Construction of α-Amido-indanones via Formal Allenamide Hydroacylation /Nazarov Cyclization

Yen-Ku Wu, Tianmin Niu and F. G. West*

Department of Chemistry, University of Alberta, E3-43 Gunning-Lemieux Chemistry Centre, Edmonton, AB, Canada T6G 2G2

frederick.west@ualberta.ca

Supporting Information: Experimental procedures, characterization data for all compounds and synthetic intermediates.

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General Information. Reactions were carried out in flame-dried glassware under a positive argon atmosphere unless otherwise stated. Transfer of anhydrous solvents and reagents was accomplished with oven-dried syringes. Solvents were distilled before use: methylene chloride and 1,2-dichloroethane from calcium hydride, tetrahydrofuran from sodium/benzophenone ketyl. All other solvents and commercially available reagents were either purified by standard procedures or used without further purification. Thin layer chromatography was performed on glass plates precoated with 0.25 mm silica gel; the stains for TLC analysis were conducted with 2.5 % *p*-anisaldehyde in AcOH-H₂SO₄-EtOH (1:3:85) and further heating until development of color. Flash chromatography was performed on 230-400 mesh silica gel with the indicated eluents. Nuclear magnetic resonance (NMR) spectra were recorded in indicated deturated solvents and are reported in ppm in the presence of TMS as internal standard and coupling constants (*J*) are reported in Hertz (Hz). Infrared (IR) spectra were recorded neat and reported in cm⁻¹. Mass spectra were recorded by

using EI or ESI as specified in each case.

Standard Procedure for the Synthesis of 4:

To a flame-dried round bottom flask containing a magnetic stirring bar was added allenamide 1 (0.5 mmol) and anhydrous THF (6 mL) under Ar. The temperature of the solution was reduced to -78 °C (acetone/dry ice bath). *n*-BuLi (1.2 equiv, 2.27 M in hexane, 0.26 mL) was added dropwise using 1 mL syringe. After stirring for 45 min, a solution of aldehydes 2 (1.1 equiv, 0.55 mmol) was added. The reaction mixture was stirred at -78 °C for 5 min, then allowed to warm to rt for 10 min. The reaction was quenched with sat. aq NH₄Cl solution. The organic layer was separated and the aqueous layer was extracted with Et₂O (3 x 20 mL). The combined organic layer was washed with brine (1 x 20 mL), dried over anhydrous MgSO₄, and then filtered through a pad of cellite. The solvent was removed under reduced pressure and the residue was purified on a silica gel column to afford the desired product.

In several cases, especially those reactions with lower yields, side product **4'** inconsistently appeared in the crude mixture and could not be obtained in pure form. The crude ¹H NMR spectrum clearly indicated the presence of vinyl protons and ethylene unit. The ¹³C NMR spectrum showed the existence of only two aliphatic carbons and several other sp² carbons. Also, the IR spectrum revealed the presence of a hydroxy group. The structure of **4'** was tentatively assigned based on these data. This side reaction will be further explored in detail.



Characterization Data for 4a-i and *d*-4a:



(Z)-3-(1-Oxo-1-phenylbut-2-en-2-yl) oxazolidin-2-one (4a)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4a** in 65% yield as a colorless oil: IR (film) 3062, 2960, 2916, 1756, 1654, 1598, 1413, 1247 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.74-7.71 (m, 2H), 7.58-7.53 (m, 1H), 7.47-7.27 (m, 2H), 6.63 (q, *J* = 6.8 Hz, 1H), 4.52-4.48 (m, 2H), 3.86-3.82 (m, 2H), 1.96 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.0, 156.6, 142.0, 137.0, 135.6, 132.0, 128.8, 128.0, 62.5, 45.5, 14.1; HRMS (EI, M⁺) Calcd for C₁₃H₁₃NO₃ 231.0895; found m/z 231.0889.



(Z)-3-(1-Oxo-1-(p-tolyl)but-2-en-2-yl) oxazolidin-2-one (4b)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4b** in 44% yield as a colorless oil: IR (film) 2959, 2921, 2852, 1755, 1653, 1606, 1412, 1246 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.65-7.63 (m, 2H), 7.27-7.23 (m, 2H), 6.60 (q, *J* = 7.2 Hz, 1H), 4.50-4.46 (m, 2H), 3.84-3.80 (m, 2H), 2.41 (s, 3H), 1.95 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.7, 156.5, 142.8, 141.1, 135.5, 134.3, 129.1, 128.7, 62.4, 45.5, 21.2, 14.0; HRMS (EI, M⁺) Calcd for C₁₄H₁₅NO₃ 245.1052; found m/z 245.1050.



(Z)-3-(1-(4-Methoxyphenyl)-1-oxobut-2-en-2-yl) oxazolidin-2-one (4c)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4c** in 18% yield as a pale yellow oil: IR (film) 2915, 2842, 1755, 1648, 1600, 1510, 1414, 1252 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.78-7.74 (m, 2H), 6.95-6.92 (m, 2H), 6.55 (q, *J* = 6.8 Hz, 1H), 4.50-4.46 (m, 2H), 3.87 (s, 3H), 3.84-3.80 (m, 2H), 1.95 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 190.7, 162.9, 156.5, 139.7, 135.4, 131.3, 129.4, 113.3, 62.4, 55.1, 45.6, 13.9; HRMS (EI, M⁺) Calcd for C_{14H15}NO₄ 261.1001; found m/z 261.0999.



(Z)-3-(1-(4-(Methylthio) phenyl)-1-oxobut-2-en-2-yl) oxazolidin-2-one (4d)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4d** in 52% yield as a pale yellow oil: IR (film) 2958, 2920, 2850, 1753, 1649, 1589, 1411, 1246 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.71-7.69 (m, 2H), 7.30-7.27 (m, 2H), 6.61 (q, *J* = 7.0 Hz, 1H), 4.53-4.50 (m, 2H), 3.87-3.83 (m, 2H), 2.55 (s, 3H), 1.98 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.4, 156.9, 145.4, 141.0, 135.8, 133.4, 129.9, 125.0, 62.8, 46.0, 14.9, 14.4; HRMS (EI, M⁺) Calcd for C₁₄H₁₅NO₃S 277.0773; found m/z 277.0772.



(Z)-3-(1-(4-Chlorophenyl)-1-oxobut-2-en-2-yl) oxazolidin-2-one (4e)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4e** in 46% yield as a colorless oil: IR (film) 3068, 2960, 2917, 1755, 1657, 1588, 1413, 1247 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.71-7.70 (m, 2H), 7.47-7.44 (m, 2H), 6.61 (q, *J* = 7.0 Hz, 1H), 4.54-4.51 (m, 2H), 3.87-3.84 (m, 2H), 1.99 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.3, 157.0, 142.2, 138.9, 135.9, 135.7, 130.7, 128.8, 62.9, 46.0, 14.5; HRMS (EI, M⁺) Calcd for C₁₃H₁₂ClNO₃ 265.0506; found m/z 265.0504.



(Z)-Methyl 4-(2-(2-oxooxazolidin-3-yl) but-2-enoyl) benzoate (4f)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4f** in 46% yield as a colorless oil: IR (film) 2954, 2916, 1755, 1724, 1659, 1610, 1408, 1281 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.15-8.13 (m, 2H), 7.80-7.77 (m, 2H), 6.65 (q, *J* = 7.0 Hz, 1H), 4.56-4.53 (m, 2H), 3.98 (s, 3H), 3.90-3.87 (m, 2H), 2.00 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.9, 166.2, 156.9, 143.6, 141.2, 136.1, 133.3, 129.6, 129.1, 62.9, 52.5, 45.9, 14.6; HRMS (EI, M⁺) Calcd for C₁₅H₁₅NO₅ 289.0950; found m/z 289.0949.



(Z)-3-(1-(Naphthalen-2-yl)-1-oxobut-2-en-2-yl) oxazolidin-2-one (4g)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4g** in 52% yield as a colorless oil: IR (film) 3053, 2960, 2915, 1755, 1662, 1591, 1414, 1246 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.12-8.01 (m, 1H), 7.98 (d, *J* = 8.5 Hz, 1H), 7.91-7.89 (m, 1H), 7.63-7.61 (m, 1H), 7.56-7.54 (m, 2H), 7.52-7.50 (m, 1H), 6.64 (q, *J* = 7.0 Hz, 1H), 4.58-4.55 (m, 2H), 3.98-3.95 (m, 2H), 1.93 (d, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 194.0, 157.2, 145.5, 138.1, 135.6, 133.6, 131.2, 130.7, 128.4, 127.4, 127.0, 126.6, 125.4, 124.3, 63.0, 46.1, 14.6; HRMS (EI, M⁺) Calcd for C₁₇H₁₅NO₃ 281.1052; found m/z 281.1050.



(Z)-3-(1-(Furan-2-yl)-1-oxobut-2-en-2-yl) oxazolidin-2-one (4h)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4h** in 33% yield as a yellow oil: IR (film) 2920, 1754, 1644, 1564, 1466, 1414, 1252 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) 7.65-7.64 (m, 1H), 7.27-7.26 (m, 1H), 7.12 (q, J = 7.0 Hz, 1H), 6.58-6.57 (m, 1H), 4.54-4.50 (m, 2H), 3.85-3.82 (m, 2H), 2.00 (d, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.4, 157.0, 151.7, 146.9, 140.9, 135.0, 119.9, 112.3, 62.8, 46.0, 14.5; HRMS (EI, M⁺) Calcd for C₁₁H₁₁NO₄ 221.0688; found m/z 221.0688.



3-((2Z, 5E)-5-Methyl-4-oxo-6-phenylhexa-2, 5-dien-3-yl) oxazolidin-2-one (4i)

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave **4i** in 35% yield as a colorless oil: IR (neat) 3056, 2985, 2917, 1757, 1642, 1575, 1411, 1243 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.46-7.42 (m, 4H), 7.38-7.36 (m, 1H), 7.29 (s, 1H), 6.67 (q, *J* = 7.0 Hz, 1H), 4.54-4.50 (m, 2H), 3.87-3.84 (m, 2H), 2.18 (br s, 3H), 1.97 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 194.8, 157.1, 139.9, 139.4, 136.2, 135.7, 135.5, 129.7, 128.6, 128.5, 62.9, 46.1, 14.6, 14.3; HRMS (EI, M⁺) Calcd for C₁₆H₁₇NO₃ 271.1209; found m/z 271.1205.



Deuterated 4a

Following the representative procedure, flash chromatography (hexane/EtOAc = 1/1) gave *d*-4a in 44% yield as colorless oil: IR (film) 3060, 2958, 2924, 2854, 1756, 1656, 1598, 1411, 1250 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.74-7.70 (m, 2H), 7.58-7.53 (m, 1H), 7.47-7.42 (m, 2H), 4.52-4.48 (m, 2H), 3.86-3.83 (m, 2H), 1.96 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 192.4, 156.9, 142.1 (t, *J*_{CD} = 23.9 Hz), 137.5, 135.9, 132.4, 129.3, 128.4, 62.8, 45.9, 14.4; HRMS (ESI, [M+Na]⁺) Calcd for C₁₃H₁₂DNNaO₃ 255.0850; found m/z 255.0848.

Standard Procedure for the Synthesis of 5:

To a stirred solution of 4 (0.1 mmole) in 1,2-dichloroethane (10 mL) at the indicated temperature

was added Lewis or Brønsted acid (reagent and equivalent specified below) under argon atmosphere. After starting material was completely consumed as monitored by TLC, the reaction was quenched with sat. aq NaHCO₃ (5 mL). The organic layer was separated and the aqueous layer was extracted with dichloromethane (2 x 5 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and then filtered through a pad of cellite. The solvent was removed under reduced pressure and the residue was purified on a silica gel column to afford the desired product.

Characterization Data for 5a-i



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5a** in 98% yield (cis:trans >20:1): IR (film) 2963, 2924, 1748, 1722, 1607, 1429, 1261, 1207 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.81-7.79 (m, 1H), 7.75-7.71 (m, 1H), 7.57-7.55 (m, 1H), 7.49-7.46 (m, 1H), 4.50 (ddd, *J* = 8.5, 8.5, 7.0 Hz, 1H), 4.45 (ddd, *J* = 9.0, 9.0, 7.0 Hz, 1H), 4.42 (d, *J* = 6.5 Hz, 1H), 3.67 (ddd, *J* = 9.0, 8.0, 7.0 Hz, 1H), 3.52 (ddd, *J* = 9.0, 8.0, 7.0 Hz, 1H), 3.36 (app. quintet, *J* = 6.5 Hz, 1H), 1.64 (d, *J* = 6.5 Hz, 3H)); ¹³C NMR (125 MHz, CDCl₃) δ 199.9, 158.9, 154.8, 135.9, 134.3, 128.2, 124.9, 124.0, 67.0, 62.3, 41.9, 36.6, 17.7; HRMS (EI, M⁺) Calcd for C₁₃H₁₃NO₃ 231.0895; found m/z 231.0900.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5b** in 87% yield (cis:trans >20:1); IR (film) 3018, 2966, 2919, 1749, 1719, 1611, 1430, 1263, 1216 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.66 (d, *J* = 8.5 Hz, 1H), 7.31 (br s, 1H), 7.24 (d, *J* = 8.5 Hz, 1H), 4.48-4.39 (m, 2H), 4.37 (d, *J* = 6.0 Hz, 1H), 3.62 (ddd, *J* = 9.5, 8.0, 7.0 Hz, 1H), 3.46 (ddd, *J* = 9.5, 8.0, 7.0 Hz, 1H), 3.26 (app. quintet, *J* = 6.5 Hz, 1H), 2.47 (s, 3H), 1.58 (d, *J* = 6.5 Hz, 3H);); ¹³C NMR (125 MHz, CDCl₃) δ 199.3, 159.0, 155.3, 147.4, 132.0, 129.4, 125.3, 123.8, 67.0, 62.3, 41.8, 36.5, 22.3, 17.6; HRMS (ESI, [M+Na]⁺) Calcd for C₁₄H₁₅NNaO₃ 268.0944; found m/z 268.0941.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5c** in 88% yield (cis:trans 1:4, inseparable mixture): IR

(film) 3019, 2969, 2917, 1750, 1713, 1601, 1429, 1263, 1217 cm⁻¹; trans isomer: ¹H NMR (500 MHz, CDCl₃) δ 7.70 (d, *J* = 8.5 Hz, 1H), 6.94 (dd, *J* = 8.5, 2.0 Hz, 1H), 6.92 (br s, 1H), 4.48-4.37 (m, 2H), 4.36 (d, *J* = 6.0 Hz, 1H), 3.91 (s, 3H), 3.64-3.59 (m, 1H), 3.48-3.43 (m, 1H), 3.25 (app. quintet, *J* = 7.0 Hz, 1H), 1.57 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 197.9, 166.3, 158.9, 157.9, 127.6, 125.9, 115.8, 108.5, 66.8, 62.2, 55.8, 41.8, 36.7, 17.7; cis isomer: ¹H NMR (500 MHz, CDCl₃) δ 7.72 (d, *J* = 8.5 Hz, 1H), 6.94 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.9 (d, *J* = 2.5 Hz, 1H), 4.81 (d, *J* = 7.5 Hz, 1H), 4.45-4.35 (m, 2H), 4.04 (ddd, *J* = 7.5, 7.5, 7.0 Hz, 1H), 3.91 (s, 3H), 3.78 (app. quintet, *J* = 8.0 Hz, 1H), 3.39 (ddd, *J* = 8.5, 8.5, 7.0 Hz, 1H), 1.28 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 198.3, 166.3, 160.6, 159.2, 126.9, 126.2, 116.2, 109.1, 62.7, 62.0, 43.5, 37.8, 17.6 [one aliphatic carbon is missing due to peak overlapping.]; HRMS (ESI, [M+Na]⁺) Calcd for C₁₄H₁₅NNaO₄ 284.0893; found m/z 284.0889.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5d** in 97% yield (cis:trans >20:1): IR (film) 3058, 2965, 2924, 1747, 1713, 1594, 1431, 1262, 1198 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.68 (d, *J* = 8.5 Hz, 1H), 7.31 (br s, 1H), 7.28 (d, *J* = 8.5 Hz, 1H), 4.49 (ddd, *J* = 8.5, 8.5, 6.5 Hz, 1H), 4.44 (ddd, *J* = 9.0, 9.0, 6.5 Hz, 1H), 4.41 (d, *J* = 6.0 Hz, 1H), 3.65 (ddd, *J* = 9.5, 8.0, 6.5 Hz, 1H), 3.51 (ddd, *J* = 9.0, 8.0, 7.0 Hz, 1H), 3.30 (app. quintet, *J* = 6.5 Hz, 1H), 2.59 (s, 3H), 1.62 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 198.5, 158.9, 155.4, 150.1, 130.9, 125.1, 124.0, 120.4, 66.9, 62.3, 41.9, 36.6, 17.6, 14.9; HRMS (ESI, [M+Na]⁺) Calcd for C₁₄H₁₅NNaO₃S 300.0665; found m/z 300.0660.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5e** in 94% yield (cis:trans >20:1): IR (film) 3061, 2965, 2923, 1748, 1725, 1600, 1431, 1259, 1210 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.70 (d, *J* = 8.5 Hz, 1H), 7.51 (br s, 1H), 7.42 (d, *J* = 8.5 Hz, 1H), 4.47 (ddd, *J* = 9.0, 9.0, 7.0 Hz, 1H), 4.42 (ddd, *J* = 8.5, 8.5, 7.0 Hz, 1H), 4.36 (d, *J* = 6.5 Hz, 1H), 3.66-3.61 (m, 1H), 3.53-3.48 (m, 1H), 3.32 (app. quintet, *J* = 6.5 Hz, 1H), 1.59 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 198.5, 158.8, 156.2, 142.6, 132.7, 129.0, 125.4, 125.2, 67.0, 62.3, 42.1, 36.6, 17.5; HRMS (ESI, [M+Na]⁺) Calcd for C₁₃H₁₂ClNNaO₃ 288.0398; found m/z 288.0395.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5f** in 91% yield (cis:trans >20:1): IR (film) 3020, 2955, 2927, 1751, 1726, 1615, 1436, 1262, 1216 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.23 (br s, 1H), 8.14-8.12 (m, 1H), 7.86-7.85 (m, 1H), 4.55-4.46 (m, 2H), 4.44 (d, *J* = 6.5 Hz, 1H), 4.01 (s, 3H), 3.69 (ddd, *J* = 9.0, 8.0, 7.0 Hz, 1H), 3.56 (ddd, *J* = 9.0, 8.0, 7.0 Hz, 1H), 3.41 (app. quintet, *J* = 7.0 Hz, 1H), 1.68 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 199.6, 166.0, 158.8, 154.5, 137.4, 136.6, 129.3, 126.3, 123.9, 67.4, 62.3, 52.7, 42.2, 36.6, 17.6; HRMS (ESI, [M+Na]⁺) Calcd for C₁₅H₁₅NNaO₅ 312.0842; found m/z 312.0841.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5g** in 97% yield (cis:trans >20:1): IR (film) 3014, 2966, 2925, 1744, 1704, 1628, 1428, 1269, 1219 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 9.07 (d, *J* = 8.5 Hz, 1H), 8.20 (d, *J* = 8.5 Hz, 1H), 7.96-7.94 (m, 1H), 7.76-7.73 (m, 1H), 7.65-7.61 (m, 2H), 4.55-4.44 (m, 2H), 4.54 (d, *J* = 5.5 Hz, 1H), 3.71 (ddd, *J* = 9.0, 8.0, 7.0, 1H), 3.55 (ddd, *J* = 9.5, 7.5, 6.5 Hz, 1H), 3.46 (app. quintet, *J* = 6.0 Hz, 1H), 1.70 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 200.2, 159.1, 157.8, 137.1, 132.8, 129.6, 128.9, 128.8, 128.3, 127.2, 124.3, 121.8, 67.1, 62.3, 42.0, 36.8, 17.9; HRMS (EI, M⁺) Calcd for C₁₇H₁₅NO₃ 281.1052; found m/z 281.1049.



Following the representative procedure using 5 equiv triflic acid at 65 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5h** in 65% yield (cis:trans 1:3, inseparable mixture): IR (film) 3059, 2972, 2918, 1749, 1715, 1579, 1426, 1268, 1246 cm⁻¹; trans isomer: ¹H NMR (500 MHz, CDCl₃) δ 7.87 (d, J = 2.0 Hz, 1H), 6.59 (d, J = 2.0 Hz, 1H), 4.52 (d, J = 3.5 Hz, 1H), 4.50-4.40 (m, 2H), 3.66-3.61 (m, 1H), 3.57-3.52 (m, 1H), 3.21 (qd, J = 7.5, 3.5 Hz, 1H), 1.53 (d, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 182.3, 158.5, 157.6, 154.5, 152.0, 109.2, 70.1, 62.2, 42.2, 33.1, 18.0; cis isomer: ¹H NMR (500 MHz, CDCl₃) δ 7.87 (d, J = 2.0 Hz, 1H), 6.60 (d, J = 2.0 Hz, 1H), 4.50-4.40 (m, 3H), 4.00-3.95 (m, 1H), 3.69 (app. quintet, J = 7.0 Hz, 1H), 3.52-3.47 (m, 1H), 1.31 (d, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 181.9, 160.0, 158.7, 154.6, 151.8, 109.6, 65.3, 62.6, 42.1, 33.0, 16.1; HRMS (ESI, [M+Na]⁺) Calcd for C₁₁H₁₁NNaO₄ 244.0580; found m/z 244.0583.



Following the representative procedure using 20 mol% Sc(OTf)₃ at 80 °C (oil bath), flash chromatography (hexane/EtOAc = 1/1) gave **5i** in 96% yield: IR (film) 3336 (br), 3027, 2977, 1700, 1656, 1600, 1406, 1360, 1206 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.38-7.35 (m, 2H), 7.32-7.29 (m, 1H), 7.16-7.14 (m, 2H), 5.37 (br s, 1H), 3.32 (app. quintet, *J* = 1.9 Hz, 1H), 2.32 (qdd, *J* = 8.0, 2.2, 0.8 Hz, 1H), 1.82 (dd, *J* = 2.3, 0.8 Hz, 3H), 1.29 (d, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 204.3, 148.2, 143.3, 141.2, 128.9, 127.5, 127.2, 54.7, 48.6, 15.0, 12.5; HRMS (EI, M⁺) Calcd for C₁₃H₁₄O₂ 202.0994; found m/z 202.0995.

Stereochemical Assignment for Nazarov Products 5a-5h

The trans isomers of 2,3-substituted-1-indanone were generally considered as thermodynamically favored products over the cis counterpart. In the case of **5c** obtained as isomeric products (ratio ca 1:4), the coupling constant (*J*) between H-2 and H-3 for the major isomer is 6.0 Hz and for the minor isomer is 7.5 Hz. That mixture was subjected to 10 mol% DBU in CDCl₃ (0.01M in **5c**) at room temperature, where the epimerization was envisioned to be guided by the thermodynamic control. In the event, the minor isomer was no longer detected in the proton NMR after 4h; therefore the disappearing isomer was assigned as cis-**5c** and the predominant isomer as trans-**5c**. With that, we presumed that J_{H2H3} for trans isomers of closely related indanones would be similar to the value of 6.0 Hz. Based on the evaluation of J_{H2H3} , **5a**, **5b**, **5c**-**g** were referred to as trans. Similar approach was then applied to the stereochemical assignment for heteroaromatic substrate **5h** isolated as a ca 1:3 isomeric mixture, and the trans isomer could be readily assigned, whose J_{H2H3} equals 3.5 Hz.



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¹H and ¹³C NMR Spectra

























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udd 499.815 MHz H1 1D in cdcl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.7 C -> actual temp = 27.0 C, colddual probe date: Apr 26 2012 sweep width: 6010Hz acq.time: 5.0s relax.time: 0.1s # scans: 16 dig.res.: 0.1 Hz/pt hz/mm:25.0 Pulse Sequence: s2pul spectrometer:ibdw file:/mnt/d600/homel3/westnmr/nmrdata/DATA_FROM_NMKSEKVICE/Yen-Ku/03-Tianmin/5a-1H 0 н 3.36 ⊐ N ო 十十十 1.101.00 1.09 4 二 2.18 1.11 ഗ ဖ 5 ዛዛ ዛዛ 1.00.08 1.11.11 ω 5a Me ი 10



udd 498.122 MHz H1 ID in cdcl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.2 C -> actual temp = 27.0 C, autoxdb probe date: Apr 2 2012 sweep width: 6001Hz acq.time: 5.0s relax.time: 0.1s # scans: 12 dig.res.: 0.1 Hz/pt hz/mm:25.0 Pulse Sequence: s2pul spectrometer:ibdw file:/mnt/d600/homel3/westnmr/nmrdata/DATA_FROM_NMKSEKVICE/Yen-KU/03-Tianmin/5b-1H 0 3.72 -N 3.41 ო 上 1.06 2.21 ഹ ဖ , 1.07 1.12 1.08 ω **5b** Me თ 10











udd 498.122 MHz H1 ID in cdcl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.2 C -> actual temp = 27.0 C, autoxdb probe date: Apr 16 2012 sweep width: 6001Hz acq.time: 5.0s relax.time: 0.1s # scans: 16 dig.res.: 0.1 Hz/pt hz/mm:25.0 Pulse Sequence: s2pul spectrometer:ibdw file:/mnt/d600/homel3/westnmr/nmrdata/DATA_FROM_NMKSEKVICE/Yen-KU/03-Tianmin/5e-1H 0 3.32 ⊤ N ო 나나 나 1.08 1.071.00 0.96 2.36 Ŋ ဖ Г ∀ ∀ ∀ 1.111.10 1.02 ω Me თ 5e ÷ 10







udd 499.815 MHz H1 1D in cdcl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.7 C -> actual temp = 27.0 C, colddual probe date: Apr 26 2012 sweep width: 6010Hz acq.time: 5.0s relax.time: 0.1s # scans: 16 dig.res.: 0.1 Hz/pt hz/mm:25.0 Pulse Sequence: s2pul spectrometer:ibdw file:/mnt/d600/homel3/westnmr/nmrdata/DATA_FROM_NMKSEKVICE/Yen-Ku/03-Tianmin/5g-1H 0 н 3.04 ₽ N ო 1.0**1**.00 1.02 4 3.08]-ഹ ဖ 5 부 부 부부 1.031.95 1.01 1.11 ω ⊢ 0.90 ი . Ме **5**g 5









