Supplementary Information for Catalyric Asymmetric Mannich-Type Reactions of α-Cyano α-Sulfonyl Carbanions

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General Information: Infrared spectra were recorded on a Shimadzu IRAffinity-1 spectrometer. ¹H NMR spectra were recorded on a JEOL JNM-ECS400 (400 MHz) spectrometer. Chemical shifts are reported in ppm from the tetramethylsilane (0.0 ppm) resonance as the internal standard (CDCl₃). Data are reported as follows: chemical shift, integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and br = broad) and coupling constants (Hz). ¹³C NMR spectra were recorded on a JEOL JNM-ECS400 (101 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from the solvent resonance as the internal standard (CDCl₃; 77.16 ppm). Optical rotations were measured on a HORIBA SEPA-500 polarimeter. The high resolution mass spectra were conducted on Thermo Fisher Scientific Exactive. Analytical thin layer chromatography (TLC) was performed on Merck precoated TLC plates (silica gel 60 GF254, 0.25 mm). Flash column chromatography was performed on PSQ60AB (spherical, av. 55 µm; Fuji Silysia Chemical ltd.). Enantiomeric excesses were determined by HPLC analysis using chiral columns (φ 4.6 mm x 250 mm, DAICEL CHIRALCEL OD-H (ODH), CHIRALCEL OD-3 (OD3), CHIRALPAK AD-H (ADH), CHIRALPAK AD-3 (AD3), CHIRALPAK AY-3 (AY3), CHIRALPAK IC (IC), CHIRALPAK IA (IA), and CHIRALPAK AS-H (ASH)) with hexane (H) isopropyl alcohol (IPA) and ethanol (EtOH), as eluent.

Toluene was supplied from Kanto Chemical Co., Inc. as "Dehydrated" and further purified by both A2 alumina and Q5 reactant using a GlassContour solvent dispensing system. *N*-Boc imines¹, and 1,2,3-triazolium salts $1 \cdot X^2$ were synthesized by following the literature methods. α -Cyano- α -sulfones were prepared via monoalkylation reactions of aryl- or alkylsulfonyl acetonitriles.³

¹ (a) Wenzel, A. G.; Jacobsen, E. N. J. Am. Chem. Soc. 2002, 124, 12964. (b) Song, J.; Wang, Y.; Deng, L. J. Am. Chem. Soc. 2006, 128, 6048.

² Ohmatsu, K.; Kiyokawa, M.; Ooi, T. J. Am. Chem. Soc. 2011, 133, 1307.

³ Ono, N.; Tamura, R.; Tanikaga, R.; Kaji, A. Synthesis 1977, 690.

Experimental Section:

Characterization of 1,2,3-Triazolium Bromide:



1a Br: ¹H NMR (400 MHz, CDCl₃) δ 10.25 (1H, brs), 8.71 (1H, brs), 8.25 (2H, brs), 7.83 (2H, brs), 7.67 (2H, m), 7.52-7.36 (8H, m), 7.29-7.05 (12H, m), 6.89 (2H, brs), 6.64 (2H, d, J = 7.3 Hz), 5.04 (2H, brs), 1.86 (2H, brm), 1.07 (3H, d, J = 6.4Hz), 0.91 (1H, m), 0.63 (3H, d, J = 6.9 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 167.9, 142.7, 140.4, 138.5, 133.5, 132.3, 132.1, 131.8, 130.8, 130.5, 129.7, 129.3, 129.0, 128.9, 128.7,

128.6, 128.5, 128.4, 128.3, 128.1, 127.7, 127.6, 127.1, 69.1, 68.7, 55.6, 38.4, 25.5, 23.2, 22.0, five peaks for aromatic carbons were not found probably due to overlapping; IR 3030, 2957, 1674, 1519, 1485, 1287, 1146 cm⁻¹; HRMS (ESI) Calcd for $C_{46}H_{43}N_4O^+$ ([M–Br]⁺) 667.3431. Found 667.3429.; $[\alpha]_D^{19} = +35.4$ (c = 1.0, CHCl₃).



1b·Br: ¹H NMR (400 MHz, CDCl₃) δ 9.74 (1H, brs), 8.52 (1H, brs), 8.19 (2H, brd, J =6.0 Hz), 7.83 (1H, br), 7.73 (1H, br), 7.68 (1H, t, *J* = 7.8 Hz), 7.53 (1H, t, *J* = 7.8 Hz), 7.48 (1H, d, J = 7.8 Hz), 7.43-7.35 (8H, m), 7.30-7.25 (5H, m), 7.20 (3H, br), 7.11 (2H, t, J = 7.8 Hz), 6.90 (2H, d, J = 7.3 Hz), 6.81 (2H, d, J = 8.2 Hz), 5.45-5.15 (2H, brm), 1.87 (1H, dd, J = 14.6, 9.6 Hz), 1.73 (1H, brm), 1.06 (3H, d, J = 6.4 Hz), 0.88 (1H, m), $(Ar^1 = p - CF_3 - C_6H_4)$ 0.60 (3H, d, J = 6.4 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 168.0, 142.6, 141.0, 140.9, 138.4, 136.1, 134.2, 133.6, 132.8, 132.6, 132.2, 131.9, 131.8 (q, J_{CF} = 33.5 Hz), 130.9, 129.4, 129.3, 129.0, 128.8, 128.7, 128.5, 128.3, 128.2, 127.8, 127.4, 125.9 (q, $J_{C-F} = 3.8$ Hz), 123.5 (q, $J_{C-F} = 274.0$ Hz), 119.9, 69.1, 68.8, 55.0, 38.6, 25.6, 23.2, 21.9, three peaks for aromatic carbons were not found probably due to overlapping; IR 3059, 3003, 1672, 1568, 1531, 1325, 1128 cm⁻¹; HRMS (ESI) Calcd for C₄₇H₄₂F₃N₄O⁺ $([M-Br]^+)$ 735.3305. Found 735.3302.; $[\alpha]_D^{19} = +40.6$ (c = 0.7, CHCl₃).



1c Br: ¹H NMR (400 MHz, CDCl₃) δ 10.12 (1H, brs), 8.39 (1H, brs), 7.79 (2H, brs), 7.72-7.69 (4H, m), 7.56-7.50 (3H, m), 7.44 (2H, d, *J* = 8.4 Hz), 7.39 (2H, t, *J* = 8.4 Hz), 7.32-7.26 (3H, m), 7.16-7.12 (6H, brm), 6.93 (2H, brd, J = 6.4 Hz), 6.78 (2H, d, J = 7.6 Hz), 5.11 (2H, brs), 1.89-1.74 (2H, m), 1.07 (3H, d, J = 6.8 Hz), 0.80-0.81 (1H, brm), 0.61 (3H, d, J = 6.8 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 166.8, 142.6, 141.0, 140.4, 138.3, 136.6, 135.5, 134.0, 133.3 (q, J_{C-F} = 32.6 Hz), 132.9, 132.7, 132.0, 132.0 (q, J_{C-F} = 33.5 Hz), 131.0, 129.3, 129.1, 129.0, 129.0, 128.7, 128.5, 128.2, 127.9, 127.8,

127.2, 126.0 (q, $J_{C-F} = 3.8 \text{ Hz}$), 125.4 (q, $J_{C-F} = 3.8 \text{ Hz}$), 123.8 (q, $J_{C-F} = 274.0 \text{ Hz}$), 123.3 (q, $J_{C-F} = 274.0 \text{ Hz}$), 119.8, 69.4, 68.8, 54.8, 38.4, 25.5, 23.1, 21.8, two peaks for aromatic carbons were not found probably due to overlapping; IR 3024, 2959, 1682, 1530, 1476, 1288, 1128 cm⁻¹; HRMS (ESI) Calcd for C₄₈H₄₁F₆N₄O⁺ $([M-Br]^+)$ 803.3179. Found 803.3169.; $[\alpha]_D^{19} = +18.6$ (c = 1.0, CHCl₃).



1d Br: ¹H NMR (400 MHz, CDCl₃) δ 10.34 (1H, brs), 9.78 (1H, brs), 8.61 (2H, s), 7.93 (1H, s), 7.88 (2H, d, J = 7.8 Hz), 7.74-7.70 (1H, m), 7.55-7.51 (4H, brm), 7.45 (2H, d, J = 8.2 Hz), 7.40 (2H, t, J = 7.8 Hz), 7.33-7.24 (4H, m), 7.18-7.14 (5H, brm), 7.01 (2H, d, J = 7.8 Hz), 6.73 (2H, d, J = 7.8 Hz), 5.00 (1H, brd, J = 14.6 Hz), 4.96 (1H, brd, J = 14.6 Hz), 1.90-1.77 (2H, m), 1.09 (3H, d, J = 6.4 Hz), 0.87 (1H, m),0.63 (3H, d, J = 6.9 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 165.6, 142.4, 141.2, 140.1, 138.4, 135.9, 135.3, 133.8, 133.0, 132.8, 132.0 (q, $J_{C-F} = 33.5$ Hz), 132.0, 131.4 (q,

 $J_{C-F} = 33.5 \text{ Hz}$, 131.1, 129.2, 129.0, 128.7, 128.7, 128.5, 128.2, 128.0, 127.8, 127.2, 126.0 (q, $J_{C-F} = 3.8 \text{ Hz}$), 125.2 (q, $J_{C-F} = 3.8 \text{ Hz}$), 123.3 (q, $J_{C-F} = 274.0 \text{ Hz}$), 123.1 (q, $J_{C-F} = 274.0 \text{ Hz}$), 119.7, 69.8, 69.0, 54.6, 38.2, 25.6, 23.1, 21.8, four peaks for aromatic carbons were not found probably due to overlapping; IR 3022, 2961, 1684, 1522, 1279, 1134 cm⁻¹; HRMS (ESI) Calcd for $C_{49}H_{40}F_9N_4O^+$ ([M–Br]⁺) 871.3053. Found 871.3044.; $[\alpha]_D^{19} = +10.3 \text{ (c} = 1.0, \text{CHCl}_3).$



1e Br: ¹H NMR (400 MHz, CDCl₃) δ 9.43 (1H, brs), 8.12 (1H, brs), 8.02 (1H, brs), 7.73 (2H, brs), 7.67 (1H, t, J = 7.8 Hz), 7.67-7.59 (2H, m), 7.55 (1H, t, J = 7.8 Hz), 7.46 (1H, d, J = 7.3 Hz), 7.41 (2H, d, J = 8.2 Hz), 7.35 (2H, d, J = 7.8 Hz), 7.33-7.25 (8H, brm), 7.13 (2H, d, J = 7.8 Hz), 7.07 (1H, s), 6.88-6.81 (4H, brm), 5.50 (1H, br), 5.32 (1H, brd, *J* = 14.2 Hz), 2.34 (6H, s), 1.88 (1H, dd, *J* = 14.6, 9.6 Hz), 1.69 (1H, br), $(Ar^1 = p - CF_3 - C_6H_4)$ 1.02 (3H, d, J = 6.4 Hz), 0.90-0.83 (1H, brm), 0.59 (3H, d, J = 6.4 Hz); ¹³C NMR (101 $(Ar^2 = 3,5-Me_2-C_6H_3)$ MHz, CDCl₃) δ 168.5, 142.5, 141.0, 138.3, 138.2, 136.3, 134.2, 133.6, 133.6, 132.6, 132.2, 131.7 (q, $J_{C-F} =$ 33.5 Hz), 130.8, 129.5, 129.1, 128.9, 128.7, 128.7, 128.4, 128.1, 127.9, 127.7, 127.5, 125.8 (q, $J_{C-F} = 3.8$ Hz), 125.6, 123.4 (q, $J_{C-F} = 273.5$ Hz), 119.8, 69.0, 68.6, 55.1, 38.6, 25.5, 23.1, 21.8, 21.1, three peaks for aromatic carbons were not found probably due to overlapping; IR 3021, 2959, 1674, 1516, 1474, 1325, 1128 cm⁻¹; HRMS (ESI) Calcd for $C_{49}H_{46}F_3N_4O^+$ ([M-Br]⁺) 763.3618. Found 763.3613.; $[\alpha]_D^{19} = +14.0$ (c = 1.0, CHCl₃).



1f·Br: ¹H NMR (400 MHz, CDCl₃) δ 9.11 (1H, brs), 8.40 (1H, brs), 7.86 (1H, d, J =7.8 Hz), 7.78-7.73 (2H, m), 7.64-7.30 (16H, brm), 7.16 (2H, d, J = 7.8 Hz), 7.07 (1H, s), 5.68 (2H, br), 2.35 (6H, s), 2.06-1.99 (2H, m), 1.11 (3H, br), 0.87 (1H, m), 0.65 (3H, br); ¹³C NMR (101 MHz, CDCl₃) δ169.7, 142.2, 138.7, 138.3, 137.5, 135.3, 134.0, 133.6, 132.5, 131.8 (q, J_{C-F} = 32.6 Hz), 131.4, 130.4, 130.0, 129.7, 129.3 (q, J_{C-F} = 37.4 $(Ar^2 = 3,5-Me_2-C_6H_3)$ Hz), 128.8, 128.4, 128.3, 128.1, 128.0, 126.6, 125.8, 125.3, 123.6 (q, *J*_{C-F} = 273.0 Hz), 123.1 (q, J_{C-F} = 275.0 Hz), 125.6, 119.4, 68.6, 67.6, 57.0, 39.9, 25.8, 23.0, 21.7, 21.2, one peak for aromatic carbon was not found probably due to overlapping; IR 3013, 2959, 1670, 1497, 1491, 1319, 1128 cm⁻¹; HRMS (ESI) Calcd for $C_{44}H_{41}F_{3}N_{4}O^{+}([M-Br]^{+})$ 755.3179. Found 755.3164.; $[\alpha]_{D}^{19} = +12.8$ (c = 1.0, CHCl₃).



 $(Ar^2 = 3, 5 - Me_2 - C_6H_3)$

1g·Br: ¹H NMR (400 MHz, CDCl₃) δ9.83 (1H, brs), 8.49 (1H, brs), 7.75 (4H, br), 7.58 (2H, t, J = 8.0 Hz), 7.58-7.49 (1H, m), 7.49 (2H, d, J = 8.2 Hz), 7.41-7.34 (9H, m), 7.31 (1H, t, J = 7.3 Hz), 7.05 (1H, s), 6.99 (2H, d, J = 8.2 Hz), 5.60 (1H, br), 5.42 (1H, brd, J = 15.6 Hz), 2.34-2.27 (1H, m), 2.32 (6H, s), 2.07-1.95 (2H, m), 1.32-1.26 (1H, m), 1.16 (3H, d, *J* = 6.9 Hz), 1.02 (3H, d, *J* = 6.9 Hz), 0.79 (3H, d, *J* = 6.9 Hz), 0.74

(3H, d, J = 6.9 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 168.7, 148.6, 140.4, 138.3, 136.8, 134.6, 133.6, 133.3, 132.8, 132.3, 131.8 (q, $J_{C-F} = 33.5$ Hz), 131.2, 129.2, 128.9, 128.8, 128.2, 127.9, 127.8, 127.7, 127.0, 126.5, 126.0 (q, $J_{C-F} = 3.8$ Hz), 125.6, 123.4 (q, $J_{C-F} = 273.5$ Hz), 119.2, 69.3, 69.2, 54.9, 38.4, 30.8, 25.6, 23.1, 22.0, 21.5, 21.1, one peak for aromatic carbon was not found probably due to overlapping; IR 3021, 2963, 1670, 1508, 1489, 1325, 1128 cm⁻¹; HRMS (ESI) Calcd for C₄₆H₄₈F₃N₄O⁺ ([M–Br]⁺) 729.3775. Found 729.3763.; $[\alpha]_D^{19} = +15.3$ (c = 1.0, CHCl₃).

General Procedure for 1g·Br-Catalyzed Asymmetric Mannich-Type Reaction, and Characterization of 4:



To a test tube was added $1g \cdot Br$ (1.62 mg, 0.002 mmol), Cs_2CO_3 (0.65 mg, 0.002 mmol), α -cyanosulfone **3a** (19.5 mg, 0.1 mmol) and the tube was degassed by alternating vaccum evacuation/Ar backfill. Then toluene (500 µL) was added, and the resulting mixture was cooled to -40 °C. To this solution was added *N*-Boc imine **2a** (24.6 mg, 0.12 mmol) and stirred for 24 h. The reaction mixture was quenched by NH₄Cl, and the extractive workup was performed with EtOAc. After drying over Na₂SO₄, filtration, and removal of solvent, the resulting crude residue was purified by column chromatography (H/EtOAc = 10:1 as eluent) to afford **4a** (40.0 mg, 0.099 mmol, 99% yield) as a white solid.

4a: ¹H NMR (400 MHz, CDCl₃) δ 8.04 (2H, d, J = 8.0 Hz), 7.75 (1H, t, J = 7.4 Hz), 7.62 (2H, dd, J = 8.0, 7.4 Hz), 7.41-7.32 (5H, m), 5.46 (1H, br), 5.29 (1H, brd, J = 8.7 Hz), 1.38 (3H, s), 1.37 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 153.9, 137.0, 135.6, 135.1, 130.5, 129.4, 129.0, 129.0, 116.3, 80.7, 65.4, 57.2, 28.2, 19.9; IR 2978, 1717, 1499, 1449, 1368, 1331, 1246, 1152 cm⁻¹; HRMS (ESI) Calcd for C₂₁H₂₄N₂NaO₄S⁺ ([M+Na]⁺) 423.1349. Found 423.1346.; HPLC ASH, H/IPA/EtOH = 97:1.5:1.5, flow rate = 0.5 mL/min, λ = 210 nm, 36.7 min (major *anti* isomer), 41.3 min (major *syn* isomer), 71.2 min (minor *anti* isomer), 81.1 min (minor *syn* isomer).

4b: ¹H NMR (400 MHz, CDCl₃) δ 7.94 (2H, d, J = 8.7 Hz), 7.61 (2H, d, J = 8.7 Hz), ^{NHBoc} ^{NHBOC}

 $\begin{array}{l} \textbf{4c: }^{1}\text{H NMR (400 MHz, CDCl_{3}) } \delta 7.95 (2\text{H}, d, J = 9.2 \text{ Hz}), 7.41-7.33 (5\text{H}, m), 7.06 \\ (2\text{H}, d, J = 9.2 \text{ Hz}), 5.53 (1\text{H}, br), 5.23 (1\text{H}, d, J = 8.9 \text{ Hz}), 3.92 (3\text{H}, s), 1.38 (9\text{H}, s), \\ (\text{R}^{1} = p\text{-MeO-C}_{6}\text{H}_{4}) \\ (\text{R}^{1} = p\text{-MeO-C}_{6}\text{H}_{4}) \\ \textbf{126.4, 116.5, 114.6, 80.6, 65.3, 57.3, 55.8, 28.2, 20.0, one peak for aromatic carbon was not found probably due to overlapping; IR 2978, 1716, 1593, 1497, 1456, 1331, 1314, 1267, 1148 cm^{-1}; HRMS \\ (\text{ESI) Calcd for } \text{C}_{22}\text{H}_{26}\text{N}_{2}\text{KO}_{5}\text{S}^{+} ([\text{M}+\text{K}]^{+}) 469.1194. Found 469.1192.; HPLC ADH, H/IPA = 5:1, flow rate \\ \end{array}$

= 0.5 mL/min, λ = 210 nm, 32.6 min (minor *anti* isomer), 39.2 min (minor *syn* isomer), 45.8 min (major *anti* isomer), 67.7 min (major *syn* isomer).

4d: ¹H NMR (400 MHz, CDCl₃) δ 8.19 (2H, d, J = 8.5 Hz), 7.88 (2H, d, J = 8.5 Hz), ^{NHBoc} ^{NHBOC}

HBoc Ph $(R^1 = i$ -Pr) 4f: ¹H NMR (400 MHz, CDCl₃) δ7.46-7.39 (5H, m), 5.83 (1H, br), 5.35 (1H, brd, J = 7.8 Hz), 3.33 (1H, m), 1.59 (3H, s), 1.54 (3H, d, J = 6.4 Hz), 1.45 (3H, d, J = 6.4 Hz), 1.41 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ154.6, 136.5, 129.3, 129.2, 127.7, 117.3, 81.2, 63.9, 57.6, 56.4, 28.3, 19.0, 17.8, 16.6; IR 2980, 1699, 1506, 1489, 1456, 1364, 1315, 1246, 1163 cm⁻¹; HRMS (ESI) Calcd for C₁₈H₂₆N₂NaO₄S⁺ ([M+Na]⁺) 389.1505. Found 389.1506.; HPLC IC, H/EtOH = 95:5, flow rate = 1.0 mL/min, $\lambda = 210$ nm, 8.6 min (major *syn* isomer), 9.0 min (minor *anti* isomer), 11.6 min (minor *syn* isomer), 12.6 min (major *anti* isomer).

4g: ¹H NMR (400 MHz, CDCl₃) δ 8.10 (2H, d, *J* = 7.8 Hz), 7.76 (1H, t, *J* = 7.8 Hz), 7.64 (2H, t, *J* = 7.8 Hz), 7.40 (1H, d, *J* = 7.8 Hz), 7.26-7.22 (2H, m), 7.18 (1H, d, *J* = 8.2 Hz), 5.77 (1H, d, *J* = 9.2 Hz), 5.26 (1H, br), 2.46 (3H, s), 1.36 (9H, s), 1.34 (3H, s); ¹³C NMR (R² = o-Me-C₆H₄) (101 MHz, CDCl₃) δ 154.0, 136.5, 135.7, 135.2, 131.1, 130.8, 129.5, 128.7, 127.3, 125.8, 116.7, 80.7, 66.4, 51.2, 28.4, 19.9, 18.4, one peak for aromatic carbon was not found probably due to overlapping; IR 2978, 1717, 1506, 1489, 1449, 1368, 1331, 1246, 1153 cm⁻¹; HRMS (ESI) Calcd for C₂₂H₂₆N₂KO₄S⁺ ([M+K]⁺) 453.1245. Found 453.1241.; HPLC AY3, H/IPA = 4:1, flow rate = 1.0 mL/min, λ = 210 nm, 12.1 min (minor *syn* isomer), 16.5 min (minor *anti* isomer), 26.7 min (major *syn* isomer), 62.8 min (major *anti* isomer).

MHBoc4h: ¹H NMR (400 MHz, CDCl₃) δ 8.11 (2H, d, J = 7.8 Hz), 7.73 (1H, t, J = 7.8 Hz), R^2 R^2 R

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MHz, CDCl₃) δ156.7, 154.1, 135.6, 134.8, 130.8, 130.1, 129.2, 125.2, 121.1, 116.8, 110.9, 80.2, 66.3, 55.0, 28.3, 18.8, one peak for aromatic carbon was not found probably due to overlapping; IR 2978, 1717, 1495, 1449, 1368, 1331, 1248, 1153 cm⁻¹; HRMS (ESI) Calcd for $C_{22}H_{26}N_2KO_5S^+$ ([M+K]⁺) 469.1194. Found 469.1192.; HPLC IA, H/IPA = 10:1, flow rate = 1.0 mL/min, λ = 210 nm, 16.2 min (major syn isomer), 17.4 min (minor svn isomer), 20.0 min (major anti isomer), 34.0 min (minor anti isomer).

4i: ¹H NMR (400 MHz, CDCl₃) δ 8.09 (2H, d, J = 7.8 Hz), 7.78 (1H, t, J = 7.8 Hz), 7.66 NHBoc (2H, t, *J* = 7.8 Hz), 7.53 (1H, d, *J* = 7.8 Hz), 7.40-7.25 (3H, m), 5.99 (1H, d, *J* = 8.2 Hz), CN SO₂Ph 5.53 (1H, br), 1.43 (3H, s), 1.37 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ153.7, 135.3, 135.3, $(R^2 = o-CI-C_6H_4)$ 133.8, 130.7, 130.0, 129.5, 129.3, 127.9, 127.1, 115.9, 81.1, 65.9, 52.3, 28.2, 18.3, one peak

for aromatic carbon was not found probably due to overlapping; IR 2978, 1705, 1506, 1449, 1368, 1333, 1246, 1155 cm⁻¹; HRMS (ESI) Calcd for $C_{21}H_{23}ClN_2KO_4S^+$ ([M+K]⁺) 473.0699. Found 473.0698.; HPLC ADH, H/IPA = 4:1, flow rate = 1.0 mL/min, λ = 254 nm, 9.0 min (major syn isomer), 10.4 min (minor syn isomer), 11.8 min (major anti isomer), 38.1 min (minor anti isomer).

4j: ¹H NMR (400 MHz, CDCl₃) δ 8.10 (2H, d, J = 7.8 Hz), 7.78 (1H, t, J = 7.8 Hz), 7.66 NHBoc (2H, t, J = 7.8 Hz), 7.57 (1H, d, J = 7.8 Hz), 7.52 (1H, d, J = 7.8 Hz), 7.39 (1H, t, J = 7.8 Hz), 7.20 (1H, t, J = 7.8 Hz), 5.97 (1H, d, J = 7.8 Hz), 5.51 (1H, brs), 1.45 (3H, s), 1.38 $(R^2 = o - Br - C_6 H_4)$ (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ153.7, 137.1, 135.3, 134.9, 133.3, 130.7, 130.3, 129.5, 128.6, 128.1, 124.5, 115.8, 81.0, 66.1, 55.3, 28.2, 18.5; IR 2978, 1705, 1506, 1491, 1449, 1368, 1333, 1246, 1155 cm⁻¹; HRMS (ESI) Calcd for C₂₁H₂₃BrN₂KO₄S⁺ ([M+K]⁺) 517.0193. Found 517.0193.; HPLC IA, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 11.6 min (major syn isomer), 13.9 min (minor syn isomer), 16.6 min (major anti isomer), 39.8 min (minor anti isomer).

4k: ¹H NMR (400 MHz, CDCl₃) δ 8.04 (2H, d, J = 7.8 Hz), 7.75 (1H, t, J = 7.8 Hz), NHBoo 7.62 (2H, t, J = 7.8 Hz), 7.30 (2H, d, J = 8.7 Hz), 6.89 (2H, d, J = 8.7 Hz), 5.42 (1H, .CN SO₂Ph br), 5.23 (1H, d, J = 8.7 Hz), 3.80 (3H, s), 1.38 (3H, s), 1.38 (9H, s); ¹³C NMR (101 Mè $(R^2 = p - MeO - C_6H_4)$ MHz, CDCl₃) δ163.3, 157.2, 139.0, 138.4, 133.8, 132.7, 132.0, 119.7, 117.6, 83.9, 68.9, 60.1, 58.6, 31.6, 23.2, one peak for aromatic carbon was not found probably due to overlapping; IR 2978, 1717, 1512, 1448, 1368, 1329, 1246, 1153 cm⁻¹; HRMS (ESI) Calcd for C₂₂H₂₆N₂KO₅S⁺ ([M+K]⁺) 469.1194. Found 469.1189.; HPLC IC, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 12.3 min (minor *anti* isomer), 16.9 min (major syn isomer), 22.4 min (major anti isomer), 34.8 min (minor syn isomer).

4I: ¹H NMR (400 MHz, CDCl₃) δ 8.02 (2H, d, J = 7.8 Hz), 7.77 (1H, t, J = 7.8 Hz), 7.63 NHBoc .CN (2H, t, J = 7.8 Hz), 7.51 (1H, d, J = 8.2 Hz), 7.27 (1H, d, J = 8.2 Hz), 5.55 (1H, br), 5.23 SO₂Pł (1H, brd, J = 8.2 Hz), 1.39 (3H, s), 1.38 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 153.9, $(R^2 = p - Br - C_6 H_4)$ 135.3, 134.5, 132.1, 131.8, 130.4, 129.9, 129.5, 129.3, 123.2, 116.1, 80.9, 64.9, 56.9, 28.2, 19.9; IR 2978, 1717, 1506, 1489, 1449, 1368, 1331, 1246, 1153 cm⁻¹; HRMS (ESI) Calcd for $C_{21}H_{23}BrN_2KO_4S^+$ ([M+K]⁺) 517.0193. Found 517.0194.; HPLC IC, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 6.9 min (minor anti isomer), 10.2 min (major syn isomer), 13.4 min (major anti isomer), 17.4 min

(minor syn isomer).

4m: ¹H NMR (400 MHz, CDCl₃) δ 8.22 (1H, d, J = 8.2 Hz), 8.14 (2H, d, J = 7.8 Hz), ^{NHBOC} ^{CN} ^{NN} ^{SO₂Ph ^{SO₂Ph ^{SO₂Ph ^{Am}: ¹H NMR (400 MHz, CDCl₃) δ 8.22 (1H, d, J = 8.2 Hz), 8.14 (2H, d, J = 7.8 Hz), 7.87 (2H, t, J = 7.8 Hz), 7.78 (1H, t, J = 7.8 Hz), 7.67-7.63 (3H, m), 7.58 (1H, t, J = 7.8 Hz), 7.52 (2H, t, J = 7.8 Hz), 6.46 (1H, brd, J = 7.8 Hz), 5.50 (1H, br), 1.32 (9H, s), 1.25 (3H, s); ¹³C NMR (101 MHz, CDCl₃) δ 153.9, 135.5, 135.1, 134.2, 133.6, 131.3, 130.6, 129.5, 129.4, 129.1, 127.3, 126.2, 125.6, 124.2, 122.5, 116.4, 80.7, 66.6, 49.8, 28.2, 19.0; IR 2979, 1717, 1506, 1491, 1449, 1368, 1331, 1246, 1153 cm⁻¹; HRMS (ESI) Calcd for C₂₅H₂₆N₂KO₄S⁺ ([M+K]⁺) 489.1245. Found 489.1247.; HPLC IA, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 10.5 min (major *syn* isomer), 13.8 min (major *anti* isomer).}}}

4n: ¹H NMR (400 MHz, CDCl₃) δ 8.07 (2H, d, J = 7.8 Hz), 7.87-7.73 (4H, m), 7.75 (1H, d, J = 7.8 Hz), 7.62 (2H, d, J = 7.8 Hz), 7.53 (3H, m), 5.57 (1H, br), 5.46 (1H, brd, J = 8.7 Hz), 1.42 (3H, s), 1.36 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 153.9, 135.5, 135.1, 134.2, 133.3, 133.0, 130.5, 129.4, 129.1, 128.2, 127.7, 127.4, 126.8, 126.7, 124.3, 116.5, 80.7, 65.4, 57.3, 28.2, 19.9; IR 2978, 1716, 1506, 1491, 1449, 1368, 1329, 1246, 1153 cm⁻¹; HRMS (ESI) Calcd for C₂₅H₂₆N₂KO₄S⁺ ([M+K]⁺) 489.1245. Found 489.1246.; HPLC IC, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 10.3 min (minor *anti* isomer), 13.9 min (major *syn* isomer), 17.2 min (major *anti* isomer), 24.5 min (minor *syn* isomer).

40: ¹H NMR (400 MHz, CDCl₃) δ 7.98 (2H, d, J = 7.8 Hz), 7.73 (1H, t, J = 7.8 Hz), 7.59 (2H, t, J = 7.8 Hz), 7.49 (1H, s), 7.37 (1H, s), 6.49 (1H, br), 5.39 (1H, brd, J = 9.4 Hz), 5.33 (1H, brd, J = 9.4 Hz), 1.58 (3H, s), 1.42 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 154.1, 144.0, 140.9, 135.0, 130.4, 129.3, 129.2, 121.8, 116.8, 108.6, 80.9, 64.9, 50.5, 28.3, 19.6; IR 2980, 1717, 1499, 1491, 1449, 1368, 1329, 1246, 1155 cm⁻¹; HRMS (ESI) Calcd for C₁₉H₂₂N₂KO₅S⁺ ([M+K]⁺) 429.0881. Found 429.0881.; HPLC IA, H/IPA = 10:1, flow rate = 1.0 mL/min, λ = 210 nm, 24.7 min (minor *syn* isomer), 28.2 min (minor *anti* isomer), 34.0 min (major *syn* isomer), 37.2 min (major *anti* isomer).

4p: ¹H NMR (400 MHz, CDCl₃) δ 8.05 (2H, d, J = 7.8 Hz), 7.74 (1H, t, J = 7.8 Hz), 7.64 (2H, t, J = 7.8 Hz), 4.92 (1H, d, J = 11.0 Hz), 4.32-4.24 (1H, m), 2.34-2.22 (1H, m), 1.98-1.65 (4H, m), 1.55 (3H, s), 1.46 (9H, s), 1.37-1.01 (6H, m); ¹³C NMR (101 MHz, CDCl₃) δ 155.3, 135.5, 135.1, 130.6, 129.4, 117.5, 80.2, 63.9, 58.1, 40.7, 31.7, 28.3, 26.1, 25.8, 21.2; IR 2978, 1701, 1506, 1491, 1449, 1368, 1329, 1244, 1155 cm⁻¹; HRMS (ESI) Calcd for C₂₁H₃₀N₂KO₄S⁺ ([M+K]⁺) 445.1558. Found 445.1560.; HPLC IC, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 6.0 min (major *syn* isomer), 10.1 min (major *anti* isomer), 13.7 min (minor *anti* isomer), 39.6 min (minor *syn* isomer).

 $\begin{array}{c} \begin{array}{c} & \text{HBoc} \\ \text{Ph} \overbrace{\text{R}^{3}}^{\text{NHBoc}} & \text{fg}^{1} \text{H NMR} (400 \text{ MHz, CDCl}_{3}) \ \delta 7.91 (2\text{H}, \text{d}, J = 7.8 \text{ Hz}), 7.69 (1\text{H}, \text{t}, J = 7.8 \text{ Hz}), 7.54 (2\text{H}, \text{t}, J = 7.8 \text{ Hz}), 7.46 - 7.43 (2\text{H}, \text{brm}), 7.37 - 7.31 (3\text{H}, \text{m}), 5.82 (1\text{H}, \text{br}), 5.41 (1\text{H}, \text{brd}, J = 8.2 \text{Hz}), 7.46 - 7.43 (2\text{H}, \text{brm}), 7.37 - 7.31 (3\text{H}, \text{m}), 5.82 (1\text{H}, \text{br}), 5.41 (1\text{H}, \text{brd}, J = 8.2 \text{Hz}), 1.97 (1\text{H}, \text{td}, J = 13.0, 4.8 \text{ Hz}), 1.83 (1\text{H}, \text{td}, J = 13.0, 4.8 \text{ Hz}), 1.39 (9\text{H}, \text{s}), 1.28 - 1.19 (4\text{H}, \text{m}), 1.15 - 1.06 (6\text{H}, \text{brs}), 0.84 (3\text{H}, \text{t}, J = 7.3 \text{ Hz}); {}^{13}\text{C} \text{NMR} (101 \text{ MHz, CDCl}_{3}) \ \delta 154.1, 136.4, 134.8, 130.3, 129.1, 128.8, 128.6, 128.5, 127.9, 116.2, 80.7, 69.9, 56.2, 32.4, 31.4, 29.2, 28.5, 28.2, 24.6, 22.5, 14.0; \text{IR 2928, 1719, 1497, 1449, 1329, 1242, 1153 cm}^{-1}; \text{HRMS} (\text{ESI}) \text{ Calcd for } C_{27}\text{H}_{36}\text{N}_{2}\text{KO}_{4}\text{S}^{+} \end{array}$

 $([M+K]^+)$ 523.2027. Found 523.2026.; HPLC AD3, H/IPA = 5:1, flow rate = 1.0 mL/min, λ = 210 nm, 7.2 min (minor *syn* isomer), 10.1 min (major *anti* isomer), 13.7 min (major *syn* isomer), 49.6 min (major *anti* isomer).

 $\begin{array}{l} \begin{array}{l} \label{eq:hermitian} \text{MHBoc} \\ \text{Ph} & \begin{array}{l} & \text{Ar:} \ ^{1}\text{H NMR} \ (400 \ \text{MHz}, \ \text{CDCl}_{3}) \ \delta \ 7.96 \ (2\text{H}, \ d, \ J = 7.8 \ \text{Hz}), \ 7.70 \ (1\text{H}, \ t, \ J = 7.8 \ \text{Hz}), \ 7.57 \\ (2\text{H}, \ t, \ J = 7.8 \ \text{Hz}), \ 7.49 \ -7.44 \ (2\text{H}, \ m), \ 7.36 \ -7.33 \ (3\text{H}, \ m), \ 5.90 \ (1\text{H}, \ br), \ 5.32 \ (1\text{H}, \ brd, \ J = 5.0 \ \text{Hz}), \ 1.80 \ (2\text{H}, \ br), \ 1.38 \ (9\text{H}, \ s), \ 0.92 \ (1\text{H}, \ d, \ J = 6.4 \ \text{Hz}), \ 0.82 \ (3\text{H}, \ d, \ J = 6.4 \ \text{Hz}), \ 0.65 \\ (3\text{H}, \ d, \ J = 6.4 \ \text{Hz}); \ ^{13}\text{C NMR} \ (101 \ \text{MHz}, \ \text{CDCl}_{3}) \ \delta \ 154.1, \ 136.1, \ 135.2, \ 135.0, \ 130.6, \ 129.2, \ 129.0, \ 128.6, \ 128.3, \ 116.5, \ 80.7, \ 68.8, \ 57.3, \ 39.9, \ 28.3, \ 25.5, \ 23.5, \ 22.7; \ \text{IR} \ 2968, \ 1717, \ 1497, \ 1449, \ 1393, \ 1327, \ 1244, \ 1153 \ \text{cm}^{-1}; \ \text{HRMS} \ (\text{ESI}) \ \text{Calcd} \ \text{for} \ C_{24}\text{H}_{30}\text{N}_{2}\text{KO}_{4}\text{S}^{+} \ ([\text{M}+\text{K}]^{+}) \ 481.1558. \ \text{Found} \ 481.1559.; \ \text{HPLC} \ \text{IC}, \ \text{H/IPA} = 95:5, \ \text{flow} \ \text{rate} = 1.0 \ \text{mL/min}, \ \lambda = 210 \ \text{nm}, \ 14.6 \ \text{min} \ (\text{minor} \ anti \ \text{isomer}), \ 30.9 \ \text{min} \ (\text{major} \ anti \ \text{isomer}), \ 30.9 \ \text{min} \ (\text{major} \ anti \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ anti \ \text{minor} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{isomer}). \ 30.9 \ \text{min} \ (\text{major} \ syn \ \text{major} \ syn \ syn \ \text{major} \ syn \$

Crystallographic Structure Determination:

Recrystallization of major diastereomer of 4a (CCDC 867728) and minor diastereomer of 4i (CCDC 867729): 4a was recrystallized from EtOAc/hexane solvent system at -5 °C, and minor isomer of 4i was recrystallized from MeOH/hexane solvent system at -5 °C.

The single crystals thus obtained were mounted on CryoLoop. Data of X-ray diffraction were collected at 153 K on a Bruker SMART APEX CCD diffractometer with graphite-monochromated Mo K α radiation ($\lambda = 0.71073$ Å). An absorption correction was made using SADABS. The structures were solved by direct methods and Fourier syntheses, and refined by full-matrix least squares on F^2 by using SHELXTL.⁴ All non-hydrogen atoms were refined with anisotropic displacement parameters. Hydrogen atoms bonded to nitrogen atoms were located from a difference synthesis and their coordinates and isotropic thermal parameters refined. The other hydrogen atoms were placed in calculated positions and isotropic thermal parameters refined. The crystallographic data were summarized in the following table.

Table S1. Crystal data and	structure refinement for 4a.
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Empirical formula	C21 H24 N2 O4 S	C21 H24 N2 O4 S		
Formula weight	400.48			
Temperature	153(2) K			
Wavelength	0.71073 Å			
Crystal system	Monoclinic			
Space group	P2 ₁			
Unit cell dimensions	a = 13.817(2) Å	$\alpha = 90^{\circ}$.		
	b = 10.8098(16) Å	$\beta = 99.403(3)^{\circ}$		
	c = 14.445(2) Å	$\gamma = 90^{\circ}$.		
Volume	2128.4(5) Å ³			
Z	4			
Density (calculated)	1.250 Mg/m ³	1.250 Mg/m ³		
Absorption coefficient	0.180 mm ⁻¹	0.180 mm ⁻¹		
F(000)	848	848		
Crystal size	0.40 x 0.10 x 0.05 mm ³	0.40 x 0.10 x 0.05 mm ³		
Theta range for data collection	1.43 to 28.30°.	1.43 to 28.30°.		
Index ranges	-16<=h<=18, -12<=k<=1	-16<=h<=18, -12<=k<=14, -19<=l<=19		
Reflections collected	15602			
Independent reflections	8724 [R(int) = 0.0283]	8724 [R(int) = 0.0283]		
Completeness to theta = 28.30°	99.4 %	99.4 %		
Absorption correction	Empirical			
Max. and min. transmission	0.9911 and 0.9315	0.9911 and 0.9315		
Refinement method	Full-matrix least-squares	Full-matrix least-squares on F ²		
Data / restraints / parameters	8724 / 1 / 513			
Goodness-of-fit on F ²	1.161			
Final R indices [I>2sigma(I)]	R1 = 0.0584, WR2 = 0.13	R1 = 0.0584, wR2 = 0.1356		
R indices (all data)	R1 = 0.0648, wR2 = 0.13	R1 = 0.0648, wR2 = 0.1385		
Absolute structure parameter	0.01(7)	0.01(7)		
Largest diff. peak and hole	0.563 and -0.305 e.Å-3	0.563 and -0.305 e.Å ⁻³		

⁴ Sheldrick, G. M. SHELXTL 5.1, Bruker AXS Inc., Madison, Wisconsin, 1997.

Table S2	Crystal	data an	d structure	refinement	for 4	i
1 abic 52.	Crystar	uata an	u su ucture	Termement	. 101 -	۰.

	· · j:		
Empirical formula	C21 H23 Cl N2 O4 S		
Formula weight	434.92		
Temperature	153(2) K		
Wavelength	0.71073 Å		
Crystal system	Orthorhombic		
Space group	$P2_{1}2_{1}2_{1}$		
Unit cell dimensions	$a = 8.911(4) \text{ Å}$ $\alpha = 9$		
	b = 9.919(4) Å	β= 90°.	
	c = 24.452(11) Å	$\gamma = 90^{\circ}$.	
Volume	2161.1(16) Å ³		
Ζ	4		
Density (calculated)	1.337 Mg/m ³		
Absorption coefficient	0.303 mm ⁻¹		
F(000)	912		
Crystal size	0.30 x 0.20 x 0.10 mm ³		
Theta range for data collection	1.67 to 28.24°.		
Index ranges	-8<=h<=11, -12<=k<=13, -32<	<=l<=32	
Reflections collected	15486		
Independent reflections	5319 [R(int) = 0.0395]		
Completeness to theta = 28.24°	99.8 %		
Absorption correction	Empirical		
Max. and min. transmission	0.9704 and 0.9147		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	5319 / 0 / 266		
Goodness-of-fit on F ²	1.071		
Final R indices [I>2sigma(I)]	R1 = 0.0329, $wR2 = 0.0842$		
R indices (all data)	R1 = 0.0339, $wR2 = 0.0850$		
Absolute structure parameter	0.05(4)		
Largest diff. peak and hole	0.319 and -0.326 e.Å ⁻³		



Figure S1. ORTEP diagrams of *anti*-4a (left) and *syn*-4j (right). Calculated hydrogen atoms and solvent molecules are omitted for clarity. Purple = sulfur, blue = nitrogen, red = oxygen, green = chlorine, and gray = carbon. Only one of the two independent molecules of 4a is shown. The thermal ellipsoids are at the 50% probability level.























































1Det A Ch1

1PDA Multi 1

1Det.A Ch1







