

## Supporting information

### Signal-on electrochemical Y or junction probe detection of nucleic acid

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## Experimental Section

### Materials

All unlabeled DNA oligonucleotides were synthesized in house whereas labeled oligonucleotides were purchased from Biosearch Technology. Restriction endonucleases were purchased from Fermenetas Inc. USA. 6-Mercaptohexanol and Tris-(2-carboxyethyl) phosphine hydrochloride (TCEP) were purchased from Sigma-Aldrich. All chemical reagents were of analytical grade or higher. Ultrapure water (18.2 MΩ·cm) was used.

### Au Electrode surface cleaning and modification

Au electrodes (2mm in diameter, CH Instrument, Inc.) were cleaned before DNA self-assembly. The cleaning procedures were done according to published protocol (Nature Protocols, 2007, 2, 2875-2879). 200 μM Probe A (1 μL) was mixed with 10 mM TCEP (2 μL) at room temperature in the dark for 45 min. The mixture was then diluted with PBS buffer (10 mM phosphate-buffered saline pH 7.4 with 1M NaCl and 1mM Mg<sup>2+</sup>) to a final Probe A concentration of 200 nM. Each electrode was immersed with 100 μL of 200 nM stock Probe A in PBS for 30 min. After rinsing with deionized water, the electrodes were incubated in 2 mM 6-Mercaptohexanol solution for 3-5 h at room temperature in the dark. The electrodes were rinsed for 3 min using deionized water to remove any remaining 6-Mercaptohexanol solution. The electrodes were then soaked in PBS for 5 min before any electrochemical measurements.

### Square wave voltammetry measurements (SWV) of signal-off JP and different restriction endonucleases screening

All electrochemical measurements were performed with a CHI 660 electrochemical workstation (CH Instrument, Inc.). The normal three-electrode system consisted of Au working electrode, platinum wire counter electrode, and 3M NaCl saturated Ag/AgCl reference electrode. SWV was carried out in the 10 mM PBS (pH 7.4) from -0.1V to -0.4V with 0.001V interval, 60Hz frequency and 0.025 amplitude. Reaction condition: 1μL of restriction endonuclease (stock solution from company), 1 μL of 20 μM Probe B, 1 μL of 2 μM template, 10 μL of 10 x restriction endonuclease buffer, 87 μL of deionized water. Reaction time and temperature were 4 h and 31 °C respectively. Final concentrations: 200 nM Probe B, 20 nM template.

### Alternating current voltammetry measurements (ACV) of signal-on JP

System set up was the same as the SWV measurements. ACV was performed in the 10 mM PBS (pH 7.4) from -0.1V to -0.45V with 0.01 V interval, 50Hz frequency, 0.025 amplitude, 1 sec sample period.

**Table S1** Sequences of oligonucleotides used in this study.

Oligonucleotide	Sequences (5'->3')
Probe A1	5'-SH-C <sub>6</sub> H <sub>12</sub> -TTTCCACCGCCAATATTT <u>GATCTGTGG</u> (MB)-3'
Probe A2	5'-SH-C <sub>6</sub> H <sub>12</sub> -TTTCCT(MB)CCGCCAATATTTT <u>GATCTGAGG</u> -3'
Probe B (helper probe)	5'TG(S) <u>GATCGGAAAACC</u> (S) <u>GATCCA</u> (S) <u>GATCATATA</u> ACGTGCTGCTA-3'
Full match template	5'-TAGCAGCACGTA-AATATTGGCG-3'
Mismatch A template	5'-TAGCAGCACGTA-AATA <u>ATGGCG</u> -3'
Mismatch G template	5'-TAGCAGCACGTA-AATA <u>GTGGCG</u> -3'
Mismatch C template	5'-TAGCAGCACGTA-AATA <u>ACTGGCG</u> -3'

The locations of mismatched bases are colored blue.

Endonuclease cognate recognition site are highlighted in italic and underlined.

(S) denotes phosphorthioate linkage.