_

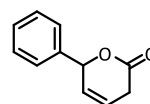
**6-phenyl-3,6-dihydro-2H-pyran-2-one (8).**



File C:\DOCUMENTS AND SETTINGS\WTAKI\W  
KII\デスクトップ\WMMR\LITERATUR  
EWLACTONE\6-PHENYL-3,6-DIHYDRO  
-2H-PYRAN-2-ONE\1H.RM1  
Date 02/Sep/2011 17:52:28  
Comment  
1  
ObsNuc 1H  
ExMode single\_pulse.ex2  
ObsFreq 500.16 MHz  
ObsSet 0.0 kHz  
ObsPine -88.4525 Hz  
Point 8558  
Frequency(Span) 8884.885 Hz  
Scan 8  
AcqTime 6.9895 s  
PD 5.0 s  
Pulse1 6.2 μs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFO  
Reference 5.0042 ppm  
Broad.Factor 0.0716 Hz  
Window Exponential  
RGain 46  
Operator \_\_\_\_\_

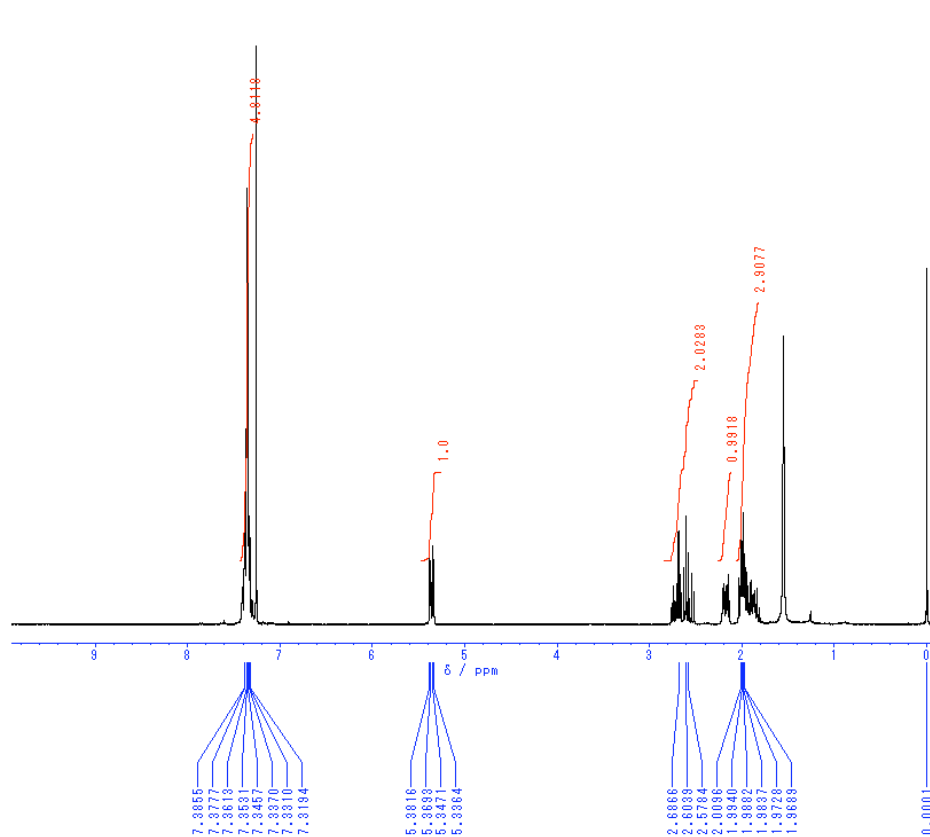


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デスクトップ\WMMR\LITERATURE\W  
LACTONE\6-PHENYL-3,6-DIHYDRO-2H-PYRAN-2-ON  
EW13C.RM1  
Date 26/Dec/2011 08:40:57  
Comment  
1  
ObsNuc 13C  
ExMode single\_pulse\_dec  
ObsFreq 125.77 MHz  
ObsSet -5.0 kHz  
ObsPine 301.0403 Hz  
Point 8192  
Frequency(Span) 39308.18 Hz  
Scan 3051  
AcqTime 0.2084 s  
PD 2.0 s  
Pulse1 5.0 μs  
IrrNuc 1H  
ProbeHead  
Instrument ECA 500  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent CHLOROFORM-D  
Reference 77.0 ppm  
Broad.Factor 0.25 Hz  
Window Exponential  
RGain 50  
Operator \_\_\_\_\_

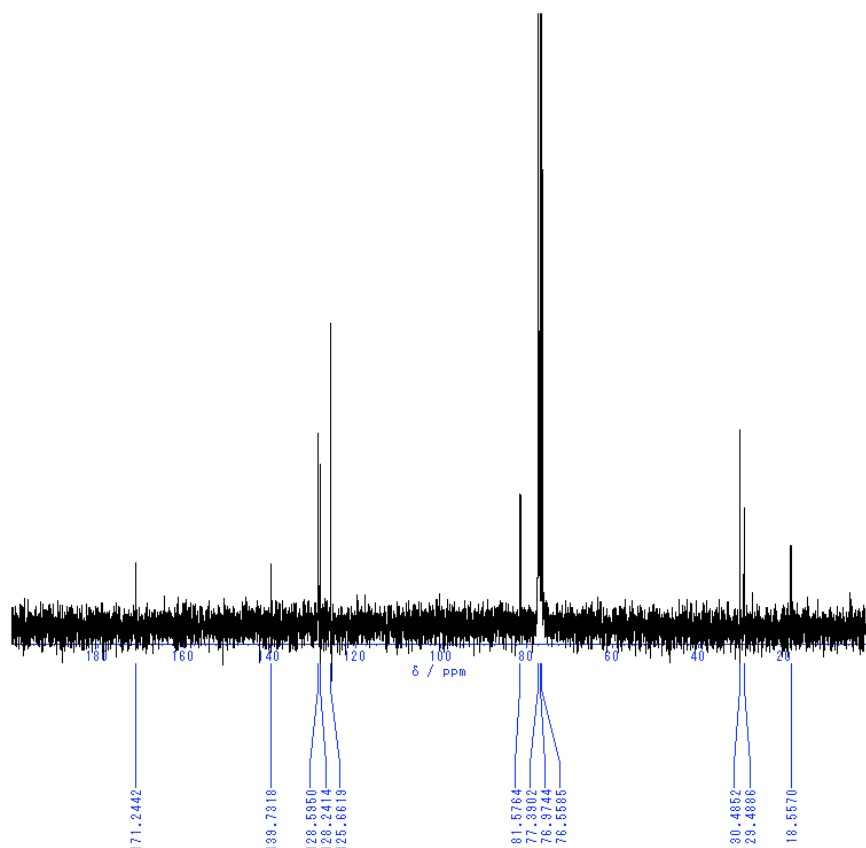
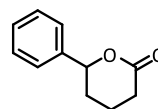




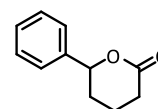
6-phenyltetrahydro-2H-pyran-2-one (10).



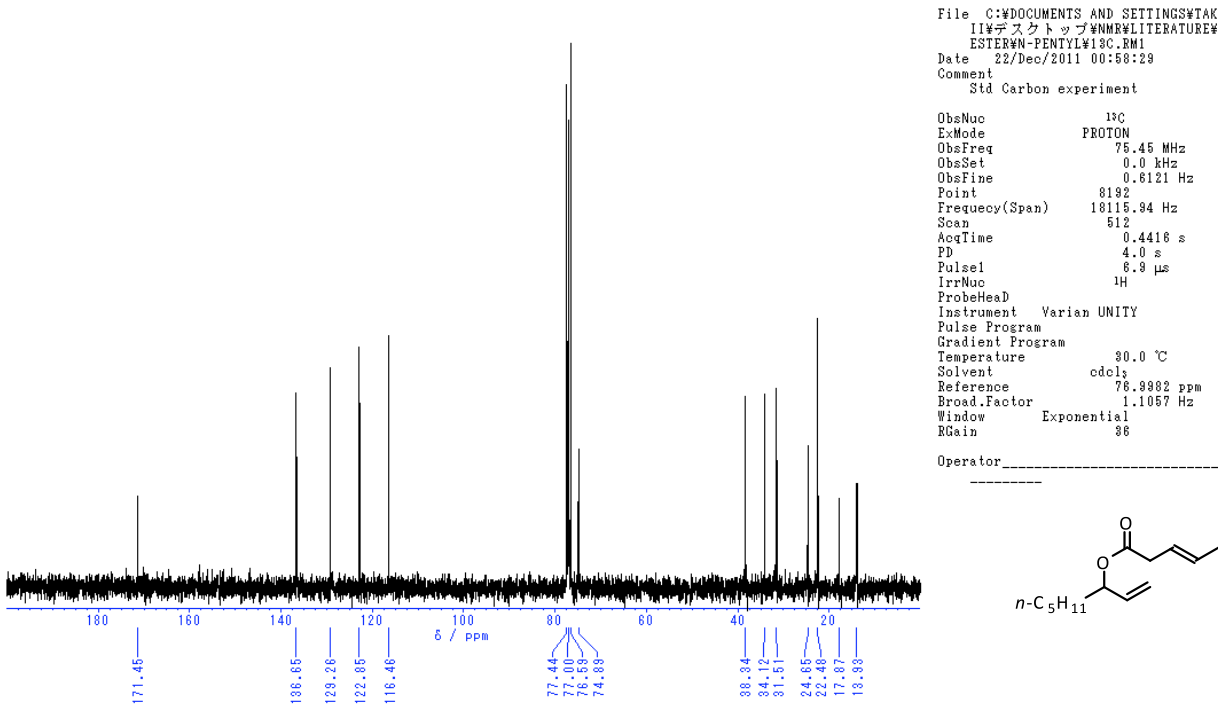
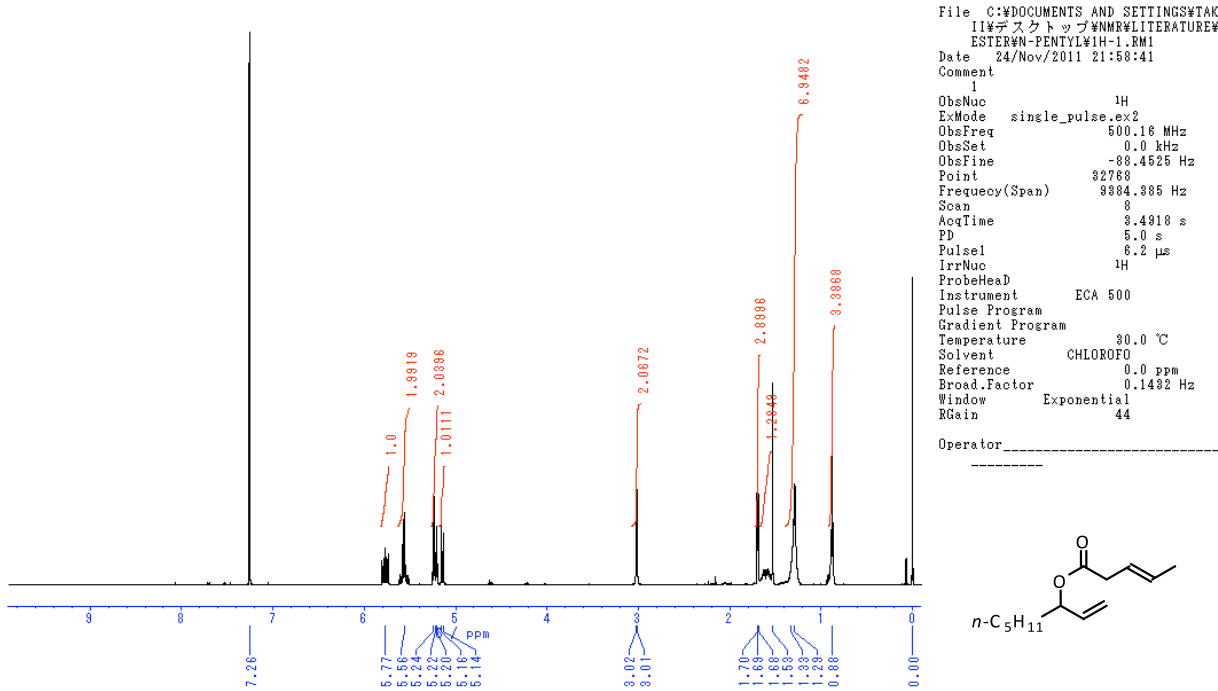
File C:\DOCUMENTS AND SETTINGS\TAKII\デスクトップ\NMR\LITERATURE\LACTONE\6-PHENYL-TETRAHYDRO-2H-PYRAN-2-ONE\1H.RM1  
Date 04/Sep/2011 00:47:34  
Comment  
Std Proton parameters  
ObsNuc <sup>1</sup>H  
ExMode NON  
ObsFreq 300.04 MHz  
ObsSet -1.0 kHz  
ObsFine 994.528 Hz  
Point 18384  
Frequency(Span) 4803.074 Hz  
Scan 8  
AcqTime 3.4095 s  
PD 6.5872 s  
Pulse1 4.7 μs  
IrrNuc <sup>13</sup>C  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl<sub>3</sub>  
Reference 0.0001 ppm  
Broad.Factor 0.1466 Hz  
Window Exponential  
RGain 36  
Operator\_\_\_\_\_



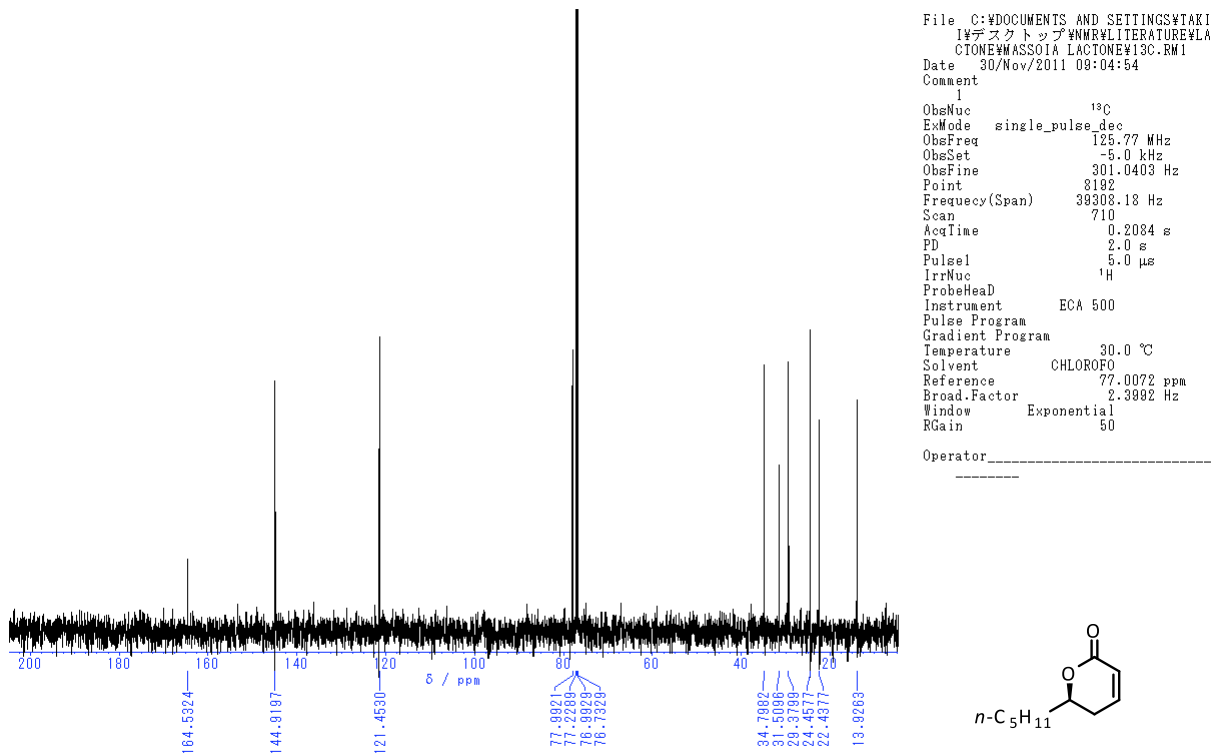
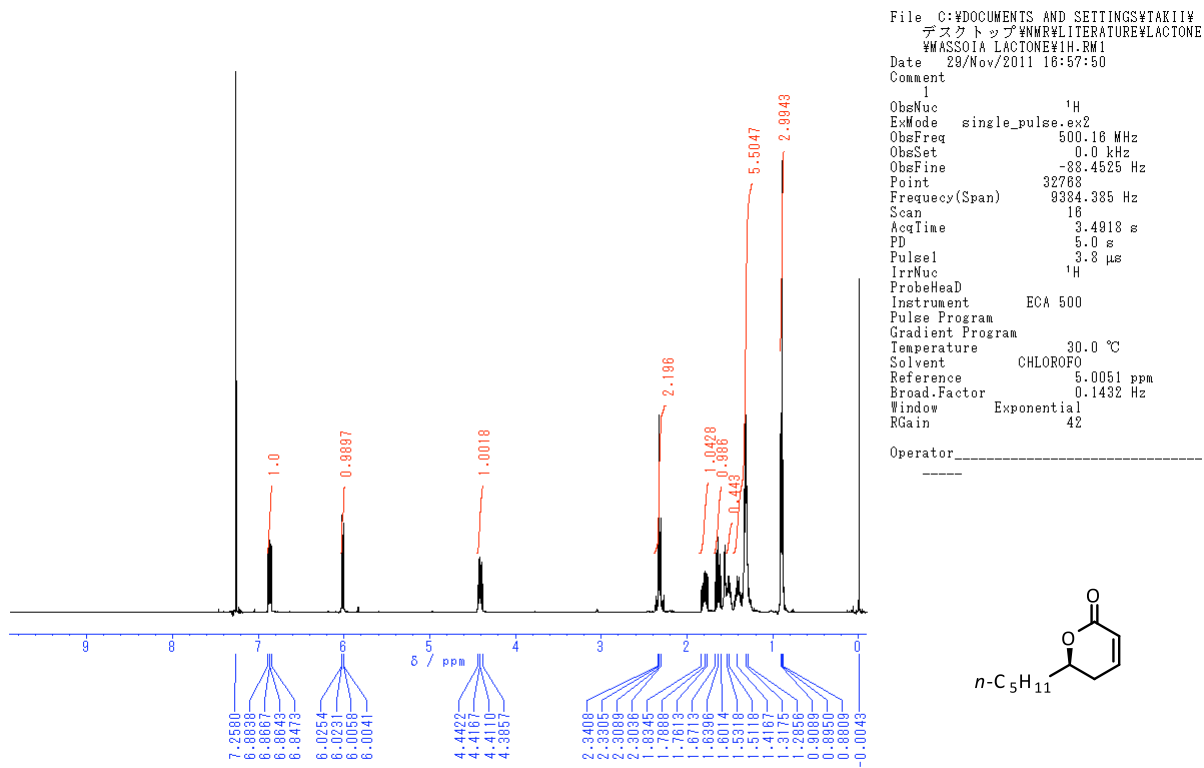
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Date 13/Oct/2011 23:45:15  
Comment  
Std Carbon experiment  
ObsNuc <sup>13</sup>C  
ExMode PROTON  
ObsFreq 75.46 MHz  
ObsSet -1.0 kHz  
ObsFine 998.4968 Hz  
Point 8192  
Frequency(Span) 18115.94 Hz  
Scan 7600  
AcqTime 0.4416 s  
PD 4.0 s  
Pulse1 8.9 μs  
IrrNuc <sup>1</sup>H  
ProbeHead  
Instrument Varian UNITY  
Pulse Program  
Gradient Program  
Temperature 30.0 °C  
Solvent cdcl<sub>3</sub>  
Reference 105.0261 ppm  
Broad.Factor 1.1057 Hz  
Window Exponential  
RGain 36  
Operator\_\_\_\_\_



(E)-oct-1-en-3-yl pent-3-enoate (4gd).

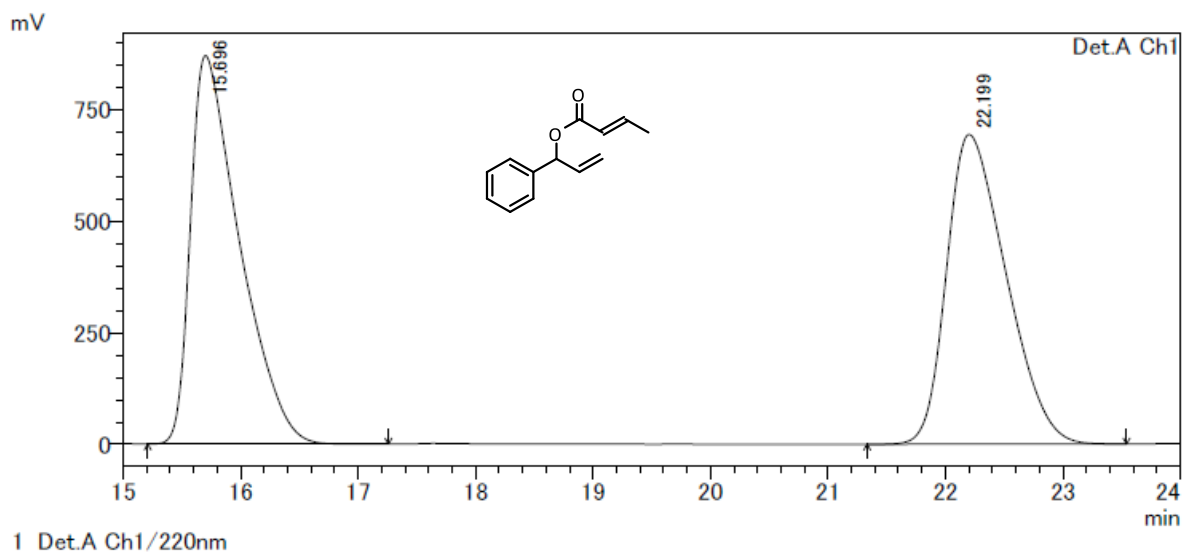


**(R)-6-pentyl-5,6-dihydro-2H-pyran-2-one ((R)-(-)-massoialactone) (14).**



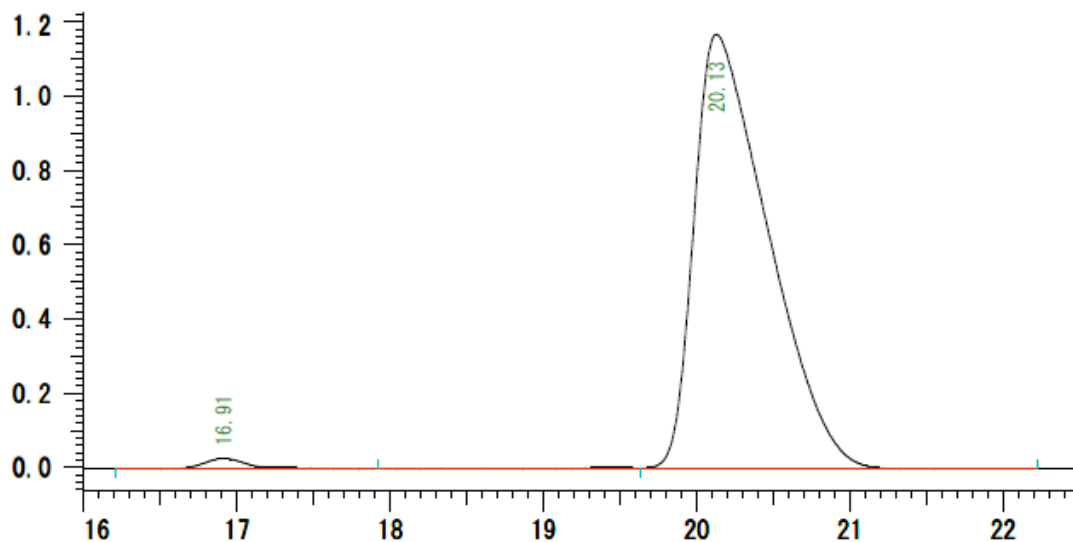
## HPLC data

### (*E*)-1-phenylallyl but-2-enoate (4ab)



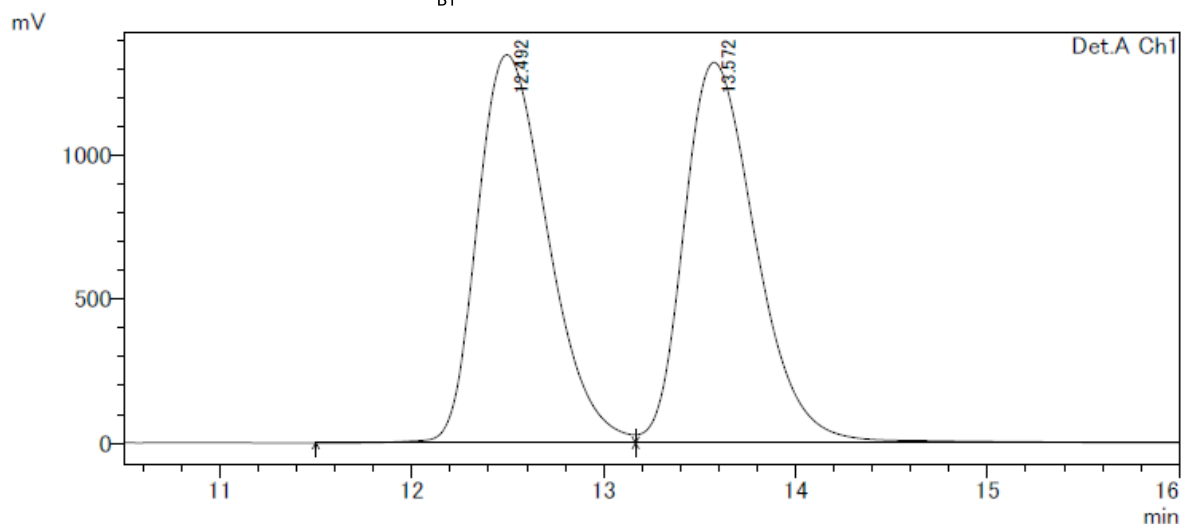
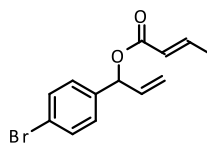
検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	15.696	25277961	50.841
2	22.199	24441514	49.159
合計		49719475	100.000



	Retention Time	Area	Area%
1	16.91	319441	1.600
2	20.13	19639764	98.400
		19959205	100.000

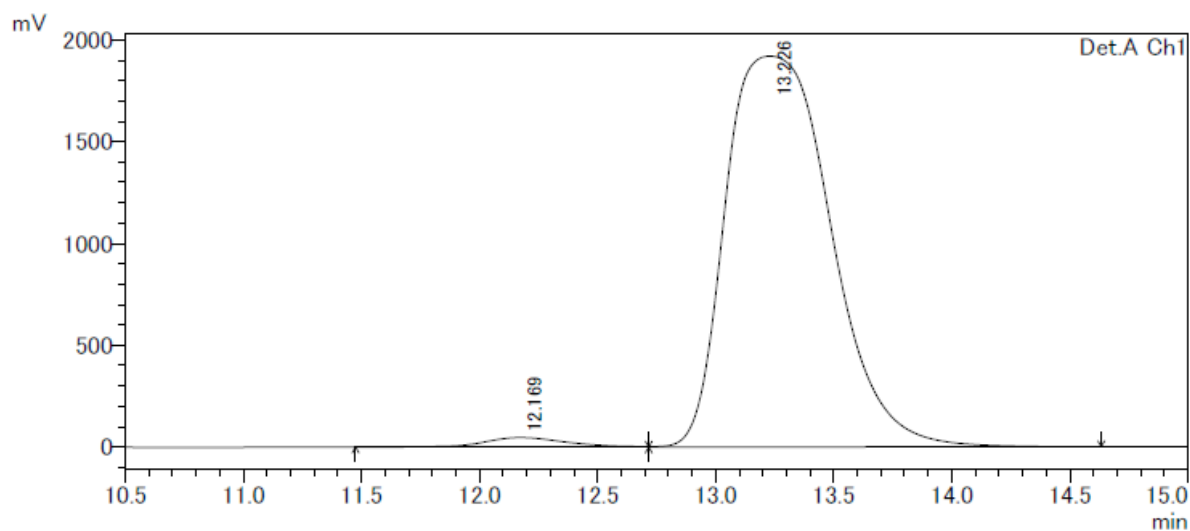
**(E)-1-(4-bromophenyl)allyl but-2-enoate (4bb).**



1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	12.492	34361132	49.776
2	13.572	34670327	50.224
合計		69031459	100.000

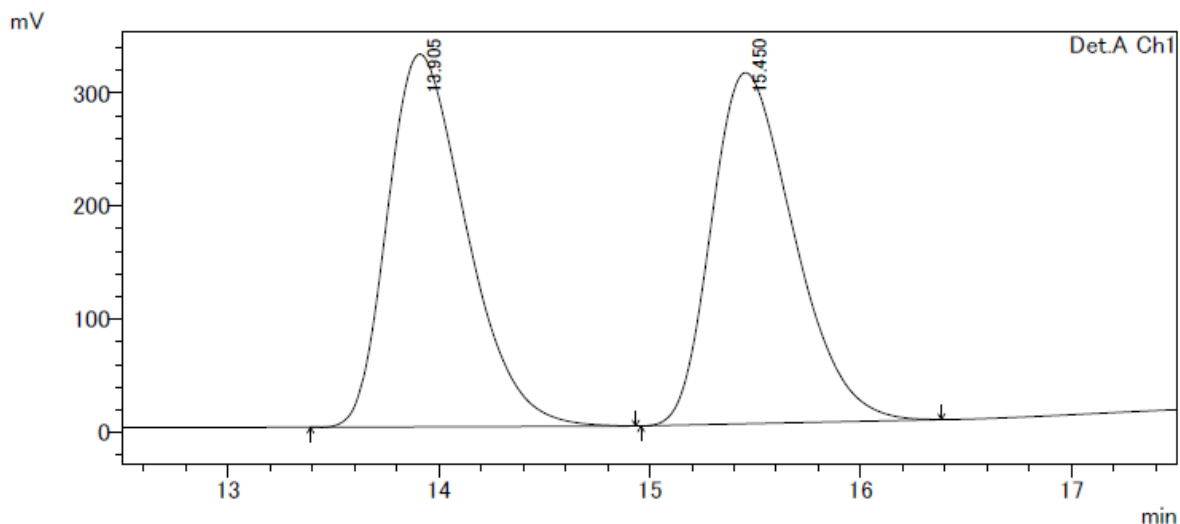
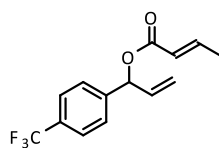


1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	12.169	1113180	1.841
2	13.226	59364144	98.159
合計		60477324	100.000

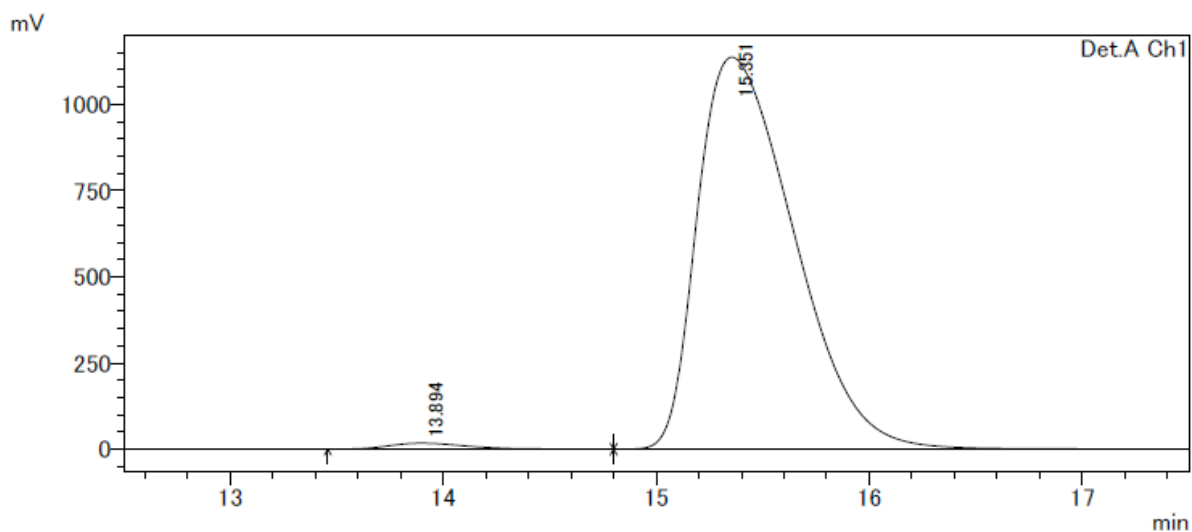
**(E)-1-(4-trifluoromethylphenyl)allyl but-2-enoate (4cb).**



1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	13.905	8574623	50.389
2	15.450	8442226	49.611
合計		17016849	100.000

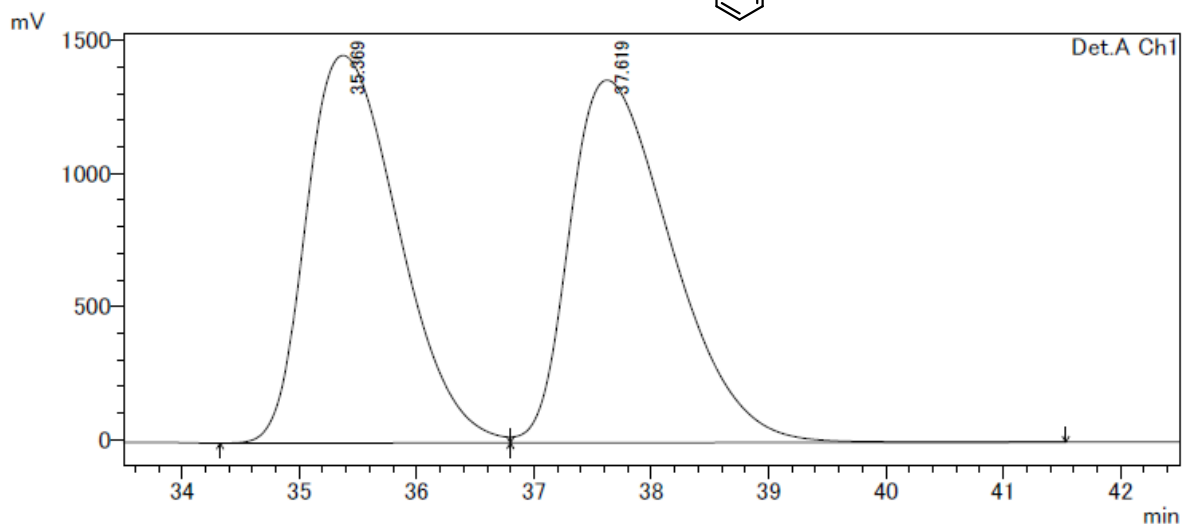
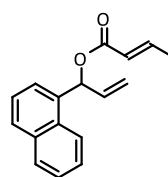


1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retentin Time	Area	Area%
1	13.894	427370	1.181
2	15.351	35762861	98.819
合計		36190232	100.000

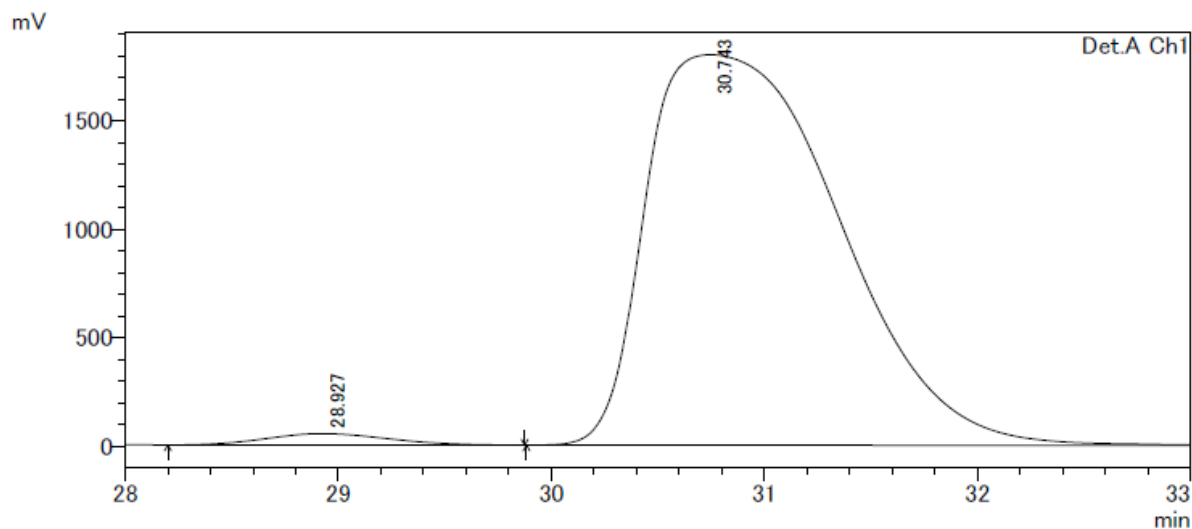
**(E)-1-(naphthalene-1-yl)allyl but-2-enoate (4db).**



1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	35.369	79658155	49.102
2	37.619	82572388	50.898
合計		162230544	100.000

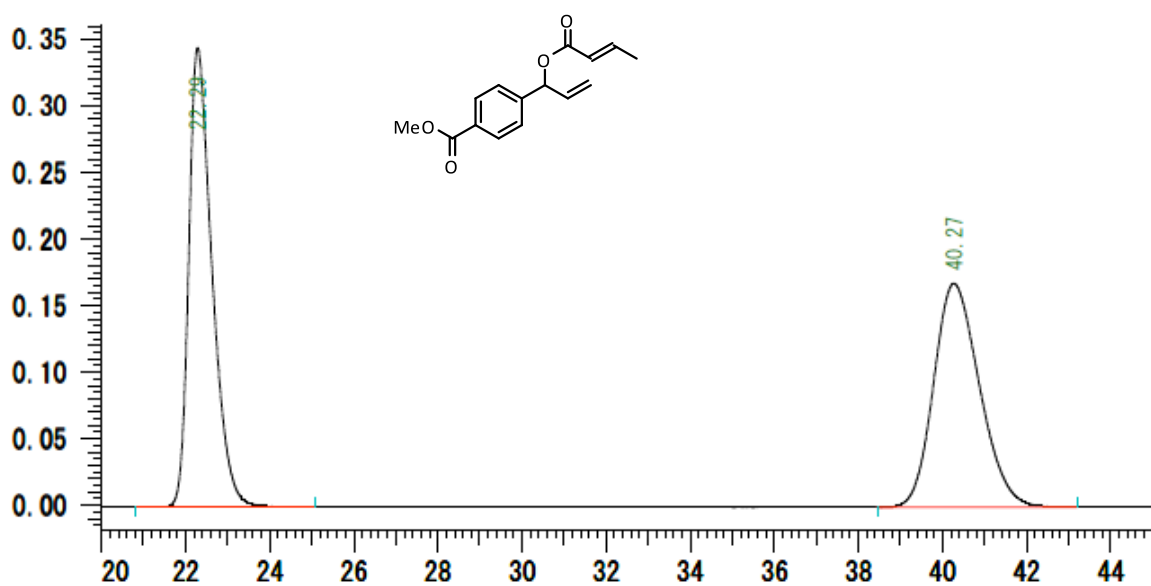


1 Det.A Ch1/220nm

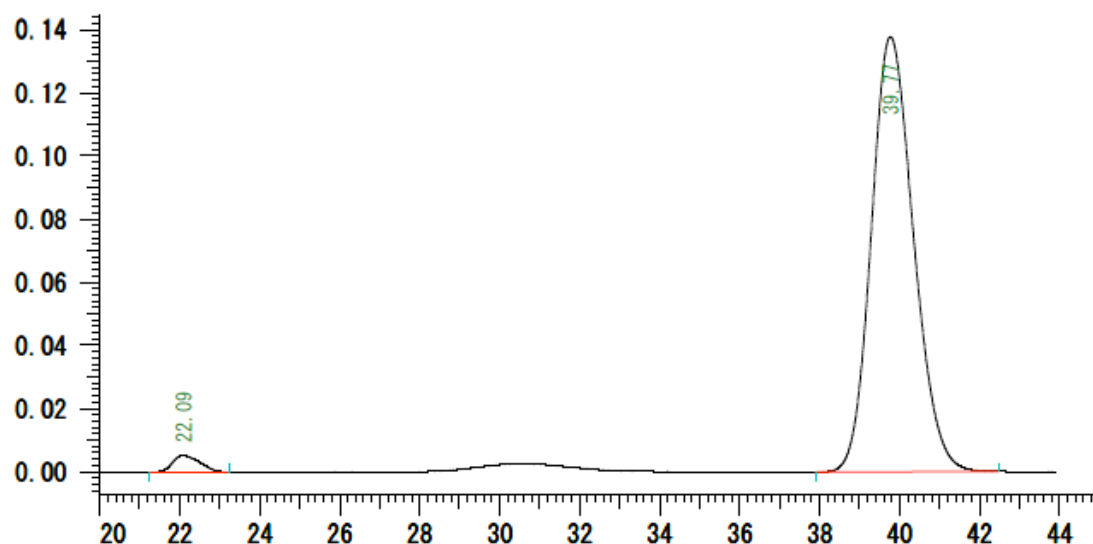
検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	28.927	2075457	1.804
2	30.743	112965443	98.196
合計		115040900	100.000

**(E)-methyl 4-(1-(but-2-enyloxy)allyl)benzoate (4eb)**

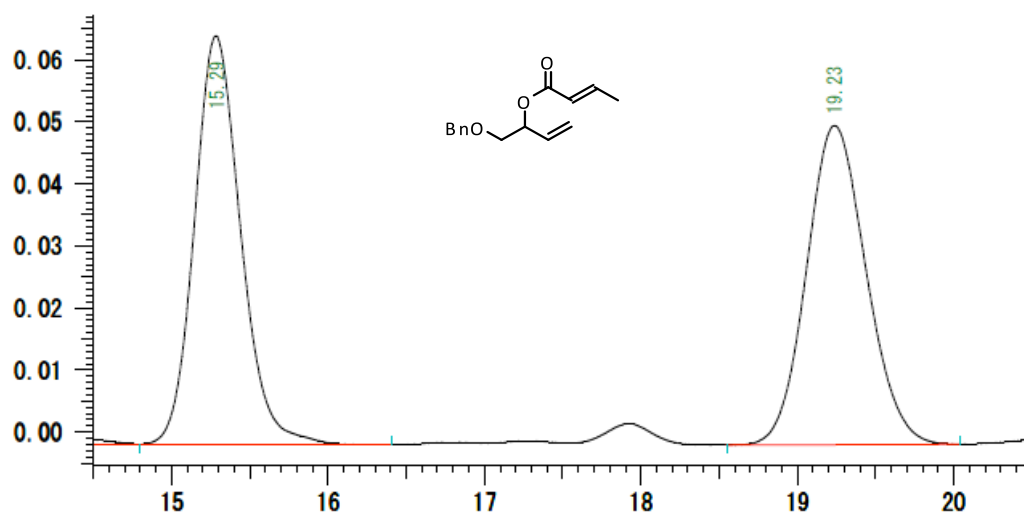


	Retention Time	Area	Area%
1	22.29	6587865	50.788
2	40.27	6383555	49.212
		12971420	100.000

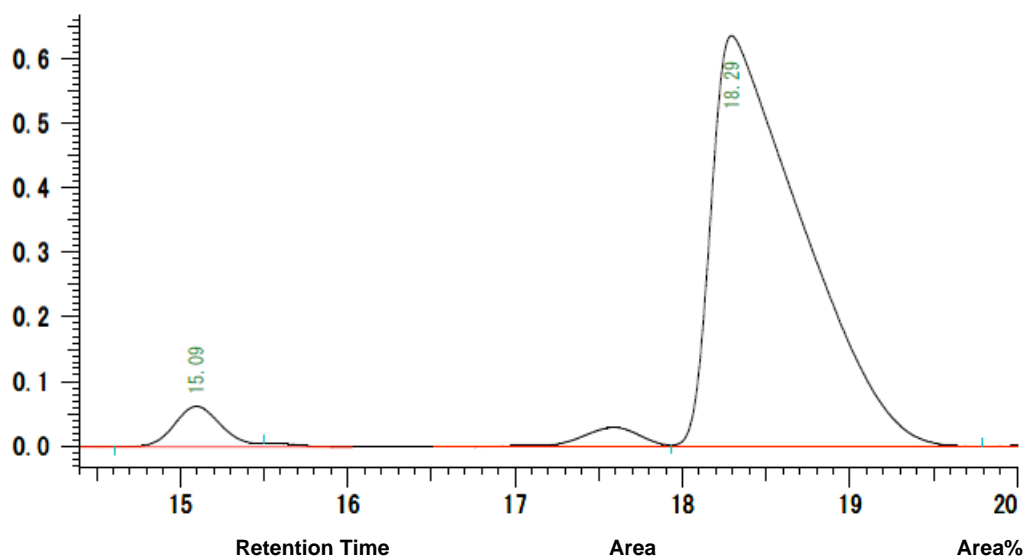


	Retention Time	Area	Area%
1	22.09	129310	2.495
2	39.77	5053089	97.505
		5182399	100.000

**(*E*)-1-(benzyloxy)but-3-en-2-yl but-2-enoate (4fb).**

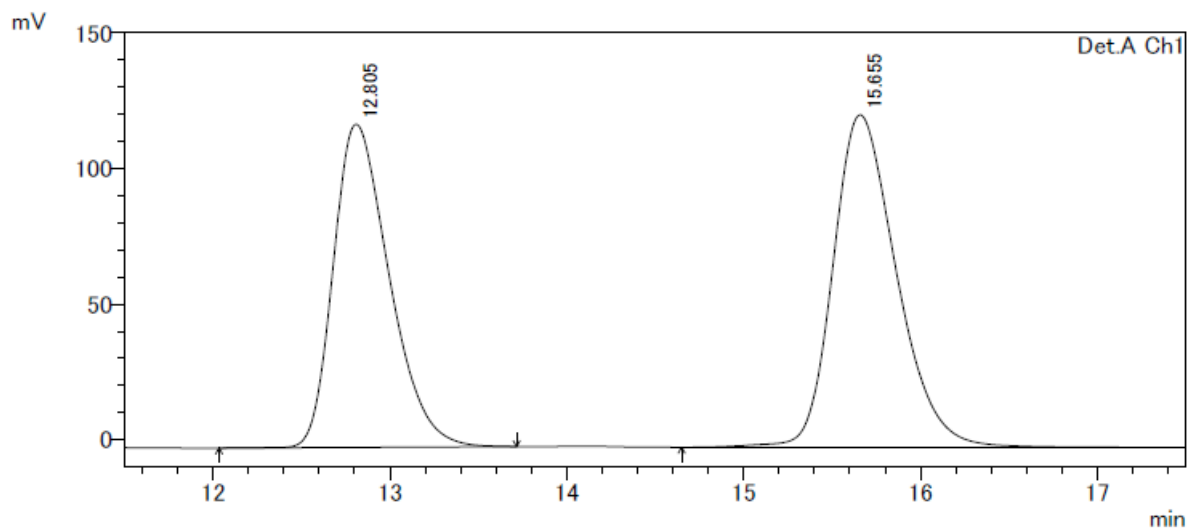
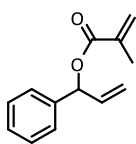


	Retention Time	Area	Area%
1	15.29	679055	49.930
2	19.23	680972	50.070
		1360027	100.000



	Retention Time	Area	Area%
1	15.09	619329	4.874
2	18.29	12087422	95.126
		12706751	100.000

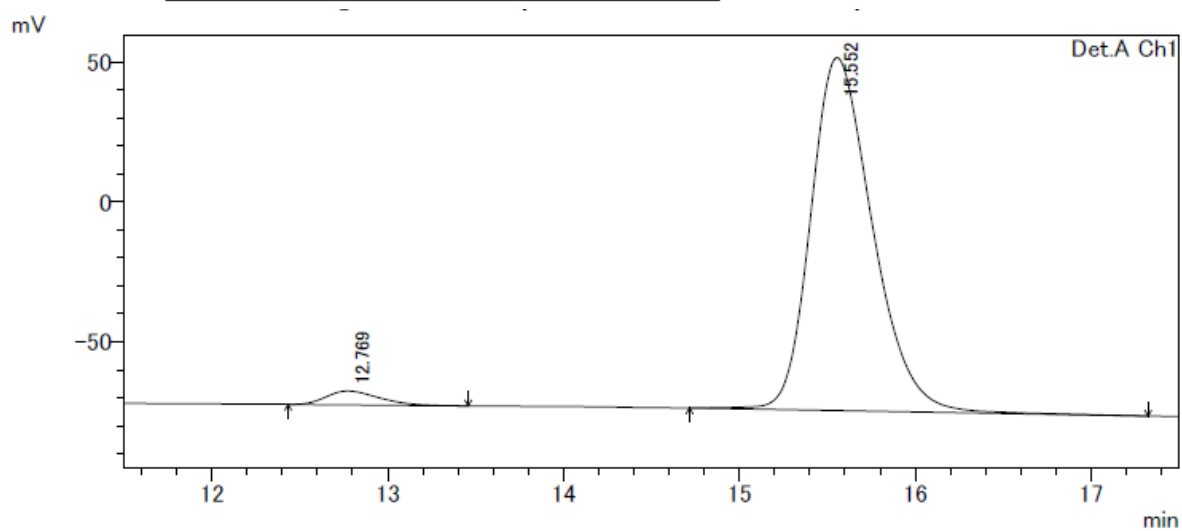
**(*E*)-1-phenylallyl methacrylate (4ac).**



1 Det.A Ch1/220nm

UV-Vis Ch1 220nm

Peak	Retention Time	Area	Area%
1	12.805	2619011	46.047
2	15.655	3068721	53.953
合計		5687732	100.000

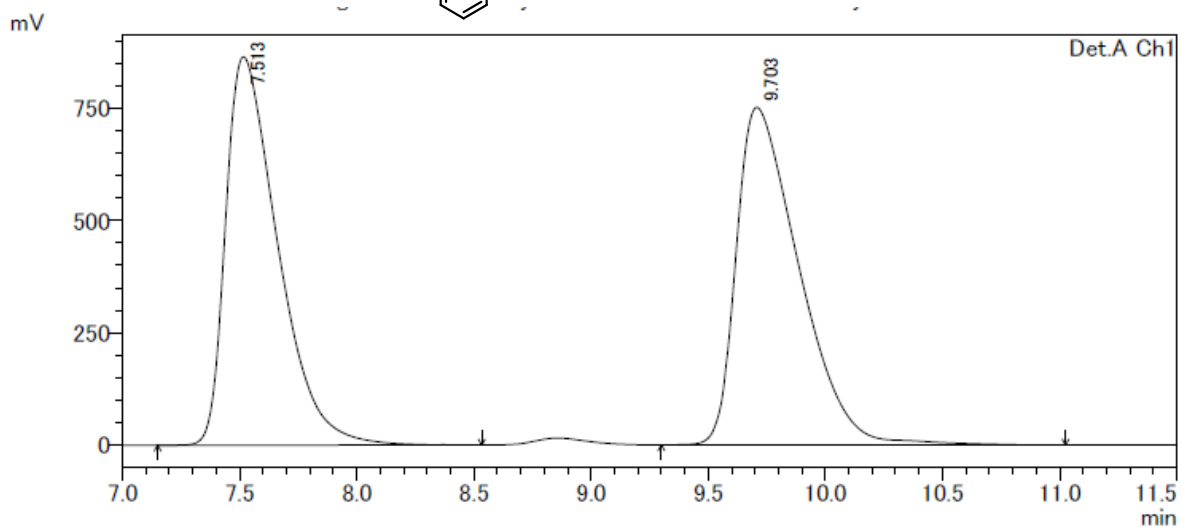
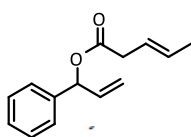


1 Det.A Ch1/220nm

UV-Vis Ch1 220nm

Peak	Retention Time	Area	Area%
1	12.769	103542	3.245
2	15.552	3087444	96.755
合計		3190986	100.000

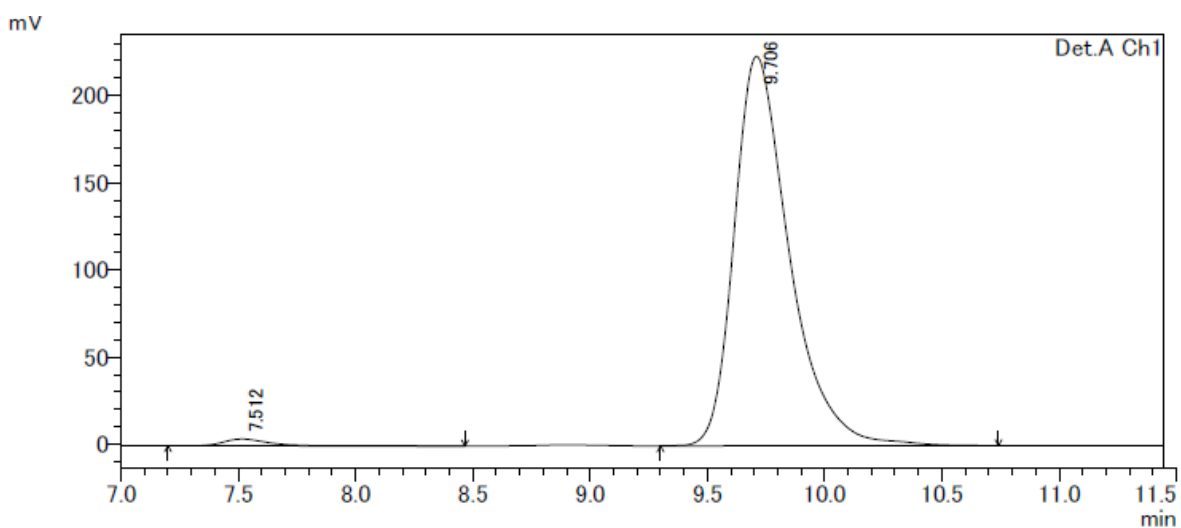
**(E)-1-phenylallyl pent-3-enoate (4ad).**



1 Det.A Ch1/220nm

検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	7.513	13362232	48.855
2	9.703	13988448	51.145
合計		27350680	100.000

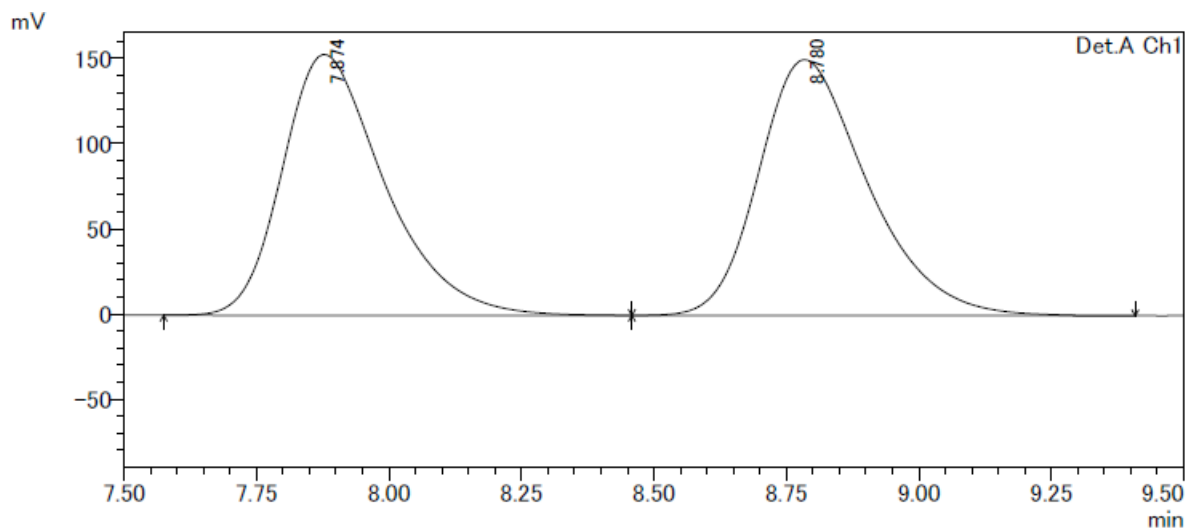
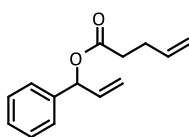


1 Det.A Ch1/220nm

検出器A Ch1 220nm

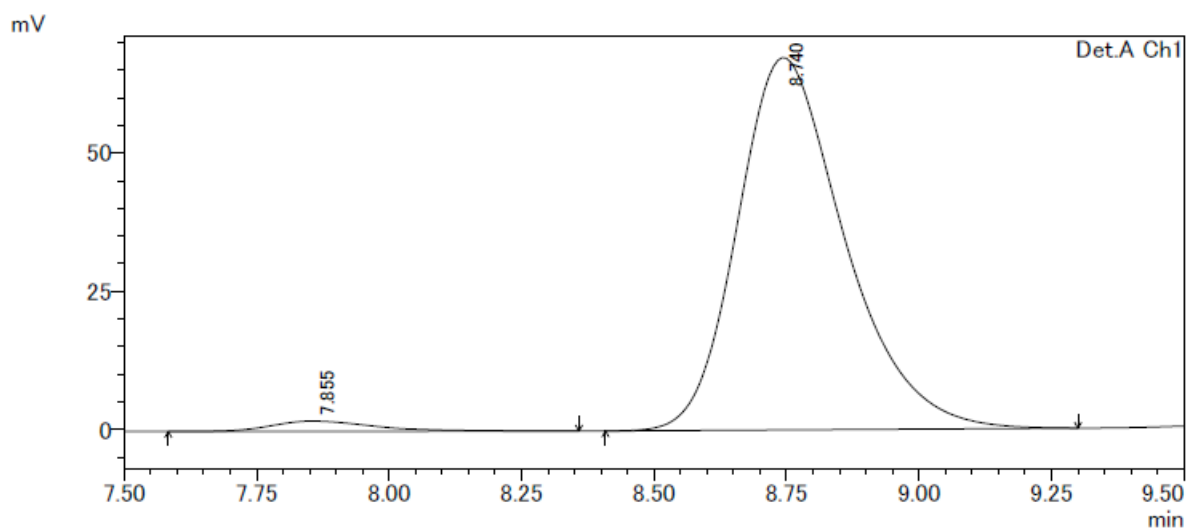
Peak	Retention Time	Area	Area%
1	7.512	53148	1.435
2	9.706	3650678	98.565
合計		3703826	100.000

**1-phenylallyl pent-4-enoate (4ae).**



検出器A Ch1 220nm

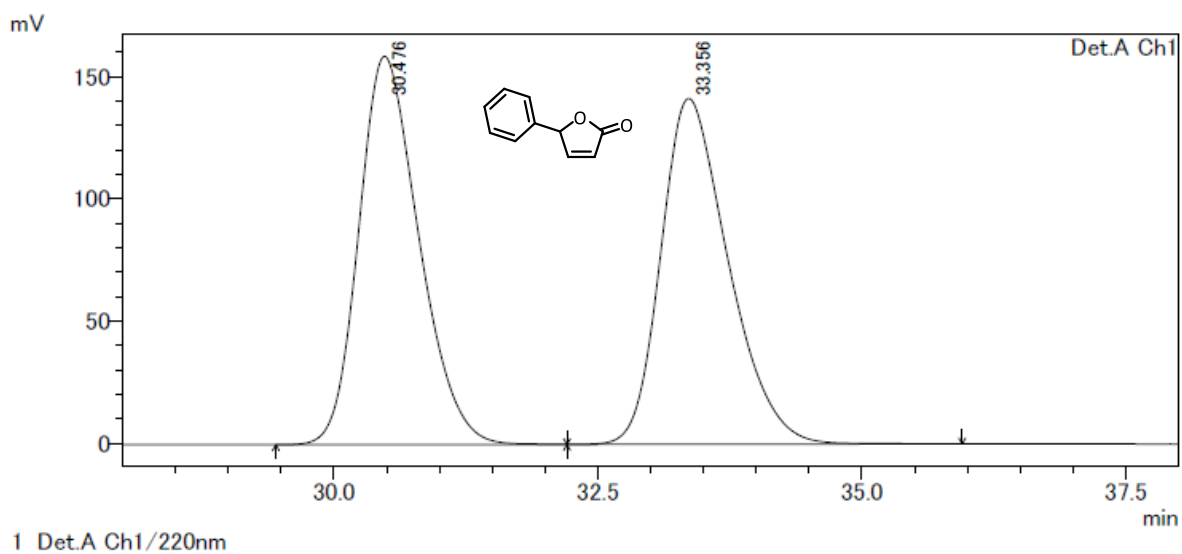
Peak	Retention Time	Area	Area%
1	7.874	2055447	48.861
2	8.780	2151307	51.139
合計		4206753	100.000



検出器A Ch1 220nm

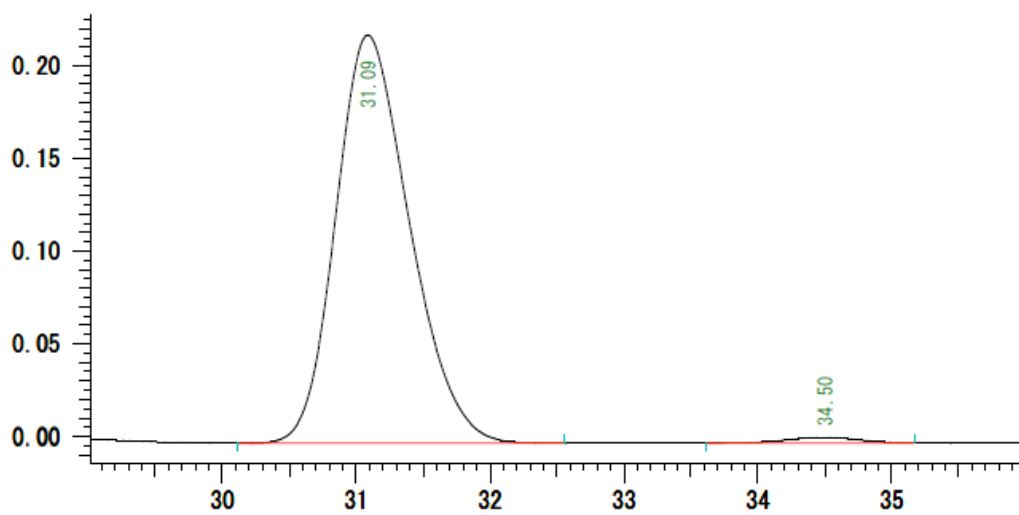
Peak	Retention Time	Area	Area%
1	7.855	25338	2.604
2	8.740	947785	97.396
合計		973123	100.000

**5-phenylfuran-2(5H)-one (6ab).**



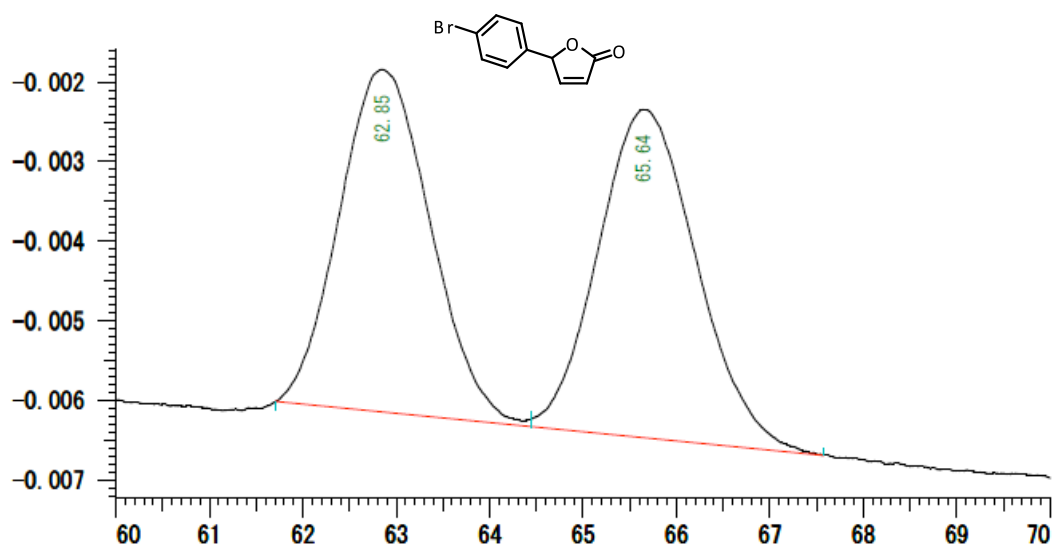
検出器A Ch1 220nm

Peak	Retention Time	Area	Area%
1	30.476	6285087	50.013
2	33.356	6281706	49.987
合計		12566794	100.000

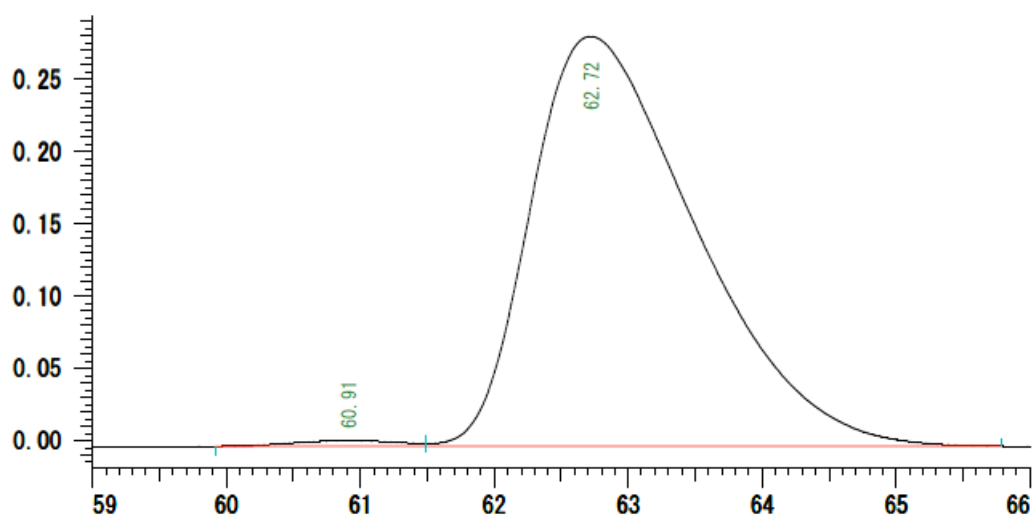


	Retention Time	Area	Area%
1	31.09	4217922	98.584
2	34.50	60568	1.416
		4278490	100.000

**5-(4-bromophenyl)furan-2(5H)-one (6bb).**

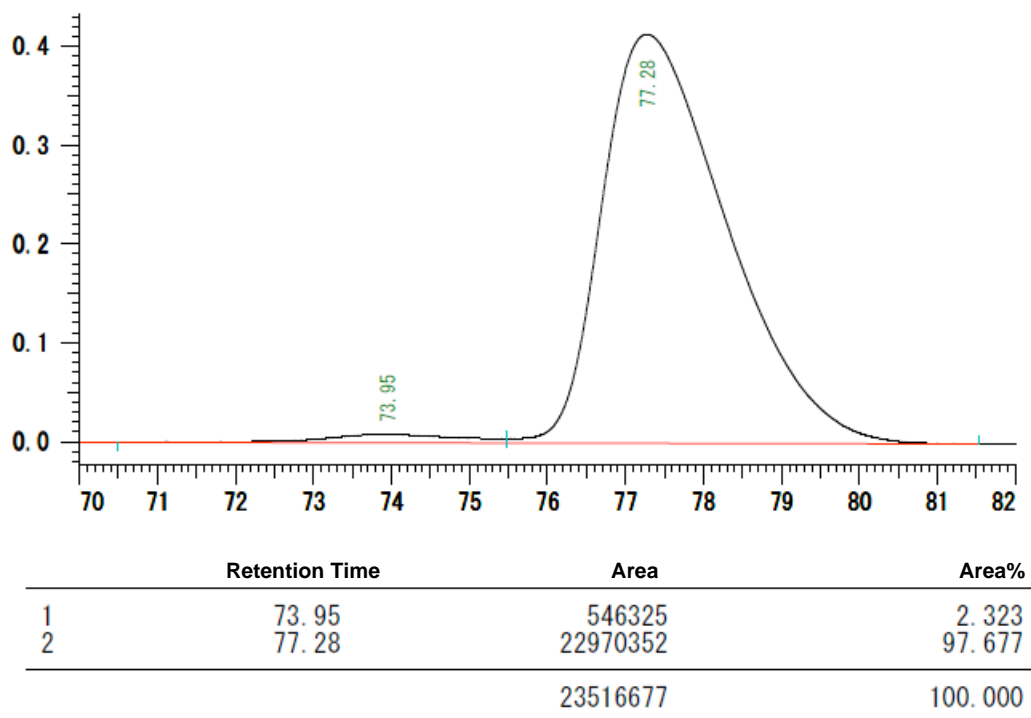
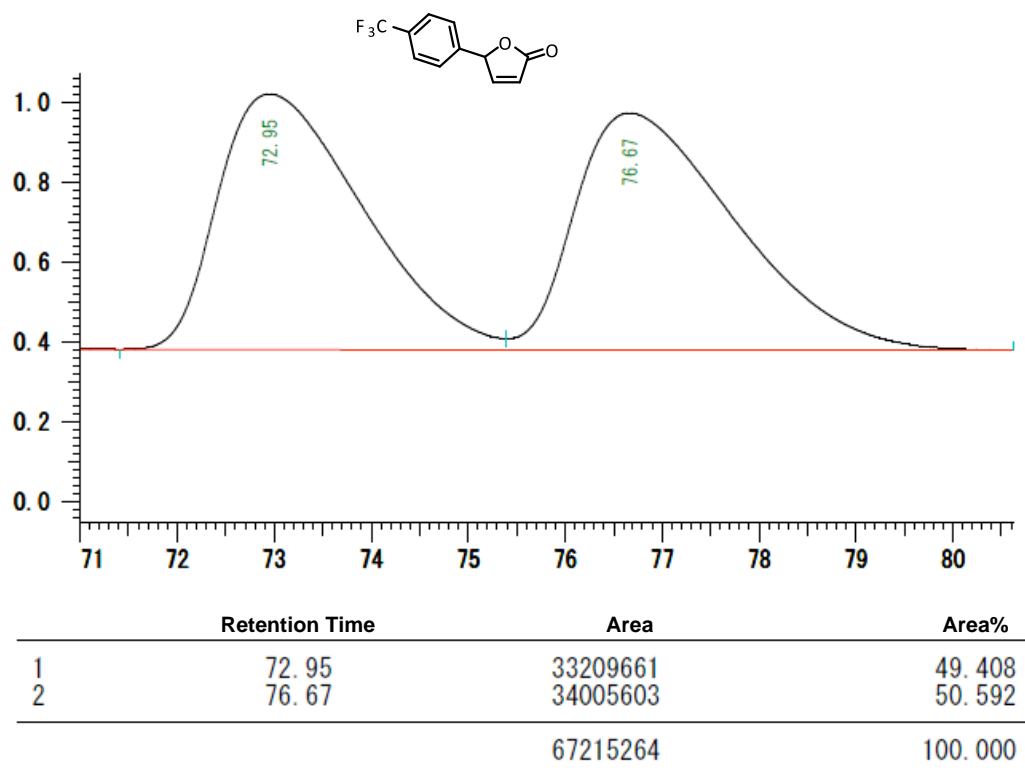


	Retention Time	Area	Area%
1	62.85	146131	49.121
2	65.64	151360	50.879
		297491	100.000

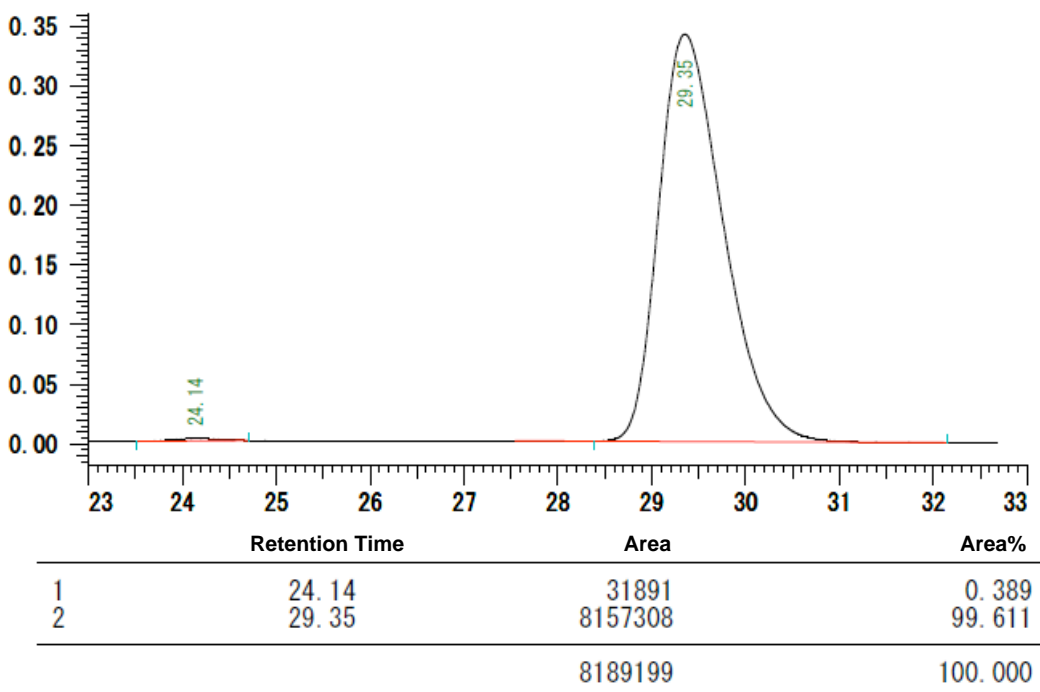
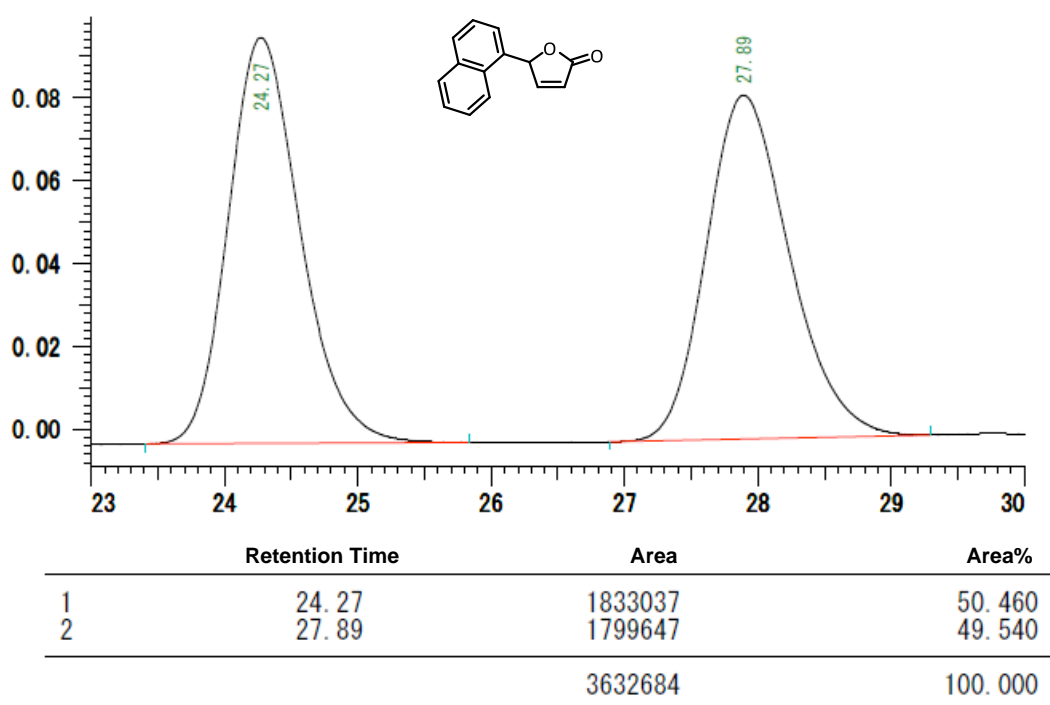


	Retention Time	Area	Area%
1	60.91	111406	0.911
2	62.72	12117867	99.089
		12229273	100.000

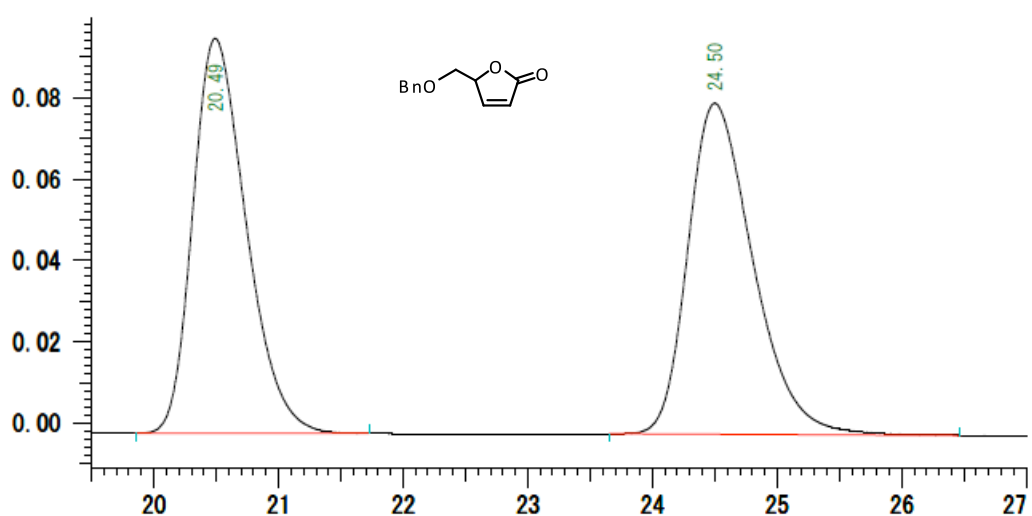
**5-(4-trifluoromethylphenyl)furan-2(5H)-one (6cb).**



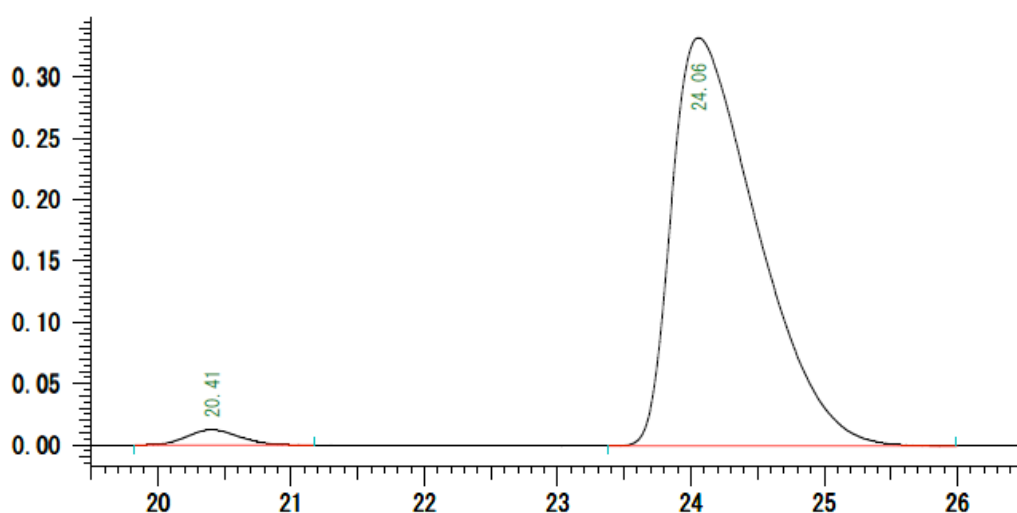
**5-(naphthalene-1-yl)furan-2(5H)-one (6db).**



**5-(benzyloxymethyl)furan-2(5H)-one (6fb).**

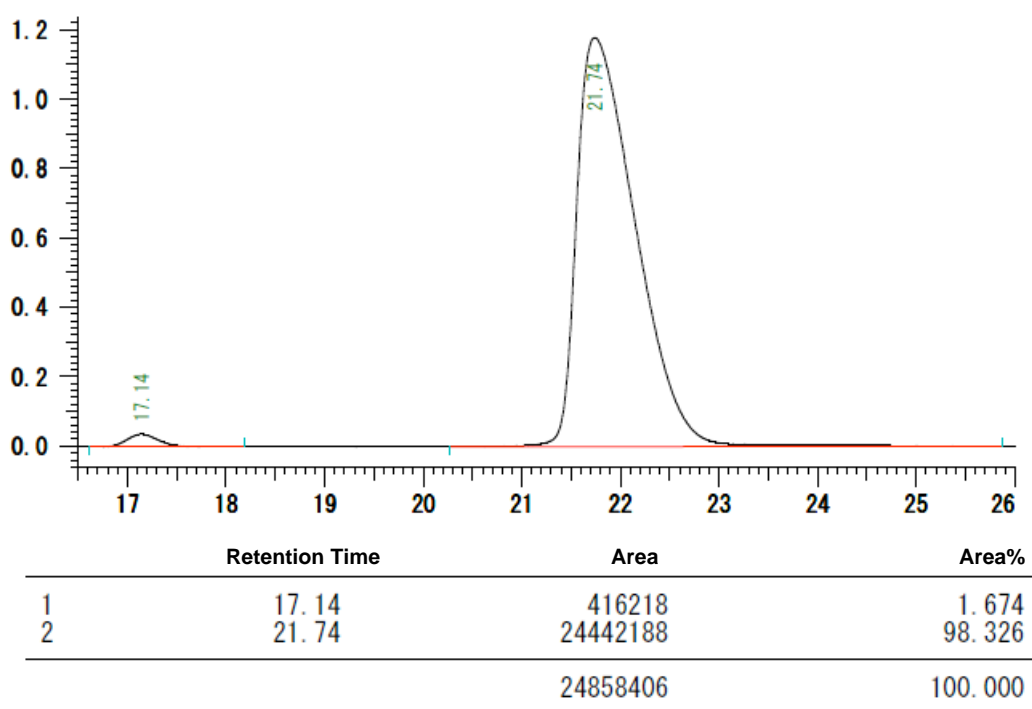
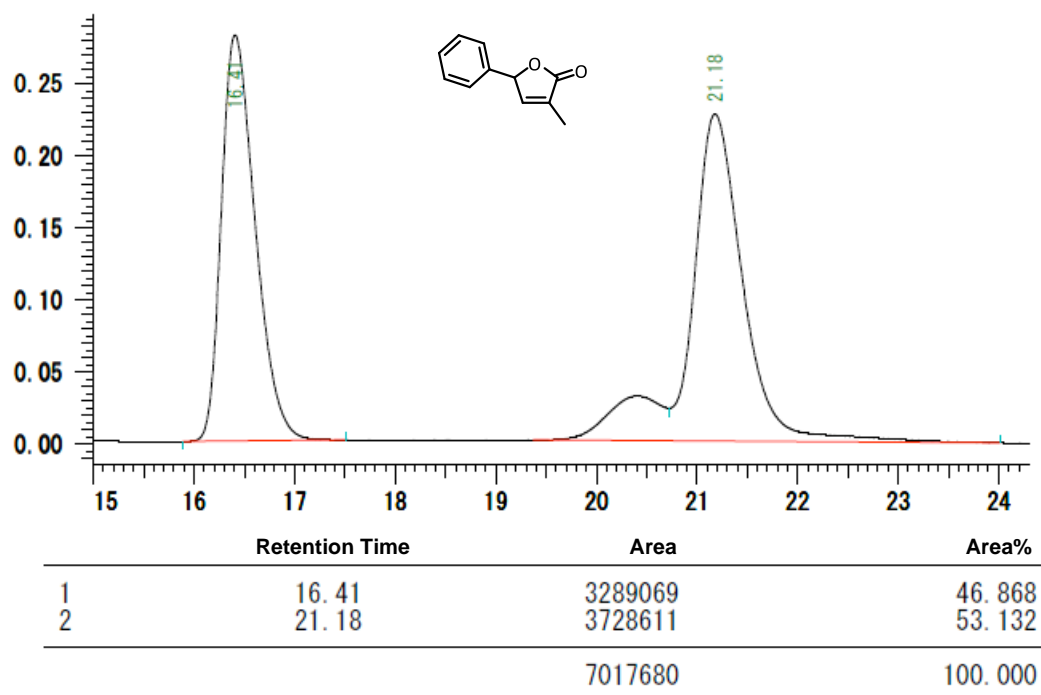


	Retention Time	Area	Area%
1	20.49	1445719	49.454
2	24.50	1477660	50.546
		2923379	100.000

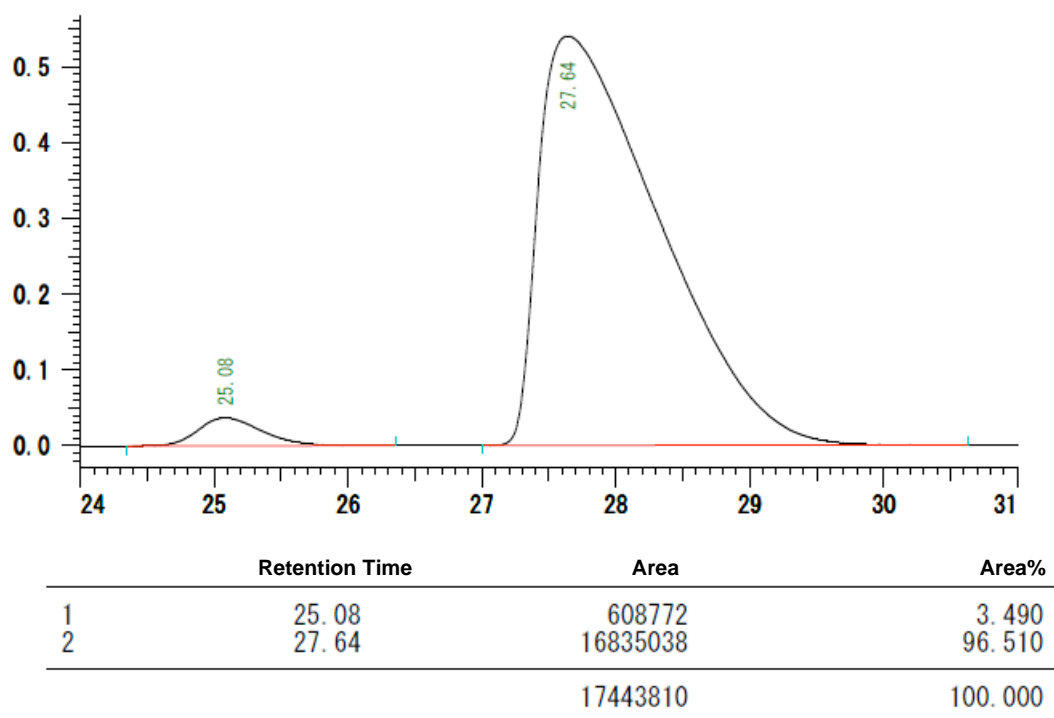
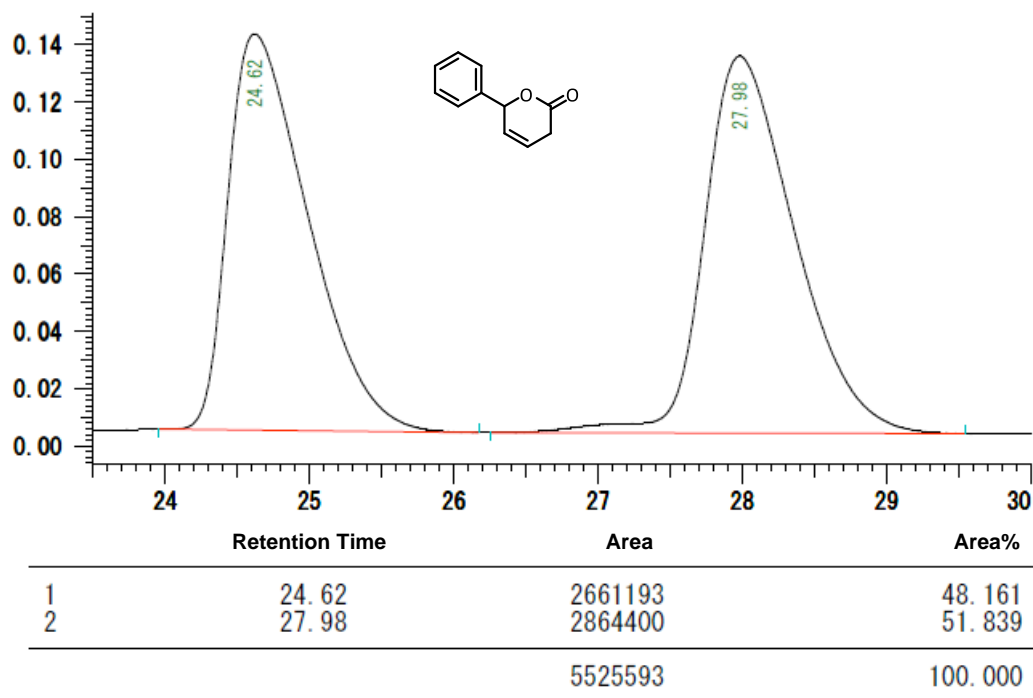


	Retention Time	Area	Area%
1	20.41	173573	2.287
2	24.06	7415562	97.713
		7589135	100.000

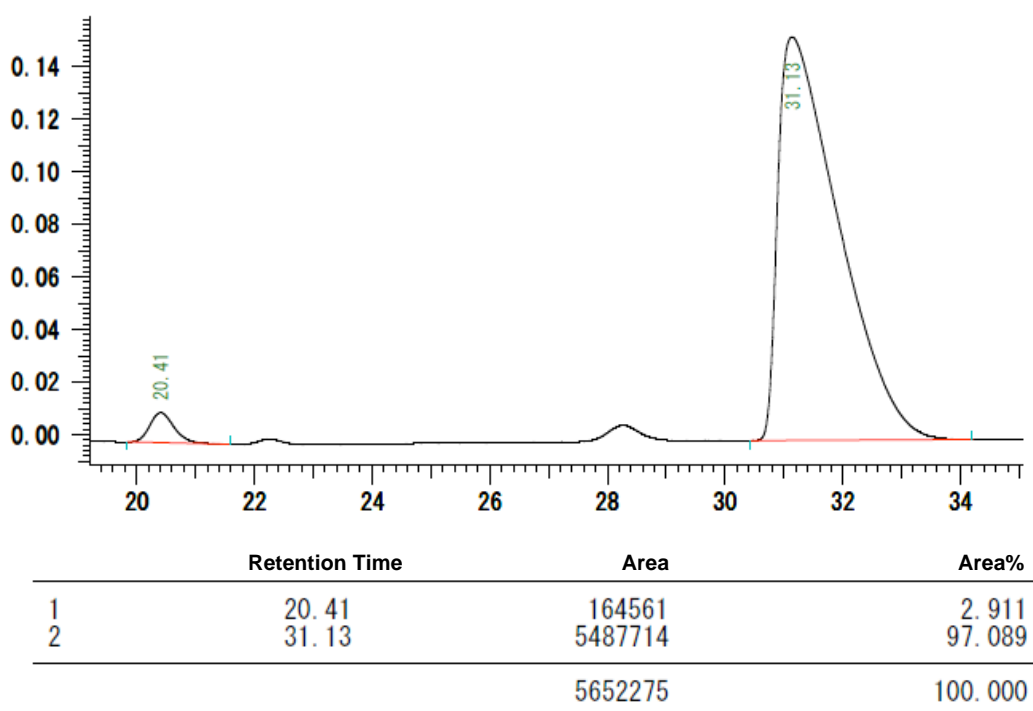
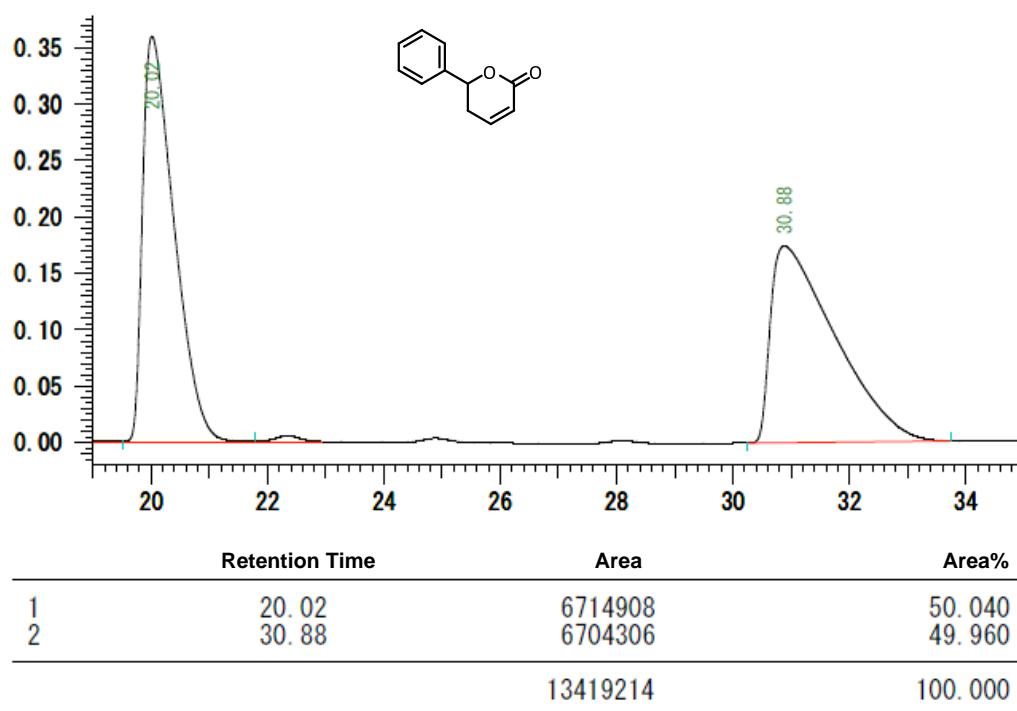
**3-methyl-5-phenylfuran-2(5H)-one (6ac).**



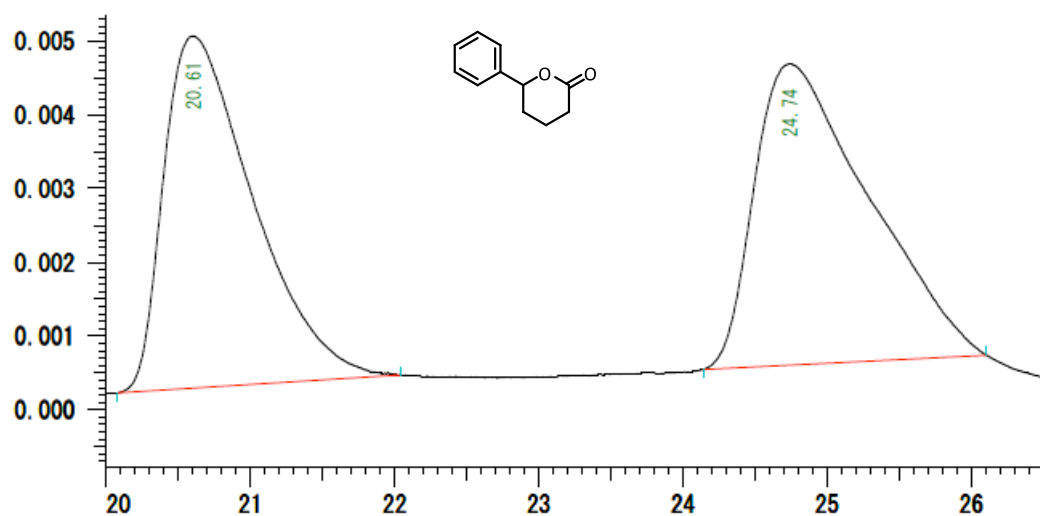
**6-phenyl-3,6-dihydro-2H-pyran-2-one (8).**



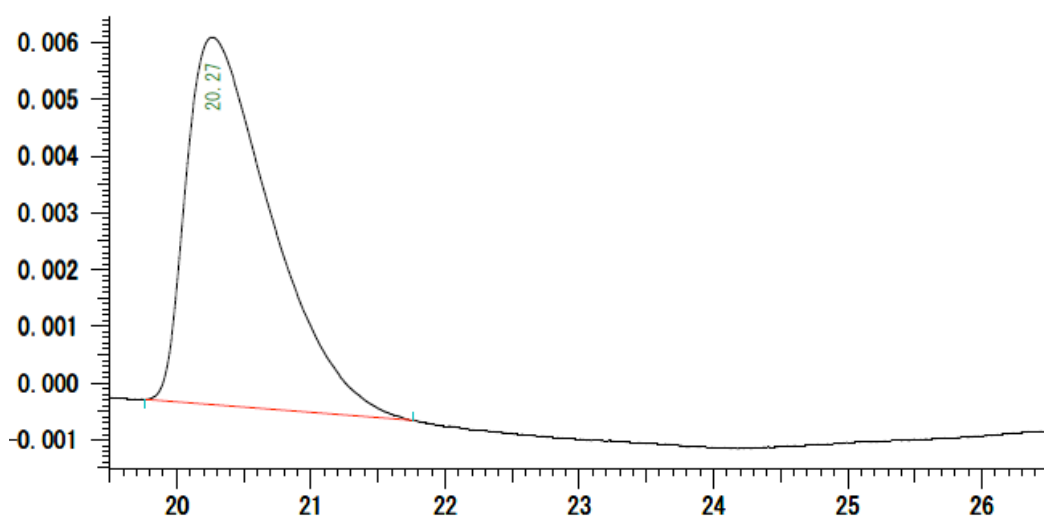
**6-phenyl-5,6-dihydro-2H-pyran-2-one (9).**



**6-phenyltetrahydro-2H-pyran-2-one (10).**



	Retention Time	Area	Area%
1	20.61	102700	47.868
2	24.74	111846	52.132
		214546	100.000



	Retention Time	Area	Area%
1	20.27	142881	100.000
		142881	100.000

**(*R*)-6-pentyl-5,6-dihydro-2H-pyran-2-one, (*R*)-(-)-massoialactone, (14).**

