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Rapid Thermally Assisted Donor-Acceptor Catenation

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Electronic Supporting Information

Table of Contents

1. General Methods
2. Synthetic Methods
3. X-Ray Crystallography
4. References

Experimental Section

1. General Methods

Starting materials and reagents were purchased from commercial suppliers and used as received. Compounds **S1**,^{S1} **1a**,^{S1} **1b**,^{S2} **1c**,^{S3} **1d**,^{S4} 1,1-[1,4-phenylenebis(methylene)]-bis(4,4-bipyridinium) bis(hexafluorophosphate)^{S5} (**2**•2PF₆) and the azido-bromomethyl-xylene derivative^{S6} **4** were all synthesised according to procedures reported in the literature. All reactions were performed in dry solvents. Both analytical and preparative high performance liquid chromatography (HPLC) were performed on reversed phase-HPLC (RP-HPLC) instruments, using C₁₈ columns and a binary solvent system, i.e., MeCN and H₂O with 0.1% (v / v) trifluoroacetic acid. All ¹H and ¹³C NMR spectra were recorded on a Bruker Avance 500 MHz spectrometer at ambient temperature with working frequencies of 500 MHz or 125 MHz, respectively. The chemical shifts are listed in ppm on the δ scale relative to the signals corresponding to the residual non-deuterated solvents (CD₃CN: δ 1.94 ppm; CDCl₃: 7.26 ppm), and the coupling constants are recorded in Hertz (Hz). The following abbreviations were used to explain the multiplicities: s, singlet; d, doublet; t, triplet; b, broad peaks; m, multiplet or overlapping peaks. High resolution electrospray ionisation (HRESI) mass spectra were measured on a Micromass Q-TOF Ultima mass spectrometer. Matrix assisted laser desorption and ionisation – time of flight (MALDI-TOF) mass spectrometry was performed, using a Bruker Autoflex III instrument. Low resolution electrospray ionisation (LRESI) mass spectra were measured on an Agilent 1100 MSD tandem liquid chromatography mass spectrometer.

2. Synthetic Methods

General synthetic procedure for preparing compounds 5/6a-d: Macrocycle **1a**, **1b**, **1c**, or **1d** (0.1 mmol) was mixed with a five-fold excess of 1,1-[1,4-phenylenebis-(methylene)]bis(4,4-bipyridinium) bis(hexafluorophosphate) **2**·2PF₆ (354 mg, 0.5 mmol) and 1,4-bis(bromomethyl)benzene **3** or its azide derivative **4** (0.5 mmol) in DMF (5 ml), and heated while stirring at 80°C for 1 h. The solution was filtered and injected into preparative RP-HPLC (C₁₈; MeCN – H₂O / 0 – 50 % in 45 min, with 0.1% trifluoroacetic acid) for purification. The pure fractions collected were subjected to counterion exchange in order to afford products as crystalline solids.

5a·4PF₆: Purple solid (152.5mg, 88%). ¹H NMR (500 MHz, CD₃CN, 298 K) δ = 8.86 (b, 4H), 8.39 (b, 4H), 8.05 (b, 4H), 7.87 (b, 4H), 7.14 (m, 4H), 6.89 (b, 8H), 6.35 (d, *J* = 7.8 Hz, 2H), 6.06 (d, *J* = 7.8 Hz, 2H), 5.81 (t, *J* = 7.8 Hz, 2H), 5.72 (m, 8H), 4.21–3.70 (b, 32H), 2.26 (d, *J* = 7.8 Hz, 2H) ppm. HRMS (ESI) calcd for C₇₂H₇₆F₂₄N₄O₁₀P₄ [*M* – 2PF₆]²⁺. *m/z*: 723.2422, found: 723.2449 [*M* – 2PF₆]²⁺.

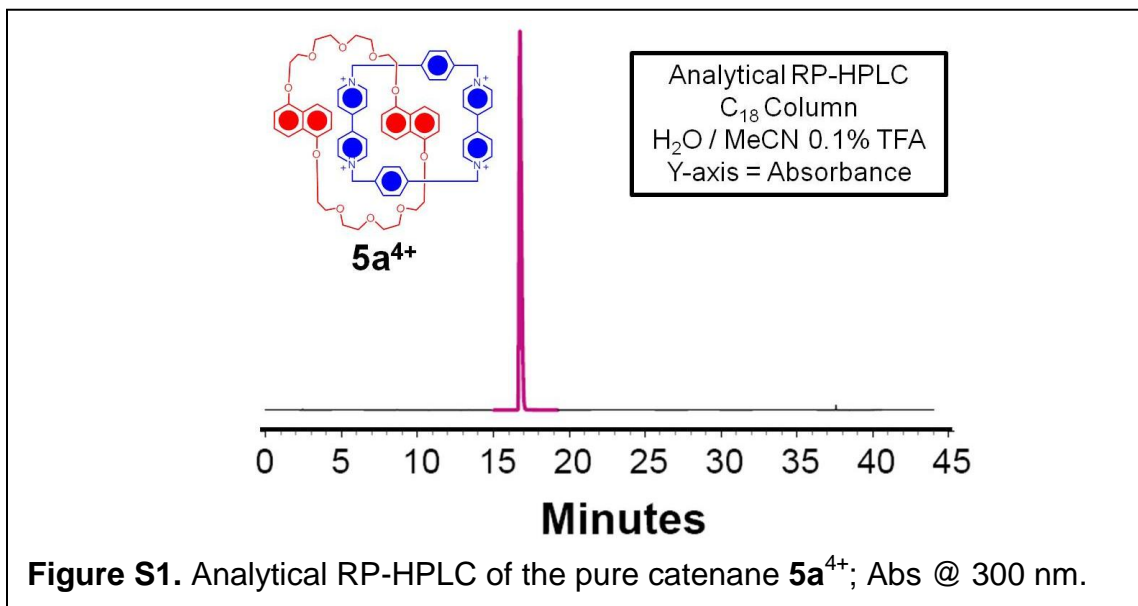
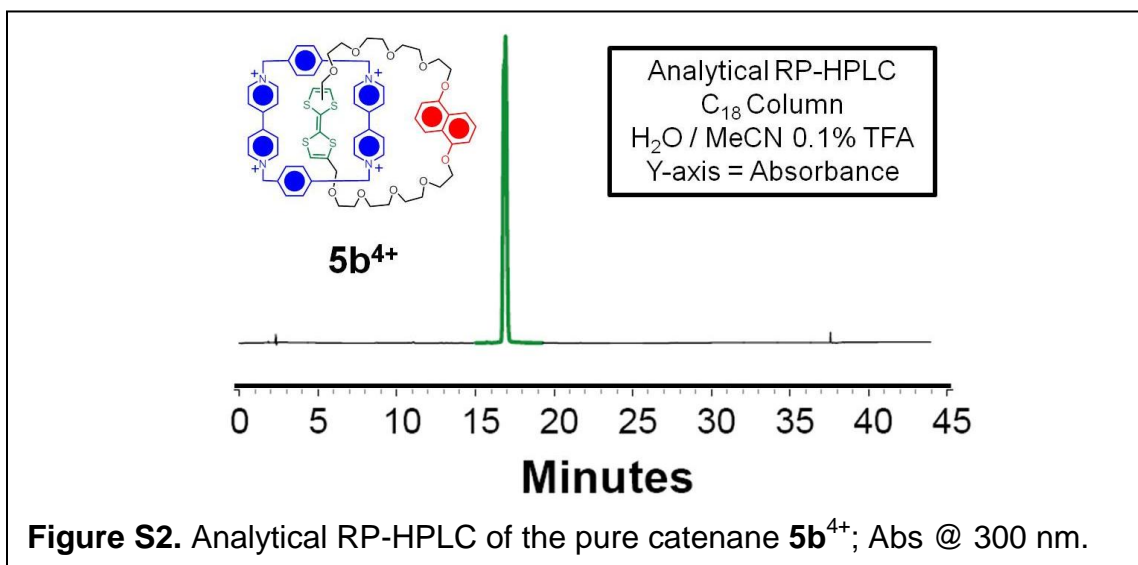
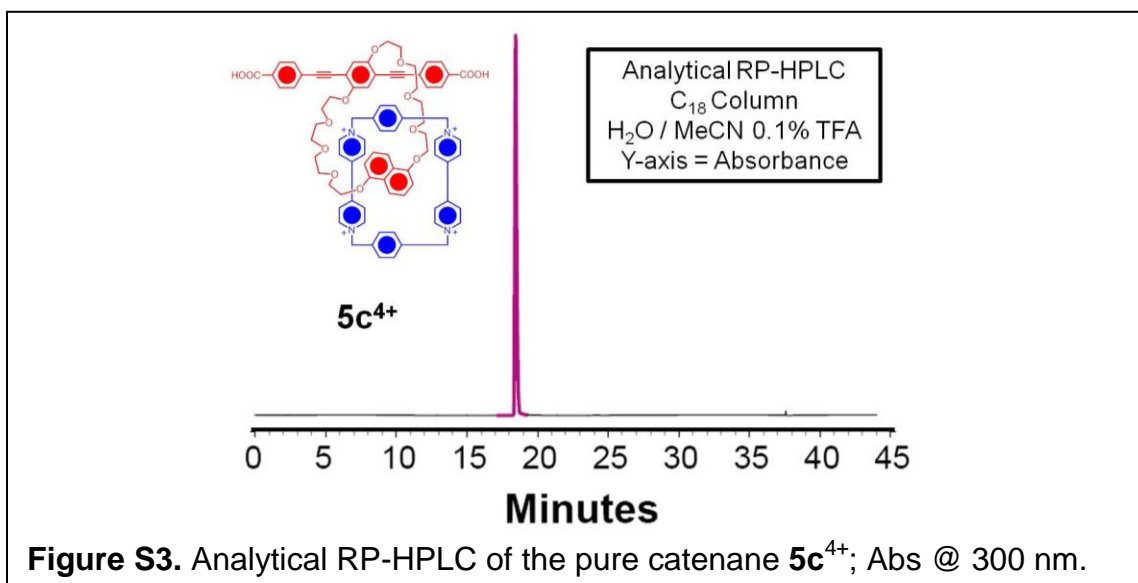


Figure S1. Analytical RP-HPLC of the pure catenane **5a**⁴⁺; Abs @ 300 nm.

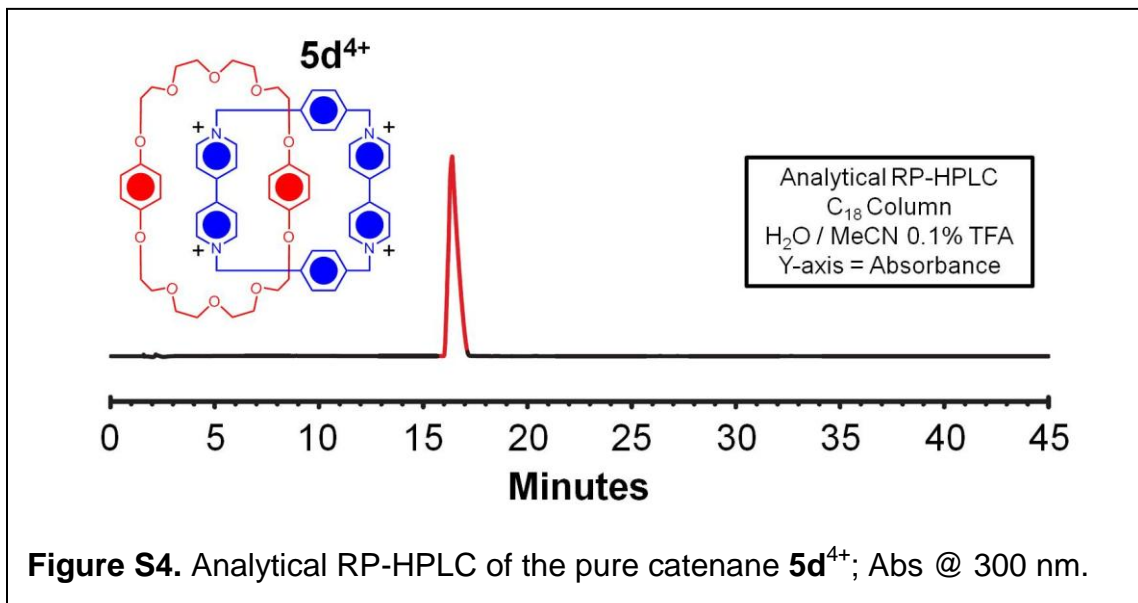


5b•4PF₆: Green solid (115 mg, 63%). ¹H NMR (500 MHz, CD₃CN, 298 K) δ = 8.91 (d, J = 6.7 Hz, 4H), 8.71 (d, J = 6.7 Hz, 4H), 7.61 (b, 8H), 7.45 (d, J = 6.3 Hz, 4H), 7.36 (m, 6H), 7.30 (t, J = 8.0 Hz, 2H), 6.71 (d, J = 8.0 Hz, 2H), 6.09 (s, 2H), 5.96 (m, 8H), 4.81 (s, 4H), 3.97–3.60 (b, 32H) ppm. HRMS (ESI) calcd for C₇₀H₇₆F₂₄N₄O₁₀P₄S₄ [$M - 2PF_6$]²⁺, m/z : 775.1863, found: 775.1846 [$M - 2PF_6$]²⁺.

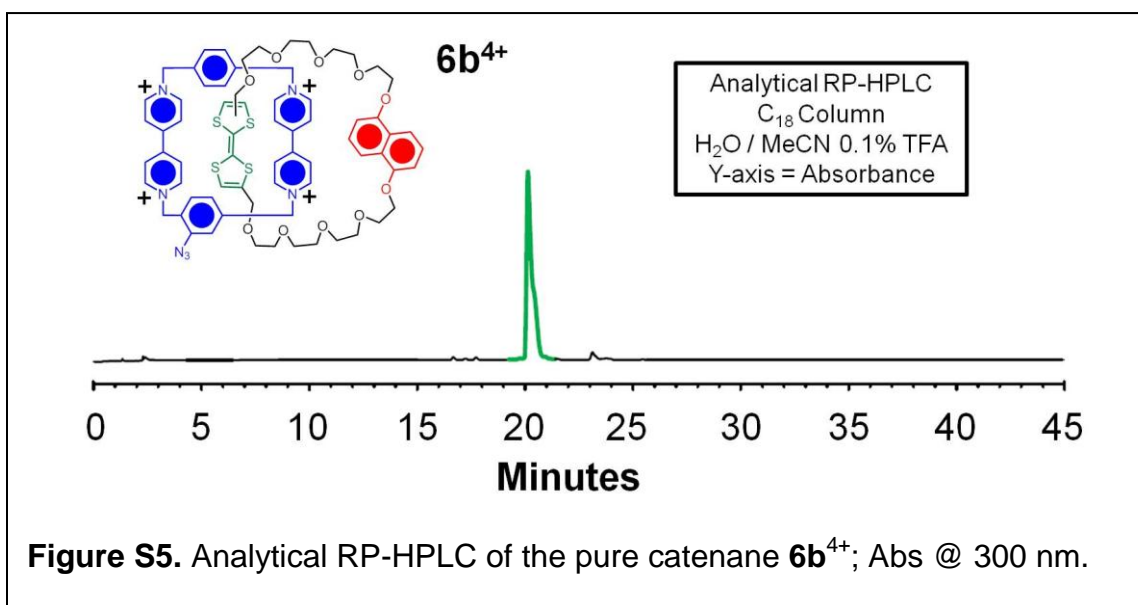


5c•4PF₆: Deep red solid (165 mg, 84%). ¹H NMR (500 MHz, CD₃SOCD₃, 298 K) δ = 9.08 (b, 8H), 8.09 (b, 12H), 7.54 (b, 12H), 6.39 (s, 2H), 6.03 (d, J = 7.5 Hz, 2H), 5.75 (b,

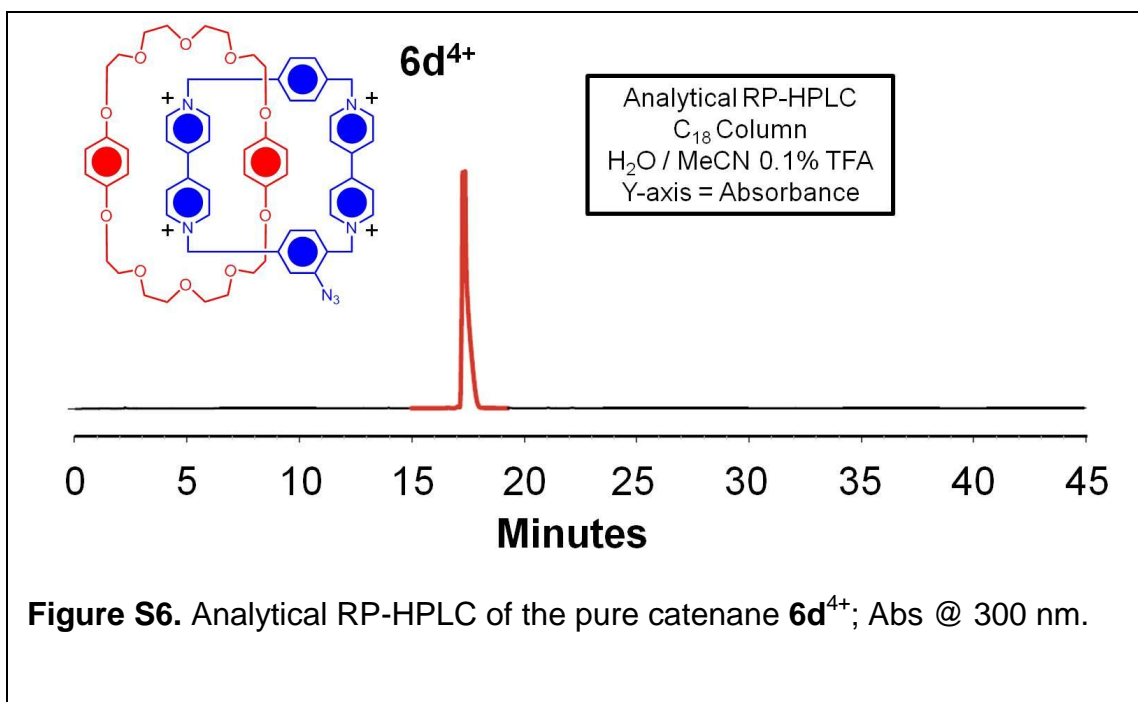
10H), 4.24–3.63 (b, 32H), 2.25 (d, $J = 7.5$ Hz, 2H) ppm. HRMS (ESI) calcd for $C_{86}H_{82}F_{24}N_4O_{14}P_4 [M - 4PF_6 - 2H]^{2+}$, m/z : 696.2835, found: 696.2830 $[M - 4PF_6 - 2H]^{2+}$.



5d•4PF₆: Red solid (55 mg, 34%). ¹H NMR (500 MHz, CD₃CN, 298 K) $\delta = 8.86$ (d, $J = 7.1$ Hz, 8H), 7.80 (s, 8H), 7.67 (d, $J = 7.1$ Hz, 8H), 6.17 (broad s, 4H), 3.98–3.82 (broad m, 16 H), 3.74 (broad m, 4H), 3.62 (broad m, 4H), 3.56 (broad m, 4H), 3.47 (broad s, 4H), 3.36 (broad m, 4H) ppm. LRMS (ESI) calcd for $C_{64}H_{72}F_{12}N_4O_{10}P_2 [M - 2PF_6]^{2+}$, m/z : 673.3, found: 673.3 $[M - 2PF_6]^{2+}$.



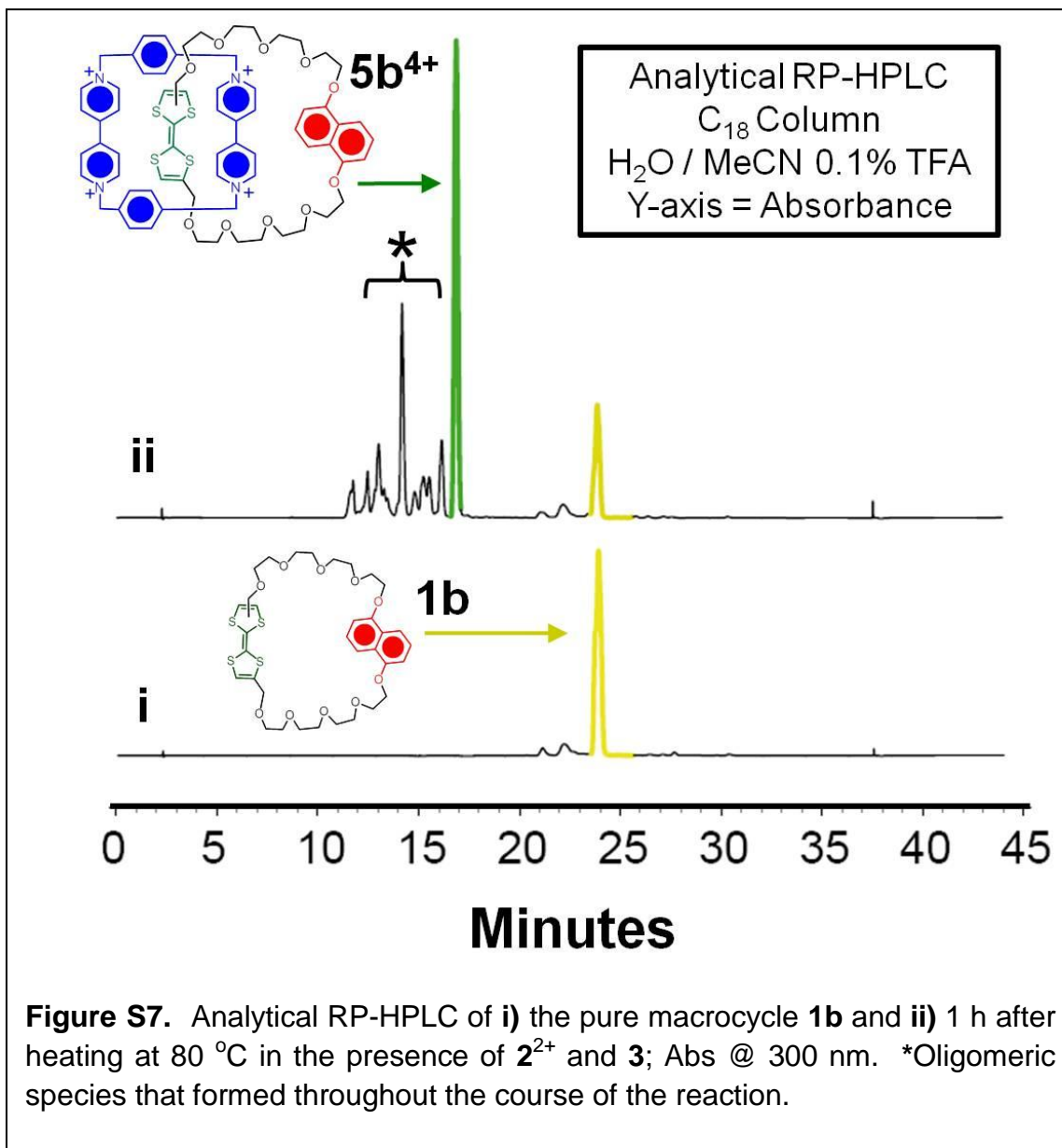
6b•4PF₆: Green solid (123 mg, 65%). ¹H NMR (500 MHz, CD₃CN, 298 K) δ = 9.14–8.42 (broad m, 8H), 7.89–7.18 (broad m, 13H), 6.83–6.55 (broad m, 2H), 6.17–5.97 (broad m, 2H), 5.95–5.28 (broad m, 8H), 4.38–3.44 (broad m, 36H) ppm. ¹³C NMR (125 MHz, CD₃CN, 298 K) δ = 206.23, 153.51, 144.53, 130.36, 125.98, 125.70, 125.16, 125.05, 124.22, 113.48, 105.48, 105.40, 70.82, 70.54, 70.44, 70.24, 70.01, 69.66, 69.49, 69.32, 67.36, 64.05, 63.64, 29.56 ppm. HRMS (ESI) calcd for C₇₀H₇₅F₁₂N₇O₁₀P₂S₄ [*M* – 2PF₆]²⁺, *m/z*: 795.6865, found: 795.6868 [*M* – 2PF₆]²⁺.

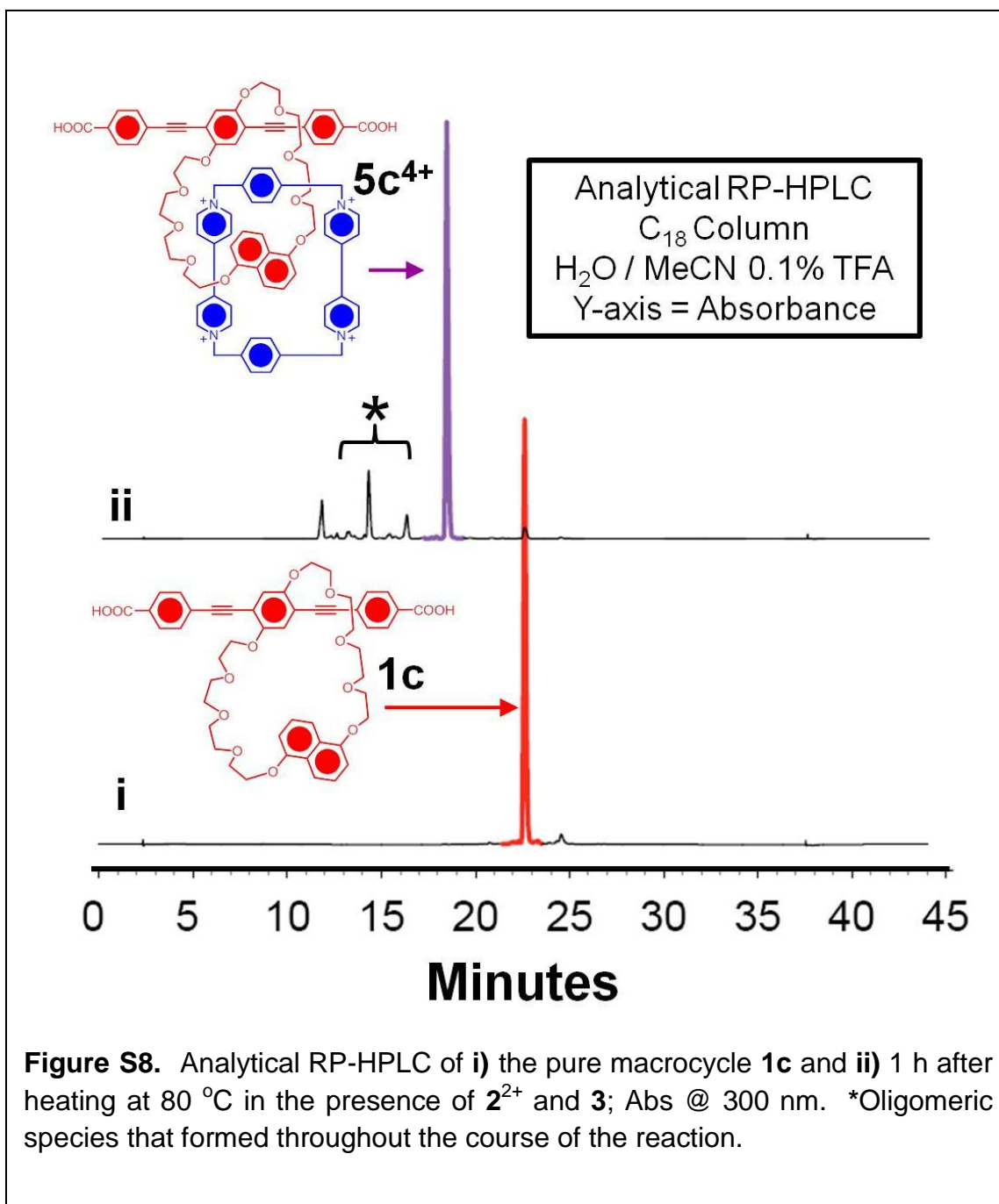


6d•4PF₆: Red solid (55 mg, 34%). ¹H NMR (500 MHz, CD₃CN, 298 K) δ = 8.90 (m, 8H), 7.90–7.57 (m, 15H), 6.18 (s, 4H), 5.69 (m, 8H), 4.01–3.52 (broad m, 32H), 3.56 (broad s, 4H) ppm. ¹³C NMR (125 MHz, CD₃CN, 298 K) δ = 151.67, 149.69, 145.67, 145.42, 145.23, 144.57, 144.35, 144.25, 139.90, 138.10, 136.27, 136.24, 133.15, 130.53, 130.41, 126.68, 126.40, 125.20, 125.06, 124.99, 119.46, 114.60, 70.37, 70.05, 69.56, 69.43, 69.19,

68.71, 67.23, 66.24, 64.31, 63.81, 59.29 ppm. HRMS (ESI) calcd for $C_{64}H_{72}F_{17}N_6O_{12}P_3$

$[M - PF_6]^+$ m/z : 1532.415, found: 1532.419 $[M - PF_6]^+$.





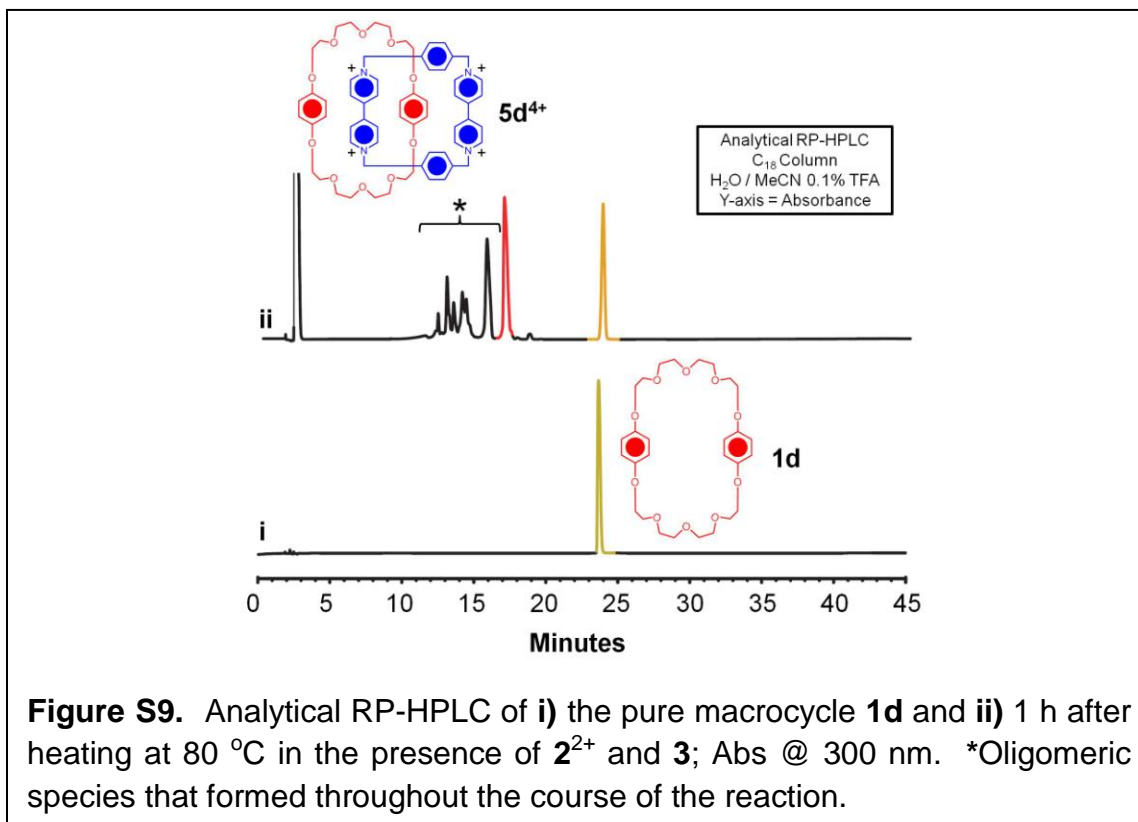


Figure S9. Analytical RP-HPLC of **i**) the pure macrocycle **1d** and **ii**) 1 h after heating at 80 °C in the presence of **2²⁺** and **3**; Abs @ 300 nm. *Oligomeric species that formed throughout the course of the reaction.

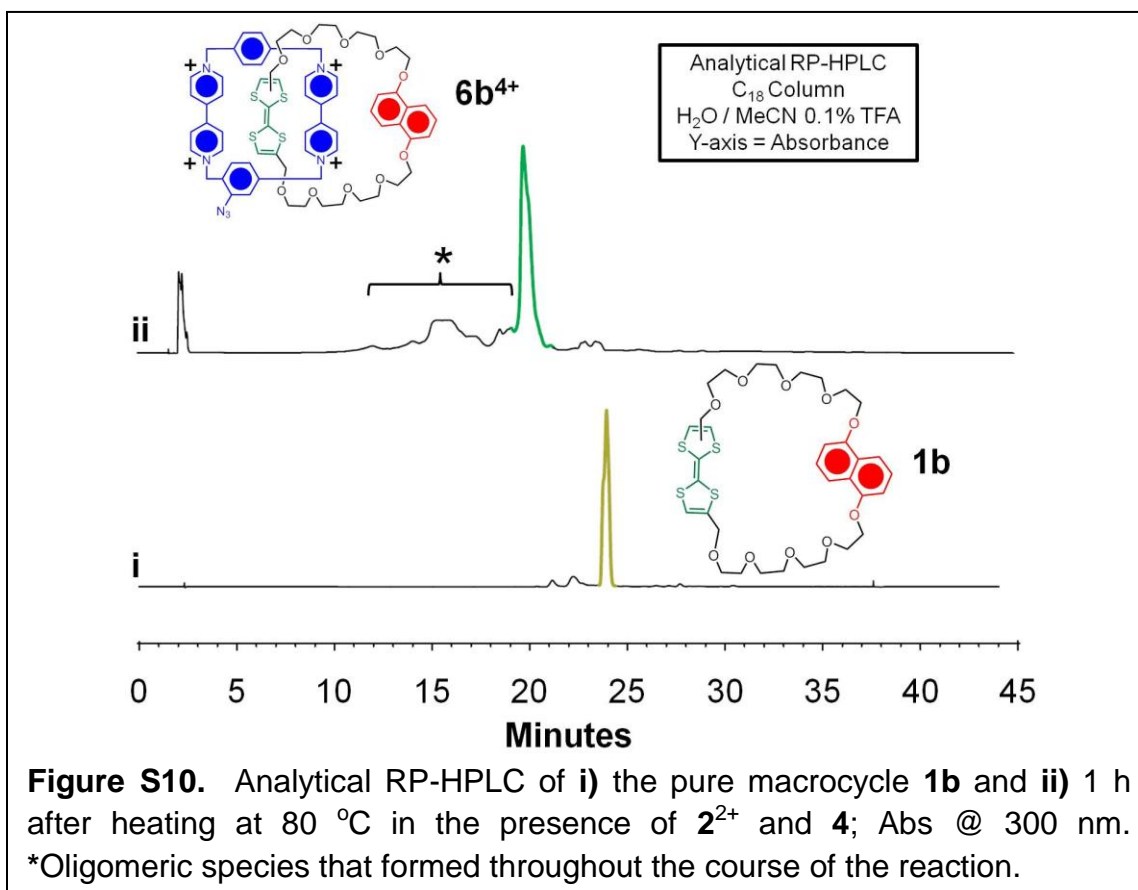


Figure S10. Analytical RP-HPLC of **i**) the pure macrocycle **1b** and **ii**) 1 h after heating at 80 °C in the presence of **2²⁺** and **4**; Abs @ 300 nm. *Oligomeric species that formed throughout the course of the reaction.

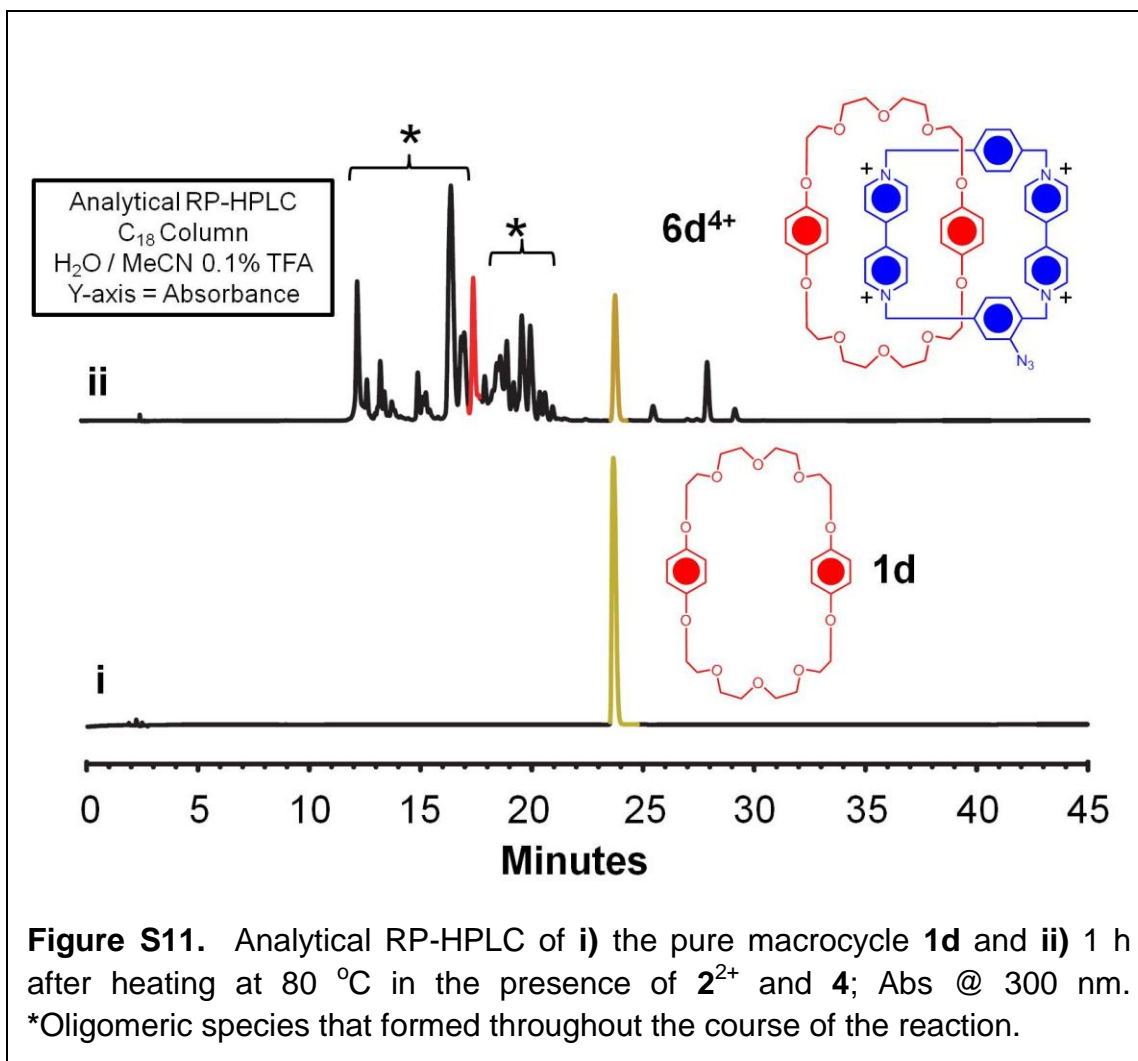
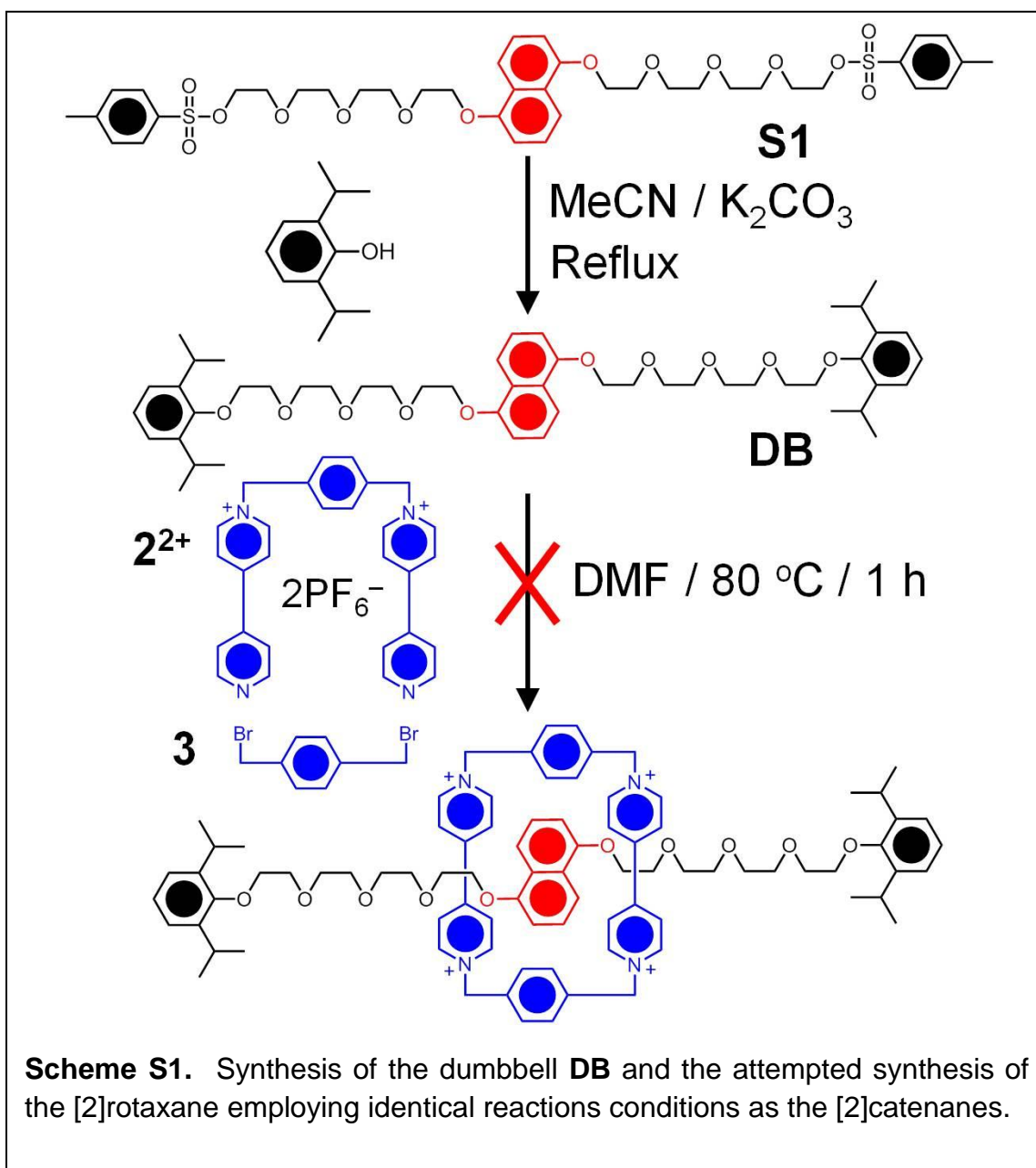
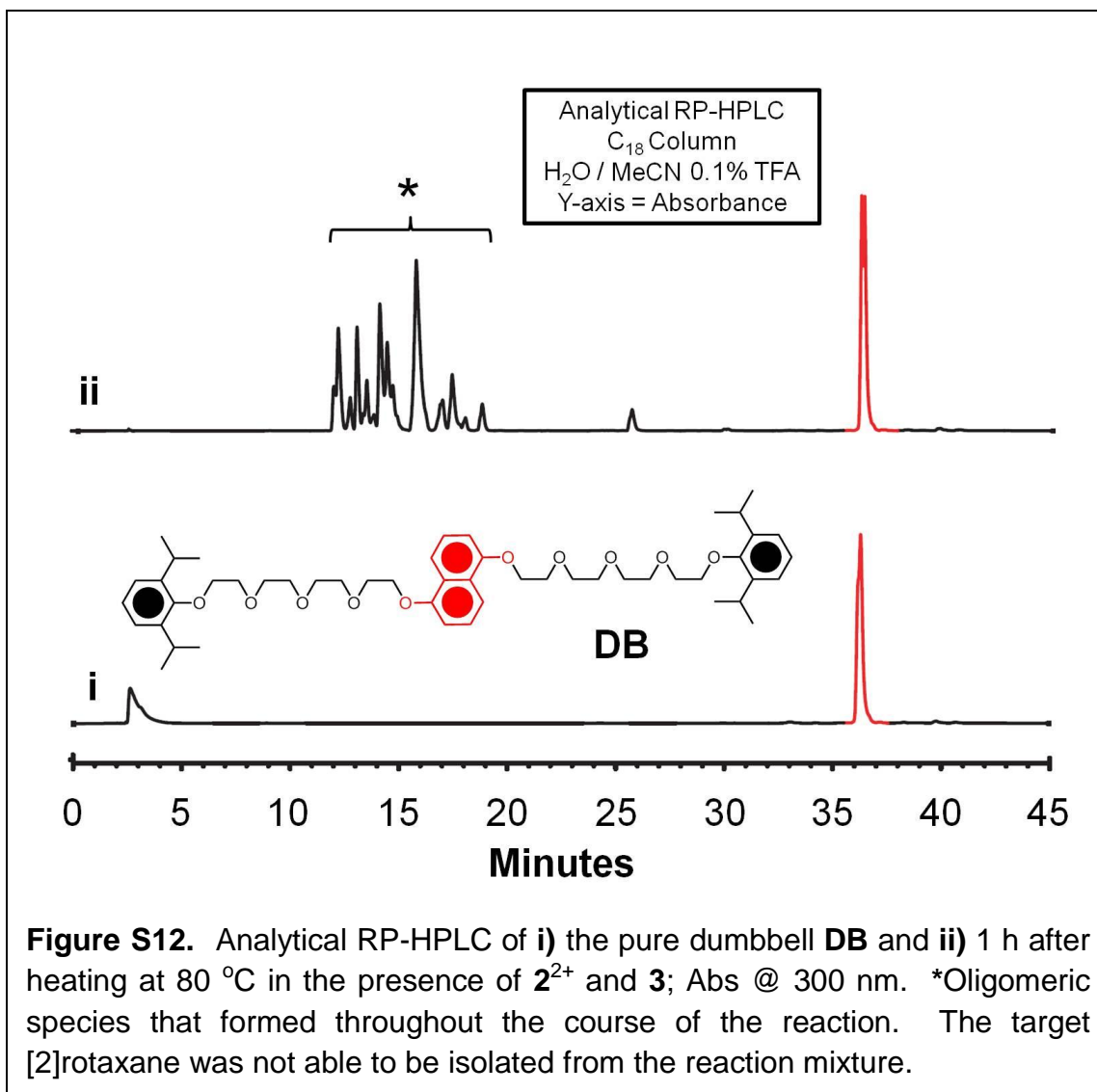


Figure S11. Analytical RP-HPLC of **i**) the pure macrocycle **1d** and **ii**) 1 h after heating at 80 °C in the presence of **2²⁺** and **4**; Abs @ 300 nm. *Oligomeric species that formed throughout the course of the reaction.

DB: A mixture of **S1** (300 mg, 0.37 mmol), 2,6-diisopropylphenol (400 mg, 2.2 mmol), and K₂CO₃ (500 mg, 3.6 mmol) was heated under reflux in MeCN (10 ml) for 18 h. After cooling to ambient temperature, the reaction mixture was added to water (50 ml) and extracted into CH₂Cl₂ (50 ml). The organic layers were dried (MgSO₄), filtered, concentrated under reduced pressure, and purified by flash column chromatography on SiO₂ (EtOAc:hexanes, 1:1 v/v) to afford the dumbbell **DB** as a viscous yellow liquid (73 mg, 24%). ¹H NMR (500 MHz, CDCl₃, 298K): δ = 7.86 (d, *J* = 8.5 Hz, 2H), 7.34 (t, *J* = 8.0 Hz, 2H), 7.1 (s, 6H), 6.84 (d, *J* = 7.5 Hz, 2H), 4.30 (t, *J* = 5.0 Hz, 4H), 4.01 (t, *J* = 5.0



Hz, 4H), 3.92–3.90 (m, 4H), 3.87–3.82 (m, 8H), 3.79–3.74 (m, 12H), 3.39 (hept, $J = 7.0$ Hz, 4H), 1.22 (d, $J = 7.0$ Hz, 24H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298K): $\delta = 154.3$, 153.0, 141.8, 127.8, 125.0, 124.6, 124.0, 114.6, 105.6, 73.8, 71.0, 71.0, 70.8, 70.7, 70.5, 69.8, 67.8, 26.2, 24.1 ppm. LRMS (ESI) calcd for C₅₀H₇₃O₁₀ [$M + H^+$]⁺ m/z : 833.5, found: 833.5 [$M + H^+$]⁺.



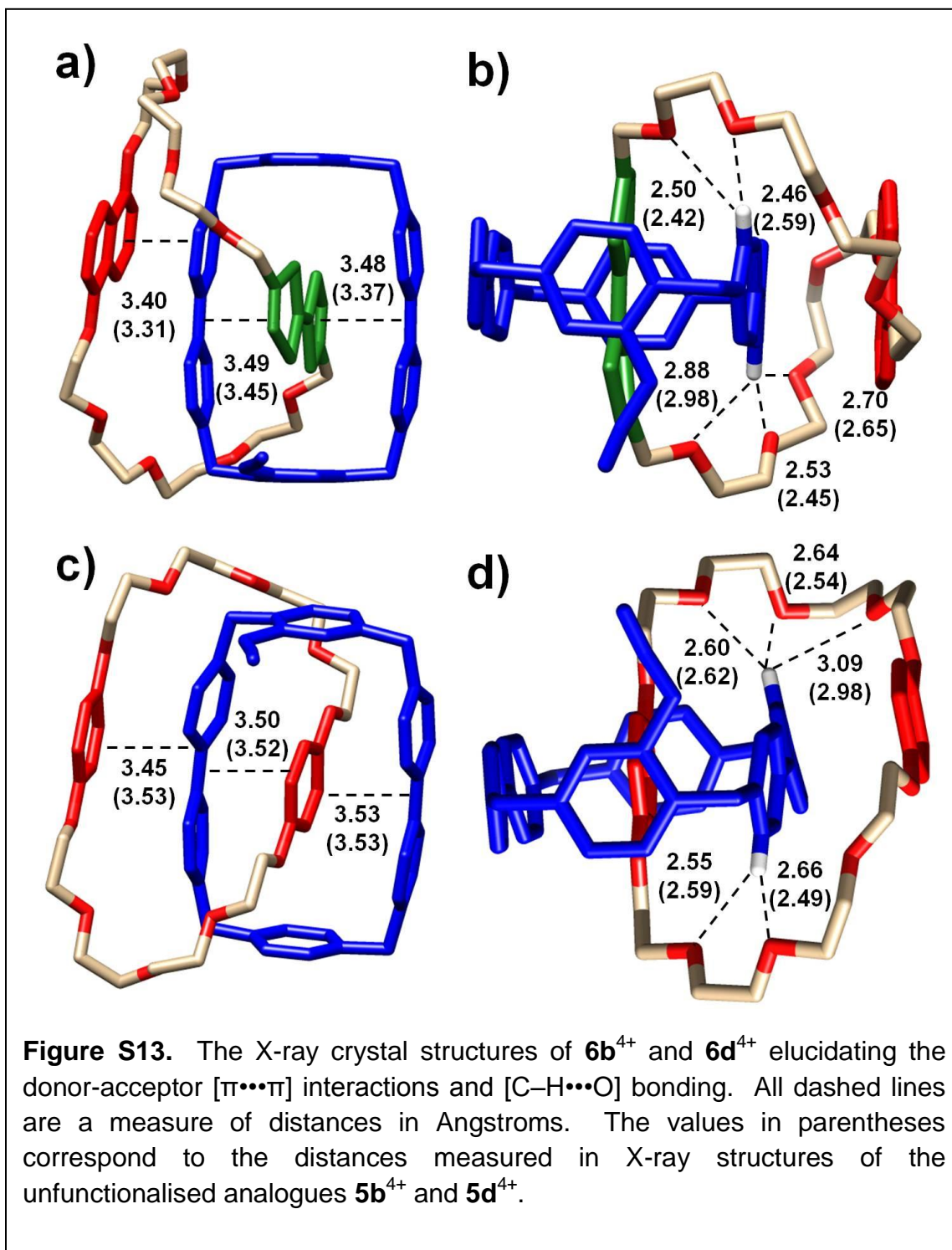
3. X-Ray Crystallography

Single crystals of **6b**•**4PF₆** and **6d**•**4PF₆** suitable for single-crystal X-ray crystallography were both grown by slow-vapor diffusion of *i*Pr₂O into individual MeCN solutions of the catenanes at 298 K. After a week, large crystals were observed and used subsequently for structural determination.

Crystal data for **6b**•**4PF₆**: C₃₄H₃₉O₁₀S₄, C₃₆H₃₀N₇, 4(PF₆), 3(C₂H₃N), $M_r = 1999.60$, triclinic, $P\bar{1}$, $a = 13.6925(4)$, $b = 14.1263(4)$, $c = 28.1027(9)$ Å, $\alpha = 96.491(2)$, $\beta = 103.206(2)$, $\gamma = 107.826(2)^\circ$. $V = 4938.8(3)$ Å³, $T = 100(2)$ K, $Z = 2$, $D_c = 1.345$ g cm⁻³, $\mu(\text{Cu-K}\alpha) = 2.389$ mm⁻¹, $F(000) = 2048$. Independent measured reflections 37696. $R_1 = 0.1181$, $wR_2 = 0.3394$ for 15377 independent observed reflections [$2\theta \leq 124.6^\circ$, $I > 2\sigma(I)$]. CCDC 887695. The SQUEEZE program was used to remove contributions of disordered solvent molecules from the unit cell.

Crystal data for **6d**•**4PF₆**: C₃₆H₃₁N₇, C₂₈H₄₀O₁₀, 4(F₆P), 5(C₂H₃N), $M_r = 1883.43$, triclinic, $P1$, $a = 13.5802(6)$, $b = 13.7732(6)$, $c = 26.2234(11)$ Å, $\alpha = 87.017(3)$, $\beta = 81.927(3)$, $\gamma = 60.811(2)^\circ$. $V = 4238.9(3)$ Å³, $T = 100(2)$ K, $Z = 2$, $D_c = 1.476$ g cm⁻³, $\mu(\text{Cu-K}\alpha) = 0.205$ mm⁻¹, $F(000) = 1940$. Independent measured reflections 99182. $R_1 = 0.1198$, $wR_2 = 0.3430$ for 24299 independent observed reflections [$2\theta \leq 124.6^\circ$, $I > 2\sigma(I)$]. CCDC 887694.

In the case of both **6b**⁴⁺ and **6d**⁴⁺, the X-ray structures reveal (Figure S13) that the azide groups do not significantly disrupt the noncovalent bonding interactions in comparison to their unfunctionalised derivatives **5b**⁴⁺ and **5d**⁴⁺. In particular, neither the donor-acceptor [$\pi \cdots \pi$] interactions or the [C–H \cdots O] bonding between the alpha protons of the encircled BIPY²⁺ unit and some of the oxygen atoms of the macrocyclic polyether are not observed to vary by more than a few tenths of an angstrom. These X-ray structures support the hypothesis that the azide group does not interrupt these noncovalent bonding interactions during the template-directed synthesis, and so as a consequence, the yields do not suffer.



4. References

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