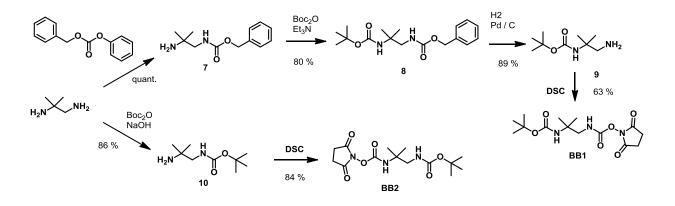
Supporting information

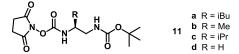
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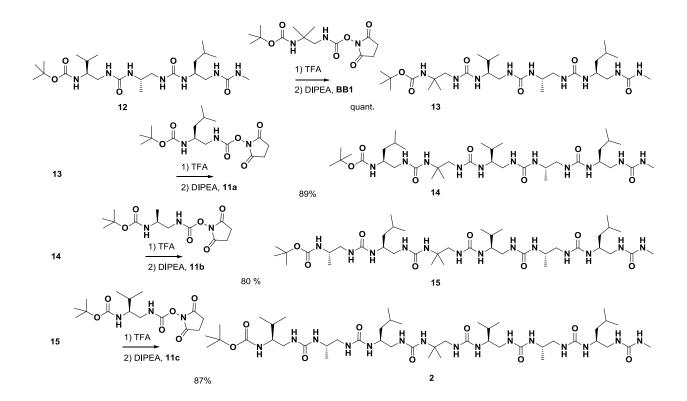
Synthesis of oligoureas



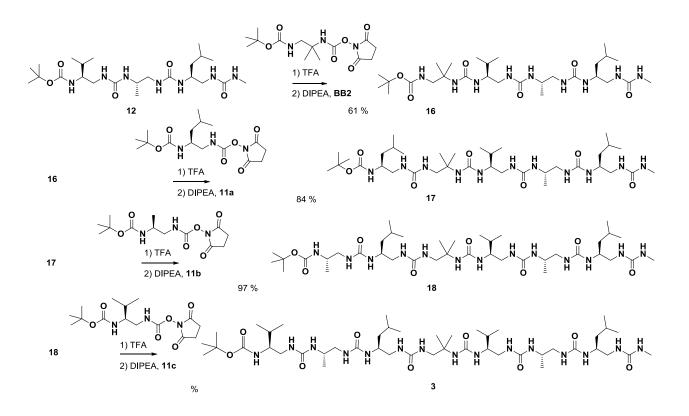
Scheme S1: Synthesis of activated monomers BB1 and BB2



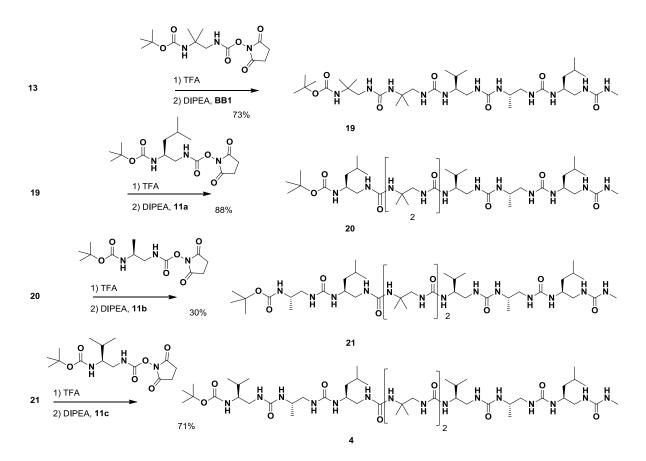
Scheme S2: Activated monomers 11a, 11b, 11c and 11d



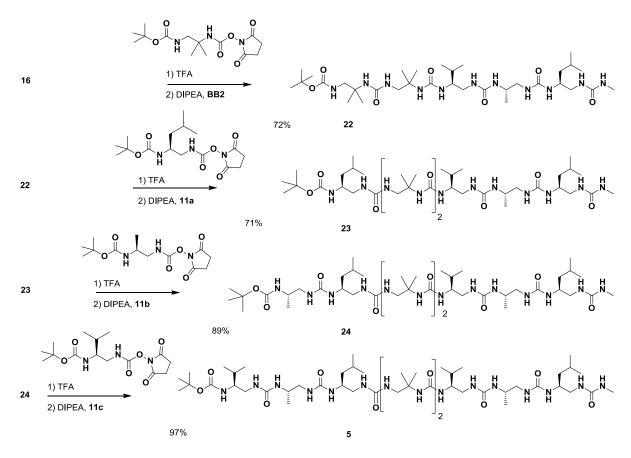
Scheme S3: Synthesis of oligomer 2



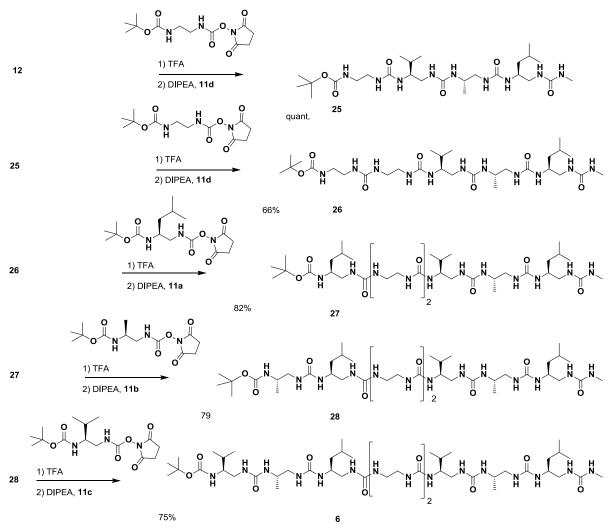
Scheme S4: Synthesis of oligomer 3



Scheme S5: Synthesis of oligomer 4



Scheme S6: Synthesis of oligomer 5



Scheme S7: Synthesis of oligomer 6

General. Thin layer chromatography (TLC) was performed on silica gel 60 F254 (Merk) with detection by UV light and charring with 1% w/w ninhydrin in ethanol followed by heating. Flash column chromatography was carried out on silica gel (40-63 μ m). ¹H NMR and ¹³C NMR spectra were recorded on an Avance II NMR spectrometer (Bruker Biospin) with a vertical 7.05T narrow-bore/ultrashield magnet operating at 300 MHz for 1H observation and 75 MHz for 13C observation by means of a 5-mm direct BBO 1H/19F_XBB_H probe with Z gradient capabilities. Chemical shifts are reported in parts per million (ppm) relative to the 1H residual signal of the deuterated solvent used. ¹H NMR splitting patterns with observed first-order coupling are designated as singlet (s), doublet (d), triplet (t), or quartet (q). Coupling constants (*J*) are reported in hertz. ESI-MS analyses were carried out on a ThermoElectron LCQ Advantage spectrometer equipped with an ion trap mass analyzer and coupled with a ThermoElectron Surveyor HPLC system.

Compound 7¹, activated monomers 11 (a R = *i*Bu; b R = Me; c R = *i*Pr; d R= H)² and oligomer 1 and $12^{3, 4}$ were prepared using a previously described procedure.

(2-Benzyloxycarbonylamino-1,1-dimethyl-ethyl)-carbamic acid tert-butyl ester, (8)

To a stirred solution of **7** (3 g, 13.5 mmol) in CH₂Cl₂ (25 mL), triethylamine (2.3 mL, 16.2 mmol) and di-*tert*-butyl dicarbonate (3.54 g, 16.2 mmol) were successively added and the reaction mixture was allowed to stir at room temperature overnight. CH₂Cl₂ was concentrated under reduced pressure and the resulting crude mixture was purified by flash chromatography on silica gel (CH₂Cl₂-MeOH (v/v), 95:5). Compound **7** was obtained as a colorless oil (4.03 g, 92% yield). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.25$ (s, 6H), 1.42 (s, 9H), 3.40 (d, J = 6.4 Hz, 2H), 4.57 (br s, 1H), 5.11 (s, 2H), 5.36 (br s, 1H), 7.30-7.40 (m, 5H). ¹³C NMR (75 MHz, CDCl₃): $\delta = 24.8$, 28.1, 49.3, 53.0, 66.4, 78.8, 127.7, 128.1, 136.3, 154.6, 156.8 ESI-MS (MW 322.19): m/z 345.1 [M + Na]⁺

(2-Amino-1,1-dimethyl-ethyl)-carbamic acid tert-butyl ester, (9)

To a stirred solution of **8** (3.88 g, 12.0 mmol) in EtOH (30 mL) at room temperature, 10% Pd/C (390 mg) was added and the reaction mixture was stirred under H₂ gas atmosphere for 20 h. The reaction mixture was then filtered over glass microfiber filter, the filtrate was concentrated under reduced pressure and dried under vacuum to afford amine **9** as a colorless oil (2.02 g, 89% yield), which was used for the next step without further purification. ¹H **NMR**: (300 MHz, CDCl₃): $\delta = 1.24$ (s, 6H), 1.42 (s, 9H), 1.53 (s, 2H), 2.74 (s, 2H), 4.79 (br

¹ M. Pittelkow, R. Lewinsky, J. B. Christensen, *Synthesis* **2002**, 2195-2202.

² G. Guichard, V. Semetey, C. Didierjean, A. Aubry, J.-P. Briand, M. Rodriguez, J. Org. Chem. 1999, 64, 8702.

³ L.Fischer, P. Claudon, N. Pendem, E. Miclet, C. Didierjean, E. Ennifar, G. Guichard, *Angew. Chem. Int.Ed.* **2010**, *49*, 1067

⁴ J.Fremaux, L. Fischer, T. Arbogast, B. Kauffmann, G. Guichard, *Angew. Chem. Int.Ed.* **2011**, *50*, 11382.

s, 1H). ¹³C NMR: (75 MHz, CDCl₃): δ = 24.6, 28.0, 49.6, 52.4, 78.6, 154.7 ESI-MS (Mw 188.15): m/z 188.9 [M+H]⁺

(2-*tert*-Butoxycarbonylamino-2-methyl-propyl)-carbamic acid 2,5-dioxo-pyrrolidin-1-yl ester (BB1)

To a stirred solution of **9** (1.8 g, 9.6 mmol) in dry CH₂Cl₂ (30 mL), a solution of *N*,*N*'-disuccinimidyl carbonate (2.94 g, 11.5 mmol) in dry CH₂Cl₂ (30 mL) was added dropwise and the reaction mixture was stirred at room temperature overnight. CH₂Cl₂ was removed under reduced pressure, the residue was dissolved in EtOAc (50 mL) and washed with an aqueous solution of 1M KHSO₄ (3 × 20 mL). The organic phase was dried over MgSO₄, filtered and concentrated under vacuum to afford compound **BB1** as a white solid (2.1 g, 67% yield). ¹**H NMR:** (300 MHz, CDCl₃): δ = 1.27 (s, 6H), 1.43 (s, 9H), 2.81 (s, 4H), 3.46 (d, *J* = 6.1 Hz, 2H), 4.52 (br s, 1H), 6.50 (br s, 1H). ¹³C NMR: (75 MHz, CDCl₃): δ = 25.4, 25.6, 28.3, 50.3, 53.0, 79.9, 151.9, 155.2, 169.8 **HRMS** calcd. for C₁₄H₂₃N₃O₆ Na 352.1479 [M+Na]⁺, found 352.1477, [2M+Na]⁺, found 681.3060

(2-Amino-2-methyl-propyl)-carbamic acid tert-butyl ester, (10)

To a solution of 1,2-diamino-2-methylpropane (1 eq) and sodium hydroxide (0.8 eq) in dioxane/water (1:1 v/v) at 0°C was added a solution of di-*tert*-butyl dicarbonate (1 eq) in dioxane dropwise. The reaction was stirred at 0°C for 3.5 h and at room temperature for 2.5 h. The reaction was concentrated under reduced pressure then water added. The product was extracted into CH₂Cl₂, the organic layers combined, dried over MgSO₄ and the solvent removed under reduced pressure. This afforded **6** as a white solid (77%).¹**H** NMR: (500 MHz, CDCl₃) $\delta = 4.95$ (br s, 1H, NH), 3.00 (d, J = 6.3 Hz, 2H, CH₂N), 1.44 (s, 9H, Boc), 1.12 (br s, 2H, NH₂), 1.08 (s, 6H, CH₃). ¹³C NMR (101 MHz, CDCl₃) $\delta = 156.5$, 79.1, 52.2, 50.1, 28.4, 28.2 **ESI-MS** (Mw 189.15): m/z 189.16 [M + Na]⁺

(2-*tert*-Butoxycarbonylamino-1,1-dimethyl-ethyl)-carbamic acid 2,5-dioxo-pyrrolidin-1-yl ester, (BB2)

To a suspension of disuccinimidyl carbonate (1.2 eq) in CH₂Cl₂ was added **10** (1 eq). The reaction was stirred at room temperature overnight. A small quantity of white solid was removed by filtration. The organic phase was washed with aqueous 1M KHSO₄ solution and water, dried over MgSO₄ and the solvent removed under reduced pressure. The residue was re-dissolved in CH₂Cl₂ and diethyl ether added. The solid precipitate formed was isolated by filtration and dried under vacuum to afford **BB2** as a white crystalline solid (56%).¹H NMR (400 MHz, CDCl₃) δ = 6.23 (br s, 1H, NH), 4.98 (br s, 1H, NH), 3.26 (d, *J* = 6.8 Hz, 2H, CH₂N), 2.81 (s, 4H, CH₂), 1.46 (s, 9H, Boc), 1.35 (s, 6H, CH₃) ¹³C NMR (75 MHz, CDCl₃) δ = 169.96, 157.33, 149.72, 80.23, 77.25, 56.00, 49.37, 28.33, 25.49, 24.06. **ESI-MS** (Mw

329.16): m/z 347 [M + Na]⁺ **HRMS** calcd. for C₁₄H₂₃N₃O₆ Na 352.1479 [M+Na]⁺, found 352.1475, [2M+Na]⁺, found 681.3057

General procedure for oligourea coupling.

Boc-protected oligourea (1.0 eq) was dissolved in TFA (3 ml / g) and stirred for 45 min. The reaction mixture was then concentrated under reduced pressure and coevaporated 3 times with cyclohexane. The crude product was then dissolved in CH₃CN (5 ml / g). DIPEA (3.0 eq) was then added and the mixture was cooled to 0° C prior to the dropwise addition of the desired carbamate, **BB1**, **BB2**, **11a**, **11b**, **11c** or **11d** dissolved in CH₃CN. The completion of reaction was controlled by TLC.

Boc-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (13)

13 was prepared from **BB1** (125 mg, 0.378 mmol) and **12** (200 mg, 0.398 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **13** as a white product. (>99%) ¹**H NMR** : (300MHz, CD₃OH) δ = 6.32 (m, 1H, NH), 6.25 (m, 1H, NH), 6.17-5.99 (m, 4H, NH), 5.98-5.79 (m, 3H, NH), 4.03-3.80 (m, 2H, CHN), 3.77-3.63 (m, 1H, CHN), 3.59-3.37 (m, 4H, CH₂N), 2.30-2.21 (m, 1H, CH₂N), 2.81-2.75 (m, 1H, CH₂N), 2.72 (d, J = 4.7 Hz, 1H, CH₃N), 2.60-2.48 (m, 1H, CH₂N), 1.77-1.62 (m,2H, CH), 1.45 (s, 9H, Boc), 1.35-1.28 (m, 2H, CH₂), 1.27-1.21 (m, 6H, CH₃), 1.06 (d, J = 6.8 Hz, 3H, CH₃), 0.97-0.89 (m, 12H, CH₃) **ESI-MS** (Mw 615.81): *m*/*z* 616.2 [M+H]⁺, 638.3 [M + Na]⁺, 1253.0 [2M+Na]⁺

Boc-Leu^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (14)

14 was prepared from **10a** (55 mg, 0.154 mmol) and **13** (100 mg, 0.162 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **14** as a white product. (116 mg, 89%) ¹H NMR : (300MHz, CD₃CN) δ = 6.16 (m, 1H, NH), 5.98 (m, 1H, NH), 5.83 (m, 2H, NH), 5.65 (m, 1H, NH), 5.59 (m, 1H, NH), 5.52-5.38 (m, 4H, NH), 5.34 (m,1H,NH), 44.02-3.92 (m, 1H, CHN), 3.86-3.68 (m, 2H, CHN), 3.68-3.49 (m, 7H, CHN-CH₂N), 3.38-3.27 (1H, CH₂N), 2.64 (d, J = 4.7 Hz, 3H, CH₃N), 2.56-2.43 (m, 2H, CH₃N), 1.71-1.56 (m, 3H, CH), 1.45 (s, 9H, Boc), 1.35 (s, 3H, CH₃), 1.29-1.15 (m, 4H, CH₂), 1.13 (s, 3H, CH₃), 1.02-0.97 (m, 3H, CH₃), 0.94-0.80 (m, 18H, CH₃). ¹H NMR : (300MHz, CD₃OH) δ = 66.47 (d, J = 9.3 Hz, 1H, NH), 6.37 (m, 1H, NH), 6.21-6.12 (m, 2H, NH), 6.09-6.00 (m,

2H, NH), 5.97-5.81 (m, 5H, NH), 4.04-3.93 (m, 1H, CHN), 3.92-.379 (m, 1H, CHN), 3.78-3.62 (m, 2H, CHN), 3.61-3.40 (m, 5H, CH₂N), 3.28-3.10 (m, 2H, CH₂N), 2.84-2.75 (m, 1H, CH₂N), 2.72 (d, J = 4.7 Hz, 3H, CH₃N), 2.70-2.58 (m, 1H, CH₂N), 2.55-2.41 (m, 1H, CH₂N), 1.78-1.57 (m, 3H, CH), 1.48 (s, 9H, CH₃), 1.31 (s, 3H, CH₃), 1.28-1.22 (m, 4H, CH₂), 1.19 (s, 3H, CH₃), 1.07 (d, J = 6.8 Hz, 3H, CH₃), 0.97-0.89 (m, 18H, CH₃). **ESI-MS** (Mw 758.01): m/z 780 [M + Na]⁺

Boc-Ala^u-Leu^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (15)

15 was prepared from **11b** (27 mg, 0.077 mmol) and **14** (50 mg, 0.081 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **15** as a white product (56 mg, 80%). ¹H NMR : (400MHz, CD₃CN) δ = 6.21 (d, J = 8.0 Hz, 1H, NH), 6.11 (m, 1H, NH), 5.99-5.90 (m, 2H, NH), 5.85-5.77 (m, 2H, NH), 5.66 (m, 1H, NH), 5.62-5.56 (m, 2H, NH), 5.52 (d, J = 9.7 Hz, 1H, NH), 5.48 (d, J = 8.0 Hz, 1H, NH), 5.46 (d, J = 9.4 Hz, 1H, NH), 5.12 (d, J = 9.8 Hz, 1H, NH), 3.95-3.84 (m, 1H, CHN), 3.29-3.19 (m, 1H, CH₂N), 2.82-2.74 (m, 1H, CH₂N), 2.55 (d, J = 4.7 Hz, 3H, CH₃N), 2.46-2.36 (m, 1H, CH₂N), 2.34-2.26 (m, 1H, CH₂N), 2.24-2.15 (m, 1H, CH₂N), 1.62-1.49 (m, 2H, CH), 1.47-1.37 (m, 1H, CH), 1.36 (s, 9H, Boc), 1.28 (s, 3H, CH₃), 1.14-1.04 (m, 4H, CH₂), 0.99 (s, 3H, CH₃), 0.97 (d, J = 7.0 Hz, 3H, CH₃), 0.92 (d, J = 6.8 Hz, 3H, CH₃), 0.83-0.73 (m, 18H, CH₃). **ESI-MS** (Mw 857,62): *m/z* 880 [M + Na]⁺

Boc-Val^u-Ala^u-Leu^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (2)

2 was prepared from **11c** (19 mg, 0.055 mmol) and **15** (50 mg, 0.058 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **2** as a white product (50mg, 87%).¹**H NMR** : (400MHz, CD₃OH) δ = 6.65 (d, *J* = 10.6 Hz, 1H, NH), 6.53 (dd, J = 10.3, 2.9 Hz, 1H, NH), 6.45 (dd, 1H, NH), 6.37 (s, 1H, NH), 6.34 (dd, J = 9.9, 3.8 Hz, 1H, NH), 6.29 (d, J = 9.9 Hz, 1H, NH), 6.20 (m, 1H, NH), 6.17 (d, J = 10.5 Hz, 1H, NH), 6.03-5.98 (m, 3H, NH), 5.97 (d, J = 11.0 Hz, 1H, NH), 5.90 (dd, J = 8.6, 4.5 Hz, 1H, NH), 5.86 (d, J = 10.3 Hz, 1H, NH), 5.83 (dd, J = 9.9, 3.6 Hz, 1H, NH), 4.10-3.95 (m, 3H, CHN), 3.92-3.79 (m, 1H,CHN), 3.75-3.41 (m, 9H, 2 CHN-7 CH₂N), 3.19 (dd, J = 12.8, 9.3 Hz, 1H, CH₂N), 2.70 (d, J = 4.8 Hz, 3H, CH₃N), 2.67-2.24 (m, 6H, CH₂N), 1.76-1.57 (m, 4H, CH), 1.48 (s, 9H, Boc), 1.33 (s, 3H, CH₃ (Aib)), 1.31-1.14 (m, 4H, CH₂), 1.12 (s, 3H, CH₃ (Aib)), 1.07-1.01 (m, 6H, CH₃), 0.98-0.88 (m, 24H, CH₃). ¹³C NMR (400 MHz, CD₃OH) δ = 162.84, 162.26, 161.13, 160.66, 160.64, 160.00, 159.91, 158.76, 79.39, 56.88, 55.51, 52.52, 47.11,

46.05, 45.72, 45.31, 45.24, 43.94, 43.16, 42.94, 41.95, 31.12, 31.09, 27.85, 26.52, 25.94, 25.28, 25.06, 24.90, 22.79, 22.70, 21.50, 21.21, 19.21, 19.18, 18.10, 17.76, 17.72, 17.47.**ESI-MS** (Mw 985,71): *m/z* 1009 [M + Na]⁺

Boc- (Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (16)

16 was prepared from **BB2** (197 mg, 0.598 mmol) and **12** (300 mg, 0.598 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **16** as a white product (225 mg, 61%). ¹H NMR : (300MHz, CD₃OH) δ = 6.88 (m, 1H, NH), 6.41 (m, 1H, NH), 6.29 (m, 1H, NH), 6.23-6.11 (m, 2H, NH), 5.94 (m, 1H, NH), 5.86-5.75 (m, 2H, NH), 5.63 (d, J = 9.8 Hz, 1H, NH), 4.10-3.95 (m, 1H, CHN), 3.94-3.83 (m, 1H, CHN), 3.77 (dd, J = 13.7, 7.9 Hz, 1H, CHN), 3.69-3.45 (m, 4H, CH₂N), 2.85 (dd, J = 14.0, 5.3 Hz, 1H, CH₂N), 2.73 (d, J = 4.6 Hz, 3H, CH₃N), 2.70-2.63 (m, 1H, CH₂N), 2.56-2.34 (m, 2H, CH₂N), 1.78-1.56 (m, 2H, CH), 1.49 (s, 9H, Boc), 1.38 (s, 3H, CH₃(Aib)), 1.32-1.20 (m, 2H, CH₂), 1.16 (s, 3H, CH₃(Aib)), 1.06 (d, J = 6.8 Hz, 2H, CH₃), 0.97-0.86 (m, 12H, CH₃) **ESI-MS** (Mw 615.81): *m/z* 616.2 [M + H]⁺, 639.3 [M + Na]⁺, 1230.8 [2M + H]⁺, 1256.0 [2M + Na]⁺

Boc- Leu^u-(Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (17)

17 was prepared **11a** (34 mg, 0.097 mmol) and **16** (60 mg, 0.097 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **16** as a white product (62 mg, 84%). ¹H NMR : (300MHz, CD₃OH) δ = 6.51 (d, J = 9.4 Hz, 1H, NH), 6.47-6.38 (m, 2H, NH), 6.31-6.10 (m, 5H, NH), 5.96 (d, J = 9.9 Hz, 1H, NH), 5.76 (d, J = 10.0 Hz, 1H, NH), 5.42 (s, 1H, NH), 4.05-3.78 (m, 3H, CHN), 3.79-3.66 (m, 2H, CHN), 3.64-3.49 (m, 4H, CH₂N), 3.42-3.35 (m, 1H, CH₂N), 3.28-3.16 (m, 1H, CH₂N), 2.71 (d, J = 4.0 Hz, 3H, CH₃N), 2.68-2.57 (m, 2H, CH₂N), 2.56-2.45 (m, 1H, CH₂N), 2.43-2.31 (m, 1H, CH₂N), 1.75-1.50 (m, 3H, CH), 1.45 (s, 9H, CH₃), 1.42 (s, 3H, CH₃), 1.35-1.17 (m, 4H, CH₂), 1.047 (s, 3H, CH₃), 1.04 (d, J = 6.8 Hz, 3H, CH₃), 0.95-0.83 (m, 18H, CH₃). **ESI-MS** (MW 758.01): *m*/z 758.3 [M + H]⁺, 781.4 [M + Na]⁺, 1514.9 [2M + H]⁺, 1538.9 [2M + Na]⁺

Boc- Ala^u-Leu^u-(Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (18)

18 was prepared from 11b (24 mg, 0.077 mmol) and 17 (62 mg, 0.081 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous

solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **18** as a white product (84 mg, 97%). ¹**H NMR** : (400MHz, CD₃OH) δ = 6.59 (d, J = 8.8 Hz, 1H, NH), 6.51-6.41 (m, 2H, NH), 6.36-6.28 (m, 2H, NH), 6.27-6.10 (m, 3H, NH), 6.07 (d, J = 10.1 Hz, 1H, NH), 6.00 (t, J = 6.1 Hz, 1H, NH), 5.89 (d, J = 10.0 Hz, 1H, NH), 5.83 (d, J = 9.5 Hz, 1H, NH), 5.75 (s, 1H, NH), 4.07-3.94 (m, 2H, CHN), 3.94-3.78 (m, 4H, CHN), 3.71-3.46 (m, 8H, CH₂N), 2.91-2.77 (m, 1H, CH₂N), 2.74 (d, J = 4.7 Hz, 3H, CH₃N), 2.71-2.33 (m, 5H, CH₂N), 1.80-1.62 (m, 4H, CH), 1.47 (s, 9H, Boc), 1.46 (s, 3H, CH₃), 1.32-1.21 (m, 4H, CH₂), 1.14 (s, 3H, CH₃), 1.12-1.03 (m, 6H, CH₃), 0.99-0.87 (m, 24H, CH₃)

Boc-Val^u-Ala^u-Leu^u-(Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (3)

3 was prepared from 11c (32 mg, 0.093 mmol) and 18 (84 mg, 0.097 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 90:10) over silica gel gave 3 as a white product (75 mg, 78%). ¹**H NMR** : (400MHz, CD₃OH) δ = 6.71 (m, 1H, NH), 6.61 (d, *J* = 10.4 Hz, 1H, NH), 6.53-6.45 (m, 2H, NH), 6.37-6.20 (m, 4H, NH), 6.05 (d, J = 10.3 Hz, 1H, NH), 6.02-5.97 (m, 2H, NH), 5.95 (d, J = 10.7 Hz, 1H, NH), 5.88 (d, J = 9.8 Hz, 1H, NH), 5.77 (m, 1H, NH), 5.68 (s, 1H, NH), 4.18-3.97 (m, 2H, CHN), 3.93-3.81 (m, 2H, CHN), 3.70-3.46 (m, 9H, CH₂N), 2.72 (d, J = 4.6 Hz, 3H, NH), 2.69-2.53 (m, 2H, CH₂N), 2.55-2.31 (m, 1H, CH₂N), 1.76-1.52 (m, 4H, CH), 1.49 (s, 9H, CH₃), 1.27-1.17 (m, 4H, CH₂), 1.14 (m, 3H, CH₃), 1.08-1.01 (m, 6H, CH₃), 0.99-0.82 (m, 24H, CH₃). ¹³C NMR (400 MHz, CD₃OH) δ 161.23, 161.12, 160.67, 160.48, 160.38, 159.97, 159.89, 158.68, 79.31, 56.65, 54.85, 53.40, 46.29, 45.60, 45.38, 45.32, 43.73, 43.16, 42.85, 42.32, 31.23, 31.06, 27.86, 26.23, 25.94, 25.29, 24.95, 24.88, 22.78, 22.73, 21.86, 21.52, 19.14, 19.12, 18.01, 17.62, 17.43. ESI-MS $(Mw 985,71): m/z 1008.7 [M + Na]^+$

Boc-Aib^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (19)

19 was prepared from **BB1** (50 mg, 0.154 mmol) and **13** (100 mg, 0.162 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂ -MeOH (v/v), 95:5) over silica gel gave **19** as a white product (90 mg, 73%). ¹H NMR : (300MHz, DMSO d₆) δ = 6.46 (s, 1H, NH), 6.10 (m, 1H, NH), 6.02-5.86 (m, 8H, NH), 5.82 (m, 1H, NH), 3.75-3.65 (m, 1H, CHN), 3.64-3.57 (m, 1H, CHN), 3.57-3.46 (m, 1H, CHN), 3.28-3.15 (m, 6H, CH₂N), 3.09-3.04 (m, 2H, CH₂N), 2.71-2.60 (m, 2H, CH₂N), 2.54 (d, J = 4.5 Hz, 3H, CH₃N), 1.67-1.54 (m, 2H, CH), 1.36 (s, 9H, Boc), 1.21-1.15 (m, 2H, CH₂), 1.17-1.07 (m, 12H, CH₃), 0.93 (d, J = 6.6 Hz, 3H, CH₃), 0.88-0.75 (m, 12H, CH₃). **ESI-MS** (Mw 729,95): *m/z* 752 [M + Na]⁺

Boc-Leu^u-Aib^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (20

20 was prepared from **11a** (27 mg, 0.078 mmol) and **19** (60 mg, 0.082 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **20** as a white product (63 mg, 88%). ¹H NMR : (400MHz, CD₃CN) δ = 6.05 (m, 1H, NH), 5.86-5.78 (m, 2H, NH), 5.74-5.54 (m, 4H, NH), 5.52-5.38 (m, 5H, NH), 5.33 (m, 1H, NH), 3.97-3.87 (m, 1H, CHN), 3.85-3.74 (m, 1H, CHN), 3.69-3.50 (m, 5H, CHN-CH₂N), 3.46-3.34 (m, 1H, CH₂N), 3.27-2.98 (m, 2H, CH₂N), 2.65 (d, J = 4.7 Hz, 3H, CH₃N), 2.59-2.48 (m, 1H, CH₂N), 1.70-1.56 (m, 3H, CH), 1.43 (s, 9H, Boc), 1.36-1.14 (m, 16H, CH₂-CH₃), 1.01 (d, J = 6.8 Hz, 3H, CH₃), 0.94-0.85 (m, 18H, CH₃). **ESI-MS** (Mw 872,15): *m/z* 894 [M + Na]⁺

Boc-Ala^u-Leu^u-Aib^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (21)

21 was prepared **11b** (10 mg, 0.032 mmol) and **20** (30 mg, 0.034 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **21** as a white product (10 mg, 30%). ¹H NMR : (400MHz, CD₃CN) δ = 6.13 (m, 1H, NH), 6.02 (m, 1H, NH), 5.96 (m, 1H, NH), 5.87 (m, 1H, NH), 5.83-5.65 (m, 4H, NH), 5.63-5.47 (m, 1H, NH), 5.37 (m, 1H, NH), 5.23 (m, 1H, NH), 4.99 (m, 1H, NH), 3.99-3.75 (m, 4H, CHN), 3.71-3.33 (m, 10H, CHN-CH₂N), 3.15-3.01 (m, 2H, CH₂N), 2.65 (s, 3H, CH₃N), 2.63-2.42 (m, 2H, CH₂N), 1.73-1.55 (m, 3H, CH), 1.47 (s, 9H, Boc), 1.35-1.27 (m, 6H, CH₃), 1.26-1.17 (m, 4H, CH₂), 1.16-1.07 (m, 6H, CH₃), 1.06-0.97 (m, 6H, CH₃), 0.96-0.84 (m, 18H, CH₃). **ESI-MS** (Mw 972,27): *m/z* 994 [M + Na]⁺

Boc-Val^u-Ala^u-Leu^u-Aib^u-Aib^u-Val^u-Ala^u-Leu^u-NHMe, (4)

4 was prepared from **11c** (3 mg, 0.009 mmol) and **21** (10 mg, 0.010 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 95:5) over silica gel gave **4** as a white product (8 mg, 71%) . ¹H NMR : (400MHz, CD₃OH) δ = 6.57 (d, J = 11.6 Hz, 1H, NH), 6.39 (m, 1H, NH), 6.28 (m, 1H, NH), 6.25-6.17 (m, 2H, NH), 6.15-6.02 (m, 5H, NH), 5.93 (m, 1H, NH), 5.86-5.68 (m, 6H, NH), 3.97-3.83 (m, 3H, CHN), 3.76 (m, 1H, CHN), 3.65-3.32 (m, 10H, CHN-CH₂N), 3.23-3.08 (m, 2H, CH₂N), 2.61 (d, J = 4.5 Hz, 3H, CH₃), 2.59-2.03 (m, 6H, CH₂N), 1.67-1.46 (m, 4H, CH), 1.37 (s, 9H, Boc), 1.21 (s, 3H, CH₃), 1.19 (s, 6H, CH₃), 1.11

(s, 3H, CH₃), 0.99-0.91 (m, 6H, CH₃), 0.88-0.76 (m, 24H, CH₃). **ESI-MS** (Mw 1099,79): m/z 573 $[M/2 + Na]^+$, 1122 $[M + Na]^+$

Boc-(Aib^u)_{rev}-(Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (22)

22 was prepared from **BB2** (63 mg, 0.193 mmol) and **16**(125 mg, 0.203 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 90:10) over silica gel gave **22** as a white product (110 mg, 72 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.74 (m, 1H, NH), 6.58-6.45 (m, 2H, NH), 6.33 (m, 1H, NH), 6.27-6.19 (m, 2H, NH), 6.07-5.98 (m, 2H, NH), 5.88 (s, 1H, NH), 5.72 (d, J = 10.0 Hz, 1H, NH), 5.65 (s, 1H, NH), 4.09-3.95 (m, 1H, CHN), 3.94-3.81 (m, 2H, CHN), 3.68-3.49 (m, 4H, CH₂N), 3.36-3.32 (m, 2H, CH₂N), 3.09-3.00 (m, 1H, CH₂N), 2.74 (d, J = 3.8 Hz, 3H, CH₃N), 2.73-2.62 (m, 1H, CH₂N), 2.59-2.39 (m, 2H, CH₂N), 1.79-1.55 (m, 2H, CH), 1.48 (s, 9H, CH₃), 1.42 (s, 3H, CH₃), 1.33 (s, 3H, CH₃), 1.29-1.23 (m, 2H, CH₂), 1.22 (s, 3H, CH₃), 1.11 (s, 3H, CH₃), 1.06 (d, J = 6.8 Hz, 3H, CH₃), 0.97-0.87 (m, 12H, CH₃). **ESI-MS** (Mw 729.95): *m/z* 730.3 [M + H]⁺, 753.4 [M + Na]⁺, 1459.9 [2M + H]⁺, 1484.1 [2M + Na]⁺

Boc-Leu^u-(Aib^u)_{rev}-(Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (23)

23 was prepared from **11a** (51 mg, 0.144 mmol) and **22** (100 mg, 0.137 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 90:10) over silica gel gave **22** as a white product (85 mg, 71 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.49 (m, 1H, NH), 6.40 (m, 2H, NH), 6.33 (m, 1H, NH), 6.27-6.16 (m, 3H, NH), 6.13-6.02 (m, 3H, NH), 5.90-5.83 (m, 2H, NH), 5.79 (d, J = 10.0 Hz, 1H, NH), 4.05-3.94 (m, 1H, CHN), 3.94-3.80 (m, 2H, CHN), 3.76-3.66 (m, 1H, CHN), 6.65-3.48 (m, 5H, CH₂N), 3.33-3.21 (m, 1H, CH₂N), 3.07-2.96 (m, 2H, CH₂N), 2.74 (d, J = 3.8 Hz, 3H, CH₃N), 2.72-2.65 (m, 2H, CH₂N), 2.59-2.37 (m, 2H, CH₂N), 1.77-1.56 (m, 3H, CH), 1.47 (s, 9H, CH₃), 1.42 (s, 3H, CH₃), 1.34 (s, 3H, CH₃), 1.31-1.25 (m, 4H, CH₂), 1.23 (s, 3H, CH₃), 1.13 (s, 3H, CH₃), 1.06 (d, J = 6.8 Hz, 3H, CH₃), 0.99-0.88 (m, 18H, CH₃).

Boc-Ala^u-Leu^u-(Aib^u)_{rev}- (Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (24)

24 was prepared from **11b** (30 mg, 0.096 mmol) and **23** (80 mg, 0.091 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 90:10) over silica gel gave **24** as a white

product (80 mg, 89 %). ¹**H NMR** : (300MHz, CD₃OH) δ = 6.60 (d, J = 9.1 Hz, 1H, NH), 6.53 (m, 1H, NH), 6.50-6.41 (m, 2H, NH), 6.36 (m, 1H, NH), 6.29 (d, J = 9.1 Hz, 1H, NH), 6.26-6.19 (m, 2H, NH), 6.16-6.00 (m, 3H, NH), 5.95 (s, 1H, NH), 5.88 (s, 1H, NH), 5.83 (m, 2H, NH), 4.07-3.79 (m, 4H, CHN), 3.78-3.65 (m, 1H, CHN), 3.64-6.40 (m, 5H, CH₂N), 3.33-3.21 (m, 2H, CH₂N), 2.97-2.84 (m, 1H, CH₂N), 2.83-2.76 (m, 1H, CH₂N), 2.75 (d, J = 3.9 Hz, 3H, CH₃N), 2.71-2.40 (m, 4H, CH₂N), 1.79-1.54 (m, 3H, CH), 1.47 (s, 9H, CH₃), 1.43 (s, 3H, CH₃), 1.39 (s, 3H, CH₃), 1.35-1.25 (m, 4H, CH₂), 1.23 (s, 3H, CH₃), 1.12 (s, 3H, CH₃), 1.09-1.04 (m, 6H, CH₃), 0.98-0.88 (m, 18H, CH₃). **ESI-MS** (Mw 972.27): *m/z* 973.4 [M + H]⁺, 995.5 [M + Na]⁺, 1102.0 [M + K]⁺

Boc-Val^u-Ala^u-Leu^u-(Aib^u)_{rev}- (Aib^u)_{rev}-Val^u-Ala^u-Leu^u-NHMe, (5)

5 was prepared from 11c (29 mg, 0.086 mmol) and 24 (80 mg, 0.082 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH2Cl2-MeOH (v/v), 90:10) over silica gel gave 5 as a white product (88 mg, 97 %). ¹**H** NMR : (300MHz, CD₃OH) δ = 6.66 (m, 1H, NH), 6.61 (d, J = 10.1 Hz, 1H, NH), 6.57-6.51 (m, 2H, NH), 6.50-6.43 (m, 2H, NH), 6.35 (m, 1H, NH), 6.30 (d, J = 9.1 Hz, 1H, NH), 6.24 (m, 1H, NH), 6.15 (d, J = 10.0 Hz, 1H, NH), 6.01-5.93 (m, 2H, NH), 5.92-5.92 (m, 2H, NH), 5.88 (m, 1H, NH), 5.85 (m, NH), 5.82 (m, 1H, NH), 4.17-3.82 (m, 4H, CHN), 3.81-3.71 (m, 1H, CHN), 3.70-3.46 (m, 9H, CHN-CH₂N), 2.91-2.82 (m, 1H, CH₂N), 2.79-2.69 (m, 1H, CH₂N), 2.75 (d, J = 3.8 Hz, 3H, CH₃N), 2.68-2.42 (m, 5H, CH₂N), 2.42-2.29 (m, 1H, CH₂N), 1.79-1.56 (m, 4H, CH), 1.50 (s, 9H, CH₃), 1.43 (s, 3H, CH₃), 1.39 (s, 3H, CH₃), 1.31-1.25 (m, 4H, CH₂), 1.24 (s, 3H, CH₃), 1.12 (s, 3H, CH₃), 1.08-1.01 (m, 6H, CH₃), 0.98-0.88 (m, 24H, CH₃). ¹³C NMR (300 MHz, CD₃OH) δ 160.68, 160.64, 160.21, 160.02, 159.66, 159.45, 159.20, 158.32, 78.93, 56.23, 56.13, 54.56, 54.46, 52.99, 52.89, 52.54, 52.45, 46.62, 46.34, 45.49, 45.25, 44.96, 44.78, 43.15, 42.82, 42.72, 42.29, 42.18, 30.68, 30.42, 27.50, 25.98, 25.93, 25.63, 25.50, 25.07, 24.72, 24.57, 24.48, 24.44, 22.39, 21.14, 21.12, 18.85, 18.78, 17.60, 17.50, 17.28, 17.18. ESI-MS (Mw 1100.44): m/z 1100.5 [M $+ H^{+}$, 1122.6 [M + Na]⁺

Boc Gly^u-Val^u-Ala^u-Leu^u-NHMe, (25)

25 was prepared from **11d** (171 mg, 0.568 mmol) and **12**(300 mg, 0.598 mmol) as described in the general procedure. After completion (12h), the desired compound **25** was precipitated upon addition of water and washed with 1M KHSO₄ aqueous solution and water. Finally **25** was dried over vacuum. (360 mg, quantitative). ¹H NMR : (300MHz, CD₃OH) δ = 6.72 (m, 1H,NH), 6.36 (m, 1H, NH), 6.27 (m, 1H, NH), 6.17 (m, 1H, NH), 6.09 (m, 1H, NH), 5.99 (d, J = 9.4 Hz, 1H, NH), 5.91-5.79 (m, 3H, NH), 4.08-3.96 (m, 1H, CHN), 3.94-3.83 (m, 1H, CHN), 3.69-3.44 (m, 4H, CHN- CH₂N), 3.11-2.94 (m, 2H, CH₂N), 2.73 (d, J = 4.1 Hz, 3H, CH₃N), 2.70-2.42 (m, 5H, CH₂N), 1.78-1.58 (m, 2H, CH), 1.48 (s, 9H, Boc), 1.37-1.21 (m, 2H, CH₂), 1.06 (d, J = 6.8 Hz, 3H, CH₃), 0.99-0.89 (m, 12H, CH₃). **ESI-MS** (Mw 587.76) : m/z 588.3 [M+H]⁺, 611.4 [M+Na]⁺

Boc Gly^u-Gly^u-Val^u-Ala^u-Leu^u-NHMe, (26)

26 was prepared from **11d** (112 mg, 0.371 mmol) and **25** (230 mg, 0.391 mmol) as described in the general procedure. After completion (12h), the desired compound **26** was precipitated upon addition of water and washed with 1M KHSO₄ aqueous solution and water. Finally **26** was dried over vacuum. (175 mg, 66 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.70 (m, 1H,NH), 6.40 (m, 1H, NH), 6.32 (m, 1H, NH), 6.28-6.14 (m, 3H, NH), 6.12-5.95 (m, 4H, NH), 5.87 (d, J = 9.8 Hz, 1H, NH), 4.08-3.96 (m, 1H, CHN), 3.95-3.84 (m, 1H, CHN), 3.68-3.43 (m, 7H, CHN-CH₂N), 3.30-2.80 (m, 6H, CH₂N), 2.73 (d, J = 4.3 Hz, 3H, CH₃N), 2.65-2.54 (m, 1H, CH₂N), 2.51-2.40 (m, 1H, CH₂N), 1.78-1.58 (m, 2H, CH), 1.48 (s, 9H, Boc), 1.34-1.22 (m, 2H, CH₂), 1.06 (d, J = 6.8 Hz, 3H, CH₃), 1.00-0.86 (m, 12H, CH₃). **ESI-MS** (Mw 673.85) : *m/z* 674.3 [M+H]⁺, 697.4 [M+Na]⁺

Boc Leu^u-Gly^u-Gly^u-Val^u-Ala^u-Leu^u-NHMe, (27)

27 was prepared from **11a** (80 mg, 0.225 mmol) and **26** (160 mg, 0.237 mmol) as described in the general procedure. After completion (12h), the desired compound **27** was precipitated upon addition of water and washed with 1M KHSO₄ aqueous solution and water. Finally **27** was dried over vacuum. (160 mg, 82 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.52 (d, J = 9.3 Hz, 1H, NH), 6.46-6.35 (m, 3H, NH), 6.31 (m, 1H, NH), 6.22 (m, 1H, NH), 6.20-6.00 (m, 6H, NH), 5.97(m, 1H, NH), 4.13-3.98 (m, 1H, CHN), 3.95-3.85 (m, 1H, CHN), 3.84-3.75 (m, 1H, CHN), 3.71-3.38 (m, 9H, CHN-CH₂N), 3.01-2.77 (m, 4H, CH₂N), 2.74 (d, J = 4.1 Hz, 3H, CH₃N), 2.71-2.62 (m, 2H, CH₂N), 2.60-2.49 (m, 1H, CH₂N), 2.48-2.37 (m, 1H, CH₂N), 1.79-1.57 (m, 3H, CH), 1.49 (s, 9H, Boc), 1.37-1.19 (m, 4H, CH₂), 1.07 (d, J = 6.8 Hz, 3H, CH₃), 1.00-0.86 (m, 18H, CH₃). **ESI-MS** (Mw 816.5) : *m/z* 817.5 [M+H]⁺, 839.5 [M+Na]⁺

Boc Ala^u-Leu^u-Gly^u-Gly^u-Val^u-Ala^u-Leu^u-NHMe, (28)

28 was prepared from **11b** (51 mg, 0.163 mmol) and **27** (140 mg, 0.172 mmol) as described in the general procedure. After completion (12h), the desired compound **28** was precipitated upon addition of water and washed with 1M KHSO₄ aqueous solution and water. Finally **28** was dried over vacuum. (125 mg, 79 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.70-6.57 (m, 2H, NH), 6.49-6.34 (m, 5H, NH), 6.26-6.19 (m, 2H, NH), 6.16-5.98 (m, 6H, NH), 4.14-3.98 (m, 1H, CHN), 3.96-3.85 (m, 1H, CHN), 3.83-3.73 (m, 1H, CHN), 3.70-3.38 (m, 10H, CHN-CH₂N), 3.14-2.77 (m, 4H, CH₂N), 2.74 (d, J = 4.5 Hz, 3H, CH₃N), 2.72-2.62 (m, 2H, CH₂N), 2.59-2.35 (m, 4H, CH₂N), 1.80-1.55 (m, 3H, CH), 1.49 (s, 9H, Boc), 1.36-1.20 (m, 4H, CH₂), 1.14-1.04 (m, 6H, CH₃), 1.00-0.86 (m, 18H, CH₃). **ESI-MS** (Mw 916.17): *m/z* 918.5 [M+H]⁺, 940.7 [M+Na]⁺

Boc Val^u-Ala^u-Leu^u-Gly^u-Gly^u-Val^u-Ala^u-Leu^u-NHMe, (6)

6 was prepared from **11c** (35 mg, 0.104 mmol) and **28** (100 mg, 0.109 mmol) as described in the general procedure. After completion (12h), the reaction mixture was evaporated, dissolved in ethyl acetate and treated with saturated NaHCO₃ aqueous solution, 1M KHSO₄ aqueous solution and brine. The organic layer was then dried over MgSO₄ and evaporated. Flash column chromatography (CH₂Cl₂-MeOH (v/v), 90:10) over silica gel gave **6** as a white product (85 mg, 75 %). ¹H NMR : (300MHz, CD₃OH) δ = 6.73-6.62 (m, 3H, NH), 6.53 (m, 1H, NH), 6.50-6.39 (m, 3H, NH), 6.30-6.20 (m, 3H, NH), 6.18-5.96 (m, 5H, NH), 5.92 (d, J = 9.7 Hz, 1H, NH), 5.84 (m, 1H, NH), 4.15-3.84 (m, 4H, CHN), 3.80-3.46 (m, 12H, CHN-CH₂N), 2.93-2.79 (m, 3H, CH₂N), 2.74 (d, J = 4.0 Hz, 3H, CH₃N), 2.71-2.31 (m, 7H, CH₂N), 1.81-1.57 (m, 4H, CH), 1.50 (s, 9H, Boc), 1.35-1.21 (m, 4H, CH₂), 1.11-1.04 (m, 6H, CH₃), 1.01-0.87 (m, 24H, CH₃). ¹³C NMR: (101 MHz, CD₃OH) δ 161.39, 161.24, 161.12, 161.03, 160.71, 160.58, 159.92, 158.72, 79.36, 56.81, 55.37, 46.40, 45.83, 45.36, 45.31, 43.74, 43.19, 42.90, 42.09, 41.59, 41.36, 39.64, 39.59, 31.07, 30.96, 27.86, 25.96, 25.29, 24.91, 24.29, 22.67, 21.72, 21.51, 19.16, 18.10, 17.83, 17.57, 17.31. **ESI-MS** (Mw 1044.34) : *m/z* 1044.7 [M+H]⁺, 1066.7 [M+Na]⁺

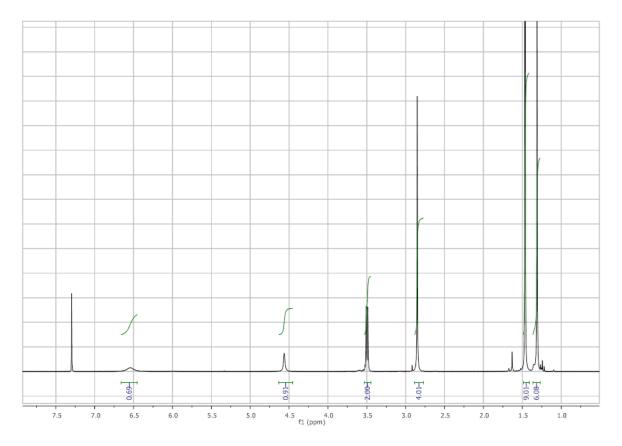


Figure S1: ¹H NMR spectra of carbamate BB1, recording in CDCl₃, (300MHz)

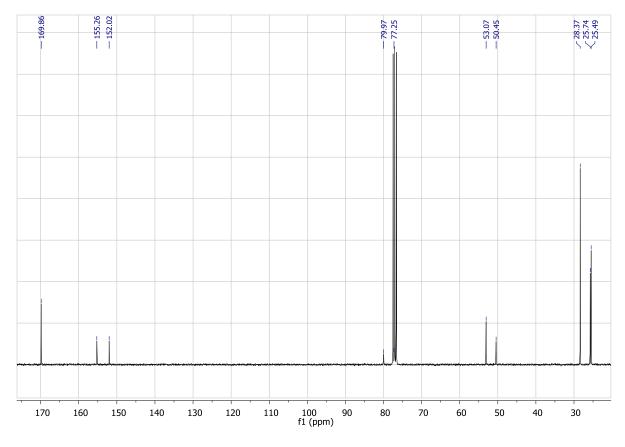


Figure S2: ¹³C NMR spectra of carbamate BB1, recording in CDCl₃, (75MHz)

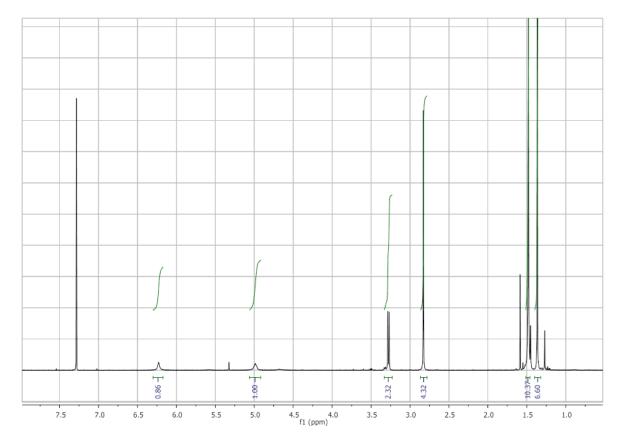


Figure S3: ¹H NMR spectra of carbamate **BB2**, recording in CDCl₃, (300MHz)

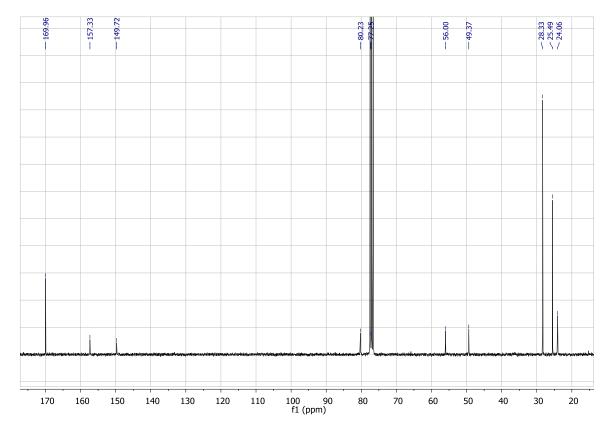


Figure S4: ¹³C NMR spectra of compound BB2, recording in CDCl₃, (75MHz)

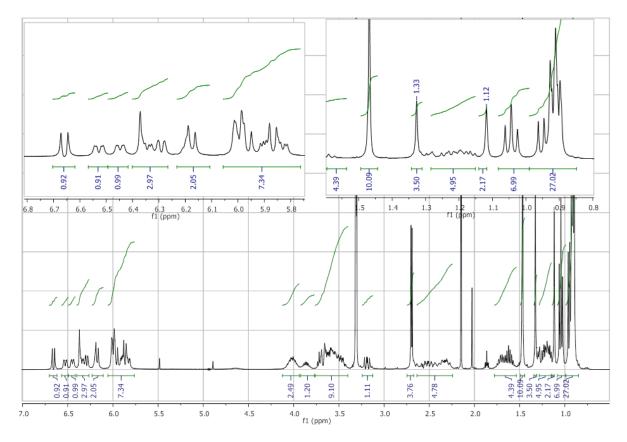


Figure S5: ¹H NMR spectra of compound **2**, recording in CD₃OH, (400MHz)

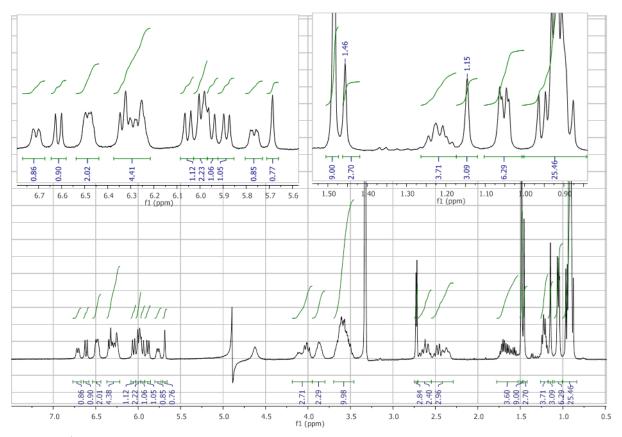


Figure S6: ¹H NMR spectra of compound 3, recording in CD₃OH, (400MHz)

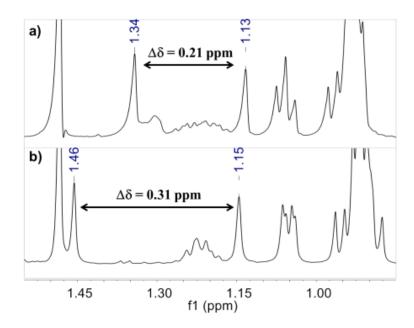


Figure S7: ¹H NMR region of methyl protons of DADME residues in a) oligourea 2 and b) oligourea 3

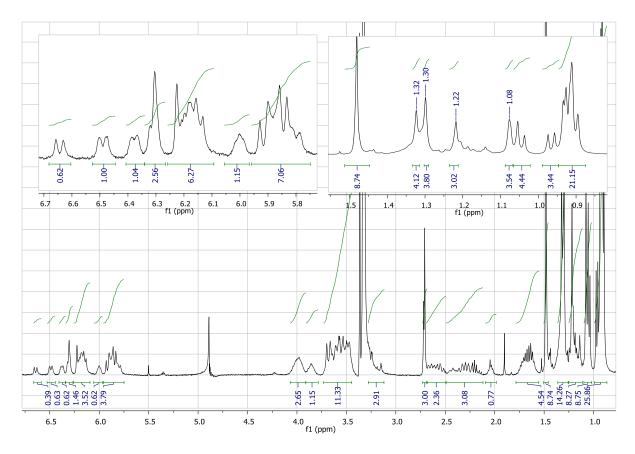


Figure S8: ¹H NMR spectra of compound 4 recording in CD₃OH (400MHz)

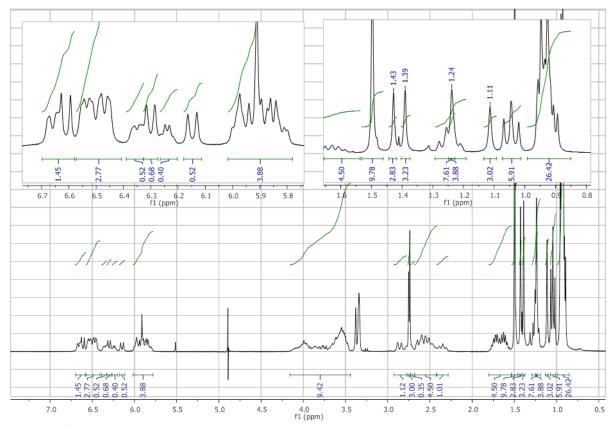


Figure S9: ¹H NMR spectra of compound 5, recording in CD₃OH, (300MHz)

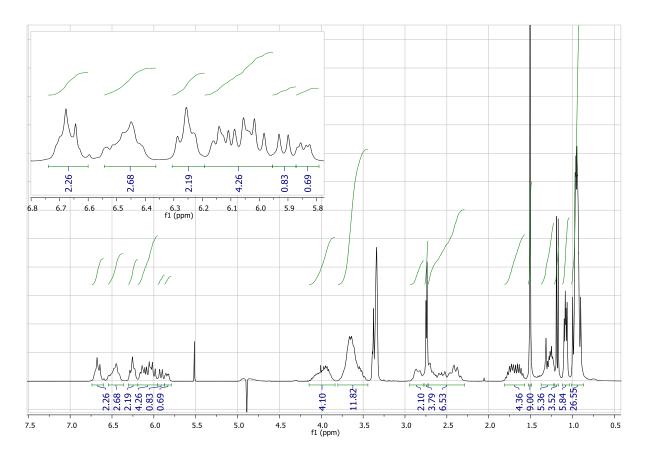
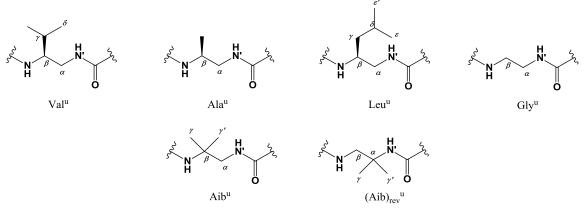


Figure S10: ¹H NMR spectra of compound 6, recording in CD₃OH, (300MHz)

NMR Characterization

Experiments were recorded on a DPX-400 NMR spectrometer (Bruker Biospin) with a vertical 9.4T narrow-bore/ultrashield magnet operating at 400 MHz for ¹H observation by means of a 5-mm direct QNP 1H_13C/31P/19F_2H probe with Z gradient capabilities. The four oligomers were dissolved in CD₃OH at room temperature.



Scheme S8: Nomenclature used for the description of the various protons for each residue type

	res. 7	re	es. 6	res. 5	res. 4	t i	res. 3 res. 2	' i	res. 1	
\downarrow_{c}			N:			NH NH		N H H		E.
Re	sidue	N'H	NH	^α CH ¹	^α CH ²	^β CH	γCH	^δ CH	٤CH	term CH
	NH-Me		6,20							2,71
Leu ^u	U1	6,44	5,99	3,58	2,66	3,88	1,23	1,70	0,93	
Ala ^u	U2	6,34	6,00	3,51	2,40	4,01	1,06			
Val ^u	U3	5,89	6,17	3,56	2,49	6,67	1,61	0,92		
Aib ^u	U4	6,30	6,37	3,69	3,20		1,34 1,13			
Leu ^u	U5	6,53	5,97	3,61	2,32	4,03	1,19	1,72	0,92	
Ala ^u	U6	5,83	5,87	3,59	2,34	4,05	1,04			
Val ^u	U7	6,00	6,65	3,47	2,56	3,63	1,64	0,94		
	Boc									1,48

Table S1: ¹H NMR chemical shifts (in ppm) of **2** in CD₃OH (400 MHz)

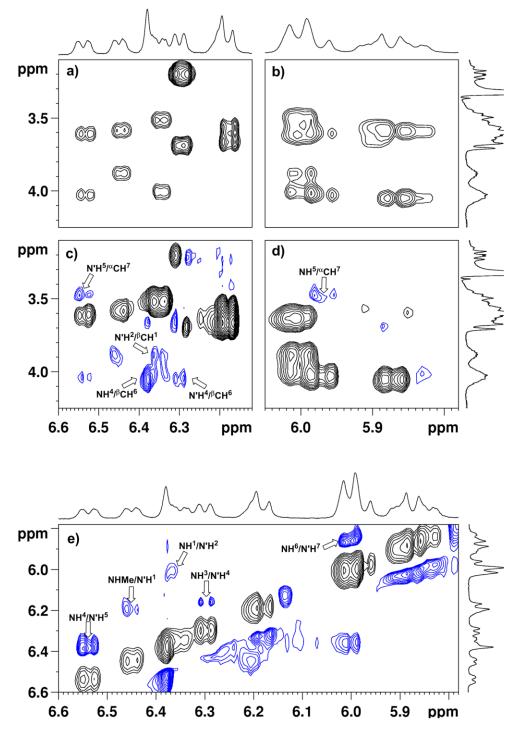
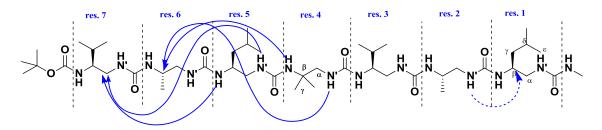


Figure S11: Representative sets of NOE connectivities observed for **2** in CD₃OH (400MHz). a) and b) part of the TOCSY plot shown for comparison c) and d) Part of the NH/CH region of the ROESY. e) Part of the NH/NH region of the ROESY plot of **2** in CD₃OH



Scheme S9: NOE connectivities observed for 2 in CD₃OH. N'H $(i+1)^{\beta}$ CH(i) represented in dashed arrows have been attributed previously as *Z*-*E* isomerization⁵

Table S2: ¹H NMR chemical shifts (in ppm) of 3 in CD₃OH (400 MHz)

Resi	due	N'H	NH	^α CH ¹	^a CH ²	^β CH	^γ CH	^ð CH	٤СН	term CH
	NH-Me		6,25							2,73
Leu ^u	U1	6,29	6,34	3,60	2,65	3,87	1,22	1,7	0,92	
Ala ^u	U2	6,47	6,05	3,57	2,38	4,02	1,06			
Val ^u	U3	6,49	5,95	3,55	2,45	3,62	1,58	0,90		
(Aib) _{rev} ^u	U4	5,69	6,27			4,01 2,60	1,46 1,15			
Leu ^u	U5	6,71	5,99	3,64	2,48	3,86	1,21	1,71	0,91	
Ala ^u	U6	5,76	5,88	3,58	2,37	4,11	1,05			
Val ^u	U7	5,98	6,61	3,51	2,59	3,59	1,64	0,93		
	Boc									

⁵ A. Violette, M. C. Averlant-Petit, V. Semetey, C. Hemmerlin, R. Casimir, R. Graff, M. Marraud, J.-P. Briand, D. Rognan, G. Guichard, *JACS*, **2005**, *127*, 2156-2164.

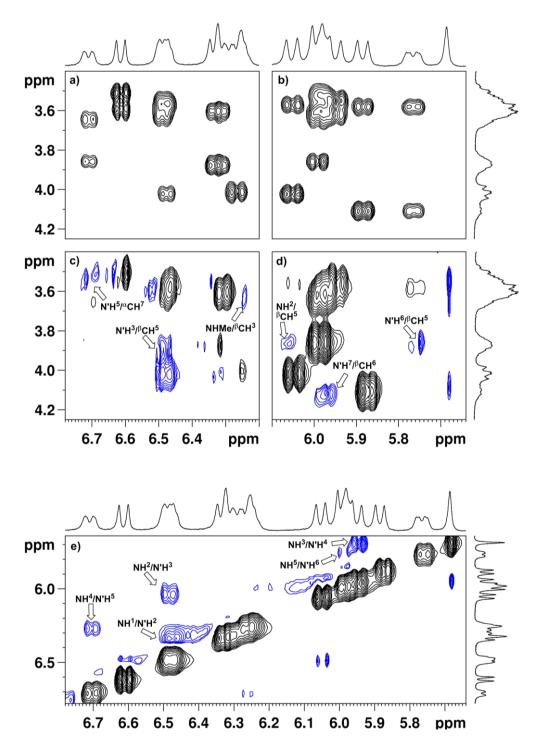
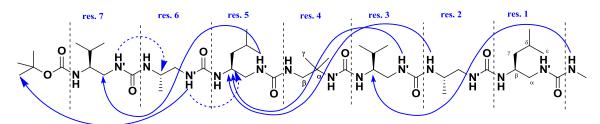


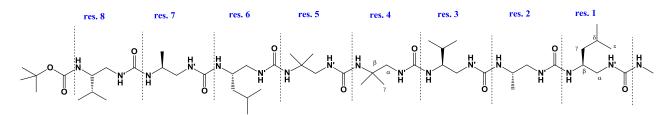
Figure S12: Representative sets of NOe connectivities observed for **3** in CD_3OH (400MHz). a) and b) part of the TOCSY plot shown for comparison c) and d) Part of the NH/CH region of the ROESY e) Part of the NH/NH region of the ROESY plot of **3**.

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Scheme S10: NOE connectivities observed for **3** in CD₃OH. N'H(i+1)/^{β}CH(i) represented in dashed arrows have been attributed previously as *Z*-*E* isomerization⁶

Table S3: ¹H NMR chemical shifts (in ppm) of **4** in CD₃OH (400 MHz)



Re	sidue	N'H	NH	^a CH ¹	^a CH ²	^β CH	γ CH	⁸ СН	٤СН	term CH
	NH-Me		6,18							2,71
Leu ^u	U1	6,37	6,14	3,56	2,67	3,85	1,68	1,20	0,90	
Ala ^u	U2	6,20	5,84	3,49	2,41	3,96	1,04			
Val ^u	U3	5,87	5,84	3,50	2,61	3,69	1,63	0,91		
Aib ^u	U4	6,16	6,22	3,55	3,32		1,30 1,22			
Aib ^u	U5	6,32	6,30	3,68	3,26		1,31 1,07			
Leu ^u	U6	6,49	5,90	3,57	2,26	3,98	1,71	1,18	0,91	
Ala ^u	U7	5,80	5,87	3,61	2,32	4,02	1,06			
Val ^u	U8	5,99	6,64	3,47	2,55	3,63	1,64	0,93		
	Boc									1,47

⁶ A. Violette, M. C. Averlant-Petit, V. Semetey, C. Hemmerlin, R. Casimir, R. Graff, M. Marraud, J.-P. Briand, D. Rognan, G. Guichard, *JACS*, **2005**, *127*, 2156-2164.

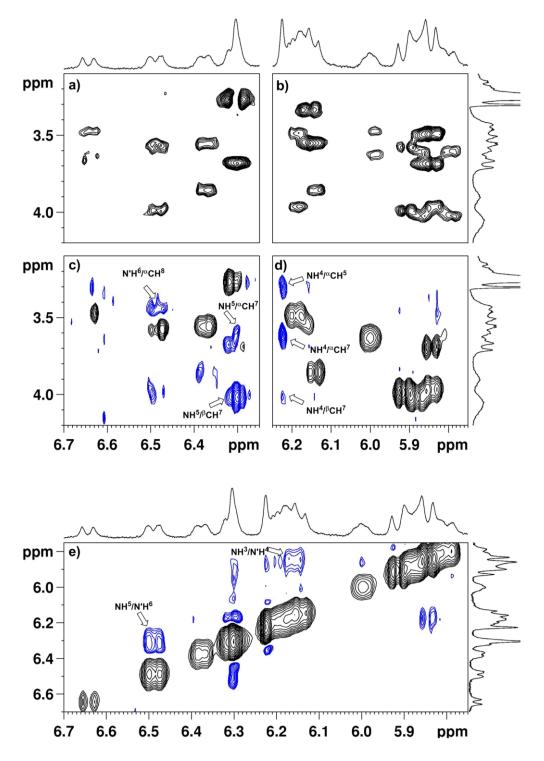
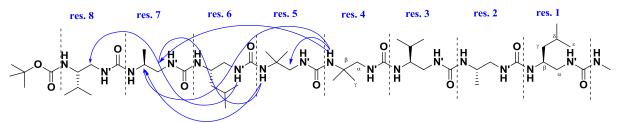


Figure S13: Representative sets of NOe connectivities observed for **4** in CD_3OH (400MHz). a) and b) part of the TOCSY plot shown for comparison c) and d) Part of the NH/CH region of the ROESY e) Part of the NH/NH region of the ROESY plot of **4**.



Scheme S11: NOE connectivities observed for 4 in CD₃OH.

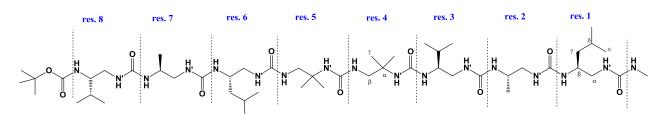


Table S4: ¹H NMR chemical shifts (in ppm) of 5 in CD₃OH (700 MHz)

Resi	idue	N'H	NH	^a CH ¹	^α CH ²	^β CH	γCH	^δ CH	٤СН	term CH
	NH-Me		6,23							2,73
Leu ^u	U1	6,34	6,29	3,57	2,73	3,85	1,71	1,25	0,93	
Ala ^u	U2	6,526	6,14	3,55	2,45	4,00	1,05			
Val ^u	U3	6,46	5,86	3,56	2,51	3,55	1,61	0,91		
(Aib) _{rev} ^u	U4	5,904	6,53			3,96 2,63	1,41 1,08			
(Aib) _{rev} ^u	U5	5,899	6,46			3,76 2,86	1,38 1,22			
Leu ^u	U6	6,65	5,94	3,65	2,54	3,98	1,72	1,22	0,94	
Ala ^u	U7	5,81	5,84	3,50	2,34	4,08	1,02			
Val ^u	U8	5,96	6,61	3,53	2,58	3,53	1,63	0,93		
	Boc									1,48

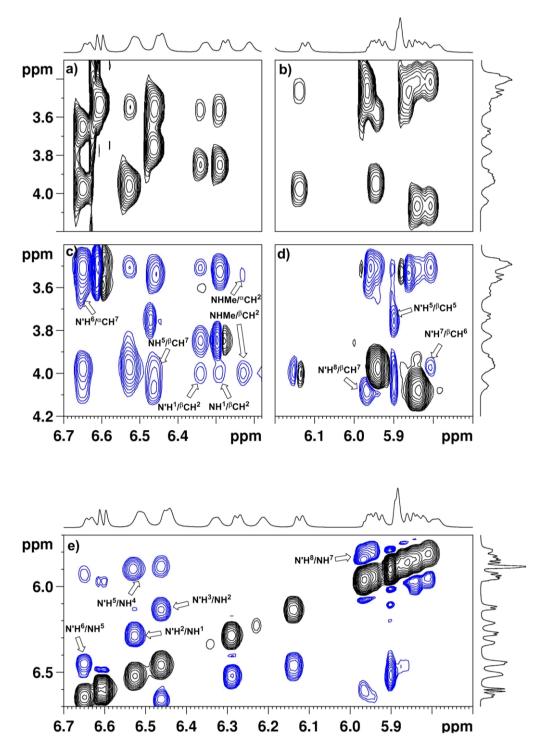
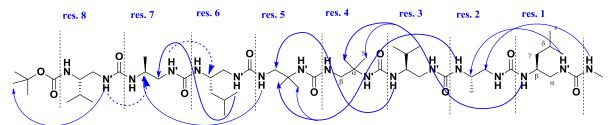


Figure S14: Representative sets of NOe connectivities observed for **5** in CD_3OH (700MHz). a) and b) part of the TOCSY plot shown for comparison c) and d) Part of the NH/CH region of the ROESY e) Part of the NH/NH region of the ROESY plot of **5**.



Scheme S12: NOE connectivities observed for **5** in CD₃OH. N'H(i+1)/^{β}CH(i) represented in dashed arrows have been attributed previously as *Z*-*E* isomerization⁷

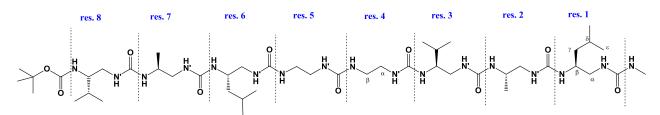


Table S5: ¹H NMR chemical shifts (in ppm) of 6 in CD₃OH (600 MHz)

Re	esidue	N'H	NH	^a CH ¹	^α CH ²	^β CH	γCH	^δ CH	٤CH	term CH
	NH-Me		6.23							2.72
Leu ^u	U1	6.44	6.05	3.57	2.68	3.89	1.70	1.24	0.93	
Ala ^u	U2	6.45	6.10	3.55	2.40	4.03	1.04			
Val ^u	U3	6.41	6.26	3.63	2.50	3.63	1.59	0.90		
Gly ^u	U4	6.51	6.13	3.62	2.81	3.69 2.71				
Gly ^u	U5	6.66	6.22	3.64	2.84	3.65 2.68				
Leu ^u	U6	6.68	5.98	3.65	2.38	3.98	1.72	1.22	0.93	
Ala ^u	U7	5.82	5.90	3.61	2.35	3.92	1.06			
Val ^u	U8	6.02	6.65	3.51	2.57	3.63	1.65	0.95		
	Boc									1.48

Table S6: ¹H NMR anisochronicity ($\Delta\delta$) values for main chain methylene protons in compounds **1**, **2** and **3**

Compound	P7	P6	P5	P4	P3	P2	P1
1	0,94	1,16	1,24	1,37	1,22	1,21	0,96
2	0,92	1,11	1,07	0,49	1,29	1,25	0,91
3	0,95	1,19	1,10	1,41	1,16	1,21	0,92

⁷ A. Violette, M. C. Averlant-Petit, V. Semetey, C. Hemmerlin, R. Casimir, R. Graff, M. Marraud, J.-P. Briand, D. Rognan, G. Guichard, *JACS*, **2005**, *127*, 2156-2164.

Compound	P8	P7	P6	P5	P4	P3	P2	P1
4	0,89	1,08	0,89	0,42	0,23	1,31	1,29	0,92
5	0,84	1,10	1,05	1,33	0,90	1,11	1,16	0,95
6	0,94	1,26	1,27	0.97-0.80	0.98-0.81	1,13	1,15	0,89

Table S7: ¹H NMR anisochronicity ($\Delta\delta$) values for main chain methylene protons in compounds 4, 5 and 6.

Circular dichroism (CD) measurements

All Circular dichroism (CD) spectra were recorded on a J-815 Jasco dichrographe (Jasco France, Nantes, France).

Spectra were acquired between 300 and 180 nm at a concentration of 0.2 mM in 2,2,2-trifluoroethanol (NMR grade, \geq 99.5%) using a quartz cell with a path length of 1 mm (Hellma 110-QS 1mm, Paris, France).

Sample temperature was regulated at 20°C. Data were collected in continuous scan mode with a data pitch of 0.1 nm, a scanning speed of 50 nm.min⁻¹, 2 nm bandwith and 2 accumulations per sample. Sample Data were collected as raw ellipticity (ψ in mdeg) and converted to mean residue ellipticity (MRE or [θ]) in deg.cm².dmol⁻¹.residue⁻¹ using the following equation:

$$[\theta] = \frac{\psi \times 10^{-3}}{res \times l \times c}$$

Where *res* is the number of residues in the oligomer, l is the pathlength in cm, and c is the Oligourea concentration in dmol.cm⁻³.

X-Ray diffraction studies

Data collections were performed at the IECB X-ray facility at room temperature on rotating anode sources. Crystal structures of compounds **2** and **3** were solved from data collected on a Rigaku micromax MM07 equiped with a partial chi goniometer and a detector IP RAPID .The data for the crystal structure of compound **5** were collected on a Bruker microstar X8 PROTEUM with a classical kappa geometry and Platinum135 CCD camera. All the statistics are compiled in table S7. All structures were solved by the ab-initio method implemented in SHELXD and refined with SHELXL (Sheldrick, G.M. Acta Cryst. A64, 2008, 112-122). Fullmatrix least-squares refinement was performed on F² for all unique reflections, minimizing $w(Fo^2 - Fc^2)^2$, with anisotropic displacement parameters for non-hydrogen atoms. The positions of the H atoms were deduced from coordinates of the non-H atoms. The non-H atoms were refined with anisotropic temperature parameters. H atoms were included for structure factor calculations but not refined

CCDC 922576 (2), CCDC 922577 (3) and CCDC 922578 (5) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via <u>www.ccdc.cam.ac.uk/data_request/cif</u>.

Compound	2	3	5
CCDC code			
Formula	C23.50 H48.50 N7.50 O5 S0.50	C90 H183 N30 O19	C25 H50.50 N8.50 O6
Μ	532.23	1989.66	566.24
Crystal system	triclinic	monoclinic	monoclinic
Space group	P1	P2(1)	P2(1)
a/Å	9.866(2)	14.2758(7)	10.658(2)
b/Å	10.472(2)	10.5020(7)	18.807(4)
c/Å	16.893(3)	39.927(3)	17.672(4)
α/ο	96.60(3)	90.00	90.00
β/ο	95.35(3)	90.983(6)	106.39(3)
γ/ο	113.94(3)	90.00	90.00
$V/\text{\AA}^3$	1565.7(5)	5985.1(6)	3398.2(12)
Т /К	293(2)	293(2)	293(2)
Ζ	2	2	4
ρ/g cm ⁻¹	1.129	1.104	1.107
size (mm)	0.2x 0.02x 0.01	0.05x 0.05x 0.01	0.1x0.02x0.01
λ/ Å	1.54178	1.54178	1.54178
μ/mm ⁻¹	0.951	0.641	0.656
Independent reflections	8949	9534	5521
measured reflections	18961	56320	13718
parameters/restraints	677/3	1284/1	708/4
<i>R</i> 1, <i>wR</i> 2	0.1196/ 0.3192	0.0459/ 0.0833	0.0676/0.1834
goodness of fit	1.113	0.874	1.072

Table S8: Crystallographic data for compounds 2, 3 and 5