

SUPPORTING INFORMATION FOR

***n*Bu₄NI-catalyzed C3-formylation of indoles with N-methylaniline**

Lan-Tao Li, Juan Huang, Hong-Ying Li, Li-Juan Wen, Peng Wang and Bin Wang

*College of Pharmacy, State Key Laboratory of Medicinal Chemical Biology and
Tianjin Key Laboratory of Molecular Drug Research, Nankai University, 94 Weijin
Road, Tianjin 300071, China.*

Corresponding author: Bin Wang

wangbin@nankai.edu.cn

Table of Contents

- 1. General experimental methods**
- 2. General procedure for the *n*Bu₄NI-catalyzed formylation of indoles with N-methylaniline**
- 3. Optimization of reaction conditions**
- 4. Preparation of 3-formylindole with compound 5**
- 5. The characterization of the products**
- 6. ¹H and ¹³C NMR charts of the products**
- 7. References**

1. General experimental methods

^1H NMR and ^{13}C NMR spectra were recorded on a Bruker AVANCE AV400 (400MHz and 100MHz). Signal positions were recorded in ppm with the abbreviations s, d, t, m and b denoting singlet, doublet, triplet, multiplet and broadened respectively. All NMR chemical shifts were referenced to residual solvent peaks or to $\text{Si}(\text{CH}_3)_4$ as an internal standard, spectra recorded in CDCl_3 were referenced to residual CHCl_3 at 7.26 ppm for ^1H or 77.0 ppm for ^{13}C , spectra recorded in CD_3OD were referenced to residual CD_2HOD at 3.31 ppm for ^1H or 49.15 ppm for ^{13}C , spectra recorded in $(\text{CD}_3)_2\text{SO}$ were referenced to residual $(\text{CD}_2\text{H})\text{SO}(\text{CD}_3)$ at 2.50 ppm for ^1H or 39.52 ppm for ^{13}C . All coupling constants J were quoted in Hz. Data were reported as follows: chemical shift, multiplicity, coupling constant and integration. Reactions were monitored by thin-layer chromatography (TLC) on 0.25mm silica gel glass plates coated with 60 F₂₅₄. Column chromatography was performed on silica gel (200-300 mesh) using a mixture of petroleum ether (60-90°C)/ethyl acetate as eluant. Reactions were carried out under a nitrogen atmosphere. Commercially available reagents were used as received without purification.

2. General procedure for the Bu_4NI -catalyzed formylation of indoles with N-methylaniline

To a mixture of indole (0.5 mmol, 1 equiv), PivOH (255.3 mg, 2.5 mmol, 5 equiv), and $n\text{BuNI}$ (18.5 mg, 0.05 mmol, 0.1 equiv) in a 50 mL Schlenk tube were added DMSO (1 mL) under N_2 . Then N-methylaniline (107 μL , 1.0 mmol, 2 equiv) and TBPB (373 μL , 2.0 mmol, 4 equiv) were added separately. The mixture was stirred at 80°C for 8h, quenched with saturated NaHCO_3 solution (10 mL). The mixture was extracted by ethyl acetate for 3 times. The combined organic phase was washed with brine and dried with anhydrous Na_2SO_4 , filtrated, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to give the formylation products.

3. Optimization of reaction conditions^a

entry ^a	1a (mmol)	2a (mmol)	catalyst (mmol)	oxidant (mmol)	Yield (%)
1	0.5	1	<i>n</i> Bu ₄ NCl (0.05)	TBPB (2)	37 ^b
2	0.5	1	<i>n</i> Bu ₄ NBr (0.05)	TBPB (2)	29 ^b
3	0.5	1	<i>n</i> Bu ₄ NI (0.05)	TBHP(2) ^c	0
4	0.5	1	<i>n</i> Bu ₄ NI (0.05)	IBX (2)	trace
5	0.5	1	<i>n</i> Bu ₄ NI (0.05)	NBS (2)	trace
6	0.5	1	<i>n</i> Bu ₄ NI (0.05)	NIS (2)	0
7	0.5	1	<i>n</i> Bu ₄ NI (0.05)	PhI(OAc) ₂ (2)	trace
8	0.5	1	<i>n</i> Bu ₄ NI (0.05)	Oxone (2)	0
9	0.5	1	<i>n</i> Bu ₄ NI (0.05)	K ₂ S ₂ O ₈ (2)	0
10	0.5	1	<i>n</i> Bu ₄ NI (0.05)	<i>m</i> -CPBA (2)	0
11	0.5	1	<i>n</i> Bu ₄ NI (0.05)	<i>p</i> -BQ (2)	0
12	0.5	1	<i>n</i> Bu ₄ NI(0.05)	DDQ (2)	0

^ageneral reaction conditions: to a mixture of indole (0.5mmol), catalyst (0.05mmol), PivOH (2.5mmol), oxidant (2mmol) in a schlank tube was added DMSO (1mL) under N₂. N-methylaniline (1.0mmol) was added, then the mixture was heated at 80°C for 8h. ^bGC yield. ^csolution of TBHP in decane (5.0-6.0M) was used.

4. Preparation of 3-formylindole with compound **5**

To a mixture of compound **5** (0.5 mmol, 1 equiv), PivOH (255.3 mg, 2.5 mmol, 5 equiv), and *n*BuNI (18.5 mg, 0.05 mmol, 0.1 equiv) in a 50 mL Schlenk tube were added DMSO (1 mL) under N₂. Then TBPB (373 μ L, 2.0 mmol, 4 equiv) were added separately. The mixture was stirred at 80°C for 8h, quenched with saturated NaHCO₃ solution (10 mL). The mixture was extracted by ethyl acetate for 3 times. The combined organic phase was washed with brine and dried with anhydrous Na₂SO₄, filtrated, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to give the formylation product **3a** in 70% yield.

5. The characterization of the products

3a¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.1 (b, 1H), 9.94(s, 1H), 8.29(s, 1H), 8.10(d, 7.2Hz, 1H), 7.51(d, 8.0Hz, 1H), 7.28-7.20(m, 2H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.00, 138.53, 137.07, 124.14, 123.49, 122.16, 120.85, 118.18, 112.45

3b¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.2(b, 1H), 9.92(s, 1H), 8.35(d, 3.2Hz, 1H), 7.76(dd, 2.4Hz, 9.6Hz, 1H), 7.53(dd, 4.8Hz, 8.8Hz, 1H), 7.11(td, 2.8Hz, 9.2Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.04, 158.74 (239.6Hz), 139.63, 133.61, 124.73 (7Hz), 118.14, 113.77 (9.5Hz), 111.61 (25.8Hz), 105.70 (24.3Hz)

3c¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.3(b, 1H), 9.93(s, 1H), 8.36(d, 3.2Hz, 1H), 8.06(d, 2.2Hz, 1H), 7.54(d, 8.4Hz, 1H), 7.28(dd, 2.2Hz, 8.6Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.10, 139.39, 135.48, 126.77, 125.27, 123.47, 119.87, 117.55, 114.11

3d²

¹H-NMR (CD₃OD, 400MHz): 9.86(s, 1H), 8.05(s, 1H), 7.99(d, 8.0Hz, 1H), 7.16(t, 7.6Hz, 1H), 7.09(d, 7.2Hz, 1H), 2.90(q, 7.6Hz, 2H), 1.31(t, 7.6Hz, 3H)

¹³C-NMR (CD₃OD, 100MHz): 187.57, 139.56, 137.63, 129.50, 125.84, 124.13, 124.00, 120.57, 120.20, 25.21, 15.10

¹H-NMR (DMSO-*d*₆, 400MHz): 12.1(b, 1H), 9.93(s, 1H), 8.27(d, 3.2Hz, 1H), 7.93(d, 7.6Hz, 1H), 7.15(t, 7.6Hz, 1H), 7.08(d, 7.2Hz, 1H), 2.89(t, 7.4Hz, 2H), 1.26(t, 7.4Hz, 3H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 184.92, 138.12, 135.74, 128.04, 124.08, 122.42,

122.25, 118.56, 118.43, 23.52, 14.55

3e³

¹H-NMR (DMSO-*d*₆, 400MHz): 12.2(b, 1H), 9.93(s, 1H), 8.32(d, 3.2Hz, 1H), 8.02(d, 8.4Hz, 1H), 7.71(d, 1.6Hz, 1H), 7.36(dd, 1.6Hz, 8.4Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.15, 139.18, 137.91, 125.05, 123.14, 122.46, 117.97, 115.91, 115.15

3f¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.3(b, 1H), 9.91(s, 1H), 8.43(d, 1.6Hz, 1H), 8.30(d, 3.2Hz, 1H), 7.54(dd, 1.6Hz, 8.6Hz, 1H), 7.37(d, 8.4Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.15, 138.89, 136.15, 131.53, 129.12, 126.60, 117.14, 114.92, 86.58

3g¹

¹H-NMR (CD₃OD, 400MHz): 10.0(s, 1H), 8.09(d, 7.2Hz, 1H), 7.34(d, 7.2Hz, 1H), 7.18(t, 4.4Hz, 2H), 2.70(s, 3H)

¹³C-NMR (CD₃OD, 100MHz): 186.55, 151.03, 137.34, 127.38, 124.33, 123.57, 121.68, 115.60, 112.33, 11.80

3h⁴

¹H-NMR (DMSO-*d*₆, 400MHz): 12.3(b, 1H), 9.93(s, 1H), 8.19(d, 3.2Hz, 1H), 7.67(d, 7.6Hz, 1H), 7.57(d, 7.2Hz, 2H), 7.42(t, 7.4Hz, 2H), 7.35(d, 7.2Hz, 1H), 7.12(t, 8.0Hz, 1H), 6.93(d, 8.0Hz, 1H), 5.29(s, 2H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.12, 145.26, 137.81, 137.03, 128.43, 127.90, 127.68, 127.18, 125.83, 122.95, 118.68, 113.57, 105.54, 69.38

3i¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.6(b, 1H), 10.00(s, 1H), 8.55(d, 3.2Hz, 1H), 8.24(d, 8.0Hz, 1H), 8.04(s, 1H), 7.59(dd, 1.4Hz, 8.2Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.44, 141.30, 135.95, 127.38, 125.08, 121.74, 119.81, 118.04, 117.43, 105.13

3j¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.0(b, 1H), 9.89(s, 1H), 8.21(d, 3.2Hz, 1H), 7.58(d, 2.8Hz, 1H), 7.40(d, 8.8Hz, 1H), 6.88(dd, 2.4Hz, 8.8Hz, 1H), 3.78(s, 3H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 184.78, 155.59, 138.38, 131.76, 124.86, 118.00, 113.26, 113.15, 102.45, 55.23

3k¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.4(b, 1H), 9.98(s, 1H), 8.50(d, 3.2Hz, 1H), 8.18(d, 8.4Hz, 1H), 8.14(s, 1H), 7.83(d, 8.4Hz, 1H), 3.87(s, 3H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.25, 166.64, 140.94, 136.47, 127.76, 124.53, 122.77, 120.68, 118.04, 114.18, 52.06

3l¹

¹H-NMR (CD₃OD, 400MHz): 9.86(s, 1H), 8.06(s, 1H), 7.98(d, 7.6Hz, 1H), 7.13(t, 7.4Hz, 1H), 7.06(d, 7.2Hz, 1H), 2.51(s, 3H)

¹³C-NMR (CD₃OD, 100MHz): 187.63, 139.57, 138.43, 125.71, 125.60, 123.99, 123.05, 120.57, 120.13, 16.90

¹H-NMR (DMSO-*d*₆, 400MHz): 12.2(b, 1H), 9.95(s, 1H), 8.29(d, 3.2Hz, 1H), 7.93(d, 7.6Hz, 1H), 7.13(t, 7.6Hz, 1H), 7.06(d, 7.2Hz, 1H), 2.50(s, 3H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.01, 138.17, 136.55, 124.01, 123.92, 122.32, 121.75, 118.56, 118.38, 16.67

3m¹

¹H-NMR (DMSO-*d*₆, 400MHz): 12.3(b, 1H), 9.93(s, 1H), 8.34(d, 3.2Hz, 1H), 8.22(d, 1.6Hz, 1H), 7.49(d, 8.4Hz, 1H), 7.39(dd, 2.0Hz, 8.8Hz, 1H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.13, 139.23, 135.74, 126.04, 125.89, 122.91, 117.43, 114.80, 114.55

3n⁵

¹H-NMR (CDCl₃, 400MHz): 9.99(s, 1H), 8.33(d, 6.4Hz, 1H), 7.66(s, 1H), 7.37(s, 3H), 3.86(s, 3H)

¹³C-NMR (CDCl₃, 100MHz): 184.48, 139.35, 137.91, 125.28, 124.04, 122.95, 122.02, 118.05, 109.90, 33.68

3o⁶

¹H-NMR (DMSO-*d*₆, 400MHz): 9.94(s, 1H), 8.48(s, 1H), 8.12(d, 6.8Hz, 1H), 7.59(d, 8.0Hz, 1H), 7.34-7.26(m, 7H), 5.55(s, 2H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 184.68, 140.97, 136.94, 136.75, 128.71, 127.77, 127.33, 124.78, 123.60, 122.54, 121.06, 117.38, 111.39, 49.78

3p⁷

¹H-NMR (DMSO-*d*₆, 400MHz): 10.0(s, 1H), 8.61(s, 1H), 8.24-8.20(m, 1H), 7.70-7.63(m, 4H), 7.56-7.54(m, 2H), 7.37-7.34(m, 2H)

¹³C-NMR (DMSO-*d*₆, 100MHz): 185.29, 140.57, 137.61, 136.74, 130.04, 128.15, 124.97, 124.70, 124.49, 123.23, 121.39, 118.80, 111.32

3q⁸

¹H-NMR (CDCl₃, 400MHz): 10.1(s, 1H), 8.34-8.32(m, 2H), 7.95(s, 1H), 7.80-7.78(m, 2H), 7.70(t, 7.4Hz, 1H), 7.60(t, 7.6Hz, 2H), 7.50-7.45(m, 2H)

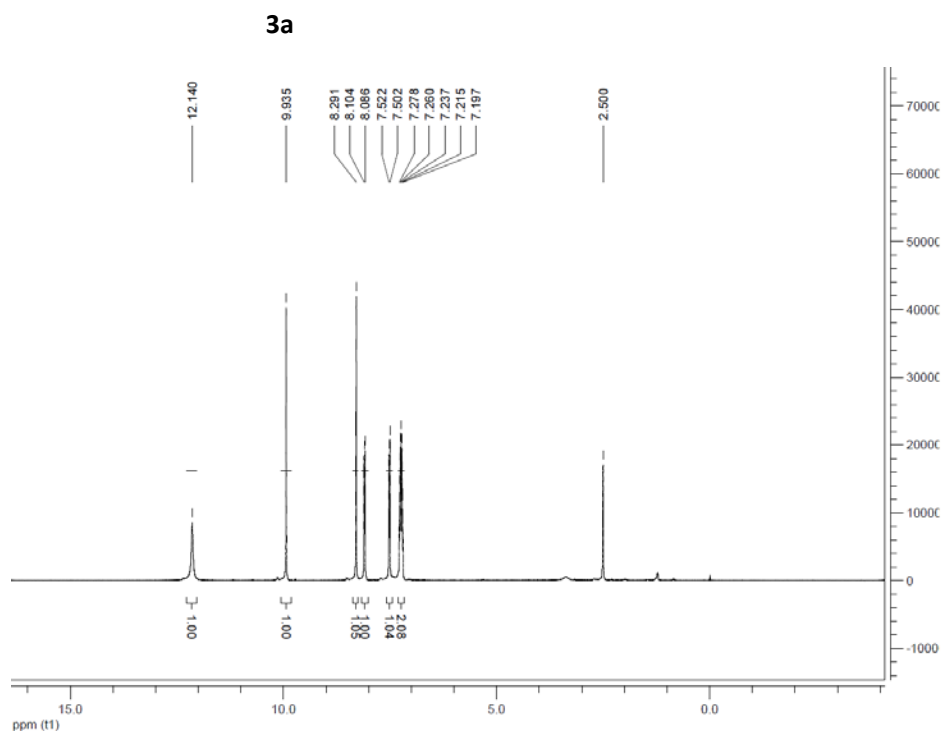
¹³C-NMR (CDCl₃, 100MHz): 185.69, 168.50, 137.59, 136.81, 132.99, 129.44, 129.01, 126.59, 126.24, 125.58, 122.18, 122.04, 116.08

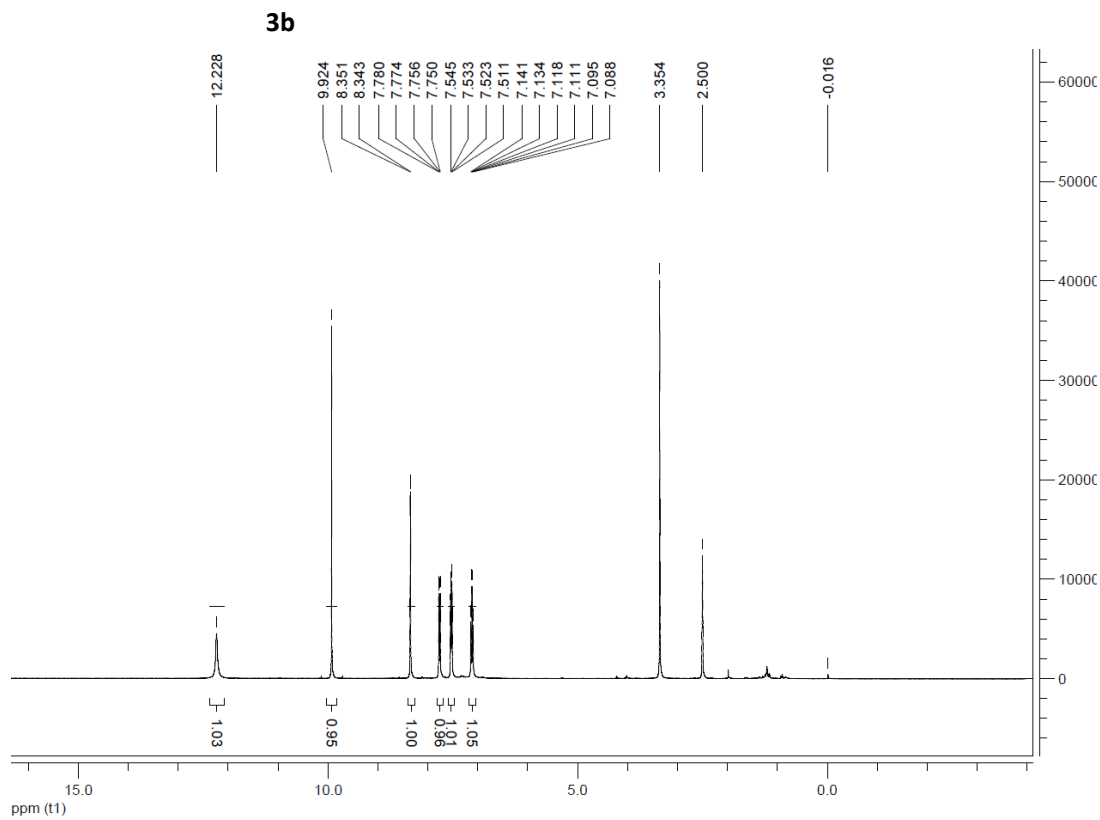
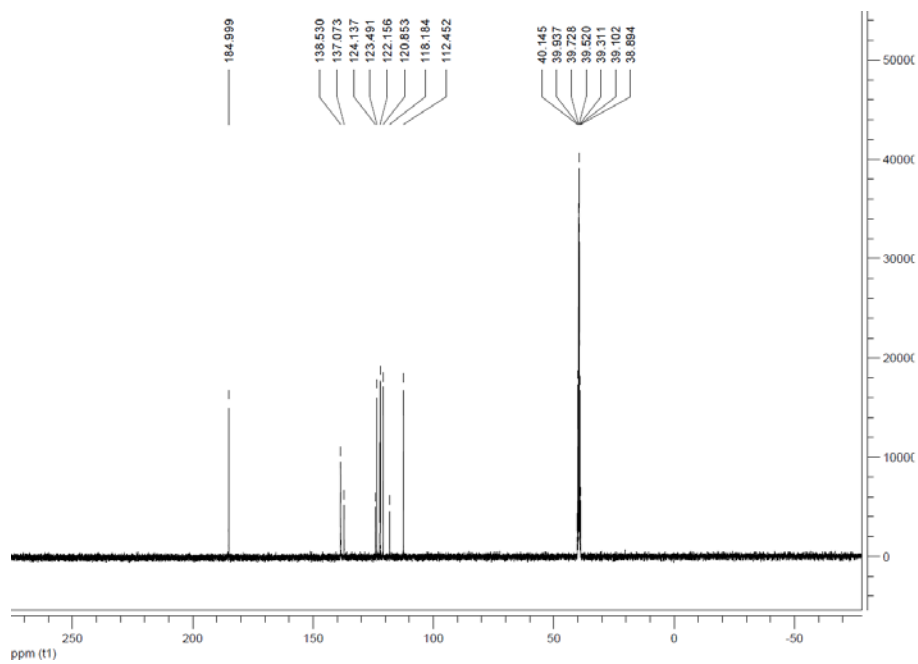
4⁹

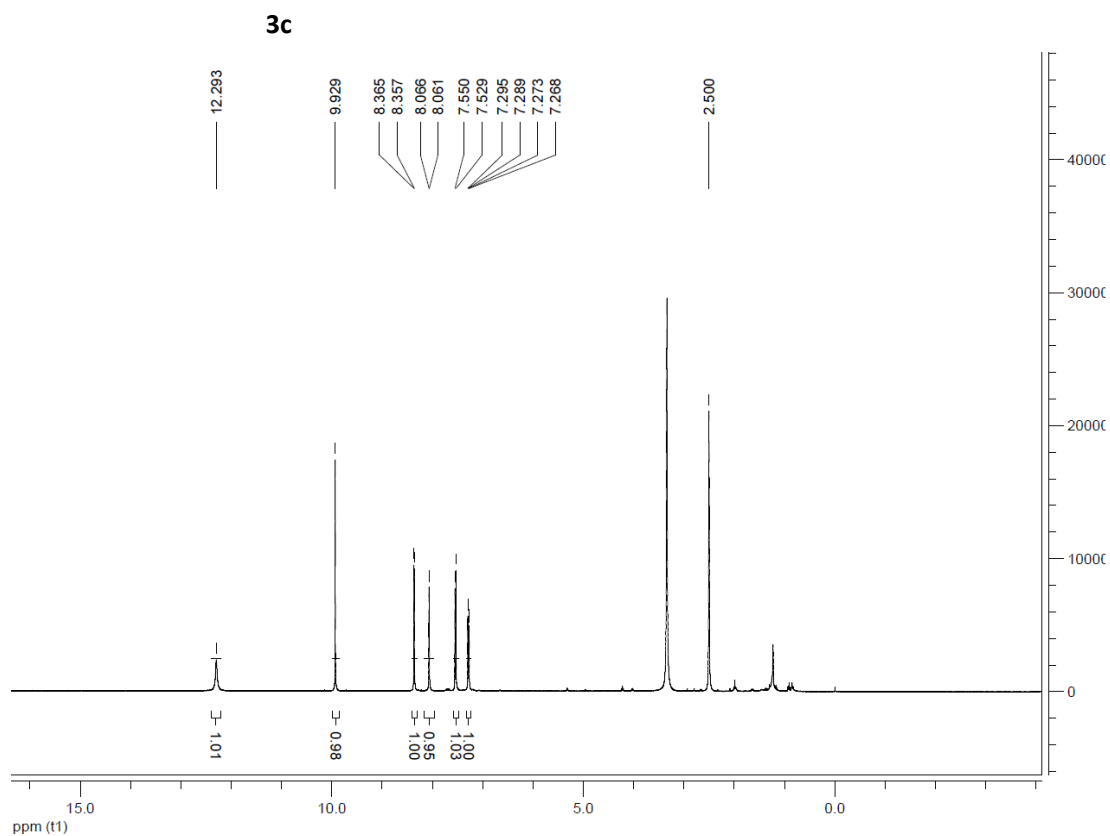
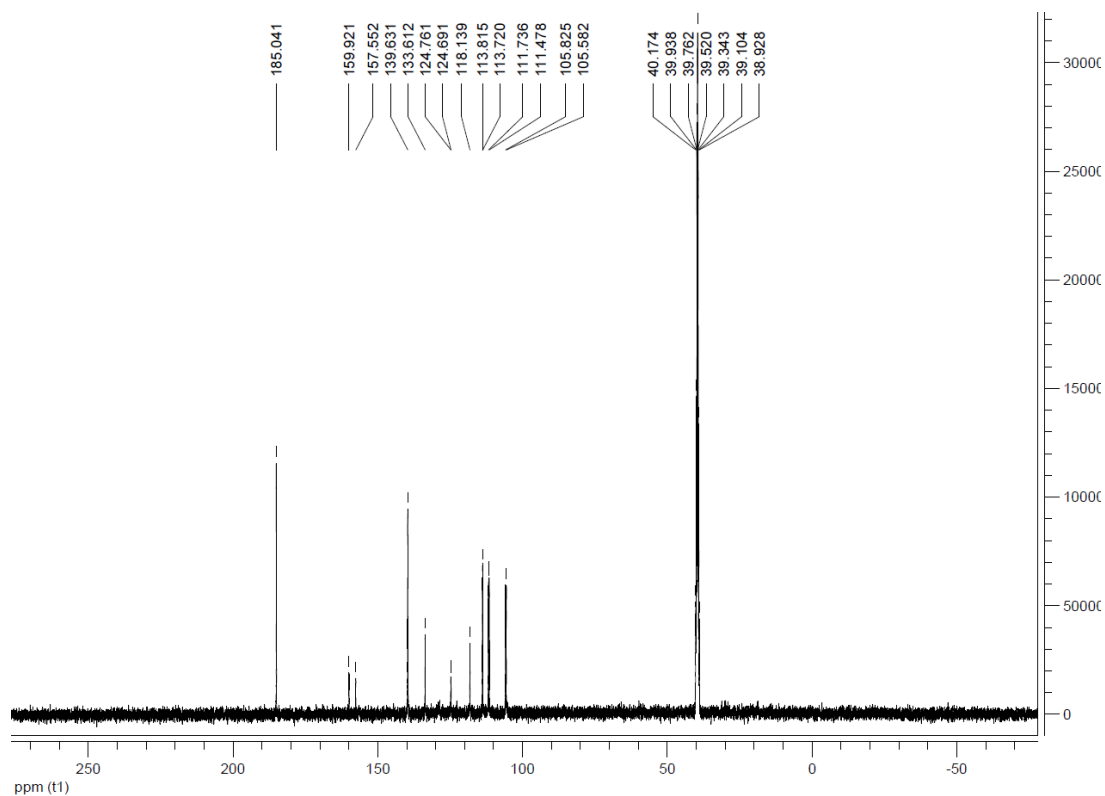
¹H-NMR (CDCl₃, 400MHz): 7.06 (d, 8.4Hz, 2H), 6.71(d, 5.6Hz, 2H), 3.81(s, 2H), 2.91(s, 12H)

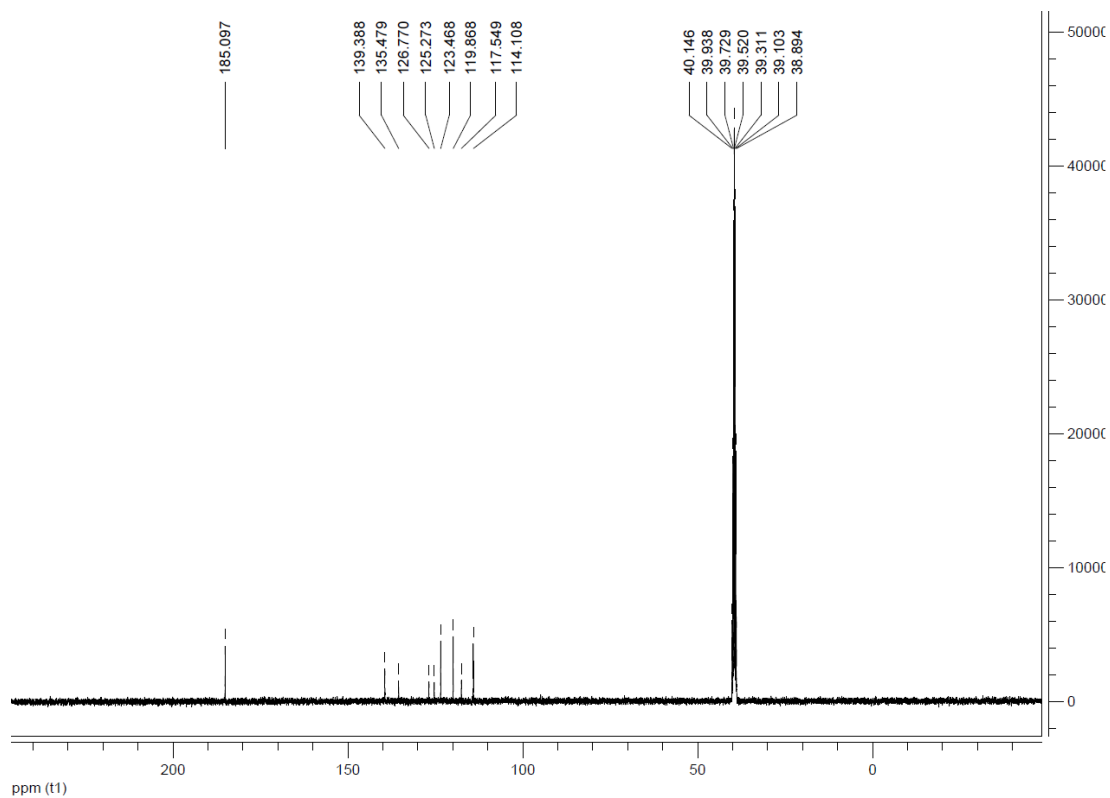
¹³C-NMR (CDCl₃, 100MHz): 129.41, 113.12, 40.97, 39.88

6. ¹H and ¹³C NMR charts of the products

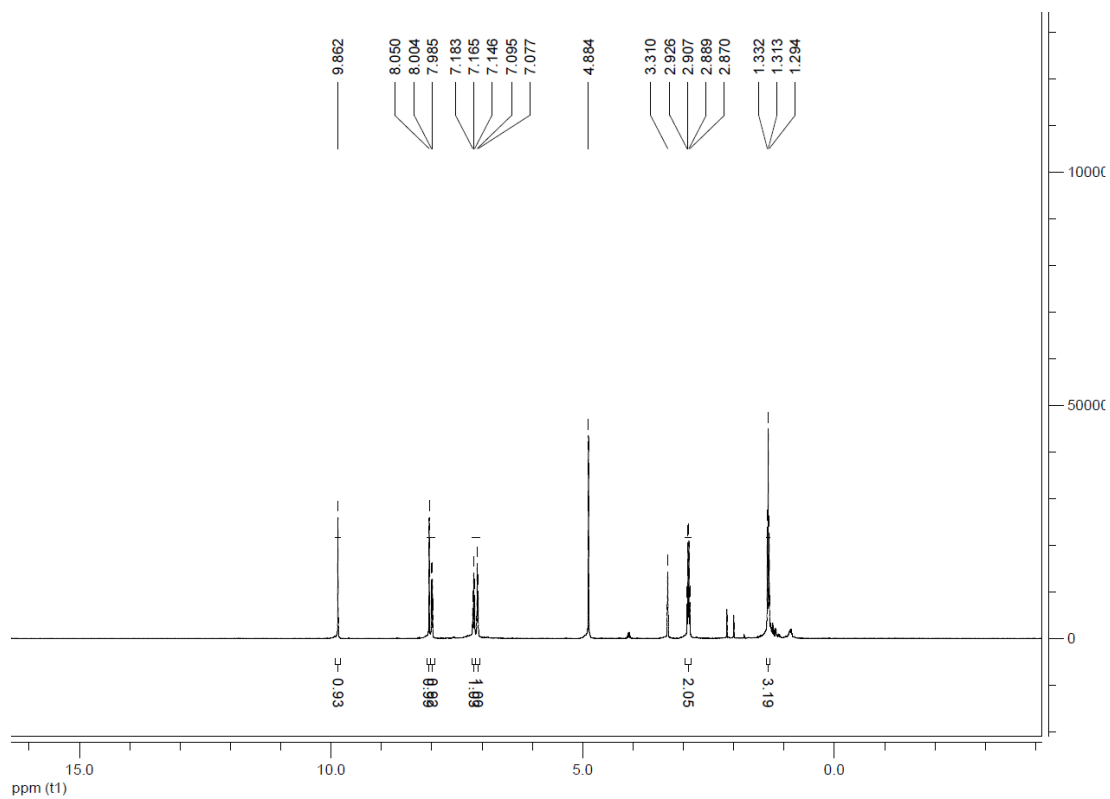


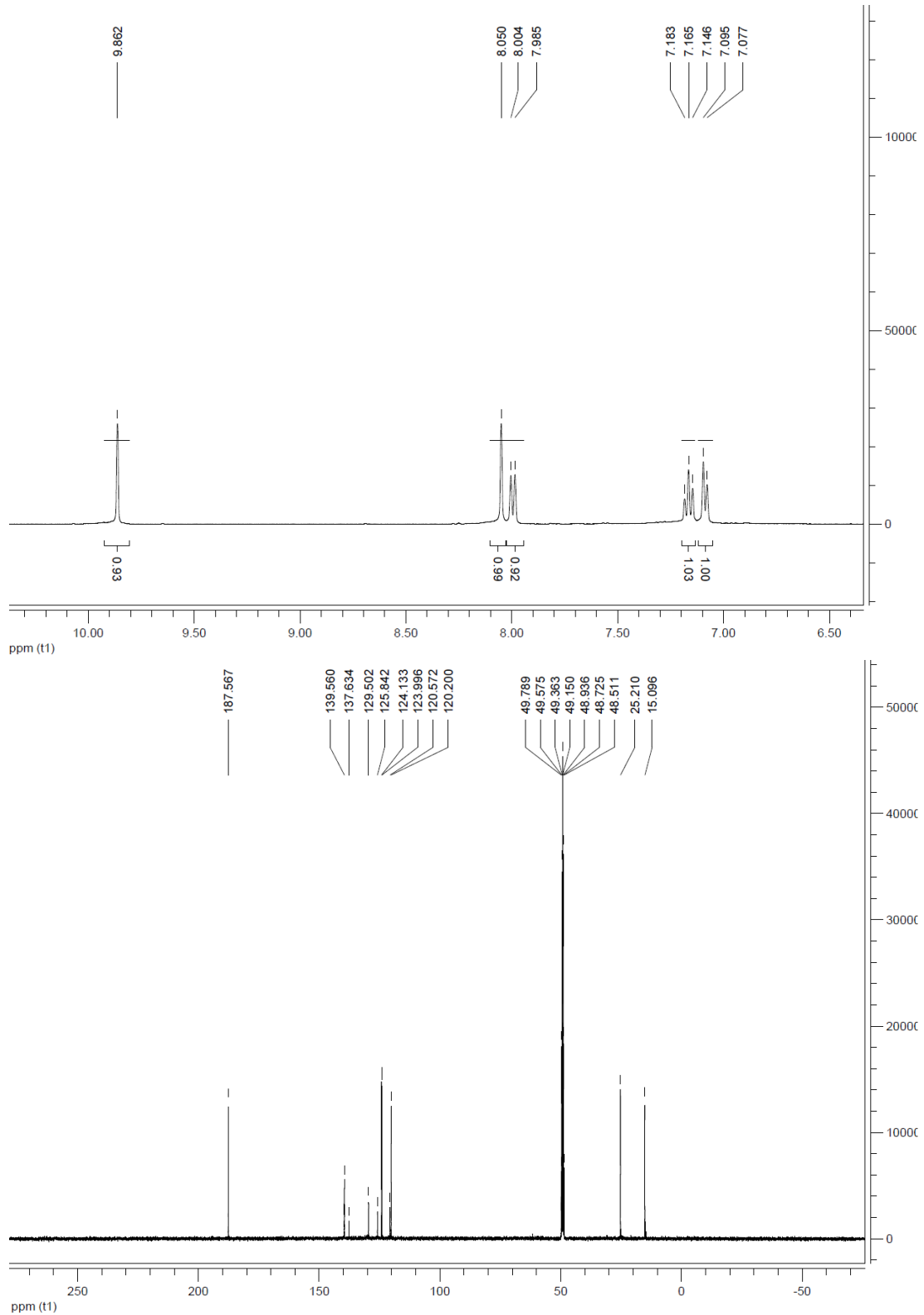




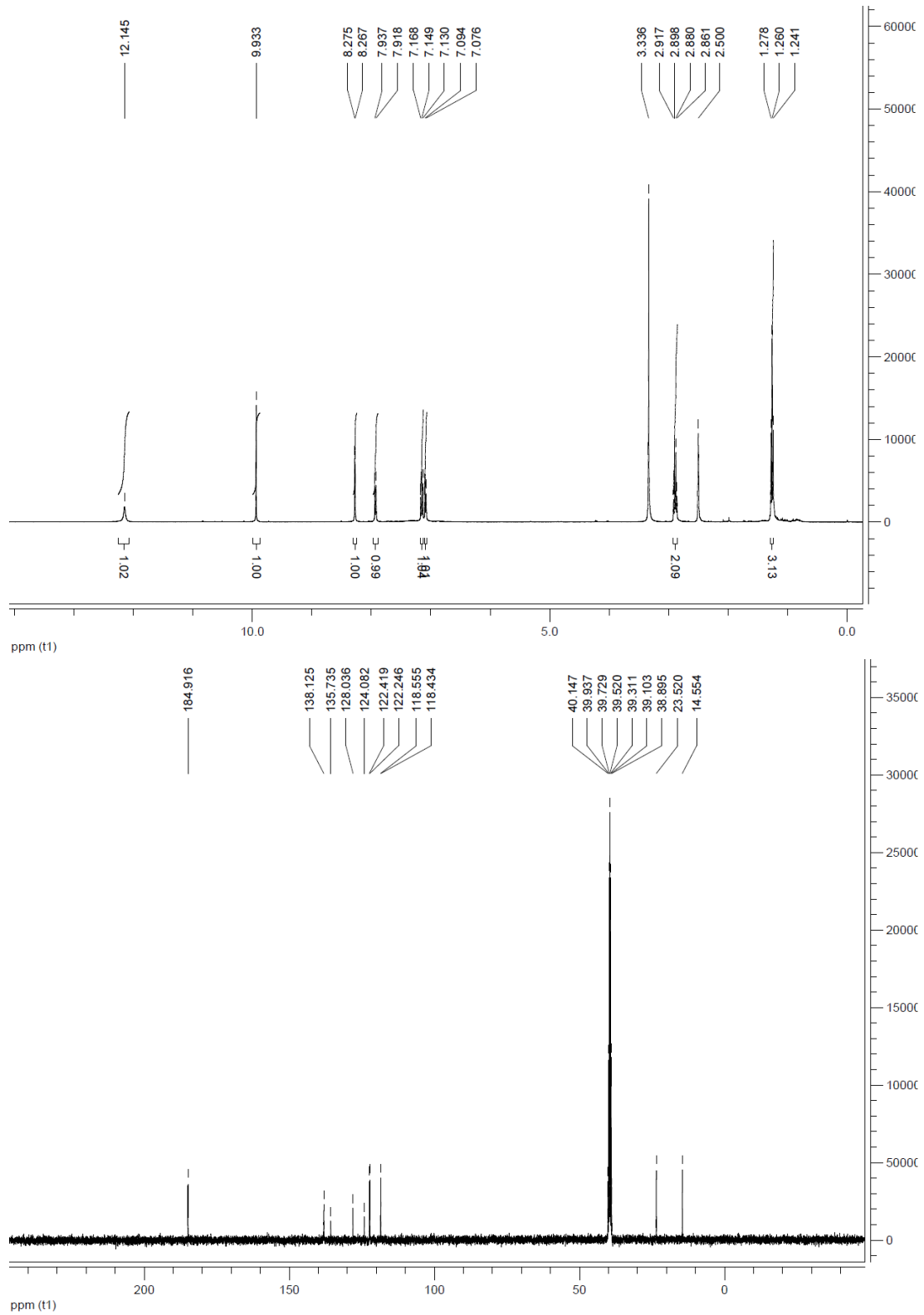


3d in methanol-*d*₄

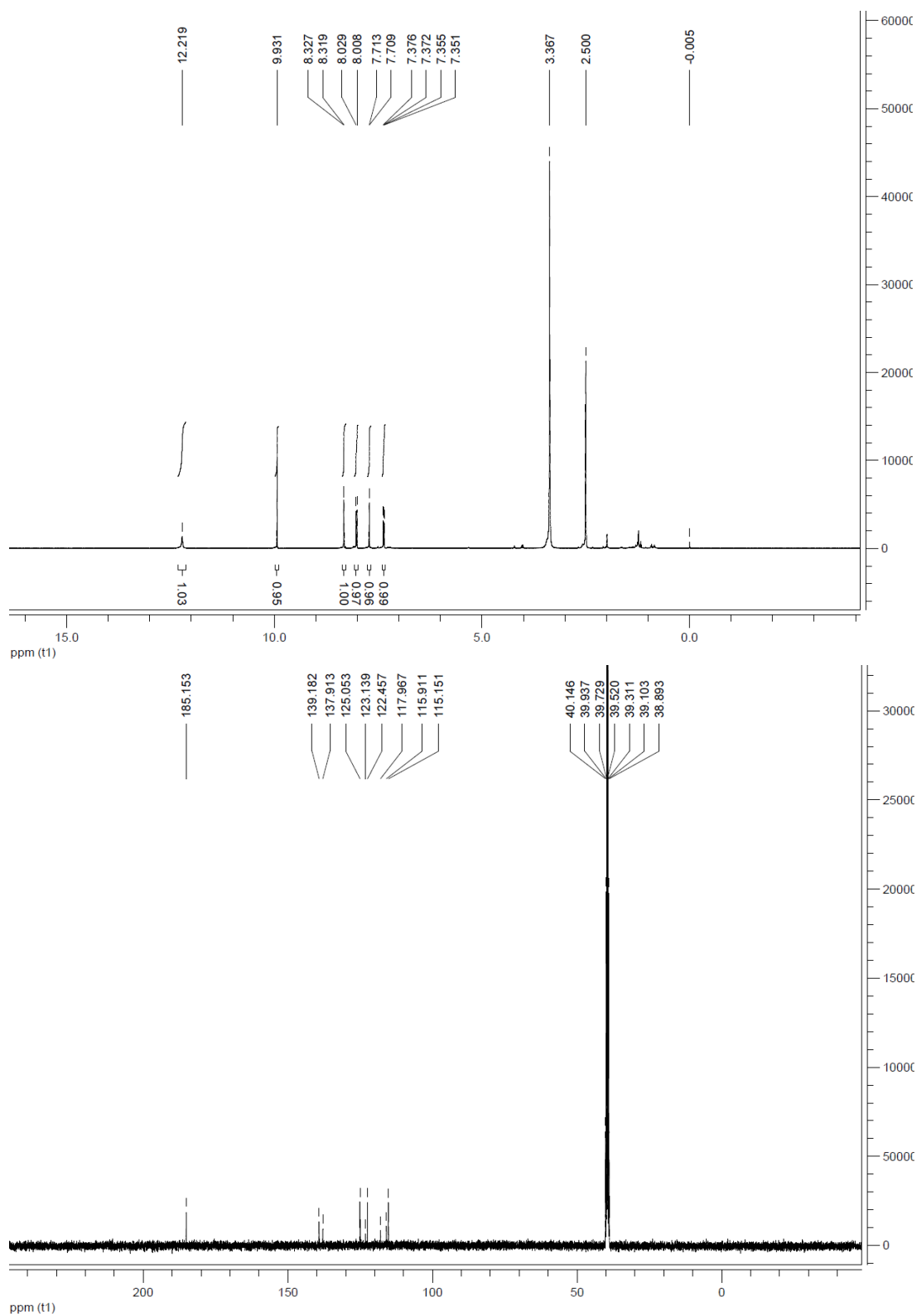




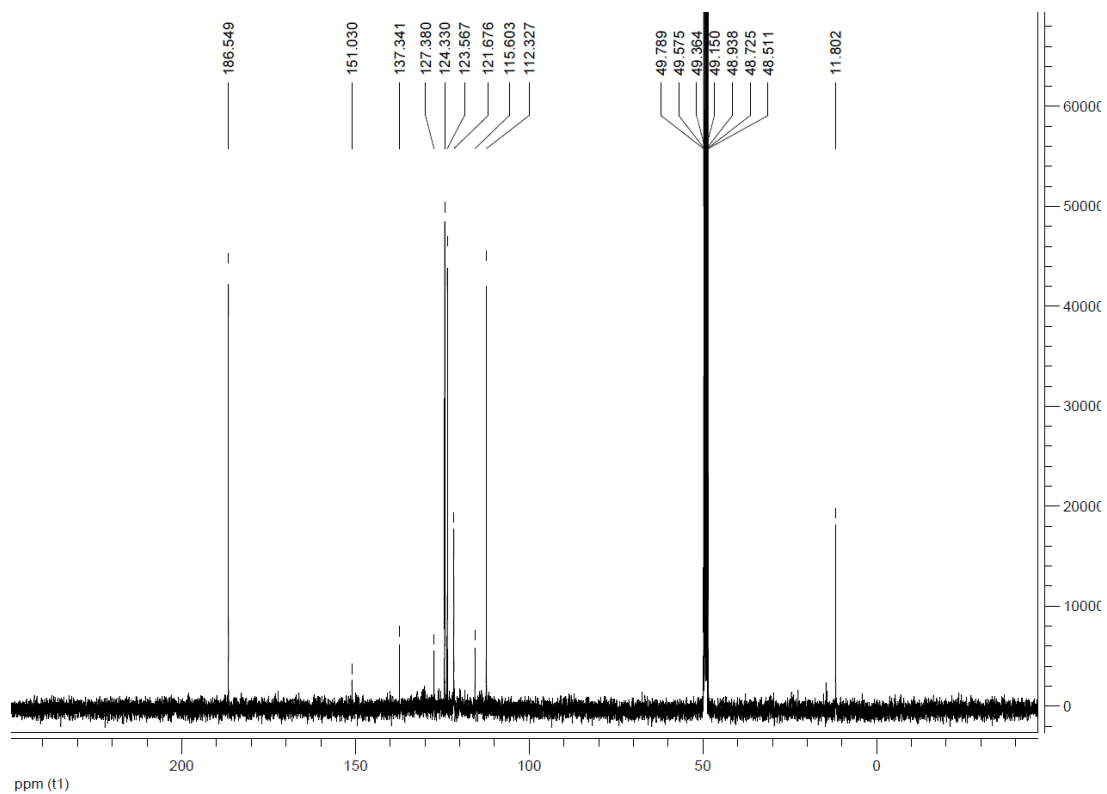
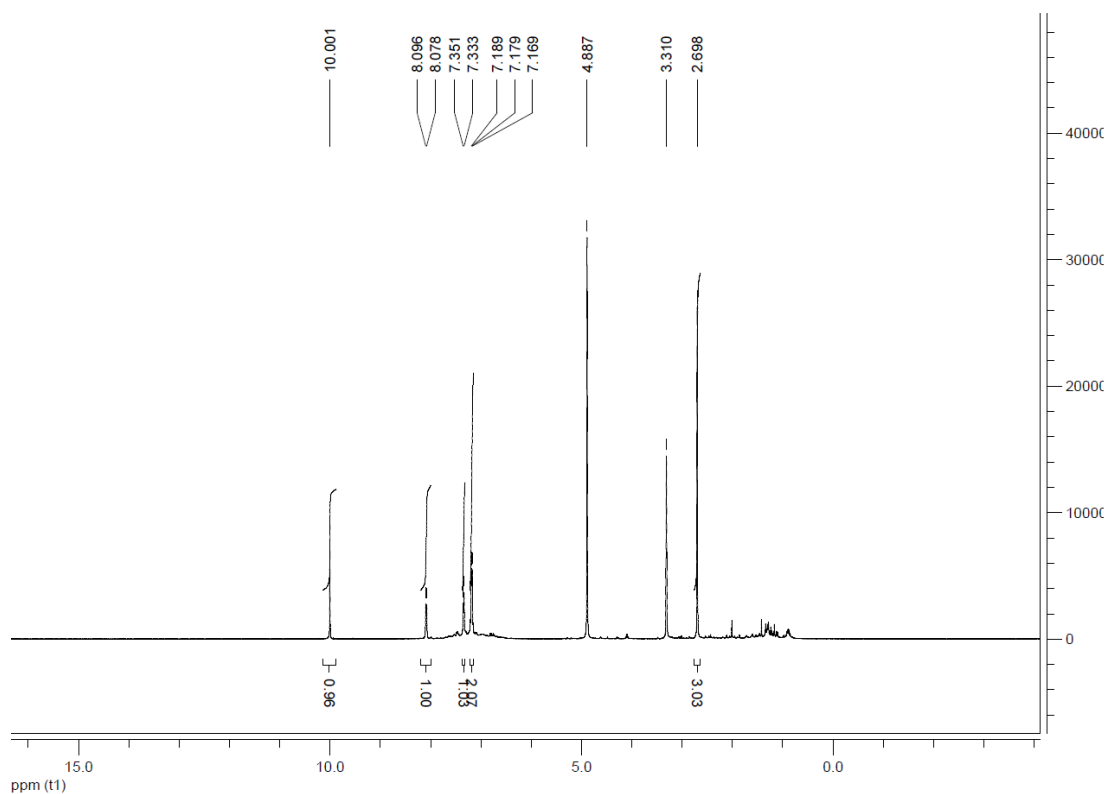
3d in DMSO-*d*₆



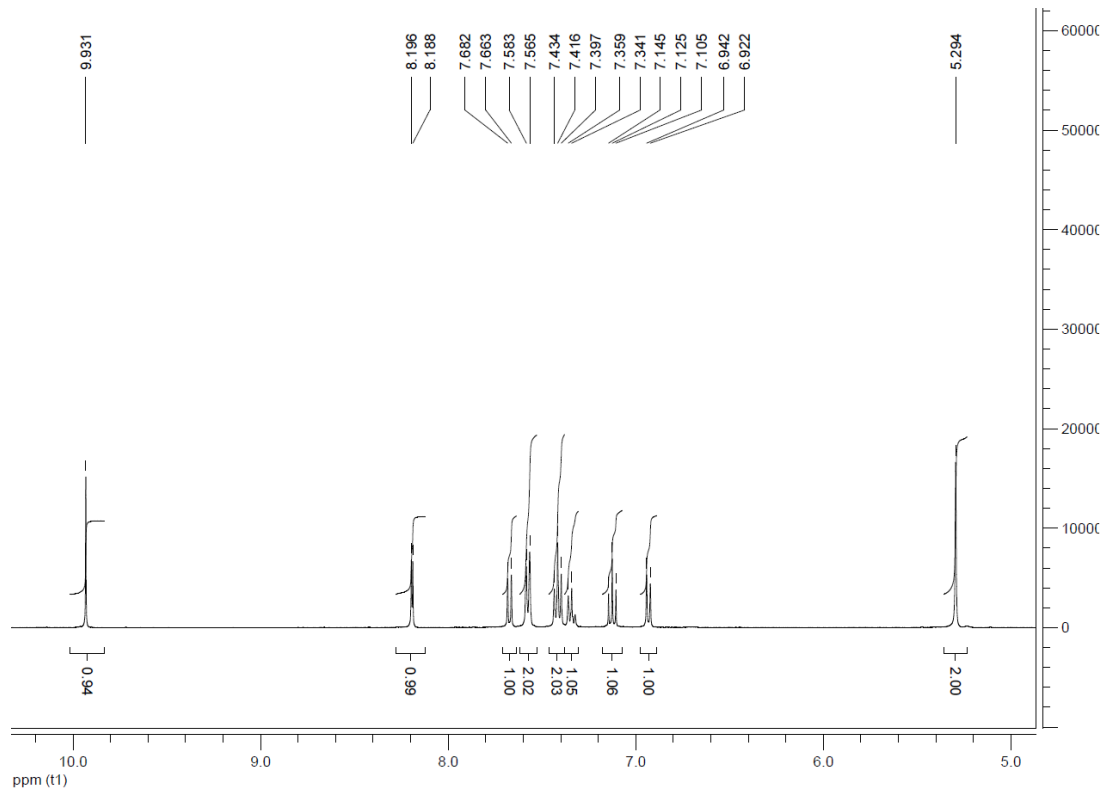
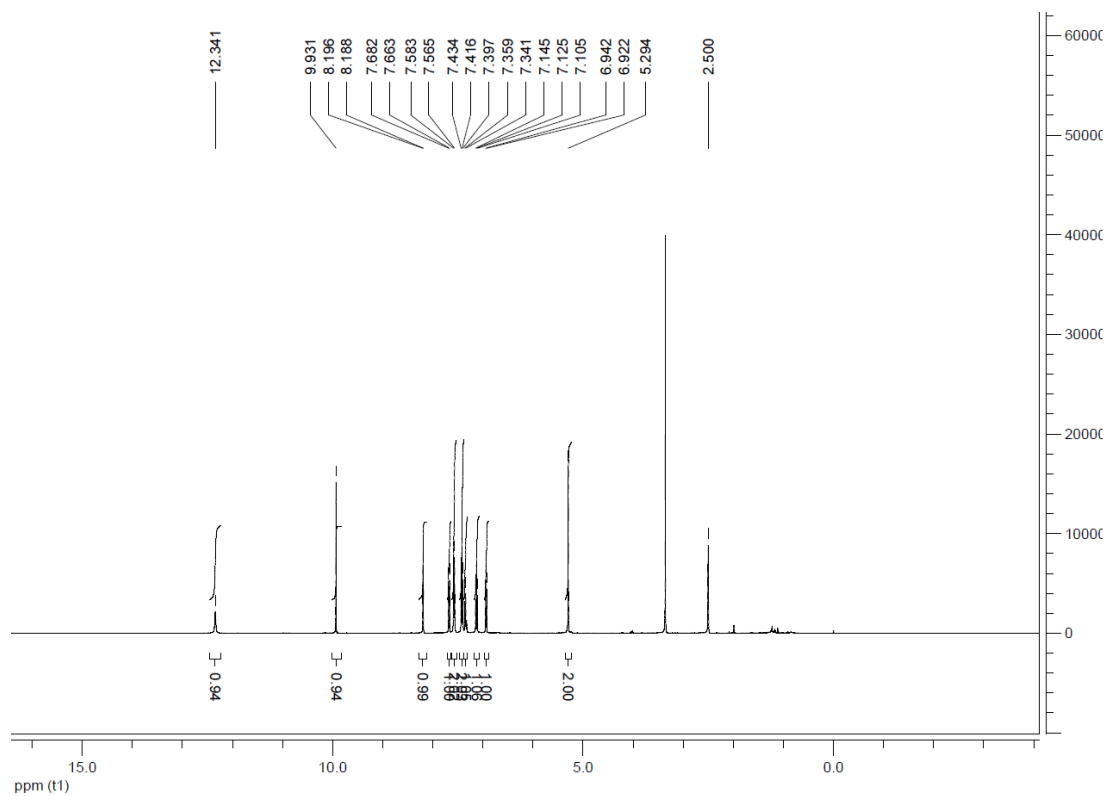
3e

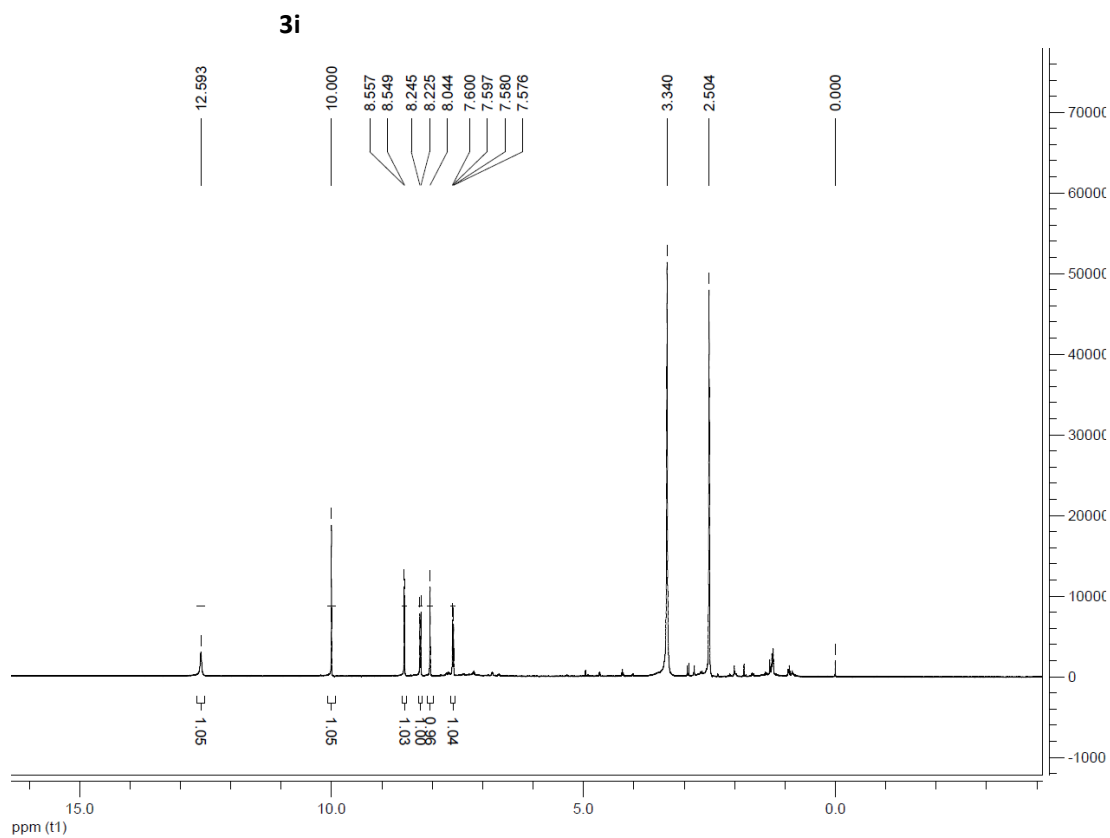
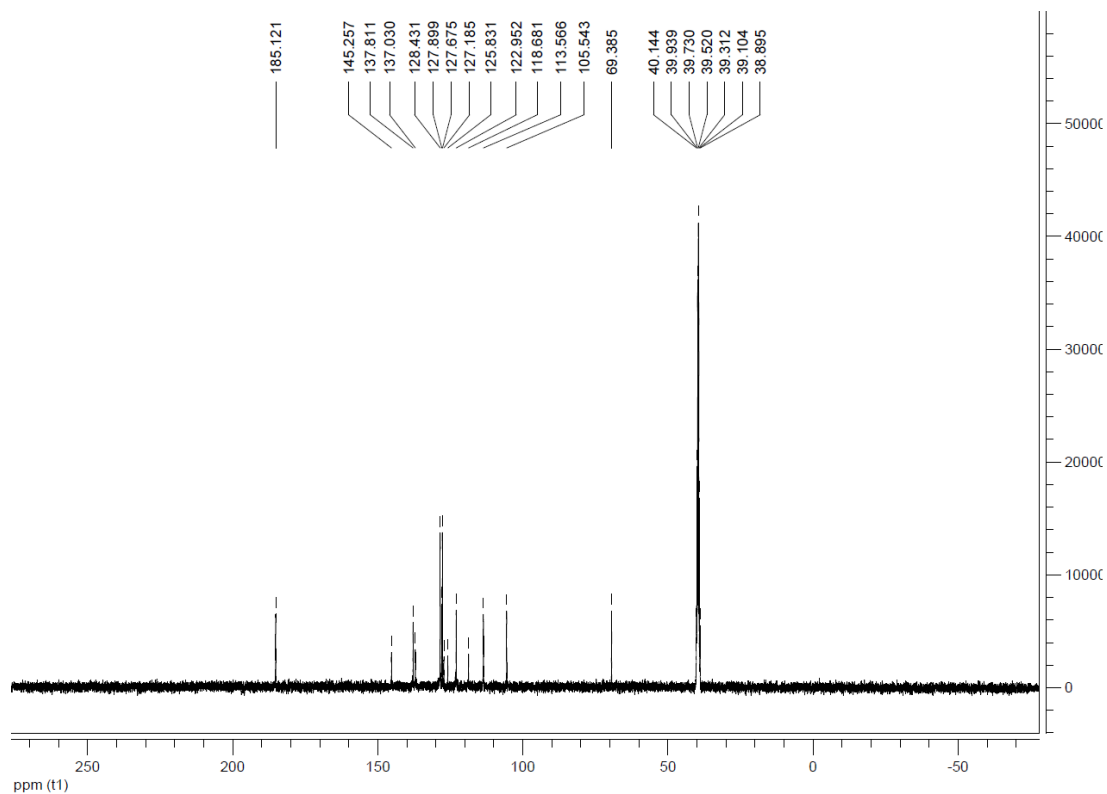


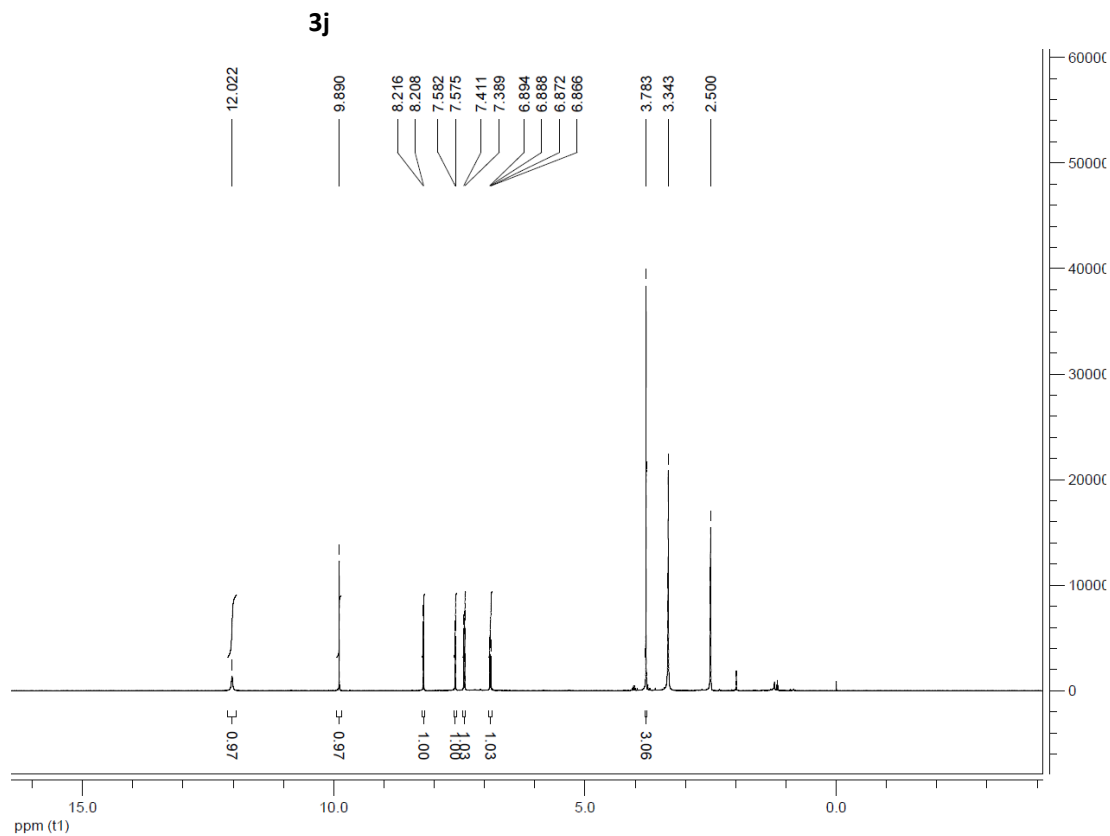
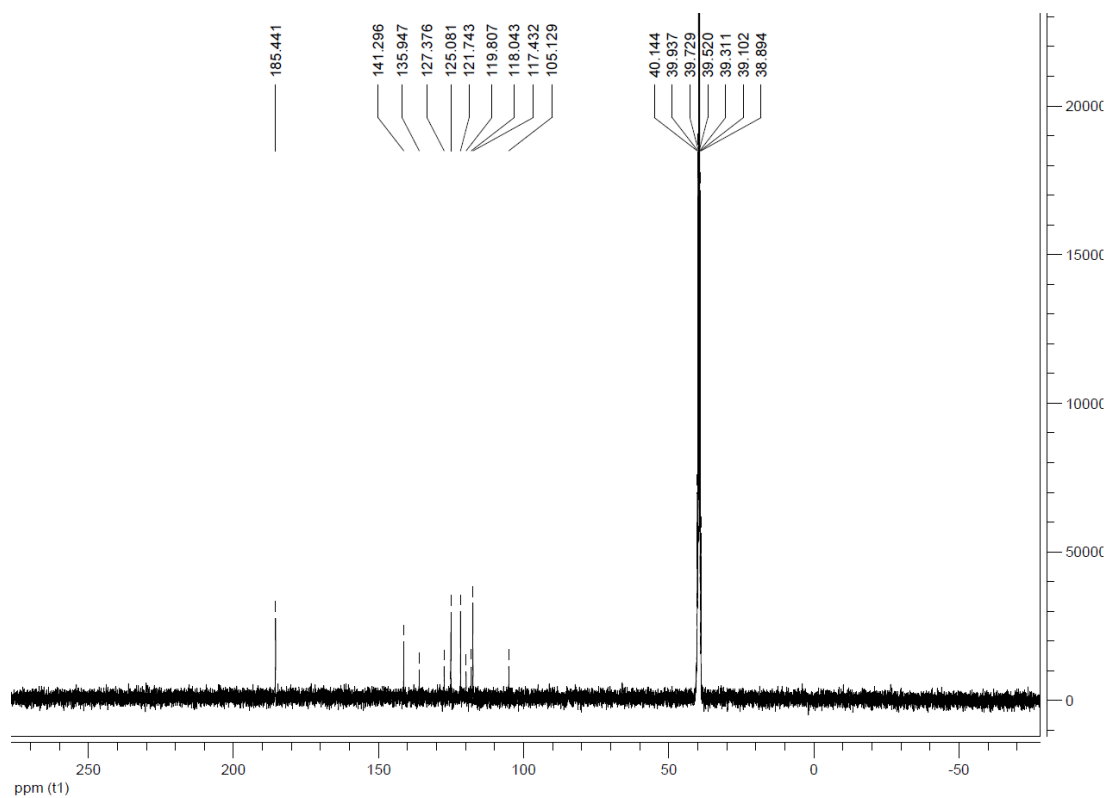
3f

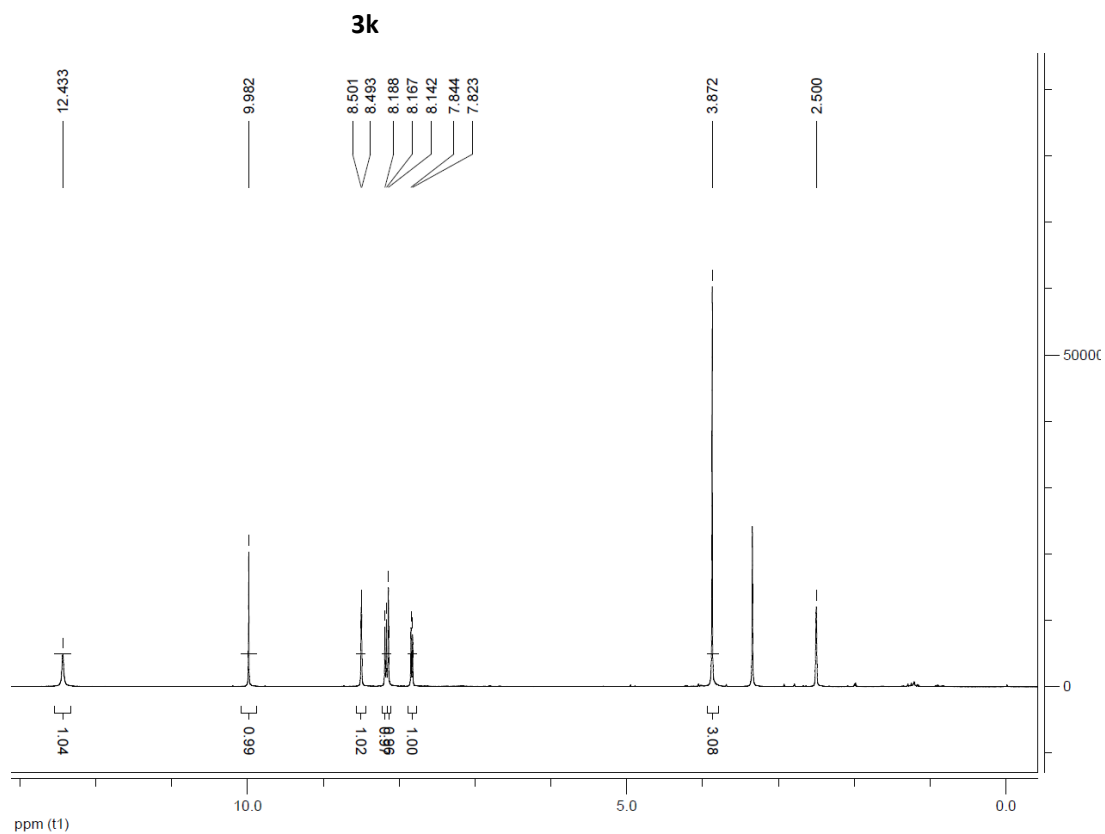
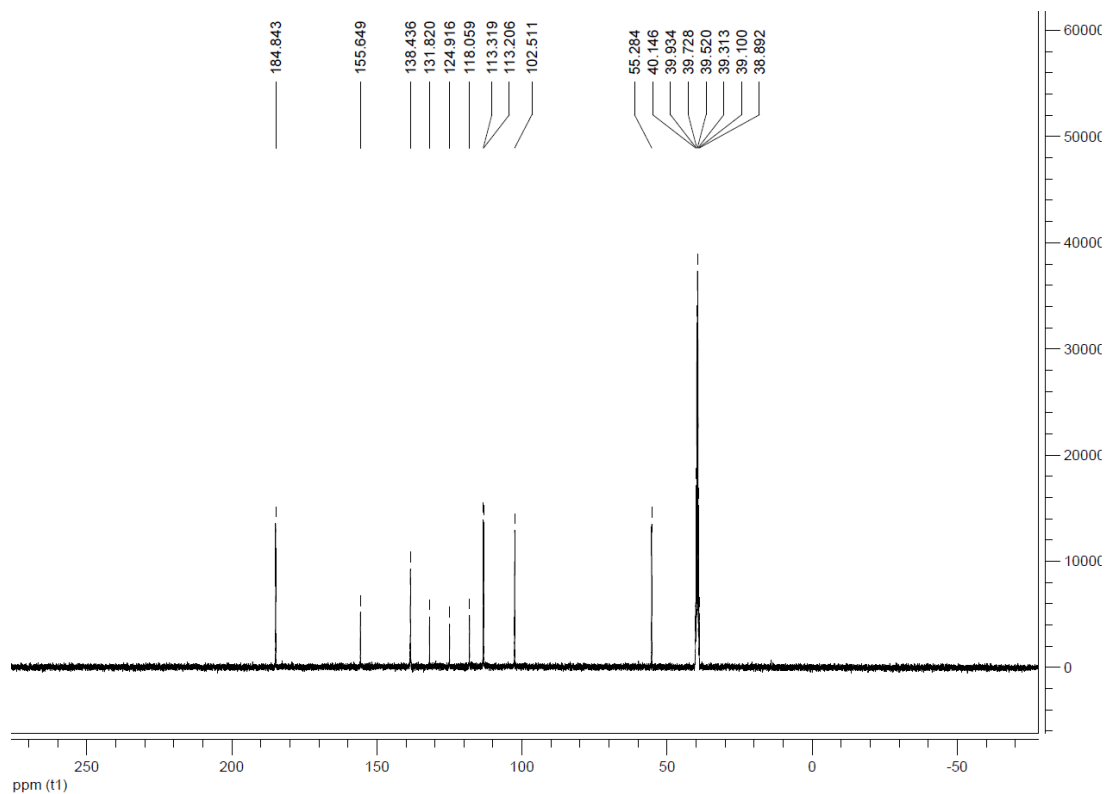


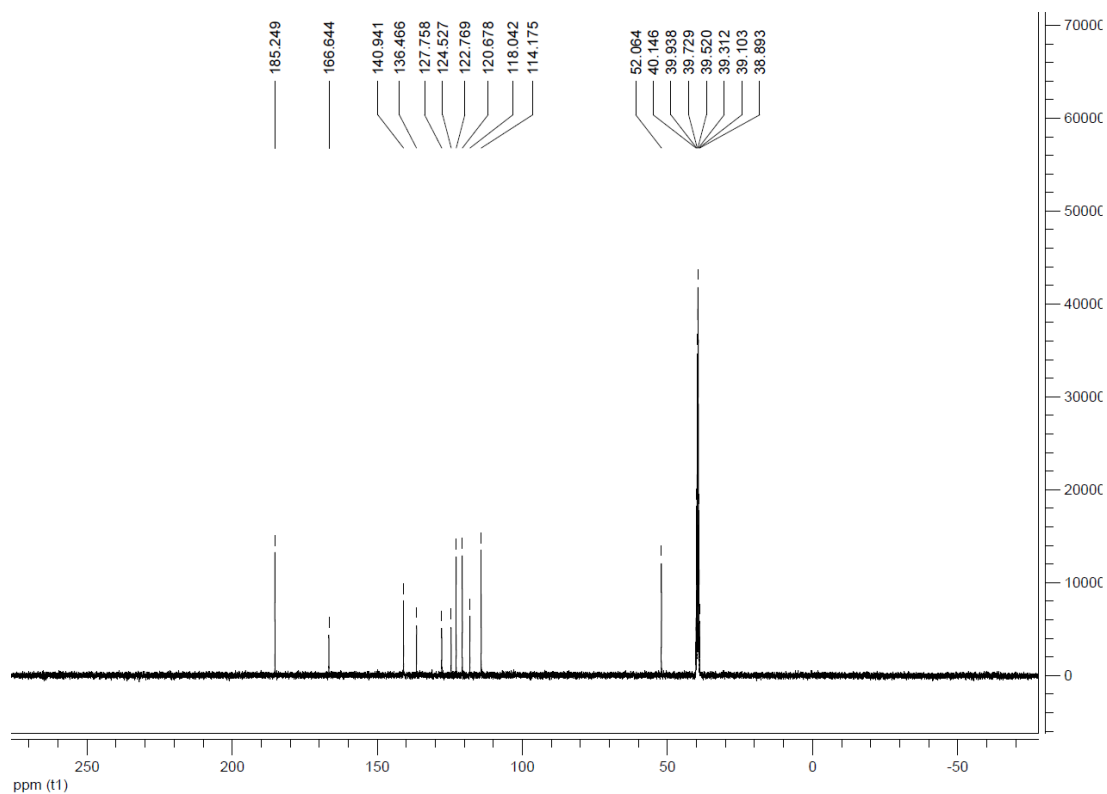
3h



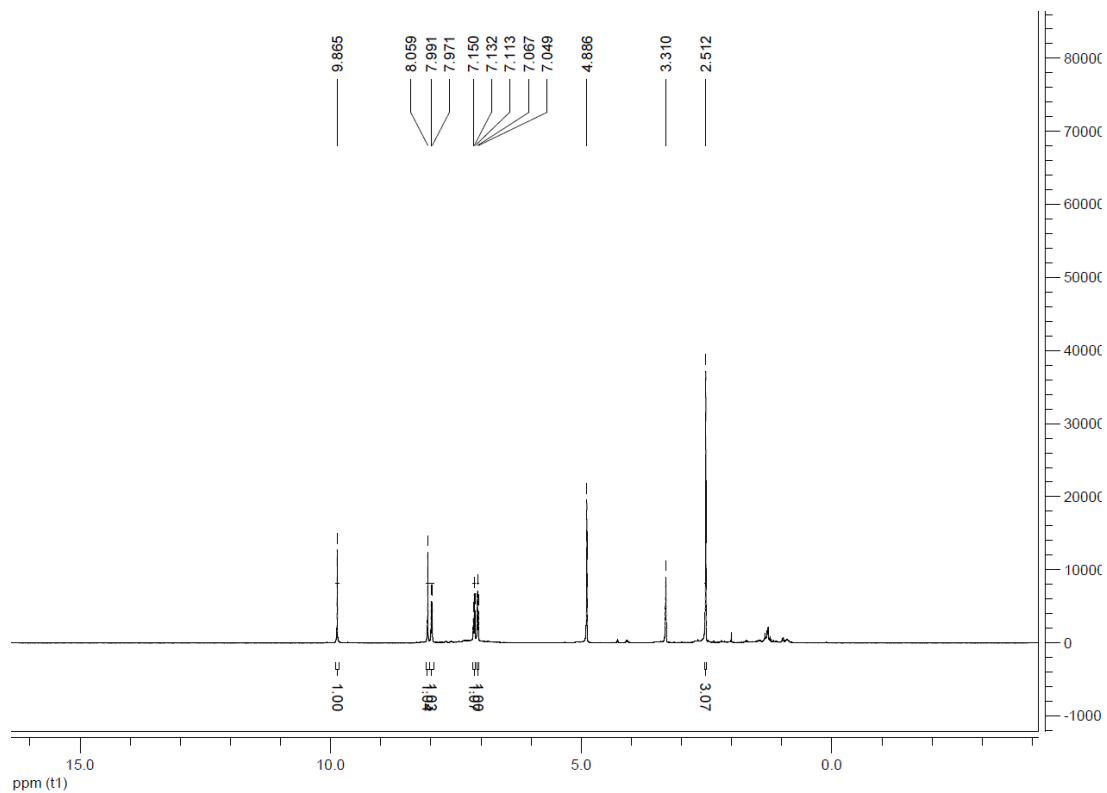


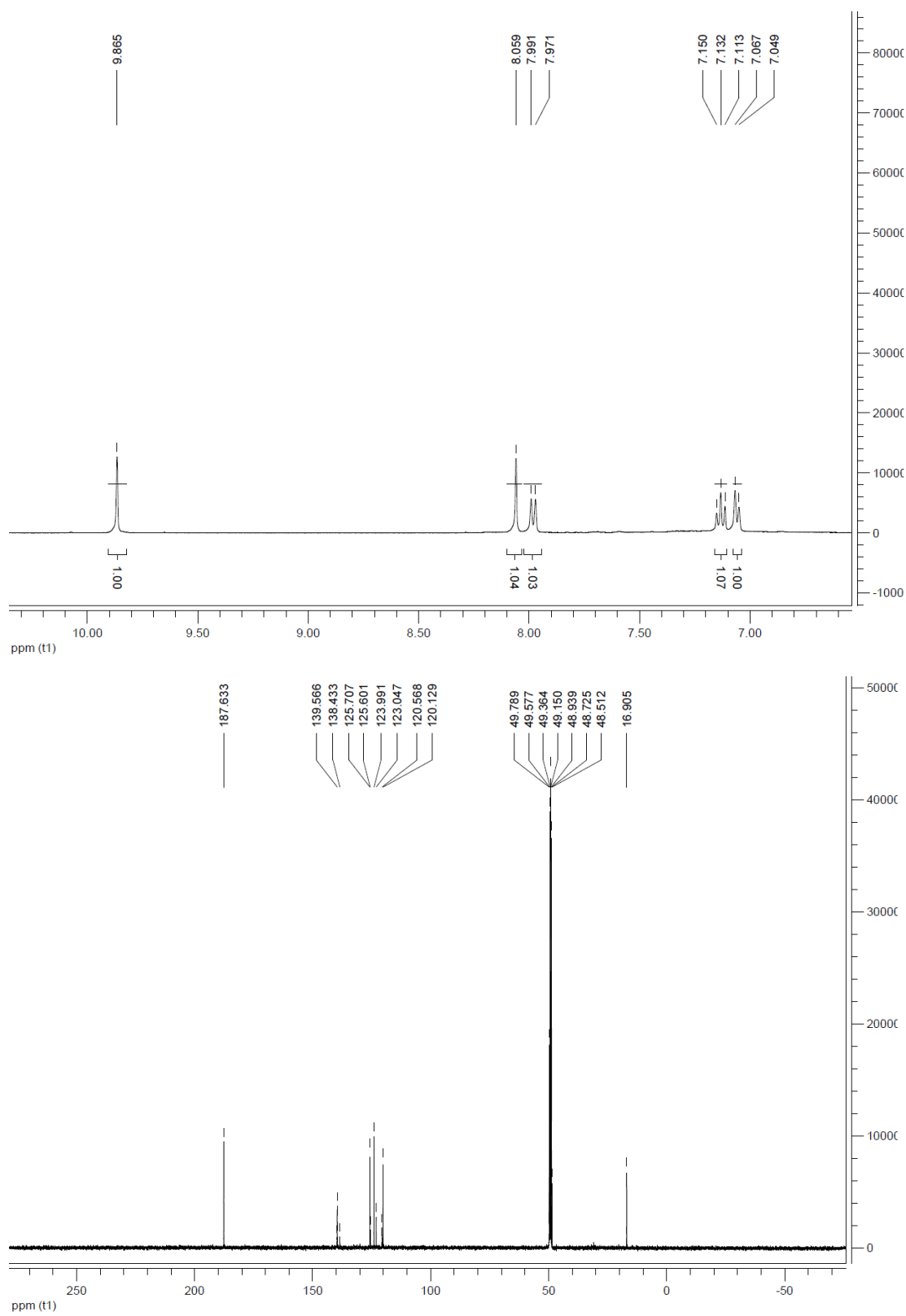




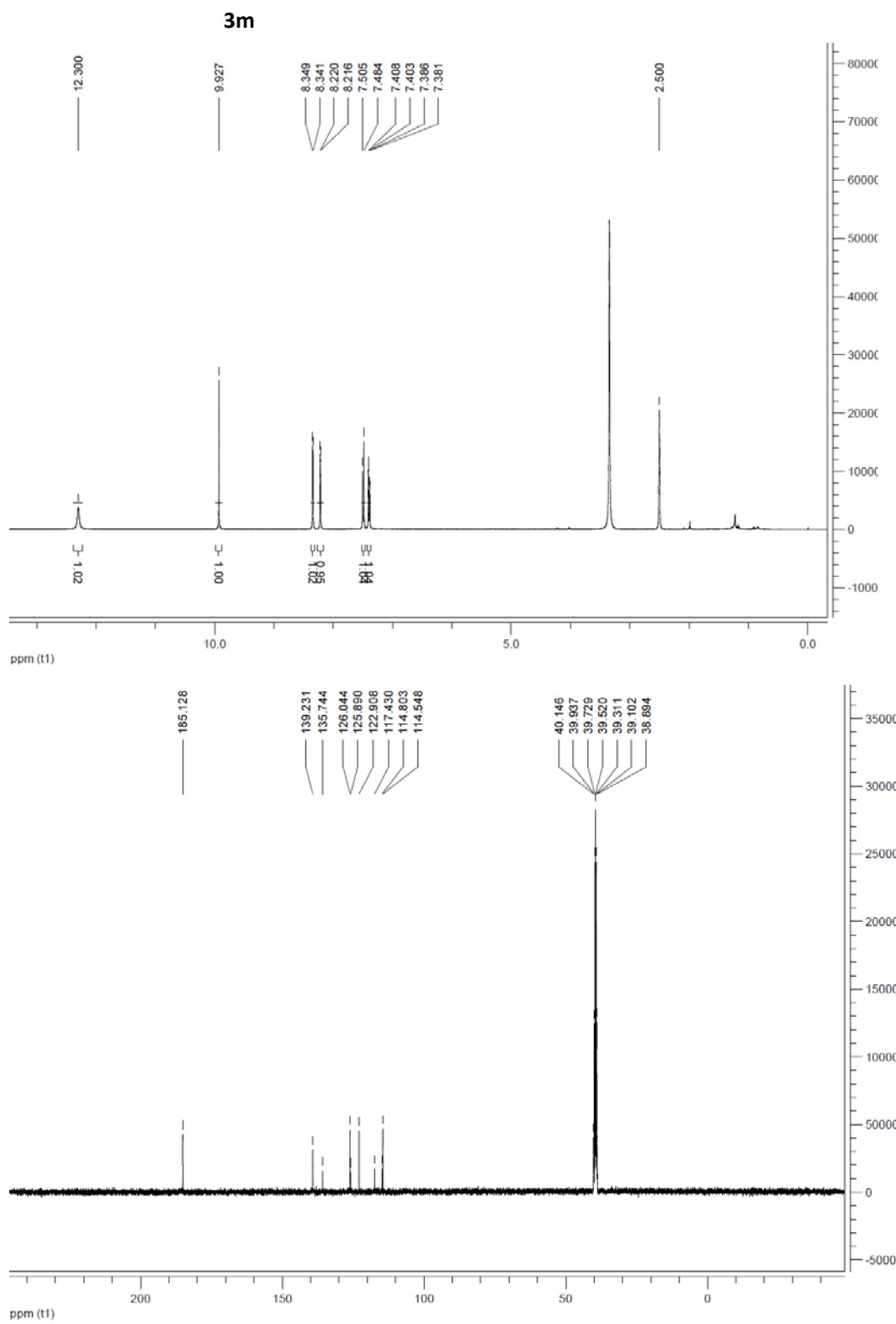


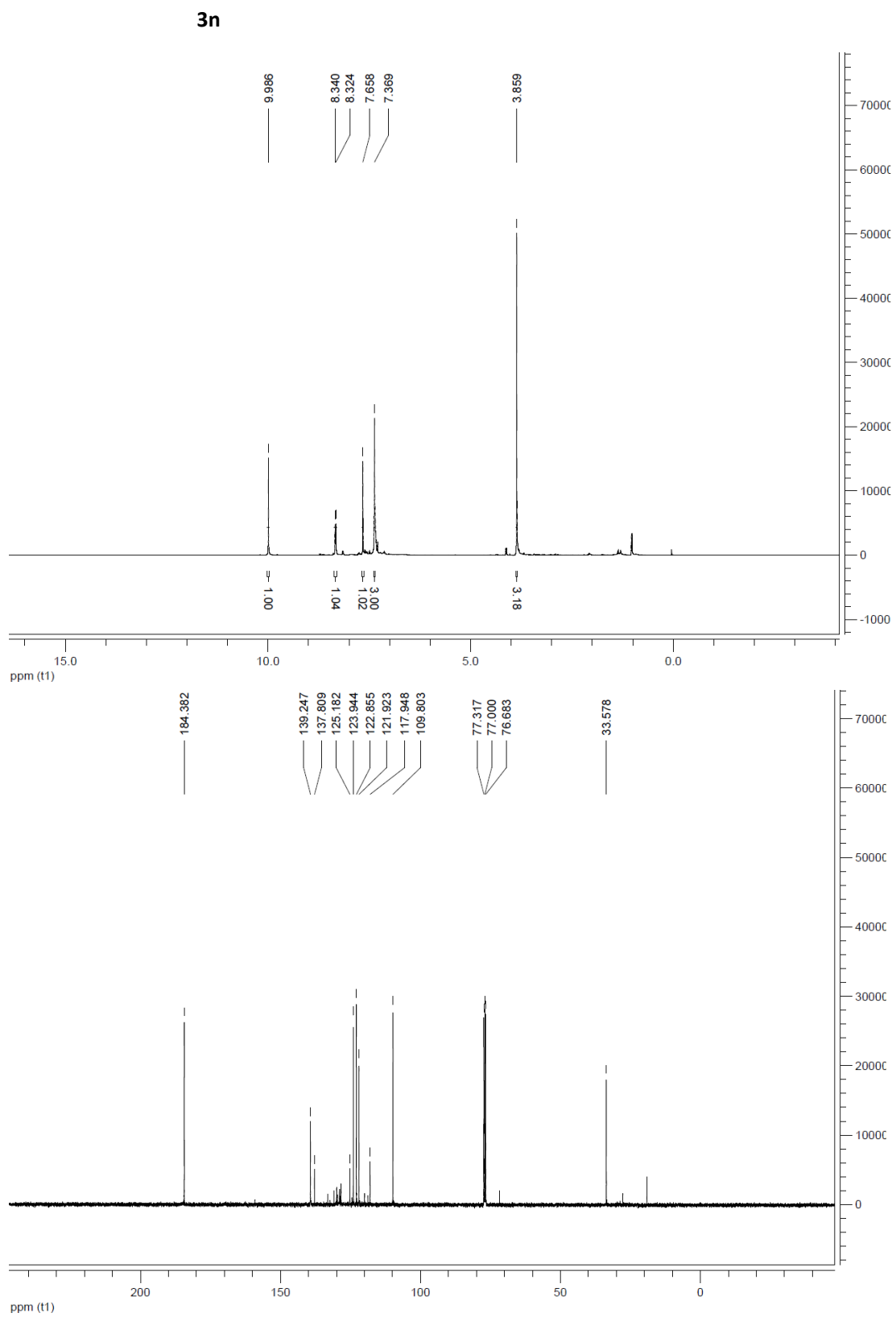
31 in methanol-*d*₄



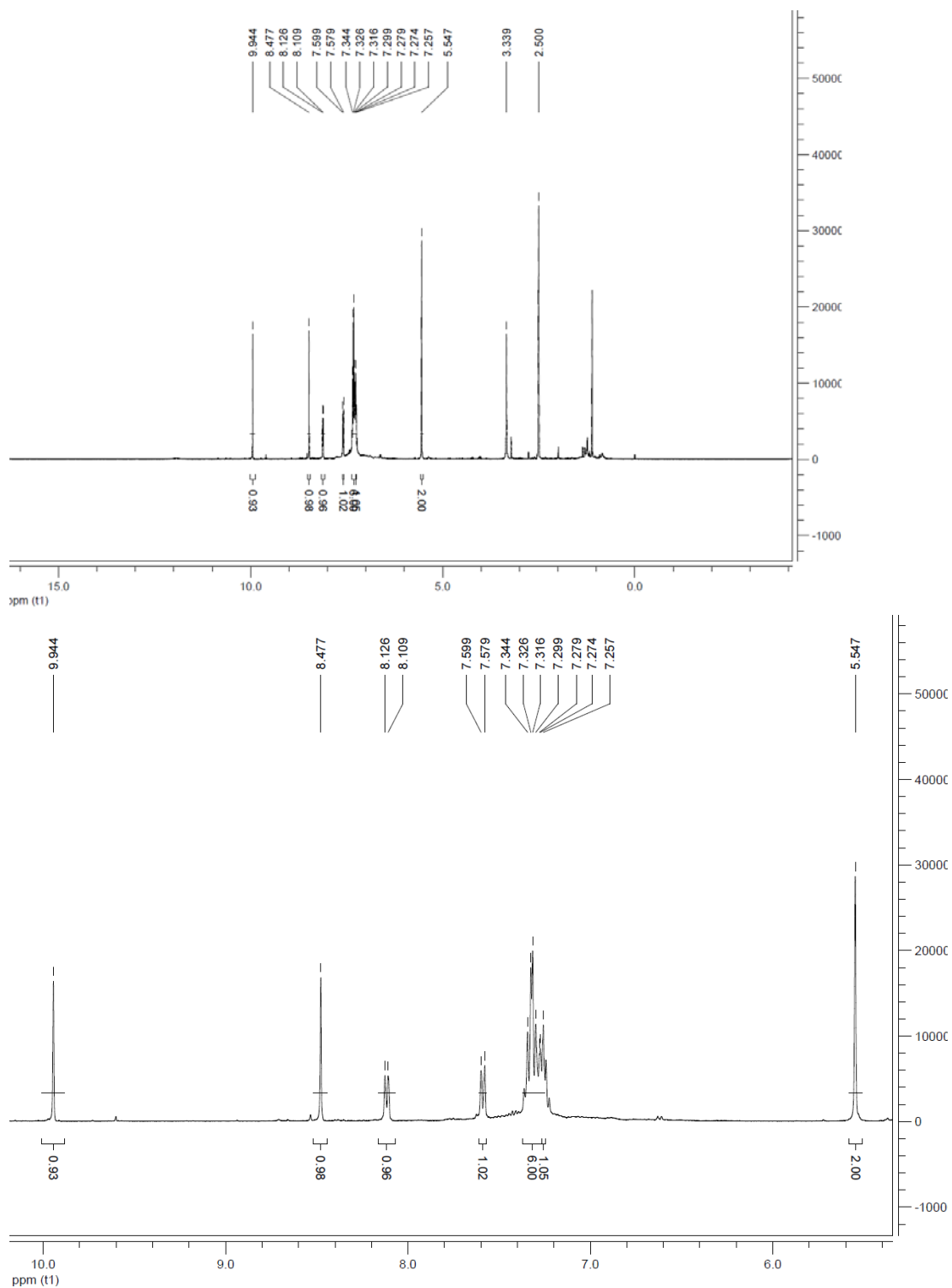


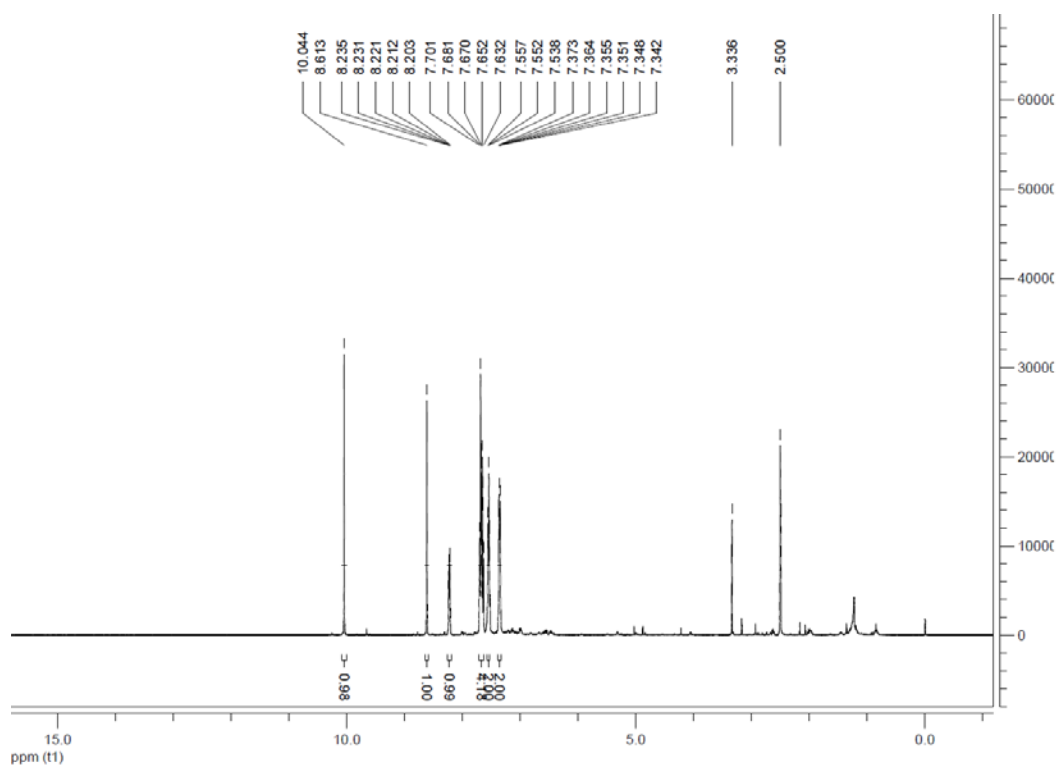
3I in DMSO-*d*₆



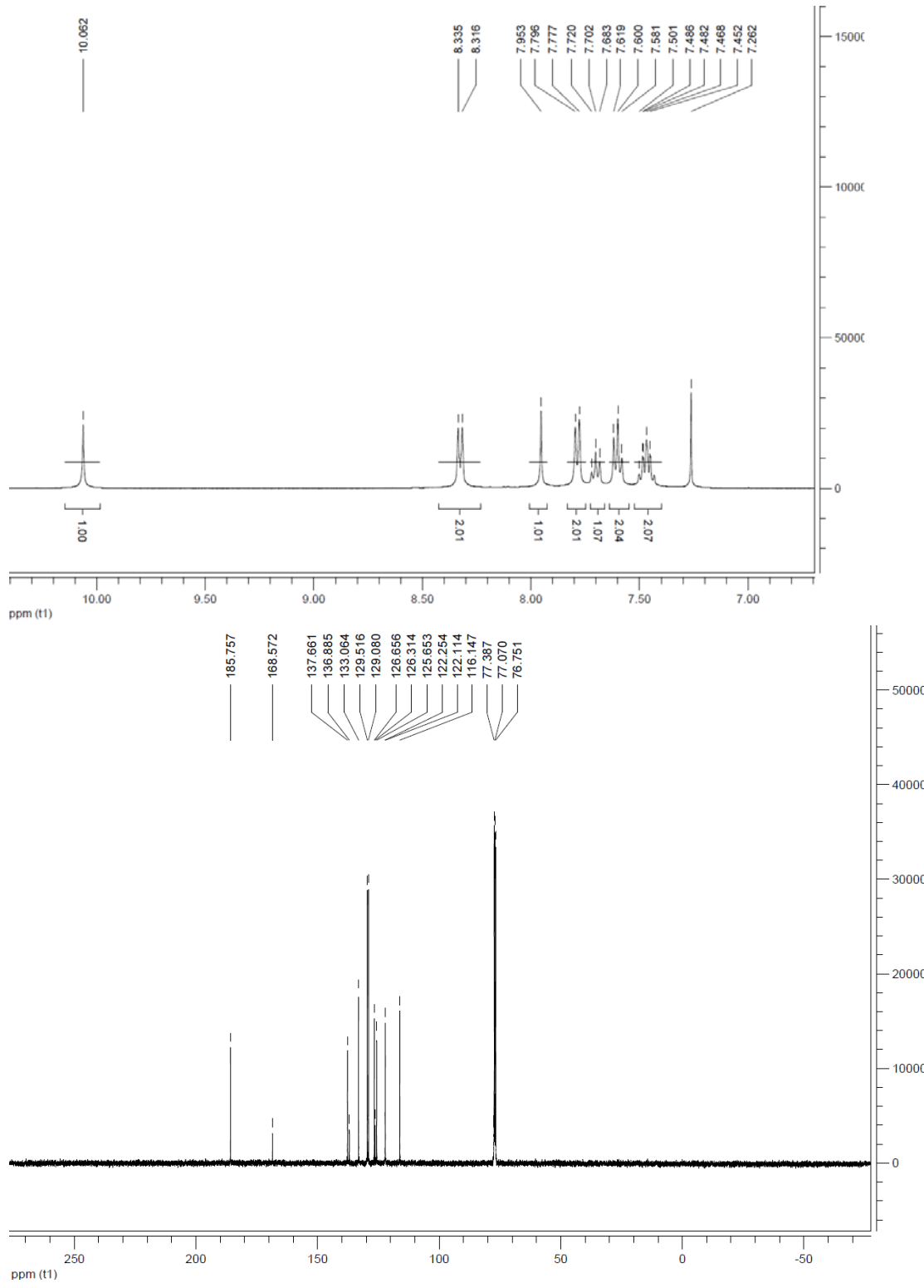


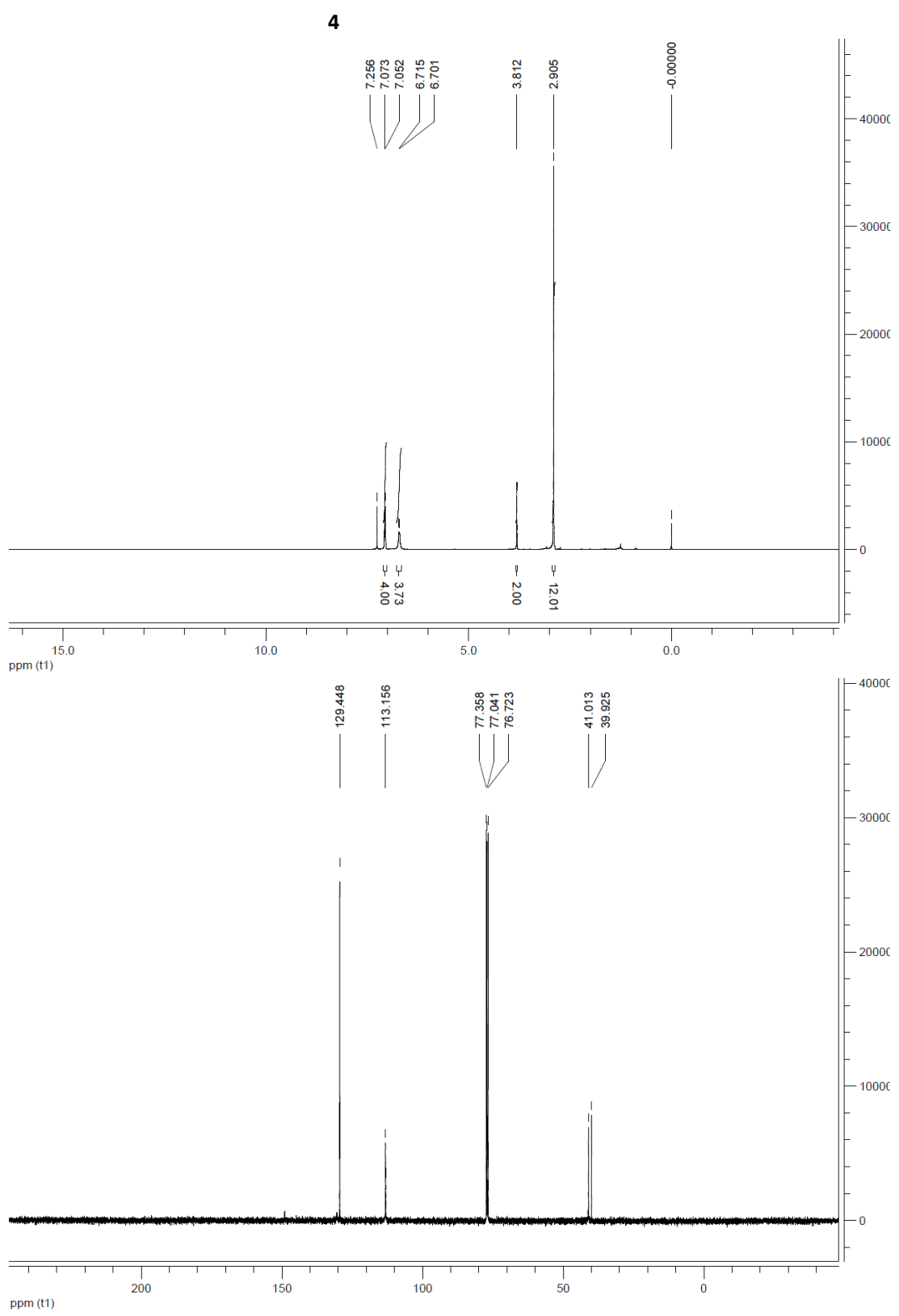
30











7. References

1. W. Wu and W. Su, *J. Am. Chem. Soc.*, 2011, **133**, 11924.
2. S.-C. Lin, F.-D. Yang, J.-S. Shiue, S.-M. Yang and J.-M. Fang, *J. Org. Chem.*, 1998, **63**, 2909.
3. D. F. Cummings, D. C. Canseco, P. Sheth, J. E. Johnson and J. A. Schetz, *Bioorg. Med. Chem.*, 2010, **18**, 4783.
4. H. Ueda, H. Satoh, K. Matsumoto, K. Sugimoto, T. Fukuyama and H. Tokuyama, *Angew. Chem. Int. Ed.*, 2009, **48**, 7600.
5. M. G. Bursavich, N. Brooijmans, L. Feldberg, I. Hollander, S. Kim, S. Lombardi, K. Park, R. Mallon and A. M. Gilbert, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 2586.
6. N. R. Penthala, T. R. Yerramreddy and P. A. Crooks, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 1411.
7. C. Sagnes, G. Fournet and B. Joseph, Synlett, 2009, 433.
8. R. Aggarwal, F. Benedetti, F. Berti, S. Buchini, A. Colombatti, F. Dinon, V. Galasso and S. Norbedo, *Chem. Eur. J.*, 2003, **9**, 3132.
9. S. Murata, M. Miura and M. Nomura, *J. Org. Chem.*, 1989, **54**, 4700.