

Discrimination of colon cancer stem cells using noncanonical amino acid

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1. CSCs proportion in HCT-116

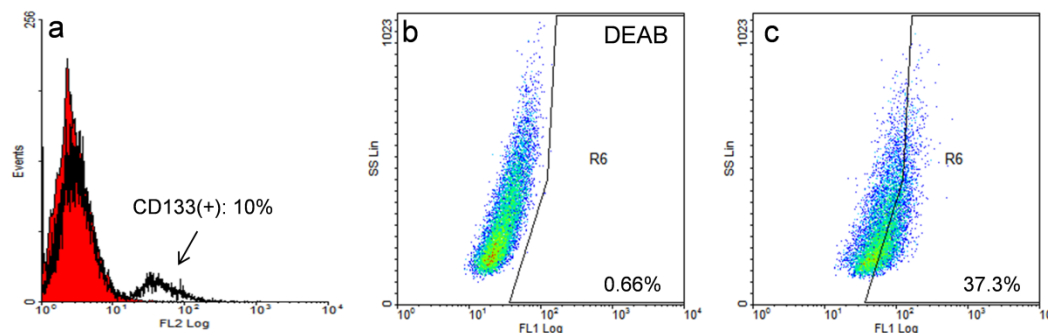


Figure S1 CSCs proportion in HCT-116 by using CD133 (a) and ALDEFLOUR assay (b, c).

2. Experimental section

Cell culture:

Colon cancer cell line HCT-116 was obtained from ATCC. Cells were maintained in Dulbecco's modified Eagle's medium (DMEM) and supplemented with 10% fetal bovine serum (FBS), 100 U/mL penicillin, 100 µg/mL streptomycin and 2 mM L-glutamine in a 5% CO₂ humidified incubator at 37°C. All media components were obtained from Hyclone laboratories Inc.

HPG labeling:

Cells were seeded at estimated 50% coverage on 6-well plate the 12 hours before labeling. HPG Labeling is accounting to our previous reports. In brief, Methionine-free medium supplemented with 2 mM L-glutaMAX replaced the culture medium 30 min before adding 100 µM HPG. After HPG incubation, 100 µM Met was added to medium for chasing and incubated for 2 hours.

Fluorescence microscopy:

After HPG labeling, Cells were rinsed twice with PBS, fixed with 4% paraformaldehyde and permeabilized with 0.1% Triton X-100 for 10 min. Cells were treated with blocking solution (1% BSA in PBS) at least 1 hour and rinsed twice with PBS. Cells were stained with 3-azido-7-hydroxycoumarin in the presence of 200 µM CuSO₄, 400µM TBTA, and 500 µM Sodium ascorbate in PBS(pH 7.4). The fluorescence was visualized under microscope (Olympus IX81) with DAPI filter set. Mercury lamp was used as a light source.

FACS and Flow cytometry:

FACS Cells were detached by Accutase (Invitrogen Inc). CD133 and ALDH1 enzyme activity was tested by using PE conjugated Anti-CD133 (Miltenyi Biotec) and ALDEFLOUR assay kit (STEMCELL Technologies, BC, Canada) according to manufacturer's manual. Each sample was filtered through a 40 µm Cell Strainers (Fisher Scientific Inc, GA) before sorting. Cells were sorted on BD FACSaria II cell sorter at 4 °C.

Flow cytometry

In brief, after HPG labelling cells were detached by Accutase (Invitrogen Inc). cells were rinsed with PBS and stain with primary anti-CD133 antibody and secondary anti-mouse FITC antibody. Then cells were fixed with 4% paraformaldehyde and were permeabilized with 0.1% Triton X-100 for 15 min each. Cells were treated with blocking solution (100 mM glycine in PBS) at least 1 hour. Cells were stained with 2 μM azido-Cy5 in the presence of 50 μM CuSO_4 , 50 μM L-histidine, 500 μM Sodium ascorbate in PBS (pH 7.4) ². Then samples were washed with PBS with 0.5% tween 20 three times. Each sample was filtered through a 40 μm Cell Strainers (Fisher Scientific Inc, GA) before analysis on Beckman Coulter FC500 Flow Cytometers.

3. Reference

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