Supporting Information

A Bowl-Shaped Organic Host Using Bispyridine Ligands: Selective Encapsulation of Carbonyl Guests in Water

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Materials and methods

NMR: Bruker AVANCE-400 (400 MHz), MALDI-TOF MS: Shimadzu AXIMA-CFR Plus, ESI-TOF MS: Bruker micrOTOF II, FT-IR: JASCO FT/IR-4200, X-ray single crystal structural analysis: Bruker APEXII ULTRA/CCD diffractometer, Elemental analysis: LECO CHNS-932 VTF-900.

Solvents and reagents: TCI Co., Ltd., WAKO Pure Chemical Industries Ltd., KANTO CHEMICAL CO., INC., Sigma-Aldrich Co., and Cambridge Isotope Laboratories, Inc. Bispyridine ligand **2** was synthesized according to previously reported procedures (M. Yoshizawa *et al.*, *J. Am. Chem. Soc.* **2011**, *133*, 11438–11441).



Bispyridine ligand 2 (0.101 g, 0.138 mmol), diiodomethane (2.66 g, 9.93 mmol), and MeCN (70 mL) ware added to a 2-necked 100 mL glass flask containing a magnetic stirring bar under N_2 . The mixture was stirred at 80 °C for 10 d and then concentrated under reduce pressure. The crude product was washed with MeOH, CHCl₃, and acetone to afford bowl **1a** as a red solid (40.7 mg, 20.3 µmol, 29% yield)

¹H NMR (400 MHz, DMSO- d_6 , r.t.): δ 9.97 (d, J = 6.4 Hz, 4H), 9.81 (s, 4H), 8.94 (d, J = 8.0 Hz, 4H), 8.79 (dd, J = 6.4, 8.0 Hz, 4H), 7.86 (d, J = 7.2 Hz, 8H), 7.72 (br, 2H) 7.56-7.47 (m, 24H), 7.30 (s, 2H), 7.16 (br, 2H), 6.84 (s, 2H), 4.17 (t, J = 4.5 Hz, 8H), 3.26 (t, J = 4.5 Hz, 8H), 2.76 (s, 12H).

¹³C NMR (100 MHz, DMSO- d_6 , r.t.): δ 158.5 (C_q), 151.8 (CH), 146.8 (CH), 146.1 (CH), 140.1 (C_q), 136.5 (C_q), 134.4 (CH), 130.1 (CH, 2 x C_q), 127.5 (CH), 127.2 (CH, C_q), 126.2 (CH), 126.0 (CH), 118.8 (C_q), 100.5 (CH), 78.2 (CH₂), 70.6 (CH₂), 68.9 (CH₂), 58.4 (CH₃).

DOSY NMR (400 MHz, DMSO- d_6 , 298 K): $D = 2.40 \text{ x} 10^{-10} \text{ m}^2 \text{ s}^{-1}$.

FT-IR (KBr, cm⁻¹): 3047, 3012, 2929, 1607, 1576, 1506, 1457, 1456, 1387, 1312, 1267, 1194, 1157, 1127, 1102, 1053, 1028, 982, 950, 905, 852.

ESI-TOF MS (CH₃CN): m/z Calcd. 373.4, Found 373.4 $[M - 4I^{-}]^{4+}$.

E.A.: Calcd. for $C_{102}H_{84}O_8N_4I_7$ •1.5H₂O: C, 50.85; H, 3.64; N, 2.33. Found: C, 50.57; H, 3.26; N, 2.36.





Fig. S3a. HH COSY (400 MHz, DMSO-*d*₆, 10 mM, r.t.) spectrum of 1a (aliphatic region).



Fig. S3b. HH COSY (400 MHz, DMSO-d₆, 10 mM, r.t.) spectrum of 1a (aromatic region).



Fig. S4a. NOESY (400 MHz, DMSO-*d*₆, 10 mM, r.t.) spectrum of 1a.



Fig. S4b. NOESY (400 MHz, DMSO-d₆, 10 mM, r.t.) spectrum of 1a (aromatic region).



Fig. S5a. HSQC (400 MHz, DMSO-d₆, 10 mM, r.t.) spectrum of 1a (aliphatic region).



Fig. S5b. HSQC (400 MHz, DMSO- d_6 , 10 mM, r.t.) spectrum of 1a (aromatic region).



Fig. S6. DOSY NMR (400 MHz, DMSO- d_6 , 10 mM, 298 K) spectrum of bowl 1a.



Fig. S7. ESI-TOF MS spectrum of bowl 1a.

Synthesis of Bowl-shaped Host 1b

KY215



Bowl **1a** (250.6 mg, 125.2 μ mol), Amberlite IRA-400 (8.11 g), MeOH (50 mL), and H₂O (20 mL) ware added to a 200 mL glass flask containing a magnetic stirring bar. The mixture was stirred at r.t. for 8 h and then filtrated. The filtrate was concentrated under reduce pressure. An excess amount of HCl aq. was added to the solution. The obatined precipitates were collected and then dissolved in water. An excess amount of acetone was added to the solution to yield a yellow solid. The solid was washed with acetone to afford bowl **1b** as a yellow solid (108.8 mg, 66.52 μ mol, 53% yield).

¹H NMR (400 MHz, D₂O, r.t.): δ 9.89 (br, 4H), 9.48 (br, 4H), 8.96 (br, 4H), 8.71 (br, 4H), 7.93–7.23 (br, 38H), 6.95 (br, 2H), 4.09 (br, 8H), 3.23 (br, 8H), 2.57 (br, 12H).

¹³C NMR (100 MHz, D₂O, r.t.): δ 158.1 (C_q), 153.2 (CH), 145.9 (CH), 141.3 (CH), 136.3 (C_q), 130.6 (C_q), 129.8, 127.5, 126.8, 126.4, 126.0, 124.8, 119.9, 101.2 (CH), 78.5 (CH₂), 70.5 (CH₂), 68.9 (CH₂), 57.8 (CH₃).

DOSY NMR (400 MHz, DMSO, 300 K): $D = 7.4 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$.

FT-IR (KBr, cm⁻¹): 3410, 3060, 2935, 1627, 1605, 1574, 1505, 1443, 1387, 1312, 1268, 1197, 1160, 1126, 1103, 1054, 1031, 982, 906, 852, 817, 771.

ESI-TOF MS (H₂O): m/z 373.4 [**1b** – 4Cl⁻]⁴⁺, 509.5 [**1b** – 3Cl⁻]³⁺, 782.3 [**1b** – 2Cl⁻]²⁺. E.A.: Calcd. for C₁₀₂H₈₄O₈N₄Cl₄•2CHCl₃•3H₂O: C, 64.77; H, 4.81; N, 2.91. Found: C, 64.49; H, 4.72; N, 3.05.



Fig. S9. DOSY NMR (400 MHz, D₂O, 4 mM, 300 K) spectrum of bowl 1b.



Benzoin (3; 2.1 mg, 9.9 μ mol) was added to a D₂O solution (0.3 mL) of bowl **1b** (2.0 mg, 1.2 μ mol) and the mixture was stirred at r.t. for 3 h. After filtration, the quantitative formation of a **1b** \supset **3** compound was confirmed by NMR and ESI-TOF MS analyses.

¹H NMR (400 MHz, D_2O , r.t.): δ 9.93 (d, J = 6.2 Hz, 4H), 9.72 (s, 4H), 8.99 (d, J = 7.0 Hz, 4H), 8.72 (dd, J = 6.2, 7.0 Hz, 4H), 7.85–7.49 (br, 38H), 7.22 (br, 2H), 6.25 (br, 2H), 5.88 (br, 1H), 5.69 (br, 2H), 5.64 (br, 1H), 5.47 (br, 4H) , 5.27 (br, 2H), 4.10 (br, 8H), 3.25 (br, 8H), 2.57 (br, 12H).

¹³C NMR (100 MHz, D₂O, r.t.): δ 197.6 (C=O), 158.2 (C_q), 153.3 (CH), 145.9 (CH), 145.7 (CH), 141.6 (C_q), 137.0 (**3**), 136.4 (C_q), 135.3 (**3**), 131.9 (**3**), 130.6 (CH), 129.8, 128.0, 127.6, 127.1, 126.6, 126.4, 126.0, 125.8, 124.8, 119.9 (C_q), 101.1 (CH), 78.9 (CH₂), 75.4 (**3**), 70.2 (CH₂), 69.2 (CH₂), 57.9 (CH₃).

DOSY NMR (400 MHz, D₂O, 300 K): $D = 5.89 \text{ x } 10^{-10} \text{ m}^2 \text{ s}^{-1}$.

FT-IR (KBr, cm⁻¹): 3409, 3061, 2934, 1683 (C=O), 1627, 1605, 1577, 1505, 1445, 1388, 1313, 1268, 1196, 1160, 1127, 1103, 1056, 1030, 974, 905, 852, 817,770. ESI-TOF MS (H₂O): m/z 426.4 [**1b** \supset **3** – 4Cl⁻]⁴⁺.





Fig. S12a. HH-COSY (400 MHz, D₂O, 4 mM, r.t.) spectrum of 1b⊃3 (aromatic region).



Fig. S12b. HH-COSY (400 MHz, D₂O, 4 mM, r.t.) spectrum of 1b⊃3 (aromatic region).



Fig. S13a. NOESY (400 MHz, D_2O , 4 mM, r.t.) spectrum of $1b\supset 3$.



Fig. S13b. NOESY (400 MHz, D₂O, 4 mM, r.t.) spectrum of 1b⊃3. (host-guest region).



Fig. S14. DOSY (400 MHz, D₂O, 4 mM, 300 K) spectrum of 1b⊃3.

Encapsulation of Benzil (4) by Bowl 1b KY251



Benzil (4; 1.1 mg, 5.2 μ mol) was added to a D₂O solution (0.28 mL) of capsule **1b** (1.9 mg, 1.2 μ mol) and the mixture was stirred at r.t. for 3 h. After filtration, the quantitative formation of a **1b**⊃**4** compound was confirmed by NMR.

¹H NMR (400 MHz, D₂O, r.t.): δ 9.92 (d, *J* = 5.8 Hz, 4H), 9.51 (s, 4H), 9.02 (d, *J* = 7.4 Hz, 4H), 8.74 (dd, *J* = 7.4, 5.8 Hz, 4H), 7.83–7.27 (br, 38H), 6.65 (br, 2H), 6.30 (br, 2H), 6.04 (br, 4H), 5.65 (br, 4H), 4.18 (br, 8H), 3.35 (br, 8H), 2.69 (br, 12H).

DOSY NMR (400 MHz, D₂O, 301 K): $D = 6.5 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$.

FT-IR (KBr, cm⁻¹): 3399, 3062, 2934, 1960, 1680 (C=O), 1662 (C=O), 1628, 1604, 1577, 1505, 1448, 1388, 1313, 1268, 1197, 1160, 1127, 1103, 1055, 1030, 982, 905, 853, 815, 770, 725. ESI-TOF MS (H₂O): m/z 425.9 [**1b** \supset **4** – 4Cl⁻]⁴⁺.



Fig. S16. DOSY (400 MHz, D_2O , 4 mM, 301 K) spectrum of **1b** \supset **4**.



Benzanilide (5; 1.5 mg, 7.6 μ mol) was added to a D₂O solution (0.3 mL) of bowl **1b** (2.1 mg, 1.3 μ mol) and the mixture was stirred at r.t. for 3 h. After filtration, the selective formation of **1b**⊃**5** compound was confirmed by NMR (~80% yield).

¹H NMR (400 MHz, D₂O, r.t.): δ 9.93 (br, 4H), 9.96 (br, 4H), 8.86 (br, 4H), 8.74 (br, 4H), 7.92–7.25 (br, 38H), 6.85 (br, 2H), 6.36 (br), 5.98 (br), 5.57 (br), 4.12 (br, 8H), 3.26 (br, 8H), 2.58 (br, 12H).

DOSY NMR (400 MHz, D₂O, 305 K): $D = 6.6 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$.



Fig. S17a.¹H NMR (400 MHz, D_2O , 4 mM, r.t.) spectrum of **1b** \supset **5**.



Fig. S17b. ¹H NMR (400 MHz, D_2O , 4 mM, 333 K) spectrum of **1b** \supset **5**.







2-Methyl-1,4-naphthoquinone (6; 2.0 mg, 11.6 µmol) was added to a D_2O solution (0.3 mL) of bowl **1b** (2.0 mg, 1.2 µmol) and the mixture was stirred at r.t. for 3 h. After filtration, the quantitative formation of a **1b** \supset (6)₂ compound was confirmed by NMR and ESI-TOF MS analyses.

¹H NMR (400 MHz, D_2O , r.t.): δ 10.01 (d, J = 6.6 Hz, 4H), 9.88 (s, 4H), 9.03 (d, J = 7.6 Hz, 4H), 8.77 (dd, J = 6.6, 7.6 Hz, 4H), 7.95 (br, 4H), 7.79 (d, J = 8.4 Hz, 8H) 7.57–7.41 (br, 24H), 7.19 (s, 2H), 6.30 (br, 2H), 6.18 (br, 2H), 5.99 (br, 2H), 5.85 (br, 4H), 5.16 (br, 2H), 4.11 (br, 8H), 3.30 (br, 8H), 2.66 (br, 12H), 0.86 (br, 6H).

DOSY NMR (400 MHz, D₂O, 300 K): $D = 5.24 \text{ x } 10^{-10} \text{ m}^2 \text{ s}^{-1}$.

FT-IR (KBr, cm⁻¹): 3384, 3068, 2935, 1655 (C=O), 1626, 1606, 1505, 1442, 1387, 1355, 1302, 1266, 1196, 1160, 1030, 982, 941, 903, 770.

ESI-TOF MS (H₂O): m/z 416.4 [**1b** \supset **6** – 4Cl⁻]⁴⁺, 459.4 [**1b** \supset (**6**)₂ – 4Cl⁻]⁴⁺.



Fig. S19. ¹H NMR (400 MHz, D_2O , 4 mM, r.t.) spectrum of **1b** \supset (**6**)₂.



Fig. S20a. HH-COSY (400 MHz, D₂O, 4 mM, r.t.) spectrum of **1b**⊃(**6**)₂.



Fig. S20b. HH-COSY (400 MHz, D₂O, 4 mM, r.t.) spectrum of **1b**⊃(**6**)₂ (aromatic region).



Fig. S21a. NOESY (400 MHz, D₂O, 4 mM, r.t.) spectrum of 1b⊃(6)₂ (aromatic region).







Fig. S21c. NOESY (400 MHz, D_2O , 4 mM, r.t.) spectrum of **1b** \supset (**6**)₂ (host-guest region).



Fig. S22. DOSY (400 MHz, D₂O, 4 mM, 300 K) spectrum of 1b⊃(6)₂.



Fig. S23. FT-IR spectra (KBr) of $1b \supset (6)_2$ and 6.

Encapsulation of *N*-methylphthalimide (7) by Bowl 1b KY243



N-Methylphthalimide (7; 1.0 mg, 6.2 μ mol) was added to a D₂O solution (0.64 mL) of bowl **1b** (4.3 mg, 2.6 μ mol) and the mixture was stirred at r.t. for 3 h. After filtration, the quantitative formation of a **1b** \supset (7)₂ compound was confirmed by NMR and ESI-TOF MS analyses.

¹H NMR (400 MHz, D₂O, r.t.): δ 9.99 (d, *J* = 6.2 Hz, 4H), 9.77 (s, 4H), 9.04 (d, *J* = 7.8 Hz, 4H), 8.77(dd, *J* = 6.2, 7.8 Hz, 4H), 7.91–7.46 (br, 36H), 7.27 (s, 2H), 6.71 (s, 2H), 6.16 (br, 4H), 5.40 (br, 4H), 4.17 (br, 8H), 3.34 (s, 8H), 2.69 (s, 12H), 1.74 (br, 6H). DOSY NMR (400 MHz, D₂O, 304 K): *D* = 6.9 x 10⁻¹⁰ m² s⁻¹. ESI-TOF MS (H₂O): *m*/*z* 413.7 [**1**b \supset 7 – 4Cl⁻]⁴⁺, 453.9 [**1**b \supset (7)₂ – 4Cl⁻]⁴⁺.



Fig. S25. DOSY (400 MHz, D₂O, 4 mM, 300 K) spectrum of 1b⊃(7)₂.

Table.	S1.	Crystal	data	and	structure	refinement	t for	1a
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Identification code	KY84			
Empirical formula	C225 H210 I14.45 N8 O23			
Formula weight	5227.71			
Temperature	90 K			
Wavelength	0.71073 Å			
Crystal system	Triclinic			
Space group	<i>P</i> -1			
Unit cell dimensions	<i>a</i> = 23.135(3) Å	$\alpha = 102.793(2)^{\circ}$		
	<i>b</i> = 23.325(3) Å	$\beta = 91.669(2)^{\circ}$		
	c = 23.341(3) Å	$\gamma = 90.170(2)^\circ$		
Volume	12277.(3) Å ³			
Z	2			
Density (calculated)	1.414 Mg/m ³			
Absorption coefficient	1.879 mm ⁻¹			
F(000)	5132			
Crystal size	0.27 x 0.26 x 0.01 mm ³			
Theta range for data collection	2.04 to 25.03°.			
Index ranges	-27<=h<=27, -13<=k<=27, -27<=l<= 27			
Reflections collected	58449			
Independent reflections	42423 [R(int) = 0.0510]			
Completeness to theta = 25.03°	97.8 %			
Absorption correction	Semi-empirical from equivalents			
Max. and min. transmission	0.8344 and 0.7051			
Refinement method	Full-matrix least-squares on F ²			
Data / restraints / parameters	42423 / 633 / 2503			
Goodness-of-fit on F ²	1.319			
Final R indices [I>2sigma(I)]	$R_1 = 0.1302, wR_2 = 0.3669$			
R indices (all data)	$R_1 = 0.1711, wR_2 = 0.3907$			
Largest diff. peak and hole	5.669 and -2.438 e.Å ⁻³			

The supplementary crystallographic data of bowl **1a** can be obtained free of charge (under CCDC 893651) by containing the Cambridge Crystallographic Data Centre via www.ccdc.cam. ac.uk/data_request/cif.



Fig. S26. ORTEP drawing of bowl 1a.



Fig. S27. Hydrogen bonding interactions between the carbonyl oxygen atoms of acetones and α -hydrogen atoms of the bispyridinium moieties of bowl 1a.



Fig. S28. Crystal structures of bowl 1a in bowl (left) and tube-shape (right) conformations.



Fig. S29. Torsion and bite angles of the aromatic rings of 1a in the crystal structures.





Fig. S30. Optimized structures of the bowl- and tube-shaped conformations of $1 (R = CH_3)$.



Fig. S31. ¹H NMR (400 MHz, D₂O:acetone- d_6 (10:1), 4 mM, r.t.) spectrum of **1b** \supset **3**.



Fig. S32. ¹H NMR (400 MHz, D₂O:acetone- d_6 (10:1), 4 mM, r.t.) spectrum of **1b** \supset (**6**)₂.