Supporting Information

Organocatalytic direct Mannich/cyclization cascade as [3 + 2] annulations: Asymmetric synthesis of 2, 3-substituted pyrrolidines

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General Experimental Methods:

All reactions under standard conditions were monitored by thin-layer chromatography (TLC) on SiO₂ gel F254 plates. The column chromatography was performed on silica gel (100-200 meshes) using EtOAc and petroleum ether with the distillation range of 60-80 °C. All other reagents were of analytical grade and used without further purification. ¹H and ¹³C NMR spectra were recorded in CDCl₃ solution and spectral data were reported in ppm relative to tetramethylsilane (TMS) as internal standard. High resolution mass spectra were recorded using quadrupole electrospray ionization (ESI) technique. HPLC was performed on Thermo Finnigan instrument using chiral Pack OD-H column and *i*-PrOH/Hexane solvent system.

General procedure for the organocatalytic Mannich/intramolecular cyclization /aromatization reaction cascade:

Succinaldehyde **3** (0.3 mL, 0.9 mmol, 3M solution) was added to a mixture of preformed *N*-PMP aldimine **2** (0.3 mmol) and L-proline (7.0 mg, 0.06 mmol) in DMSO (3.0 mL) at room temperature. The reaction mixture was stirred at 5 $^{\circ}$ C until the aldimine was consumed as monitored by TLC. The reaction was worked up by addition of saturated NaHCO₃ solution (3 mL) and extracted with ethyl acetate with three times. The combined organic extracts were washed with brine one time, dried over anhydrous Na₂SO₄ and concentrated in vacuum after filtration.

The crude adduct was taken in MeOH (3 mL) and CH_3CO_2H (50 mol%, 9 µL) and then NaBH₄ was cautiously at 0 °C and further stirred for 3 h and allows it come to room temperature. The reaction was subsequently quenched with saturated NaHCO₃ solution (3 mL). The aqueous solution was extracted with ethyl acetate twice and combined organic extracts were washed with brine once and dried over anhydrous Na₂SO₄ and concentrated in vacuum after filtration. Purification by silica gel column chromatography (hexane: EtOAc) gave *trans*-2,3-disubstituted pyrrolidines **6** with 56-78% yields.

The enantiomeric excess (ee) of products was determined by HPLC analysis using chiral OD-H column and absolute configuration could not be established by the single crystal X-ray of **6b**. The model chosen for refinement has C2-S, and C3-S stereochemistry, as expected through the well documented *syn*-selective direct Mannich reaction catalyzed by L-proline.

((2S, 3S)-1-(4-methoxyphenyl)-2-(2-nitrophenyl)pyrrolidin-3-yl)methanol (6a)

6a: ¹H NMR (400 MHz, CDCl₃) δ 2.03-2.14 (m, 2H), 2.31-2.36 (m, 1H), 3.49-3.53 (m, 1H), 3.56-3.61 (m, 1H), 3.66-3.86 (m, 1H), 3.70 (s, 3H), 3.88-3.92 (m, 1H), 5.08 (d, *J* = 3.0 Hz, 1H), 6.37 (d, *J* = 8.8 Hz, 2H), 6.76 (d, *J* = 8.8 Hz, 1H), 7.39 (t, *J* = 7.7 Hz, 2H), 7.50 (t, *J* = 7.7 Hz, 2H), 8.02 (d, *J* = 8.5 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 24.59, 47.39, 50.14, 55.74, 62.21, 63.80, 112.71 (2C), 114.91 (2C), 124.84, 128.25, 129.58, 134.00, 139.60, 140.68, 147.85, 151.18;

HRMS (ESI): Calcd for C₁₈H₂₀N₂O₄ (MH⁺) 329.1501, Found 329.1497.

 $[\alpha]_D^{22} = +18.6 (c \ 1.0, \text{CHCl}_3, 98\% \text{ ee})$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 85:15), 1.0 mL/min; minor enantiomer $t_R = 11.347$ min, major enantiomer $t_R = 14.273$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(3-nitrophenyl)pyrrolidin-3-yl)methanol (6b)

6b: ¹H NMR (400 MHz, CDCl₃) δ 1.88-1.95 (m, 1H), 2.13-2.23 (m, 1H), 2.30-2.37 (m, 1H), 3.45 (dd, J = 16.3 Hz, 8.8 Hz, 1H), 3.69 (m, 2H), 3.71 (s, 3H), 3.79 (dt, J = 8.5 Hz, 3.2 Hz, , 1H), 4.63 (d, J = 2.7 Hz, 1H), 6.41 (d, J = 9.0 Hz, 2H), 6.75 (d, J = 9.1 Hz, 2H), 7.47 (t, J = 8.1 Hz, 2H), 7.60 (d, J = 7.8 Hz, 1H), 8.08 (d, J = 8.0 Hz, 1H), 8.14 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 25.49, 48.56, 51.49, 55.77, 63.84, 65.21, 113.43 (2C), 114.86(2C), 121.02, 121.95, 129.54, 132.27, 141.39, 147.11, 148.68, 151.39; HRMS (ESI): Calcd for C₁₈H₂₀N₂O₄ (MH⁺) 329.1501, Found 329.1512.

 $[\alpha]_{D}^{22} = -50.9 (c \ 1.0, \text{CHCl}_{3}, 92\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 70:30), 1.0 mL/min; minor enantiomer $t_R = 7.223$ min, major enantiomer $t_R = 10.713$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(4-nitrophenyl)pyrrolidin-3-yl)methanol (6c)

6c: ¹H NMR (400 MHz, CDCl₃) δ 1.88-1.94 (m, 1H), 2.11-2.21 (m, 1H), 2.31-2.37 (m, 1H), 3.46 (dd, J = 16.3 Hz, 8.8 Hz, 1H), 3.68 (m, 2H), 3.70 (s, 3H), 3.75 (dt, J = 8.8 Hz, 3.3 Hz, 1H), 4.63 (d, J = 2.8 Hz, 1H), 6.38 (d, J = 9.1 Hz, 2H), 6.74 (d, J = 9.0 Hz, 2H), 7.42 (d, J = 8.8 Hz, 2H), 8.14 (d, J = 8.8 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 25.51, 48.43, 51.37, 55.75, 63.79, 65.22, 113.29 (2C), 114.84 (2C), 123.88 (2C), 126.86 (2C), 141.27, 146.84, 151.28, 152.52;

HRMS (ESI): Calcd for C₁₈H₂₀N₂O₄ (MH⁺) 329.1501, Found 329.1505.

 $[\alpha]_D^{22} = -79.5 (c \ 1.0, \text{CHCl}_3, 96\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 70:30), 1.0 mL/min; minor enantiomer $t_R = 8.060$ min, major enantiomer $t_R = 11.660$ min.

((2S, 3S)- 2-(2-chlorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6d)

6d: ¹H NMR (300 MHz, CDCl₃) δ 2.01-2.12 (m, 1H), 2.37-2.39 (m, 1H), 3.51 (dd, J = 16.5 Hz, 9.2 Hz, 1H), 3.61-3.67 (m, 1H), 3.70 (s, 3H), 3.72-3.78 (m, 1H), 3.84-3.89 (m, 1H), 4.74 (d, J = 2.4 Hz, 1H), 6.35 (d, J = 8.8 Hz, 2H), 6.75 (d, J = 8.8 Hz, 2H), 7.16-7.21 (m, 3H), 7.38 (d, J = 6.9 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 24.88, 47.72, 49.57, 55.82, 63.08, 64.20, 112.87 (2C), 114.91 (2C), 127.05, 127.55, 128.14, 129.75, 132.20, 140.75, 141.16, 151.07;

HRMS (ESI): Calcd for C₁₈H₂₀ClNO₂ (MH⁺) 318.1261, Found: 318.1259.

 $[\alpha]_D^{22} = +11.6 (c \ 1.0, \text{CHCl}_3, >99\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 9.726$ min, major enantiomer $t_R = 14.433$ min.

((2S, 3S)- 2-(3-chlorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6e)

6e: ¹H NMR (300 MHz, CDCl₃) δ 1.84-1.89 (m, 1H), 2.14-2.21 (m, 1H), 2.28-2.37 (m, 1H), 3.40 (dd, 16.5 Hz, J = 8.6 Hz, 1H), 3.60-3.65 (m, 2H), 3.70 (s, 4H), 4.47 (d, J = 2.1 Hz, 1H), 6.40 (d, J = 8.8 Hz, 2H), 6.74 (d, J = 8.8 Hz, 2H), 7.12-7.25 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 25.43, 48.32, 51.45, 55.79, 63.96, 65.29, 113.23 (2C), 114.79 (2C), 124.14, 126.03, 126.90, 129.85, 134.47, 141.65, 146.80, 151.07;

HRMS (ESI): Calcd for C₁₈H₂₀ClNO₂ (MH⁺) 318.1261, Found: 318.1265.

 $[\alpha]_D^{22} = -33.5 (c \ 1.0, \text{CHCl}_3, 92\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 92:08), 1.3 mL/min; minor enantiomer $t_R = 13.105$ min, major enantiomer $t_R = 33.857$ min.

((2S, 3S)- 2-(4-chlorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6f)

6f: ¹H NMR (300 MHz, CDCl₃) δ 1.84-1.91 (m, 1H), 2.11-2.20 (m, 1H), 2.26-2.33 (m, 1H), 3.42 (dd, J = 16.3 Hz, 8.8 Hz, 1H), 3.61-3.67 (m, 2H), 3.68-3.72 (m, 1H) 3.70 (s, 3H), 4.47 (d, J = 2.7 Hz, 1H), 6.38 (d, J = 9.1 Hz, 2H), 6.73 (d, J = 9.1 Hz, 2H), 7.17 (d, J = 8.6 Hz, 2H), 7.24 (d, J = 8.6 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 25.53, 48.38, 51.58, 55.82, 64.12, 65.10, 113.28 (2C), 114.82 (2C), 127.39 (2C), 128.73 (2C), 132.30, 141.65, 142.90, 151.12;

HRMS (ESI): Calcd for C₁₈H₂₀ClNO₂ (MH⁺) 318.1261, Found: 318.1273.

 $[\alpha]_D^{22} = -49.0 (c \ 1.0, \text{CHCl}_3, 90\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 92:08), 1.3 mL/min; minor enantiomer $t_R = 12.338$ min, major enantiomer $t_R = 29.488$ min.

((2S, 3S)- 2-(2-fluorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6g)

6g: ¹H NMR (300 MHz, CDCl₃) δ 1.96-2.00 (m, 1H), 2.10-2.18 (m, 1H), 2.36-2.39 (m, 1H), 3.44-3.47 (m, 1H), 3.62-3.67 (m, 2H), 3.70 (s, 3H), 3.73-3.78 (m, 1H), 4.76 (d, *J* = 2.4 Hz, 1H), 6.41 (d, *J* = 8.8 Hz, 2H), 6.75 (d, *J* = 8.8 Hz, 2H), 7.01 (t, *J* = 8.0 Hz, 1H), 7.06 (t, *J* = 8.0 Hz, 1H), 7.13 (t, *J* = 7.6 Hz, 1H), 7.19 (t, *J* = 7.6 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 25.43, 47.89, 50.31, 55.82, 59.55, 64.21, 113.02 (2C), 114.86 (2C), 124.21, 126.26, 127.64, 128.27, 130.80, 133.00, 141.46, 151.10; HRMS (ESI): Calcd for C₁₈H₂₀FNO₂ (MH⁺) 302.1556, Found 302.1548.

 $[\alpha]_{D}^{22} = -11.5 (c \ 0.5, \text{CHCl}_{3}, 93\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 9.290$ min, major enantiomer $t_R = 18.738$ min.

((2S, 3S)- 2-(4-fluorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6h)

6h: ¹H NMR (300 MHz, CDCl₃) δ 1.84-1.89 (m, 1H), 2.12-2.19 (m, 1H), 2.26-2.32 (m, 1H), 3.42 (dd, J = 16.5 Hz, 8.8 Hz, 1H), 3.60-3.68 (m, 2H), 3.70 (s, 4H), 4.48 (d, J = 2.8 Hz, 1H), 6.41 (d, J = 8.8 Hz, 2H), 6.75 (d, J = 8.8 Hz, 2H), 6.98 (t, J = 8.5 Hz, 2H), 7.21 (dd, J = 5.5 Hz, 8.8 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 25.45, 48.30, 51.58, 55.78, 64.02, 64.96, 113.21 (2C), 114.77 (2C), 115.20, 115.41, 127.42, 139.87, 141.71, 150.95, 160.42, 160.85;

HRMS (ESI): Calcd for $C_{18}H_{20}FNO_2$ (MH⁺) 302.1556, Found 302.1562.

 $[\alpha]_D^{22} = -37.9 (c \ 1.0, \text{CHCl}_3, 91\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 11.775$ min, major enantiomer $t_R = 28.242$ min.

((2S, 3S)- 2-(4-bromophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6i)

6i: ¹H NMR (300 MHz, CDCl₃) δ 1.83-1.90 (m, 1H), 2.10-2.19 (m, 1H), 2.26-2.31 (m, 1H), 3.43 (dd, *J* = 16.3 Hz, 8.6 Hz, 1H), 3.60-3.67 (m, 2H), 3.70 (s, 4H), 4.46 (d, *J* = 2.2 Hz, 1H), 6.40 (d, *J* = 9.1 Hz, 2H), 6.75 (d, *J* = 9.1 Hz, 2H), 7.12 (d, *J* = 8.3 Hz, 2H), 7.40 (dd, *J* = 5.5 Hz, 8.3 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 25.53, 48.39, 51.54, 55.83, 64.10, 65.15, 113.29 (2C), 114.84 (2C), 127.80 (2C), 128.59, 131.66 (2C), 141.63, 143.46, 151.13;

HRMS (ESI): Calcd for C₁₈H₂₀BrNO₂ (MH⁺) 362.0755, Found 362.0759.

 $[\alpha]_D^{22} = -43.2 (c \ 1.0, \text{CHCl}_3, 90\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 92:08), 1.3 mL/min; minor enantiomer $t_R = 12.542$ min, major enantiomer $t_R = 33.592$ min.

((2S, 3S)- 2-(3-bromo-4-fluorophenyl)-1-(4-methoxyphenyl)pyrrolidin-3-yl)methanol (6j)

6j: ¹H NMR (300 MHz, CDCl₃) δ 1.83-1.88 (m, 1H), 2.10-2.17 (m, 1H), 2.25-2.28 (m, 1H), 3.40 (dd, J = 18.1 Hz, 9.5 Hz, 1H), 3.61 (d, J = 7.3 Hz, 2H), 3.70 (s, 4H), 4.46 (bs, 1H), 6.40 (d, J = 8.8 Hz, 2H), 6.75 (d, J = 8.8 Hz, 2H), 7.03 (t, J = 8.5 Hz, 1H), 7.14-7.17 (m, 1H), 7.45 (dd, J = 6.5 Hz, 2.0 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 25.27, 48.23, 51.37, 55.73, 63.40, 64.58, 113.21 (2C), 114.76 (2C), 126.29, 127.25, 130.68, 131.72, 141.49, 150.99, 156.45, 158.89;

HRMS (ESI): Calcd for C₁₈H₁₉BrFNO₂ (MH⁺) 380.0661, Found 380.0667.

 $[\alpha]_D^{22} = -19.2 (c \ 1.0, \text{CHCl}_3, 92\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 92:08), 1.3 mL/min; minor enantiomer $t_R = 12.560$ min, major enantiomer $t_R = 33.893$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-phenylpyrrolidin-3-yl)methanol (6k)

6k: ¹H NMR (300 MHz, CDCl₃) δ 1.85-1.90 (m, 1H), 2.18-2.24 (m, 1H), 2.31-2.37 (m, 1H), 3.41-3.48 (dd, *J* = 17.0 Hz, 8.8 Hz, 1H), 3.62-3.73 (m, 6H), 4.49 (d, *J* = 2.8 Hz, 1H), 6.41 (d, *J* = 8.8 Hz, 2H), 6.72 (d, *J* = 8.8 Hz, 2H), 7.18-7.31 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ 25.56, 48.34, 51.62, 55.83, 64.31, 65.65, 113.18 (2C), 114.79 (2C), 125.94 (2C), 126.70, 128.57 (2C), 141.87, 144.28, 150.89;

HRMS (ESI): Calcd for C₁₈H₂₁NO₂ (MH⁺) 284.1650, Found: 284.1660.

 $[\alpha]_D^{22} = -41.2 (c \ 1.0, \text{CHCl}_3, 90\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 13.180$ min, major enantiomer $t_R = 29.432$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(naphthalen-1-yl)pyrrolidin-3-yl)methanol (6l)

61: ¹H NMR (300 MHz, CDCl₃) δ 1.88-1.93 (m, 1H), 2.13-2.23 (m, 1H), 2.48-2.53 (m, 1H), 3.523.59 (dd, J = 16.1 Hz, 9.1 Hz, 1H), 3.68 (s, 3H), 3.71-3.86 (m, 3H), 5.39 (bs, 1H), 6.38 (d, J = 8.8 Hz, 2H), 6.71 (d, J = 8.8 Hz, 2H), 7.24 (d, J = 6.8 Hz, 1H), 7.34 (t, J = 7.2 Hz, 1H), 7.50-7.60 (m, 2H), 7.72 (d, J = 8.0 Hz, 1H), 7.89 (d, J = 8.6 Hz, 1H), 8.40 (d, J = 8.3 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 25.19, 47.23, 49.10, 55.85, 62.39, 64.49, 112.64 (2C), 114.93 (2C), 123.24, 123.61, 125.53, 125.95, 127.40, 128.82, 128.92, 130.65, 134.16, 138.18, 141.40, 150.75;

HRMS (ESI): Calcd for C₂₂H₂₃NO₂ (MH⁺): 334.1807, Found 334.1826.

 $[\alpha]_{D}^{22} = +7.3 (c \ 1.0, \text{CHCl}_{3}, 97\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 15.240$ min, major enantiomer $t_R = 27.638$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(naphthalen-2-yl)pyrrolidin-3-yl)methanol (6m)

6m: ¹H NMR (300 MHz, CDCl₃) δ 1.89-1.94 (m, 1H), 2.21-2.26 (m, 1H), 2.40-2.44 (m, 1H), 3.46 (dd, J = 16.5 Hz, 8.5 Hz, 1H), 3.68 (bs, 4H), 3.71-3.77 (m, 2H), 4.64 (d, J = 2.2 Hz, 1H), 6.47 (d, J = 8.8 Hz, 2H), 6.71 (d, J = 8.8 Hz, 2H), 7.40-7.46 (m, 3H), 7.75-3.81 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ

25.63, 48.51, 51.49, 55.82, 64.28, 65.96, 113.31 (2C), 114.81 (2C), 124.38, 124.49, 125.44, 126.00,

127.64, 127.78, 128.31, 128.49, 132.67, 133.48, 141.99, 150.98;

HRMS (ESI): Calcd for $C_{22}H_{23}NO_2$ (MH⁺): 334.1807, Found 334.1815.

 $[\alpha]_D^{22} = -19.6 (c \ 0.5, \text{CHCl}_3, 90\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.0 mL/min; minor enantiomer $t_R = 18.290$ min, major enantiomer $t_R = 37.092$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(pyridin-2-yl)pyrrolidin-3-yl)methanol (6n)

6n: ¹H NMR (300 MHz, CDCl₃) δ 1.77-1.85 (m, 1H), 2.06-2.14 (m, 1H), 2.47-2.54 (m, 1H), 3.56 (dd, *J* = 16.3 Hz, 8.4 Hz, 1H), 3.71 (bs, 4H), 3.78-3.80 (m, 2H), 4.64 (d, *J* = 5.5 Hz, 1H), 6.40 (d, *J* = 8.8 Hz, 2H), 6.74 (d, *J* = 8.8 Hz, 2H), 7.13 (d, *J* = 8.1 Hz, 2H), 7.57-3.81 (dt, *J* = 7.8 Hz, 1.4 Hz, 1H), 8.55 (d, *J* = 8.7 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 26.49, 48.95, 49.80, 55.73, 64.91, 68.01, 113.46 (2C), 114.75 (2C), 120.24, 121.89, 137.31, 141.66, 148.83, 151.14, 163.33;

HRMS (ESI): Calcd for $C_{17}H_{20}N_2O_2$ (MH⁺) 285.1603, Found: 285.1635.

 $[\alpha]_D^{22} = -37.7 (c \ 1.0, \text{CHCl}_3, 95\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 80:20), 1.0 mL/min; major enantiomer $t_R = 10.297$ min, minor enantiomer $t_R = 14.238$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(pyridin-4-yl)pyrrolidin-3-yl)methanol (60)

60: ¹H NMR (300 MHz, CDCl₃) δ 2.26-2.34 (m, 1H), 2.37-2.43 (m, 1H), 2.96-2.98 (m, 1H), 3.33 (dd, J = 16.1 Hz, 9.6 Hz 1H), 3.71 (s, 4H), 3.74-3.82 (m, 2H), 5.02 (d, J = 2.2 Hz, 1H), 6.40 (d, J = 8.8 Hz, 2H), 6.75 (d, J = 8.8 Hz, 2H), 7.21 (d, J = 5.9 Hz, 2H), 8.53 (d, J = 4.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 23.88, 29.64, 48.15, 55.71, 59.88, 61.60, 113.58 (2C), 114.87 (2C), 121.28 (2C), 140.65, 149.99 (2C), 151.83, 152.70;

HRMS (ESI): Calcd for $C_{17}H_{20}N_2O_2$ (MH⁺) 285.1603, Found: 285.1608.

 $[\alpha]_D^{22} = +7.2 (c \ 1.0, \text{CHCl}_3, 98\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 80:20), 1.0 mL/min; minor enantiomer $t_R = 12.457$ min, major enantiomer $t_R = 16.562$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(thiophen-2-yl)pyrrolidin-3-yl)methanol (6p)

6p: ¹H NMR (300 MHz, CDCl₃) δ 1.87-1.94 (m, 1H), 2.25-2.34 (m, 1H), 2.42-2.48 (m, 1H), 3.36 (dd, J = 16.5 Hz, 8.3 Hz 1H), 3.62-3.68 (m, 3H), 3.71 (s, 3H), 4.76 (d, J = 2.2 Hz, 1H), 6.53 (d, J = 8.8 Hz, 2H), 6.75 (d, J = 8.8 Hz, 2H), 6.91-6.93 (m, 2H), 7.12 (dd, J = 1.7 Hz, 4.7 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 2 432, 33.76, 45.13, 56.78, 59.46, 62.76, 113.58, 114.87, 124.54, 124.88, 130.24, 135.04, 142.08, 151.76;

HRMS (ESI): Calcd for C₁₆H₁₉NO₂S (MH⁺) 290.1212, Found 390.1225.

 $[\alpha]_D^{22} = -15.8 (c \ 1.0, \text{CHCl}_3, 95\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 92:08), 1.3 mL/min; minor enantiomer $t_R = 14.488$ min, major enantiomer $t_R = 32.368$ min.

((2S, 3S)-1-(4-methoxyphenyl)-2-(5-nitrofuran-2-yl)pyrrolidin-3-yl)methanol (6q)

6q: ¹H NMR (400 MHz, CDCl₃) δ 1.94-2.00 (m, 1H), 2.16-2.23 (m, 1H), 2.60-2.66 (m, 1H), 3.38 (dd, J = 16.1 Hz, 8.5 Hz, 1H), 3.56-3.68 (m, 3H), 3.72 (s, 3H), 4.70 (bs, 1H), 6.29 (d, J = 3.8 Hz, 1H), 6.48 (d, J = 8.8 Hz, 2H), 6.78 (d, J = 8.8 Hz, 2H), 7.19 (d, J = 3.8 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 25.74, 47.52, 48.04, 55.77, 59.49, 63.62, 109.90, 112.79, 113.36 (2C), 114.92 (2C), 140.91, 151.61, 151.81, 160.91;

HRMS (ESI): Calcd for $C_{16}H_{18}N_2O_4$ (MH⁺) 319.1294, Found 334.1296.

 $[\alpha]_D^{22} = -58.8 (c \ 0.5, \text{CHCl}_3, 96\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 85:15), 1.3 mL/min; minor enantiomer t_R = 19.330 min, major enantiomer t_R = 26.727 min.

((2S, 3S)-1-(4-methoxyphenyl)-2-((E)-styryl)pyrrolidin-3-yl)methanol (6r)

6r: ¹H NMR (400 MHz, CDCl₃) δ 1.84-1.90 (m, 1H), 2.18-2.24 (m, 1H), 2.32-2.36 (m, 1H), 3.34 (dd, J = 16.5 Hz, 8.3 Hz, 1H), 3.52-3.68 (m, 3H), 3.73 (s, 3H), 4.10 (bs, 1H), 6.20 (dd, J = 15.9 Hz, 5.8 Hz, 1H), 6.48 (d, J = 15.9 Hz, 1H), 6.59 (d, J = 8.8 Hz, 2H), 6.80 (d, J = 8.8 Hz, 2H), 7.26-7.37 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ 25.65, 48.10, 55.75, 60.32, 63.42, 63.77, 113.17 (2C), 114.67 (2C), 126.22 (2C), 127.16, 128.37 (2C), 129.69, 131.61, 136.77, 142.20, 150.81;

HRMS (ESI): Calcd for $C_{20}H_{23}NO_2$ (MH⁺) 310.1807, Found 310.1819.

 $[\alpha]_D^{22} = -54.3 (c \ 1.0, \text{CHCl}_3, 95\% \text{ ee});$

Enantiomeric excess was determined by HPLC with a Chiralpak OD-H column (*n*-Hexane: *i*-PrOH = 90:10), 1.3 mL/min; minor enantiomer $t_R = 13.307$ min, major enantiomer $t_R = 27.058$ min.

Single Crystal X-ray data for ((2*S*, 3*S*)-1-(4-methoxyphenyl)-2-(3-nitrophenyl)pyrrolidin-3-yl)methanol (**6b**): [(CCDC no: **864411**)]

The title compound, ((2S, 3S)-1-(4-methoxyphenyl)-2-(3-nitrophenyl)pyrrolidin-3yl)methanol (C₁₈H₂₀N₂O₄), crystallizes in the monoclinic space group P2₁ with the following unit-cell parameters: a= 7.2761(4), b=7.6312(5), c = 14.6614(10)Å, β = 95.224(5)^o and Z = 2. The crystal structure was solved by direct methods and refined by full-matrix least-squares procedures to a final R-value of 0.0467 for 2266 observed reflections. The pyrrolidine ring is in envelope conformation and makes a dihedral angle of 11.42(8)^o with the phenyl ring (C6-C11). Phenyl ring (C15-C20) is almost perpendicular to the pyrrolidine ring (dihedral angle 88.78(8)^o). The molecules in the unit cell are arranged in layers and are stabilized by O-H...O, C-H...O hydrogen bonds and C-H... π interactions.

The X-ray intensity data of a well defined crystal (0.30 x 0.20 x 0.10 mm) were collected at room temperature (293K) by using a CCD area-detector diffractometer (*X'calibur system* – *Oxford diffraction make*, *U.K.*) which is equipped with graphite monochromated MoK α radiation (λ =0.71073 Å). The cell dimensions were determined by least-squares fit of angular settings of 4103 reflections in the θ range 3.7698 to 29.0804 °. A total number of 15385 reflections were collected of which 2266 reflections were treated as observed (I > 2 σ (I)). Data were corrected for Lorentz and polarization and absorption factors.

The structure was solved by direct methods using SHELXS97 (Sheldrick, 2008). All nonhydrogen atoms of the molecule were located from the E-map. Full-matrix least-squares refinement was carried out by using SHELXL97 software (Sheldrick, 2008). All the hydrogen atoms were located from a difference electron density map and their positional and isotropic thermal parameters were included in the refinement. The final refinement cycle yielded an Rfactor of 0.0467 (wR (F^2) = 0.0920) for the observed data. The residual electron density ranges from -0.145 to 0.189 eÅ⁻³. Atomic scattering factors were taken from International Tables for Xray Crystallography (1992, Vol. C, Tables 4.2.6.8 and 6.1.1.4). The crystallographic data are summarized in Table 1. The CIF for this structure has been deposited at Cambridge Crystal Data Centre (**CCDC no: 864411**).



Fig. 1: *ORTEP* view of the molecule, showing the atom-labelling scheme. Displacement ellipsoids are drawn at the 40% probability level and H atoms are shown as small spheres of arbitrary radii.



Fig. 2: *ORTEP* view of the molecule, showing the atom-labelling scheme. Displacement ellipsoids are drawn at the 40% probability level and H atoms are shown as small spheres of arbitrary radii.



Fig. 3 The packing arrangement of molecules viewed down the b-axis.



Fig. 4 The packing arrangement of molecules viewed down the a-axis.

Crystal data and other experimental details are given in Table 1. The final atomic coordinates with equivalent isotropic displacement parameters are presented in Table 2. An ORTEP view of the title compound with atomic labeling at different rotations is shown in Fig.1, and Fig.2 (Farrugia, 1997). Bond distances and angles in ((2S, 3S)-1-(4-methoxyphenyl)-2-(3-nitrophenyl)pyrrolidin-3-yl)methanol are in agreement with the theoretical values (Allen, et al., 1987). The pyrrolidine ring is in envelope conformation and makes a dihedral angle of 11.42(8)° with the phenyl ring (C6-C11). Phenyl ring (C15-C20) is almost perpendicular to the pyrrolidine ring (dihedral angle 88.78(8)°). Pyrrolidine has *envelope* conformation with the best mirror plane passing through C3 bisecting the opposing bond (N1-C5). The asymmetry parameter is: ΔC_s (C3) = 5.6.

CCDC	864411
Crystal description	Red coloured block shaped
Crystal size	0.30 x 0.20 x 0.10 mm
Empirical formula	$C_{18}H_{20}N_2O_4$
Formula weight	328.36
Measurement	X'calibur system–Oxford diffraction make, U.K.
Radiation, Wavelength	MoKα, 0.71073 Å
Cell measurement Temperature	293(2) K
Unit cell dimensions	a= 7.2761(4), b= 7.6312(5)
	$c = 14.6614(10)$ Å; $\beta = 95.224(5)^{\circ}$
Crystal system	Monoclinic
Space group	P21
Unit cell volume	810.70(9)Å ³
Density (calculated)	1.345Mgm ⁻³
No. of molecules per unit cell, Z	2

Table 1. Crystal data and other experimental details

Absorption coefficient	0.096 mm^{-1}
F(000)	348
Refinement of unit cell	4103 reflections for 3.7698< θ < 29.0804 °
Scan mode	w scans
θ range for entire data collection	$3.78 < \theta < 26.00$ °
Range of indices	h = -8 to 8, $k = -9$ to 9, $l = -18$ to 18
Reflections collected / unique	15385 / 3157
Reflections observed (I > $2\sigma(I)$)	2266
R _{int}	0.0568
R _{sigma}	0.0510
Structure determination	Direct methods
Refinement	Full-matrix least-squares on F ²
No. of parameters refined	218
Final R- factor	0.0467
$wR(F^2)$	0.0920
Goodness-of-fit	1.041
Final residual electron density	$-0.145 < \Delta \rho < 0.189 \text{ eÅ}^{-3}$
Software for structure solution:	SHELXS97 (Sheldrick, 2008)
Software for refinement:	SHELXL97 (Sheldrick, 2008)
Software for molecular plotting:	ORTEP-3 (Farrugia,1997)
	PLATON (Spek, 2003)
Software for geometrical calculations	PLATON (Spek, 2003)
	PARST (Nardelli, 1995)

Atom	x	у	Z	U _{eq} *
N1	1.0787(2)	0.0975(3)	0.68520(13)	0.0424(5)
N2	0.7352(3)	0.4990(4)	0.89068(19)	0.0657(7)
01	0.7742(4)	0.5841(3)	0.82457(18)	0.0891(8)
O2	0.6562(4)	0.5637(4)	0.95255(17)	0.1014(9)
O3	0.5440(2)	0.0632(2)	0.38814(12)	0.0471(5)
C2	1.0778(3)	-0.0012(4)	0.76919(15)	0.0403(6).
C3	1.2822(3)	0.0047(4)	0.80865(17)	0.0480(7)
C4	1.3448(4)	0.1843(4)	0.77611(19)	0.0547(7)
C5	1.2409(3)	0.2070(4)	0.68191(18)	0.0515(7)
C6	0.9425(3)	0.0850(3)	0.61276(16)	0.0337(5)
C7	0.7851(3)	-0.0171(3)	0.61785(16)	0.0379(6)
C8	0.6508(3)	-0.0272(3)	0.54415(16)	0.0395(6)
C9	0.6699(3)	0.0638(3)	0.46454(17)	0.0370(6)
C10	0.8273(3)	0.1650(3)	0.45874(17)	0.0410(6)
C11	0.9602(3)	0.1765(3)	0.53124(16)	0.0397(6)
C12	0.3709(3)	-0.0208(4)	0.39648(19)	0.0562(8)
C13	1.3947(4)	-0.1399(4)	0.7705(2)	0.0570(8)
O14	1.3184(3)	-0.3062(3)	0.78774(14)	0.0715(6)
C15	0.9513(3)	0.0747(4)	0.83694(17)	0.0407(6)
C16	0.8968(3)	0.2480(4)	0.83184(18)	0.0455(7)
C17	0.7884(3)	0.3143(4)	0.89726(19)	0.0490(7)
C18	0.7304(4)	0.2131(5)	0.9660(2)	0.0598(8)
C19	0.7860(4)	0.0413(5)	0.9714(2)	0.0660(9)
C20	0.8957(3)	-0.0275(4)	0.90732(18)	0.0523(7)

Table 2. Atomic coordinates and equivalent isotropic thermal parameters ($Å^2$) for non-hydrogen atoms (e.s.d.'s are given in parenthesis)

N1- C6	1.388(3)		N1 -C2	1.444(3)
N1 -C5	1.450(3)		N2 -O1	1.221(3)
N2 -O2	1.222(3)		N2 -C17	1.463(4)
O3 -C9	1.381(3)		O3 -C12	1.429(3)
C2 -C15	1.528(3)		C2 -C3	1.546(3)
C3 -C13	1.511(4)		C3 -C4	1.535(4)
C4 -C5	1.523(4)		C6 -C7	1.393(3)
C6 -C11	1.400(3)		C7 -C8	1.391(3)
C8 -C9	1.376(3)		C9 -C10	1.390(3)
C10- C11	1.373(3)		C13 -O14	1.418(4)
C15 -C16	1.381(4)		C15 -C20	1.383(4)
C16 -C17	1.391(3)		C17 -C18	1.367(4)
C18 -C19	1.373(4)		C19 -C20	1.389(4)
Bond angles				
C6- N1- C2	123.94(19)		C6 -N1 -C5	122.7(2)
C2 -N1 -C5	113.31(19)		O1 -N2 -O2	122.2(3)
O1 -N2 -C17	119.1(3)		O2 -N2 -C17	118.7(3)
C9 -O3 -C12	117.20(18)		N1 -C2 -C15	114.0(2)
N1 -C2 -C3	103.00(18)		C15- C2 -C3	111.17(19)
C13 -C3 -C4	110.5(2)		C13 -C3 -C2	112.0(2)
C4 -C3 -C2	102.3(2)		C5 -C4 -C3	104.2(2)
N1 -C5 -C4	104.1(2)		N1 -C6 -C7	122.0(2)
N1 -C6 -C11	120.3(2)		C7 -C6 -C11	117.7(2)
C8 -C7 -C6	120.8(2)		C9 -C8 -C7	120.9(2)
C8 -C9 -O3	125.1(2)		C8 -C9 -C10	118.7(2)
		C40		

Table 3. Bond lengths (Å), Bond angles and torsion angles for non hydrogen atoms (e.s.d.'s are given in parentheses)

O3 -C9 -C10	116.2(2)	C11- C10- C9	120.9(2)
C10 -C11 -C6	121.0(2)	O14 -C13 -C3	110.7(2)
C16 -C15 -C20	118.6(2)	C16 -C15 -C2	121.0(2)
C20 -C15 -C2	120.4(2)	C15 -C16 -C17	119.3(3)
C18 -C17 -C16	122.4(3)	C18 -C17 -N2	119.7(3)
C16 -C17 -N2	117.9(3)	C17 -C18 -C19	118.2(3)
C18 -C19 -C20	120.5(3)	C15 -C20 -C19	121.1(3)
Torsion angles			
C5- N1 -C2 -C3	-18.0(3)	N1- C2- C3- C4	32.4(2)
C2 -C3 -C4 -C5	-35.7(2)	C2- N1- C5- C4	-4.4(3)
C3 -C4- C5 -N1	25.2(3)		

Table 3 Geometry of Intra and inter molecular hydrogen bond

D-HA D-	-H(Å)	HA(Å)	DA(Å)	θ [D–HA(°)]
C3-H3O2 ⁱ	0.9800	2.559(2)	3.518(4)	165.9(2)
C12-H12AO1 ⁱⁱ	0.9600	2.522(3)	3.411(4)	154.1(2)
C12 -H12CCg(2	$2)^{ii}$ 0.9600	2.7331	3.5128(3)	138.80
O14-H14O3 ⁱⁱⁱ	0.8200	2.414(2)	3.019(3)	131.3(1)
C4 -H4BCg(3) ^{i}	v 0.9700	3.1348	3.916(3)	138.65
C11 -H11Cg(2)	0.9300	2.8678	3.669(3)	145.10
C2-H2O14	0.9800	2.461(2)	2.910(3)	107.5(1)
C16-H16N1	0.9300	2.539(2)	2.866(3)	101.0(2)
Symmetry code (i)	-x+2,+y-1/2,-	z+2 (ii) $-x+1,+y$	z-1/2,-z+1 (iii) -x+2,	+y-1/2,-z+1





Racemic 6a

Retention Time	Area	Area %
11.513	39186688	51.15
14.593	37424555	48.85
Totals	76611243	100.00









































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