In situ Formation of β-Glycosyl Imidinium Triflate from Participating Thioglycosyl Donors: Elaboration to Disarmed-Armed Iterative Glycosylation

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Section A: Experimental section

Reagent-grade chemicals were purchased from commercial vendors and used without purification. Dichloromethane (CH₂Cl₂) was dried by Asianwong solvent system (AWS-1000). *N*,*N*-Dimethylformamide (DMF) was stocked with flame-dried molecular sieves (MS) under N₂. Progress of reactions was monitored by thinlayer chromatography on silica gel 60 F-254 plate and visualized under UV illumination and/or by staining with acidic ceric ammonium molybdate or *p*-anisaldehyde. Silica gel (Geduran Si-60, 0.063-0.200 mm) for chromatography was obtained from Merck. NMR spectra were recorded at 300 MHz and 75 MHz spectrometers in Brüker console or 400/500/600 MHz and 100/125/150 MHz in Varian console as specified. Coupling constants in Hz was calculated from chemical shifts of ¹H NMR spectra. Preparations of glycosyl substrates 1,^{\$1} 7,^{\$2} 8,^{\$2} 9,^{\$3} 10,^{\$4} 11a,^{\$5} 11b,^{\$6} 12,^{\$7} \$13,^{\$8} s13ba,^{\$9} s14a,^{\$9} 16,^{\$6} 32,^{\$10} s33a,^{\$3} and 34^{\$3} were referred to literature methods. Diacetonide galactose 3 was purchased from common venders. Naming of the saccharide molecules follows the nomenclature rules of carbohydrate from IUPAC.^{\$11}

p-Tolyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α-D-thiomannopyranoside 2:



A solution of per-*O*-acetyl mannosyl acetate **s2a** (33.6 g, 86.0 mmol) in CH₂Cl₂ (35 mL) was cooled to 0 °C and treated with 33% HBr in AcOH (34 mL) under N₂. After stirring at 0 °C for 30 min, the reaction mixture was brought to RT. Upon the completion of the reaction as checked by TLC (*ca.* 2 h), the reaction mixture was diluted with EtOAc (20 mL \times 2) and poured into a separatory funnel. The organic phase was then washed with satd. NaHCO₃ (15 mL \times 2), brine (50 mL), dried (over $^{-3}$ -

MgSO₄), filtered, concentrated, and dried under *vacuo* for 5h. The crude was taken up in dried CH₃CN (98 mL), treated with tetrabutyl ammonium bromide (TBAB) (5 g, 15.7 mmol), 2,6-lutidine (18.2 mL, 157 mmol), and MeOH (9.5 mL). The reaction was stirred overnight at RT under N₂. Upon completion of the reaction, the solvent was reduced by rotary evaporator. The residue was taken up by EtOAc (100 mL \times 2), which was washed with satd. NaHCO₃ (150 mL \times 2), H₂O (50 mL) and brine (50 mL), dried over MgSO₄, filtered, and concentrated to give crude per-O-acetylated orthoester s2b (28.3 g, 78.4 mmol). The per-O-acetylated orthoester s2b was dissolved in MeOH/CH₂Cl₂ solution mixture, then treated with Na(s) at RT. Upon completion of deacetylation, the reaction mixture was concentrated and dried under vacuo for 4h. The crude mixture was dissolved in DMF and cooled at 0 °C bath under N₂. The reaction mixture was treated with sodium hydride (NaH, 13.2 g, 331 mmol) (60% in mineral oil) and benzyl bromide (BnBr, 40 mL, 331 mmol). Upon completion of the reaction, ice was added to quench excessive NaH. The reaction crude was subsequently diluted with EtOAc and H₂O in the separation funnel, then washed with satd. NaHCO₃ and brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: CH₂Cl₂/EtOAc/Hexane 1/0/3 stepwise to 1/1/3) to give per-O-benzyl mannosyl orthoester s2c (35.1g). The per-O-benzylated mannosyl orthoester s2c (10.12 g, 20 mmol) in CH_2Cl_2 was treated with p-thiocresol (6.21 g, 50 mmol) was cooled to 0 °C, then BF₃.OEt₂ (6.3 mL, 50 mmol) was added to the mixture. Upon completion of the reaction, the reaction crude was diluted with EtOAc, washed with satd. NaHCO₃ and brine in chill, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: EtOAc/Hexane 1/19 stepwise to 3/17) to give 2-O-acetyl thiomannoside s2d (5.2 g, 8.7 mmol) as yellow syrup. The 2-O-acetyl thiomannoside s2d (7.8 g, 13 mmol) in CH₂Cl₂/MeOH mixed solution was treated with Na(s). Upon completed deacetylation, the reaction was neutralized with IR-120 (H⁺), filtered, and concentrated for column chromatography purification (Elution: recycled EtOAc/Hexane 0~25% stepwise to 25~50%) to give

the 2-hydroxyl unprotected thiomannoside (6.08g, 10.9mmol). The 2-hydroxyl unprotected thiomannoside (6.08g, 10.9mmol) in CH₂Cl₂ was subsequently treated with benzoyl chloride (BzCl) (2.56 mL, 21.8 mmol), Et₃N (4.5 mL, 32.7 mmol) and DMAP (133 mg, 1.09 mmol) under N_2 . Upon completion of the benzoylation, the reaction mixture was diluted with satd. NaHCO₃, then washed with EtOAc, H₂O and brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: recycled EtOAc/Hexane 0~25%) to yield target thiomannoside donor **2** (6.11 g, 9.27 mmol). For thiomannoside **2**: $[\alpha]_D^{35}$ +77.11 (*c* 0.42, CHCl₃); *R*_f 0.35 (EtOAc/Hexane 1:3); ¹H NMR (300 MHz, CDCl₃): δ 8.05 (dd, J = 7.8, 1.5 Hz, 2 H), 7.54 (t, J = 7.5 Hz, 1 H), 7.39-7.25 (m, 17 H), 7.23-7.20 (m, 2 H), 7.05 (d, J = 8.1 Hz, 2 H), 5.87 (dd, J = 2.7, 1.8 Hz, 1 H, H-2), 5.58 (d, J = 1.5 Hz, 1 H, H-1), 4.90 (d, J = 10.8 Hz, 1 H), 4.81 (d, J = 11.4 Hz, 1 H), 4.70 (d, J = 11.7 Hz, 1 H), 4.58 (t, J = 11.7 11.4 Hz, 2 H), 4.49 (d, J = 11.7 Hz, 1 H), 4.40 (dd, J = 9.3, 2.1 Hz, 1 H), 4.16 (t, J = 9.6 Hz, 1 H), 4.06 (dd, J = 9, 2.7 Hz, 1 H), 3.95 (dd, J = 10.8, 4.2 Hz, 1 H), 3.79 (dd, J = 10.5, 1.5 Hz, 1 H), 2.29 (s, 3 H, CH₃); ¹³C NMR (75 MHz, CDCl₃): δ 166.0 (C=O), 138.8, 138.7, 138.4, 138.1, 133.7, 132.8, 130.4, 130.3, 130.2, 128.9, 128.81, 128.77, 128.6, 128.5, 128.3, 128.2, 128.0, 127.9, 87.1 (C-1'), 79.0, 75.8, 75.0, 73.8, 73.0, 72.1, 71.0, 69.5, 21.6; HRMS (m/z): $[M + Na]^+$ calcd. for $C_{41}H_{40}NaO_6S^+$, 683.2438; found, 683.2450.

6-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-1,2:3,4-di-*O*-isopropyli dene-α-D-galactopyranose 4



Disaccharide 4 was prepared from thioglucoside 1 and diacetonide galactose acceptor 3 by general modulated glycosylation procedure. Purification of 4 was

performed by column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:2:10 stepwise to 1:2:7). For **4**: $[\alpha]_D^{35}$ –30.00 (*c* 0.36, CHCl₃); *R*_f 0.275 (EtOAc/CH₂Cl₂/Hexane 1:2:5); ¹H NMR (300 MHz, CDCl₃): δ 8.03 (d, *J* = 7.5 Hz, 2 H), 7.53 (t, *J* = 7.2 Hz, 1 H), 7.42-7.27 (m, 10 H), 7.20-7.17 (m, 2 H), 7.12 (s, 5 H), 5.38 (d, *J* = 4.8 Hz, 1 H, H-1), 5.30 (t, *J* = 8.1 Hz, 1 H, H-2'), 4.82 (d, *J* = 10.8 Hz, 1 H), 4.46 (d, *J* = 11.1 Hz, 1 H), 4.68-4.63 (m, 3 H), 4.59-4.55 (m, 2 H), 4.39 (dd, *J* = 7.8, 1.8 Hz, 1 H), 4.19-4.17 (m, 1 H), 4.10 (d, *J* = 8.1 Hz, 1 H), 4.02 (dd, *J* = 10.8, 5.1 Hz, 1 H), 3.82-3.71 (m, 6 H), 3.56 (d, *J* = 1.8 Hz, 1 H), 1.38 (s, 3 H, CH₃), 1.21 (s, 3 H, CH₃), 1.17 (s, 6 H, CH₃ × 2); ¹³C NMR (75 MHz, CDCl₃): δ 165.6 (*C*=O), 138.5, 138.3, 138.2, 133.2, 130.4, 130.3, 128.8, 128.7, 128.6, 128.34, 128.31, 128.19, 128.15, 128.1, 109.5 (quaternary-C), 108.8 (quaternary-C), 101.8, 96.6, 83.3, 78.4, 75.7, 75.5, 74.1, 74.0, 69.1, 68.4, 67.7, 26.4, 26.1, 25.3, 24.6; HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₄₆H₅₁NaO₁₂, 819.3351; found, 819.3343.

2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-D-mannopyranosyl-α(1→6)-1,2:3,4-di-*O*-isoprop ylidene-α-D-galactopyranose 5



Disaccharide **5** was prepared from thiomannoside **2** and galactose acceptor **3** by general modulated glycosylation procedure. Purification of **5** was performed by column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:2:15 stepwise to 1:2:10). For disaccharide **5**: $[\alpha]_D^{35}$ –18.34 (*c* 0.52, CHCl₃); R_f 0.375 (EtOAc/CH₂Cl₂/Hexane 1:2:5); ¹H NMR (300 MHz, CDCl₃): δ 8.07 (d, *J* = 8.1 Hz, 2 H), 7.53 (t, *J* = 7.5 Hz, 1 H), 7.40-7.18 (m, 17 H), 5.67 (s, 1 H), 5.52 (d, *J* = 4.8 Hz, 1 H, H-1), 5.04 (s, 1 H), 4.86 (d, *J* = 10.8 Hz, 1 H), 4.78 (t, *J* = 11.1 Hz, 2 H), 4.63-4.51 (m, 4 H), 4.31 (dd, *J* =

4.8, 1.8 Hz, 1 H), 4.24 (d, J = 8.1 Hz, 1 H), 4.12 (s, 2 H), 3.99-3.91 (m, 3 H), 3.87-3.71 (m, 3 H), 1.53 (s, 3H, CH₃), 1.43 (s, 3 H, CH₃), 1.34 (d, J = 6.6 Hz, 6 H, CH₃ × 2); ¹³C NMR (75 MHz, CDCl₃): δ 166.0 (*C*=O), 138.9, 138.8, 138.5, 130.4, 130.3, 128.8, 128.73, 128.71, 128.68, 128.4, 128.03, 127.97, 127.93, 127.86, 109.8 (quaternary-C), 109.0 (quaternary-C), 98.4, 96.7, 78.6, 75.7, 74.7, 73.8, 72.2, 72.0, 71.2, 71.02, 70.97, 69.4, 69.3, 66.6, 66.4, 26.6, 26.4, 25.3, 24.9; HRMS-ESI (*m/z*): [M + Na]⁺ calcd. for C₄₆H₅₁NaO₁₂, 819.3351; found, 819.3339.

p-Tolyl 6-*O*-Acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-thio-β-D-glucopyranoside 6:



Known thioglucoside **s6** (1.0 g, 1.71 mmol) in EtOAc (4 mL) was treated with Ac₂O (0.2 mL, 2.05 mmol), Et₃N (0.35 mL, 2.56 mmol), and DMAP (20 mg, 0.171 mmol). Upon completion of acetylation, the reaction mixture was diluted with EtOAc and washed with satd. NaHCO₃, H₂O, brine, dried (over MgSO₄), filtered, and concentrated for column chromatography purification (Elution: EtOAc/Hexane 1:4) to furnish **6** (quantitative). For compound **6**: $[\alpha]_D^{35}$ +63.1 (*c* 0.30, CHCl₃); *R*_f 0.4 (EtOAc/Hexane 3/7); ¹H-NMR (400 MHz, CDCl₃): δ 7.96–7.91 (m, 4 H), 7.53–7.48 (m, 2 H), 7.39–7.34 (m, 6 H), 7.20-7.17 (m, 3 H), 7.13–7.08 (m, 4 H), 5.73 (t, *J* = 9.6 Hz, 1 H), 5.32 (t, *J* = 10.0 Hz, 1 H), 4.84 (d, *J* = 10.0 Hz, 1 H, H-1), 4.56 (d, *J* = 11.2 Hz, 1 H), 4.50–4.46 (m, 2H), 4.25 (dd, *J* = 12.0, 4.8 Hz, 1 H), 3.83–3.74 (m, 2 H), 2.33 (s, 3 H, CH₃), 2.09 (s, 3 H, CH₃C=O); ¹³C-NMR (100 MHz, CDCl₃): δ 171.1 (*C*=O), 166.1 (*C*=O), 165.8 (*C*=O), 139.0, 137.3, 134.1, 133.73, 133.70, 130.3, 130.2, 130.1, 129.8, 128.90, 128.88, 128.8, 128.7, 128.6, 86.7 (C-1), 77.6, 76.9, 76.1, 75.3, 71.2, 63.3, 21.7, 21.4; HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₃₆H₃₄NaO₈S, 649.1867; found, 649.1900.



p-Tolyl 2,4,6-Tri-*O*-benzyl-thio-α-D-mannopyranoside 13a:

Unprotected thiomannoside s13aa^[s8] (2 g, 7 mmol) was treated with Bu₂SnO (2.3 g, 9.3 mmol) in MeOH (23.3 mL) and heated to 85 °C. Upon Until a clear the reaction reaction mixture solution clearlywas observed, the reaction mixture was cooled to RT, concentrated, and dried high vacuum ca 2h. The reaction residue was treated with cesium fluoride (1.3 g, 8.4 mmol), p-methoxybenzyl chloride (1.4 mL, 10.2 mmol) in DMF. Upon completion of the alkylation, the reaction mixture was filtered and the filtrate was concentrated for column chromatography purification (Elution: EtOAc/Hexane 1/2 stepwise to 1/0) to furnish crude 3-OH protected thiomannoside s13ab (1.78 g). The thiomannoside s13ab (1.78 g, 4.38 mmol) in dried in DMF was treated with benzyl bromide (BnBr) (2.4 mL, 19.7 mmol) and sodium hydride (NaH) (800 mg in 60% mineral oil, 19.7 mmol). Upon completion of the benzylation, the DMF was removed by high *vacuo* rotary evaporator and the residue was taken up with EtOAc. The organic phase was washed with water and, brine, dried over MgSO₄, concentrated, and dried under vacuo for ca 4 h to furnish fully protected thiomannoside s13ac. p-Methoxybenzyl ether (PMB) protection in s13ac was removed by treatment with 2,3-dichloro-5,6-dicyano quinone (DDQ) (835 mg, 3.68 mmol) in 10:1 EtOAc/H₂O (10/1) mixture. Upon completion of the PMB deprotection, the crude reaction mixture was diluted with EtOAc, and the organic phase was washed with Na₂S₂O_{3(aq)}, NaOH_(aq), water and brine. After drying (over MgSO₄) and filtration, the EtOAc solution was concentrated for column chromatography

purification (Elution: EtOAc/CH₂Cl₂/Hexane 1/2/15 stepwise to 1/2/13) to furnish target thiomannoside acceptor **13a** (473 mg, 12%, three steps unoptimized). For thiomannoside **13a**: $[\alpha]_D^{35}$ +115.1 (*c* 0.40, CHCl₃); R_f 0.27 (EtOAc/CH₂Cl₂/Hexane 1:2:6); ¹H NMR (300 MHz, CDCl₃): δ 7.38-7.21 (m, 17H), 7.04 (d, *J* = 8.1 Hz, 2H), 5.61 (s, 1H, H-1), 4.87 (d, *J* = 10.8 Hz, 1H), 4.74 (d, *J* = 11.4 Hz, 1H), 4.64 (d, *J* = 12 Hz, 1H), 4.54 (d, *J* = 11.1 Hz, 1H), 4.48 (dd, *J* = 11.7, 4.8 Hz, 2H), 4.33-4.28 (m, 1H), 4.03-3.96 (m, 2H), 3.86-3.72 (m, 3H), 2.44 (d, *J* = 9 Hz, 1H), 2.30 (s, 3H); ¹³C-NMR (75 MHz, CDCl₃): δ 138.8, 138.7, 132.8, 130.7, 130.2, 129.0, 128.8, 128.7, 128.5, 128.43, 128.37, 128.2, 127.9, 85.7 (C-1), 80.0, 77.3, 75.3, 73.8, 72.7, 72.5, 72.4, 69.6, 21.6 (*C*H₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₃₄H₃₆NaO₅S, 579.2176; found, 579.2192.





Per-*O*-acetyl thiomannoside $s13ba^{[s9]}$ (15.2 g, 39 mmol) in MeOH/CH₂Cl₂ mixture (150 mL) was treated with Na_(s) (120 mg) for deacetylation. Upon completion of deacetylation, the reaction solution was neutralized with IR-120 (H⁺), and the mixture was filtered to remove resin, followed by concentration (by rotary evaporator) and dried (under *vacuo*) to give crude unprotected thiomannoside s13bb. The thiomannoside s13bb (11.1 g, 39 mmol) in anhydrous CH₃CN (300 mL) was treated

with benzaldehyde dimethyl acetal (11.7 mL, 78.0 mmol) and p-toluenylsulfonic acid (TsOH, 740 mg, 3.9 mmol) to form the 4,6-O-benzylidene protected thiomannoside. Upon completion of the acetalation, the reaction was quenched by neutralization with Et₃N. The 4,6-O-benzylidene protected thiomannoside was precipitated in the reaction solution, and it was obtained by filtration and washing the precipitate with minimal amount of hexane/Et₂O mixture. The washed precipitate was dried under vacuo to give benzylidene acetal derivative s13bc (11.9 g). The acetal derivative s13bc (11.9 g, 32.0 mmol) suspended in toluene (160 mL) was treated with dibutyl tin oxide (Bu₂SnO, 11.9 g, 48.0 mmol) and refluxed at 145 °C for 8–10 h under Dean-Stark trap. The volume of toluene was then reduced approximately by half and the reaction mixture was cooled to RT. After then, 2-(bromomethyl)-naphthalene (2-NAP-Br, 10.54 g, 48.0 mmol), cesium fluoride (CsF, 7.2 g, 48.0 mmol), and CH₃CN (80 mL) were added and resulting mixture was stirred at 70 °C for ca. 11 h. Upon the completion of alkylation, the reaction mixture was filtered (over celite), and concentrated for column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 0.1:1:3 stepwise to 0.3:2:2) to furnish amorphous white solid of the 2-naphthylmethyl derivative s13bd (12.3 g). The derivative s13bd (10.24 g, 19.9 mmol) in BH₃.THF solution (1 M, 59.7 mL, 59.7 mmol) was treated with trimethylsilyl triflate (TMSOTf) (180 µL) at 0 °C. Upon completion of the reductive ring opening, the mixture was neutralized with Et₃N, followed by addition of MeOH. The resulting mixture was then concentrated for column chromatography purification (Elution: EtOAc/Hexane 1/5 stepwise to 1/1) to yield 2,6-dihydroxyl thiomannoside s13be (8.8 g). The 2,6-dihydroxyl thiomannoside s13be (9.44 g, 18.3 mmol) in 10% NaOH (13.3 mL) THF solution was treated with BnBr (2.2 mL, 18.3 mmol) and TBAB (5.9 g, 18.3 mmol) and the mixture was stirred vigorously. Upon the completion of benzylation, THF was removed by rotary evaporator. The residual aqueous mixture was extracted with EtOAc, which was washed with H₂O, brine, dried over MgSO₄, and concentrated for column chromatography purification (Elution:

EtOAc/CH₂Cl₂/Hexane 1/1/5) to give surprisingly 2-O-benzyl thiomannoside. The C2 position of benzyl function was confirmed by ¹H, ¹³C, HSQC, and HMBC spectroscopy (HSQC and HMBC spectra were unpublished data). To the 2-O-benzyl thiomannoside in CH₂Cl₂ was added BzCl (2.6 mL, 22.0 mmol), Et₃N (4 mL, 29.4 mmol), and DMAP (180 mg, 1.47 mmol). Upon completion of the benzoylation, satd. NaHCO₃ was added to the mixture and the product was extracted with EtOAc. The EtOAc solution was washed with H2O, brine, dried over MgSO4, filtered, and concentrated for column chromatography purification to give fully protected thiomannoside s13bf. The thiomannoside s13bf (10.45 g, 14.7 mmol) was treated with DDQ (10 g, 44.1 mmol) in 10/1 v/v EtOAc/H₂O mixture. Upon completion of the deprotection, the reaction mixture was diluted with EtOAc, which was washed with satd. Na₂S₂O₃, water, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: EtOAc/Hexane 1:10) to furnish thiomannoside **13b** (6.2 g, 68% from **s13be**). For thiomannoside **13b**: $[\alpha]_D^{35} + 126.4$ (*c* 0.44, CHCl₃); $R_{\rm f}$ 0.3 (EtOAc/Hexane 1:4); ¹H-NMR (300 MHz, CDCl₃): δ 7.99 (d, J = 7.8 Hz, 2H), 7.54 (t, J = 7.5 Hz, 1H), 7.39-7.19 (m, 14H), 6.98 (d, J = 7.8 Hz, 2H), 5.62 (s, 1H), 4.94 (d, J = 11.1 Hz, 1H), 4.73 (d, J = 11.4 Hz, 1H), 4.64 (d, J = 8.1 Hz, 1H), 4.58-4.47 (m, 4H), 1.11-4.04 (m, 2H), 3.80 (t, J = 9 Hz, 1H), 2.58 (d, J = 9.3 Hz, 1H, OH), 2.26 (s, 3H, CH₃); ¹³C-NMR (75 MHz, CDCl₃): δ166.7 (C=O), 138.4, 138.2, 137.8, 133.3, 132.6, 130.3, 130.2, 129.0, 128.9, 128.7, 128.53, 128.48, 128.3, 128.2, 85.4 (C-1), 70.1, 77.0, 75.4, 72.8, 72.5, 70.7, 64.4, 21.5 (CH₃); HRMS-EI (m/z): [M + Na]⁺ calcd. for C₃₄H₃₄NaO₆S, 593.1968; found, 593.1979.

p-Tolyl 2-*O*-Benzoyl-4,6-di-*O*-benzyl-thio-α-D-mannopyranoside 14:



Fully protected thiomannoside s14a^[s9] in BH₃ THF solution (10.8 mL, 10.8

mmol) was treated with TMSOTf (78.0 μL, 0.43 mmol) at 0 °C. Upon completion of the reductive cleavage of acetal function, the reaction was quenched with Et₃N, followed by addition of MeOH to react with excess borane reagent. The resulting mixture was concentrated for column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 1:2.5:10 stepwise to 1:2.5:7) to produce thiomannoside **14**. For thiomannoside **14**: $[\alpha]_D^{35}$ +81.4 (*c* 0.52, CHCl₃); *R*_f 0.19 (EtOAc/Hexane 1/3); ¹H NMR (300 MHz, CDCl₃): δ7.34-7.23 (m, 17 H), 7.09 (d, *J* = 8.1 Hz, 2 H), 5.43 (s, 1 H, H-1), 4.95 (d, *J* = 11.1 Hz, 1 H), 4.73-4.58 (m, 5 H), 4.13 (d, *J* = 9.3 Hz, 1 H), 4.06-3.98 (m, 2 H), 3.89 (dd, *J* = 9, 2.7 Hz, 2 H), 3.80 (s, 2 H), 2.32 (s, 3 H), 1.99 (s, 1 H, OH); ¹³C NMR (75 MHz, CDCl₃): δ138.8, 138.5, 138.4, 138.2, 132.9, 130.5, 130.3, 128.9, 128.5, 128.4, 128.3, 128.21, 128.19, 86.8 (C-1), 80.5, 76.8, 75.7, 75.2, 73.6, 72.7, 72.6, 62.6, 21.6 (CH₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₃₄H₃₆NaO₅S, 579.2176; found, 579.2179.

p-Tolyl 2-*O*-Benzoyl-2-*O*-benzyl-2-(2,2,2-trichloroethoxycarbonylamino)-2deoxy-thio-β-D-glucopyranoside 15:



Per-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy- β -D-thioglucopyr anoside **7** (9.44 g, 16.1 mmol)^[s2] was treated with Na(s) in CH₂Cl₂/MeOH mixture at 0 °C. Upon the completion of the reaction, the reaction was neutralized with IR-120 (H⁺), filtered, concentrated, and dried under *vacuo* to obtain the thioglucoside **s15a** (7.24 g, 98%). **s15a** (5.24 g, 11.4 mmol) was treated with benzaldehyde dimethyl acetal (2.2 mL, 14.8 mmol) and TsOH (230 mg, 1.2 mmol) in dried CH₃CN (48 mL).

Upon completion of the acetal formation, the reaction was neutralized by Et_3N , filtered, concentrated to produce 4,6-O-benzylidene-2-(2,2,2-trichloroethoxycarbonyl amino)-2-deoxy-β-D-thioglucopyranoside **s15b** (4.67 g, 75%). **s15b** (2.5 g, 4.55 mmol) in dried CH₂Cl₂ (23 mL) was treated with BzCl (0.8 mL, 6.83 mmol), Et₃N (1.3 mL, 9.1 mmol), and DMAP (56 mg, 0.46 mmol). Upon completion of benzoylation, the reaction mixture was diluted with EtOAc. The EtOAc solution was washed with satd. NaHCO₃, H₂O, brine, dried (over MgSO₄), filtered, and concentrated. The fully protected $2-(2,2,2-\text{trichloroethoxycarbonylamino})-2-\text{deoxy}-\beta-D-\text{thioglucopyranoside}$ s15c (2 g, 68%) was obtained by precipitation of the concentrated solution in Hexane/EtOAc mixture. The 2-amino-2-deoxy-β-D-thioglucopyranoside s15c (2 g, 3.1 mmol) in BH₃ THF (15.5 mL, 15.5 mmol) was treated with TMSOTf (112 µL, 0.62 mmol) at 0 °C. Upon completion of the reductive clealvage of acetal function, excessive borane reagent was quenched with Et₃N and MeOH. The resulting mixture was concentrated for column chromatography purification (Elution: EtOAc/Hexane 1/4) to furnish C6 hydroxyl unprotected thioglucosaminyl acceptor 15 (1.41 g, 70%). For 15: $[\alpha]_D^{35}$ -6.2 (c 0.42, CHCl₃); R_f 0.25 (EtOAc/Hexane 1/2); ¹H NMR (300 MHz, CDCl₃): δ 7.99 (d, J = 7.8 Hz, 2 H), 7.58 (t, J = 7.2 Hz, 1 H), 7.44-7.38 (m, 4 H), 7.12-7.00 (m, 7 H), 5.98 (d, J = 9.9 Hz, 1 H, NH), 5.60 (t, J = 9.9 Hz, 1 H, H-2), 4.74 (d, J = 10.5 Hz, 1 H), 4.63 (q, J = 11.7 Hz, 2 H), 4.50 (s, 2 H), 3.98 (q, J = 10.2 Hz, 1 H), 3.85-3.66 (m, 3 H), 3.48 (d, J = 9.3 Hz, 1 H), 2.28 (s, 3 H, CH₃), 2.08 (s, 1 H, OH); ¹³C NMR (75 MHz, CDCl₃): δ167.2 (C=O), 154.9 (C=O), 138.8, 137.5, 134.0, 130.4, 130.2, 129.5, 129.3, 129.0, 128.8, 128.7, 128.4, 95.8 (CCl₃), 88.2 (C-1), 79.6, 75.8, 75.3, 74.8, 62.0, 55.8, 21.6 (CH₃); HRMS-EI (m/z): $[M + Na]^+$ calcd. for C₃₀H₃₀Cl₃NNaO₇S, 676.0701; found, 676.0719.

(4-t-Butyl-2-methylphenyl) 2,3-O-Isopropylidene-thio-α-L-rhamnopyranoside 17



L-Rhamnose (5 g, 30.5 mmol) in acetic anhydride (Ac₂O, 17.3 mL, 183 mmol) was treated with dried TsOH (580 mg, 3.05 mmol) in ice bath. Upon completion of acetylation, the anhydride reagent was quenched with MeOH under ice bath ca 1 h and concentrated. After then, the concentrated residue was diluted with EtOAc, washed with ca. 10% NaOH, brine, ice, and dried over MgSO₄. The solution was filtered, concentrated, and dried under vacuo to give crude per-O-acetyl rhamnosyl acetate. The crude acetate in dried CH2Cl2 was cooled to -10 °C and treated with (4-t-butyl-2-methyl)-thiophenol (3.2 mL, 17.6 mmol) and BF₃·OEt₂ (3 mL, 23.4 mmol). Upon completion of thioglycosidation, the reaction was quenched with 10% NaOH_(aq) at 0 °C and the mixture was washed with 10% NaOH_(aq), chilled H₂O, brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: EtOAc/Hexane 1/8) to furnish per-O-acetyl thiorhamnoside s17a (4.34 g, 74%). s17a (4.34 g, 8.6 mmol) in MeOH/CH₂Cl₂ mixture (35 mL, MeOH/CH₂Cl₂ 1/2) was treated with Na(s). Upon completion of the deacetylation, the reaction mixture was neutralized with IR-120 (H⁺), and filtered to remove resin. The filtrate was concentrated and dried under vacuo to give crude deacetylated thiorhamnoside intermediate. The crude thiorhamnoside intermediate in dried acetone (29 mL) was then treated with 2,2-dimethoxypropane (2.1 mL, 17.2 mmol) and TsOH (170 mg, 0.86 mmol). Upon completion of the acetalation, the reaction was neutralized with Et₃N, and followed by concentration for column chromatograph purification (Elution: EtOAc/CH₂Cl₂/Hexane 1:1:6 stepwise to 1:1:5) to furnish thiorhamnoside acceptor 17 (2.17 g, 69%, α/β 10:1). For thiorhamnoside 17: $[\alpha]_D^{35}$ -170.0 (c 0.50, CHCl₃); R_f 0.423 (EtOAc/CH₂Cl₂/Hexane 1:1:2); ¹H-NMR (300 MHz, CDCl₃): δ 7.59 (d, J = 2.1 Hz, 1 H, ArH), 7.21 (dd, J = 7.8, 1.8 Hz, 1 H, ArH), 7.14 (d, J = 8.1 Hz, 1 H, ArH),

5.74 (s, 1 H, H-1), 4.40 (d, J = 5.4 Hz, 1 H), 4.18-4.06 (m, 2 H), 3.50-3.43 (m, 1 H), 2.88 (d, J = 3.6 Hz, 1 H), 1.55 (s, 3 H, CH_3), 1.39 (s, 3 H, CH_3), 1.30 (s, 9 H, tBu-H), 1.24 (d, J = 6 Hz, 3 H, $CH_3 \times 2$); ¹³C-NMR (75 MHz, CDCl₃): δ 150.1, 136.9, 132.5, 130.4, 130.1, 125.3, 110.2 (quaternary-C), 83.3 (C-1), 79.0, 77.4, 75.7, 67.4, 34.9, 31.8, 28.6, 26.9, 20.6, 17.6; HRMS-ESI (m/z): [M + Na]⁺ calcd. for C₂₀H₃₀NaO₄S, 389.1757; found, 389.1761.

Entry	Donor	DMF	$T(^{\circ}\mathrm{C})^{a}$	Time (h)	Product, 4		
(equiv	(equiv)	(equiv)			Yield%	α/β ratio	
1	1 (1.2)	1.2	0	4	50	β only	
2	1 (1.2)	1.2	-10	4	75	β only	
3	1 (1.2)	1.2	-20	5	75	β only	
4	1 (1.2)	1.2	-40	5	71	β only	
5	1 (1.2)	1.2	-60	6	60	β only	
^{<i>a</i>} T = temperature at pre-activation and coupling reaction							

Table S1: Temperature optimization for DMF-modulated glycosylation of diacetonidegalactose acceptor 3 with thioglucoside 1.

Table S2: Characteristic NMR data for reaction intermediates 40α , 40β , and 41

Entry	Selected NMR data	Observed reaction intermediates			
		40α	40β	41	
1	¹ H signal at C-1/ppm (${}^{3}J_{\rm HH}/\rm Hz$)	6.19 (s)	5.79 (7.8)	5.22 (9.0)	
2	¹ H signal at C-2/ppm	5.22	5.30	5.87	
3	¹ H signal at C-1/ppm ($^{1}J_{CH}/Hz$)	103.0 (183)	102.0 (174)	78.6 (ND)	
4	¹ H signal of imidate /ppm	8.75	9.17	-	

General DMF-modulated glycosylation procedure for participating thioglycosyl donors (refer to Table 2 in main text): 1.2 Equiv of thioglycoside donor (1, 2, 6, 7, or 8), DMF (1.2 equiv), and activated molecular sieve (4 Å) were suspended in dried CH₂Cl₂. The mixture was stirred at RT for 5 min and cooled to -30, -20 or -10 °C at -15least 15 min in cooling reactor (Eyela model: PCL 1810 or PCL 1800). Subsequently, 1.2 equiv of NIS and 1.2 (or 1.8 for thiomannoside donors) equiv of TMSOTf (for most of thioglycoside acceptors used) or TfOH (for glycosylation of **13b**) were added. Upon completion of pre-activation at -30, -20 or -10 °C, acceptors (**9**–**17**, 1 equiv) was added to reaction solution. The temperature for each coupling reaction needed optimization. Upon completion of glycosylation, the reaction was quenched with satd. NaHCO₃ and solid Na₂S₂O₃, then stirred vigorously at RT until the dark red coloration of the solution turning to pale yellow. The mixture was dried by MgSO₄ powder, followed by filtration and concentration to give crude concentrate for column chromatography purification giving the glycosylation products **18–32**. Exact amount of substrates and reagents used were given in Table S3



F	90-F-02	1. 1.2 equiv DN STol 2. NIS, TMSO	//F, ^{[f} 9–17	PgO-E-	-0	OR/SR
	R-	T₁ ºC, CH₂Cl₂	T₂°C) P		
	ر 1, 2, 6 ,7 or 8			<u>к_//</u>	18–32	
Entry	Donor, (mg,	Acceptor, (mg,	NIS (mg,	TMSOTf	T_{1}, T_{2}	Product, mg,
	mmol)	mmol)	mmol)	(µL, mmol)	$(^{\circ}C)^{[a]}$	yield (%)
1	1, 150, 0.23	9 , 88, 0.19	55.8, 0.25	41, 0.23	-10, -10	18 , 125, 65
2	1, 250, 0.38	10 , 176, 0.32	93, 0.41	68, 0.38	-10, -10	19 , 216, 67
3	1 , 172, 0.26	11a , 120, 0.22	64, 0.28	47, 0.260	-10, -10	20 , 164, 70
5	1, 143, 0.22	16 , 100, 0.18	53, 0.23	39, 0.22	-10, -10	21 , 137, 70
6	1, 150, 0.227	17, 69, 0.19	55.8, 0.25	41, 0.23	-10, -10	22 , 84, 55
7	6, 200, 0.3	11a , 141, 0.25	72, 0.3	86, 0.48	-20, -10	23 , 178, 66
8	6, 200, 0.3	11b , 149, 0.25	72, 0.3	86, 0.48	0, 10	24 , 172, 63
9	2 , 500, 0.758	12 , 384, 0.69	188, 0.828	206, 1.14	-20, 0	25 ,339, 61
10	2 , 200, 0.303	13a , 141, 0.253	74.7, 0.329	82.1, 0.46	-20, -20	26 , 166, 60
11	2 , 2000, 3.03	13b , 1440, 2.53	717, 3.16	767, 4.56 ^[b]	-20, 0	27 , 2050, 73
12	2 , 150, 0.23	14 , 105, 0.19	55.8, 0.25	61, 0.34	-10, -10	28 , 128, 62
13	7, 182, 0.31	11a , 144, 0.26	76.5, 0.34	56.1, 0.31	-10, -10	29 , 183, 64
14	7, 100, 0.17	15 , 94, 0.14	42, 0.19	31, 0.17	-10, -10	30 , 64, 40

15	8, 200, 0.30	16 , 134, 0.24	68, 0.30	65, 0.36	-30, -30	31 , 186, 57	
				1			

^a T_1 = pre-activation temperature; T_2 = coupling temperature. ^b2.5 M stock solution of triflic acid (TfOH in Et₂O) was used as acid promoter.

Methyl 4-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-2,3,6-tri-*O*benzyl-α-D-glucopyranoside 18:



Disaccharide 18 was prepared from thioglucoside 1 and methyl glucoside 9 (Table S3, entry 1). Purification of 18 was achieved by standard column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 0.5:4:7 stepwise to 1:4:5). For disaccharide **18**: $[\alpha]_D^{35}$ +30.6 (*c* 0.36, CHCl₃); *R*_f 0.35 (EtOAc/CH₂Cl₂/Hexane 1/4/5); ¹H-NMR (300 MHz, CDCl₃): δ 7.88 (d, J = 7.2 Hz, 2H, ArH), 7.56 (t, J = 7.5 Hz, 1H, ArH), 7.43–7.35 (m, 6H, ArH), 7.32–7.18 (m, 20H, ArH), 7.14–7.05 (m, 6H, ArH), 5.23 (dd, J = 9.3, 8.1 Hz, 1H), 5.08 (d, J = 11.4 Hz, 1H), 4.81 (s, 1H), 4.77 (s, 1H), 4.75–4.68 (m, 2H), 4.66–4.63 (m, 1H), 4.60–4.54 (m, 3H), 4.50 (dd, J = 9.9, 6.3 Hz, 2H), 4.45 (d, J = 12.3 Hz, 1H), 4.36 (d, J = 12.3 Hz, 1H), 4.26 (d, J = 12.0 Hz, 1H), 3.91-3.82 (m, 2H), 3.76-3.68 (m, 2H), 3.63-3.59 (m, 2H), 3.56-3.49 (m, 2H), 3.45–3.35 (m, 3H), 3.25 (s, 3H, OCH₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.3, 140.1, 138.9, 138.8, 138.5, 138.4, 138.2, 133.6, 130.2, 129.0, 128.90, 128.87, 128.79, 128.76, 128.70, 128.61, 128.57, 128.5, 128.44, 128.37, 128.3, 128.24, 128.20, 128.12, 128.08, 128.07, 127.9, 127.5, 100.8 (C-1'), 98.8 (C-1), 83.3, 80.7, 79.3, 78.6, 77.2, 75.9, 75.8, 75.6, 75.3, 74.7, 74.0, 73.9, 70.0, 69.2, 68.2, 55.7; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₆₂H₆₄NaO₁₂, 1023.4290; found, 1023.4347.

p-Tolyl 4-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-2,3,6-tri-*O*-benzyl-thio-β-D-glucopyranoside 19:



Disaccharide **19** was prepared from thioglucosides **1** and **10** (Table S3, entry 2). Purification of **19** was achieved by standard column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:1:15). For disaccharide **19**: $[\alpha]_D^{35}$ +10.1 (*c* 0.34, CHCl₃); *R*_f 0.47 (EtOAc/CH₂Cl₂/Hexane 1:1:4); ¹H-NMR (300 MHz, CDCl₃): δ 7.92 (d, *J* = 7.8 Hz, 2 H, Ar*H*), 7.55 (t, *J* = 7.2 Hz, 1H, Ar*H*), 7.43-7.22 (m, 25H, Ar*H*), 7.20-7.16 (m, 5H, Ar*H*), 7.10-7.09 (m, 5H, Ar*H*), 6.97 (d, *J* = 7.8 Hz, 2H, Ar*H*), 5.25 (t, *J* = 8.4 Hz, 1 H), 5.14 (d, *J* = 11.4 Hz, 1H), 4.80-4.65 (m, 6H), 4.60 (s, 1H), 4.55 (d, *J* = 10.2 Hz, 2H), 4.47 (d, *J* = 9.9 Hz, 1H), 4.38-4.32 (m, 3H), 3.96 (t, *J* = 9.6 Hz, 1H), 3.85-3.48 (m, 8H), 3.42-3.35 (m, 2 H), 3.2 (d, *J* = 9.3 Hz, 1H), 2.26 (s, 3H, C*H*₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.3 (*C*=O), 139.7, 138.74, 138.7, 138.6, 138.4, 138.2, 138.0, 133.6, 133.1, 130.1, 130.0, 128.92, 128.88, 128.8, 128.70, 128.69, 128.66, 128.6, 128.5, 128.32, 128.29, 128.21, 128.16, 128.1, 128.03, 128.01, 127.9, 127.5, 100.9 (C-1'), 88.1 (C-1), 85.3, 83.3, 80.7, 79.0, 78.6, 77.0, 75.78, 75.74, 75.5, 75.3, 74.7, 73.9, 73.8, 69.1, 68.6, 21.5 (CH₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4436.

p-Tolyl 6-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-2,3,4-tri-*O*-benzyl-thio-β-D-glucopyranoside 20:



Disaccharide **20** was prepared from thioglucosides **1** and **11a** (Table S3, entry 3). Purification of **20** was achieved by standard column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:1:6). For disaccharide **20**: $[\alpha]_D^{35}$ +4.9 (*c* 0.32, CHCl₃); *R*_f 0.45 (EtOAc/Hexane 1:2); ¹H-NMR (300 MHz, CDCl₃): δ 7.92 (d, J = 6.9 Hz, 2H, Ar*H*), 7.50–7.43 (m, 3H, Ar*H*), 7.38–7.18 (m, 25H, Ar*H*), 7.13–7.11 (m, 7H, Ar*H*), 7.07–7.03 (m, 2H, Ar*H*), 5.36 (t, J = 7.8 Hz, 1H), 4.84 (d, J = 3.3 Hz, 1H), 4.81 (d, J = 2.4 Hz, 1H), 4.78 (s, 1H), 4.69-4.64 (m, 3H), 4.62 (s, 1H), 4.60 (s, 1H), 4.57 (s, 1 H), 4.54-4.48 (m, 2H), 4.40 (d, J = 10.8 Hz, 1H), 4.13 (d, J = 10.8 Hz, 1H), 3.85–3.71 (m, 5H), 3.57 (t, J = 7.2 Hz, 2H), 3.41 (p, J = 9.6 Hz, 3H), 2.30 (s, 3H, C*H*₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.5 (*C*=O), 138.8, 138.6, 138.4, 138.3, 138.21, 138.17, 133.4, 133.3, 130.2, 130.1, 130.1, 128.9, 128.80, 128.76, 128.70, 128.6, 128.4, 128.2, 128.1, 128.0, 101.3 (C-1'), 87.9 (C-1), 87.0, 83.3, 80.7, 79.2, 78.5, 77.8, 76.0, 75.8, 75.7, 75.5, 75.4, 75.2, 74.2, 74.0, 69.3, 68.0, 21.6 (CH₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4436.

p-Tolyl 6-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-2,3,4-tri-*O*benzyl-thio-β-D-galactopyranoside 21:



Disaccharide **21** was prepared from thioglucoside **1** and thiogalactoside **16** (Table S3, entry 5). Purification of **21** was achieved by standard column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:3:11). For disaccharide **21**: $[\alpha]_D^{35}$ +17.5 (*c* 0.42, CHCl₃); R_f 0.41 (EtOAc/CH₂Cl₂/Hexane 1:2:5); ¹H NMR (300 MHz, CDCl₃): δ 7.95 (d, *J* = 7.2 Hz, 2H, Ar*H*), 7.50 (t, *J* = 7.2 Hz, 1H, Ar*H*), 7.42 (t, *J* = 8.4 Hz, 3H, Ar*H*), 7.37–7.21 (m, 24H, Ar*H*), 7.20–7.15 (m, 2H, Ar*H*), 7.11 (s, 5H, Ar*H*), 6.97 (d, *J* = 7.8 Hz, 2H, Ar*H*), 5.27 (t, *J* = 8.7 Hz, 1H), 4.82 (dd, *J* = 11.4, 9 Hz, 2H), 4.75–4.64 (m, 5 H), 4.61–4.58 (m, 2H), 4.54 (s, 1H), 4.47 (d, *J* = 5.7 Hz, 1H), 4.43 (d, *J* = 3.3 Hz, 1 H), 4.39 (d, *J* = 3.6 Hz, 2H), 4.01 (dd, *J* = 7.5 Hz, 1H), 3.84–3.71 (m, 7H), 3.52–3.47 (m, 1H), 3.41 (t, *J* = 6.0 Hz, 1H), 3.33 (dd, *J* = 9.3,2.7 Hz, 1H), 2.25

(s, 3H, *CH*₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.6 (*C*=O), 139.2, 138.8, 138.6, 138.40, 138.36, 138.2, 137.8, 133.7, 133.0, 130.3, 130.2, 130.1, 130.0, 129.0, 128.84, 128.82, 128.8, 128.73, 128.71, 128.7, 128.5, 128.4, 128.35, 128.30, 128.2, 128.09, 128.07, 128.0, 127.9, 127.7, 101.5 (C-1'), 88.3 (C-1), 84.5, 83.1, 78.2, 77.3, 76.0, 75.55, 75.47, 75.45, 74.8, 74.3, 73.9, 73.4, 72.6, 68.9, 67.8, 21.6 (*C*H₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4465.

4-(t-Butyl)-2-methylphenyl4-O-(2-O-Benzoyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl)-2,3-O-isopropylidene-thio-α-L-rhamnopyranoside 22:



Disaccharide **22** was prepared from thioglucoside **1** and thiorhamnoside **17** (Table S3, entry 6). Purification of **22** was achieved by column chromatography purification (Elution: EtOAc/Hexane 1:15). For disaccharide **22**: $[\alpha]_D^{35}$ –77.9 (*c* 0.83, CHCl₃); *R*_f 0.3 (EtOAc/Hexane 1:4); ¹H-NMR (300 MHz, CDCl₃): δ 8.11 (d, *J* = 6.9 Hz, 2H, Ar*H*), 7.61-7.56 (m, 1H, Ar*H*), 7.51 (d, *J* = 2.1 Hz, 1H, Ar*H*), 7.45 (t, *J* = 7.8 Hz, 2H, Ar*H*), 7.33-7.09 (m, 17H, Ar*H*), 5.63 (s, 1H), 5.26 (t, *J* = 8.1 Hz, 1H), 4.99 (d, *J* = 7.8 Hz, 1H), 4.84 (d, *J* = 10.8 Hz, 1H), 4.76–4.68 (m, 2H), 4.65–4.53 (m, 3H), 4.22 (d, *J* = 5.7 Hz, 1H), 4.07–4.00 (m, 2H), 3.89–3.74 (m, 4H), 3.63 (dd, *J* = 9.9, 7.5 Hz, 1H), 3.52 (d, *J* = 9.3 Hz, 1H), 2.34 (s, 3H, C*H*₃), 1.48 (s, 3H, C*H*₃), 1.28–1.26 (m, 15H); ¹³C-NMR (75 MHz, CDCl₃): δ 166.0 (*C*=O), 150.1, 138.6, 138.5, 138.4, 137.1, 133.5, 132.4, 130.8, 130.4, 128.9, 128.84, 128.76, 128.7, 128.44, 128.41, 128.3, 128.09, 128.06, 125.3, 109.7 (quaternary-C), 101.0 (C-1'), 83.4 (C-1), 80.8, 78.4, 77.4, 75.8, 75.4, 74.5, 74.0, 69.1, 66.3, 34.9, 31.8, 28.4, 26.8, 20.6, 18.0; HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₅₄H₆₂NaO₁₀S, 925.3956; found, 925.4029.

p-Tolyl 6-*O*-(6-*O*-Acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-β-D-glucopyranosyl)-2,3,4tri-*O*-benzyl-thio-β-D-glucopyranoside 23:



Disaccharide 23 was prepared from thioglucosides 6 and 11a by the general DMF-modulated glycosylation procedure (Table S3, entry 7). The disaccharide 23 was obtained as white semisolid after column chromatography purification (Elution: EtOAc/Hexane 1:4). For disaccharide 23: $[\alpha]_D^{35}$ +28.0 (c 0.80, CHCl₃); R_f 0.3 (EtOAc/Hexane 3:7); ¹H NMR (400 MHz,CDCl₃): δ 7.97 (d, J = 8.0 Hz, 2H, ArH), 7.81 (d, J = 8.0 Hz, 2H, ArH), 7.53–7.47 (m, 4H, ArH), 7.40-7.37 (m, 5H), 7.34–7.13 (m, 19H, Ar*H*), 7.09–7.07 (m, 2H, Ar*H*), 5.70 (t, *J* = 9.2 Hz, 1H), 5.44–5.39 (m, 1H), 4.87-4.71 (m, 5H), 4.66 (d, J = 10.4 Hz, 1H), 4.60-4.53 (m, 3H), 4.50 (d, J = 1.6 Hz, 1H), 4.45 (d, J = 12.0 Hz, 1H), 4.39 (d, J = 11.2 Hz, 1H), 4.27 (dd, J = 12.4, 4.0 Hz, 1H), 3.78 (d, J = 11.2 Hz, 1H), 3.67 (d, J = 9.2 Hz, 1H), 3.60-3.56 (m, 1H), 3.43-3.35 (m, 3H), 2.34 (s, 3H, CH₃), 2.05 (s, 3H, CH₃CO); ¹³C NMR (100 MHz, CDCl₃): δ 171.2 (C=O), 166.1 (C=O), 165.7 (C=O), 138.8, 138.53, 138.48, 138.3, 137.3, 133.7, 133.5, 133.3, 130.4, 130.22, 130.18, 130.0, 129.82, 129.76, 128.90, 128.87, 128.85, 128.83, 128.75 128.7, 128.6, 128.3, 128.24, 128.18, 128.13, 128.10, 127.9, 127.8, 101.2 (C-1'), 88.2 (C-1), 81.0, 79.2, 76.2, 76.1, 75.8, 75.7, 75.3, 75.2, 75.1, 73.5, 72.5, 68.2, 63.1, 21.6, 21.4; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₆₃H₆₁NaO₁₃S, 1081.3803; found, 1081.3784.

p-Tolyl 6-*O*-(6-*O*-Acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-β-D-glucopyranosyl)-2,3-di-*O*-benzoyl-4-*O*-benzyl-thio-β-D-glucopyranoside 24:



Disaccharide 24 was prepared from thioglucosides 6 and 11b by the general DMF-modulated glycosylation procedure (Table S3, entry 8). Purification of 24 was achieved by column chromatography purification (Elution: EtOAc/Hexane 1:4). For disaccharide 24: $[\alpha]_{D}^{35}$ +41.5 (c 0.13, CHCl₃); R_f 0.2 (EtOAc/Hexane 1:5 two times); ¹H-NMR (400 MHz, CDCl₃): δ 7.98 (d, J = 6.8 Hz, 2H, ArH), 7.90 (dd, J = 14.4, 7.2Hz, 4H, ArH), 7.83 (d, J = 7.2 Hz, 2H, ArH), 7.54–7.46 (m, 4H, ArH), 7.43-7.30 (m, 9H), 7.27 (t, *J* = 7.6 Hz, 2H, Ar*H*), 7.21-7.20 (m, 3H, Ar*H*), 7.17-7.10 (m, 6H, Ar*H*), 6.95-6.93 (m, 2H, Ar*H*), 5.74 (t, *J* = 9.2 Hz, 1H), 5.61 (t, *J* = 9.2 Hz, 1H), 5.46 (dd, *J* = 9.6, 1.6 Hz, 1H), 5.27 (t, J = 10.0 Hz, 1H), 4.80 (d, J = 7.6 Hz, 1H), 4.76 (d, J = 8.0Hz, 2H), 4.63-4.50 (m, 4H), 4.34-4.27 (m, 3H), 4.16 (d, J = 11.2 Hz, 1H), 3.94 (t, 9.6 Hz, 1H), 3.84 (dd, J = 8.7, 3.3 Hz, 1H), 3.75-3.69 (m, 2H), 3.64 (dd, J = 10.0, 6.4 Hz, 1H), 2.33 (s, 3H, CH₃), 2.08 (s, 3H, CH₃CO); ¹³C NMR (100 MHz, CDCl₃): δ 171.1 (C=O), 166.1 (C=O), 166.0 (C=O), 165.7 (C=O), 165.6 (C=O), 139.0, 137.6, 137.3, 134.1, 133.8, 133.6, 130.3, 130.24, 130.21, 130.18, 130.16, 129.9, 129.8, 128.93, 128.89, 128.8, 128.74, 128.69, 128.6, 128.3, 101.3 (C-1), 86.7 (C-1), 79.4, 76.9, 76.2, 76.1, 75.7, 75.22, 75.15, 73.6, 72.5, 71.2, 68.2, 63.1, 21.7, 21.4; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₆₃H₅₇NaO₁₅S, 1109.3389; found, 1109.3373.

p-Tolyl 2-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-3,4,6-tri-*O*-benzyl-thio-α-D-mannopyranoside 25:



Disaccharide **25** was prepared from thiomannosides **2** and **12** by the general DMF-modulated glycosylation procedure for participating donor (Table S3, entry 9). **25** was purified by standard column chromatography (Et₂O/CH₂Cl₂/Hexane 0.5:2:10 stepwise to 0.1:2:8). For disaccharide **25**: $[\alpha]_D^{35}$ +42.5 (*c* 0.33, CHCl₃); R_f 0.4 (Et₂O/CH₂Cl₂/Hexane 1:1:4); ¹H-NMR (300 MHz, CDCl₃): δ 8.07 (d, *J* = 7.5 Hz, 2H, Ar*H*), 7.54 (t, *J* = 7.2 Hz, 1H, Ar*H*), 7.39-7.12 (m, 34H, Ar*H*), 7.00 (d, *J* = 8.1 Hz, 2H, Ar*H*), 5.76 (s, 1H), 5.58 (s, 1 H), 5.17 (s, 1H), 4.90 (d, *J* = 10.8 Hz, 1H), 4.82 (d, *J* = 10.8 Hz, 1H), 4.76-4.61 (m, 5 H), 4.57 (s, 1H), 4.49 (s, 1H), 4.46-4.40 (m, 3H), 4.30 (s, 1H), 4.23 (s, 1H), 4.08-4.06 (m, 1H), 4.02-3.96 (m, 2H), 3.93-3.91 (m, 2H), 3.85-3.71 (m, 3H), 3.63 (d, *J* = 10.5 Hz, 1H), 2.54 (s, 3H, C*H*₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.9 (*C*=O), 138.94, 138.85, 138.8, 138.5, 138.1, 133.5, 132.8, 130.7, 130.4, 130.2, 128.9, 128.81, 128.75, 128.70, 128.67, 128.52, 128.46, 128.33, 128.25, 128.2. 128.1, 128.0, 127.9, 127.8, 100.2 (C-1'), 88.0 (C-1), 80.4, 78.5, 75.7, 75.6, 75.3, 74.8, 73.6, 73.3, 72.8, 72.6, 72.1, 69.7, 69.5, 69.3, 21.5 (CH₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4451.

p-Tolyl 3-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-2,4,6-tri-*O*-benzyl-thio-α-D-mannopyranoside 26:



Disaccharide **26** was prepared from thiomannosides **2** and **13a** by the general DMF-modulated glycosylation procedure (Table S3, entry 10). Purification of **26** was performed with column chromatography (Elution: Et₂O/CH₂Cl₂/Hexane 1:1:16 stepwise to 1:1:11). For disaccharide **26**: $[\alpha]_D^{35}$ +46.9 (*c* 0.35, CHCl₃); *R*_f 0.4 (Et₂O/CH₂Cl₂/Hexane 1:1:4); ¹H NMR (300 MHz, CDCl₃): δ 8.06 (d, *J* = 7.2 Hz, 2H,

Ar*H*), 7.53 (t, J = 7.5 Hz, 1H, Ar*H*), 7.38–7.16 (m, 34H, Ar*H*), 7.03 (d, J = 8.1 Hz, 2H, Ar*H*), 5.78 (t, J = 2.1 Hz, 1H), 5.56 (s, 1H), 5.38 (d, J = 1.2 Hz, 1H), 4.89 (d, J = 10.8 Hz, 1H), 4.78 (dd, J = 12.3, 10.8 Hz, 2H), 4.72–4.63 (m, 3H), 4.58–4.45 (m, 6H), 4.30 (dd, J = 9, 3.6 Hz, 1H), 4.20–4.12 (m, 4H), 4.04 (t, J = 9.6 Hz, 1H), 4.97 (d, J = 9.6 Hz, 1H), 3.87–3.70 (m, 4H), 2.28 (s, 3H, C*H*₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.8 (*C*=O), 139.0, 138.8, 138.7, 138.3, 138.2, 137.9, 133.5, 132.6, 130.9, 130.3, 130.2, 130.1, 128.84, 128.79, 128.76, 128.71, 128.67, 128.6, 128.44, 128.42, 128.2, 128.01, 127.97, 127.89, 127.87, 127.85, 100.2 (C-1'), 85.9 (C-1), 79.2, 78.5, 75.6, 75.4, 74.8, 73.9, 73.7, 73.2, 72.8, 71.8, 69.6, 69.5, 21.5 (*C*H₃); HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4472.

p-Tolyl 3-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-6-*O*-benzoyl-2,4-di-*O*-benzyl-thio-α-D-mannopyranoside 27:



Disaccharide **27** was prepared from thiomannosides **2** and **13b** by the general DMF-modulated glycosylation procedure (Table S1, entry 11). Purification of **27** was performed with column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 0.2:2:8 stepwise to 0.4:2:8). For disaccharide **27**: $[\alpha]_D^{35}$ +50.2 (*c* 0.33, CHCl₃); *R*_f 0.44 (EtOAc/Hexane 1/2); ¹H NMR (300 MHz, CDCl₃): δ 8.08 (d, *J* = 7.2 Hz, 2H, Ar*H*), 8.00 (d, *J* = 7.2 Hz, 2H, Ar*H*), 7.53 (dd, *J* = 13.8, 7.2 Hz, 2H, Ar*H*), 7.39-7.16 (m, 31H, Ar*H*), 7.00 (d, *J* = 8.1 Hz, 2H, Ar*H*), 5.79 (s, 1 H), 5.57 (s, 1 H), 5.40 (s, 1H), 4.89 (t, *J* = 10.8 Hz, 2H), 4.76 (d, *J* = 11.4 Hz, 1H), 4.72 (t, *J* = 12.3 Hz, 2H), 4.60-4.52 (m, 7H), 4.50-4.46 (m, 1H), 4.24-4.10 (m, 4H), 4.06 (d, *J* = 5.7 Hz, 2H), 3.83-3.75 (m, 2H), 2.26 (s, 3H); ¹³C-NMR (75 MHz, CDCl₃): δ 166.7 (*C*=O), 165.9 (*C*=O), 138.9, 138.7, 138.24, 138.17, 138.1, 137.8, 133.6, 133.3, 132.4, 130.6, 130.3,

130.22, 130.17, 128.94, 128.87, 128.8, 128.74, 128.71, 128.69, 128.65, 128.4, 128.2, 128.03, 127.95, 127.9, 100.4 (C-1'), 85.6 (C-1), 80.8, 79.4, 78.4, 75.9, 75.4, 75.1, 74.8, 74.0, 72.9, 72.0, 71.7, 71.4, 69.7, 69.5, 64.1, 21.5 (*C*H₃); HRMS-ESI (*m/z*): [M + Na]⁺ calcd. for $C_{68}H_{66}NaO_{12}S$, 1129.4167; found, 1129.4195.

p-Tolyl 6-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-2,3,4-tri-*O*-benzyl-thio-α-D-mannopyranoside 28:



Disaccharide 28 was prepared from thiomannosides 2 and 14 by the general DMF-modulated glycosylation procedure for participating donors (Table S3, entry 12). Purification of 28 was performed with column chromatography (Elution: EtOAc/CH₂Cl₂/Hexane 1:2:20 stepwise to 1:2:10). For disaccharide 28: $[\alpha]_D^{35}$ +30.8 (c 0.34, CHCl₃); R_f 0.325 (EtOAc/CH₂Cl₂/Hexane 1:2:10); ¹H-NMR (300 MHz, CDCl₃): δ 8.08 (dd, J = 8.1, 0.9 Hz, 2H, ArH), 7.52 (t, J = 7.5 Hz, 1H, ArH), 7.39–7.12 (m, 34H, Ar*H*), 7.07 (d, *J* = 8.1 Hz, 2H, Ar*H*), 5.74 (s, 1H), 5.53 (d, *J* = 1.5 Hz, 1H), 5.07 (d, J = 1.8 Hz, 1H), 4.95 (d, J = 1.1 Hz, 1H), 4.89 (d, J = 1.1 Hz, 1H), 4.79–4.69 (m, 3H), 4.64–4.59 (m, 3H), 4.54 (s, 1H), 4.50–4.43 (m, 3H), 4.27 (dd, J = 9.6, 3.6 Hz, 1H), 4.11 (d, J = 6 Hz, 2H), 4.04–3.97 (m, 3H), 3.89–3.79 (m, 3H), 3.70 (t, J = 9.9 Hz, 2H), 2.16 (s, 3H, CH₃); ¹³C-NMR (75 MHz, CDCl₃): δ 165.9 (C=O), 139.1, 138.9, 138.8, 138.5, 138.3, 137.9, 133.4, 131.9, 131.3, 130.4, 130.3, 128.86, 128.84, 128.79, 128.74, 128.71, 128.6, 128.5, 128.32, 128.3, 128.20, 128.16, 128.02, 127.96, 127.90, 127.86, 98.7 (C-1'), 86.3 (C-1), 80.7, 78.2, 76.5, 75.7, 75.6, 75.0, 74.6, 72.6, 72.3, 72.3, 72.0, 71.7, 69.3, 69.1, 67.3, 21.4 (CH₃); HRMS-ESI (m/z): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4374; found, 1115.4421.

p-Tolyl 6-*O*-[3,4,6-Tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxyβ-D-glucopyranosyl]-2,3,4-tri-*O*-benzyl-thio-β-D-glucopyranoside 29:



Disaccharide 29 was prepared from thioglucosaminyl donor 7 and thioglucoside acceptor **11a** by the general DMF-modulated glycosylation procedure for participating donors (Table S3, entry 13). Purification of 29 was performed by column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 0.2:1:2 stepwise to 0.5:1:2). For disaccharide **29**: $[\alpha]_D^{35}$ -0.6 (*c* 0.33, CHCl₃); R_f 0.36 (EtOAc/CH₂Cl₂/Hexane 1:1:2); ¹H-NMR (300 MHz, CDCl₃): δ 7.48 (d, J = 8.1 Hz, 2H, ArH), 7.39–7.30 (m, 13H, ArH), 7.25–7.21 (m, 4H, ArH), 5.05–4.98 (m, 2H), 4.89 (dd, J = 10.8, 6.3 Hz, 2H), 4.84 (s, 1H), 4.81 (s, 1H), 4.74–4.67 (m, 3H), 4.62 (d, 12.6 Hz, 2H), 3.76–3.66 (m, 3H), 3.63–3.59 (m, 1H), 3.52–3.44 (m, 2H), 3.41–3.32 (m, 1H), 2.34 (s, 3H, CH₃), 2.07 (s, 3H, CH₃C=O), 2.03 (s, 3H, CH₃C=O), 2.01 (s, 3H, $CH_3C=O$; ¹³C-NMR (75 MHz, CDCl₃): δ 171.2 (C=O), 170.9 (C=O), 170.0 (C=O), 154.6 (OC=O), 138.8, 138.6, 138.3, 138.2, 133.2, 130.7, 129.8, 129.0, 128.95, 128.9, 128.6, 128.5, 128.4, 128.3, 101.4 (C-1'), 95.9 (CCl₃), 88.1 (C-1), 87.1, 81.2, 80.1, 78.2, 76.3, 75.8, 75.4, 74.9, 72.7, 72.0, 69.2, 68.3, 62.4, 56.3, 21.5 (CH₃), 21.2 $(CH_3C=O)$, 21.12 $(CH_3C=O)$, 21.10 $(CH_3C=O)$; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₄₉H₅₄Cl₃NaNO₁₄S, 1040.2223; found, 1040.2437.

p-Tolyl 6-*O*-[3,4,6-Tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxyβ-D-glucopyranosyl]-3-*O*-benzoyl-4-*O*-benzyl-2-(2,2,2-trichloroethoxycarbonyl amino)-2-deoxy-thio-β-D-glucopyranoside 30:

TrocHN STo VHTroc

prepared from thioglucosaminyl donor 7 Disaccharide **30** was and thioglucosaminyl acceptor 15 by the general DMF-modulated glycosylation procedure for participating donors (Table S1, entry 14). Purification of **30** was performed by column chromatography (EtOAc/CH₂Cl₂/Hexane 1:3:2). For disaccharide **30**: $[\alpha]_D^{35}$ -5.3 (c 0.35, CHCl₃); R_f 0.36 (EtOAc/CH₂Cl₂/Hexane 1:3:2); ¹H NMR (300 MHz, CDCl₃): δ 8.01 (d, J = 7.5 Hz, 2H, ArH), 7.58 (t, J = 7.2 Hz, 1H, ArH), 7.46-7.40 (m, 4H, ArH), 7.17-7.00 (m, 7H, ArH), 6.06 (bs, 1H, carbamate-H), 5.59 (t, J = 8.7 Hz, 1H), 5.40 (s, 1H, carbamate-H), 5.28 (t, J = 9.6 Hz, 1H), 5.06 (t, J = 9.3 Hz, 1H), 4.79-4.65 (m, 4 H), 4.58-4.40 (m, 4H), 4.28 (dd, J = 12.3, 3.9 Hz, 1H), 4.15 (d, J = 12.3, 311.4 Hz, 1H), 4.00-3.91 (m, 2H), 3.74-3.66 (m, 5H), 2.32 (s, 3H, CH₃), 2.06 (s, 3H, CH₃C=O), 2.03 (s, 3H, CH₃C=O), 2.1 (s, 3H, CH₃C=O); ¹³C NMR (75 MHz, CDCl₃): δ 171.2 (C=O), 170.9 (C=O), 169.9 (C=O), 167.0 (C=O), 154.7 (C=O), 154.5 (C=O), 138.8, 137.4, 134.0, 133.6, 130.3, 129.3, 128.9, 128.76, 128.5, 128.4, 100.5 (C-1'), 95.8 (CCl₃), 95.7 (CCl₃), 87.6 (C-1), 79.0, 76.3, 75.0, 74.6, 72.5, 72.0, 69.1, 67.3, 62.3, 56.0, 55.4, 30.0, 21.5 (CH₃), 21.1 (CH₃C=O), 21.00 (CH₃C=O), 20.97 $(CH_3C=O)$; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for $C_{45}H_{48}Cl_6N_2NaO_{16}S$, 1137.0748; found, 1139.0757.

p-Tolyl 6-*O*-(2-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl)-2,3,4-tri-*O*-benzyl-thio-β-D-galactopyranoside 31:



Disaccharide **31** was prepared from 2-*O*-benzoyl thiogalactoside **8** and thiogalactoside **16** by general DMF-modulated glycosylation procedure (Table S1, entry 15). For disaccharide **31**: $[\alpha]_D^{35}$ +17.6 (*c* 0.75, CHCl₃); *R*_f 0.4 (EtOAc/Hexane 1:4); ¹H-NMR (400 MHz, CDCl₃): δ 7.97 (d, *J* = 7.6 Hz, 2H, Ar*H*), 7.51 (t, *J* = 7.2 Hz, 1H, Ar*H*), 7.40 (d, *J* = 8.0 Hz, 4H, Ar*H*), 7.36-7.21 (m, 25H, Ar*H*), 7.14 (bs, 5H, Ar*H*), 6.95 (d, *J* = 7.6 Hz, 2H, Ar*H*), 5.61 (t, *J* = 8.8 Hz, 1H), 4.98 (d, *J* = 11.6 Hz, 1H), 4.81 (d, *J* = 11.2 Hz, 1H), 4.65-4.60 (m, 5H), 4.53 (d, *J* = 7.6 Hz, 1H), 4.48-4.31 (m, 6H), 4.02-3.98 (m, 2H), 3.85 (s, 1H), 3.77 (t, *J* = 9.6 Hz, 1H), 3.72 (dd, *J* = 10.0, 4.8 Hz, 1H), 3.63-3.57 (m, 3H), 3.54-3.51 (m, 1H), 3.41 (t, *J* = 6.0 Hz, 1H), 3.32 (d, *J* = 9.2 Hz, 1H), 2.25 (s, 3H, *CH*₃); ¹³C-NMR (100 MHz, CDCl₃): δ 165.7 (*C*=O), 139.4, 138.91, 138.87, 138.67, 138.2, 138.0, 137.7, 133.6, 132.8, 130.5, 130.4, 130.2, 130.0, 129.9, 128.74, 128.71, 128.68, 128.46, 128.40, 128.3, 128.12, 128.09, 128.05, 127.94, 127.90, 127.6, 101.8 (C-1'), 88.3 (C-1), 84.5, 80.3, 77.3, 76.0, 75.0, 74.8, 74.03, 73.99, 73.3, 72.9, 72.6, 72.5, 72.1, 68.7, 67.4, 21.6 (*C*H₃). HRMS-ESI (*m*/*z*): [M + Na]⁺ calcd. for C₆₈H₆₈NaO₁₁S, 1115.4375; found, 1115.4376.

p-Tolyl 2-O-Benzoyl-3,4-di-O-benzyl-thio-α-D-mannopyranoside 33



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Acetal derivative s13bc (5.0 g, 13.4 mmol) suspended in toluene (67 mL) was treated with di-butyl tin oxide (Bu₂SnO, 5.0 g, 20.1 mmol) and the solution was heated to reflux with Dean-Stark trap for 4 h (at 145 °C). The volume of toluene was then reduced approximately by half and the reaction mixture was cooled to RT. After then, benzyl bromide (2.4 ml, 20.1 mmol), cesium fluoride (CsF, 3.0 g, 20.1 mmol), and CH₃CN (33 mL) were added and the resulting mixture was stirred at 70 °C for ca. 16 h. Upon the completion of alkylation, the reaction mixture was filtered (over celite), and concentrated for column chromatography purification (Elution EtOAc/Hexane 1/8 stepwise to 1/3) to afford s33a (4.72 g, 10.17 mmol). s33a (3.36 g, 7.2 mmol) was treated with BzCl (1.3 mL, 10.8 mmol), Et₃N (2 mL, 14.4 mmol) and DMAP (85 mg, 0.7 mmol) in CH₂Cl₂. Upon completion of the reaction, the mixture was diluted with satd. NaHCO₃, then the mixture was extracted with EtOAc (\times 2). The EtOAc solution was then washed with H₂O and brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: Ether/CH₂Cl₂/Hexane 0.08/1/4 stepwise to 0.1/1/4) to furnish fully protected mannoside derivative s33b (3.18 g, 5.6 mmol). The derivative s33b (0.755 g, 1.33 mmol) in BH₃.THF solution (1 M, 5.32 mL, 5.32 mmol) was treated with trimethylsilyl triflate (TMSOTf) (49 µL) at 0 °C. Upon completion of the reductive acetal cleavage, the mixture was neutralized with Et₃N, excess borane was quenched with addition of MeOH. The resulting mixture was then concentrated for column chromatography purification (Elution: EtOAc/Hexane 1:8 stepwise to 1:6) to afford thiomannoside acceptor **33** (0.71 g, 1.24 mmol). For monosaccharide **33**: $[\alpha]_D^{35}$ +40.8 $(c \ 0.34, \text{CHCl}_3); R_f \ 0.34 \text{ (EtOAc/Hexane 1/2); }^1\text{H-NMR} (300 \text{ MHz}, \text{CDCl}_3): \delta 8.06 \text{ (d,})$ J = 6.9 Hz, 2 H), 7.56 (t, J = 7.5 Hz, 1 H), 7.46-7.41 (m, 2 H), 7.37-7.25 (m, 12 H), 7.09 (d, J = 8.1 Hz, 2 H), 5.85-5.83 (m, 1 H), 5.50 (d, J = 1.5 Hz, 1 H), 4.94 (d, J = 10.8 Hz, 1 H), 4.79 (d, J = 11.4 Hz, 1 H), 4.67 (d, J = 11.1 Hz, 1 H), 4.60 (d, J = 11.4Hz, 1 H), 4.27-4.24 (m, 1 H), 4.11-4.01 (m, 2 H), 3.85 (s, 2 H), 2.30 (s, 3 H), 1.97 (s, 1 H); 13 C-NMR (75 MHz, CDCl₃): δ 166.0, 138.7, 138.5, 138.0, 133.8, 133.2, 130.4,

130.3, 130.1, 129.8, 128.93, 128.85, 128.8, 128.6, 128.5, 128.3, 128.2, 87.1, 78.8, 75.7, 74.5, 73.3, 72.1, 71.1, 62.4, 21.6; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₃₄H₃₄NaO₆S, 593.1968; found, 593.1977.

Methyl2-O-Benzoyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-O-benzyl- α -D-mannopyranoside 35



Trisaccharide 35 was prepared from one pot disarmed-armed glycosylation method. In the first step, thiomannoside donor 2 (200 mg, 0.303 mmol) and DMF (23.5 µL, 0.303 mmol) were treated with NIS (74.7 mg, 0.329 mmol) and TMSOTf (83 μ L, 0.455 mmol) in dried CH₂Cl₂ at -20 °C. Upon the completion of the pre-activation (ca 0.5 h), thiomannoside acceptor 12 (141 mg, 0.253 mmol) was added to the mixture, which was stirred at -20 °C for ca. 7.5 h. Progress of the reaction was monitored by TLC ($R_{\rm f}$ of disaccharide product = 0.4, developed with 1/1/4 v/v Et₂O/CH₂Cl₂/Hexane). Upon the completion of glycosylation (ca 7.5 h), methyl mannoside 32 (117 mg, 0.253 mmol), DMF (55 µL, 0.708 mmol), NIS (68.8 mg, 0.303 mmol) and TMSOTf (82 µL, 0.455 mmol) was added to the mixture and the reaction temperature was stirred raised at 10 °C for ca. 14.5 h. Followed by standard workup (as described in general modulated glycosylation procedure) and column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 1/2/12 stepwise to 1/1/8), target trisaccharide 35 was obtained as white glassy solid (126.7 mg, 35%). For **35**: $[\alpha]_D^{35}$ +10.0 (*c* 0.32, CHCl₃); *R*_f 0.47 (EtOAc/CH₂Cl₂/Hexane - 30 -

1/1/4); ¹H-NMR (300 MHz, CDCl₃): δ 8.09 (d, J = 7.8 Hz, 2 H), 7.55 (t, J = 7.5 Hz, 1 H), 7.39-7.10 (m, 49 H), 7.05 (t, J = 8.1 Hz, 1 H), 5.77 (s, 1 H), 5.25 (s, 1 H), 5.12 (s, 1 H), 4.88-4.82 (m, 4 H), 4.77 (d, J = 11.1 Hz, 1 H), 4.70-4.45 (m, 14 H), 4.37 (d, J = 12.0 Hz, 1 H), 4.12-4.06 (m, 3 H), 4.01-3.93 (m, 4 H), 3.87-3.69 (m, 9 H), 3.60 (d, J = 1 H), 3.23 (s, 3 H); ¹³C NMR (75 MHz, CDCl₃): δ 165.9, 139.0, 138.9, 138.8, 138.5, 133.5, 130.5, 130.4, 128.84, 128.82, 128.76, 128.7, 128.6, 128.4, 128.3, 128.14, 128.08, 128.0, 127.94, 127.86, 101.1, 100.2, 99.9, 80.1, 79.7, 78.5, 75.7, 75.52, 75.45, 75.3, 75.2, 74.8, 73.8, 73.7, 72.7, 72.6, 72.5, 72.1, 72.0, 70.0, 69.8, 69.5, 55.1; HRMS-ESI (m/z): [M + Na]⁺ calcd. for C₈₉H₉₂NaO₁₇, 1455.6227; found, 1455.6222.

 $\label{eq:metric} Methyl 2-O-Benzoyl-3,4,6-tri-O-benzyl-\alpha-D-mannopyranosyl-(1\to6)-2-O-benzyl-3,4-di-O-benzyl-\alpha-D-mannopyranosyl-(1\to2)-3,4,6-tri-O-benzyl-\alpha-D-mannopyranoside 36$



Trisaccharide **36** was prepared from one pot disarmed-armed glycosylation method. In the first step, thiomannoside donor **2** (200 mg, 0.303 mmol) and DMF (23.5 μ L, 0.303 mmol) were treated with NIS (74.7 mg, 0.329 mmol) and TMSOTf (83 μ L, 0.455 mmol) in dried CH₂Cl₂ at -20 °C. Upon the completion of donor activation (*ca* 0.5 h), thiomannoside acceptor **33** (141 mg, 0.253 mmol) was added to the mixture, which was stirred at 0 °C for *ca*. 4.5 h. Progress of the reaction was monitored by TLC examination (*R*_f of disaccharide product = 0.55, developed with 1/1/3 v/v Et₂O/CH₂Cl₂/Hexane). Upon the completion of the glycosylation (*ca* 4.5 h), the reaction temperature was decreased to -20 °C, methyl mannoside acceptor **32**

(105.8 mg, 0.228 mmol), NIS (68.8 mg, 0.303 mmol) and TMSOTf (68.6 µL, 0.380 mmol) were added to the mixture and the mixture was stirred at -20 °C for *ca*. 2 h. Followed by standard workup (as described in general modulated glycosylation procedure) and column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 1/2/10 stepwise to 1/2/7), trisaccharide **36** was obtained as a white glassy solid (181.2) mg, 55%). For trisaccharide **36**: $[\alpha]_D^{35}$ +21.1 (*c* 1.05, CHCl₃); R_f 0.30 (EtOAc/Hexane 1/3); ¹H-NMR (400 MHz, CDCl₃): δ 8.12 (dd, J = 7.2, 1.6 Hz, 2 H), 8.08 (d, J = 7.6 Hz, 2 H), 7.55-7.46 (m, 5 H), 7.36-7.17 (m, 36 H), 7.15-7.06 (m, 7 H), 5.79 (s, 2 H), 5.24 (s, 1 H), 5.15 (s, 1 H), 4.88-4.82 (m, 4 H), 4.79-4.75 (m, 3 H), 4.70-4.59 (m, 5 H), 4.52-4.42 (m, 6 H), 4.14-4.09 (m, 4 H), 4.03-3.98 (m, 2 H), 3.93-3.86 (m, 3 H), 3.83-3.69 (m, 7 H), 3.64 (d, J = 10.4 Hz, 1 H), 3.33 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃): *δ*165.97, 165.95, 138.94, 138.88, 138.6, 138.5, 138.2, 133.6, 133.5, 130.6, 130.44, 130.37, 130.3, 129.0, 128.83, 128.79, 128.77, 128.75, 128.70, 128.69, 128.6, 128.5, 128.4, 128.3, 128.1, 128.03, 128.02, 127.97, 127.9, 100.3, 99.7, 98.5, 80.4, 78.9, 78.6, 75.7, 75.6, 75.5, 74.9, 74.61, 74.59, 74.2, 73.82, 73.80, 72.6, 72.3, 72.16, 72.15, 71.8, 69.6, 69.4, 69.3, 69.1, 66.7, 55.2; HRMS MALDI-TOF (*m/z*): [M + Na]⁺ calcd. for C₈₉H₉₀NaO₁₈, 1469.6019; found, 1469.5842.



Trisaccharide 37 was prepared from one pot disarmed-armed glycosylation

method. In first step, thioglucoside 6 (200 mg, 0.319 mmol) and DMF (30 µL, 0.389 mmol) were treated with NIS (72 mg, 0.319 mmol) and TMSOTf (86 µL, 0.479 mmol) in dried CH₂Cl₂ at -10 °C. Upon the completion of pre-activation, thioglucoside acceptor 11b was added to the mixture and the reaction temperature was raised to 0 °C. The mixture was stirred at 0 °C for *ca* 6 h and the reaction was monitored by TLC examination (R_f of disaccharide = 0.27, developed by 1/4 v/v EtOAc/Hexane \times 2). Upon the completion of the first glycosylation, DMF (20 µL, 0.319 mmol), NIS (72 mg, 0.319 mmol) and TNSOTf (85 µL, 0.479 mmol) were added to the reaction mixture to react with the disaccharide for ca. 1 h at 0 °C. Upon the completion of the disaccharide activation, methyl glycoside acceptor 34 (151 mg, 0.256 mmol) was added to the mixture and the reaction temperature was raised to 10 °C. The reaction mixture was stirred at 10 °C for ca. 8 h. After standard workup (as described in general modulated glycosylation procedure) and column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 1/22), target trisaccharide 37 was obtained as a light yellow glassy substance (255 mg, 52%). For trisaccharide **37**: $\left[\alpha\right]_{D}^{35}$ +35.7 (c 2.1, CHCl₃); $R_f 0.23$ (EtOAc/Hexane: 1/4 × 2); ¹H NMR (400 MHz,CDCl₃): δ 7.86 (d, J =7.2 Hz, 2 H), 7.82 (d, J = 7.2 Hz, 2 H, ArH), 7.74 (t, J = 8.4 Hz, 4 H, ArH), 7.41-7.34 (m, 3 H), 7.31-7.14 (m, 22 H, ArH), 7.09-7.02 (m, 10 H), 6.91-6.86 (m, 4 H, ArH), 5.63 (t, J = 9.6 Hz, 1 H), 5.52 (t, J = 9.6 Hz, 1 H), 5.35-5.26 (m, 2 H), 4.80-4.78 (m, 2 H), 4.61 (t, J = 12.8 Hz, 2 H), 4.51-4.48 (m, 3 H), 4.44-4.38 (m, 3 H), 4.29-4.17 (m, 4 H), 4.09-3.95 (m, 3 H), 3.83-3.69 (m, 3 H), 3.64-3.48 (m, 5 H), 3.35-3.27 (m, 2 H), 3.17 (s, 3 H), 2.00 (s, 3 H, CH₃); ¹³C NMR (100 MHz, CDCl₃): δ 171.0, 166.0, 165.6, 165.4, 139.2, 138.62, 138.57, 137.4, 137.2, 133.7, 133.6, 133.3, 130.07, 130.05, 130.0, 129.8, 129.6, 128.84, 128.78, 128.70, 128.67, 128.6, 128.5, 128.3, 128.2, 127.83, 127.77, 101.5, 101.1, 98.4, 82.2, 80.1, 77.6, 76.4, 76.0, 75.9, 75.7, 75.5, 75.4, 75.2, 75.03, 74.96, 73.69, 73.65, 72.5, 72.2, 69.8, 68.3, 68.2, 63.0, 55.6, 21.3; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₈₄H₈₂NaO₂₁, 1449.5241; found, 1449.5242.



p-Tolyl 2-O-Benzoyl 3,4,6-tri-O-methyl-thio-β-D-glucopyranoside 38

Per-O-acetyl glucosyl acetate s38a^[s2] (31.1g, 79.7mmol) in CH₂Cl₂ (20 mL) was cooled to 0 °C and treated with 33% HBr in AcOH (31.0 mL) under N2. After 30 min, the reaction mixture was brought to RT. Upon completion of the bromination, the reaction mixture was extracted with EtOAc, and the EtOAc solution was washed with H₂O, 8% NaOH_(aq), NaCl, dried (over MgSO₄), filtered, and concentrated, and dried under vacuo to give crude bromide derivative. The bromide derivative dissolved in CH₃CN (75 mL) was treated with TBAB (4.9 g, 15.2 mmol), 2,6-lutidine (17.6 mL, 152 mmol), and MeOH (9.2 mL). The reaction mixture was stirred at RT under N₂ overnight. Upon completion of the orthoester formation, the solvent was reduced by rotary evaporator. The residue was dissolved by EtOAc, which was washed with satd. NaHCO₃, H₂O, brine, followed by dried over MgSO₄, filtered, and concentrated to give per-O-acetyl glucosyl orthoester s38b (32.5 g). s38b (32.5 g) in MeOH/CH₂Cl₂ mixture (120 mL, MeOH/CH₂Cl₂ 2/1) was treated with Na(s) (120 mg) at RT. Upon completion of deacetylation, the reaction mixture was concentrated and dried under vacuo for 4 h. The crude deacetyl compound (14.3 g, 60.5 mmol) was then dissolved in dried DMF (120 mL) and the solution was cooled to 0 °C under N₂. The solution was treated with (7.3 g, 181.5 mmol) (60% in mineral oil), followed by iodomethane (MeI) (11.3 mL, 182 mmol). Upon completion of methylation, H₂O was added to quench the reaction. The reaction mixture was diluted with EtOAc, and the resulting solution was washed with H₂O, brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (EtOAc/Hexane 3/7) to furnish per-O-methyl

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glucosyl orthoester s38c (7.20 g). s38c (7.20 g, 25.9 mmol) in dried CH_2Cl_2 (52 mL) was treated with p-thiocresol (4.8 g, 38.9 mmol) at 0°C, followed by treatment with BF₃.OEt₂ (4.93 mL, 39 mmol). Upon completion of the reaction, the reaction mixture was diluted with 8% NaOH_(aq), and the reaction solution was washed with satd. NaHCO₃, EtOAc, brine, dried over MgSO₄, filtered, and concentrated for column chromatography purification (Elution: EtOAc/CH₂Cl₂/Hexane 1/4/9 stepwise to 1/3/7) to give 2-O-acetyl-3,4,6-tri-O-methyl thioglucoside s33d (7.1 g, 74%) as yellow syrup. s33d (6.8 g, 18.4 mmol) in CH₂Cl₂/MeOH (120 mL) was treated with Na(s) (150 mg). Upon completion of deacetylation, the reaction was neutralized with IR-120 (H⁺), filtered, concentrated, and column chromatography purification (EtOAc/Hexane 1/3 stepwise to 1/2) to furnish (4.6 g, 76%). The preceding derivative in CH₂Cl₂ was treated with BzCl (2.4 mL, 20.1 mmol), Et₃N (3.7 mL, 26.8 mmol) and DMAP (170 mg, 1.4 mmol) under N₂. Upon completion of benzoylation, the reaction solution was diluted with satd. NaHCO₃, followed by extraction with EtOAc. The EtOAc solution was washed with H₂O, brine, dried over MgSO₄, filtered, and recrystalized to obtain target 2-O-benzoyl-3,4,6-tri-O-methyl-β-D-thioglucoside **38** (4.02g, 69%). For thioglucoside **38**: $[\alpha]_D^{35}$ +33.8 (c 0.5, CHCl₃); R_f 0.375 (EtOAc/Hexane 1/2); ¹H NMR (300 MHz, CDCl₃): δ 8.09 (d, J = 7.5 Hz, 2H, ArH), 7.56 (t, J = 7.2 Hz, 1H, Ar*H*), 7.44 (t, *J* = 7.5 Hz, 1H, Ar*H*), 7.35 (d, *J* = 7.8 Hz, 2H, Ar*H*), 7.05 (d, *J* = 7.8 Hz, 2H, ArH), 5.15 (t, J = 9.6 Hz, 1H), 4.70 (d, J = 10.2 Hz, 1H), 3.70-3.60 (m, 2H), 3.54-3.41 (m, 11H), 3.35-3.29 (m, 1H), 2.28 (s, 3H, CH₃); ¹³C-NMR (75 MHz, $CDCl_3$: δ 165.4 (C=O), 138.1, 133.5, 133.0, 130.2, 130.0, 129.8, 129.7, 128.7, 87.0 (C-1), 86.6, 79.5, 79.3, 72.7, 71.6, 60.8, 60.7, 59.7, 21.3; HRMS-ESI (m/z): $[M + Na]^+$ calcd. for C₂₃H₂₈NaO₆S, 455.1499; found, 455.1493.
6-Chlorohexyl 2-*O*-benzoyl-3,4,6-tri-*O*-methyl β-D-glucopyranoside 42:



6-Chlorohexyl glucoside 42 was prepared from glycosylation of chlorohexanol **39** with thioglucoside **38** under the general DMF-modulated glycosylation procedure. 42 purified column chromatography The glucoside was by (Elution: EtOAc/CH₂Cl₂/Hexane 1/2/15). For 42: $[\alpha]_D^{35}$ +8.2 (c 0.41, CHCl₃); R_f 0.5 (EtOAc/CH₂Cl₂/Hexane 1:2:5); ¹H NMR (300 MHz, CDCl₃): δ 8.08 (d, J = 7.2 Hz, 2H, ArH), 7.59 (t, J = 7.2 Hz, 1H, ArH), 7.46 (t, J = 7.8 Hz, 2H. ArH), 5.13 (dd, J = 9.3,7.8 Hz, 1H, H-2), 4.46 (d, J = 8.1 Hz, 1H, H-1), 3.91–3.86 (m, 1H), 3.71–3.67 (m, 1 H), 3.64 (d, J = 4.5 Hz, 1 H), 3.61-3.56 (m, 4 H), 3.50 (s, 3 H), 3.47-3.38 (m, 6 H), 3.35-3.29 (m, 3 H), 1.50–1.43 (m, 4H), 1.26–1.16 (m, 4H); ¹³C-NMR (75 MHz, CDCl₃): δ 165.6 (C=O), 133.5, 130.5, 130.1, 128.8, 101.5 (C-1), 85.2, 79.7, 75.4, 74.0, 71.7, 69.9, 60.9, 60.8, 59.8, 45.3, 32.8, 29.6, 26.8, 25.5; HRMS (m/z): $[M + Na]^+$ calcd. for C₂₂H₃₃ClNaO₇, 467.1807; found, 467.1801.

Determination of RRV for thioglycosides:^[s2]

The RRV calculation formula:
$$\frac{K_1}{K_2} = \frac{\ln([A_1, I]/[A_1, 0])}{\ln([A_2, I]/[A_2, 0])}$$

0.3 g of particle molecular sieve was flame-dried for 5 times. Tested thioglycoside A_2 (0.02 mmol) and reference thioglycoside A_1 (0.02 mmol) were dissolved in 2 mL CH₂Cl₂ and stirred in RT for 10 min, 100 µL of reaction solution was removed for determination of time-zero absorbance [$A_{1,0}$] and [$A_{2,0}$]. The remainder of the reaction solution was treated with 4 µL MeOH (0.1 mmol) and particle molecular sieve stirred in 0 °C. 0.5 M NIS solution (40 µL, 0.02 mmol) and 0.1 M TMSOTf solution (20 µL, 0.002 mmol) were added to the reaction solution and stirred for 2 h. Upon the reaction completed, the reaction mixture was treated with few drops of 10% NaHCO₃/Na₂SO₃

solution. After stirring for 15 min, the reaction temperature was raised to RT. The mixture was dried over MgSO₄, filtered and concentrated. The crude residue was dissolved in 2 mL CH₂Cl₂ for determination of final absorbance $[A_{1,t}]$ and $[A_{2,t}]$ at 2 h reaction. As an example, the HPLC traces from thioglucoside **11b** and reference compound **1** before and after the reaction were given (**Figure s1**). The RRV of tested thioglycoside was calculated by substitution of the absorbance values (before and after the reaction) and the RRV value of the reference compound from literature into the equation above (Table S4).



Figure S1. HPLC traces of thioglucoside 11b and 1 before and after the glycosylation in RRV determination

reference thioglycosides A 1	thioglycosides for RRV measurement A 2	reference thioglycosides A₁	thioglycosides for RRV measurement A 2
BnO BnO OBz	6, 10, 11a,17	BnO OBn BnO STol OBn	16
BnO BnO OBn OBn	1, 11b, 12	HO Levo NHTroc	15
BnO BnO BnO STol	2, 13a, 13b, 14	2	33

Table S4. Reference thioglycosides and thioglycosides for RRV measurement

Section B: NMR spectra:

¹H spectrum of *p*-tolyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-thio-α-D-mannopyranoside **2**





¹³C spectrum of *p*-tolyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-thio-α-D-mannopyranoside **2**

¹H spectrum of 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-1,2:3,4-

di-O-isopropylidene- α -D-galactopyranose 4



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¹³C spectrum of 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-1,2:3,4-

di-O-isopropylidene- α -D-galactopyranose 4



¹H spectrum of 6-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-1,2:3,4-

di-O-isopropylidene- α -D-galactopyranose 5



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 $^{13}C \ spectrum \ of \ 6-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-\alpha-D-mannopyranosyl)-1,2:3,4-interval \ (2-O-benzoyl-3,4,6-tri-O-benzyl-\alpha-D-mannopyranosyl)-1,2:3,4-interval \ (2-O-benzoyl-3,4,6-tri-O-benzyl-3,6-tri-O-benzyl-3,4,6-tri-O-benzyl-3,4,6-t$







¹H spectrum of *p*-tolyl 6-*O*-acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-thio-β-D-gluco-

¹³C spectrum of *p*-tolyl 6-O-acetyl-2,3-di-O-benzoyl-4-O-benzyl thio-β-D-gluco-





¹H spectrum of *p*-tolyl 2,4,6-tri-*O*-benzyl-thio- α -D-mannopyranoside **13a**



¹³C spectrum of *p*-tolyl 2,4,6-tri-*O*-benzyl-thio- α -D-mannopyranoside **13a**



¹H spectrum of *p*-tolyl 6-*O*-benzoyl-2,4-di-*O*-benzyl-thio-α-D-mannopyranoside **13b**







¹H spectrum of *p*-tolyl 2,3,4-tri-*O*-benzyl-thio- α -D-mannopyranoside 14



¹³C spectrum of *p*-tolyl 2,3,4-tri-*O*-benzyl-thio- α -D-mannopyranoside 14

¹H spectrum of *p*-tolyl 3-*O*-benzoyl-4-*O*-benzyl-2-(2,2,2-trichloroethoxycarbonyl

amino)-2-deoxy-thio- β -D-glucopyranoside 15



¹³C spectrum of *p*-tolyl 3-O-benzoyl-4-O-benzyl-2-(2,2,2-trichloroethoxycarbonyl

amino)-2-deoxy-thio- β -D-glucopyranoside 15





¹H spectrum of 4-(*t*-butyl)-2-methylphenyl 2,3-*O*-isopropylidene-thio-α-L-rhamno





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¹H spectrum of methyl 4-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-

2,3,6-tri-O-benzyl-α-D-glucopyranoside 18



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¹³C spectrum of methyl 4-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl)-

2,3,6-tri-O-benzyl-α-D-glucopyranoside 18



¹H spectrum of *p*-tolyl 4-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl- β -D-glucopyranosyl)-

2,3,6-tri-O-benzyl-thio- β -D-glucopyranoside 19



¹³C spectrum of *p*-tolyl 4-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl)-



2,3,6-tri-O-benzyl-thio-β-D-glucopyranoside 19

¹H spectrum of *p*-tolyl 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-



2,3,4-tri-*O*-benzyl-thio-β-D-glucopyranoside 20

¹³C spectrum of *p*-tolyl 6-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl)-



2,3,4-tri-O-benzyl-thio-β-D-glucopyranoside 20

¹H spectrum of *p*-tolyl 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-



2,3,4-tri-O-benzyl-thio-β-D-galactopyranoside 21

¹³C spectrum of *p*-tolyl 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl)-





¹H spectrum of 4-(*t*-butyl)-2-methylphenyl 4-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-

 β -D-glucopyranosyl)-2,3-O-isopropylidene-thio- α -L-rhamnopyranoside 22



¹³C spectrum of 4-*t*-butyl-2-methylphenyl 4-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-



 β -D-glucopyranosyl)-2,3-O-isopropylidene-thio- α -L-rhamnopyranoside 22

¹H spectrum of *p*-tolyl 6-O-(6-O-acetyl-2,3-di-O-benzoyl-4-O-benzyl-β-D-gluco





13 C spectrum of *p*-tolyl 6-*O*-(6-*O*-acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl- β -D-gluco

pyranosyl)-2,3,4-tri-O-benzyl-thio-β-D-glucopyranoside 23



¹H spectrum of *p*-tolyl 6-*O*-(6-*O*-acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-β-D-gluco

pyranosyl)-2,3-di-O-benzoyl-4-O-benzyl-thio-β-D-glucopyranoside 24



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¹³C spectrum of *p*-tolyl 6-*O*-(6-*O*-acetyl-2,3-di-*O*-benzyl-4-*O*-benzyl-β-D-

glucopyranosyl)-2,3-di-O-benzoyl-4-O-benzyl-thio-β-D-glucopyranoside 24



¹H spectrum of *p*-tolyl 2-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-

3,4,6-tri-O-benzyl-thio-α-D-mannopyranoside 25


¹³C spectrum of *p*-tolyl 2-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-





¹H spectrum of *p*-tolyl 3-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-





¹³C spectrum of *p*-tolyl 3-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-

2,4,6-tri-O-benzyl-thio- α -D-mannopyranoside **26**



¹H spectrum of *p*-tolyl 3-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-

6-O-benzoyl-2,4-di-O-benzyl-thio-α-D-mannopyranoside 27



¹³C spectrum of *p*-tolyl 3-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-

-6-*O*-benzoyl-2,4-di-*O*-benzyl-thio-α-D-mannopyranoside **27**



¹H spectrum of *p*-tolyl 6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-

2,3,4-tri-O-benzyl-thio-α-D-mannopyranoside 28



¹³C spectrum of *p*-tolyl 6-O-(2-O-benzoyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl)-

2,3,4-tri-O-benzyl-thio-α-D-mannopyranoside 28



¹H spectrum of *p*-tolyl 6-*O*-[3,4,6-tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonyl

amino)-2-deoxy-β-D-glucopyranosyl]-2,3,4-tri-O-benzyl-thio-β-D-glucopyranoside 29



¹³C spectrum of *p*-tolyl 6-*O*-[3,4,6-tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonyl

amino)-2-deoxy-β-D-glucopyranosyl]-2,3,4-tri-O-benzyl-thio-β-D-glucopyranoside 29



¹H spectrum of *p*-tolyl 6-*O*-[3,4,6-tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonyl amino)-2-deoxy- β -D-glucopyranosyl]-3-*O*-benzoyl-4-*O*-benzyl-2-(2,2,2-trichloroetho xycarbonylamino)-2-deoxy-thio- β -D-glucopyranoside **30**



¹³C spectrum of *p*-tolyl 6-*O*-[3,4,6-tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonyl amino)-2-deoxy-β-D-glucopyranosyl]-3-*O*-benzoyl-4-*O*-benzyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-thio-β-D-glucopyranoside **30**



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¹H spectrum of *p*-tolyl 6-*O*-(2-benzoyl-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl)-



2,3,4-tri-O-benzyl-thio- β -D-galactopyranoside **31**

¹³C spectrum of *p*-tolyl 6-*O*-(2-benzoyl-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl)-

2,3,4-tri-O-benzyl-thio-β-D-galactopyranoside 31



¹H spectrum of *p*-tolyl 2-*O*-benzoyl-3,4-di-*O*-benzyl-thio-α-D-mannopyranoside **33**



 13 C spectrum of *p*-tolyl 2-*O*-benzoyl-3,4-di-*O*-benzyl-thio- α -D-mannopyranoside **33**



¹H spectrum of methyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl- α -D-mannopyrano side **35**



¹³C spectrum of methyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl-α-D-mannopyrano side **35**



¹H spectrum of methyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2-*O*-benzoyl-3,4-di-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl- α -D-m



¹³C spectrum of methyl 2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl-(1→6)-2-*O*-benzoyl-3,4-di-*O*-benzyl-α-D-mannopyranosyl-(1→2)-3,4,6-tri-*O*-benzyl-α-D-m annopyranoside **36**



¹H spectrum of methyl 6-*O*-acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-β-D-glucopyranosyl- $(1\rightarrow 6)$ -2,3-di-*O*-benzoyl-4-benzyl-β-D-glucopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-*O*-benzyl-β-D-glucopyranoside **37**



¹³C spectrum of methyl 6-*O*-acetyl-2,3-di-*O*-benzoyl-4-*O*-benzyl-β-D-gluco

pyranosyl- $(1\rightarrow 6)$ -2,3-di-O-benzoyl-4-benzyl- β -D-glucopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-





¹H spectrum of *p*-tolyl 2-*O*-benzoyl-3,4,6-tri-*O*-methyl-thio-β-D-glucopyranoside **38**



¹³C spectrum of *p*-tolyl 2-*O*-benzoyl-3,4,6-tri-*O*-methyl-thio-β-D-glucopyranoside **38**





¹H spectrum of 6-chlorohexyl 2-O-benzoyl-3,4,6-tri-O-methyl-β-D-glucopyranoside



 $^{13}C\ spectrum\ of\ 6-chlorohexyl\ 2-O-benzoyl-3,4,6-tri-O-methyl-\beta-D-glucopyranoside$

¹H NMR spectrum of crude pre-activation mixture of **38** (refer to Fig 2 and scheme 3

in main context)



¹³C NMR spectrum of crude pre-activation mixture of **38** (refer to Fig 2 and scheme 3

in main context)



¹³C non-decoupling NMR spectrum of crude pre-activation mixture of **38** (refer to Fig

2 and scheme 3 in main context)



HSQC spectrum of crude pre-activation mixture of $\mathbf{38}$ (refer to Fig 2 and scheme 3 in

main context)



HMBC spectrum of crude pre-activation mixture of 38 (see Fig S1 in following page

for selected expansion)





Figure S2. Selected 1H NMR spectrum of pre-activation mixture of 38



Figure S3. HMBC spectrum of β -glucosyl imidinium triflate 40 β in reaction mixture



Figure S4. Proposed mechanism for modulated glycosylations



Figure S5. (a) Selected ¹H NMR spectrum of pre-activation of **38**. (b) Selected ¹H NMR spectrum of the crude reaction after adding acceptor **39**.



Crude ¹H spectrum of the reaction mixture after addition of 39 (formation of 42)

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