

Supplementary Information

A Convenient Synthesis of Anthranilic Acids by Pd-Catalyzed Direct Intermolecular *ortho*-C-H Amidation of Benzoic Acids

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1. General Information

General. All the amidation reactions were performed in a 10 mL Schlenk tube under an atmosphere of nitrogen. Thin layer chromatography was performed on silica gel plates. Flash column chromatography was performed on 230-400 mesh silica gel (NA Chemical). GC-MS analyses were performed on a 6890N-GC (Agilent Technology) with 5973Network-MS (Agilent Technology). ^1H , ^{13}C , DEPT 135° NMR analyses were performed on a Bruker (400 MHz) spectrometer. Chemical shifts (δ) were given in ppm, and the signals were referenced with the solvent residual peak(s). NMR yields and conversions were determined with dibromomethane (0.1 mmol) as the internal standard, with reference to a singlet signal (2H) at δ_{H} 4.9 ppm (in CDCl_3) δ_{H} 5.3 ppm (in d_6 -acetone). IR spectra were obtained by a Nicolet-380 FT-IR spectrometer. Melting points were recorded on a BÜCHIB- 545 instrument and were uncorrected. High resolution mass spectra were obtained using a VG MICROMASS Fison VG platform and with an electrospray ionization mode.

Materials. Palladium(II) acetate, benzoic acid, 3,4-dimethylbenzoic acid, 3-methoxybenzoic acid, 3-chlorobenzoic acid, 3-bromobenzoic acid, 3-trifluoromethylbenzoic acid, 2-benzylbenzoic acid, 2,4-dimethylbenzoic acid, 2-methyl-4-fluorobenzoic acid, 2-methoxy-4-methylbenzoic acid, 1,4-benzodioxane-6-carboxylic acid, 2-napthoic acid, 1-napthoic acid, sodium hydroxide, lithium hydroxide monohydrate, potassium hydroxide, tetrabutylammonium hydroxide, hydroxylamine hydrochloride, ethyl chloroformate, triethylamine, *p*-nitrobenzenesulfonyl chloride, *p*-toluenesulfonyl chloride, 2-mesitylenesulfonyl chloride, pentafluorobenzoyl chloride, pivaloyl chloride, potassium acetate, potassium pivalate, potassium hydrogenphosphate dibasic, potassium carbonate, potassium hydrogencarbonate and all the solvents (except 1,4-dioxane) were obtained commercially, and used as received without purification. All the lithium benzoates were synthesized from commercially available benzoic acids (except **1f**) by reacting with lithium hydroxide in methanol. Ethyl *N*-carboxylate carbamates,¹ ethyl *N*-sulfonyloxycarbamates,¹ 2,6- d_2 -benzoic acid² and 3-phenylbenzoic acid³ (**1f**) were prepared according to the literature procedures. 1,4-Dioxane was sodium-dried and distilled before use.

2. Experimental Procedures

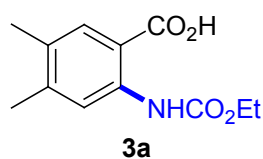
2.1 General Procedures for Preparation of Lithium Benzoates

Lithium benzoates (3 mmol) were prepared by treating benzoic acids with LiOH·H₂O (0.9 equiv) in MeOH (10 mL) at room temperature for 24 h. After removal of the solvent by rotary evaporation, the residue was rinsed with acetone (3 × 5 mL). The crude products were dried under vacuum for 24 h and were used directly without further purification.

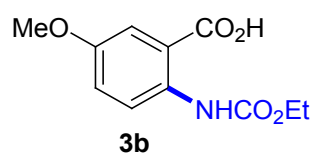
2.2 General Procedure for the Pd-Catalyzed C-H Amidation of Lithium Benzoates

A mixture of lithium benzoate (0.2 mmol), Pd(OAc)₂ (10 mol%) and KOAc (1 equiv) was dissolved in 1,4-dioxane (0.5 mL) in a 10 mL Schlenk tube, and ethyl *N*-mesitylsulfonyloxycarbamate (1.5 equiv) dissolved in dioxane (1 mL) was added dropwise by a syringe pump at the rate of 0.3 equiv / h. The reaction was stirred at 90 °C for 6 h under a N₂ atmosphere. After cooling to room temperature, the reaction mixture was diluted with EtOAc (ca. 4 mL), and the mixture was acidified with 2 M HCl (2 mL) solution. The aqueous layer was extracted with EtOAc (5 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and then filtered through a short plug of Celite, and the filtrate was evaporated to dryness by rotary evaporation. The residue was redissolved with dichloromethane (DCM). A minimum amount of MeOH may be added (when the reaction crude was sparingly-soluble in DCM). The dissolved mixture was then purified by flash column chromatography on silica gel by gradient elution with 10-50% EtOAc in hexanes (5% increment of EtOAc).

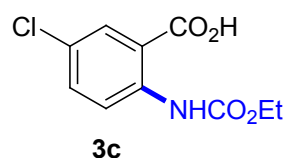
3. Physical Characterization Data



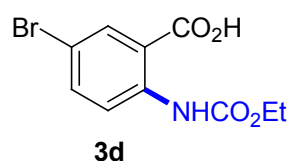
3a was isolated as a brown solid (30.0 mg, 63% yield). mp = 140-142 °C; ¹H NMR (CDCl₃, 400 MHz): δ_H 10.1 (s, 1H), 8.26 (s, 1H), 7.84 (s, 1H), 4.23 (q, *J* = 7.2 Hz, 2H), 2.32 (s, 3H), 2.23 (s, 3H), 1.34 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ_C 172.6 (C), 153.7 (C), 145.8 (C), 140.4 (C), 132.3 (CH), 130.2 (C), 119.9 (CH), 111.1 (C), 61.2 (CH₂), 20.6 (CH₃), 18.9 (CH₃), 14.5 (CH₃); IR (KBr, cm⁻¹): 3324, 2980, 1735, 1670, 1584, 1521, 1263, 1226, 1093, 1034; HRMS *m/z* (ESI): calculated for C₁₂H₁₅NO₄Na⁺: 260.0899, found: 260.0907.



3b was isolated as a pale yellow solid (33.5 mg, 70% yield). mp = 146-148 °C; ¹H NMR (*d*₆-acetone, 400 MHz): δ_H 10.4 (s, 1H), 8.36 (d, *J* = 9.2 Hz, 1H), 7.57 (d, *J* = 3.2 Hz, 1H), 7.20 (dd, *J* = 9.2 Hz, 2.8 Hz, 1H), 4.16 (q, *J* = 7.2 Hz, 2H), 3.81 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₆-acetone, 100 MHz): δ_C 170.1 (C), 154.9 (C), 154.2 (C), 136.8 (C), 121.9 (CH), 120.8 (CH), 116.4 (C), 115.9 (CH), 61.5 (CH₂), 56.0 (CH₃), 14.9 (CH₃); IR (KBr, cm⁻¹): 3348, 1738, 1536, 1280, 1250, 1210, 1091, 1063, 1042; HRMS *m/z* (ESI): calculated for C₁₁H₁₃NO₅Na⁺: 262.0685, found: 262.0685.

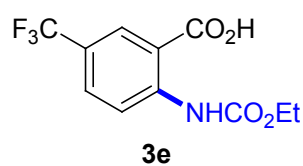


3c was isolated as a pale yellow solid (30.6 mg, 63% yield). mp = 146-148 °C; ¹H NMR (*d*₆-DMSO, 400 MHz): δ_H 12.9 (s, 1H), 8.19 (d, *J* = 8.8 Hz, 1H), 7.97 (d, *J* = 2.8 Hz, 1H), 7.38 (dd, *J* = 8.8 Hz, 2.8 Hz, 1H), 4.10 (q, *J* = 7.2 Hz, 2H), 1.22 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₆-DMSO, 100 MHz): δ_C 169.5 (C), 153.6 (C), 140.1 (C), 131.2 (CH), 131.0 (CH), 124.91 (C), 124.8 (C), 119.2 (CH), 60.7 (CH₂), 15.0 (CH₃); IR (KBr, cm⁻¹): 3217, 3201, 1706, 1664, 1428, 1282, 1224; HRMS *m/z* (ESI): calculated for C₁₀H₁₀NO₄ClNa⁺: 265.0118, found: 265.0119.

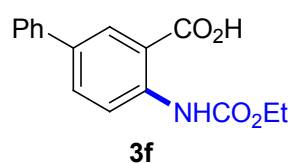


3d was isolated as a pale yellow solid (34.3 mg, 60% yield). mp = 146-148 °C; ¹H NMR (CDCl₃, 400 MHz): δ_H 10.2 (s, 1H), 8.42 (d, *J* = 9.2 Hz, 1H), 8.22 (d, *J* = 2 Hz, 1H), 7.67 (dd, *J* = 9.2 Hz, 2 Hz, 1H), 4.25 (q, *J* = 7.2 Hz, 2H), 1.34 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ_C 171.3 (C), 154.4 (C), 142.6 (C), 139.3 (CH), 135.1 (CH), 121.7 (CH), 115.7 (C), 114.8 (C), 62.6 (CH₂), 15.4 (CH₃); IR (KBr,

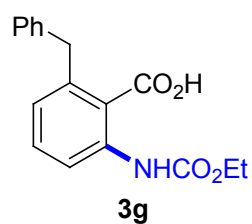
cm⁻¹): 3233, 1742, 1686, 1606, 1532, 1508, 1246, 1198, 1154, 1037; HRMS *m/z* (ESI): calculated for C₁₀H₁₀NO₄BrNa⁺: 309.9691, found: 309.9687.



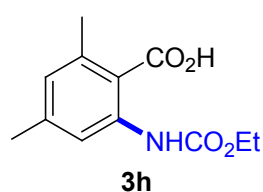
3e was isolated as a pale yellow solid (30.5 mg, 55% yield). mp = 187-188 °C; ¹H NMR (*d*₆-acetone, 400 MHz): δ_H 10.9 (s, 1H), 8.64 (d, *J* = 8.8 Hz, 1H), 8.33 (s, 1H), 7.90 (d, *J* = 7.6 Hz, 1H), 4.23 (q, *J* = 7.2 Hz, 2H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₆-acetone, 100 MHz): δ_C 169.7 (C), 154.0 (C), 146.3 (C), 131.6 (CH), 129.4 (CH), 129.1, 126.4, 123.8, 121.1 (q, *J*_{CF} = 269 Hz, CF₃), 124.0, 123.6, 123.3, 121.1 (q, *J*_{CF} = 33 Hz, C), 119.6 (CH), 116.2 (C), 62.2 (CH₂), 14.7 (CH₃); IR (KBr, cm⁻¹): 3332, 1732, 1689, 1592, 1542, 1409, 1249, 1121, 1087; HRMS *m/z* (ESI): calculated for C₁₁H₁₀NO₄F₃Na⁺: 322.0279, found: 322.265.



3f was isolated as a pale yellow solid (26.8 mg, 47% yield). mp = 187-188 °C; ¹H NMR (*d*₄-methanol, 400 MHz): δ_H 8.43 (d, *J* = 8.8, 1H), 8.29 (s, 1H), 7.79 (d, *J* = 8.8, 1H), 7.59 (d, *J* = 7.2 Hz, 2H), 7.43 (t, *J* = 7.6 Hz, 2H), 7.32 (t, *J* = 7.2 Hz, 1H), 4.22 (q, *J* = 7.2 Hz, 2H), 1.32 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₄-methanol, 100 MHz): δ_C 153.7 (C), 140.7 (C), 139.5 (C), 134.3 (C), 132.4 (C), 132.0 (CH), 129.2 (CH), 128.5 (CH), 126.9 (CH), 126.5 (C), 126.1 (CH), 118.6 (CH), 60.8 (CH₂), 13.3 (CH₃); IR (KBr, cm⁻¹): 3339, 3028, 2485, 1742, 1670, 1247; HRMS *m/z* (ESI): calculated for C₁₆H₁₅NNaO₄⁺: 308.0899, found: 308.0890.

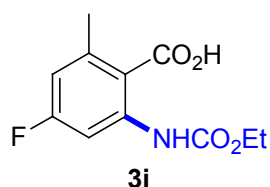


3g was isolated as a pale yellow solid (42.5 mg, 71% yield). mp = 117-118 °C; ¹H NMR (CDCl₃, 400 MHz): δ_H 8.78 (s, 1H), 8.06 (d, *J* = 6.8 Hz, 1H), 7.39 (t, *J* = 8.0 Hz, 1H), 7.26-7.10 (m, 5H), 6.92 (d, *J* = 7.6 Hz, 1H), 4.26-4.20 (m, 4H), 1.30 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ_C 154.0 (C), 142.1 (C), 140.4 (C), 139.2 (C), 138.7 (C), 132.2 (CH), 128.9 (CH), 128.3 (CH), 126.2 (CH), 126.1 (CH), 119.2 (CH), 61.5 (CH₂), 40.5 (CH₂), 14.4 (CH₃); IR (KBr, cm⁻¹): 3360, 3027, 1739, 1653, 1528, 1266, 1216, 1089; HRMS *m/z* (ESI): calculated for C₁₇H₁₇NO₄Na⁺: 322.1055, found: 322.1059.

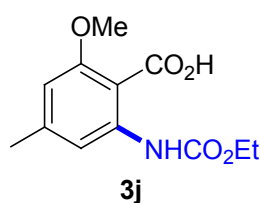


3h was isolated as a yellow solid (34.6 mg, 73% yield). mp = 145-146 °C; ¹H NMR (*d*₄-methanol, 400 MHz): δ_H 7.69 (s, 1H), 6.80 (s, 1H), 4.17 (q, *J* = 7.2 Hz, 2H), 2.44 (s, 3H), 2.30 (s, 3H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₄-methanol, 100 MHz): δ_C

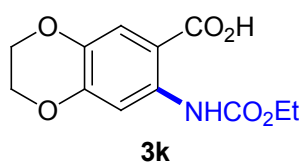
171.4 (C), 155.3 (C), 142.6 (C), 139.8 (C), 139.4 (C), 127.6 (CH), 119.6 (CH), 61.8 (CH₂), 22.1 (CH₃), 21.1 (CH₃), 14.4 (CH₃); IR (KBr, cm⁻¹): 3217, 2372, 1706, 1664, 1428, 1224; HRMS *m/z* (ESI): calculated for C₁₂H₁₅NO₄Na⁺: 260.0899, found: 260.0898.



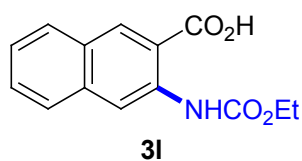
3i was isolated as a brown solid (22.2 mg, 46% yield). mp = 133-134 °C; ¹H NMR (CDCl₃, 400 MHz): δ_H 9.71 (s, 1H), 8.03 (dd, *J* = 11.2 Hz, 2 Hz, 1H), 6.66 (dd, *J* = 8.8 Hz, 2 Hz, 1H), 4.24 (q, *J* = 7.2 Hz, 2H), 1.33 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ_C 172.4 (C), 166.2, 163.7 (d, *J* = 250 Hz, CF), 153.4 (C), 144.5, 144.4 (d, *J* = 11 Hz, C), 143.5, 143.4 (d, *J* = 13 Hz, C), 113.1, 112.9 (d, *J* = 22 Hz, CH), 104.9, 104.7 (d, *J* = 28 Hz, CH), 61.5 (CH₂), 23.8 (CH₃), 14.4 (CH₃); IR (KBr, cm⁻¹): 3256, 1725, 1606, 1523, 1446, 1245, 1197; HRMS *m/z* (ESI): calculated for C₁₁H₁₂NO₄FNa⁺: 264.0648, found: 264.0659.



3j was isolated as a yellow solid (25.3 mg, 50% yield). mp = 102-103 °C; ¹H NMR (*d*₄-methanol, 400 MHz): δ_H 7.64 (s, 1H), 6.64 (s, 1H), 4.17 (q, *J* = 7.2 Hz, 2H), 3.90 (s, 3H), 2.34 (s, 3H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₄-methanol, 100 MHz): δ_C 169.9 (C), 159.9 (C), 154.9 (C), 145.6 (C), 141.5 (C), 114.0 (CH), 107.7 (CH), 106.6 (C), 61.9 (CH₂), 56.6 (CH₃), 21.9 (CH₃), 14.4 (CH₃); IR (KBr, cm⁻¹): 3199, 2990, 1735, 1707, 1583, 1442, 1366, 1254, 1223, 1049; HRMS *m/z* (ESI): calculated for C₁₂H₁₅NO₅Na⁺: 276.0848, found: 276.0843.

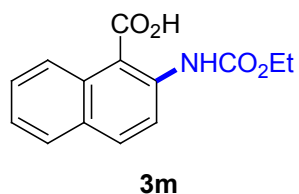


3k was isolated as a white solid (21.4 mg, 40% yield). mp = 210-211 °C; ¹H NMR (*d*₄-methanol, 400 MHz): δ_H 7.83 (s, 1H), 7.50 (s, 1H), 4.31-4.29 (m, 2H), 4.23-4.21 (m, 2H), 4.18 (q, *J* = 7.2 Hz, 2H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₆-DMSO, 100 MHz): δ_C 170.5 (C), 154.2 (C), 149.7 (C), 139.1 (C), 137.4 (C), 120.4 (CH), 109.8 (C), 107.9 (CH), 66.3 (CH₂), 65.2 (CH₂), 62.1 (CH₂), 15.8 (CH₃); IR (KBr, cm⁻¹): 1736, 1646, 1565, 1308, 1266, 1064; HRMS *m/z* (ESI): calculated for C₁₂H₁₃NO₆Na⁺: 290.0641, found: 290.0650.



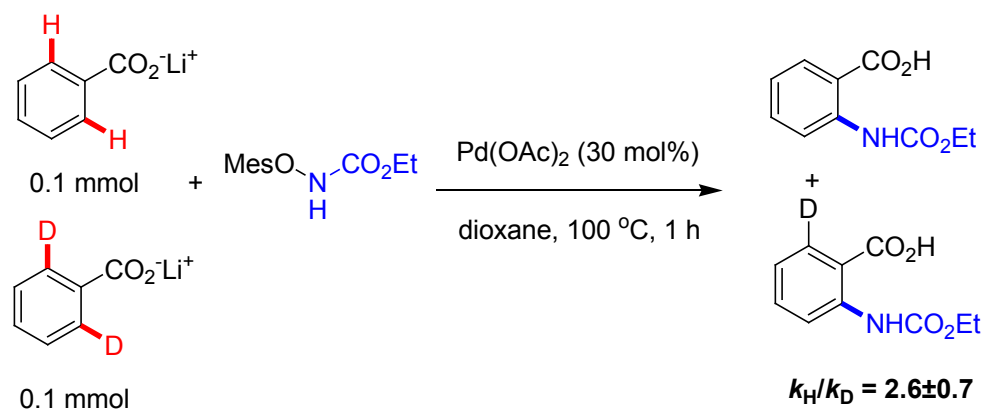
3l was isolated as a yellow solid (28.5 mg, 55% yield). mp = 197-198 °C; ¹H NMR (*d*₆-acetone, 400 MHz): δ_H 10.7 (bs, 1H), 8.82 (s, 1H), 8.76 (s, 1H), 7.98 (d, *J* = 8.0 Hz, 1H), 7.86 (d, *J* = 8.4 Hz, 1H), 7.61 (t, *J* = 8.0 Hz, 1H), 7.45 (t, *J* = 8.0 Hz, 1H), 4.23 (q, *J* = 7.2, 2H), 1.31 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (*d*₆-acetone, 100

MHz): δ_C 169.4 (C), 153.2 (C), 137.1 (C), 136.5 (C), 133.6 (CH), 129.2 (CH), 129.0 (CH), 128.0 (C), 127.0 (CH), 125.0 (CH), 115.6 (C), 114.6 (CH), 60.6 (CH₂), 13.9 (CH₃); IR (KBr, cm⁻¹): 3330, 2987, 1723, 1678, 1541, 1284, 1247, 1220, 1041; HRMS m/z (ESI): calculated for C₁₄H₁₃NO₄Na⁺: 282.0742, found: 282.0748.



3m was isolated as a brown solid (10.9 mg, 21% yield). mp = 140-142 °C; ¹H NMR (CDCl₃, 400 MHz): δ_H 9.58 (s, 1H), 8.55 (d, J = 8.4 Hz, 1H), 8.46 (d, J = 9.2 Hz, 1H), 7.98 (d, J = 9.2 Hz, 1H), 7.81 (d, J = 8.0 Hz, 1H), 7.56 (t, J = 8.0 Hz, 1H), 7.44 (t, J = 7.6 Hz, 1H), 4.27 (q, J = 7.2 Hz, 2H), 1.35 (t, J = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ_C 172.9 (C), 153.8 (C), 140.0 (C), 134.3 (CH), 131.4 (C), 129.9 (C), 128.4 (CH), 128.0 (CH), 125.7 (CH), 124.9 (CH), 119.3 (CH), 61.6 (CH₂), 14.4 (CH₃); IR (KBr, cm⁻¹): 1729, 1654, 1577, 1508, 1251, 1214, 1085, 825; HRMS m/z (ESI): calculated for C₁₄H₁₃NO₄Na⁺: 282.0742, found: 282.0747.

4. Kinetic Isotope Effect (KIE) Study



To a 8 mL-vial, Pd(OAc)₂ (30 mol%) , benzoic acid (A) (0.1 mmol) and 2,6-*d*₂-benzoic acid (*d*₂-A) (0.1 mmol) were added, and the vial was sealed with a Teflon® liner cap, the vial was evacuated and back filled with N₂ for three times. Freshly distilled dioxane (1 mL) was added to the reaction vial. The reaction mixture was stirred at 100 °C for 0.5 h, and ethyl *N*-mesitylsulfonyloxycarbamate (30 mol% in 0.5 mL dioxane) was added. The reaction was stirred at 100 °C for another 0.5 h. After cooling to room temperature, the reaction mixture was diluted with EtOAc (5 mL) and treated with 2 M HCl (2 mL). The organic layer was collected, and the aqueous layer was further extracted with EtOAc (5 mL × 3). The combined organic fractions were dried with MgSO₄. Solvent was removed with rotary evaporation. The residue was redissolved in a minimum amount of DCM and then filtered through a plug of silica gel using a 20% EtOAc in hexanes mixture as eluent. The filtrate was then evaporated to dryness for NMR analysis. The conversions of benzoic acids were determined by ¹H NMR using dibromomethane as the internal standard.

Table S1. Experimental Data of the KIE Study

runs	% conv. of A ^a	R _{unused} ^a = integral ratio of d ₂ - A / A unused	amount of A unused ^b (mol)	amount of d ₂ - A unused ^c (mol)	ratio of A / d ₂ - A consumed ^d (mol ratio)	<i>k</i> _H / <i>k</i> _D
1 st run	48	1.60/1.00	0.0515	0.0816	0.0490/0.0184	2.66
2 nd run	46	1.60/1.00	0.0540	0.8640	0.0460/0.0136	3.38
3 rd run	51	1.46/1.00	0.0490	0.0715	0.0510/0.0285	1.79
average						2.6±0.7

^adetermined by ¹H NMR using dibromomethane as internal standard.

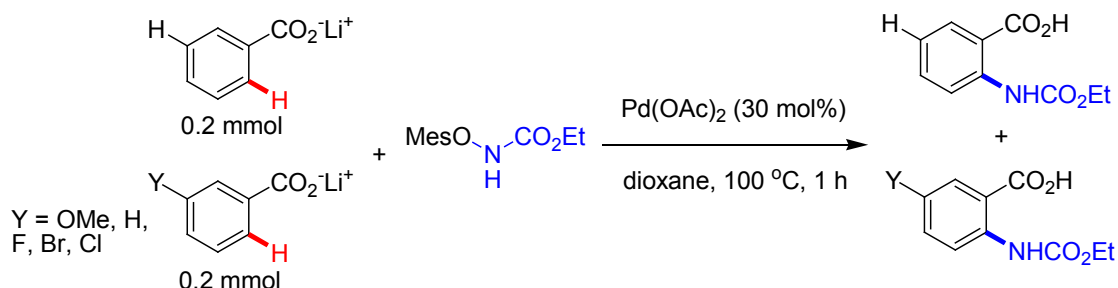
^bamount of **A** unused = 0.1 × (1 – % conv. of **A**)

^camount of **d**₂**A** unused = **R**_{unused} × 0.1 × (1 – % conv. of **A**)

^damount of **d**₂**A** consumed = 0.1 – amount of **d**₂**A** unused

^eamount of **A** consumed = 0.1 – amount of **A** unused

5. Hammett Correlation Studies



To a 8 mL-vial, Pd(OAc)₂ (30 mol%) , benzoic acid (**A**) (0.2 mmol) and 3-Y-benzoic acid (0.2 mmol) (**YA**) were added and the vial was sealed with a Teflon® liner cap, the vial was evacuated and back filled with N₂ for three times. Freshly distilled dioxane (1 mL) was added to the reaction vial. The reaction mixture was stirred at 100 °C for 0.5 h and ethyl *N*-mesitylsulfonyloxycarbamate (30 mol% in 0.5 mL dioxane) was added. The reaction was stirred at 100 °C for another 0.5 h. After cooling to room temperature, the reaction mixture was diluted with EtOAc (5 mL) and treated with 2 M HCl (2 mL). The organic layer was collected, and the aqueous layer was further extracted by EtOAc (5 mL × 3). The combined organic fractions were dried with MgSO₄. Solvent was removed with rotary evaporation. The residue was redissolved in a minimum amount of DCM and then filtered through a plug of silica gel using a 20% EtOAc in hexanes mixture as eluent. The filtrate was then evaporated to dryness for NMR analysis. The conversions of benzoic acids were determined by ¹H NMR using dibromomethane as the internal standard. Each experiment was duplicated and the average values were listed below:

Table S2. Data for the Hammett Correlation Study

Y	% conv. of A ^a	R _{unused} ^a = integral ratios of YA / A unused	amount of A unused ^b (mol)	amount of YA unused ^c (mol)	ratio of YA / A consume ^{d,e} (mol/mol)	<i>k</i> _Y / <i>k</i> _H	log (<i>k</i> _Y / <i>k</i> _H)	σ ⁺
OMe	7.4	0.65/1.00	0.185	0.116	0.084/0.015	5.60	0.75	-0.78
F	7.8	0.78/1.00	0.185	0.144	0.056/0.015	1.63	0.21	-0.07
H	/	1.00/1.00	/	/	1.000/1.000	1.00	0.00	0.00
Cl	23	1.13/1.00	0.154	0.173	0.027/0.046	0.58	-0.24	0.11
Br	13	1.05/1.00	0.174	0.182	0.018/0.026	0.67	-0.17	0.15

^adetermined by ¹H NMR using dibromomethane as internal standard.

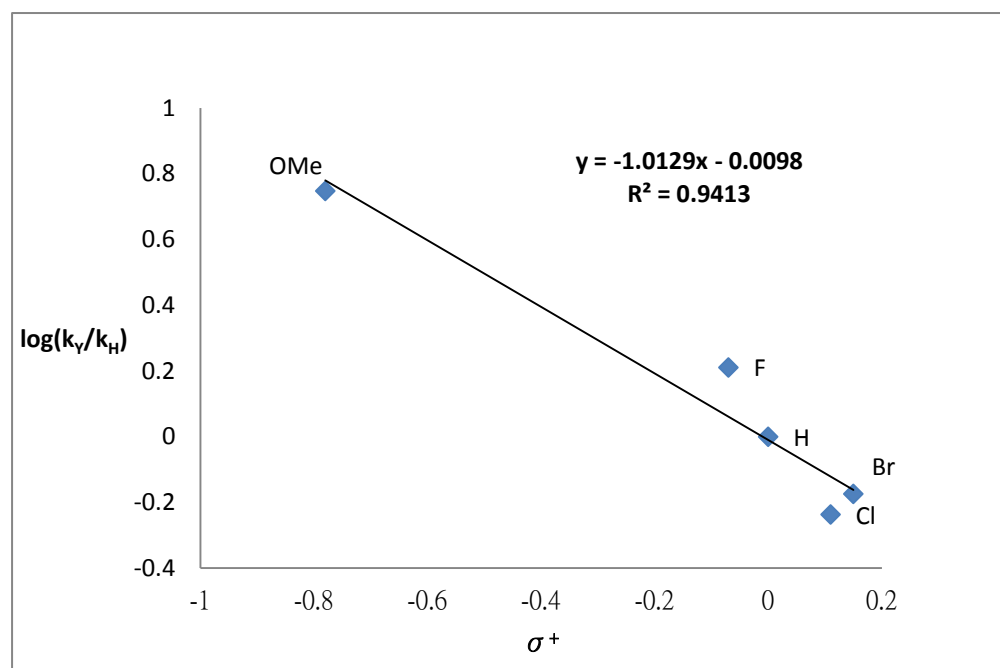
^bamount of **A** unused = 0.2 × (1 – % conv. of **A**)

^camount of **YA** unused = **R**_{unused} × 0.2 × (1 – % conv. of **A**)

^damount of **YA** consumed = 0.2 – amount of **YA** unused

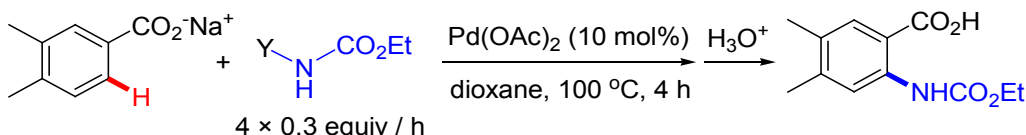
^eamount of **A** consumed = 0.2 – amount of **A** unused

Figure 1. Result of the Hammett Correlation Studies



6. Reaction Optimizations

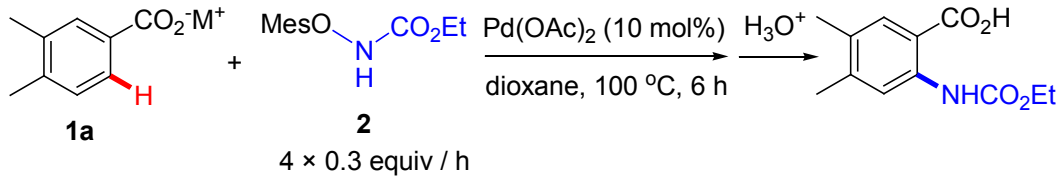
Table S3. Effect of *N*-leaving Groups



entry	Y	conversion (%)	yield (%)
1	NsO	50	5
2	TsO	n.d.	27
3	MesO (2)	67	30
4	PFB	64	25
5	PivO	76	32

^a Reaction conditions: Sodium 3,4-dimethylbenzoate (0.2 mmol), carbamates (4 × 0.3 equiv / h), Pd(OAc)₂ (10 mol%), dioxane (2 mL). NsO = *p*-nitrobenzenesulfonate; TsO = *p*-toluenesulfonate; MesO = *p*-mesitylenesulfonate; PFB = pentafluorobenzoate; PivO = 2,2-dimethylpropionate; n.d. = not determined. Conversions and yields were determined by ¹H NMR with dibromomethane as internal standard.

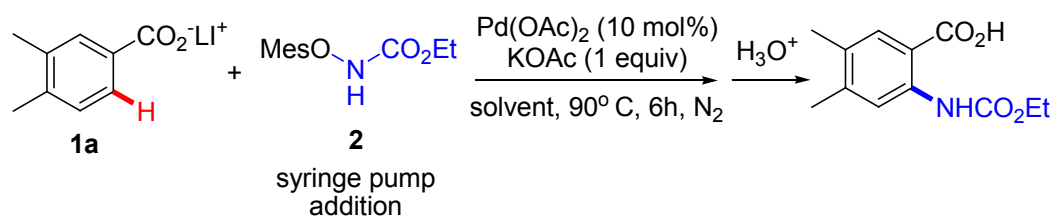
Table S4. Effect of Counter-ions ^a



entry	M ⁺	conversion (%)	yield (%)
1	H	0	0
2	Li	70	40
3	Na	66	30
4	K	56	16
5	(<i>n</i> -Bu) ₄ N	n.d.	14

^a Reaction conditions: **1a** (0.2 mmol), **2** (4 × 0.3 equiv / h), Pd(OAc)₂ (10 mol%), solvent (2 mL). n.d. = not determined. Conversions and yields were determined by ¹H NMR with dibromomethane as internal standard.

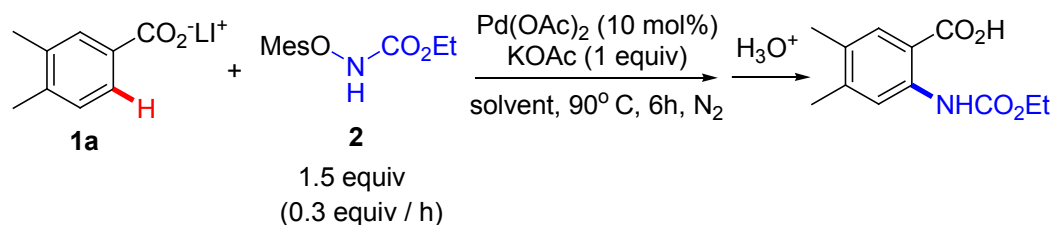
Table S5. Effect of Reagent Quantities and Addition Rates^a



entry	2 (equiv)	reaction time (h)	addition rate (equiv / h)	conversion (%)	yield (%)
1	1.2	6	0.3	95	45
2	1.5	6	0.3	95	47
3	2	6	0.3	95	41
4	1.5	4	0.6	91	45
5	1.5	2	1.2	87	40
6 (under N ₂)	1.5	6	0.3	83	65

^a Reaction conditions: **1a** (0.2 mmol), **2** (syringe pump addition), Pd(OAc)₂ (10 mol%), KOAc (1 equiv), dioxane (1.5 mL). Conversions and yields were determined by ¹H NMR with dibromomethane as internal standard.

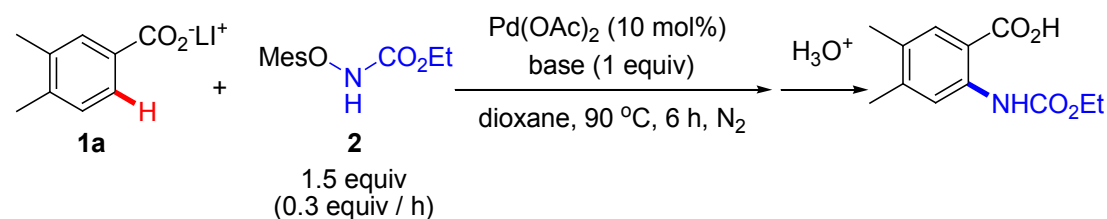
Table S6. Effect of Solvents^a



entry	solvent	conversion (%)	yield (%)
1	DMF	50	20
2	DMSO	80	5
3	DMA	21	13
4	Diglyme	67	45
5 ^b	<i>t</i> -BuOH	90	57
6 ^c	DCE	87	45
7	dioxane	83	65(63) ^d

^a Reaction conditions: **1a** (0.2 mmol), **2** (1.5 equiv, 0.3 equiv / h, syringe pump addition), Pd(OAc)₂ (10 mol%), KOAc (1 equiv), solvent (1.5 mL) under N₂. Conversions and yields were determined by ¹H NMR with dibromomethane as internal standard. ^b 20% dioxane was added. ^c 10% dioxane was added to dissolve the reagent. ^d isolated yield in parentheses.

Table S7 . Effect of Bases^a



entry	base (equiv)	conversion (%)	yield (%)
1	KOAc	83	65(63) ^b
2	KOPiv	67	41
3	K ₂ HPO ₄	80	54
4	KHCO ₃	80	59
5	K ₂ CO ₃ (0.5)	71	60
6	K ₂ CO ₃ (0.75)	80	56
7	K ₂ CO ₃	76	48

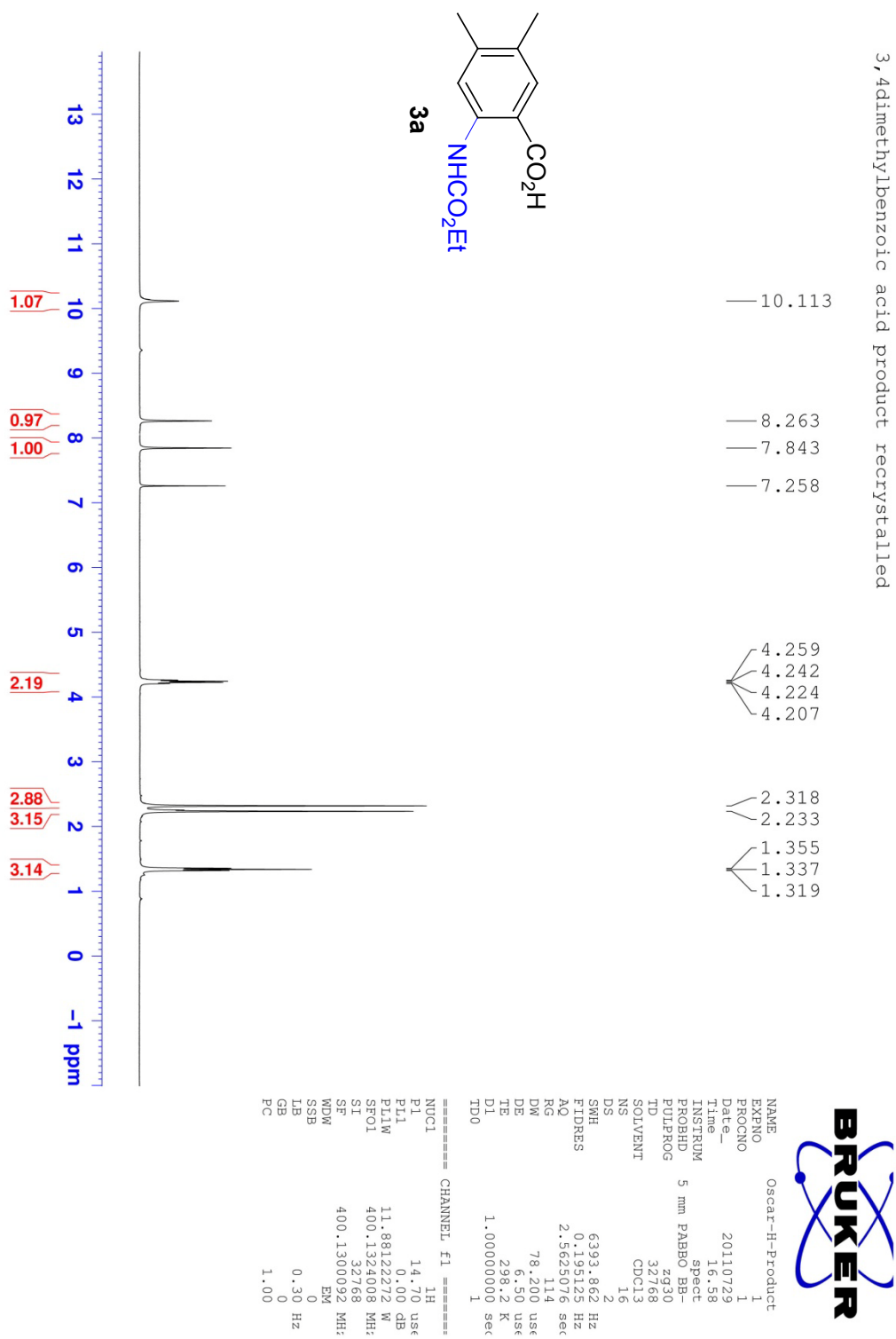
^a Reaction conditions: **1a** (0.2 mmol), **2** (1.5 equiv, 0.3 equiv / h, syringe pump addition), Pd(OAc)₂ (10 mol%), base (0.5 – 1 equiv), dioxane (1.5 mL) under N₂. Conversions and yields were determined by ¹H NMR with dibromomethane as internal standard. ^b isolated yield in parentheses.

7. References

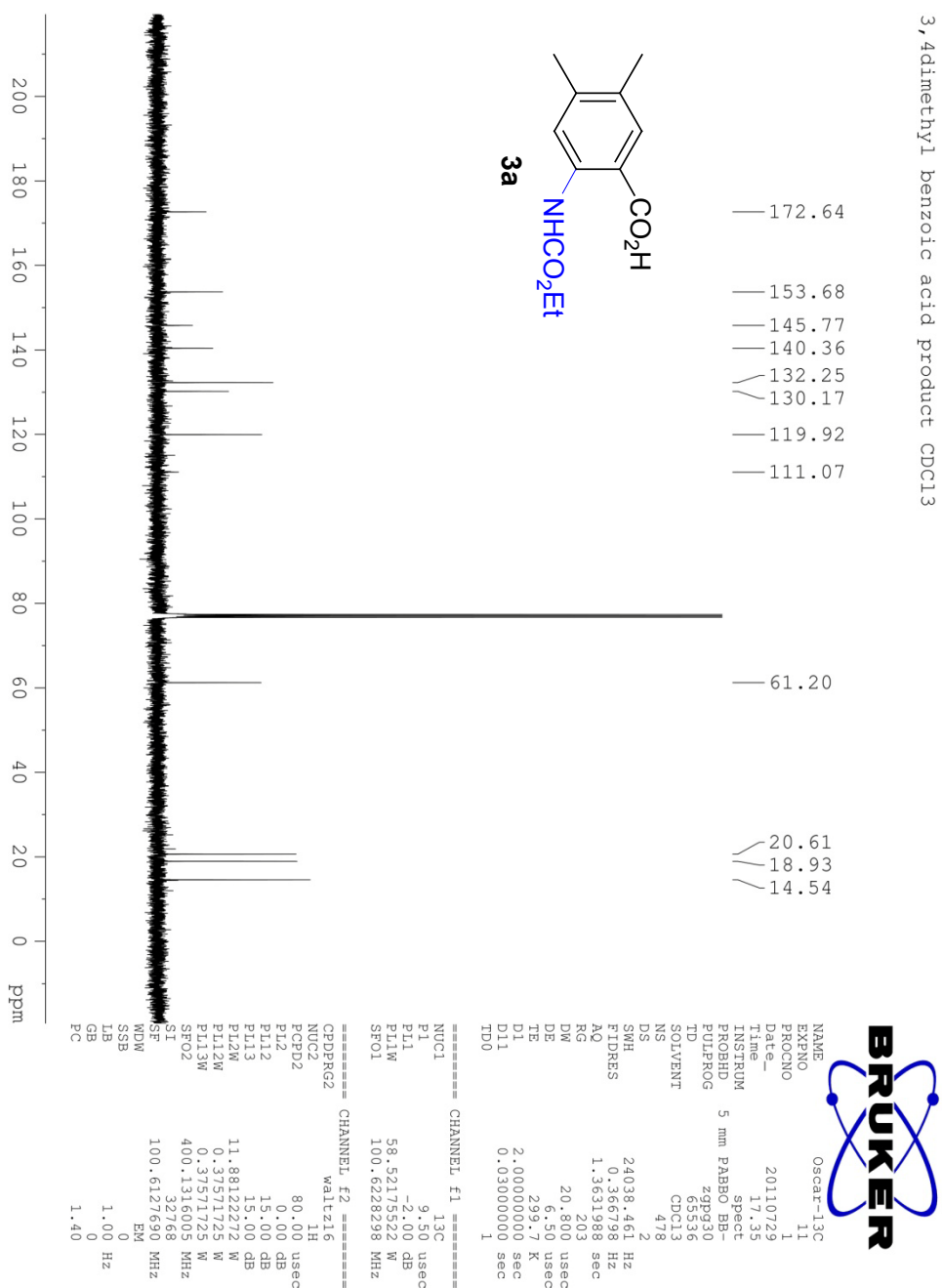
1. K.-H. Ng, A. S. C. Chan, and W.-Y. Yu, *J. Am. Chem. Soc.*, 2010, **132**, 12864.
2. D. R. Boyd, N. D. Sharma, J. S. Harrison, J. F. Malone, W. C. McRoberts, J. T. G. Hamilton, and D. B. Harper, *Org. Biomol. Chem.*, 2008, **6**, 1251.
3. A. Herschhorn, L. Lerman, M. Weitman, I. O. Gleenberg, A. Nudelman, and A. Hizi, *J. Med. Chem.*, 2007, **50**, 2370.

8. ^1H NMR and ^{13}C NMR Spectra

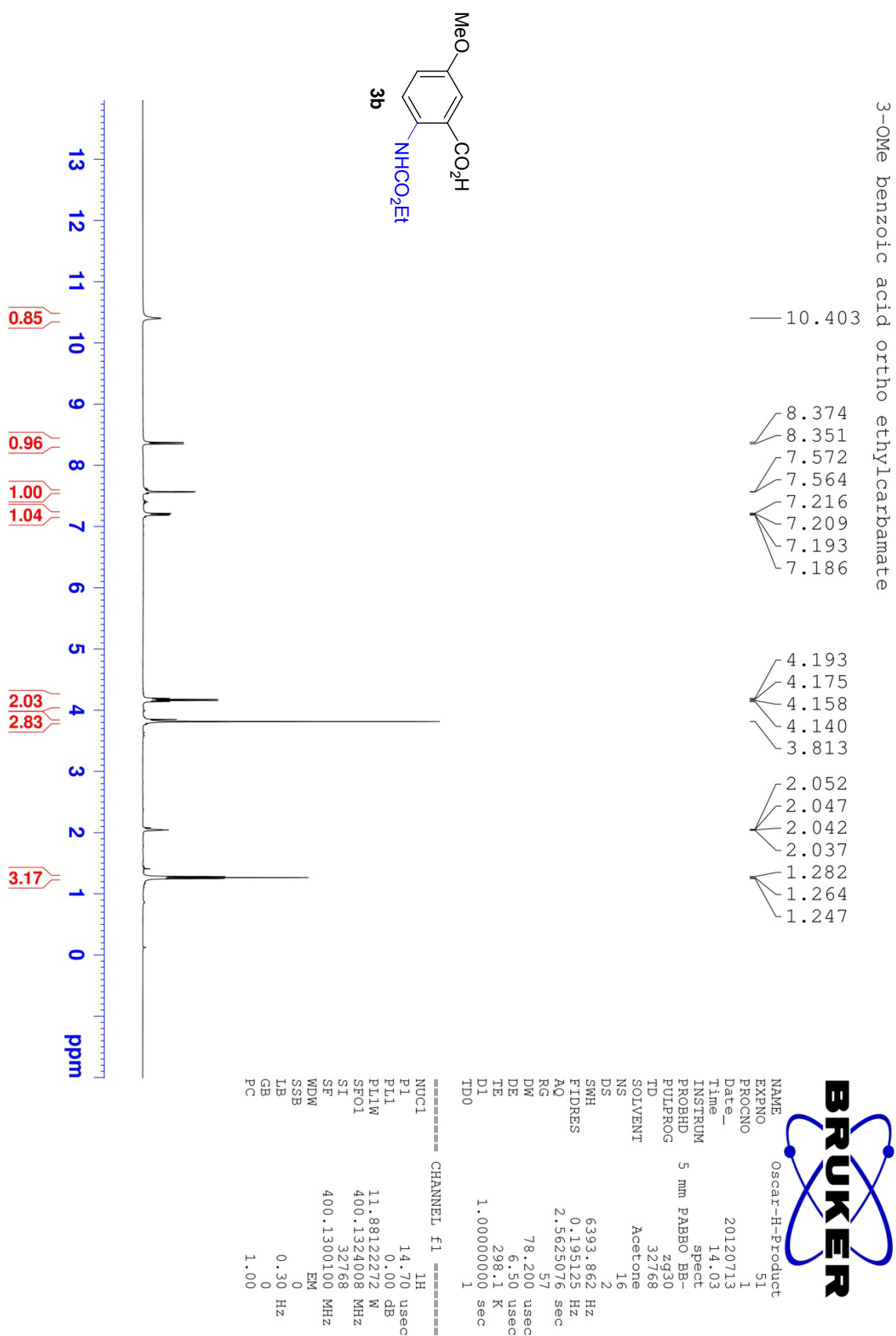
^1H NMR spectrum of **3a**



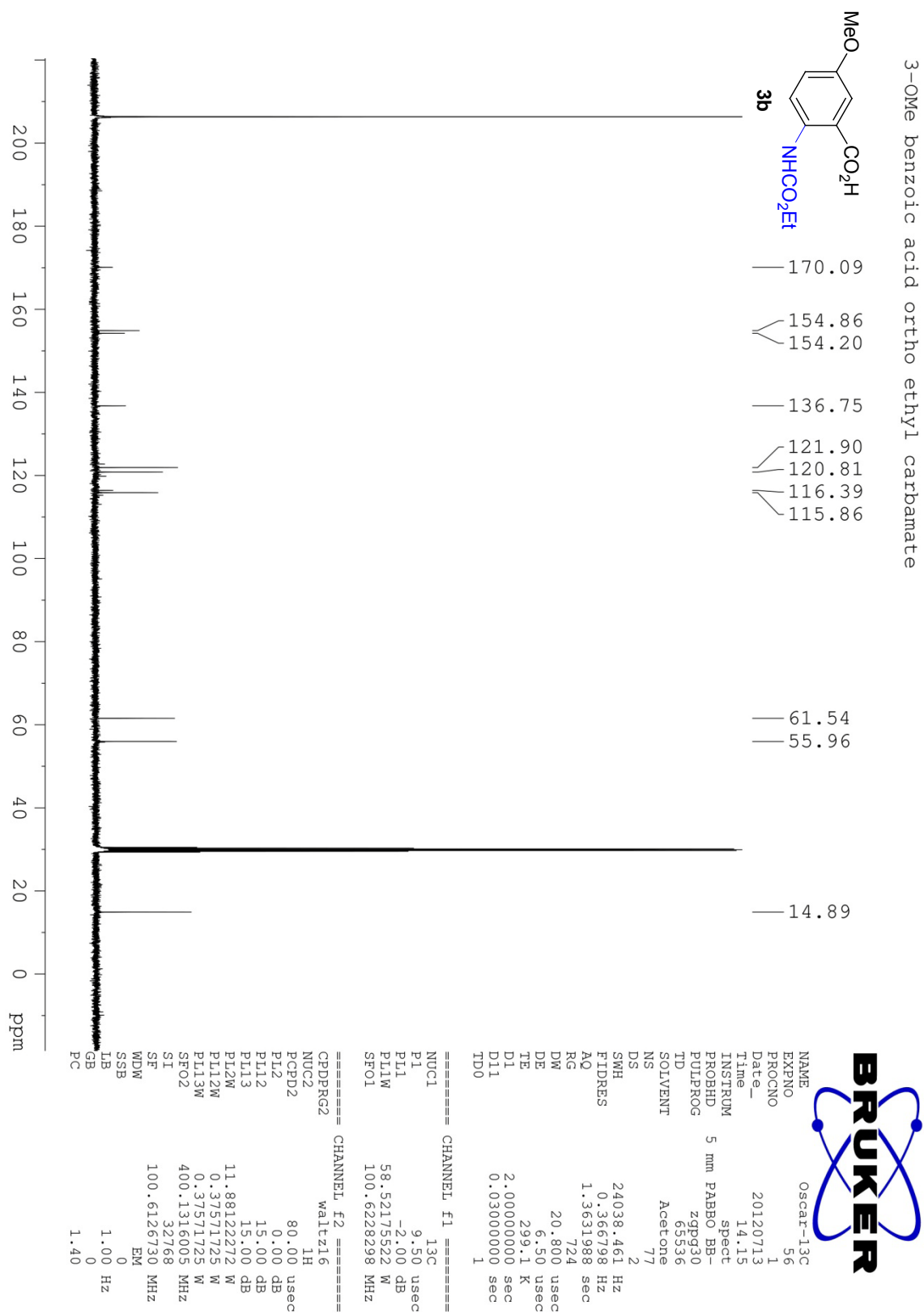
¹³C NMR spectrum of **3a**



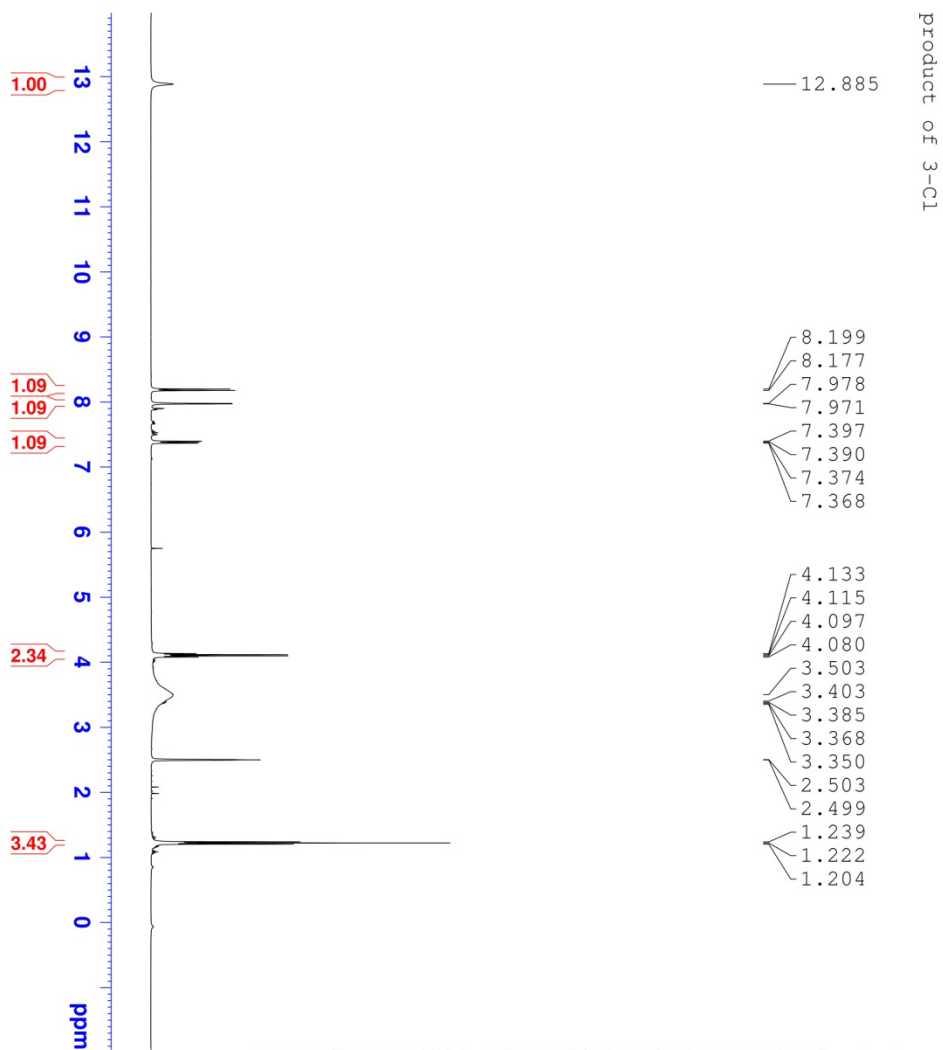
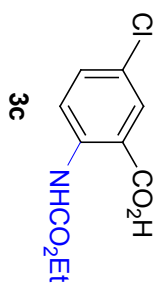
¹H NMR spectrum of **3b**



¹³C NMR spectrum of **3b**



¹H NMR spectrum of **3c**



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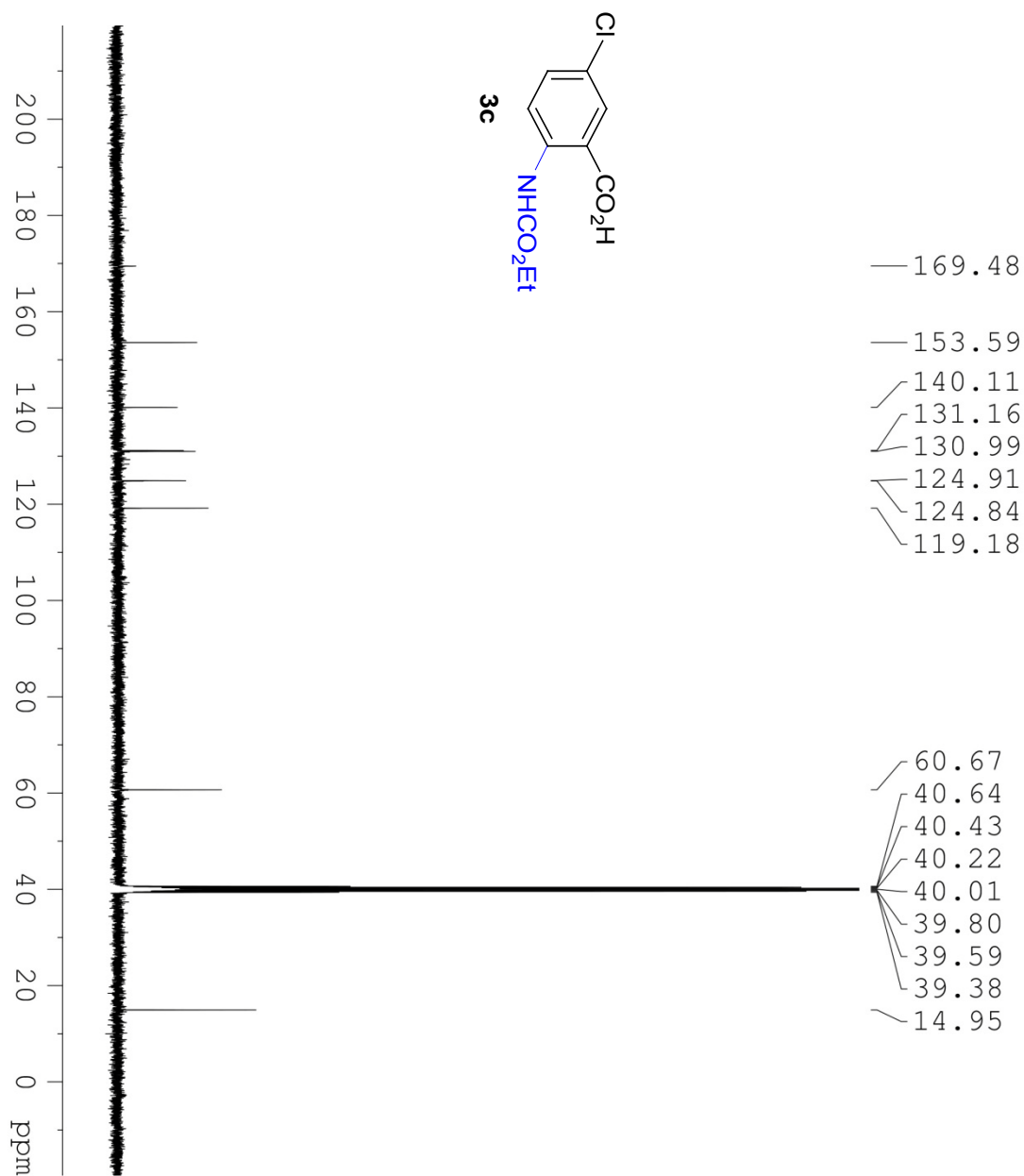
Oscar-H-column

```

NAME          Oscar-H-column
EXPNO         23
PROCNO        1
Date_         20110402
Time         19.30
INSTRUM       5 mm PABBO BR-
PROBHD        zg30
PULPROG       32768
TD            16
SOLVENT       DMSO
NS            16
DS            2
SMH           6393.862 Hz
FIDRES        0.195125 Hz
AQ            2.5625076 sec
RG            25.4
DW            78.200 usec
DE            6.50 usec
TE            299.6 K
D1            1.00000000 sec
TD0           1

===== CHANNEL f1 =====
NUC1          1H
P1            14.70 usec
PL1           0.00 dB
PL1W          11.88122272 W
SFO1          400.1324008 MHz
SI            32768
SE           400.1300036 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

¹³C NMR spectrum of **3c**



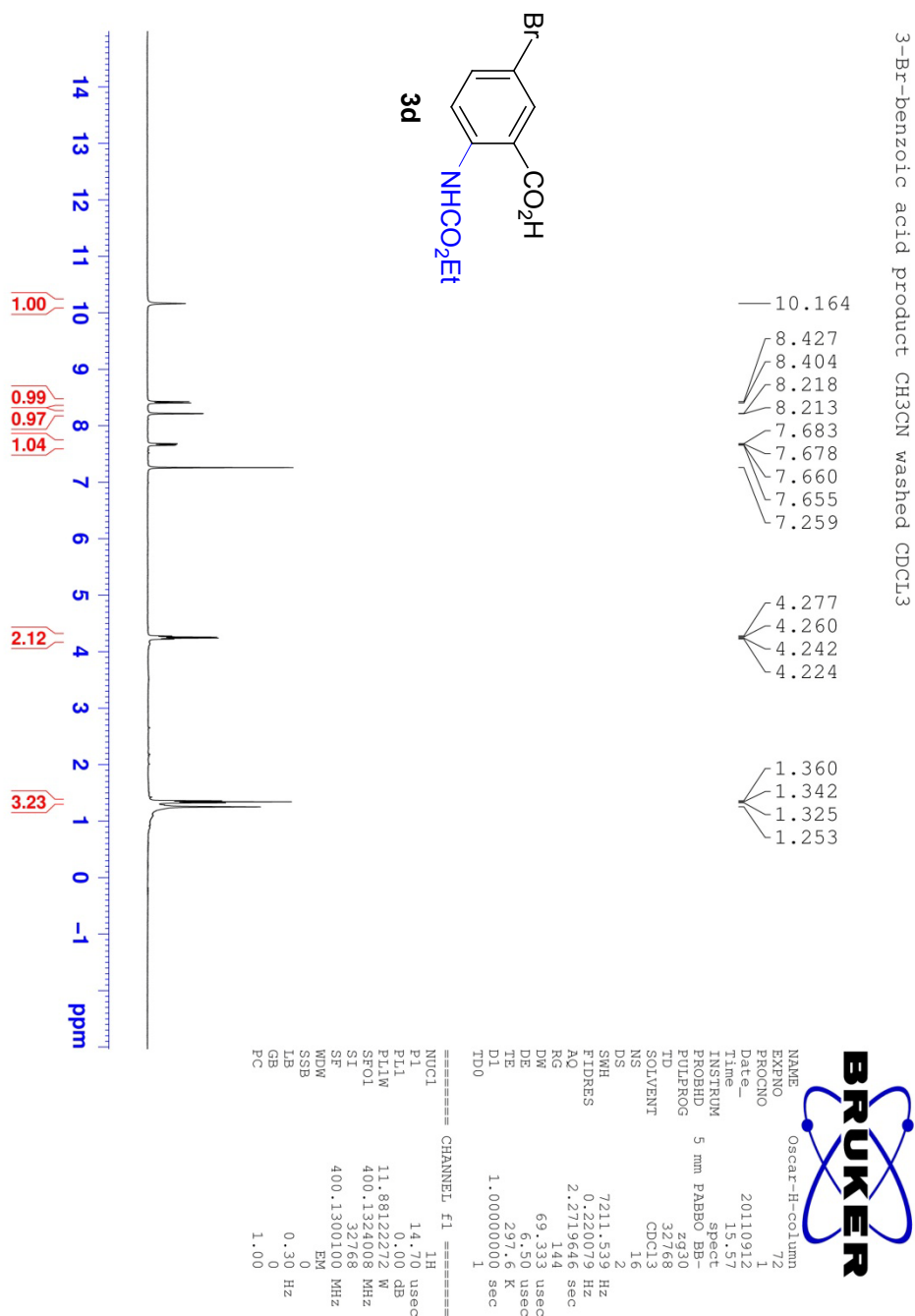
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===== OSCAR-13C =====
NAME          OSCAR-13C
EXPNO         6
PROCNO        1
Date_         20110402
Time          19.57
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       zgpg30
TD            65536
SOLVENT       DMSO
NS            413
DS            2
SWH           24038.461 Hz
FIDRES        0.366798 Hz
AQ            1.3631988 sec
RG            40.3
DW            20.800 usec
DE            6.50 usec
TE            301.1 K
D1            2.00000000 sec
D11           0.03000000 sec
TD0           1

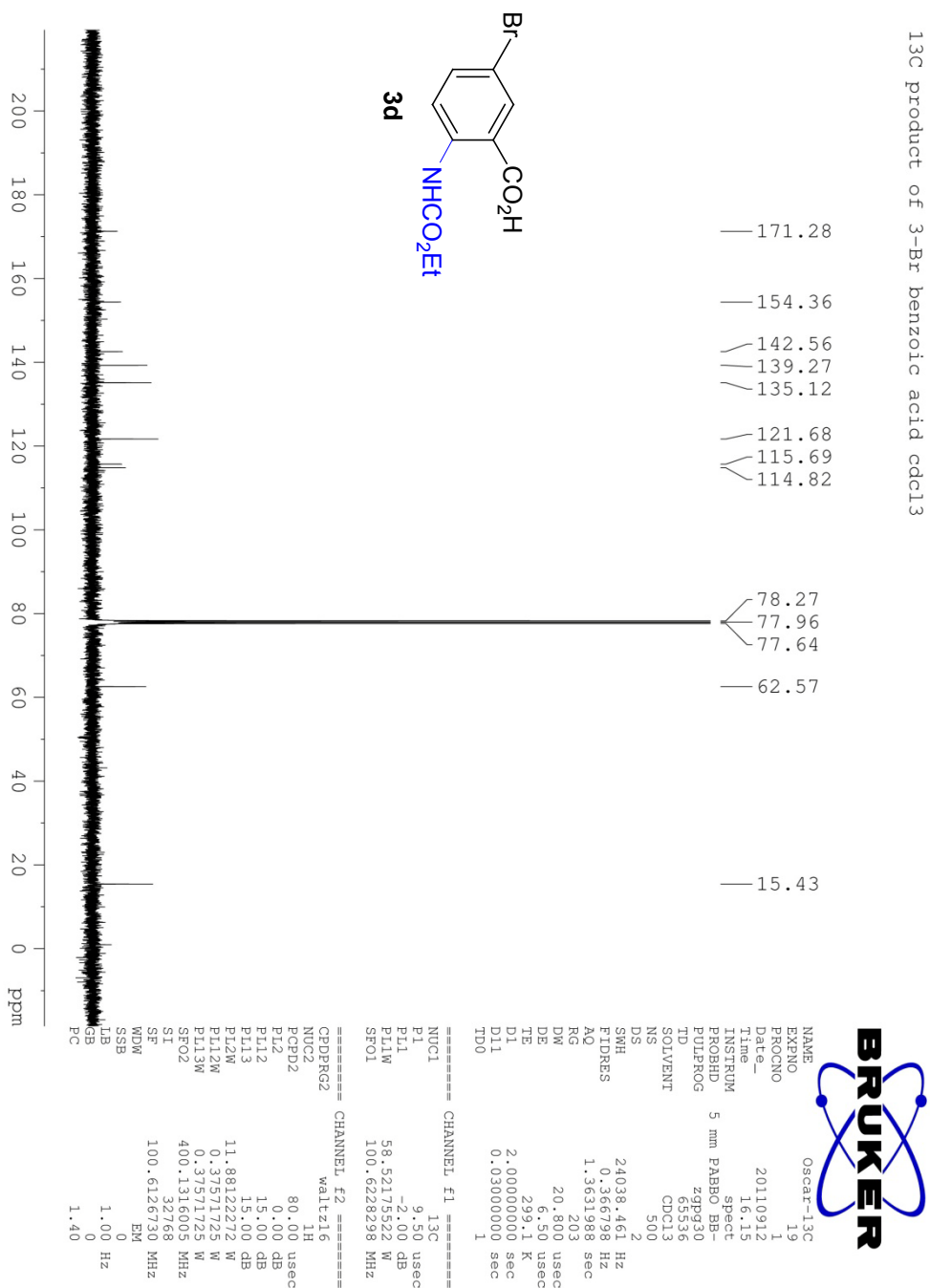
===== CHANNEL f1 =====
NUC1          13C
P1            9.50 usec
PL1          -2.00 dB
PL1W         58.52175522 W
SFO1         100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        80.00 usec
PL2           0.00 dB
PL12         15.00 dB
PL13         15.00 dB
PL2W         11.88132272 W
PL12W        0.37571725 W
PL13W        0.37571725 W
SFO2         400.1316005 MHz
SI           32768
SF           100.6127690 MHz
WDW          EM
SSB           0
LB           1.00 Hz
GB           0
PC           1.40
    
```

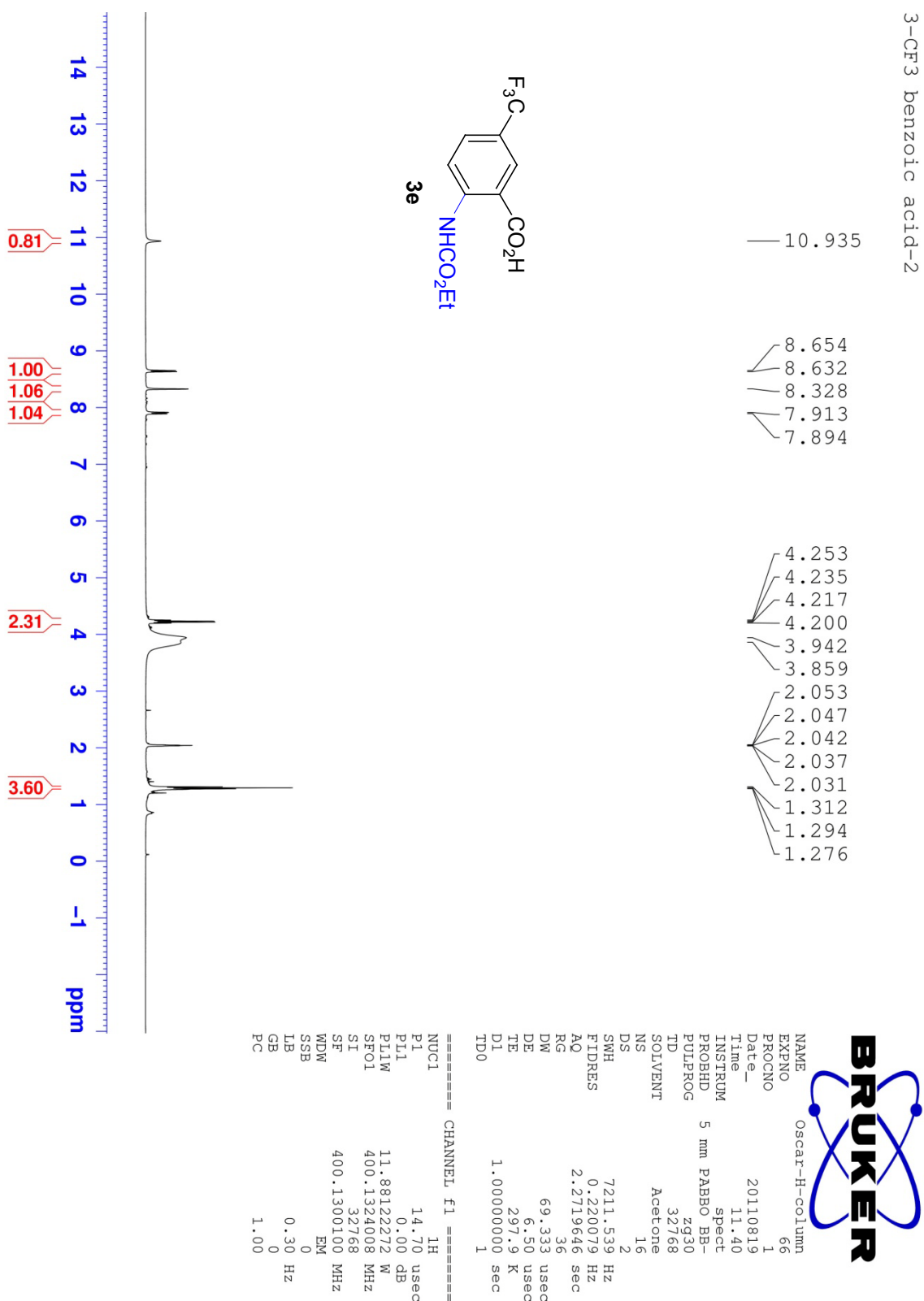
¹H NMR spectrum of **3d**



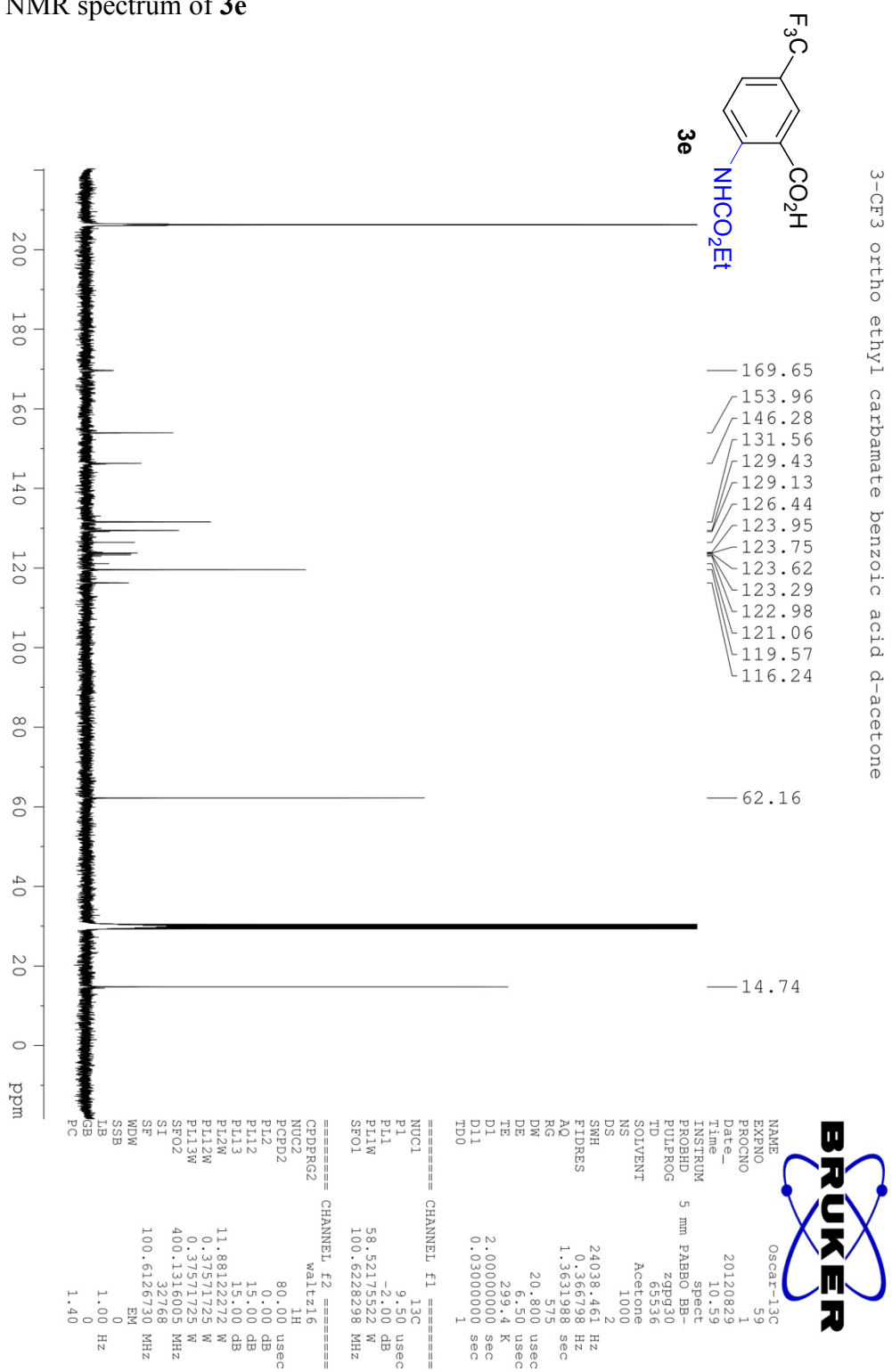
¹³C NMR spectrum of **3d**



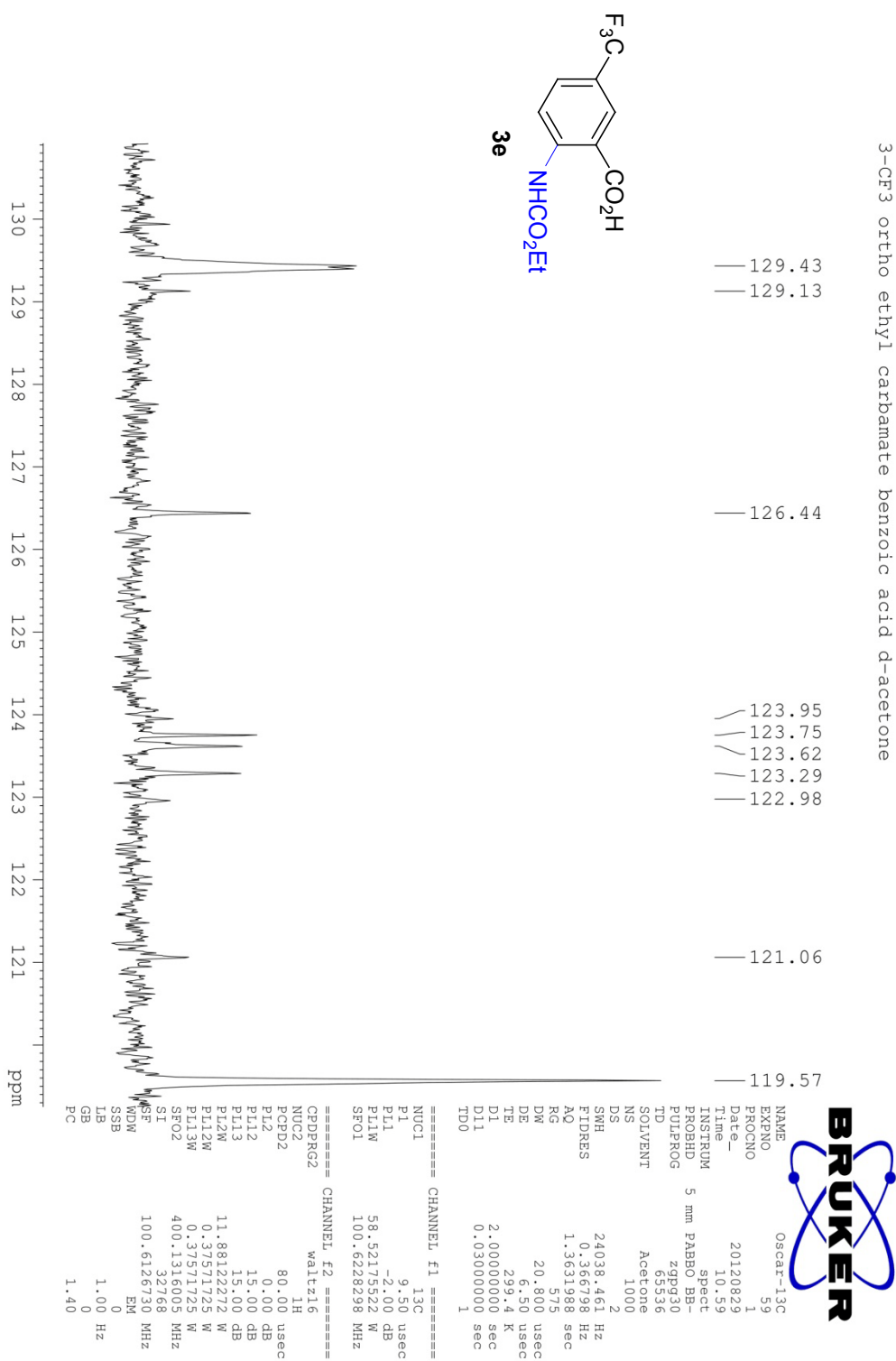
¹H NMR spectrum of **3e**



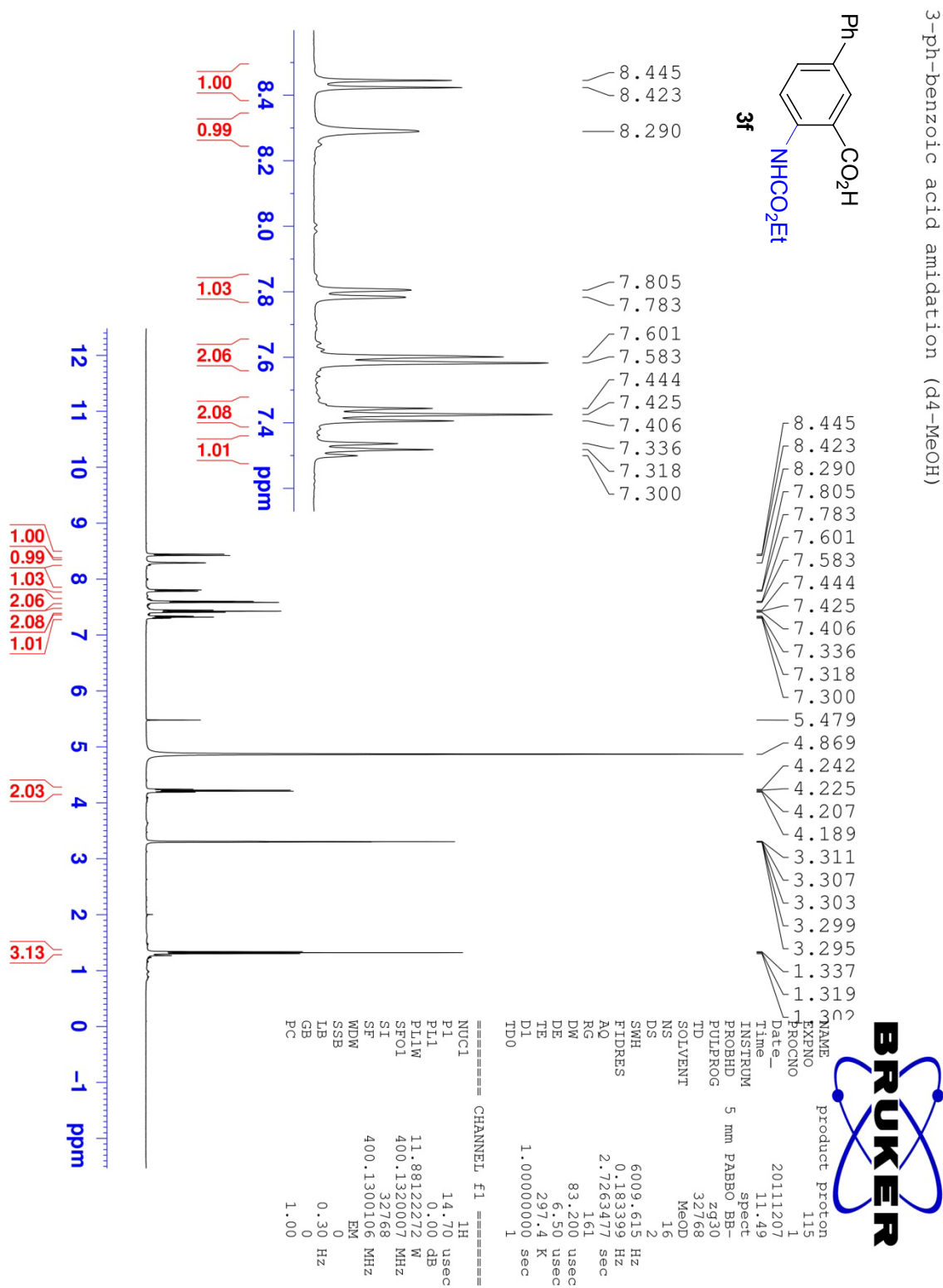
¹³C NMR spectrum of **3e**



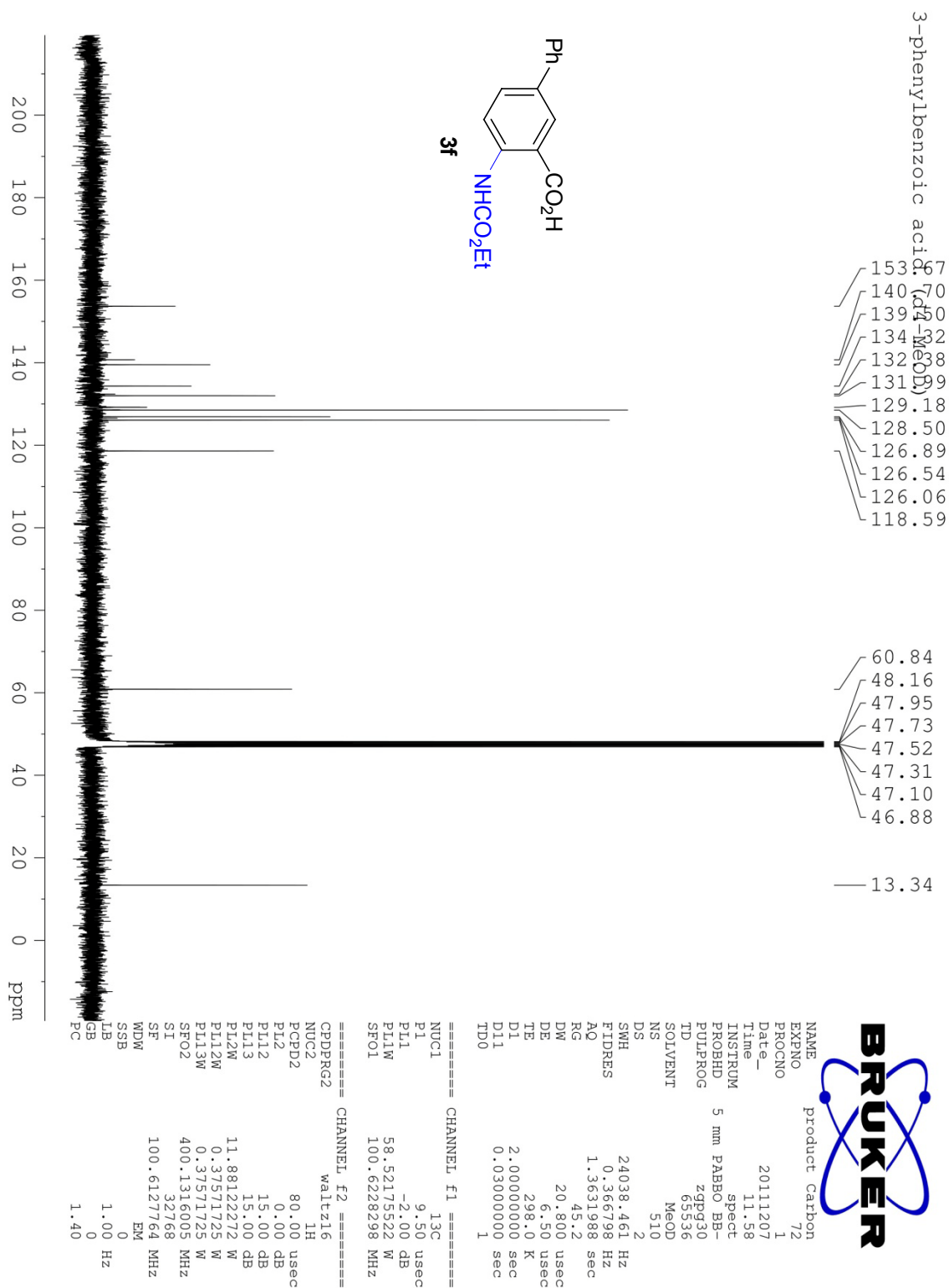
¹³C NMR spectrum of **3e** (magnified)



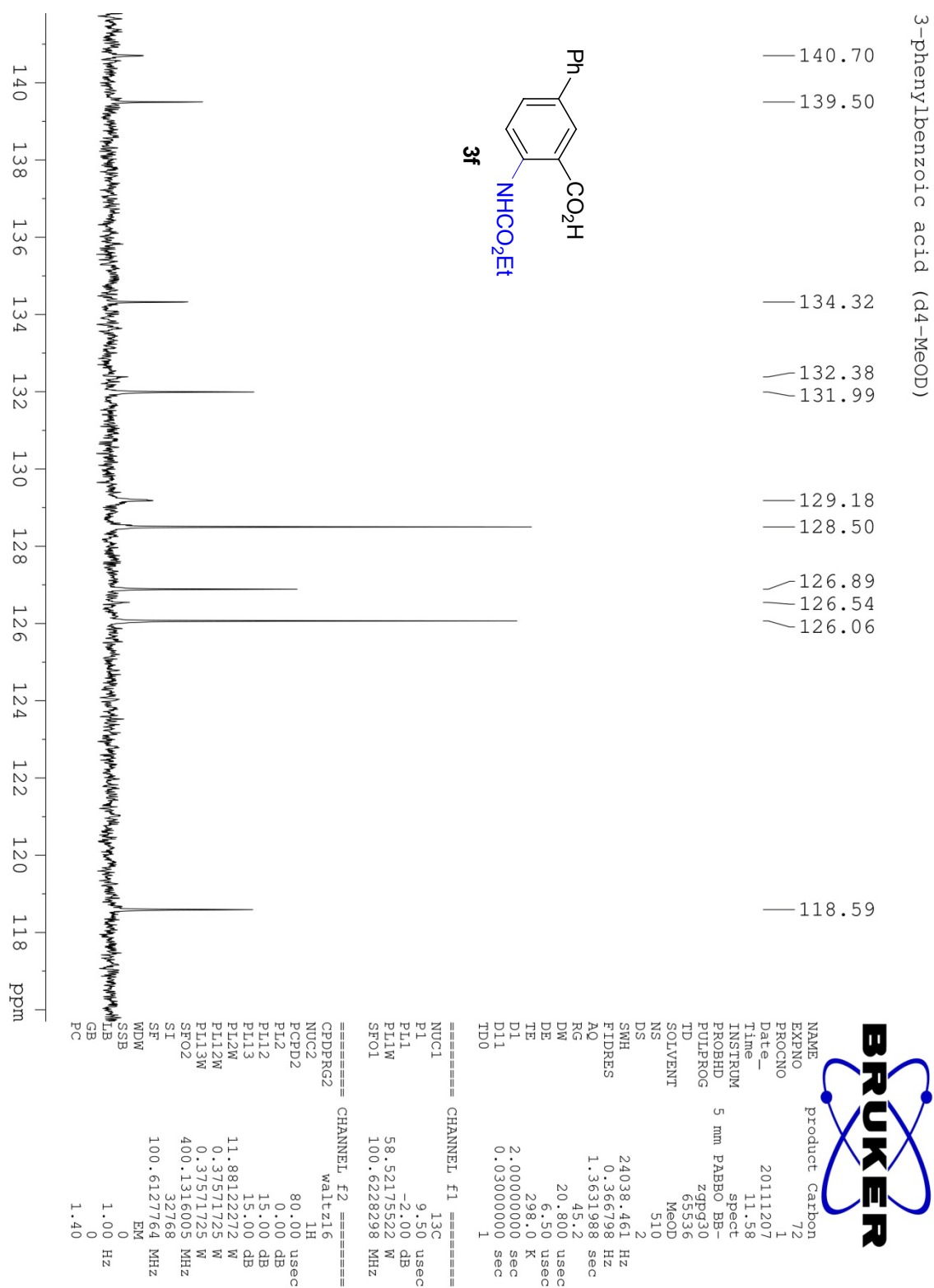
¹H NMR spectrum of **3f**



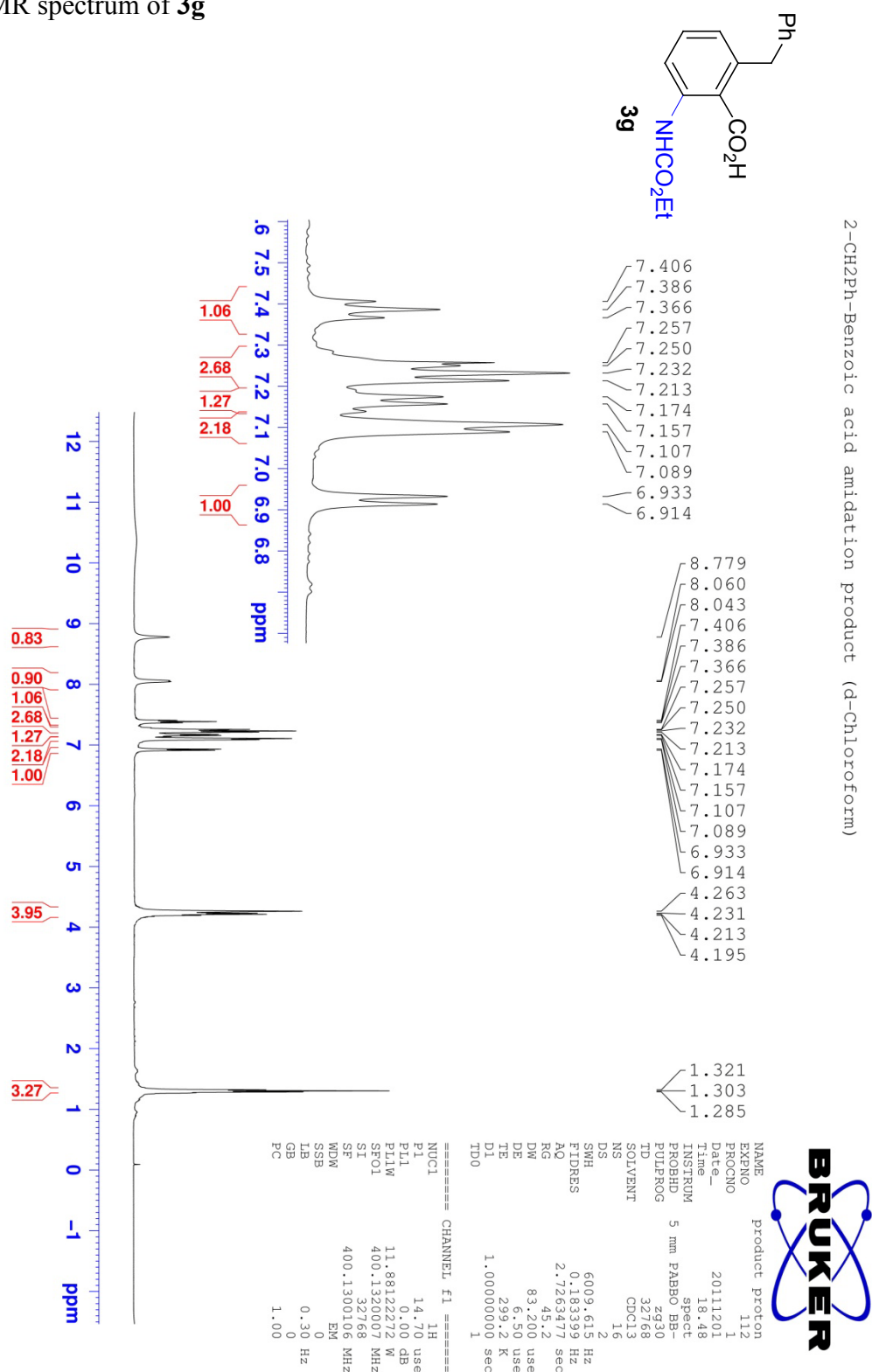
¹³C NMR spectrum of **3f**



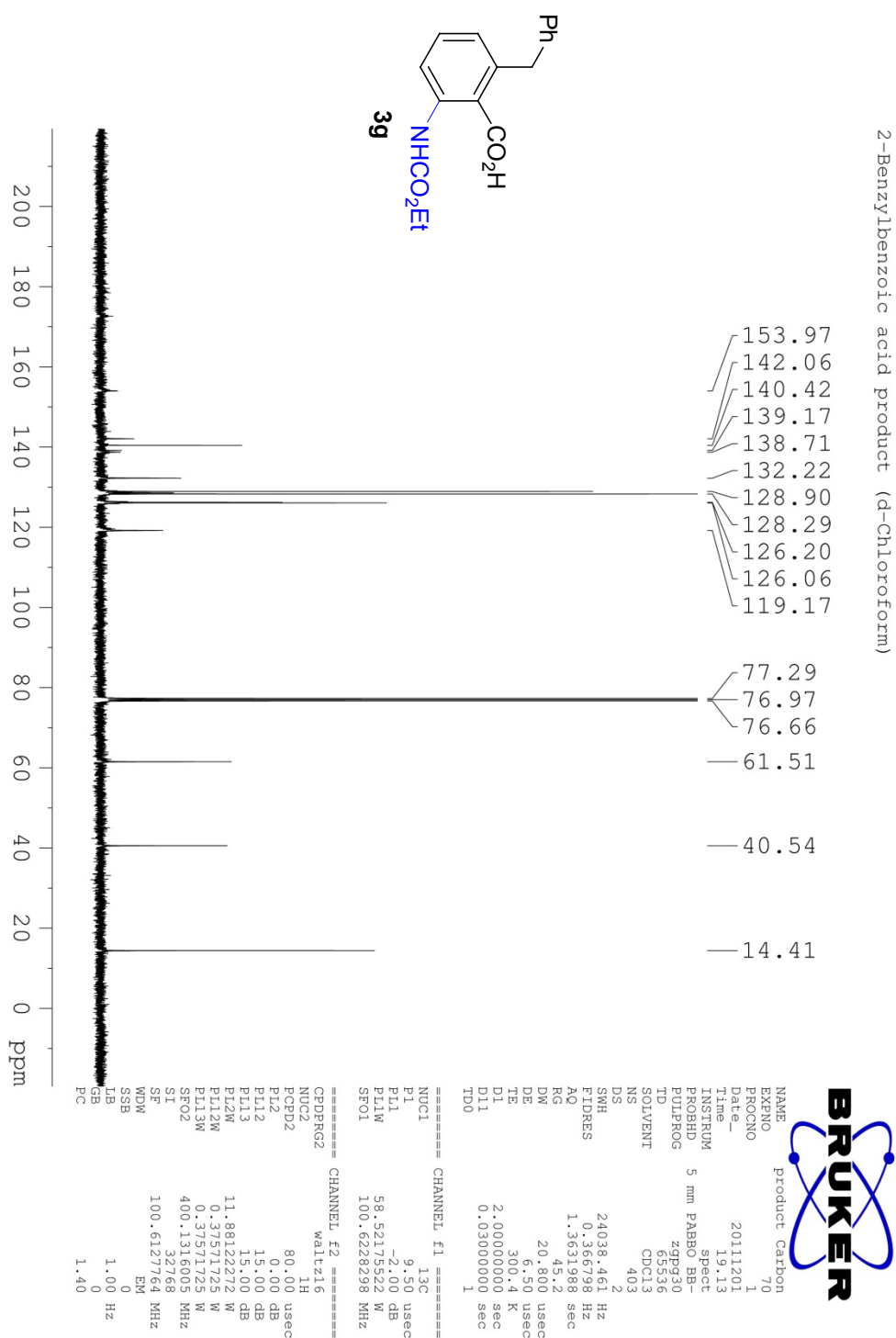
^{13}C NMR spectrum of **3f** (magnified)



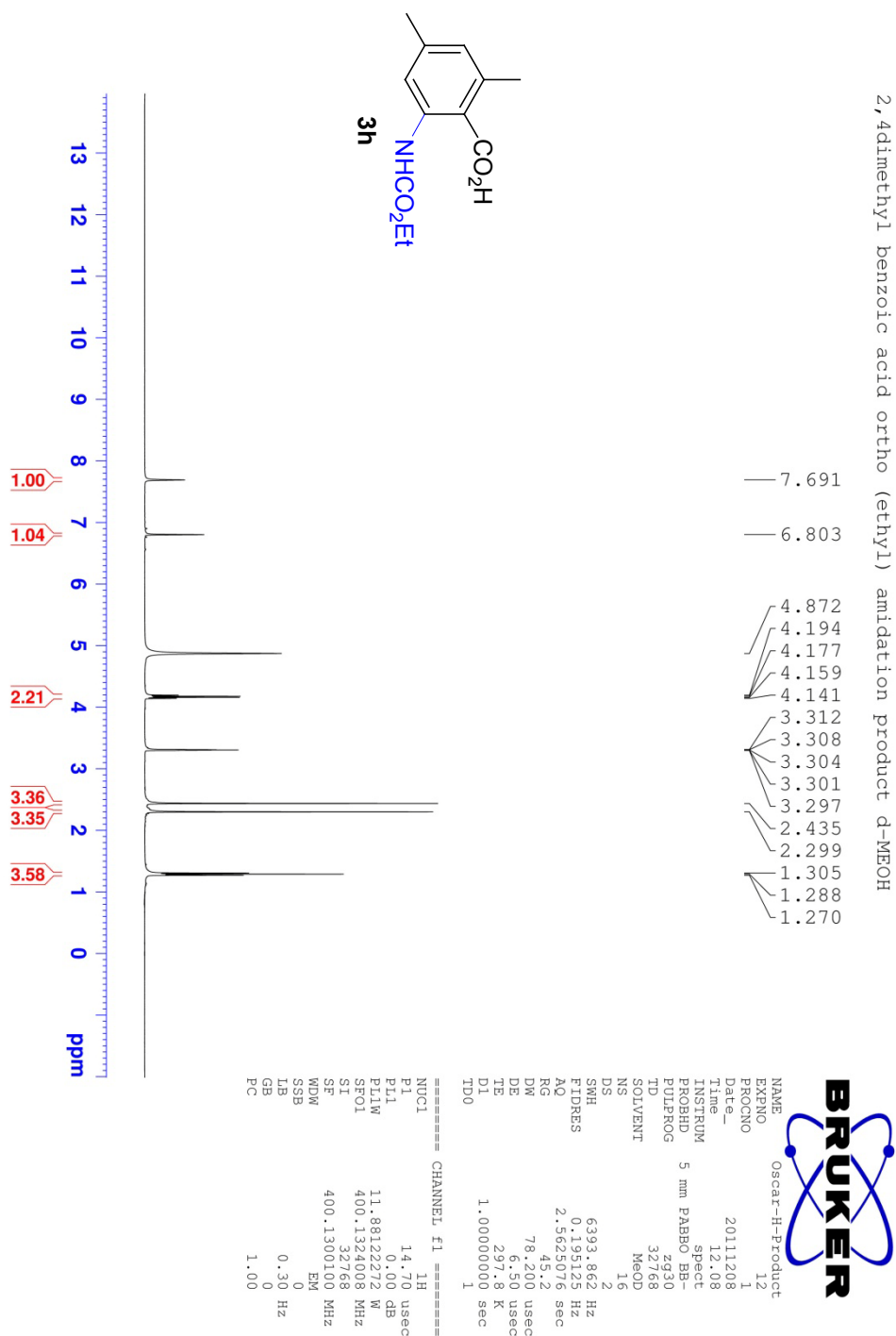
¹H NMR spectrum of **3g**



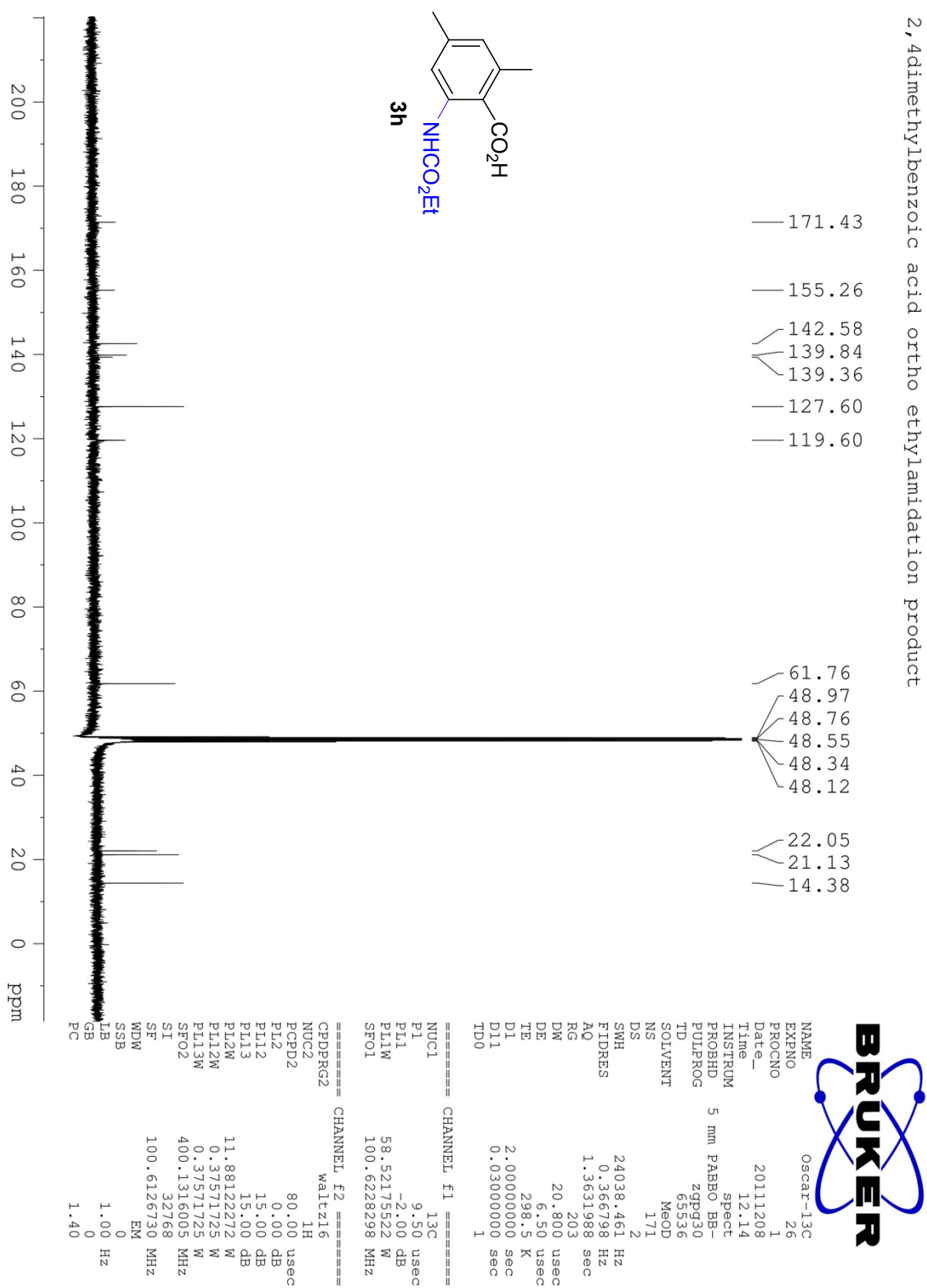
¹³C NMR spectrum of **3g**



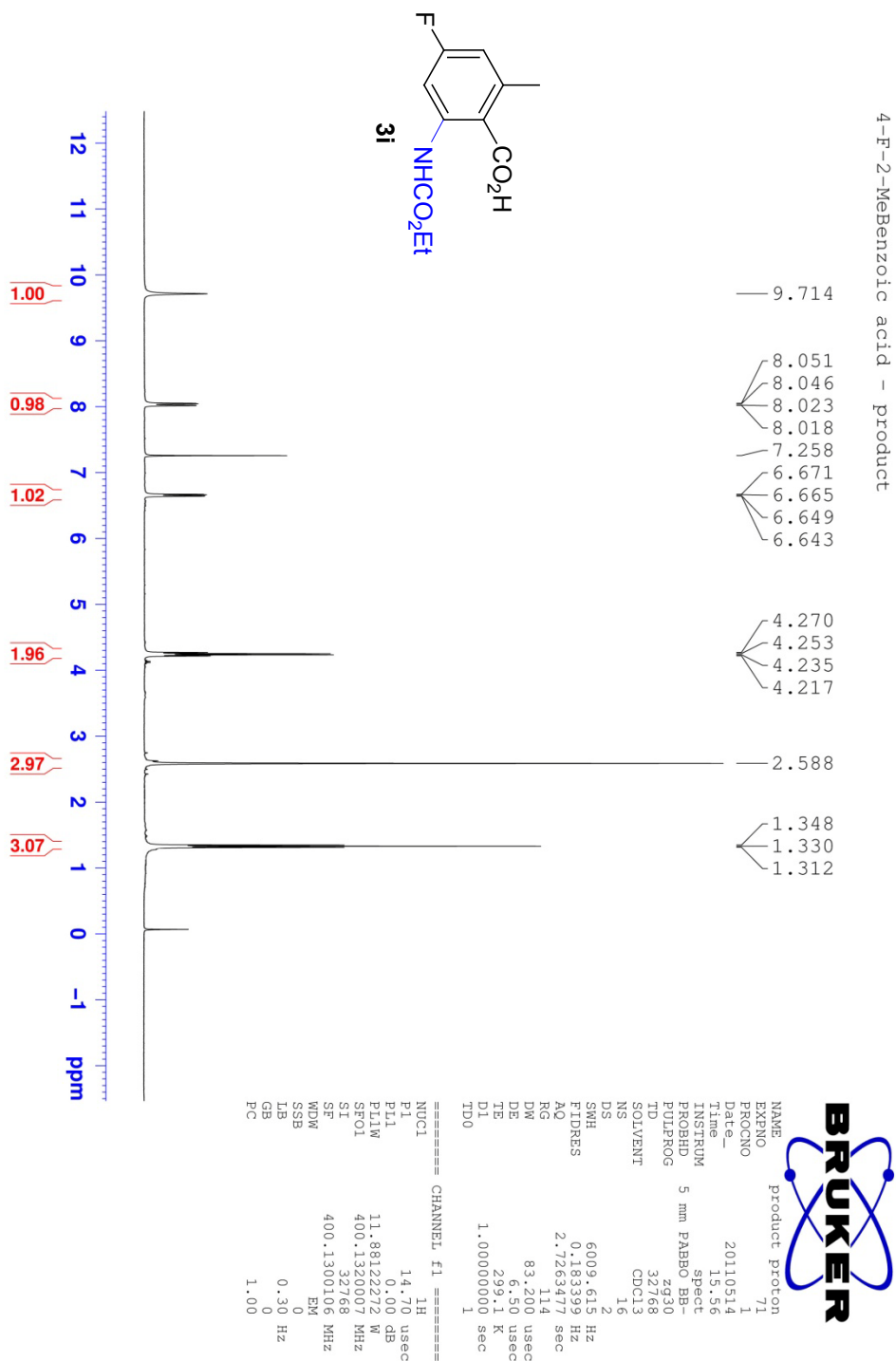
¹H NMR spectrum of **3h**



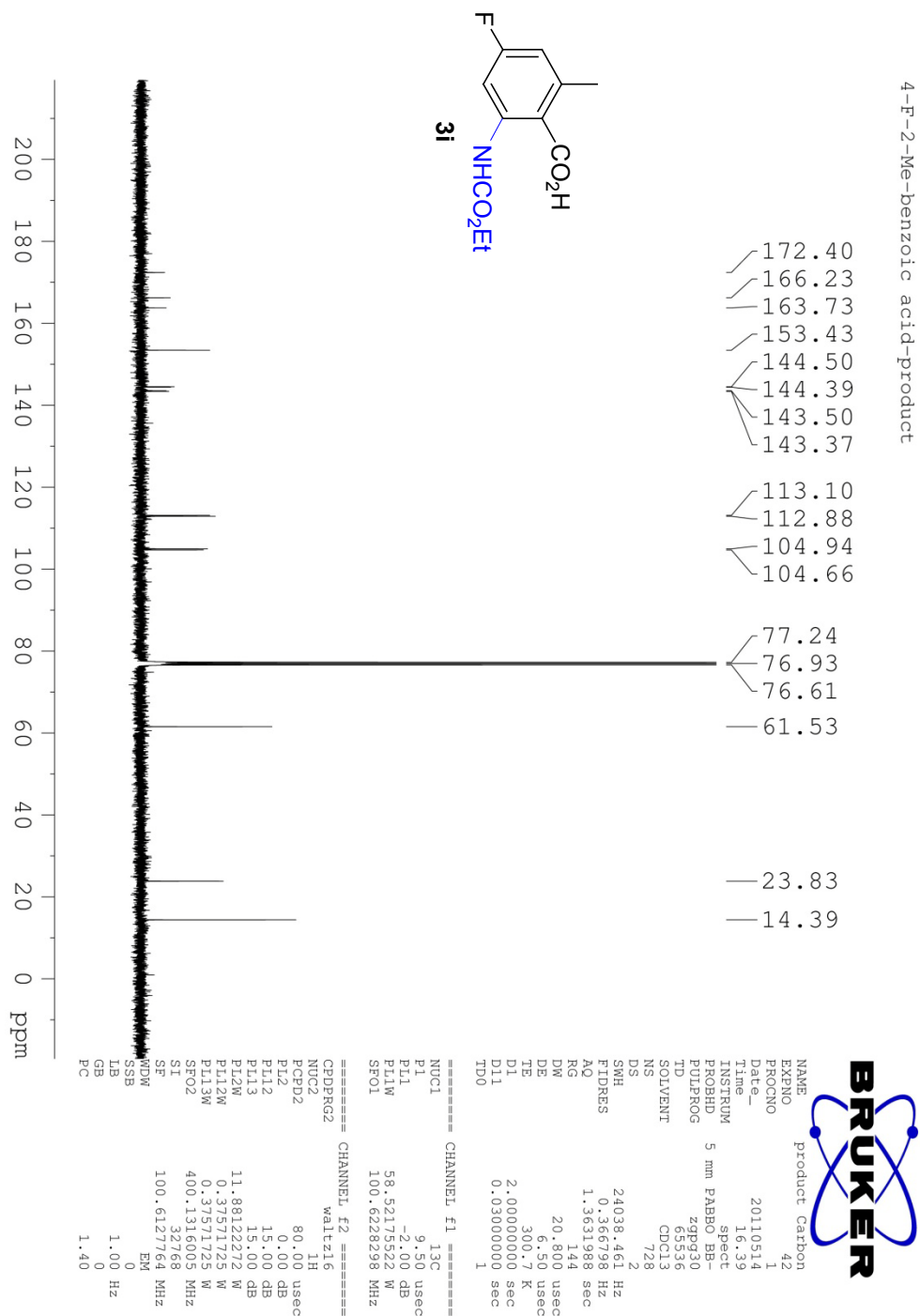
¹³C NMR spectrum of **3h**



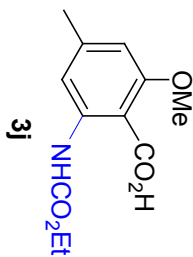
¹H NMR spectrum of **3i**



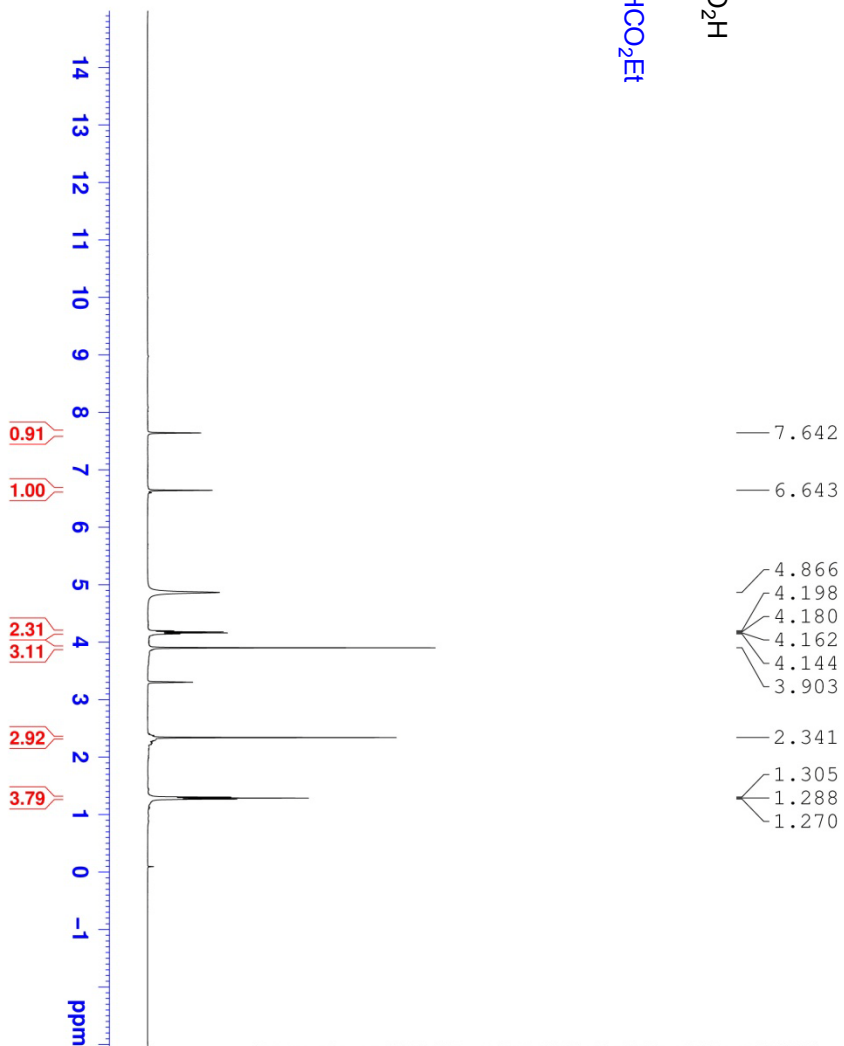
¹³C NMR spectrum of **3i**



¹H NMR spectrum of **3j**



Pd-substrate 2-Me-4Mbenzoic acid after LC

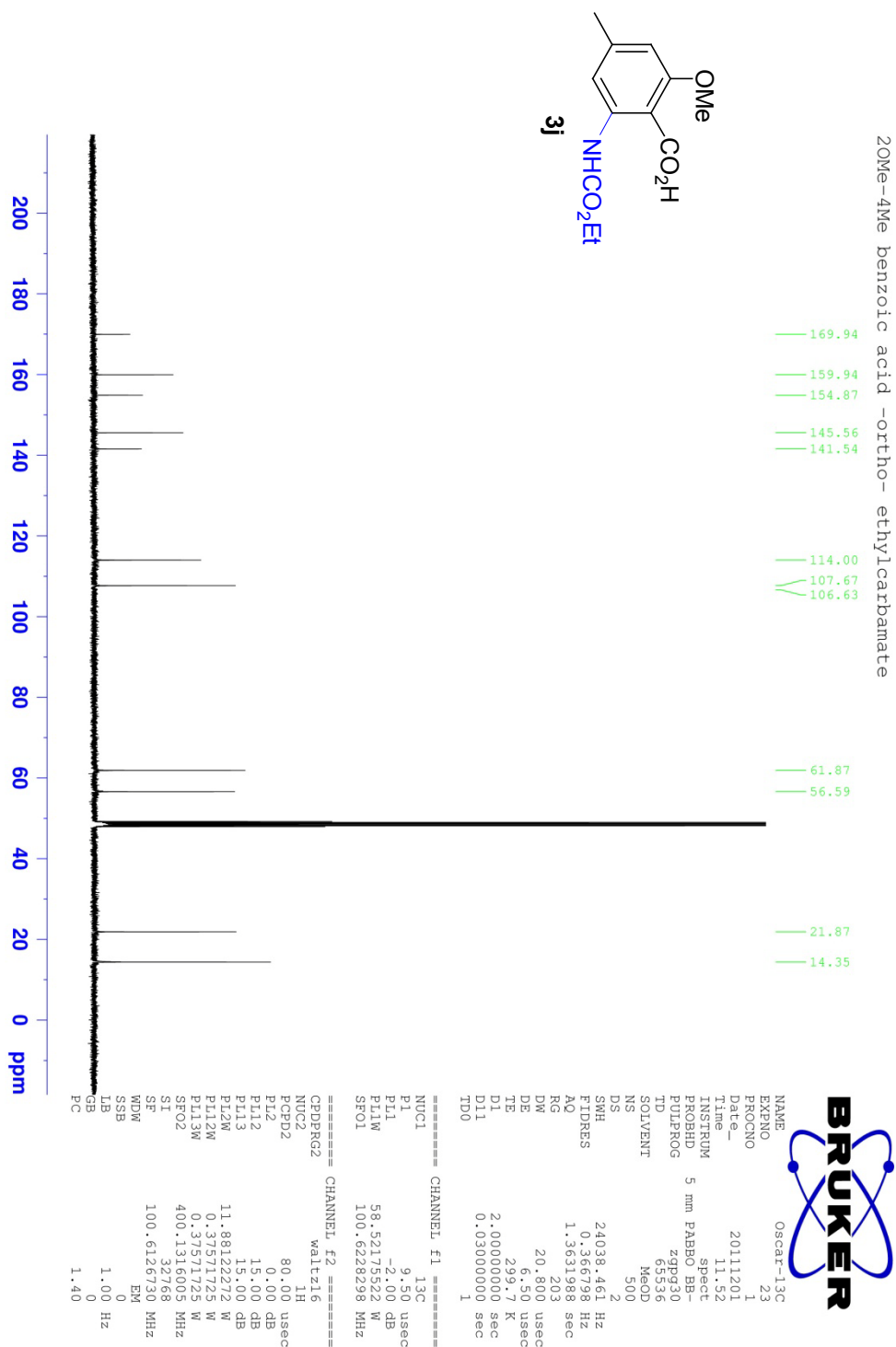


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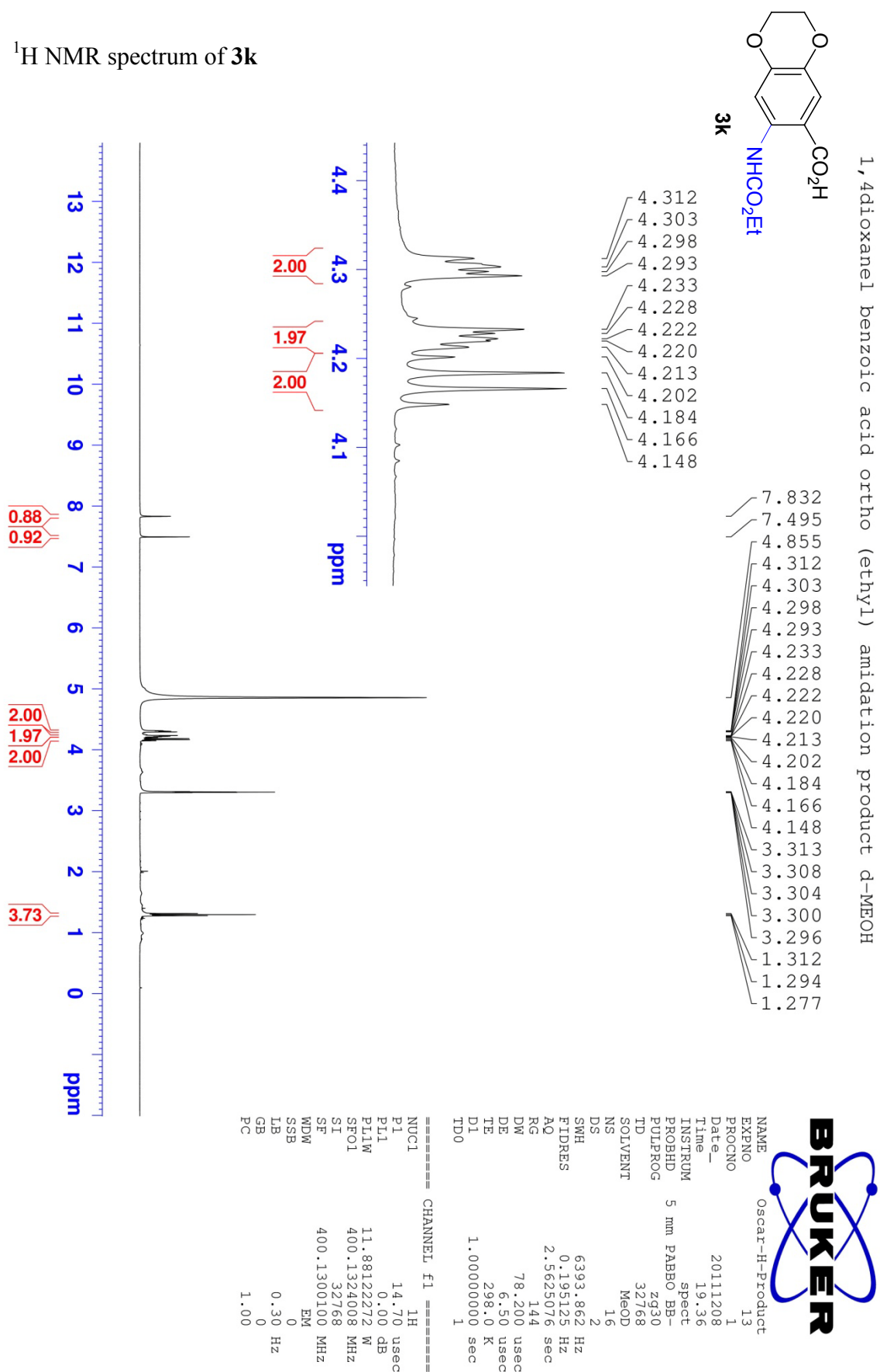
NAME Oscar-H-column
 EXPNO 100
 PROCNO 1
 Date_ 20111201
 Time 11.46
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 7211.539 Hz
 FIDRES 0.220079 Hz
 AQ 2.2719646 sec
 RG 57
 DW 69.333 usec
 DE 6.50 usec
 TE 298.9 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 14.70 usec
 PL1 0.00 dB
 PL1W 11.88122272 W
 SFO1 400.1324008 MHz
 SI 32768
 SF 400.1300100 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

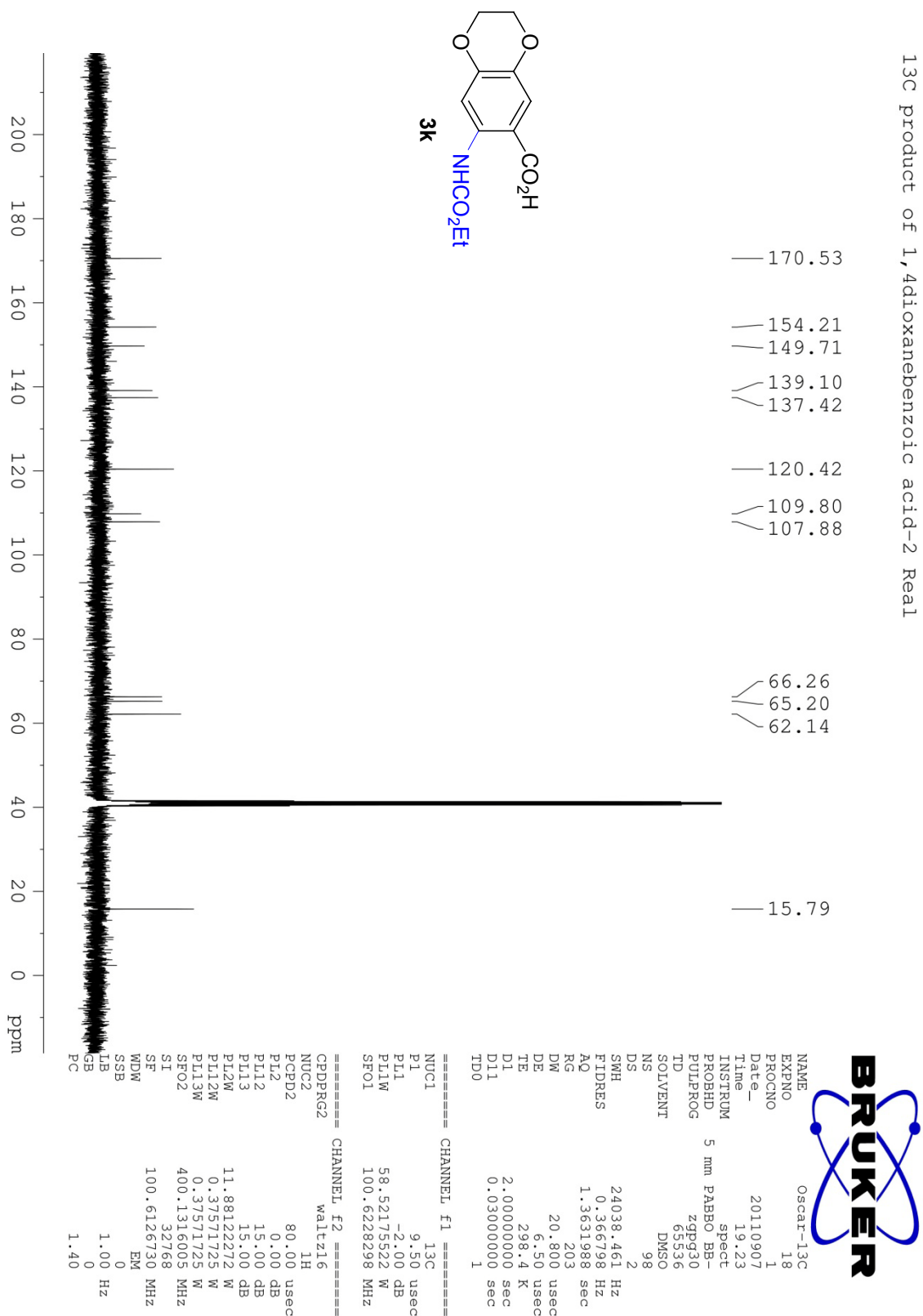
¹³C NMR spectrum of **3j**



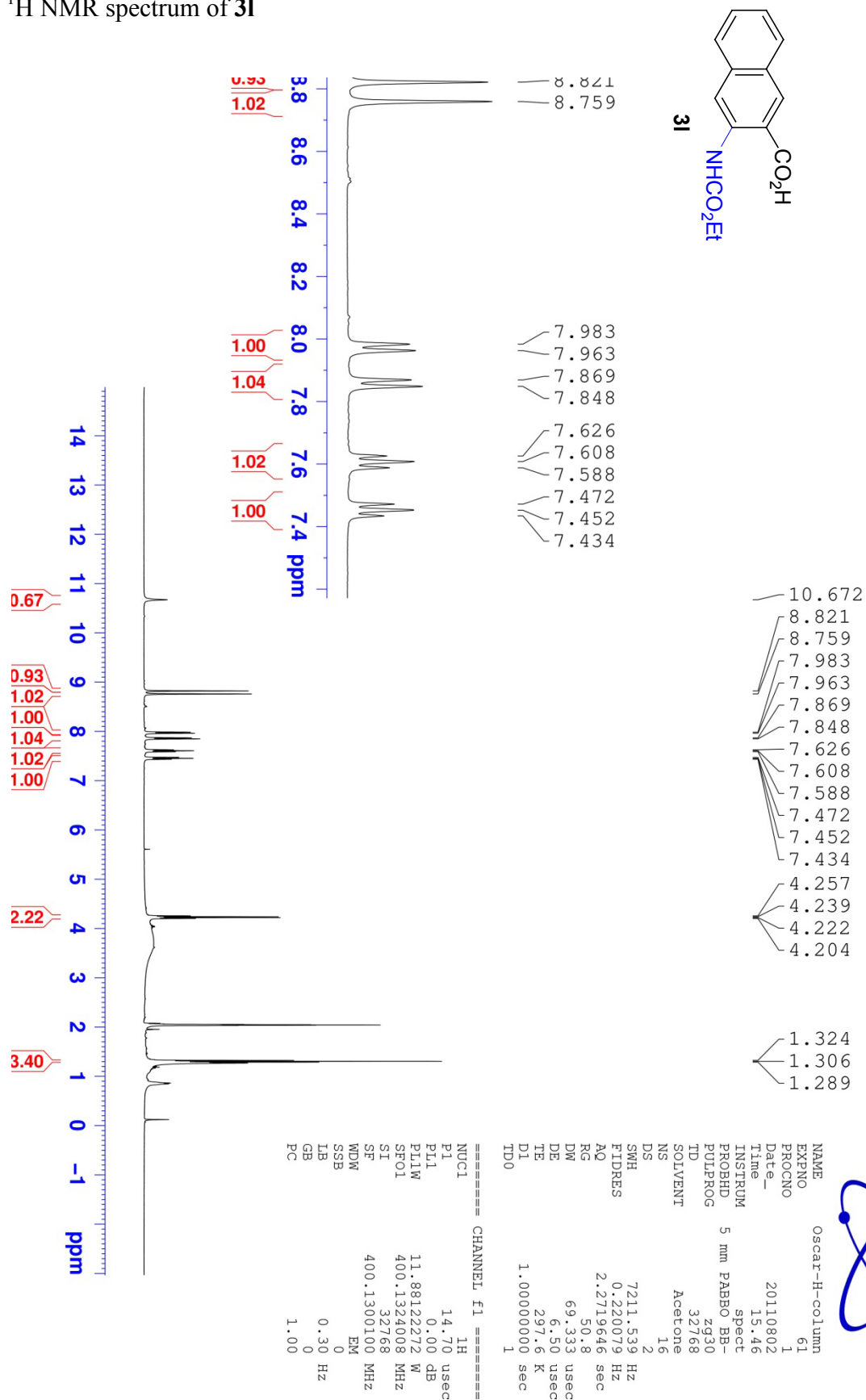
¹H NMR spectrum of **3k**



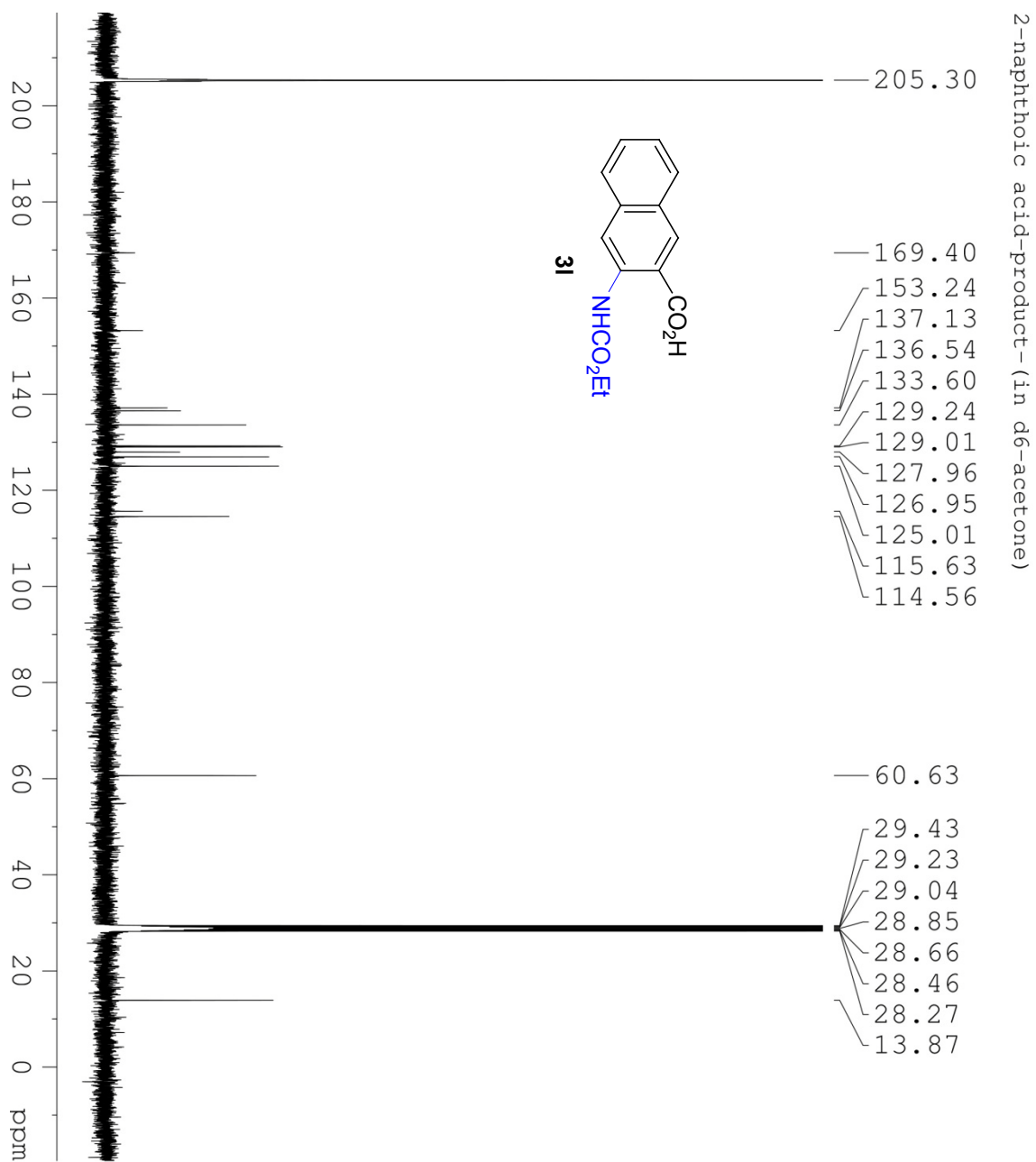
¹³C NMR spectrum of **3k**



¹H NMR spectrum of **31**



¹³C NMR spectrum of **31**



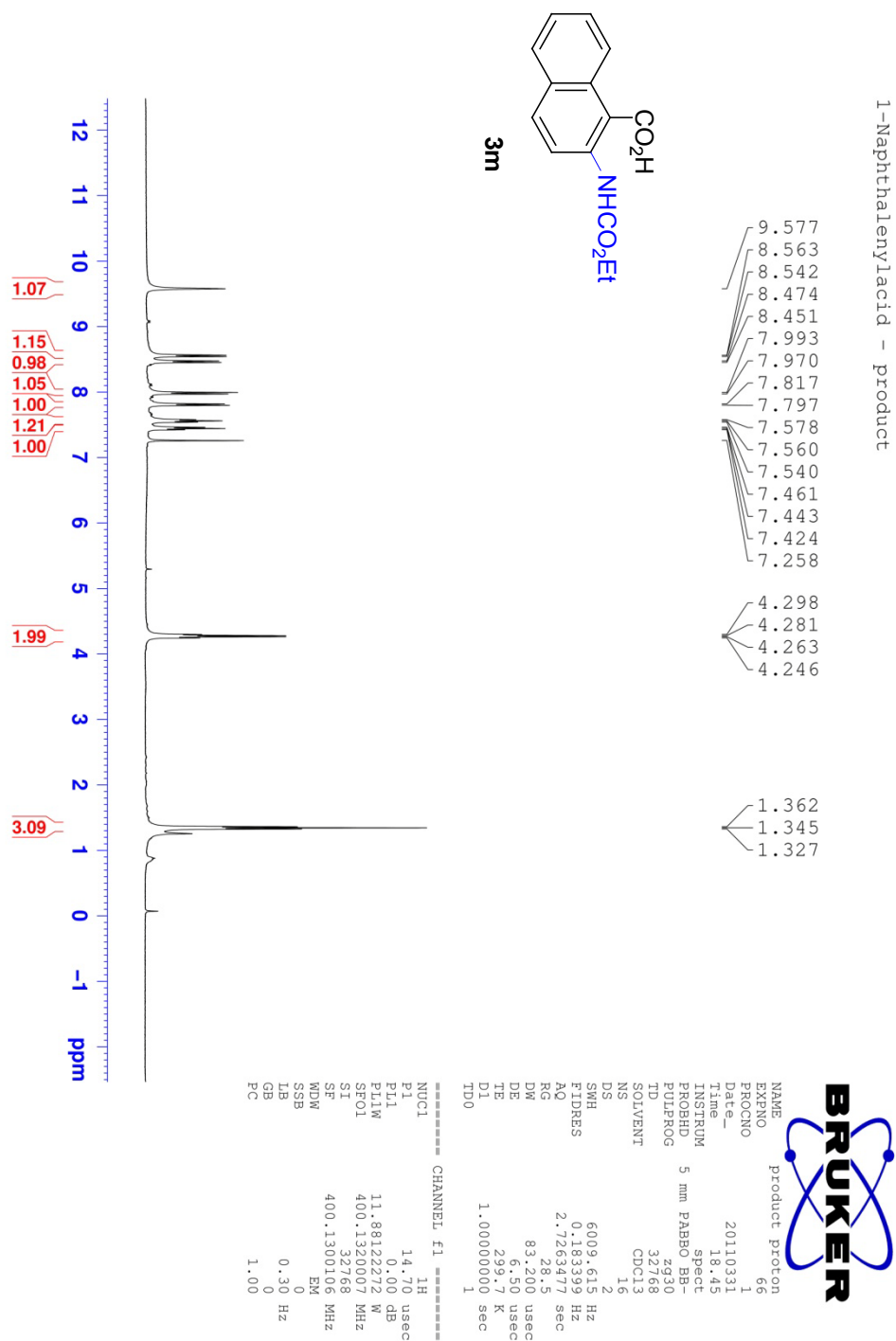
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NAME product Carbon
EXPNO 41
PROCNO 1
Date_ 20110506
Time 15.28
INSTRUM spect
PROBHD 5 mm PABBO B5-
PULPROG zgpg30
TD 65536
SOLVENT Acetone
NS 375
DS 2
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 181
DM 20.800 usec
DE 6.50 usec
TE 300.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

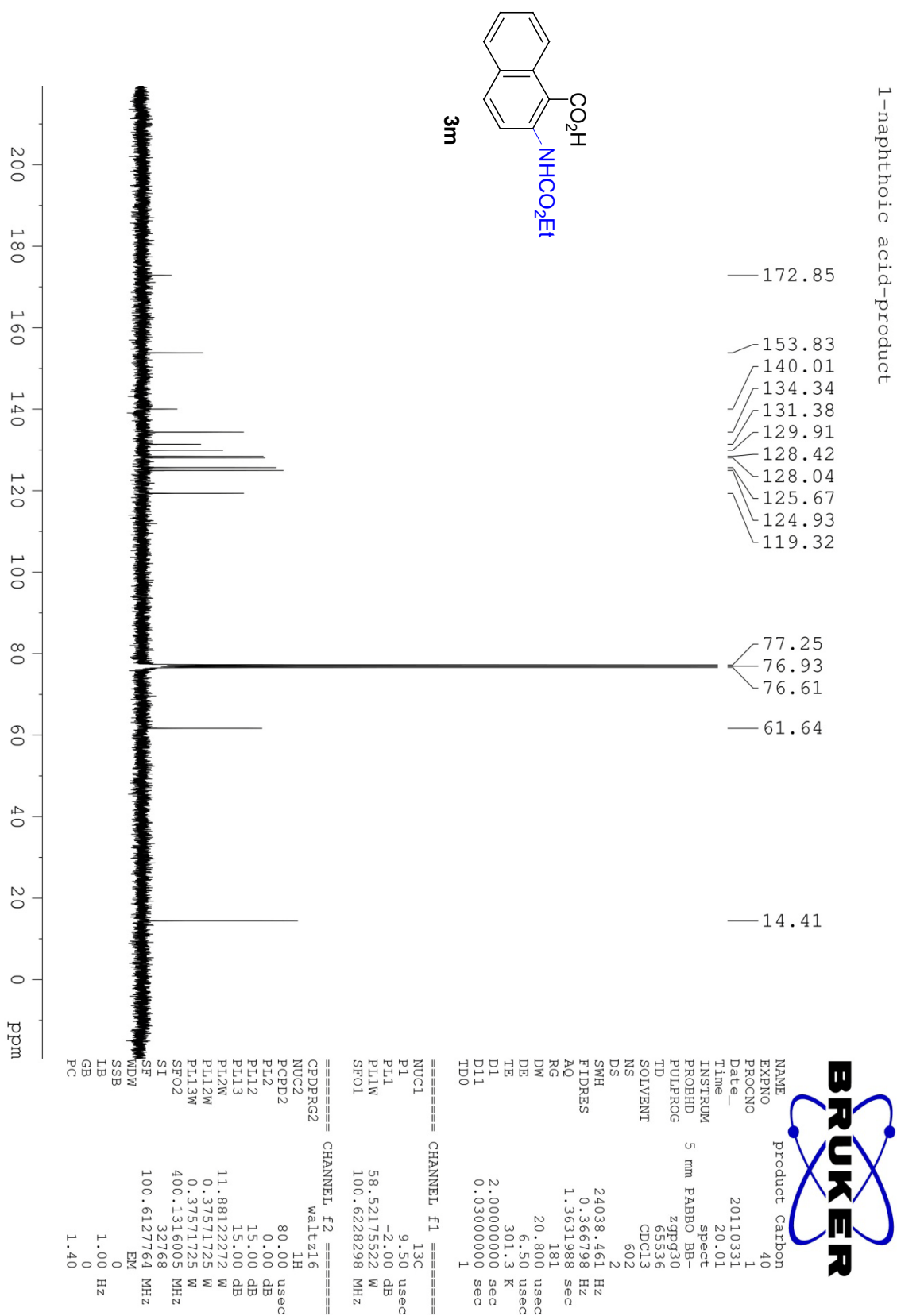
===== CHANNEL F1 =====
NUC1 13C
P1 9.50 usec
PL1 -2.00 dB
PL1W 58.52175522 W
SFO1 100.6228298 MHz

===== CHANNEL F2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 15.00 dB
PL13 15.00 dB
PL2W 11.88122272 W
PL12W 0.37571725 W
PL13W 0.37571725 W
SFO2 400.1316005 MHz
SI 32758
SF 100.6127764 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40
    
```


¹H NMR spectrum of **3m**

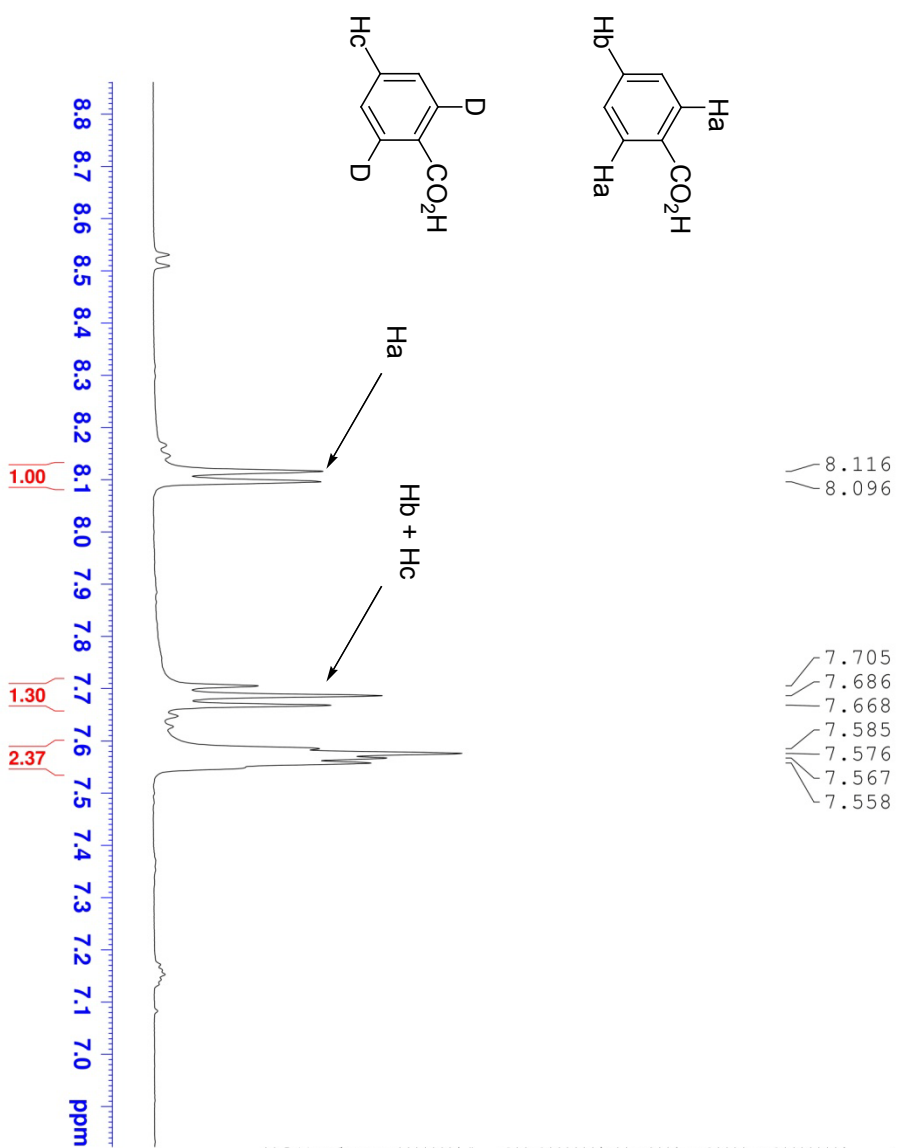


¹³C NMR spectrum of **3m**



KIE NMR data

k_H/k_D 1st run



F0A 386 Pd benzoic acid KIE 30 min one pot-purified-3

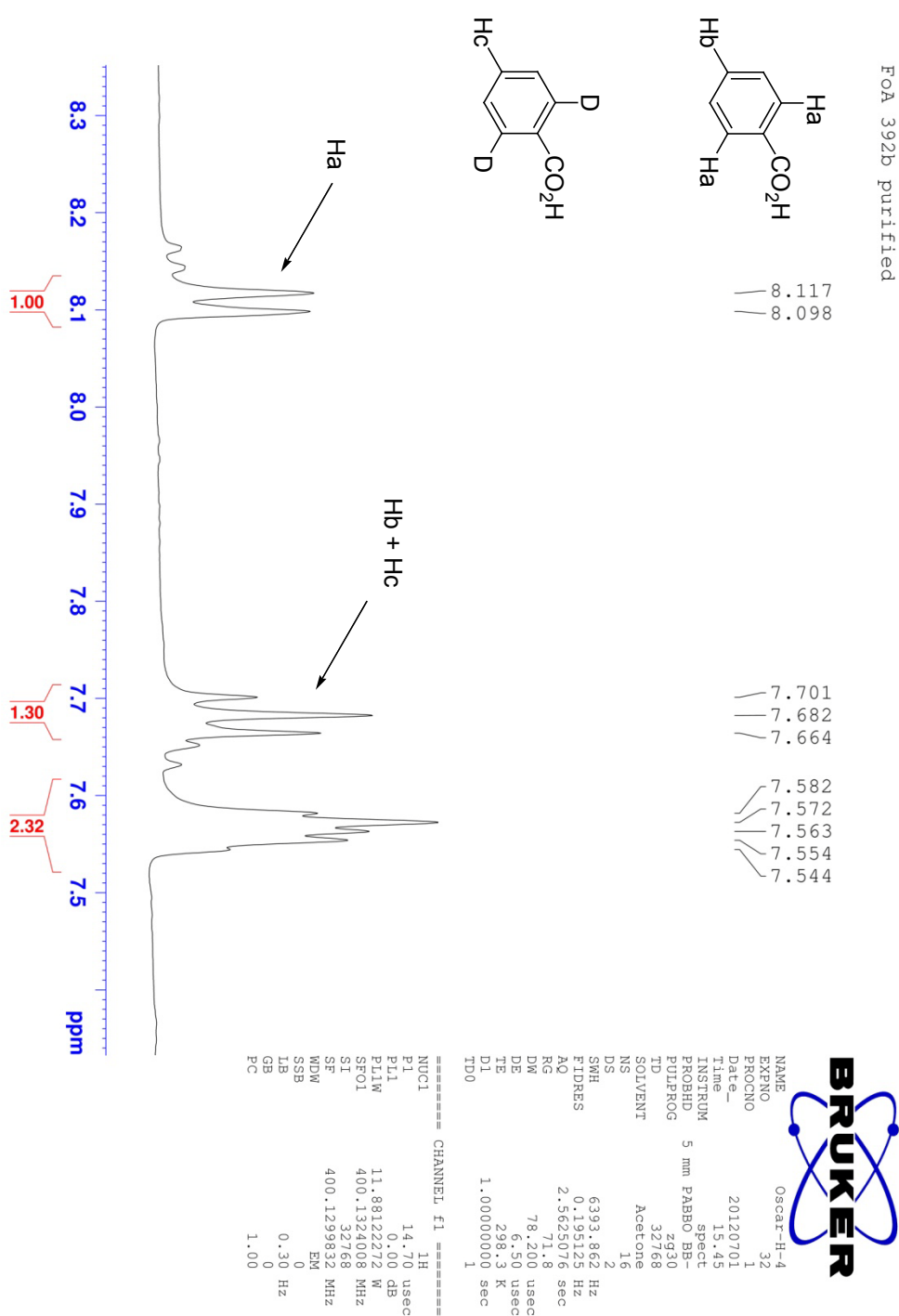
8.116
 8.096
 7.705
 7.686
 7.668
 7.585
 7.576
 7.567
 7.558



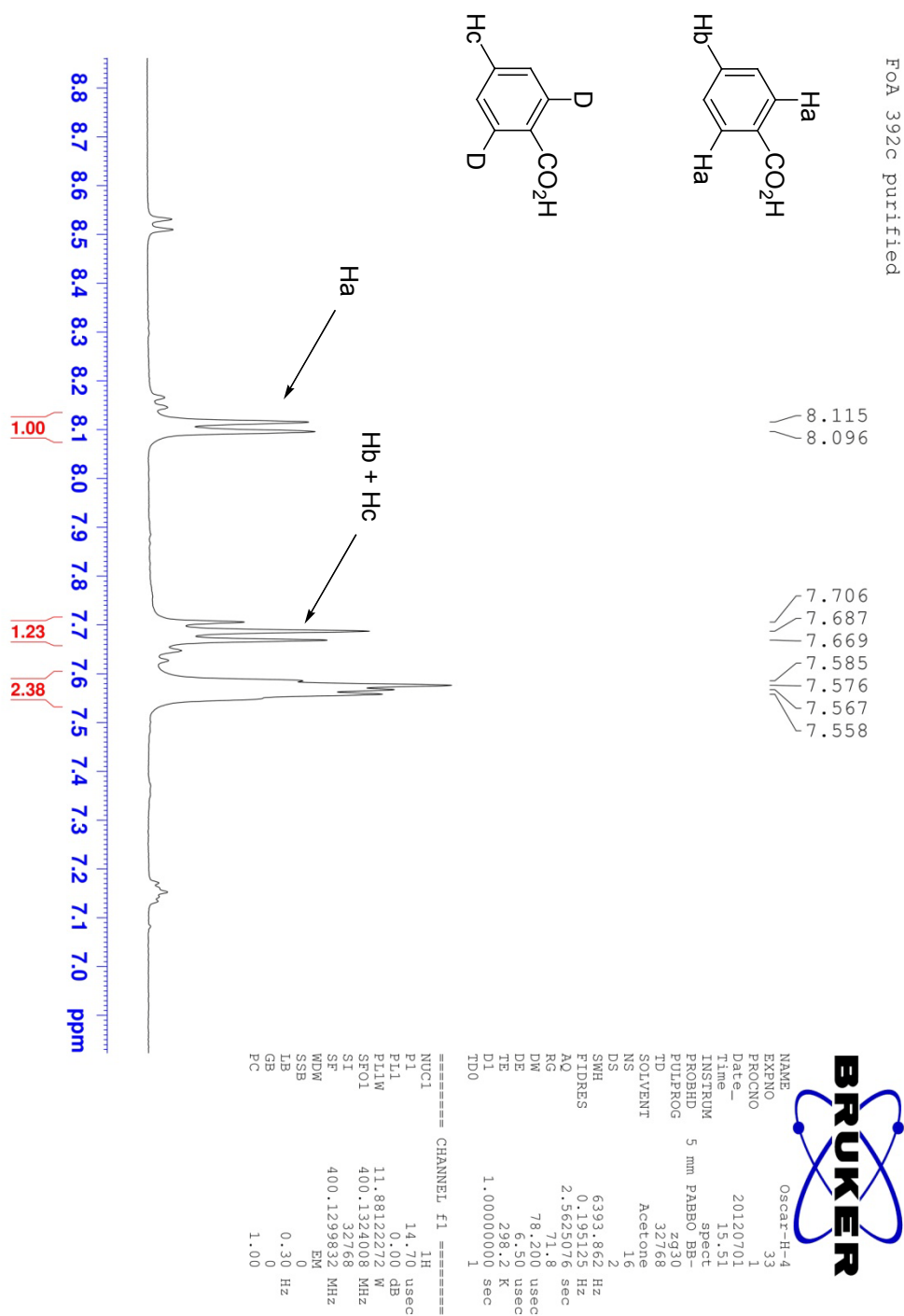
NAME Oscar-H-4
 EXXNO 29
 PROCNO 1
 Date_ 20120629
 Time_ 20.03
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT Acetone
 NS 16
 DS 2
 SMH 6393.862 Hz
 FIDRES 0.193125 Hz
 AQ 2.3623076 sec
 RG 71.8
 DE 78.200 usec
 TE 298.5 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL F1 =====
 NUC1 1H
 P1 14.70 usec
 PL1 0.00 dB
 PL1W 11.88122272 W
 SFO1 400.1324008 MHz
 SF 400.1324008 MHz
 ST 32768
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

k_H/k_D 2nd run

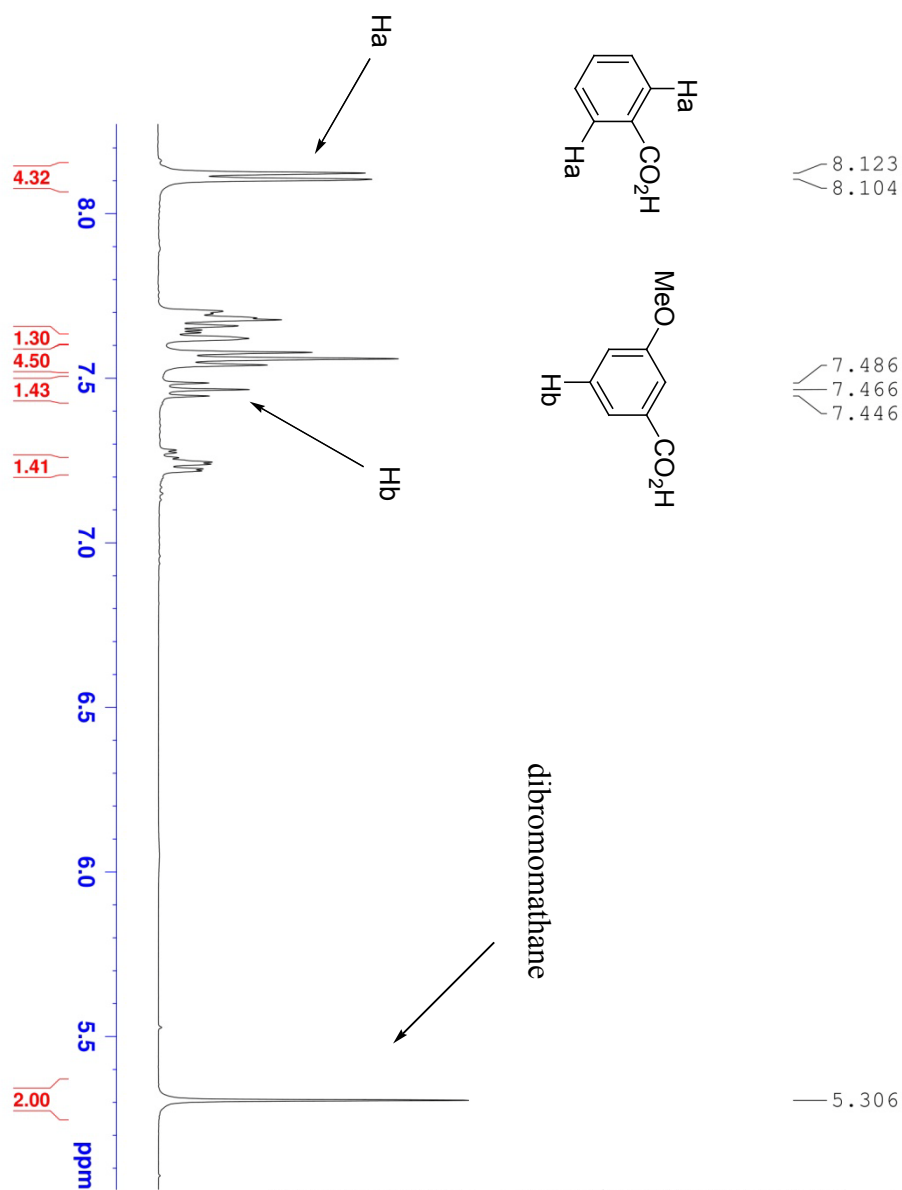


k_H/k_D 3rd run



Experimental Data of the Hammett Correlation Study

k_{OMe}/k_H 1st run

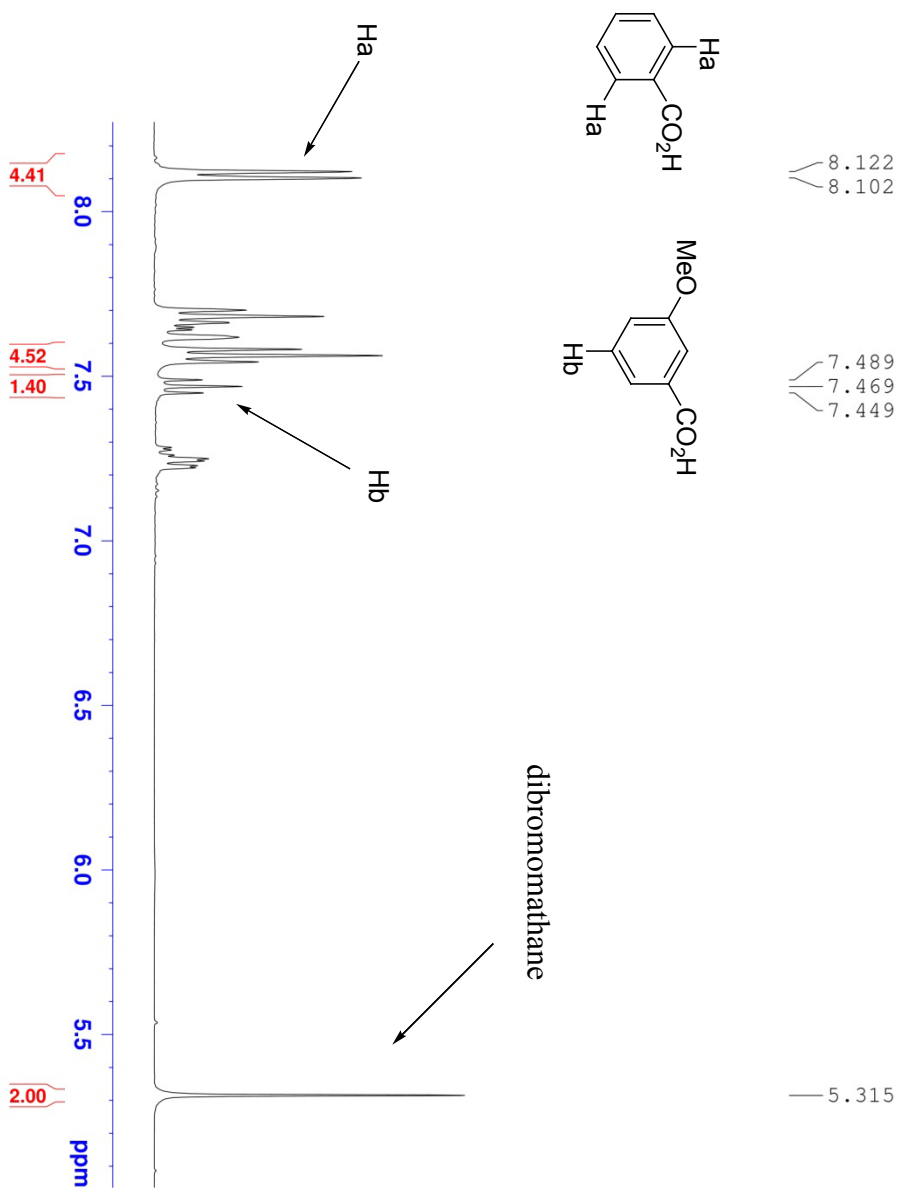


NAME Oscar-H-4
 EXPNO 50
 PROCNO 1
 Date_ 20120709
 Time 15.00
 INSTRUM spect
 PROBD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT Acetone
 NS 16
 DS 2
 SMH 6393.862 Hz
 FIDRES 0.195125 Hz
 AQ 2.5625076 sec
 RG 36
 DW 78.200 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUCL1 1H
 P1 14.70 usec
 PL1 0.00 dB
 PL1W 11.89122272 W
 SFOL 400.1324008 MHz
 SI 32768
 SE 400.1299832 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



kOMe/k_H 2nd run



KOME/KH purified-2

8.122
8.102

7.489
7.469
7.449

5.315

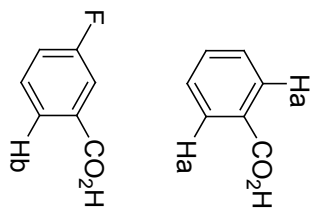


```

NAME          Oscar-H-4
EXPNO         51
PROCNO        1
Date_         20120709
Time_         15.44
INSTRUM       5 mm PABBO BB-
PROBHD        zg30
PULPROG       32768
TD            32768
SOLVENT       Acetone
NS            12
DS            2
SWH           6393.862 Hz
FIDRES        0.195125 Hz
AQ            2.5625076 sec
RG            45.2
DE            78.200 usec
TE            298.2 K
D1            1.00000000 sec
TD0           1

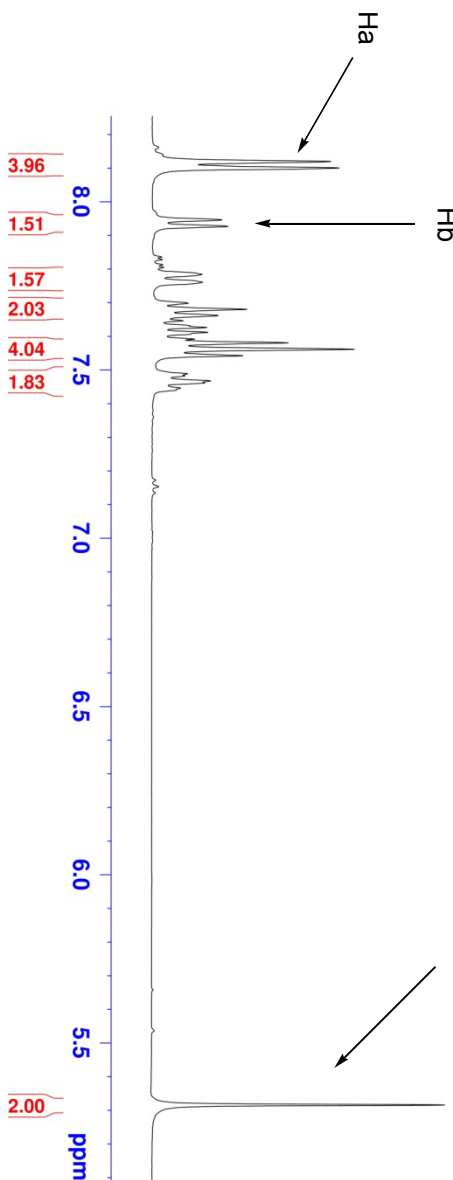
===== CHANNEL f1 =====
NUC1          1H
P1            14.70 usec
PL1           0.00 dB
PL1W          11.88122272 W
SFO1          400.1324008 MHz
SI            32768
SF            400.1239832 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

k_F/k_H 1st run (rejected as conversion <5%)



KF/KH purified 1

8.119
 8.100
 7.946
 7.927



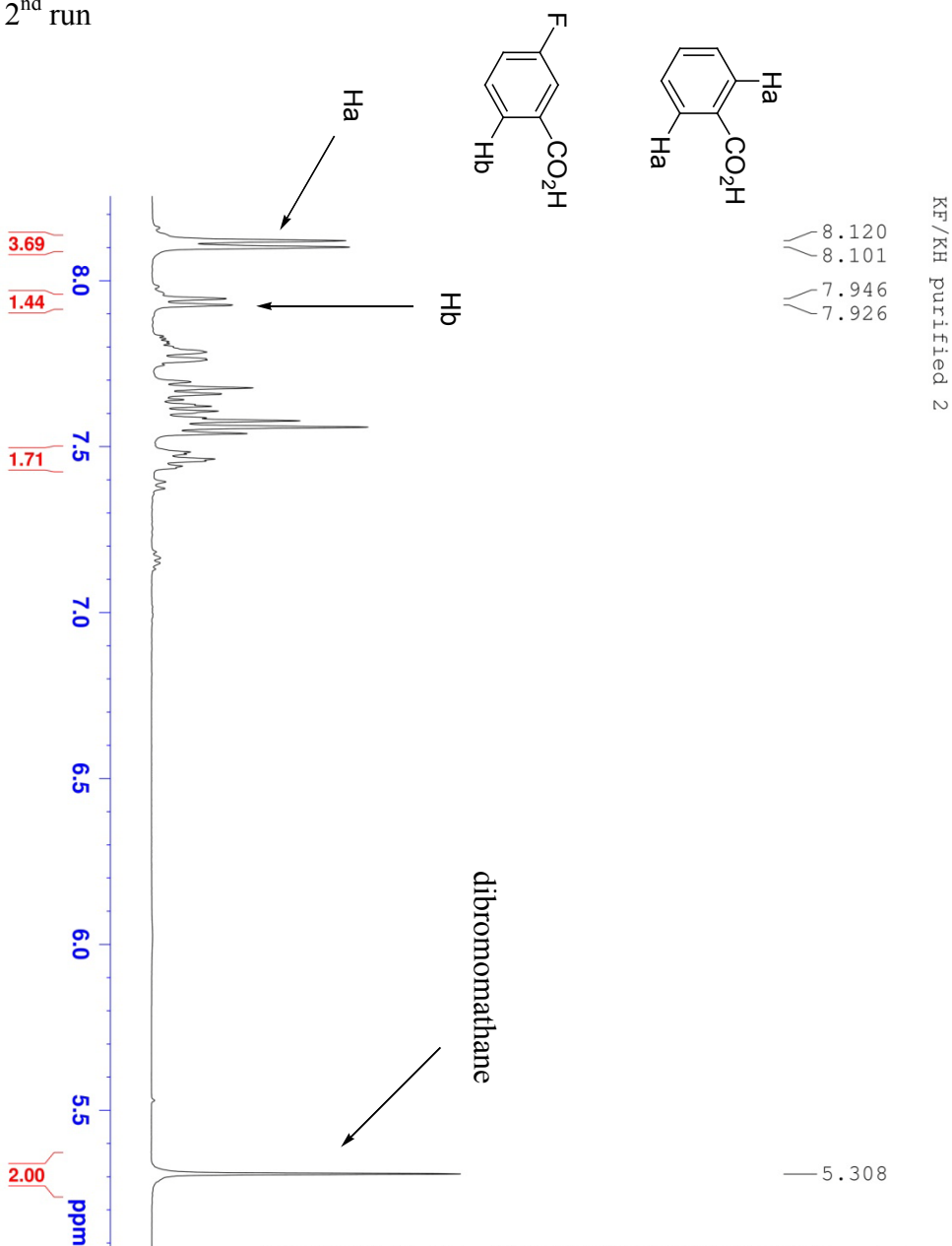
dibromomethane



NAME Oscar-H-4
 EXPNO 52
 PROCNO 1
 Date_ 20120709
 Time 22.02
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT Acetone
 NS 16
 DS 2
 SMH 6393.862 Hz
 FIDRES 0.195125 Hz
 AQ 2.5625076 sec
 RG 45.2
 DW 78.200 usec
 DE 6.50 usec
 TE 298.3 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUCL1 1H
 P1 14.70 usec
 PL1 0.00 dB
 PL1W 11.8812272 W
 SF01 400.1324008 MHz
 SI 32768
 SF 400.1299832 MHz
 WDW EM
 SSB 0
 IB 0.30 Hz
 GB 0
 PC 1.00

k_F/k_H 2nd run



KF/KH purified 2

8.120
8.101
7.946
7.926

5.308

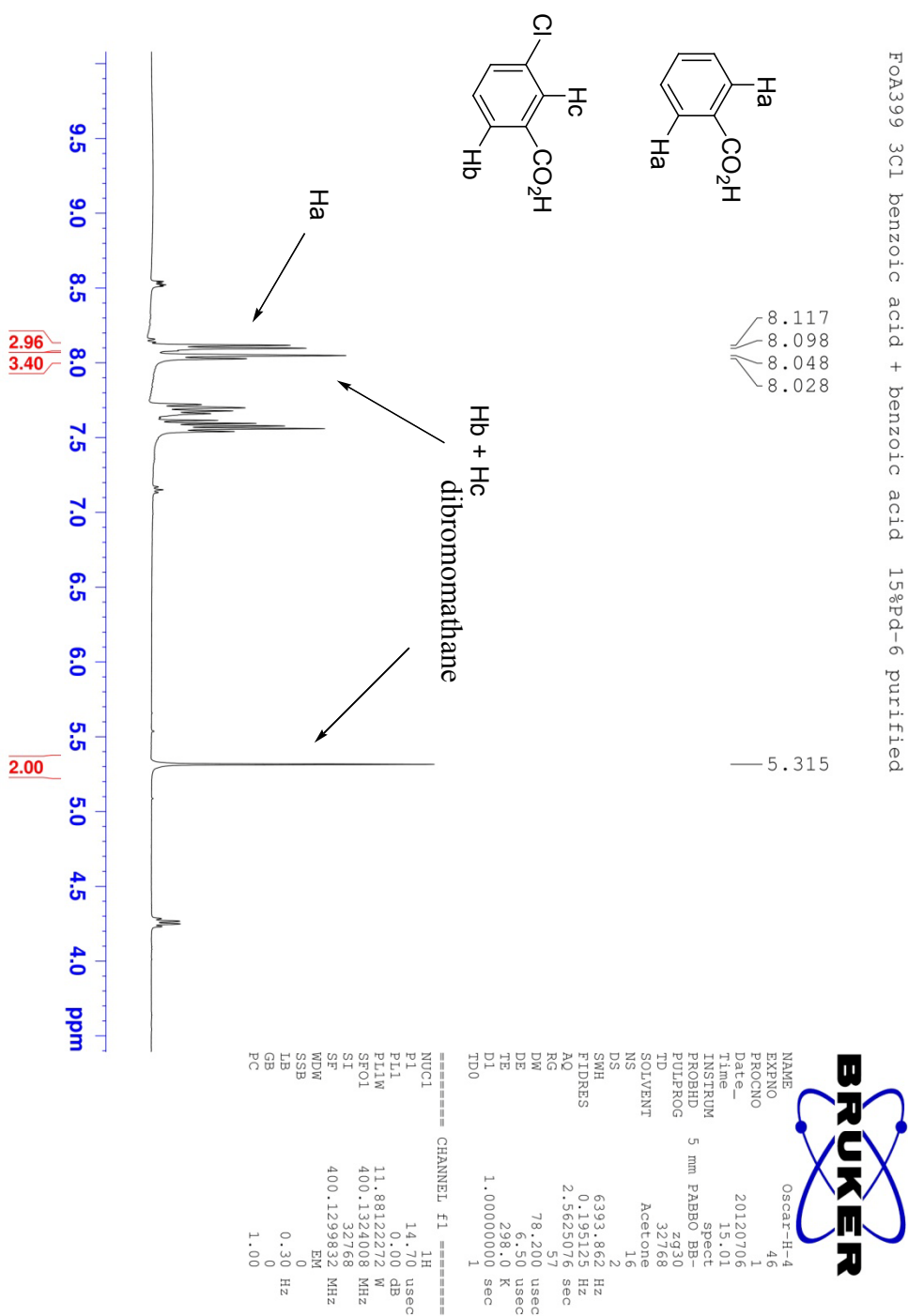


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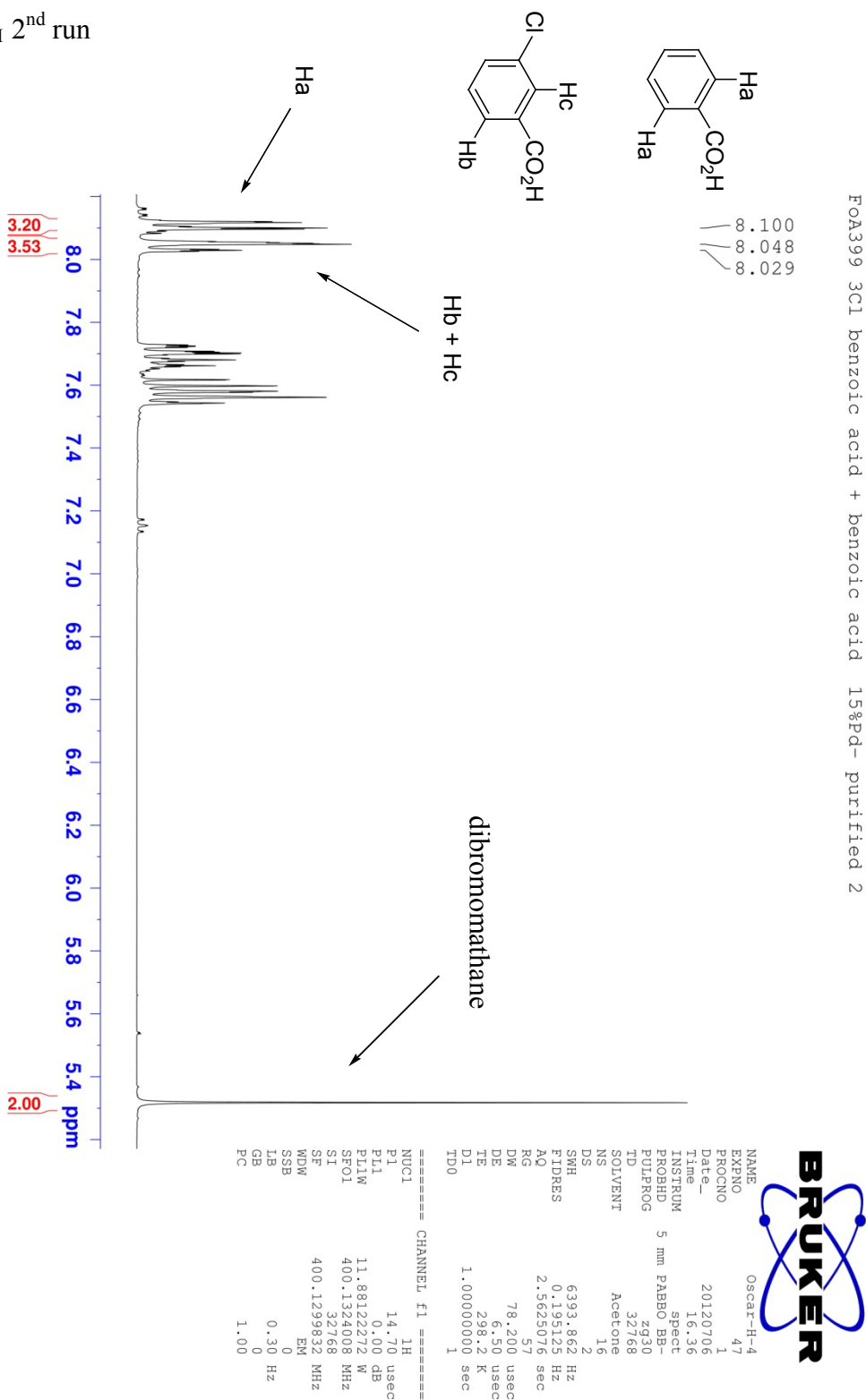
NAME      Oscar-H-4
EXPNO     53
PROCNO    1
Date_     20120709
Time      22.09
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD         32768
SOLVENT   Acetone
NS         16
DS         2
SWH        6393.862 Hz
FIDRES     0.199125 Hz
AQ         2.5629076 sec
RG         45.2
DE         78.200 usec
TE         298.4 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1       1H
P1         14.70 usec
PL1        0.00 dB
PL1W       11.8812272 W
SFO1       400.1324008 MHz
SI         32768
SF         400.1299932 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```

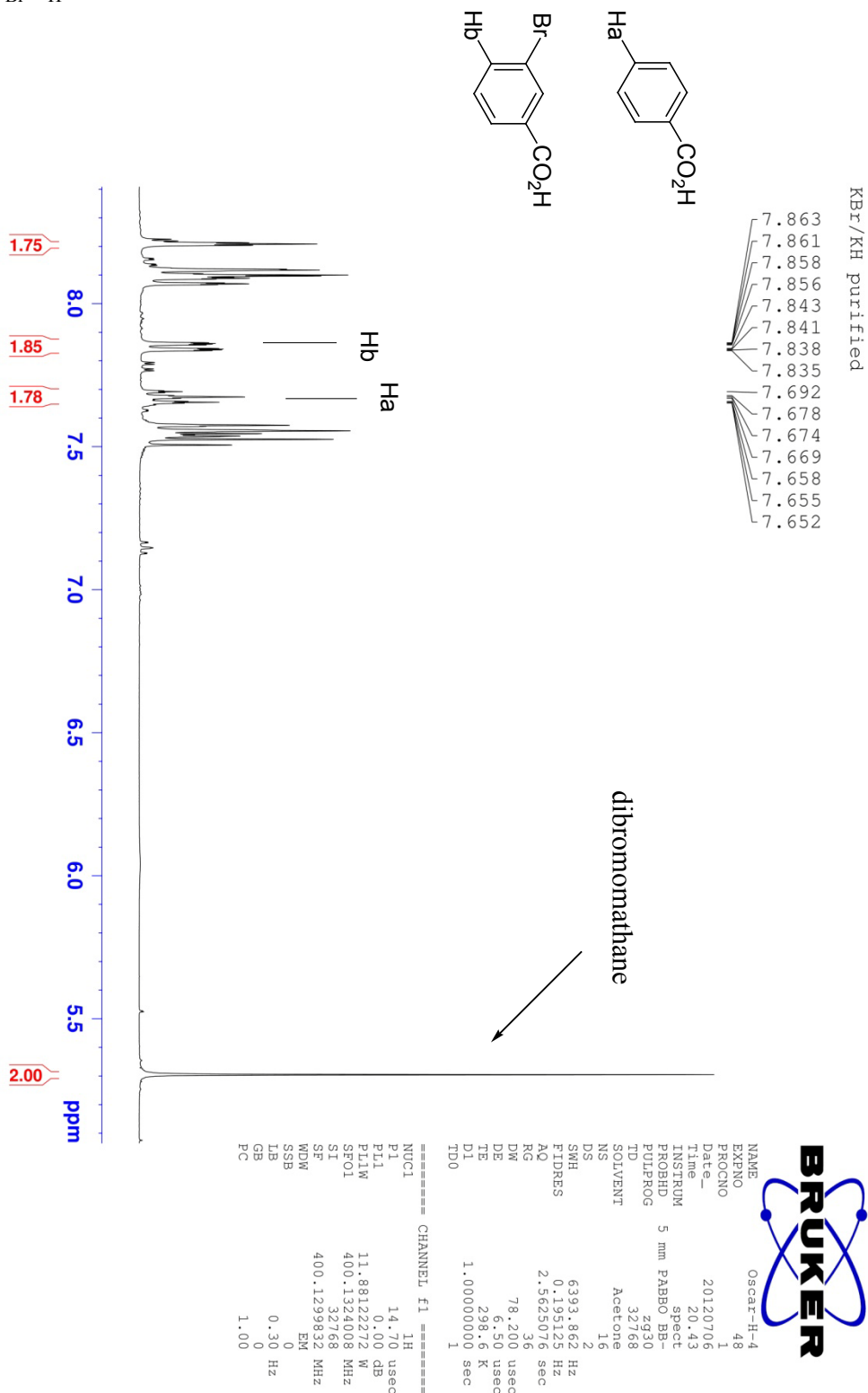
k_{Cl}/k_H 1st run



k_{Cl}/k_H 2nd run



k_{Br}/k_H 1st run



k_{Br}/k_H 2nd run

