Enantioselective Organocatalytic Reductive Amination of Aliphatic Ketones by Benzothiazoline as Hydrogen Donor

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General. NMR spectra were recorded on Unity Inova-400 instrument (Varian Inc., 400 MHz for ¹H, 100 MHz for ¹³C) using CDCl₃ as a solvent. Tetramethylsilane (TMS) ($\delta = 0$) or CHCl₃ ($\delta = 7.27$) served as an internal standard for ¹H NMR, and CDCl₃ was used as an internal standard ($\delta = 77.0$) for ¹³C NMR. Optical rotations were measured on a HORIBA SEPA-500 polarimeter. Purification of the products was performed by preparative TLC on silica gel (Wako gel B-5F). All solvents were purified according to the standard procedures. Benzothiazoline **1a** was synthesized according to our previous report.¹

1. General procedure for the chiral phosphoric acid catalyzed enantioselective reductive amination using benzothiazoline

A typical procedure for the preparation of 3a is described for the reaction of 4-phenyl-2-butanone with *p*-anisidine:

A magnetic stirrer bar and powdered molecular sieves 5A (MS 5A) (100 mg) were placed in a test tube under nitrogen atmosphere. The MS 5A were then dried with a heat gun under reduced pressure and the test tube was refilled with nitrogen. p-Anisidine

(12.3 mg, 0.100 mmol), phosphoric acid **2g** (3.60 mg, 5 μ mol), and benzothiazoline **1a** (30.0 mg, 0.140 mmol) were added to the test tube successively under nitrogen atmosphere at room temperature. 4-Phenyl-2-butanone (15.0 μ L, 0.100 mmol) was added to the test tube using benzene (2 mL) at 20 °C. After being stirred for 2 days at the same temperature, the mixture was filtered and the filtrate was concentrated under reduced pressure, and the residue was purified by preparative thin layer chromatography on silica gel (AcOEt / hexane = 1 / 10) to give 22.1 mg (0.0870 mmol, 87%) of **3a** as pale yellow oil.



 $3a (87\%, 97\% ee)^2$

¹H NMR (400 MHz, CDCl₃) δ : 1.19 (d, J = 6.4 Hz, 3 H), 1.67 – 1.79 (m, 1H), 1.81 – 1.92 (m, 1H), 2.72 (dd, J = 8.4, 8.4 Hz, 2 H), 3.39 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 3.73 (s, 3H), 6.48 – 6.53 (m, 2 H), 6.72 – 6.78 (m, 2 H), 7.15 – 7.21 (m, 3 H), 7.24 – 7.31 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 20.8, 32.5, 38.8, 48.9, 55.8, 114.7, 114.9, 125.8, 128.3, 128.4, 141.7, 142.0, 151.9.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 95/5, flow rate = 0.5 mL min⁻¹, major enantiomer: t_R =21.8 min; minor enantiomer: t_R = 24.4 min.

 $[\alpha]_D^{24}$ - 7.0 (c 2.3, CHCl₃) [lit., $[\alpha]_D^{27}$ -2.8 (c 1.2, CHCl₃)]²



3b $(97\%, 95\% \text{ ee})^2$

¹H NMR (400 MHz, CDCl₃) δ : 1.17 (d, J = 6.4 Hz, 3 H), 1.45 – 1.57 (m, 1H), 1.62 – 1.74 (m, 1H), 2.10 – 2.22 (m, 2 H), 3.40 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 3.74 (s, 3H), 4.97 (dd, J = 10.0, 1.2 Hz, 1 H), 5.02 (ddd, J = 16.4, 3.2, 1.6 Hz, 1 H), 5.75 – 5.88 (m, 1 H), 6.59 (d, J = 8.8 Hz, 2 H), 6.74 – 6.81 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 20.5, 30.4, 36.0, 49.5, 55.8, 114.8, 114.9, 115.2, 138.3, 141.1, 152.2.

HPLC conditions: Daicel CHIRALPAK[®] IE column, hexane/2-propanol = 20/1, flow rate = 0.5 mL min⁻¹, major enantiomer: $t_R = 11.1$ min; minor enantiomer: $t_R = 10.6$ min. $[\alpha]_D^{24} - 9.9$ (c 1.5, CHCl₃) [lit., $[\alpha]_D^{28} - 3.8$ (c 0.4, CHCl₃)]²



 $3c (64\%, 91\% ee)^2$

¹H NMR (400 MHz, CDCl₃) δ: 1.32 (d, *J* = 6.4 Hz, 3 H), 3.74 (s, 3H), 3.83 (ddq, *J* = 6.4, 5.8, 5.8 Hz, 1 H), 4.21 (dd, *J* = 10.8, 5.8 Hz, 1 H), 4.45 (dd, *J* = 10.8, 5.8 Hz, 1 H), 6.67 (d, *J* = 8.8 Hz, 2 H), 6.75 – 6.81 (m, 2 H), 7.43 (t, *J* = 7.6 Hz, 2 H), 7.53 – 7.59 (m, 1 H), 7.99 -8.04 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 18.2, 48.9, 55.7, 67.9, 115.0, 115.1, 128.4, 129.6, 130.0, 133.0, 140.9, 152.4, 166.5.

HPLC conditions: Daicel CHIRALPAK[®] IE column, hexane/2-propanol = 20/1, flow rate = 1.0 mL min⁻¹, major enantiomer: t_R =22.1 min; minor enantiomer: t_R = 19.0 min. $[\alpha]_D^{24}$ + 12.0 (c 1.8, CHCl₃) [lit., $[\alpha]_D^{26}$ +12.8 (c 0.2, CHCl₃)]²



3d $(80\%, 95\% \text{ ee})^2$

¹H NMR (400 MHz, CDCl₃) δ : 0.89 (t, J = 6.8 Hz, 3 H), 1.15 (d, J = 6.4 Hz, 3 H), 1.24 – 1.45 (m, 9 H), 1.51 – 1.62 (m, 1H), 3.36 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 3.75 (s, 3H), 6.54 – 6.60 (m, 2 H), 6.75 – 6.81 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 14.1, 20.7, 22.6, 26.1, 29.4, 31.8, 37.2, 49.6, 55.8, 114.8, 114.9, 141.7, 151.8.

HPLC conditions: Daicel CHIRALPAK[®] IE column, hexane/2-propanol = 20/1, flow rate = 0.5 mL min⁻¹, major enantiomer: $t_R = 10.6$ min; minor enantiomer: $t_R = 9.9$ min. $[\alpha]_D^{24} - 6.9$ (c 1.8, CHCl₃) [lit., $[\alpha]_D^{27} - 8.0$ (c 1.1, CHCl₃)]²

 $3e(77\%, 94\% ee)^3$

¹H NMR (400 MHz, CDCl₃) δ : 0.89 (t, J = 6.8 Hz, 3 H), 1.18 (d, J = 6.4 Hz, 3 H), 1.24 – 1.48 (m, 7 H), 1.50 – 1.63 (m, 1H), 3.36 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 6.55 – 6.61 (m, 2 H), 6.63 – 6.69 (m, 1 H), 7.13 – 7.20 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 14.0, 20.7, 22.6, 25.8, 31.9, 37.1, 48.5, 113.1, 116.8, 129.2, 147.6.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 99/1, flow rate = 0.5 mL min⁻¹, major enantiomer: $t_R = 11.1$ min; minor enantiomer: $t_R = 12.1$ min.

 $[\alpha]_D^{24}$ - 17.2 (c 1.4, CHCl₃).



 $3f(99\%, 86\% ee)^2$

¹H NMR (400 MHz, CDCl₃) δ : 0.94 (t, J = 7.6 Hz, 3 H), 1.15 (d, J = 6.4 Hz, 3 H), 1.38 – 1.50 (m, 1 H), 1.54 – 1.66 (m, 1H), 3.31 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 3.74 (s, 3H), 6.55 – 6.61 (m, 2 H), 6.74 – 6.80 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 10.3, 20.1, 29.5, 51.0, 55.8, 114.89, 114.94, 141.6, 151.9.

HPLC conditions: Daicel CHIRALCEL[®] AS-H column, hexane/2-propanol = 95/5, flow rate = 1.0 mL min⁻¹, major enantiomer: $t_R = 10.5$ min; minor enantiomer: $t_R = 8.2$ min. $[\alpha]_D^{24} - 17.9$ (c 1.6, CHCl₃) [lit., $[\alpha]_D^{28} - 25.2$ (c 0.2, CHCl₃)]⁵



 $3g (98\%, 97\% ee)^3$

¹H NMR (400 MHz, CDCl₃) δ : 1.18 (d, J = 6.4 Hz, 3 H), 1.63 – 1.74 (m, 1 H), 1.75 – 1.87 (m, 1H), 2.64 (dd, J = 7.8, 7.8 Hz, 2 H), 3.38 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 3.74 (s, 3H), 5.91 (s, 2 H), 6.48 – 6.55 (m, 2 H), 6.59 – 6.64 (m, 1 H), 6.65 – 6.68 (m, 1 H), 6.72 (d, J = 8.0 Hz, 1 H)), 6.72 – 6.79 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 20.8, 32.2, 39.0, 48.9, 55.8, 100.7, 108.1, 108.8, 114.8, 114.9, 121.1, 135.9, 141.6, 145.6, 147.5, 151.9.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 98/2, flow rate = 1.0 mL min⁻¹, major enantiomer: t_R =32.9 min; minor enantiomer: t_R = 26.9 min.

 $[\alpha]_{D}^{24}$ - 16.2 (c 1.9, CHCl₃).



3h $(85\%, 97\% \text{ ee})^3$

¹H NMR (400 MHz, CDCl₃) δ : 1.20 (d, J = 6.4 Hz, 3 H), 1.66 – 1.77 (m, 1 H), 1.77 – 1.88 (m, 1H), 2.64 (dd, J = 7.8, 7.8 Hz, 2 H), 3.47 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 5.91 (s, 2 H), 6.51 – 6.57 (m, 2 H), 6.59 – 6.64 (m, 1 H), 6.64 – 6.70 (m, 2 H), 6.72 (d, J = 8.0 Hz, 1 H), 7.11 – 7.18 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 20.8, 32.2, 39.0, 47.9, 100.7, 108.1, 108.8, 113.3, 117.1, 121.1, 129.3, 135.8, 145.6, 147.4, 147.5.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 98/2, flow rate = 1.0 mL min⁻¹, major enantiomer: t_R =22.2 min; minor enantiomer: t_R = 20.2 min.

 $[\alpha]_D^{24}$ - 11.4 (c 2.2, CHCl₃).



3h $(86\%, 96\% \text{ ee})^3$

¹H NMR (400 MHz, CDCl₃) δ: 1.18 (d, *J* = 6.4 Hz, 3 H), 1.64 – 1.86 (m, 2 H), 2.62 (dd, *J* = 7.8, 7.8 Hz, 2 H), 3.40 (ddq, J = 6.4, 6.4, 6.4 Hz, 1 H), 5.91 (s, 2 H), 6.36 – 6.42 (m, 2 H), 6.57 – 6.62 (m, 1 H), 6.63 – 6.66 (m, 1 H), 6.71 (d, *J* = 7.6 Hz, 1 H), 7.18 – 7.24 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 20.6, 32.1, 38.8, 47.8, 100.7, 108.1, 108.4, 108.8, 114.7, 121.1, 131.9, 135.5, 145.6, 146.4, 147.6.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 99/1, flow rate = 1.0 mL min⁻¹, major enantiomer: t_R =47.3 min; minor enantiomer: t_R = 42.7 min.

 $[\alpha]_D^{24}$ - 16.8 (c 2.7, CHCl₃).



3h $(82\%, 97\% \text{ ee})^2$

¹H NMR (400 MHz, CDCl₃) δ : 1.08 (d, J = 6.4 Hz, 3 H), 0.97 – 1.32 (m, 5 H), 1.38 – 1.49 (m, 1 H), 1.62 – 1.85 (m, 5 H), 3.22 (dq, J = 6.4, 6.0 Hz, 1 H), 3.74 (s, 3 H), 6.54 (d, J = 8.8 Hz, 2 H), 6.76 (d, J = 9.2 Hz, 1 H).

¹³C NMR (100 MHz, CDCl₃) δ: 17.3, 26.3, 26.5, 26.6, 28.2, 29.8, 42.8, 54.1, 55.8, 114.5, 114.9, 142.2, 151.6.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 99/1, flow rate = 0.5 mL min⁻¹, major enantiomer: t_R =13.3 min; minor enantiomer: t_R = 14.8 min.

 $[\alpha]_{D}^{26}$ - 6.9 (c 1.5, CHCl₃) [lit., $[\alpha]_{D}^{28}$ -4.3 (c 1.1, CHCl₃)]²



 $3i (55\%, 97\% ee)^3$

¹H NMR (400 MHz, CDCl₃) δ: 0.90 (d, *J* = 6.4 Hz, 3 H), 0.93 (d, *J* = 6.4 Hz, 3 H), 1.13 (d, *J* = 6.4 Hz, 3 H), 1.18 – 1.27 (m, 1 H), 1.41 – 1.50 (m, 1 H), 1.75 (dq, *J* = 6.8, 6.4 Hz, 1 H), 3.43 (ddq, *J* = 6.4, 6.4, 6.4 Hz, 1 H), 3.74 (s, 3 H), 6.53 – 6.59 (m, 2 H), 6.74 – 6.79 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 21.0, 22.5, 23.0, 25.1, 46.9, 47.6, 55.8, 114.7, 114.9, 141.8, 151.8.

HPLC conditions: Daicel CHIRALCEL[®] AS-H column, hexane/2-propanol = 99/1, flow rate = 0.3 mL min⁻¹, major enantiomer: t_R =21.8 min; minor enantiomer: t_R = 23.2 min. $[\alpha]_D^{24}$ - 12.7 (c 1.1, CHCl₃).



3j $(35\%, 68\% \text{ ee})^3$

¹H NMR (400 MHz, CDCl₃) δ : 0.89 (t, J = 7.6 Hz, 3 H), 0.91 (t, J = 7.6 Hz, 3 H), 1.24 – 1.62 (m, 8 H), 3.10 (brs, 1 H), 3.18 (dddd, J = 6.0, 6.0, 6.0, 6.0 Hz, 1 H), 3.73 (s, 3 H), 6.51 – 6.57 (m, 2 H), 6.73 – 6.79 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 10.0, 14.1, 22.8, 27.1, 28.1, 34.0, 55.0, 55.8, 114.3, 114.9, 142.4, 151.5.

HPLC conditions: Daicel CHIRALCEL[®] AS-H column, hexane/2-propanol = 99/1, flow rate = 0.5 mL min⁻¹, major enantiomer: $t_R = 11.4$ min; minor enantiomer: $t_R = 10.3$ min. $[\alpha]_D^{24} + 11.5$ (c 2.5, CHCl₃).

2. General procedure for reductive amination (syntheses of racemic products)

A typical procedure for the preparation of rac-3a is described for the reaction of 4-phenyl-2-butanone and *p*-anisidine:

A magnetic stirrer bar and powdered molecular sieves 5A (MS 5A) (100 mg) were placed in a test tube under nitrogen atmosphere. The MS 5A were then dried with a heat gun under reduced pressure and the test tube was refilled with nitrogen. *p*-Anisidine (24.6 mg, 0.200 mmol), 4-phenyl-2-butanone (30.0 μ L, 0.200 mmol), and benzene (1 mL) were added to the test tube successively under nitrogen atmosphere at room temperature. After the mixture was stirred for 1 days, the mixture was filtered and the filtrate was concentrated under reduced pressure. The residue was diluted by MeOH (2 mL), and NaBH₄ (24.0 mg, 0.634 mmol) was added to the mixture at room temperature. After 5 min, AcOH (6.00 μ L, 0.105 mmol) was slowly added to the mixture at the same time and stirred for 1 day. Saturated aqueous NaHCO₃ solution (5 mL) was added to the mixture and extracted with CH₂Cl₂ (10 mL × 3), dried over anhydrous Na₂SO₄. The filtrate was evaporated under reduced pressure. The residue was purified by preparative thin layer chromatography on silica gel (AcOEt / hexane = 1 / 10) to give 19.0 mg (0.0744 mmol, 37%) of *rac*-**3a** as pale yellow oil.

NaBH₃CN was used instead of NaBH₄ for the preparation of *rac*-3c.

3. Deprotection of PMP group on $3a^4$

The reaction was carried out under air. To a solution of **3a** (51.0 mg, 0.200 mmol), MeCN (2 mL), and H₂O (2 mL) were added trichloroisocyanuric acid (TCCA) (0.230 g, 0.100 mmol) and 1 M aqueous H₂SO₄ (0.2 mL). The mixture was stirred for 20 h at room temperature and then washed CH₂Cl₂ (10 mL × 3). The resulting aqueous phase was subsequently brought to pH 10.5 by addition of 5 M aqueous KOH (3 mL) and extracted with AcOEt (15 mL × 4). The combined organic layers were dried over anhydrous Na₂SO₄. The organic material was filtered and the filtrate was concentrated under reduced pressure. The crude material was diluted with CH₂Cl₂ (1 mL), NEt₃ (0.140 mL, 1.0 mmol), and Boc₂O (0.140 mL, 0.6 mmol) successively added to the mixture at room temperature. After 12 h, the mixture was concentrated under reduced pressure. The crude material by preparative thin layer chromatography on silica gel (AcOEt / hexane = 1 / 10) to give 39.1 mg (0.157 mmol, 79%) of **5** as white solid.

HN O

 $5(79\%, 97\% \text{ ee})^5$

¹H NMR (400 MHz, CDCl₃) δ: 1.15 (d, *J* = 6.4 Hz, 3 H), 1.45 (s, 9 H), 1.66 – 1.78 (m, 2 H), 2.60 – 2.70 (m, 2 H), 3.71 (brs, 1 H), 4.37 (brs, 1 H), 7.15 – 7.21 (m, 3 H), 7.24 – 7.31 (m, 2 H).

¹³C NMR (100 MHz, CDCl₃) δ: 21.4, 28.4, 32.5, 39.2, 46.4, 79.0, 125.8, 128.3, 128.4, 141.9, 155.3.

HPLC conditions: Daicel CHIRALCEL[®] OD-H column, hexane/2-propanol = 96/4, flow rate = 0.5 mL min⁻¹, major enantiomer: t_R =14.96 min; minor enantiomer: t_R = 13.6 min.

 $[\alpha]_{D}^{24}$ + 12.0 (c 3.5, CHCl₃) [lit., $[\alpha]_{D}^{25}$ +13.9 (c 0.9, CHCl₃)]⁵

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