

## Electronic Supplementary Information

### Enantioselective Bromolactonization of *cis*-1,2-disubstituted olefinic acids using amino-thiocarbamate catalyst

Chong Kiat Tan, Chencheng Le, and Ying-Yeung Yeung\*

Department of Chemistry, National University of Singapore, 3 Science Drive 3, Singapore 117543, Singapore

E-mail: chmyyy@nus.edu.sg; Fax: +65-6779-1691; Tel: +65-6516-7760

## A. General Information

All reactions that required anhydrous conditions were carried out under nitrogen atmosphere. Commercially available reagents were used as received from Alfa Aesar, Sigma-Aldrich and TCI-Tokyo Kasei Kogyo Co. Ltd. Dichromethane and acetonitrile were dried by distillation over the appropriate drying reagents. Anhydrous THF was purchased from EM Science and used without further purification. HPLC grade chloroform was purchased from Fischer Chemicals while HPLC grade hexane and isopropanol were purchased from Tedia and used as received. Analytical thin layer chromatography (TLC) was performed with Merck pre-coated TLC plates, silica gel 60 F-254, layer thickness 0.25 mm. The plate were visualized under 254 nm UV light and stained with potassium permanganate. Flash chromatography separations were performed on Merck 60 (0.040-0.0063 mm) mesh silica gel. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Bruker ACF 300 (300MHz) and AMX500 (500 MHz) spectrometer. Chemical shift ( $\delta$ ) are reported in parts per million (ppm) relative to tetramethylsilane (TMS,  $\delta$ =0.00) for <sup>1</sup>H NMR and are referenced to the residual solvent peak chloroform ( $\delta$ =7.26). Chemical shift for <sup>13</sup>C NMR are reported in ppm downfield from TMS and are referenced to the carbon resonance of the solvent chloroform ( $\delta$ =77.0). Data are represented as follow: chemical shift, multiplicity (br. s = broad singlet, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants in Hertz (Hz). Low resolution mass spectra (ESI) were obtained on a Finnigan/MAT LCQ spectrometer. High resolution mass spectra (ESI) were obtained on Finnigan/MAT 95XL-T spectrometer. Enantiomeric excesses were determined by HPLC analysis on Shimadzu HPLC units, including the following instruments: LC-20AD pump, SPD-20A detector and Dacel Chiralpak IA, IB, IC or ODH columns. Optical rotations were recorded on a Jasco DIP-1000 polarimeter. Infrared spectra were recorded on a Varian 3100 FTIR spectrophotometer and reported in wave numbers ( $\text{cm}^{-1}$ ). Melting points were determined on a BÜCHI B-540b melting point apparatus.

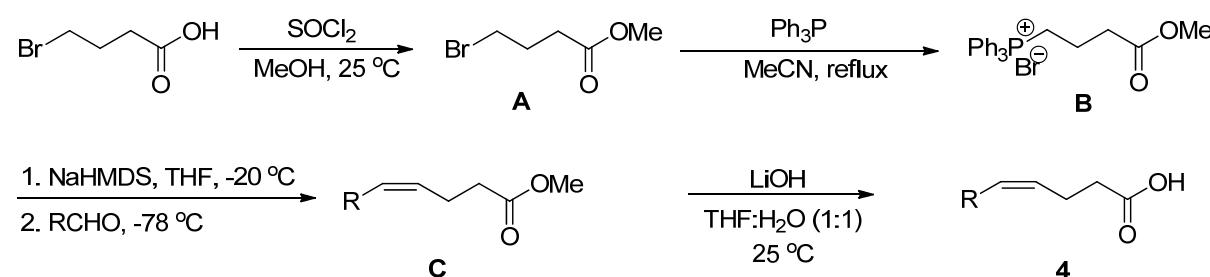
## B. Experimental Procedures and Physical Data

All the amino-thiocarbamate catalysts were prepared according to our previously reported procedure. Spectral data of other catalysts were reported in previous reports.<sup>1-4</sup>

### Amino-thiocarbamate 6a

White solid, mp 68.5-69.6 °C;  $[\alpha]_D^{25} +140.8$  (*c* 1.0, CHCl<sub>3</sub>); IR (KBr): 2952, 2868, 1619, 1510, 1464, 1308, 1284, 1208, 1159, 1033 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 8.75 (br, 1H), 8.34 (s, 1H), 8.02 (d, *J* = 9.5 Hz, 1H), 7.66-7.44 (m, 1H), 7.38-7.34 (m, 3H), 6.50 (d, *J* = 7.6 Hz, 2H), 5.82 (br, 1H), 5.03-5.00 (m, 2H), 4.15 (br, 1H), 4.04 (br, 1H), 3.85-3.74 (m, 6H), 3.49 (br, 1H), 3.18 (br, 1H), 3.05 (br, 1H), 2.67 (m, 2H), 2.28 (br, 1H), 1.92-1.50 (m, 9H), 1.02 (d, *J* = 6.3 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 188.0, 158.6, 157.2, 152.0, 147.1, 144.6, 143.0, 141.6, 131.3, 127.0, 124.8, 123.8, 122.2, 119.4, 119.2, 114.3, 103.8, 102.5, 98.6, 81.5, 66.7, 59.5, 56.4, 55.4, 42.4, 39.6, 37.7, 27.4, 25.0, 24.0, 22.5, 22.5; HRMS (ESI) calcd for C<sub>33</sub>H<sub>42</sub>N<sub>3</sub>O<sub>4</sub>S [M + H]<sup>+</sup>: 576.2891; found: 576.2908.

### Preparation of *cis*-olefinic acid 4



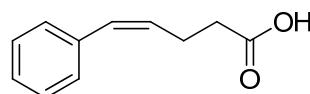
**General Procedure for the preparation of *cis*-olefinic acid 4:** To a solution of 4-bromobutyric acid (1.0 eq) in MeOH was added thionyl chloride (1.5 eq) dropwise at 0 °C and the reaction mixture was stirred at 25 °C overnight. The reaction was quenched by adding sat'd NaHCO<sub>3</sub> solution and was then extracted with Et<sub>2</sub>O. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. The residue was subjected to flash chromatography to afford the desired ester product **A**.

A mixture of methyl 4-bromobutanoate **A** (1.0 eq) and triphenylphosphine (1.0 eq) was refluxed in MeCN for 6 hours to yield salt **B**. The crude phosphonium bromide **B** was recrystallized from MeCN and Et<sub>2</sub>O to give the Wittig salt as white crystals.

The Wittig salt **B** (1.0 eq) was suspended in anhydrous THF at -20 °C. Sodium bis(trimethylsilyl)amide (1.0 eq) was added dropwise into the suspension and stirred for 20 min. The reaction mixture was further cooled to -78 °C, to which a solution of the aldehyde (1.0 eq) was added dropwise. The reaction mixture was allowed to rise to room temperature

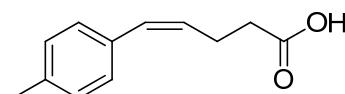
and stirred overnight. The reaction was then quenched with sat'd NH<sub>4</sub>Cl solution and the product was then extracted with Et<sub>2</sub>O. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and the concentrated *in vacuo*. The residue was separated by flash column chromatography on silica gel to afford the *cis*-alkenoate **C**.

To a solution of *cis*-alkenoate **C** (1.0 eq.) in THF:H<sub>2</sub>O (1:1) was added LiOH (5.0 eq.). The reaction mixture was stirred at room temperature overnight. Upon termination, H<sub>2</sub>O was added and an extraction with CH<sub>2</sub>Cl<sub>2</sub> was performed. The organic layer was discarded. The aqueous layer was then acidified with 1 M HCl before it was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and filtered. The solvent was removed *in vacuo* to yield *cis*-olefinic acid **4** in quantitative yield.



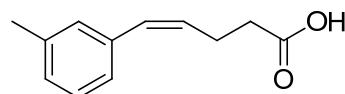
**(Z)-5-Phenyl-4-pentenoic acid (4a)<sup>2</sup>**

Colorless oil; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.37 (br, 1H), 7.44-7.31 (m, 5H), 6.57 (d, *J* = 11.52 Hz, 1H), 5.76-5.67 (m, 1H), 2.79-2.71 (m, 2H), 2.62-2.54 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.4, 137.2, 130.5, 129.9, 128.7, 126.9, 34.2, 23.8.



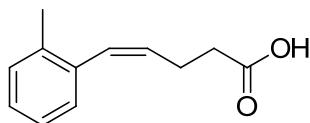
**(Z)-5-p-toylpent-4-enoic acid (4b)**

White solid; mp 54.4–55.0 °C; IR (KBr): 3009, 2927, 1700, 1508, 1425, 1296, 1212 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 11.57 (br, 1H), 7.28-7.24 (m, 4H), 6.55 (d, *J* = 11.4 Hz, 1H), 5.71-5.66 (m, 1H), 2.79-2.76 (m, 2H), 2.59-2.56 (m, 2H), 2.44 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 179.6, 136.4, 134.2, 130.2, 129.1, 128.9, 128.5, 34.1, 23.7, 21.0; HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>O<sub>2</sub> [M – H]<sup>-</sup>: 189.0921; found: 189.0919.



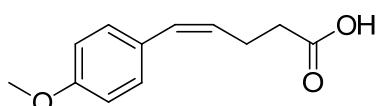
**(Z)-5-m-toylpent-4-enoic acid (4c)**

Yellow oil; IR (Neat): 3018, 2922, 1709, 1413, 1281, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.92 (br, 1H), 7.41-7.36 (m, 1H), 7.26-7.19 (m, 2H), 6.62 (d, *J* = 11.7 Hz, 1H), 5.81-5.73 (m, 1H), 2.82-2.79 (m, 2H), 2.65-2.50 (m, 2H), 2.51 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.6, 137.6, 136.9, 130.4, 129.6, 129.3, 128.0, 127.5, 125.6, 34.1, 23.6, 21.3; HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>O<sub>2</sub> [M – H]<sup>-</sup>: 189.0921; found: 189.0918.



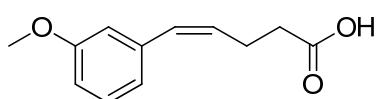
**(Z)-5-o-toylpent-4-enoic acid (4d)**

Pale yellow oil; IR (Neat): 3019, 1710, 1413, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.74 (br, 1H), 7.33 (s, 4H), 6.68 (d, *J* = 11.3 Hz, 1H), 5.91-5.82 (m, 1H), 2.71-2.54 (m, 4H), 2.42 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.9, 136.3, 129.9, 129.9, 129.8, 128.9, 127.2, 125.5, 34.2, 23.6, 19.9; HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>O<sub>2</sub> [M - H]<sup>-</sup>: 189.0921; found: 189.0915.



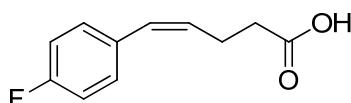
**(Z)-5-(4-methoxyphenyl)pent-4-enoic acid (4e)**

White solid; mp 64.2-65.0 °C; IR (KBr): 3031, 2837, 1698, 1508, 1251 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.25 (br, 1H), 7.30 (d, *J* = 8.7 Hz, 2H), 6.96 (d, *J* = 8.7 Hz, 2H), 6.50 (d, *J* = 11.7 Hz, 1H), 5.66-5.57 (m, 1H), 3.87 (s, 3H), 2.78-2.71 (m, 2H), 2.59-2.54 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.4, 158.3, 129.8, 129.7, 128.2, 113.6, 55.0, 34.0, 23.7; HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>O<sub>3</sub> [M - H]<sup>-</sup>: 205.0870; found: 205.0862.



**(Z)-5-(3-methoxyphenyl)pent-4-enoic acid (4f)**

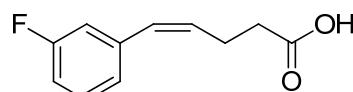
Colorless oil; IR (Neat): 3019, 1709, 1645, 1599, 1215 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.40 (br, 1H), 7.36-7.31 (m, 1H), 6.97-6.86 (m, 3H), 6.54 (d, *J* = 11.7 Hz, 1H), 5.76-5.57 (m, 1H), 3.87 (s, 3H), 2.80-2.72 (m, 2H), 2.59-2.54 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.4, 158.3, 129.8, 129.7, 128.2, 113.6, 55.0, 34.0, 23.7; HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>O<sub>3</sub> [M - H]<sup>-</sup>: 205.0870; found: 205.0862.



**(Z)-5-(4-fluorophenyl)pent-4-enoic acid (4g)**

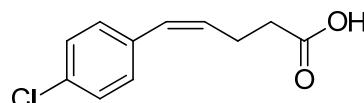
White solid; mp 45.6-46.9 °C; IR (KBr): 3030, 2927, 1699, 1604, 1405, 1241 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.79 (br, 1H), 7.32-7.27 (m, 2H), 7.12-7.06 (m, 2H), 6.50 (d, *J* = 11.7 Hz, 1H), 5.72-5.64 (m, 1H), 2.74-2.67 (m, 2H), 2.60-2.53 (m, 2H); <sup>13</sup>C NMR (75 MHz,

$\text{CDCl}_3$ ):  $\delta$  179.5, 161.6 (d,  $J_{C,F} = 246$  Hz), 133.1 (d,  $J_{C,F} = 3$  Hz), 130.2 (d,  $J_{C,F} = 8$  Hz), 129.7 (d,  $J_{C,F} = 1$  Hz), 129.2, 115.1 (d,  $J_{C,F} = 21$  Hz), 34.0, 23.6; HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{11}\text{FO}_2$   $[\text{M} - \text{H}]^-$ : 193.0670; found: 193.0672.



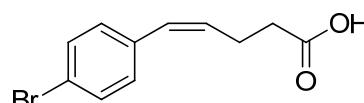
**(Z)-5-(3-fluorophenyl)pent-4-enoic acid (4h)**

Colorless oil; IR (Neat): 3019, 1708, 1611, 1580, 1487, 1440, 1234, 1136  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  11.51 (br, 1H), 7.39-7.32 (m, 1H), 7.13-6.98 (m, 3H), 6.51 (d,  $J = 11.7$  Hz, 1H), 5.79-5.70 (m, 1H), 2.77-2.69 (m, 2H), 2.62-2.53 (m, 2H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  179.5, 162.6 (d,  $J_{C,F} = 245$  Hz), 139.2 (d,  $J_{C,F} = 8$  Hz), 131.0, 129.6 (d,  $J_{C,F} = 6$  Hz), 129.2 (d,  $J_{C,F} = 2$  Hz), 124.3 (3 Hz), 115.3 (d,  $J_{C,F} = 21$  Hz), 113.6 (d,  $J_{C,F} = 21$  Hz), 33.9, 23.6; HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{11}\text{FO}_2$   $[\text{M} - \text{H}]^-$ : 193.0670; found: 193.0668.



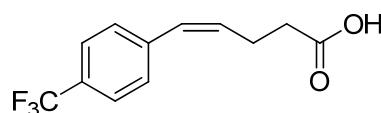
**(Z)-5-(4-chlorophenyl)pent-4-enoic acid (4i)**

White solid; mp 48.1-49.1  $^\circ\text{C}$ ; IR (KBr): 2927, 1700, 1491, 1424, 1216, 1097  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  11.64 (br, 1H), 7.35 (d,  $J = 8.6$  Hz, 2H), 7.24 (d,  $J = 8.6$  Hz, 2H), 6.46 (d,  $J = 11.7$  Hz, 1H), 5.74-5.66 (m, 1H), 2.72-2.65 (m, 2H), 2.59-2.51 (m, 2H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  178.9, 135.3, 132.3, 130.4, 129.8, 129.0, 128.1, 67.6, 33.8, 33.8, 25.3, 23.5; HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{11}\text{ClO}_2$   $[\text{M} - \text{H}]^-$ : 209.0375; found: 209.0366.



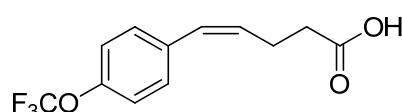
**(Z)-5-(4-bromophenyl)pent-4-enoic acid (4j)**

Pale yellow solid; mp 62.2-63.5  $^\circ\text{C}$ ; IR (KBr): 2926, 1700, 1488, 1425, 1287, 1217, 1009  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.46 (d,  $J = 8.2$  Hz, 2H), 7.13 (d,  $J = 8.2$  Hz, 2H), 6.41 (d,  $J = 11.4$  Hz, 1H), 5.67-5.65 (m, 1H), 2.64-2.60 (m, 1H), 2.54-2.48 (m, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  179.2, 135.9, 131.3, 130.6, 130.3, 129.3, 34.0, 23.6; HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{11}\text{BrO}_2$   $[\text{M} - \text{H}]^-$ : 252.9870; found: 252.9861.



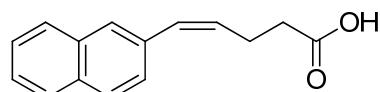
**(Z)-5-[(4-trifluoromethyl)phenyl]pent-4-enoic acid (4k)**

White semi-solid; IR (Neat): 3020, 1711, 1326, 1215, 1167, 1128, 1068 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.79 (br, 1H), 7.66 (d, *J* = 8.2 Hz, 2H), 7.43 (d, *J* = 8.2 Hz, 2H), 6.56 (d, *J* = 11.7 Hz, 1H), 5.86-5.77 (m, 1H), 2.76-2.68 (m, 2H), 2.63-2.54 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.6, 140.7 (q, *J*<sub>C-F</sub> = 1 Hz), 132.0, 130.5, 129.2, 128.9, 125.2 (q, *J*<sub>C-F</sub> = 3 Hz), 124.2 (q, *J*<sub>C-F</sub> = 272 Hz), 33.9, 23.6; HRMS (ESI) calcd for C<sub>12</sub>H<sub>11</sub>F<sub>3</sub>O<sub>2</sub> [M - H]<sup>-</sup>: 243.0638; found: 243.0631.



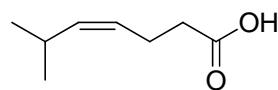
**(Z)-5-[(4-trifluoromethoxy)phenyl]pent-4-enoic acid (4l)**

Yellow oil; IR (Neat): 3020, 1712, 1507, 1415, 1262, 1215, 1170, cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.37 (br, 1H), 7.34 (d, *J* = 8.7 Hz, 2H), 7.24 (d, *J* = 8.1 Hz, 2H), 6.51 (d, *J* = 11.5 Hz, 1H), 5.77-5.69 (m, 1H), 2.71-2.69 (m, 2H), 2.58-2.53 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.6, 147.9 (q, *J*<sub>C-F</sub> = 2 Hz), 135.8, 130.7, 130.0, 129.0, 120.7, 120.5 (q, *J*<sub>C-F</sub> = 257 Hz), 34.0, 23.6; HRMS (ESI) calcd for C<sub>12</sub>H<sub>11</sub>F<sub>3</sub>O<sub>3</sub> [M - H]<sup>-</sup>: 259.0588; found: 259.0581.



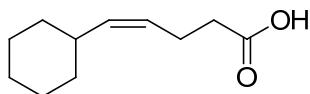
**(Z)-5-(naphthalen-2-yl)pent-4-enoic acid (4m)**

White solid; mp 87.7-88.4 °C; IR (KBr): 3010, 2921, 1700, 1632, 1405, 1272, 1253, 1180 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.88 (br, 1H), 7.97-7.62 (m, 4H), 7.61-7.55 (m, 3H) 6.78 (d, *J* = 11.7 Hz, 1H), 5.90-5.81 (m, 1H), 2.95-2.88 (m, 2H), 2.71-2.63 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.7, 134.5, 133.2, 132.2, 130.3, 130.2, 127.9, 127.7, 127.5, 127.4, 126.9, 126.0, 125.7, 34.1, 23.7; HRMS (ESI) calcd for C<sub>15</sub>H<sub>14</sub>O<sub>2</sub> [M - H]<sup>-</sup>: 225.0921; found: 225.0914.



**(Z)-6-methylhept-4-enoic acid (4n)**

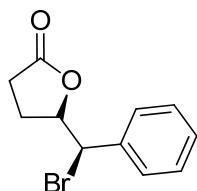
Colorless oil; IR (Neat): 3019, 2961, 1708, 1645, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 10.62 (br, 1H), 5.29-5.18 (m, 2H), 2.67-2.55 (m, 1H), 2.39 (s, 4H), 0.94 (d, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.9, 139.3, 124.5, 34.4, 26.5, 23.0, 22.6; HRMS (ESI) calcd for C<sub>8</sub>H<sub>14</sub>O<sub>2</sub> [M - H]<sup>-</sup>: 141.0921; found: 141.0925.



**(Z)-5-cyclohexylpent-4-enoic acid (4o)**

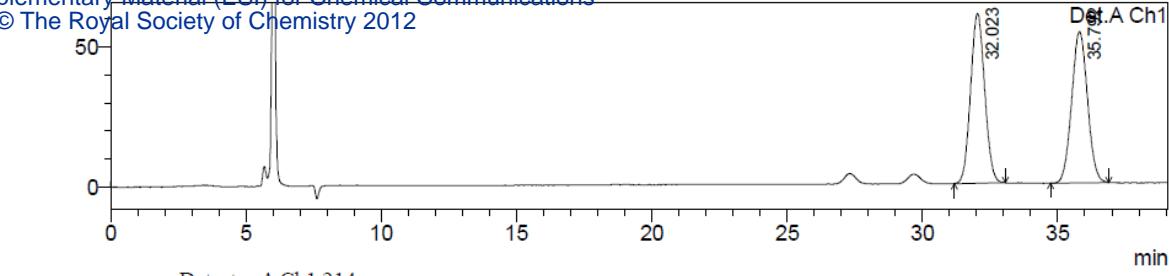
Colorless oil; IR (Neat): 3020, 2928, 1708, 1644, 1215 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 11.36 (br, 1H), 5.30-5.17 (m, 2H), 2.38 (s, 4H), 2.37-2.21 (m, 1H), 1.71-1.55 (m, 5H), 1.35-0.98 (m, 5H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 180.0, 137.8, 125.0, 36.3, 26.0, 25.9, 22.7; HRMS (ESI) calcd for C<sub>11</sub>H<sub>18</sub>O<sub>2</sub> [M - H]<sup>-</sup>: 181.1234; found: 181.1231.

**General procedure for Bromolactonization:** The corresponding *cis*-alkenoic acid **4** (0.05 mmol, 1.0 eq) and catalyst **7** (3.0 mg, 0.005 mmol, 0.1 eq) were dissolved in CHCl<sub>3</sub> (0.5 ml) and *n*-hexane (1.0 ml) and stirred at -78 °C in the dark under nitrogen atmosphere. *N*-bromosuccinimide (10.6mg, 0.06 mmol, 1.2 eq) was added. The reaction mixture was stirred at the corresponding temperature and monitored by TLC. Sat'd Na<sub>2</sub>SO<sub>3</sub> (3.0 ml) was added to quench the reaction mixture. The resulting biphasic mixture was warmed to 25 °C before it was diluted with water and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 5 ml). The combined organic extracts were dried over MgSO<sub>4</sub>, filtered and concentrated *in vacuo*. The residue was purified by flash column chromatography over silica gel to provide the corresponding product **5**.



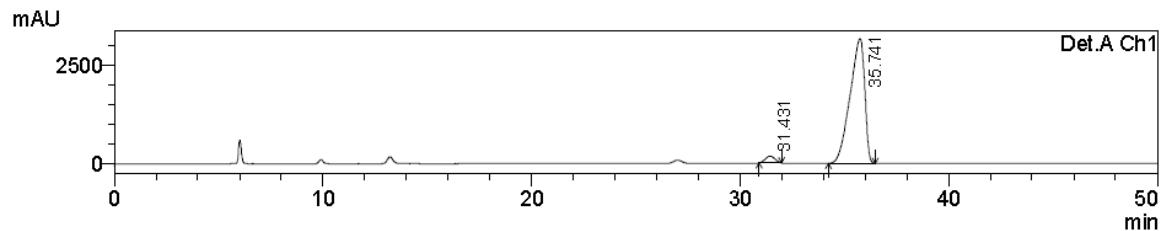
**5-bromo(phenyl)methyldihydrofuran-2(3H)-one (5a)<sup>2</sup>**

White solid; [α]<sub>D</sub><sup>25</sup> -101.8 (c 0.8, CHCl<sub>3</sub>, 93% ee); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.39-7.27 (m, 5H), 4.91 (d, *J* = 5.61 Hz, 1H), 4.86-4.80 (m, 1H), 2.47-2.26 (m, 2H), 2.21-2.09 (m, 1H), 2.02-1.89 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 175.8, 136.9, 129.1, 128.9, 128.5, 81.9, 55.1, 28.3, 25.6; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 31.4 min (minor), t<sub>2</sub> = 35.7 min (major).



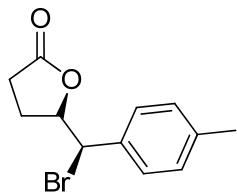
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	32.023	2190304	60568	49.932	52.891
2	35.792	2196230	53948	50.068	47.109
Total		4386534	114516	100.000	100.000



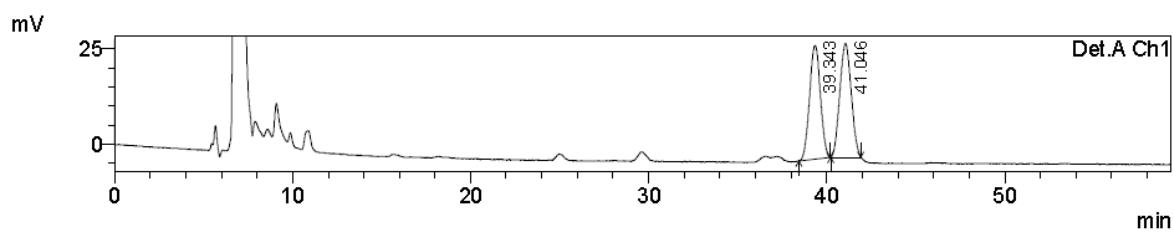
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	31.431	5562444	174883	3.475	5.236
2	35.741	154513311	3164890	96.525	94.764
Total		160075755	3339773	100.000	100.000



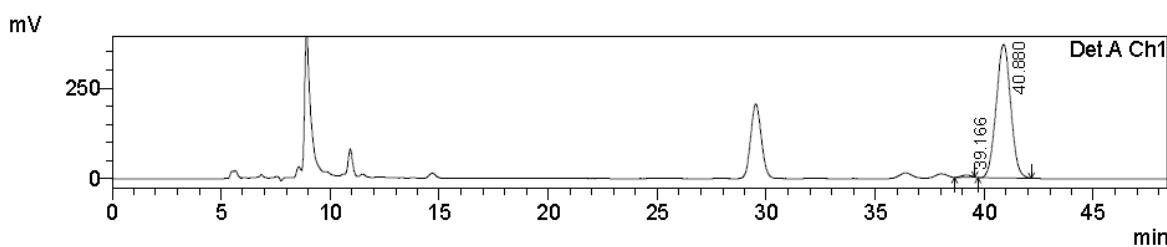
### 5-bromo(p-toyl)methylidihydrofuran-2(3H)-one (5b)

Colorless oil;  $[\alpha]_D^{25} -34.0$  (*c* 0.5, CHCl<sub>3</sub>, 98% ee); IR (Neat): 3019, 1776, 1739, 1713, 1215 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  7.34 (d, *J* = 8.0 Hz, 2 H), 7.16 (d, *J* = 7.8 Hz, 2 H), 4.97 (d, *J* = 5.5 Hz, 1 H), 4.92-4.88 (m, 2H), 2.50-2.41 (m, 1H), 2.39 (s, 1H), 2.39-2.35 (m, 1H), 2.27-2.01 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  176.0, 139.2, 133.9, 129.5, 128.3, 82.1, 55.2, 28.3, 25.6, 21.1; HRMS (EI) calcd for C<sub>12</sub>H<sub>13</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 268.0099; found: 268.0089; HPLC (Daicel Chiraldapak IC, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 39.2 min (minor), t<sub>2</sub> = 40.9 min (major).



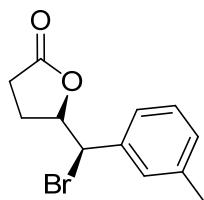
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	39.343	1254300	29695	48.603	49.909
2	41.046	1326395	29803	51.397	50.091
Total		2580695	59498	100.000	100.000



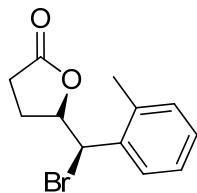
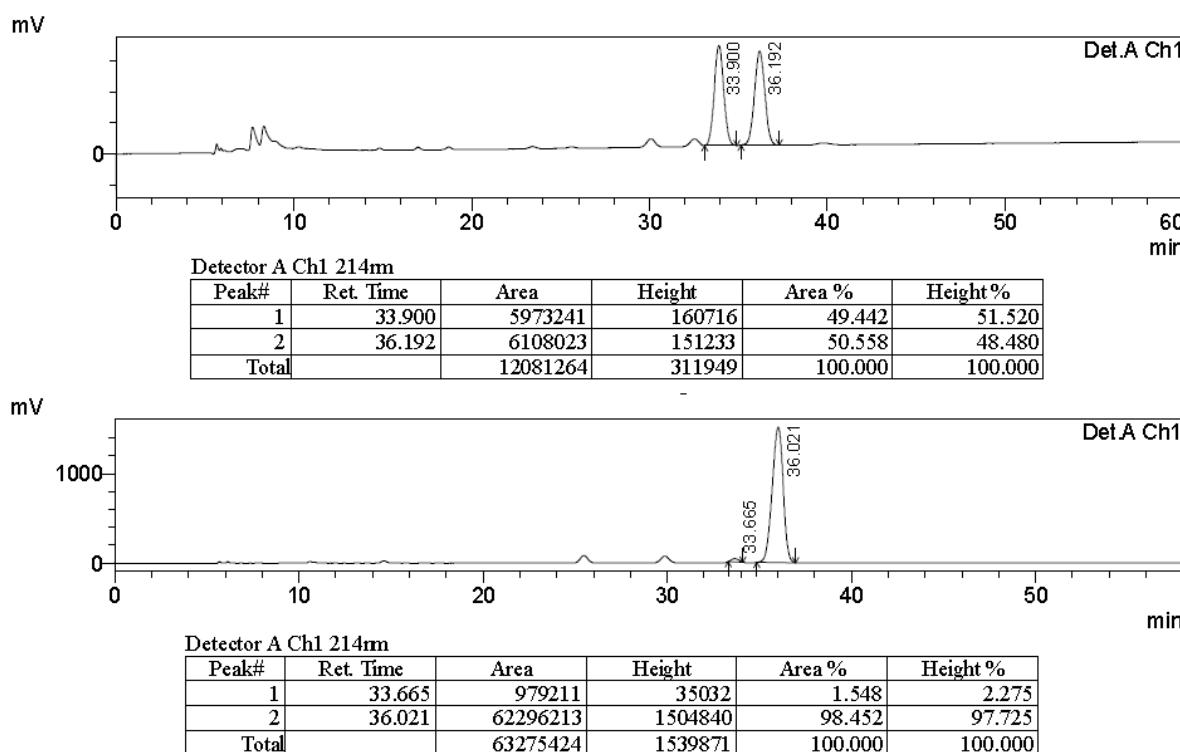
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	39.166	177592	5880	1.030	1.572
2	40.880	17060333	368158	98.970	98.428
Total		17237924	374038	100.000	100.000



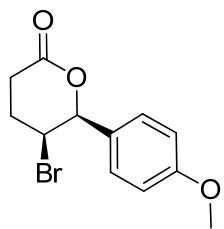
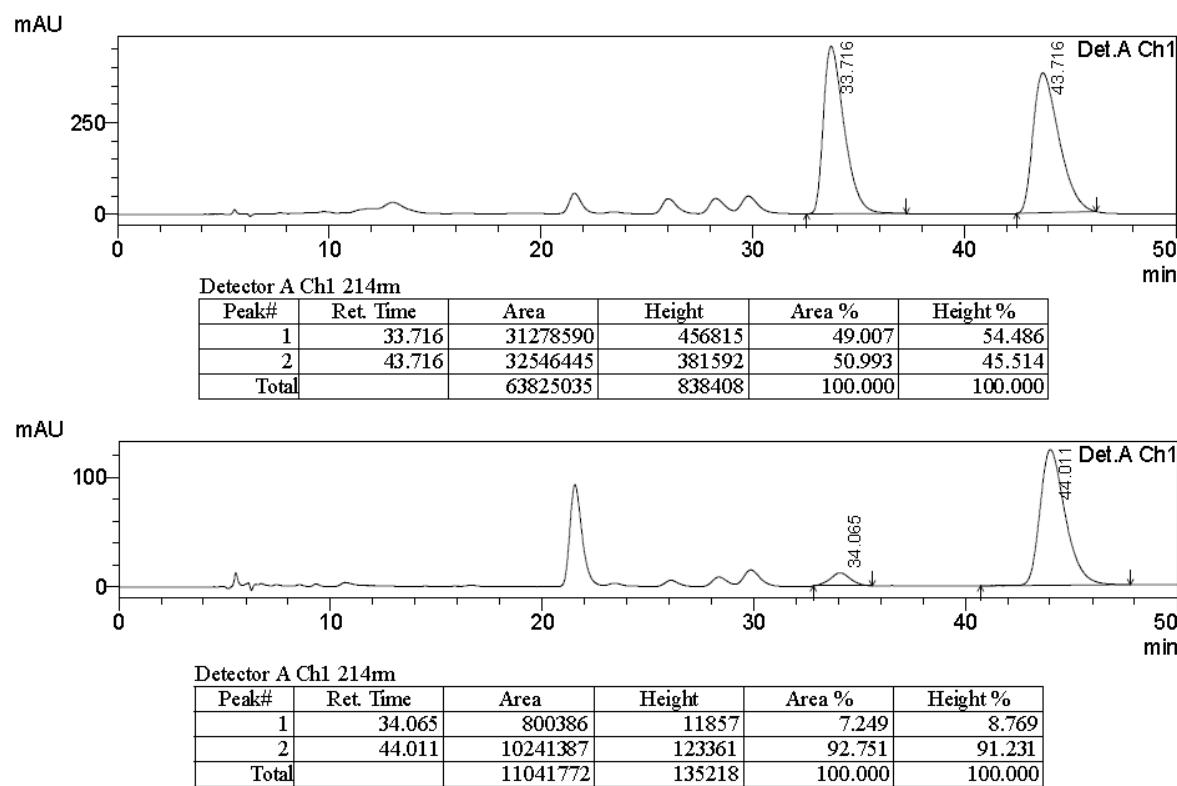
### 5-bromo(m-toyl)methyldihydrofuran-2(3H)-one (**5c**)

Colorless oil;  $[\alpha]_D^{25} -111.7$  (*c* 1.1, CHCl<sub>3</sub>, 97% ee); IR (Neat): 3020, 1780, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.26-7.15 (m, 4H), 4.96-4.88 (m, 2H), 2.52-2.41 (m, 2H), 2.36 (s, 3H), 2.30-2.17 (m, 1H), 2.11-1.99 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 179.3, 138.7, 136.8, 129.9, 129.0, 128.8, 125.5, 121.8, 82.1, 55.3, 28.4, 25.8, 21.4; HRMS (EI) calcd for C<sub>12</sub>H<sub>13</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 268.0099; found: 268.0091; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 33.7 min (minor), t<sub>2</sub> = 36.0 min (major).



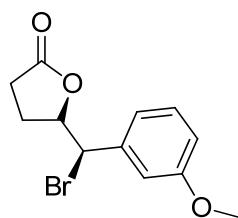
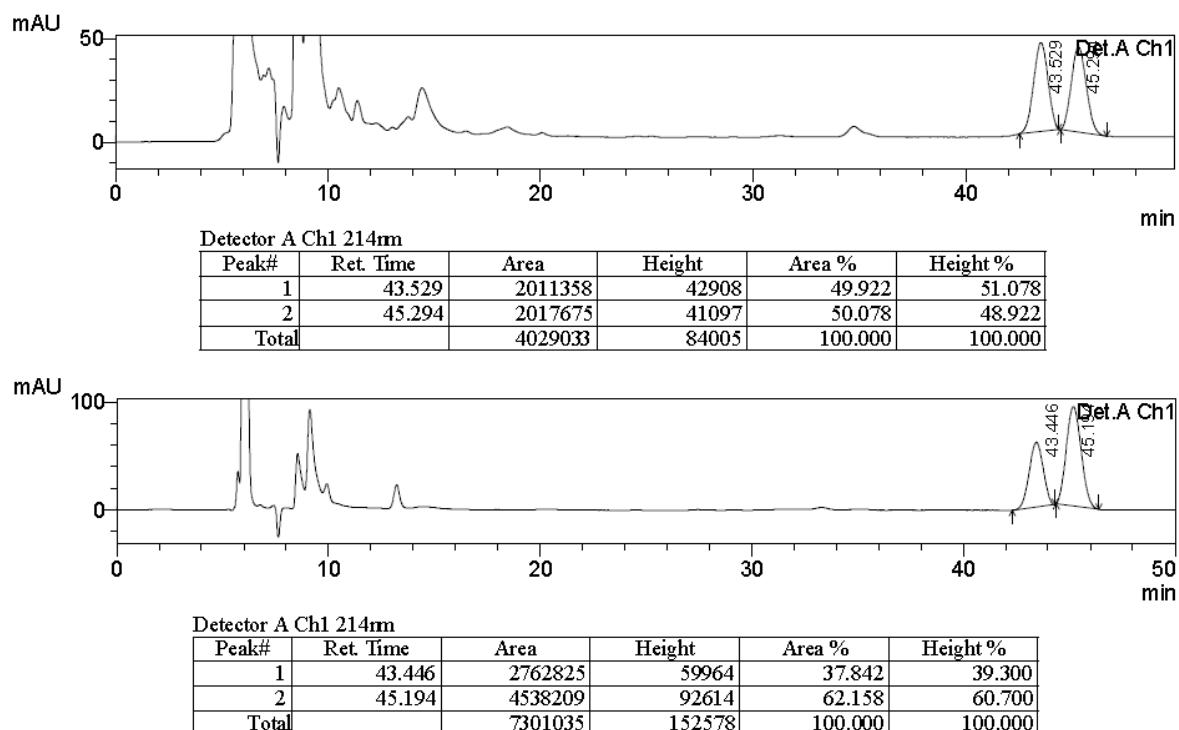
### 5-bromo(o-toyl)methylidihydrofuran-2(3H)-one (**5d**)

(*5-exo* : *6-endo* = 11:1) Colorless oil;  $[\alpha]_D^{25} -20.1$  (*c* 0.6, CHCl<sub>3</sub>, 86% ee); IR (Neat): 3018, 2972, 1732, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  7.56-7.54 (m, 1H), 7.26-7.16 (m, 3H), 5.21 (d, *J* = 6.4 Hz, 1H), 5.04-5.00 (m, 1H), 2.55-2.52 (m, 2H), 2.40 (s, 3H), 2.30-2.26 (m, 1H), 1.97-1.93 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  175.7, 135.6, 130.9, 128.9, 128.2, 126.9, 82.0, 51.5, 28.5, 26.3, 19.4; HRMS (EI) calcd for C<sub>12</sub>H<sub>13</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 268.0099; found: 268.0087; HPLC (Daicel Chiralpak ODH, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 34.1 min (minor), t<sub>2</sub> = 44.0 min (major).



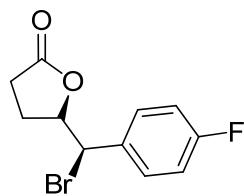
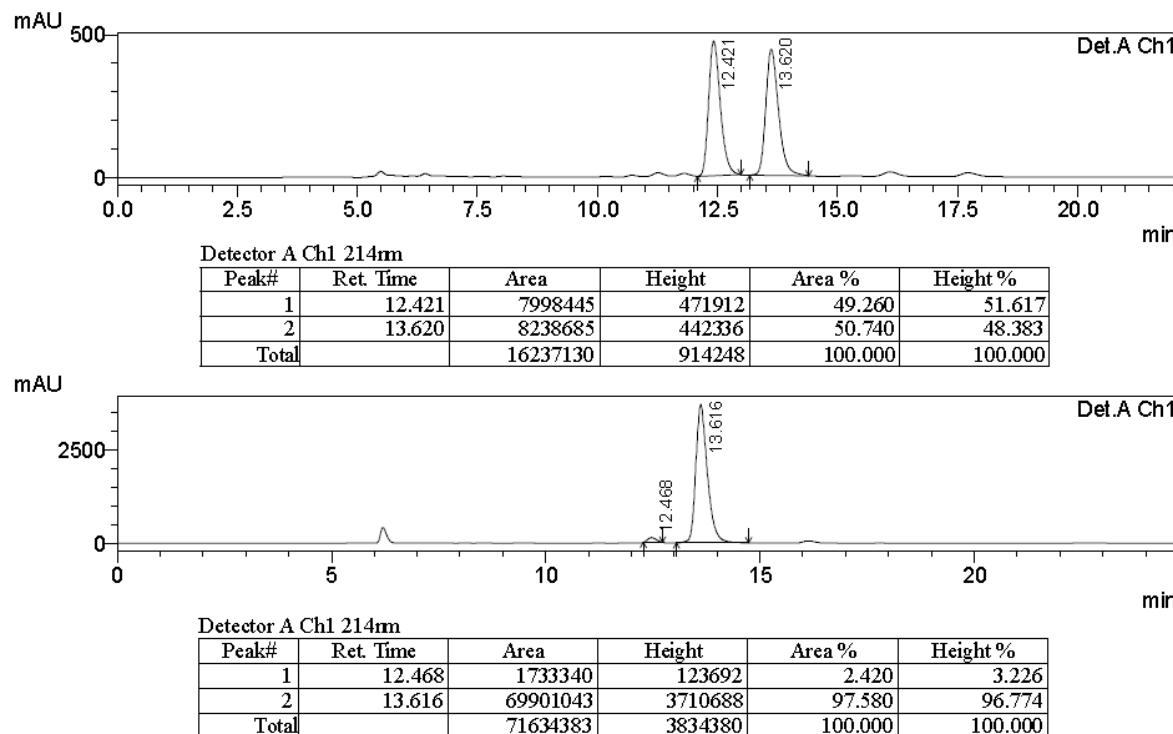
**(5R,6S)-5-bromo-6-(4-methoxyphenyl)tetrahydro-2H-pyran-2-one (8e)**

White solid;  $[\alpha]_D^{25} -7.8$  (*c* 0.2,  $\text{CHCl}_3$ , 24% ee);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.25 (d, *J* = 8.4 Hz, 2H), 6.92 (d, *J* = 8.7 Hz, 2H), 5.49 (d, *J* = 6.8 Hz, 1H), 4.37-4.31 (m, 1H), 3.82 (s, 3H), 2.99-2.88 (m, 1H), 2.76-2.65 (m, 1H), 2.49-2.40 (m, 1H), 2.34-2.23 (m, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ):  $\delta$  169.2, 160.1, 129.3, 127.8, 114.1, 85.4, 55.3, 47.4, 28.5, 28.0; HRMS (EI) calcd for  $\text{C}_{12}\text{H}_{13}\text{BrO}_3$  [ $\text{M}]^+$ : 284.0048; found: 284.0046; HPLC (Daicel Chiralpak IC, *i*- $\text{PrOH}/n\text{-hexane} = 25/75$ , 0.6 mL/min, 214 nm)  $t_1 = 43.4$  min (minor),  $t_2 = 45.2$  min (major).



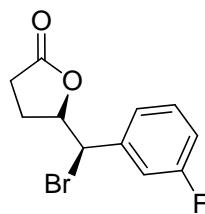
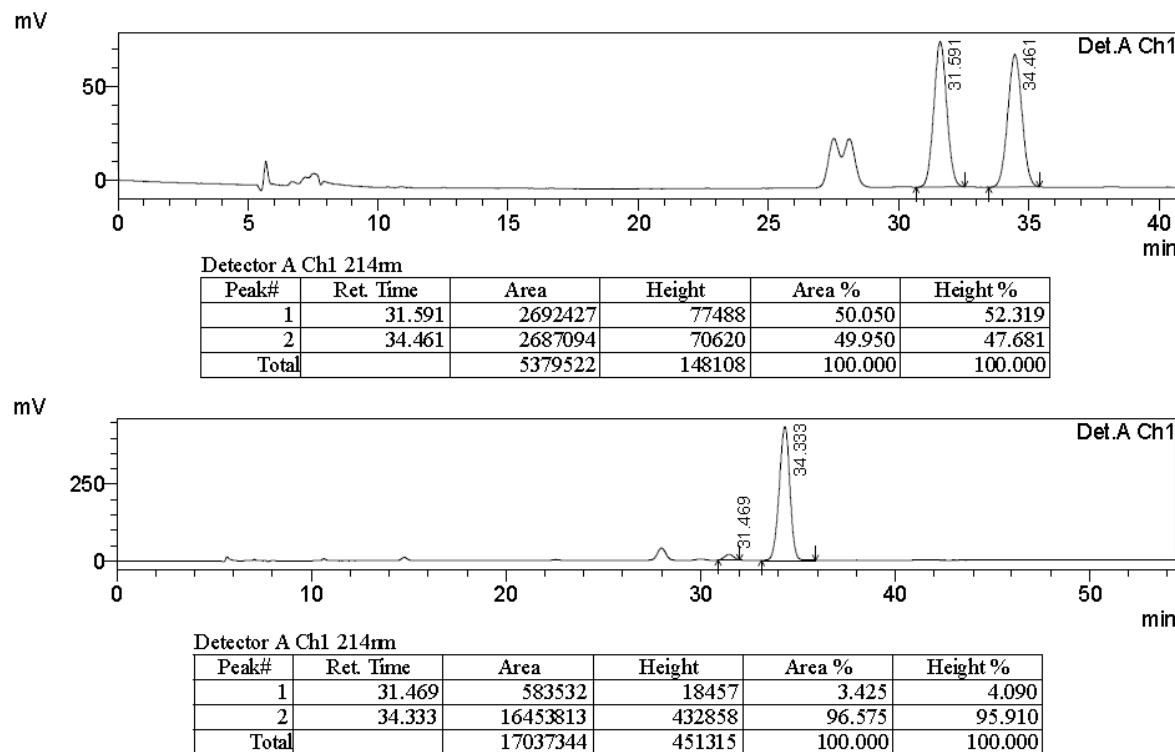
**5-bromo(o-toyl)methylidihydrofuran-2(3H)-one (5f)**

Pale yellow oil;  $[\alpha]_D^{25} -122.9$  (*c* 1.0, CHCl<sub>3</sub>, 95% ee); IR (Neat): 3020, 1780, 1215 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.30-7.25 (m, 1H), 7.03-6.98 (m, 2H), 6.89-6.85 (m, 1H), 4.96-4.87 (m, 2H), 3.82 (s, 3H), 2.55-2.42 (m, 2H), 2.40-2.18 (m, 1H), 2.11-1.98 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  175.9, 159.8, 138.2, 130.0, 120.6, 114.5, 114.2, 82.0, 55.3, 55.0, 28.3, 25.7; HRMS (EI) calcd for C<sub>12</sub>H<sub>13</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 284.0048; found: 284.0044; HPLC (Daicel Chiralpak IA, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 12.5 min (minor), t<sub>2</sub> = 13.6 min (major).



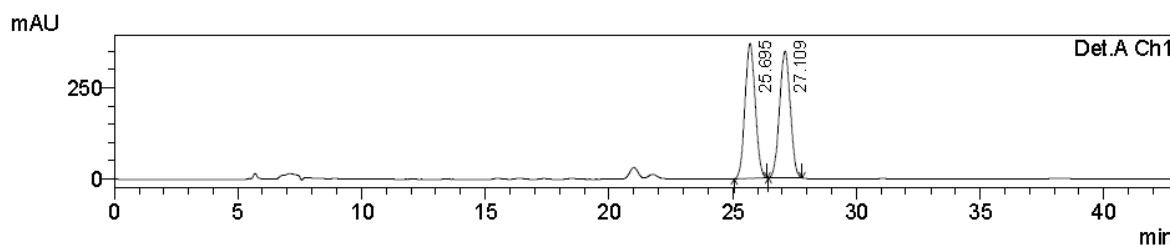
### 5-bromo(p-fluorophenyl)methyldihydrofuran-2(3H)-one (5g)

(5-*exo* : 6-*endo* = 23:1) Yellow oil;  $[\alpha]_D^{25} -100.8$  (*c* 0.6, CHCl<sub>3</sub>, 93% ee); IR (Neat): 3020, 1780, 1644, 1607, 1511, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.47-7.44 (m, 2H), 7.07-7.03 (m, 2H), 4.99 (d, *J* = 5.0 Hz, 1H), 4.88-4.84 (m, 1H), 2.52-2.44 (m, 2H), 2.31-2.23 (m, 1H), 2.11-2.04 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 162.8 (d, *J*<sub>C-F</sub> = 249 Hz), 133.0 (d, *J*<sub>C-F</sub> = 4 Hz), 130.3 (d, *J*<sub>C-F</sub> = 8 Hz), 115.8 (d, *J*<sub>C-F</sub> = 22 Hz), 81.7, 54.5, 28.2, 25.7; HRMS (EI) calcd for C<sub>11</sub>H<sub>10</sub>BrFO<sub>2</sub> [M]<sup>+</sup>: 271.9848; found: 271.9851; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 31.5 min (minor), t<sub>2</sub> = 34.3 min (major).



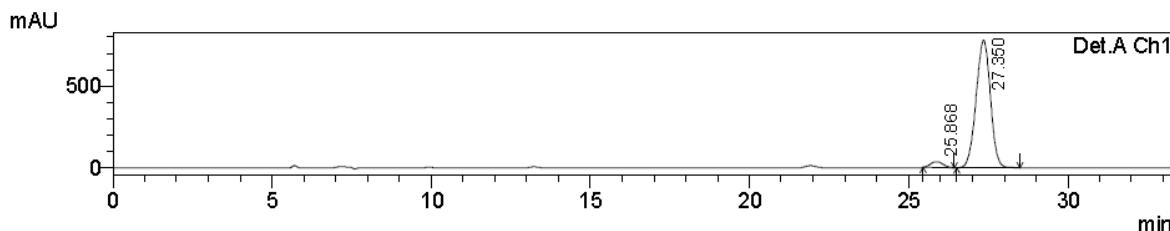
**5-bromo(m-fluorophenyl)methyldihydrofuran-2(3H)-one (**5h**)**

Colorless oil;  $[\alpha]_D^{25} -128.8$  ( $c$  0.9,  $\text{CHCl}_3$ , 92% ee); IR (Neat): 3020, 1782, 1216  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.34-7.31 (m, 1H), 7.23-7.20 (m, 2H), 7.05-7.01 (m, 1H), 4.97 (d,  $J = 5.1$  Hz, 1H), 4.88-4.84 (m, 1H), 2.55-2.44 (m, 2H), 2.32-2.25 (m, 1H), 2.11-2.05 (m, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  175.7, 162.6 (d,  $J_{\text{C-F}} = 248$  Hz), 139.4 (d,  $J_{\text{C-F}} = 7$  Hz), 130.4 (d,  $J_{\text{C-F}} = 8$  Hz), 124.1 (d,  $J_{\text{C-F}} = 3$  Hz), 116.1 (d,  $J_{\text{C-F}} = 21$  Hz), 115.7 (d,  $J_{\text{C-F}} = 23$  Hz), 81.5, 54.2 (d,  $J_{\text{C-F}} = 2$  Hz), 28.2, 25.7; HRMS (EI) calcd for  $\text{C}_{11}\text{H}_{10}\text{BrFO}_2$  [M] $^+$ : 271.9848; found: 271.9857; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm)  $t_1 = 25.9$  min (minor),  $t_2 = 27.4$  min (major).



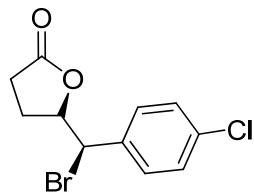
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	25.695	10593981	368772	50.338	51.574
2	27.109	10451836	346267	49.662	48.426
Total		21045817	715039	100.000	100.000



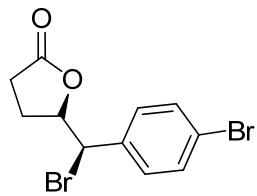
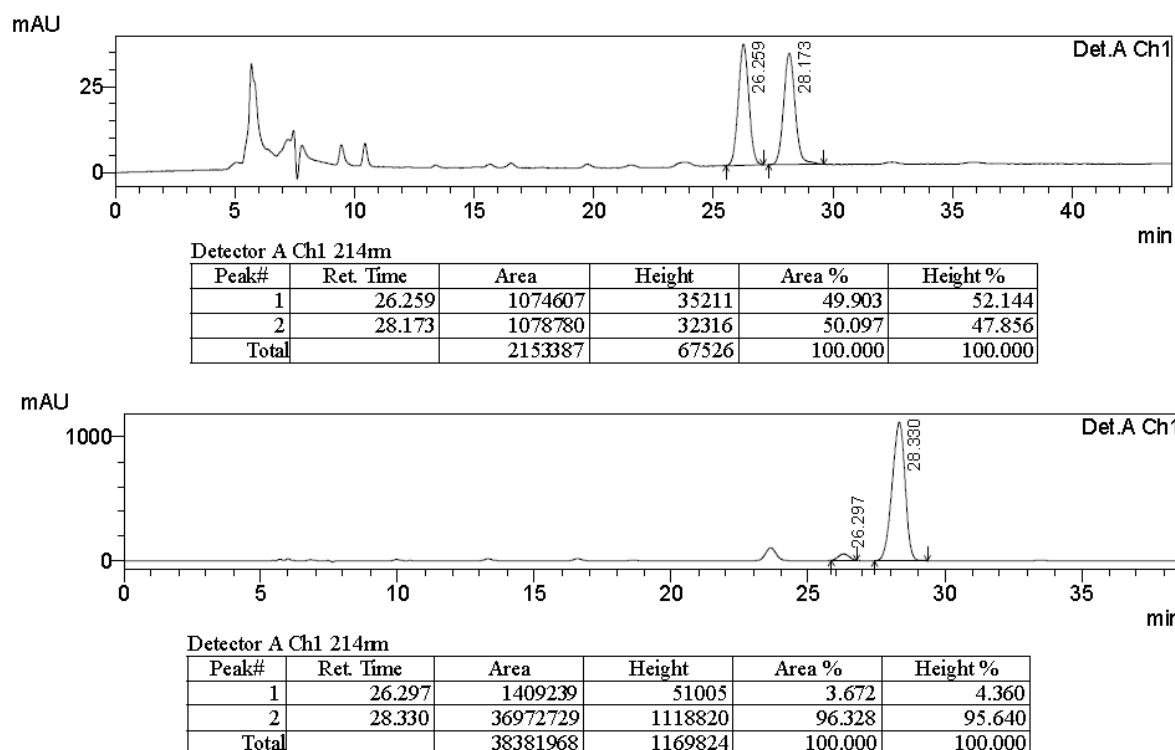
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	25.868	977775	35869	3.862	4.413
2	27.350	24342333	777010	96.138	95.587
Total		25320109	812879	100.000	100.000



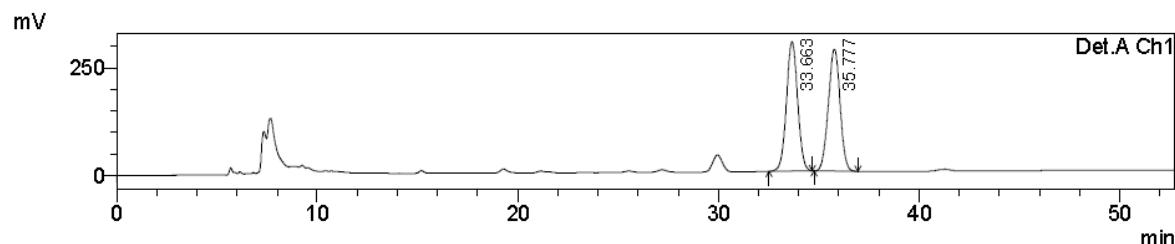
### 5-bromo(p-chlorophenyl)methyldihydrofuran-2(3H)-one (**5i**)

Colorless oil;  $[\alpha]_D^{25} -115.1$  (*c* 0.8, CHCl<sub>3</sub>, 93% ee); IR (Neat): 3020, 1780, 1215 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.42-7.28 (m, 4H), 4.97 (d, *J* = 3.0 Hz, 1H), 4.87-4.84 (m, 1H), 2.55-2.42 (m, 2H), 2.32-2.25 (m, 1H), 2.11-2.04 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 175.8, 135.5, 135.1, 129.8, 129.1, 81.5, 54.3, 28.2, 25.6; HRMS (EI) calcd for C<sub>11</sub>H<sub>10</sub>ClO<sub>2</sub> [M-Br]<sup>+</sup>: 209.0364; found: 209.0370; HPLC (Daicel Chiraldpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 26.3 min (minor), t<sub>2</sub> = 28.3 min (major).



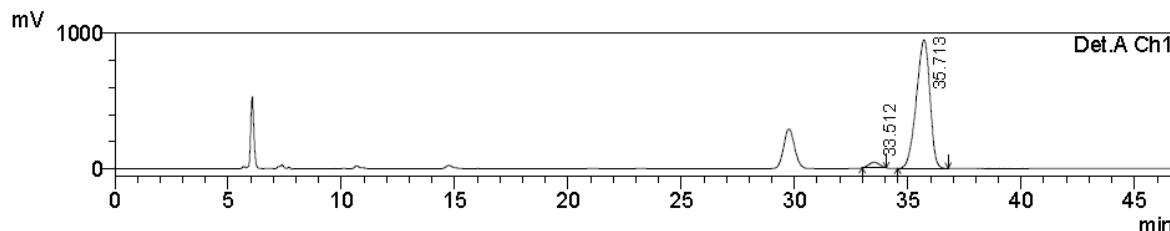
### 5-bromo(p- bromophenyl)methyldihydrofuran-2(3H)-one (5j)

(5-*exo* : 6-*endo* = 5:1) Orange semi-solid;  $[\alpha]_D^{25} -91.7$  (*c* 1.3, CHCl<sub>3</sub>, 94% ee); IR (Neat): 3020, 1780, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  7.50 (d, *J* = 8.8 Hz, 2H), 7.35 (d, *J* = 8.2 Hz, 2H), 4.95 (d, *J* = 4.9 Hz, 1H), 4.88-4.82 (m, 1H), 2.55-2.41 (m, 2H), 2.35-2.23 (m, 1H), 2.13-2.01 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  175.7, 136.1, 132.0, 130.1, 123.3, 81.4, 54.4, 28.2, 25.6; HRMS (EI) calcd for C<sub>11</sub>H<sub>10</sub>Br<sub>2</sub>O<sub>2</sub> [M]<sup>+</sup>: 331.9048; found: 331.9054; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 33.5 min (minor), t<sub>2</sub> = 35.7 min (major).



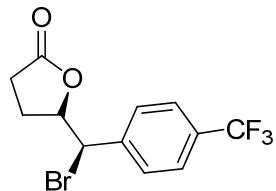
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	33.663	11653935	299599	50.345	51.478
2	35.777	11494112	282395	49.655	48.522
Total		23148046	581995	100.000	100.000



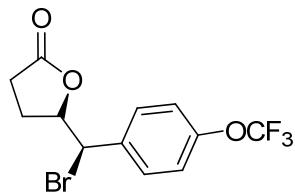
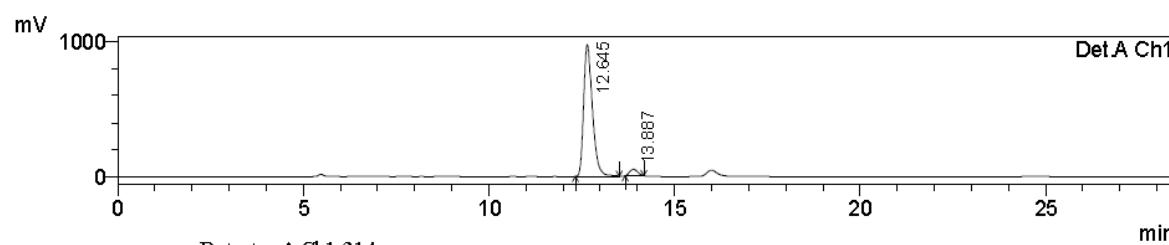
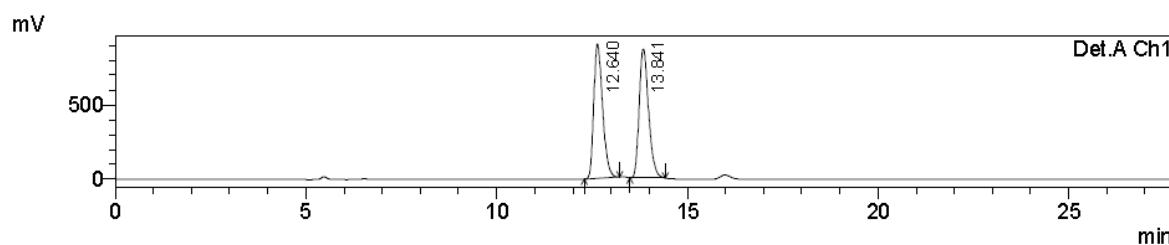
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	33.512	1303128	39621	3.206	4.017
2	35.713	39347033	946756	96.794	95.983
Total		40650160	986378	100.000	100.000



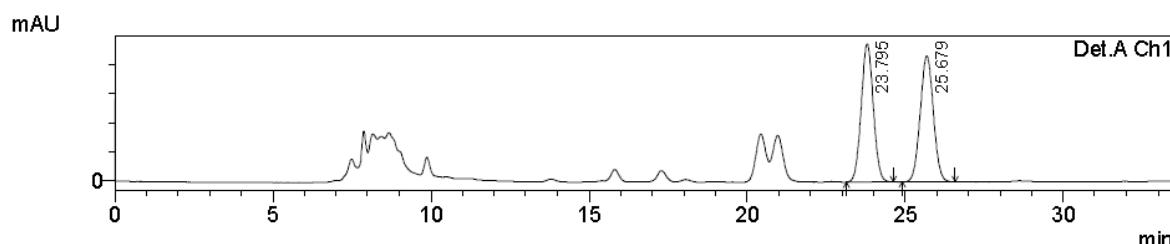
### 5-bromo-[{(p-trifluoromethyl)phenyl]methyl}dihydrofuran-2(3H)-one (5k)

Yellow semi-solid;  $[\alpha]_D^{25} -100.9$  (*c* 0.6, CHCl<sub>3</sub>, 91% ee); IR (Neat): 3021, 1781, 1326, 1216, 1170, 1115, 1068 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.66-7.59 (m, 4H), 5.03 (d, *J* = 4.6 Hz, 1H), 4.90-4.84 (m, 1H), 2.55-2.44 (m, 2H), 2.38-2.27 (m, 1H), 2.17-2.05 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  175.7, 141.0, 131.2 (q, *J*<sub>C,F</sub> = 33 Hz), 129.0, 125.8 (q, *J*<sub>C,F</sub> = 4 Hz), 123.7 (q, *J*<sub>C,F</sub> = 272 Hz), 81.2, 54.1, 28.2, 25.7; HRMS (EI) calcd for C<sub>12</sub>H<sub>10</sub>BrF<sub>3</sub>O<sub>2</sub> [M]<sup>+</sup>: 321.9816; found: 321.9803; HPLC (Daicel Chiraldak IA, *i*-PrOH/*n*-hexane = 20/80, 0.6 mL/min, 214 nm) t<sub>1</sub> = 12.6 min (major), t<sub>2</sub> = 13.9 min (minor).



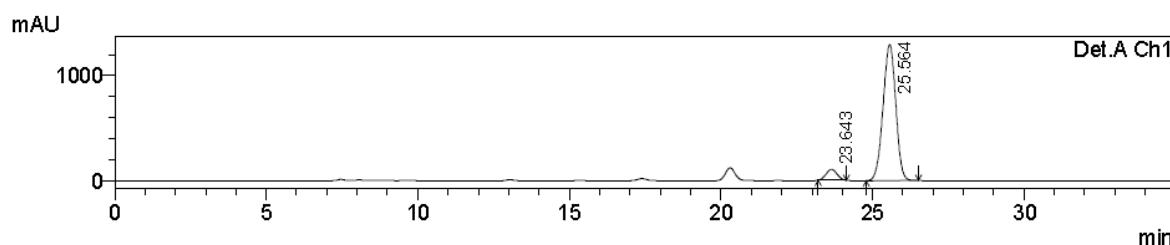
### 5-bromo-[{(p-trifluoromethoxy)phenyl]methyl}dihydrofuran-2(3H)-one (5l)

Yellow semi-solid;  $[\alpha]_D^{25} -96.2$  ( $c$  1.0, CHCl<sub>3</sub>, 87% ee); IR (Neat): 3020, 1781, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  7.53-7.50 (m, 2H), 7.21 (d,  $J$  = 8.2 Hz, 2H), 5.01 (d,  $J$  = 3.0 Hz, 1H), 4.87-4.83 (m, 1H), 2.53-2.49 (m, 2H), 2.35-2.27 (m, 1H), 2.13-2.08 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 149.4 (q,  $J_{C,F}$  = 1.8 Hz), 135.8, 130.1, 121.1, 120.3 (q,  $J_{C,F}$  = 258 Hz), 81.4, 54.2, 28.2, 25.7; HRMS (EI) calcd for C<sub>12</sub>H<sub>10</sub>BrF<sub>3</sub>O<sub>3</sub> [M]<sup>+</sup>: 337.9765; found: 337.9769; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 23.6 min (minor), t<sub>2</sub> = 25.6 min (major).



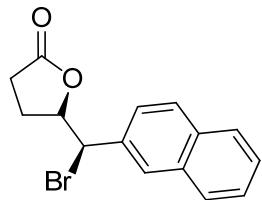
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.795	2517899	94703	50.106	52.262
2	25.679	2507247	86505	49.894	47.738
Total		5025145	181208	100.000	100.000



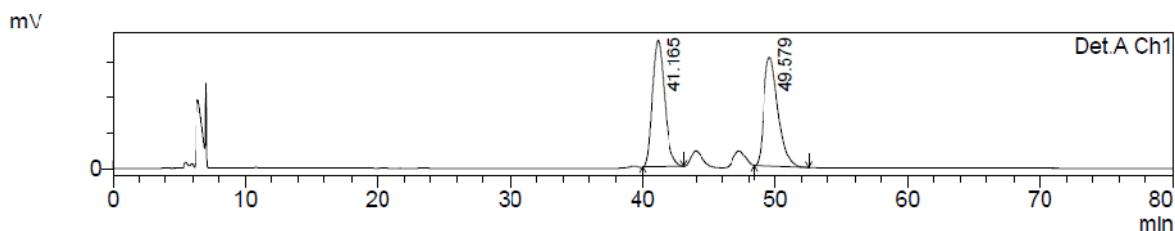
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.643	2617723	103788	6.501	7.435
2	25.564	37651339	1292232	93.499	92.565
Total		40269062	1396020	100.000	100.000



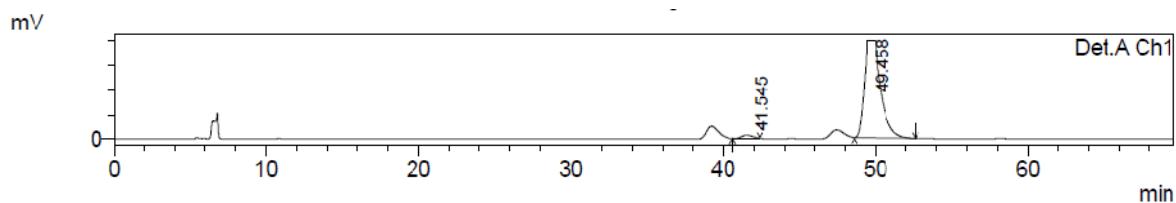
### 5-bromo(naphthalene-2-yl)methyldihydrofuran-2(3H)-one (5m)

Yellow semi-solid;  $[\alpha]_D^{25} -102.0$  (*c* 1.0, CHCl<sub>3</sub>, 95% ee); IR (Neat): 3019, 1776, 1326, 1221, 1210, 1169, 1136, 1068 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.88-7.83 (m, 4H), 7.60-7.51 (m, 3H), 5.15 (d, *J* = 5.6 Hz, 1H), 5.05-4.98 (m, 1H), 2.52-2.45 (m, 2H), 2.31-2.19 (m, 1H), 2.16-2.06 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  178.6, 134.3, 133.3, 132.9, 128.9, 128.1, 127.7, 127.6, 127.0, 126.8, 125.6, 82.0, 55.7, 28.4, 25.9; HRMS (EI) calcd for C<sub>15</sub>H<sub>13</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 304.0099; found: 304.0096; HPLC (Daicel Chiralpak IA, *i*-PrOH/*n*-hexane = 5/95, 0.6 mL/min, 214 nm) t<sub>1</sub> = 41.5 min (minor), t<sub>2</sub> = 49.5 min (major).



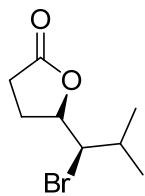
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	41.165	112834642	1786832	49.949	53.509
2	49.579	113064004	1552454	50.051	46.491
Total		225898646	3339286	100.000	100.000



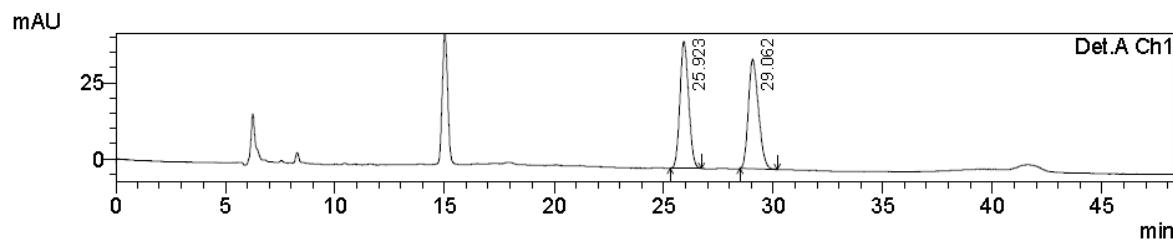
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	41.545	6226762	128075	2.149	3.143
2	49.458	283510956	3946707	97.851	96.857
Total		289737718	4074782	100.000	100.000



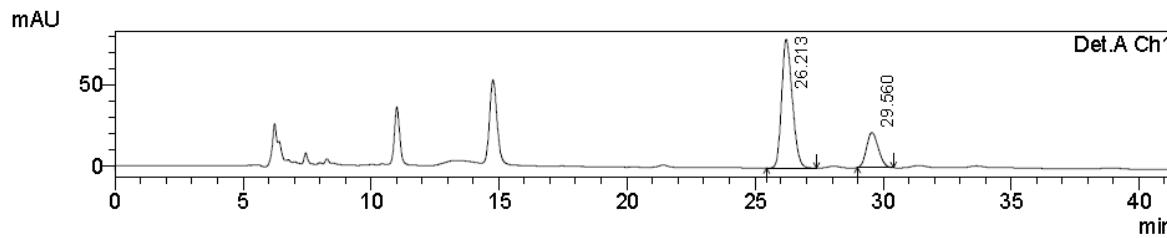
### 5-bromo(isopropyl)methyldihydrofuran-2(3H)-one (5n)

Yellow oil;  $[\alpha]_D^{25} -19.9$  (*c* 1.1, CHCl<sub>3</sub>, 53% ee); IR (Neat): 3019, 1772, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  4.76-4.70 (m, 1H), 3.88-3.84 (m, 1H), 2.77-2.65 (m, 1H), 2.58-2.43 (m, 1H), 2.41-2.44 (m, 1H), 2.22-2.05 (m, 1H), 1.09 (d, *J* = 1.0 Hz, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  176.3, 79.9, 66.8, 32.9, 28.3, 26.7, 21.2, 20.3; HRMS (EI) calcd for C<sub>8</sub>H<sub>13</sub>O<sub>2</sub> [M-Br]<sup>+</sup>: 141.0910; found: 141.0909; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 26.2 min (major), t<sub>2</sub> = 29.6 min (minor).



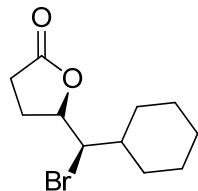
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	25.923	1152502	41618	50.049	53.650
2	29.062	1150224	35954	49.951	46.350
Total		2302727	77572	100.000	100.000



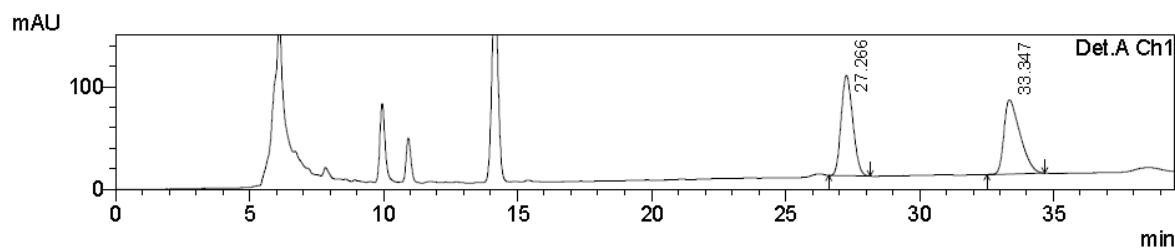
Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	26.213	2295322	79603	76.786	78.529
2	29.560	693909	21765	23.214	21.471
Total		2989231	101368	100.000	100.000



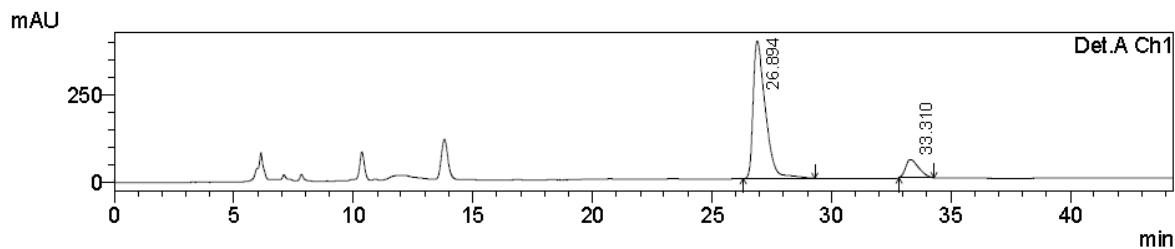
### 5-bromo(cyclohexyl)methyldihydrofuran-2(3H)-one (5o)

Colorless oil;  $[\alpha]_D^{25} -31.1$  (*c* 1.0, CHCl<sub>3</sub>, 75% ee); IR (Neat): 3020, 2933, 1775, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  4.82-4.76 (m, 1H), 3.87-3.84 (m, 1H), 2.78-2.67 (m, 1H), 2.57-2.45 (m, 1H), 2.42-2.30 (m, 1H), 2.26-2.14 (m, 1H), 2.05 (d, *J* = 12.0 Hz, 1H), 1.89-1.74 (m, 4H), 1.67 (d, *J* = 12.0 Hz, 1H), 1.35-1.05 (m, 5H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  179.3, 78.8, 66.3, 54.3, 42.4, 31.3, 31.2, 28.2, 26.6, 26.1, 26.0, 25.9; HRMS (EI) calcd for C<sub>11</sub>H<sub>17</sub>BrO<sub>2</sub> [M]<sup>+</sup>: 260.0412; found: 260.0409; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm) t<sub>1</sub> = 26.9 min (major), t<sub>2</sub> = 33.3 min (minor).



Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	27.266	2969120	97785	49.680	57.533
2	33.347	3007346	72178	50.320	42.467
Total		5976466	169962	100.000	100.000



Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	26.894	14067363	393692	87.561	88.323
2	33.310	1998503	52048	12.439	11.677
Total		16065865	445740	100.000	100.000

### C. Study on the effect on regioselectivity by various catalysts

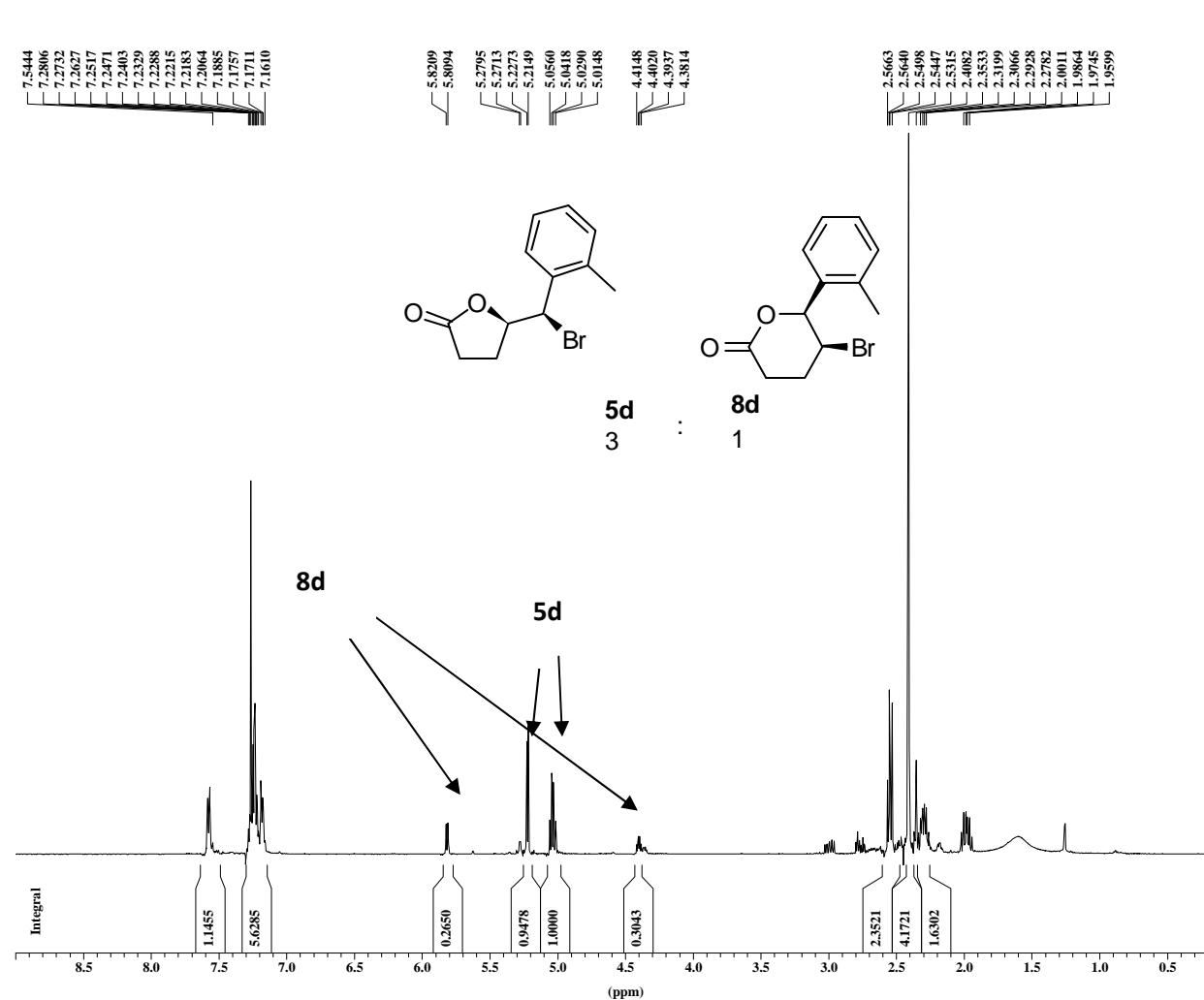
The regioselectivities of the reaction were determined via the ratio of the integration values of both the *5-exo* and *6-endo* lactone products (spectrums attached in the following pages). The <sup>1</sup>H signals corresponding to the *5-exo* and *6-endo* lactones are in agreement with the literature values.<sup>5</sup>

### General procedure for the bromolactonization of **4d**

**4d** (95 mg, 0.05 mmol, 1.0 eq) and the corresponding catalyst (3.0 mg, 0.005 mmol, 0.1 eq) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (1.5 ml) at 25 °C in the dark under nitrogen atmosphere. *N*-bromosuccinimide (10.6 mg, 0.06 mmol, 1.2 eq) was then added. The reaction mixture was stirred at 25 °C for 24 h before subjecting to the work up procedure for the bromolactonization as described above.

Scheme 2, with Ph<sub>3</sub>P=S (10 mol%) as catalyst

cc-i-156



\*\*\* Current Data Parameters \*\*\*

NAME : ck0207  
EXPNO : 1  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCMNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 2751.27 Hz  
PULPROG : zg  
SFO1 : 500.2327513 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 15.0080 ppm  
TD : 32768  
TE : 296.4 K

\*\*\* Processing Parameters \*\*\*

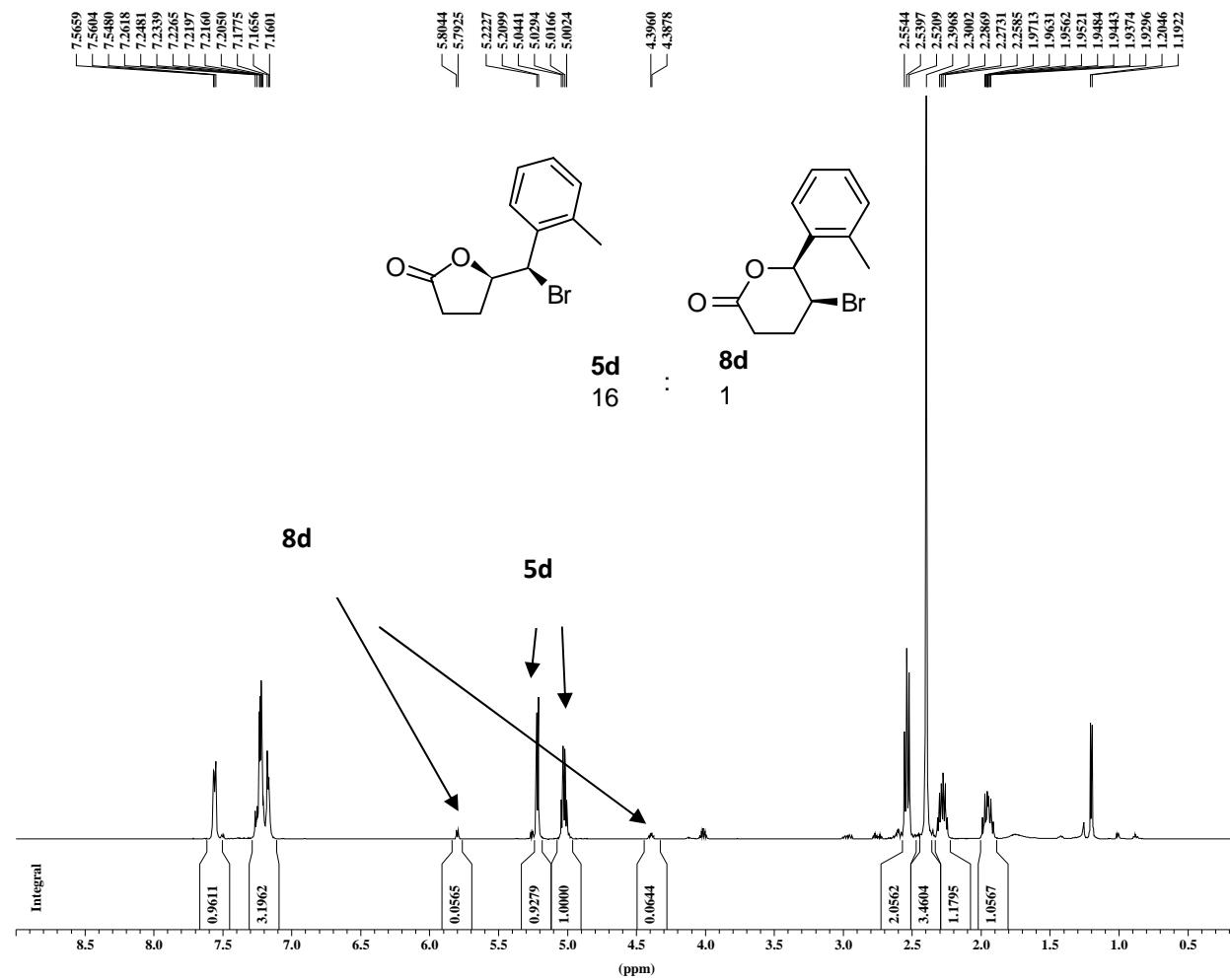
LB : 0.10 Hz  
SF : 500.2300083 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

Scheme 2, with 6b:7 (1:1, 10 mol%) as catalyst

ck-v-119



\*\*\* Current Data Parameters \*\*\*

NAME : ck0131  
EXPNO : 1  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 2751.27 Hz  
PULPROG : zg  
SFO1 : 500.2327513 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 15.0080 ppm  
TD : 32768  
TE : 298.0 K

\*\*\* Processing Parameters \*\*\*

LB : 0.10 Hz  
SF : 500.2300083 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

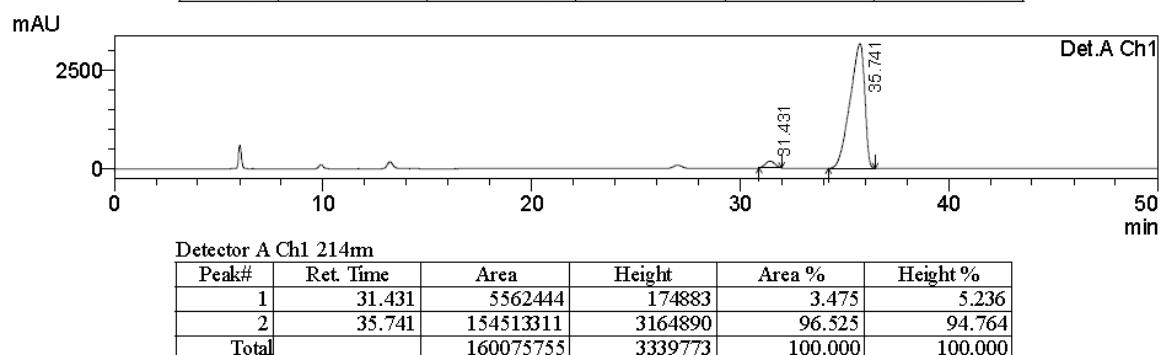
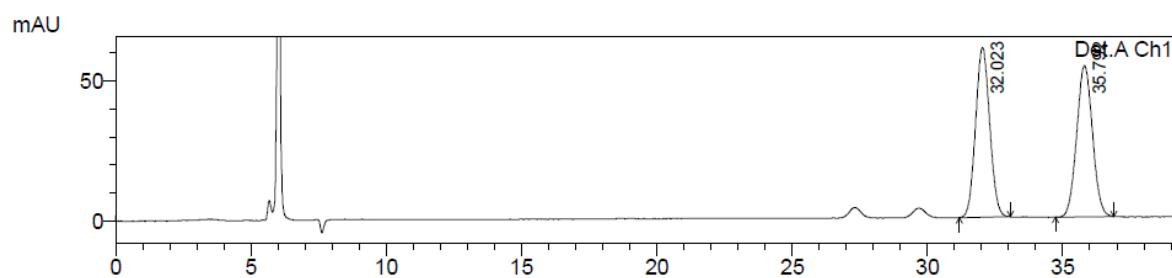
#### D. Study on the effect of various halogen sources

Olefinic acid **2** was synthesized according to our previous report.<sup>2</sup>

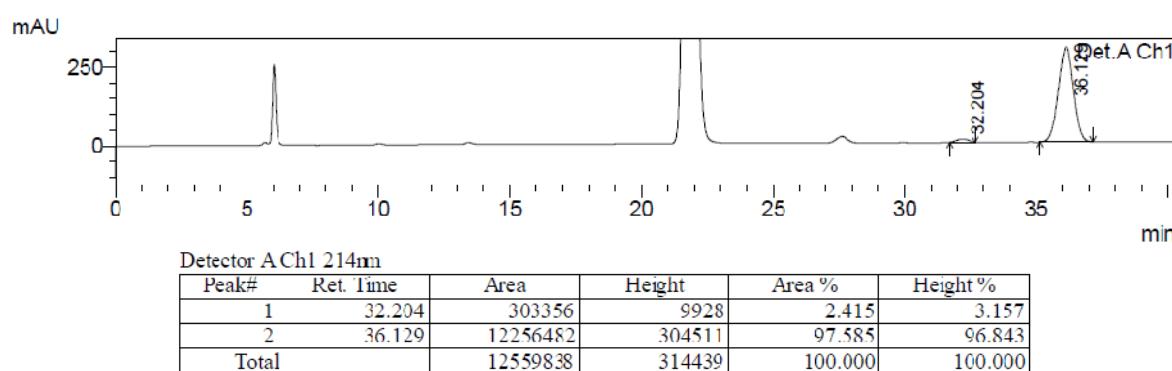
#### General procedure for the halolactonization with various halogen sources

The corresponding olefinic acid (0.05 mmol, 1.0 eq) and the corresponding catalyst (0.005 mmol, 0.1 eq) were dissolved in the stated solvent blends at -78 °C in the dark under nitrogen atmosphere. The corresponding halogen source (0.06 mmol, 1.2 eq) was then added. The reaction mixture was stirred at -78 °C and monitored using TLC until completion. Upon completion the mixture was subjected to the work up procedure for the bromolactonization as described above.

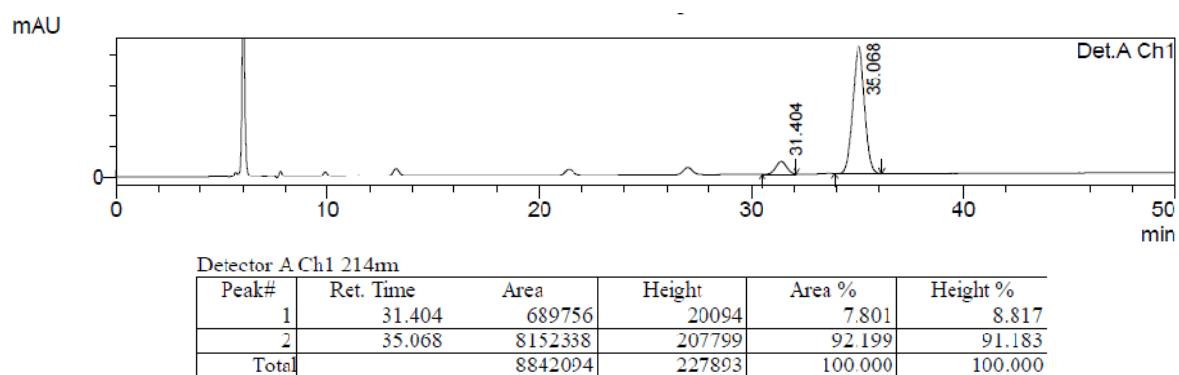
#### Bromolactonization with NBS (Table 3, entry 1)



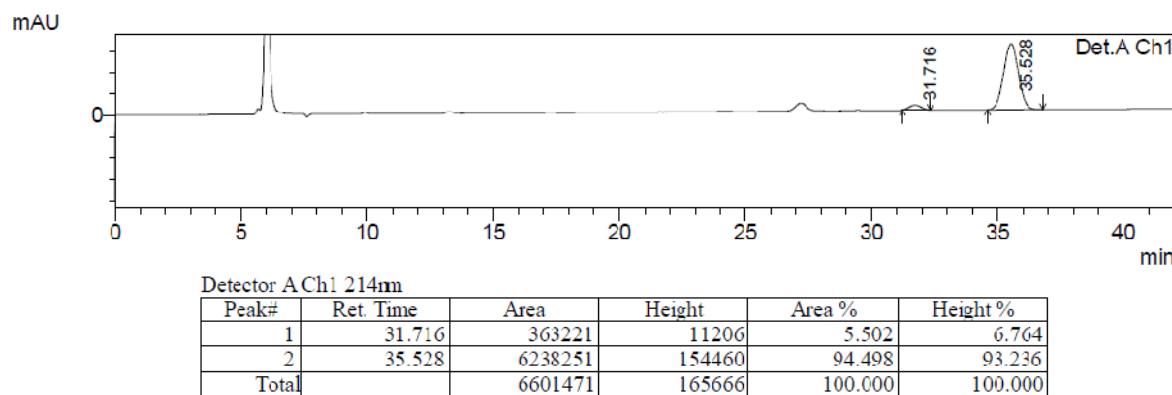
### Bromolactonization with NBP (Table 3, entry 2)



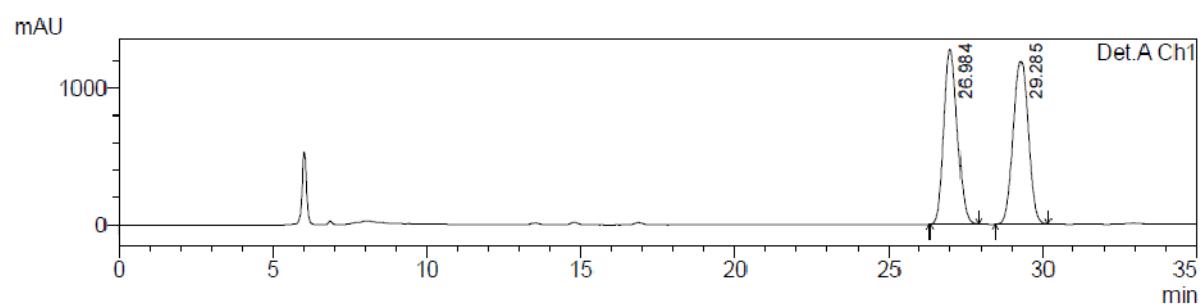
### Bromolactonization with DBH (Table 3, entry 3)



### Bromolactonization with TABCO (Table 3, entry 4)

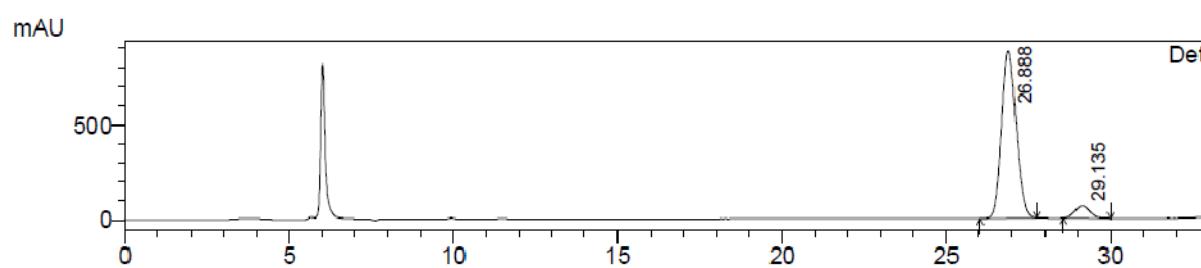


**Bromolactonization of acid 2with TABCO (Table 3, entry 5)**



Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	26.984	39751616	1271491	49.873	51.686
2	29.285	39954651	1188519	50.127	48.314
Total		79706268	2460010	100.000	100.000

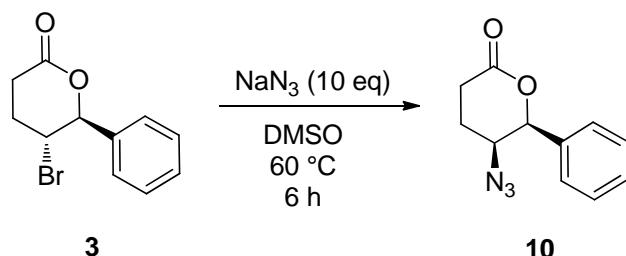


Detector A Ch1 214nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	26.888	26957774	877305	92.981	93.204
2	29.135	2034852	63972	7.019	6.796
Total		28992626	941277	100.000	100.000

## E. Synthetic application of bromolactones 2 and 5a

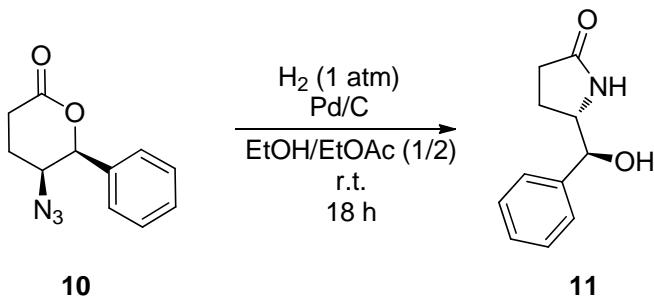
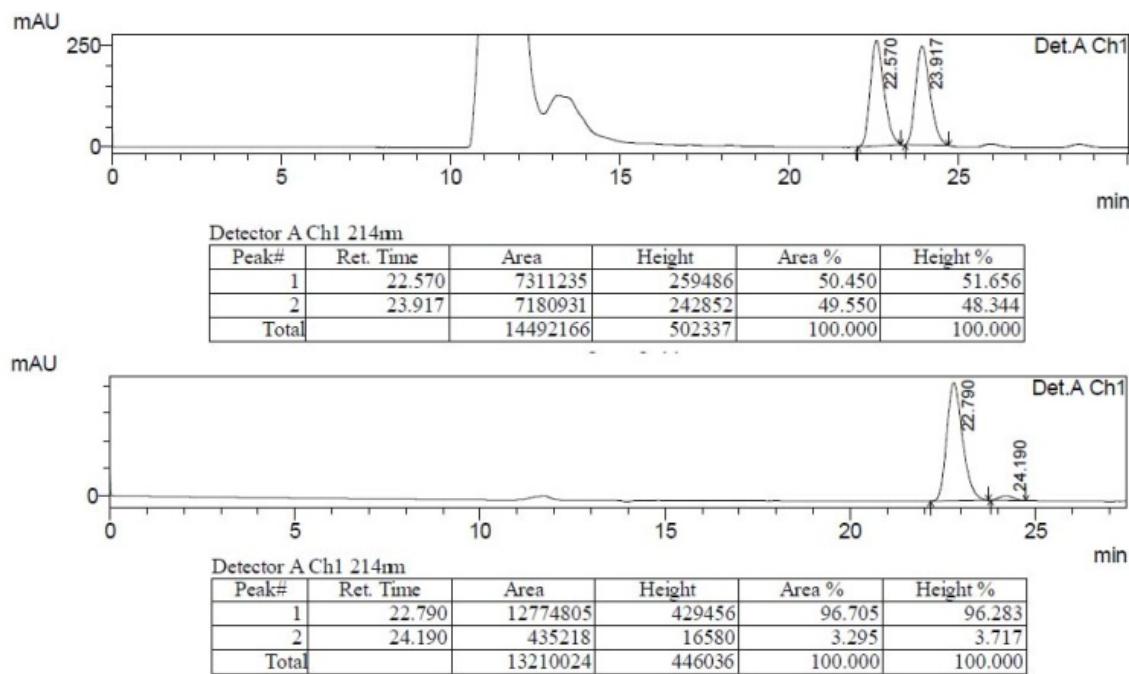
Bromolactone **3** was prepared using the procedure described in our previous report.<sup>2</sup>



### Synthesis of (5S,6S)-5-azido-6-phenyltetrahydro-2H-pyran-2-one (**10**)

Bromolactone **3** (434 mg, 2 mmol, 1.0 eq) and NaN<sub>3</sub> (1.30 g, 20 mmol, 10.0 eq) were first evacuated *in vacuo* and flushed with N<sub>2</sub> for 3 cycles. DMSO (6 ml) was injected and the reaction mixture was stirred at 60 °C for 6 hours. Upon termination, the reaction mixture was cooled to r.t. before quenching with water (30 ml). The product was extracted with ethyl acetate (3 x 20 ml). The combined ethyl acetate layer was further washed with brine (50 ml), dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The product was purified over silica gel with hexane:ethyl acetate (1:1) as the eluent.

Pale yellow oil; Yield: 63%; [α]<sub>D</sub><sup>25</sup> +36.0 (c 1.0, CHCl<sub>3</sub>, 93% ee); IR (Neat): 3020, 2110, 1712, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.40-7.36 (m, 5H), 5.48 (d, *J* = 2.0 Hz, 1H), 4.09-4.06 (m, 1H), 2.84-2.61 (m, 2H), 2.37-2.15 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.3, 135.8, 128.7, 128.6, 126.0, 81.7, 59.0, 25.3, 24.8; HPLC (Daicel Chiralpak IC, *i*-PrOH (1% Ethanolamine)/*n*-hexane = 80/20, 0.3 mL/min, 214 nm) t<sub>1</sub> = 22.8 min (major), t<sub>2</sub> = 24.2 min (minor).

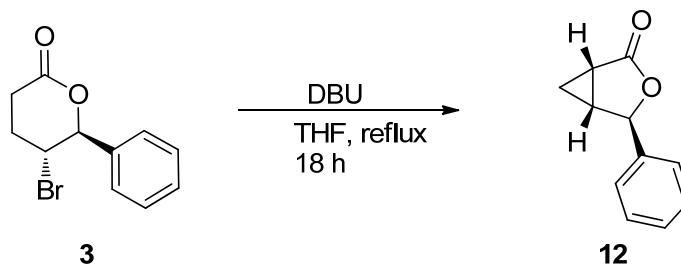
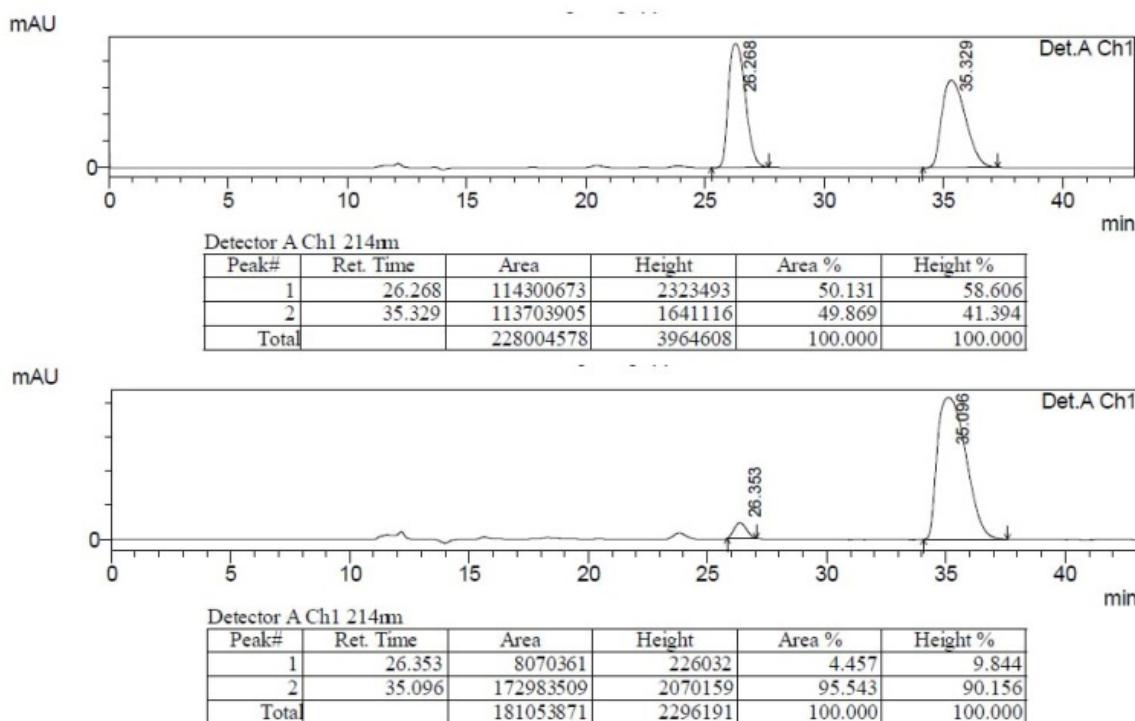


### Synthesis of (S)-5-((S)-hydroxy(phenyl)methyl)pyrrolidin-2-one (**11**)<sup>6</sup>

The azide **10** (217 mg, 1 mmol) was dissolved in ethanol (2 ml) and ethyl acetate (4 ml). Pd/C (80 mg) was added before the attachment of a hydrogen balloon. The reaction mixture was stirred at r.t. for 18 h. The reacted mixture was filtered over celite with several washings of chloroform. The filtrate was dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The product was then purified over silica gel with MeOH:chloroform (1:20) and 1% ammonia solution as the eluent.

White solid, mp 136.6-138.3; Yield: 55%;  $[\alpha]_D^{25} +27.0$  (*c* 1.0, CHCl<sub>3</sub>, 91% ee); IR (KBr): 2937, 1682, 1541, 1350, 1164 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, MeOD):  $\delta$  7.38-7.27 (m, 5H), 4.49 (d, *J* = 6.9 Hz, 1H), 3.90-3.30 (m, 1H), 2.23-2.03 (m, 2H), 1.93-1.77 (m, 2H); <sup>13</sup>C NMR (75 MHz, MeOD):  $\delta$  181.1, 142.3, 129.4, 128.9, 128.0, 78.0, 61.7, 31.0, 23.8; HRMS (ESI) calcd for C<sub>11</sub>H<sub>13</sub>NNaO<sub>2</sub> [M]<sup>+</sup>: 214.0844; found: 214.0846. HPLC (Daicel Chiraldak IC, *i*-PrOH

(1% Ethanolamine)/*n*-hexane = 80/20, 0.3 mL/min, 214 nm)  $t_1$  = 26.4 min (minor),  $t_2$  = 35.1 min (major).

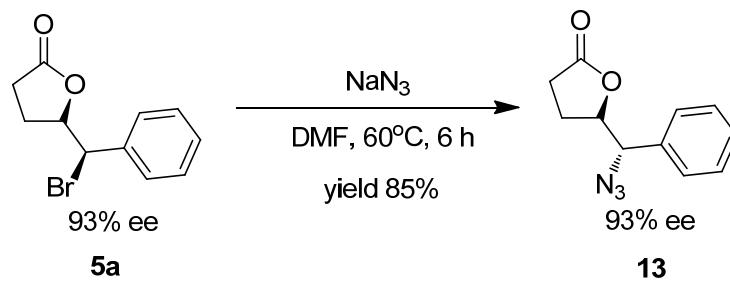
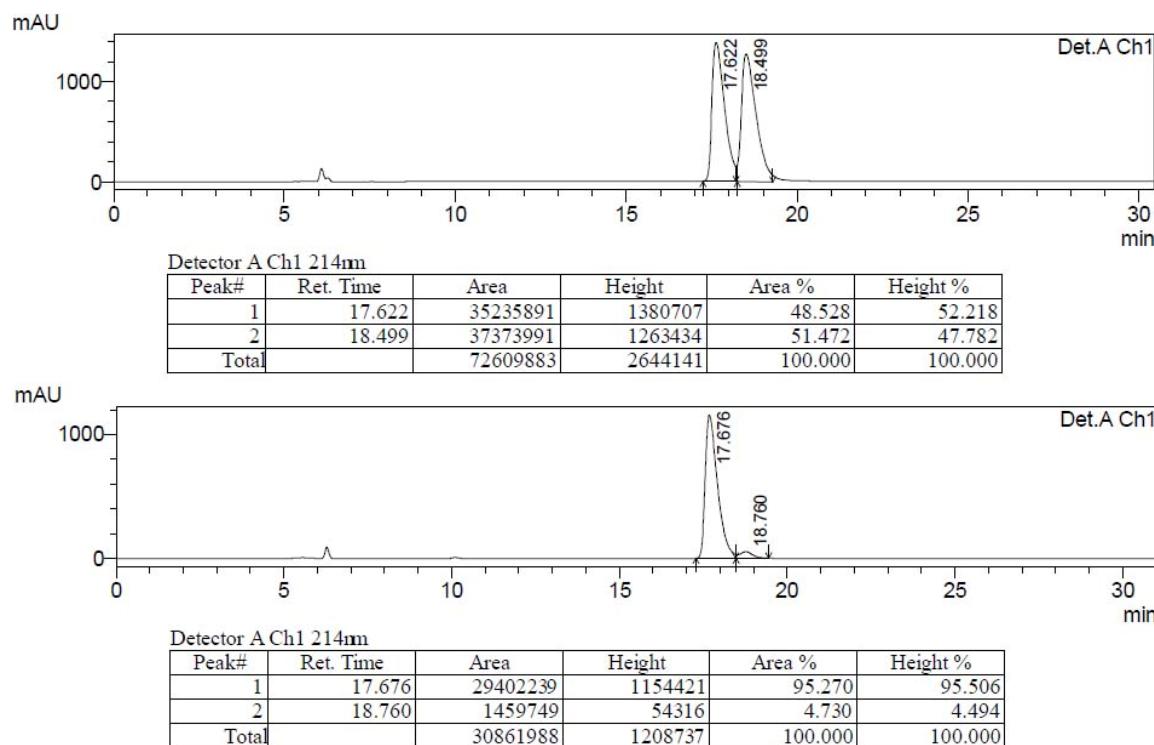


### Synthesis of (1S,4R,5R)-4-phenyl-3-oxabicyclo[3.1.0]hexan-2-one (12)<sup>7</sup>

The bromolactone **3** (147 mg, 0.67 mmol, 1.0 equiv) was evacuated *in vacuo* and flushed with N<sub>2</sub> for 3 cycles. THF (6 ml) was injected followed by DBU (110 µL, 0.74 mmol, 1.1 equiv). The reaction mixture was then refluxed for 18 h. The reacted mixture was then quenched with water (30 ml) and extracted with ethyl acetate (3 x 15 ml). The ethyl acetate layers were combined and was dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The product was then purified over silica gel with EtOAc:Hexane (1:3) as the eluent.

Colorless oil; Yield: 42%;  $[\alpha]_D^{25} +36.0$  (*c* 1.0, CHCl<sub>3</sub>, 91% ee); IR (Neat): 3020, 1767, 1641, 1216 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.39–7.33 (m, 5H), 5.31 (s, 1H), 2.28–2.19 (m, 2H),

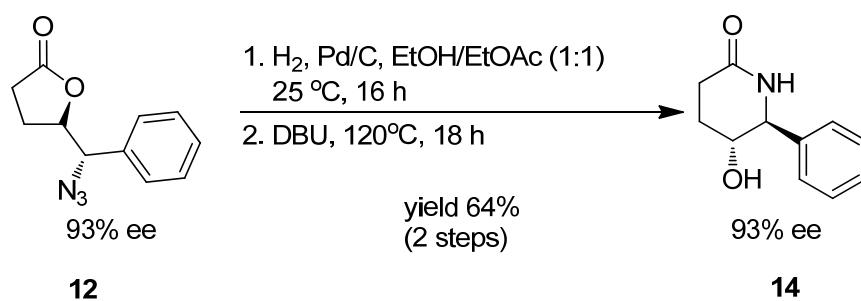
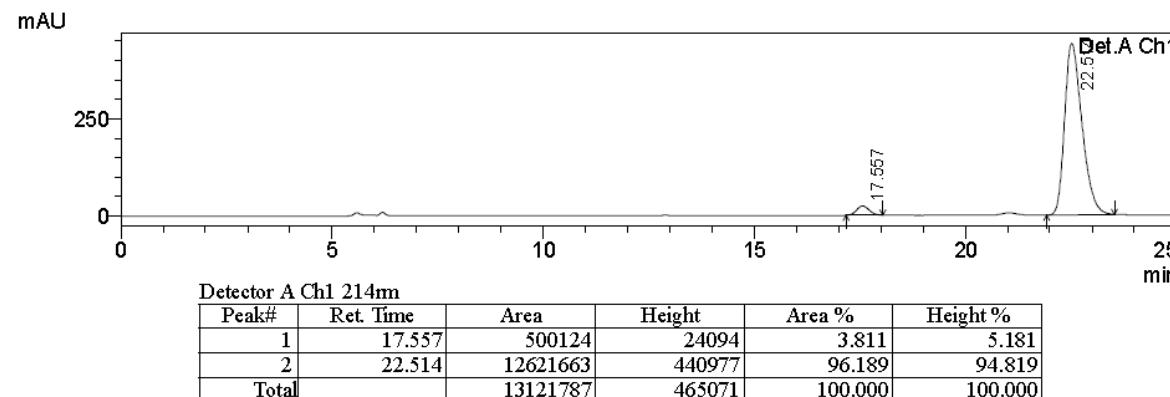
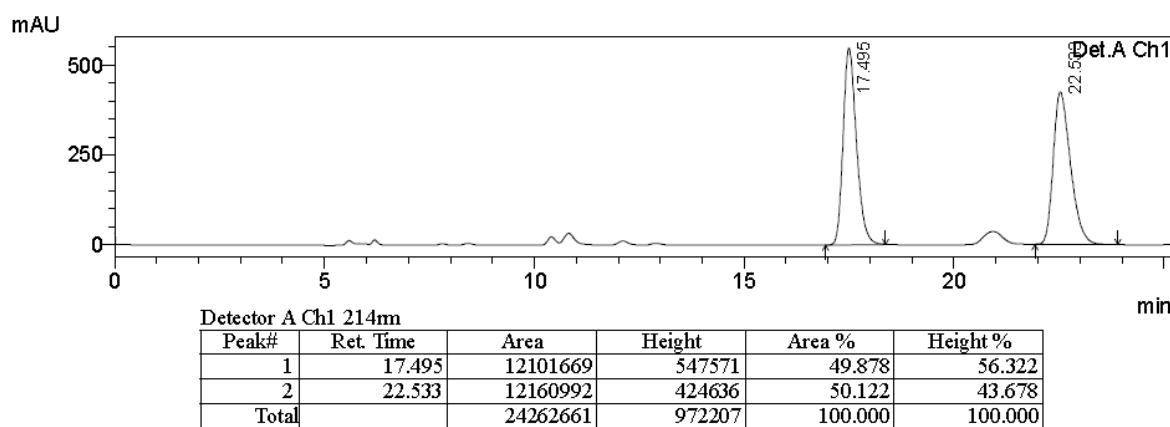
1.34 (ddd,  $J = 8.8, 7.6, 5$  Hz, 1H), 1.07-1.04 (m, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  175.9, 139.6, 128.8, 128.7, 125.5, 81.5, 24.6, 17.7, 12.8; DEPT 135 (125 MHz,  $\text{CDCl}_3$ ) 128.9 (CH), 128.8 (CH), 125.6 (CH), 81.6 (CH), 24.7 (CH), 17.8 (CH), 12.9 (CH<sub>2</sub>); HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{10}\text{NaO}_2$  [M]<sup>+</sup>: 197.0578; found: 197.0573. HPLC (Daicel Chiralpak IB, *i*-PrOH/*n*-hexane = 15/85, 0.6 mL/min, 214 nm)  $t_1 = 17.6$  min (major),  $t_2 = 18.7$  min (minor).



### Synthesis of 5-azido(phenyl)methylidihydro-furan-2(3H)-one (13)

**5a** (213 mg, 0.84 mmol, 1.0 eq) and sodium azide (82 mg, 1.26 mmol, 1.5 eq) were dissolved in DMF (15 ml) and the reaction mixture was stirred for 6 hours at 60 °C. Upon completion,  $\text{H}_2\text{O}$  (50 ml) was added before extracting with diethyl ether (3 x 50 ml). The combined organic layer were washed with brine (2 x 100 ml), dried over sodium sulfate and the solvent was removed *in vacuo*. The product was purified using flash column chromatography over silica gel with EtOAc:Hexane (1:3) as the eluent. Light brown oil;  $[\alpha]_D^{25} -146.7$  (*c* 1.0,  $\text{CHCl}_3$ ,

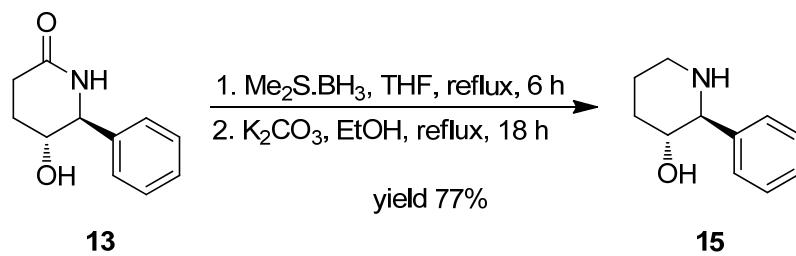
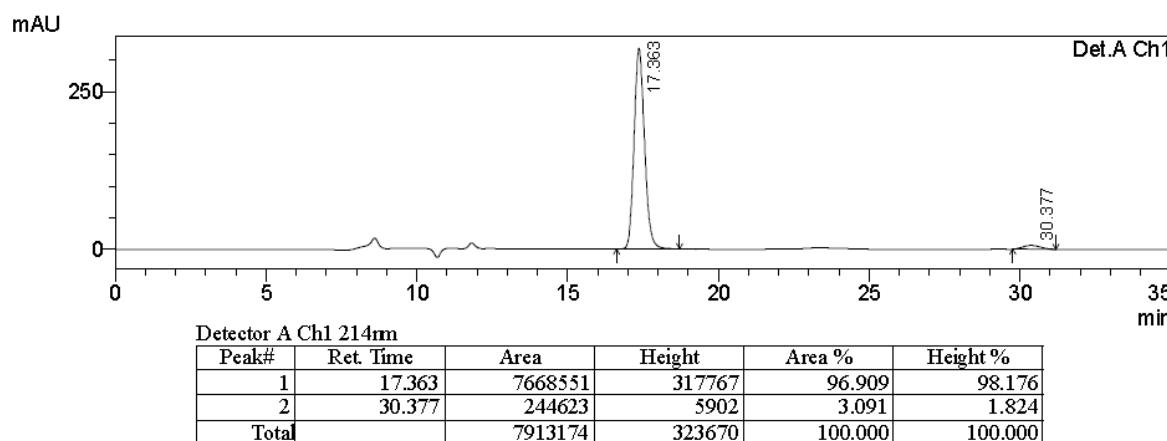
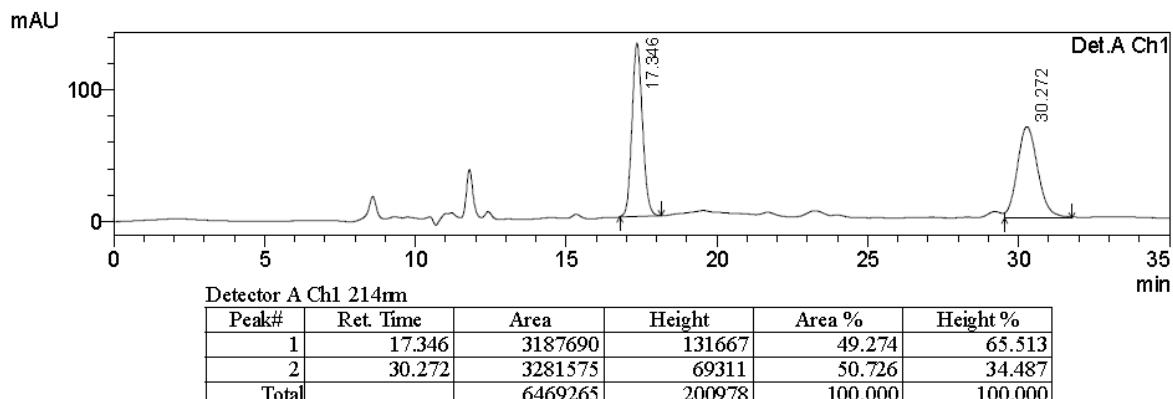
93% ee); IR (Neat): 3020, 2111, 1778, 1642, 1216  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.44-7.35 (m, 5H), 4.90 (d,  $J$  = 4.4 Hz, 1H), 4.70-4.66 (m, 1H), 2.58-2.42 (m, 2H), 2.23-2.16 (m, 1H), 2.13-2.05 (m, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  176.4, 134.5, 129.1, 128.9, 127.2, 81.4, 67.5, 28.1, 22.3; HRMS (EI) calcd for  $\text{C}_{11}\text{H}_{17}\text{BrO}_2$  [ $\text{M}]^+$ : 217.0851; found: 217.0847; HPLC (Daicel Chiralpak IB, *i*-PrOH/*n*-hexane = 25/75, 0.6 mL/min, 214 nm)  $t_1$  = 17.6 min (minor),  $t_2$  = 22.5 min (major).



### Synthesis of 5-hydroxy-6-phenylpiperidin-2-one (14)<sup>8</sup>

To a solution of azide **12** (154 mg, 0.71 mmol) in 1:1 mixture of EtOH/EtOAc (16 ml), Pd/C (100 mg, 10% on activated carbon) was added. The flask was evacuated and charged with hydrogen, and the mixture was stirred at 25 °C overnight. The mixture was filtered through a pad of Celite, and washed with copious amount of CH<sub>2</sub>Cl<sub>2</sub>. The filtrate was concentrated under reduced pressure to deliver 5-amino(phenyl)methyl-dihydrofuran-2(3H)-one as an

orange oil. The crude product was then dissolved in DBU (1.5 ml) and heated at 120 °C for 18 hours. The mixture was allowed to cool to room temperature before subjecting to silica gel column chromatography with MeOH:CH<sub>2</sub>Cl<sub>2</sub> (2:98) to deliver product **13** as a white solid. White solid, mp 176.5-177.8; [α]<sub>D</sub><sup>25</sup> -30.1 (c 1.0, MeOH, 93% ee); <sup>1</sup>H NMR (300 MHz, MeOD): δ 7.41-7.27(m, 5H), 4.46 (d, *J* = 4.8 Hz, 1H), 3.92-3.88 (m, 1H), 2.63-2.52 (m, 1H), 2.47-2.37 (m, 1H), 1.93-1.76 (m, 2H); <sup>13</sup>C NMR (125 MHz, MeOD): δ 174.8, 142.0, 129.7, 128.9, 128.0, 70.4, 64.5, 28.3, 25.8; HRMS (ESI) calcd for C<sub>11</sub>H<sub>13</sub>NO<sub>2</sub> [M – H]<sup>-</sup>: 190.0868; found: 190.0870; HPLC (Daicel Chiralpak IC, *i*-PrOH/*n*-hexane = 50/50, 0.6 mL/min, 214 nm) t<sub>1</sub>= 17.4min (major), t<sub>2</sub> = 30.4 min (minor).



## Synthesis of 2-phenyl-3-piperidinol (15)

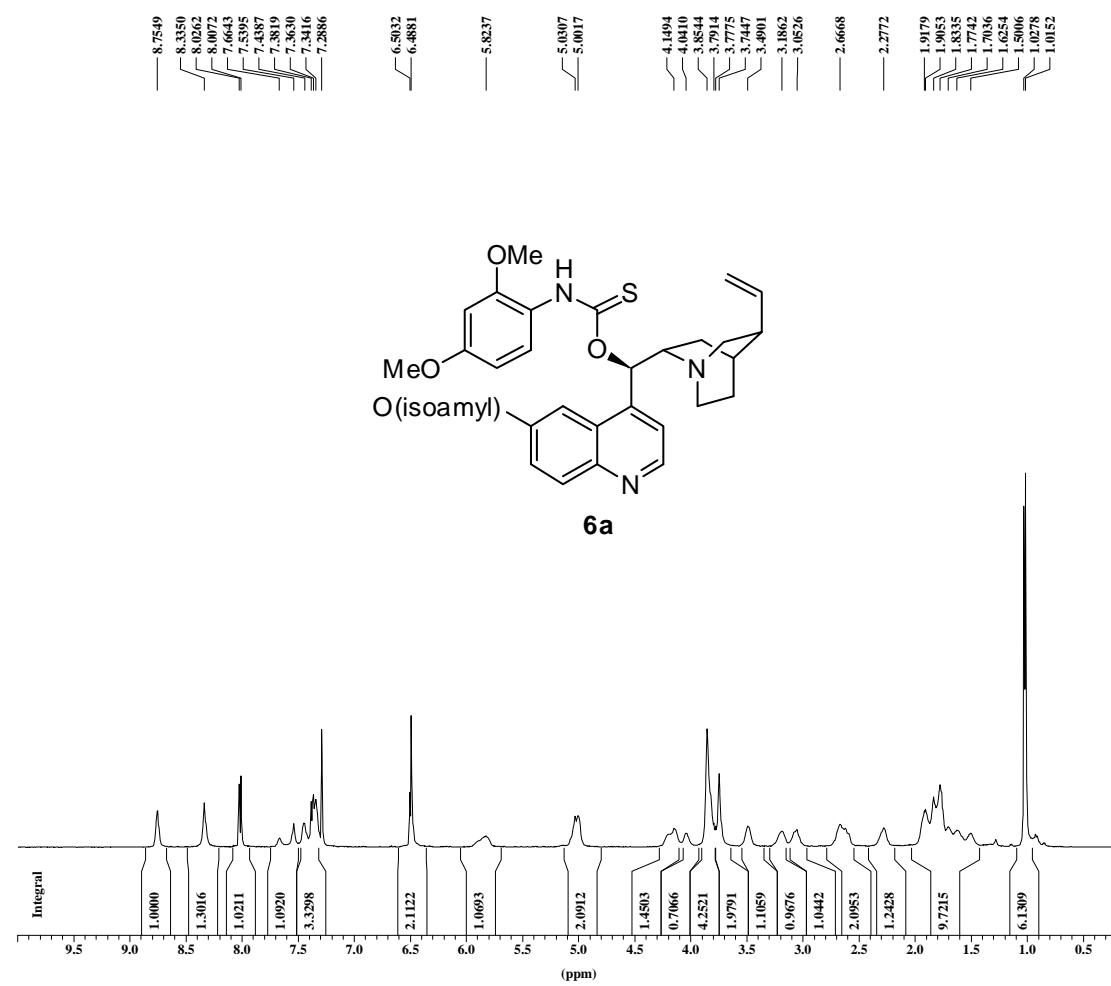
To a solution of **13** (100 mg, 0.52 mmol, 1.0 eq) in THF (3 mL) was added borane-dimethyl sulfide complex (1.30 mL, 2.6 mmol, 2 M in Et<sub>2</sub>O, 5.0 eq) under nitrogen atmosphere. The

reaction mixture was heated to reflux for 6 h. After the reaction was cooled to room temperature, MeOH (20 mL) was added cautiously to quench the excess borane-dimethyl sulfide complex. The solvent was removed *in vacuo*. The residue together with powdered K<sub>2</sub>CO<sub>3</sub> (215 mg, 1.56 mmol, 3.0 eq) were dissolved in EtOH (6 mL) and. The resulting mixture was heated to reflux for 18 h, cooled, and concentrated *in vacuo*. The residue was then dissolved in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) and H<sub>2</sub>O (15 mL). The layers were separated and further extraction with CH<sub>2</sub>Cl<sub>2</sub> (2 x 5 mL) were performed. The combined organic layer was then washed with brine (15 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo* to yield product **14** was a white solid. White solid, mp 154.4-155.1; [α]<sub>D</sub><sup>25</sup> -21.4 (c 1.0, MeOH); <sup>1</sup>H NMR (300 MHz, MeOD): δ 7.42-7.26 (m, 5H), 3.63-3.58 (m, 1H), 3.32-3.31 (d, 2H), 3.04-3.01 (d, 1H), 2.68-2.63 (m, 1H), 2.16-2.14 (d, 1H), 1.81-1.79 (d, 1H), 1.75-1.67 (m, 1H), 1.52-1.44 (m, 1H); <sup>13</sup>C NMR (125 MHz, MeOD): δ 142.9, 129.4, 129.3, 128.6, 72.7, 70.0, 35.4, 26.5; HRMS (ESI) calcd for C<sub>11</sub>H<sub>13</sub>NO<sub>2</sub> [M + H]<sup>+</sup>: 178.1232; found: 178.1223.

## F. References

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3. L. Zhou, J. Chen, C. K. Tan and Y.-Y. Yeung, *J. Am. Chem. Soc.* 2011, **113**, 9164.
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1H AMX500 ck-v-cat 6a



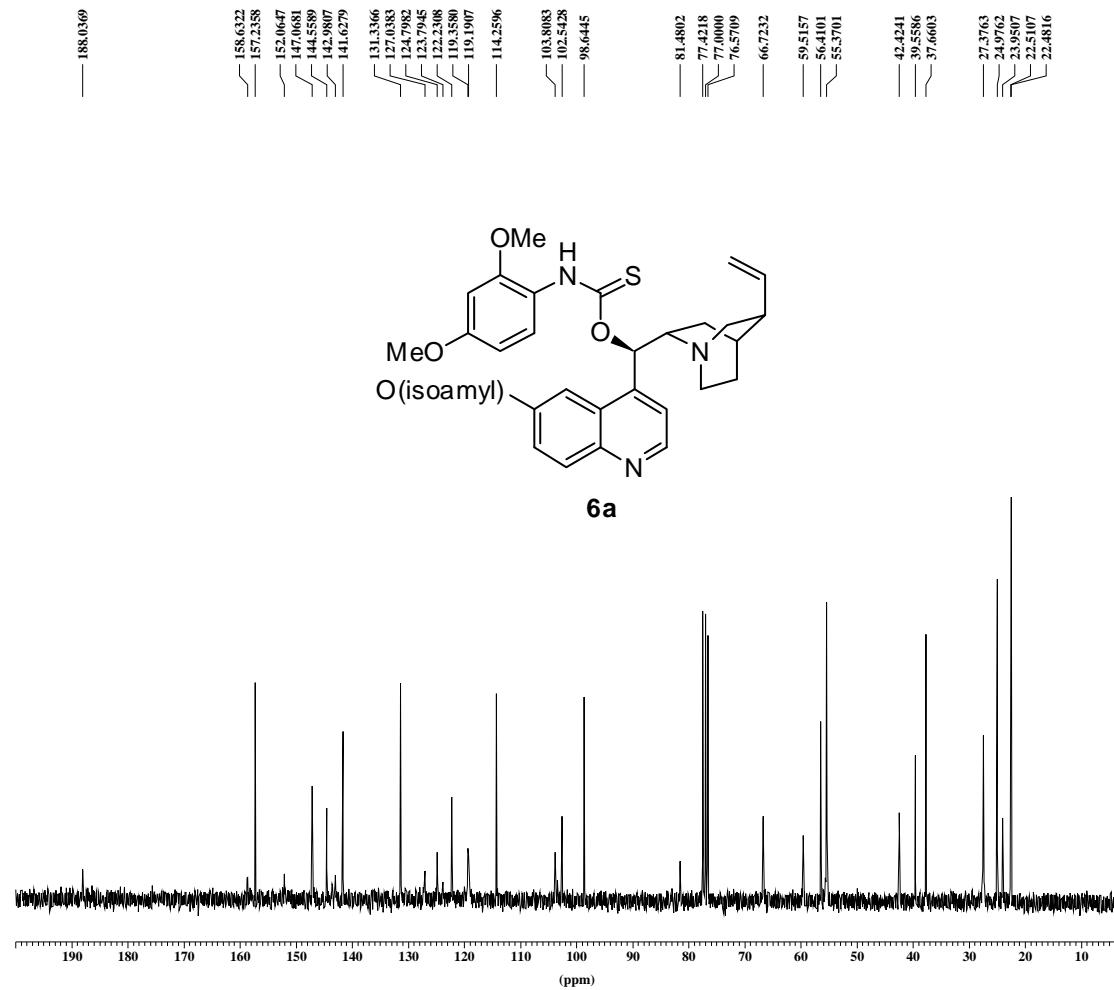
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PROCNO : 1  
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O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 297.9 K

\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300000 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

13C Standard AC300 ck-v-157 cat



\*\*\* Current Data Parameters \*\*\*

NAME : fe11ck

EXPNO : 3

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 207

NUCLEUS : off

O1 : 7924.11 Hz

PULPROG : zgpg30

SFO1 : 75.4756731 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 238.2968 ppm

TD : 32768

TE : 297.1 K

\*\*\* Processing Parameters \*\*\*

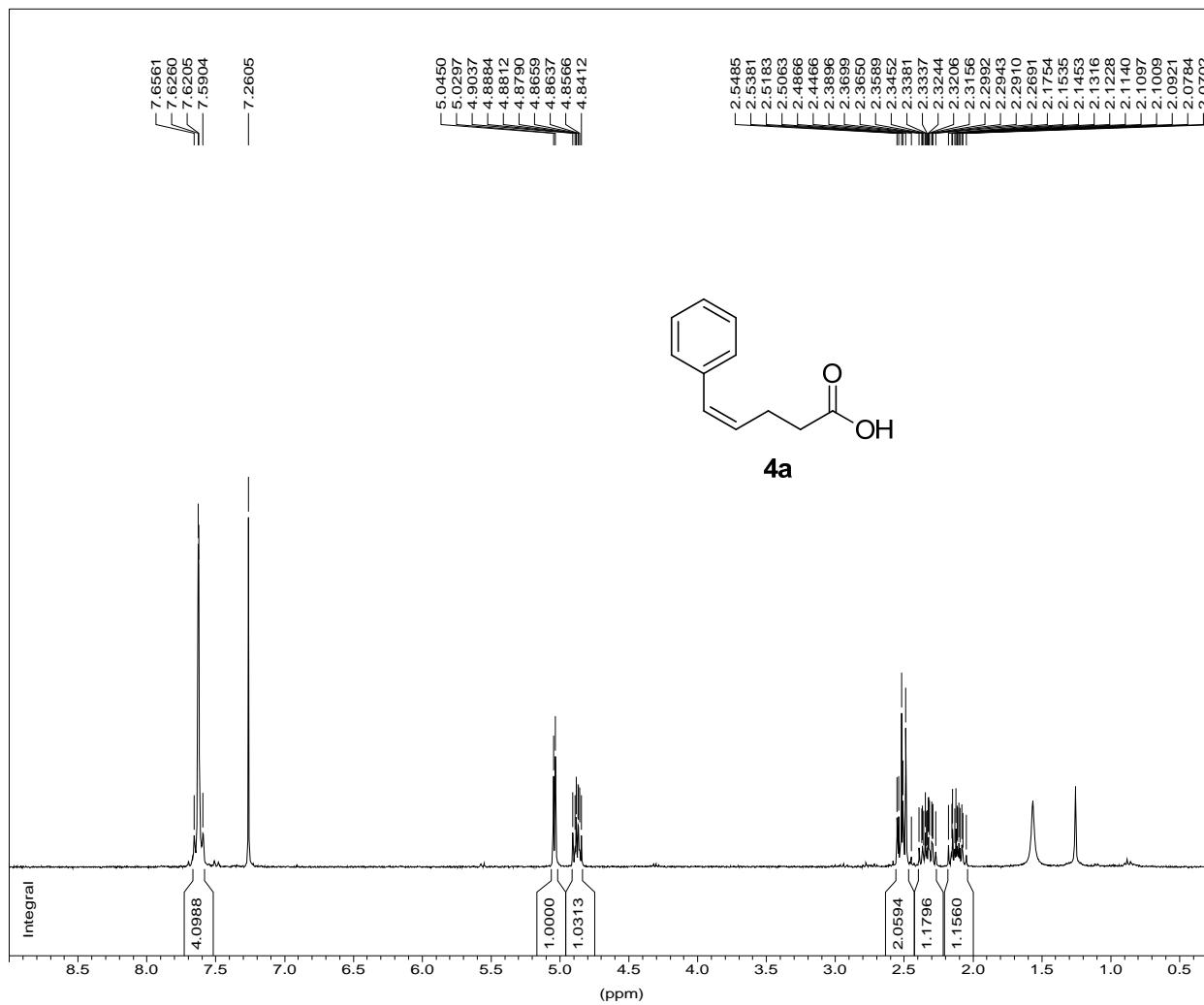
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\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>1</sup>H normal range AC300 cc-112-B



\*\*\* Current Data Parameters \*\*\*

NAME : ja05lcc

EXPNO : 4

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

BF1 : 300.1300000 MHz

LOCNUC : 2H

NS : 8

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

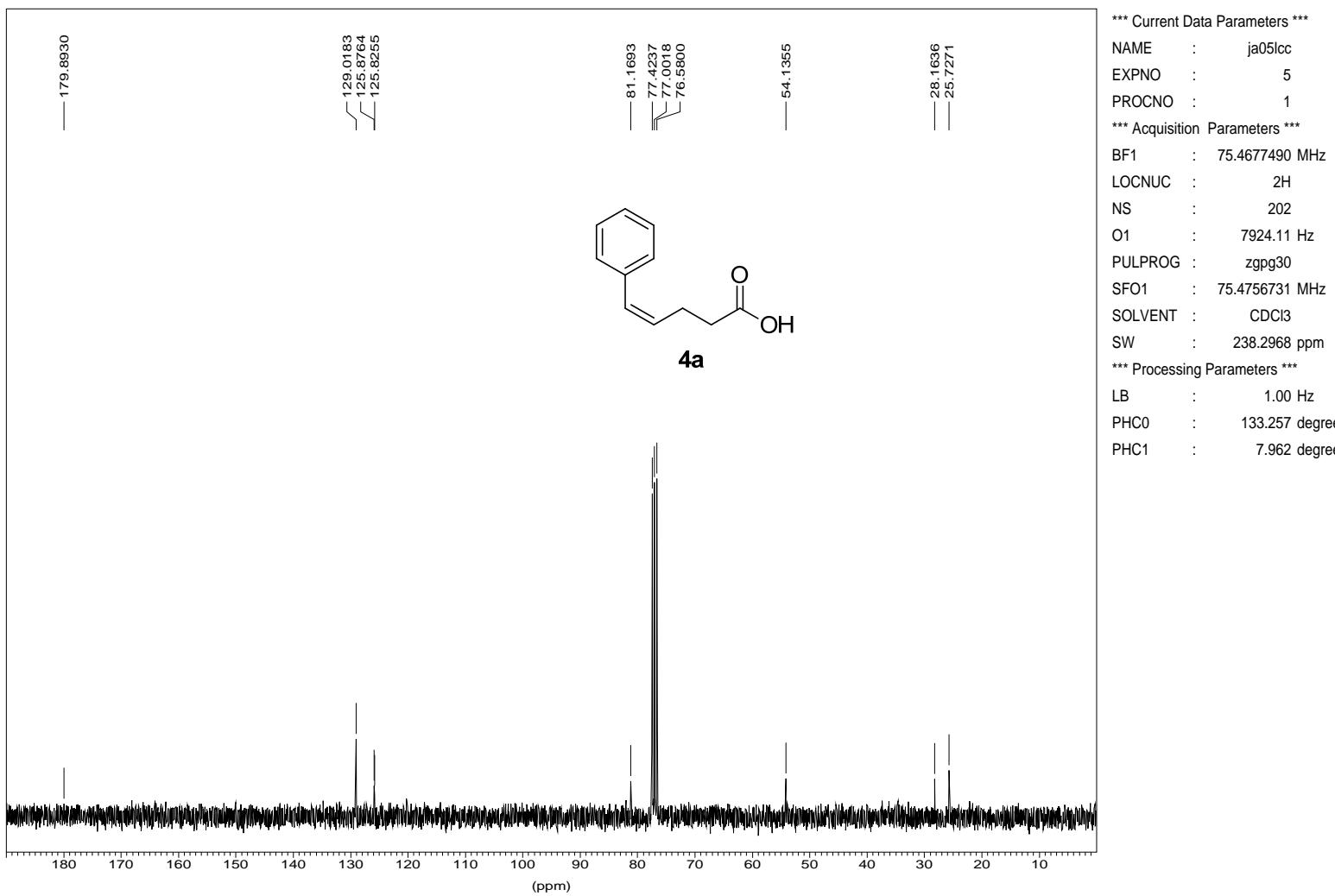
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz

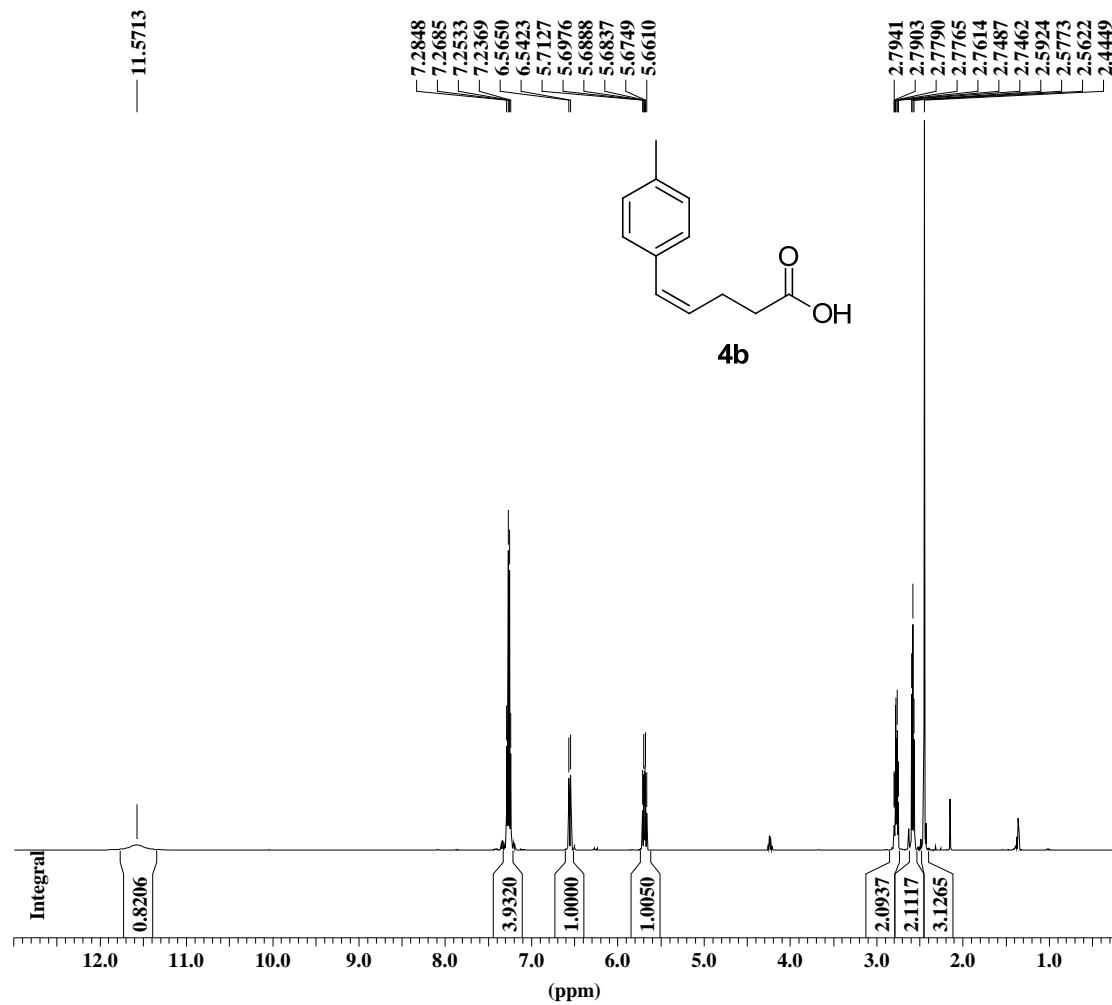
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PHC1 : 0.332 degree

13C Standard AC300 cc-112-B



<sup>1</sup>H AMX500 cc-i-33a



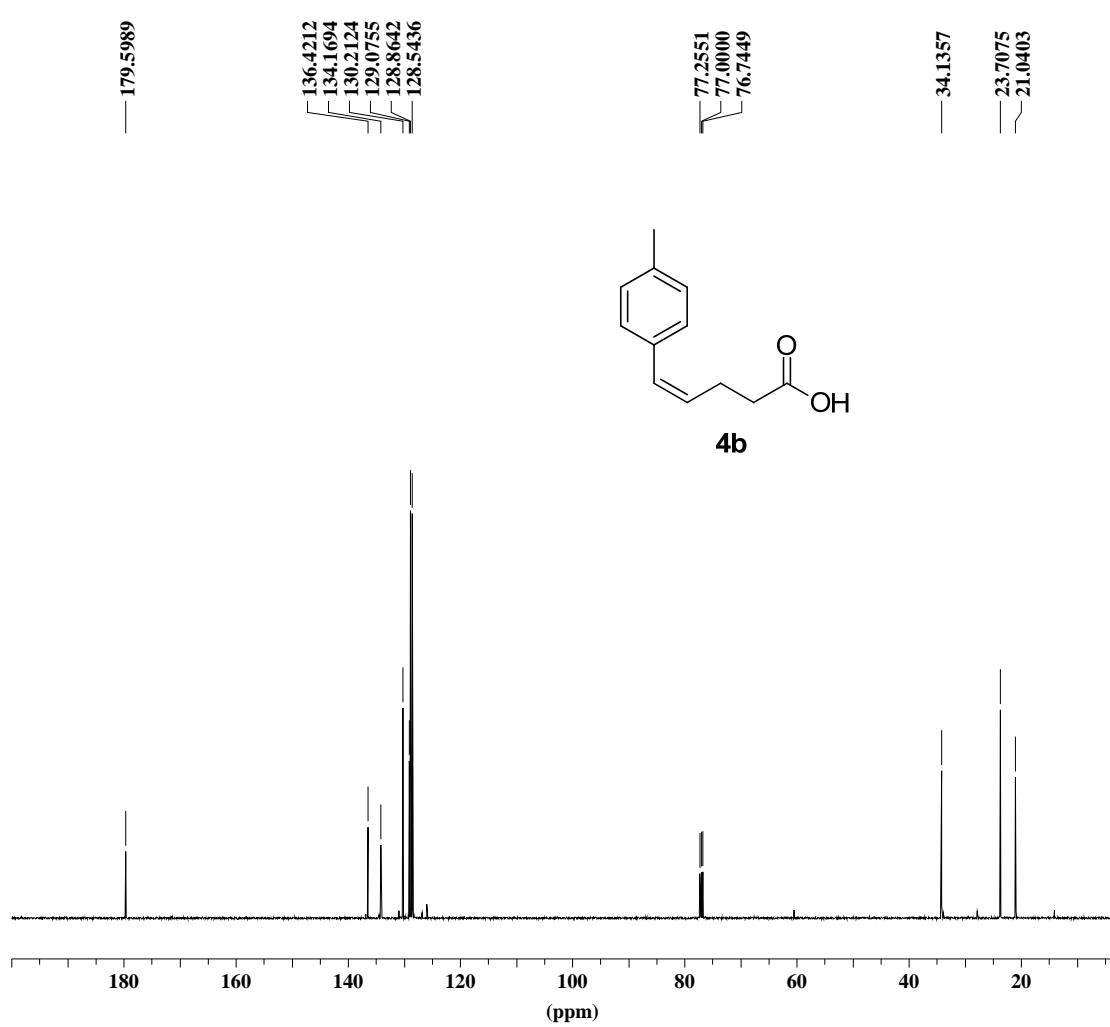
\*\*\* Current Data Parameters \*\*\*

NAME : ck0820  
EXPNO : 1  
PROCNO : 1  
LOCMNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.130085 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 300.0 K

\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300000 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

13C AMX500 cc-i-33a



\*\*\* Current Data Parameters \*\*\*

NAME : ck0820  
EXPNO : 2  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 101  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7675 ppm  
TD : 65536  
TE : 300.0 K

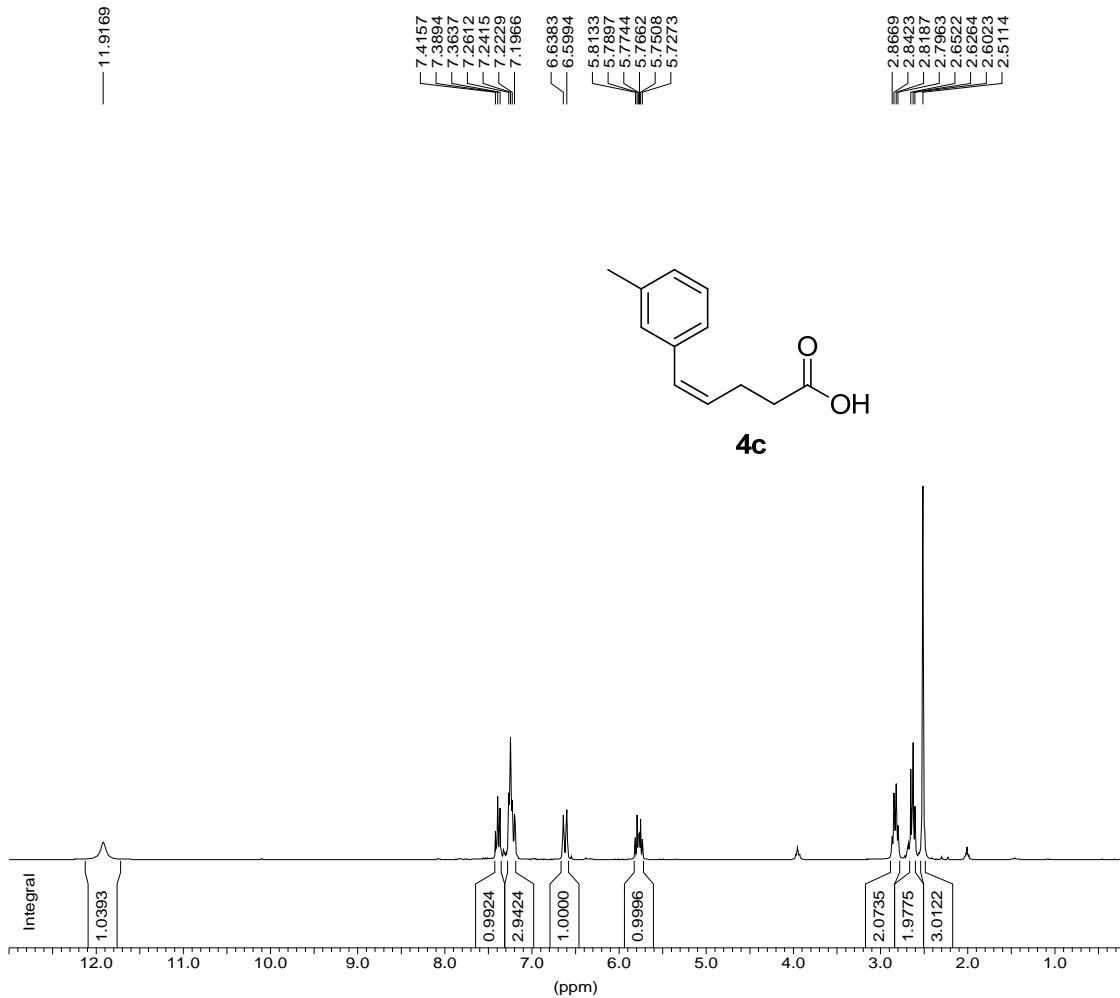
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\*\*\* 1D NMR Plot Parameters \*\*\*

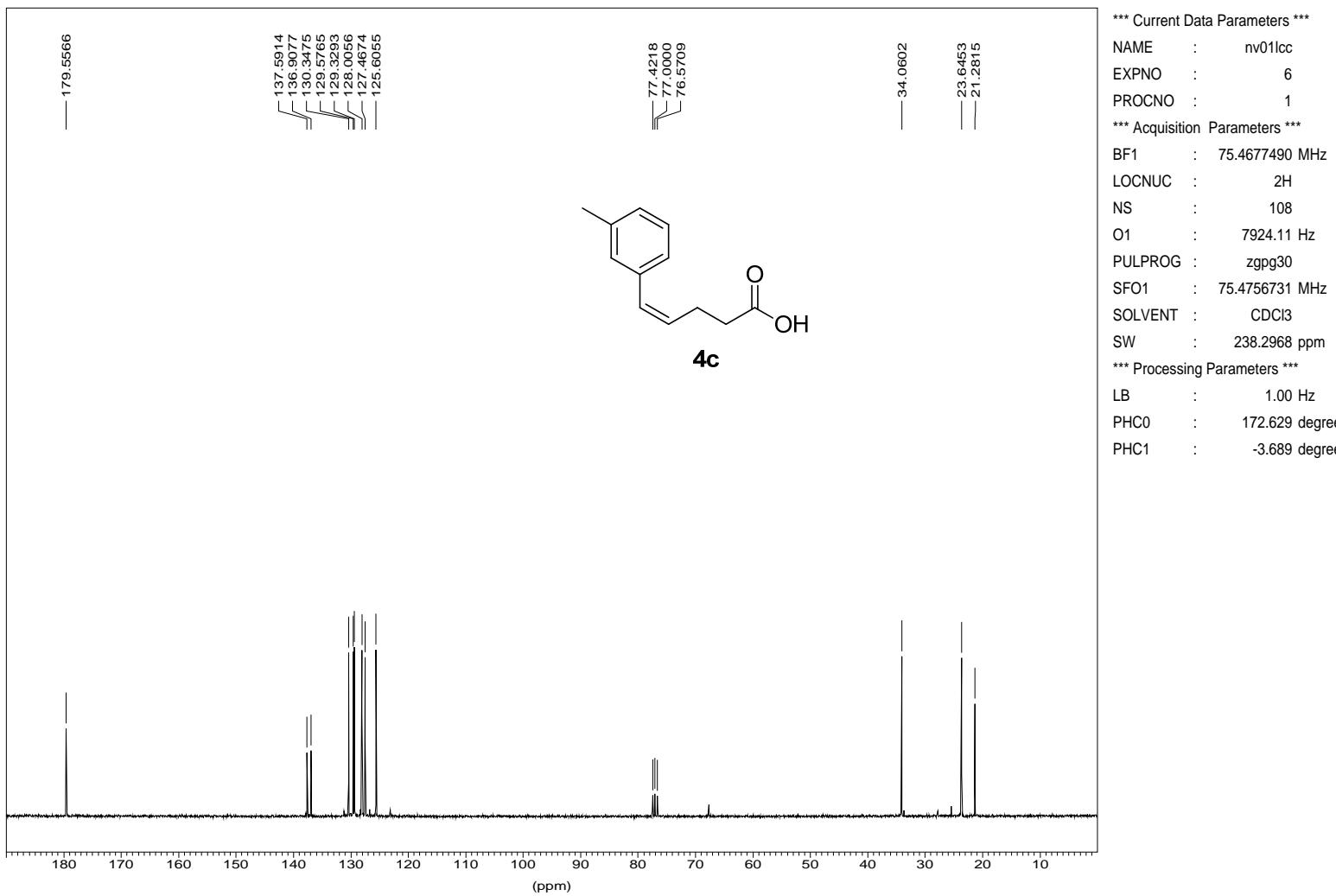
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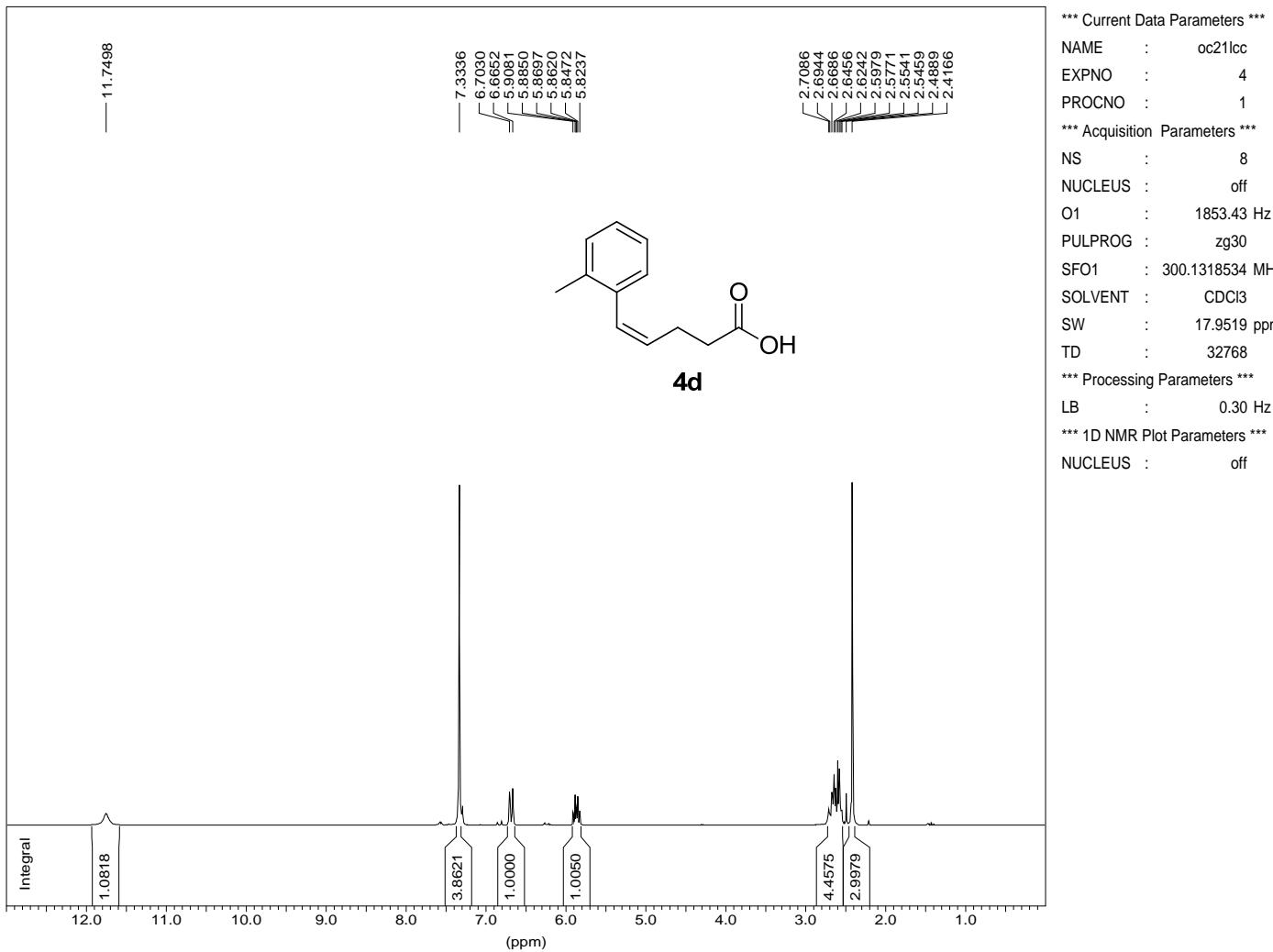


\*\*\* Current Data Parameters \*\*\*  
NAME : nv01lc  
EXPNO : 5  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

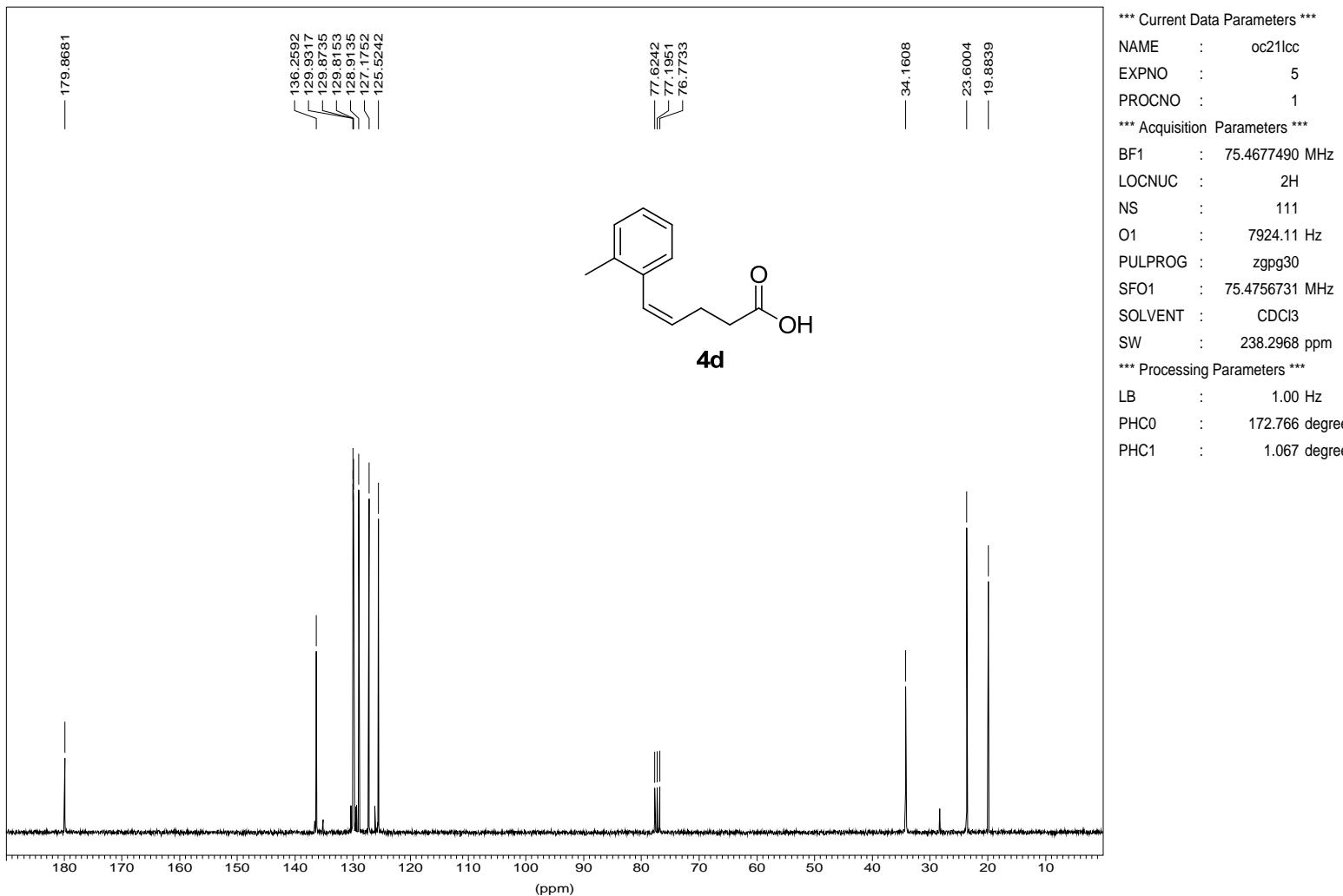
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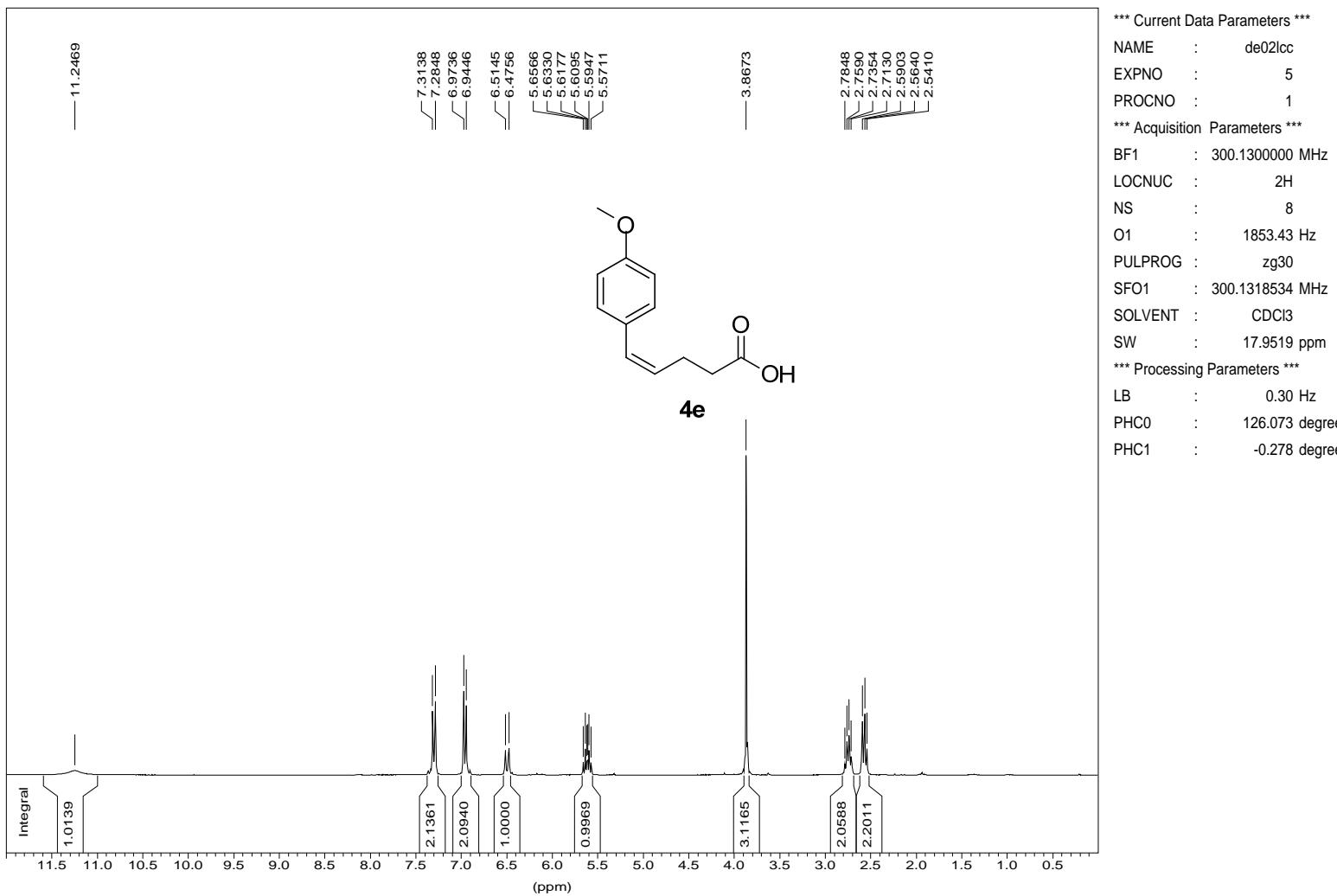
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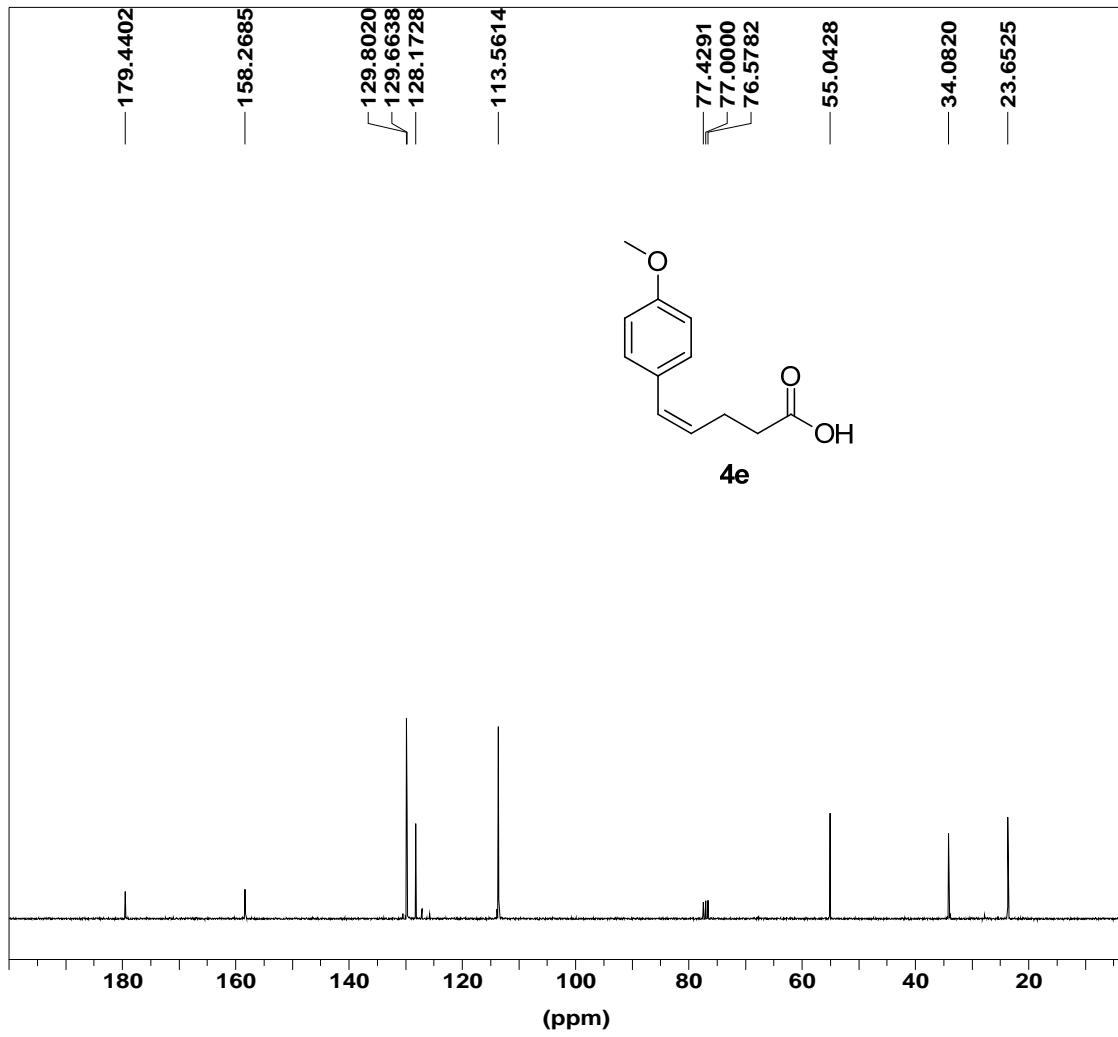
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<sup>1</sup>H normal range AC300 cc-i-103-C

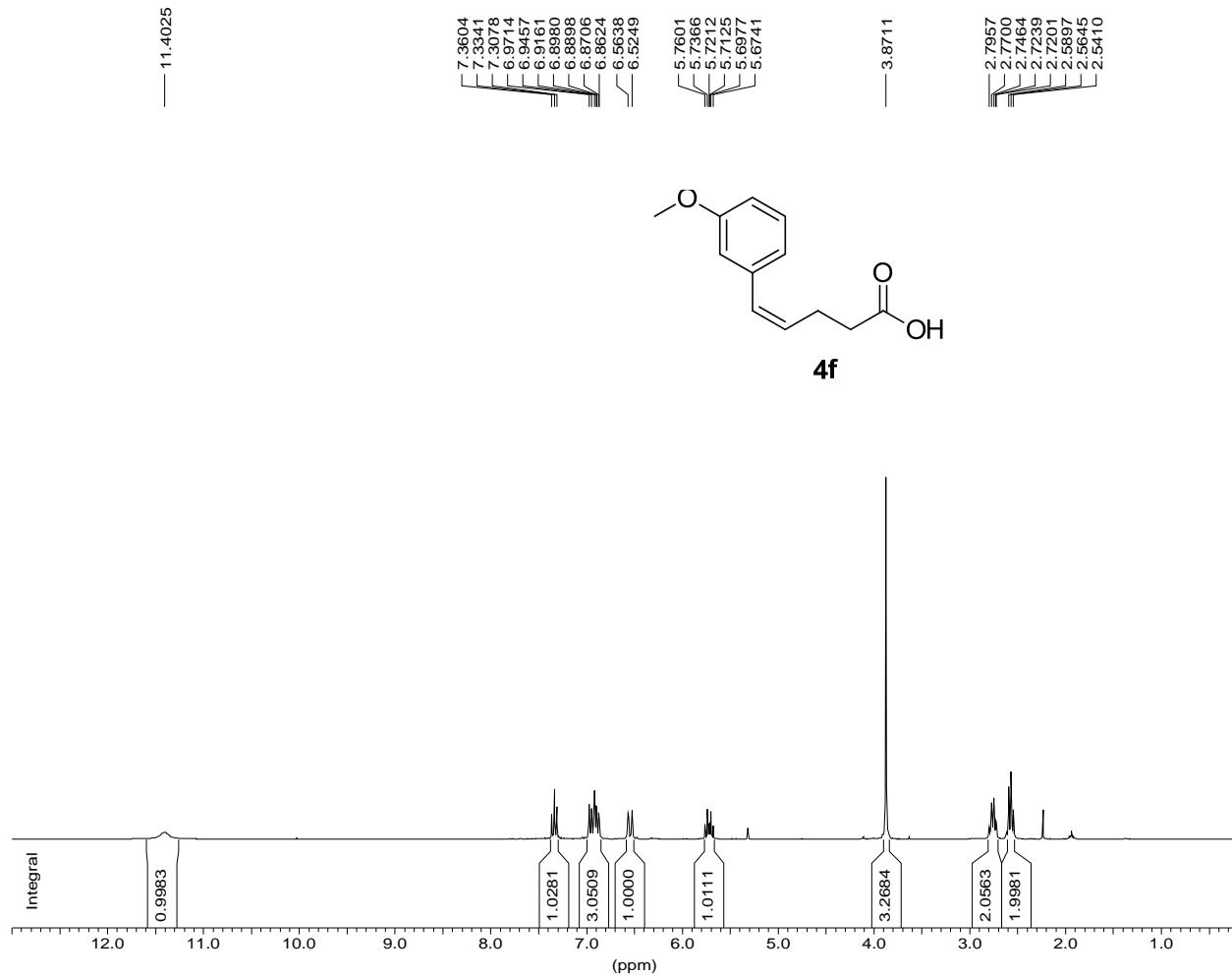


<sup>13</sup>C Standard AC300 cc-i-103-C



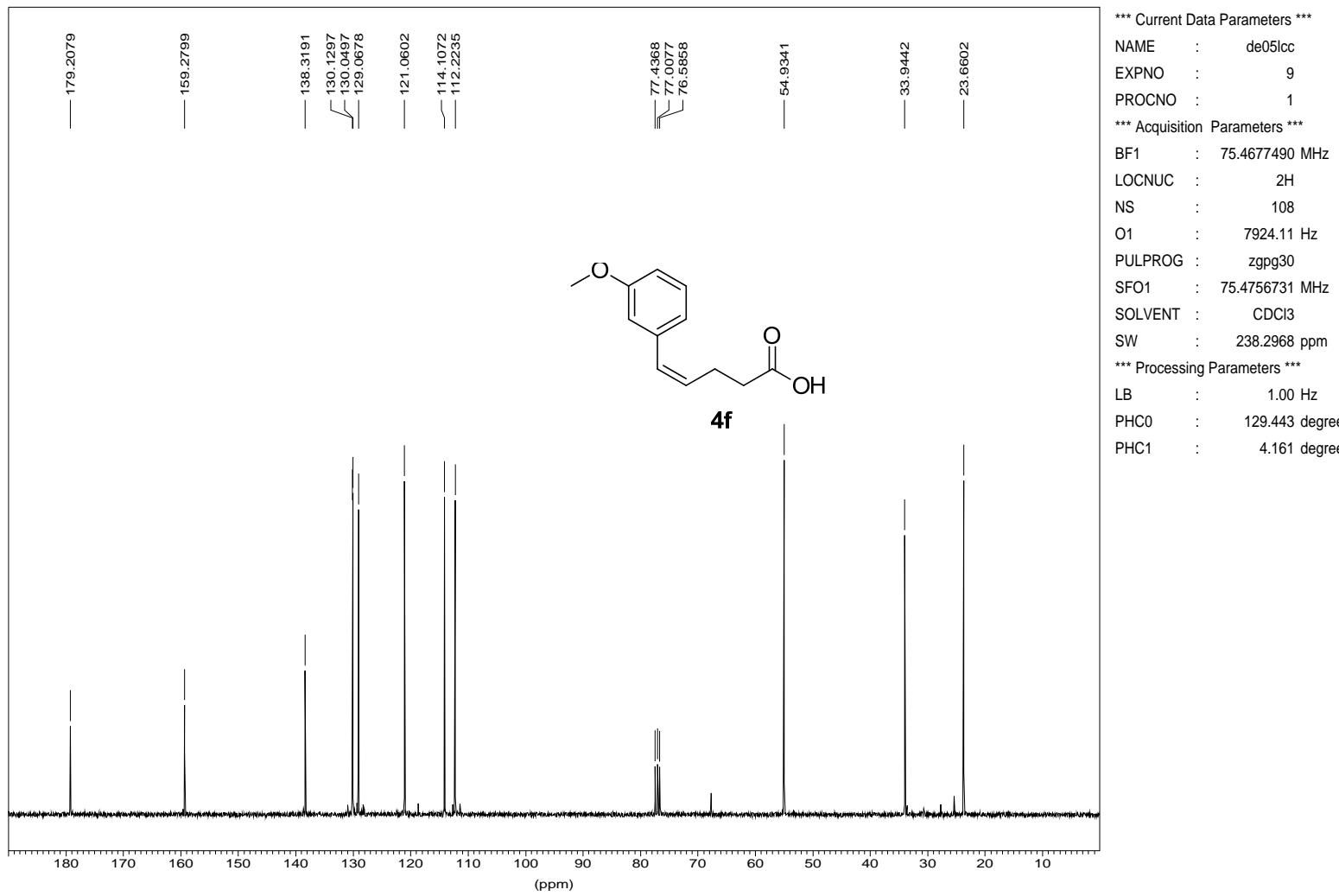
\*\*\* Current Data Parameters \*\*\*  
NAME : de02lcc  
EXPNO : 6  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
NS : 108  
O1 : 7924.11 Hz  
PULPROG : zgpg30  
SFO1 : 75.4756731 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.2968 ppm  
TD : 32768  
\*\*\* Processing Parameters \*\*\*  
LB : 1.00 Hz

1H normal range AC300 cc-090-B

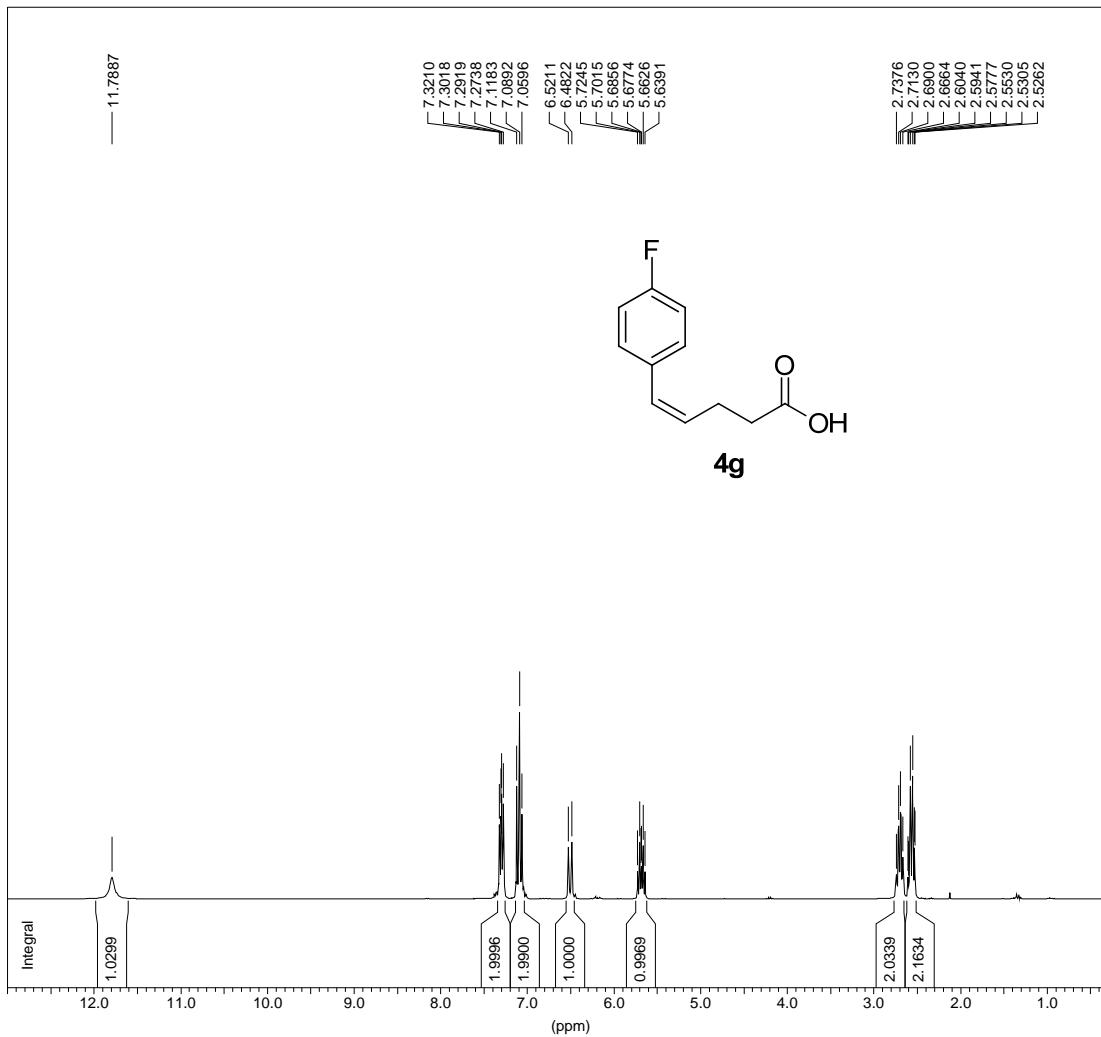


\*\*\* Current Data Parameters \*\*\*  
NAME : de05icc  
EXPNO : 8  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-i-090-B



1H normal range AC300 cc-i-073-B



\*\*\* Current Data Parameters \*\*\*

NAME : oc17cc  
EXPNO : 2  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
INSTRUM : spect  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
TE : 298.3 K

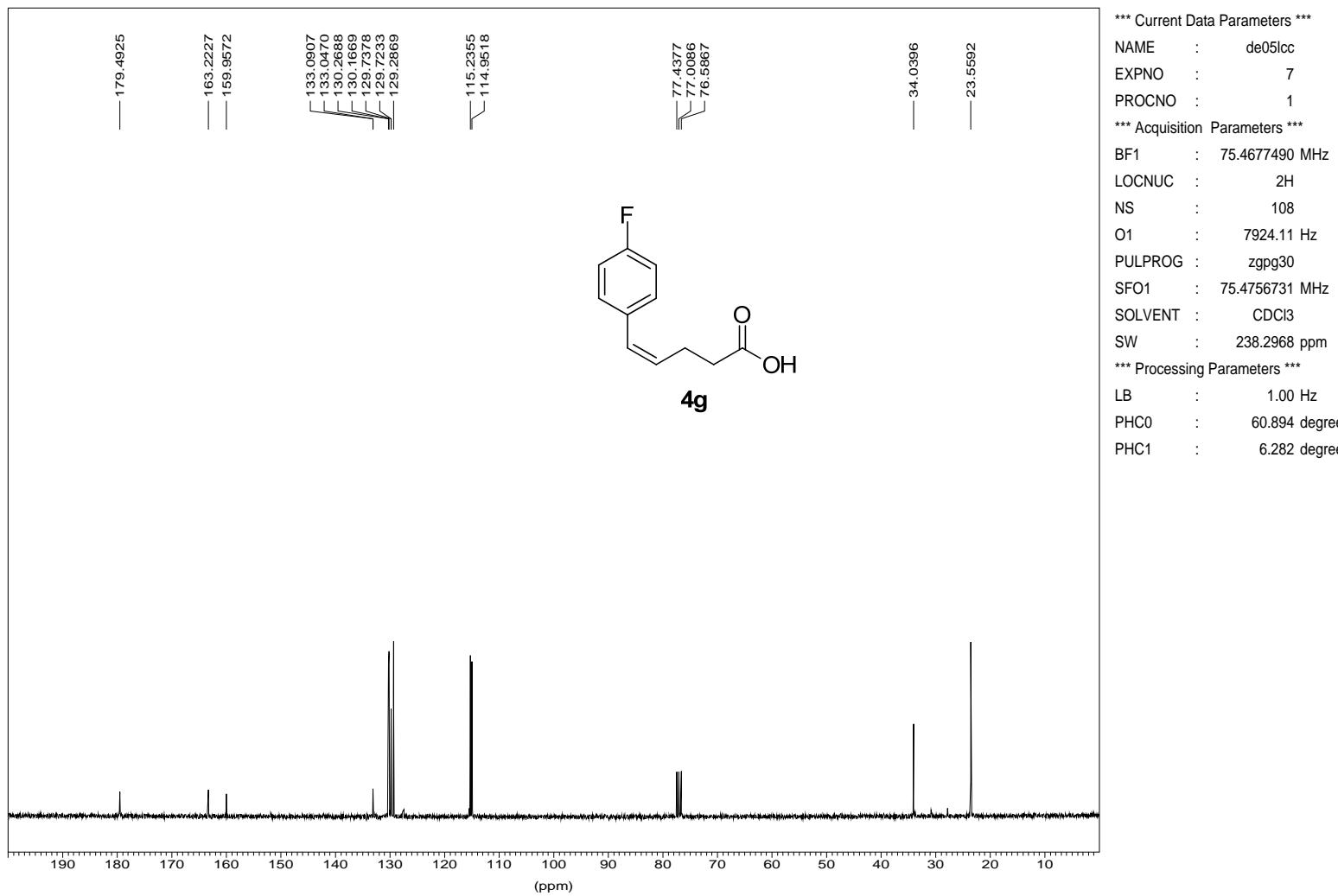
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LB : 0.30 Hz

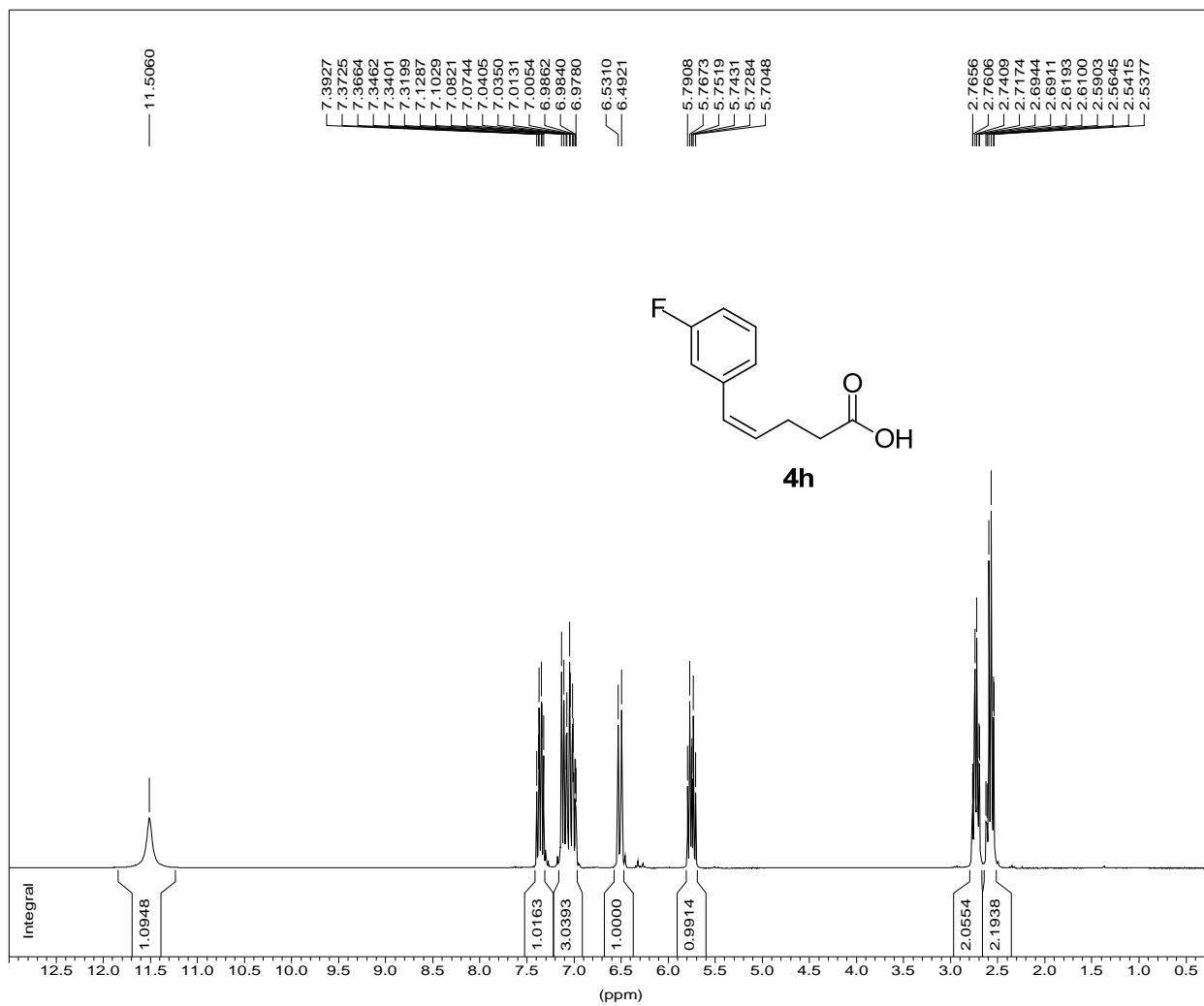
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

13C Standard AC300 cc-039-A



<sup>1</sup>H normal range AC300 cc-i-103-A



\*\*\* Current Data Parameters \*\*\*

NAME : de02lc

EXPNO : 1

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

BF1 : 300.1300000 MHz

LOCNUC : 2H

NS : 8

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

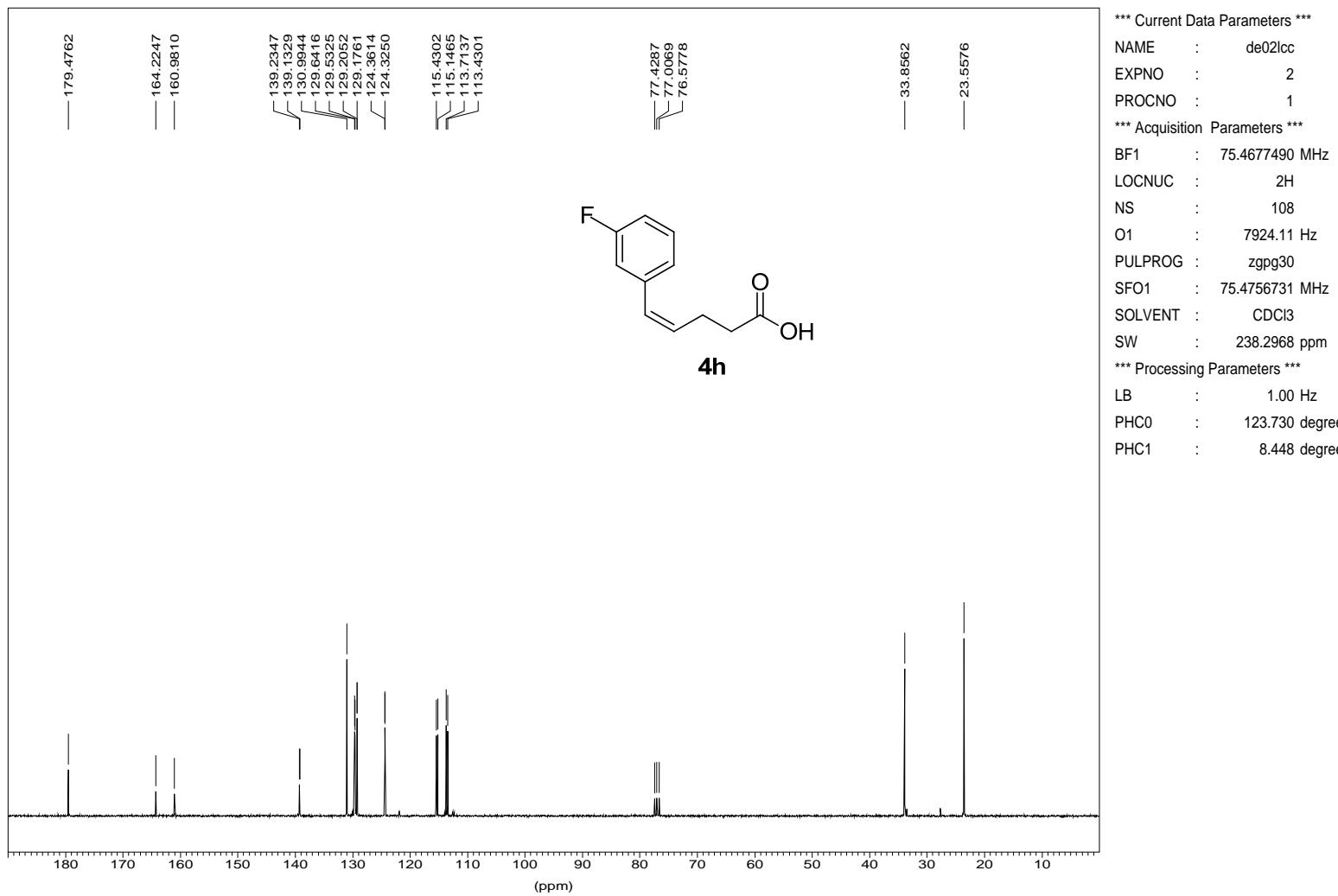
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz

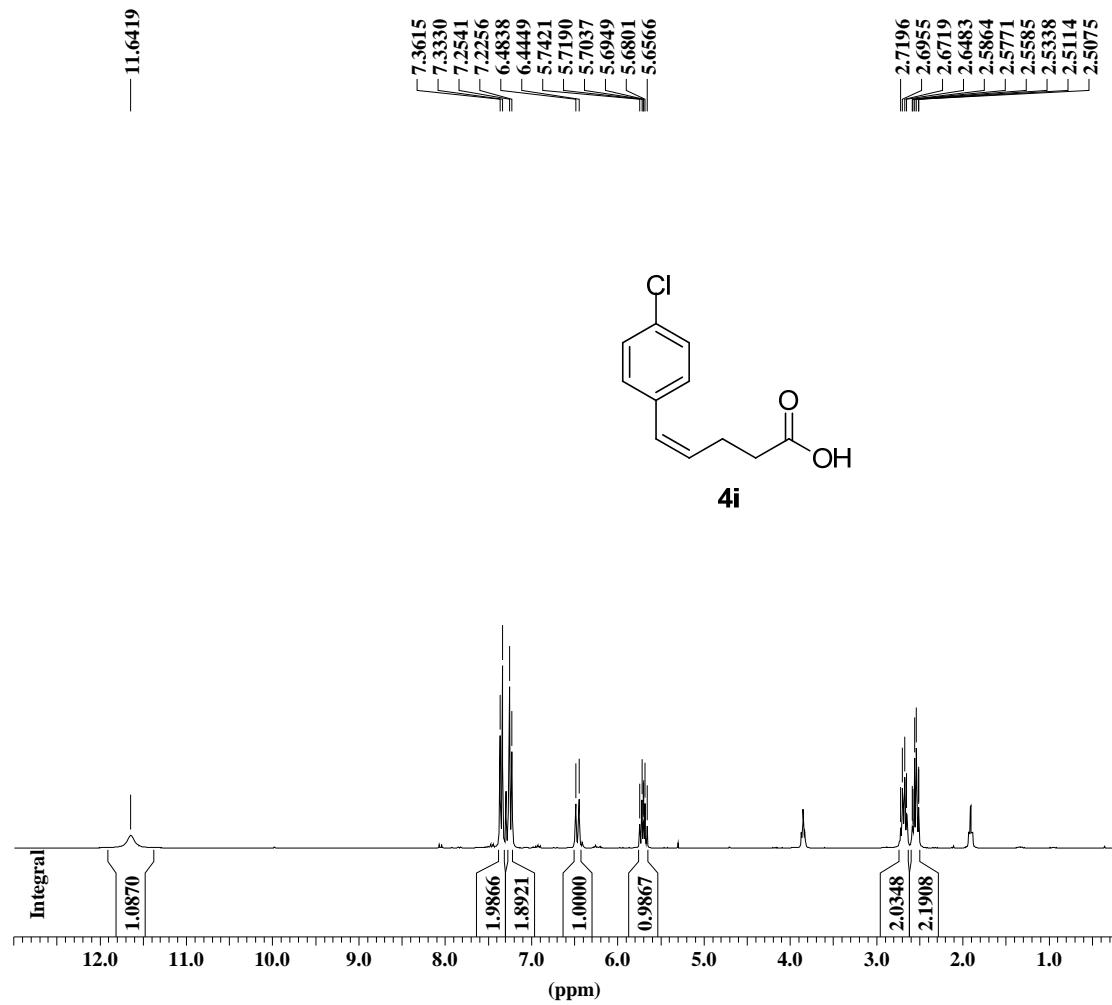
PHC0 : 123.186 degree

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<sup>13</sup>C Standard AC300 cc-i-103-A



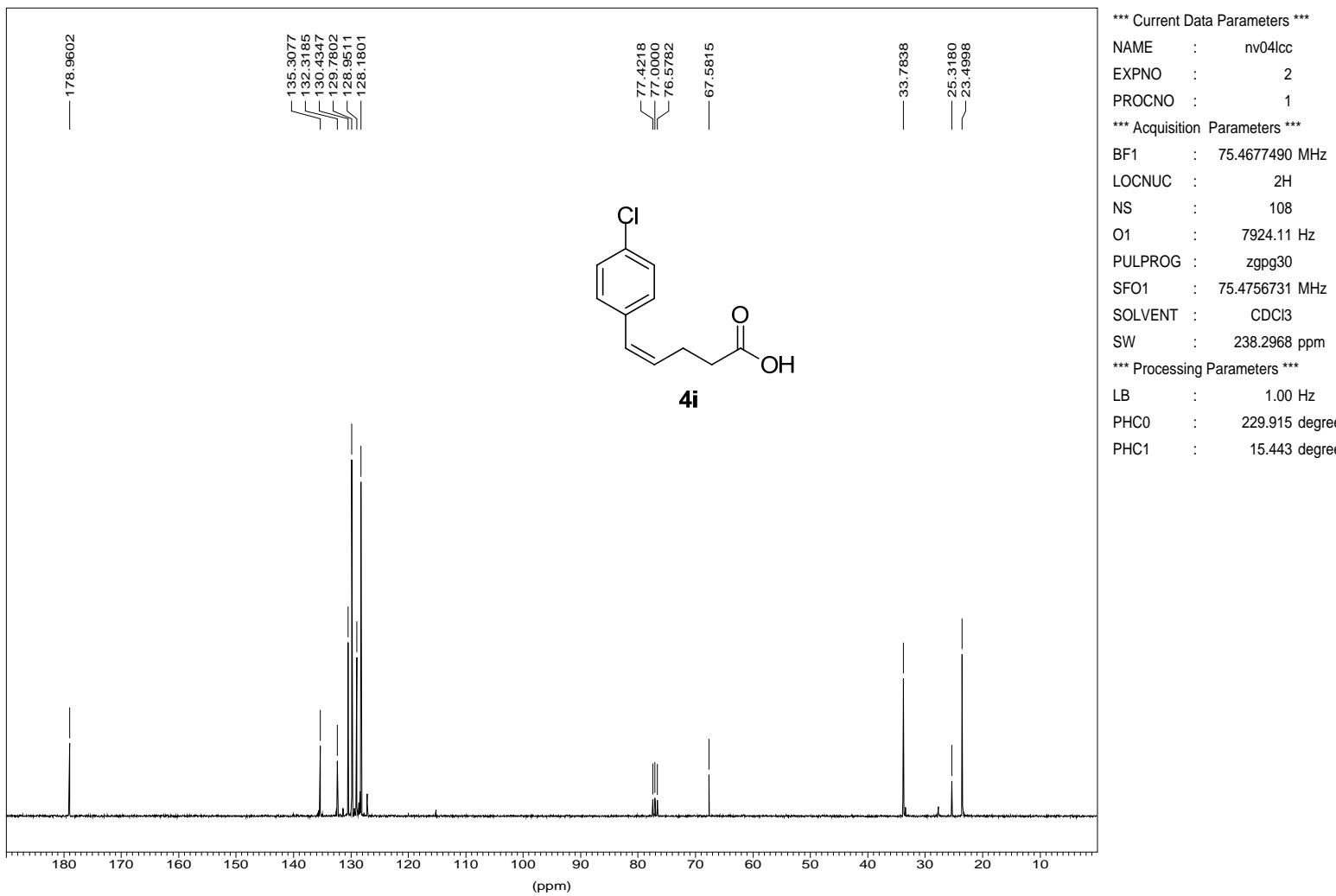
1H normal range AC300 cc-i-087-A



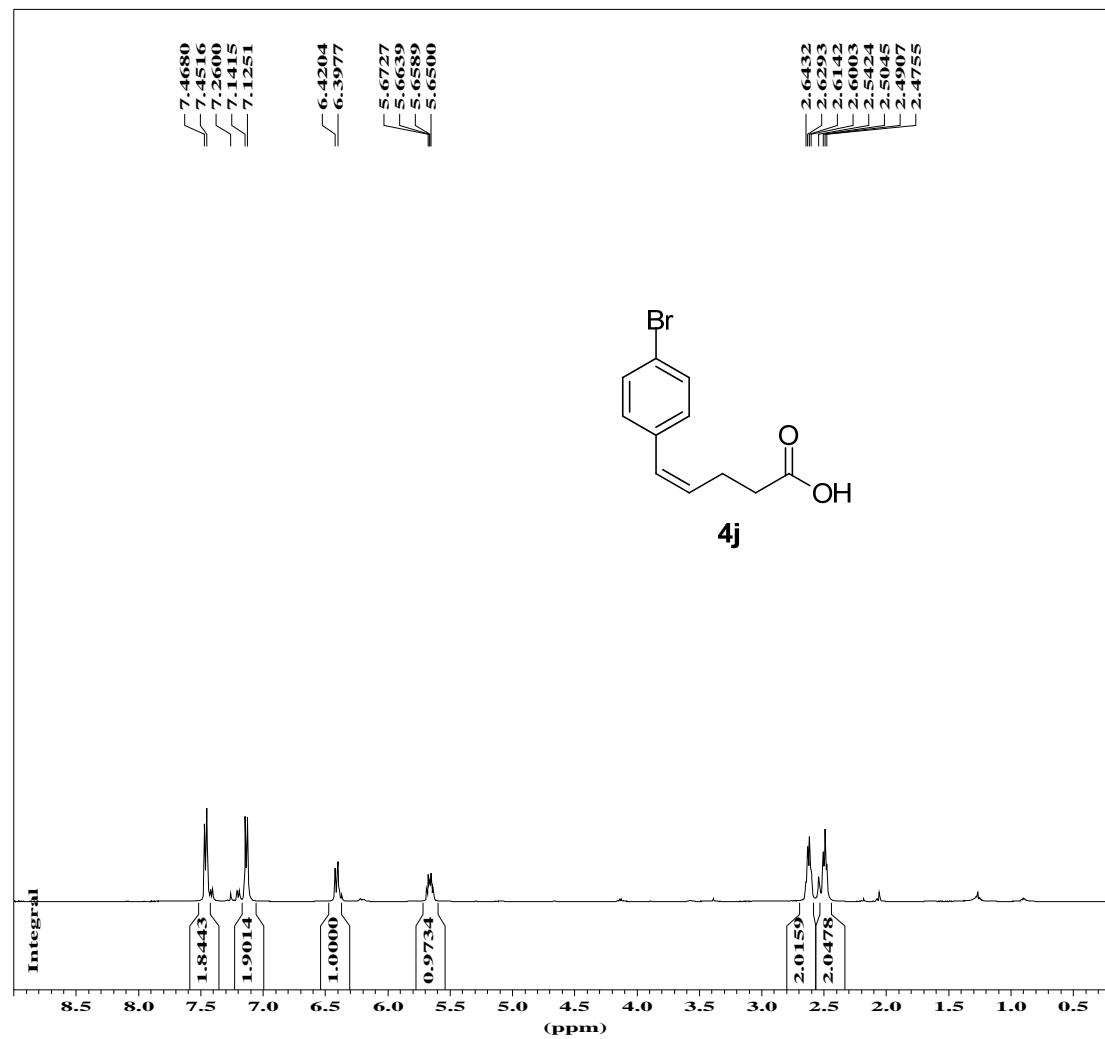
\*\*\* Current Data Parameters \*\*\*

NAME : nv04lcc  
EXPNO : 1  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
TE : 297.1 K  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
SF : 300.1300000 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-i-087-A



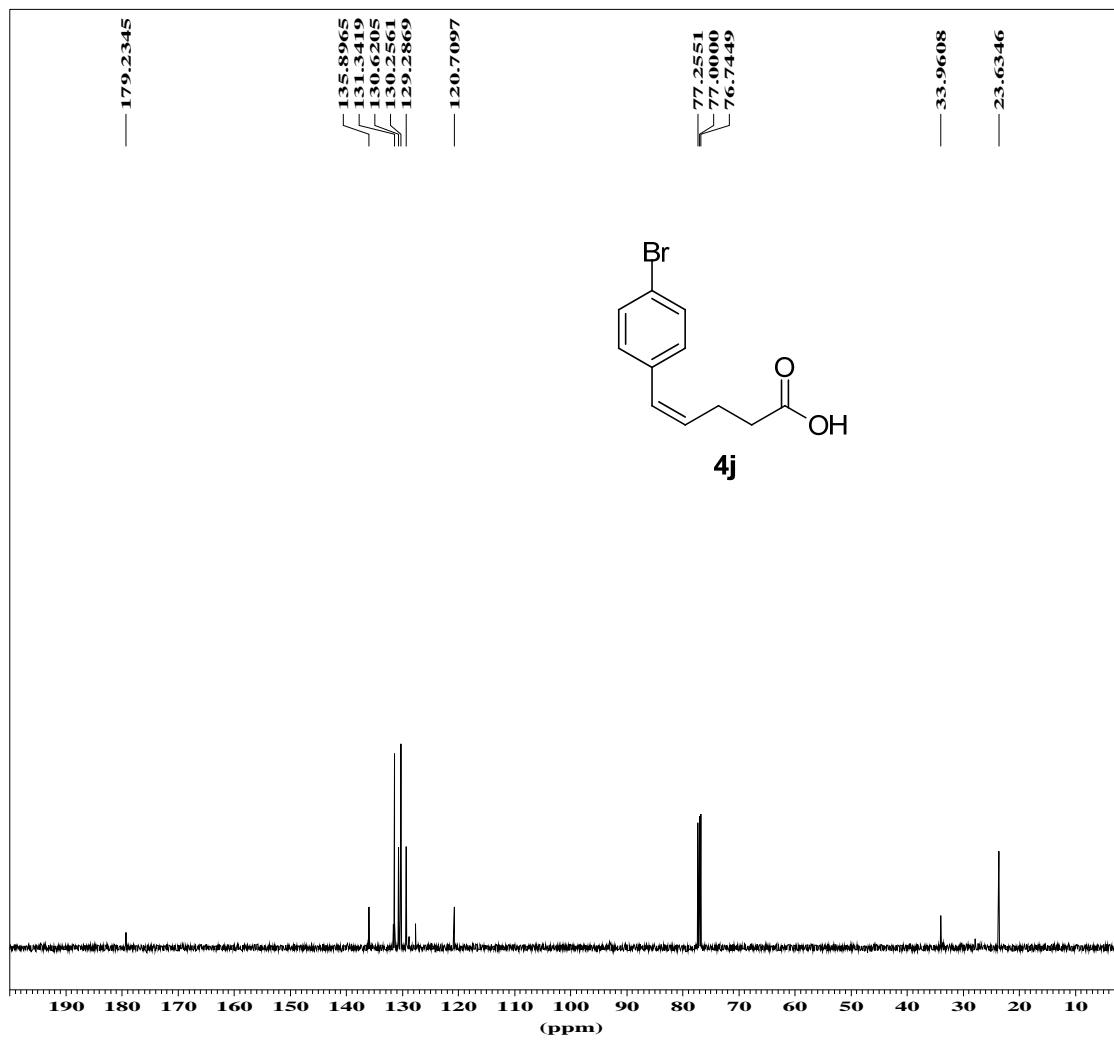
1H AMX500 4-Br acid



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214  
EXPNO : 5  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 298.0 K  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
SF : 500.1300140 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

13C AMX500 4-Br acid



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214  
EXPNO : 6  
PROCNO : 1  
LOCMNUC : 2H  
NS : 30  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7675 ppm  
TD : 65536  
TE : 298.3 K

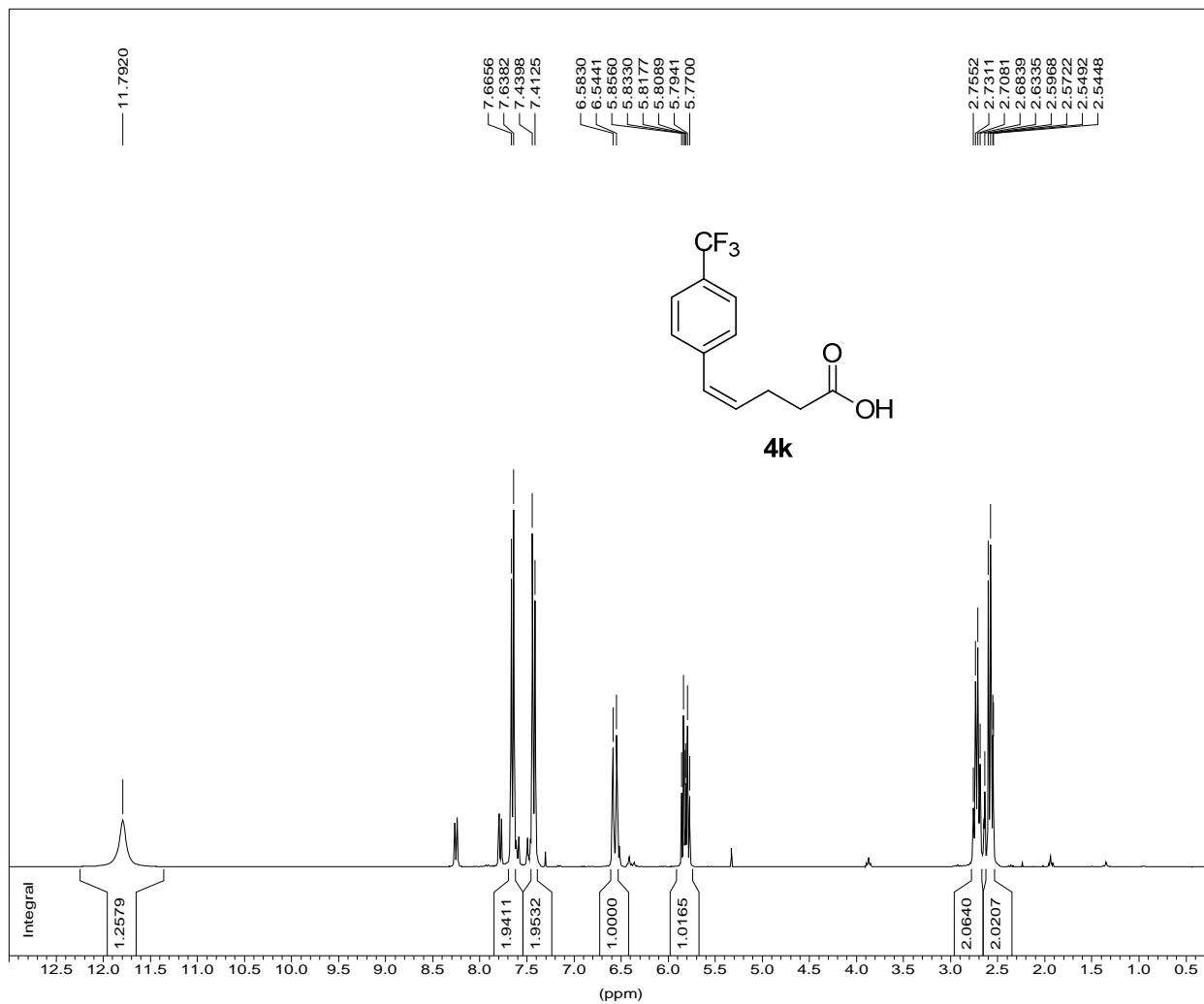
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 125.7577970 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

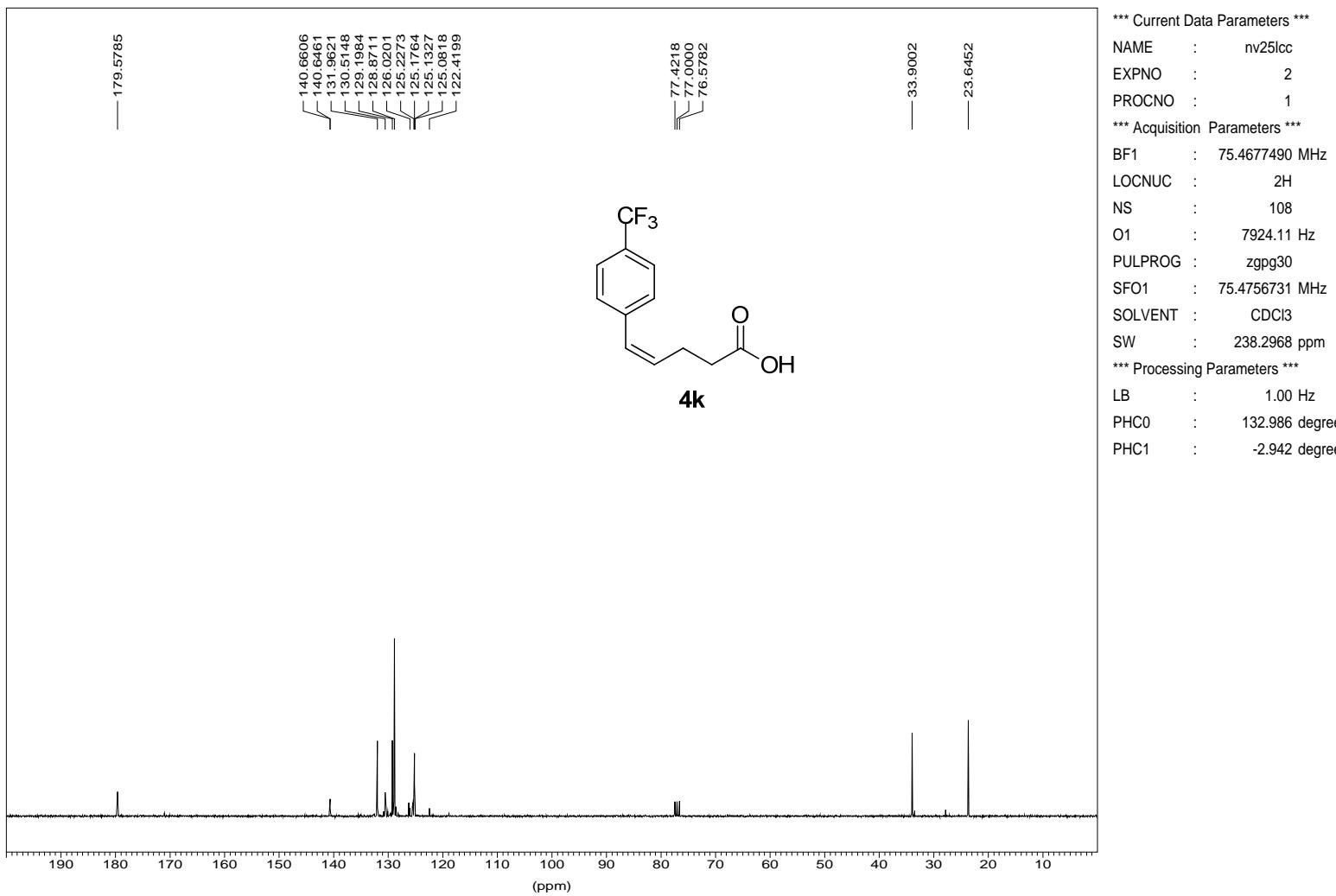
<sup>1</sup>H normal range AC300 cc-093-A



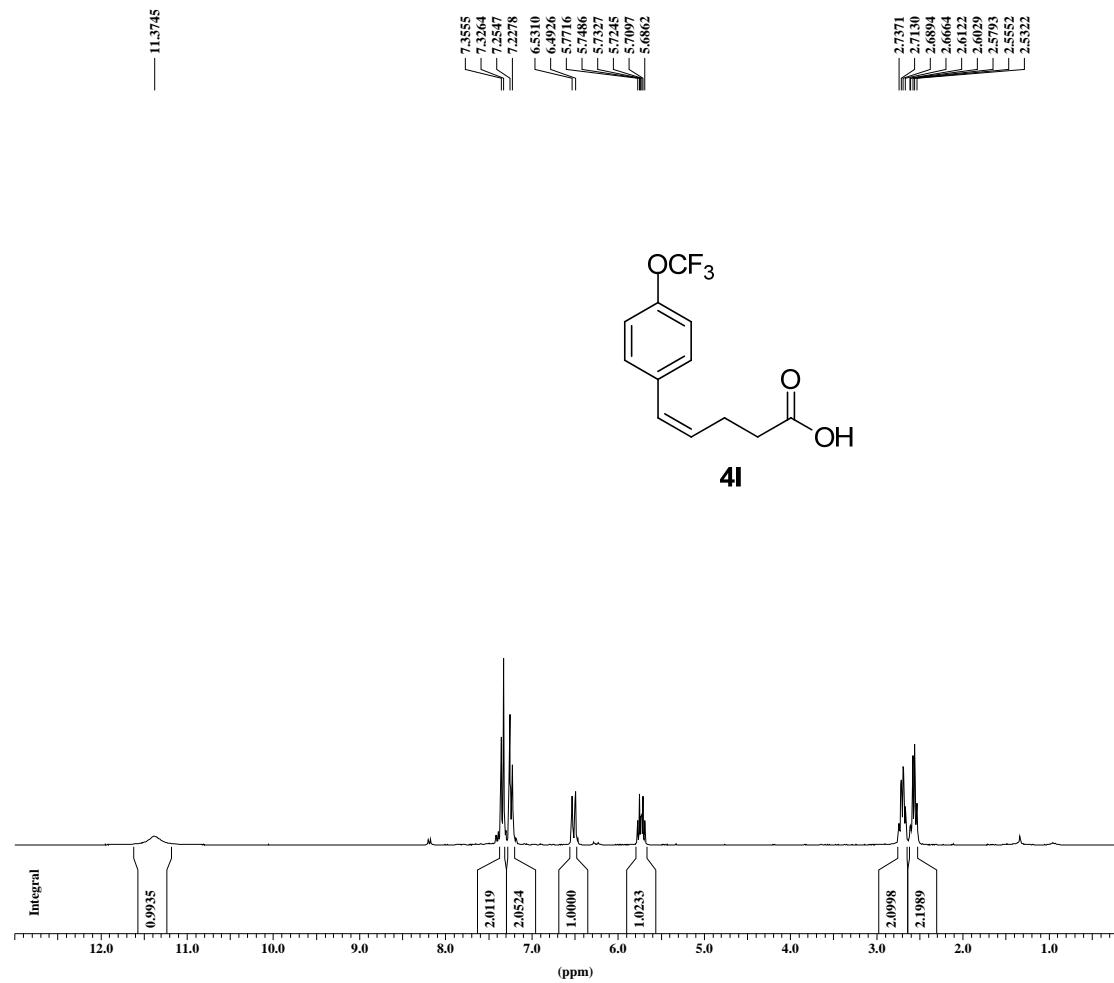
\*\*\* Current Data Parameters \*\*\*

NAME : nv25lc  
EXPNO : 1  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
BF1 : 300.1300000 MHz  
LOCNUC : 2H  
NS : 8  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
PHC0 : 126.640 degree  
PHC1 : 0.468 degree

<sup>13</sup>C Standard AC300 cc-093-A



1H normal range AC300 cc-079-A



\*\*\* Current Data Parameters \*\*\*

NAME : oc25lcc  
EXPNO : 1  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
TE : 297.4 K

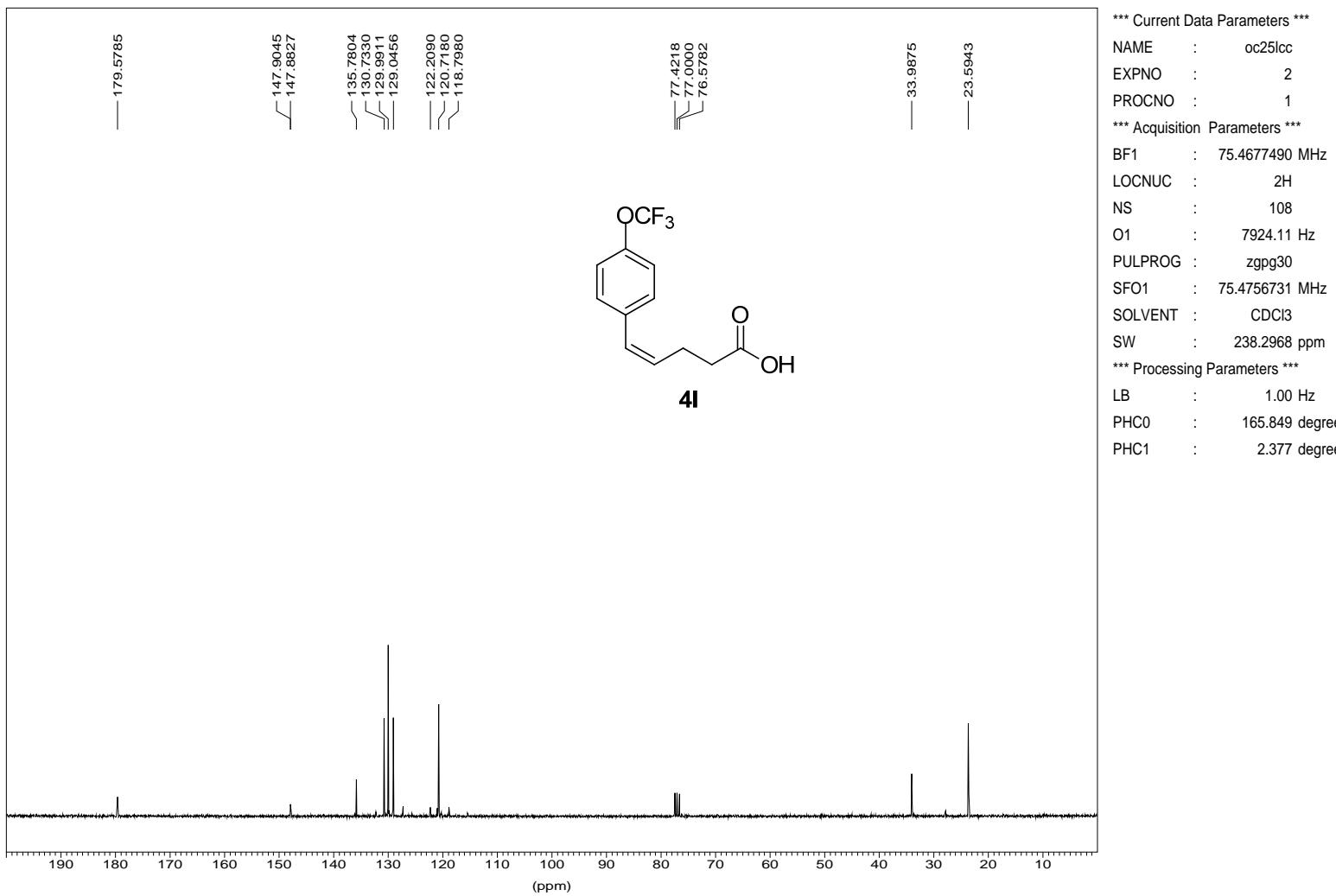
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 300.1300000 MHz

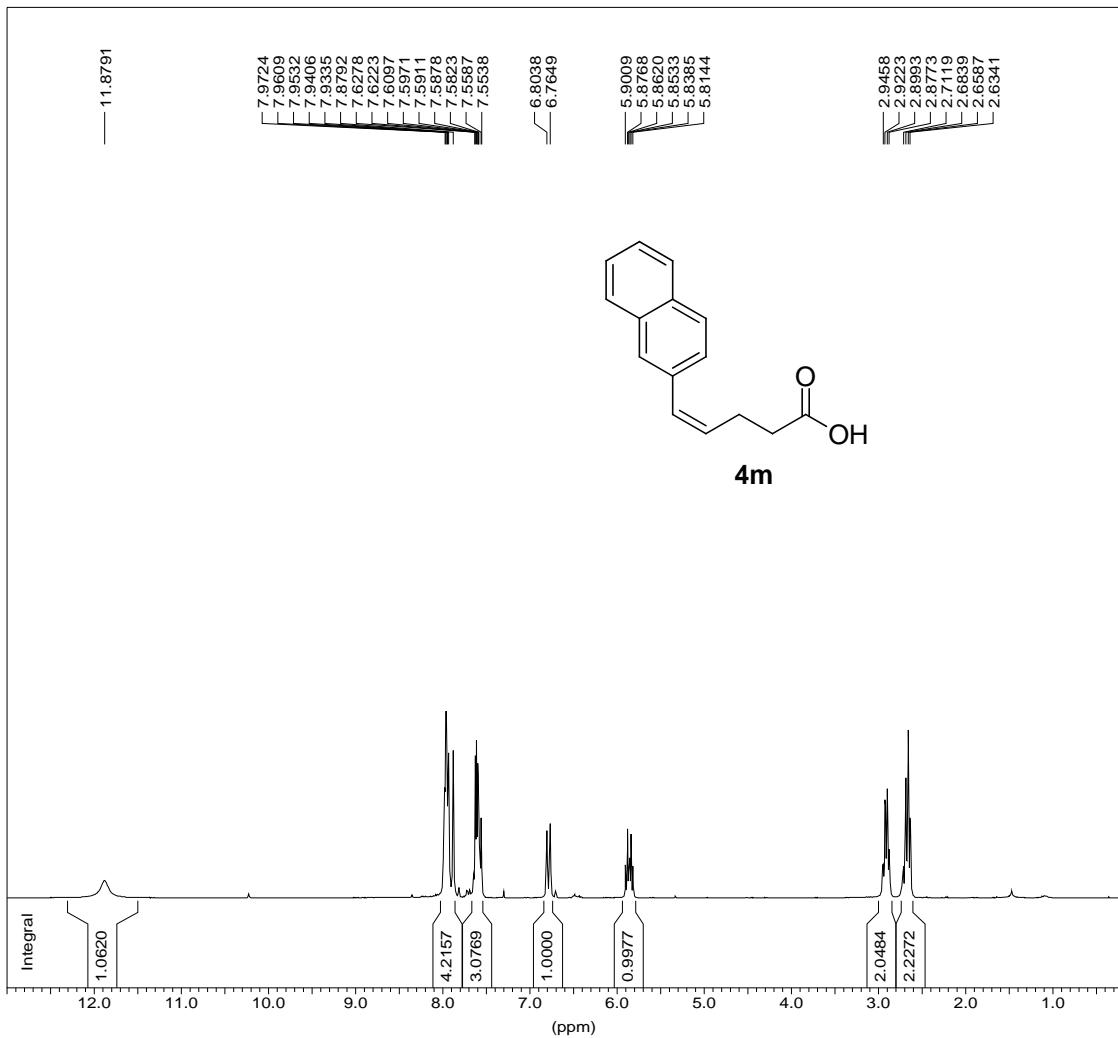
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-079-A

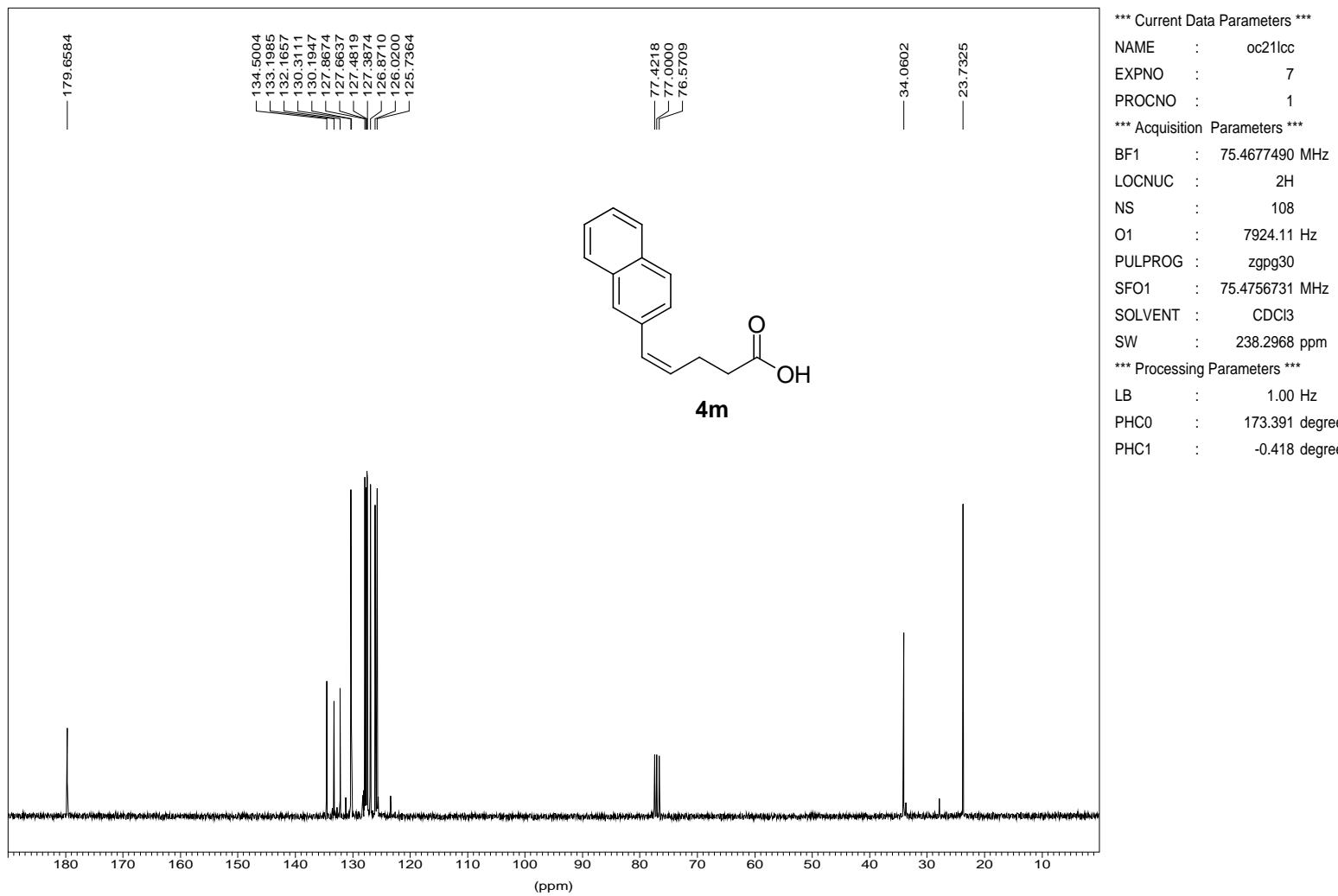


<sup>1</sup>H normal range AC300 cc-i-076-B

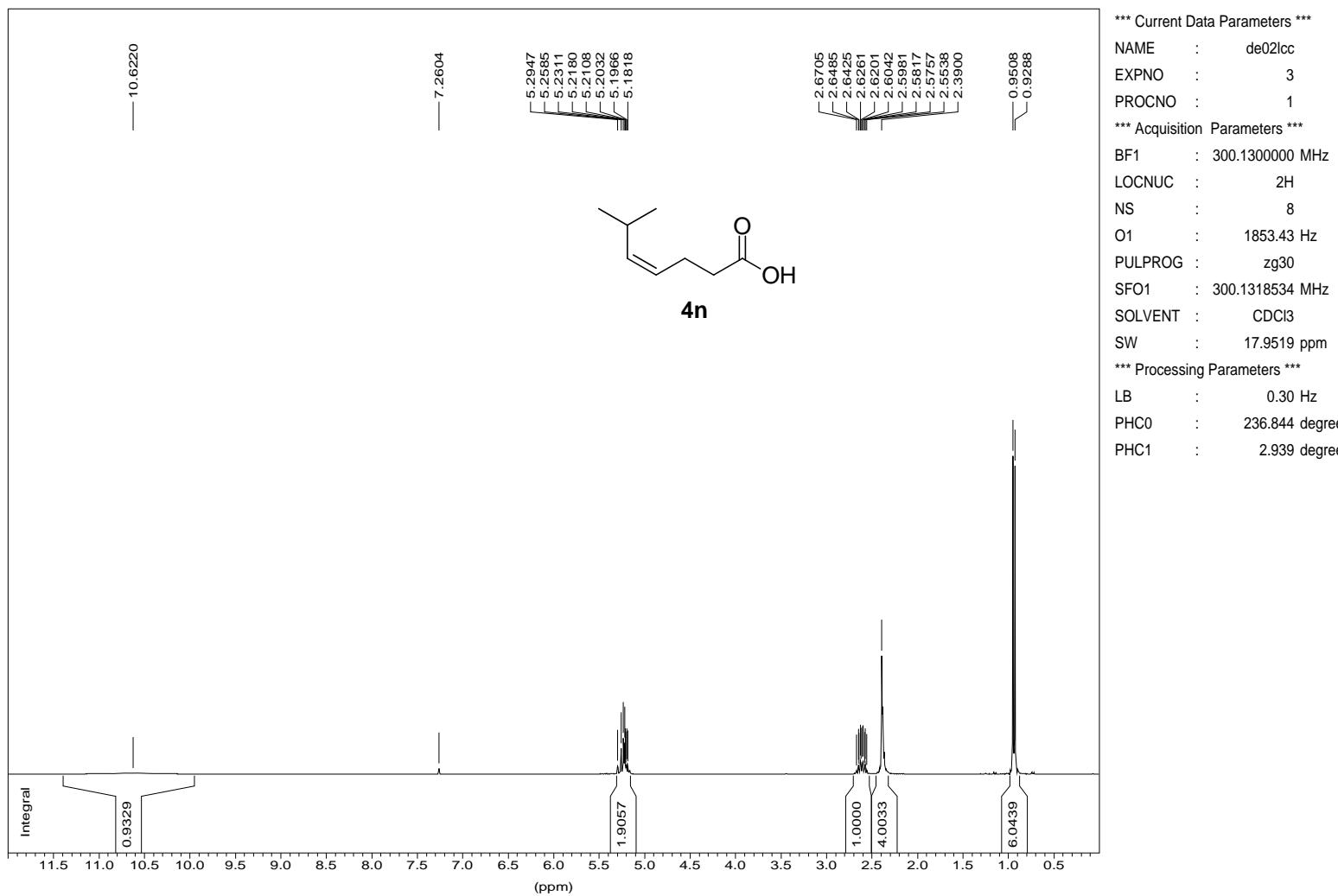


\*\*\* Current Data Parameters \*\*\*  
NAME : oc21lcc  
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PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

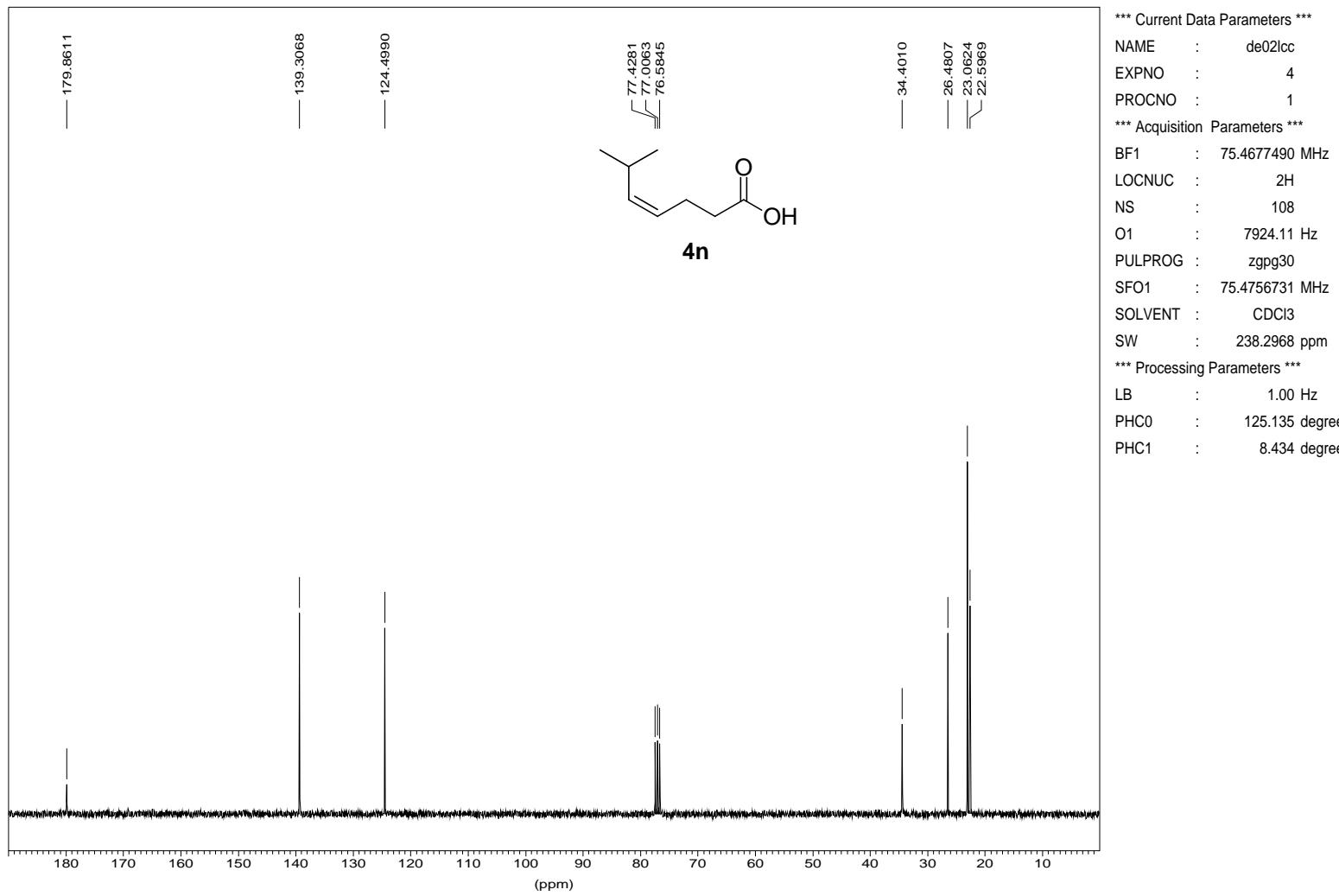
<sup>13</sup>C Standard AC300 cc-i-076-B



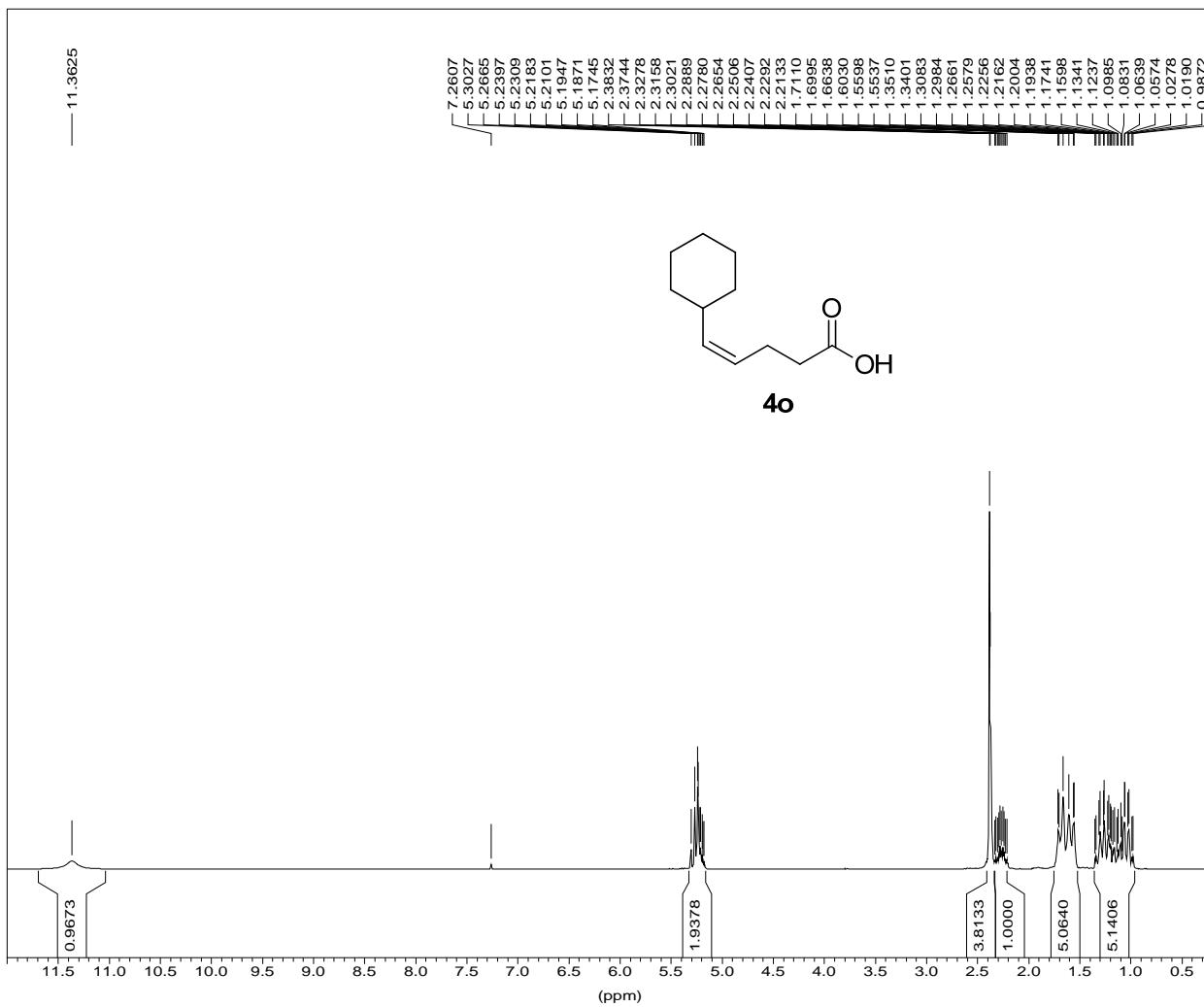
<sup>1</sup>H normal range AC300 cc-i-103-B



<sup>13</sup>C Standard AC300 cc-i-103-B



<sup>1</sup>H normal range AC300 cc-i-103-D



\*\*\* Current Data Parameters \*\*\*

NAME : de02lc  
EXPNO : 7  
PROCNO : 1

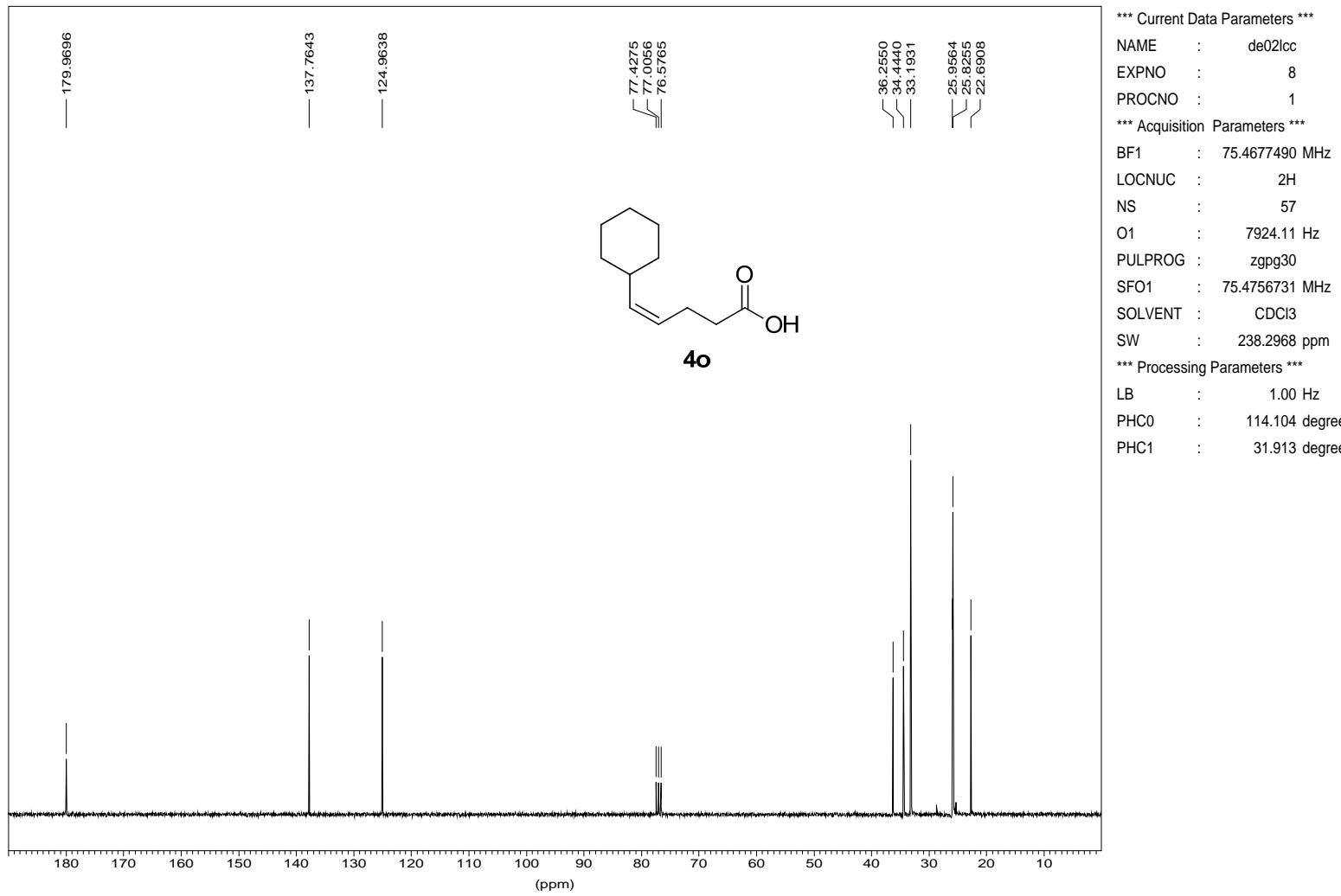
\*\*\* Acquisition Parameters \*\*\*

BF1 : 300.1300000 MHz  
LOCNUC : 2H  
NS : 8  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm

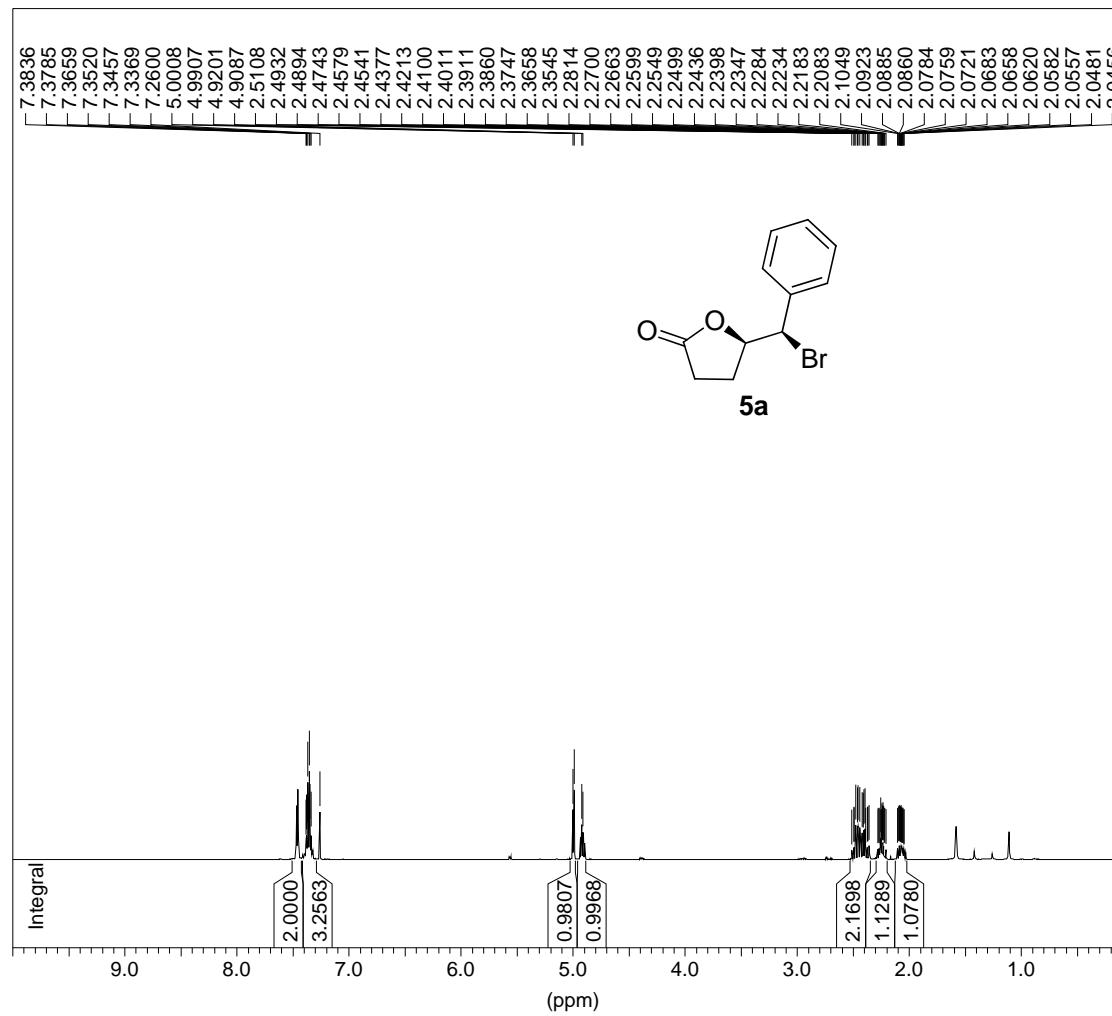
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
PHC0 : 126.063 degree  
PHC1 : 1.329 degree

<sup>13</sup>C Standard AC300 cc-i-103-C



<sup>1</sup>H AMX500 cis lactone 1



Bruker

\*\*\* Current Data Parameters \*\*\*

NAME : ck0415  
EXPNO : 3  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

INSTRUM : av500  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768

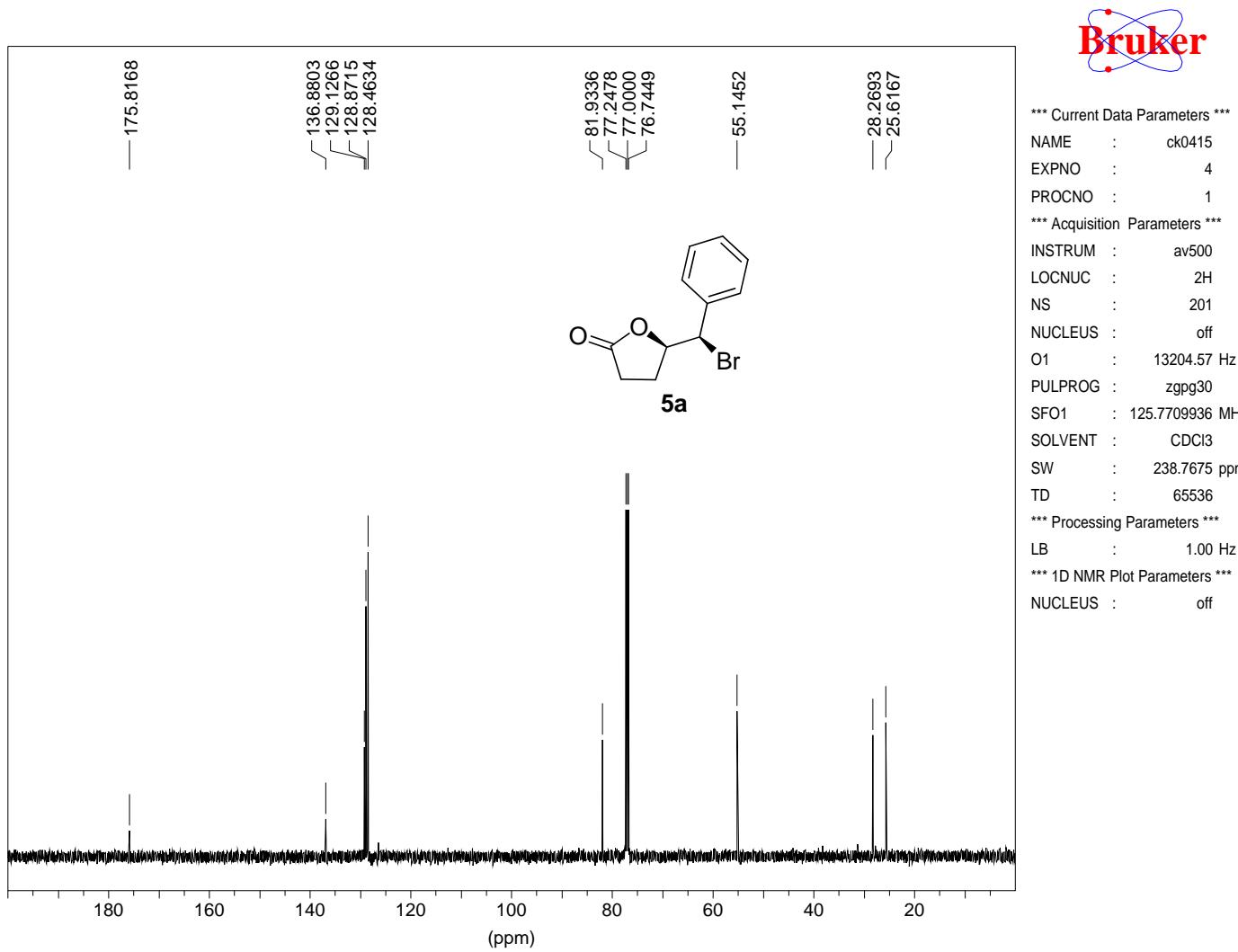
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz

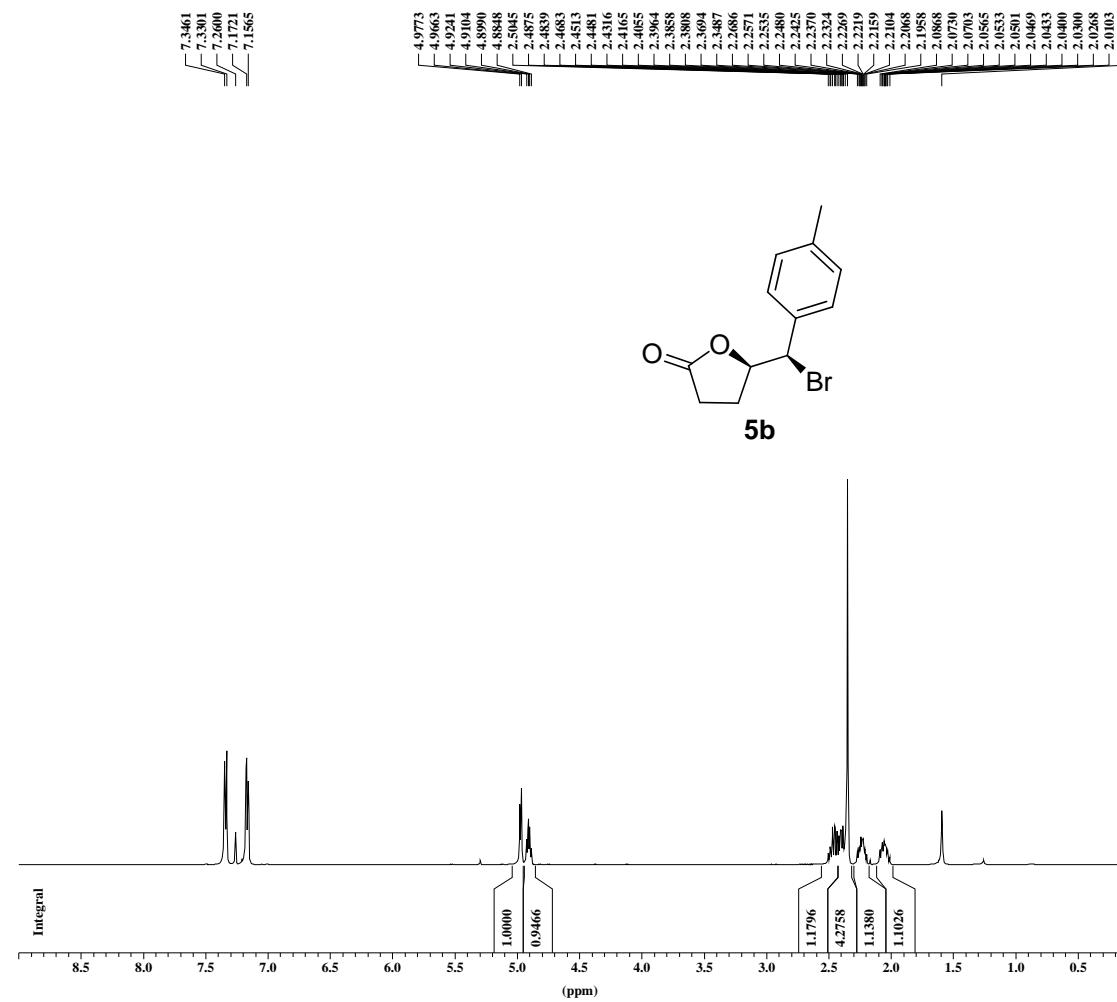
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C AMX500 cis lactone 1



cc-i-167a



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214

EXPNO : 1

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 2751.27 Hz

PULPROG : zg

SFO1 : 500.2327513 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 15.0080 ppm

TD : 32768

TE : 300.0 K

\*\*\* Processing Parameters \*\*\*

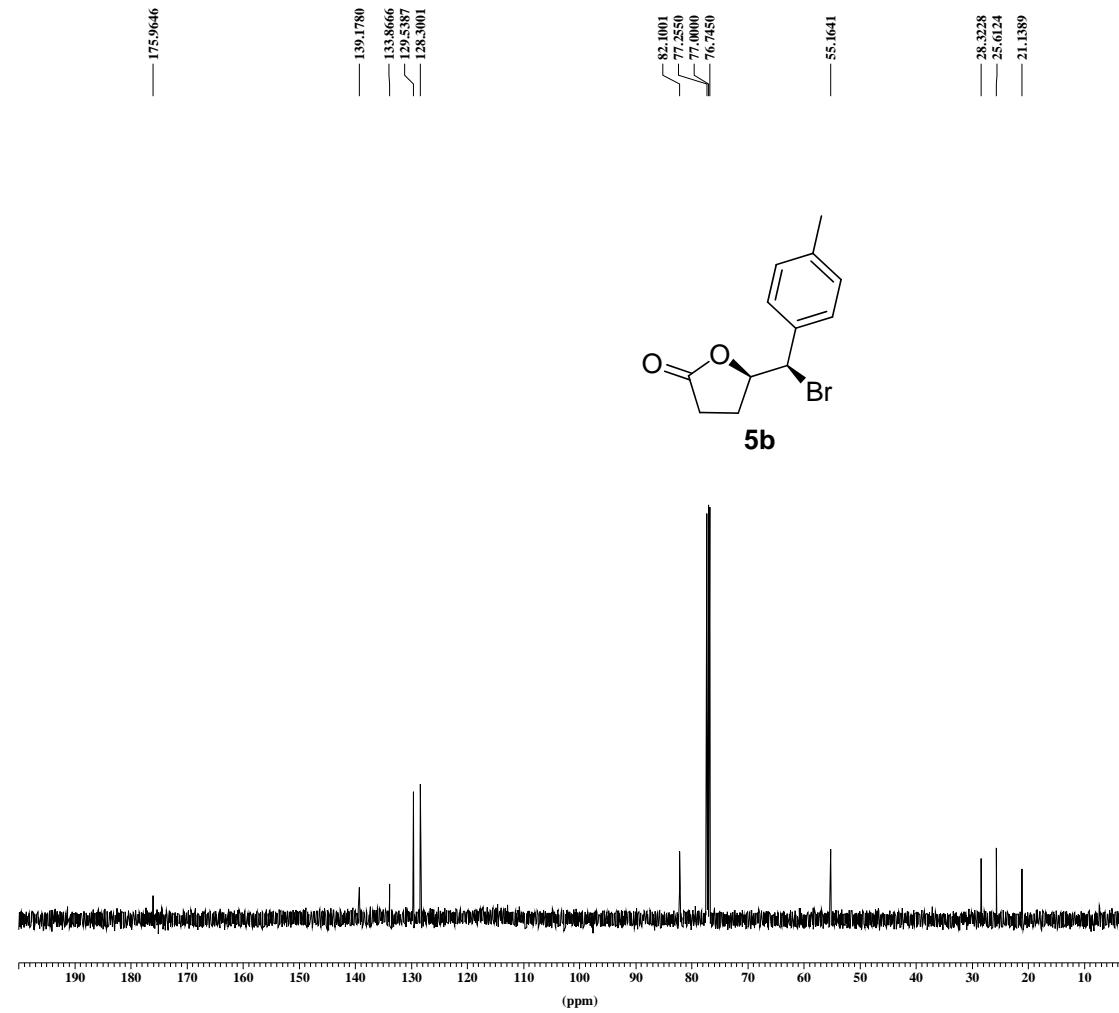
LB : 0.10 Hz

SF : 500.2300090 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

cc-i-167a



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214  
EXPNO : 2  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 210  
NUCLEUS : off  
O1 : 12577.84 Hz  
PULPROG : zgpg30  
SFO1 : 125.7955118 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7210 ppm  
TD : 65536  
TE : 300.0 K

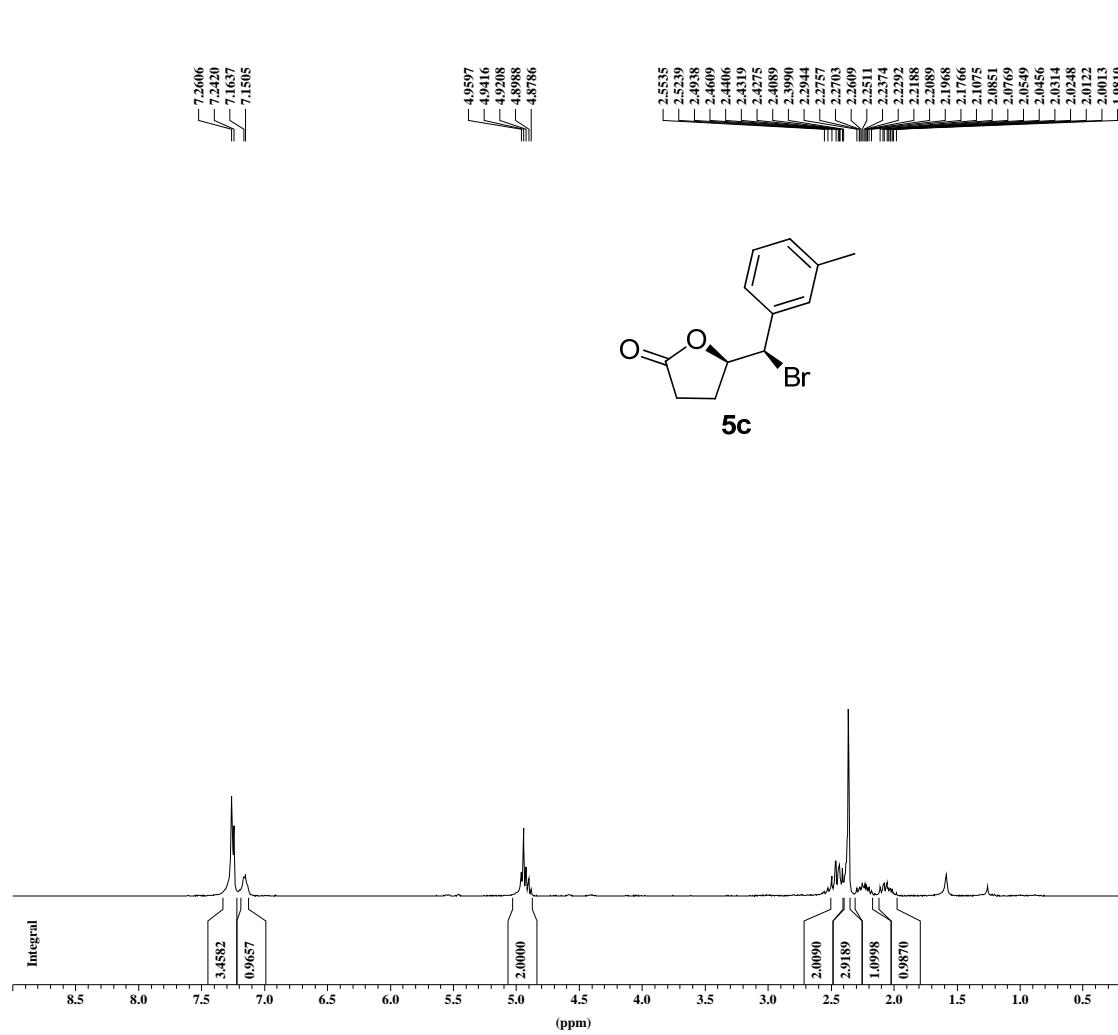
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 125.7829370 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

1H normal range AC300 cc-i-112-D



\*\*\* Current Data Parameters \*\*\*

NAME : ja05lcc

EXPNO : 6

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

TD : 32768

TE : 678.6 K

\*\*\* Processing Parameters \*\*\*

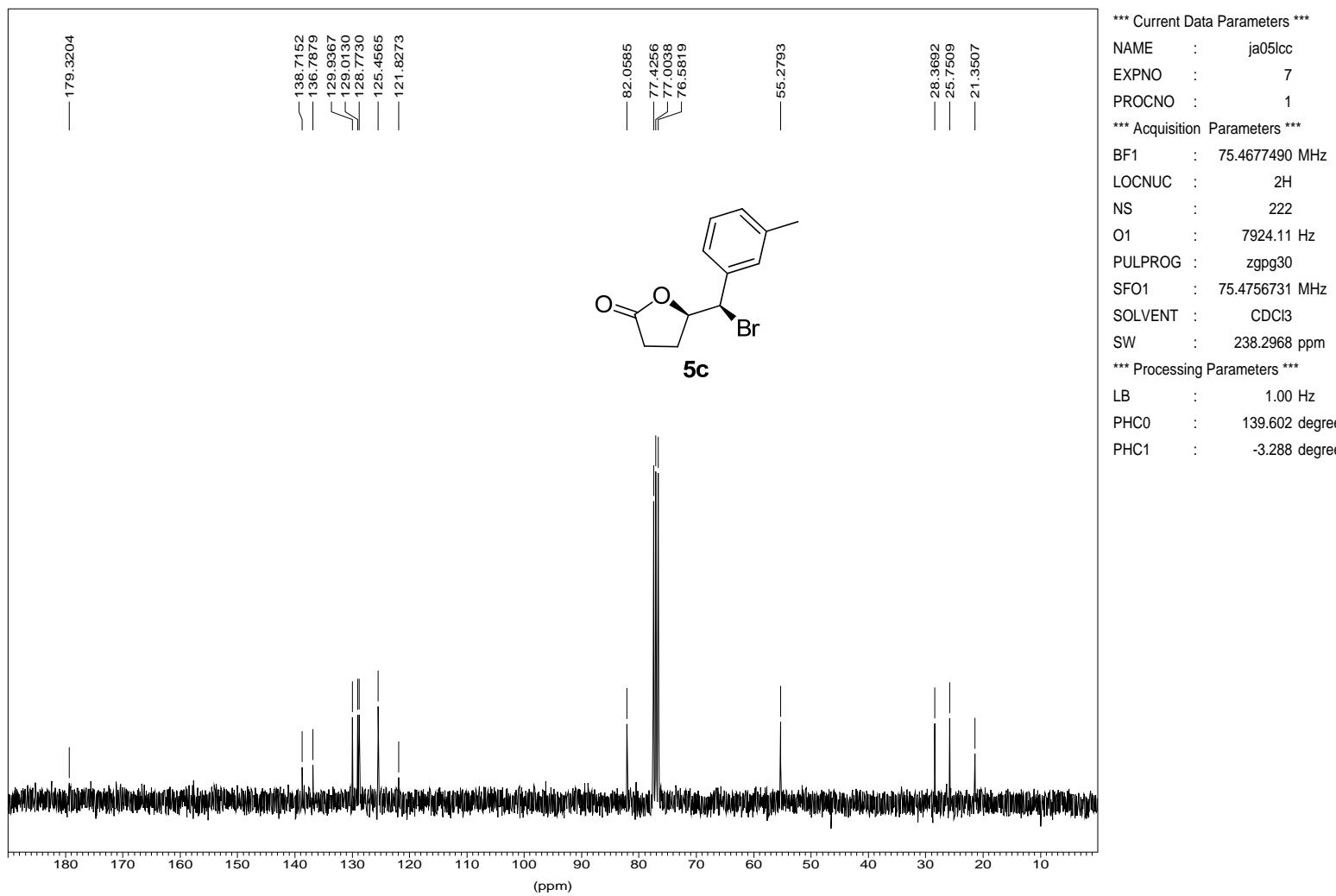
LB : 0.30 Hz

SF : 300.1300115 MHz

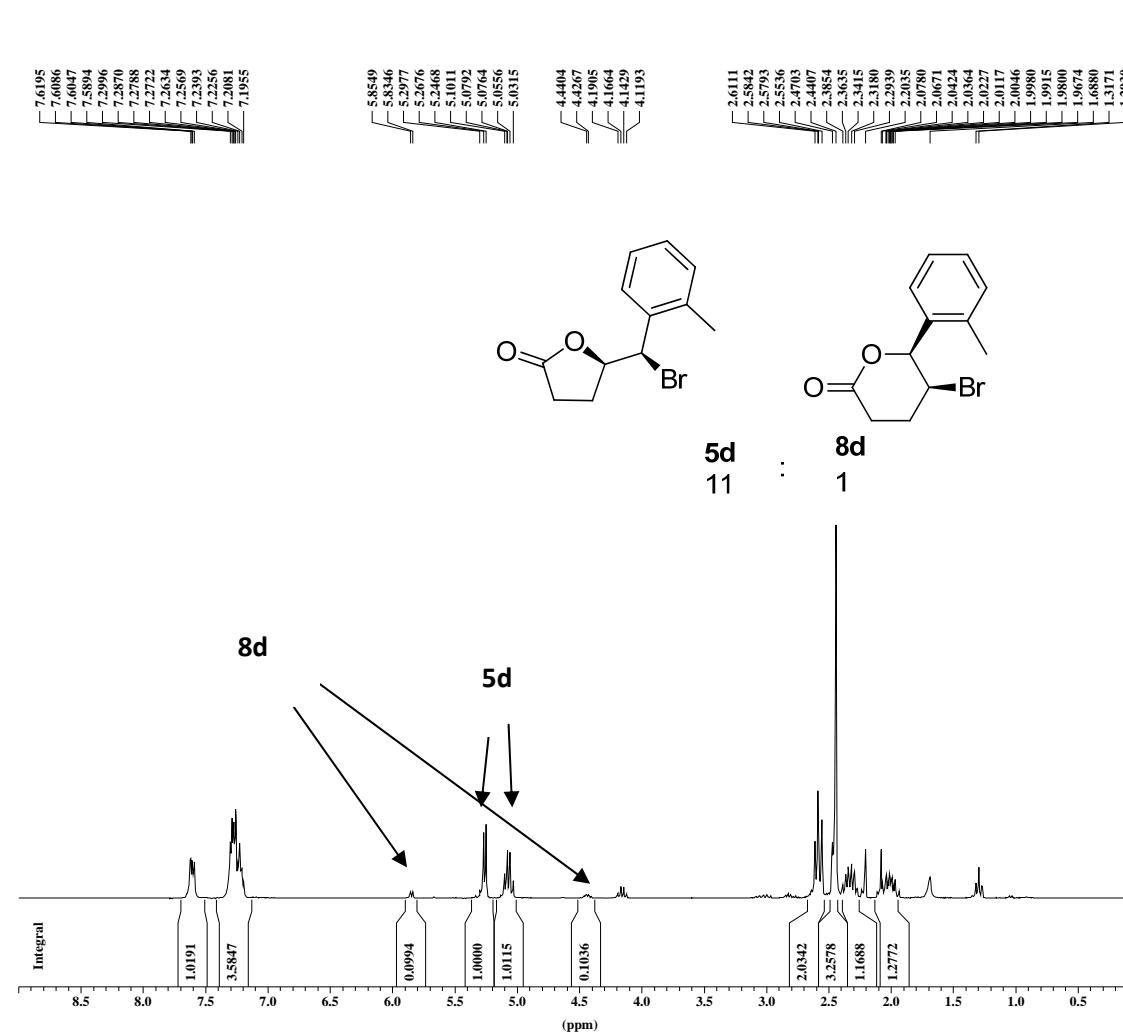
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-i-112-D



1H normal range AC300 cc-i-162b



\*\*\* Current Data Parameters \*\*\*

NAME : fe11ck

EXPNO : 1

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

TD : 32768

TE : 297.1 K

\*\*\* Processing Parameters \*\*\*

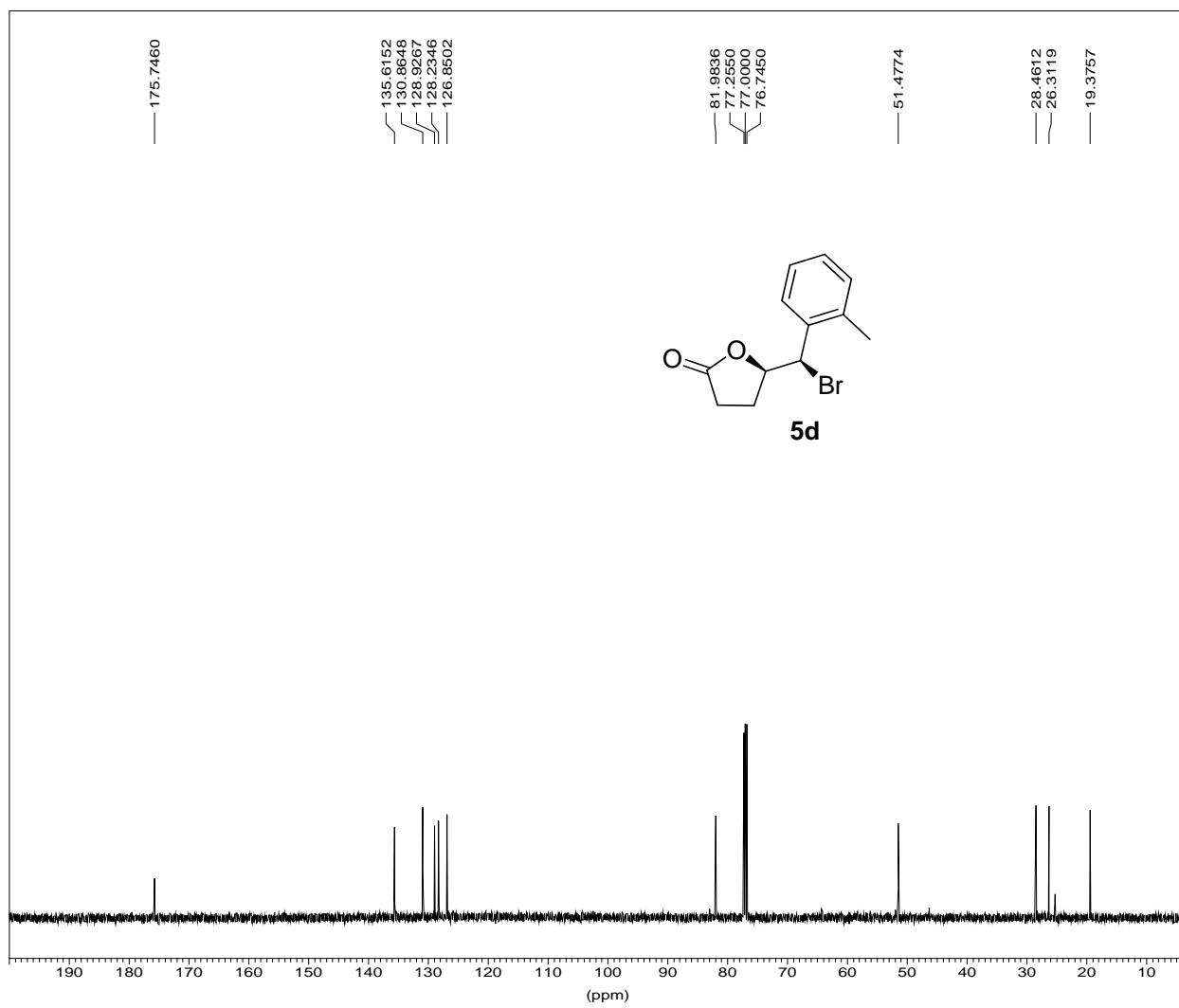
LB : 0.30 Hz

SF : 300.1300000 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

ck-v-119



\*\*\* Current Data Parameters \*\*\*

NAME : ck0131  
EXPNO : 4  
PROCNO : 1

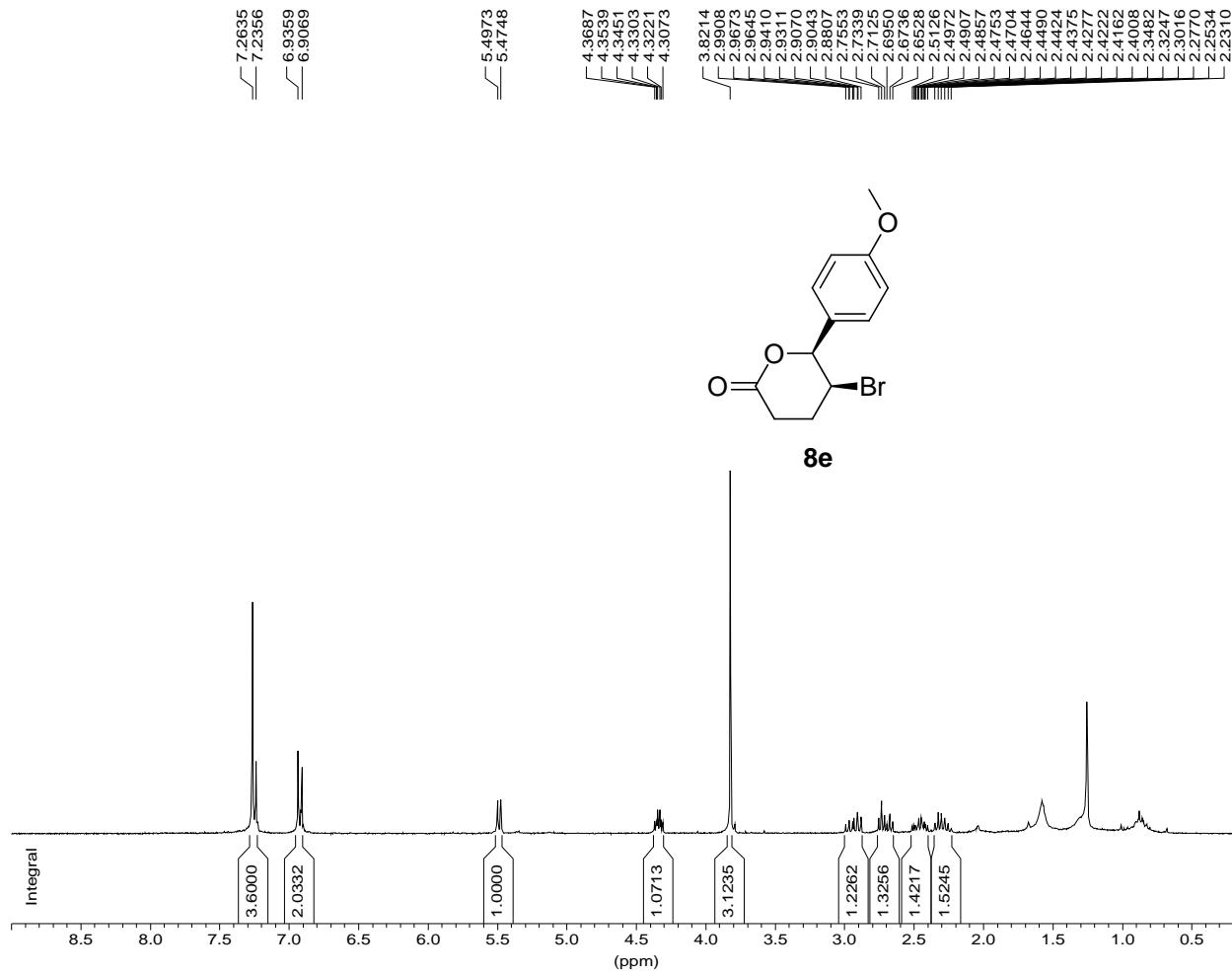
\*\*\* Acquisition Parameters \*\*\*

BF1 : 125.7829340 MHz  
LOCNUC : 2H  
NS : 140  
O1 : 12577.84 Hz  
PULPROG : zgpg30  
SFO1 : 125.7955118 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7210 ppm

\*\*\* Processing Parameters \*\*\*

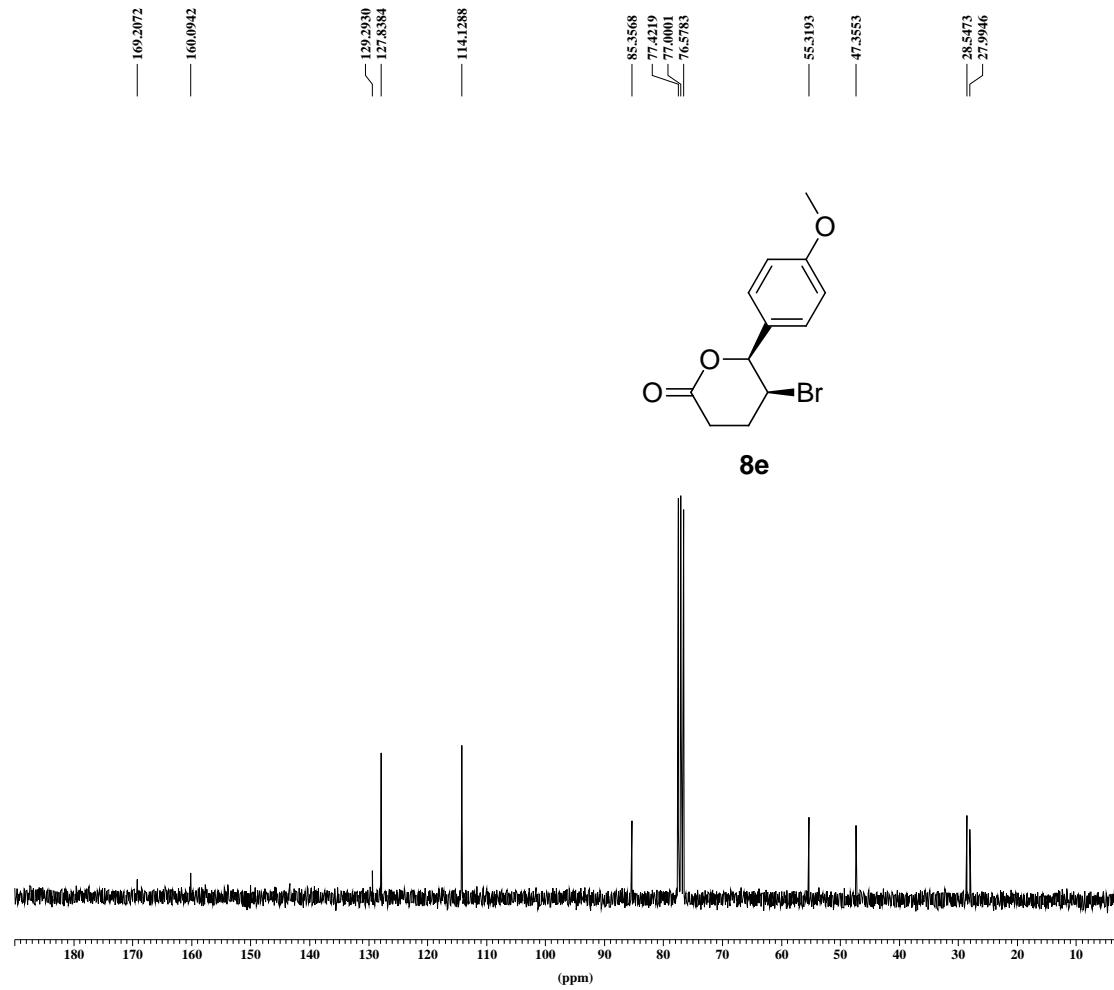
LB : 1.00 Hz  
PHC0 : 188.334 degree  
PHC1 : 11.705 degree

<sup>1</sup>H normal range AC300 cc-i-104-A



\*\*\* Current Data Parameters \*\*\*  
NAME : de06icc  
EXPNO : 1  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
\*\*\* Processing Parameters \*\*\*  
LB : 0.30 Hz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-i-166-C



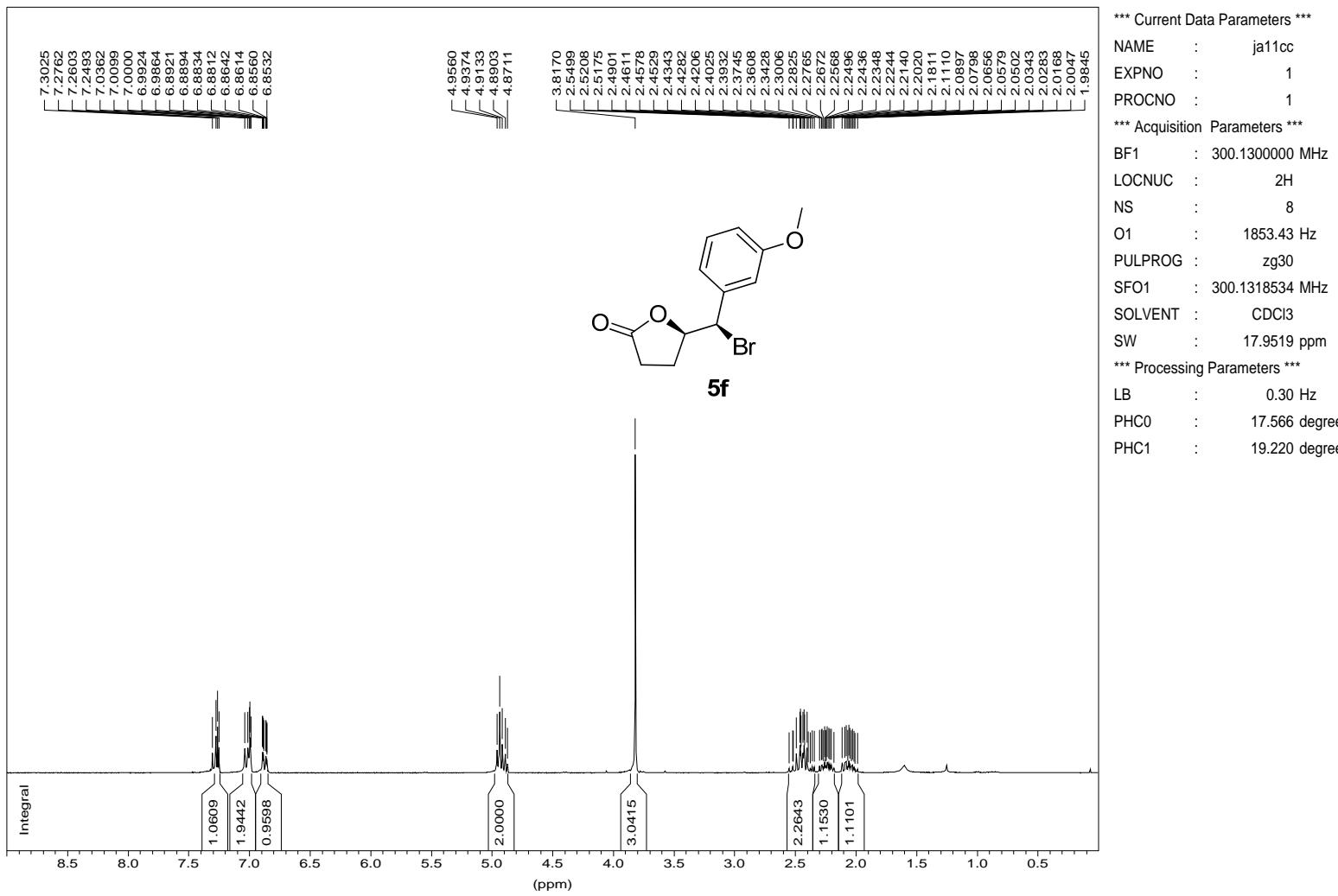
\*\*\* Current Data Parameters \*\*\*

NAME : fe11cc  
EXPNO : 6  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 657  
NUCLEUS : off  
O1 : 7924.11 Hz  
PULPROG : zgpg30  
SFO1 : 75.4756731 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.2968 ppm  
TD : 32768  
TE : 297.1 K

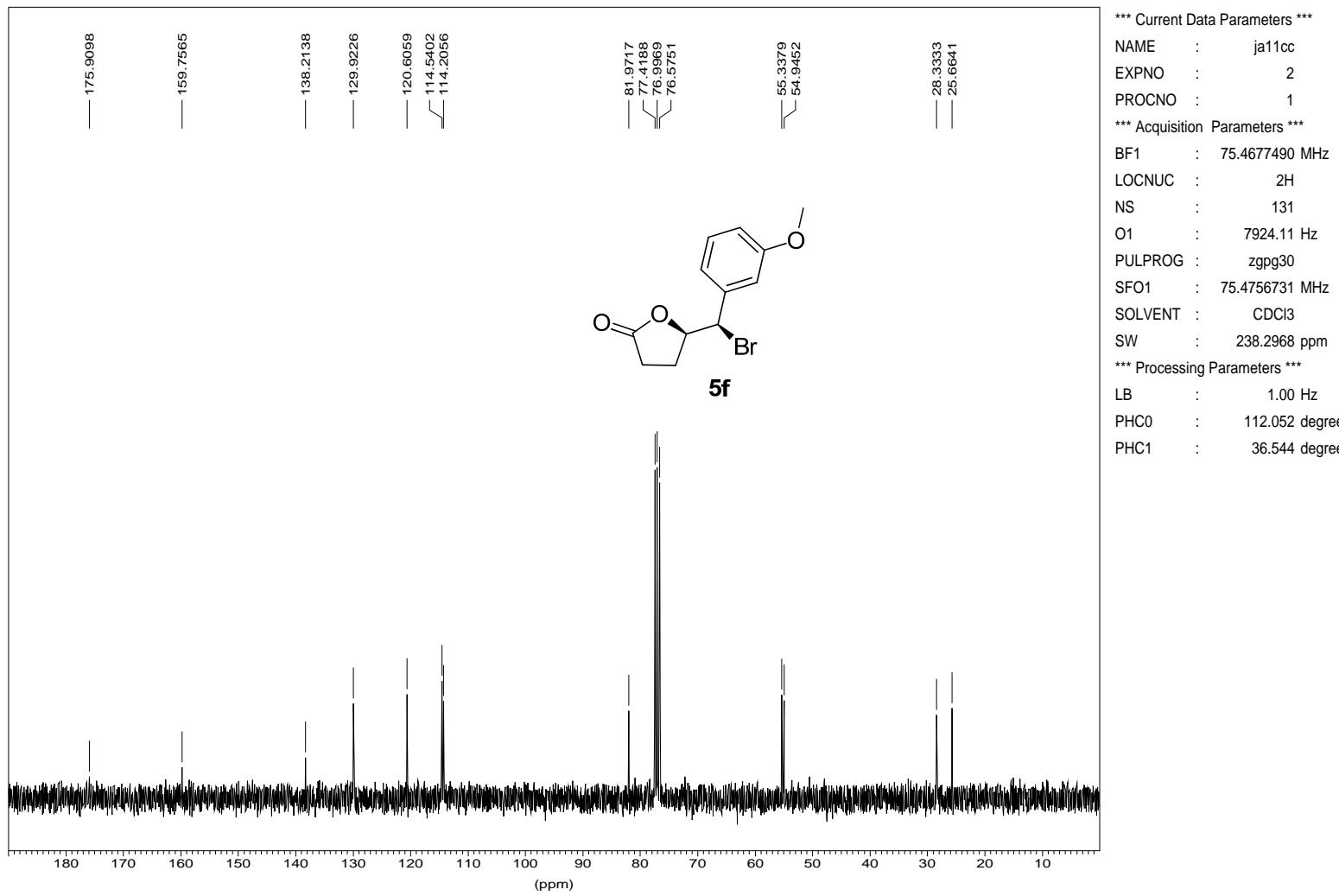
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 75.4677517 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

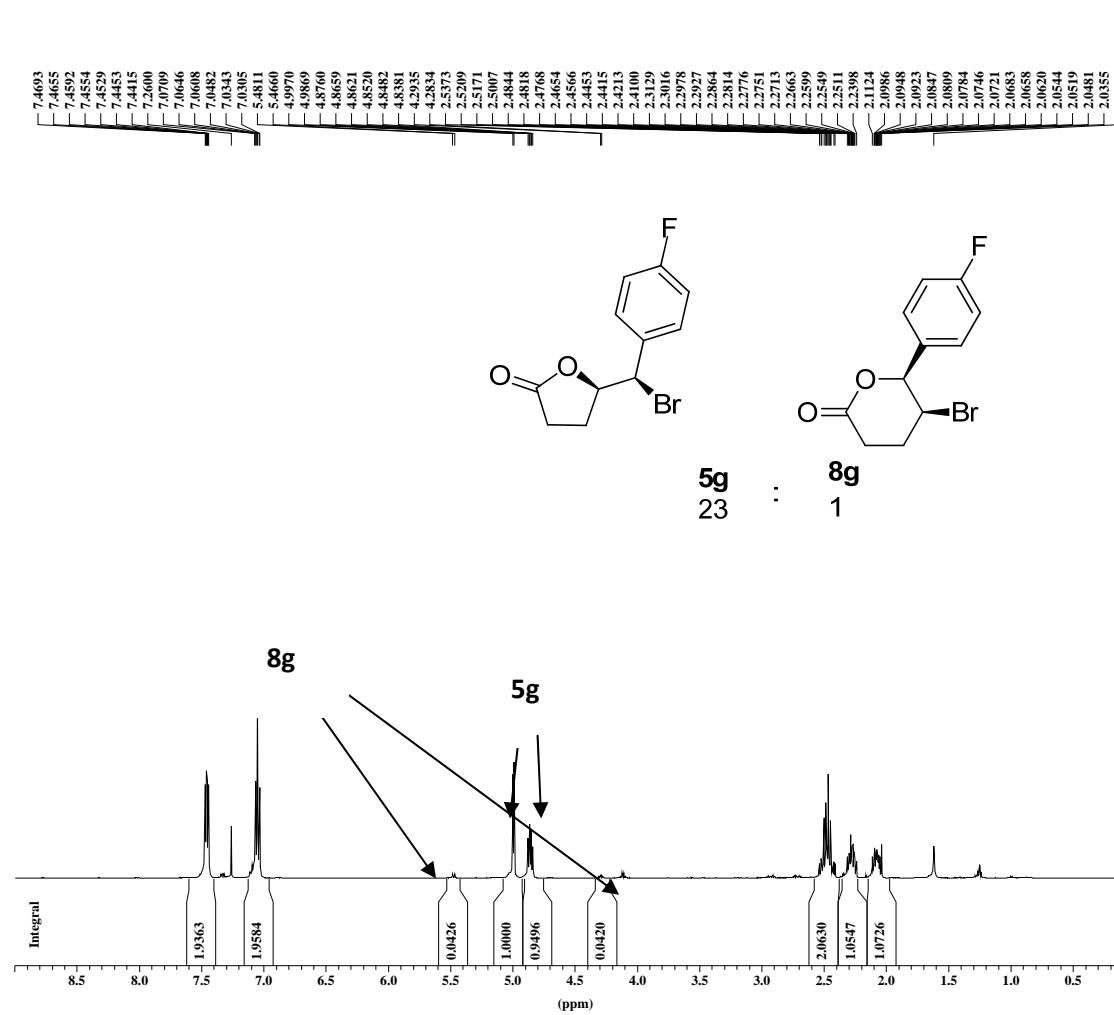
<sup>1</sup>H normal range AC300 cc-112-A



<sup>13</sup>C Standard AC300 cc-112-A



1H AMX500 cc-i-165



\*\*\* Current Data Parameters \*\*\*

NAME : ck0213  
EXPNO : 2  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 297.9 K

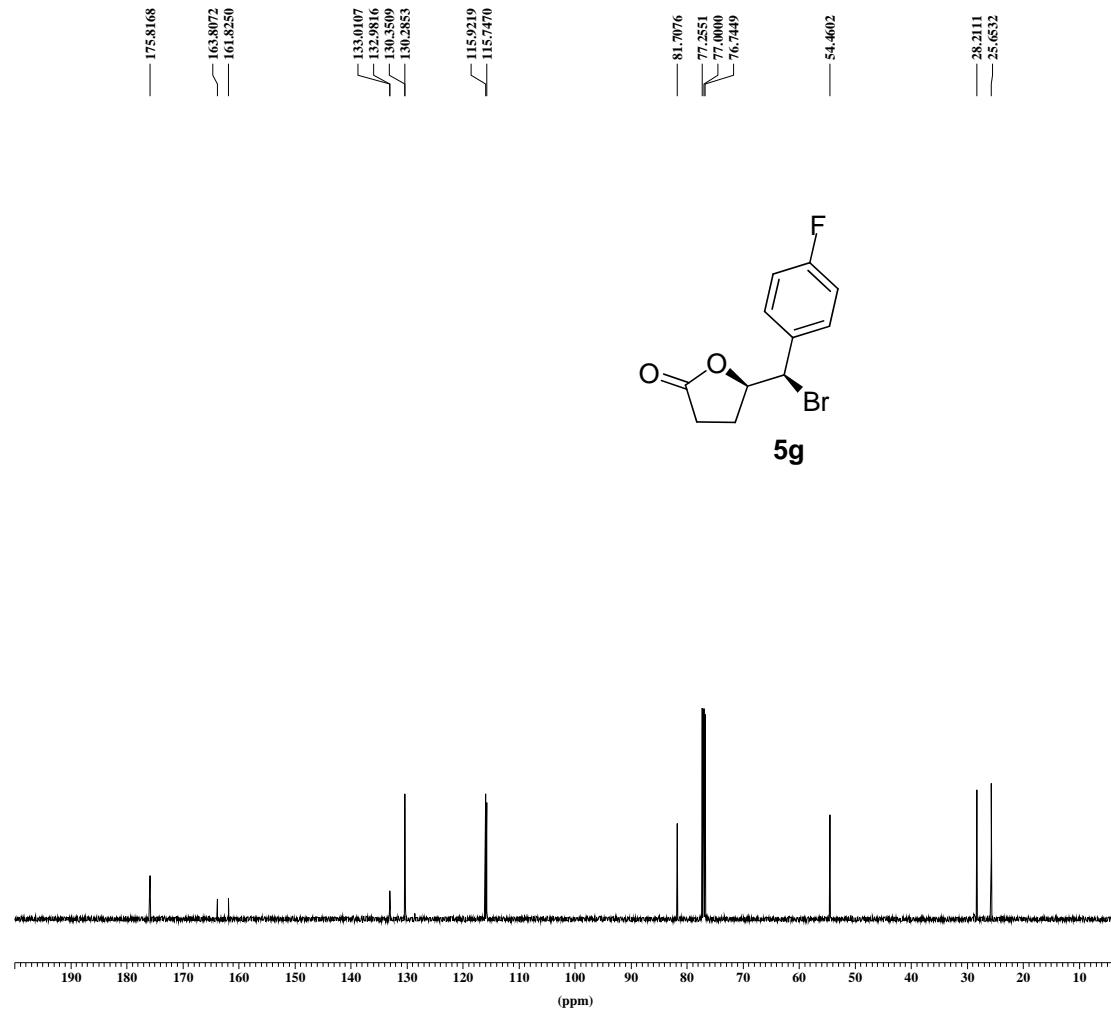
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300140 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

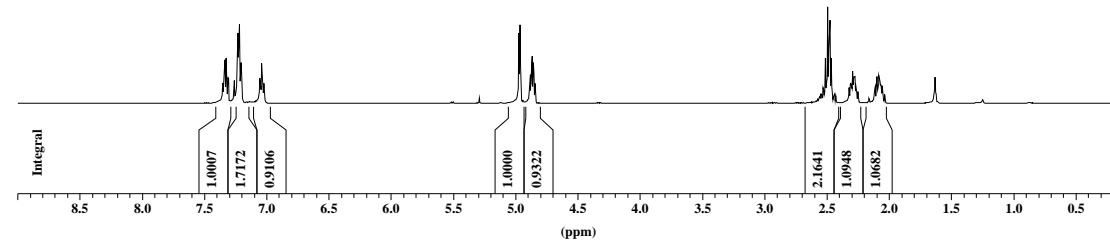
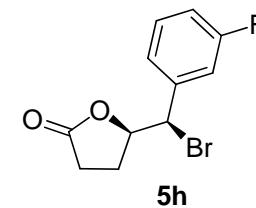
13C AMX500 cc-i-165



\*\*\* Current Data Parameters \*\*\*

NAME : ck0213  
EXPNO : 3  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 227  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7675 ppm  
TD : 65536  
TE : 298.3 K  
\*\*\* Processing Parameters \*\*\*  
LB : 1.00 Hz  
SF : 125.7577950 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

cc-i-167b



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214

EXPNO : 3

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 2751.27 Hz

PULPROG : zg

SFO1 : 500.2327513 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 15.0080 ppm

TD : 32768

TE : 300.0 K

\*\*\* Processing Parameters \*\*\*

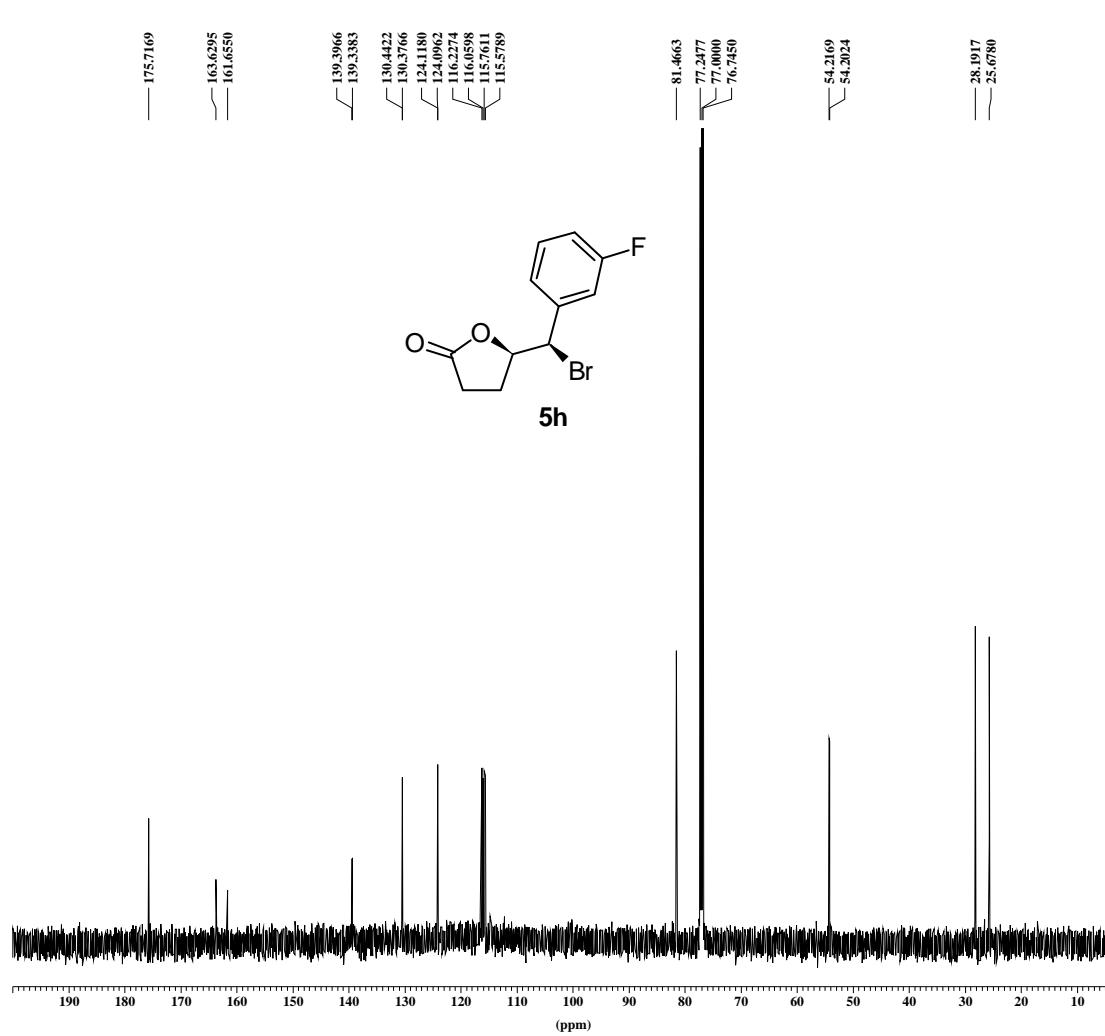
LB : 0.10 Hz

SF : 500.2300090 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

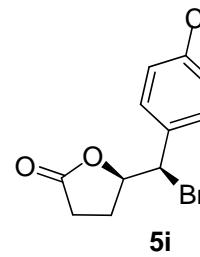
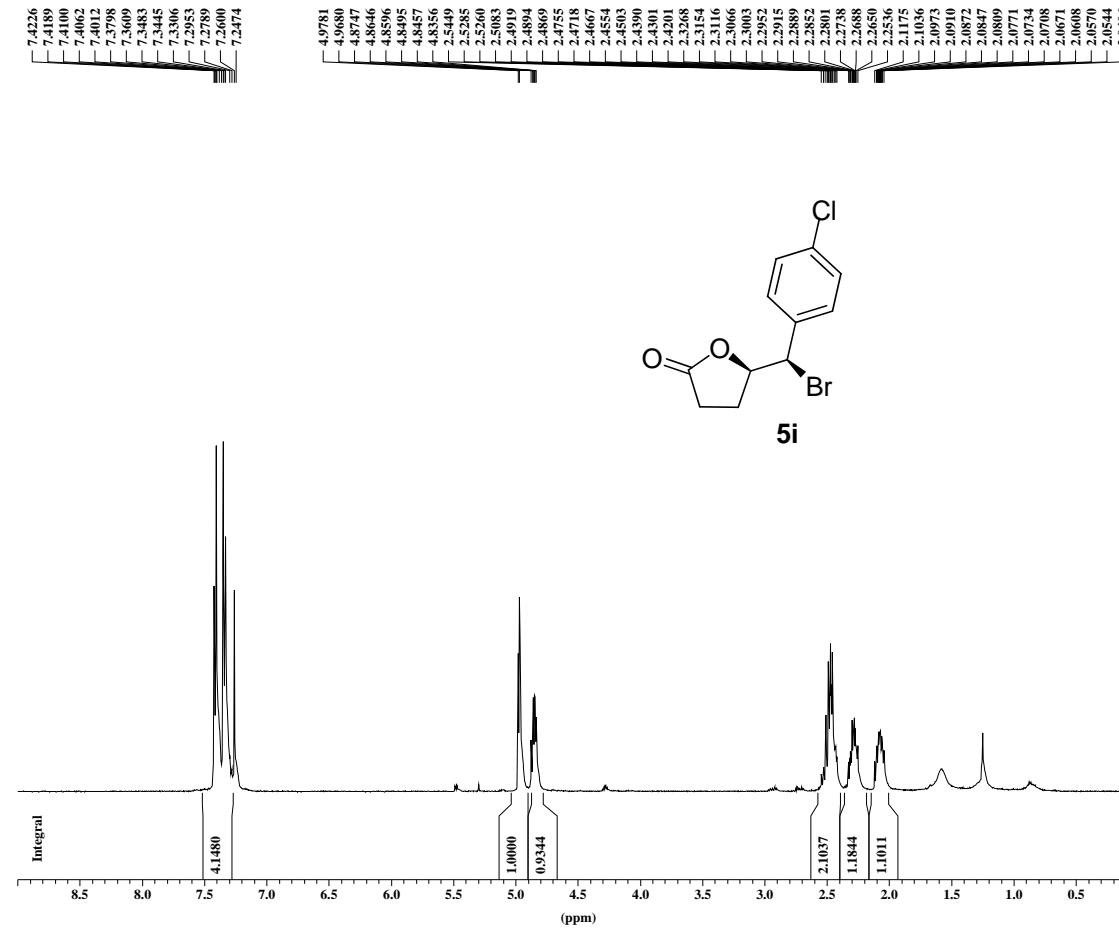
cc-i-167b



\*\*\* Current Data Parameters \*\*\*

NAME : ck0214  
EXPNO : 4  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 309  
NUCLEUS : off  
O1 : 12577.84 Hz  
PULPROG : zgpg30  
SFO1 : 125.7955118 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7210 ppm  
TD : 65536  
TE : 300.0 K  
\*\*\* Processing Parameters \*\*\*  
LB : 1.00 Hz  
SF : 125.7829400 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

1H AMX500 cc-i-104c



### \*\*\* Current Data Parameters \*\*\*

**NAME** : ck020

901 EXPNO .

PROCNO : 1

#### \*\*\* Acquisition Parameters \*\*\*

Acquisition Parameters

LOCNUC : 2H

NS : 8

**NUCLEUS :** off

O1 : 3088.51 Hz

PULPROG : zg30

**SFO1** : 500.1330885 MHz

SOLVENT : **C<sub>6</sub>H<sub>6</sub>**

SW : 20.6557 ppm

TD : 32768

TE : 285.2 N

TE : 295.3 K

### **\*\*\* Processing Parameters \*\*\***

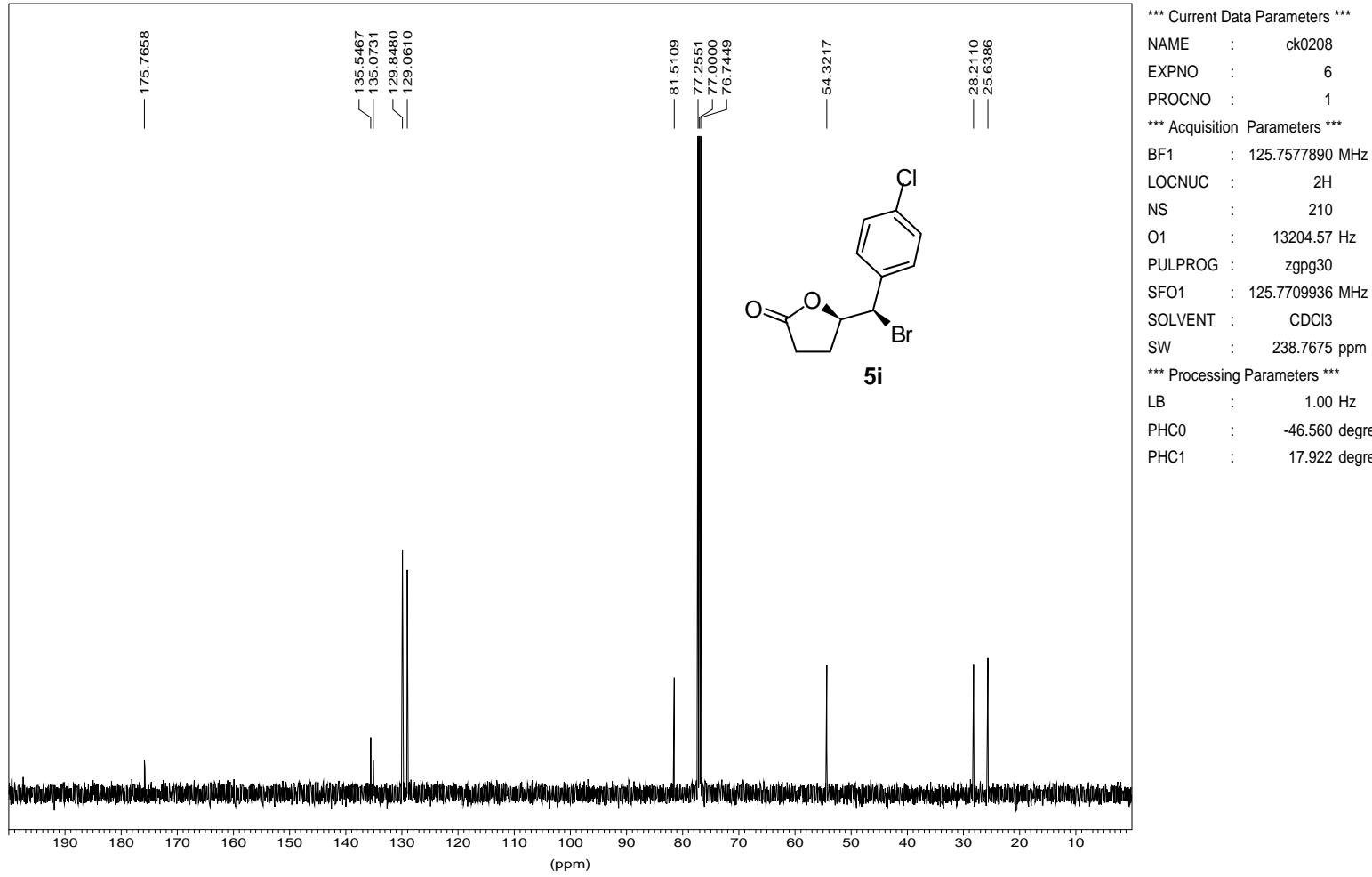
**LB** : **0.30 Hz**

SF : 500.1300140 MF

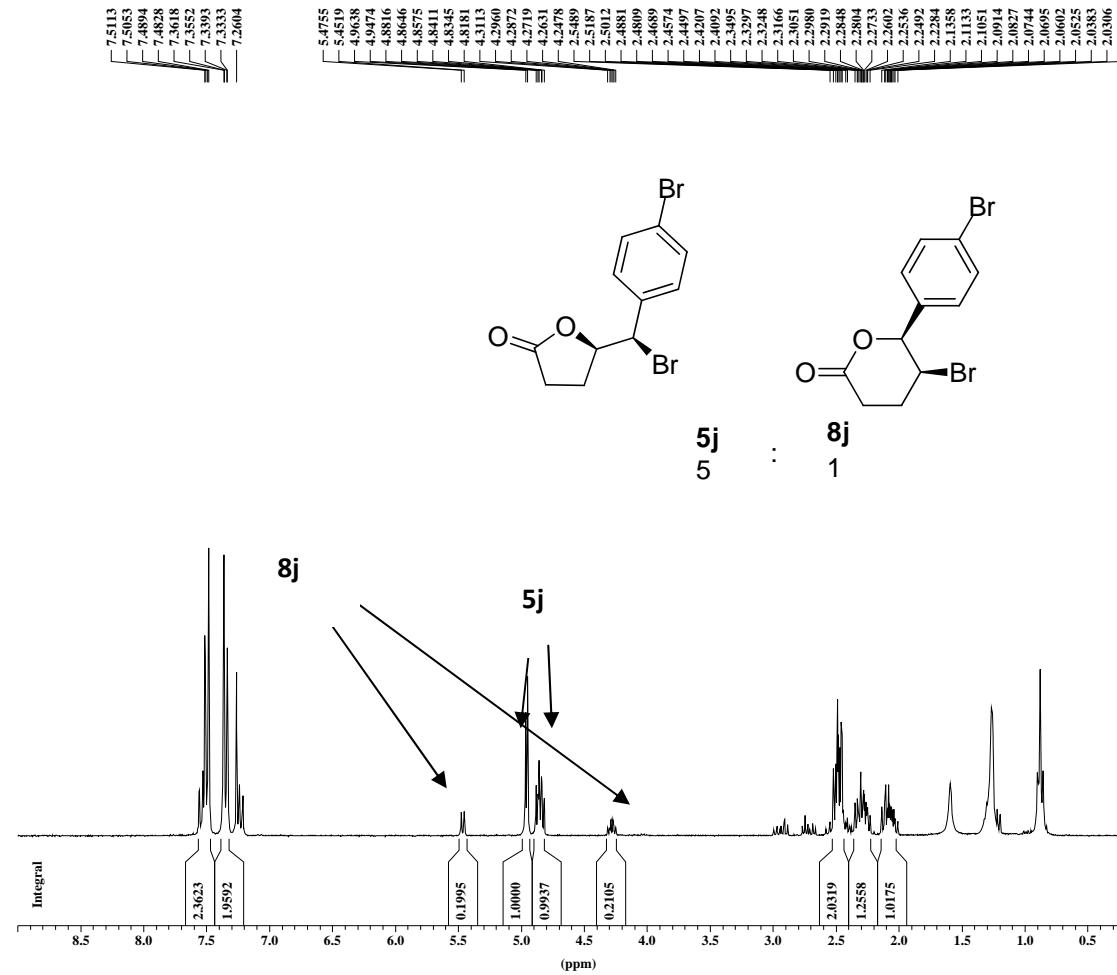
### \*\*\* 1D NMR Plot Parameters \*\*\*

**NUCLEUS :** off

<sup>13</sup>C AMX500 cc-i-104c



<sup>1</sup>H normal range AC300 cc-i-111



\*\*\* Current Data Parameters \*\*\*

NAME : de30lcc

EXPNO : 8

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

TD : 32768

TE : 678.6 K

\*\*\* Processing Parameters \*\*\*

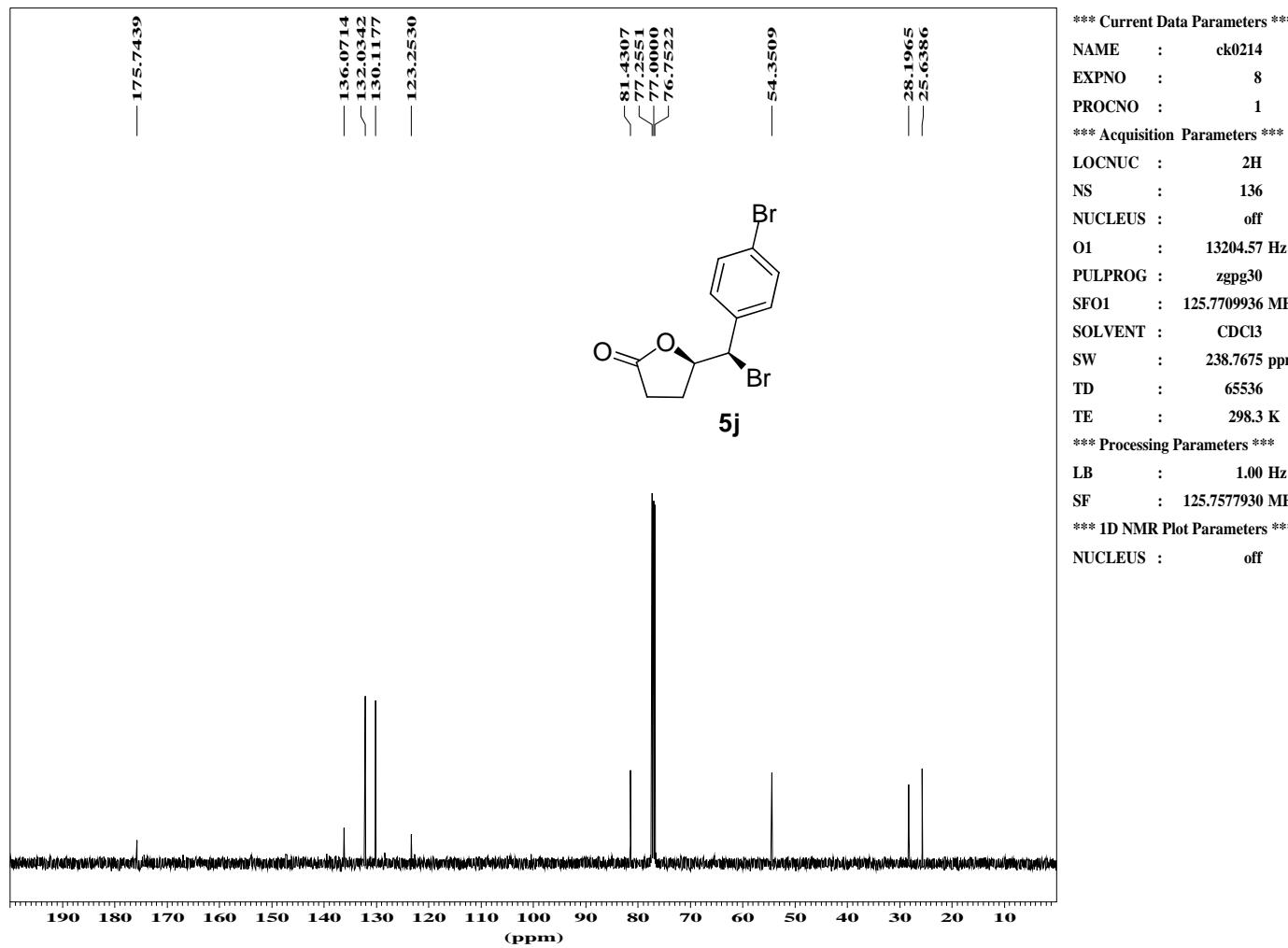
LB : 0.30 Hz

SF : 300.1300123 MHz

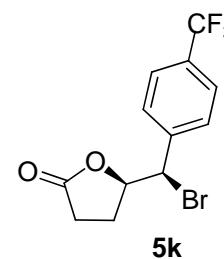
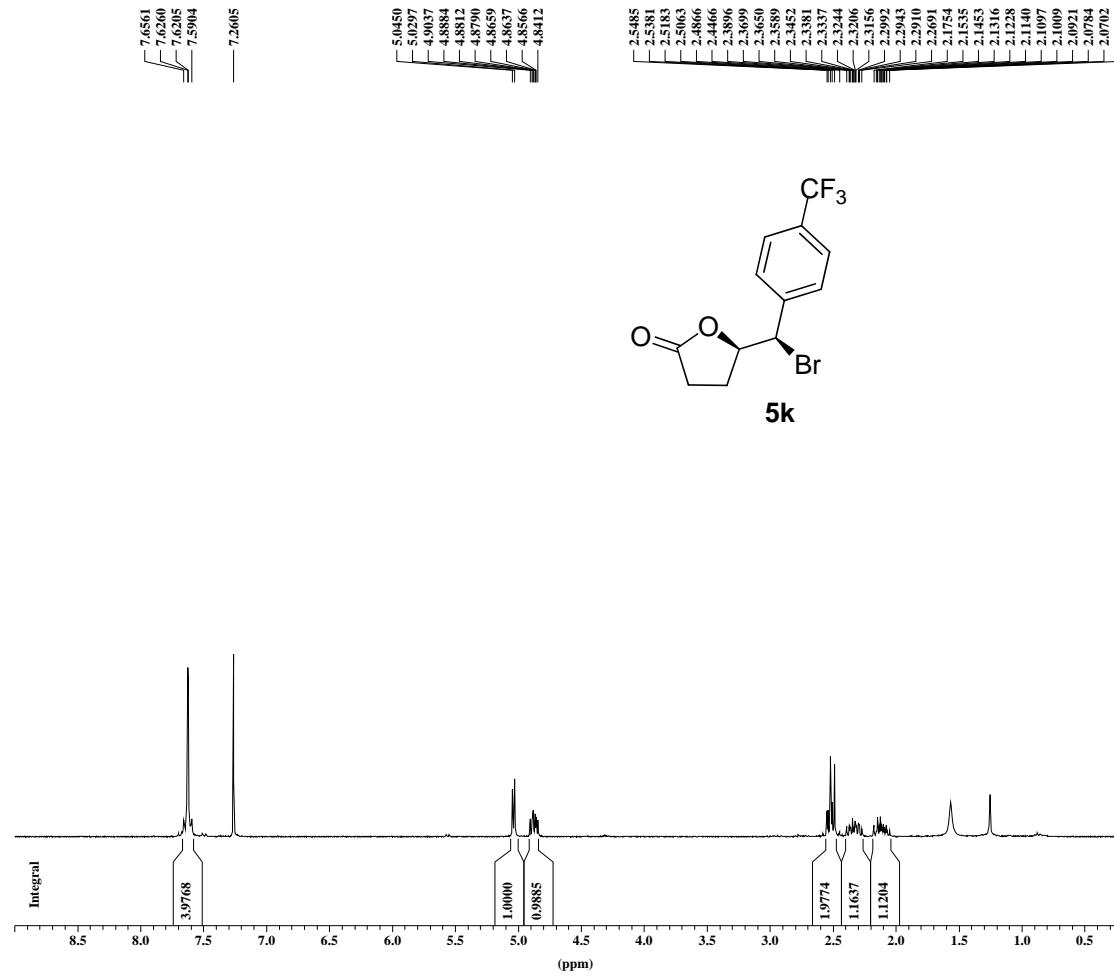
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

13C AMX500 4-Br lactone



1H normal range AC300 cc-112-B



\*\*\* Current Data Parameters \*\*\*

NAME : ja05lcc  
EXPNO : 4  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 17.9519 ppm  
TD : 32768  
TE : 678.6 K

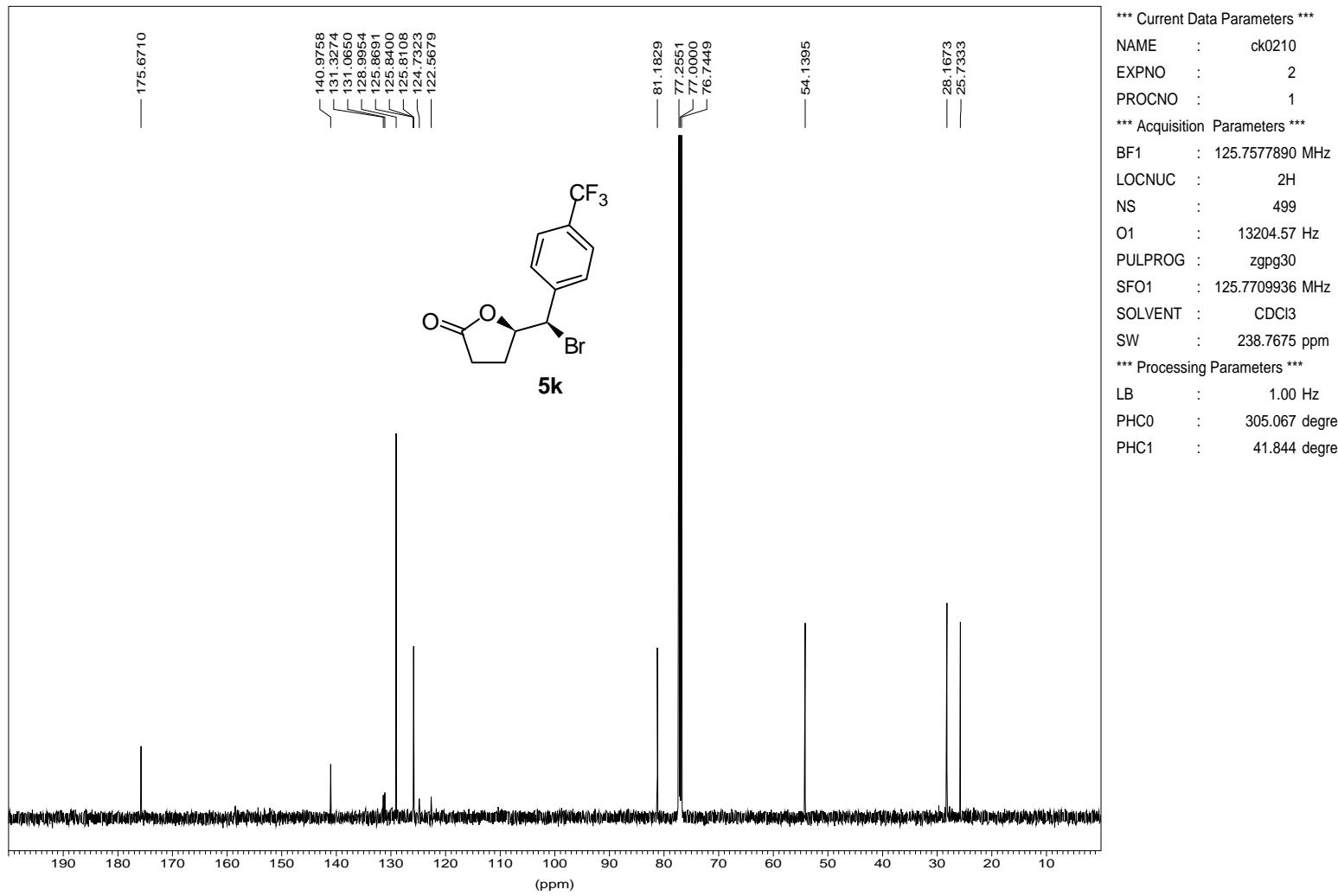
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 300.1300122 MHz

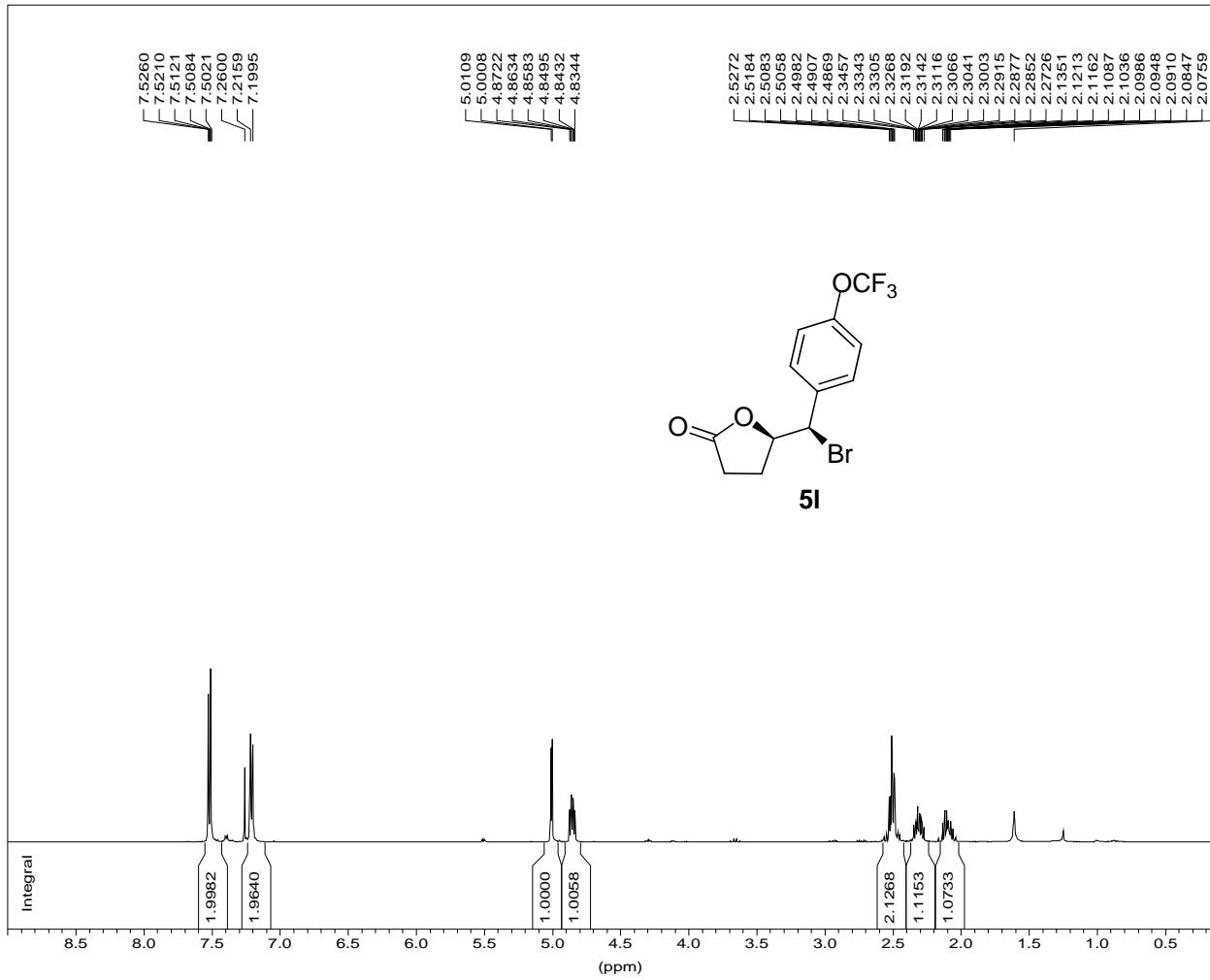
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C AMX500 cc-i-166a



<sup>1</sup>H AMX500 cc-i-166b



\*\*\* Current Data Parameters \*\*\*

NAME : ck0210

EXPNO : 3

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

BF1 : 500.1300000 MHz

LOCNUC : 2H

NS : 8

O1 : 3088.51 Hz

PULPROG : zg30

SFO1 : 500.1330885 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 20.6557 ppm

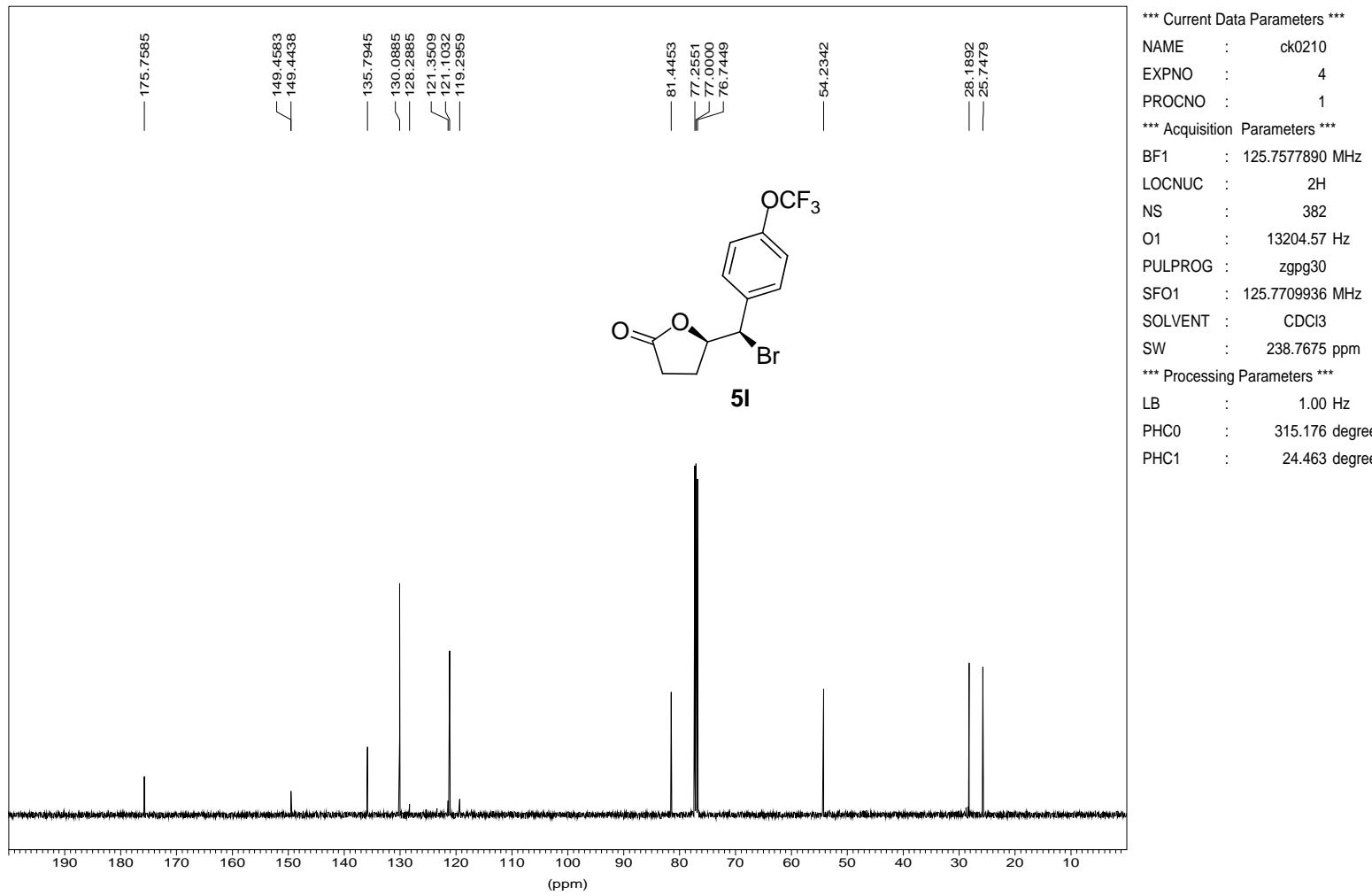
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz

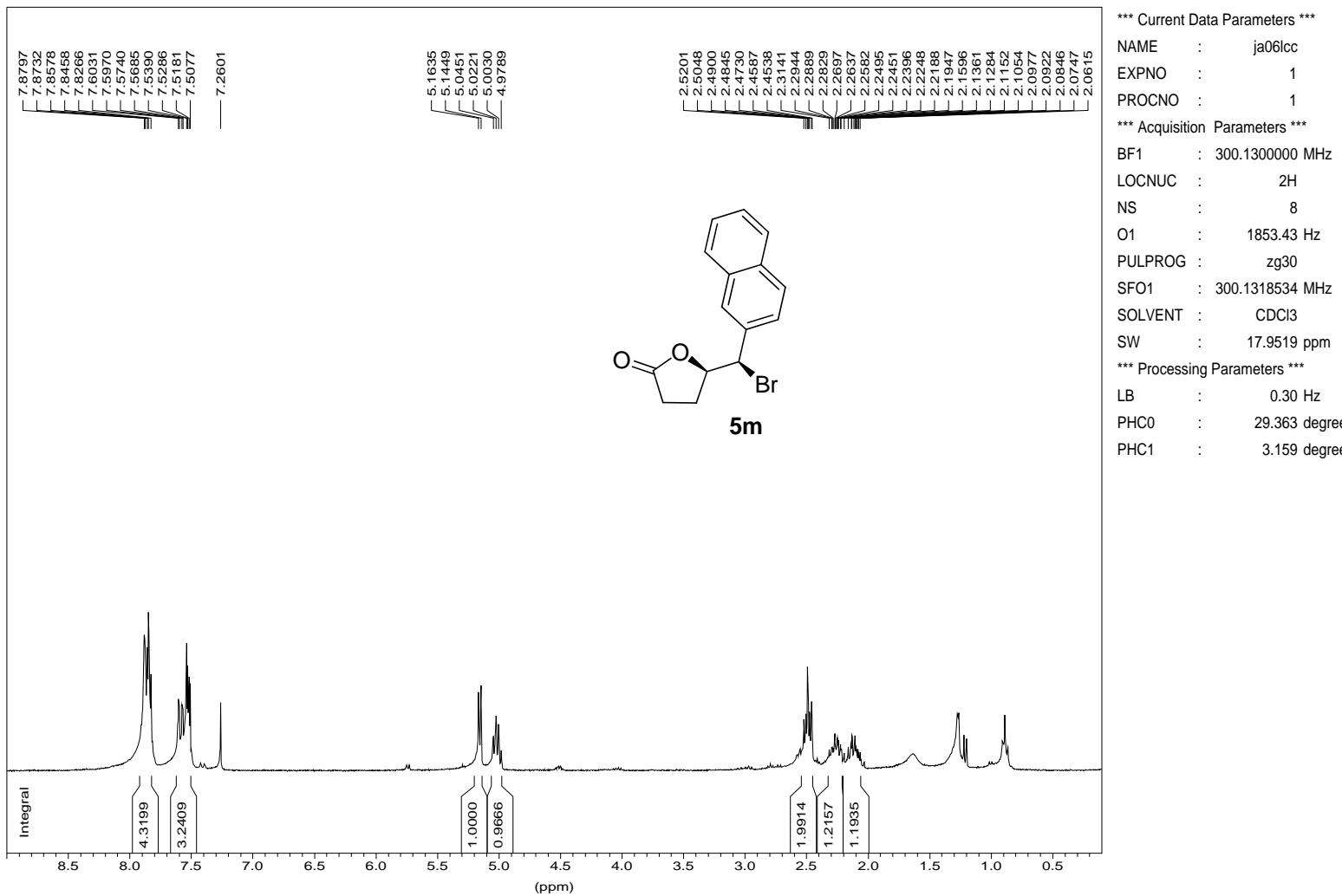
PHC0 : 274.235 degree

PHC1 : 1.980 degree

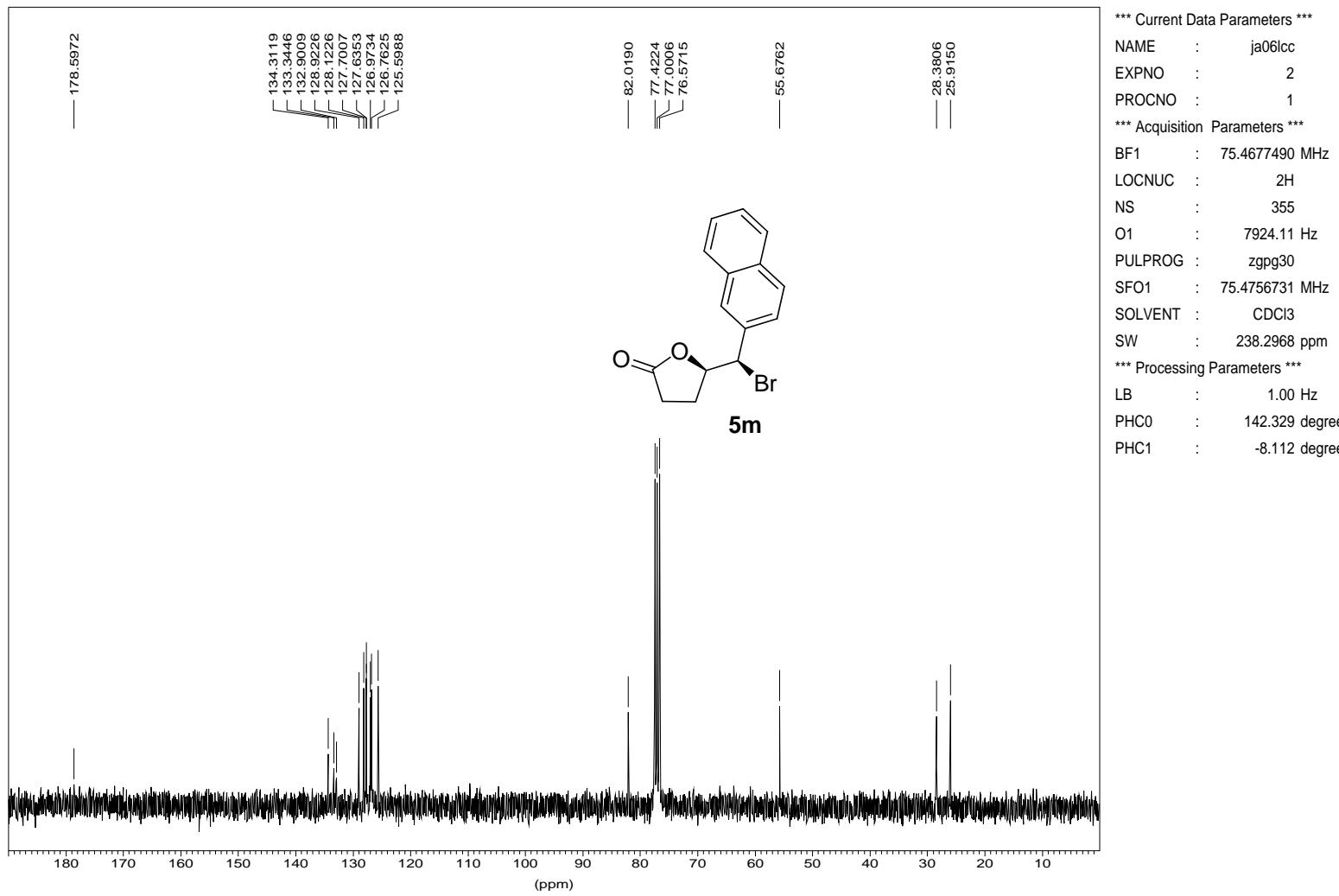
<sup>13</sup>C AMX500 cc-i-166b



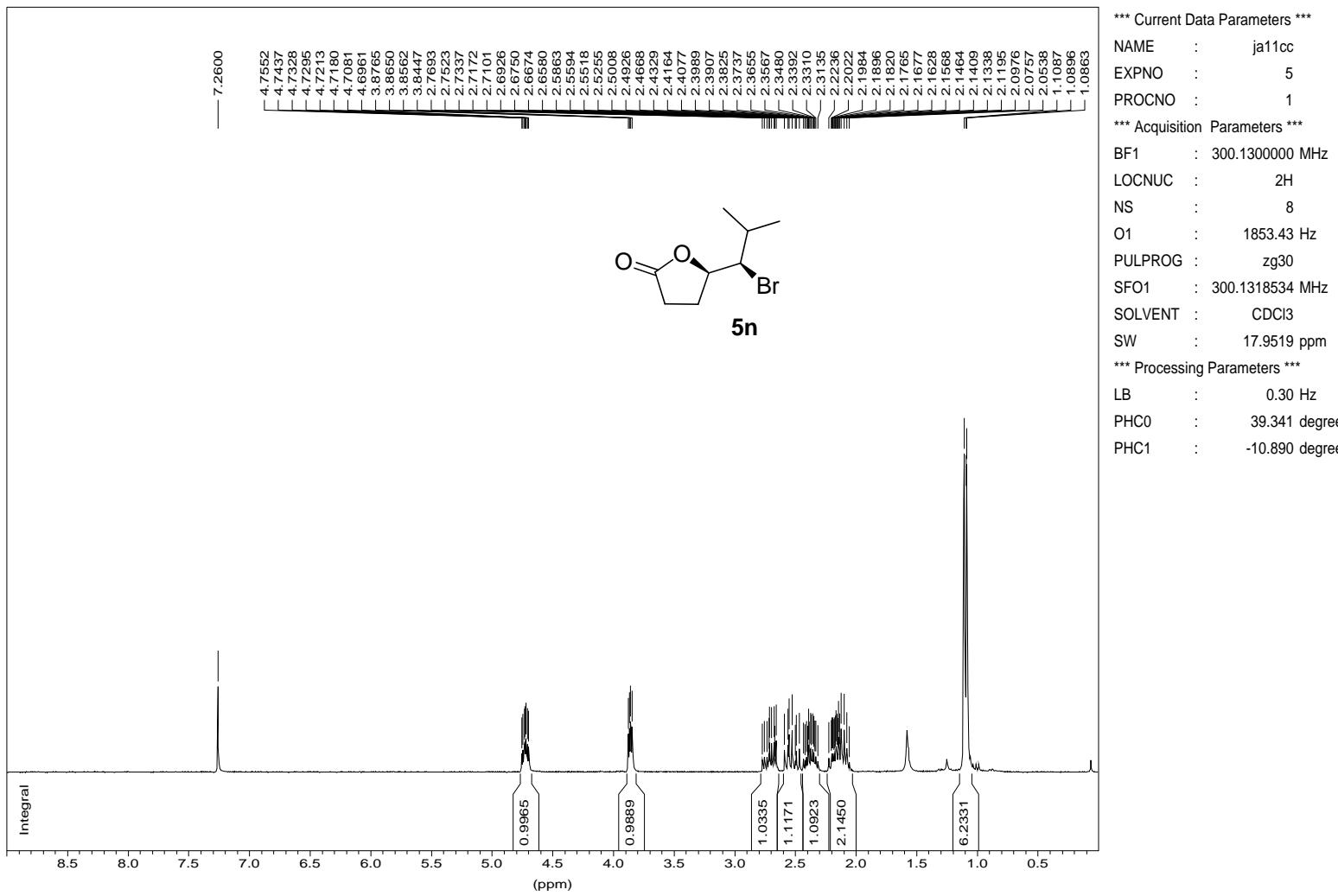
<sup>1</sup>H normal range AC300 cc-i-112-c



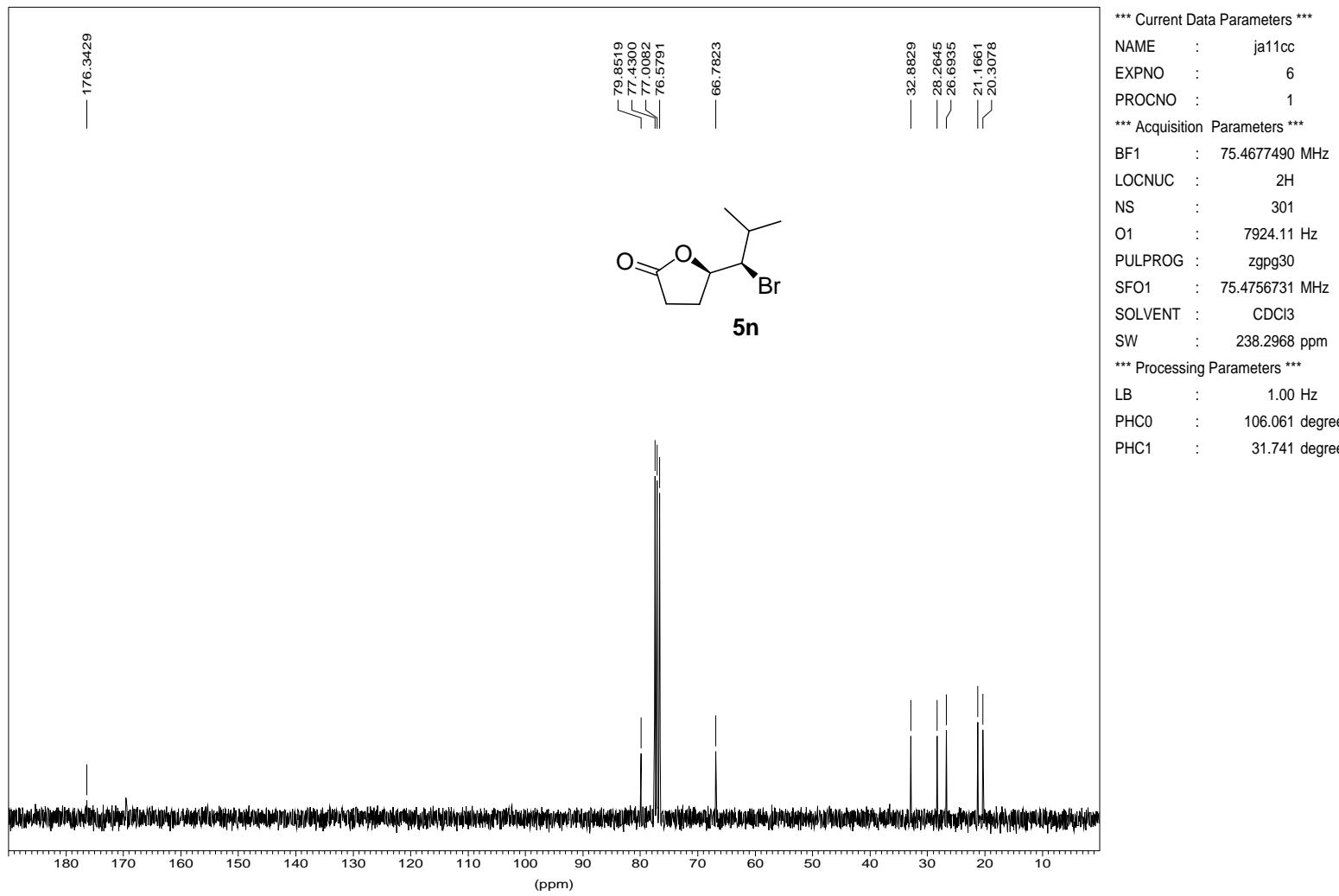
<sup>13</sup>C Standard AC300 cc-i-112-c



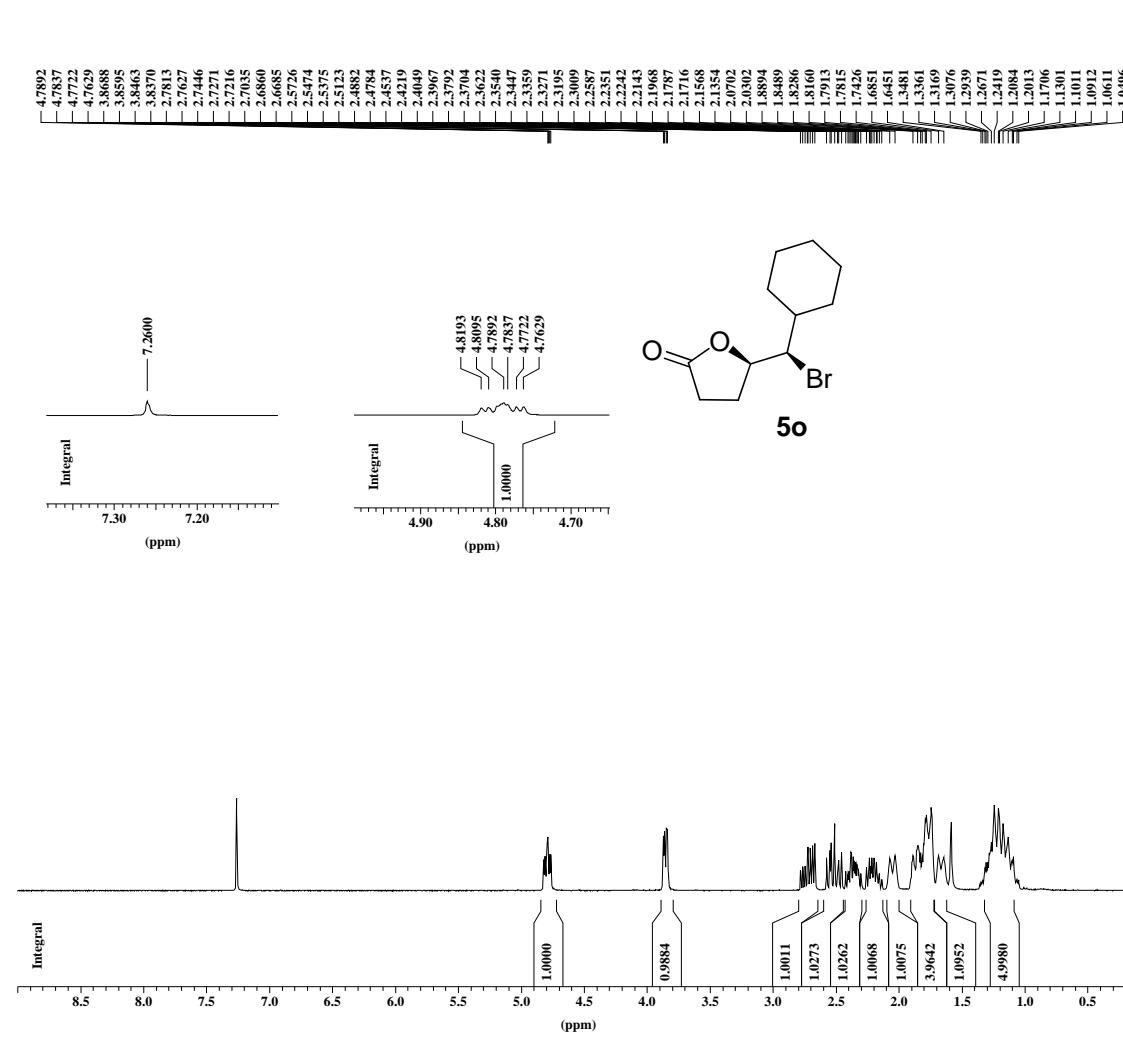
1H normal range AC300 cc-118-C



<sup>13</sup>C Standard AC300 cc-118-C



<sup>1</sup>H normal range AC300 cc-i-118-D



\*\*\* Current Data Parameters \*\*\*

NAME : ja1cc

EXPNO : 3

PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H

NS : 8

NUCLEUS : off

O1 : 1853.43 Hz

PULPROG : zg30

SFO1 : 300.1318534 MHz

SOLVENT : CDCl<sub>3</sub>

SW : 17.9519 ppm

TD : 32768

TE : 297.0 K

\*\*\* Processing Parameters \*\*\*

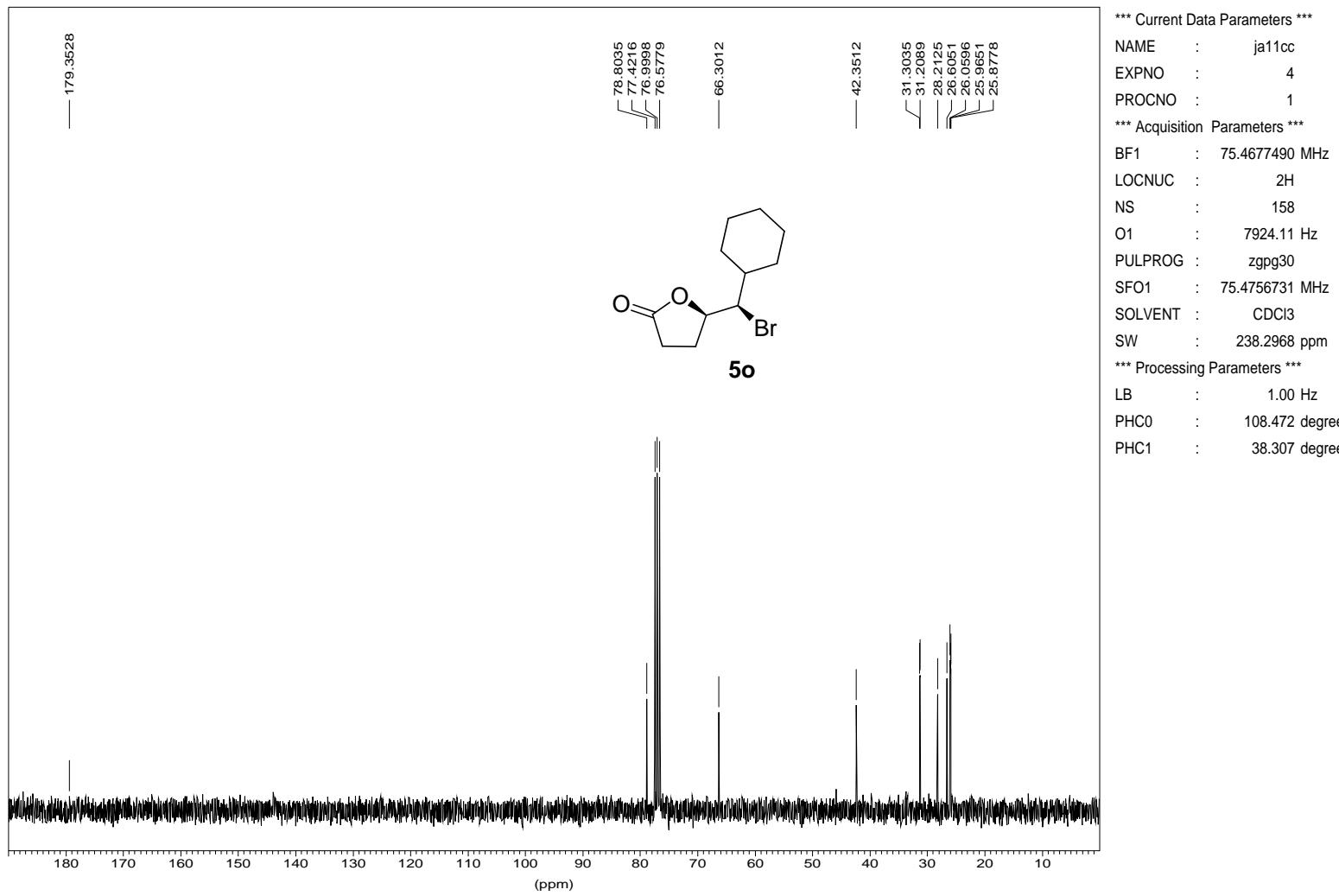
LB : 0.30 Hz

SF : 300.1300130 MHz

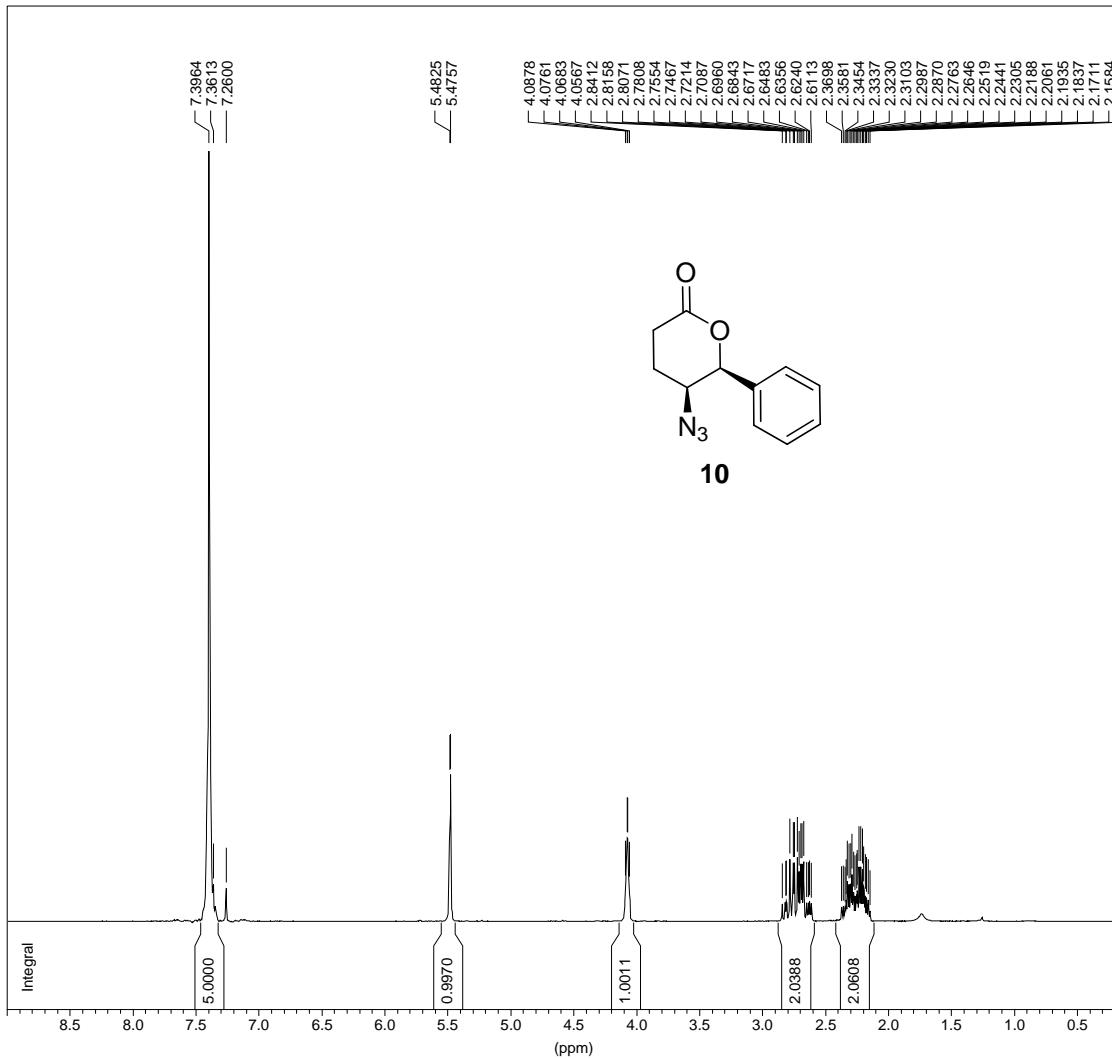
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C Standard AC300 cc-118-C



ck-ii-134



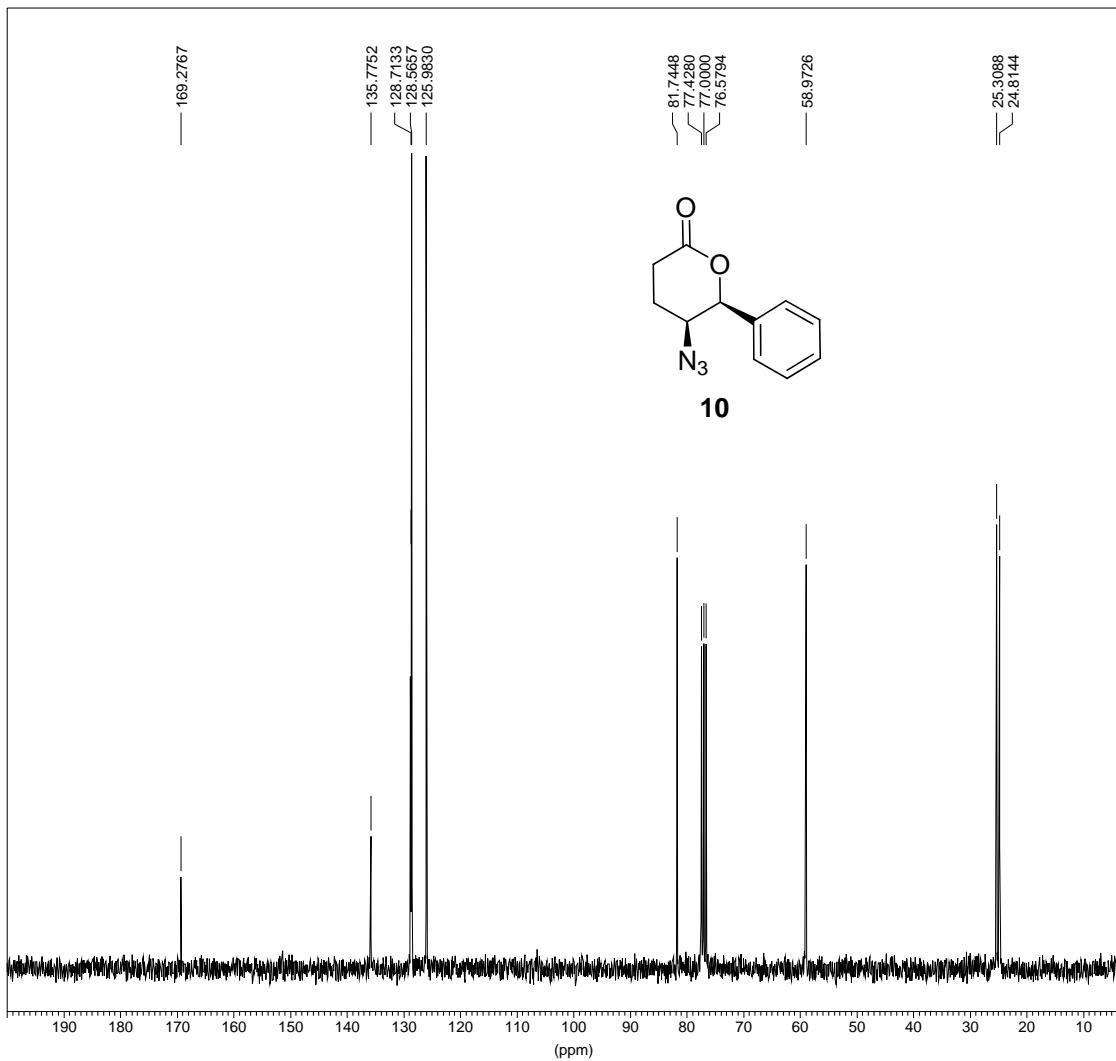
\*\*\* Current Data Parameters \*\*\*

NAME : se10ck  
EXPNO : 1  
PROCNO : 1  
INSTRUM : spect  
LOCMNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1250.00 Hz  
PULPROG : zg30  
SFO1 : 300.1312500 MHz  
SOLVENT : D2O  
SW : 15.9573 ppm  
TD : 32768  
TE : 299.2 K

\*\*\* Processing Parameters \*\*\*

LB : 0.10 Hz  
\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

ck-ii-134



\*\*\* Current Data Parameters \*\*\*

NAME : se10ck  
EXPNO : 5  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
INSTRUM : spect  
LOCMUC : 2H  
NS : 52  
NUCLEUS : off  
O1 : 8365.03 Hz  
PULPROG : zgdc  
SFO1 : 75.4760840 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 241.7743 ppm  
TD : 32768  
TE : 299.2 K

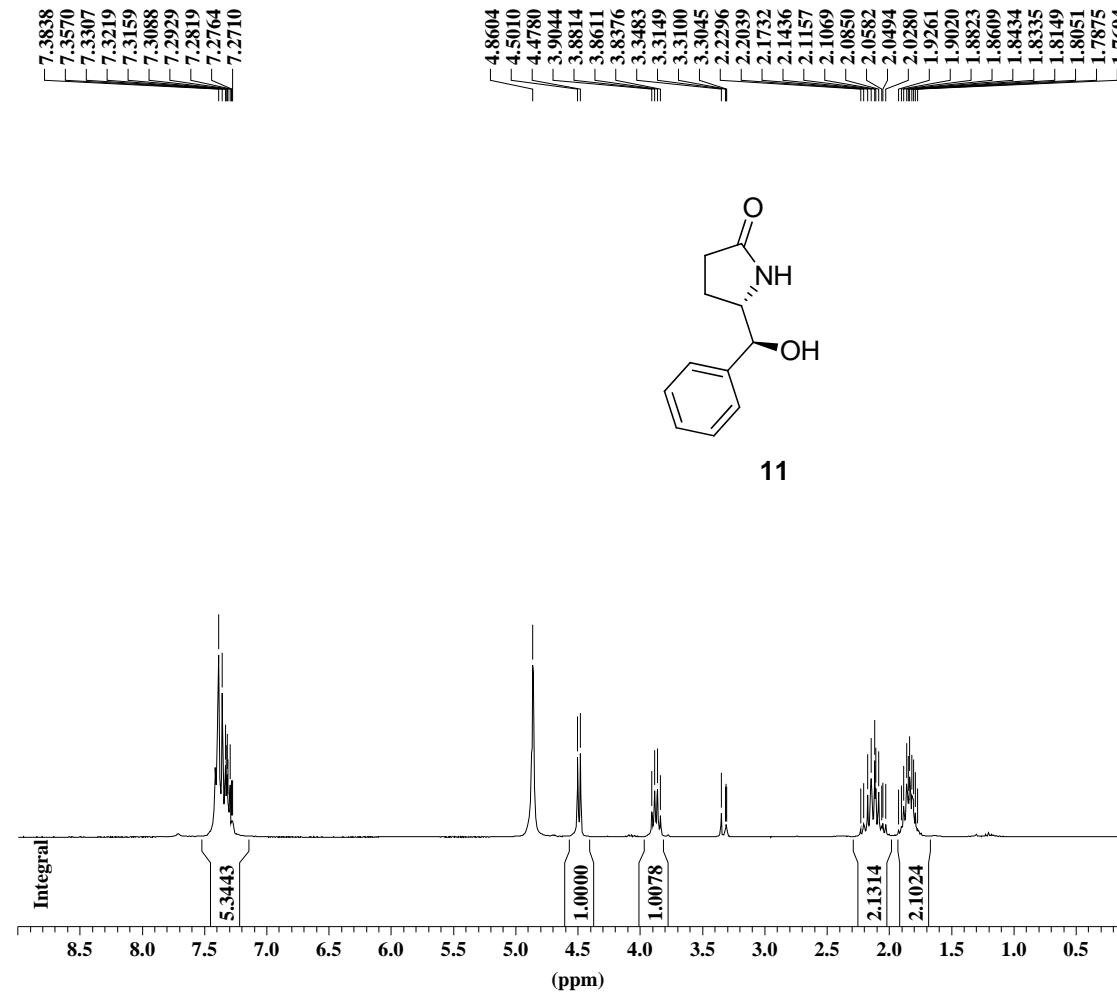
\*\*\* Processing Parameters \*\*\*

LB : 2.00 Hz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>1</sup>H normal range AC300 ck-ii-215



\*\*\* Current Data Parameters \*\*\*

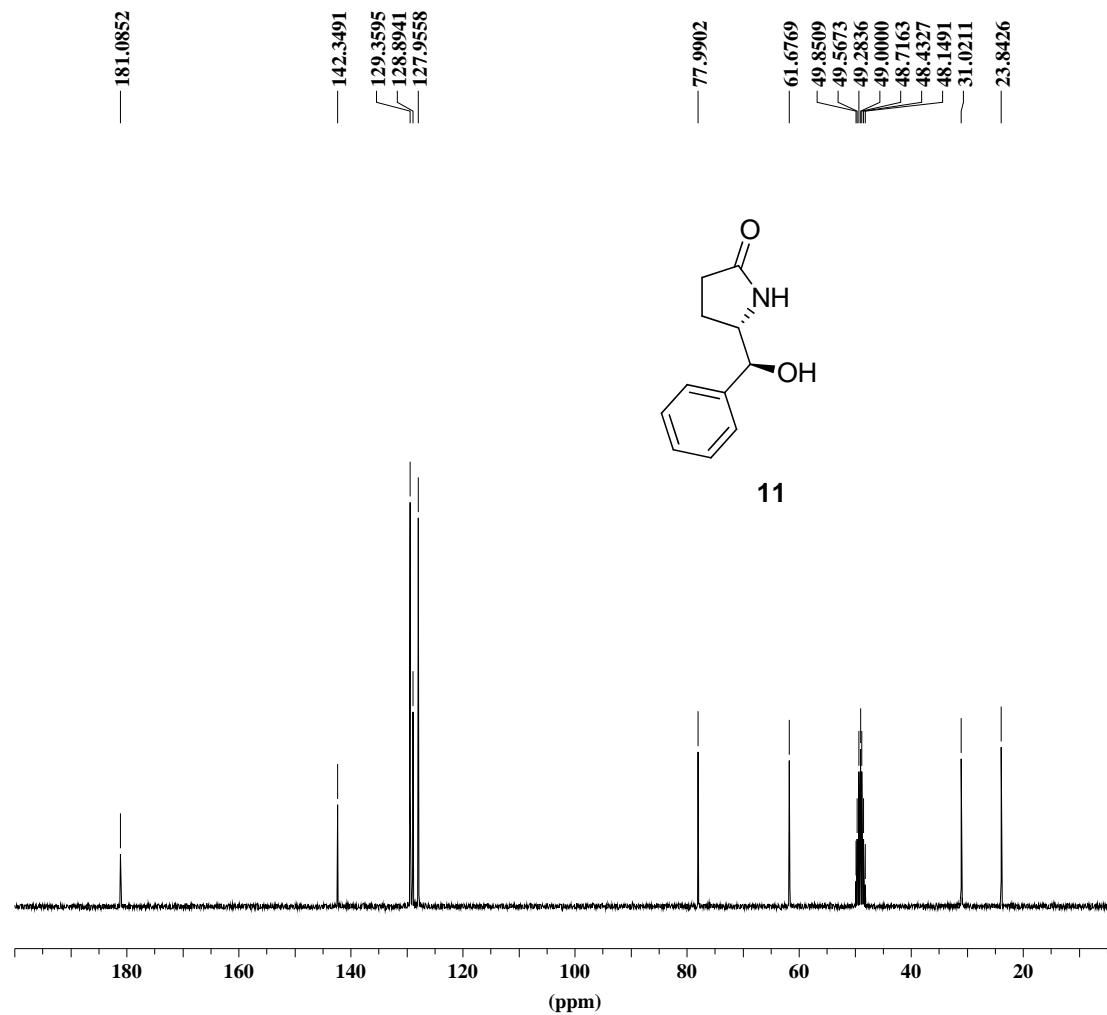
NAME : oc23ck  
EXPNO : 1  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : MeOD  
SW : 17.9519 ppm  
TD : 32768  
TE : 297.4 K

\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 300.1300050 MHz  
\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>13</sup>C Standard AC300 ck-ii-215



\*\*\* Current Data Parameters \*\*\*

NAME : oc23ck  
EXPNO : 2  
PROCNO : 1  
LOCMNUC : 2H  
NS : 202  
NUCLEUS : off  
O1 : 7924.11 Hz  
PULPROG : zgpg30  
SFO1 : 75.4756731 MHz  
SOLVENT : MeOD  
SW : 238.2968 ppm  
TD : 32768  
TE : 297.5 K

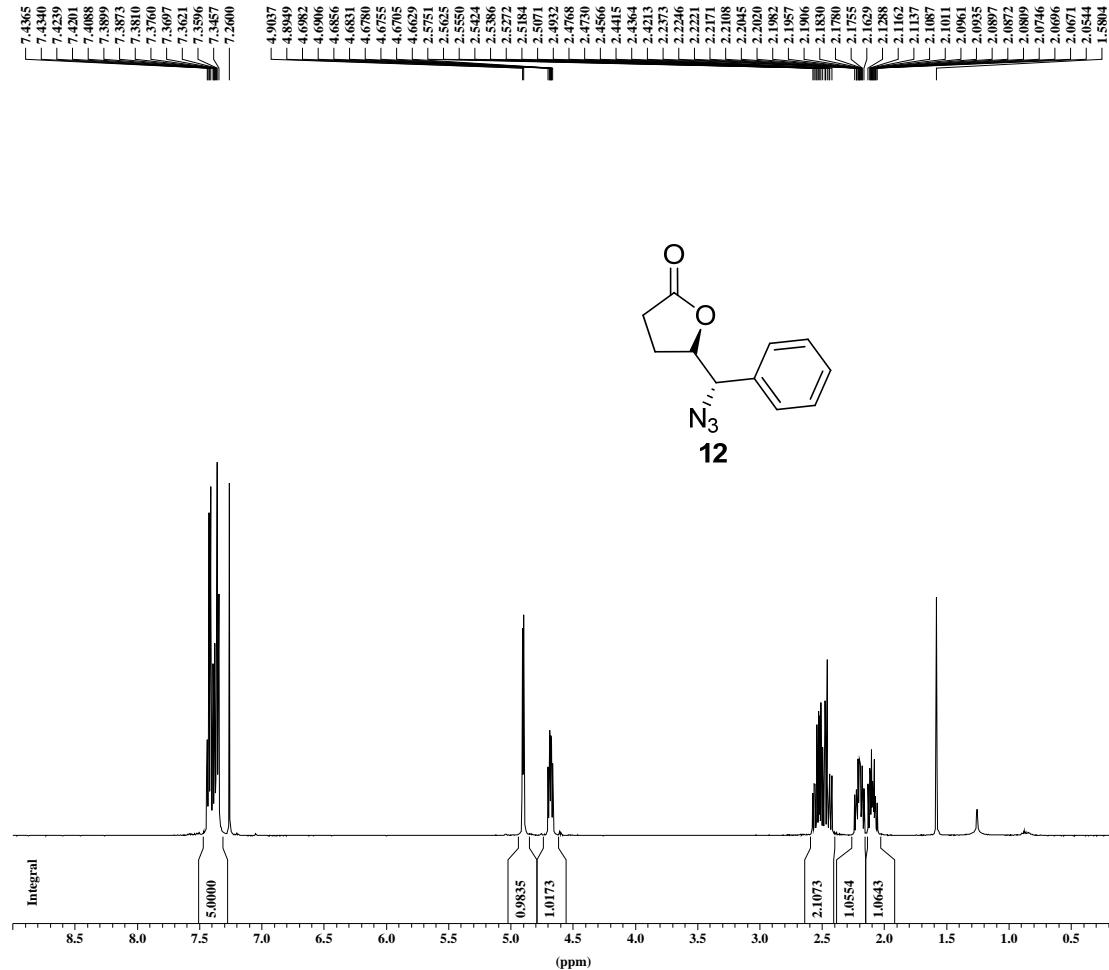
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 75.4676520 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>1</sup>H AMX500 cc-i-azide



\*\*\* Current Data Parameters \*\*\*

NAME : ck0209  
EXPNO : 1  
PROCNO : 1

\*\*\* Acquisition Parameters \*\*\*

LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 294.7 K

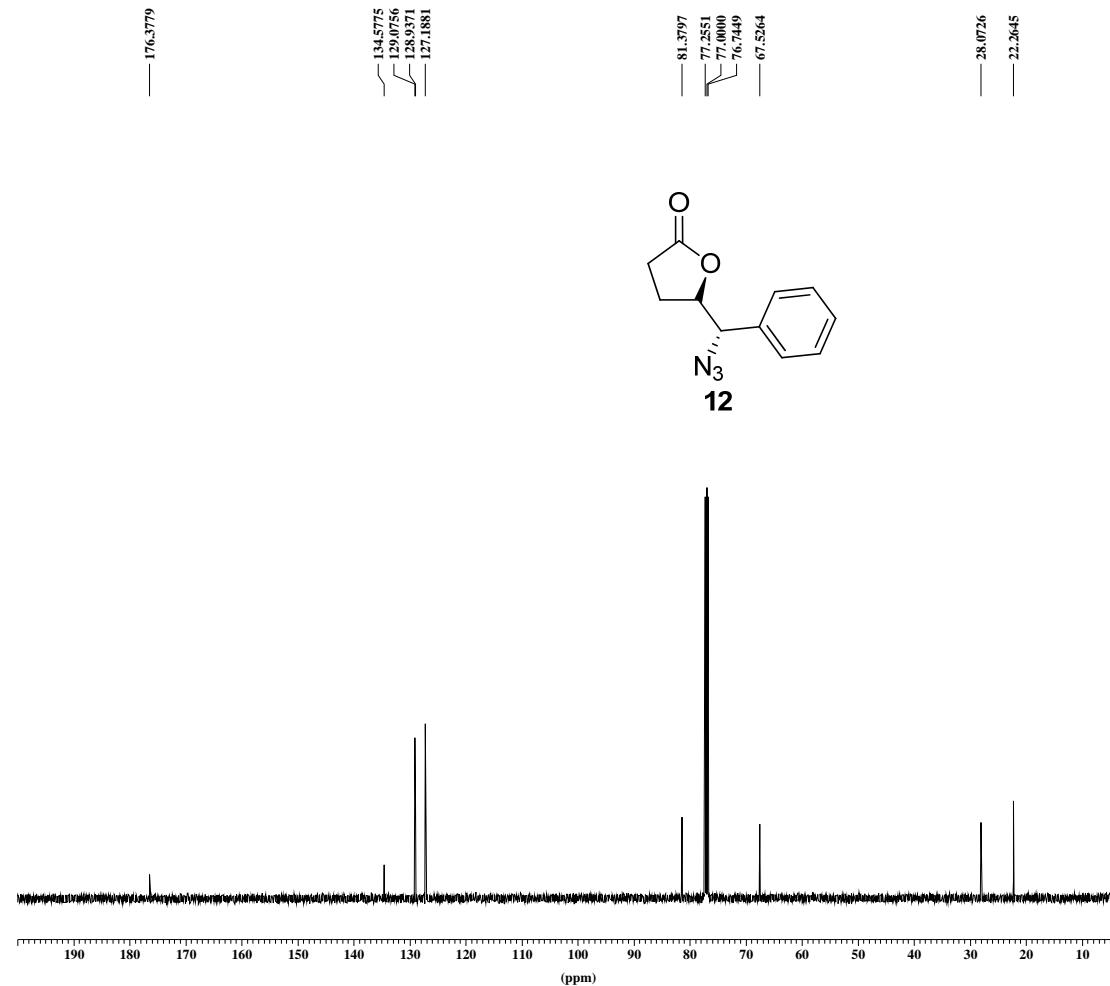
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300140 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

13C AMX500 cc-i-azide



\*\*\* Current Data Parameters \*\*\*

NAME : ck0209  
EXPNO : 2  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 201  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : CDCl3  
SW : 238.7675 ppm  
TD : 65536  
TE : 295.6 K

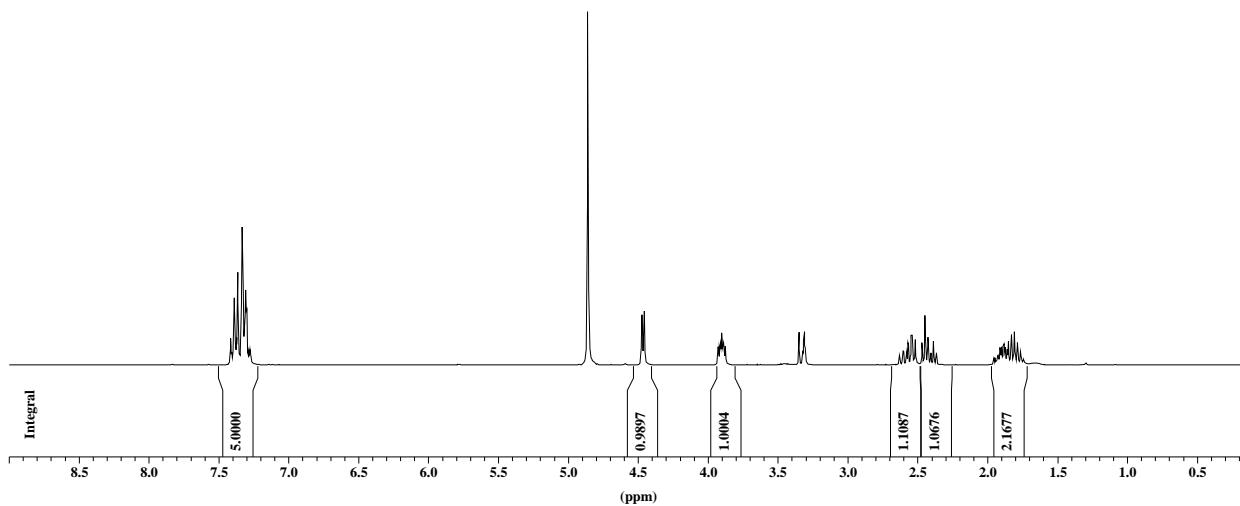
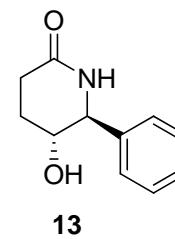
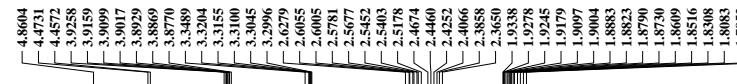
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 125.7577940 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

<sup>1</sup>H normal range AC300 ck-v-93



\*\*\* Current Data Parameters \*\*\*

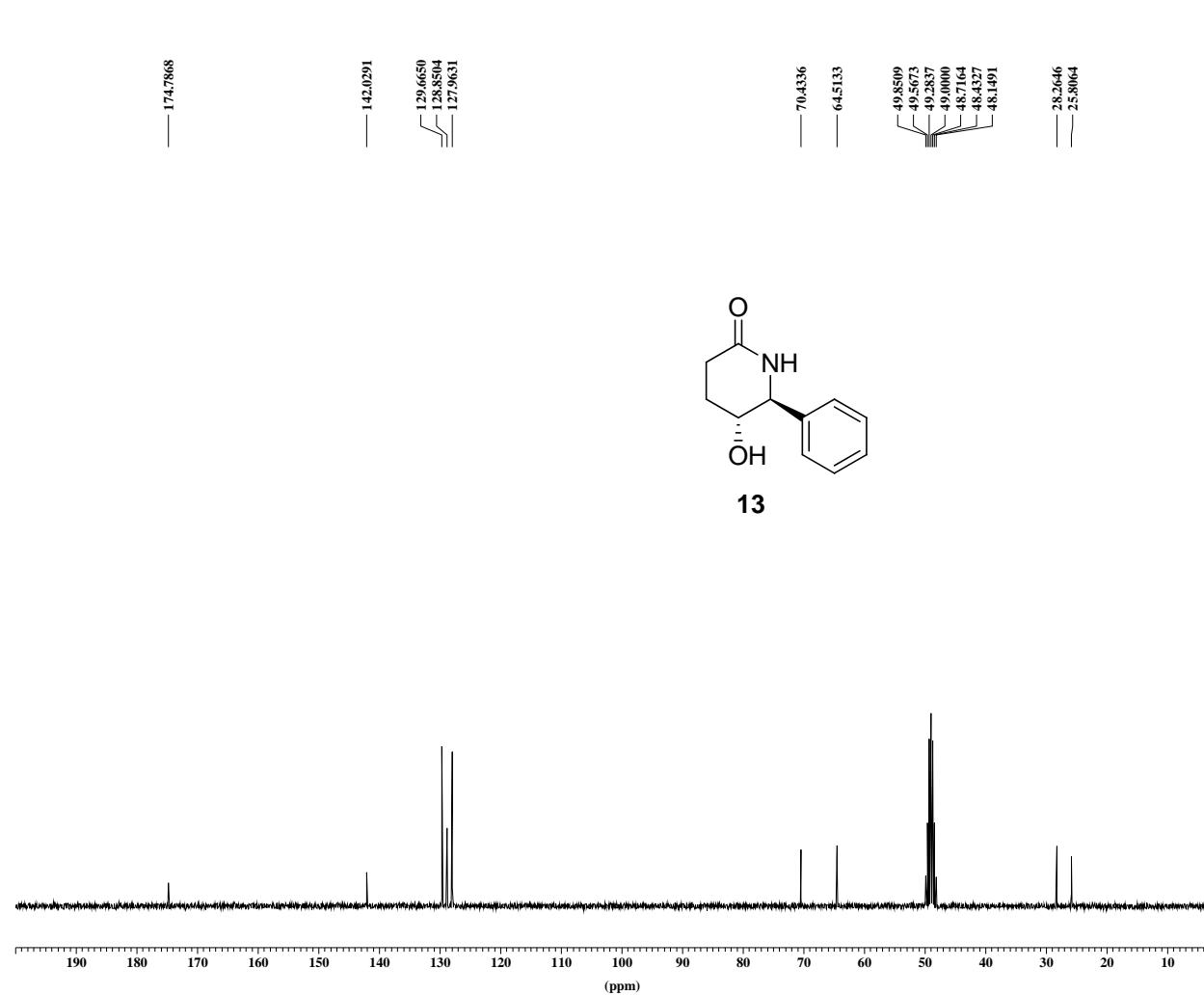
NAME : ja12ek  
EXPNO : 3  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 1853.43 Hz  
PULPROG : zg30  
SFO1 : 300.1318534 MHz  
SOLVENT : MeOD  
SW : 17.9519 ppm  
TD : 32768  
TE : 296.9 K

\*\*\* Processing Parameters \*\*\*

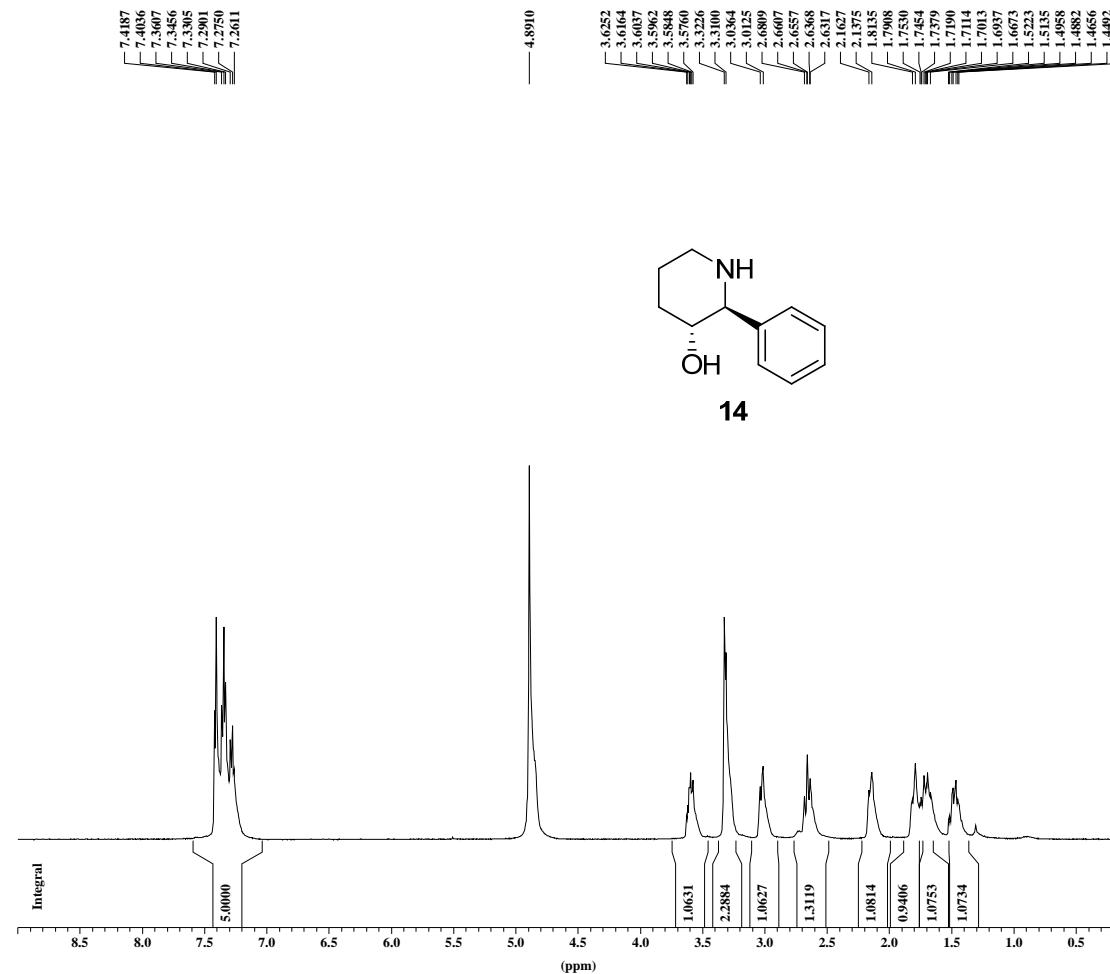
LB : 0.30 Hz  
SF : 300.1300050 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*  
NUCLEUS : off

<sup>13</sup>C Standard AC300 ck-v-93



<sup>1</sup>H AMX500 cc-i-159



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\*\*\* Current Data Parameters \*\*\*

NAME : ck0208  
EXPNO : 1  
PROCNO : 1  
LOCMNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 295.2 K

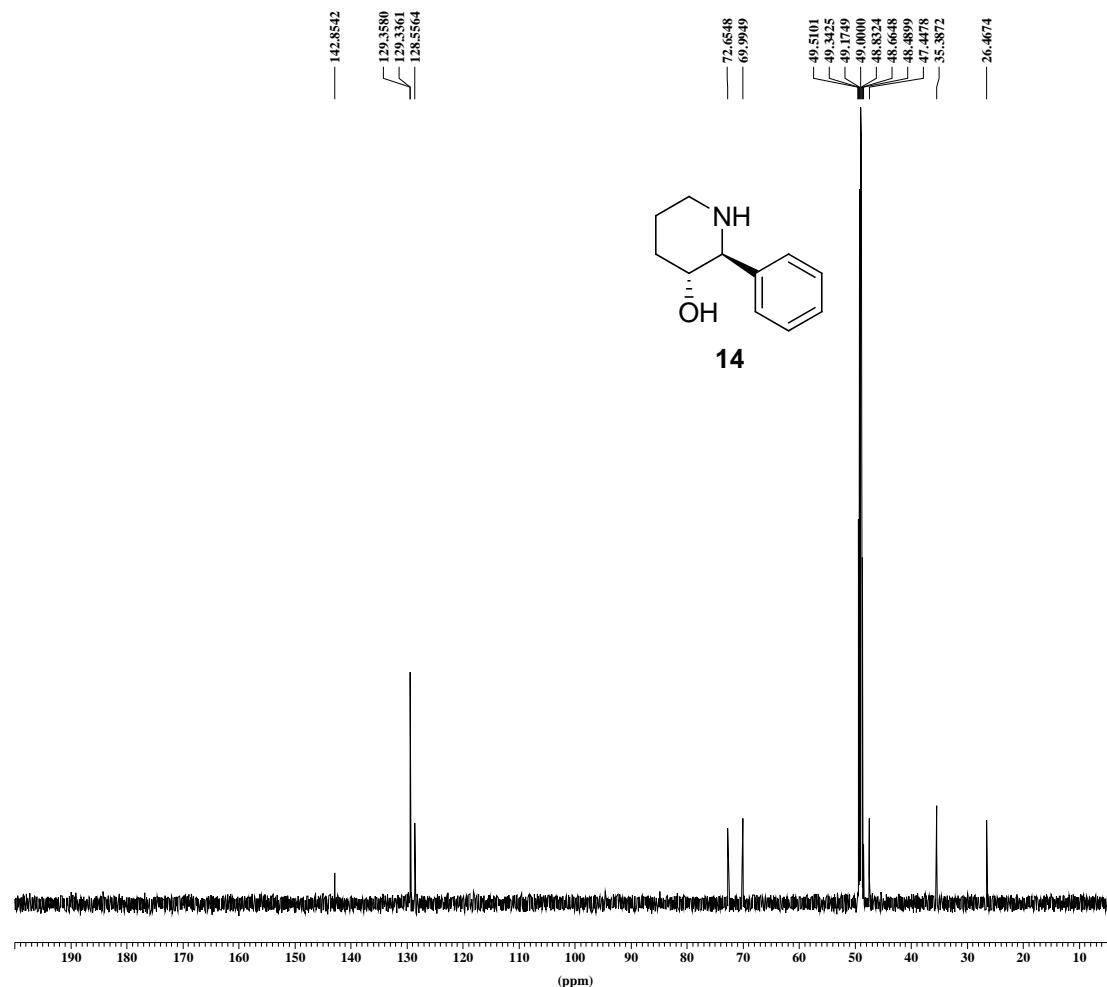
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300060 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

13C AMX500 cc-i-159



\*\*\* Current Data Parameters \*\*\*

NAME : ck0208  
EXPNO : 2  
PROCNO : 1  
LOCMUC : 2H  
NS : 165  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : MeOD  
SW : 238.7675 ppm  
TD : 65536  
TE : 295.9 K

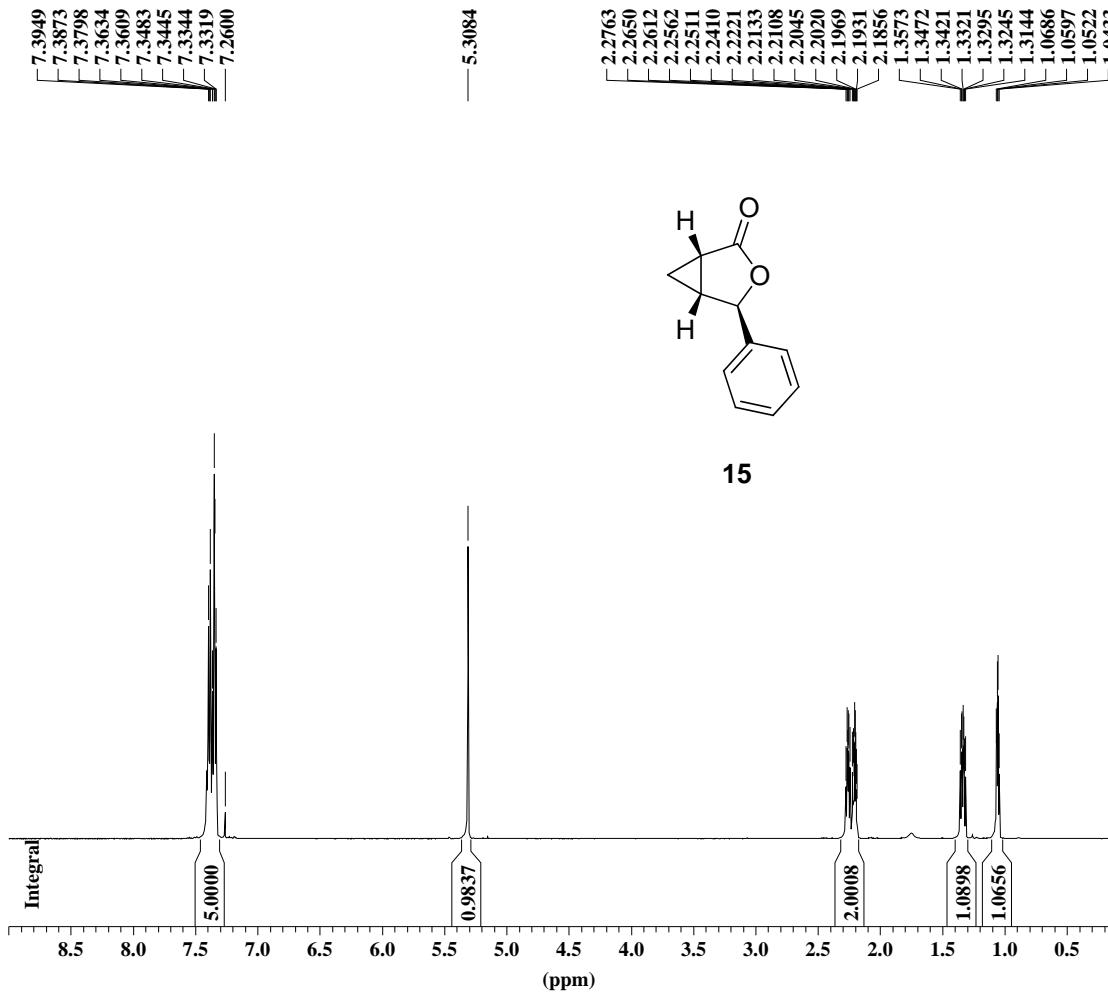
\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 125.7576130 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

1H AMX500 ck-ii-019



\*\*\* Current Data Parameters \*\*\*

NAME : ck0208  
EXPNO : 1  
PROCNO : 1  
LOCMNUC : 2H  
NS : 8  
NUCLEUS : off  
O1 : 3088.51 Hz  
PULPROG : zg30  
SFO1 : 500.1330885 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 20.6557 ppm  
TD : 32768  
TE : 303.1 K

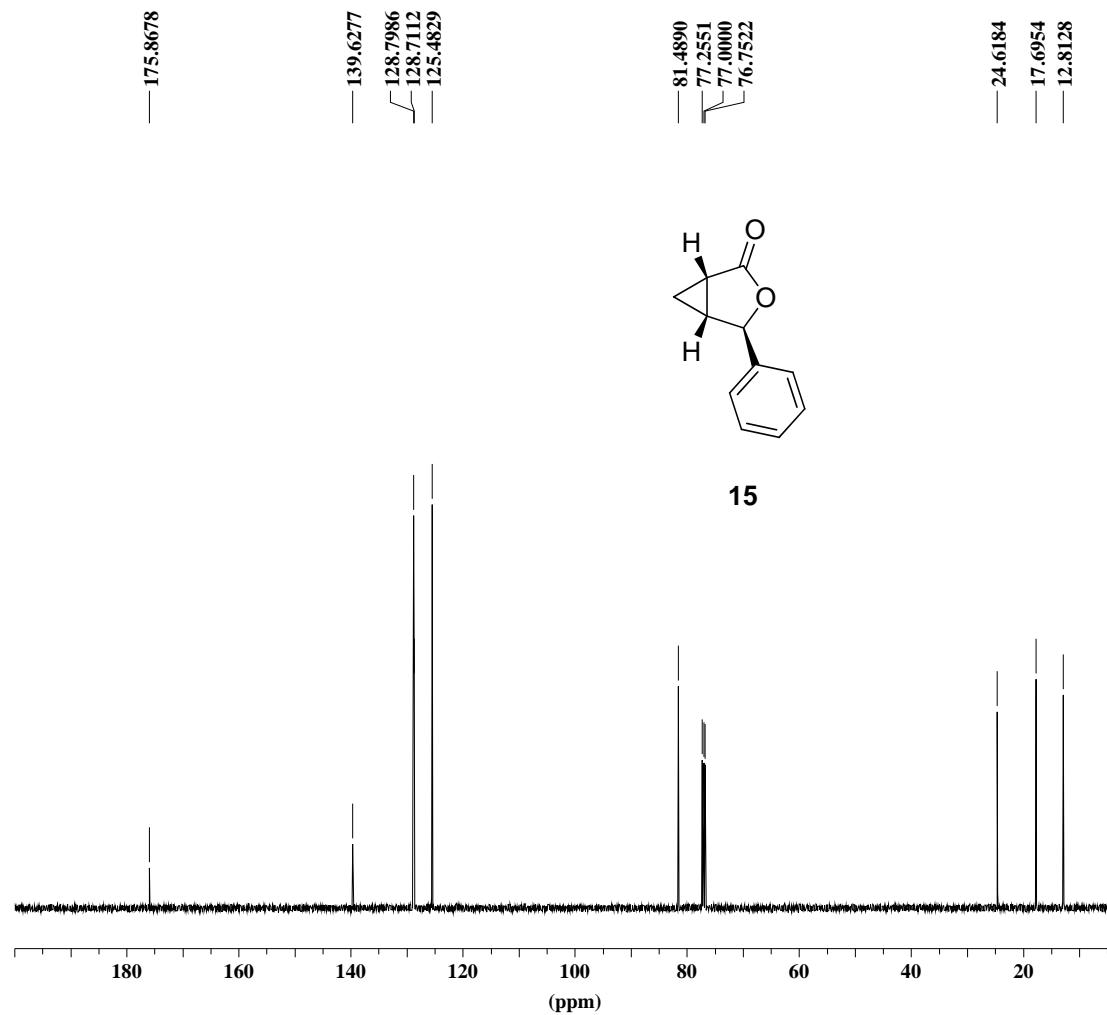
\*\*\* Processing Parameters \*\*\*

LB : 0.30 Hz  
SF : 500.1300120 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

13C AMX500 ck-iii-019



\*\*\* Current Data Parameters \*\*\*

NAME : ck0208  
EXPNO : 2  
PROCNO : 1  
\*\*\* Acquisition Parameters \*\*\*  
LOCNUC : 2H  
NS : 55  
NUCLEUS : off  
O1 : 13204.57 Hz  
PULPROG : zgpg30  
SFO1 : 125.7709936 MHz  
SOLVENT : CDCl<sub>3</sub>  
SW : 238.7675 ppm  
TD : 65536  
TE : 301.0 K

\*\*\* Processing Parameters \*\*\*

LB : 1.00 Hz  
SF : 125.7578040 MHz

\*\*\* 1D NMR Plot Parameters \*\*\*

NUCLEUS : off

X-ray Structure of **5a**

