

Supporting Information for Design of Chiral Sulfoxide–Schiff Base Hybrids and Their Application in Cu-Catalyzed Asymmetric Henry Reactions

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1. General Information

Unless otherwise noted, materials were purchased from commercial suppliers and used without further purification. All the solvents were treated according to general methods. Flash column chromatography was performed using 200-300 mesh silica gel. ¹H NMR spectra were recorded on 400 MHz spectrophotometers. Chemical shifts (δ) are reported in ppm from the solvent resonance as the internal standard (CDCl₃: 7.26 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, dd = doublet of doublets, m = multiplet), coupling constants (Hz) and integration. ¹³C NMR spectra were recorded on 100 MHz with complete proton decoupling spectrophotometers (CDCl₃: 77.0 ppm). Mass spectra were measured on a MS spectrometer. Elemental analysis was taken on an elementary analysis instrument. Enantiomeric ratios were determined by chiral HPLC with chiral columns (chiralpak AS-H column, chiralpak AD-H column, chiralpak OJ-H column or chiralcel OD-H column) with hexane and *i*-PrOH as solvents. Optical rotations were measured with a polarimeter.

2. Detailed Optimizations of Reaction Conditions

Table 1S. Metal salt screening for the asymmetric Henry reaction^[a]

Entry	Metal salt	Ligand	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	Cu(OAc)₂•H₂O	1a	36	88	80
2	Cu(OAc) ₂	1a	36	85	81
3	CuBr ₂	1a	96	trace	-
4	CuCl ₂ •2H ₂ O	1a	96	trace	-
5	CuSO ₄	1a	96	trace	-
6	CuBr	1a	96	trace	-
7	Cu(OTf) ₂	1a	96	trace	-
8	AgOAc	1a	36	69	0
9	Co(OAc) ₂	1a	36	90	0
10	Zn(OAc) ₂ •H ₂ O	1a	36	93	0
11	Cu(OAc) ₂ •H ₂ O	1a	240	94	0
12	-	1a	96	trace	-
13	-	-	96	trace	-

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a** (0.6 mL), Metal salt/**1a** (0.04 mmol), in *i*-PrOH (4 mL) at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC, the absolute configuration was established as *R* by comparison with literature data.

As shown in Table 1S, among the metal salts tested, the Cu(OAc)₂•H₂O gave the best result in terms of yield and enantioselectivity (entry 1), and was thus selected for further optimization studies.

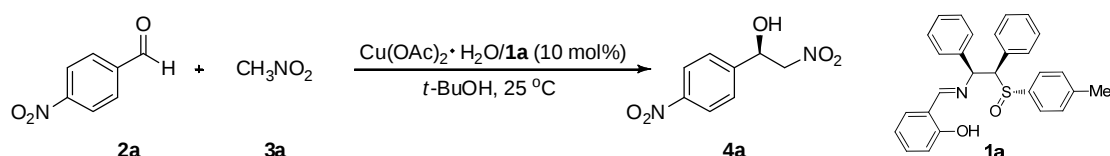
Table 2S. Solvent screening for the asymmetric Henry reaction^[a]

Entry	Solvent	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	<i>i</i> -PrOH	36	88	80
2	MeOH	48	90	38
3	EtOH	40	85	70
4	<i>t</i>-BuOH	24	94	85
5	DMF	20	94	20
6	MeCN	74	60	70
7	MeNO ₂	56	66	77
8	CH ₂ Cl ₂	70	42	68
9	toluene	72	21	53
10	THF	68	55	0

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a** (0.6 mL), Cu(OAc)₂•H₂O/**1a** (0.04 mmol) in solvent (4 mL) at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC.

As shown in Table 2S, among the solvents tested, the *t*-BuOH gave the best result in terms of yield and enantioselectivity (entry 4), and was thus selected for further optimization studies.

Table 3S. Nitromethane loading screening for the asymmetric Henry reaction^[a]

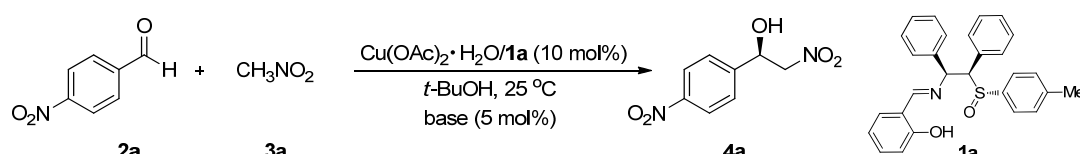


Entry	Nitromethane (mL)	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	2.0	24	89	87
2	1.2	24	92	87
3	1.0	24	94	87
4	0.9	24	94	86
5	0.6	24	94	85
6	0.5	24	95	83
7	0.4	24	91	82
8	0.2	32	91	84
9	0.1	46	76	82

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a**, Cu(OAc)₂·H₂O/**1a** (0.04 mmol) in *t*-BuOH (4 mL) at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC.

As shown in Table 3S, among the nitromethane loading tested, 1.0 mL of nitromethane gave the best result in terms of yield and enantioselectivity (entry 3), and was thus selected for further optimization studies.

Table 4S. Screening the base for the asymmetric Henry reaction^[a]

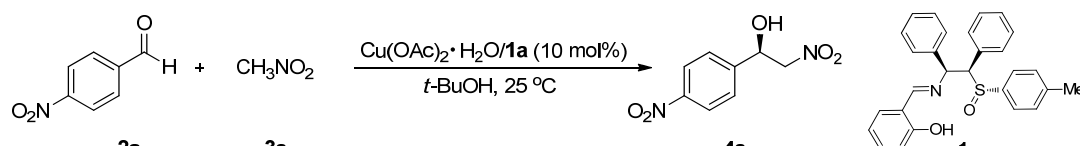


Entry	Base	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	Et ₃ N	11	92	70
2	Py	11.5	91	82
3	DIPEA	12	93	69
4	DMAP	12.5	91	60
5	DBU	13.5	96	74
6	-	24	94	87

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a** (1.0 mL), Cu(OAc)₂·H₂O/**1a** (0.04 mmol) in *t*-BuOH (4 mL) at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC.

As shown in Table 4S, the result in entry 6 was the best in terms of enantioselectivity, and was thus selected for further optimization studies.

Table 5S. Concentration screening for the asymmetric Henry reaction^[a]



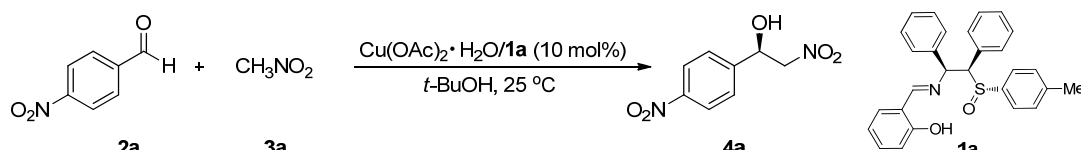
Entry	Concentration (M)	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	0.4	21.5	91	84
2	0.2	22	95	85
3	0.1	24	94	87
4	0.05	42	88	72
5 ^d	0.1	19	93	87

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a** (1.0 mL), Cu(OAc)₂·H₂O/**1a** (0.04 mmol) in *t*-BuOH at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC.

[d] 4 Å MS (100 mg) was added.

As shown in Table 5S, the result in entry 3 was the best in terms of yield and enantioselectivity, and was thus selected for further optimization studies.

Table 6S. Screening of equiv. of ligand for the the asymmetric Henry reaction^[a]

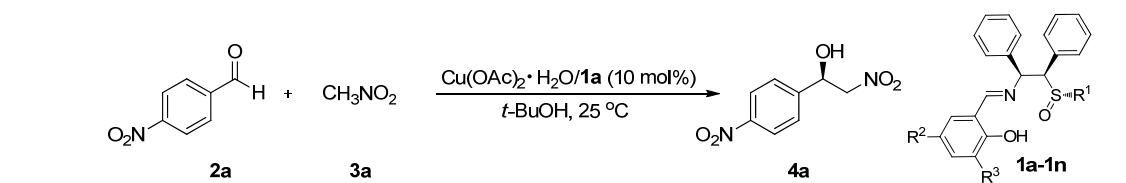


Entry	Equiv. of 1a	t (h)	Yield (%) ^[b]	ee (%) ^[c]
1	0.10	26	94	85
2	0.11	26	88	86
3	0.12	25	96	88
4	0.13	25	94	86
5 ^d	0.15	24	96	88
5 ^d	0.20	24	95	89

[a] Unless otherwise noted, reactions were carried out with **2a** (0.40 mmol), **3a** (1.0 mL), Cu(OAc)₂·H₂O (0.04 mmol) and **1a** in *t*-BuOH (4.0 mL) at 25 °C. [b] Yield of isolated product. [c] Determined by chiral HPLC.

As shown in Table 6S, among the equiv. of ligand tested, the 0.12 equiv. of ligand gave the best result (entry 3), and was thus selected for further studies.

Table 7S. Screening of ligands for the the asymmetric Henry reaction^[a]



Entry	Ligand	R ¹	R ²	R ³	Time (h)	yield (%) ^b	ee (%) ^c
1	1a	4-MePh	H	H	24	95	88
2	1b	Ph	H	H	24	92	88
3	1c	4-BrPh	H	H	24	91	87
4	1d	3,5-Me ₂ Ph	H	H	24	94	80
5	1e	2-OMePh	Cl	H	108	53	24
6	1f	Bn	H	H	20	96	90
7	1g	4-MePh	Cl	H	32	92	90
8	1h	4-MePh	CH ₃	H	24	93	86
9	1i	4-MePh	Cl	Cl	48	90	90
10	1j	4-MePh	<i>t</i> -butyl	<i>t</i> -butyl	24	94	80
11	1k	2-naphthyl	Cl	H	28	88	88
12	1l	4-MePh	Br	H	28	82	87
13	1m	4-MePh	F	H	28	91	89
14	1n	4-MePh	Br	Br	48	82	92
15^d	1f	Bn	H	H	30	94	93

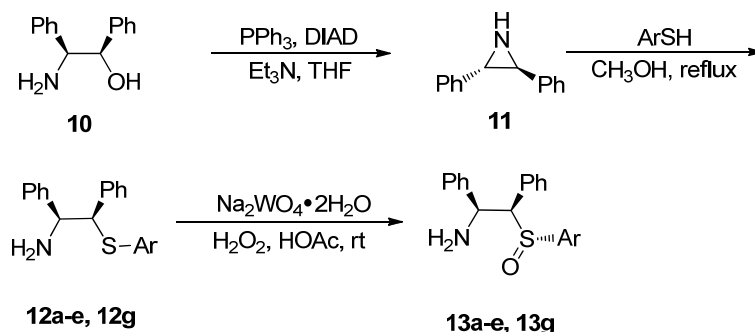
^a Unless otherwise noted, the reaction was carried out with **2a** (0.4 mmol), **3a** (1.0 mL), Cu(OAc)₂·H₂O (0.04 mmol) and **1** (0.048 mmol) in *t*-BuOH (4.0 mL) at 25 °C. ^b Isolated yield. ^c Determined by chiral HPLC, the absolute configuration was established as *R* by comparison with literature data. ^d 2.5 mol% of Cu(OAc)₂·H₂O and 3 mol% of **1f** were used.

As shown in Table 7S, among the ligands tested, 2.5 mol% of ligand **1f** gave the best result (entry 15), and was thus selected for further studies.

3. General Procedures for the Synthesis of Ligands

3.1 General Procedure for the Preparation of Aminosulfoxides 13

(1) Synthesis of Aminosulfoxides 13a-13e, 13g



a) Mitsunobu reaction

(1R,2S)-2-amino-1,2-diphenylethanol (4.26 g, 20 mmol) was treated with Ph_3P (6.30 g, 24 mmol), Et_3N (8.40 mL) and diisopropyl azodicarboxylate (DIAD, 4.45 g, 22 mmol) in THF (60 mL) at room temperature for 20 h. After the solids were filtered, the filtrate was concentrated and purified by flash chromatography to give (2S,3S)-2,3-diphenylaziridine **11** as a white solid, 3.48 g, yield: 89%.

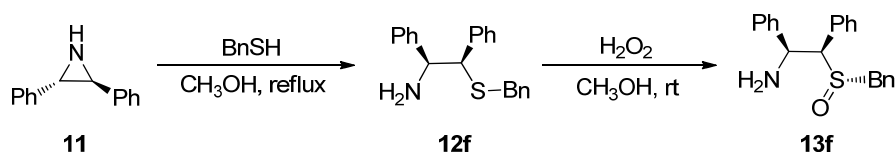
b) Ring opening reaction

Benzenethiol (11 mmol) was added to a solution of aziridine **11** (1.95 g, 10 mmol) in methanol. The solution was heated to reflux for 8 h. After the solvent was evaporated under vacuum, the crude product was purified by flash chromatography to give **12**.

c) Oxidation reaction

To a solution of **12** (8.5 mmol) and $\text{Na}_2\text{WO}_4 \cdot 2\text{H}_2\text{O}$ (85 mg) in HOAc (8.5 mL), H_2O_2 (30% in water, 0.85 mL, 8.5 mmol) was added dropwisely. The mixture was stirred for another 2 min. Then the reaction mixture was quenched with water, extracted with CH_2Cl_2 (3*50 mL), and the organic layer was washed with Sat. NaHCO_3 (aq.), dried over Na_2SO_4 . After the solvent was evaporated under vacuum, the crude product was purified by flash chromatography to give a mixture of two diastereomeric aminosulfoxides. The major diastereoisomer **13** was obtained in pure form after a simple recrystallization with petroleum ether and ethanol as a white solid.

(2) Synthesis of Aminosulfoxide 13f^[1]



b) Ring opening reaction

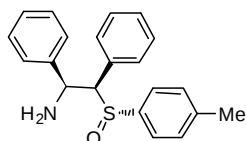
Phenylmethanethiol (1.86g, 15 mmol) was added to a solution of aziridine **11** (1.95 g, 10 mmol) in methanol. The solution was heated to reflux for 5 days. After the solvent was evaporated under vacuum, the crude product was purified by flash chromatography to give **12f** as a white solid, 2.07 g, yield: 65%.

c) Oxidation reaction

To a solution of **12f** (2.07g, 6.5 mmol) in methanol (10 mL) at room temperature, H_2O_2 (30% in water, 0.65 mL, 6.5 mmol) was added dropwisely. The mixture was stirred at room temperature for 3 days. Then the solvent was evaporated under vacuum, the crude product was purified by flash chromatography to give a mixture of two diastereomeric aminosulfoxides. The major diastereoisomer **13f** was obtained in pure form after a simple recrystallization with petroleum ether and ethanol as a white solid, 0.85 g, yield: 39%.

3.2 Characterization Data of Aminosulfoxides 13

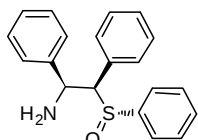
(1S,2R)-1,2-diphenyl-2-((R)-*p*-tolylsulfinyl)ethanamine (13a)



N, 4.446; S, 9.801.

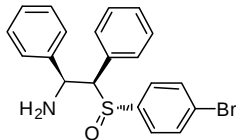
White solid, 47% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.27 – 7.23 (m, 1H), 7.20 – 7.14 (m, 7H), 7.06 (d, J = 8.2 Hz, 2H), 7.00 – 6.94 (m, 4H), 5.02 (d, J = 3.1 Hz, 1H), 3.79 (d, J = 3.6 Hz, 1H), 2.29 (s, 3H), 1.82 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.43, 141.13, 139.46, 130.88, 130.47, 129.15, 128.19, 128.14, 128.05, 127.14, 126.71, 124.70, 78.91, 53.93, 21.27. MS: m/z = 335.04 (M^+). Anal. calcd for ($\text{C}_{21}\text{H}_{21}\text{NOS}$): C, 75.19; H, 6.31; N, 4.18; S, 9.56; found: C, 75.41; H, 6.146;

(1S,2R)-1,2-diphenyl-2-((R)-phenylsulfinyl)ethanamine (13b)



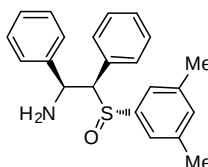
White solid, 49% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.35 – 7.31 (m, 1H), 7.27 – 7.24 (m, 5H), 7.20 – 7.15 (m, 5H), 7.00 – 6.94 (m, 4H), 5.03 (d, J = 3.3 Hz, 1H), 3.80 (d, J = 3.6 Hz, 1H), 1.83 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.76, 142.30, 130.69, 130.39, 128.34, 128.22, 128.10, 128.04, 127.12, 126.64, 124.61, 78.97, 53.82. MS: m/z = 321.14 (M^+). Anal. calcd for ($\text{C}_{20}\text{H}_{19}\text{NOS}$): C, 74.73; H, 5.96; N, 4.36; S, 9.98; found: C, 74.77; H, 5.818; N, 4.539; S, 9.752.

(1S,2R)-2-((R)-(4-bromophenyl)sulfinyl)-1,2-diphenylethanamine (13c)



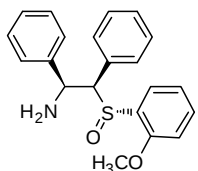
White solid, 52% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.39 – 7.36 (m, 2H), 7.31 – 7.27 (m, 1H), 7.22 – 7.18 (m, 5H), 7.10 – 7.07 (m, 2H), 6.99 – 6.95 (m, 4H), 5.01 (d, J = 3.5 Hz, 1H), 3.76 (d, J = 3.7 Hz, 1H), 1.82 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.19, 142.15, 131.56, 130.46, 130.42, 128.56, 128.31, 128.24, 127.30, 126.67, 126.23, 125.22, 79.23, 53.90. MS: m/z = 400.06 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{20}\text{H}_{18}\text{BrNOS}$): C, 60.00; H, 4.53; N, 3.50; S, 8.01; found: C, 60.11; H, 4.567; N, 3.389; S, 8.263.

(1S,2R)-2-((R)-(3,5-dimethylphenyl)sulfinyl)-1,2-diphenylethanamine (13d)



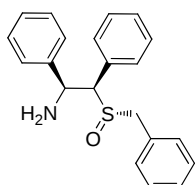
White solid, 41% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.28 – 7.24 (m, 1H), 7.21 – 7.16 (m, 5H), 7.02 – 6.93 (m, 5H), 6.81 (s, 2H), 5.01 (d, J = 2.4 Hz, 1H), 3.78 (d, J = 3.7 Hz, 1H), 2.17 (s, 6H), 1.83 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 140.78, 138.30, 137.47, 133.00, 130.91, 129.98, 128.97, 128.44, 128.34, 128.06, 127.92, 122.46, 75.74, 56.75, 20.89. MS: m/z = 349.26 (M^+). Anal. calcd for ($\text{C}_{22}\text{H}_{23}\text{NOS}$): C, 75.61; H, 6.63; N, 4.01; S, 9.17; found: C, 75.52; H, 6.910; N, 4.131; S, 9.341.

(1S,2R)-2-((R)-(2-methoxyphenyl)sulfinyl)-1,2-diphenylethanamine (13e)



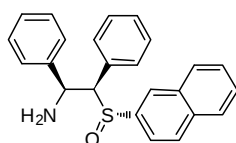
White solid, 45% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.81 (dd, J = 7.7, 1.3 Hz, 1H), 7.41 – 7.36 (m, 1H), 7.29 – 7.21 (m, 3H), 7.18 – 7.09 (m, 6H), 6.88 – 6.86 (m, 2H), 6.77 (d, J = 8.2 Hz, 1H), 4.89 (d, J = 3.5 Hz, 1H), 4.04 (d, J = 3.6 Hz, 1H), 3.61 (s, 3H), 1.85 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 155.45, 142.47, 132.58, 132.18, 130.54, 130.24, 128.00, 127.87, 127.76, 126.96, 126.59, 125.94, 121.37, 110.41, 74.41, 55.28, 53.86. MS: m/z = 351.94 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{21}\text{H}_{21}\text{NO}_2\text{S}$): C, 71.76; H, 6.02; N, 3.99; S, 9.12; found: C, 72.00; H, 5.976; N, 3.791; S, 9.056.

(1S,2R)-2-((S)-benzylsulfinyl)-1,2-diphenylethanamine (13f)



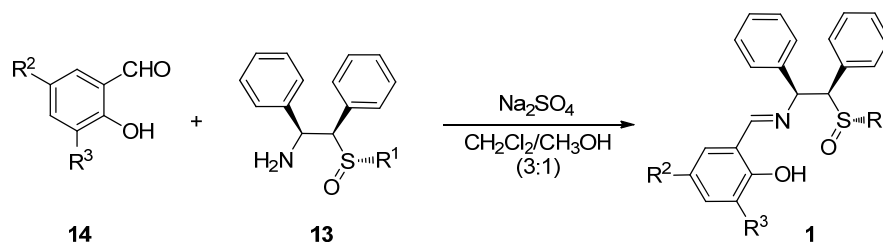
White solid, 39% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.37 – 7.25 (m, 6H), 7.19 – 7.15 (m, 3H), 7.14 – 7.11 (m, 2H), 7.02 (d, J = 7.0 Hz, 2H), 6.90 – 6.88 (m, 2H), 4.88 (d, J = 3.1 Hz, 1H), 3.84 – 3.77 (m, 2H), 3.49 (d, J = 13.4 Hz, 1H), 1.74 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.26, 131.08, 130.28, 130.08, 129.77, 128.58, 128.52, 128.40, 128.00, 127.96, 127.04, 126.57, 72.96, 55.16, 53.52. MS: m/z = 335.8 ($[\text{M}]^+$). Anal. calcd for ($\text{C}_{21}\text{H}_{21}\text{NOS}$): C, 75.19; H, 6.31; N, 4.18; S, 9.56; found: C, 75.21; H, 6.254; N, 4.029; S, 9.664.

(1S,2R)-2-((R)-naphthalen-2-ylsulfinyl)-1,2-diphenylethanamine (13g)



White solid, 48% yield. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.87 (s, 1H), 7.79 – 7.73 (m, 2H), 7.69 (d, J = 8.6 Hz, 1H), 7.53 – 7.47 (m, 2H), 7.28 – 7.24 (m, 1H), 7.18 – 7.14 (m, 6H), 7.01 – 6.97 (m, 4H), 5.07 (d, J = 3.5 Hz, 1H), 3.90 (d, J = 3.6 Hz, 1H), 1.86 (s, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.31, 140.02, 134.13, 132.38, 130.79, 130.52, 128.33, 128.17, 128.14, 127.74, 127.43, 127.20, 126.79, 126.71, 125.31, 120.65, 78.79, 54.00. MS: m/z = 373.23 ($[\text{M}+2]^+$). Anal. calcd for ($\text{C}_{24}\text{H}_{21}\text{NOS}$): C, 77.59; H, 5.70; N, 3.77; S, 8.63; found: C, 77.60; H, 5.590; N, 3.642; S, 8.892.

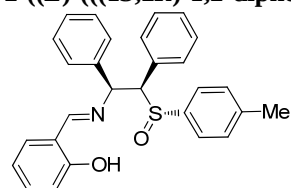
3.3 General Procedure for the Preparation of Ligands



A mixture of dry CH_2Cl_2 /methanol (3:1) (16 mL), **13** (2 mmol), **14** (2 mmol) and anhydrous Na_2SO_4 (1.1 g) was refluxed for 4 h. The solution was filtered and the solvent was evaporated under reduced pressure. Then, the residue was recrystallized with ethanol to afford the ligands.

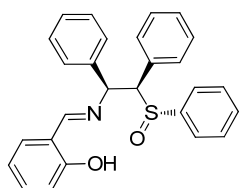
3.4 Spectral Data of Ligands

2-((*E*)-(((1*S*,2*R*)-1,2-diphenyl-2-((*R*)-*p*-tolylsulfinyl)ethyl)imino)methyl)phenol (**1a**)



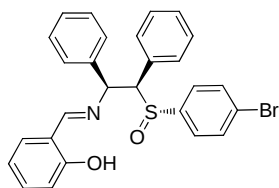
Yellow solid, yield: 93%. 100% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 85:15 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: t_r = 8.171 min for (*S*, *R*, *R*)-isomer (major), t_r = 9.918 min for (*R*, *S*, *S*)-isomer. $^1\text{H NMR}$ (400 MHz, DMSO) δ 13.16 (s, 1H), 8.86 (s, 1H), 7.60 (d, J = 7.4 Hz, 1H), 7.42 (t, J = 7.6 Hz, 1H), 7.23 – 7.12 (m, 12H), 7.01 – 6.91 (m, 4H), 5.52 (d, J = 3.4 Hz, 1H), 4.41 (d, J = 3.8 Hz, 1H), 2.24 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, DMSO) δ 167.53, 159.78, 141.26, 139.63, 138.78, 133.09, 132.19, 130.92, 130.28, 129.22, 128.28, 128.15, 127.92, 127.38, 127.20, 125.40, 119.18, 118.71, 116.40, 76.41, 70.32, 20.82. MS: m/z = 440.1 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{28}\text{H}_{25}\text{NO}_2\text{S}$): C, 76.51; H, 5.73; N, 3.19; S, 7.29; found: C, 76.29; H, 6.025; N, 3.068; S, 7.461. $[\alpha]_D^{25}$ = -177.72 (c = 0.5, CHCl_3).

2-((*E*)-(((1*S*,2*R*)-1,2-diphenyl-2-((*R*)-phenylsulfinyl)ethyl)imino)methyl)phenol (**1b**)



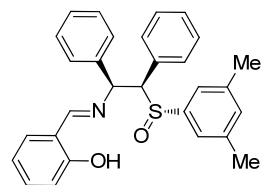
Yellow solid, yield: 94%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 13.28 (s, 1H), 8.80 (s, 1H), 7.42 – 7.36 (m, 2H), 7.33 – 7.29 (m, 1H), 7.25 – 7.22 (m, 5H), 7.20 – 7.16 (m, 5H), 7.07 – 7.04 (m, 3H), 6.96 – 6.88 (m, 3H), 5.63 (d, J = 3.0 Hz, 1H), 3.95 (d, J = 3.1 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO) δ 167.54, 159.78, 141.98, 139.59, 133.10, 132.19, 131.26, 130.87, 130.27, 128.62, 128.29, 128.20, 127.95, 127.40, 127.22, 125.30, 119.20, 118.73, 116.41, 76.57, 70.27. MS: m/z = 426.0 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{27}\text{H}_{23}\text{NO}_2\text{S}$): C, 76.21; H, 5.45; N, 3.29; S, 7.54; found: C, 76.29; H, 5.705; N, 3.244; S, 7.656. $[\alpha]_D^{25}$ = -165.04 (c = 0.5, CHCl_3).

2-((*E*)-(((1*S*,2*R*)-2-((*R*)-(4-bromophenyl)sulfinyl)-1,2-diphenylethyl)imino)methyl)phenol (**1c**)



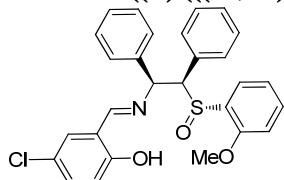
Yellow solid, yield: 93%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 13.22 (s, 1H), 8.77 (s, 1H), 7.41 – 7.38 (m, 4H), 7.30 – 7.17 (m, 6H), 7.07 – 7.04 (m, 5H), 6.96 – 6.90 (m, 3H), 5.60 (d, J = 2.0 Hz, 1H), 3.91 (d, J = 2.1 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO) δ 167.53, 160.11, 159.73, 141.44, 139.44, 133.11, 132.13, 131.56, 130.72, 130.26, 128.30, 128.06, 127.44, 127.35, 127.27, 124.81, 119.18, 118.71, 116.39, 76.20, 70.18. MS: m/z = 504.8 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{27}\text{H}_{22}\text{BrNO}_2\text{S}$): C, 64.29; H, 4.40; N, 2.78; S, 6.36; found: C, 63.99; H, 4.689; N, 2.581; S, 6.374. $[\alpha]_D^{25}$ = -162.90 (c = 0.5, CHCl_3).

2-((*E*)-(((1*S*,2*R*)-2-((*R*)-(3,5-dimethylphenyl)sulfinyl)-1,2-diphenylethyl)imino)methyl)phenol (**1d**)



Yellow solid, yield: 87%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 13.31 (s, 1H), 8.79 (s, 1H), 7.42 – 7.38 (m, 2H), 7.26 – 7.23 (m, 2H), 7.20 – 7.15 (m, 5H), 7.08 – 7.03 (m, 3H), 6.96 – 6.89 (m, 4H), 6.76 (s, 2H), 5.61 (d, J = 3.0 Hz, 1H), 3.92 (d, J = 3.0 Hz, 1H), 2.14 (s, 6H). $^{13}\text{C NMR}$ (100 MHz, DMSO) δ 167.39, 160.15, 159.77, 141.63, 139.65, 137.87, 133.06, 132.53, 132.15, 131.15, 130.42, 128.27, 128.08, 127.86, 127.40, 127.29, 122.63, 119.17, 118.73, 116.39, 76.33, 70.25, 20.59. MS: m/z = 454.1 ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{29}\text{H}_{27}\text{NO}_2\text{S}$): C, 76.79; H, 6.00; N, 3.09; S, 7.07; found: C, 76.63; H, 6.178; N, 3.008; S, 7.170. $[\alpha]_D^{25}$ = -159.42 (c = 0.5, CHCl_3).

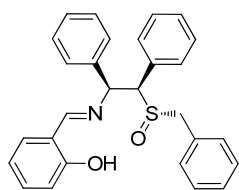
4-chloro-2-((*E*)-(((1*S*,2*R*)-2-((*R*)-(2-methoxyphenyl)sulfinyl)-1,2-diphenylethyl)imino)methyl)phenol (**1e**)



Yellow solid, yield: 85%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 13.18 (s, 1H), 8.60 (s, 1H), 7.78 – 7.74 (m, 1H), 7.32 – 7.27 (m, 3H), 7.22 – 7.14 (m, 6H), 7.05 – 7.00 (m, 5H), 6.94 (d, J = 8.7 Hz, 1H), 6.60 (d, J = 8.2 Hz, 1H), 5.55 (d, J = 3.9 Hz, 1H), 4.22 (d, J = 4.0 Hz, 1H), 3.40 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 166.20, 159.47, 156.15, 139.25, 132.57, 132.46, 131.29, 131.25, 130.53,

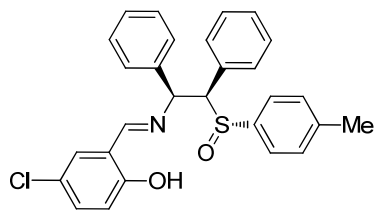
130.49, 128.36, 128.13, 127.74, 127.65, 127.45, 125.73, 123.44, 121.40, 119.54, 118.41, 110.40, 77.51, 70.93, 55.08. MS: m/z = 490.0 ($[M+1]^+$). HRMS (ESI) for $C_{28}H_{24}ClNO_3S$ $[M+H]^+$: calcd 490.1244, found 490.12382. $[\alpha]_D^{25}$ = 13.76 (c = 0.5, $CHCl_3$).

2-((E)-(((1S,2R)-2-((S)-benzylsulfinyl)-1,2-diphenylethyl)imino)methyl)phenol (1f)



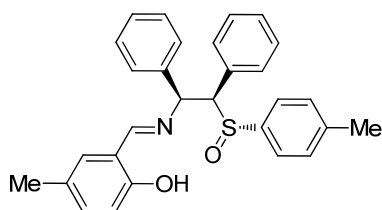
Yellow solid, yield: 93%. 1H NMR (400 MHz, $CDCl_3$) δ 13.19 (s, 1H), 8.66 (s, 1H), 7.39 – 7.31 (m, 8H), 7.19 – 7.16 (m, 3H), 7.13 – 7.11 (m, 2H), 7.03 – 6.95 (m, 5H), 6.90 (t, J = 7.5 Hz, 1H), 5.40 (d, J = 2.9 Hz, 1H), 3.97 (d, J = 3.0 Hz, 1H), 3.81 (d, J = 13.3 Hz, 1H), 3.53 (d, J = 13.3 Hz, 1H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 167.90, 160.80, 139.14, 132.94, 132.38, 130.57, 130.21, 130.06, 129.85, 129.03, 128.79, 128.71, 128.57, 128.27, 127.53, 127.24, 118.93, 118.56, 116.78, 72.95, 70.71, 55.12. MS: m/z = 439.8 (M^+). Anal. calcd for ($C_{28}H_{25}NO_2S$): C, 76.51; H, 5.73; N, 3.19; S, 7.29; found: C, 76.67; H, 5.455; N, 3.448; S, 7.104. $[\alpha]_D^{16}$ = -155.54 (c = 0.5, $CHCl_3$).

4-chloro-2-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)phenol (1g)



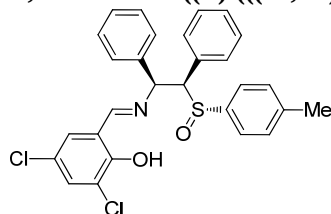
Yellow solid, yield: 91%. 1H NMR (400 MHz, $CDCl_3$) δ 13.25 (s, 1H), 8.70 (s, 1H), 7.36 (d, J = 2.5 Hz, 1H), 7.32 – 7.29 (m, 1H), 7.26 – 7.22 (m, 1H), 7.18 – 7.15 (m, 5H), 7.10 (d, J = 8.2 Hz, 2H), 7.03 – 7.00 (m, 4H), 6.97 (d, J = 8.8 Hz, 1H), 6.86 (d, J = 7.3 Hz, 2H), 5.61 (d, J = 3.1 Hz, 1H), 3.93 (d, J = 3.1 Hz, 1H), 2.26 (s, 3H). ^{13}C NMR (100 MHz, DMSO) δ 166.09, 158.82, 141.31, 139.40, 138.73, 132.71, 130.87, 130.69, 130.31, 129.25, 128.35, 128.21, 127.97, 127.49, 127.25, 125.44, 122.59, 120.01, 118.57, 76.21, 70.30, 20.85. MS: m/z = 474.9 ($[M+2]^+$). HRMS (ESI) for $C_{28}H_{24}ClNO_2S$ $[M+H]^+$: calcd 474.1295, found 474.12890. $[\alpha]_D^{25}$ = -155.94 (c = 0.5, $CHCl_3$).

2-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)-4-methylphenol (1h)



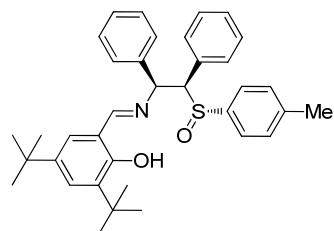
Yellow solid, yield: 92%. 1H NMR (400 MHz, $CDCl_3$) δ 13.03 (s, 1H), 8.74 (s, 1H), 7.23 – 7.15 (m, 8H), 7.11 (d, J = 8.2 Hz, 2H), 7.06 – 7.02 (m, 4H), 6.95 (d, J = 8.1 Hz, 1H), 6.88 (d, J = 7.4 Hz, 2H), 5.61 (d, J = 2.9 Hz, 1H), 3.93 (d, J = 3.0 Hz, 1H), 2.31 (s, 3H), 2.27 (s, 3H). ^{13}C NMR (100 MHz, DMSO) δ 167.66, 158.01, 141.26, 139.62, 138.82, 133.82, 132.05, 130.92, 130.30, 129.23, 128.29, 128.16, 127.94, 127.71, 127.38, 127.22, 125.42, 118.32, 116.34, 76.44, 70.29, 20.84, 19.88. MS: m/z = 454.1 ($[M+1]^+$). Anal. calcd for ($C_{29}H_{27}NO_2S$): C, 76.79; H, 6.00; N, 3.09; S, 7.07; found: C, 76.90; H, 5.999; N, 2.859; S, 7.123. $[\alpha]_D^{25}$ = -171.34 (c = 0.5, $CHCl_3$).

2,4-dichloro-6-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)phenol (1i)



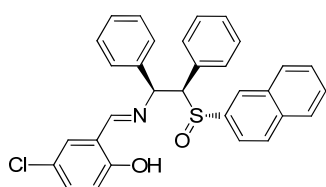
Yellow solid, yield: 88%. 1H NMR (400 MHz, $CDCl_3$) δ 14.07 (s, 1H), 8.72 (s, 1H), 7.47 (d, J = 2.1 Hz, 1H), 7.32 (d, J = 2.2 Hz, 1H), 7.28 – 7.24 (m, 1H), 7.21 – 7.16 (m, 5H), 7.11 (d, J = 8.1 Hz, 2H), 7.05 – 7.03 (m, 4H), 6.86 (d, J = 7.2 Hz, 2H), 5.65 (d, J = 2.8 Hz, 1H), 3.94 (d, J = 3.0 Hz, 1H), 2.28 (s, 3H). ^{13}C NMR (100 MHz, DMSO) δ 166.85, 155.89, 141.30, 138.70, 138.51, 132.24, 130.68, 130.36, 130.12, 129.21, 128.41, 128.26, 128.04, 127.65, 127.28, 125.35, 122.02, 121.58, 119.73, 75.66, 69.72, 20.81. MS: m/z = 507.9 ($[M+1]^+$). Anal. calcd for ($C_{28}H_{23}Cl_2NO_2S$): C, 66.14; H, 4.56; N, 2.75; S, 6.31; found: C, 66.01; H, 4.375; N, 2.696; S, 6.521. $[\alpha]_D^{25}$ = -140.36 (c = 0.5, $CHCl_3$).

2,4-di-tert-butyl-6-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)phenol (1j)



Yellow solid, yield: 93%. 1H NMR (400 MHz, $CDCl_3$) δ 13.58 (s, 1H), 8.81 (s, 1H), 7.44 (d, J = 1.8 Hz, 1H), 7.24 – 7.21 (m, 2H), 7.17 – 7.08 (m, 9H), 7.01 (d, J = 8.1 Hz, 2H), 6.88 (d, J = 7.2 Hz, 2H), 5.61 (d, J = 2.4 Hz, 1H), 3.91 (d, J = 2.6 Hz, 1H), 2.26 (s, 3H), 1.50 (s, 9H), 1.31 (s, 9H). ^{13}C NMR (100 MHz, DMSO) δ 169.10, 157.43, 141.27, 140.12, 139.59, 138.71, 135.69, 130.74, 130.39, 129.23, 128.34, 128.24, 127.86, 127.42, 127.20, 127.07, 125.98, 125.40, 117.71, 76.37, 70.03, 34.65, 33.87, 31.23, 29.25, 20.83. MS: m/z = 551.63 ($[M+1]^+$). Anal. calcd for ($C_{36}H_{41}NO_2S$): C, 78.36; H, 7.49; N, 2.54; S, 5.81; found: C, 78.55; H, 7.663; N, 2.362; S, 6.096. $[\alpha]_D^{25}$ = -131.12 (c = 0.5, $CHCl_3$).

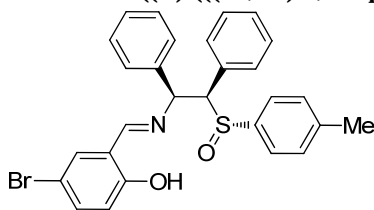
4-chloro-2-((E)-(((1S,2R)-2-((R)-naphthalen-2-ylsulfinyl)-1,2-diphenylethyl)imino)methyl)phenol (1k)



Yellow solid, yield: 90%. 1H NMR (400 MHz, $CDCl_3$) δ 13.28 (s, 1H), 8.74 (s, 1H), 7.82 – 7.66 (m, 4H), 7.53 – 7.48 (m, 2H), 7.40 (d, J = 2.3 Hz, 1H), 7.35 – 7.32 (m, 1H), 7.27 – 7.23 (m, 1H), 7.18 – 7.12 (m, 6H), 7.06 – 6.99 (m, 3H), 6.89 (d, J = 7.2 Hz, 2H), 5.68 (d, J = 2.9 Hz, 1H), 4.05 (d, J = 3.0 Hz, 1H). ^{13}C NMR (100 MHz, DMSO) δ 166.04, 158.79, 139.37, 139.13, 133.91, 132.72, 131.86, 130.96, 130.63, 130.29, 128.34, 127.98, 127.50, 127.31, 127.07, 126.35, 122.61, 121.13, 120.06, 118.58, 75.93, 70.29. MS: m/z = 509.4 (M^+). Anal. calcd for

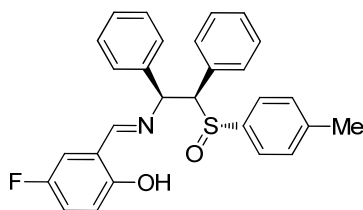
(C₃₁H₂₄ClNO₂S): C, 73.00; H, 4.74; N, 2.75; S, 6.29; found: C, 72.87; H, 5.025; N, 2.661; S, 6.320. [α]_D²⁵ = -161.70 (c = 0.5, CHCl₃).

4-bromo-2-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)phenol (1l)



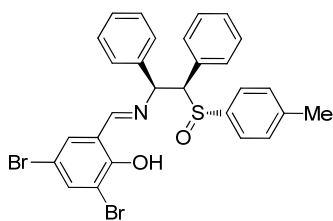
Yellow solid, yield: 89%. ¹H NMR (400 MHz, CDCl₃) δ 13.29 (s, 1H), 8.70 (s, 1H), 7.51 (d, *J* = 2.4 Hz, 1H), 7.48 – 7.43 (m, 1H), 7.28 – 7.23 (m, 1H), 7.19 – 7.16 (m, 5H), 7.12 (d, *J* = 8.2 Hz, 2H), 7.05 – 7.01 (m, 4H), 6.94 (d, *J* = 8.8 Hz, 1H), 6.87 (d, *J* = 7.4 Hz, 2H), 5.62 (d, *J* = 3.0 Hz, 1H), 3.94 (d, *J* = 3.1 Hz, 1H), 2.27 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 166.06, 159.25, 141.29, 139.38, 138.72, 135.47, 133.66, 130.85, 130.30, 129.24, 128.34, 128.20, 127.96, 127.47, 127.24, 125.41, 120.60, 118.99, 109.93, 76.25, 70.29, 20.85. MS: *m/z* = 518.0 ([*M*+1]⁺). Anal. calcd for (C₂₈H₂₄BrNO₂S): C, 64.86; H, 4.67; N, 2.70; S, 6.18; found: C, 64.81; H, 4.942; N, 2.611; S, 6.438. [α]_D²⁵ = -147.24 (c = 0.5, CHCl₃).

2-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)-4-fluorophenol (1m)



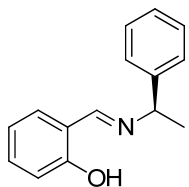
Yellow solid, yield: 90%. ¹H NMR (400 MHz, CDCl₃) δ 13.03 (s, 1H), 8.72 (s, 1H), 7.27 – 7.23 (m, 1H), 7.20 – 7.16 (m, 5H), 7.13 – 7.08 (m, 4H), 7.05 – 6.97 (m, 5H), 6.88 (d, *J* = 7.2 Hz, 2H), 5.63 (d, *J* = 2.8 Hz, 1H), 3.94 (d, *J* = 3.0 Hz, 1H), 2.27 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 166.13, 156.33, 156.10, 153.76, 141.29, 139.48, 138.76, 130.87, 130.32, 129.23, 128.31, 128.18, 127.95, 127.43, 127.24, 125.43, 119.98, 118.97, 117.87, 116.88, 76.31, 70.32, 20.84. MS: *m/z* = 457.46 (*M*⁺). Anal. calcd for (C₂₈H₂₄FNO₂S): C, 73.50; H, 5.29; N, 3.06; S, 7.01; found: C, 73.42; H, 5.555; N, 2.939; S, 7.206. [α]_D²⁵ = -157.08 (c = 0.5, CHCl₃).

2,4-dibromo-6-((E)-(((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)imino)methyl)phenol (1n)



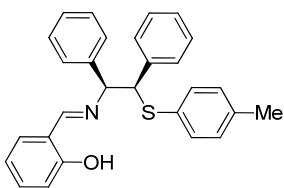
Yellow solid, yield: 89%. ¹H NMR (400 MHz, CDCl₃) δ 14.16 (s, 1H), 8.69 (s, 1H), 7.76 (d, *J* = 2.2 Hz, 1H), 7.50 (d, *J* = 2.2 Hz, 1H), 7.28 – 7.24 (m, 1H), 7.21 – 7.16 (m, 5H), 7.11 (d, *J* = 8.2 Hz, 2H), 7.05 – 7.02 (m, 4H), 6.85 (d, *J* = 7.5 Hz, 2H), 5.65 (d, *J* = 3.0 Hz, 1H), 3.94 (d, *J* = 3.2 Hz, 1H), 2.28 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 166.82, 157.29, 141.34, 138.69, 138.47, 137.56, 133.99, 130.68, 130.12, 129.25, 128.46, 128.30, 128.07, 127.70, 127.31, 125.40, 120.14, 111.45, 109.37, 75.54, 69.62, 20.85. MS: *m/z* = 598.0 ([*M*+1]⁺). Anal. calcd for (C₂₈H₂₃Br₂NO₂S): C, 56.30; H, 3.88; N, 2.34; S, 5.37; found: C, 56.53; H, 4.124; N, 2.300; S, 5.424. [α]_D²⁵ = -126.48 (c = 0.5, CHCl₃).

(R,E)-2-(((1-phenylethyl)imino)methyl)phenol (5)^[2]



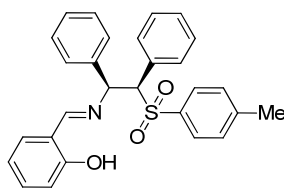
Pale yellow solid, yield: 82%. ¹H NMR (400 MHz, CDCl₃) δ 13.55 (s, 1H), 8.41 (s, 1H), 7.38 – 7.23 (m, 7H), 6.96 (d, *J* = 8.3 Hz, 1H), 6.87 (t, *J* = 7.4 Hz, 1H), 4.55 (q, *J* = 6.6 Hz, 1H), 1.63 (d, *J* = 6.7 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 163.39, 161.01, 143.78, 132.24, 131.34, 128.63, 127.22, 126.35, 118.79, 118.58, 116.92, 68.50, 24.93. MS: *m/z* = 225.1 (*M*⁺).

2-((E)-(((1S,2R)-1,2-diphenyl-2-(p-tolylthio)ethyl)imino)methyl)phenol (6)



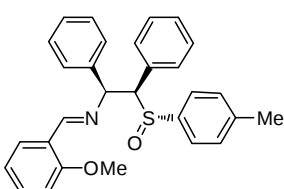
Yellow solid, yield: 93%. ¹H NMR (400 MHz, CDCl₃) δ 13.09 (s, 1H), 8.19 (s, 1H), 7.31 – 7.23 (m, 4H), 7.19 – 7.11 (m, 10H), 7.01 (d, *J* = 8.0 Hz, 2H), 6.95 (d, *J* = 8.3 Hz, 1H), 6.84 (t, *J* = 7.5 Hz, 1H), 4.73 (d, *J* = 5.8 Hz, 1H), 4.61 (d, *J* = 5.8 Hz, 1H), 2.27 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 165.56, 160.83, 140.24, 137.61, 137.25, 132.50, 132.32, 131.70, 130.84, 129.65, 129.28, 128.26, 127.87, 127.64, 127.49, 127.45, 118.62, 116.87, 60.72, 21.07. MS: *m/z* = 423.4 (*M*⁺). Anal. calcd for (C₂₈H₂₅NOS): C, 79.40; H, 5.95; N, 3.31; S, 7.57; found: C, 79.61; H, 6.064; N, 3.179; S, 7.802. [α]_D²⁵ = -84.58 (c = 0.5, CHCl₃).

2-((E)-(((1S,2R)-1,2-diphenyl-2-tosylethyl)imino)methyl)phenol (7)



Yellow solid, yield: 88%. ¹H NMR (400 MHz, DMSO) δ 12.83 (s, 1H), 8.62 (s, 1H), 7.47 – 7.39 (m, 6H), 7.22 – 7.19 (m, 10H), 6.88 (t, *J* = 7.3 Hz, 2H), 5.56 (d, *J* = 5.5 Hz, 1H), 5.30 (d, *J* = 5.6 Hz, 1H), 2.29 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 178.09, 166.65, 159.63, 143.86, 139.60, 135.92, 132.76, 132.00, 131.01, 130.79, 129.18, 128.42, 128.18, 127.88, 127.58, 127.52, 118.91, 118.64, 116.25, 73.02, 70.99, 20.96. MS: *m/z* = 455.2 (*M*⁺). Anal. calcd for (C₂₈H₂₅NO₃S): C, 73.82; H, 5.53; N, 3.07; S, 7.04; found: C, 73.83; H, 5.612; N, 2.980; S, 7.245. [α]_D²⁵ = -182.38 (c = 0.5, CHCl₃).

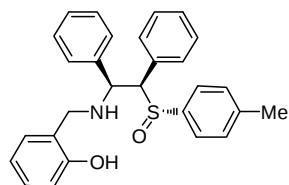
(1S,2R,E)-N-(2-methoxybenzylidene)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethanamine (8)



White solid, yield: 81%. ¹H NMR (400 MHz, CDCl₃) δ 9.11 (s, 1H), 8.28 (d, *J* = 7.7 Hz, 1H), 7.43 (t, *J* = 7.2 Hz, 1H), 7.20 – 7.04 (m, 13H), 7.00 (d, *J* = 8.0 Hz, 2H), 6.94 (d, *J* = 8.3 Hz, 1H),

5.62 (d, $J = 2.8$ Hz, 1H), 3.92 (d, $J = 3.0$ Hz, 1H), 3.86 (s, 3H), 2.27 (s, 3H). ^{13}C NMR (100 MHz, DMSO) δ 158.82, 158.75, 141.01, 140.92, 139.16, 132.96, 131.15, 131.03, 129.09, 127.98, 127.90, 127.67, 127.30, 126.92, 126.81, 125.30, 123.51, 120.76, 111.97, 77.99, 71.07, 55.66, 20.84. MS: $m/z = 454.1$ ($[\text{M}+1]^+$). Anal. calcd for ($\text{C}_{29}\text{H}_{27}\text{NO}_2\text{S}$): C, 76.79; H, 6.00; N, 3.09; S, 7.07; found: C, 76.88; H, 6.239; N, 2.934; S, 7.349. $[\alpha]_{\text{D}}^{25} = -155.74$ ($c = 0.5$, CHCl_3).

2-((((1S,2R)-1,2-diphenyl-2-((R)-p-tolylsulfinyl)ethyl)amino)methyl)phenol (9)



The title compound was prepared by reduction of ligand **5a** with NaBH_4 in methanol and purified by column chromatography (petroleum ether/ethyl acetate, 3:1) to give a white solid in 95% yield. ^1H NMR (400 MHz, CDCl_3) δ 10.27 (s, 1H), 7.29 – 7.24 (m, 4H), 7.21 – 7.12 (m, 5H), 7.06 (d, $J = 8.1$ Hz, 2H), 7.02 – 7.00 (m, 2H), 6.93 (d, $J = 8.1$ Hz, 1H), 6.84 – 6.80 (m, 3H), 6.74 (t, $J = 7.4$ Hz, 1H), 4.72 (s, 1H), 3.91 (d, $J = 3.4$ Hz, 1H), 3.81 (d, $J = 4.2$ Hz, 2H), 2.86 (s, 1H), 2.29 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 157.82, 141.41, 139.09, 137.49, 130.81, 130.14, 129.24, 128.89, 128.57, 128.47, 127.98, 127.93, 127.86, 124.71, 122.43, 119.06, 116.45, 77.88, 61.37, 50.36, 21.25. MS: $m/z = 441.31$ (M^+). Anal. calcd for ($\text{C}_{28}\text{H}_{27}\text{NO}_2\text{S}$): C, 76.16; H, 6.16; N, 3.17; S, 7.26; found: C, 76.28; H, 6.412; N, 3.121; S, 7.271. $[\alpha]_{\text{D}}^{25} = -113.38$ ($c = 0.5$, CHCl_3).

3.5 References

- [1] Petra, D. G. I.; Kamer, P. C. J.; Spek, A. L.; Schoemaker, H. E.; Van Leeuwen, P. W. N. M. *J. Org. Chem.* **2000**, 65, 3010.
- [2] Guo, J.; Mao, J. *Chirality* **2009**, 21, 619.

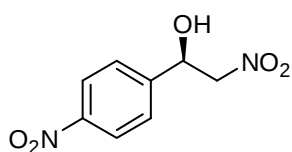
4. General Procedure for the Asymmetric Henry Reactions and Spectral Data

4.1 General Procedure for the Asymmetric Henry Reactions

Ligand **1f** (5.3 mg, 0.012 mmol, 3 mol%) and $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (2.0 mg, 0.01 mmol, 2.5 mol%) were dissolved in *t*-BuOH (4.0 mL) in a Schlenk tube. The mixture was stirred at 25 °C for 1 h, then aldehyde **2** (0.4 mmol) and nitromethane **3** (1 mL) were successively added. The resulting solution was stirred at the same temperature until the reaction completed (TLC). The crude reaction mixture was concentrated and directly purified by flash column chromatography, eluting with petroleum ether and ethyl acetate to afford the corresponding product **4**.

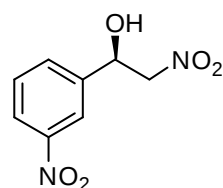
4.2 Spectral Data of Henry Products

(R)-2-nitro-1-(4-nitrophenyl)ethanol (4a)



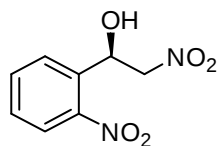
White solid, 30 h, 94% yield and 93% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 85:15 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: $t_{\text{r}} = 26.419$ min for (*R*)-isomer (major), $t_{\text{r}} = 33.402$ min for (*S*)-isomer. ^1H NMR (400 MHz, DMSO) δ 8.23 (d, $J = 8.6$ Hz, 2H), 7.74 (d, $J = 8.6$ Hz, 2H), 6.43 (d, $J = 4.9$ Hz, 1H), 5.46 – 5.44 (m, 1H), 4.95 (dd, $J = 12.7$, 3.2 Hz, 1H), 4.64 (dd, $J = 12.7$, 9.5 Hz, 1H). ^{13}C NMR (100 MHz, DMSO) δ 148.00, 147.13, 127.56, 123.43, 81.21, 69.09. $[\alpha]_{\text{D}}^{16} = -35.12$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25} = -33.1$ ($c = 0.56$, CH_2Cl_2) (85% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(R)-2-nitro-1-(3-nitrophenyl)ethanol (4b)



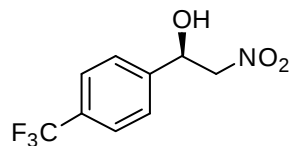
White solid, 48 h, 94% yield and 94% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: $t_{\text{r}} = 41.363$ min for (*R*)-isomer (major), $t_{\text{r}} = 48.262$ min for (*S*)-isomer. ^1H NMR (400 MHz, CDCl_3) δ 8.32 (s, 1H), 8.22 (d, $J = 8.1$ Hz, 1H), 7.78 (d, $J = 7.6$ Hz, 1H), 7.62 (t, $J = 7.9$ Hz, 1H), 5.63 – 5.61 (m, 1H), 4.67 – 4.58 (m, 2H), 3.39 (d, $J = 4.0$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ 148.50, 140.25, 131.99, 130.08, 123.75, 121.10, 80.66, 69.80. $[\alpha]_{\text{D}}^{16} = -27.47$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25} = -26.7$ ($c = 0.43$, CH_2Cl_2) (85% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(R)-2-nitro-1-(2-nitrophenyl)ethanol (4c)



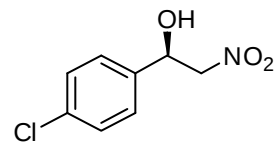
Brown solid, 42 h, 95% yield and 96% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 95:05 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: t_r = 45.211 min for (R)-isomer (major), t_r = 51.895 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, DMSO) δ 8.05 (d, J = 8.2 Hz, 1H), 7.93 (d, J = 7.8 Hz, 1H), 7.81 (t, J = 7.6 Hz, 1H), 7.61 (t, J = 7.7 Hz, 1H), 6.44 (s, 1H), 5.83 (d, J = 9.2 Hz, 1H), 4.98 (dd, J = 12.8, 2.2 Hz, 1H), 4.61 (dd, J = 12.7, 9.6 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO) δ 147.26, 135.64, 134.08, 129.49, 129.20, 124.51, 81.01, 66.29. $[\alpha]_D^{16}$ = 231.82 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{21}$ = 227.1 (c = 1.00, CH_2Cl_2) (89% ee)]. Reference: Evans, D. A.; Seidel, D.; Rueping, M.; Lam, H. W.; Shaw, J. T.; Downey, C. W. *J. Am. Chem. Soc.* **2003**, 125, 12692.

(R)-2-nitro-1-(4-(trifluoromethyl)phenyl)ethanol (4d)



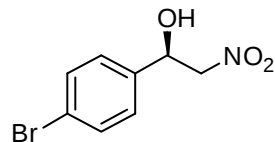
Yellow oil, 48 h, 96% yield and 94% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: t_r = 16.856 min for (R)-isomer (major), t_r = 21.680 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.65 (d, J = 8.1 Hz, 2H), 7.52 (d, J = 8.1 Hz, 2H), 5.51 (d, J = 8.8 Hz, 1H), 4.81 – 4.50 (m, 2H), 3.53 (s, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 141.97, 131.14, 130.81, 126.32, 125.89, 122.42, 80.84, 70.27. $[\alpha]_D^{17}$ = -31.84 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{23}$ = -49.2 (c = 1.25, CHCl_3) (86% ee)]. Reference: Bulut, A.; Aslan, A.; Dogan, Ö. *J. Org. Chem.* **2008**, 73, 7373.

(R)-1-(4-chlorophenyl)-2-nitroethanol (4e)



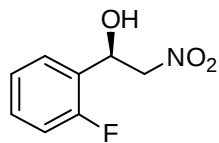
Colorless oil, 96 h, 91% yield and 92% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: t_r = 21.448 min for (R)-isomer (major), t_r = 27.463 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.37 – 7.30 (m, 4H), 5.40 (d, J = 9.2 Hz, 1H), 4.58 – 4.45 (m, 2H), 3.36 (d, J = 3.0 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 136.55, 134.64, 129.08, 127.28, 80.92, 70.20. $[\alpha]_D^{15}$ = -38.01 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{23}$ = -37.6 (c = 2.03, CH_2Cl_2) (90% ee)]. Reference: Evans, D. A.; Seidel, D.; Rueping, M.; Lam, H. W.; Shaw, J. T.; Downey, C. W. *J. Am. Chem. Soc.* **2003**, 125, 12692.

(R)-1-(4-bromophenyl)-2-nitroethanol (4f)



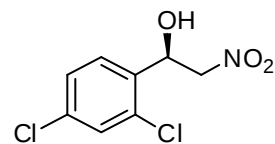
Colorless solid, 96 h, 94% yield and 92% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: t_r = 25.761 min for (R)-isomer (major), t_r = 33.990 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.51 (d, J = 8.4 Hz, 2H), 7.26 (d, J = 8.8 Hz, 2H), 5.39 (d, J = 8.8 Hz, 1H), 4.58 – 4.45 (m, 2H), 3.28 (d, J = 2.8 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 137.05, 132.06, 127.58, 122.82, 80.86, 70.25. $[\alpha]_D^{16}$ = -36.51 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{25}$ = -34.6 (c = 1.00, CH_2Cl_2) (92% ee)]. Reference: Panov, I.; Drabina, P.; Padělková, Z. k.; Šimůnek, P.; Sedlák, M. *J. Org. Chem.* **2011**, 76, 4787.

(R)-1-(2-fluorophenyl)-2-nitroethanol (4g)



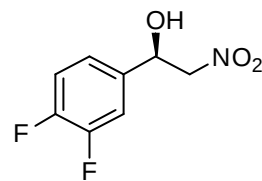
Colorless oil, 96 h, 93% yield and 93% ee, determined by chiral HPLC analysis (Chiralcel AD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: t_r = 15.054 min for (R)-isomer (major), t_r = 13.866 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.57 (t, J = 7.6 Hz, 1H), 7.37 – 7.33 (m, 1H), 7.23 (t, J = 7.5 Hz, 1H), 7.11 – 7.07 (m, 1H), 5.75 (d, J = 4.2 Hz, 1H), 4.65 – 4.56 (m, 2H), 2.98 (d, J = 4.8 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 130.42, 130.34, 127.53, 124.79, 115.62, 115.42, 79.63, 65.31. $[\alpha]_D^{16}$ = -49.58 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{23}$ = 24.6 (c = 1.2, CH_2Cl_2) (46% ee) (S) isomer]. Reference: Kowalczyk, R.; Sidorowicz, Ł.; Skarżewski, J. *Tetrahedron: Asymmetry* **2007**, 18, 2581.

(R)-1-(2,4-dichlorophenyl)-2-nitroethanol (4h)



White solid, 36 h, 98% yield and 95% ee, determined by chiral HPLC analysis (Chiralpak AD-H, hexane/isopropanol, 85:15 v/v, 0.8 mL/min, 25 °C, UV 215 nm): Retention times: t_r = 9.510 min for (R)-isomer (major), t_r = 11.137 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.59 (d, J = 8.4 Hz, 1H), 7.39 (d, J = 1.8 Hz, 1H), 7.33 (dd, J = 8.4, 1.7 Hz, 1H), 5.78 (dd, J = 6.3, 3.2 Hz, 1H), 4.64 (dd, J = 13.6, 2.1 Hz, 1H), 4.42 (dd, J = 13.6, 9.5 Hz, 1H), 3.36 (d, J = 4.3 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 135.12, 134.11, 132.00, 129.40, 128.50, 127.89, 79.03, 67.37. $[\alpha]_D^{15}$ = -50.91 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_D^{23}$ = -47.6 (c = 0.95, CH_2Cl_2) (86% ee)]. Reference: Kogami, Y.; Nakajima, T.; Ikeno, T.; Yamada, T. *Synthesis* **2004**, 1947.

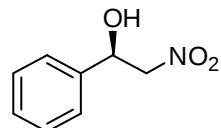
(R)-1-(3,4-difluorophenyl)-2-nitroethanol (4i)



$[\alpha]_D^{16} = -38.49$ ($c = 0.5$, CH_2Cl_2).

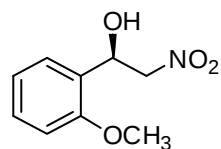
Colorless oil, 72 h, 90% yield and 94% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: $t_r = 17.619$ min for (R)-isomer (major), $t_r = 21.361$ min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.33 – 7.13 (m, 3H), 5.43 (d, $J = 8.7$ Hz, 1H), 4.58 – 4.47 (m, 2H), 3.37 (s, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 149.34, 149.20, 135.14, 122.03, 117.93, 117.75, 115.28, 115.10, 80.88, 69.74. MS: $m/z = 203.08$ (M^+). Anal. calcd for ($\text{C}_8\text{H}_7\text{F}_2\text{NO}_3$): C, 47.30; H, 3.47; N, 6.89; found: C, 47.57; H, 3.676; N, 6.637.

(R)-2-nitro-1-phenylethanol (4j)



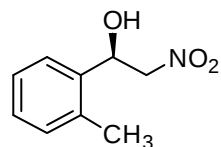
Colorless oil, 120 h, 85% yield and 93% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: $t_r = 21.602$ min for (R)-isomer (major), $t_r = 27.350$ min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.40 – 7.34 (m, 5H), 5.41 – 5.37 (m, 1H), 4.59 – 4.44 (m, 2H), 3.17 (d, $J = 3.8$ Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 138.10, 128.90, 128.84, 125.87, 81.11, 70.88. $[\alpha]_D^{16} = -40.16$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_D^{21} = -41.6$ ($c = 1.03$, CH_2Cl_2) (94% ee)]. Reference: Evans, D. A.; Seidel, D.; Rueping, M.; Lam, H. W.; Shaw, J. T.; Downey, C. W. *J. Am. Chem. Soc.* **2003**, 125, 12692.

(R)-1-(2-methoxyphenyl)-2-nitroethanol (4k)



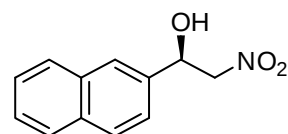
Yellow oil, 132 h, 92% yield and 94% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: $t_r = 17.698$ min for (R)-isomer (major), $t_r = 21.585$ min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.44 (d, $J = 7.5$ Hz, 1H), 7.33 (t, $J = 7.8$ Hz, 1H), 7.01 (t, $J = 7.5$ Hz, 1H), 6.91 (d, $J = 8.2$ Hz, 1H), 5.63 – 5.61 (m, 1H), 4.67 – 4.54 (m, 2H), 3.88 (s, 3H), 3.16 (d, $J = 6.0$ Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 155.91, 129.69, 127.08, 125.93, 121.03, 110.45, 79.78, 67.69, 55.32. $[\alpha]_D^{15} = -39.94$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_D^{21} = -44.5$ ($c = 1.00$, CH_2Cl_2) (93% ee)]. Reference: Evans, D. A.; Seidel, D.; Rueping, M.; Lam, H. W.; Shaw, J. T.; Downey, C. W. *J. Am. Chem. Soc.* **2003**, 125, 12692.

(R)-2-nitro-1-(o-tolyl)ethanol (4l)



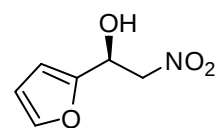
Colorless oil, 132 h, 84% yield and 91% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 85:15 v/v, 0.8 mL/min, 25 °C, UV 210 nm): Retention times: $t_r = 12.673$ min for (R)-isomer (major), $t_r = 19.695$ min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.54 – 7.52 (m, 1H), 7.30 – 7.24 (m, 2H), 7.21 – 7.18 (m, 1H), 5.71 – 5.67 (m, 1H), 4.56 (dd, $J = 13.4, 9.7$ Hz, 1H), 4.45 (dd, $J = 13.4, 2.5$ Hz, 1H), 2.74 (d, $J = 3.6$ Hz, 1H), 2.40 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 136.15, 134.40, 130.77, 128.61, 126.70, 125.53, 80.14, 67.83, 18.81. $[\alpha]_D^{15} = -45.88$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_D^{25} = -33.4$ ($c = 0.5$, CH_2Cl_2) (88% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(R)-1-(naphthalen-2-yl)-2-nitroethanol (4m)



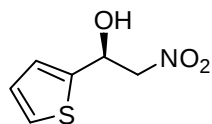
Yellow solid, 108 h, 75% yield and 93% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 85:15 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: $t_r = 43.653$ min for (R)-isomer (major), $t_r = 60.632$ min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.82 – 7.79 (m, 4H), 7.50 – 7.48 (m, 2H), 7.39 (d, $J = 8.4$ Hz, 1H), 5.53 (d, $J = 9.5$ Hz, 1H), 4.64 – 4.49 (m, 2H), 3.20 (d, $J = 2.7$ Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 135.38, 133.29, 133.07, 128.87, 127.99, 127.70, 126.60, 125.23, 123.15, 81.07, 71.03. $[\alpha]_D^{16} = -35.69$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_D^{25} = -45.0$ ($c = 0.38$, CH_2Cl_2) (95% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(S)-1-(furan-2-yl)-2-nitroethanol (4n)



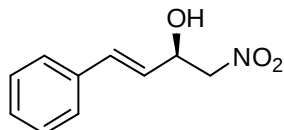
Colorless oil, 96 h, 91% yield and 95% ee, determined by chiral HPLC analysis (Chiralpak AD-H, hexane/isopropanol, 95:5 v/v, 0.5 mL/min, 26 °C, UV 210 nm): Retention times: $t_r = 52.803$ min for (S)-isomer (major), $t_r = 55.511$ min for (R)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.42 (s, 1H), 6.41 – 6.39 (m, 2H), 5.49 – 5.47 (m, 1H), 4.79 (dd, $J = 13.5, 9.1$ Hz, 1H), 4.67 (dd, $J = 13.5, 3.4$ Hz, 1H), 2.96 (s, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 150.63, 143.03, 110.54, 108.07, 78.27, 64.63. $[\alpha]_D^{16} = -33.20$ ($c = 1.00$, CH_2Cl_2). [lit.: $[\alpha]_D^{25} = -37.1$ ($c = 0.24$, CH_2Cl_2) (98% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(S)-2-nitro-1-(thiophen-2-yl)ethanol (4o)



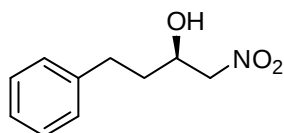
Brown oil, 96 h, 61% yield and 92% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 220 nm): Retention times: t_r = 23.948 min for (S)-isomer (major), t_r = 26.444 min for (R)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.34 (dd, J = 5.1, 1.2 Hz, 1H), 7.08 – 7.07 (m, 1H), 7.02 (dd, J = 5.0, 3.6 Hz, 1H), 5.75 – 5.71 (m, 1H), 4.72 (dd, J = 13.4, 9.3 Hz, 1H), 4.61 (dd, J = 13.4, 3.3 Hz, 1H), 3.07 (d, J = 3.8 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 141.15, 127.14, 126.04, 125.01, 80.64, 66.91. $[\alpha]_{\text{D}}^{17}$ = -39.55 (c = 0.5, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25}$ = -27.5 (c = 0.28, CH_2Cl_2) (95% ee)]. Reference: Qin, B.; Xiao, X.; Liu, X.; Huang, J.; Wen, Y.; Feng, X. *J. Org. Chem.* **2007**, 72, 9323.

(R,E)-1-nitro-4-phenylbut-3-en-2-ol (4p)



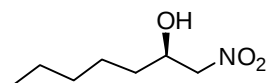
Yellow solid, 108 h, 72% yield and 91% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 80:20 v/v, 0.8 mL/min, 20 °C, UV 220 nm): Retention times: t_r = 28.155 min for (R)-isomer (major), t_r = 24.980 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.40 – 7.26 (m, 5H), 6.79 (d, J = 15.9 Hz, 1H), 6.14 (dd, J = 15.9, 6.3 Hz, 1H), 5.09 – 5.03 (m, 1H), 4.52 (d, J = 5.8 Hz, 2H), 2.72 (t, J = 4.1 Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 135.42, 133.49, 128.66, 128.43, 126.65, 124.89, 79.81, 69.53. $[\alpha]_{\text{D}}^{22}$ = -8.86 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25}$ = -32.4 (c = 0.1, CH_2Cl_2) (92% ee)]. Reference: Xiong, Y.; Wang, F.; Huang, X.; Wen, Y.; Feng, X. *Chem. Eur. J.* **2007**, 13, 829.

(R)-1-nitro-4-phenylbutan-2-ol (4q)



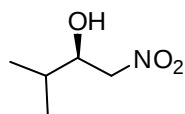
White solid, 96 h, 90% yield and 92% ee, determined by chiral HPLC analysis (Chiralpak AD-H, hexane/isopropanol, 90:10 v/v, 0.8 mL/min, 25 °C, UV 210 nm): Retention times: t_r = 28.317 min for (R)-isomer (major), t_r = 35.971 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.32 (t, J = 7.0 Hz, 2H), 7.26 – 7.20 (m, 3H), 4.41 – 4.39 (m, 2H), 4.34 – 4.28 (m, 1H), 2.90 – 2.83 (m, 1H), 2.79 – 2.71 (m, 2H), 1.92 – 1.75 (m, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 140.58, 128.54, 128.32, 126.20, 80.50, 67.77, 35.04, 31.22. $[\alpha]_{\text{D}}^{16}$ = 12.03 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25}$ = 13.1 (c = 0.51, CH_2Cl_2) (74% ee)]. Reference: Blay, G.; Climent, E.; Fernández, I.; Hernández-Olmos, V.; Pedro, J. R. *Tetrahedron: Asymmetry* **2007**, 18, 1603.

(R)-1-nitroheptan-2-ol (4r)



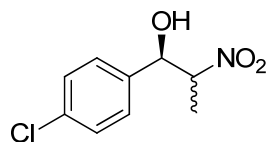
Colorless oil, 96 h, 90% yield and 92% ee, determined by chiral HPLC analysis (Chiralpak AD-H, hexane/isopropanol, 95:5 v/v, 0.8 mL/min, 26 °C, UV 210 nm): Retention times: t_r = 28.794 min for (R)-isomer (major), t_r = 40.104 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 4.48 – 4.38 (m, 2H), 4.35 – 4.32 (m, 1H), 2.59 (d, J = 4.6 Hz, 1H), 1.57 – 1.32 (m, 8H), 0.90 (t, J = 6.6 Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 80.64, 68.66, 33.63, 31.36, 24.75, 22.36, 13.83. $[\alpha]_{\text{D}}^{16}$ = -7.06 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25}$ = -2.3 (c = 0.13, CH_2Cl_2) (90% ee)]. Reference: Xiong, Y.; Wang, F.; Huang, X.; Wen, Y.; Feng, X. *Chem. Eur. J.* **2007**, 13, 829.

(R)-3-methyl-1-nitrobutan-2-ol (4s)



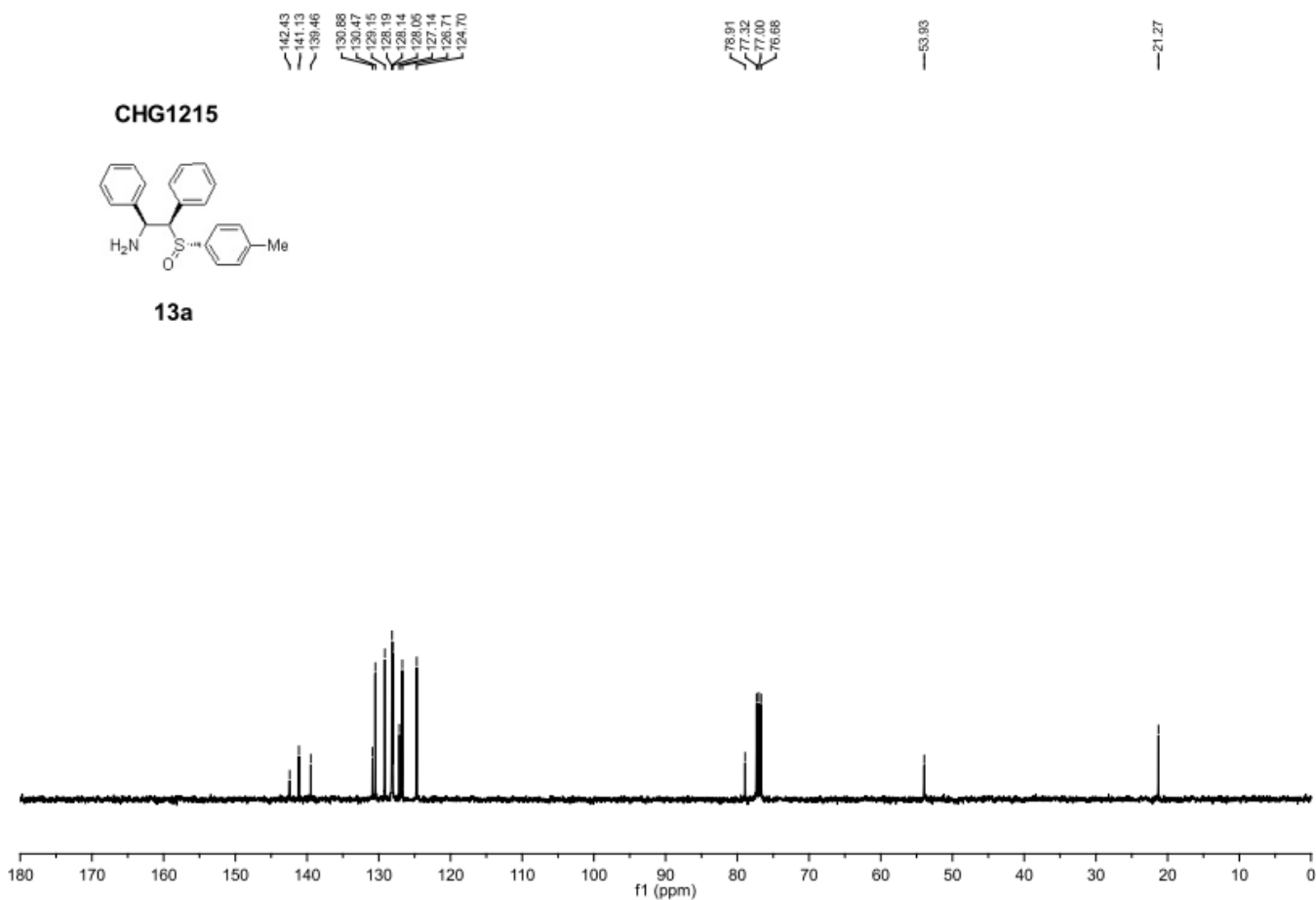
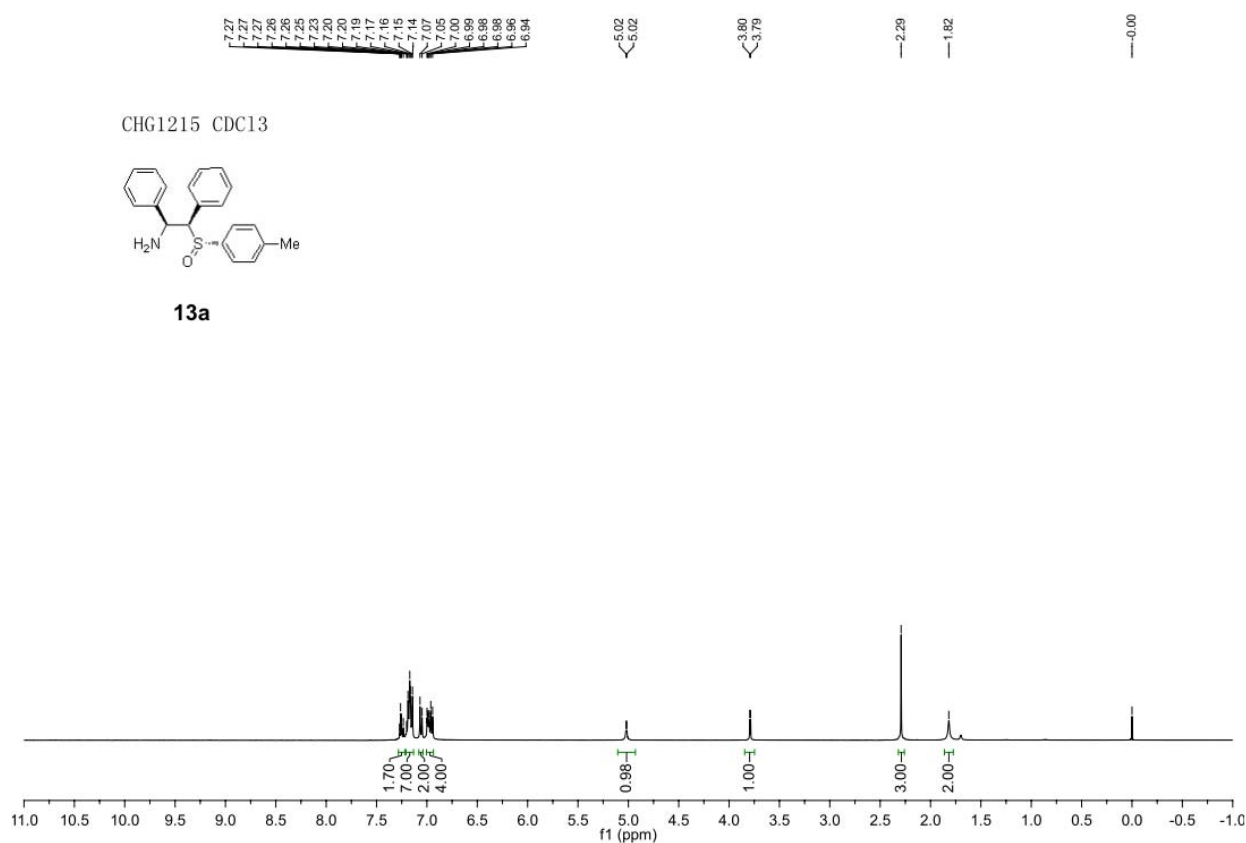
Colorless oil, 96 h, 66% yield and 92% ee, determined by chiral HPLC analysis (Chiralcel OD-H, hexane/isopropanol, 98:2 v/v, 0.5 mL/min, 25 °C, UV 210 nm): Retention times: t_r = 39.99 min for (R)-isomer (major), t_r = 45.238 min for (S)-isomer. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 4.50 – 4.38 (m, 2H), 4.14 – 4.08 (m, 1H), 2.57 (d, J = 4.2 Hz, 1H), 1.85 – 1.78 (m, 1H), 1.00 (t, J = 6.2 Hz, 6H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 79.29, 73.35, 31.68, 18.29, 17.35. $[\alpha]_{\text{D}}^{16}$ = -24.23 (c = 1.00, CH_2Cl_2). [lit.: $[\alpha]_{\text{D}}^{25}$ = -29.2 (c = 1.84, CHCl_3) (94% ee)]. Reference: Evans, D. A.; Seidel, D.; Rueping, M.; Lam, H. W.; Shaw, J. T.; Downey, C. W. *J. Am. Chem. Soc.* **2003**, 125, 12692.

(1R)-1-(4-chlorophenyl)-2-nitropropan-1-ol (4t)

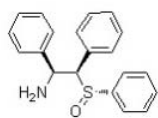


Colorless oil, 96 h, 87% yield, 3:2 *anti/syn* and 68/81% ee (*anti/syn*), determined by chiral HPLC analysis (Chiralcel AD-H, hexane/isopropanol, 95:5 v/v, 1 mL/min, 25 °C, UV 210 nm): Retention times: t_r = 18.243 min for (1R,2R)-isomer (*anti*_{major}), t_r = 16.660 min for (1S,2S)-isomer (*anti*_{minor}), t_r = 24.161 min for (1R,2S)-isomer (*syn*_{major}), t_r = 26.950 min for (1S,2R)-isomer (*syn*_{minor}). $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.39 – 7.35 (m, 2H, *anti*+*syn*), 7.32 – 7.29 (m, 2H, *anti*+*syn*), 5.38 – 5.36 (m, 0.66H, *anti*), 5.02 – 4.98 (m, 0.34H, *syn*), 4.75 – 4.69 (m, 0.34H, *syn*), 4.68 – 4.62 (m, 0.66H, *anti*), 3.01 (d, J = 3.7 Hz, 0.66H, *anti*), 2.97 (d, J = 4.1 Hz, 0.34H, *syn*), 1.47 (d, J = 6.8 Hz, 1.98H, *anti*), 1.31 (d, J = 6.8 Hz, 1.02H, *syn*). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 136.81 (*anti*), 136.64 (*syn*), 134.94 (*syn*), 134.23 (*anti*), 129.12 (*syn*), 128.84 (*anti*), 128.22 (*syn*), 127.29 (*anti*), 88.12 (*syn*), 87.12 (*anti*), 75.42 (*syn*), 73.14 (*anti*), 16.29 (*syn*), 11.88 (*anti*). $[\alpha]_{\text{D}}^{16}$ = -12.09 (c = 1.00, CH_2Cl_2 , mixture of diastereomers). [lit.: $[\alpha]_{\text{D}}^{25}$ = -15.8 (c = 1.00, CHCl_3 , for the 1R enantiomer, 71% ee, mixture of diastereomers)]. Reference: Borah, J. C.; Boruwa, J.; Kalita, B.; Hazarika, A. K.; Barua, N. C. *Indian J. Chem.* **2005**, 44B, 1961-1965.

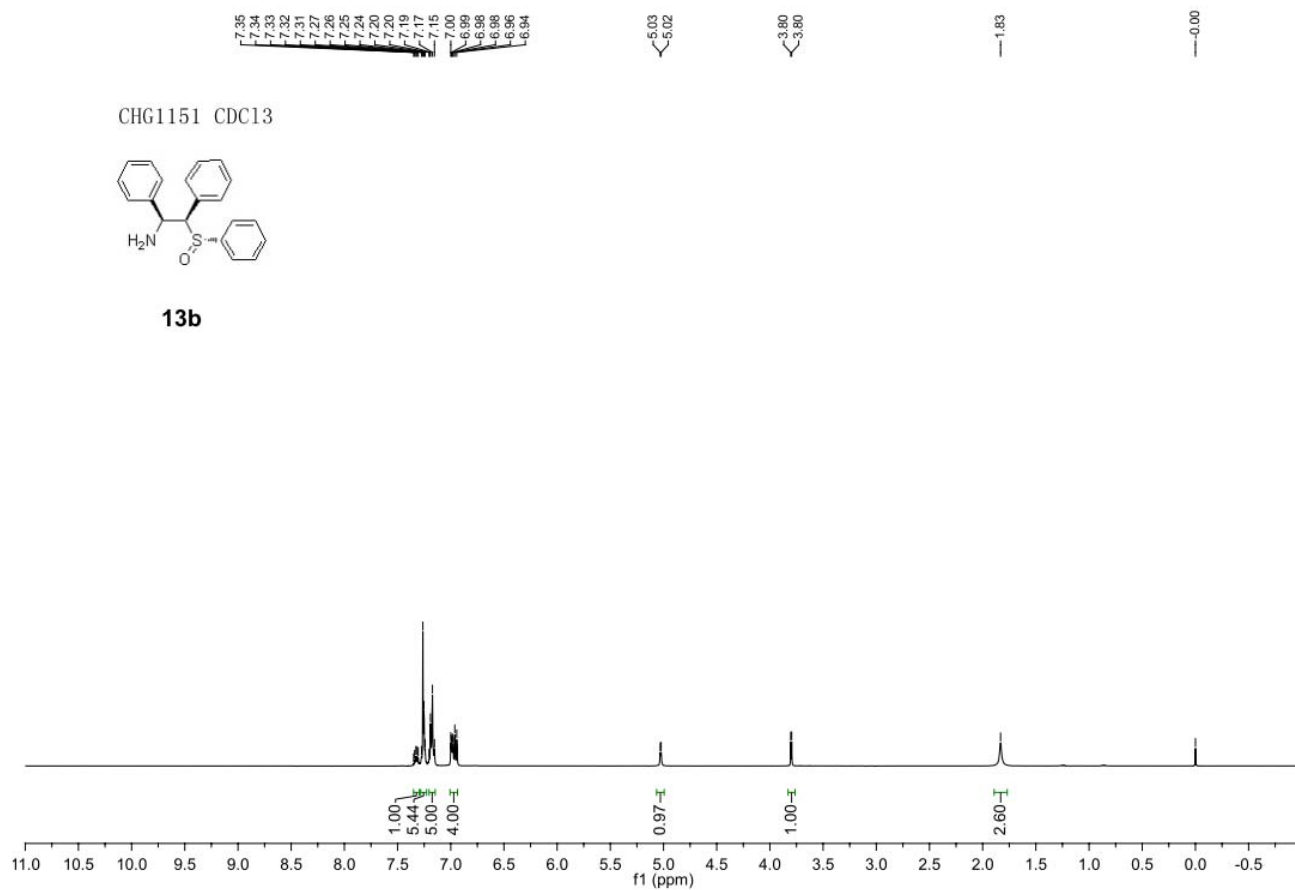
5. Copies of ^1H NMR and ^{13}C NMR Spectra



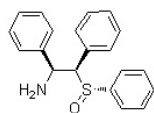
CHG1151 CDC13



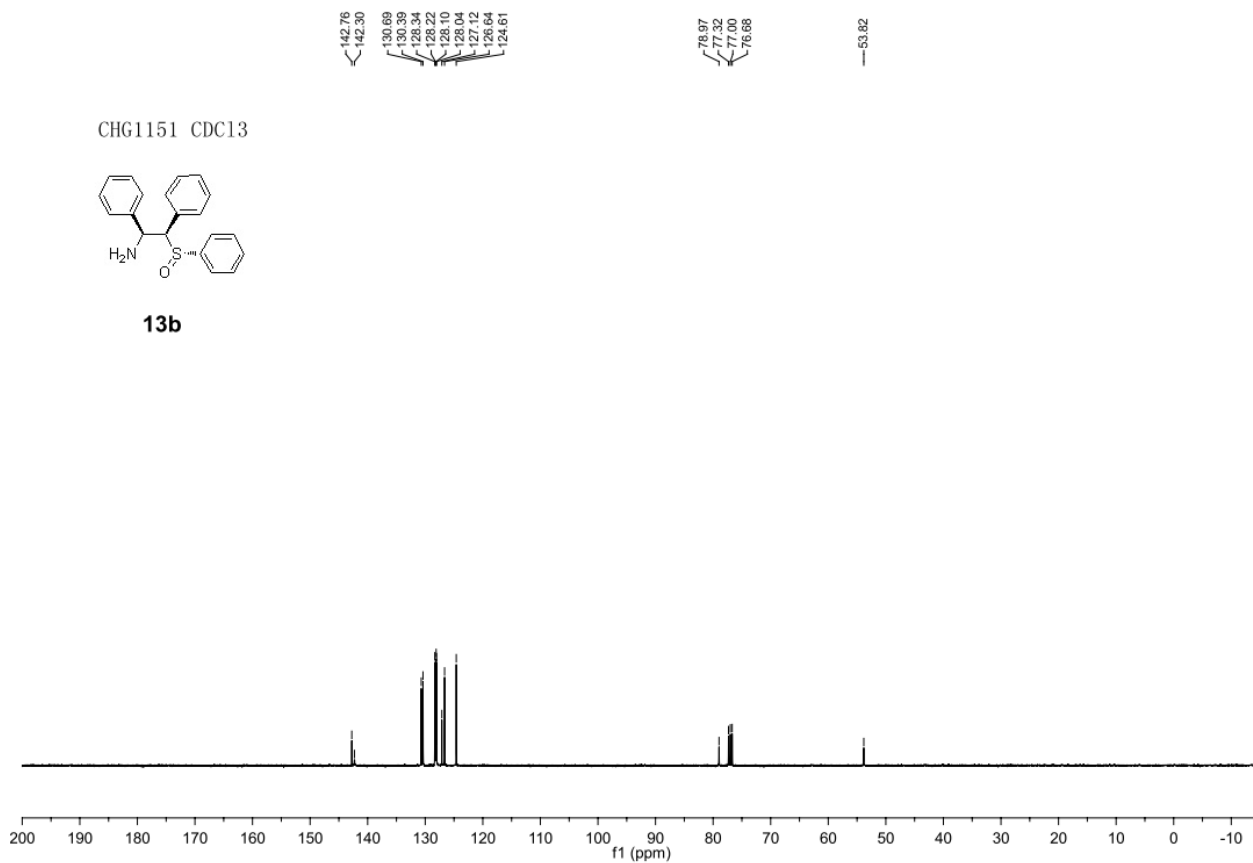
13b



CHG1151 CDC13

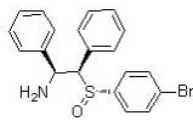


13b

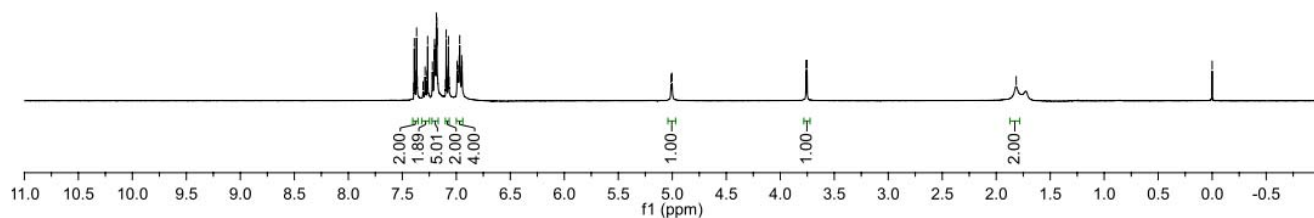




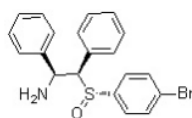
CHG1157 CDC13



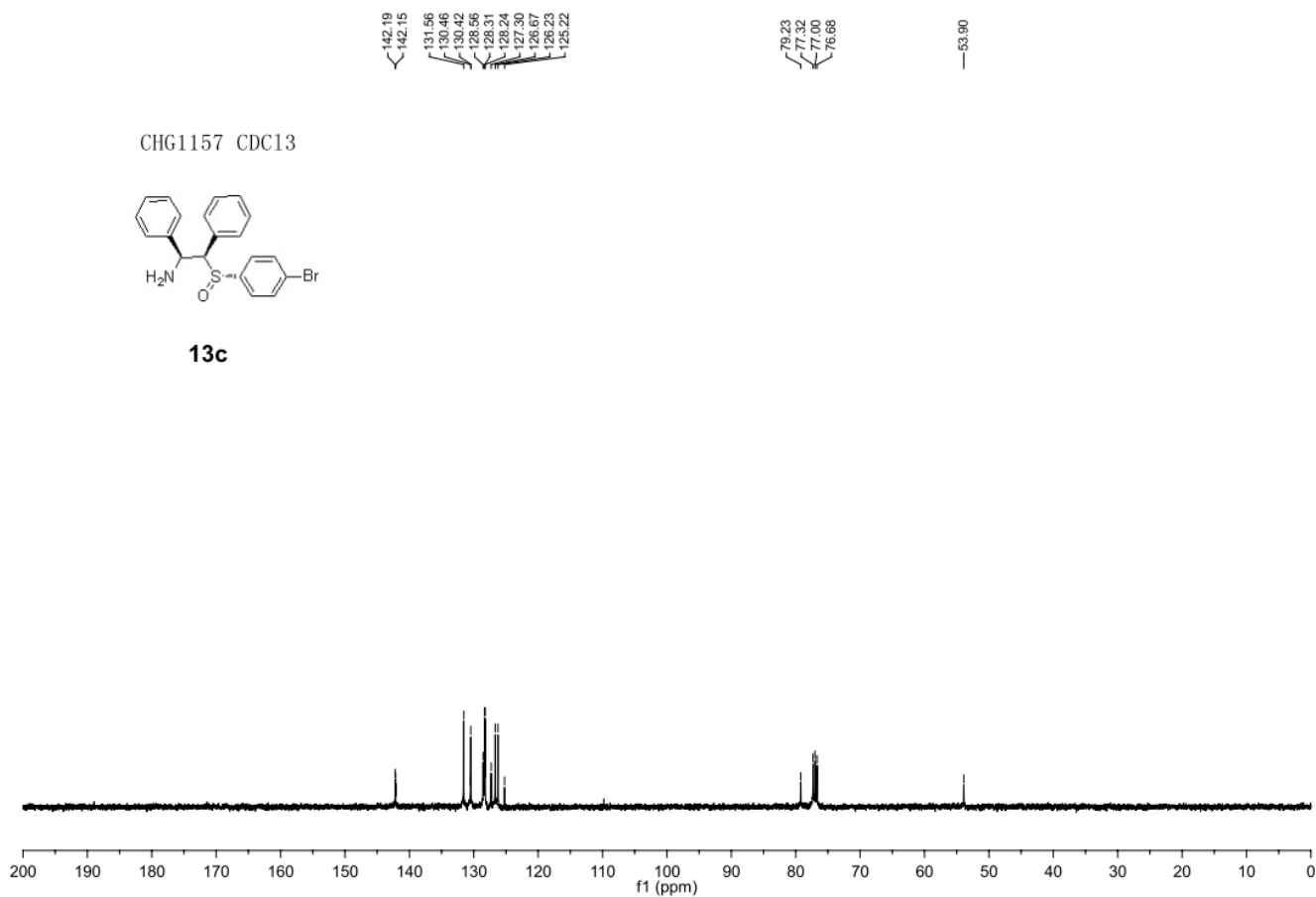
13c

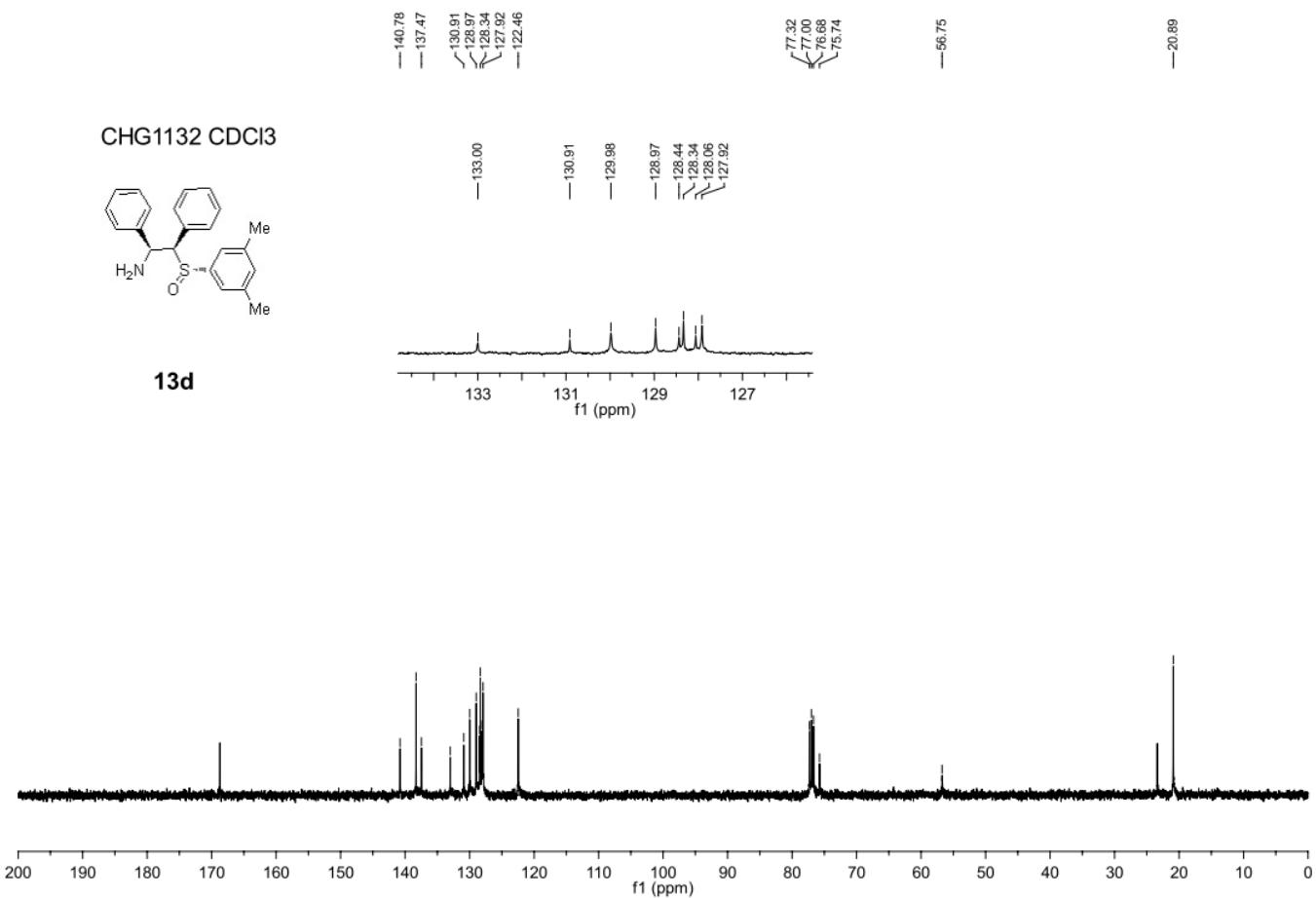
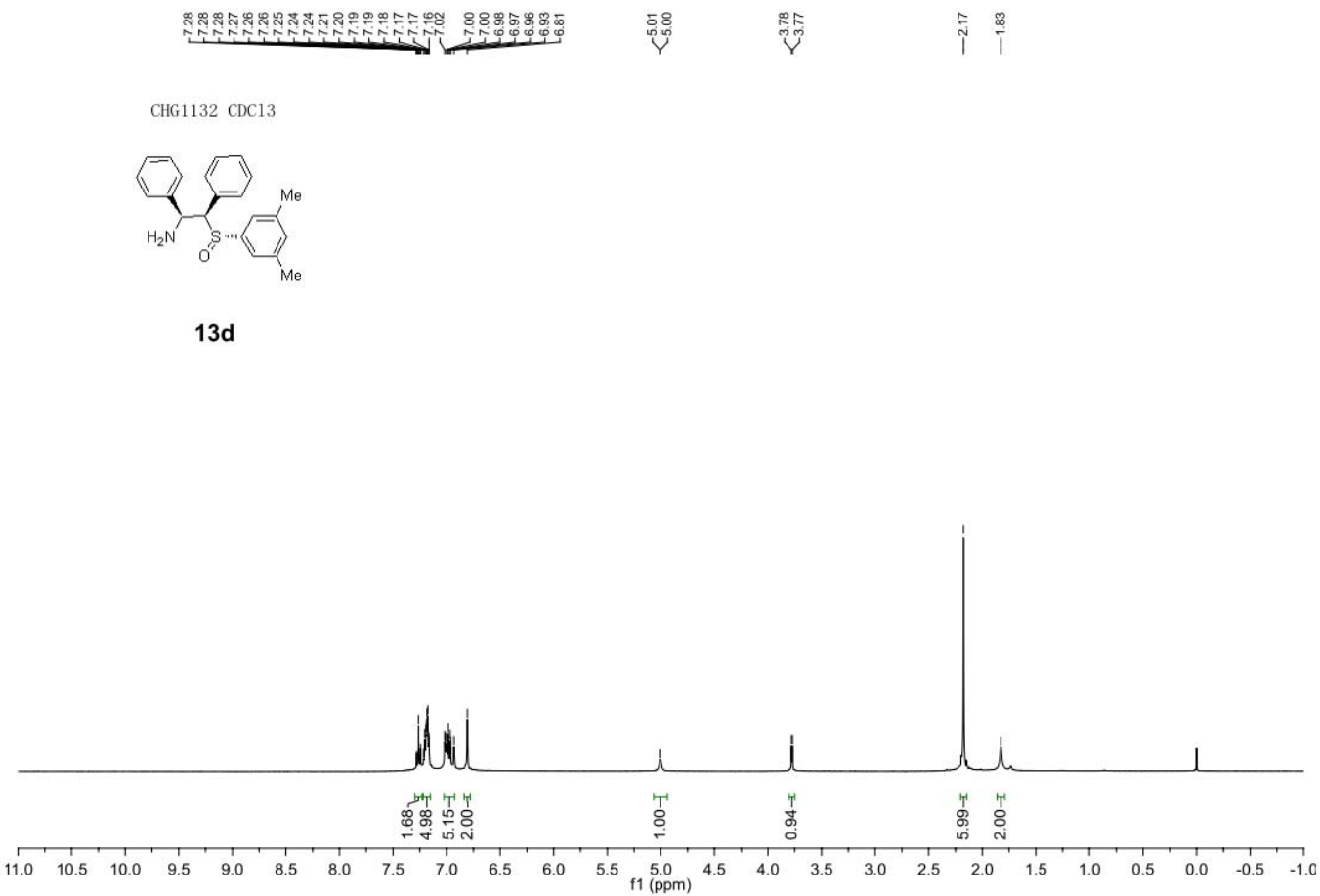


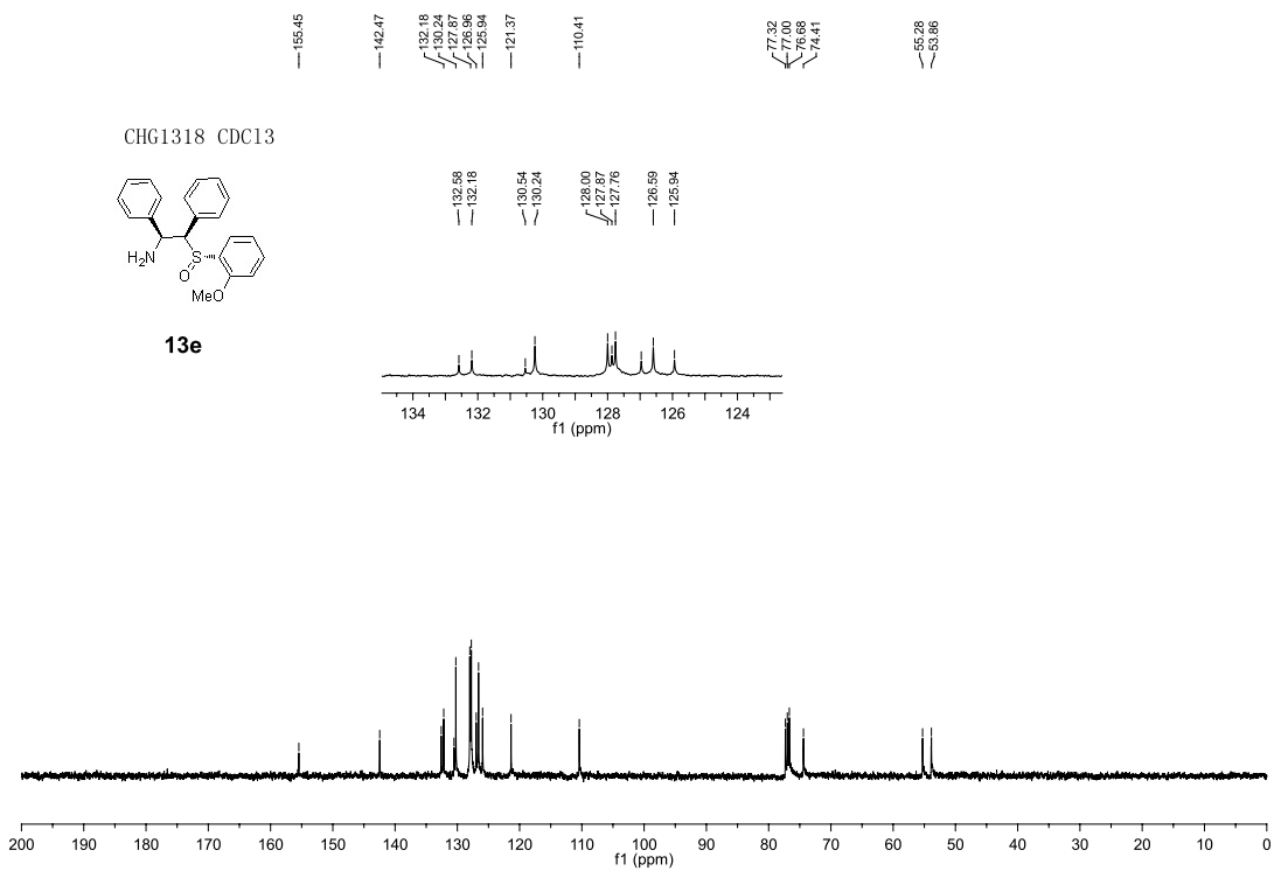
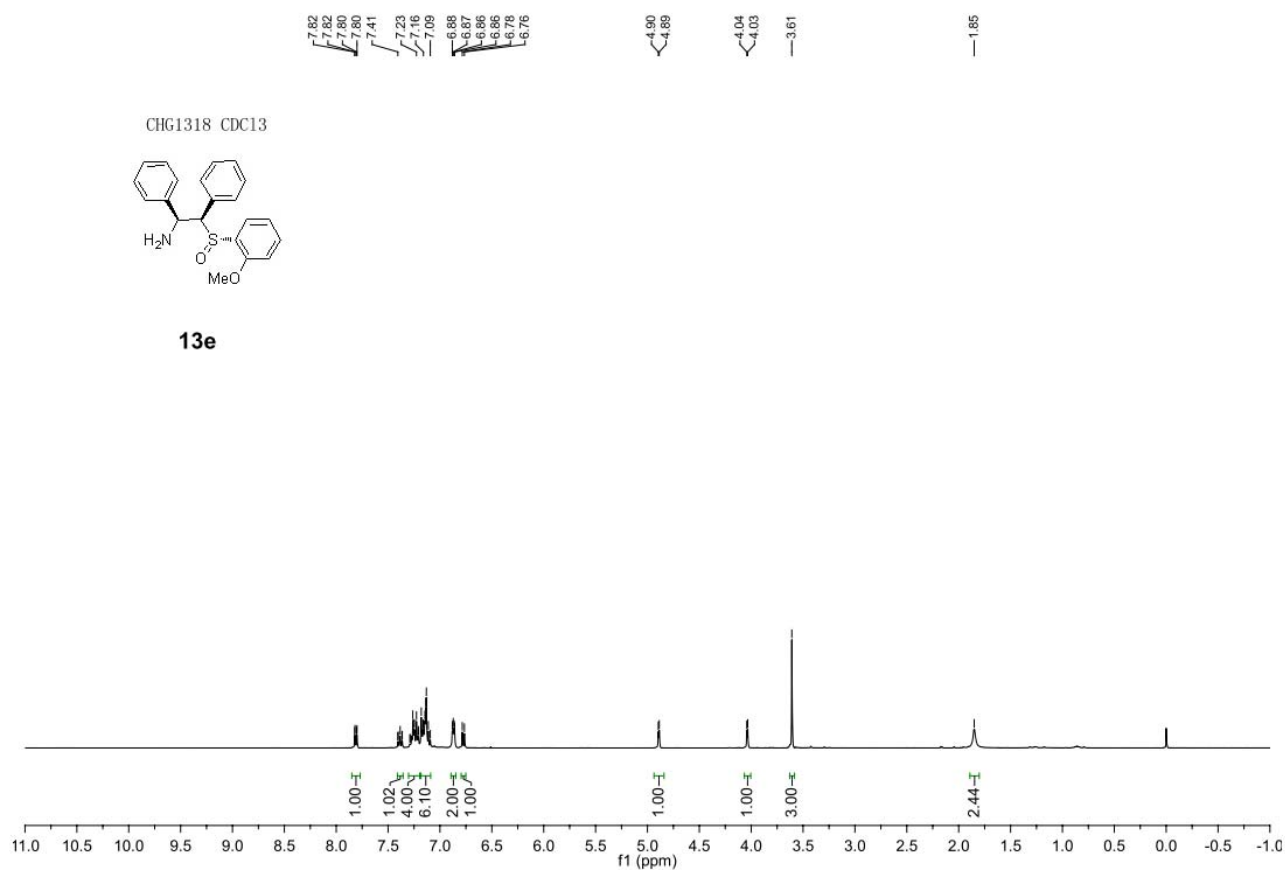
CHG1157 CDC13



13c

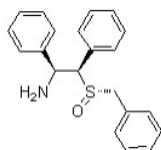




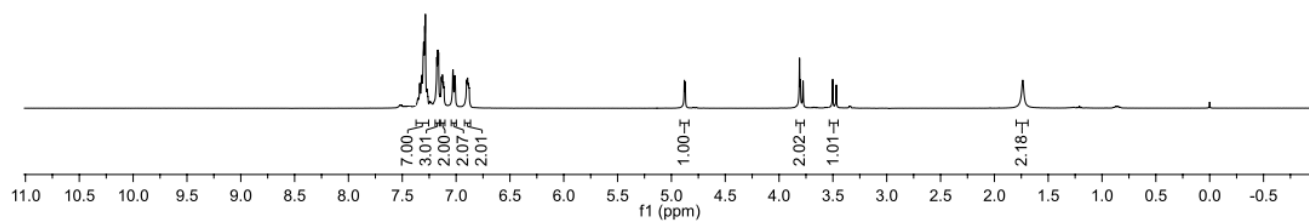




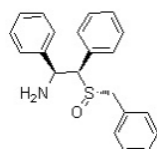
CHG1396 CDCl₃



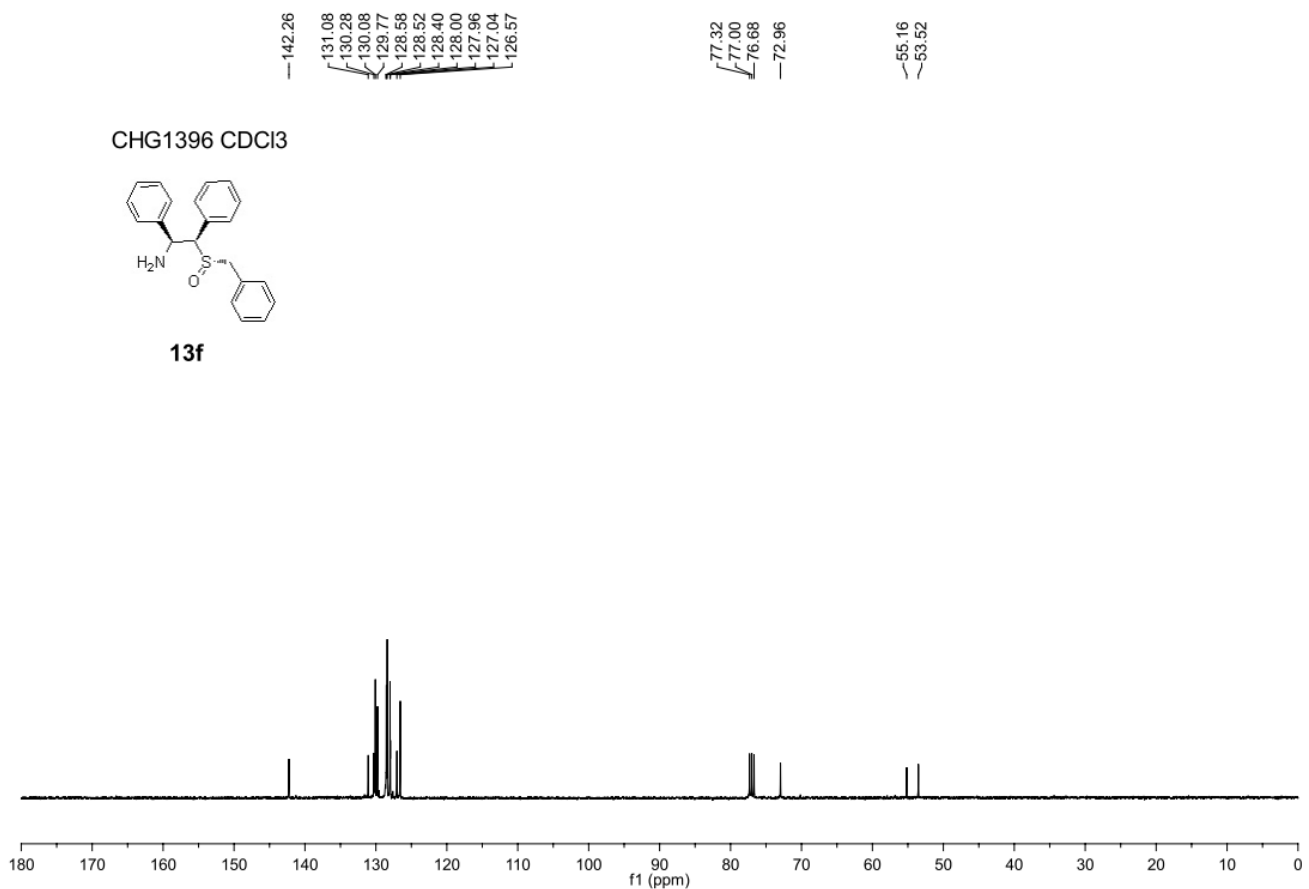
13f

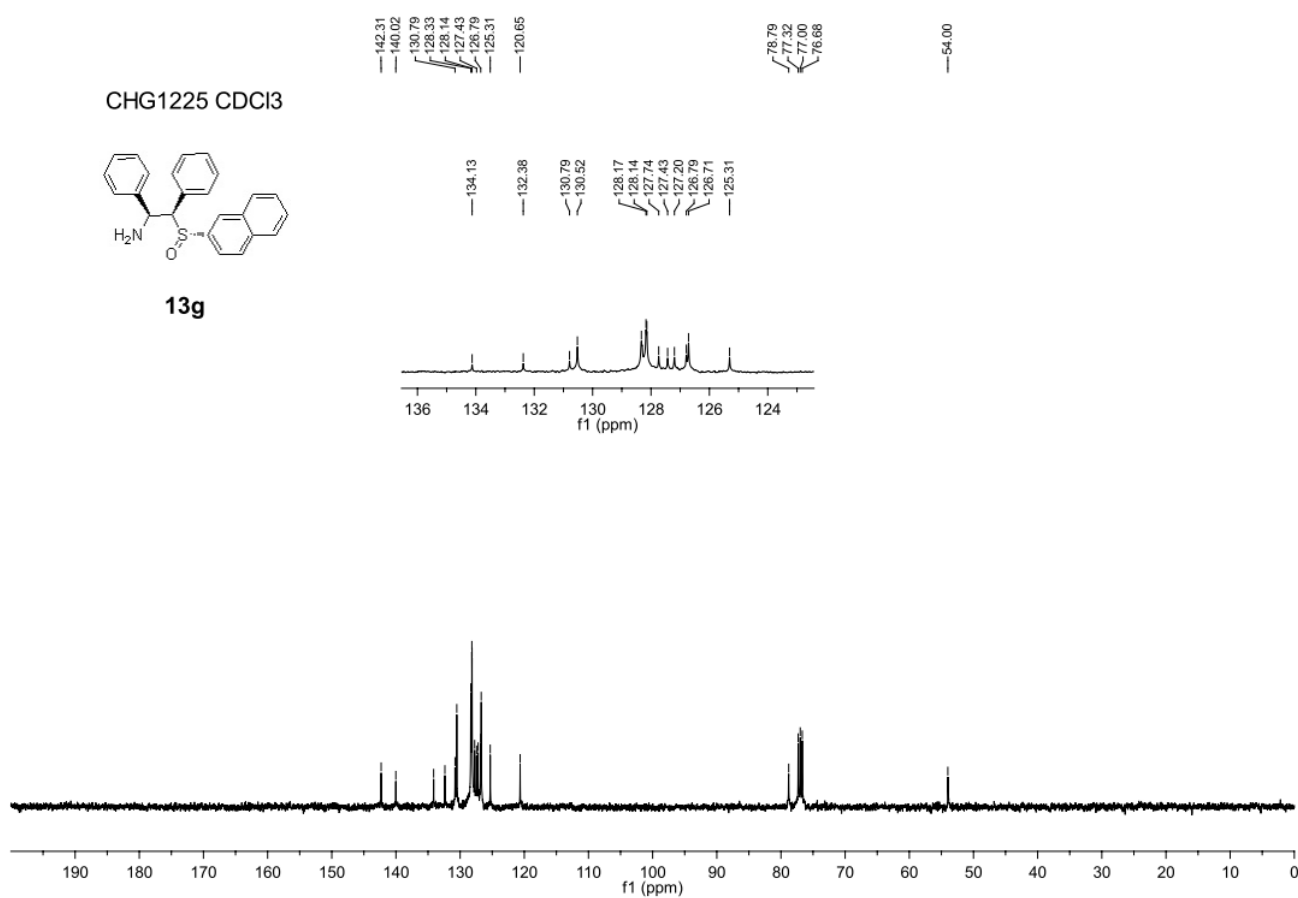
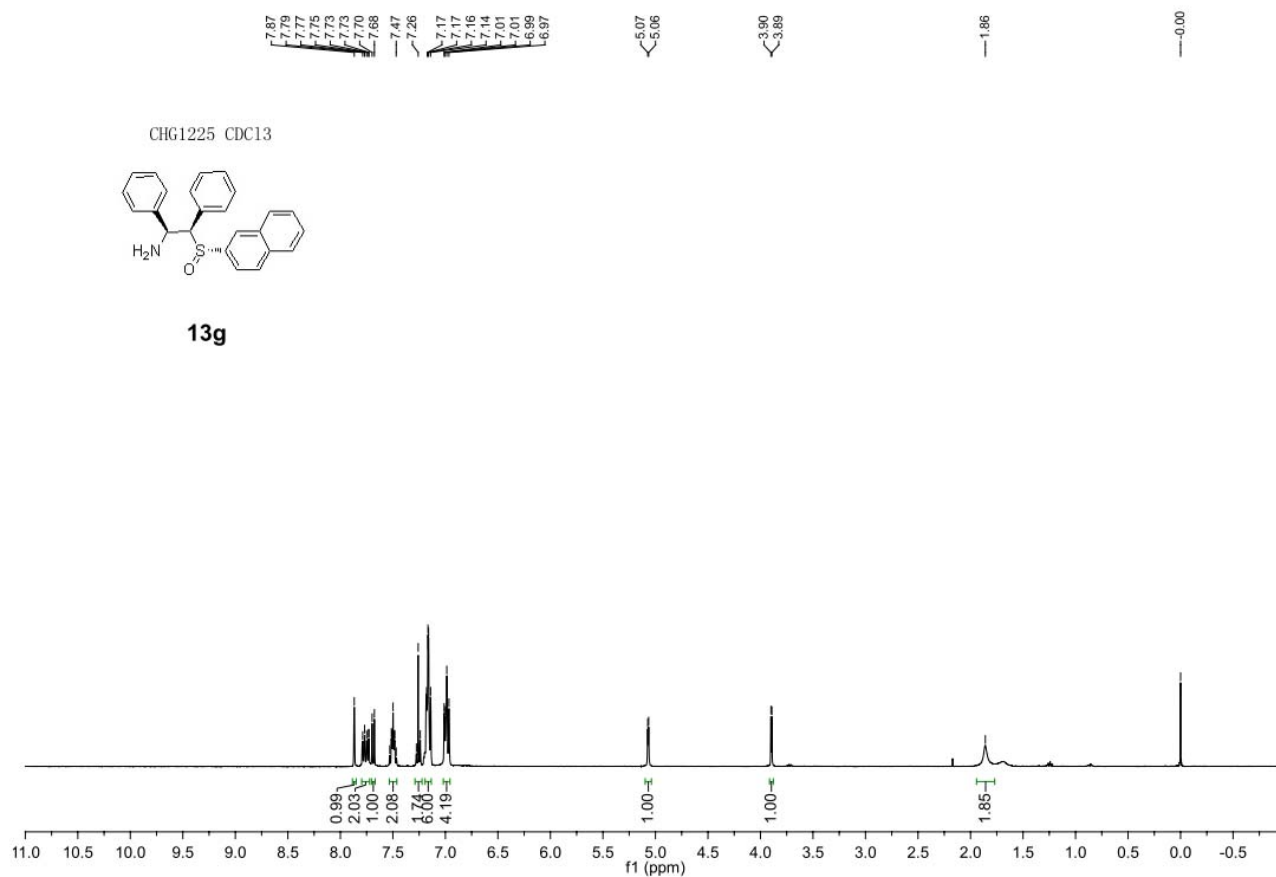


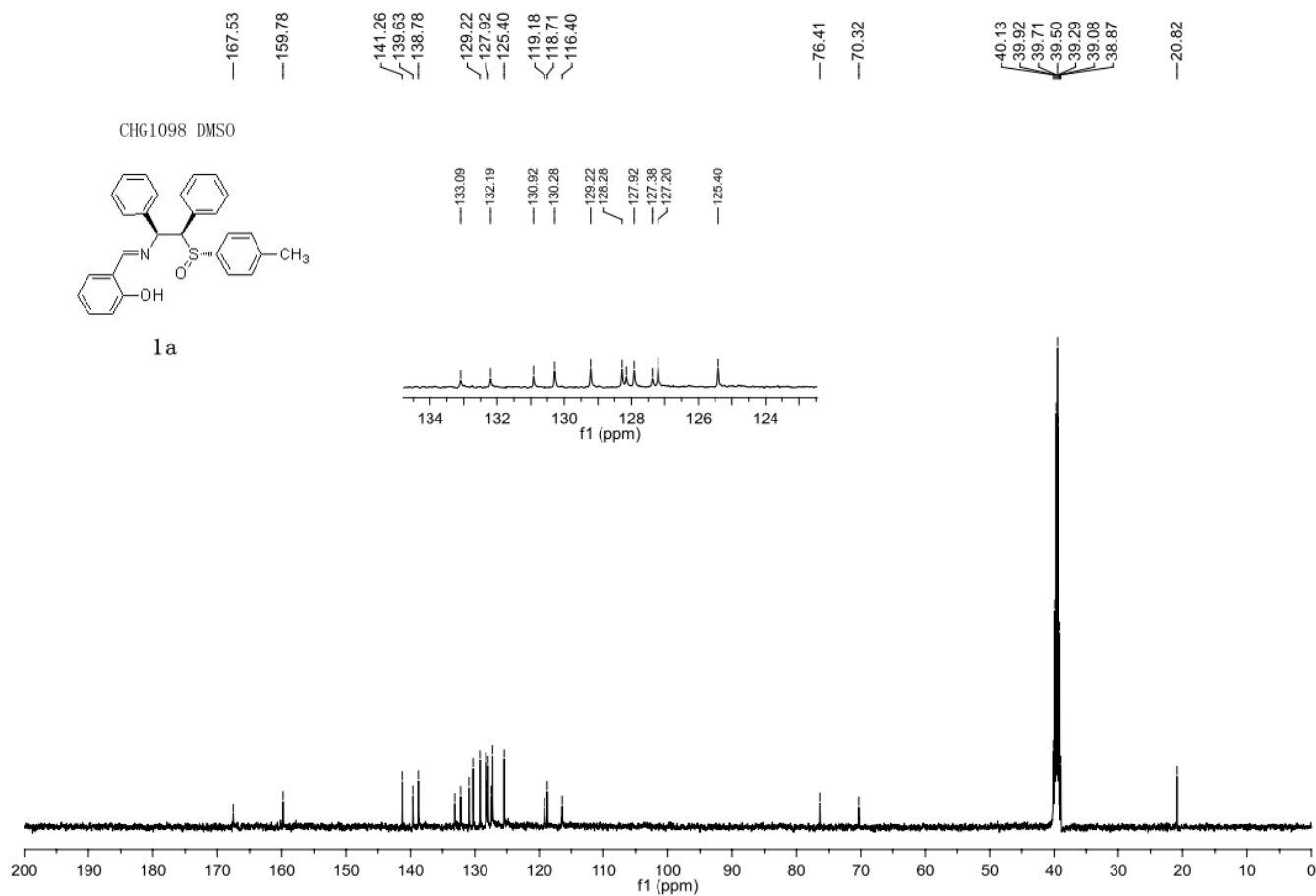
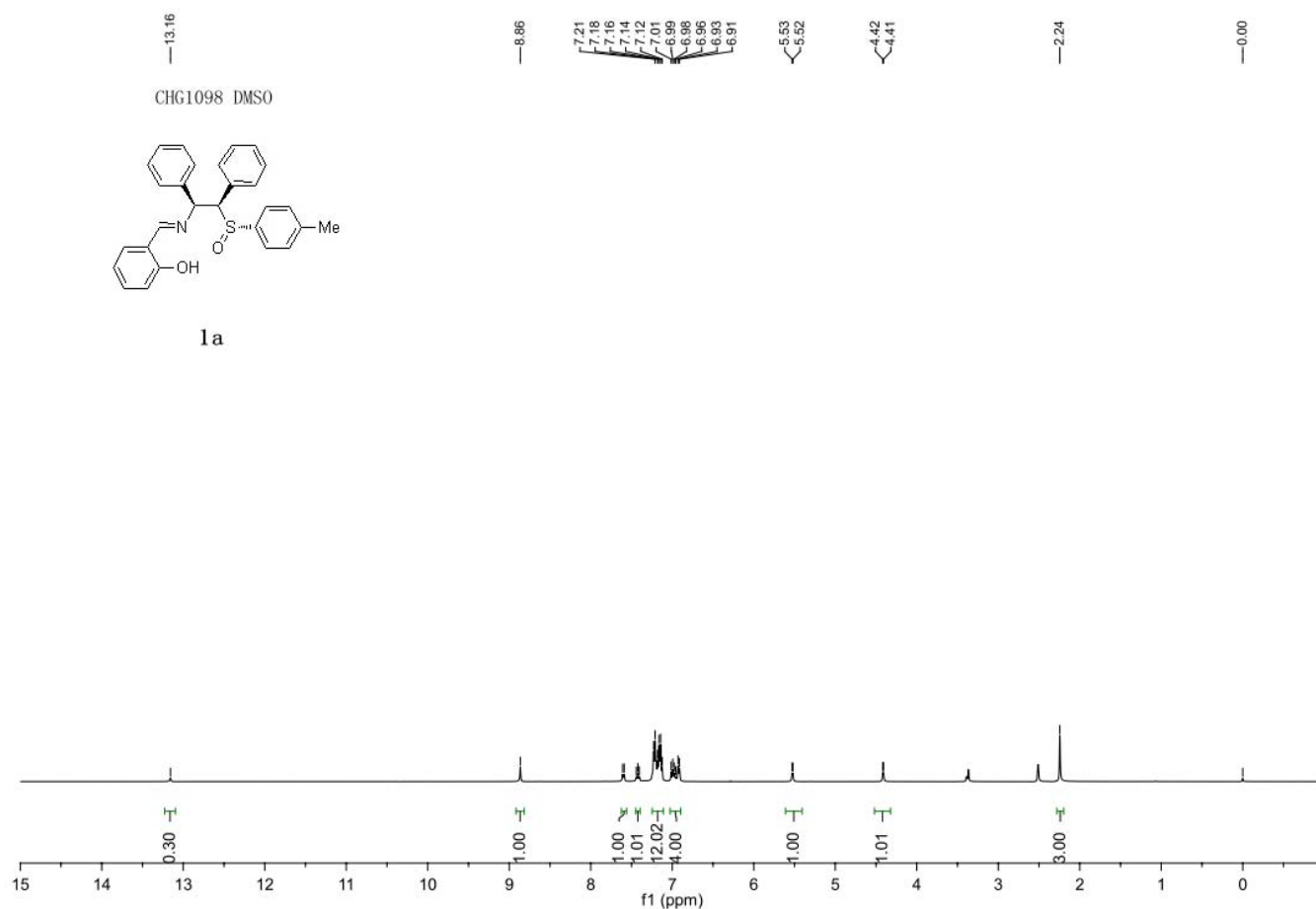
CHG1396 CDCl₃

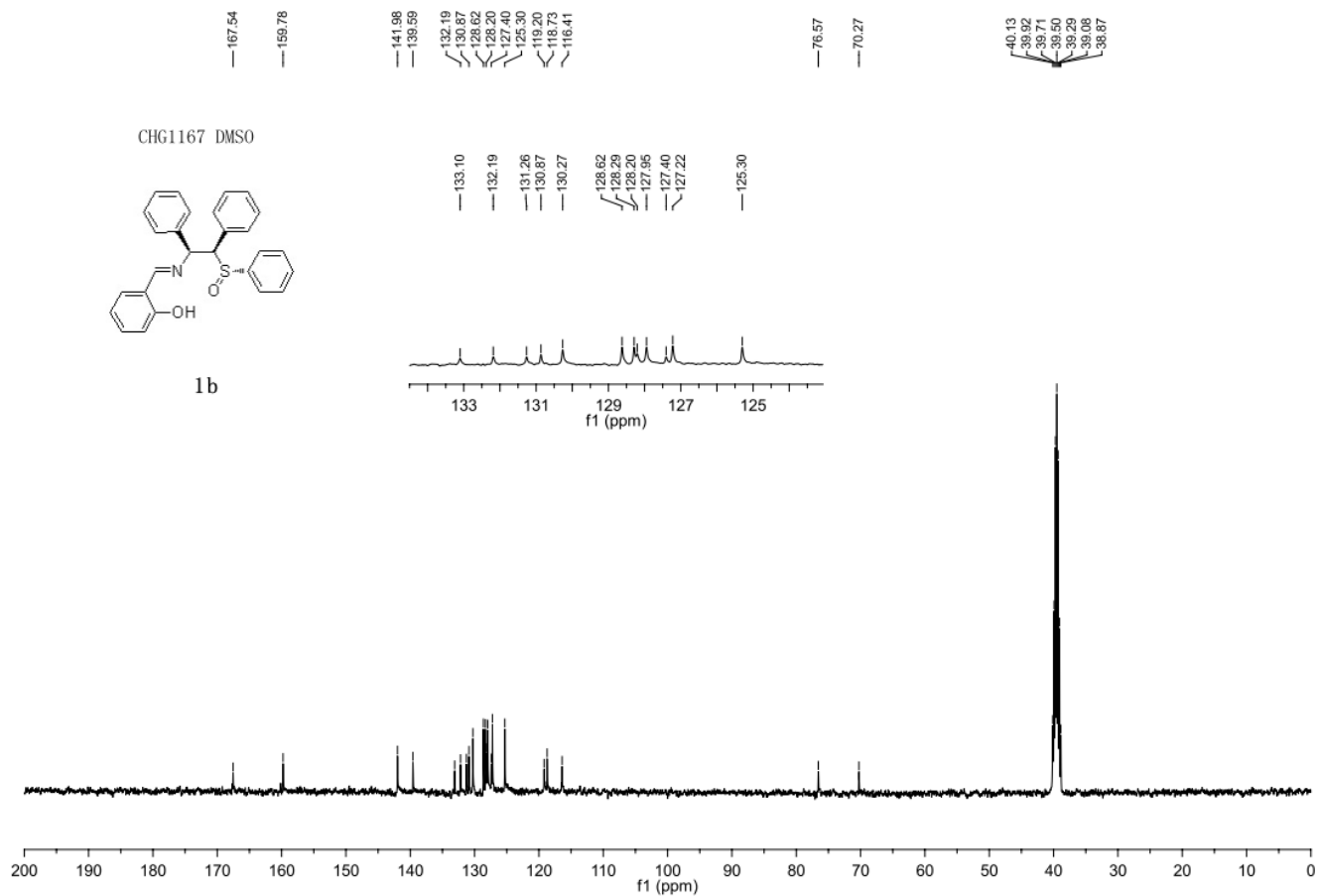
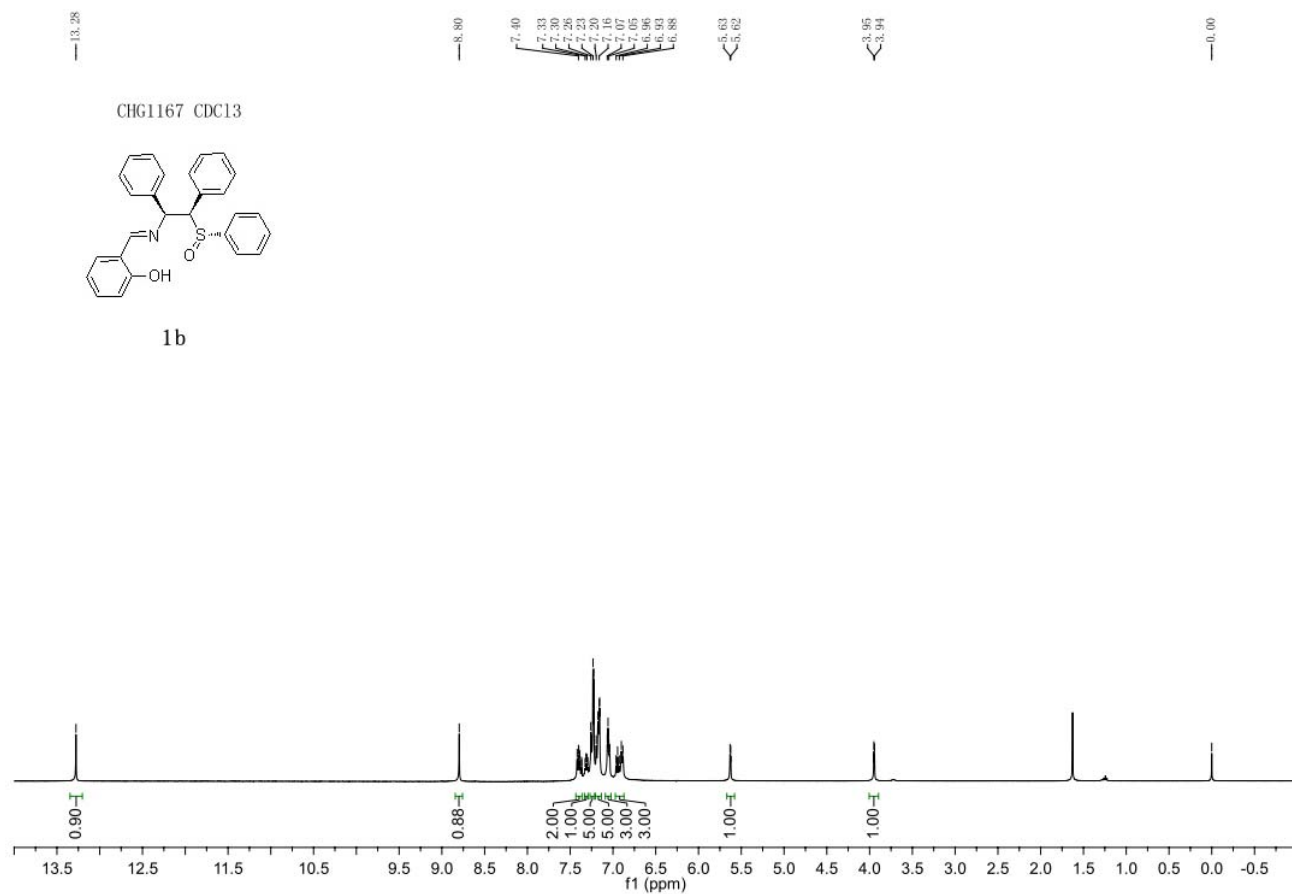


13f









— 13.22

— 8.77

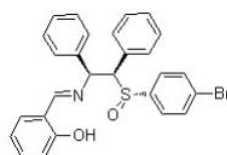
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7.04
6.96
6.94
6.92
6.90

5.60
5.59

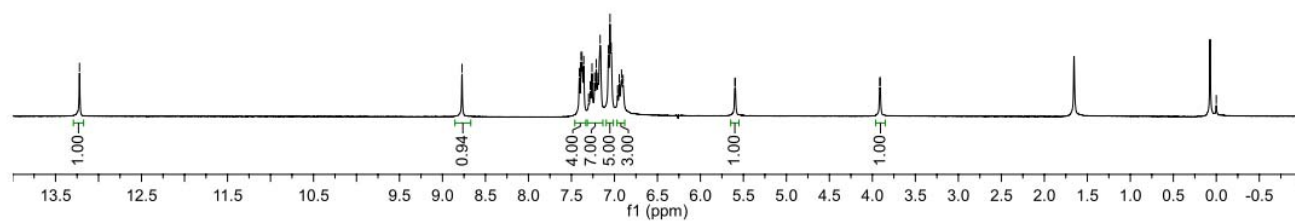
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3.91

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CHG1166 CDC13



1c



— 167.53

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159.73

141.44

139.44

130.72

128.30

127.44

127.27

124.81

119.18

118.71

116.39

76.20

70.18

40.13

39.92

39.71

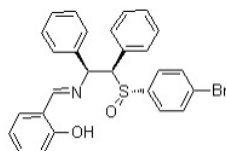
39.50

39.29

39.08

38.87

CHG1166 DMSO



1c

133.11

132.13

131.56

130.72

130.26

128.30

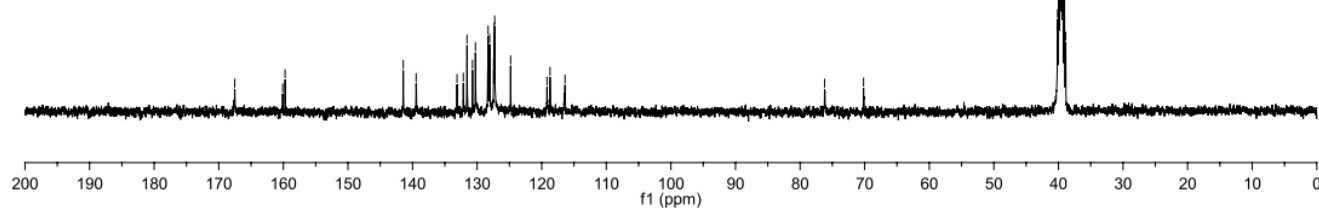
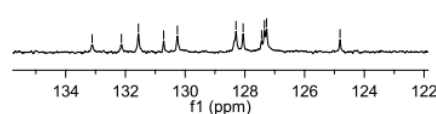
128.06

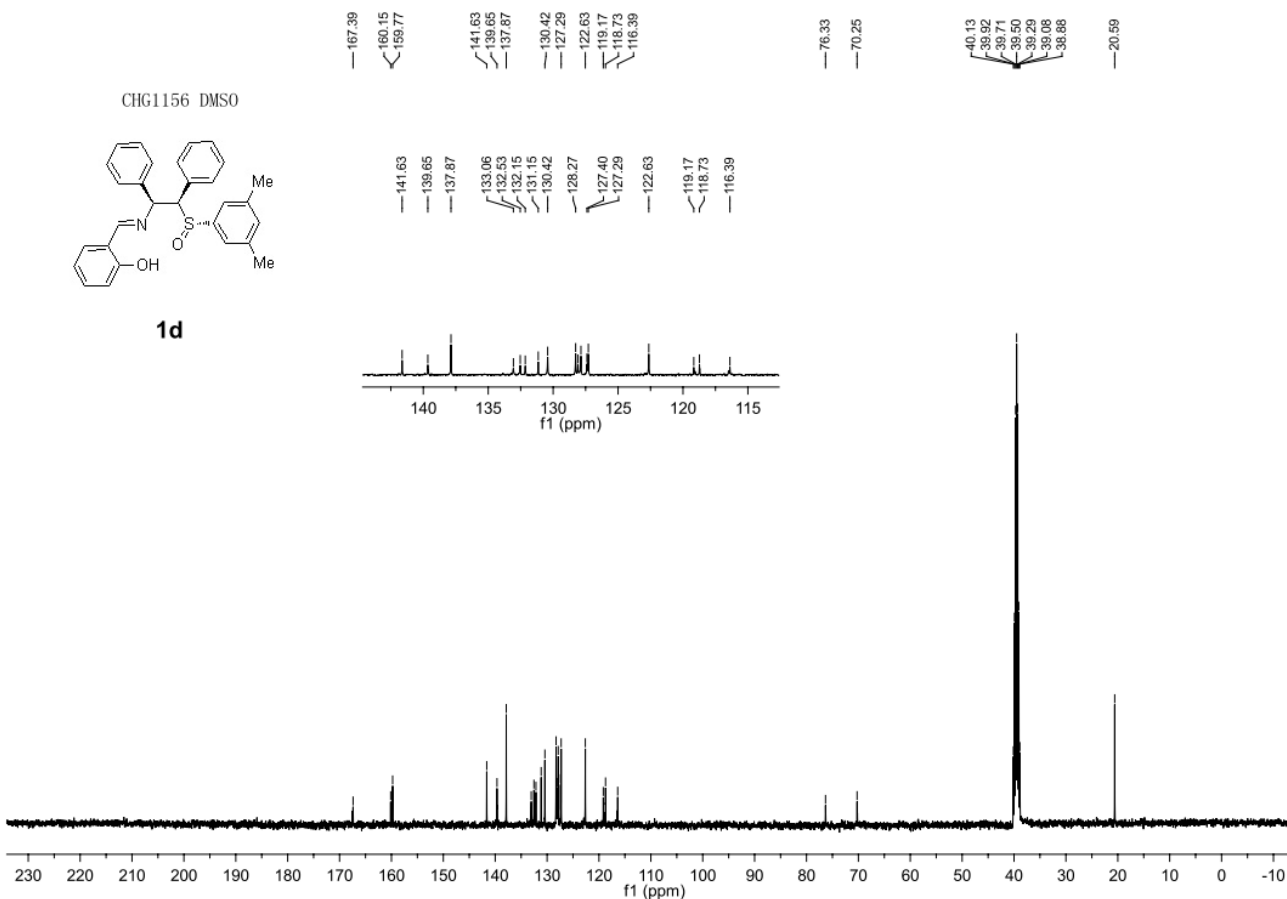
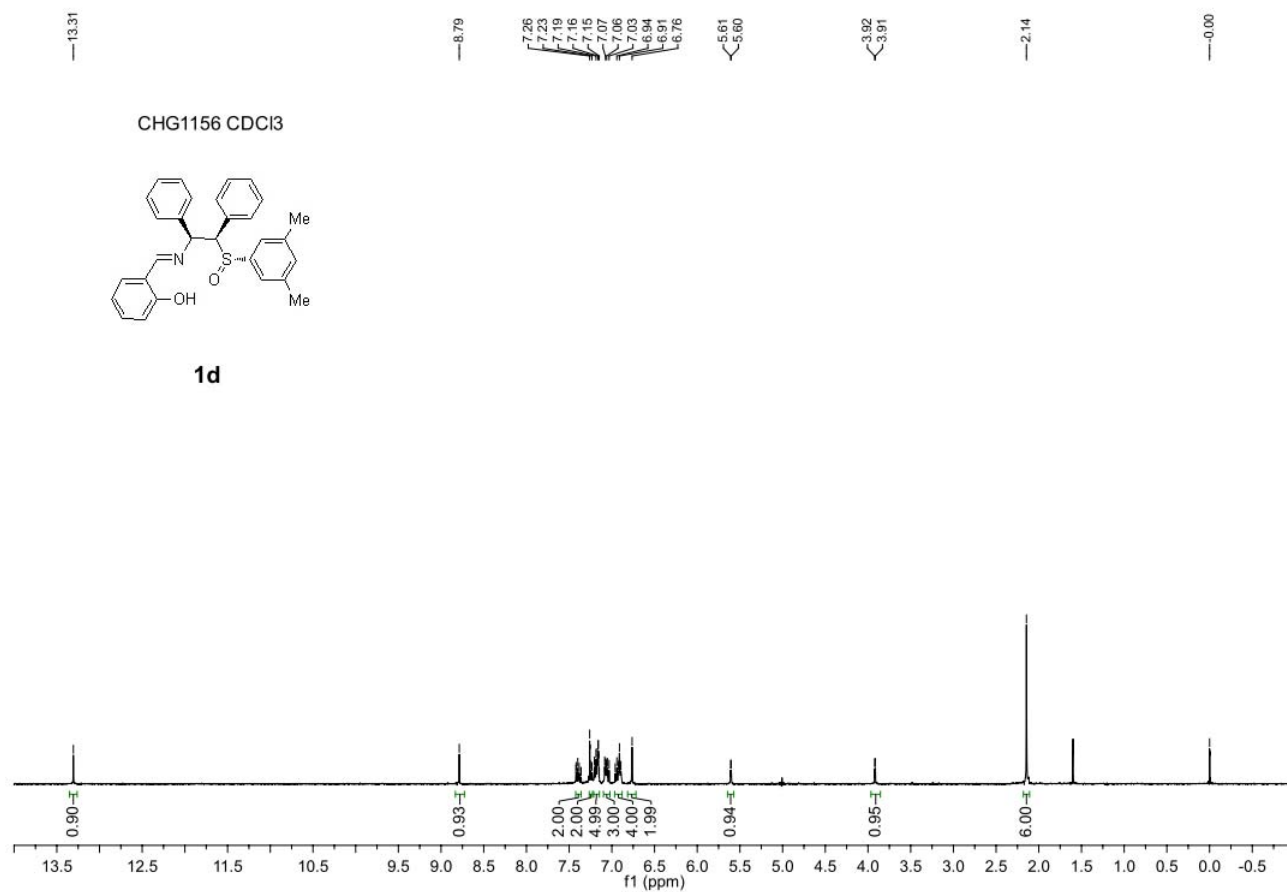
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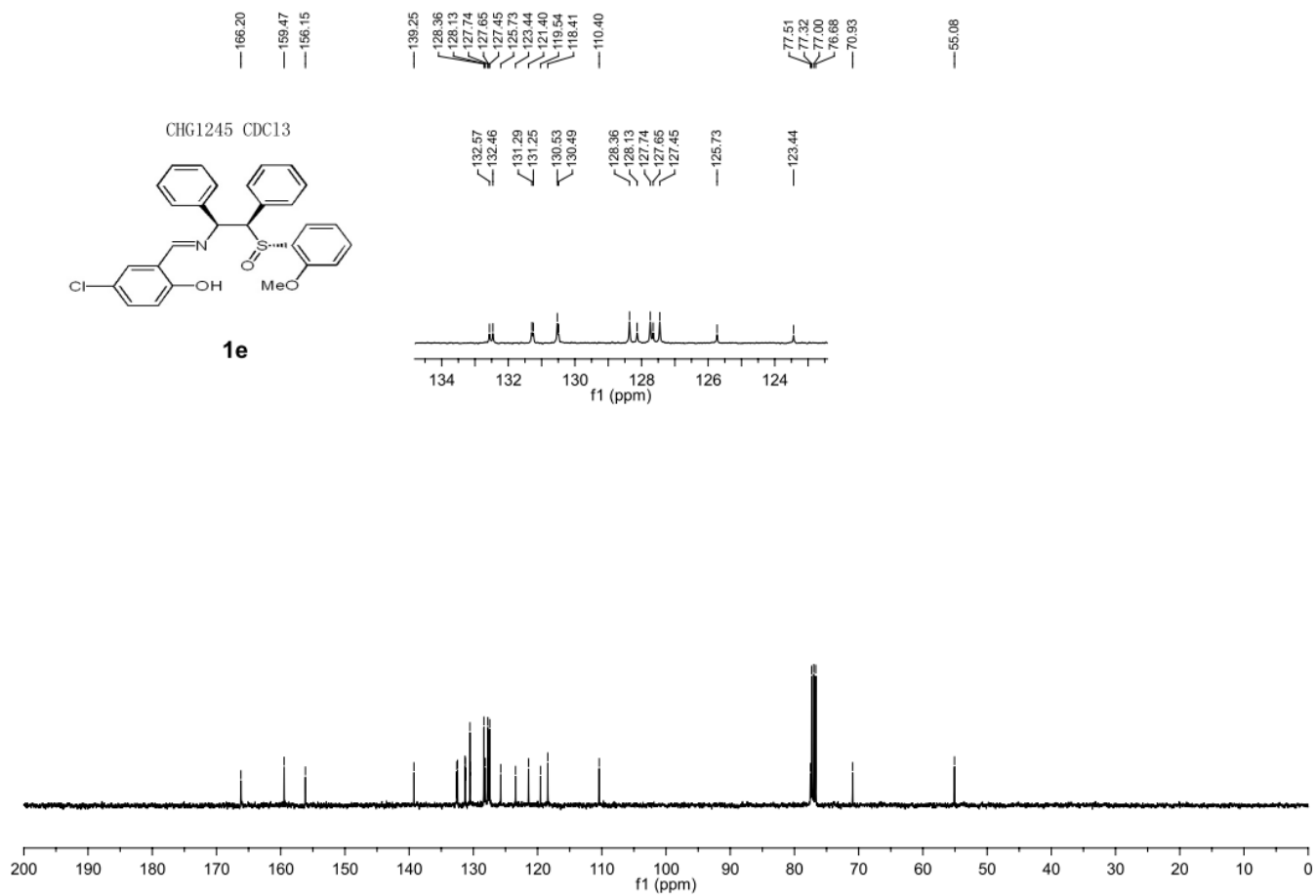
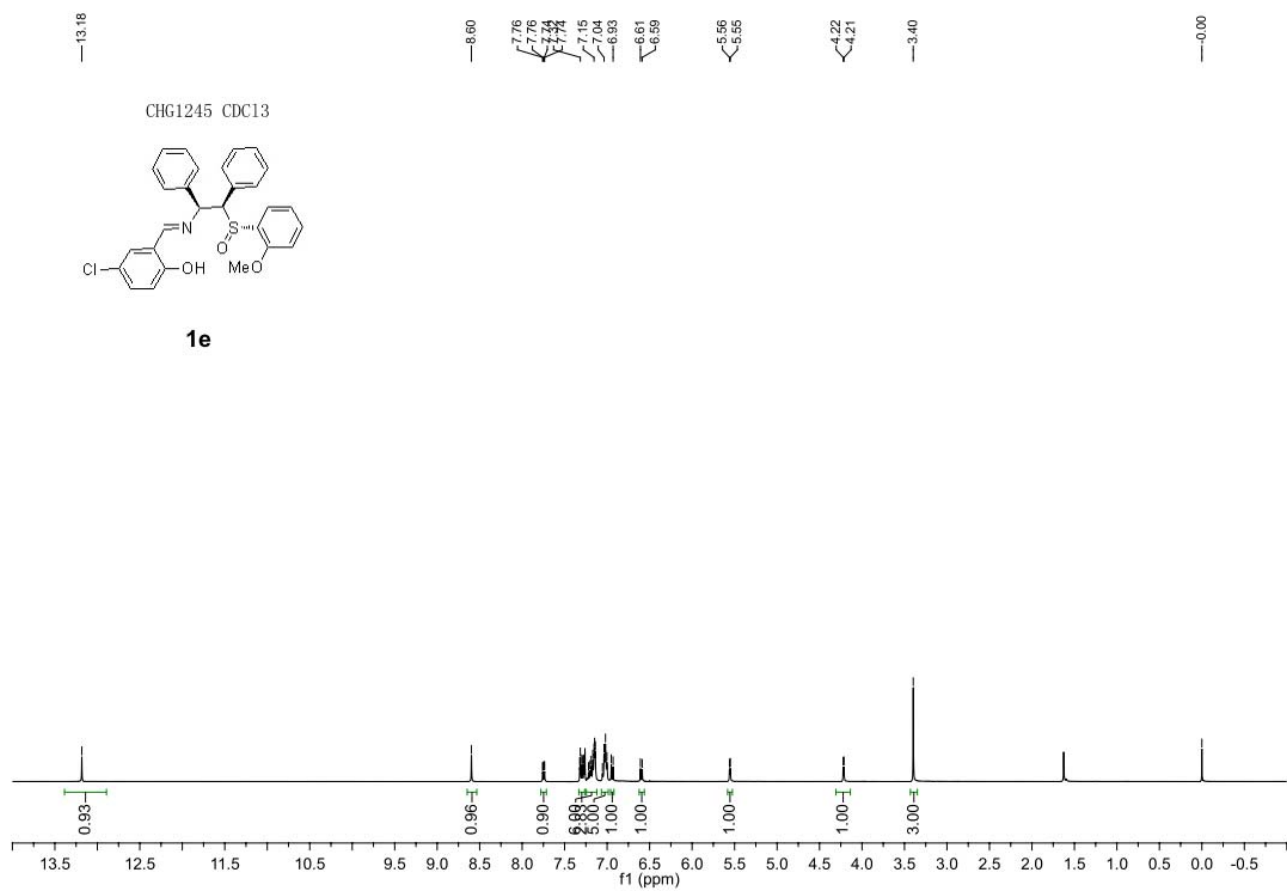
127.35

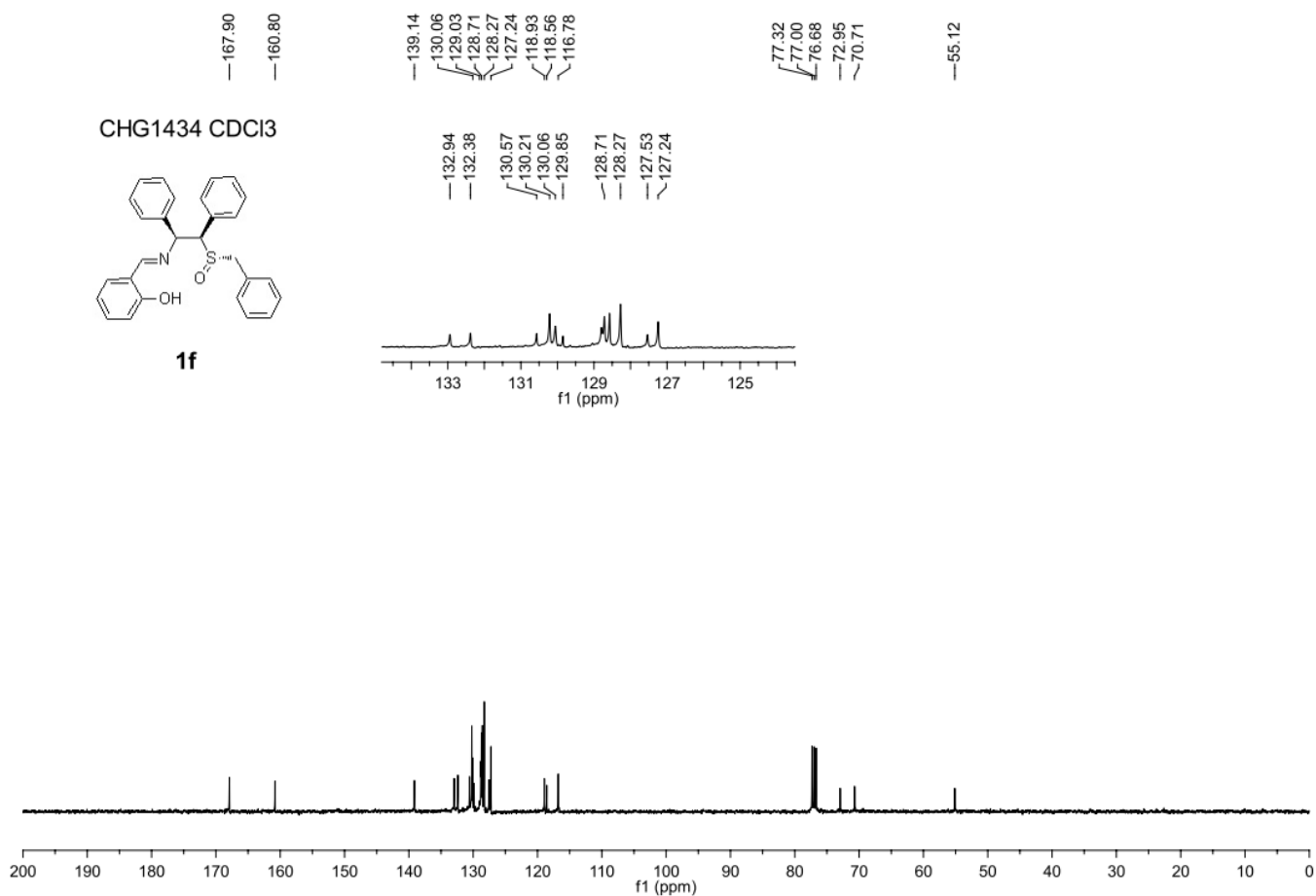
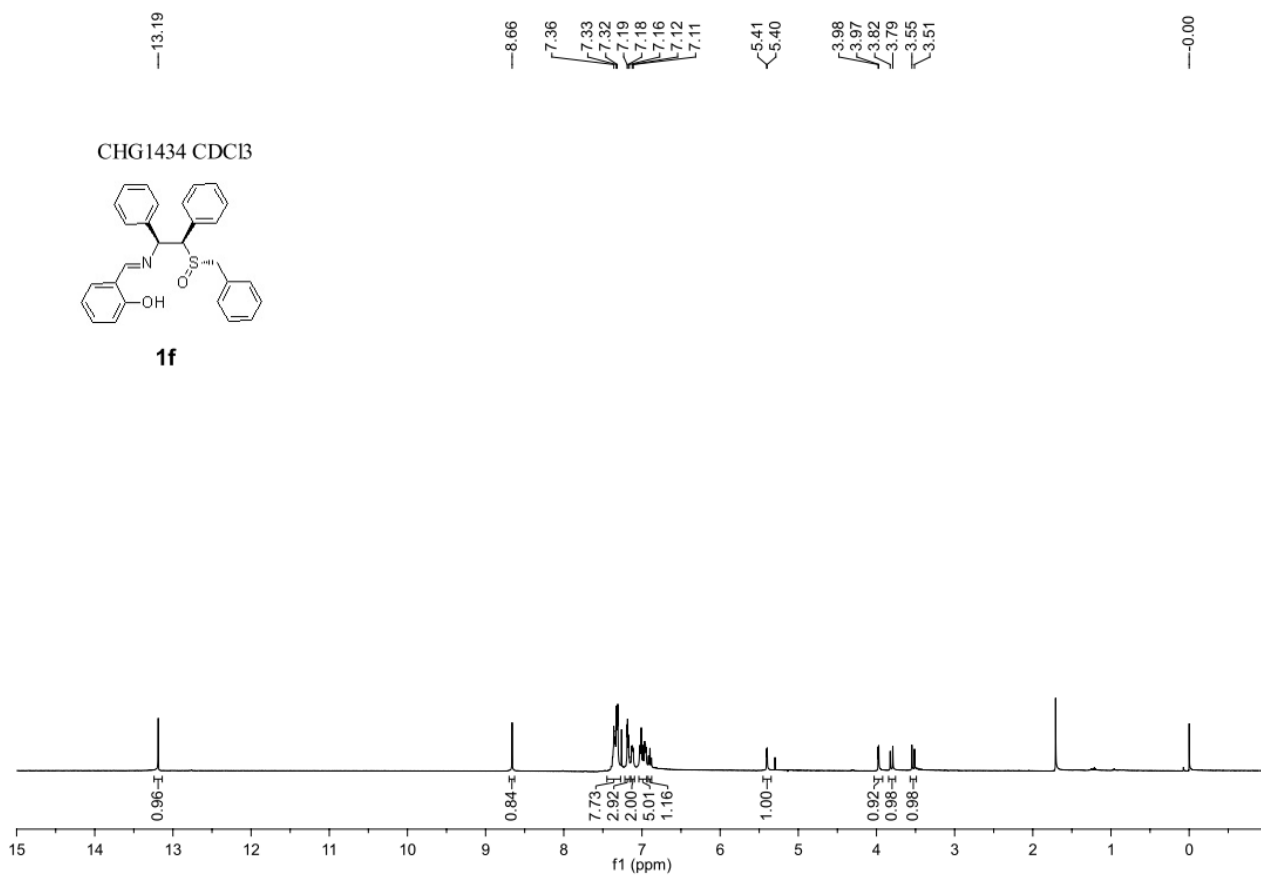
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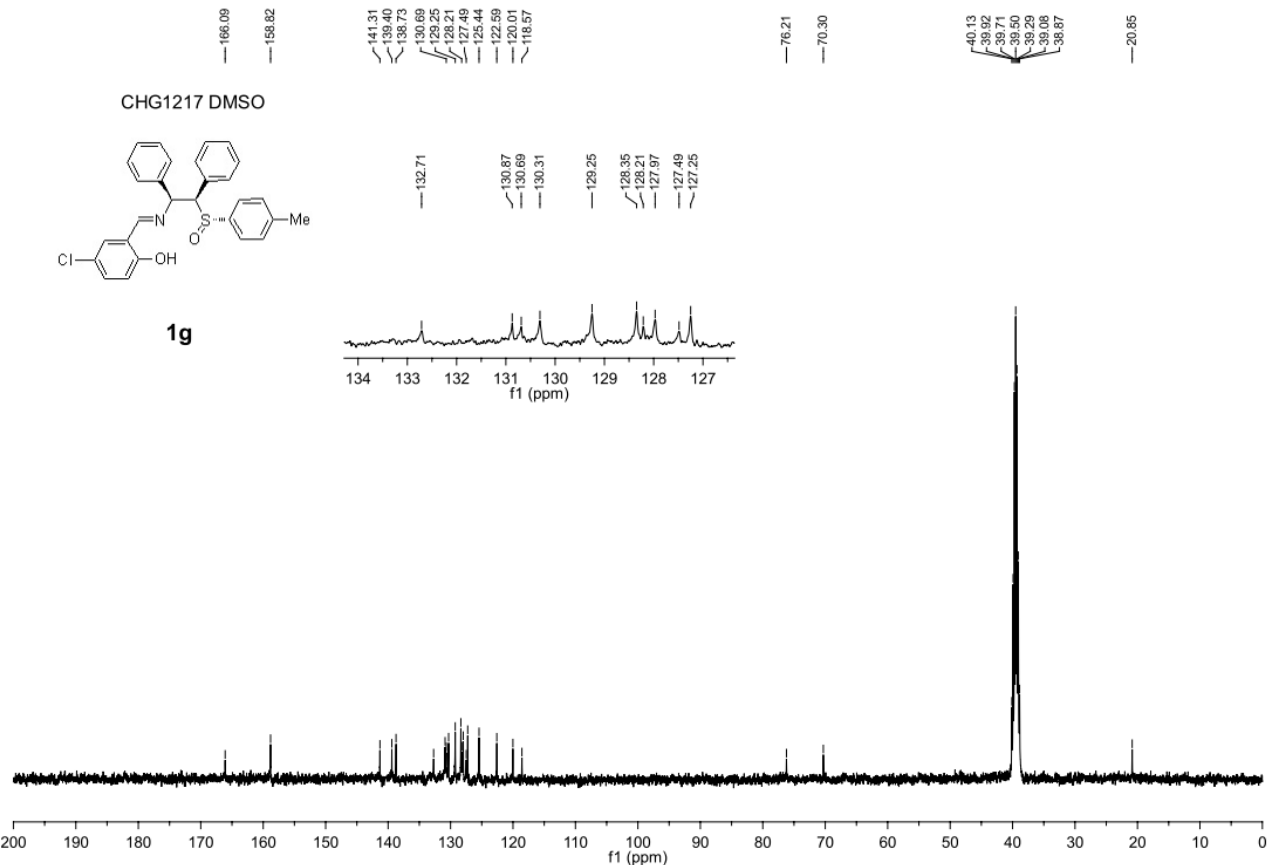
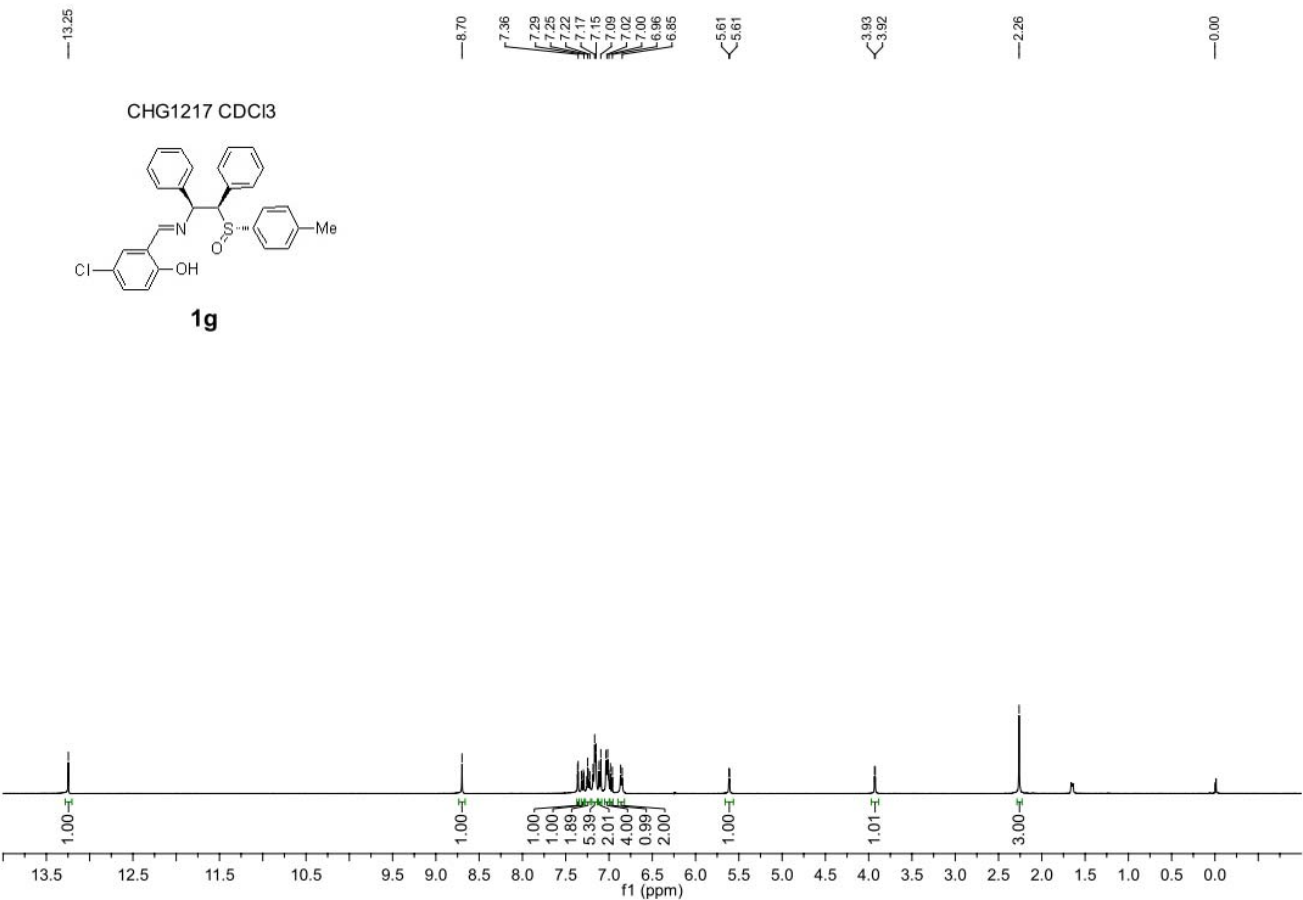
124.81

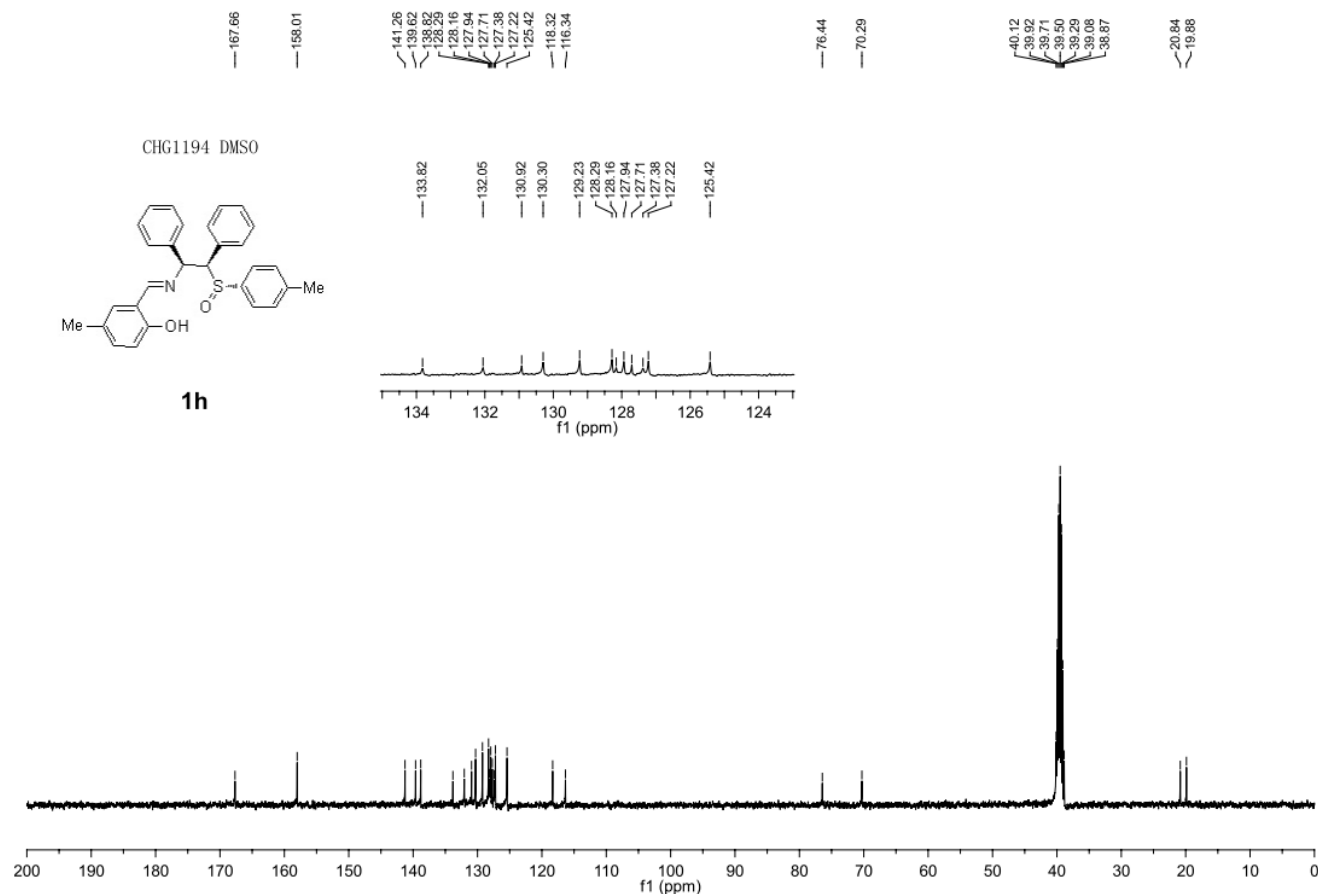
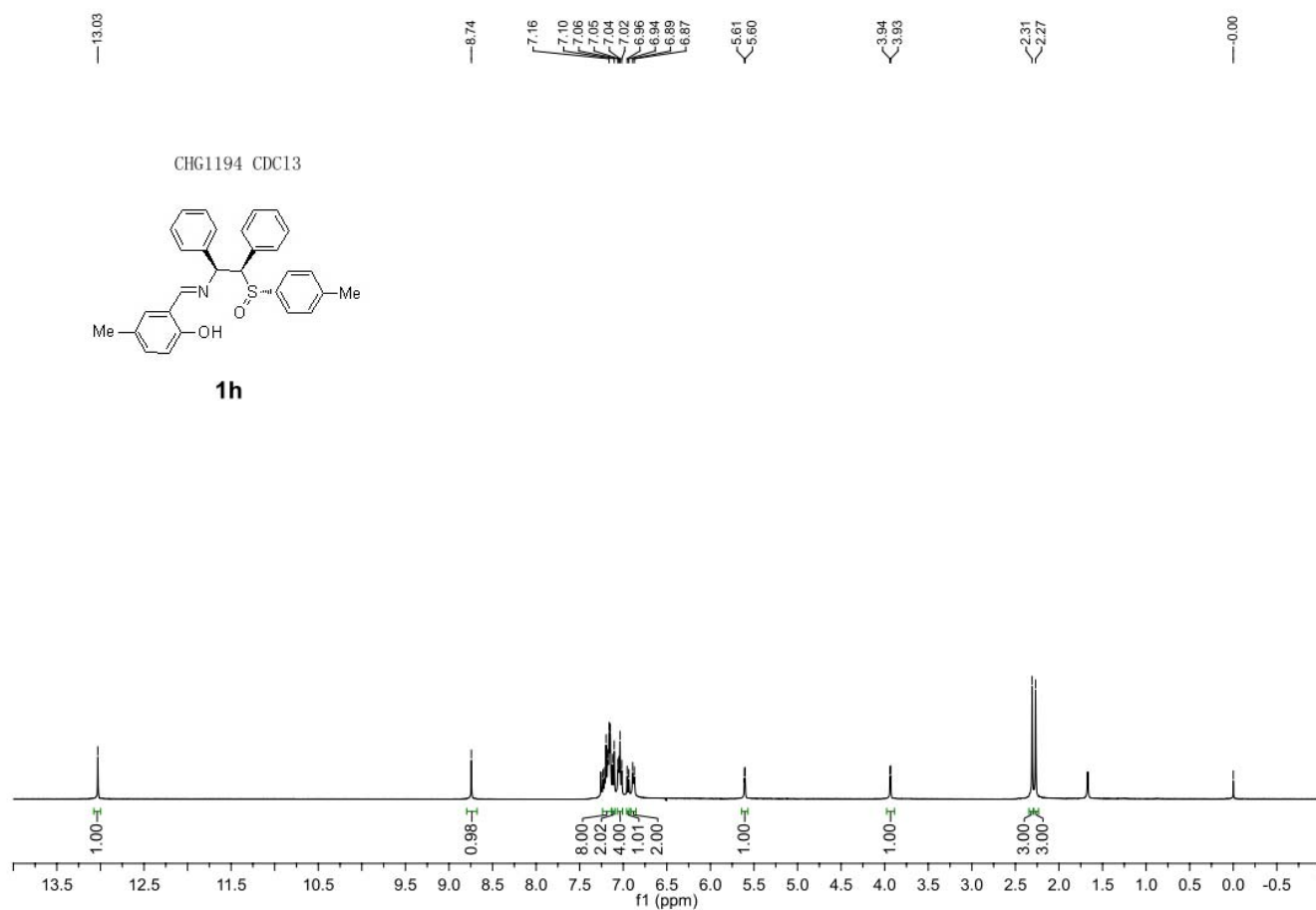


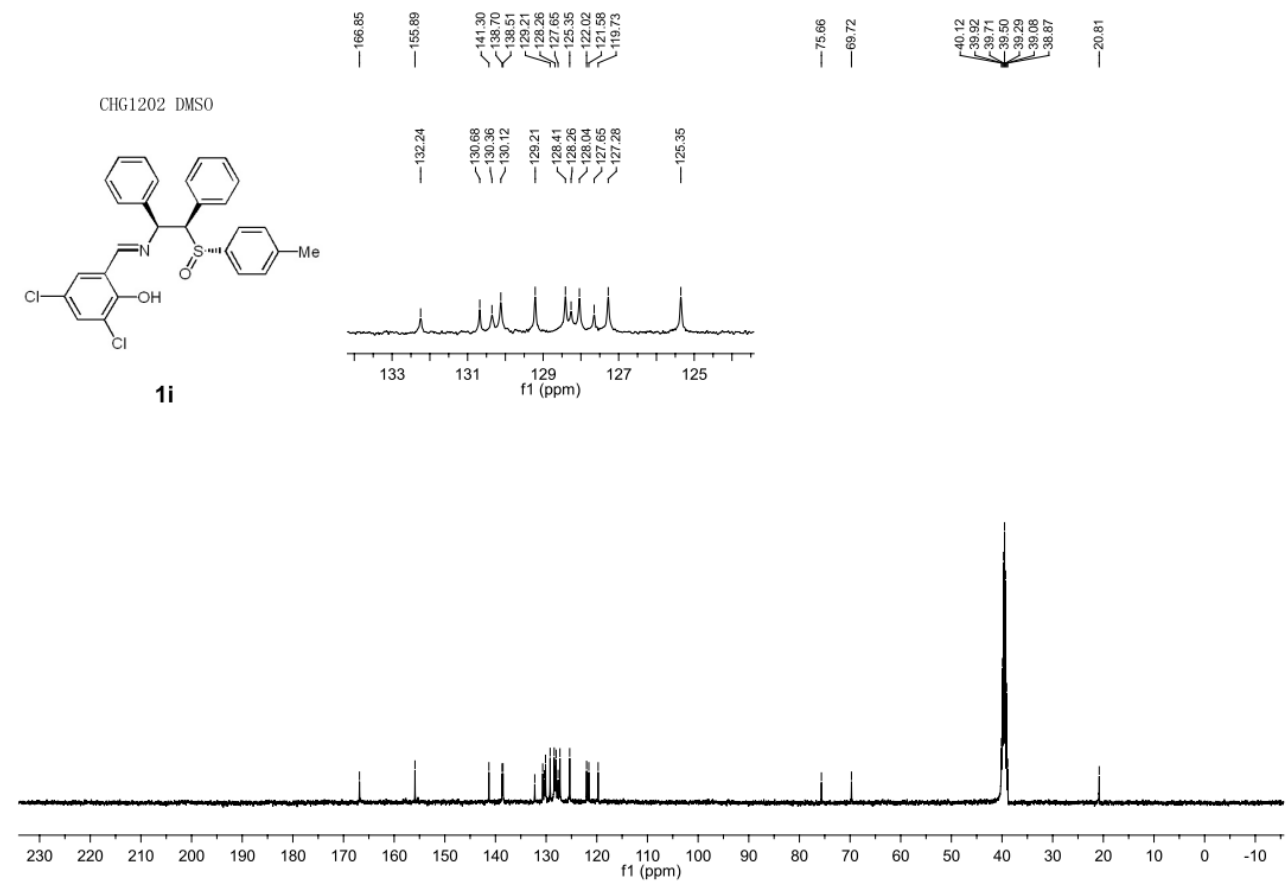
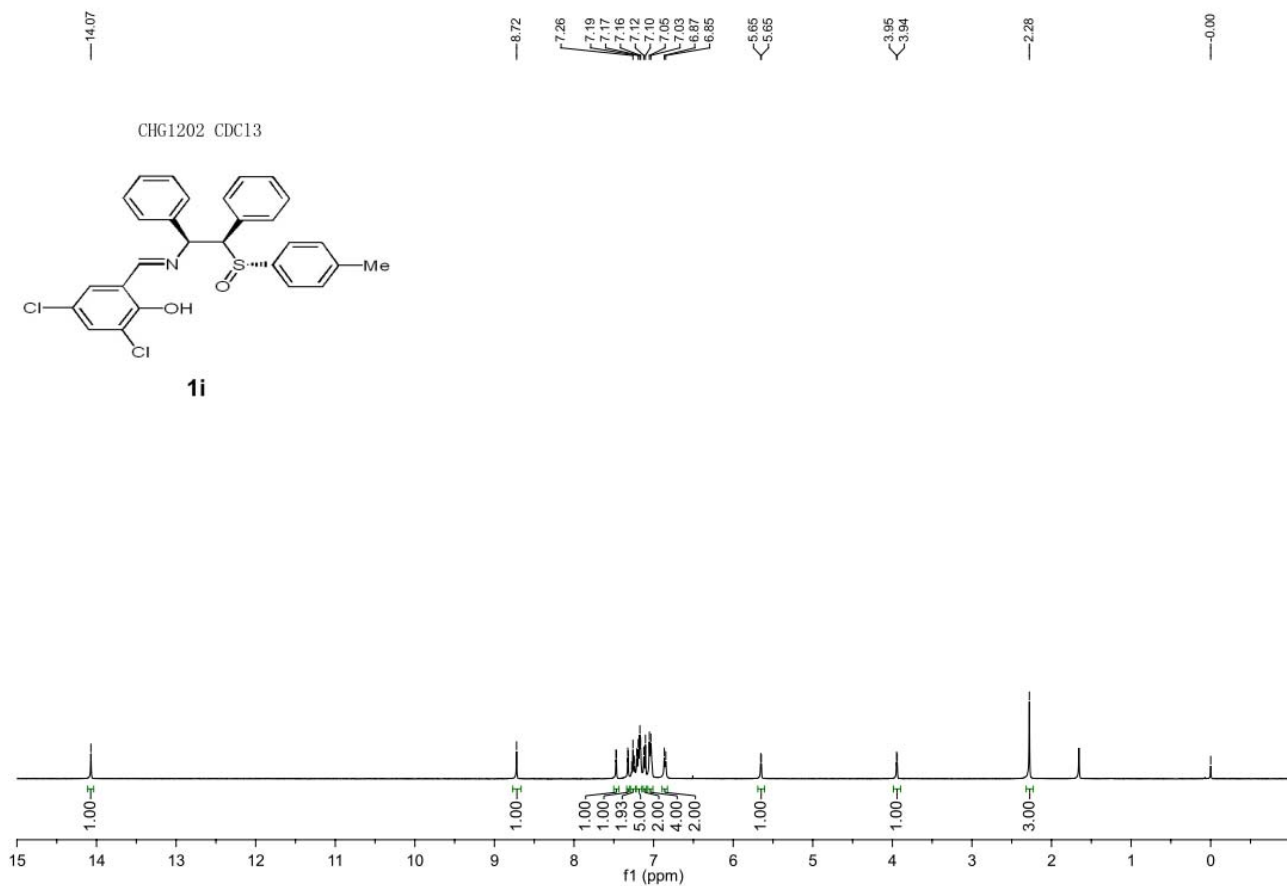


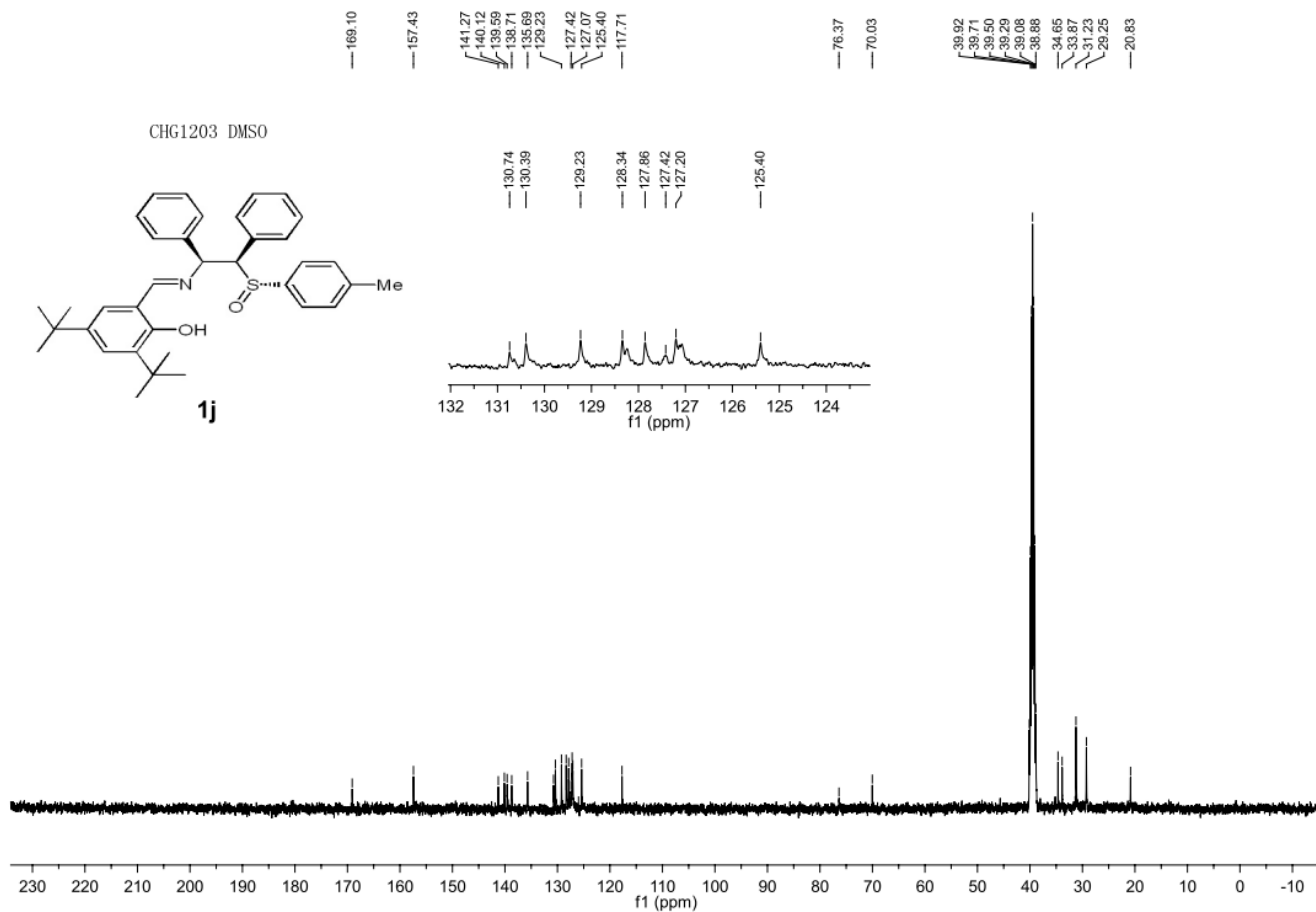
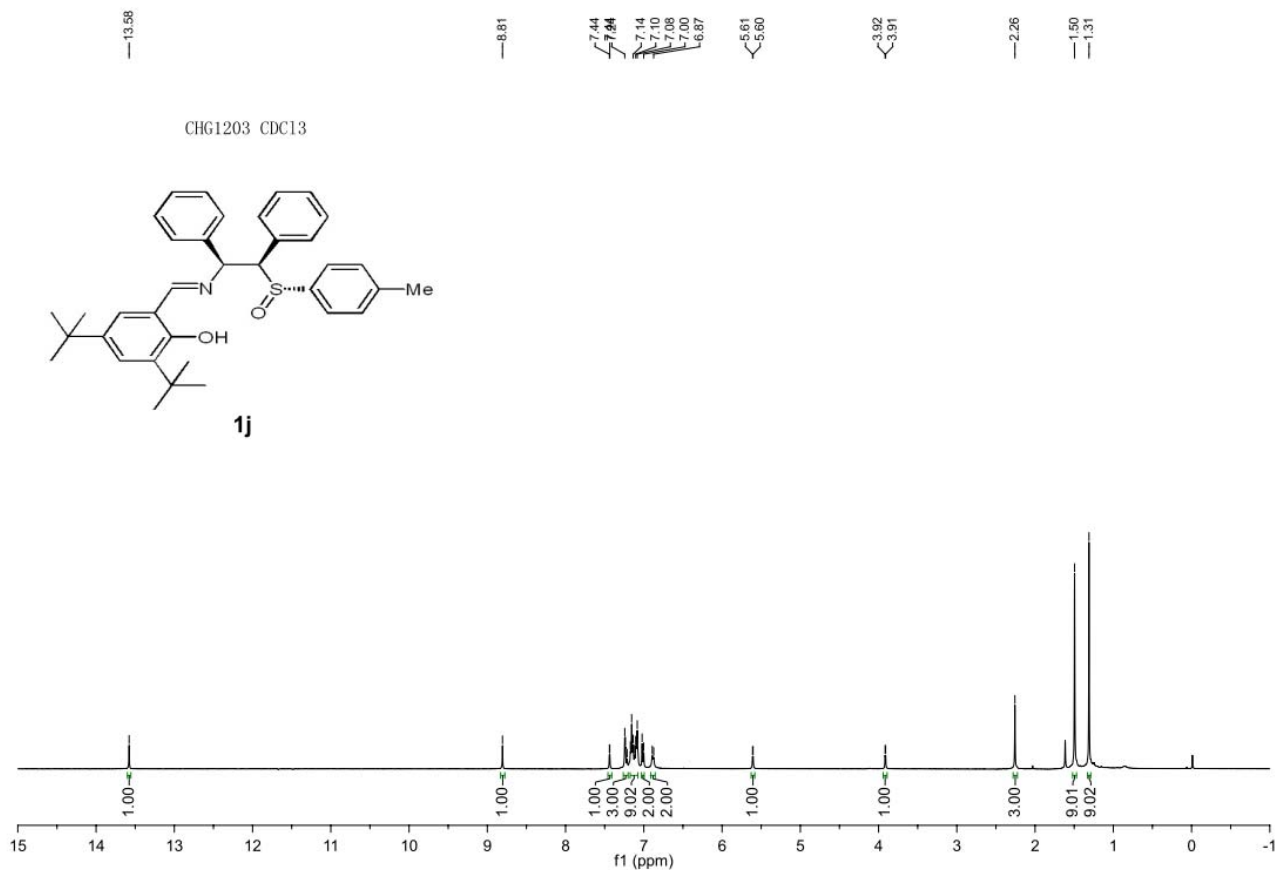


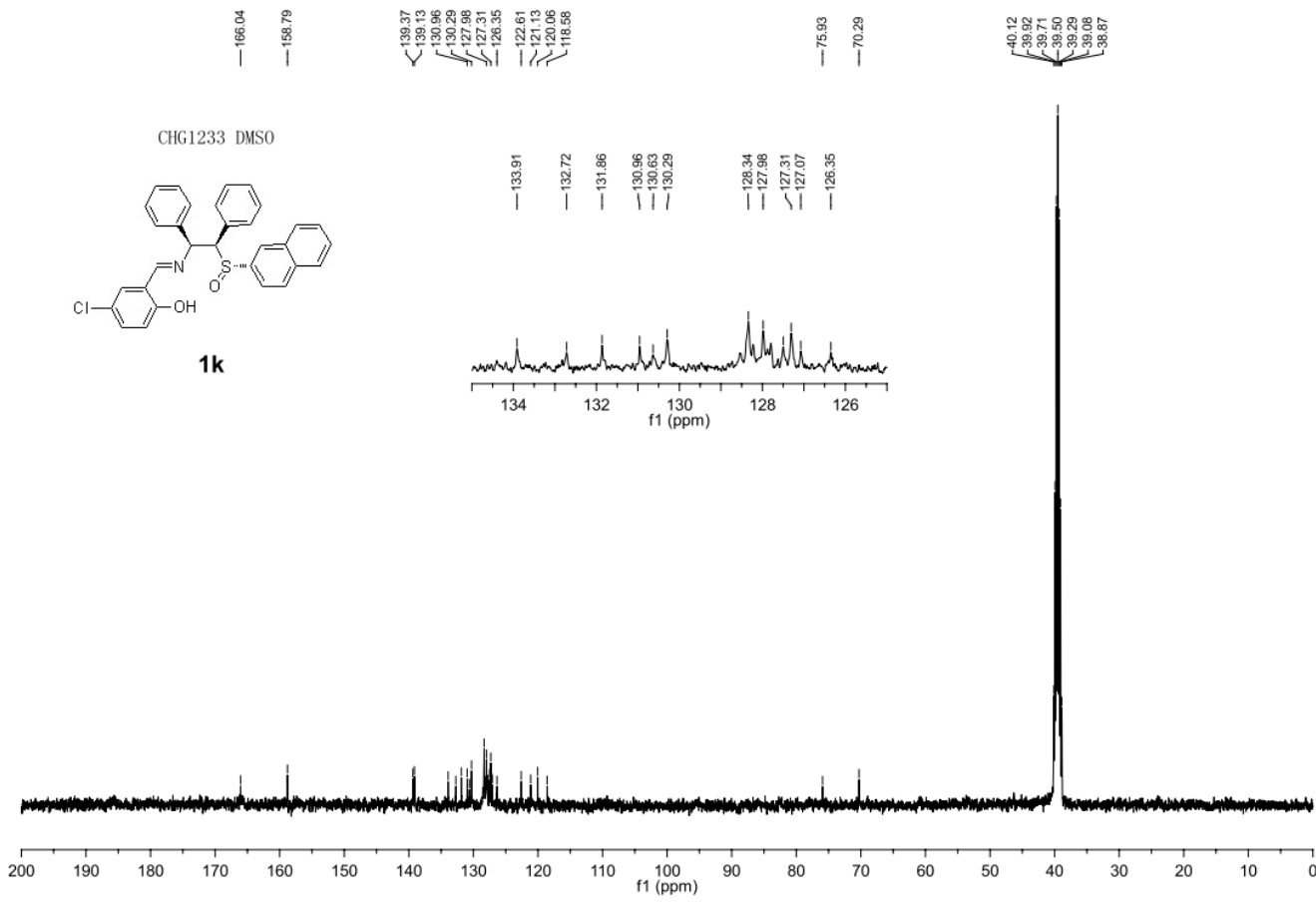
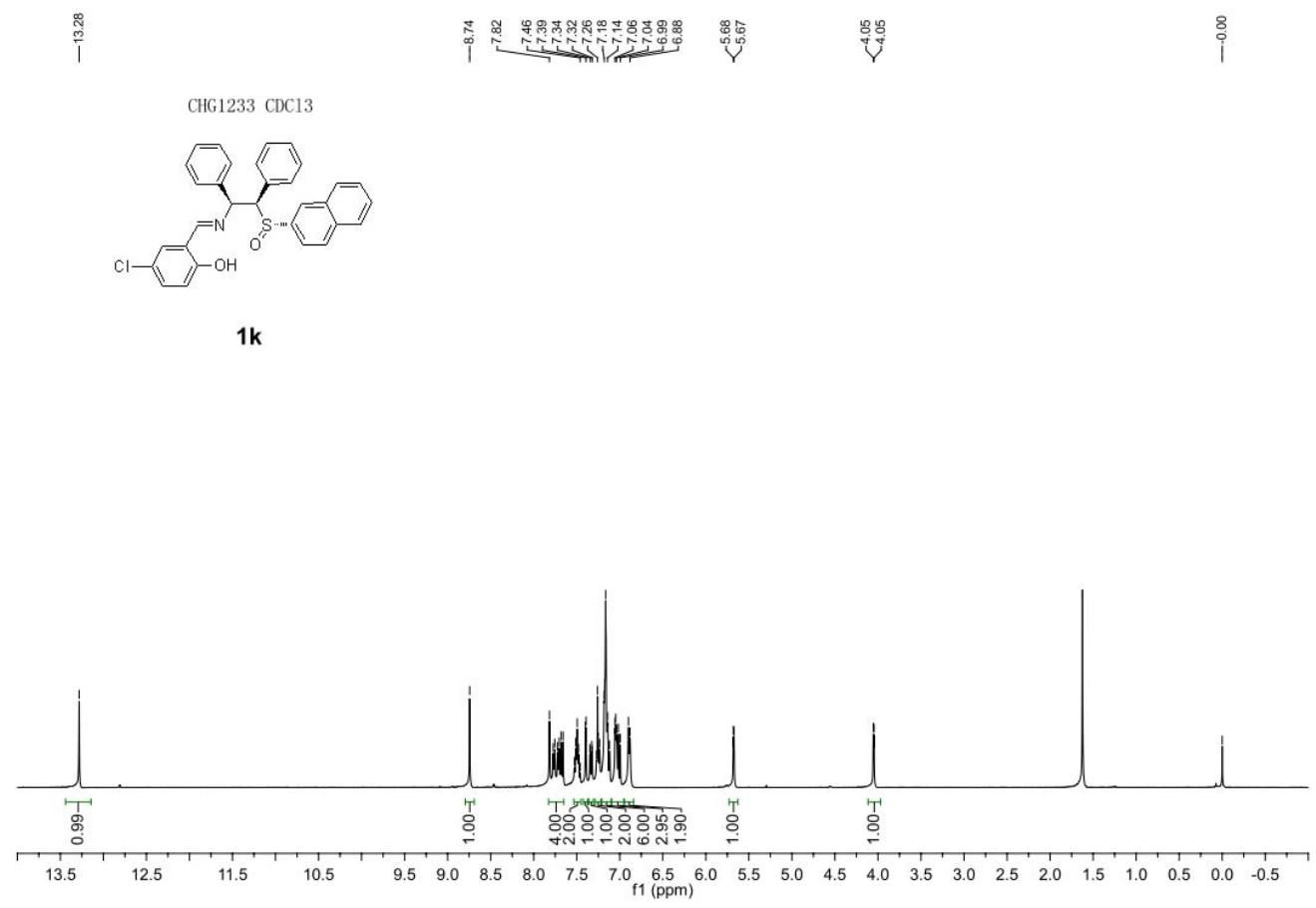


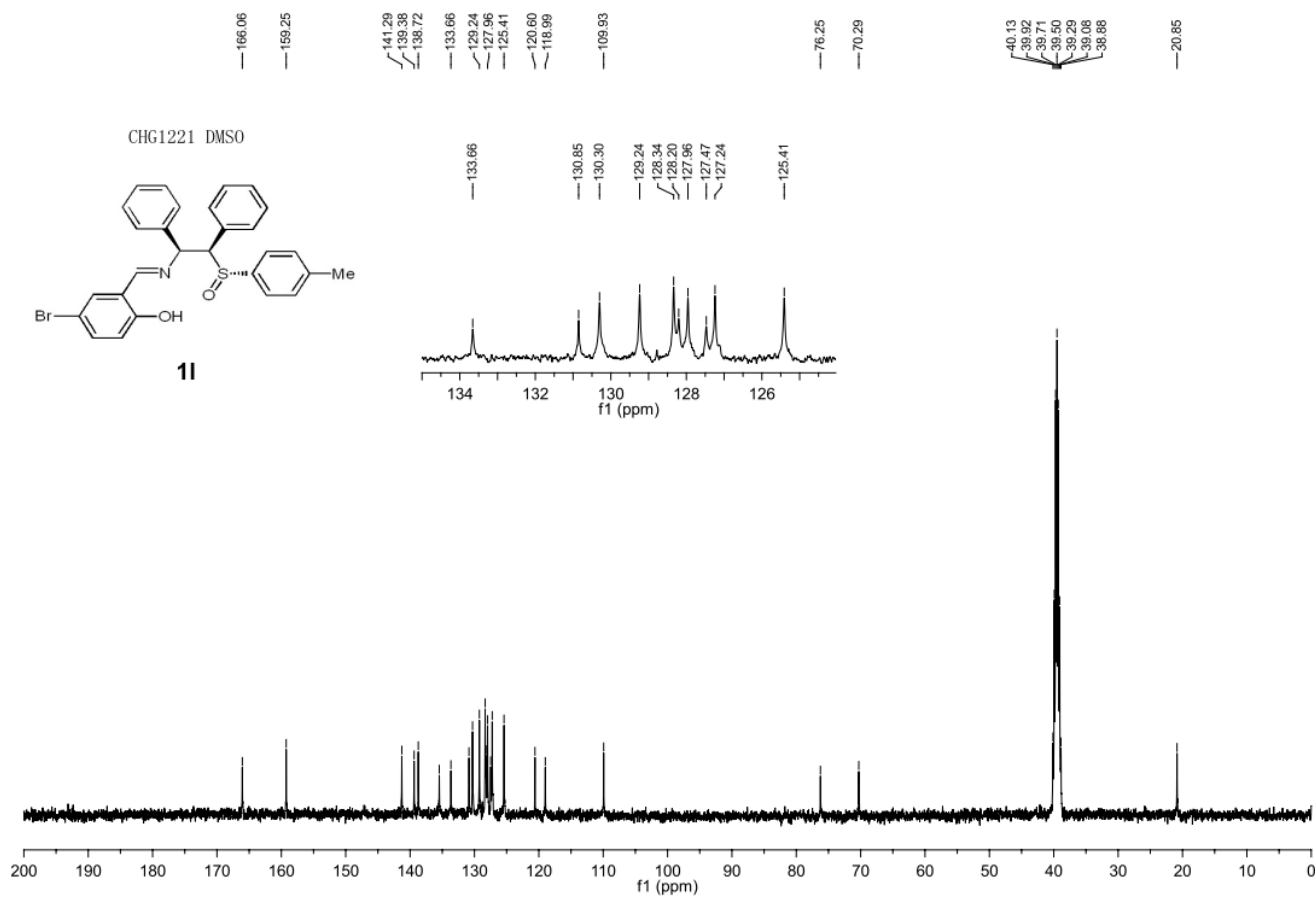
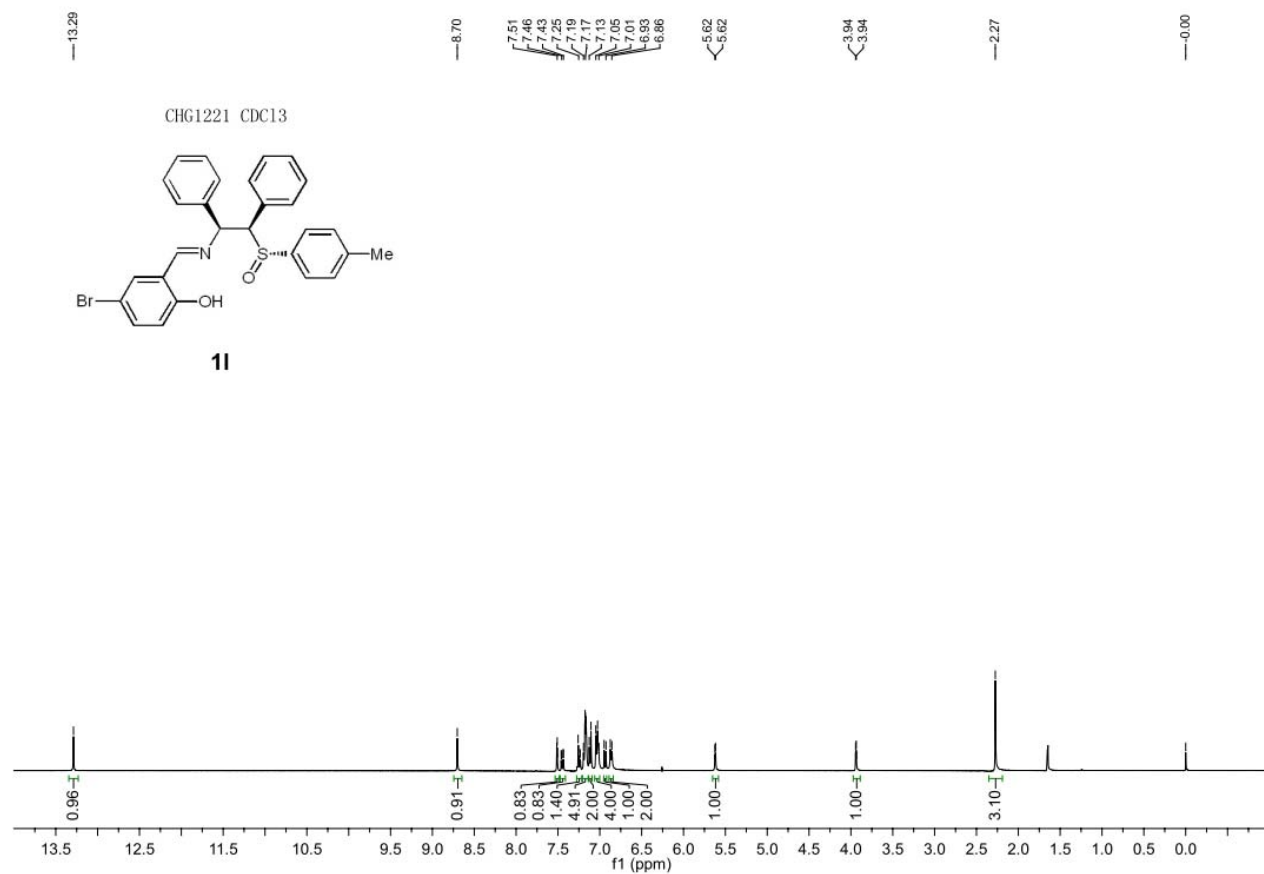


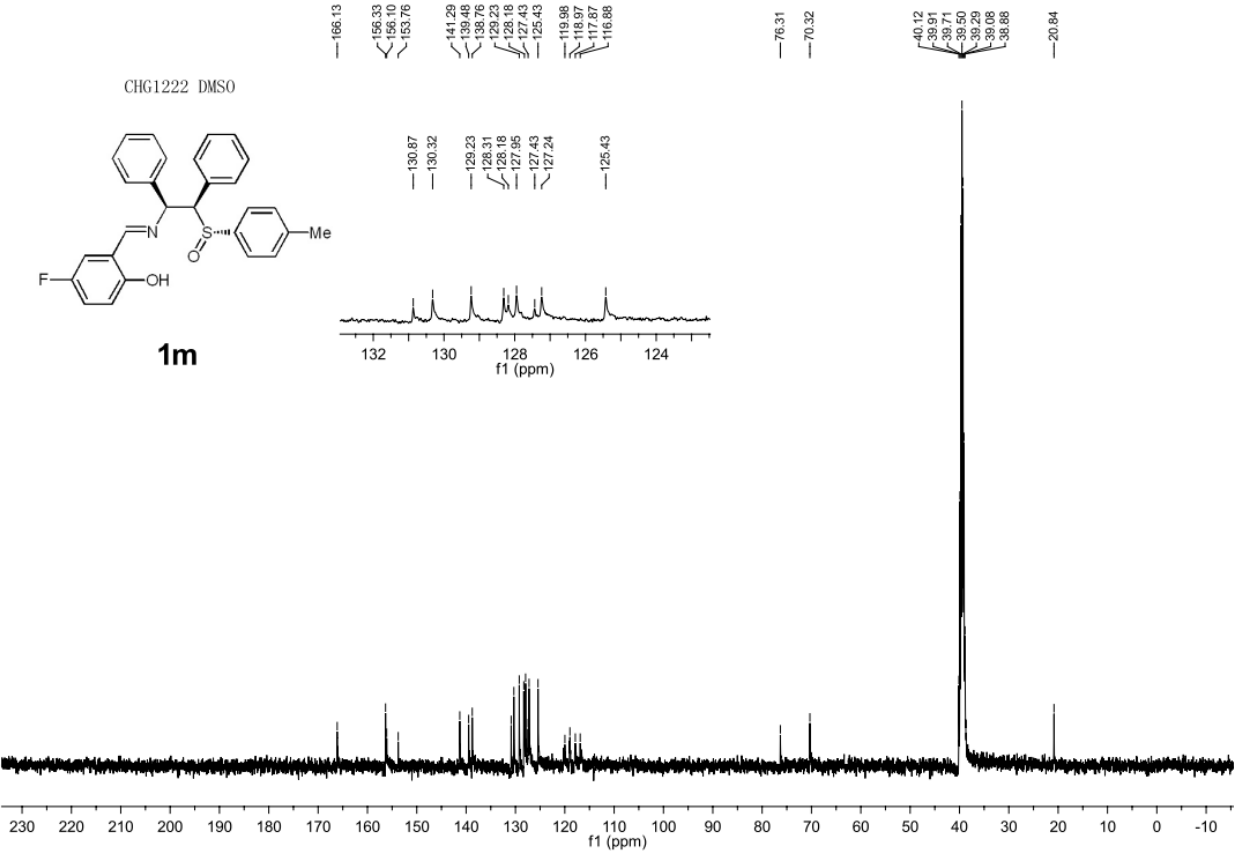
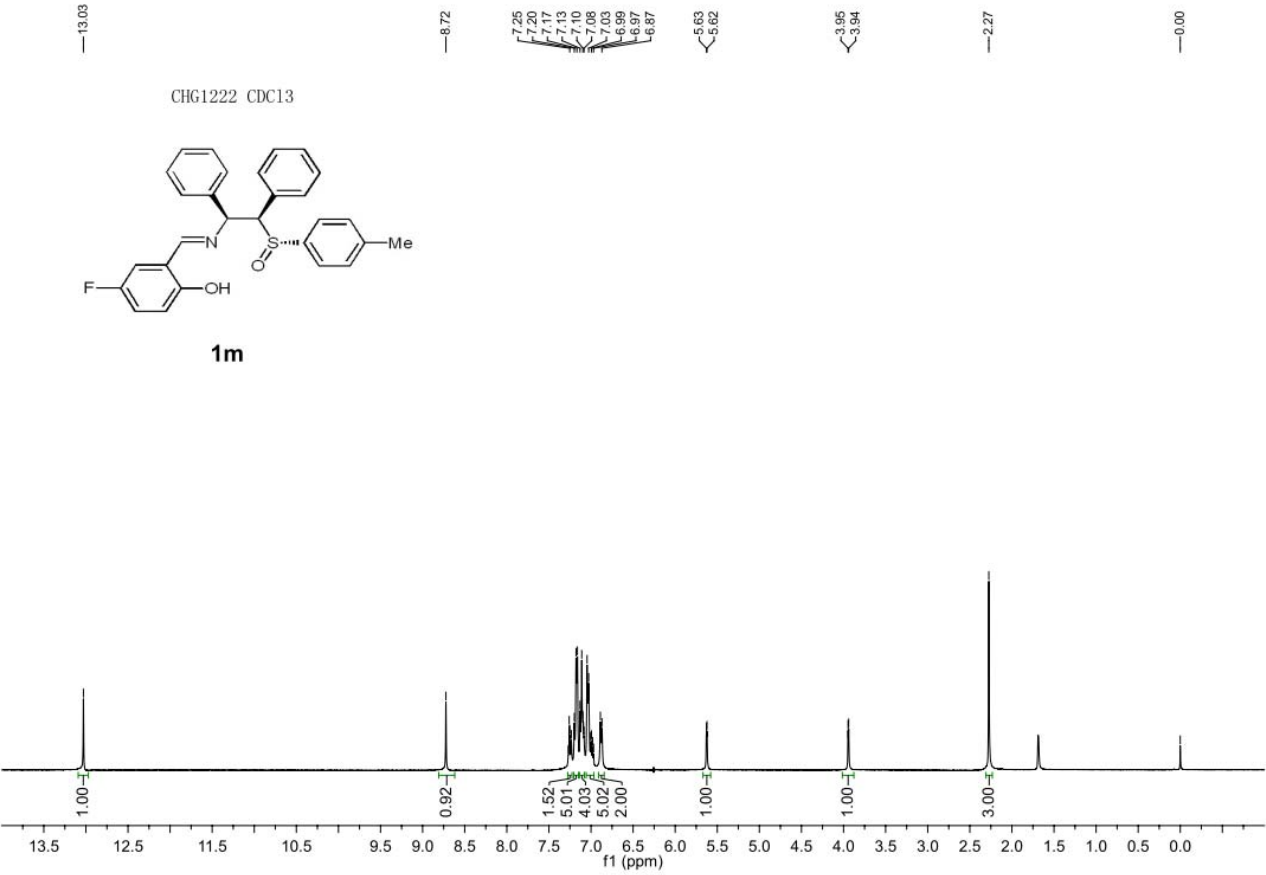


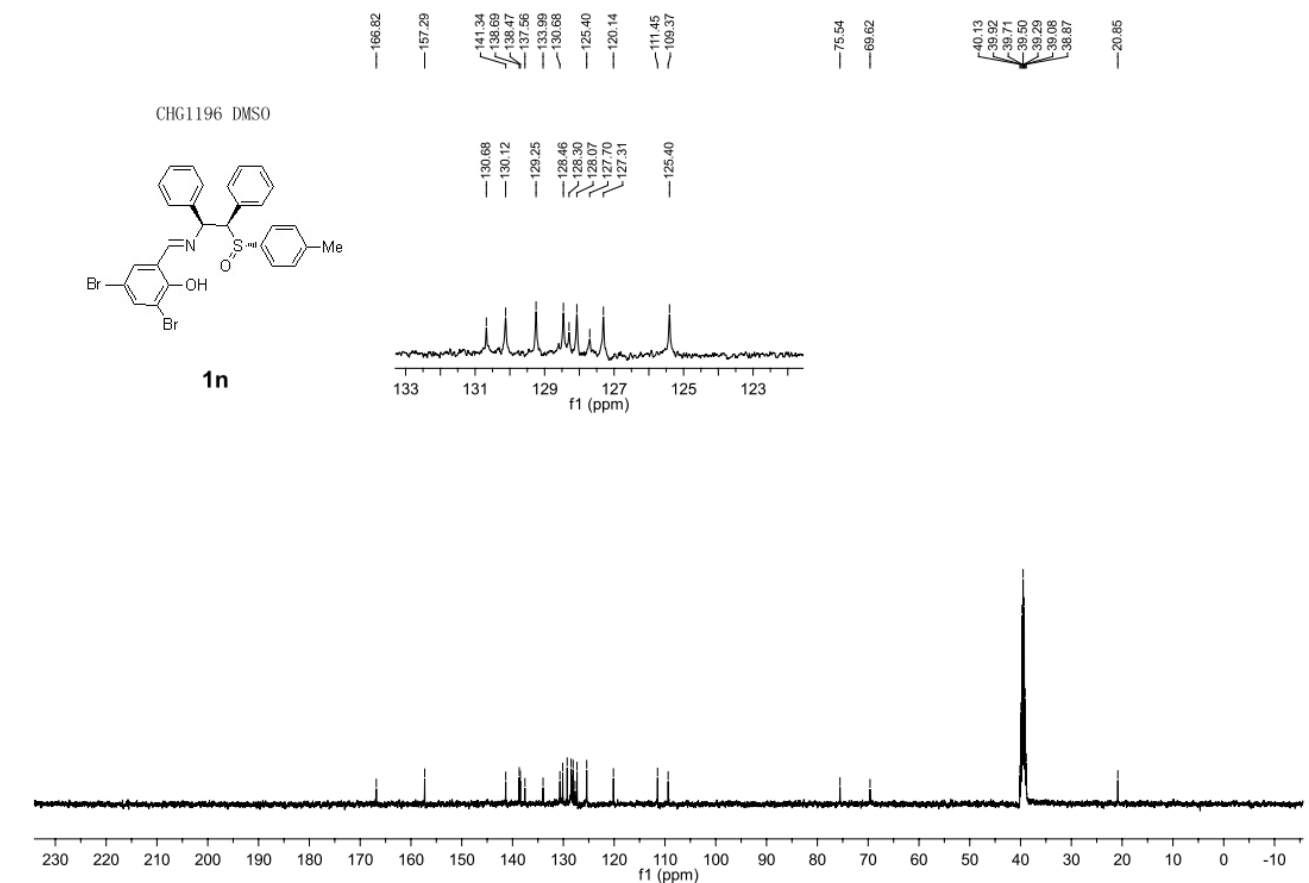
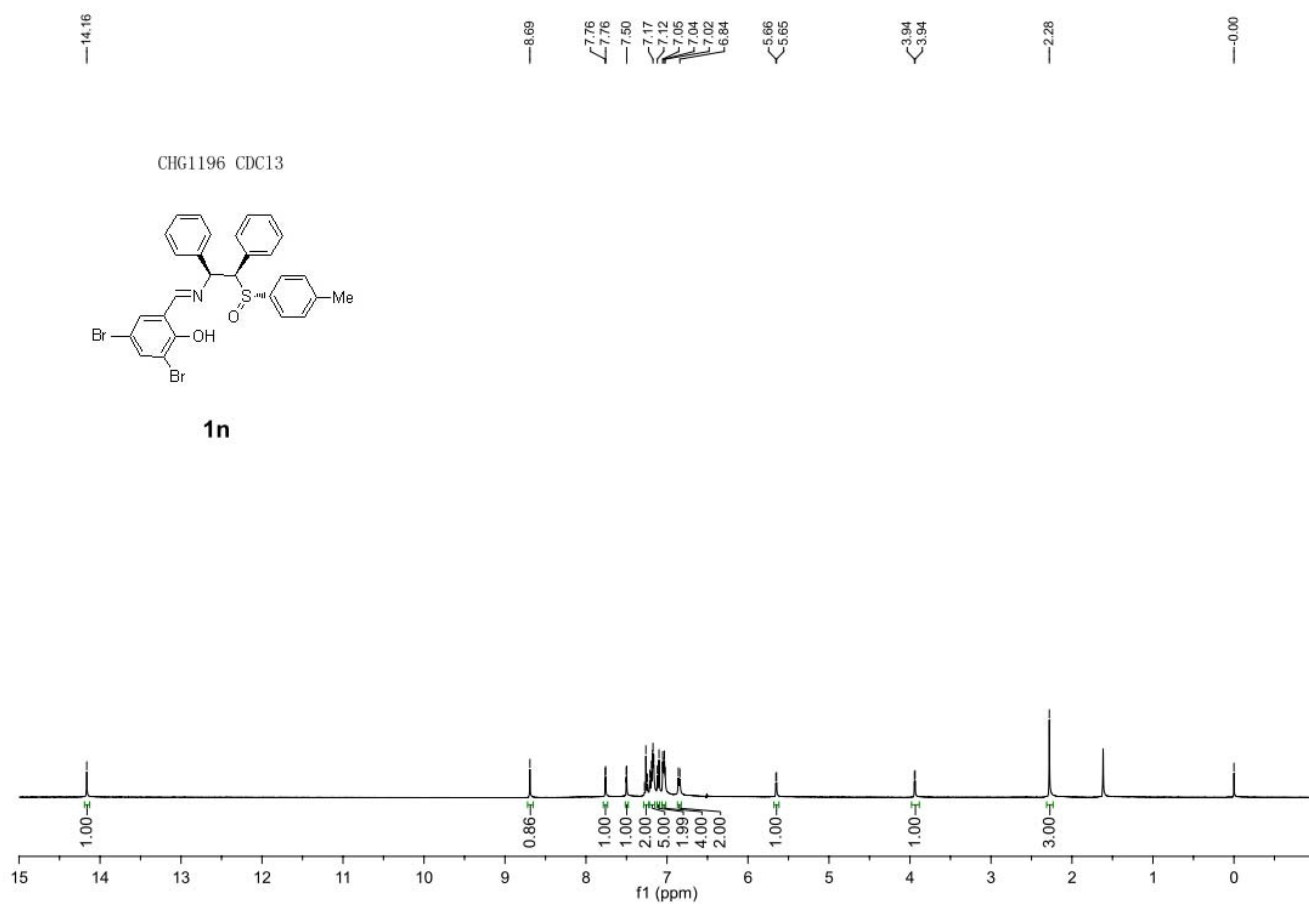


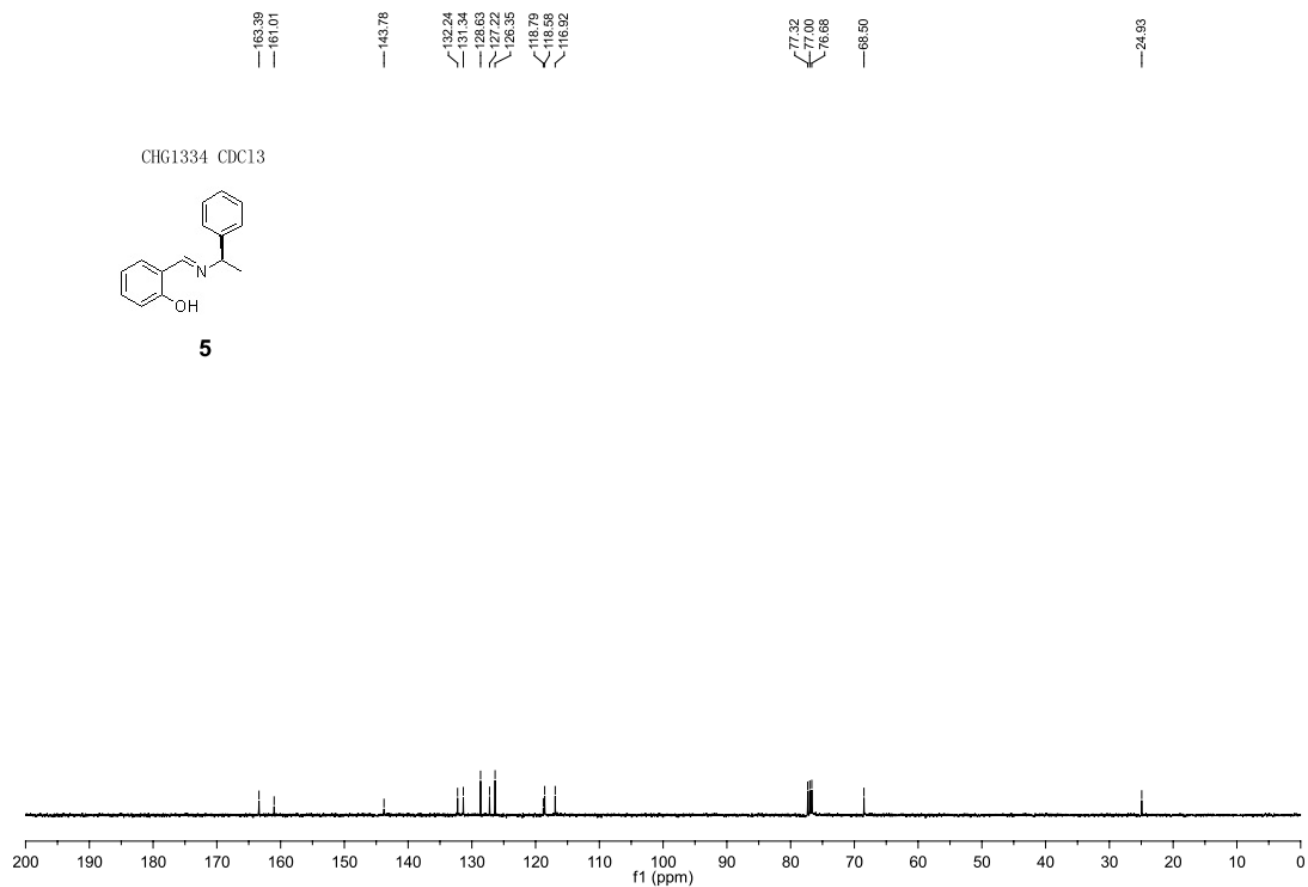
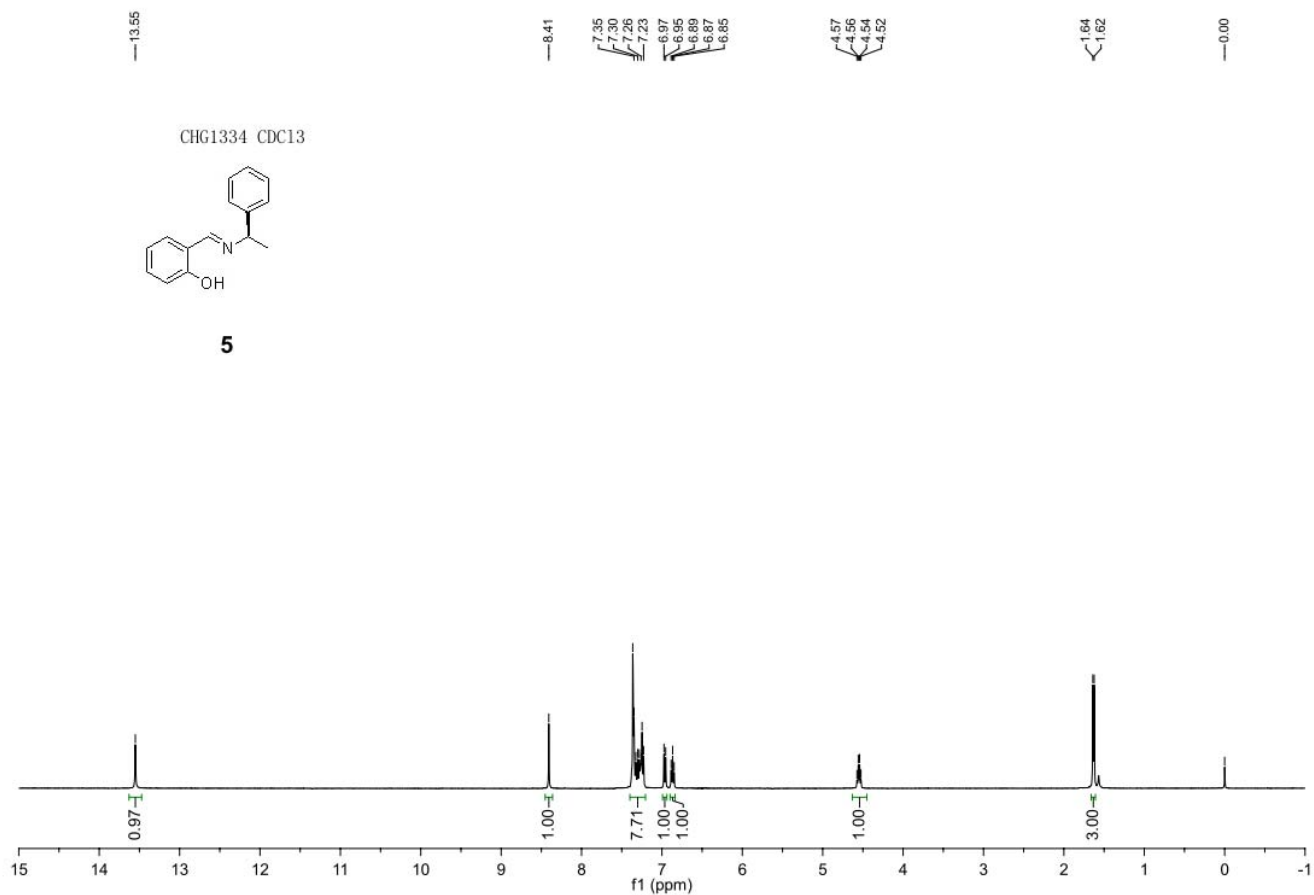


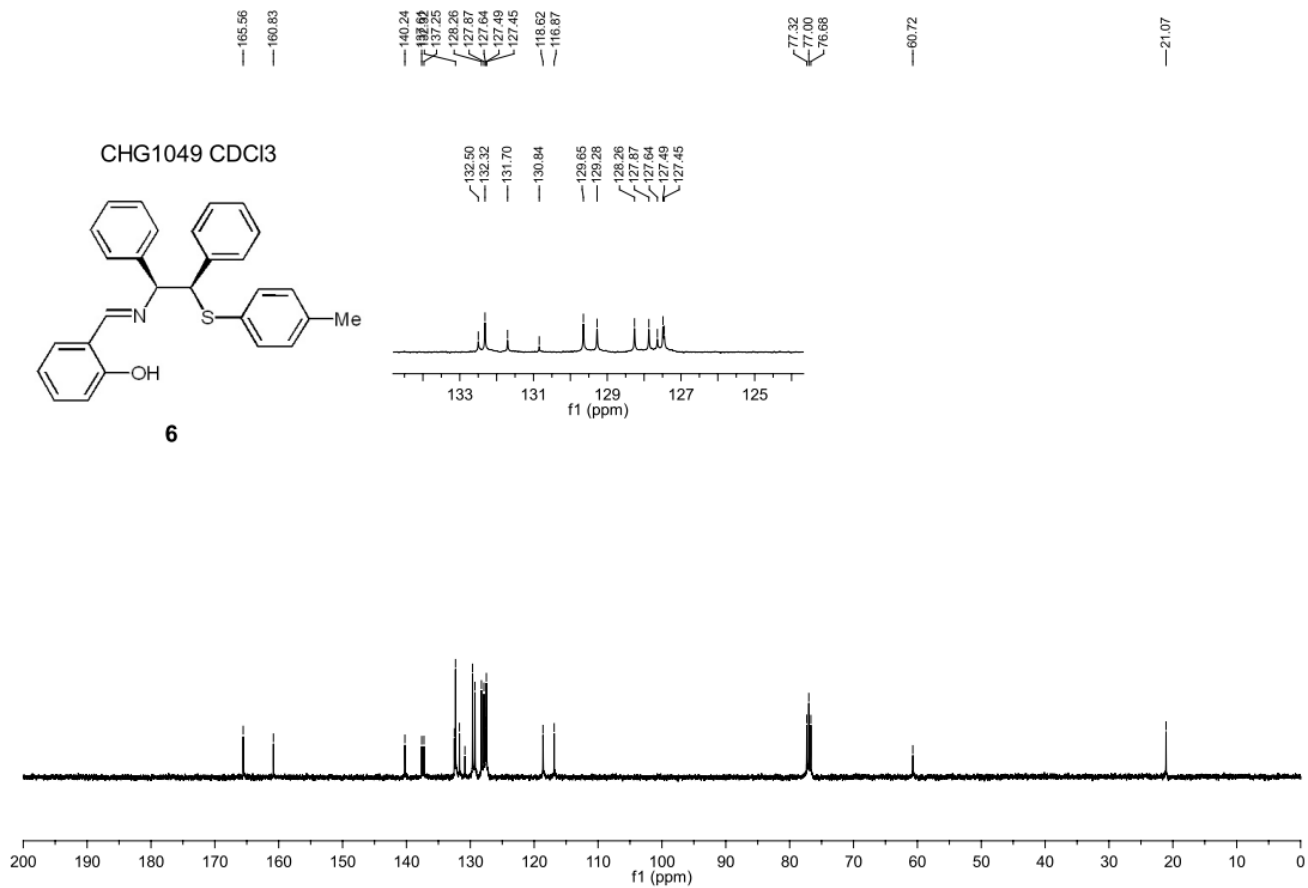
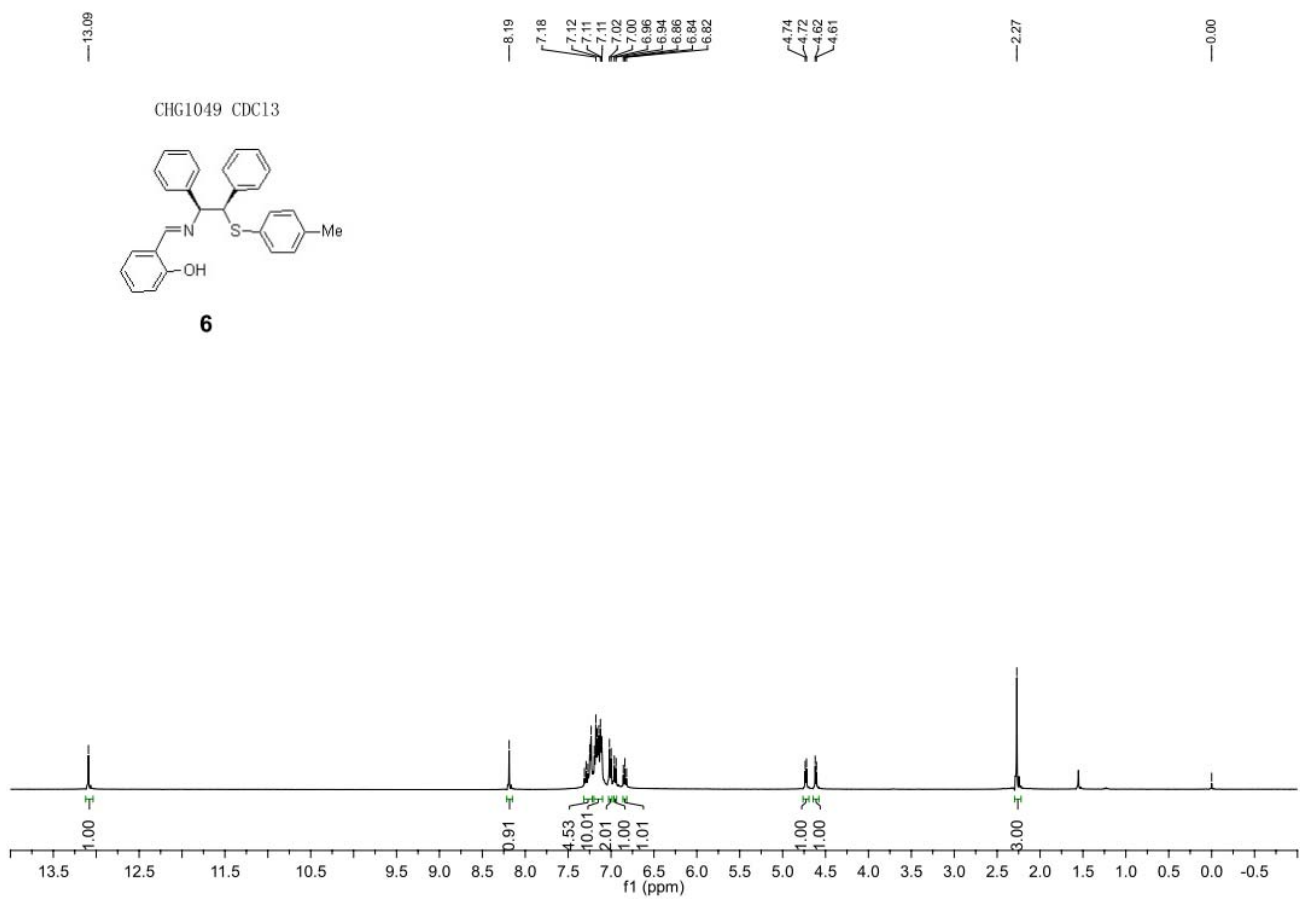


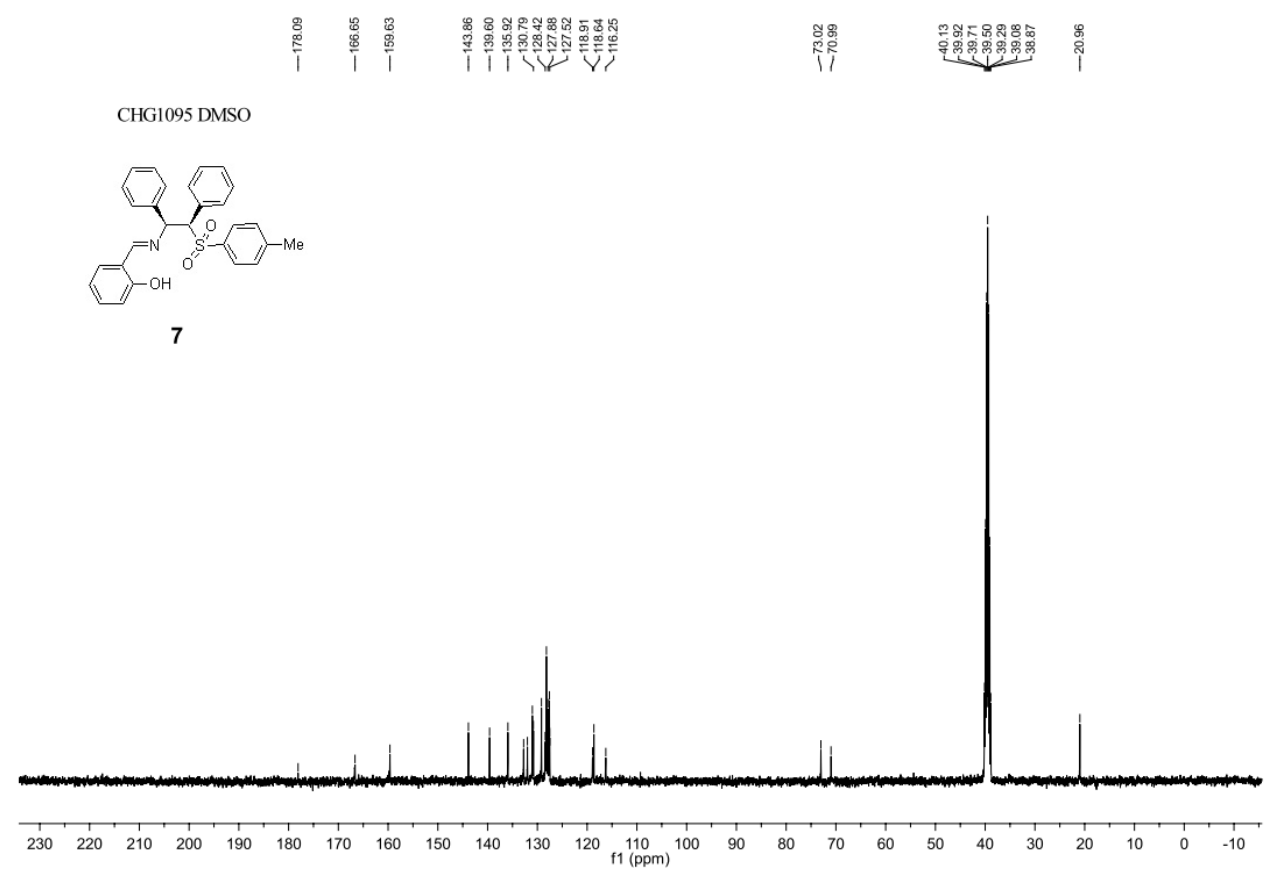
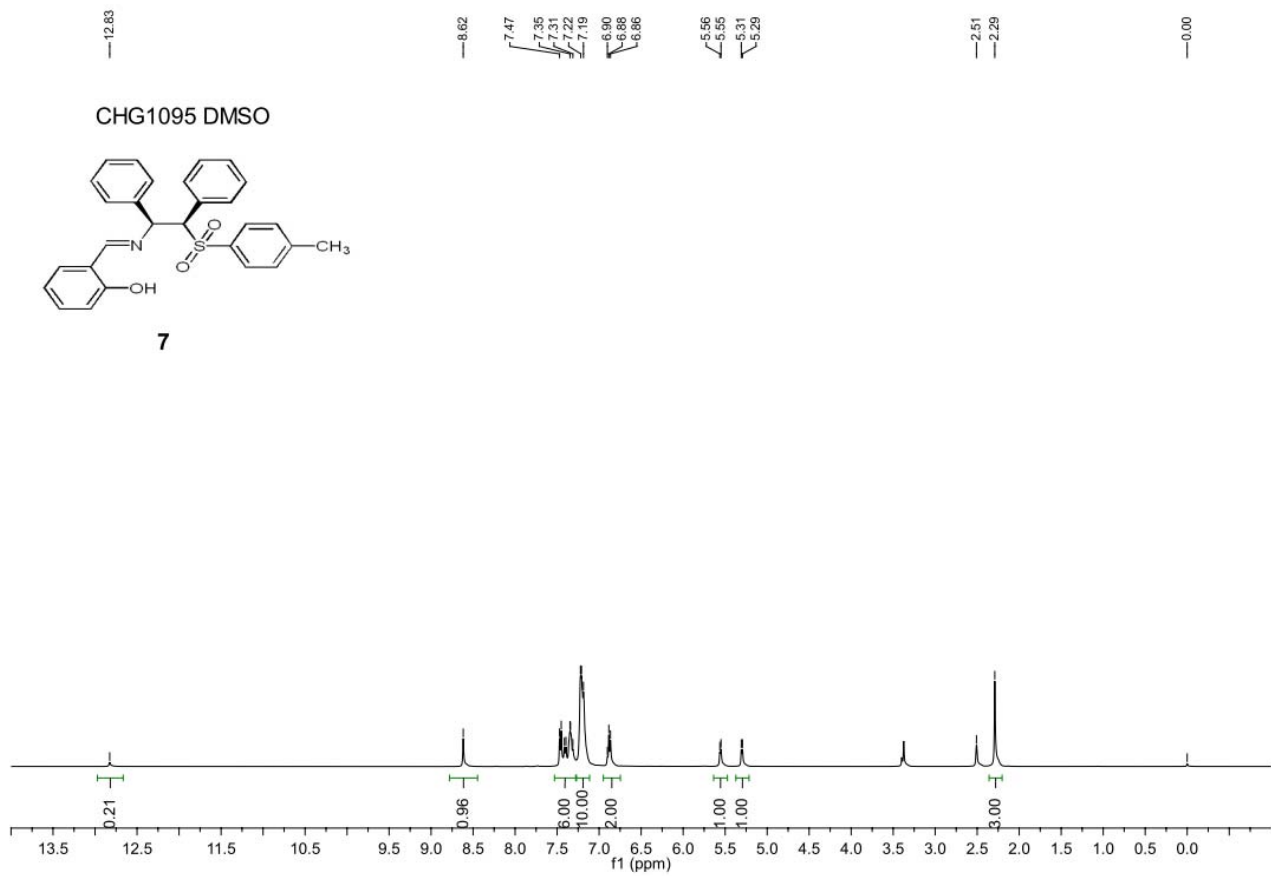


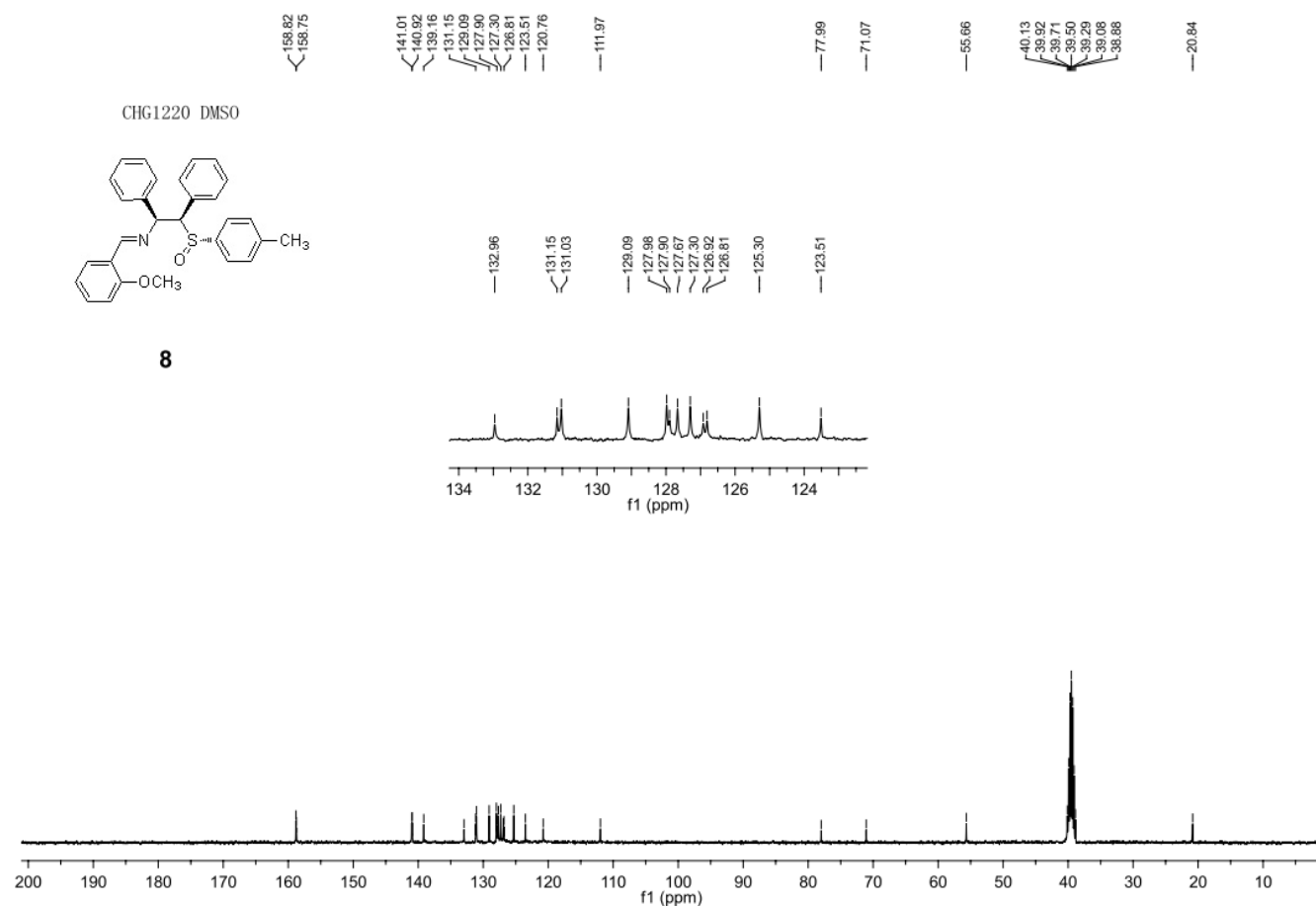
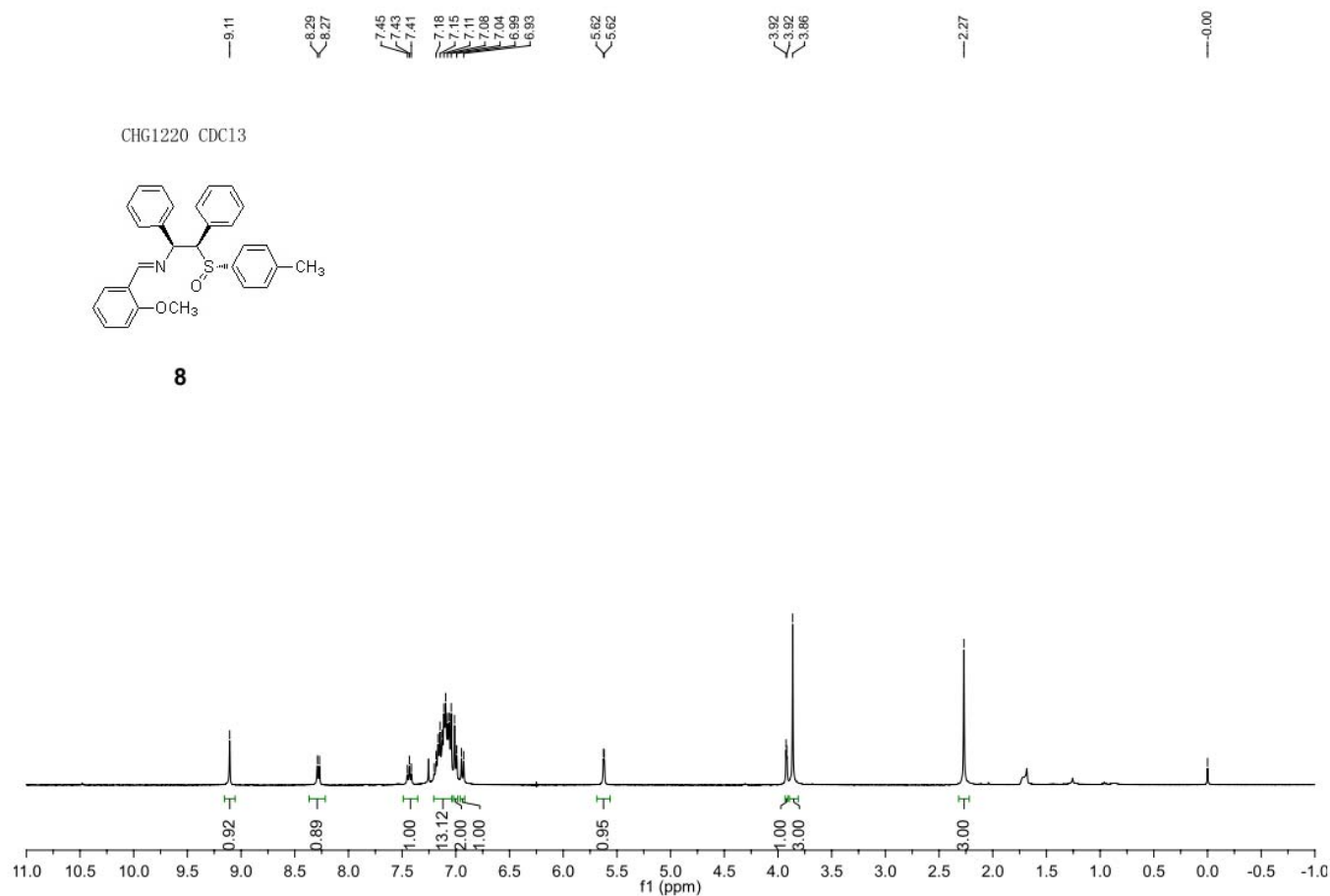


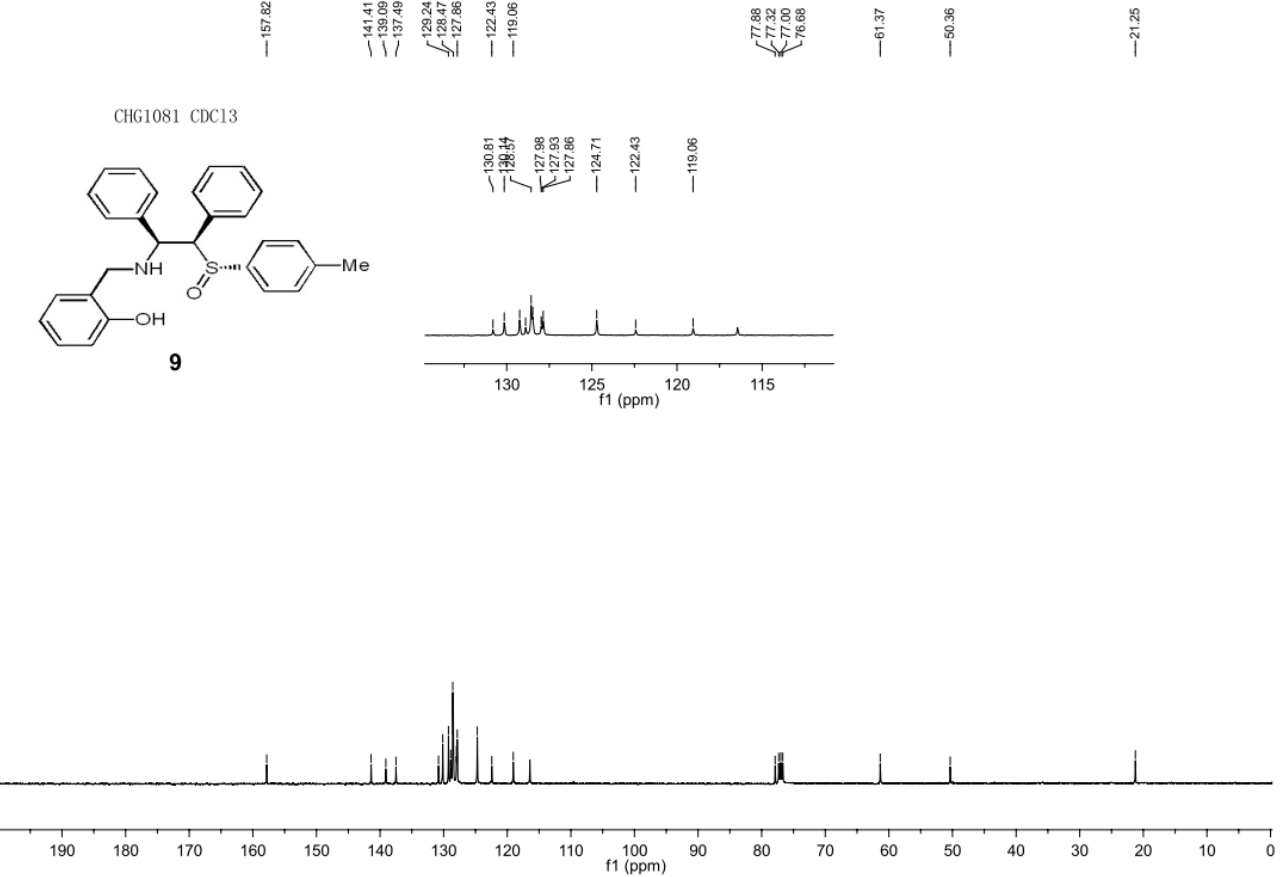
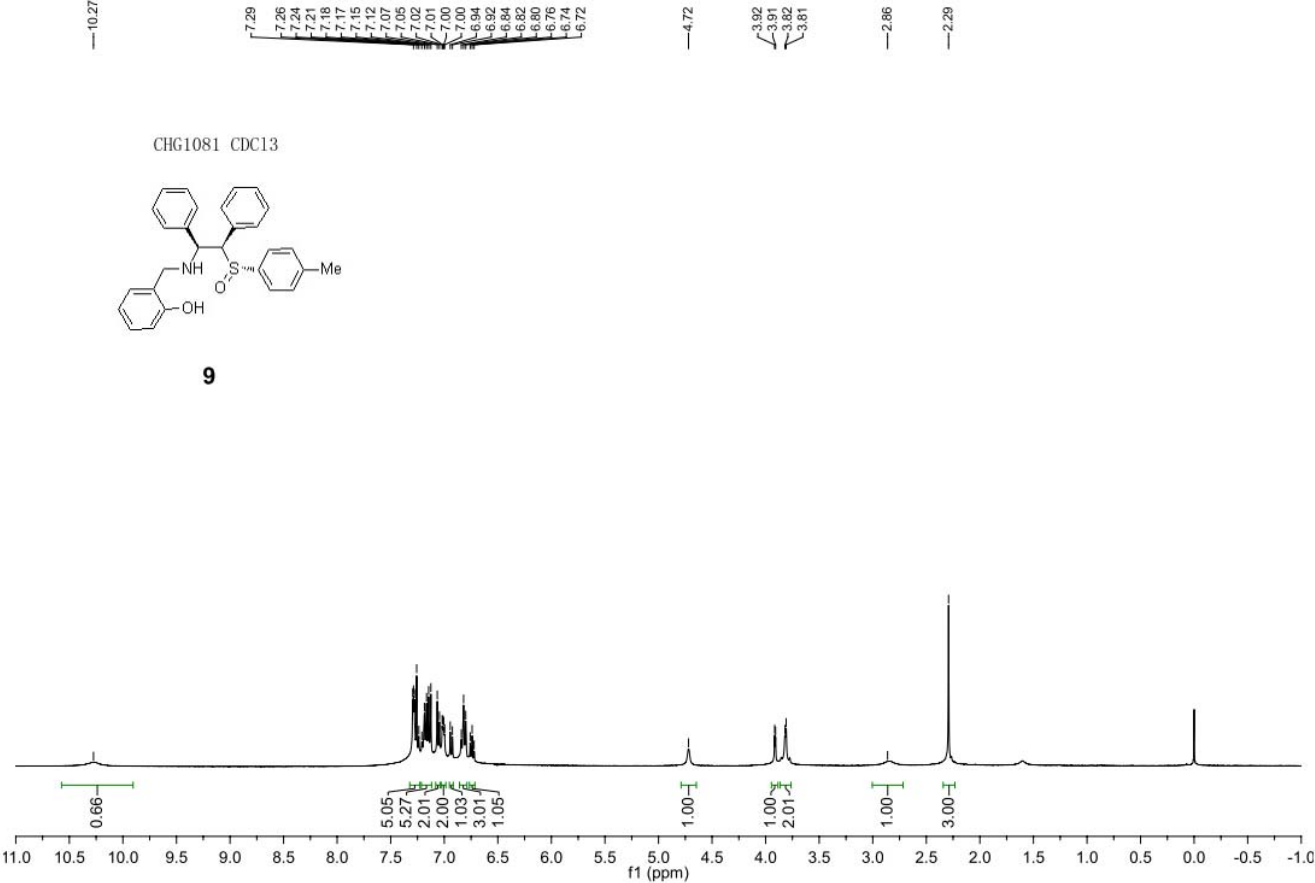


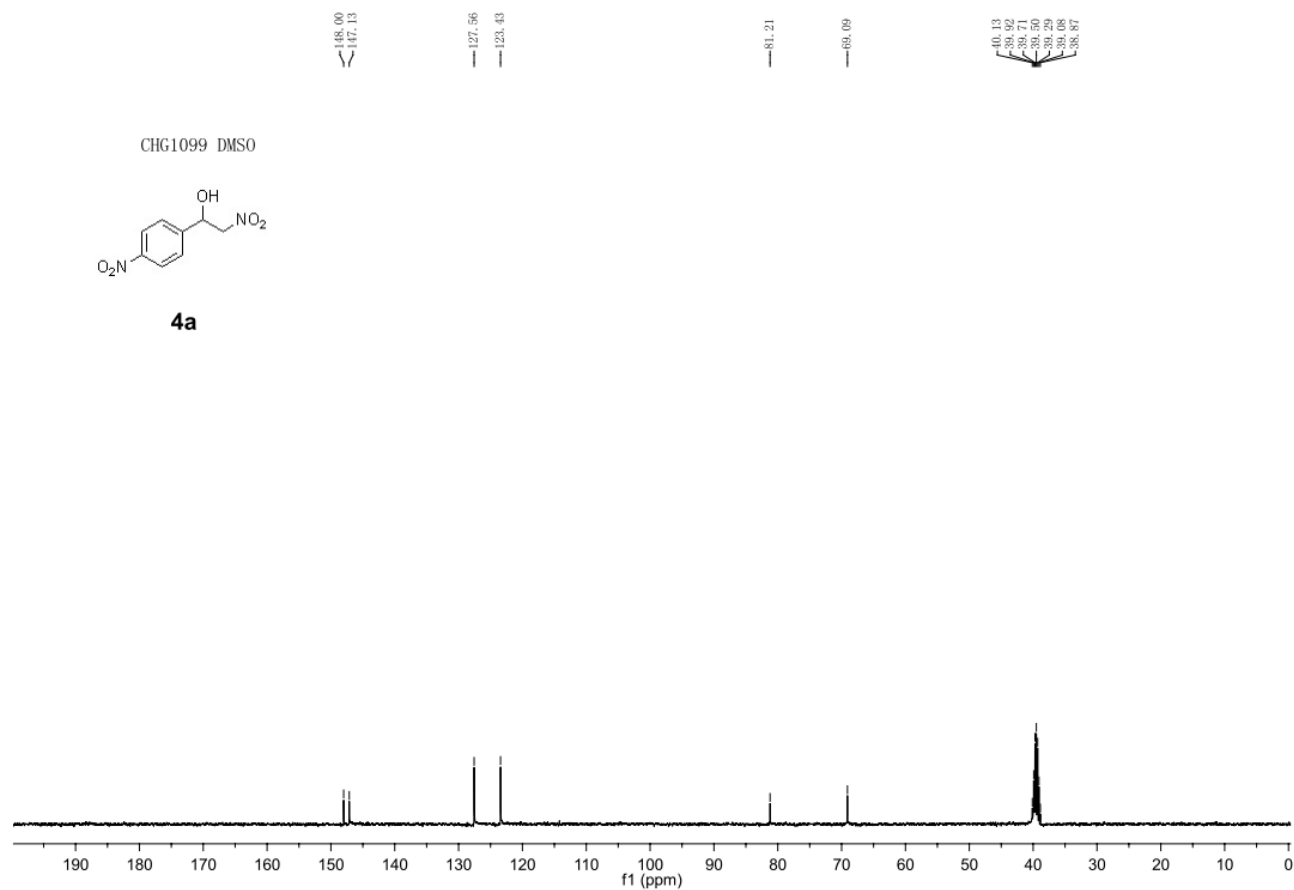
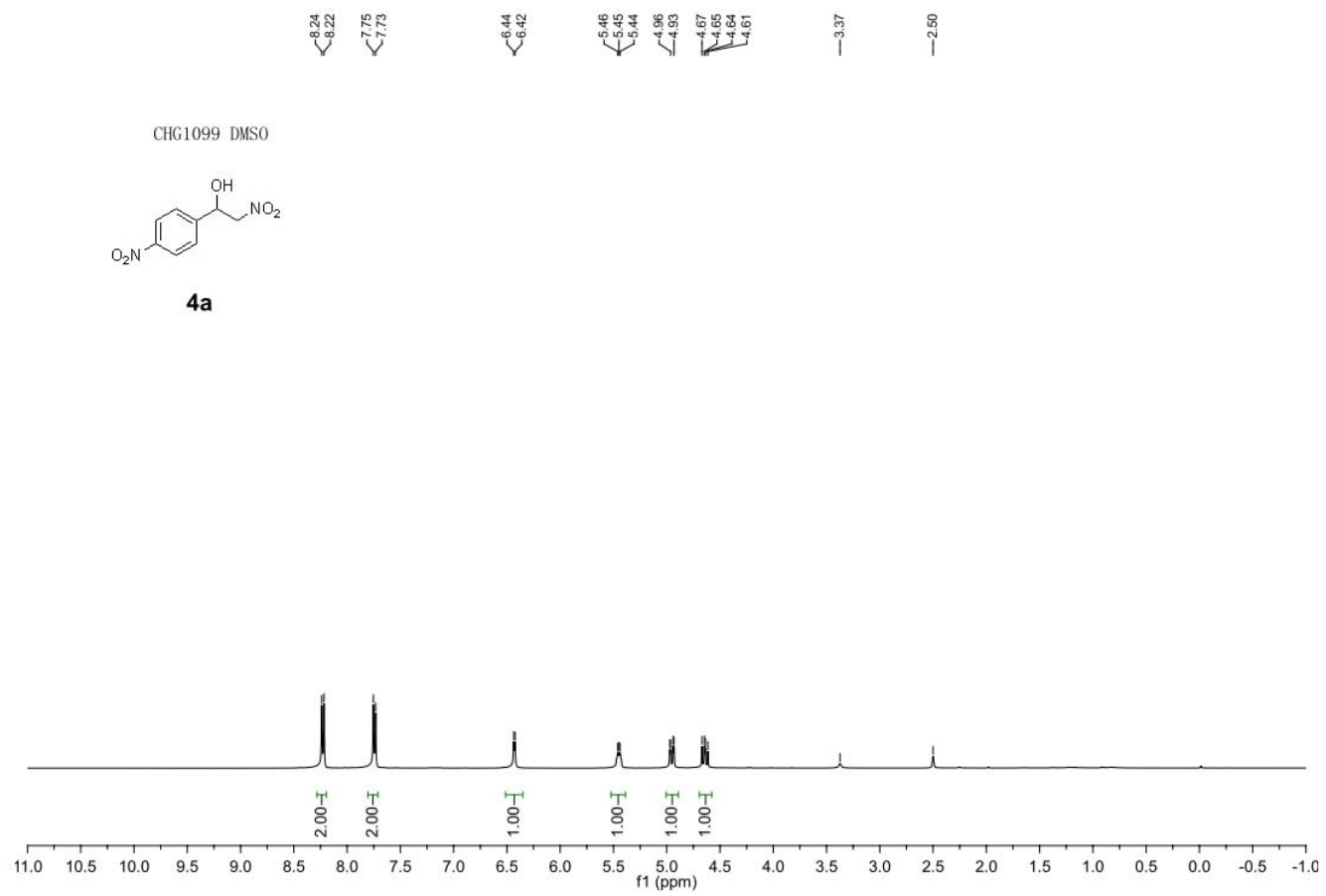


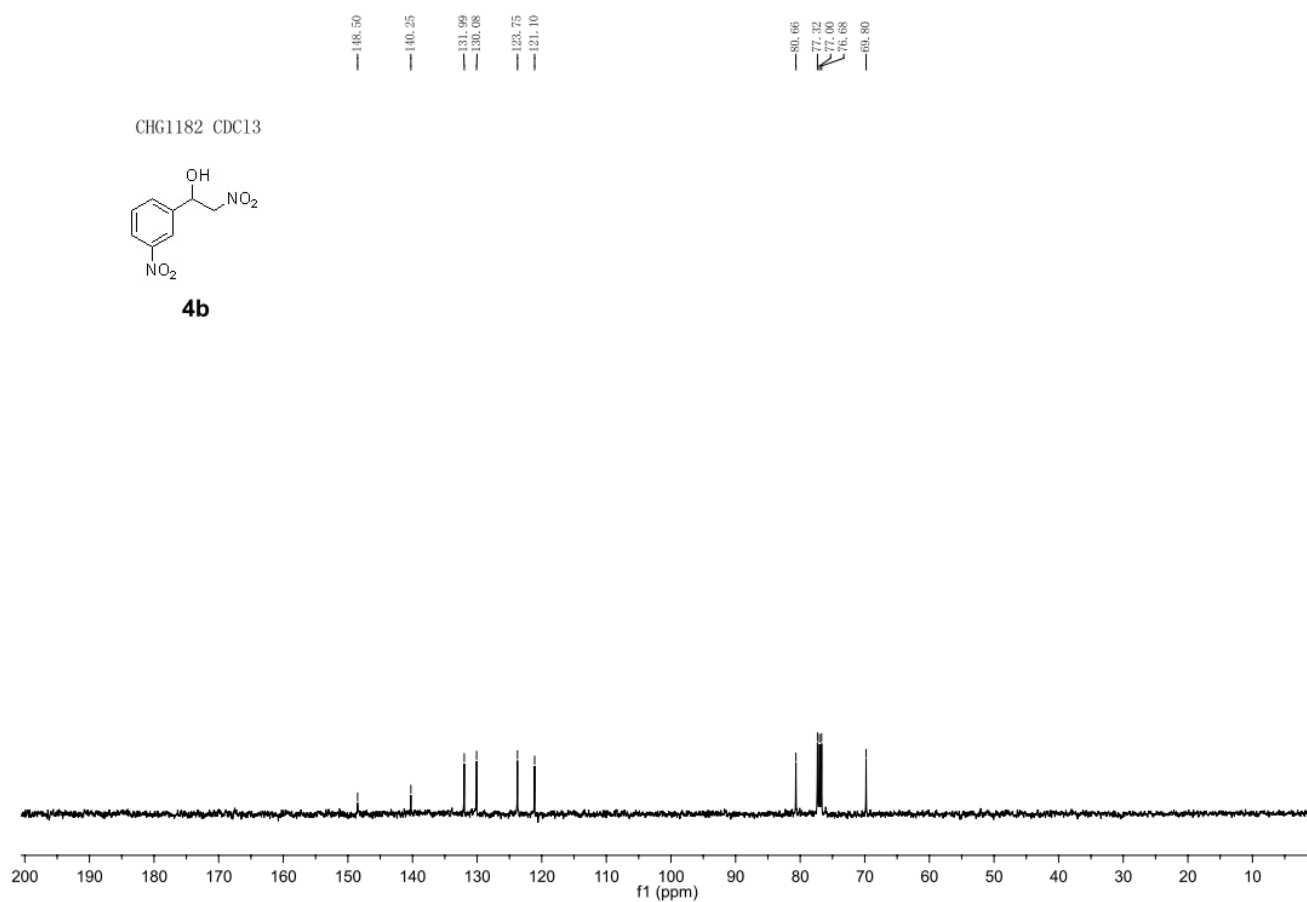
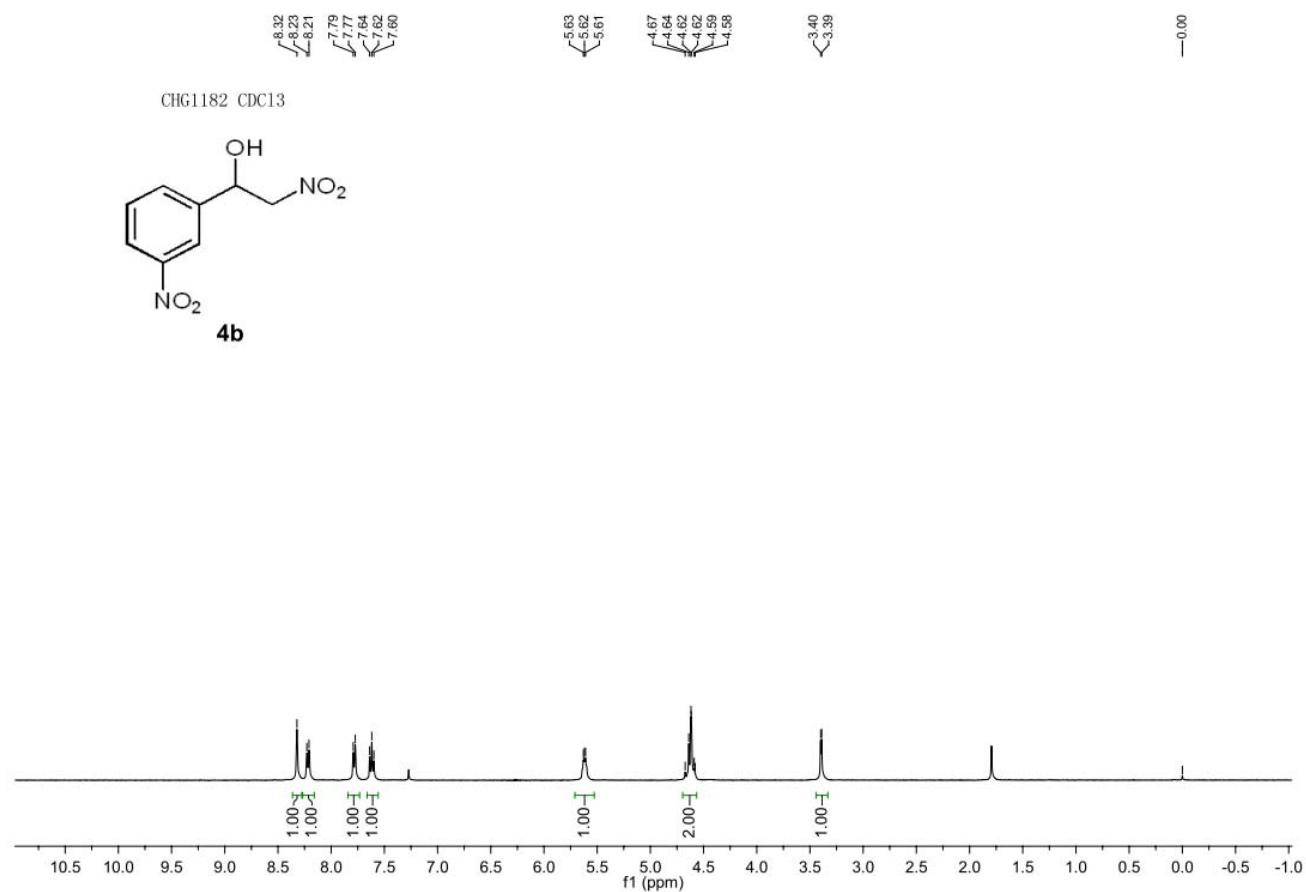




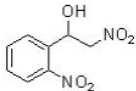




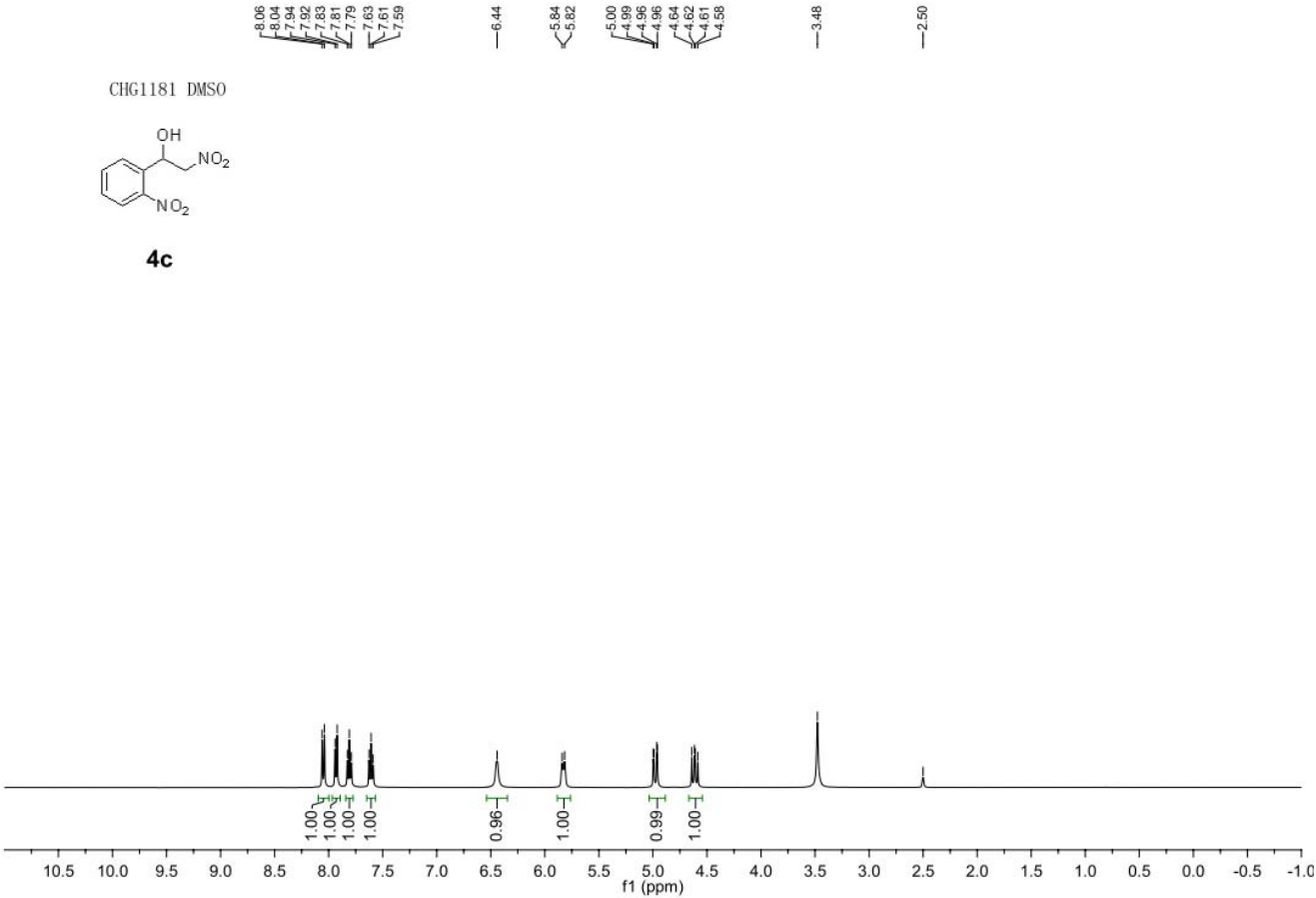




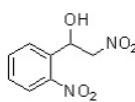
CHG1181 DMSO



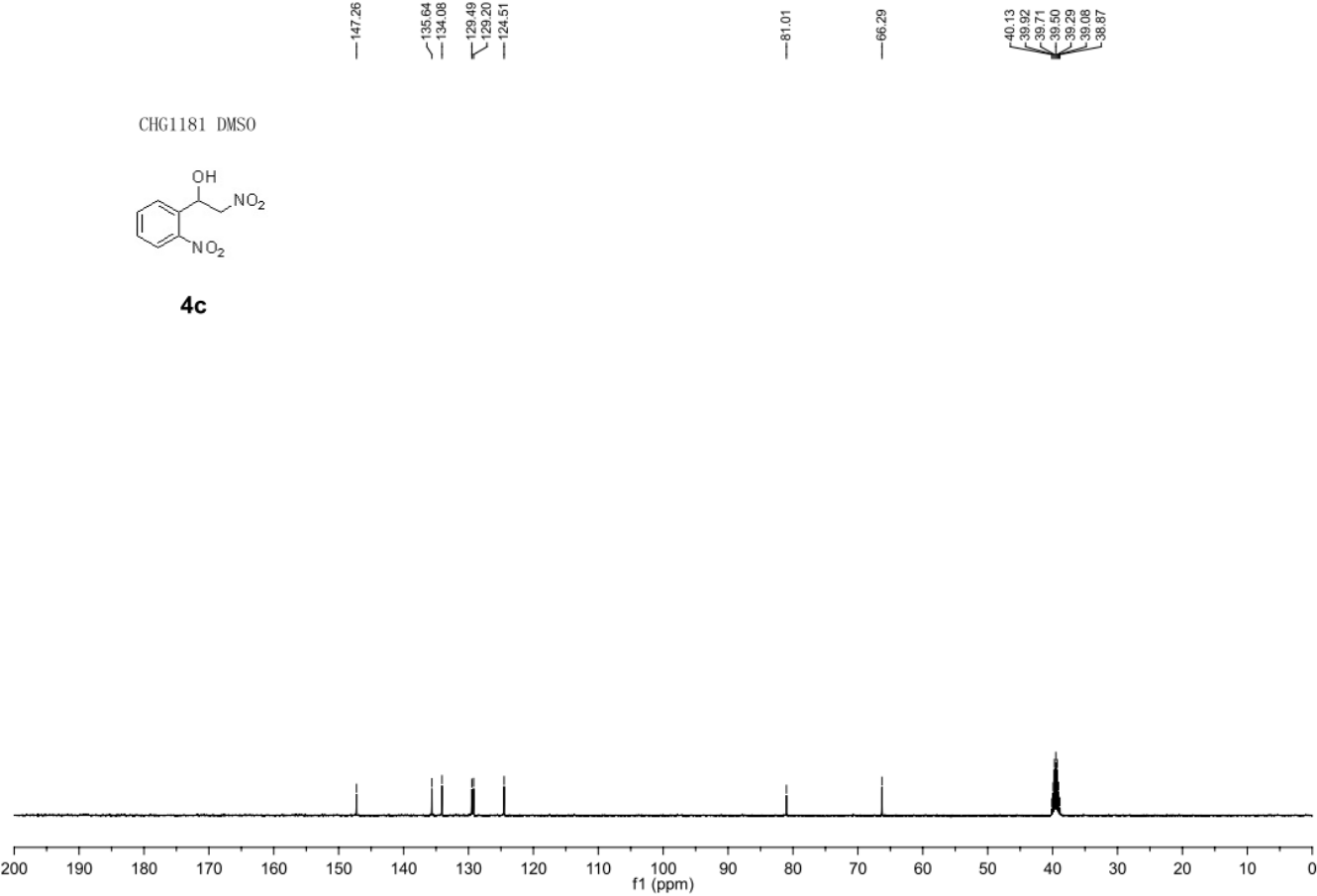
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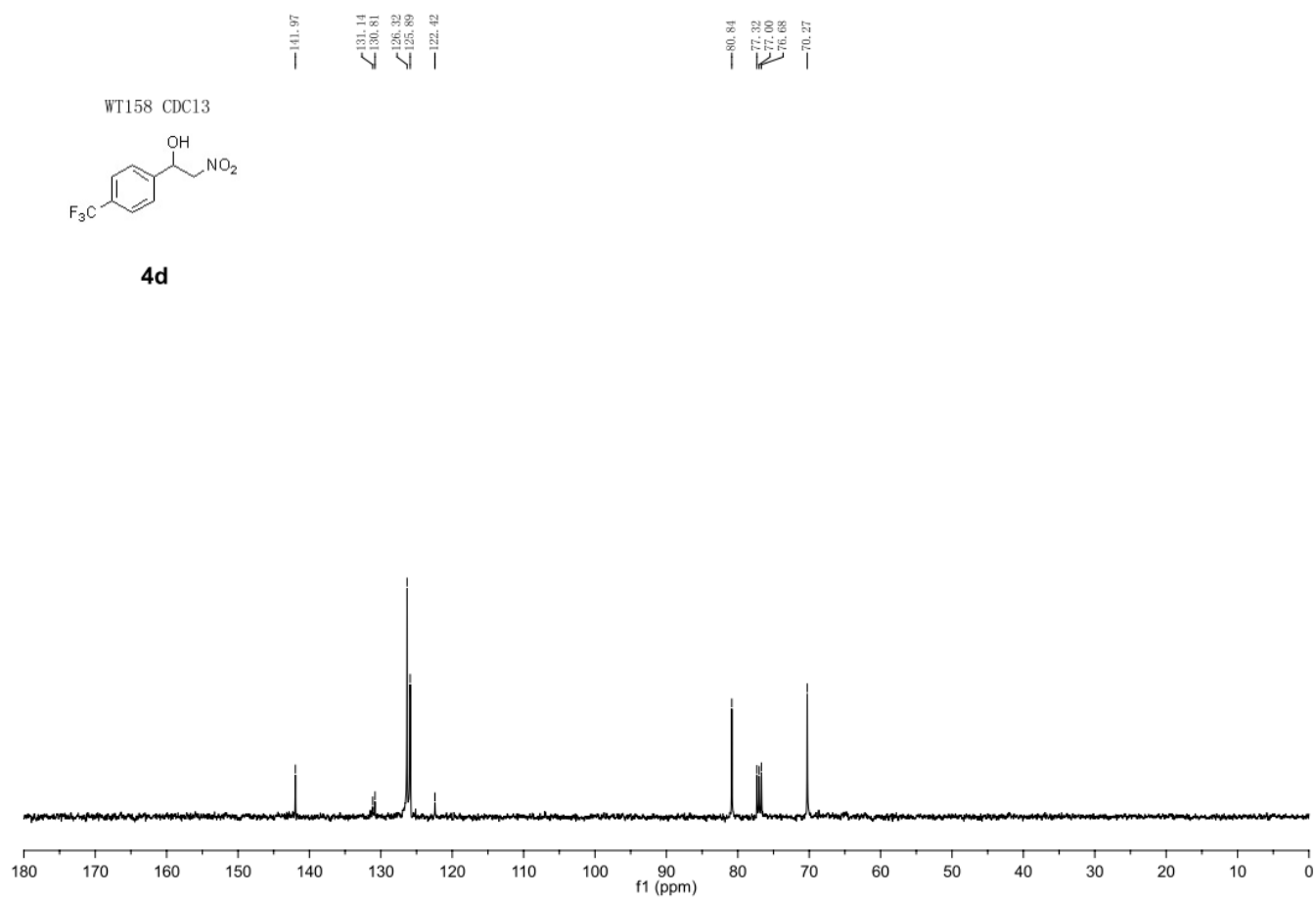
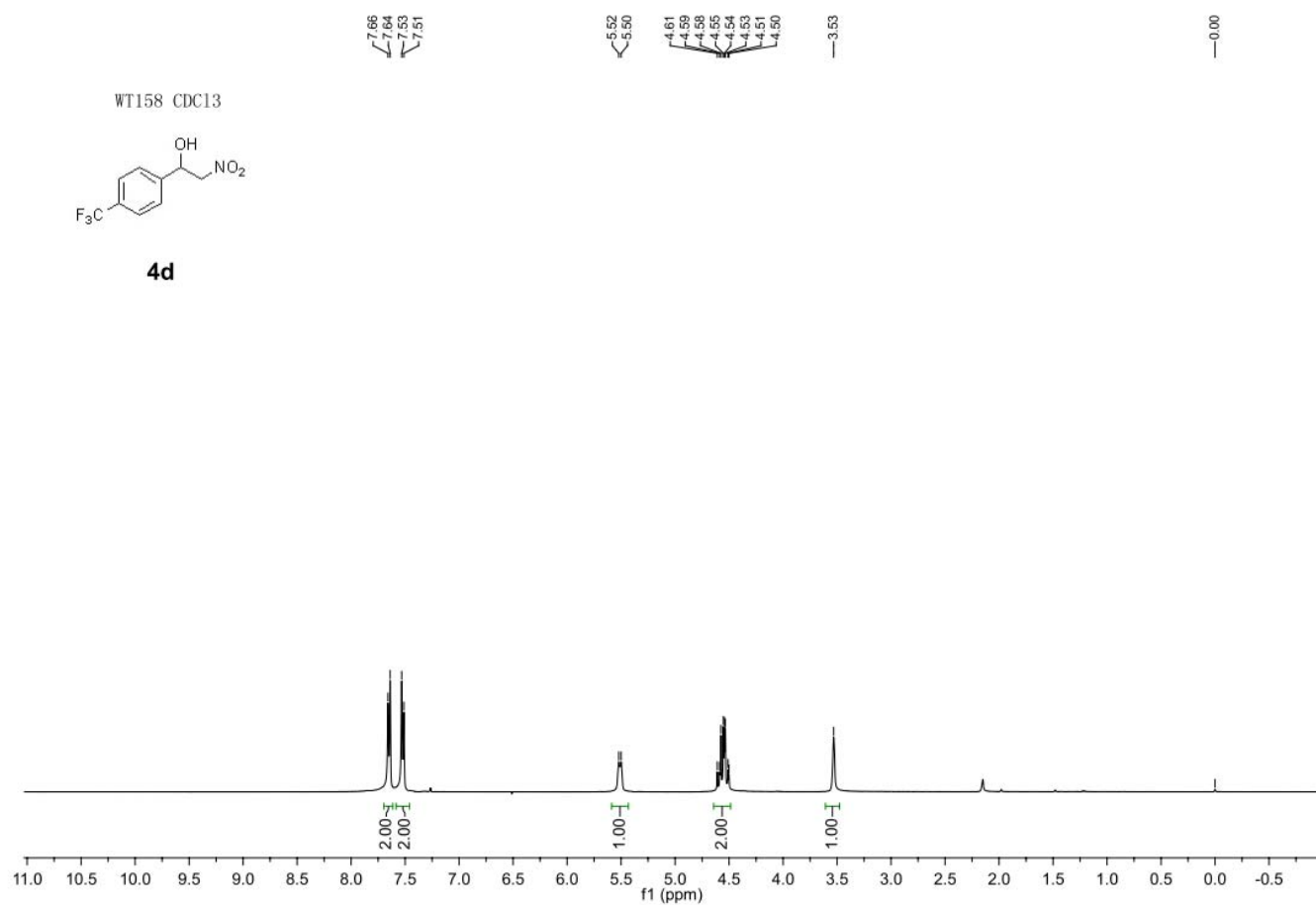


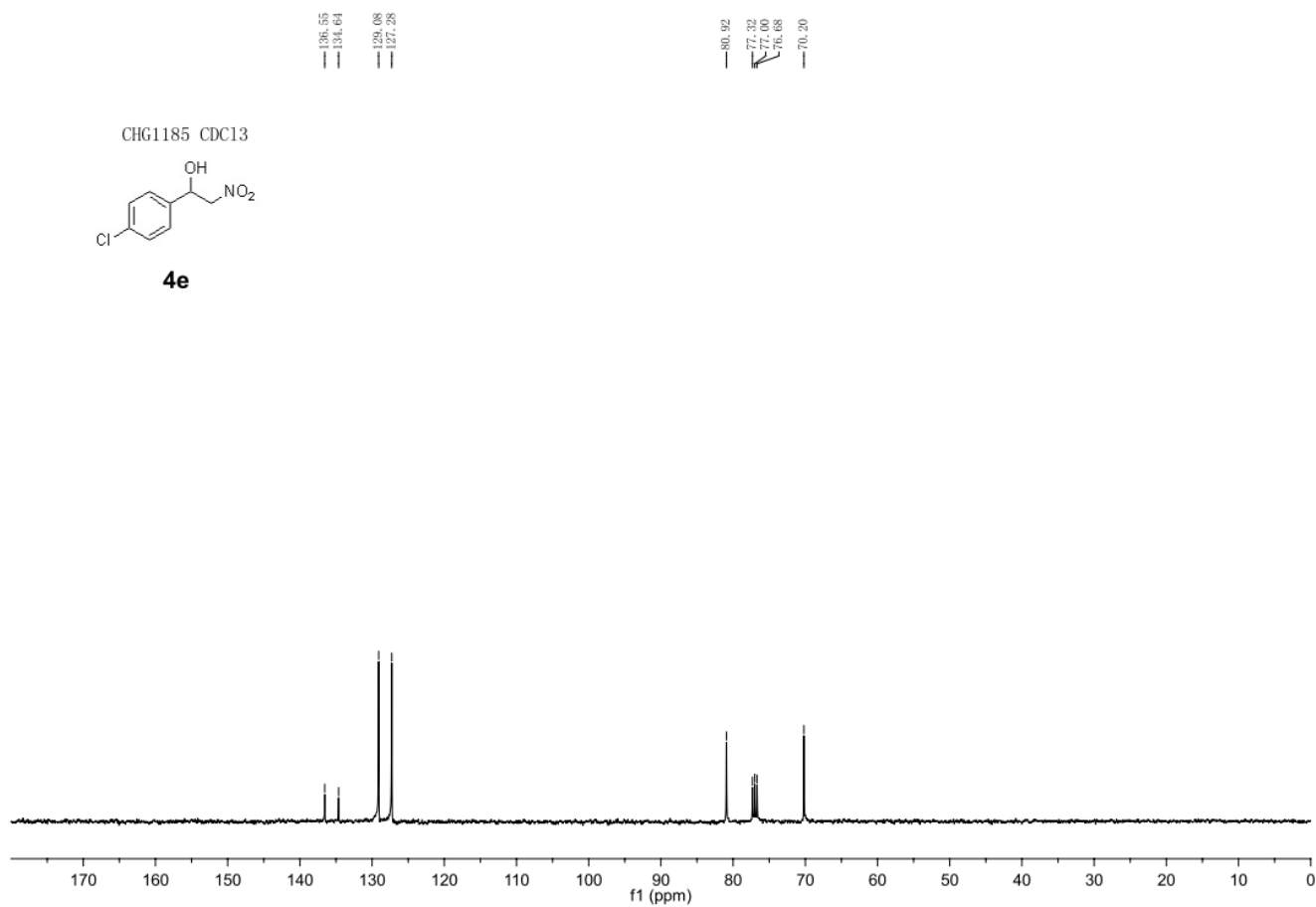
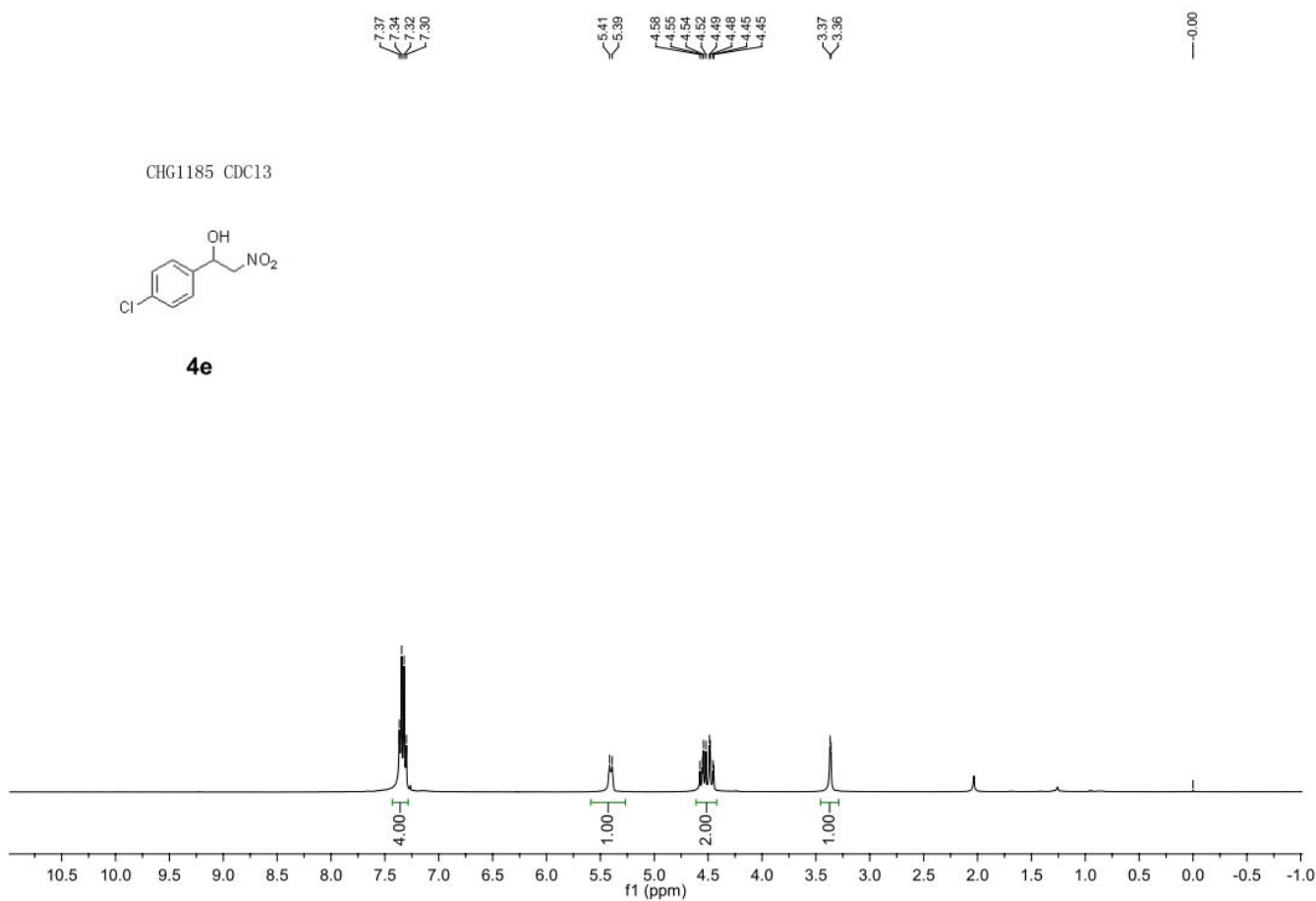
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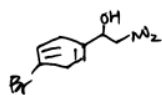
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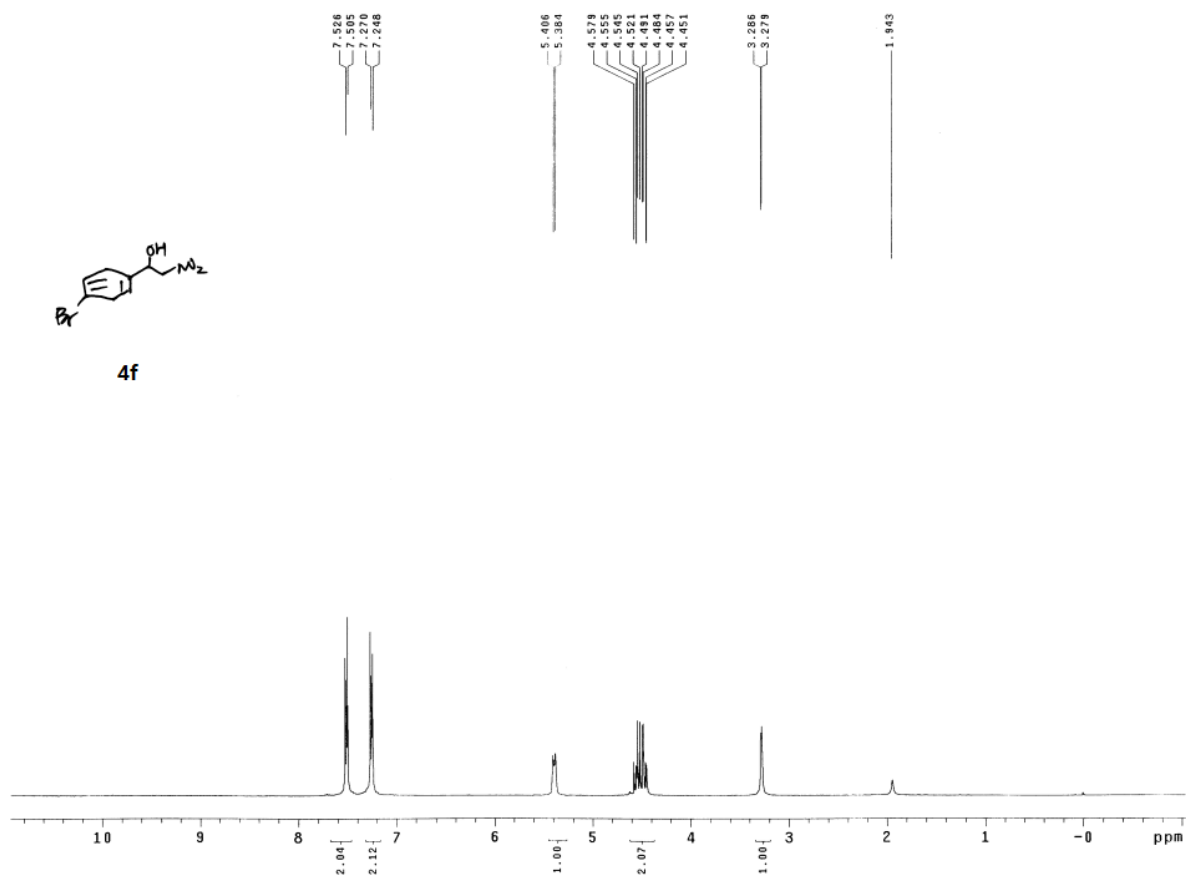




chg1186 cdc13
20110722



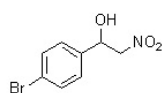
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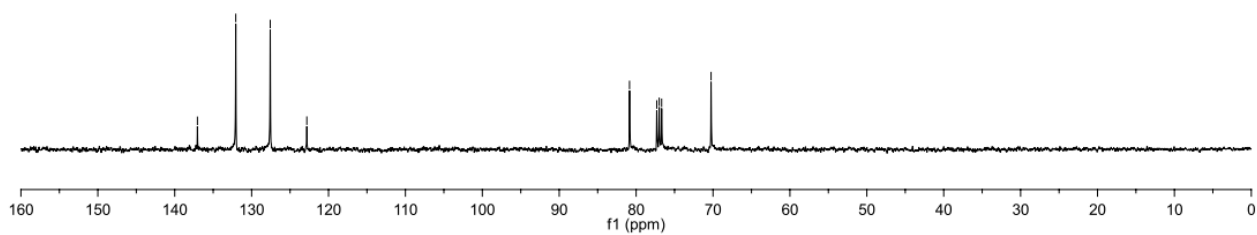
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— 132.06
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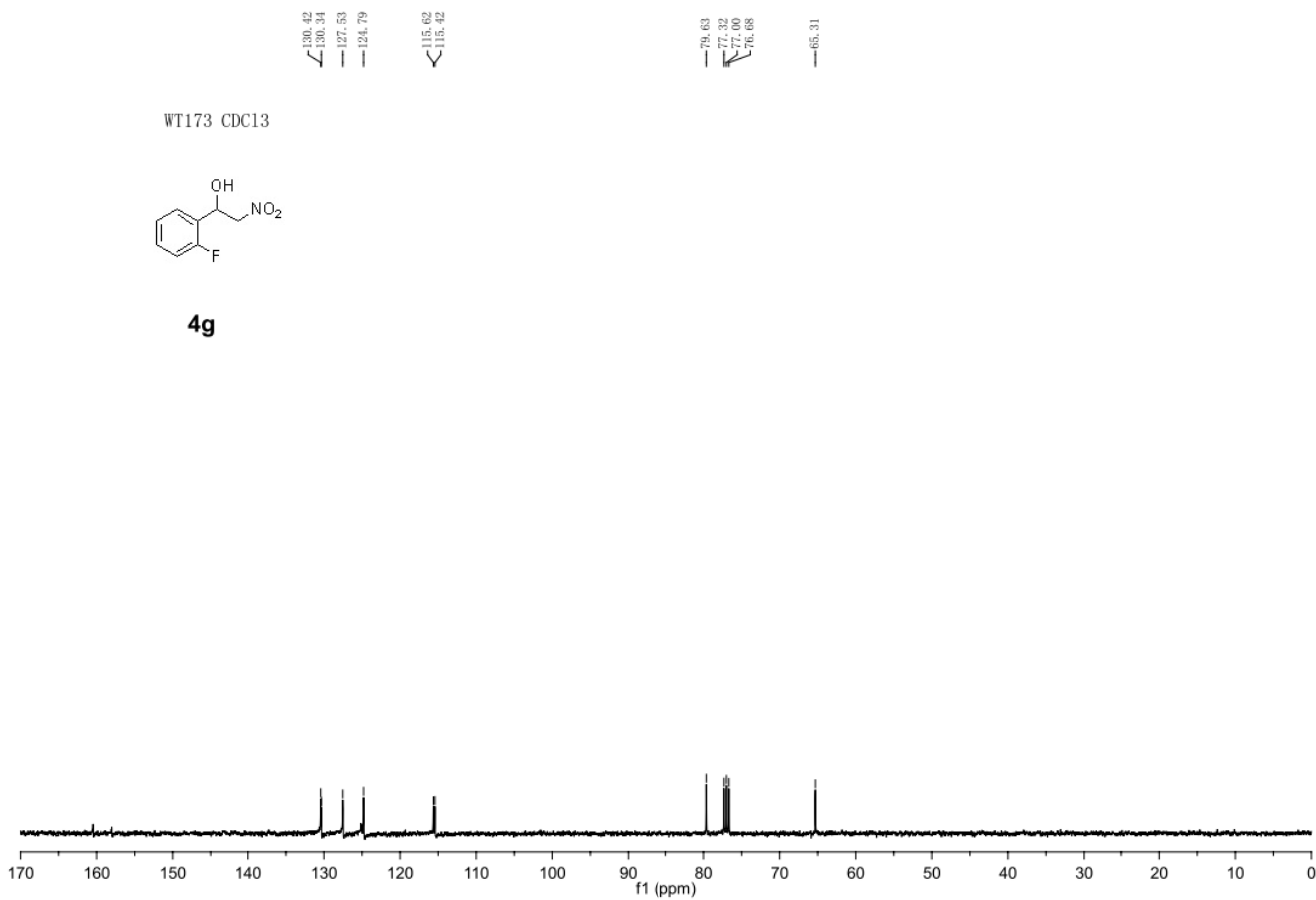
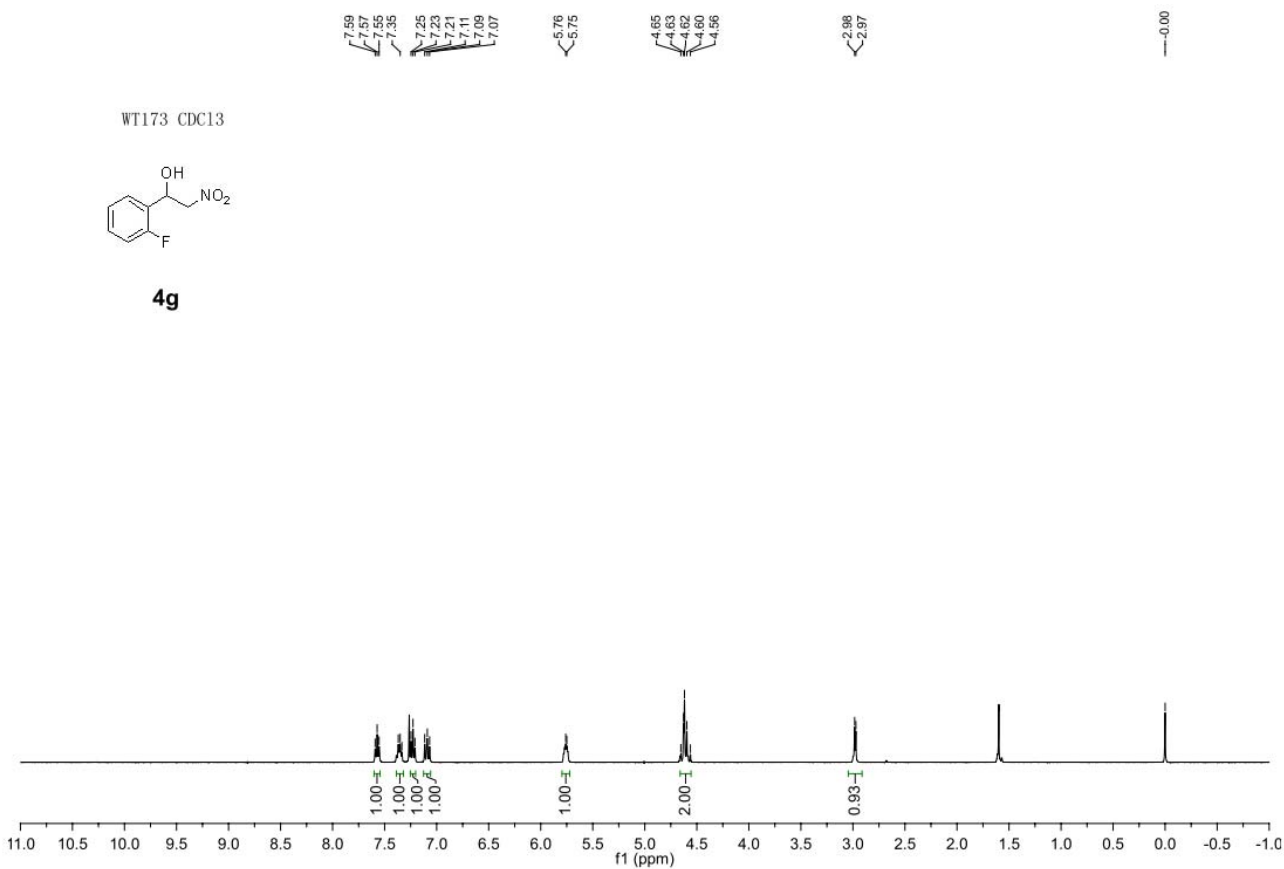
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— 77.32
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— 76.86
— 70.25

CHG1186 CDC13

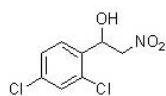


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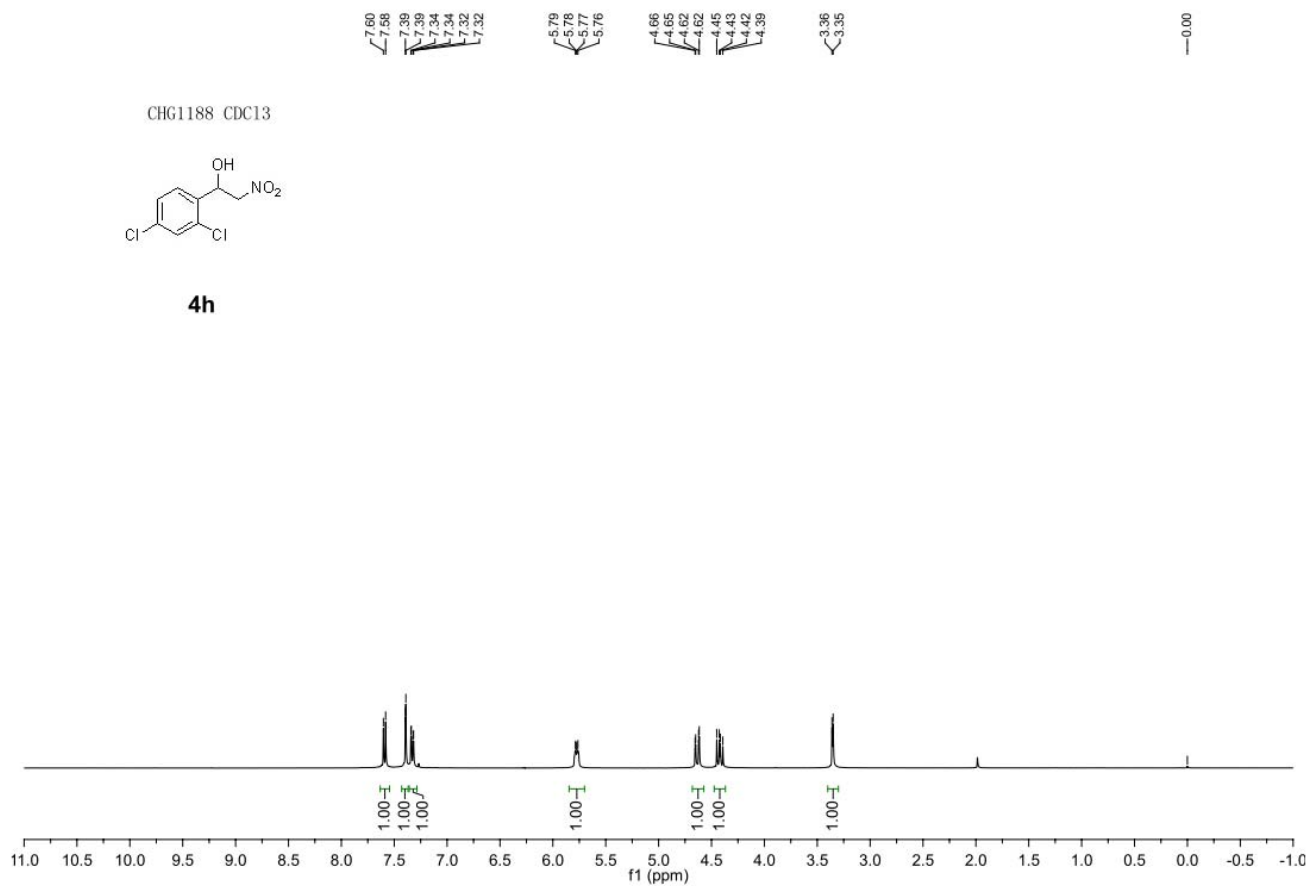




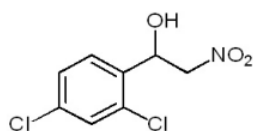
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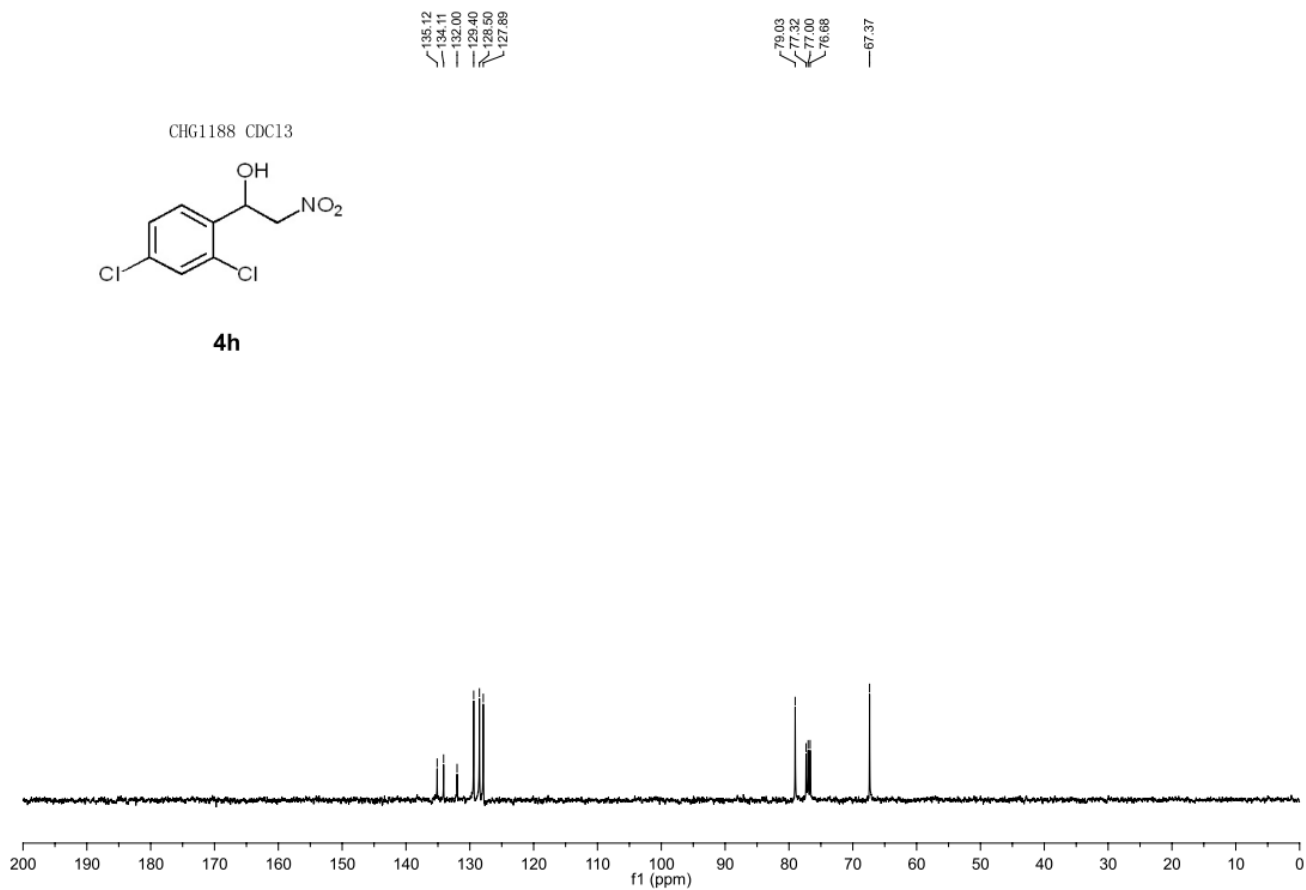
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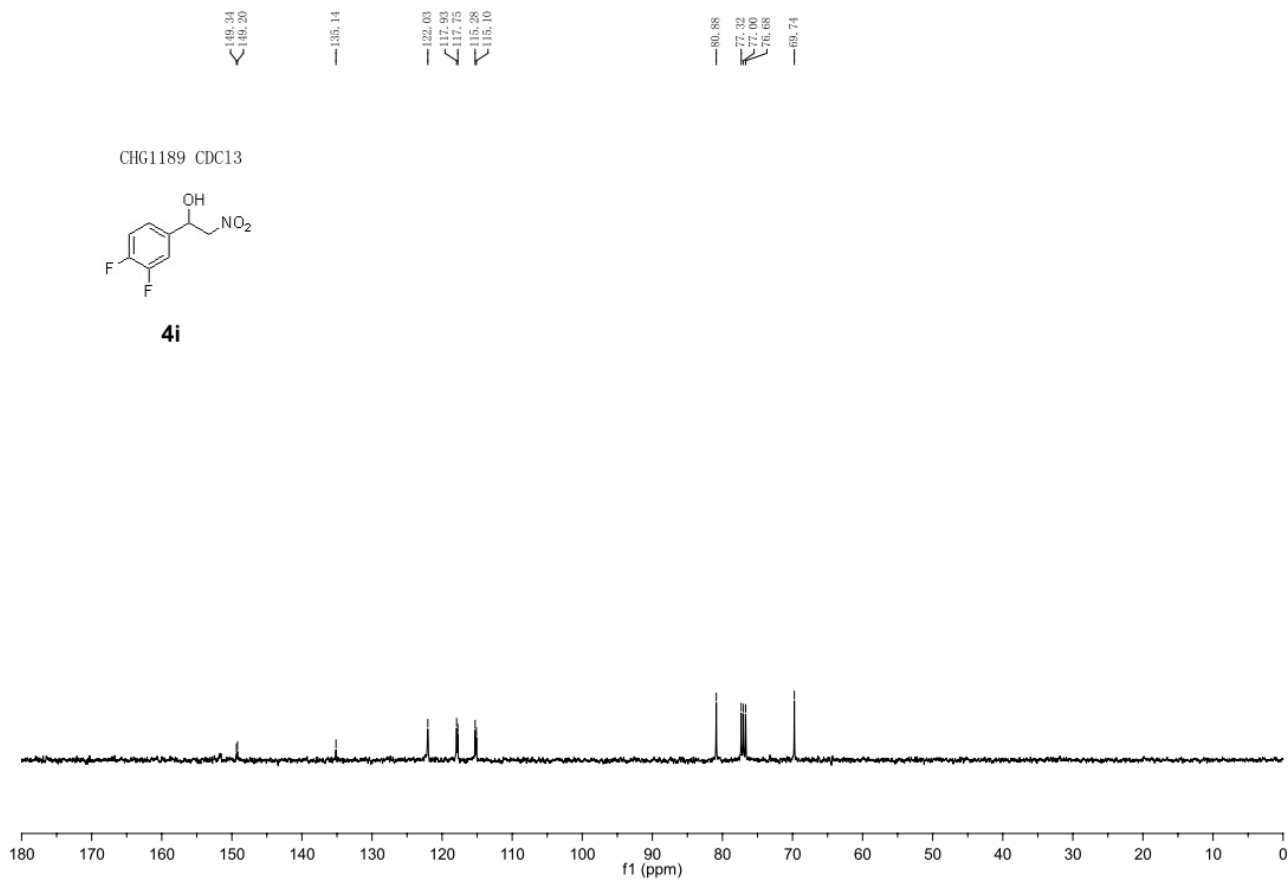
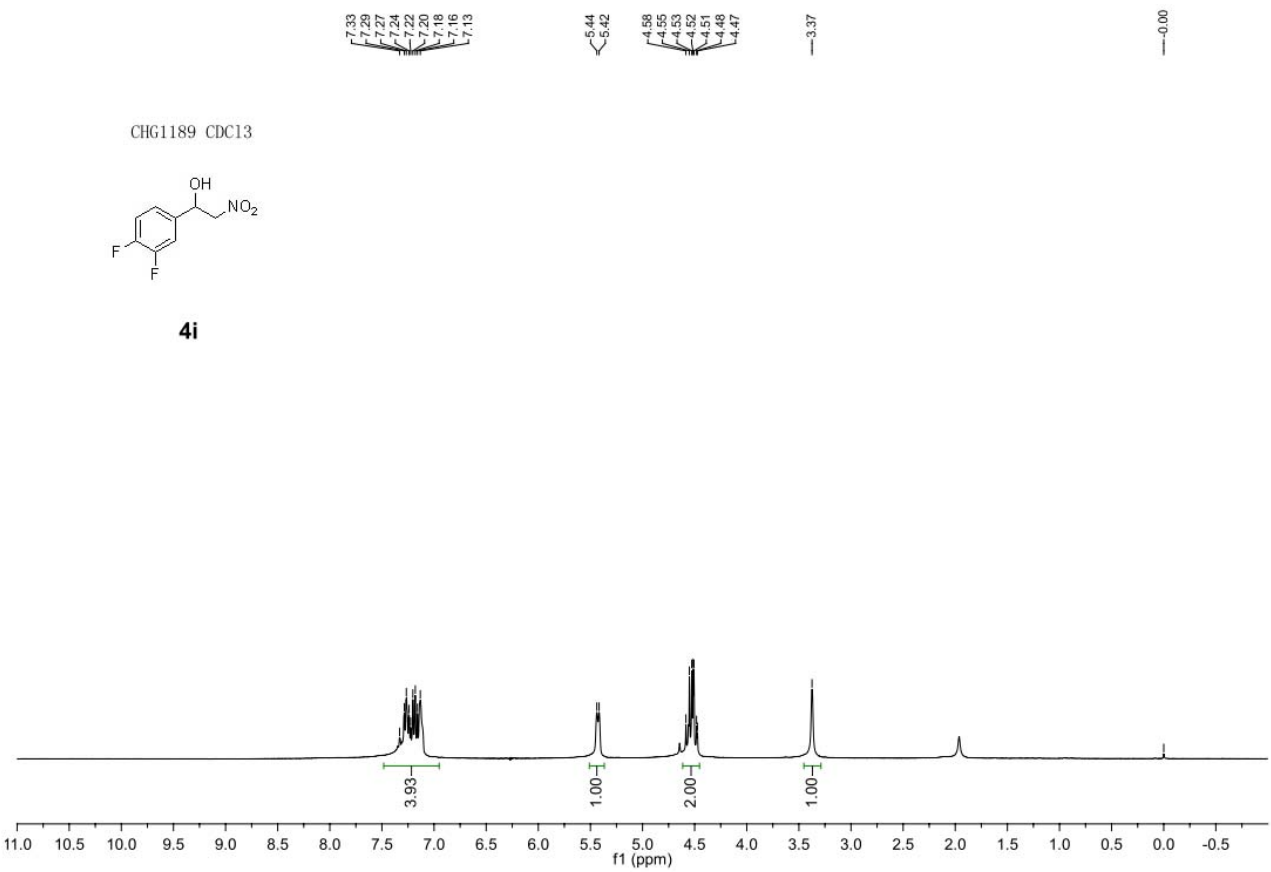


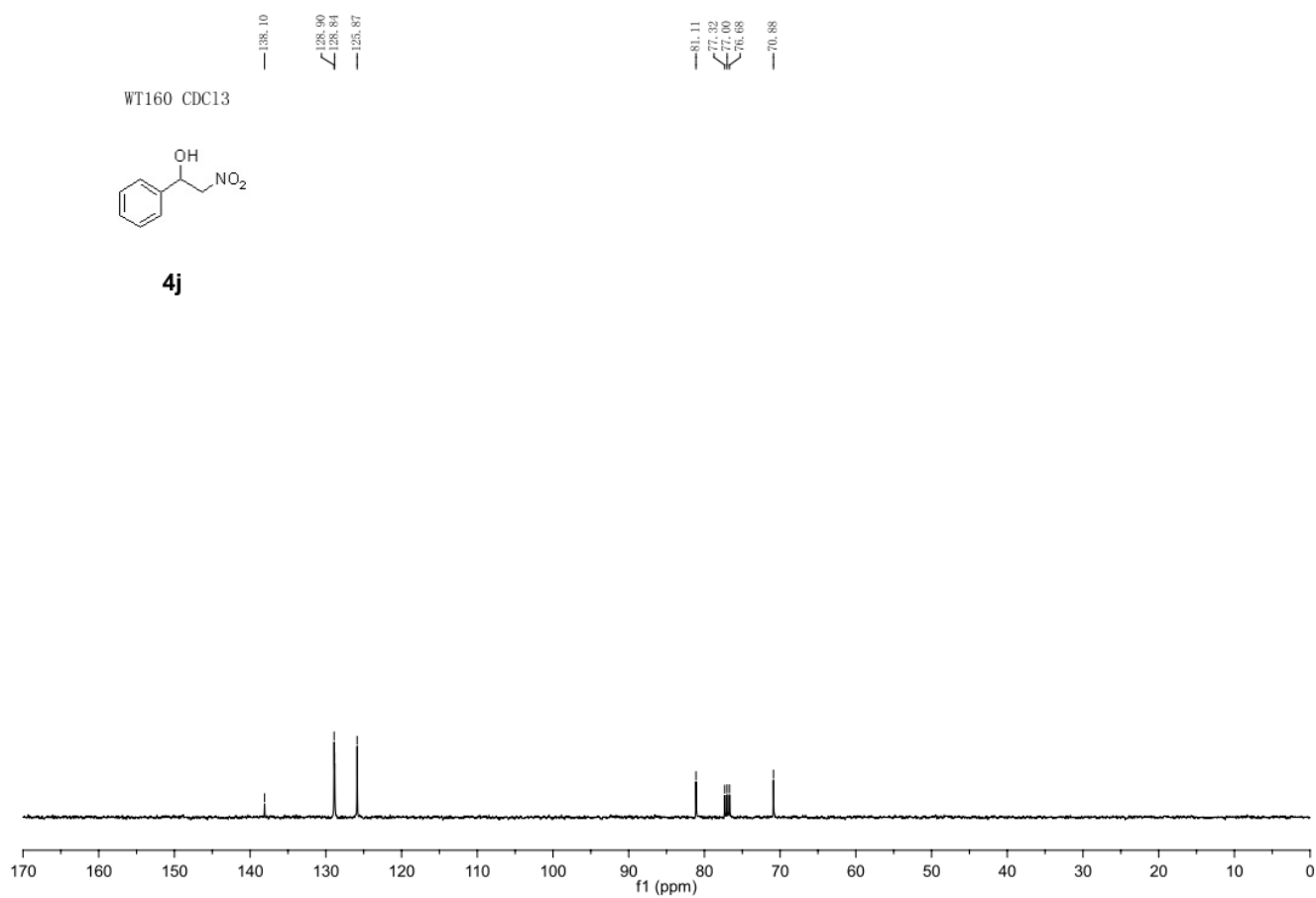
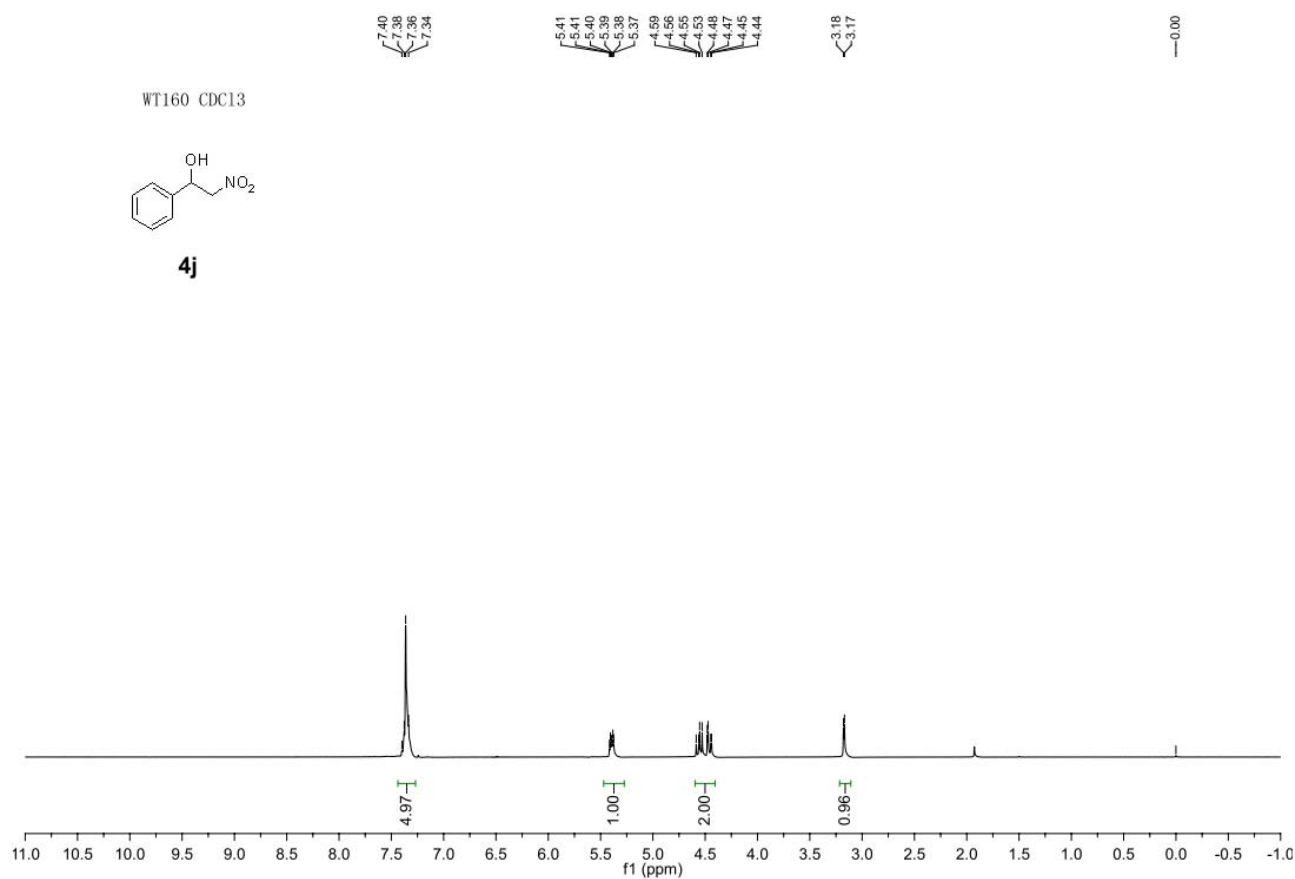
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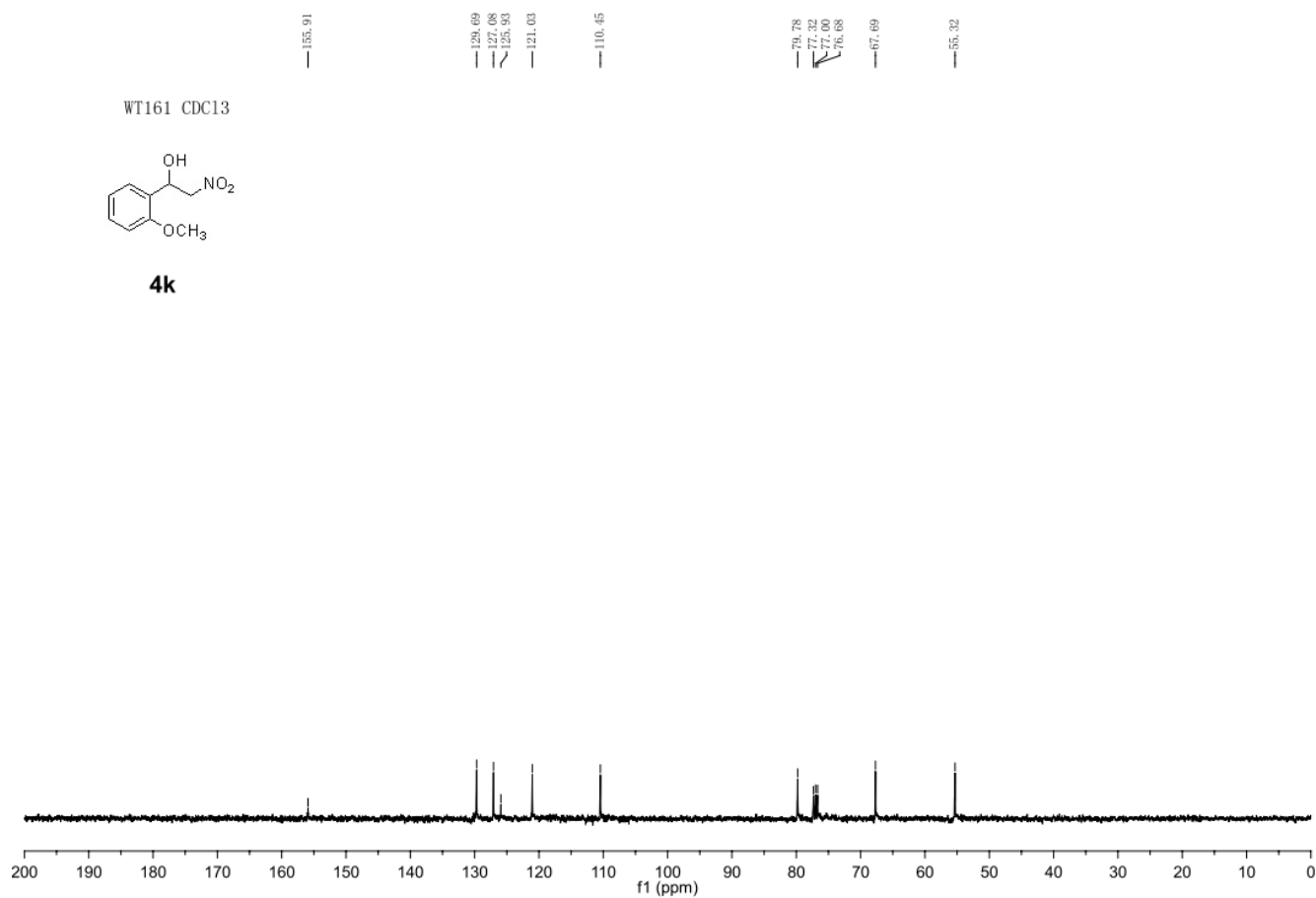
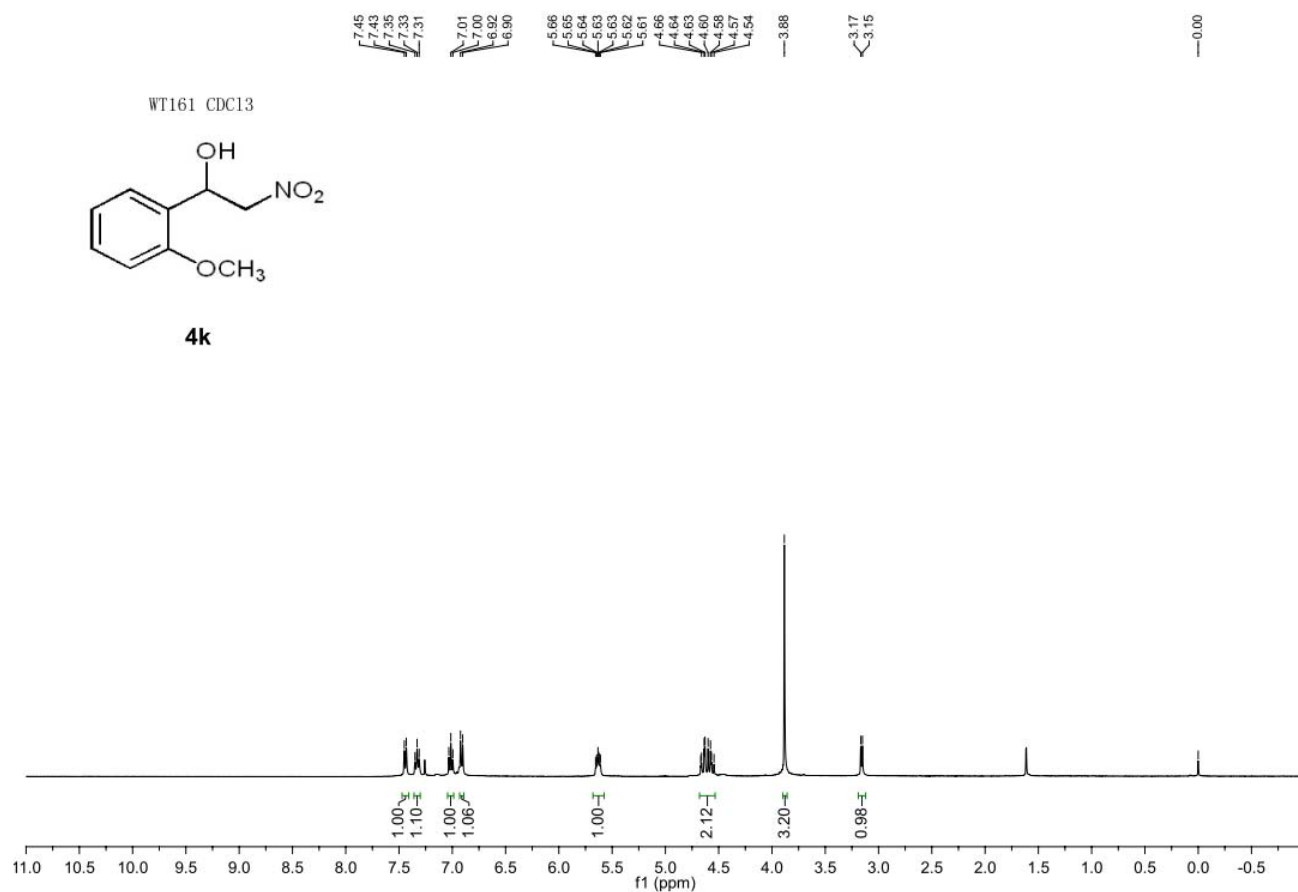


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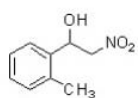




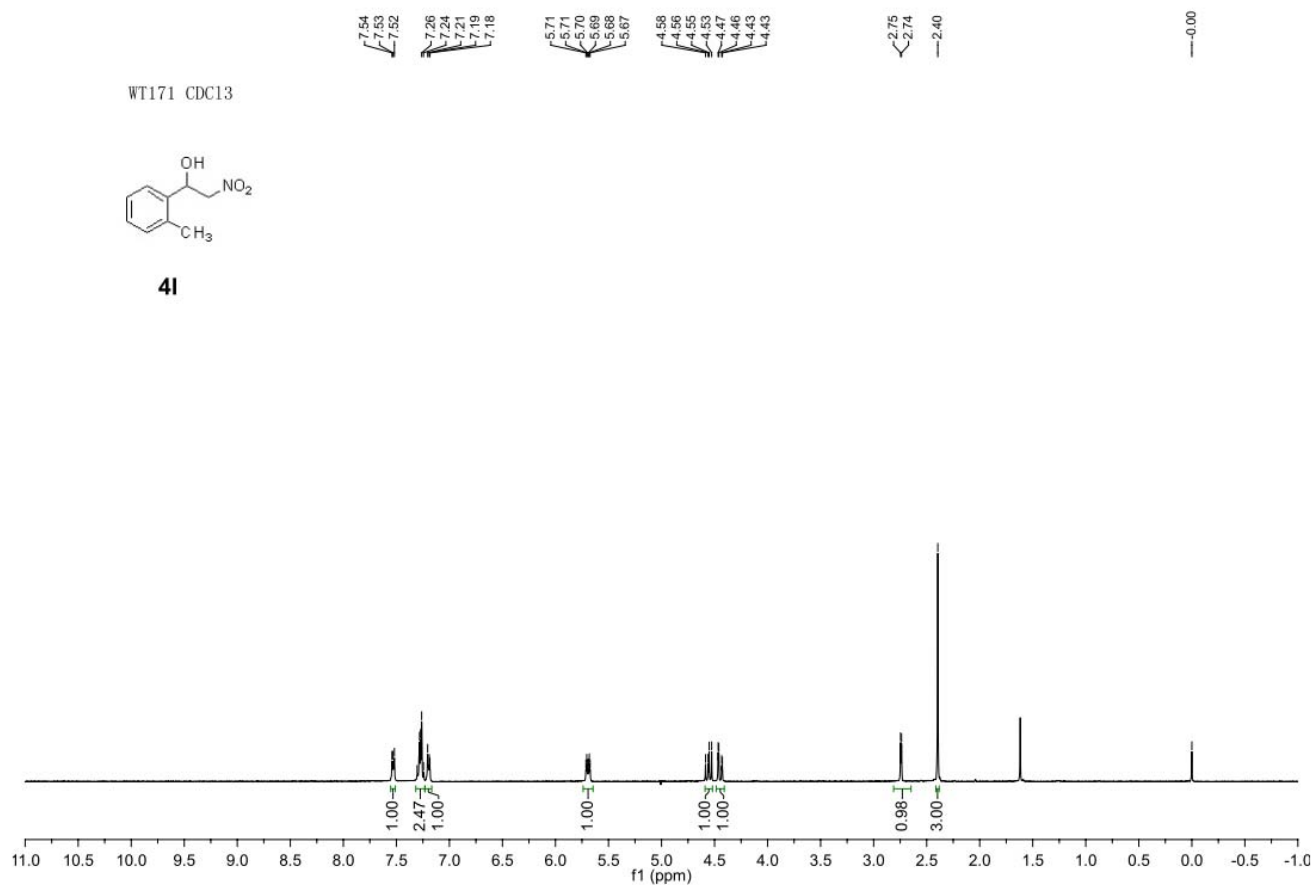




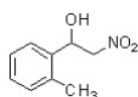
WT171 CDC13



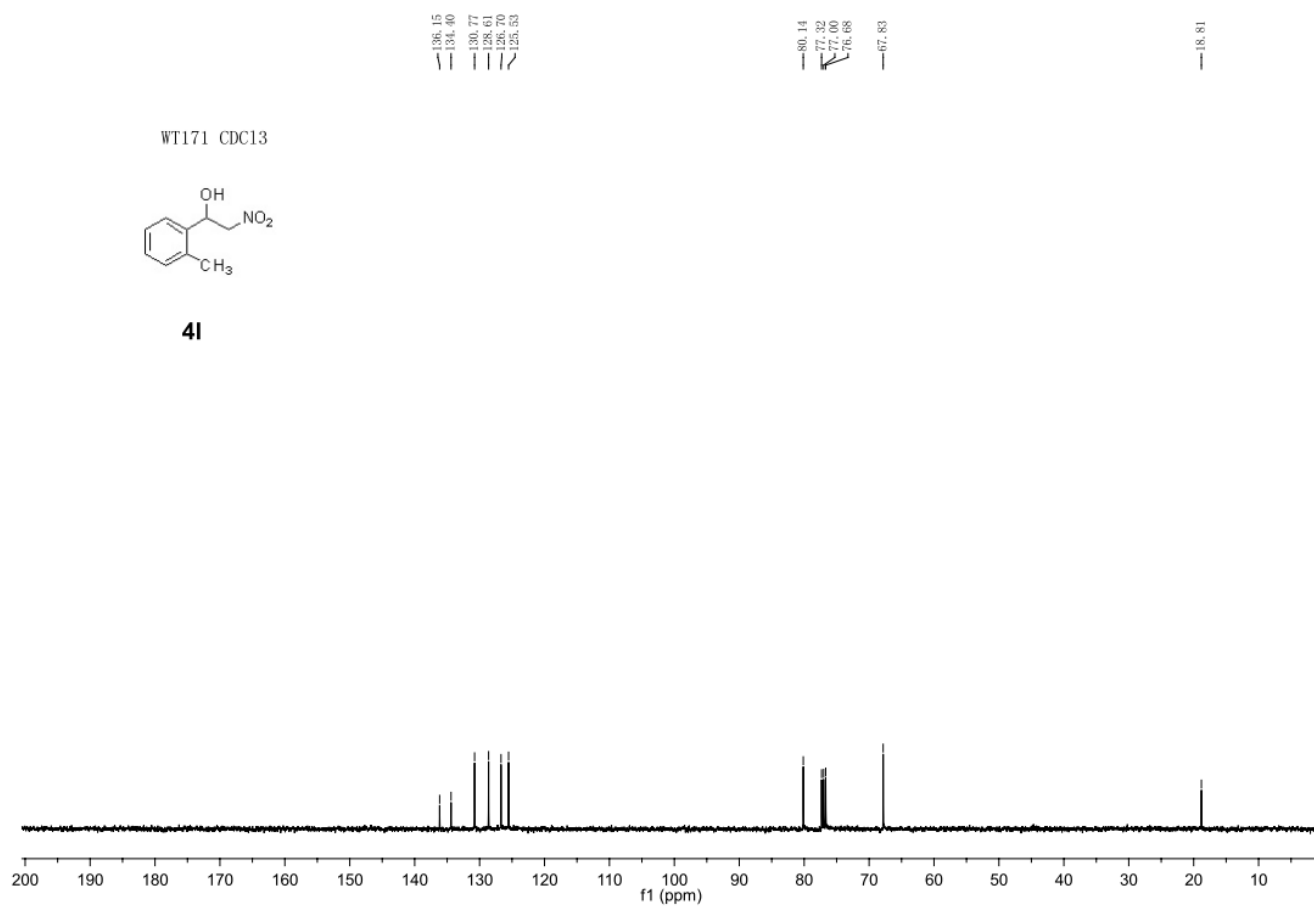
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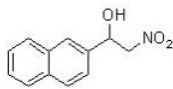
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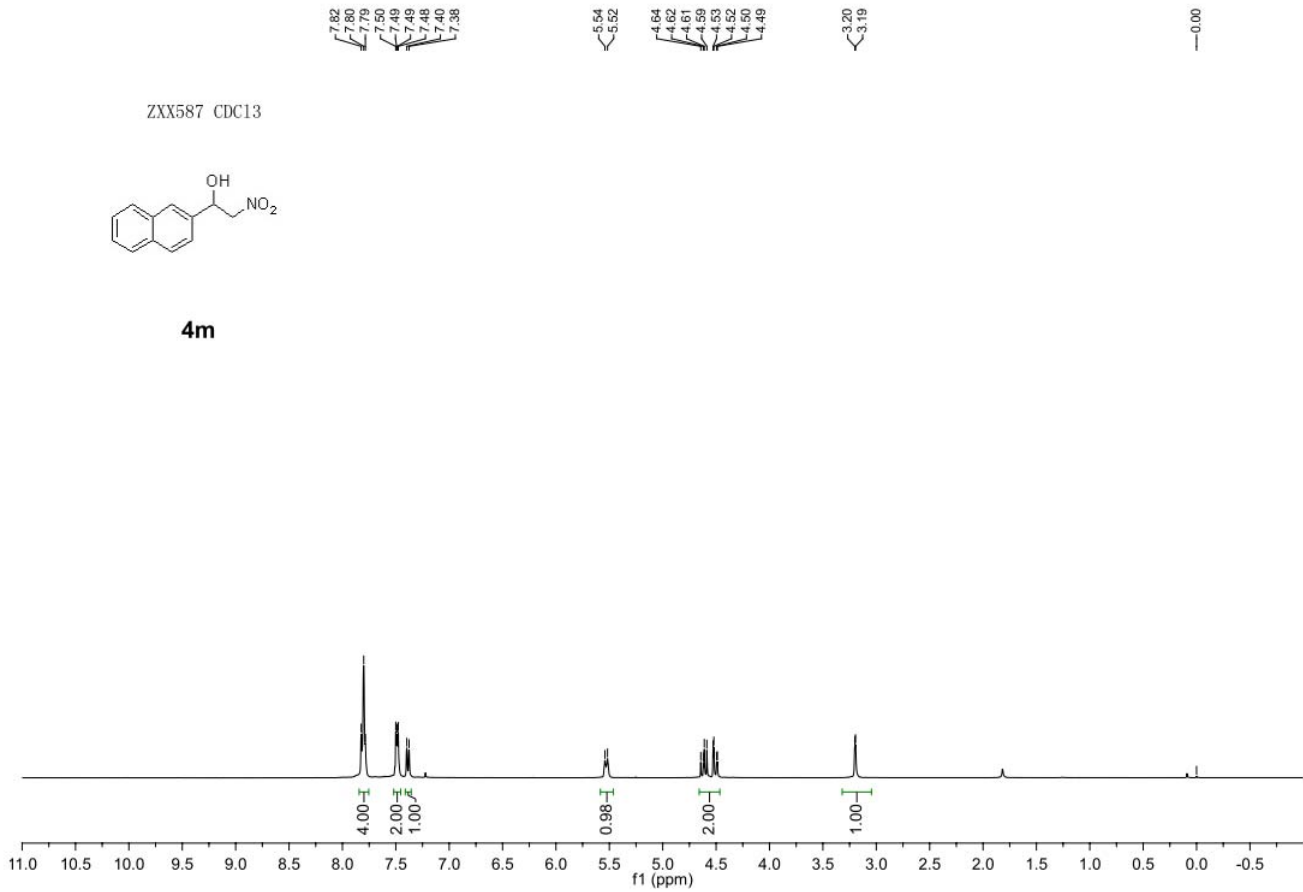
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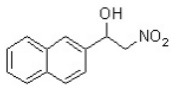
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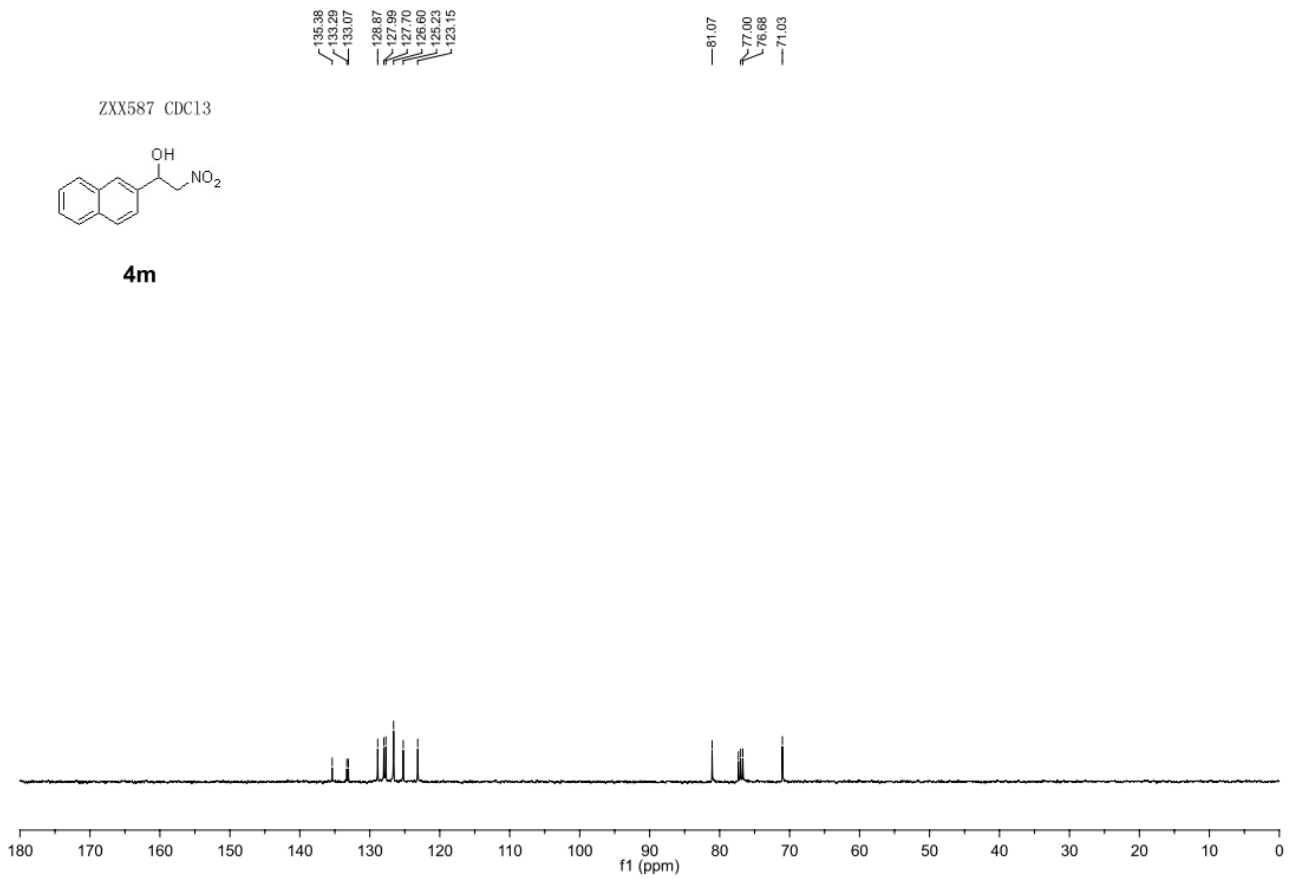
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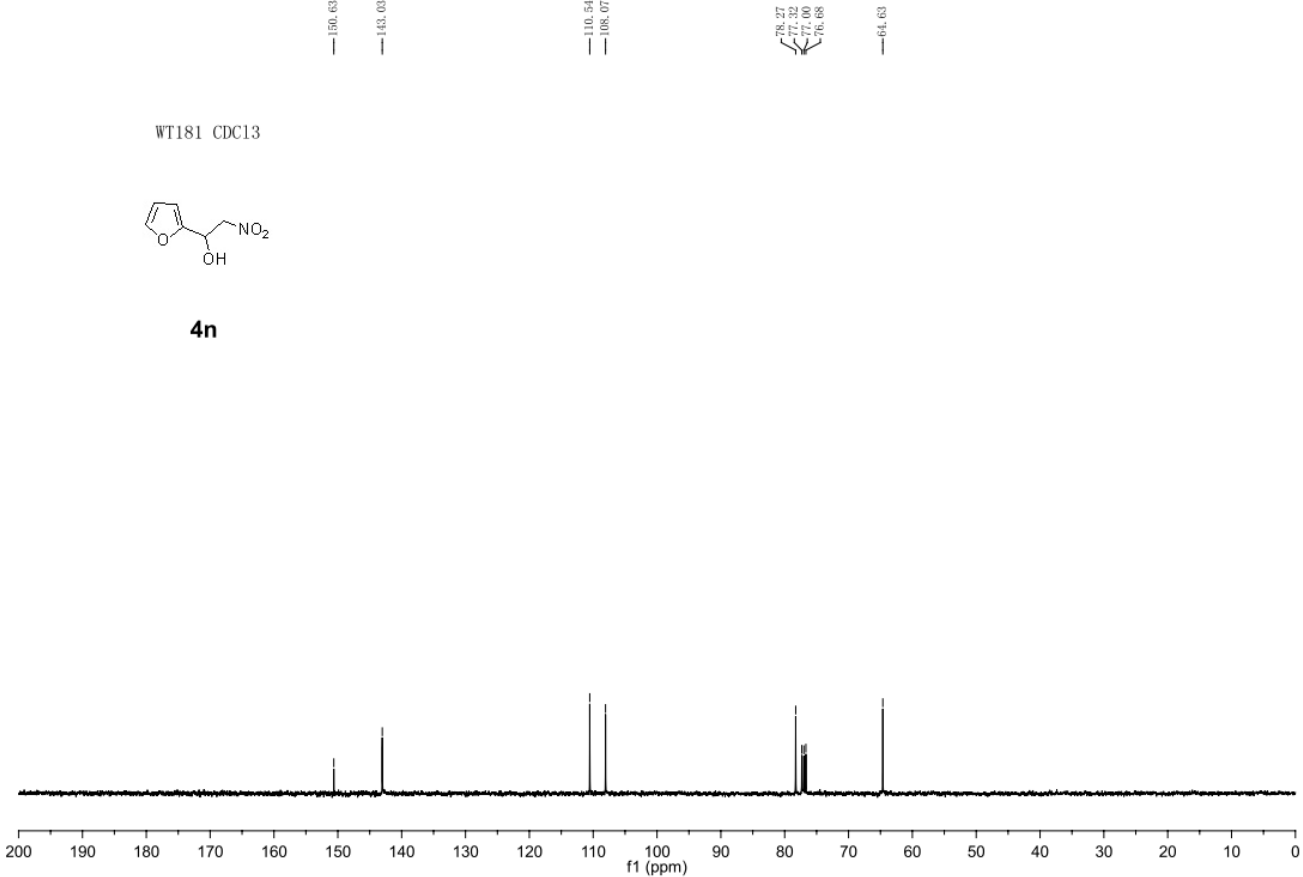
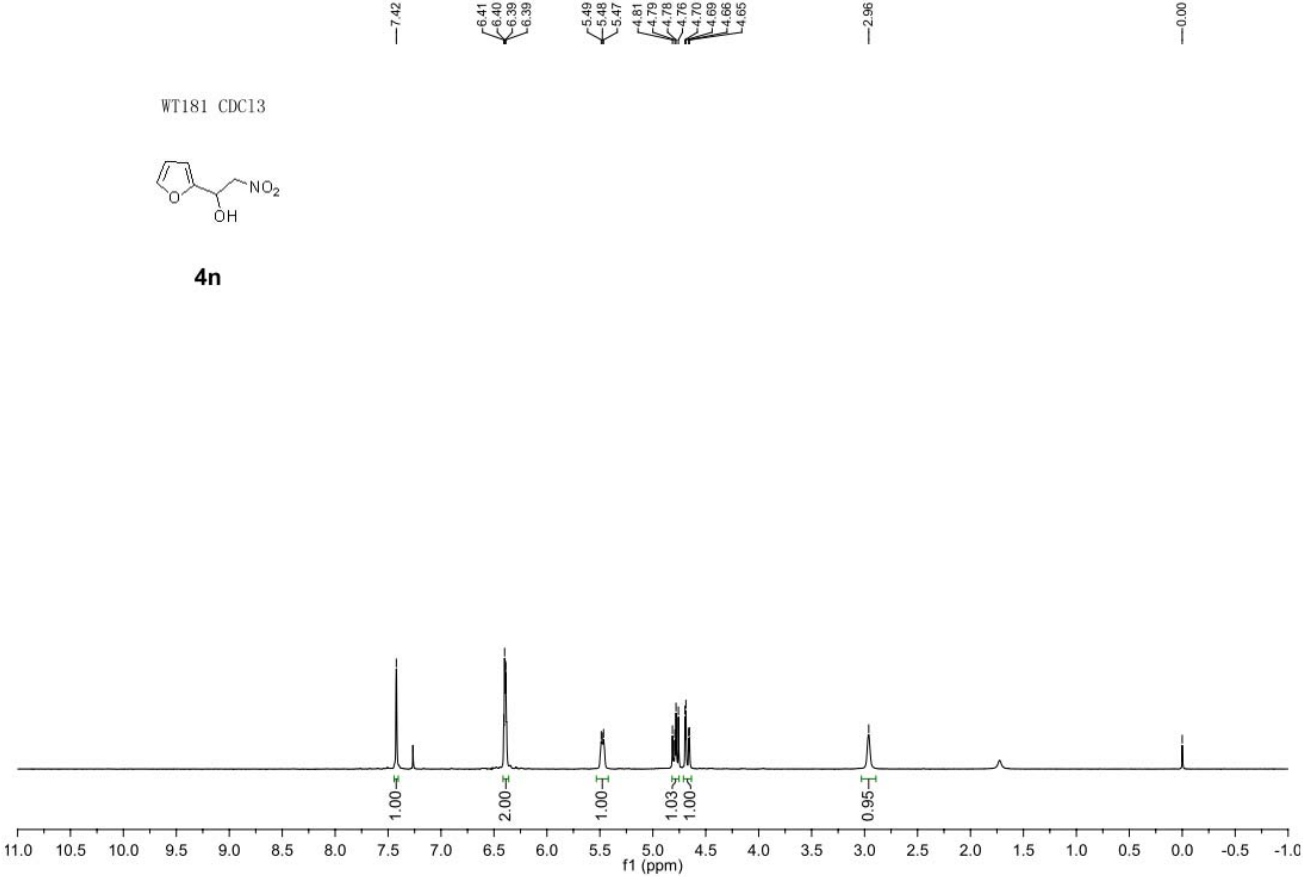


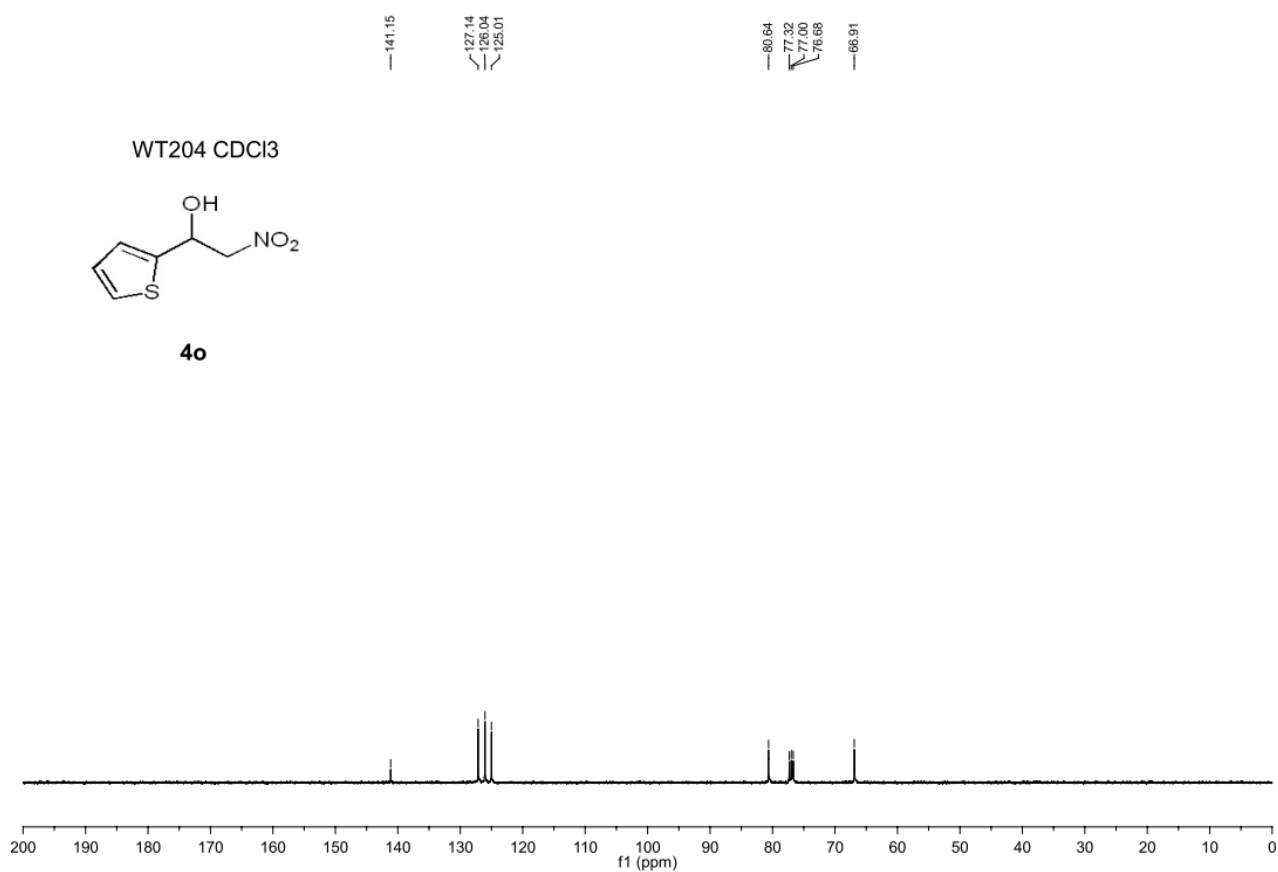
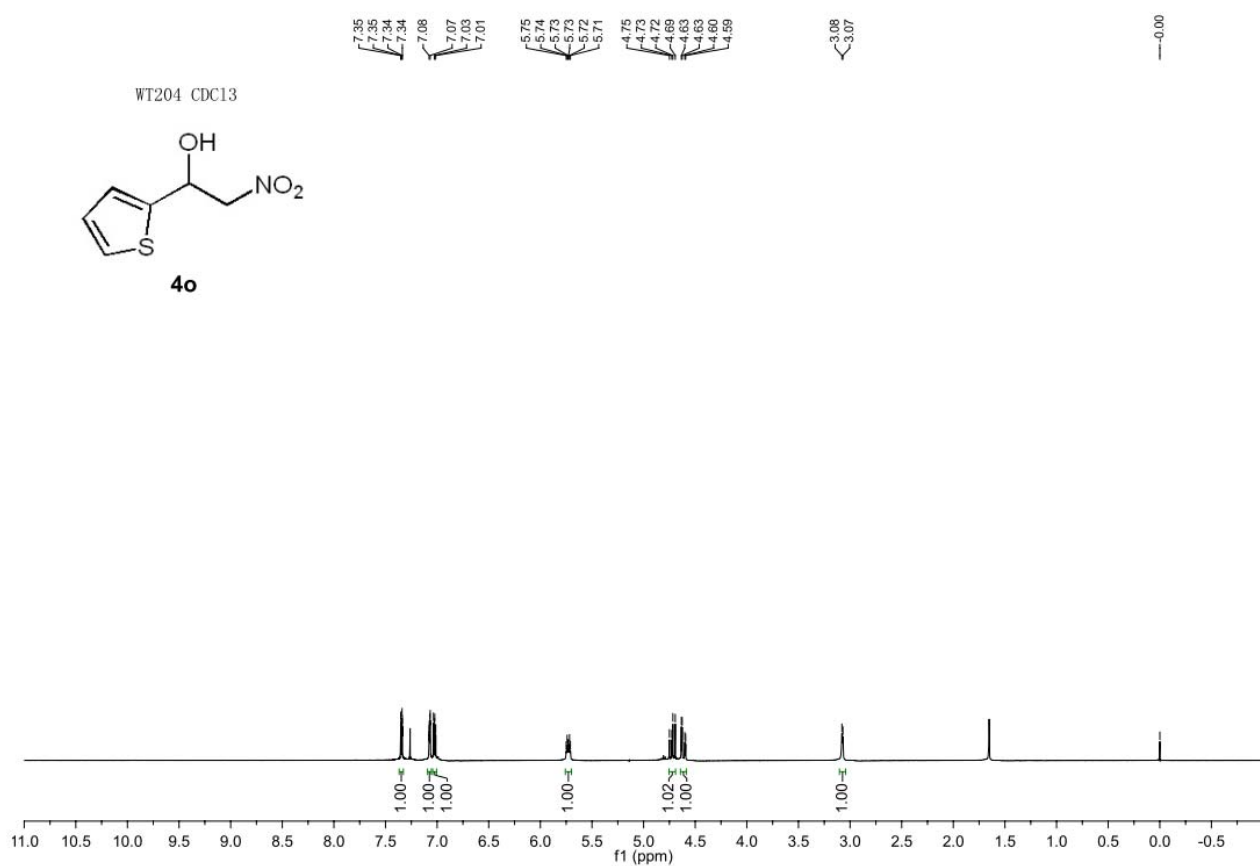
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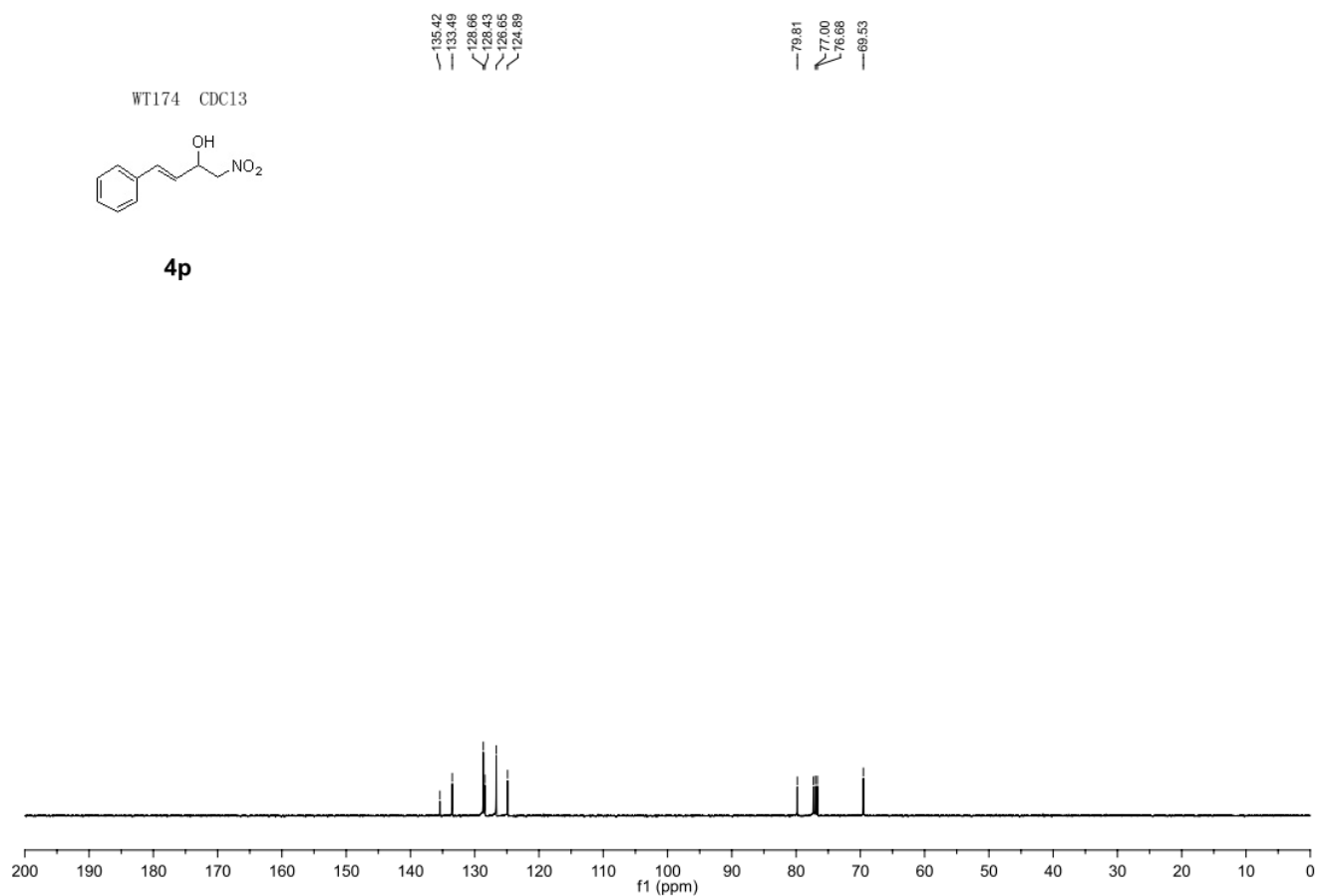
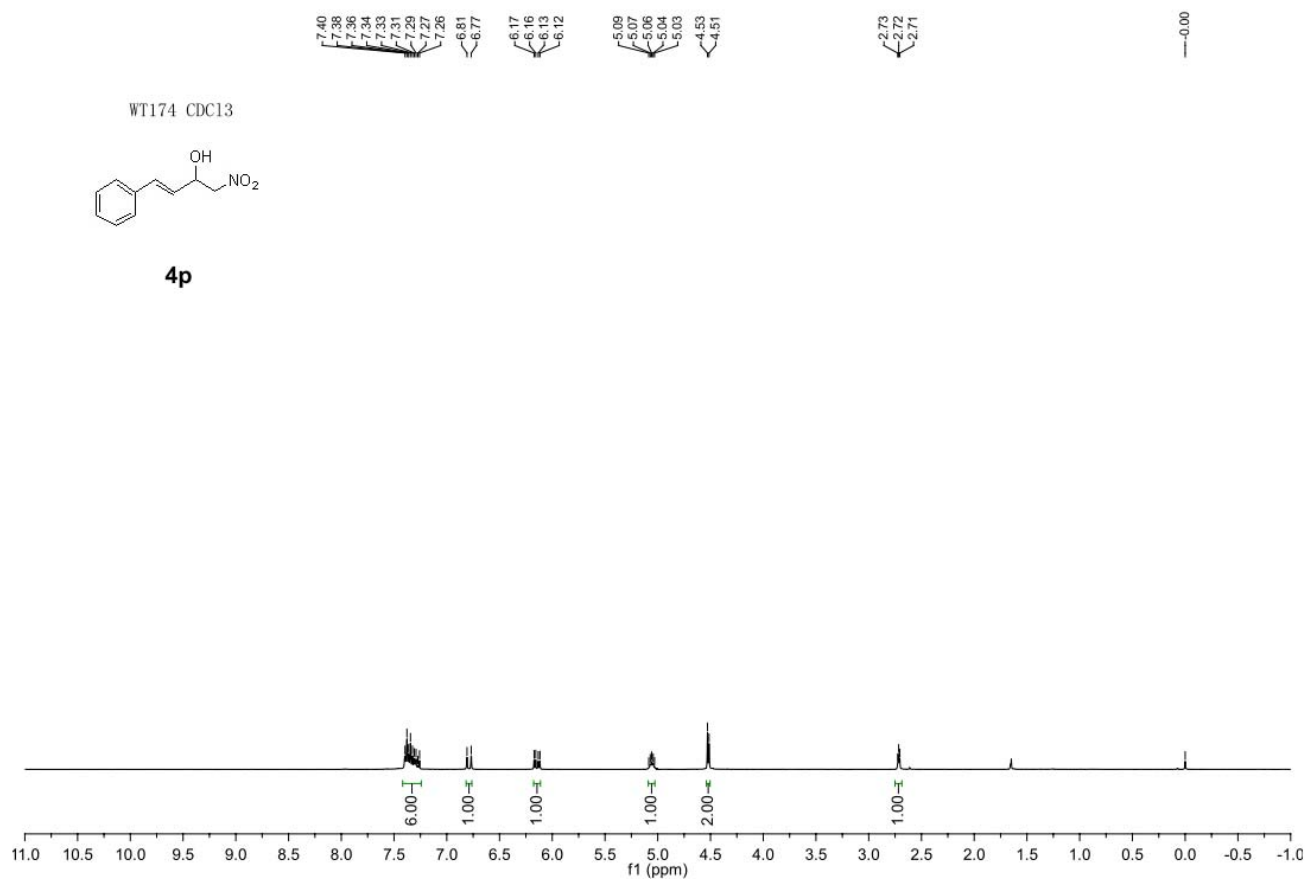


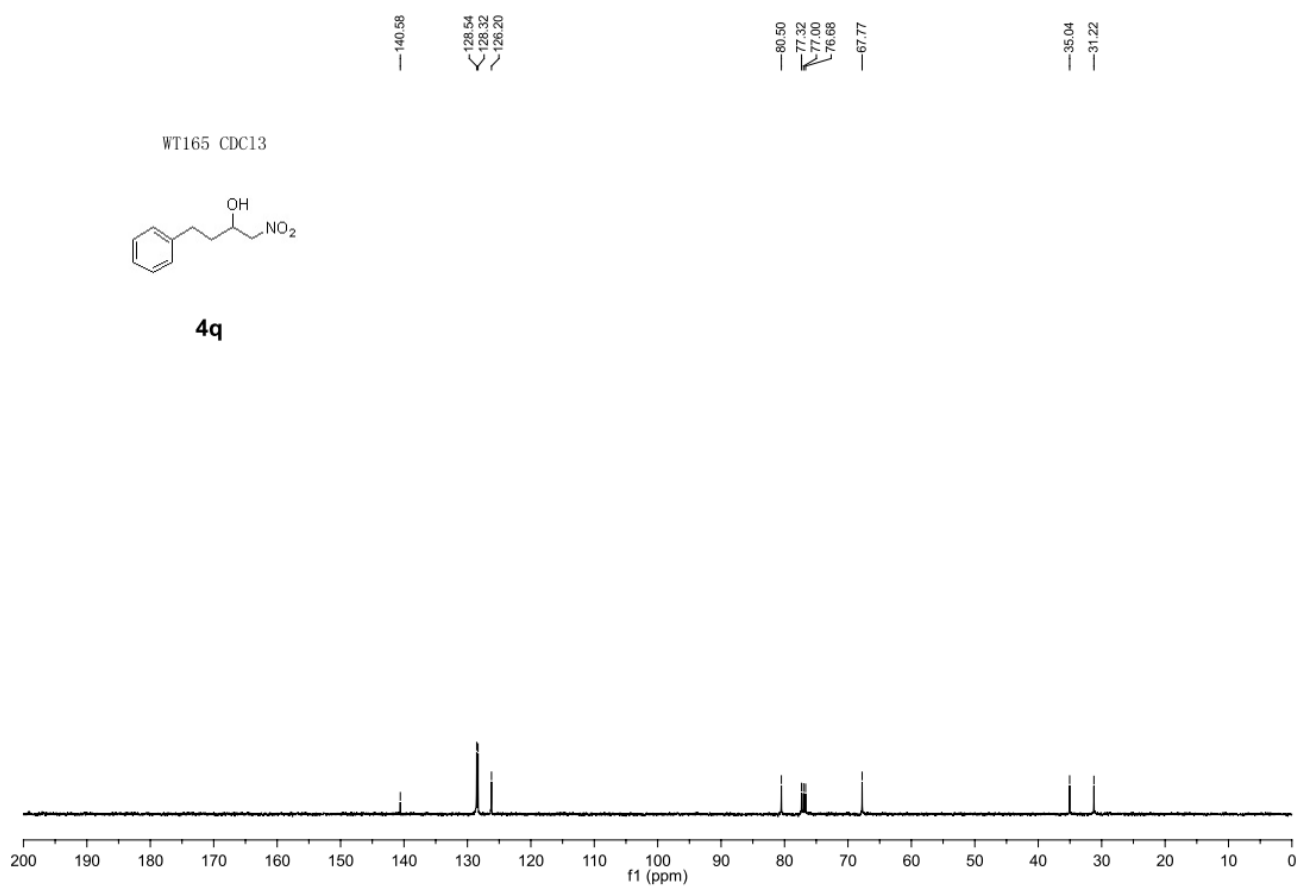
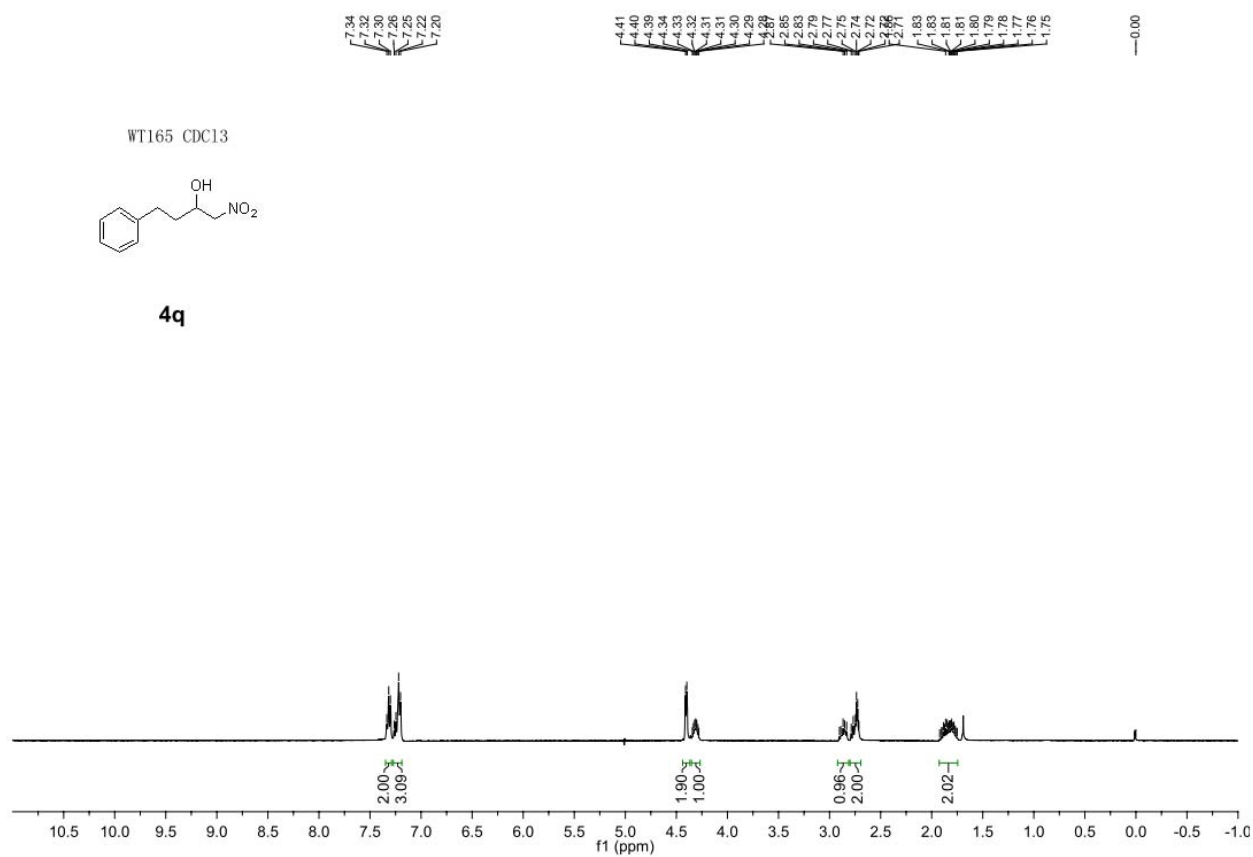
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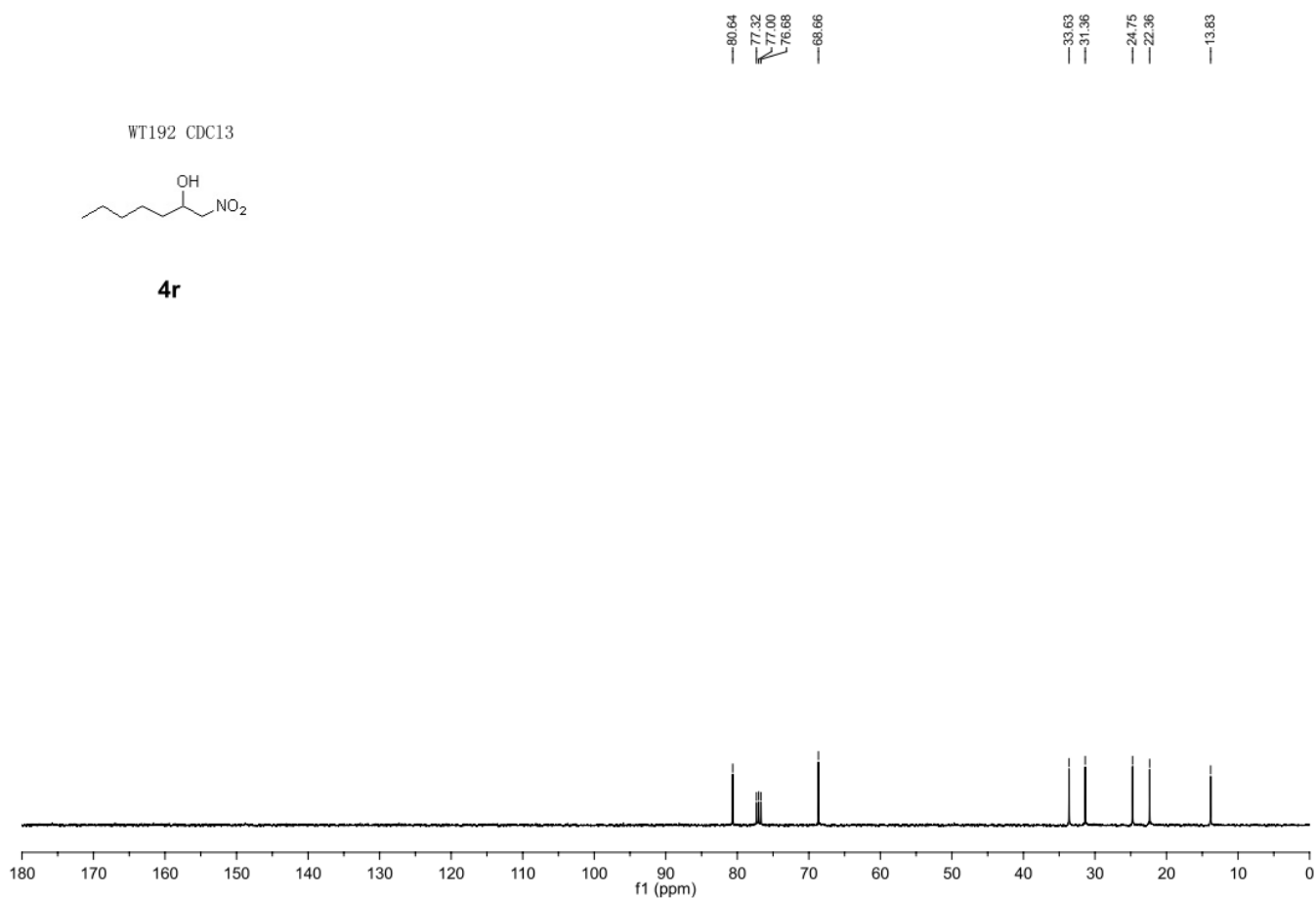
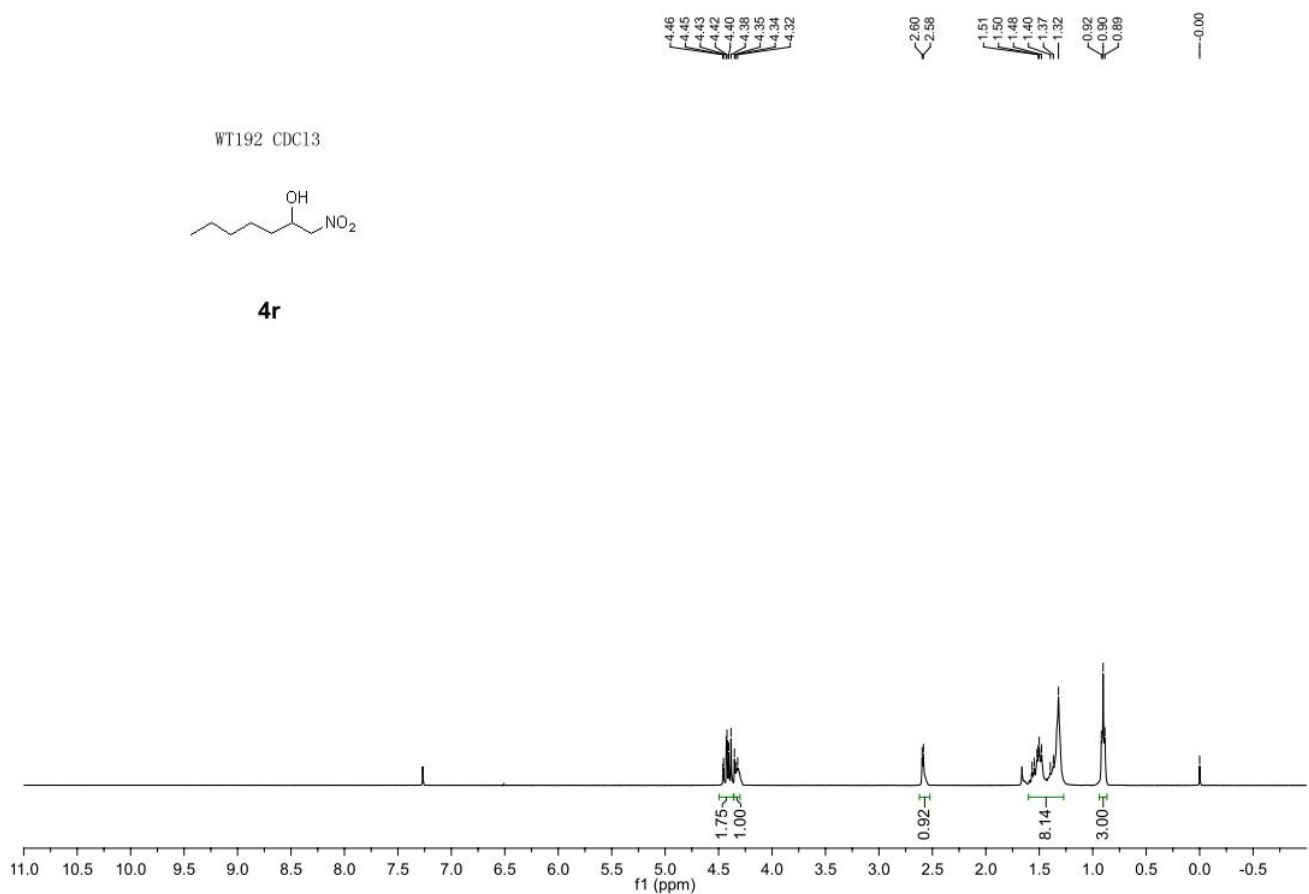


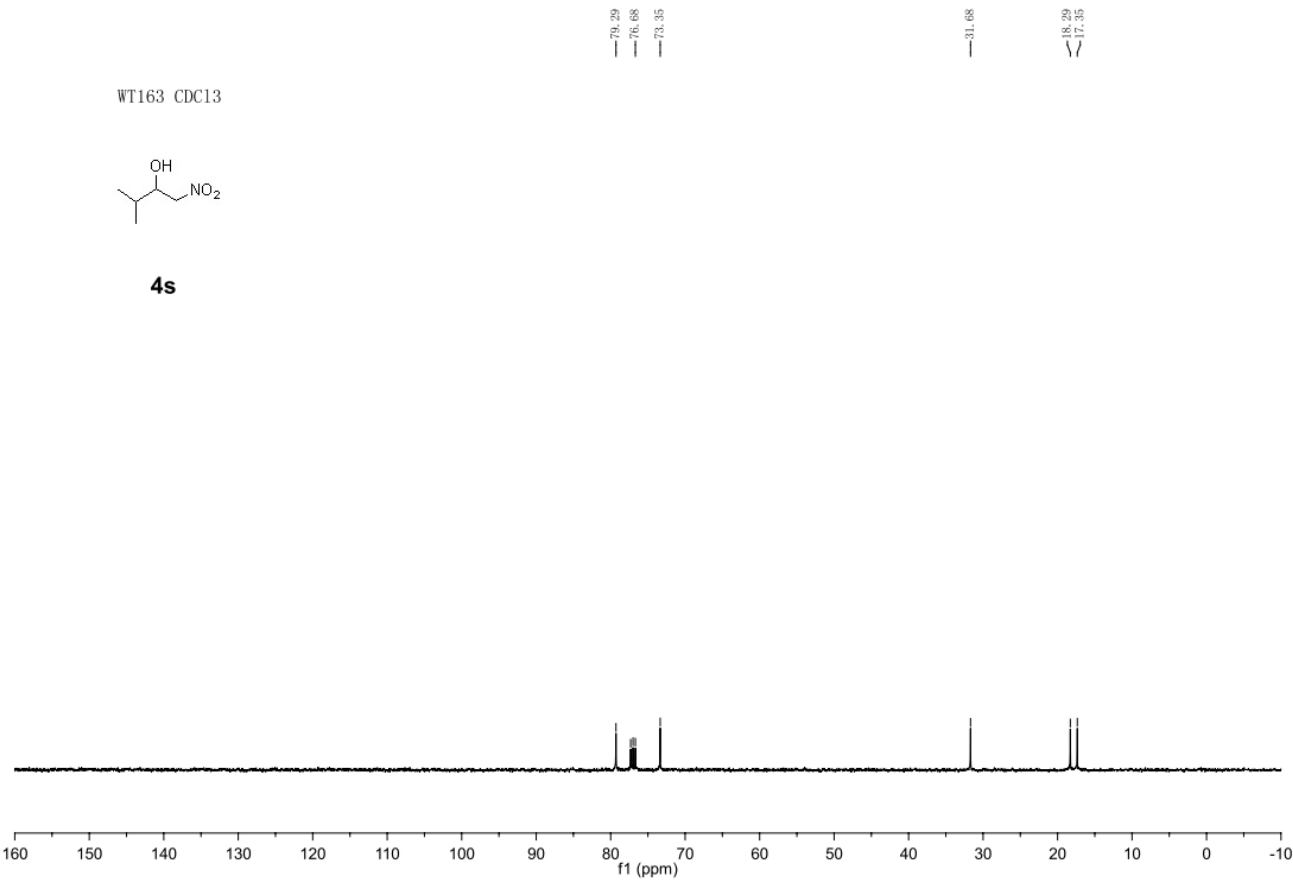
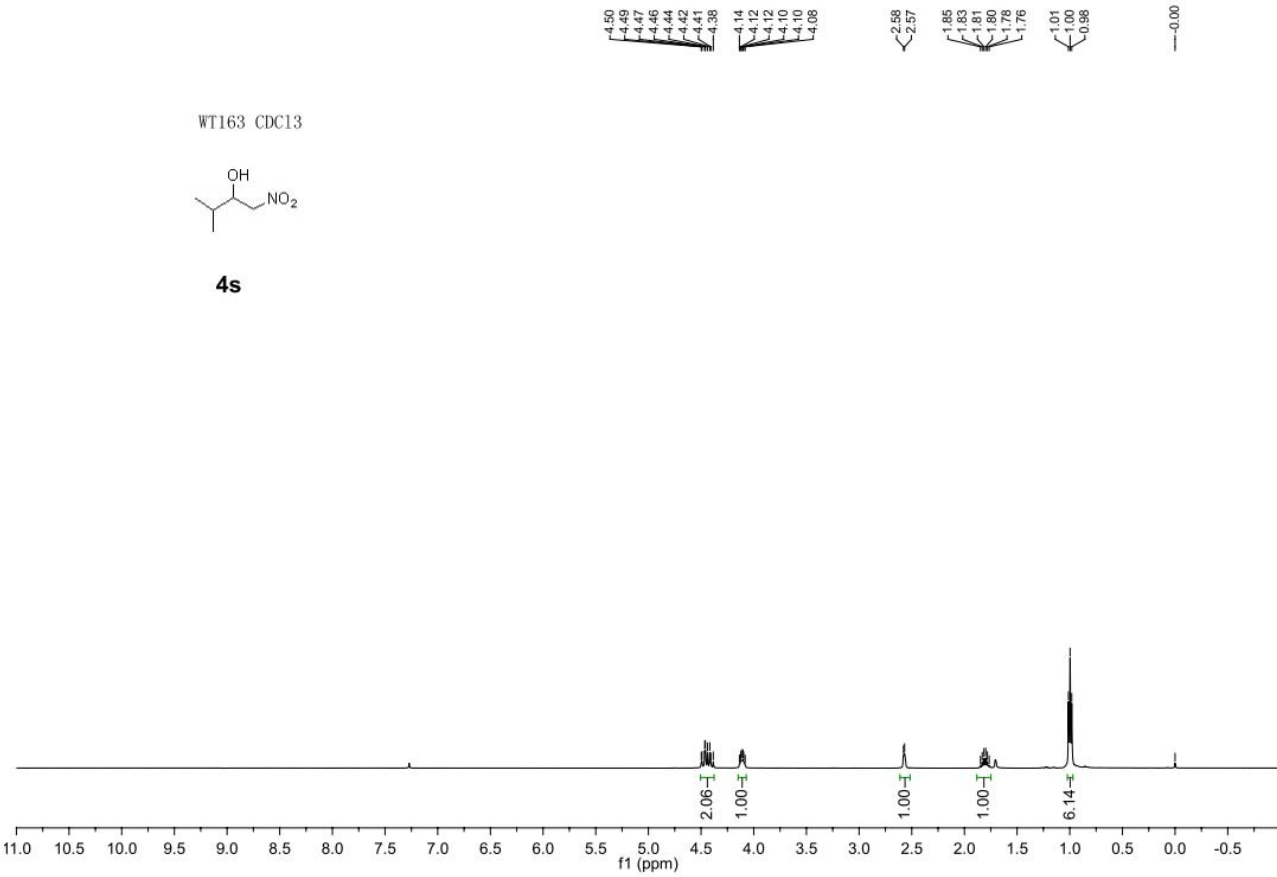


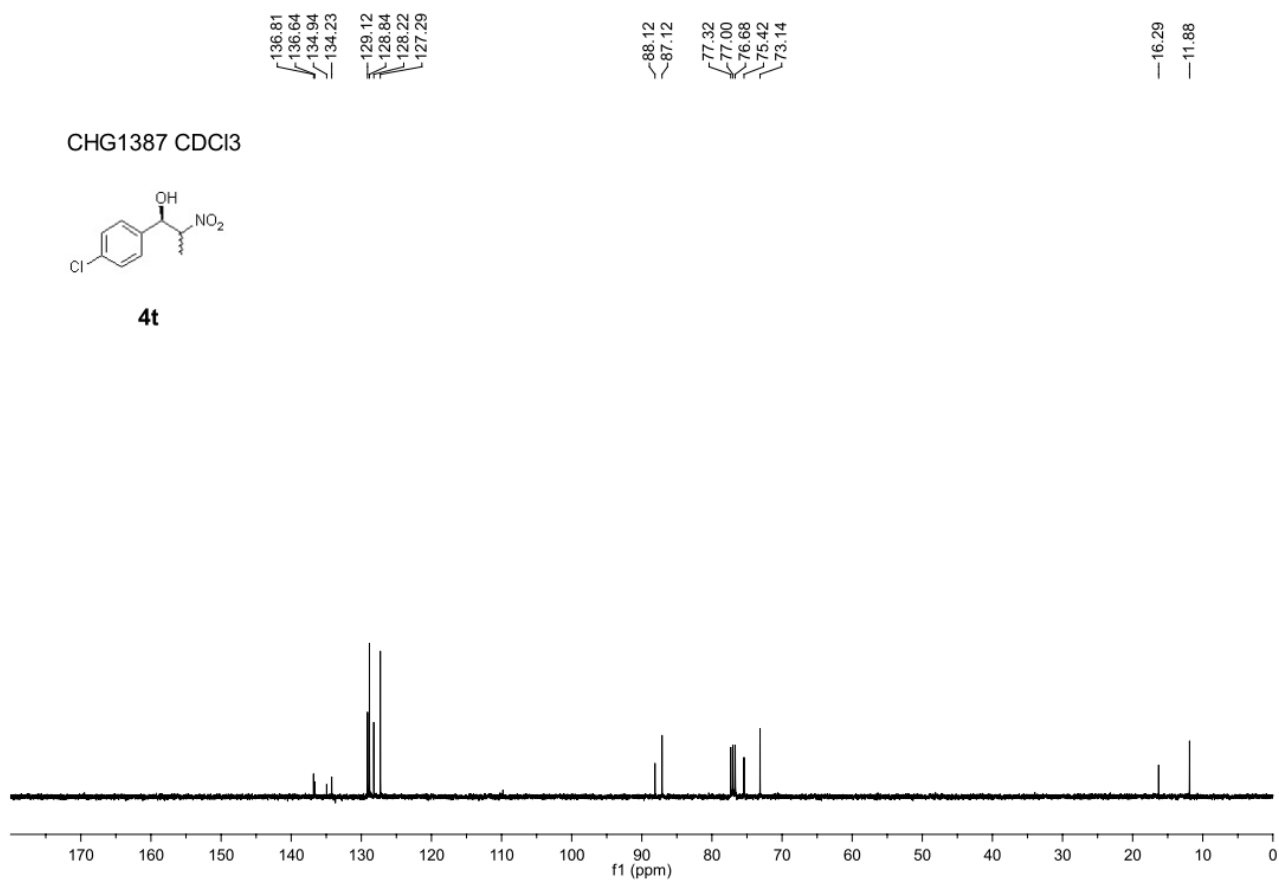
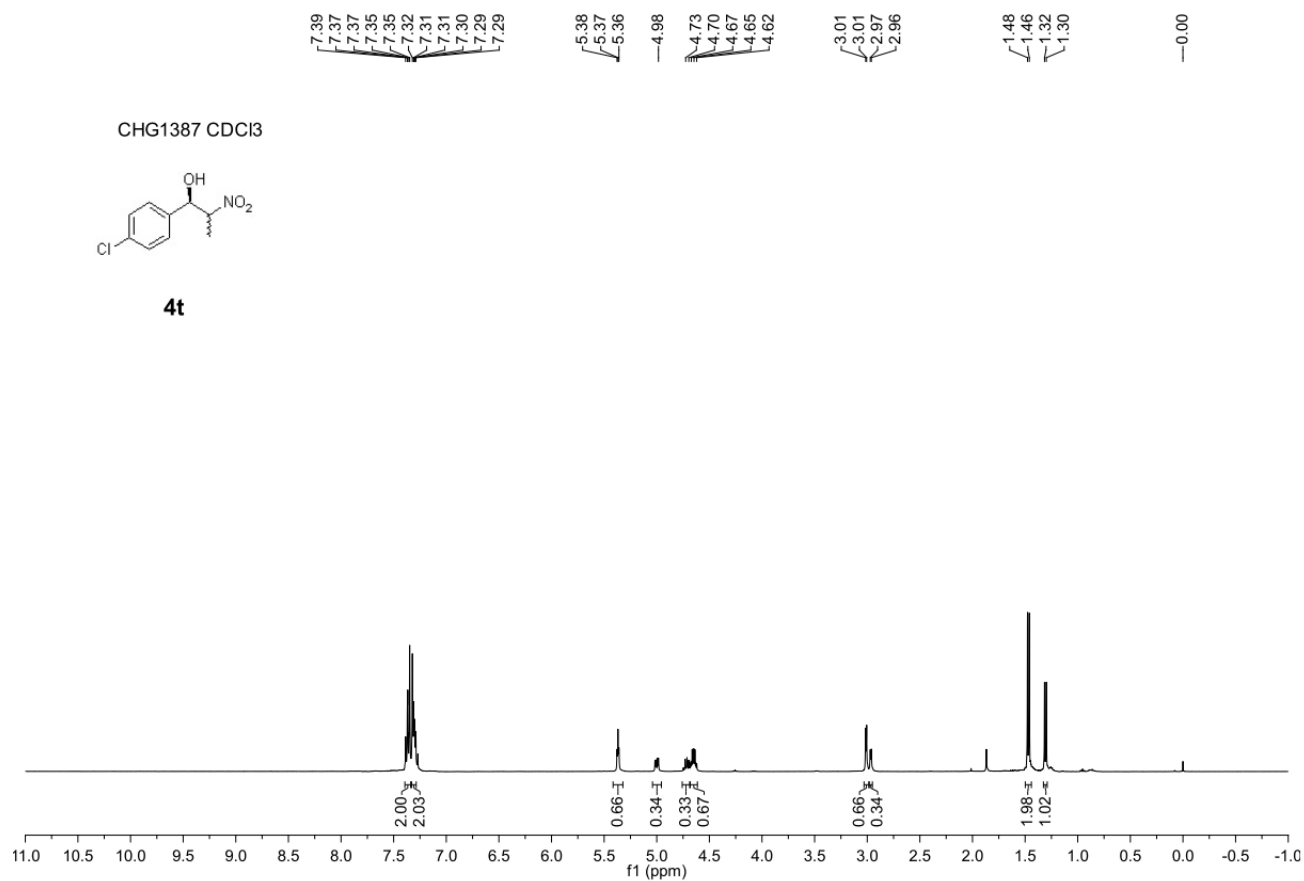




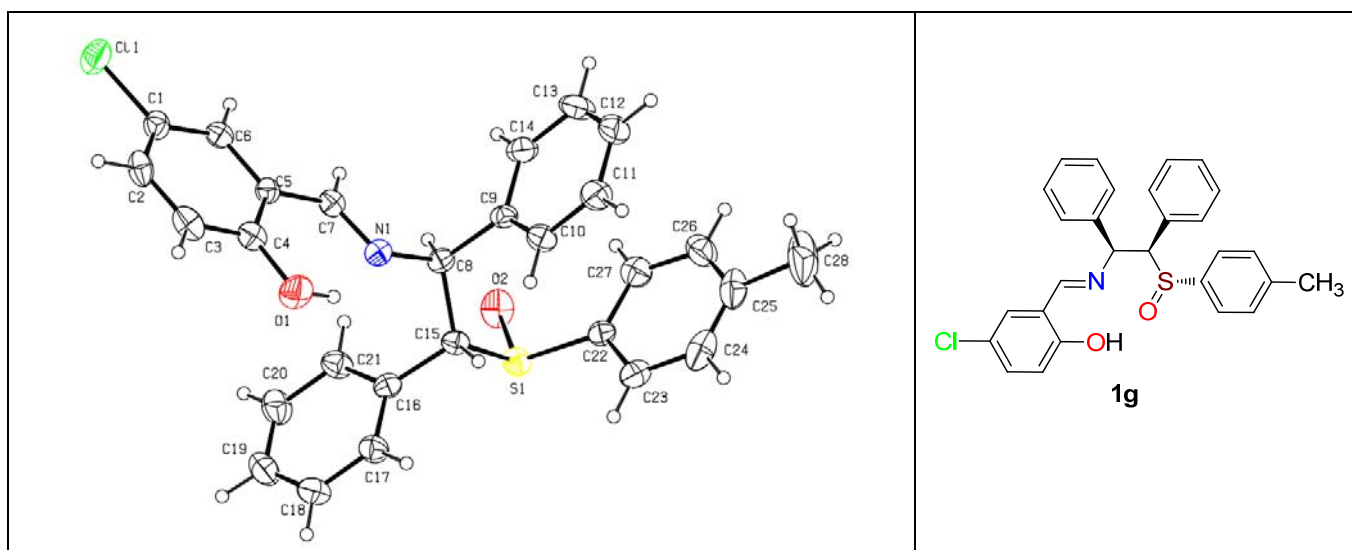




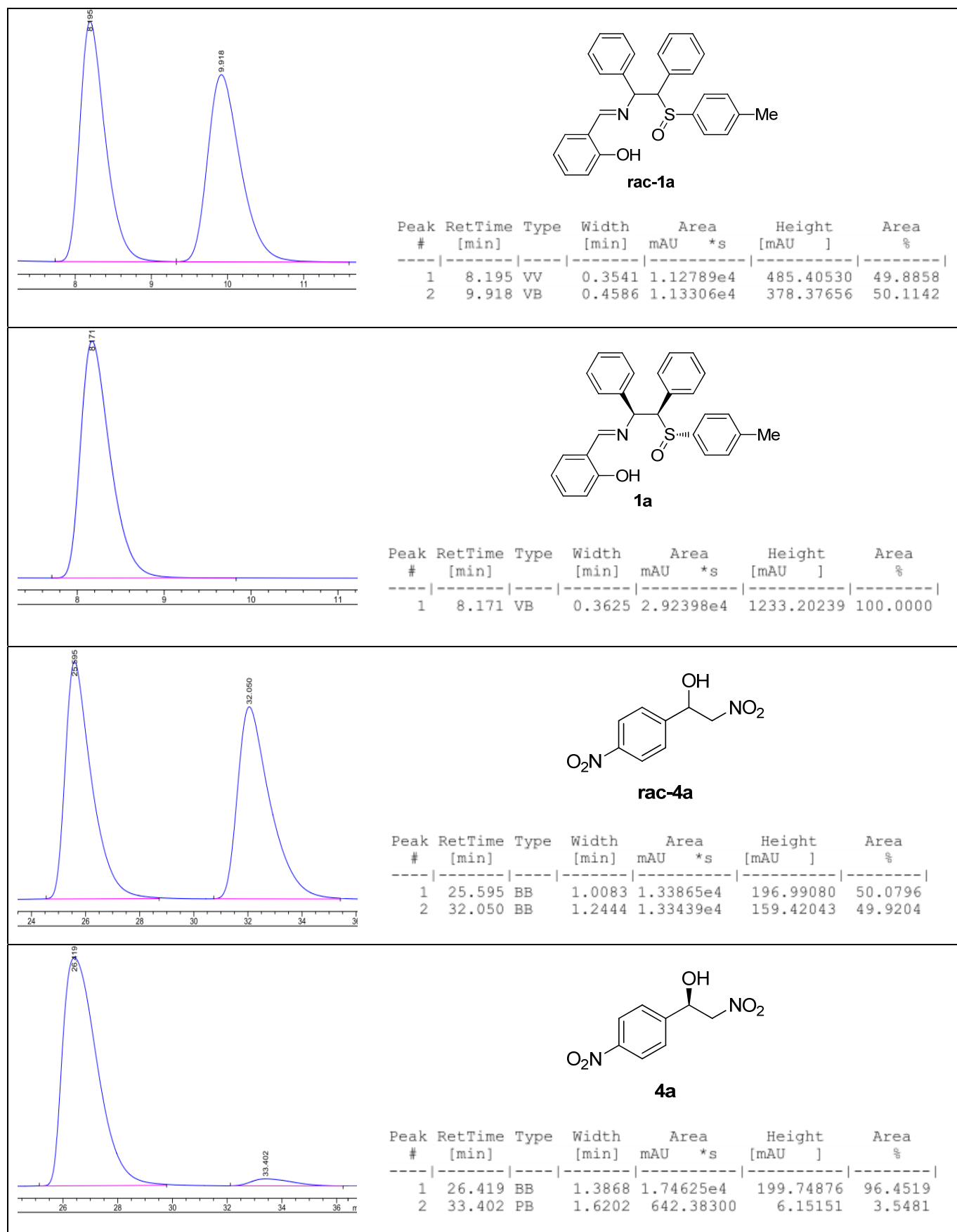


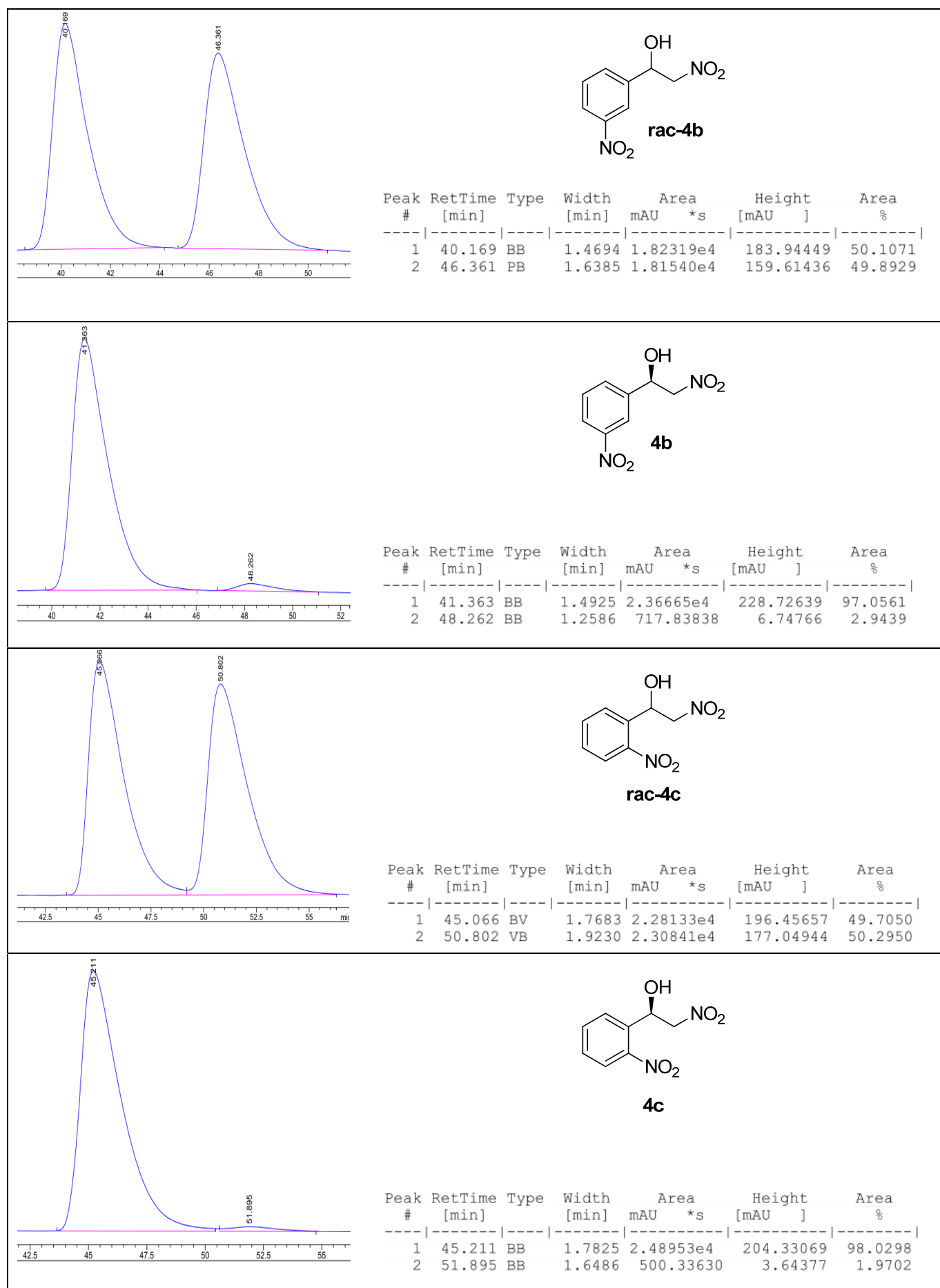


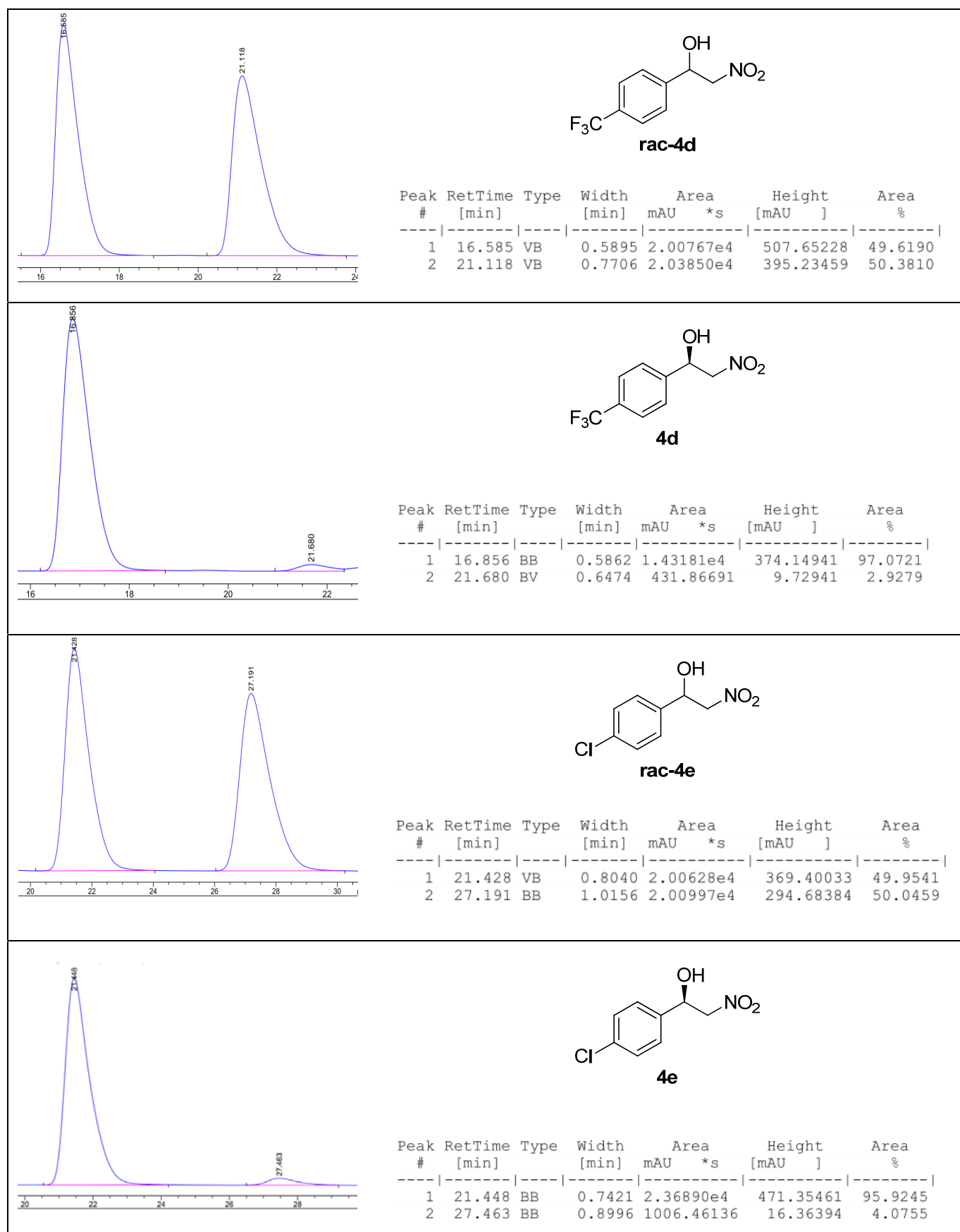
6. X-Ray Structure of Ligand 1g

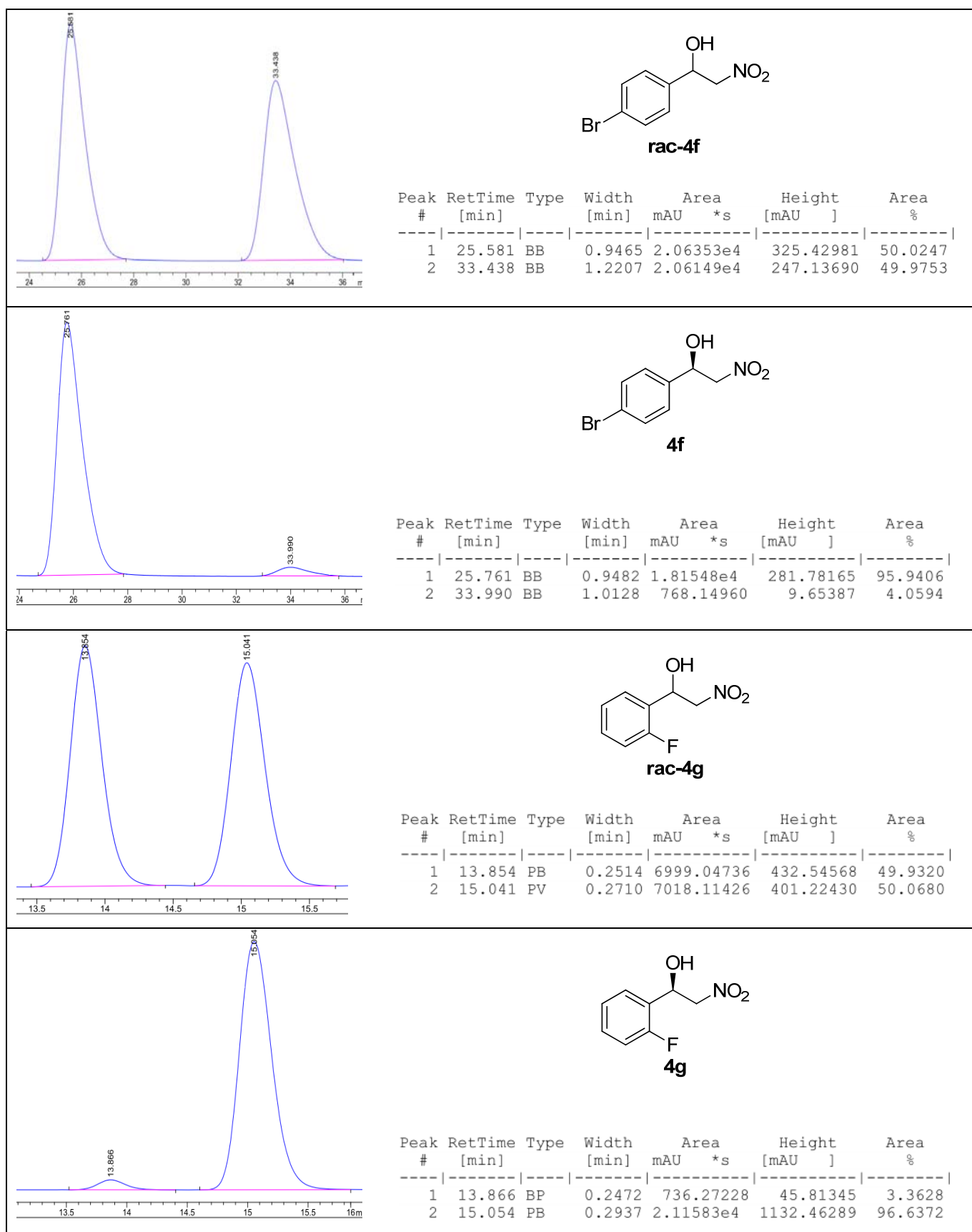


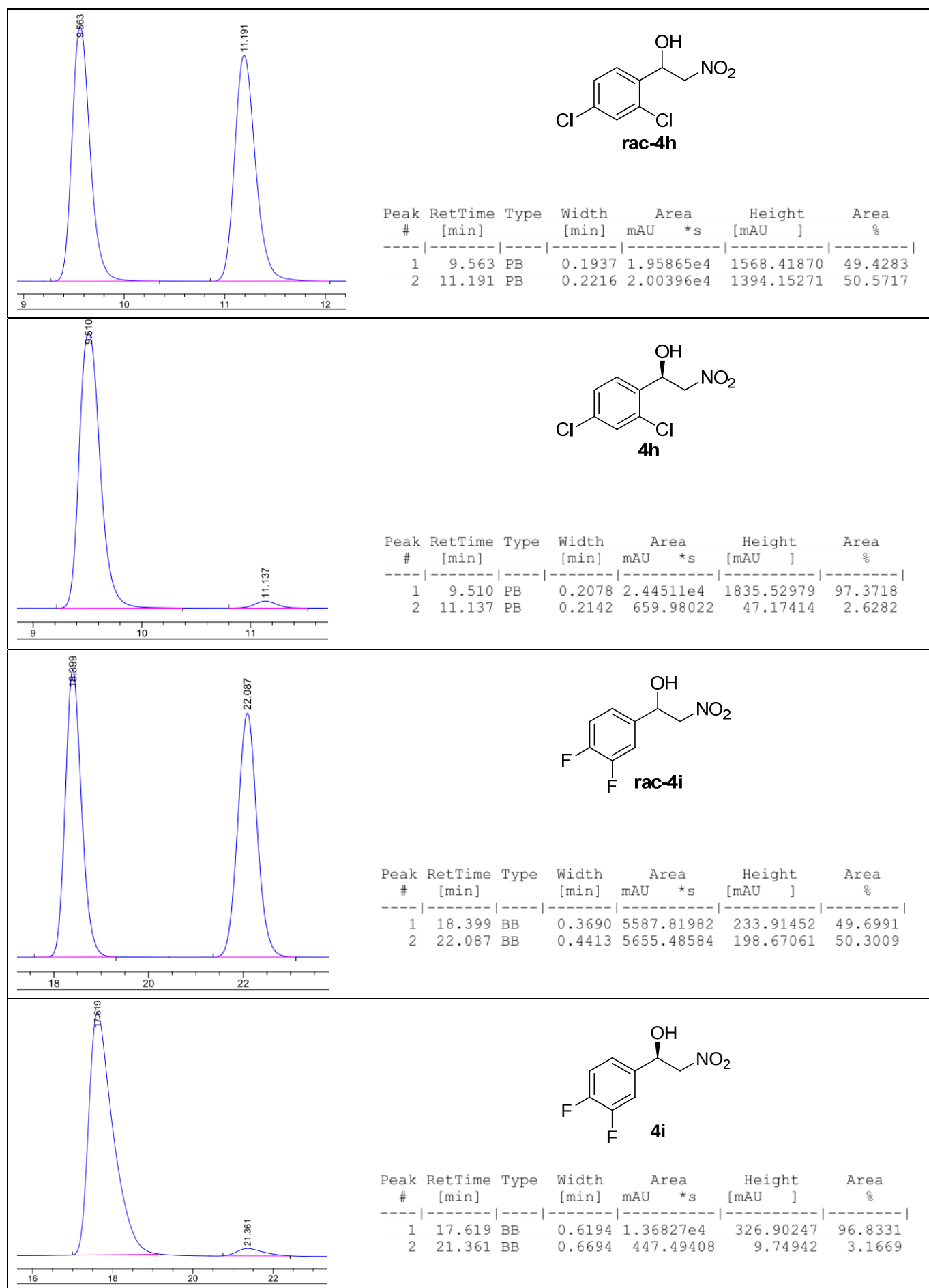
7. Copies of HPLC Chromatograms

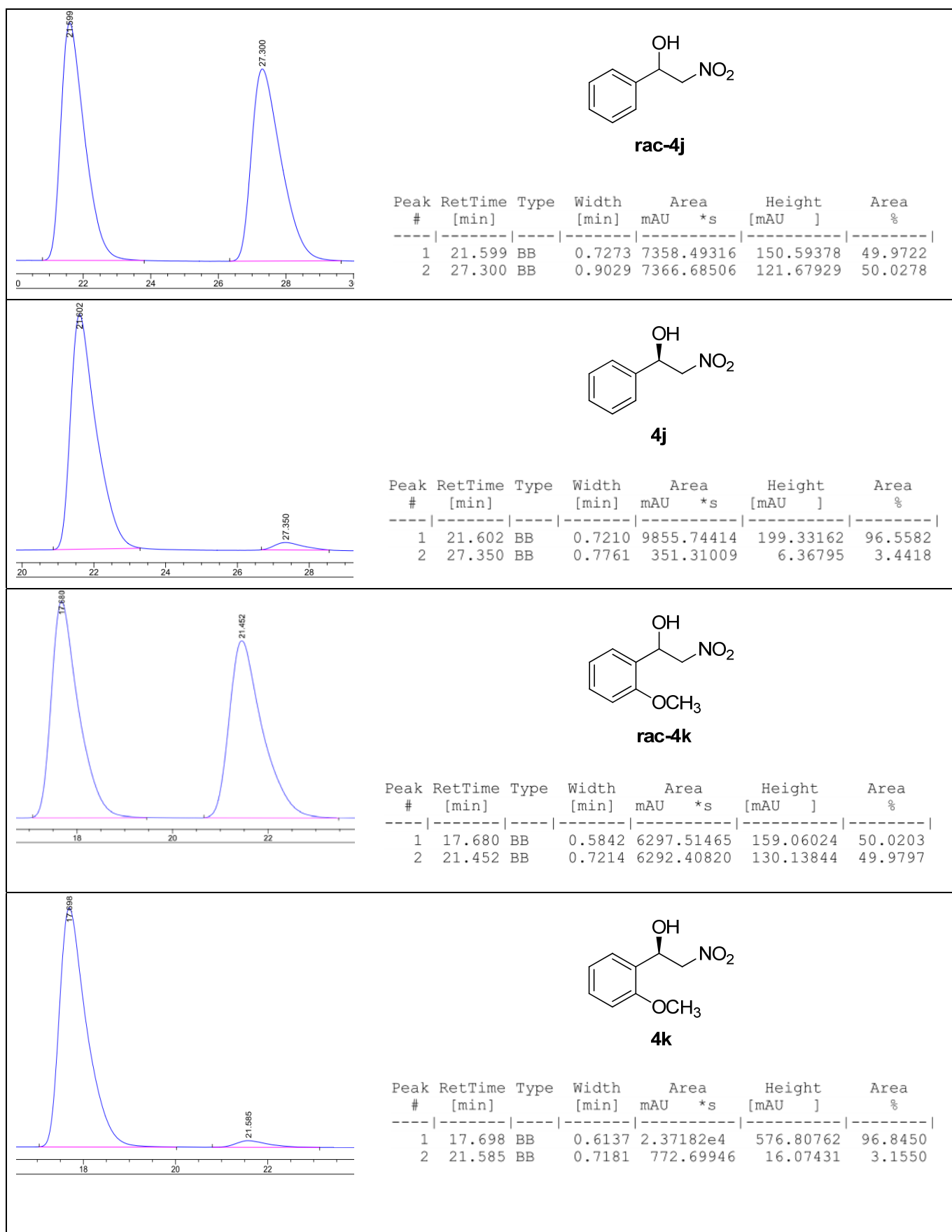


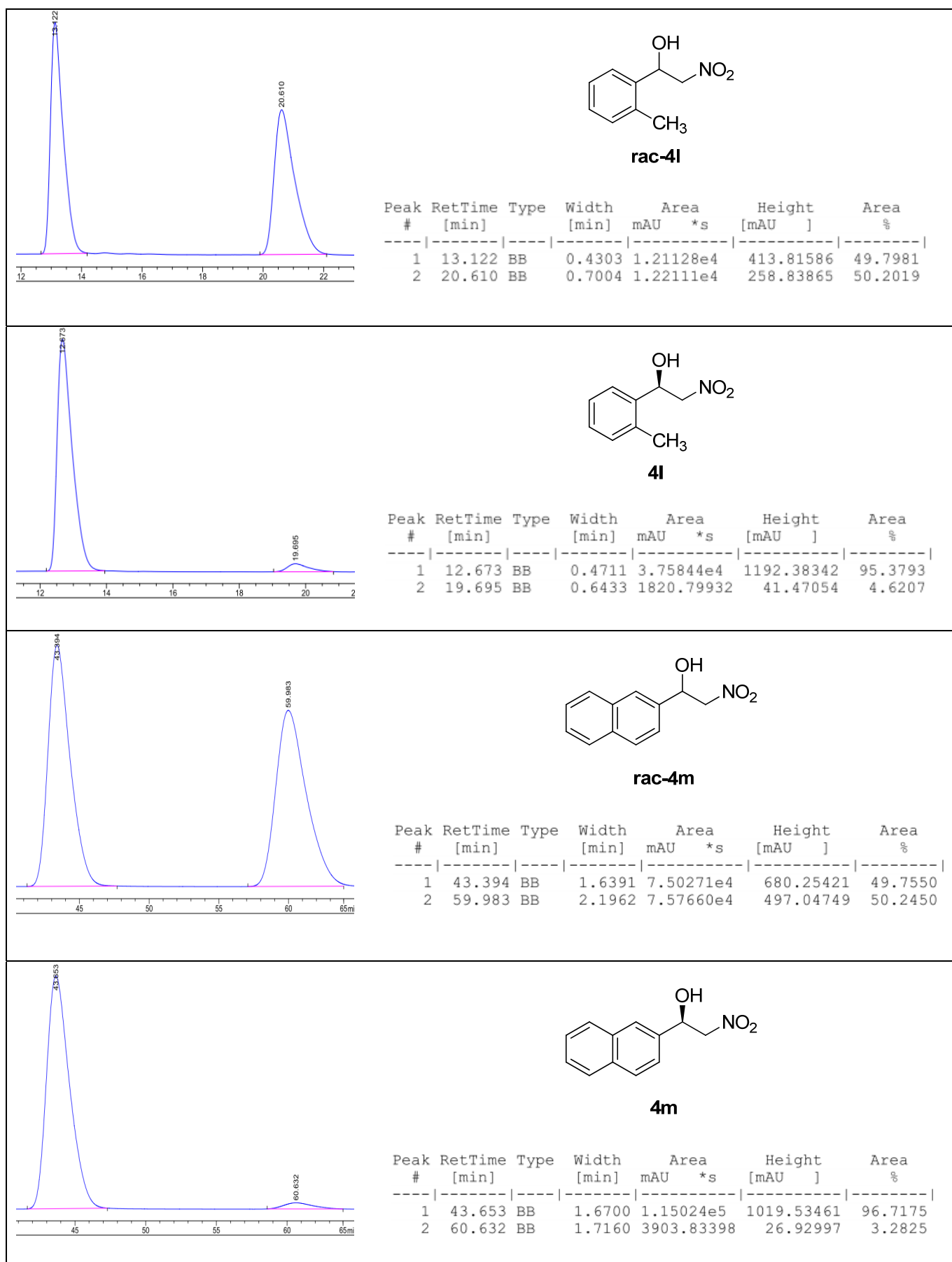


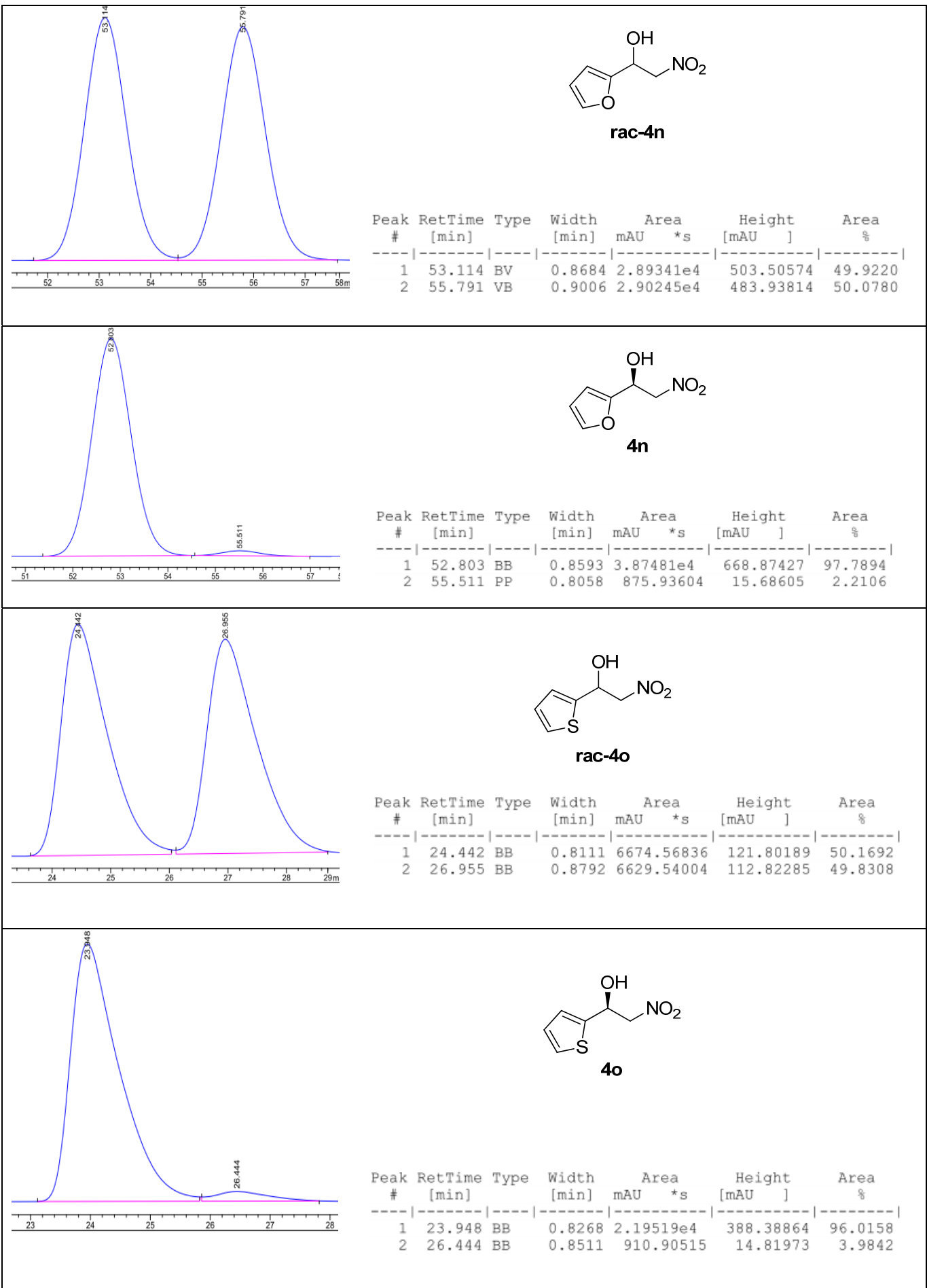


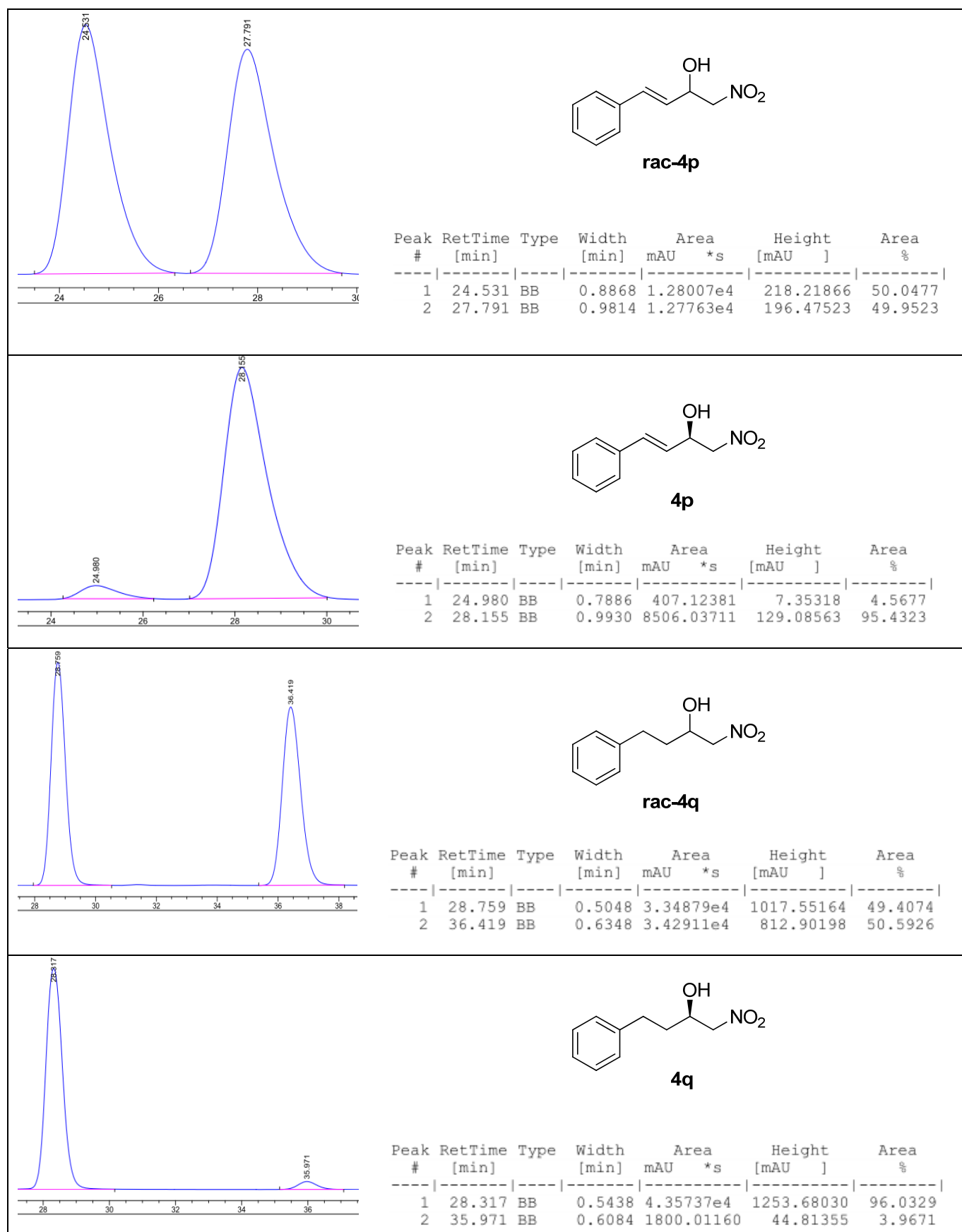


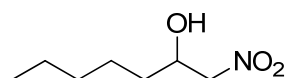
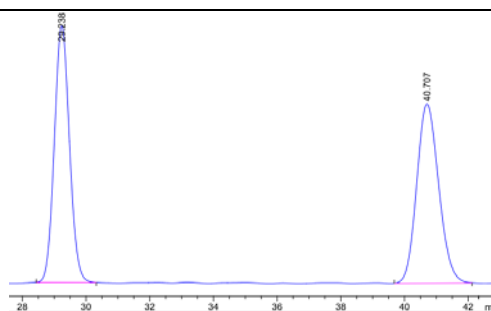






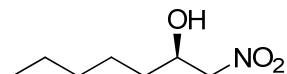
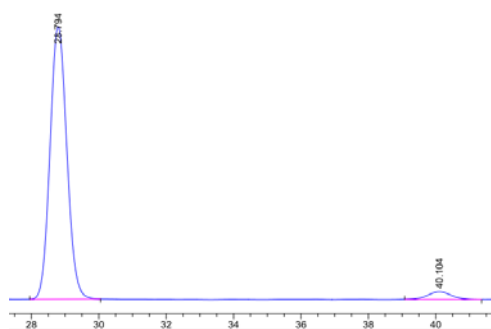






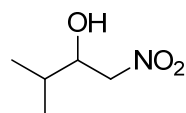
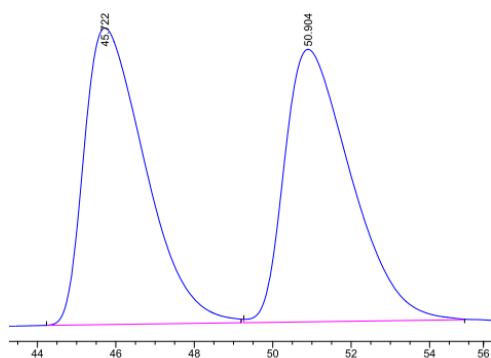
rac-4r

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	29.238	BB	0.5197	1.55708e4	470.87073	49.6969
2	40.707	BB	0.7385	1.57607e4	328.68909	50.3031



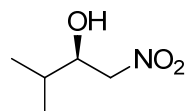
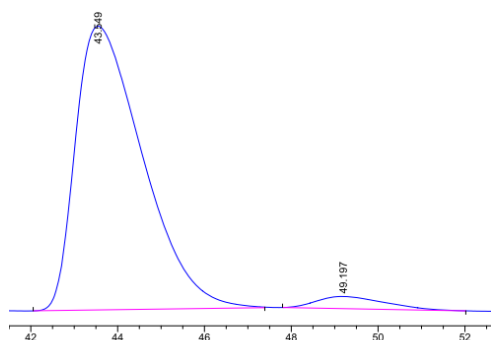
4r

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	28.794	BB	0.5420	3.19972e4	921.49512	95.9316
2	40.104	BB	0.7338	1356.98450	27.59771	4.0684



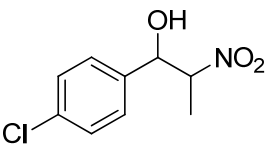
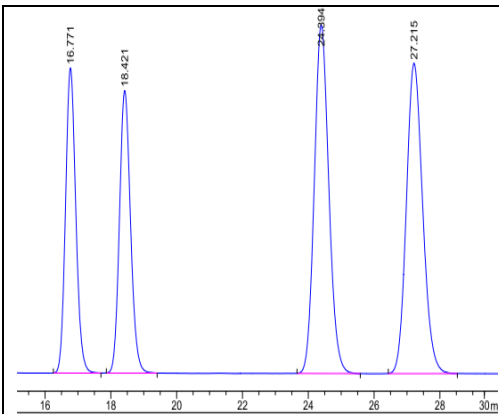
rac-4s

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	45.722	BB	1.5198	2.55413e4	236.38252	49.9692
2	50.904	BB	1.6743	2.55728e4	217.08615	50.0308



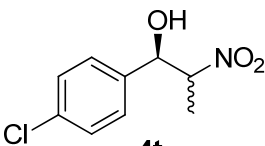
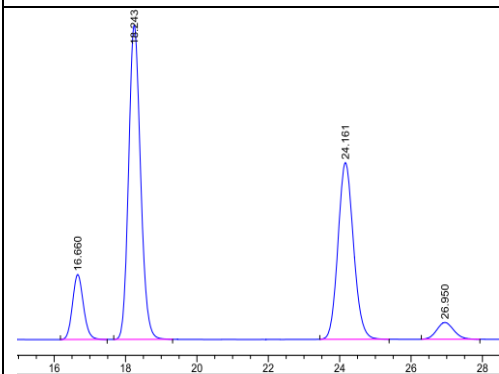
4s

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	43.549	PB	1.5540	3.38660e4	314.33649	95.8117
2	49.197	BB	1.2978	1480.42590	13.49029	4.1883



rac-4t

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	16.771	BB	0.3341	6505.79004	300.48267	19.1030
2	18.421	BB	0.3600	6502.14160	278.27350	19.0923
3	24.394	BB	0.4739	1.05168e4	343.32358	30.8806
4	27.215	BB	0.5330	1.05316e4	305.70712	30.9240



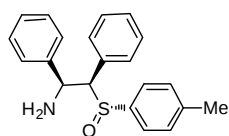
4t

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	16.660	BB	0.3269	2612.41284	122.73810	9.4990
2	18.243	BB	0.3607	1.37508e4	593.29620	49.9994
3	24.161	BB	0.4690	1.00780e4	333.52454	36.6449
4	26.950	BB	0.5153	1060.65674	31.72115	3.8567

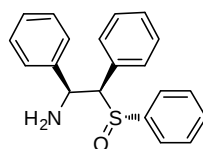
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serial number 11033032

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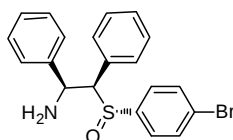
No.	Name	Weight [mg]	Content [%]	Peak Area
87	chg1278	1.2600	N: 5.264 C: 69.62 S: 12.65 H: 6.884	2390 22285 1789 4056
94	chg1215	1.3620	N: 4.446 C: 75.41 S: 9.801 H: 6.146	2179 26125 1488 3885
105	chg1151	1.4670	N: 4.539 C: 74.77 S: 9.752 H: 5.818	2400 27912 1599 3978
106	chg1068	1.3180	N: 3.778 C: 71.63 S: 9.291 H: 6.237	1785 24000 1360 3800
113	chg1157	1.1990	N: 3.389 C: 60.11 S: 8.263 H: 4.567	1450 18277 1089 2252
115	chg1318	1.2940	N: 3.791 C: 72.00 S: 9.056 H: 5.976	1758 23682 1299 3525
117	chg1132	1.2140	N: 4.131 C: 75.52 S: 9.341 H: 6.910	1798 23300 1255 3895
119	chg1225	1.2170	N: 3.642 C: 77.60 S: 8.892 H: 5.590	1585 24007 1195 3000
121	chg1050	1.4220	N: 4.597 C: 79.05 S: 10.14 H: 6.429	2355 28611 1612 4321



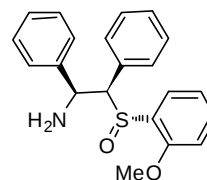
CHG1215, **13a**



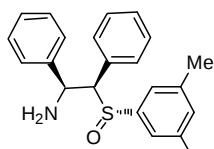
CHG1151, **13b**



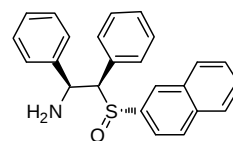
CHG1157, **13c**



CHG1318, **13e**



CHG1132, **13d**



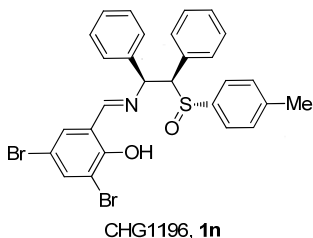
CHG1225, **13g**

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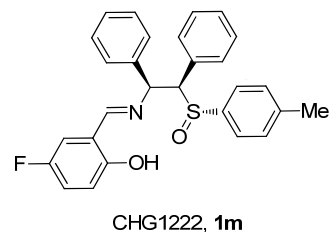
22.07.11

No.	Name	Weight [mg]	Content [%]
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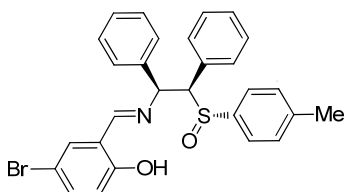
85	chg1196	1.3860	N: 2.300 C: 56.53 S: 5.424 H: 4.124
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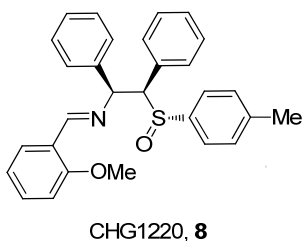
88	chg1222	1.1510	N: 2.939 C: 73.42 S: 7.206 H: 5.555
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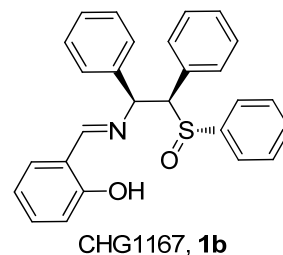
89	chg1221	1.0470	N: 2.611 C: 64.81 S: 6.438 H: 4.942
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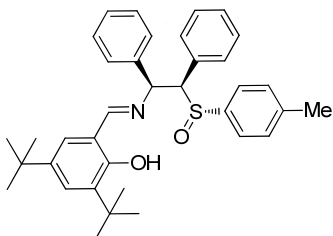
90	chg1234	1.3920	N: 2.129 C: 58.56 S: 5.313 H: 3.759
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91	chg1220	1.1250	N: 2.934 C: 76.88 S: 7.349 H: 6.239
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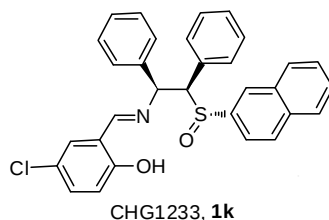


92	chg1167	1.3200	N: 3.244 C: 76.29 S: 7.656 H: 5.705
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95	chg1203	1.1340	N: 2.362 C: 78.55 S: 6.096 H: 7.663
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97	chg1218	1.0850	N: 2.694 C: 78.80 S: 6.841 H: 5.832
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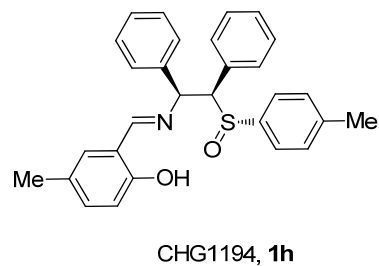
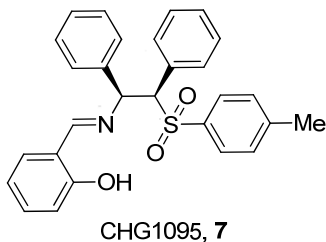
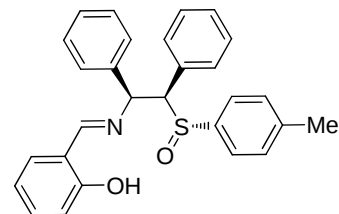
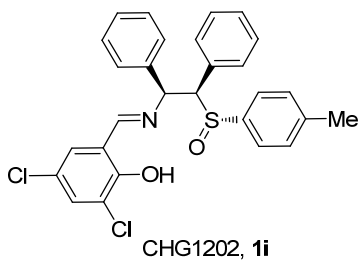


98	chg1233	1.1540	N: 2.661 C: 72.87 S: 6.320 H: 5.025
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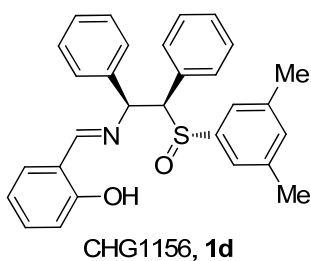
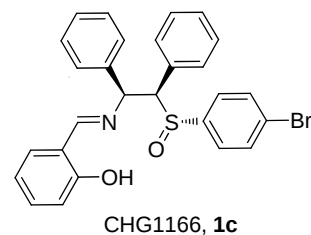
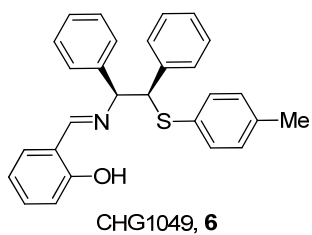
No.	Name	Weight [mg]	Content [%]
86	chg1202	1.2210	N: 2.696 C: 66.01 S: 6.521 H: 4.375
93	chg1155	1.2350	N: 3.068 C: 76.29 S: 7.461 H: 6.025
96	chg1095	1.1130	N: 2.980 C: 73.83 S: 7.245 H: 5.612
101	chg1194	1.3120	N: 2.963 C: 76.90 S: 6.880 H: 5.999
103	chg1219	1.2650	N: 2.859 C: 73.56 S: 7.123 H: 5.507



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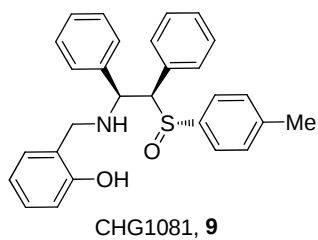
No.	Name	Weight [mg]	Content [%]
99	chg1049	1.3650	N: 3.179 C: 79.61 S: 7.802 H: 6.064
102	chg1166	1.1780	N: 2.581 C: 63.99 S: 6.374 H: 4.689
104	chg1239	1.3120	N: 2.745 C: 78.27 S: 6.716 H: 5.862
106	chg1156	1.3180	N: 3.008 C: 76.63 S: 7.170 H: 6.178



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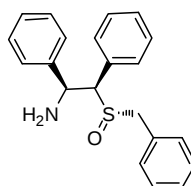
No. Name	Weight [mg]	Content [%]
41 chg1081	1.2820	N: 3.121 C: 76.28 S: 7.271 H: 6.412



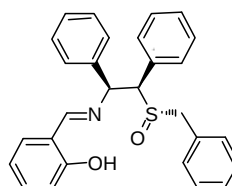
analytic functional testing
 VarioEL III CHNS
 serial number 11033032

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No.	Name	Weight [mg]	Content [%]	Peak Area
45	chg1396	1.4640	N: 4.029 C: 75.21 S: 9.664 H: 6.254	2112 28714 1630 3896
46	chg1385	1.7150	N: 2.471 C: 82.33 S: 6.233 H: 5.687	1507 36875 1217 4205
47	chg1434	1.2620	N: 3.448 C: 76.67 S: 7.104 H: 5.455	1548 25212 1011 2721



CHG1396, **13f**



CHG1434, **1f**

Shanghai Mass Spectrometry Center
Shanghai Institute of Organic Chemistry
Chinese Academic of Sciences
High Resolution MS DATA REPORT



Instrument: IonSpec 4.7 Tesla FTMS

Card Serial Number : I11 3028

Sample Serial Number: CHG1217

Operator : HuaQin Date: 2011/10/31

Operation Mode: MALDI/DHB

Elemental Composition Search Report:

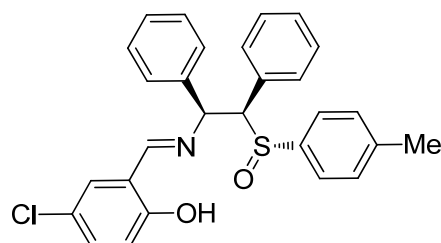
Target Mass:

Target m/z = 474.1283 ± 0.002

Charge = +1

Possible Elements:

Element:	Exact Mass:	Min:	Max:
C	12.000000	0	100
H	1.007825	0	100
N	14.003074	0	3
O	15.994915	0	3
S	31.972071	0	1
Cl	34.968853	0	1



C₂₈H₂₄ClNO₂S, CHG1217, **1g**

Additional Search Restrictions:

DBE Limit Mode = Both Integer and Half-Integer

Minimum DBE = 0

Search Results:

Number of Hits = 4

m/z	Delta m/z	DBE	Formula
474.12842	-0.00012	21.0	C ₃₁ H ₂₂ O ₃ S ⁺¹
474.12773	0.00057	30.5	C ₃₇ H ₁₆ N ⁺¹
474.12890	-0.00060	17.0	C ₂₈ H ₂₅ NO ₂ SCl ⁺¹
474.12707	0.00123	21.5	C ₂₉ H ₂₀ N ₃ O ₂ S ⁺¹

Shanghai Mass Spectrometry Center
Shanghai Institute of Organic Chemistry
Chinese Academic of Sciences
High Resolution MS DATA REPORT



Instrument: IonSpec 4.7 Tesla FTMS

Card Serial Number : I11 3030

Sample Serial Number: CHG1245

Operator : HuaQin Date: 2011/10/31

Operation Mode: MALDI/DHB

Elemental Composition Search Report:

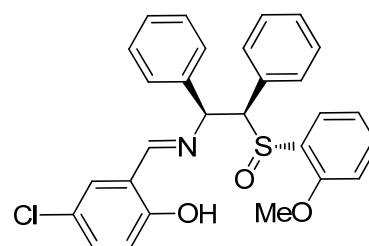
Target Mass:

Target m/z = 490.1235 ± 0.002

Charge = +1

Possible Elements:

Element:	Exact Mass:	Min:	Max:
C	12.000000	0	100
H	1.007825	0	100
N	14.003074	0	3
O	15.994915	0	3
S	31.972071	0	1
Cl	34.968853	0	1



C₂₈H₂₄ClNO₃S, CHG1245, **1e**

Additional Search Restrictions:

DBE Limit Mode = Both Integer and Half-Integer

Minimum DBE = 0

Search Results:

Number of Hits = 4

m/z	Delta m/z	DBE	Formula
490.12382	-0.00032	17.0	C ₂₈ H ₂₅ NO ₃ SCl ⁺¹
490.12313	0.00037	26.5	C ₃₄ H ₁₉ N ₂ Cl ⁺¹
490.12264	0.00086	30.5	C ₃₇ H ₁₆ NO ⁺¹
490.12199	0.00151	21.5	C ₂₉ H ₂₀ N ₃ O ₃ S ⁺¹