# - Electronic Supplementary Information -

# Adsorption and separation of poly-aromatic hydrocarbons by a hydrogen-bonded coordination polymer

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### **Experimental Section**

#### **Materials and Instrumentations**

All chemicals and solvents were purchased from Kanto Chemical Co., Ltd., Wako Pure Chemical Co., Ltd., and Tokyo Kasei Kogyo Co., Ltd., and were used as received without further purification. <sup>1</sup>H NMR (500MHz) spectra were recorded on a JEOL  $\alpha$ -500 spectrometer. Chemical shifts are quoted as parts per million (ppm) relative to tetramethylsilane (CDCl<sub>3</sub>) and dimethylsulfoxide (DMSO-*d*<sub>6</sub>). X-ray powder diffraction (XRD) patterns were collected on a Rigaku Multi-Flex X-ray diffractometer using graphite-monochromatized Cu K $\alpha$  radiation ( $\lambda$  = 1.5418 Å) with a scanning rate of 0.020 ° sec<sup>-1</sup> at room temperature.

#### X-ray Crystallography

X-ray crystallographic data of an inclusion compound of pyrene ( $H_{II} \supseteq$ (PY)) were collected on a Rigaku RAXIS-RAPID imaging plate area detector using graphite-monochromatized Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å) at 103 K. The crystal structures were solved by direct method using the *SHELXS-97* program and refined by successive differential Fourier syntheses and full-matrix least-squares procedures using the *SHELXL-97* program.<sup>1</sup> Anisotropic thermal factors were applied to all non-hydrogen atoms.

**Crystal data for H**<sub>II</sub> $\supset$ (PY):  $M_r = 1044.4$ , green block crystal, crystal dimensions  $0.32 \times 0.18 \times 0.09$  mm<sup>3</sup>, *Triclinic*, space group  $P_{\overline{1}}$  (#2), Z = 2, F(000) = 1067.8,  $2\theta_{max} = 55.8$  ° were a = 11.1430(6), b = 14.1170(8), c = 14.3720(8) Å, a = 98.177(1),  $\beta = 97.785(1)$ ,  $\gamma = 107.030(1)$  °, and V = 2101.89(10) Å<sup>3</sup>. A total of 12860 reflections were collected of which 9093 reflections were independent ( $R_{int} = 0.018$ ). The structure was refined to final  $R_1 = 0.039$  for 7541 data [ $I > 2\sigma(I)$ ] with 602 parameters, wR2 = 0.090 for all data, GOF = 1.038, and residual electron density max/min = 0.592/-0.334 eÅ<sup>-3</sup>. CCDC 867556.



*Fig.* S1. X-ray crystal structure of  $H_c \supset (NA)$  viewed along the *a* axis (left) and the *b* axis (right).<sup>2</sup> Colour scheme: blue (hydrogen-bonded CPs), green (NA). Red dotted lines denote hydrogen bonds between the carboxyl functional groups of the isoH ligands.



*Fig.* S2. (a) Time course experiment of the thermal treatment of  $H_c \supset (NA)$  at 140 °C in vacuo. ( $\circ$ ) denotes the host-to-guest ratio determined by <sup>1</sup>H NMR spectroscopy. (Host = [Ni(SCN)<sub>2</sub>(isoH)<sub>2</sub>], Guest = NA) Reaction time: (i) 0, (ii) 4 and (iii) 62 h. (b) X-ray powder diffraction (XRD) patterns of (i), (ii) and (iii). (c) <sup>1</sup>H NMR spectra (500 MHz, DMSO-*d*<sub>6</sub>, rt) of (i) and (iii). The signals of the isoH ligand are broadened due to coordination of the isoH ligand to the paramagnetic Ni<sup>2+</sup> center. ( $\circ$ ) denotes spinning side band.

Previously, we reported the TG analysis of  $\mathbf{H_c} \supset (NA)$ .<sup>2</sup>  $\mathbf{H_c} \supset (NA)$  showed an initial weight loss of 11% at 150 °C, corresponding to release of NA from the channels (13%, calculated). This means that 15 % of NA remained inside of residual-host  $\mathbf{H_{res}}$ . This is in good agreement of the time course experiment. Fig. S2(a) shows that 12 – 13% of NA remained inside of  $\mathbf{H_{res}}$ . Most likely, this is due to the change of the crystal structure upon removal of NA (Fig. S2b (i) – (iii)), which physically obstructs removal of remaining NA.



*Fig.* **S3.** <sup>1</sup>H NMR spectra (500 MHz, DMSO- $d_6$ , rt) of (a)  $\mathbf{H}_{res} \supset (BI)$ , (b)  $\mathbf{H}_{res} \supset (AZ)$  and (c)  $\mathbf{H}_{res} \supset (PY)$ . The host-to-guest ratios were determined by the relative signal intensities of remaining NA and the guest. ( $\circ$ ) denotes spinning side band.



*Fig.* **S4.** (a) Observed XRD pattern of  $H_c \supseteq$  (NA). (b), (c) and (d) observed XRD patterns of the samples prepared by immersing  $H_c \supseteq$  (NA) in CH<sub>2</sub>Cl<sub>2</sub> solutions containing three equiv. (b) BI, (c) AZ or (d) PY at rt for 12 h.



*Fig.* **S5.** <sup>1</sup>H NMR spectra (500 MHz, DMSO- $d_6$ , rt) of (a)  $\mathbf{H}_{res} \supset$  (CH) and (b)  $\mathbf{H}_{res} \supset$  (BE). The host-toguest ratios were determined by the relative signal intensities of remaining NA and the guest.



*Fig.* S6. <sup>1</sup>H NMR spectra (500 MHz, CDCl<sub>3</sub>, rt) of the CDCl<sub>3</sub> solutions after filtration of (a)  $H_{res} \supset (BE)$  and (b)  $H_{res} \supset (CH)$ .  $H_{res} \supset (CH)$  and (b)  $H_{res} \supset (CH)$  were immersed in the CDCl<sub>3</sub> solutions at rt for a week. ( $\circ$ ) denotes spinning side band.



*Fig.* **S7.** Observed XRD patterns of (a)  $H_c \supseteq$ (BI) and (b)  $H_{res} \supseteq$ (BI).



*Fig.* **S8.** <sup>1</sup>H NMR spectra (500 MHz, DMSO- $d_6$ , rt) of (a)  $H_c \supset (NA)$ , (b)  $H_{res} \supset (PY)$  and (c) isolated PY. Reaction conditions: i) 1 mol L<sup>-1</sup> of HCl (3 mL), ii) extraction of the adsorbed guests with CH<sub>2</sub>Cl<sub>2</sub> (3 mL × 3) and iii) removal of co-existing NA under reduced pressure. Green and red circles denote NA and PY, respectively. ( $\blacktriangle$ ) denotes DMSO.



*Fig.* **S9.** Observed XRD patterns of (a)  $H_{res} \supset (NA)$ , (b)  $H_{res} \supset (PY)$  and (c)  $H_{res} \supset (PY>NA)$ .



*Fig.* **S10.** <sup>1</sup>H NMR spectra (500 MHz, DMSO- $d_6$ , rt) of (a)  $H_{res} \supset (PY>NA)$  and (b)  $H_{res} \supset (NA>PY)$ . Green and red circles denote NA and PY, respectively. (**■**) denotes the isoH ligand.



*Fig. S11.* Observed XRD patterns of (a)  $H_{res}$ , (b)  $H_c \supset (NA)$  and (c) the sample prepared by co-grinding of  $H_{res}$  and three equiv. NA for 1 min followed by standing at rt for 30 min. The green circles denote the diffraction peaks of NA.



**Fig. S12.** Observed XRD patterns of (a)  $\mathbf{H}_{res}$ , (b)  $\mathbf{H}_{res} \supset$  (BI) and (c) the sample prepared by co-grinding of  $\mathbf{H}_{res}$  (30 mg) and three equiv. BI for 1 min followed by standing at rt for 30 min. The blue circles denote the diffraction peaks of BI.



*Fig.* **S13.** Observed XRD patterns of (a) a mixture of  $\mathbf{H}_{res} \supset (BI)' + BI$  (see also Fig. S12c) and (b)  $\mathbf{H}_{res} \supset (BI)'$  that was obtained by thermal treatment of the mixture of  $\mathbf{H}_{res} \supset (BI)' + BI$  at 50 °C for 6 h. The blue circles denote the diffraction peaks of BI.



*Fig.* **S14.** Observed XRD patterns of (a)  $\mathbf{H}_{res}$ , (b)  $\mathbf{H}_{res} \supset (AZ)$  and (c) the sample prepared by cogrinding of  $\mathbf{H}_{res}$  and three equiv of AZ for 5 min followed by standing at rt for 30 min. The brown circles denote the diffraction peaks of AZ.



**Fig. S15.** Observed XRD patterns of (a)  $H_{res}$ , (b)  $H_{res} \supset (PY)$  and (c) the sample prepared by cogrinding of  $H_{res}$  and three equiv of PY for 5 min followed by standing at rt for 30 min. The red circles denote the diffraction peaks of PY.



*Fig.* **S16.** Observed XRD patterns of (a)  $\mathbf{H}_{res}$ , (b)  $\mathbf{H}_{res} \supset (PY)$ , (c)  $\mathbf{H}_c \supset (NA)$  and (d) the sample prepared by co-grinding of  $\mathbf{H}_{res}$  with three equiv. NA and PY for 5 min followed by standing at rt for 30 min. The green and red circles denote the diffraction peaks of NA and PY, respectively.



*Fig.* **S17.** Observed XRD patterns of the sample prepared by exposure of  $H_{res}$  to the saturated vapour of NA in a Petri dish at rt for (a) 0, (b) 12 and (c) 24 h.



*Fig.* **S18.** Observed XRD patterns of the sample prepared by exposure of  $H_{res}$  to the saturated vapour of BI in a Petri dish at rt for (a) 0 and (b) 24 h.

## References

#### References

- (1) G. M. Sheldrick, Acta Cryst., A, 2008, 64, 112.
- (2) R. Sekiya, S. Nishikiori and K. Ogura, J. Am. Chem. Soc., 2004, 127, 16587–16600.