1	Supporting Information
2	A One-Step Homogeneous Plasmonic Circular Dichroism
3	<b>Detection of Aqueous Mercury Ions using Nucleic Acid</b>
4	Functionalized Gold Nanorods
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# 1 EXPERIMENTAL SECTION

## 2 Materials

3	Cetyltrimethylammonium bromide (CTAB, 99%) and sodium borohydride, L-ascorbic acid,
4	hydrogen tetrachloroaurate(III) trihydrate (HAuCl <sub>4</sub> ) (99%), and silver nitrate were all purchased
5	from Sigma-Aldrich and used as received. All metal ions were atomic absorption standards. Cu <sup>2+</sup> ,
6	$Hg^{2+}$ , $Cr^{6+}$ , $Fe^{2+}$ , $Ag^+$ , $Pb^{2+}$ , and $Zn^{2+}$ (1000 µg/mL in 1% HNO <sub>3</sub> or 5% HCl) were purchased from
7	the National Institute of Metrology P.R China (Beijing, China). All other reagents were available
8	commercially. Milli-Q ultra-pure water (18.2 M $\Omega$ ) was used in all experiments.
9	The oligonucleotide was synthesized by Sangon Biotechnology Inc. (Shanghai, P. R. China)
10	with this sequence:
11	DNA 1: 5'-SH-AAAAAGTGACCATTTTTGCAGTG-3'
12	DNA 2: 5'-SH-CACTGCTTTTTTGGTCACAAAAAA-3'
13	Instrumentation
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# 23 Synthesis of Gold Nanorods

GNRs with an aspect ratio of about 3 were prepared from a slightly modified seed-mediated
 growth procedure, as described by El-Sayed and Murphy.<sup>29, 30</sup>

Initially, 0.1 mL of a 5 mM HAuCl<sub>4</sub> solution was added to 1 mL of 0.20 M CTAB solution, 3 4 which was kept at a constant temperature of 28.0°C. Immediately, a deep orange color appeared. 5 Then 0.12 mL of freshly prepared 10 mM NaBH<sub>4</sub> solution was quickly added to the solution and 6 mixed by inversion. The solution was rapidly stirred for 2 min, and the color of the solution turned 7 pale brown. Upon seed production, the GNRs were formed. 5 mL of 5 mM HAuCl<sub>4</sub> was added to 8 5 mL of 0.2 M CTAB solution and then 4 mL of water was added; this represents the GNR growth 9 solution. To this mixture, 65  $\mu$ L of 0.1 M ascorbic acid was added, followed by 0.125 mL of 10 10 mM AgNO<sub>3</sub> solution, which was mixed by inversion for about 2 min. The solution became 11 colorless. Finally, 0.05 mL of seed solution was added, with gentle mixing by inversion for about 12 20 s. The GNRs were used after 4 h.

#### 13 Preparation of DNA-Functionalized GNRs.

Briefly, 5 µL 1 mM thiolated modified PEG-5000(SH-PEG) solution and 1000 µL 10 nM 14 15 purified GNR solution were mixed by stirring, then incubated for 8 h whilst shaking. The facets at 16 the ends of the GNRs bound fewer CTAB, so they were able to preferentially conjugate the thiolated modified PEG. The mixture was then collected by centrifugation at  $7500 \times g$  for 15 min. 17 Subsequently, 20 µL 100 µM thiolated modified DNA solution was added to 1000 µL 10 nM 18 PEG-thiolated GNR solution, mixed well, and incubated for another 12 h at room temperature. 19 20 Subsequently, the conjugate was collected by centrifugation at  $7500 \times g$  for 15 min. Finally, the 21 DNA-GNR probe was stored in 5 mM CTAB solution at 4°C for the next step. The GNR probes 22 are denoted as GNR-DNA1and GNR-DNA2.

## 23 Determination procedure

Firstly, 50 μL of GNR-DNA-1 was mixed with 50 μL of GNR-DNA-2 solution with gentle
shaking for 5 min. To samples of the mixed solution, Hg ion solutions at different concentrations
were separately added and mixed well. The concentrations of the Hg ion standard were 0, 0.05, 0.1,
0.5,1, 5 and 10 ng/mL. The solution was then incubated for 1 h at 40 °C to form the assemble
structure via the T-Hg<sup>2+</sup>-T interaction. After reacting, the sample was analyzed by TEM, DLS and
CD. A standard curve was established by plotting the CD signal intensity against the concentration

- of Hg ion standard. The limit of detection (LOD) was calculated as LOD = 3.3SD/S, where SD is
- the standard deviation of the response and S is the slope of the calibration curve.







