A General Approach to the Synthesis of 19-*nor*-Vitamin D₃ and its Cyclic Phosphate Analogs Prepared from Cyclohexadienyl Sulfone.

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Supporting Information

GENERAL PROCEDURES

Tetrahydrofuran (THF) was distilled from benzophenone ketyl. Sodium sulfate (Na₂SO₄) was anhydrous. Unless otherwise indicated, all reactions were carried out under in a positive pressure of Argon in anhydrous solvents and the reaction flasks were fitted with rubber septa for the introduction of substrates and reagents via syringe. Progress of reactions was monitored by thin layer chromatography (TLC) using silica gel plates. The TLC plates were visualized with a UV lamp (254 nm) and/or with TLC visualizing solutions activated with heat. The two commonly employed TLC visualizing solutions were: (i) *p*-anisaldehyde solution (1350mL absolute ethanol, 50mL concentrated H_2SO_4 , 37mL *p*-anisaldehyde), and (ii) permanganate solution (weight percents of 1% KMnO₄ and 2% Na₂CO₃ in water).

¹H NMR and ¹³C NMR spectra were recorded on 300-500 MHz. The NMR spectra were determined in CDCl3 solution. Peak multiplicities in ¹H-NMR spectra, when reported, are abbreviated as s (singlet), d (doublet), t (triplet), m (multiplet), and/or quint (quintet). Mass spectra were run by the Purdue University campus wide mass spectrometry facility.

EXPERIMENTALS

Preparation of 7:

TBSO^W 7 T

To a -78 °C solution of epoxyvinyl sulfone 6 (200 mg, 0.55 mmol) in THF (6

mL) was added LiBH₄ (3.0 equiv, 1.65 mmol, 1.65 mL of 1.0 M THF solution). The reaction mixture was slowly allowed to warm to room temperature and stirred for 8 h and then quenched with water and extracted with diethyl ether. The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residues were purified on silica gel using 1:3 Ethyl Acetate/Hexane to give the desired alcohol 7 in 81 % yield and 2.8:1 dr. (164 mg).

¹H NMR (CDCl₃, 300 MHz): 7.86 (1H, d, J= 9 Hz), 7.65 (1H, t, J= 9Hz), 7.55 (2H, t, J= 9 Hz),
4.32 (1H, s), 3.96 (1H, m), 3.37 (1H, m), 2.35 – 1.23 (6H, m), 0.85 (9H, s), 0.02 (6H, s).
¹³C NMR (CDCl₃, 75 MHz): 136.9, 136.7, 133.7, 129.1, 129.0, 128.8, 66.4, 67.0, 66.4, 66.2,
57.4, 41.5, 41.3, 34.4, 33.8, 32.2, 31.6, 25.7, 25.6, 18.0, 17.8, -4.8, -5.1, -5.3.
HRMS (CI) calculated for C₁₈H₃₀O₄SSi (M+H), 371.1712 found 371.1717.

Preparation of 10:



To a -78 °C solution of alcohol 7 (40 mg, 0.11 mmol) in THF (0.5 mL) was

added BuLi (2.0 equiv, 0.22 mmol, 88 μ L of 2.5M hexane solution). The reaction mixture was slowly allowed to warm to 0 °C over a period of 1 h and then cooled back to -78 °C. Allyl chloride **8** (1.1 equiv) dissolved in 0.2 mL HMPA was added drop wise to the reaction mixture. The reaction mixture was slowly allowed to warm to room temperature over a period of 3 h and then quenched by adding water (10 μ L). TBAF (10 equiv, 1.1 mmol, 1M in THF) was added and

the reaction mixture heated at 65 °C for 8 h after which it is quenched with 5 mL of water and extracted with diethyl ether (10 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:1 Ethyl Acetate/Hexane to give the diol **10** in 70 % yield (29 mg).

¹H NMR (CDCl₃, 500 MHz) δ 6.31 (1H, d, J= 10 Hz), 5.85 (1H, d, J= 10Hz), 4.11 (1H, m), 4.03 (1H, m), 2.73 (2H, m), 2.47 (1H, d, J= 10 Hz), 2.19 (2H, m), 2.01 – 1.01 (21H, m), 0.92 (3H, d, J= 5Hz), 0.88 (3H, d, J= 2.5 Hz), 0.87 (3H, d, J= 2.5 Hz), 0.55 (3H, s).

¹³C NMR (CDCl₃, 500 MHz) δ 143.2, 131.0, 123.9, 115.2, 67.4, 67.2, 56.6, 56.3, 45.8, 44.6, 42.2, 40.5, 39.5, 37.2, 36.1, 29.0, 28.0, 27.6, 23.8, 23.5, 22.8, 22.6, 22.3, 18.8, 12.1 HRMS (CI) calculated for $C_{26}H_{44}O_2$ (M+) 388.3341, found 388.3347

Preparation of 14:



14 1,3-dithaine (223 mg, 1.86 mmol) was dissolved in THF (5 mL) and cooled to -10 °C. BuLi (0.74 mL, 2.5 M solution in hexane, 1.86 mmol) was added and reaction mixture left stirring for 30 minutes at -10 °C and then further cooled to -78 °C. Dienyl sulfone 12 (500 mg, 1.43 mmol) was dissolved in THF (3 mL) and added dropwise to the above solution and reaction mixture left stirring at the same temperature for 1 h. Diphenyl disulfide (218 mg, 2.79 mmol) dissolved in THF (1 mL) was added to the above reaction mixture and the reaction mixture gradually warmed to room temperature over 5 h and then quenched by adding water and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:4 Ethyl Acetate/Hexane to give the vinyl sulfide 14 in 80% yield (661 mg).

¹H NMR (CDCl₃, 400 MHz): 7.83(2H, d, J = 4.0 Hz), 7.61 (1H, t, J= 8 Hz), 7.50(2H, t, J= 8Hz), 7.40- 7.31 (5H, m), 5.26 (1H, d, J= 4 Hz), 4.40 (1H, d, J = 4.0 Hz), 4.18 (1H, dd, J= 4 Hz, 8 Hz), 3.99 (1H, dd, J= 8 Hz, 16 Hz), 2.88- 2.75 (6H, m), 2.06 (2H, d, J= 8 Hz), 1.87- 1.76 (1H, M), 0.87 (9H, s), 0.10 (3H, s), -0.06 (3H, s).

¹³C NMR (CDCl₃, 100 MHz): δ 141.7, 137.9, 133.6, 133.5, 131.1, 129.3, 129.2, 128.9, 128.4, 114.7, 68.7, 66.4, 51.6, 45.5, 36.5, 31.0, 25.8, 17.9, -4.3, -4.5. HRMS (CI) calculated for C₂₈H₃₈O₃S₄Si (M+Na) 601.1371 found 601.1366.

Preparation of 15:

HO^{-///,} TBSO⁻ SPh

To a suspension of HgO (300 mg, 1.38 mmol) in THF/ H₂O (5 mL, 5:1) was added BF₃.OEt₂ (196 mg, 0.17 mL, 1.38 mmol) and reaction mixture stirred for 10 min. Vinyl sulfide **14** (400 mg, 0.69 mmol) dissolved in THF (3 mL) was added to the above suspension and reaction mixture heated to 75 °C for 30 min. and then diluted with H₂O (20 mL) and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was dissolved in MeOH (10mL) and reaction mixture cooled to 0 °C and NaBH₄ (78.7 mg, 2.07 mmol) was added in small portions over 30 minutes and reaction mixture stirred for an additional 2.5 h. The reaction mixture was then quenched by adding NH₄Cl and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:2 Ethyl Acetate/Hexane to give the alcohol **15** in 72% yield (243 mg).

¹H NMR (CDCl₃, 400 MHz): 7.80 (2H, d, J = 8.0 Hz), 7.67 (2H, t, J= 8 Hz), 7.54 (3H, t, J= 8 Hz), 7.38 (5H, s), 5.38 (1H, d, J= 4 Hz), 4.18 (1H, d, J= 8 Hz), 4.08 (1H, d, J= 8 Hz), 3.99 (1H, d, J= 12 Hz), 3.77 (1H, t, J= 4 Hz, 12 Hz), 2.06 (2H, m), 1.84 (1H, m), 0.87 (9H s), 0.07 (3H, s), 0.02 (3H, s)

¹³C NMR (CDCl₃, 100 MHz): δ 140.3, 136.4, 133.7, 133.6, 130.9, 129.3, 129.2, 128.9, 128.9, 128.6, 115.3, 68.1, 64.8, 60.3, 43.1, 37.7, 25.6, 17.8, -4.4, -5. HRMS (ESI) calculated for $C_{25}H_{34}O_4S_2Si$ (M+Na), 513.1566, found 513.1561.

Preparation of 16:

Alcohol **15** (200 mg, 0.41 mmol) was dissolved in DCM (20 mL) and Et₃N (373 mg, 0.51 mL, 3.69 mmol) followed by TBSOTf (130 mg, 0.11 mL, 0.492 mmol) was added at 0 °C and solution left stirring for 1 h. TMSOTf (456 mg, 0.37 mL, 2.05 mmol) was added drop wise and then solution refluxed for 8 h and finally quenched by adding H₂O (200 mL). The organic phase is separated and cooled to 0 °C and MCPBA (460 mg, 2.05 mmol) is added portion wise over 3 h and reaction mixture stirred for another 8 h and then quenched by adding saturated aq. NaHSO₃ (10 mL). The organic phase is separated and washed with saturated aq. NaHCO₃ (20 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:4 Ethyl Acetate/Hexane to give the dienyl sulfone **16** in 74% yield (202 mg).

¹H NMR (CDCl₃, 300 MHz): 7.87 (1H, d, J= 9 Hz), 7.49 (3H, m), 6.76 (1H, s), 6.08 (1H, d, J= 12 Hz), 5.94 (1H, dd, J= 6 Hz, 9 Hz), 4.63 (1H, dd, J= 3 Hz, 9 Hz), 3.53 (1H, m), 3.43 (1H, m), 2.57 (1H, brs), 0.89 (9H, s), 0.87 (9H, s), 0.12 (3H, s), 0.09 (3H, s), 0.02 (6H, s) ¹³C NMR (CDCl₃, 75 MHz): δ 139.4, 137.9, 136.6, 133.4, 132.7, 129.2, 127.8, 118.1, 66.7, 61.7, 45.2, 25.8, 25.7, 18.2, 17.9, - 4.36, - 4.85, -5.46, -5.56.

HRMS (ESI) calculated for C₂₅H₄₂O₄S₂Si (M+Na), 517.2240 found 517.2248.

Preparation of 17:



Dienyl sulfone **16** (510 mg, 1.03 mmol) was dissolved in MeCN/H₂O (10 mL, 10:1) and NaHCO₃ (519 mg, 6.18 mmol) was added to it and the reaction mixture cooled to 0 $^{\circ}$ C. 1,1,1 - trifluoroacetone (346 mg, 0.28 mL, 3.09 mmol) was added and Oxone (1.90 g, 3.09 mmol) added over 1 hr and reaction mixture stirred for another 2 h and then diluted by adding water (20 mL) and DCM (50 mL). The organic phase was separated and combined

extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:4 Ethyl Acetate/Hexane to give the epoxide **17** in 6:1 dr and in 75% yield (394 mg).

¹H NMR (CDCl₃, 300 MHz): 7.89(2H, d, J = 9.0 Hz), 7.64-7.48 (3H, m), 6.92 (1H, s), 4.10 (1H, d, J= 12 Hz), 3.99 (1H, dd, J= 3 Hz, 9 Hz), 3.75 (1H, dd, J= 6 Hz, 3.68 (1H, dd, J= 2 Hz, 4 Hz), 3.61 (1H, d, J= 6 Hz), 1.82 (1H, m), 0.89 (9H, s), 0.87 (9H, s), 0.10 (3H, s), 0.06 (6H, s) ¹³C NMR (CDCl₃, 75 MHz): δ 145.3, 139.1, 137.8, 133.7, 129.4, 127.9, 66.2, 61.9, 53.6, 45.7, 43.0, 25.7, 25.5, 18.1, 17.7, 4.6, 5.1, 5.6.

HRMS (ESI) calculated for C₂₅H₄₂O₅SSi₂ (M+Na) 533.2189 found 533.2192.

Preparation of sulfone 4:



Epoxide 17 (300 mg, 0.59 mmol) was dissolved in THF (10 mL) and cooled to -78 °C. LiBH₄ (1.8 mL, 1M solution in THF, 1.77 mmol) was added dropwise and reaction mixture gradually allowed to warm to room temperature over a period of 10 h and then quenched by adding NH₄Cl. The reaction mixture was diluted with ether (30 mL), organic phase separated and combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was dissolved in DCM (10 mL) and cooled to 0 °C and Et₃N (119 mg, 0.16 mL, 1.18 mmol) followed by TBSOTf (156 mg, 0.14 mL, 0.59 mmol) was added and reaction mixture quenched by adding water after 1 h. The organic phase was separated and combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:5 Ethyl Acetate/Hexane to give the sulfone **4** with >90% purity and in 56% yield (208 mg).

¹H NMR (CDCl₃, 400 MHz): 7.86 (2H, d, J= 8.0 Hz), 7.64 (1H, t, J= 8 Hz), 7.54(2H, t, J=8 Hz), 5.82 (1H, s), 5.22 (1H, s), 4.36 (1H, d, J = 2.4Hz), 4.10 (1H, s), 1.81 (1H, m), 1.06(1H, d, J = 6.9 Hz), 0.693(9H, s), -0.452(3H, s), -0.044(3H, s). ¹³C NMR (CDCl₃, 300 MHz): δ 137.2,

133.6, 129.0, 128.8, 67.0, 66.2, 60.1, 57.4, 52.1, 33.2, 33.1, 25.8, 25.6, 18.1, 17.8, 17.7, - 4.2, - 5.1, - 5.2, -5.4, -5.5 HRMS (ESI) calculated for $C_{31}H_{60}O_5SSi_3$ (M+Na) 651.3367 found 651.3359.

Preparation of allyl sulfone 19:

S SO₂Ph S TBSO^W 19

¹⁹ 1,3-dithaine (223 mg, 1.86 mmol) was dissolved in THF (5 mL) and cooled to -10 °C. n-BuLi (0.74 mL, 2.5 M solution in hexane, 1.86 mmol) was added and reaction mixture left stirring for 30 minutes at -10 °C and then further cooled to -78 °C. Dienyl sulfone **18** (500 mg, 1.43 mmol) was dissolved in THF (3 mL) and added dropwise to the above solution and reaction mixture left stirring at the same temperature for 1 h and then quenched by adding water and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:4 Ethyl Acetate/Hexane to give the allyl sulfone **19** in 83% yield (558 mg).

¹H NMR (CDCl₃, 300 MHz): 7.89 (2H, d, J= 9.0 Hz), 7.61 (1H, t, J= 9 Hz), 7.51 (2H, t, J=9 Hz), 5.96 (1H, m), 5.57 (1H, m), 4.32 (1H, d, J= 3 Hz), 4.18 (1H, t, J=3 Hz), 3.98 (1H, m), 2.75 (6H, m), 2.04 (1H, m), 1.8 (2H, m), 0.90 (9H, s), 0.15 (3H, s), 0.06 (3H, s)

¹³C NMR (CDCl₃, 75 MHz): δ 137.9, 133.6, 133.2, 129.4, 128.8, 119.7, 67.7, 64.6, 51.6, 45.5, 32.0, 31.0, 30.8, 25.9, 18.0, -4.3.

HRMS (ESI) calculated for C₂₂H₃₄O₃S₃Si (M+Na), 493.1337 found 493.1344.

Preparation of allyl sulfone 20:



To a suspension of HgO (300 mg, 1.38 mmol) in THF/ H_2O (5 mL, 5:1) was added BF₃.OEt₂ (196 mg, 0.17 mL, 1.38 mmol) and reaction mixture stirred for 10 min. Allyl sulfone **19** (324 mg, 0.69 mmol) dissolved in THF (3 mL) was added to the above suspension and reaction mixture heated to 75 °C for 30 min. and then diluted with H_2O (20 mL) and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was dissolved in MeOH (10 mL) and reaction mixture cooled to 0 $^{\circ}$ C and NaBH₄ (78.7 mg, 2.07 mmol) was added in small portions over 30 minutes and reaction mixture stirred for an additional 2.5 h. The reaction mixture was then quenched by adding NH₄Cl and extracted with ether (50 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 1:2 Ethyl Acetate/Hexane to give the alcohol **20** in 77% yield (264 mg).

¹H NMR (CDCl₃, 400 MHz): 7.85 (2H, d, J= 8.0 Hz), 7.63 (1H, t, J=8.0 Hz), 7.52 (2H, d, J= 8Hz), 5.84 (1H, t, J= 8 Hz), 5.64 (1H, d, J= 12 Hz), 4.13 (1H, d, J= 8 Hz), 4.02 (2H, d, J= 4 Hz), 3.72 (1H, dt, J= 8 Hz, 12 Hz), 2.13 (1H, td, J= 8 Hz, 16 Hz), 1.91 (1H, t, J= 12 Hz), 1.68 (1H, t, J= 12 Hz), 0.86 (9H, s), 0.07 (3H, s), 0.03 (3H, s).

¹³C NMR (CDCl₃, 100 MHz): δ 136.4, 133.8, 131.9, 129.4, 128.8, 120.4, 67.8, 64.3, 60.1, 43.6, 33.8, 25.7, 17.8, -4.3, -5.1.

HRMS (ESI) calculated for C₁₉H₃₀O₄SSi (M+H) 383.1712 found 383.1708.

Preparation of allyl sulfone 21:



Alcohol **20** (50 mg, 0.13 mmol) was dissolved in DCM (1mL) and DMAP (3 mg, 0.03 mmol) followed by Pyridine (31 mg, 31 μ l, 0.39 mmol) was added to it. The reaction mixture was cooled to 0 °C and (Boc)₂O (57 mg, 0.26 mmol) was added and the reaction mixture warmed to room temperature and left stirring for 3 h. The reaction mixture was directly loaded on the silica gel column and purified by using Ethyl Acetate/Hexane (1:4) to give the allyl sulfone **21** in 90% yield (56 mg).

¹H NMR (CDCl₃, 400 MHz): 7.86 (2H, d, J= 8 Hz), 7.65 (1H, t, J= 8 Hz), 7.54 (2H, t, J= 8 Hz), 5.86 (1H, t, J= 8 Hz), 5.65 (1H, d, J= 8Hz), 4.47 (2H, s), 4.18 (1H, d, J= 8 Hz), 3.78 (1H, dt, J= 4 Hz, 12 Hz), 2.16 (1H, td, J= 8 Hz, 16 Hz), 2.06 (1H, t, J= 8 Hz), 1.69 (1H, t, J=12 Hz), 1.48 (3H, s), 0.86 (9H, s), -0.02 (3H, s), -0.01 (3H, s)

¹³C NMR (CDCl₃, 100 MHz): δ 153.1, 136.6, 133.8, 131.8, 129.5, 128.9, 120.4, 82.0, 66.2, 63.9, 63.8, 41.3 33.8, 27.8, 25.7, 17.9, -4.4, -5.3.

HRMS (ESI) calculated for C₂₄H₃₈O₆SSi (M+Na) 505.2056 found 505.2063.

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Preparation of alcohol 22: SO₂Ph TBSO ЮH 22

² OTBS To a -10 °C suspension of trimethylsulfonium iodide (82 mg, 0.40 mmol) in

THF (0.6 mL) was added *n*-BuLi (0.15 mL of 2.5 M hexane solution, 0.37 mmol). After 30 min, the solution was cooled to -78 $^{\circ}$ C and epoxide **17** (50 mg, 0.10 mmol) in THF (0.6 mL) was introduced. The reaction was left stirring for 1 h and then quenched with water and extracted with diethyl ether (10 mL). The combined extracts were washed with brine, dried over magnesium sulfate, filtered, and concentrated under vacuum. The residues were purified on silica gel using Ethyl Acetate/Hexane (1:3) to give the desired alcohol **22** in 52 % yield (27 mg).

¹H NMR (CDCl₃, 300 MHz): 7.84 (2H, d, J= 9.0 Hz), 7.55 (1H, t, J= 9 Hz), 7.46 (2H, t, J= 9 Hz), 7.25 (1H, s), 5.91 (1H, s), 5.22 (1H, s), 4.93 (1H, s), 4.06 (1H, d, J= 3 Hz), 3.79 (1H, d, J= 6 Hz), 3.57 (1H, t, J= 9 Hz), 2.36 (1H, m), 0.89 (9H, s), 0.74 (9H, s), 0.08 (3H, s), 0.06 (3H, s), - 0.03 (3H, s), - 0.33 (3H, s).

¹³C NMR (CDCl₃, 125 MHz): δ142.0, 139.9, 136.2, 136.3, 133.1, 128.9, 127.8, 117.6, 73.9, 68.0, 61.7, 46.3, 25.7, 25.5, 18.0, 17.8, 1.0, -4.8, -5.1, -5.7.

HRMS (ESI) calculated for C₂₆H₄₄O₅SSi₂ (M+Na), 547.2346 found 547.2343.

Preparation of allyl sulfone 23:



To a 0 °C solution of alcohol 22 (25 mg, 0.05 mmol) in THF (1 mL) was added DEAD (26 mg, 24 μ l, 0.15 mmol) and PPh₃ (40 mg, 0.15 mmol) followed by HCOOH (3.5 mg, 3 μ l, 0.075 mmol). The reaction mixture was left stirring for 5 h and then quenched by adding 5 mL of saturated solution of Na₂CO₃. The solution was left stirring for an additional 2 h and then extracted with diethyl ether (10 mL). The combined extracts were washed with brine, dried over magnesium sulfate, filtered, and concentrated under vacuum. The residues were purified on silica gel column using Ethyl Acetate/Hexane (1:3) to give the desired allylic alcohol 23 in 65 % yield (16 mg). ¹H NMR (CDCl₃, 500 MHz): 7.88 (2H, d, J= 7.0 Hz), 7.58 (1H, t, J= 7 Hz), 7.49 (2H, t, 7 Hz), 7.20 (1H, d, J= 5 Hz), 6.05 (1H, s), 5.28 (1H, s), 4.28 (1H, d, J = 5 Hz), 4.24 (1H, t, J= 3.5 Hz), 3.42 (1H, d, J= 10 Hz), 2.36 (1H, m), 0.88 (9H, s), 0.75 (9H, s), 0.04 (3H, s), 0.02 (3H, s), -0.06 (3H, s), -0.33 (3H, s).

¹³C NMR (CDCl₃, 125 MHz): δ 139.6, 138.5, 137.0 134.8, 133.3, 129.0, 128.1, 119.1, 73.7, 67.4, 62.4, 48.8, 25.8, 25.6, 18.1, 17.9, -4.9, -5.4, -5.6.

HRMS (ESI) calculated for $C_{26}H_{44}O_5SSi_2$ (M+Na), 547.2346 found 547.2343.

Preparation of tetrol 1,2:



To a -78 °C solution of sulfone 4 (60 mg, 0.10 mmol) in THF

(0.3 mL) was added *n*-BuLi (44 μ L of 2.5M hexane solution, 0.11 mmol) and yellow solution left stirring for 30 min. Allyl Chloride (48 mg, 0.11 mmol) dissolved in HMPA (0.1 mL) was added drop wise to the reaction mixture. The reaction mixture was slowly allowed to warm to room temperature over a period of 6h. TBAF (1 mL, 1M solution in THF, 1.0 mmol) was added and the reaction mixture heated at 65 °C for 8 h after which it is quenched with 5 mL of water and extracted with diethyl ether (10 mL). The combined extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The residue was purified on silica gel using 4:1 Ethyl Acetate/Hexane to give the tetrol **1** and **2** in 76 % yield and 2:1 dr (32 mg).

¹H NMR (CDCl₃, 500 MHz) δ 6.39 (1Ha, d, J= 10 Hz), 6.24 (1Hb, d, J= 10Hz), 5.88 (1Hb, d, J= 10 Hz), 5.79 (1Ha, d, J= 10 Hz), 4.24 (1Ha, s), 4.16 (1Hb, s), 4.02 (3Ha, m), 3.92 (3Hb, m), 3.17 – 0.90 (64 H), 0.53 (3Ha + 3Hb, d, J= 6 Hz).

¹³C NMR (CDCl₃, 125 MHz) δ 143.2, 143.1, 130.9, 130.8, 124.2, 123.6, 115.3, 115.2, 71.4, 71.1, 68.6, 68.3, 64.4, 64.1, 56.5, 56.3, 53.7, 49.7, 45.7, 45.4, 44.4, 44.1, 40.4, 37.5, 36.3, 36.1, 35.7, 29.3, 29.1, 29.0, 28.7, 27.7, 23.5, 23.4, 22.3, 20.8, 18.8, 14.1, 12.1, 12.0.



HRMS (ESI) calculated for C₂₇H₄₆O₄ (M+Na) 457.3294, found 457.3298.

To a -20 °C solution of 1:1 mixture of tetrol **1**, **2** (50 mg, 0.12 mmol) in DCM (1.0 mL) was added Pyridine (57 mg, 58 μ L, 0.72 mmol) followed by Phenyl dichlorophosphate (25mg, 18 μ L, 0.12 mmol). The solution was left stirring for 1 h at -20 °C and at room temperature for 6 h and then directly loaded on silica gel column and purified using CH₃CN/DCM (1:5) to give the mixture of phosphate **25** and **26** (which were separated on HPLC column to give **25** (9 mg) and **26** (15.5 mg) and **27** (6.4 mg), **28** (10 mg) in 60 % yield.

Phosphonate 25:



¹H NMR (CDCl₃, 500 MHz) δ 7.34 (2H, t, J= 8 Hz), 7.27 (1H, m), 7.17 (2H, m, 7 Hz), 6.34 (1H, d, J= 11 Hz), 5.85 (1H, d, J= 11 Hz), 4.57 (2H, m), 4.34 (1H, m), 4.07 (1H, m), 3.29 (1H, dd, J= 5 Hz, 14 Hz), 2.78 (1H, dd, J= 5 Hz, 13 Hz), 2.54 (1H, d, J= 14 Hz), 2.38 (1H, d, J= 13 Hz), 2.15 (1H, t, J= 12 Hz), 2.02 (2H, d, J= J= 11 Hz), 1.90 (2H, m), 1.68 (2H, m), 1.49 – 1.27 (10 H, m), 1.23 (6H, s), 1.06 (2H, m), 0.96 (3H, d, J= 6 Hz), 0.83 (2H, m), 0.56 (3H, s). 31P (200 MHz): δ - 13.05. ¹³C NMR (CDCl₃, 125 MHz) δ 143.7, 129.9, 127.6, 125.0, 124.3, 119.5, 115.2, 79.6 (d, J (P, H) = 7 Hz), 71.1, 68.4 (d, J (P, H) = 6 Hz), 64.5, 56.5, 56.2, 45.7, 44.4, 43.7 (d, J (P, H) = 4 Hz), 41.5 (d, J (P, H) = 9 Hz), 40.4, 37.4, 36.4, 36.1, 29.7, 29.3, 29.2, 29.0, 27.7, 23.5, 22.3, 20.8, 18.8, 12.0. HRMS (ESI) calculated for C₃₃H₄₉O₆P (M+Na) 595.3168, found 595.3168.

Phosphonate 26:



¹H NMR (CDCl₃, 500 MHz) δ 7.34 (2H, t, J= 8 Hz), 7.18 (3H, m), 6.50 (1H, d, J= 11Hz), 5.75 (1H, d, J= 11 Hz), 4.69 (2H, m), 4.36 (1H, m), 4.04 (1H, s), 3.67 (1H, m), 2.98 (1H, d, J= 14 Hz), 2.79 (1H, dd, J= 5 Hz, 14 Hz), 2.33 (1H, t, J= 13 Hz), 2.10 (1H, d, J= 14 Hz), 2.02 (1H, d, J= 10 Hz), 1.87 (1H, m), 1.68 – 1.26 (12 H, m), 1.23 (6H, s), 1.06 (1H, m), 0.94 (3H, d, J= 6 Hz), 0.83 (1H, m), 0.54 (3H, s). 31P (200 MHz): δ -10.61. ¹³C NMR (CDCl₃, 125 MHz) δ 150.5, 145.1, 129.7, 126.8, 126.1, 125.4, 120.3, 114.8, 77.9 (d, J (P, H) = 6 Hz), 71.1, 70.6 (d, J (P, H) = 7 Hz), 66.5, 56.5, 56.3, 45.9, 44.8 (d, J (P, H) = 7 Hz), 44.4, 43.8, 40.4, 36.3 (d, J (P, H) = 7 Hz), 36.1, 35.3, 29.7, 29.3, 29.2, 29.0, 27.7, 23.6, 22.2, 20.8, 18.8, 12.0. HRMS (ESI) calculated for $C_{33}H_{49}O_6P$ (M+Na) 595.3168, found 595.3168.

Phosphonate 27:



¹H NMR (CDCl₃, 500 MHz) δ 7.34 (2H, t, J= 10 Hz), 7.19 (3H, m), 6.32 (1H, d, J= 11 Hz), 5.83 (1H, d, J= 11 Hz), 4.68 (1H, dt, J= 6 Hz, 12 Hz), 4.62 (1H, dt, J= 6 Hz, 11 Hz), 4.35 (1H, m), 3.99 (1H, s), 3.33 (1H, dd, J= 5 Hz, 12 Hz), 2.76 (1H, d, J= 12 Hz), 2.46 (1H, d, J= 15 Hz), 2.35 (1H, d, J= 14 Hz), 2.12 – 1.91 (4H, m), 1.68 (2H, m), 1.51 – 1.27 (10H, m), 1.23 (6H, s), 1.06 (2H, m), 0.95 (3H, d, J= 6 Hz), 0.82 (2H, m), 0.55 (3H, s). 31P (200 MHz): δ -10.40. ¹³C NMR (CDCl₃, 125 MHz) δ 150.4, 145.0, 129.8, 127.1, 126.0, 125.0, 119.6, 114.9, 78.8 (d, J(P, H) = 7 Hz), 71.2 (d, J (P, H) = 7 Hz), 71.1, 66.8, 56.5, 56.3, 46.0, 45.1 (d, J(P, H) = 7 Hz), 44.4, 44.0, 40.4, 36.3, 36.0, 35.0 (d, J (P, H) = 9 Hz), 29.7, 29.4, 29.2, 29.1, 27.6, 23.6, 22.2, 20.7, 18.8, 12.0. HRMS (ESI) calculated for $C_{33}H_{49}O_6P$ (M+Na) 595.3168, found 595.3168.

Phosphonate 28:



¹H NMR (CDCl₃, 500 MHz): δ 7.34 (2H, t, J= 8 Hz), 7.27 (1H, m), 7.17 (2H, t, J= 8 Hz), 6.50 (1H, d, J= 12 Hz), 5.76 (1H, d, J= 12 Hz), 4.66 (1H, dt, J= 5 Hz, 11 Hz), 4.61 (1H, t, J= 11 Hz), 4.37 (1H, m), 4.15 (1H, s), 3.66 (1H, m), 2.99 (1H, d, J= 15 Hz), 2.77 (1H, m), 2.45 (1H, t, J= 12 Hz), 2.17 (1H, d, J= 14 Hz), 2.01 (1H, d, J= 11 Hz), 1.90 (1H, m), 1.68 (1H, d, J= 11 Hz), 1.49 - 1.26 (10H, m), 1.23 (6H, s), 1.06 (2H, m), 0.94 (3H, d, J= 6 Hz), 0.82 (2H, m), 0.54 (3H, s). 31P (200 MHz): δ -13.04. ¹³C NMR (CDCl₃, 125 MHz) δ 150.5, 145.2, 129.8, 126.8, 125.1, 119.5, 114.6, 79.1 (d, J (P, H) = 8 Hz), 71.3 (d, J (P, H) = 7 Hz), 71.1, 67.1, 56.5, 56.3, 45.9, 45.1 (d, J (P, H) = 6 Hz), 44.4, 42.9 (d, J (P, H) = 9 Hz), 40.3, 36.3, 36.1, 35.8, 29.7, 29.4, 29.2, 29.0, 27.6, 23.4, 22.2, 20.8, 18.8, 12.1. HRMS (ESI) calculated for C₃₃H₄₉O₆P (M+Na) 595.3168, found 595.3168

Structural assignments for cyclic phosphate analogs of 19-nor-Vitamin D₃:



COSY spectra of all the four compounds 25, 26, 27 and 28 were recorded which helped identify the protons at C-1, C-2, C-3, C-4 and C-10. NOESY spectra of the compounds showed the correlation between H_7 and H_{10} . Correlation was also seen between H_4 and H_6 in the NOESY spectra. These NMR results confirmed the ring structures in the respective molecules. To confirm the stereochemistry at the phosphorus stereo-center, 31P spectra were recorded for all the four compounds. As already reported in literature,¹ the axial P-O bond results in an upfield shift in the 31P spectra, this lead to the assignment for the cyclic phosphate analogs 25, 26, 27 and 28. (Figure 1)



¹ (a) Riley, A. M.; Guedat, P.; Schlewer, G.; Spiess, B.; Potter, B. V. L. J. Org. Chem. 1998, 63, 295. (b) Xu, H.; Zhang, J.; Liu, H.; Wang, J. Synthetic communications, 2006, 36, 407. (c) Hoeve, W. T.; Wynberg, H. J. Org. Chem., 1985, 23, 4508.

Figure 1: Axial P-O vs. equatorial P-O bond (CD rings not shown).

Cell culture experiment:

Cells were seeded in 12 well dishes (100,000 cells/well for Caco-2; 50,000 cells/well for SW480-ADH) and studied when they reached 70-80% confluence (3 days in culture). Cells were treated with 1,25 dihydroxyvitamin D or analogs in high glucose DMEM supplemented with Pen/Strep but without fetal bovine serum (FBS). Cell culture conditions have been previously described by our group.² Vitamin D compounds were diluted in ethanol and used at three concentrations: 1 nM, 10 nM, and 100 nM. Ethanol control groups were included for all of the three treatments but total ethanol levels in the medium were 0.001, 0.01, and 0.1% for the three treatment groups, respectively. Four replicate wells were used for each of the vitamin D compounds. Six hours after the treatment started, medium was removed and cells were harvested directly into TriReagent (Molecular Research Center, Inc., Cincinnati, OH). RNA was isolated, converted to cDNA, and analyzed by real-time PCR for mRNA levels for the two target genes using methods previously described.³ Data were normalized to the expression level of the control gene RPLP0. Primers for human CYP24 and RPLPO have been described previously⁴ and the primers for E-Cadherin are: Forward ^{5'}GATTGCAAATTCCTGCCATT^{3'}; Reverse: ^{5'}GCTGGCTCAAGTCAAGTCC^{3'}.

Treatment	1 nM	SEM	10 nM	SEM	100 nM	SEM
Ethanol	1.00	0.08	0.92	0.10	1.19	0.20
1,25D	233.31	57.10	1479.96	305.84	1417.24	216.53
33	1.25	0.27	1.28	0.21	1.32	0.16
32	0.85	0.26	1.21	0.10	1.60	0.37
34	1.26	0.18	1.38	0.50	5.04	0.36
35	2.00	0.17	1.82	0.20	3.31	0.36

Table 1: Caco-2 Summary:

Table 2: SW480 ADH Summary:

² Fleet, J.C., DeSmet, M., Johnson, R., Li, Y. Biochemical Journal, 2012, 61-76.

³ Cui, M., Klopot, A., Jiang, Y. and Fleet, J.C. J. Cell Physiol., 2009, 113.

⁴ Kovalenko, P.L., Zhang, Z., Cui, M., Clinton, S.K., Fleet, J.C. BMC Genomics, 2009, 11, 26.

SW480 ADH Summary Table		CYP24			E- Cadherin		
Compound	Dose (nM)	Mean	SD	SEM	Mean	SD	SEM
ethanol	1	1.00	0.15	0.08	1.00	0.07	0.03
	10	1.03	0.19	0.09	1.07	0.43	0.22
	100	1.12	0.71	0.35	1.07	0.37	0.18
1,25(OH)2 D	1	653.35	92.18	46.09	6.57	0.99	0.50
	10	1380.02	211.28	105.64	11.36	2.65	1.32
	100	634.80	257.35	128.68	4.97	1.16	0.58
33	1	0.79	0.22	0.11	1.05	0.31	0.15
	10	1.45	0.34	0.17	1.09	0.33	0.16
	100	23.54	6.37	3.68	1.92	0.59	0.34
32	1	0.89	0.18	0.09	0.87	0.32	0.16
	10	1.10	0.22	0.11	0.94	0.07	0.04
	100	3.95	0.50	0.25	1.06	0.30	0.15
34	1	1.07	0.22	0.11	0.73	0.39	0.19
	10	23.51	1.79	0.89	1.17	0.23	0.11
	100	650.31	119.53	59.77	6.11	2.89	1.45
35	1	1.19	0.47	0.24	1.36	0.77	0.38
	10	6.55	1.43	0.72	1.42	0.24	0.12
	100	429.01	42.71	24.66	5.89	0.46	0.26

18



19



20



 $\frac{21}{21}$



125 MHz ¹³C NMR of compound **10** in CDCl₃

22



23



24



25



26





28









31



100 MHz ¹³C NMR of compound 4 in CDCl₃

32



33





35



36



37



38



39



125 MHz ¹³C NMR of compound 22 in CDCl₃



41



42



43



S1



45



46



47



125 MHz ¹³C NMR of compound **28** in CDCl₃

48





500 MHz $^{\rm COSY}$ NMR of compound 28 in CDCl3





51



52





53







500 MHz $^{\rm COSY}$ NMR of compound 27 in CDCl3





57



125 MHz ¹³C NMR of compound 25 in CDCl₃















63





500 MHz ^{COSY} NMR of compound 26 in CDCl₃



500 MHz $^{\rm NOESY}$ NMR of compound 26 in CDCl₃