Electronic Supplementary Information

A Reverse Strategy for Synthesis of Nucleosides

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Contents

1. General Information	S 2
2. General procedure for <i>N</i> -glycosidation of <i>n</i> -pentenyl orthoesters	S2
3. Synthesis and characterization of perbenzoylated <i>ribo</i> nucleosides.	S 3
4. Synthesis and characterization of C3' and C5' differentiated <i>ribo</i> -NPOEs.	S5
5. Assembly of nucleobases with C3' and C5' differentiated <i>ribo</i> -NPOEs.	S12
6. Preparation and N-glycosylation of xylo n-pentenyl orthoesters	S17
8. References	S18
9. ¹ H NMR and ¹³ C NMR Spectra	S19

1. General Information

Chemicals were purchased and used without further purification. Dry solvents were obtained by distillation using standard procedures. Unless stated otherwise, reactions were carried out under a dry argon atmosphere in vacuum-flame dried glassware. Detection was by examination under UV light (254 nm) and by charring with 10% sulfuric acid in ethanol. Flash column chromatography was performed using silica gel [Merck, 230–400 mesh (40–63 μ m)]. Extracts were concentrated *in vacuo* using both a Büchi rotary evaporator (bath temperatures up to 40 °C) at a pressure of either 15 mmHg (diaphragm pump) or 0.1 mmHg (oil pump), as appropriate, and a high vacuum line at room temperature. ¹H NMR and ¹³C NMR spectra were measured in the solvent stated at 200 and 50 MHz respectively. Chemical shifts are quoted in parts per million from residual solvent peak (CDCl₃: ¹H - 7.26 ppm and ¹³C - 77.16 ppm) and coupling constants (*J*) given in Hertz. Multiplicities are abbreviated as: b (broad), s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) or combinations thereof. The units of the specific rotation, (deg·mL)/(g·dm), are implicit and are not included with the reported value. Concentration *c* is given in g/100 mL. Starting *n*-pentenyl orthoesters **1**, **6** and **25** were prepared following previously described procedures^[1] and the exchange of substituents at the different hydroxyl groups was carried out following routine procedures^[2].

2. General procedure for N-glycosidation with n-pentenyl orthoesters.



General procedure for silvlation of nucleobases. A suspension of the nucleobase (1 mmol), in hexamethyldisilazane (HMDS) (6 mmol) was treated under a N₂ atmosphere with TMSOTf (10% mol), then heated to 85 °C and kept at that temperature until reaction mixture became clear (\sim 2–3 hr). The reaction mixture was then concentrated *in vacuo* and the crude was azeotroped 2–3 times with anhydrous CH₂Cl₂ under nitrogen. The obtained solids were kept under high vacuum overnight to remove traces of solvent.

General experimental procedure for *N*-glycosidation. To a stirred solution of the appropriate NPOE (1.0 mmol) in anhydrous acetonitrile (1 mL) was added a solution of silylated base (1.2 mmol) in acetonitrile (2 mL) at 0 $^{\circ}$ C and allowed to stir for 10 min at this temperature. To this reaction mixture was

added a solution of NIS (1.1 mmol) and Yb(OTf)₃ (30 mol-%) in CH₃CN (2 mL) at 0 °C and the mixture was allowed to stir at room temperature for the appropriate time (\sim 3–10 h, analysis by TLC). After which, the reaction mixture was quenched with saturated sodium thiosulphate solution (5 mL) and extracted with CH₂Cl₂ (3 x 10 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuum. The resulting crude mixture was purified by column chromatography on silica gel to afford the pure nucleoside.

3. Synthesis and characterization of perbenzoylated ribo nucleosides.



2,3,5-tri-*O***-benzoyl-β-D-ribofuranosyl-uracil (11a):** The compound was prepared according to general procedure with uracil **7a** (1.2 mmol, 134 mg) and orthoester **6** (1 mmol, 530 mg) over a course of 4 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc (2:3) to give the desired product **11a** (511 mg, 92% yield) as a colorless crystalline solid. ¹H NMR (200 MHz, CDCl₃): δ 8.60 (bs, 1H), 8.15–7.90 (m, 6H), 7.65–7.30 (m, 10H), 6.31 (d, J = 4.4 Hz, 1H), 5.74 (dd, J = 5.4 Hz, 6.0 Hz, 1H), 5.60 (d, J = 8.2 Hz, 1H), 4.87–4.62 (m, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.3, 165.6, 165.5, 163.4, 150.5, 139.9, 134.0, 134.0, 133.9, 130.2, 130.1, 129.9, 129.5, 129.0, 128.8, 128.6, 103.7, 88.4, 80.4, 74.0, 71.4, 64.0. Proton and carbon NMR were consistent with literature data.^[3]



2,3,5-tri-*O***-benzoyl-β-D-ribofuranosyl-thymine (11b):** The compound was prepared according to general procedure with thymine **7b** (0.6 mmol, 76 mg) and orthoester **6** (0.5 mmol, 265 mg) over a course of 4 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 2:3 to give the desired product **11b** (276 mg, 97% yield). ¹H NMR (200 MHz, CDCl₃): δ 8.80 (bs,1H), 8.18–8.14 (m, 2H), 8.0–7.90 (m, 4H), 7.66–7.32 (m, 9H), 6.43 (d, J = 6.2 Hz, 1H), 5.9 (dd, J = 3.2 Hz, J = 3.4 Hz, 1H), 5.75 (t, J = 5.8 Hz, 1H), 4.82–4.94 (m, 1H), 4.72–4.60 (m, 2H), 1.57 (s, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.2, 165.6, 165.6, 164.0, 150.8, 135.2, 134.0, 133.9, 130.2, 130.1, 129.9,

129.5, 129.1, 128.9, 128.8, 128.6, 112.4, 87.3, 80.8, 73.7, 71.7, 64.2, 12.3. Proton and carbon NMR were consistent with literature data.^[4]



2,3,5-tri-*O***-benzoyl-**β**-**D**-ribofuranosyl-***N***4-benzoyl cytosine (12):** The compound was prepared according to general procedure from *N***4**-benzoylcytosine **8** (0.6 mmol, 129 mg) and orthoester **6** (0.5 mmol, 265 mg) over a course of 2 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 3:7 to give the coupling product **12** (287 mg, 82% yield). ¹H NMR (200 MHz, CDCl₃): δ 8.78 (bs, 1H), 8.16–8.03 (m, 2H), 8.0–7.82 (m, 7H), 7.66–7.21 (m, 13H), 6.43 (d, *J* = 6.2 Hz, 1H), 5.9 (d, *J* = 4.4 Hz, 1H), 5.88–5.81 (m, 2H), 4.94–4.68 (m, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.3, 165.4, 144.3, 133.8, 133.4, 130.2, 130.0, 129.8, 129.5, 129.2, 128.9, 128.6, 127.8, 89.7, 80.9, 74.9, 71.2, 63.8, 29.9. Proton and carbon NMR were consistent with literature data.^[4,5]



2,3,5-tri-*O***-benzoyl-β-D-ribofuranosyl-2,6-dichloropurine** (13a): The compound was prepared according to general procedure from 2,6-dichloropurine 9a (0.6 mmol, 113 mg) and orthoester **6** (0.5 mmol, 265 mg) over a course of 4 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 85:15 to give the coupling product 13a (269 mg, 85% yield). ¹H NMR (200 MHz, CDCl₃): δ 8.28 (s, 1H), 8.07–7.90 (m, 6H), 7.64–7.33 (m, 9H), 6.48 (d, *J* = 5.2 Hz, 1H), 6.15 (m, 2H), 4.96–4.70 (m, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.2, 165.5, 165.4, 153.5, 152.8, 152.3, 144.5, 134.2, 134.1, 133.8, 131.6, 130.0, 129.8, 129.3, 128.9, 128.8, 128.8, 128.3, 87.4, 81.6, 74.5, 71.8, 63.7. Proton and carbon NMR were consistent with literature data.^[6]



2,3,5-tri-*O***-benzoyl-**β**-**D**-ribofuranosyl-2-amino-6- chloro purine (13b):** The compound was prepared according to general procedure from 2-amino-6-chloropurine **9b** (0.6 mmol, 101 mg) and orthoester **6**(0.5 mmol, 265 mg) over a course of 3 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 4:1 to give the coupling product **13b** (214 mg, 70% yield). ¹H NMR (200 MHz, CDCl₃): δ 8.01–7.80 (m, 6H),7.52–7.05 (m, 10H), 6.48–6.31 (m, 2H), 6.28 (d, J = 4.4 Hz, 1H), 5.20 (bs, 1H), 4.91–4.60 (m, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.3, 165.5, 165.3, 159.4, 153.3, 152.0, 141.3, 134.0, 133.9, 133.6, 130.0, 129.9, 129.8, 129.4, 128.9, 128.7, 128.6, 125.9, 87.5, 80.5, 73.7, 71.5, 63.4. Proton and carbon NMR were consistent with literature data.^[4]



2,3,5-tri-*O***-benzoyl-**β**-**D**-ribofuranosyl-adenine (13c):** The compound was prepared according to general procedure from adenine **9c** (0.6 mmol, 81 mg) and orthoester **6** (0.5 mmol, 265 mg) over a course of 18 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc (100% EtOAc) to give the coupling product **13c** (87 mg, 30% yield). ¹H NMR (200 MHz, CDCl₃): δ 8.36 (s, 1H), 8.12–7.84 (m, 7H), 7.62–7.21 (m, 10H), 6.46–6.32 (m, 2H), 6.30–6.22 (t, *J* = 4.8 Hz, 1H), 5.72 (bs, 2H), 4.88–4.62 (m, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 166.4, 165.5, 165.3, 156.0, 153.5, 149.9, 139.3, 133.8, 133.6, 130.0, 129.9, 129.6, 129.0, 128.7, 120.3, 87.1, 80.8, 76.6, 74.2, 71.7, 63.9. Proton and carbon NMR were consistent with literature data.^[7]

4. Preparation of differentially protected ribo n-pentenyl orthoesters.



β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (15a): To a well stirred solution of *n*-pentenyl orthoester **6** (800 mg, 1.54 mmol) in methanol (10 mL) was treated with triethylamine (1.1 mL, 7.72 mmol) and water (0.5 mL). The reaction mixture was heated to 65 °C and then left stirring at this temperature for 48 hours after which it was determined to be complete by TLC. The solvent was removed under vacuum and the residue was purified by silica gel column chromatography using hexane:EtOAc 2:3 to afford the product **15a** (342 mg, 72 %) as a colorless thick syrup. $[\alpha]_D^{25}$: + 31.7 (*c* 1.0, CHCl₃), ¹H NMR (300 MHz, CDCl₃): δ 7.60–7.55 (m, 2H), 7.31–45 7.29 (m, 3H), 5.95 (d, *J* = 4.2 Hz, 1H), 5.70 (m, 1H), 4.90 (m, 2H), 4.71 (m, 1H), 3.93 (m, 1H), 3.74 (dd, *J* = 12.5, 3.4 Hz, 1H), 3.53 (dd, *J* = 12.5, *J* = 3.4 Hz, 1H), 3.43 (m, 1H), 3.34 (m, 2H), 2.01 (m, 2H), 1.59 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 138.4, 137.2, 129.9, 128.8 (x 2), 126.4 (x 2), 124.0, 115.4, 104.5, 81.4, 80.0, 71.3, 63.1, 60.8, 53.9, 30.7, 29.0. MS (API-ES positive mode): 345.2 [M+Na]⁺.



3,5-di-*O*-(*tert***butoxycarbonyl-L-valinoyl)-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (15b)**: A solution of diol **15a** (100 mg, 0.032 mmol), EDCI (124 mg, 0.645 mmol), Fmoc valine (240.4 mg, 0.708 mmol), and DMAP (4.0 mg, 0.032 mmol) in anhydrous DMF (10 mL) was stirred under an N₂ atmosphere at room temperature overnight. The reaction mixture was diluted with water (20 mL) and then extracted with EtOAc (3 x 10 mL) and the organic phase was dried (Na₂SO₄), concentrated in vacuum. The resulting crude mixture was purified on silica gel column using hexane/EtOAc 78:22 to give a colorless syrup **15b** (230 mg, 75%): $[\alpha]_D^{25}$ + 46.0 (*c* 0.2, CHCl₃); ¹H NMR (200 MHz, CDCl₃): δ 7.75 (d, *J* = 7.2 Hz, 2H), 7.60 (m, 6H), 7.3 (m, 11H), 6.1 (d, *J* = 3.6 Hz, 1H), 5.75 (m, 1H), 5.3 (t, *J* = 7.6 Hz, 2H), 5.2–4.9 (m, 4H), 4.75 (dd, *J* = 5.2 Hz, *J* = 4.8 Hz, 1H), 4.10–4.0 (m, 10H), 3.42 (m, 2H), 2.1 (m, 4H), 1.6 (pentate, *J* = 7.4 Hz, 2H), 0.95 (d, *J* = 6.6 Hz, 6H), 0.85 (d, *J* = 7.0 Hz, 6H); ¹³C NMR (80 MHz, CDCl₃): δ 171.9, 171.3, 144.1, 144.0, 141.6, 138.2, 137.2, 129.6, 128.4, 128.0, 127.3, 126.3, 125.3, 124.3, 120.2, 115.2, 104.4, 77.9, 76.1, 72.9, 67.3, 62.7, 62.6, 59.2, 47.4, 31.5, 30.4, 28.8, 19.2, 17.8.



3,5-di-*O***-benzyl**–**β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate)** (**15c)**: Sodium hydride (774 mg, 32.25 mmol) was added to a solution of diol **15a** (2.5 g, 8.06 mmol) in dry DMF (30 mL) at 0 °C under N₂ atmosphere. The mixture was stirred at this temperature for 25–30 min, and then benzyl bromide (2.10 mL, 17.74 mmol) was added dropwise for a period of 10 min. The reaction was allowed to warm to room temperature and stirred for an additional 3 h, after which it was quenched with ice cold water (5 mL) and stirred for 30 min. Added more water (30 mL) and extracted with diethyl ether (4 x 40 mL). The organic phase was washed with brine, dried over Na₂SO₄, filtered and evaporated in vacuum. Flash chromatography using hexane/EtOAc 88:12 of the residual syrup afforded less polar compound **15c** (2.8 g, 71 %). [α]_D²⁵: +107.4 (*c* 1.0, CHCl₃) ¹H NMR (300 MHz, CDCl₃): δ 7.73–7.70 (m, 3H), 7.37–7.25 (m, 12H), 6.06 (d, *J* = 3.9 Hz, 1H), 5.80 (tdd, *J* = 16.8, *J* = 10.1, *J* = 6.5 Hz, 1H), 5.05–4.95 (m. 2H), 4.84–4.75 (m, 1H), 4.60–4.44 (m, 4H), 3.94 (dd, *J* = 9.1, *J* = 4.5 Hz, 1H), 3.86 (m, 1H), 3.70 (dd, *J* = 11.4, *J* = 1.9 Hz, 1H), 3.52 (dd, *J* = 11.3, *J* = 3.4 Hz, 1H), 3.41 (t, *J* = 6.5 Hz, 2H), 2.10 (m, 2H), 1.68 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 138.4, 138.2, 137.9, 137.3, 129.4, 128.8, 128.7 (x 2), 128.6 (x 2), 128.4 (x 2), 128.3 (x 2), 128.0 (x 2), 127.9, 126.6 (x 2), 123.9, 115.2, 104.8, 78.6, 78.0, 77.0, 73.7, 72.5, 67.7, 62.8, 30.5, 29.0; MS (API-ES positive mode): 525.5 [M+Na].



5-*O-tert***butyldiphenylsilyl-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (21a):** To a stirred solution of diol **15a** (2 g, 6.45 mmol) which had been pre-dried under vacuum in a flame-dried flask for 1 h, was dissolved in anhydrous CH₂Cl₂. The reaction mixture was cooled to 0 °C, after which Et₃N (1.8 mL, 12.9 mmol) and DMAP (78 mg, 0.64 mmol) was added and allowed to stir for 15–20 min. TBDPSCI (1.85 mL, 7.095 mmol) was then slowly added for a period of 20 min after which the resulting solution was allowed to warm to room temperature and stir for a further 3 h. The reaction mixture was washed with water (50 mL) and extracted with CH₂Cl₂ (3 x 40 mL). The combined organic layer was dried over sodium sulfate, concentrated and the residue was purified by flash chromatography using hexane/EtOAc 4:1 to afford **21a** (2.48 g, 69 %) as a colorless oil. $[\alpha]_D^{25}$: + 17.3 (*c* 0.13, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 7.72–7.63 (m, 6H), 7.44–7.35 (m, 9H), 6.12 (d, *J* = 4.0 Hz, 1H), 5.92–5.69 (tqt, *J* = 7.0 Hz, *J* = 6.6 Hz, 1H), 5.10–4.94 (m, 2H), 4.89–4.82 (t, *J* = 5.2 Hz, 1H), 4.22–4.08 (m, 1H), 3.95–3.71 (m, 2H), 3.58–3.38 (m, 3H), 2.21–2.01 (m, 3H), 1.80–1.59 (m, 2H),1.2 (s, 9H); ¹³C NMR (80 MHz, CDCl₃): δ 138.2, 135.8, 129.9, 129.6, 128.5, 127.9, 126.2, 115.2, 104.5, 81.9, 79.8, 77.9, 71.4, 62.7, 62.2, 30.4, 28.8, 19.5.



3-O-benzyl-5-O-*tert***butyldiphenylsilyl-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate)** (15d): Sodium hydride (43.8 mg, 1.82 mmol) was added to a solution of compound **21** (500 mg, 0.912 mmol) in dry N,N-dimethyl formamide (20 mL) at 0 °C under N₂ atmosphere. The mixture was stirred at this temperature for 20 min, and then benzyl bromide (0.12 mL, 1.00 mmol) was added. The reaction was allowed to warm to room temperature and stirred for an additional 2 h. The reaction mixture was quenched with cold water (30 mL) and extracted with ether (3 x 30 mL). The organic phase was washed with brine, dried over Na₂SO₄, filtered and evaporated. Flash chromatography of the residual brown syrup afforded **15d** (405 mg, 70%) as colorless liquid. $[α]_D^{25}$: 10.3 (*c* 0.25, CHCl₃), ¹H NMR (200 MHz, CDCl₃): 7.8–7.6 (m, 7H), 7.4–7.23 (m, 13H), 6.06 (d, *J* = 4.0 Hz, 1H), 5.9–5.7 (tqt, *J* = 6.8 Hz, *J* = 8.0 Hz, 1H), 5.1–4.92 (m, 2H), 4.88–4.82 (t, *J* = 4.0 Hz, 1H), 4.82–4.76 (d, *J* = 10.0 Hz, 1H), 4.64–4.56 (d, *J* = 10.0 Hz, 1H), 4.16–4.04 (dd, *J* = 3.8 Hz, 1H), 3.94–3.66 (m, 3H), 3.48–3.36 (t, *J* = 5.4 Hz, 2H), 2.2–2.0 (m, 2H), 1.78–1.69 (m, 2H), 1.0 (s, 9H).



3-*O*-(*tert*butoxycarbonyl-L-valinoyl)-5-*O*-*tert*butyldiphenylsilyl-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (15e): A solution of compound 21 (200 mg, 0.364 mmol), Boc-Valine (87.12 mg, 0.4 mmol), DCC (112.6 mg, 0.546 mmol) and DMAP (22.2 mg, 0.182 mmol) in anhydrous CH₂Cl₂ was stirred at room temperature overnight. The reaction mixture was diluted with CH₂Cl₂ (20 mL) and washed with water (25 mL). The aqueous layer was extracted with CH₂Cl₂ (2 x 10 mL) and the combined organic layer was dried over Na₂SO₄, concentrated under vacuum. The obtained crude mixture was purified by silica gel column chromatography using hexane/EtOAc 8:2 to afford **15e** (226 mg, 83%) as colorless syrup. $[\alpha]_D^{25}$: +70.5 (*c* 0.21, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 7.60 (m, 6H), 7.35 (m, 9H), 6.15 (d, *J* = 4.0 Hz, 1H), 5.8 (m, 1H). 5.0 (m, 4H), 4.3 (m, 1H), 3.85 (m, 2H), 3.6 (dd, *J* = 3.2 Hz, *J* = 3.6 Hz, 1H), 3.45 (t, *J* = 6.6 Hz, 2H), 2.1 (m, 3H), 1.7 (m, 2H), 1.46 (s, 9H), 1.05 (s, 9H), 0.93 (d, *J* = 6.6 Hz, 3H), 0.8 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (80 MHz, CDCl₃): δ 171.7, 155.7, 138.2, 137.7, 135.8, 135.7, 133.3, 133.1,

130.0, 129.4, 128.3, 128.0, 127.9, 126.4, 124.2, 115.2, 104.7, 80.0, 79.2, 78.0, 72.1, 62.6, 61.7, 58.6, 31.7, 30.4, 29.9, 28.8, 28.6, 27.0, 19.5, 19.1, 17.7.



3-O-dibenzylphosphoryl-5-O-tertbutyldiphenylsilyl-β-D-ribofuranose-1,2-(pent-4-enyl

orthobenzoate) (15f): To a well stirred solution of dibenzylchlorophosphate (194.0 mg, 0.654 mmol) in anhydrous acetonitrile (2 mL) and under N₂ atmosphere was added at -78 °C a solution of NPOE 15a (120 mg, 0.22 mmol) and DMAP (133.5 mg, 1.09 mmol) in anhydrous CH₂Cl₂ (10 mL). The mixture was allowed to warm to room temperature and then stirred overnight. The solvents were removed under vacuum and the crude was purified by silica gel column chromatography using hexane/EtOAc 2:3 to give 15f (90 mg, 51%) as thick syrup. $[\alpha]_D^{25}$: +16.83 (*c* 0.65, CHCl₃); ¹H NMR (200 MHz, CDCl₃): δ 7.6 (m, 6H), 7.40–7.24 (m, 19H), 6.05 (d, *J* = 4.0 Hz, 1H), 5.7 (m, 1H), 5.1–4.90 (m, 7H), 4.80 (m, 1H), 3.9–3.6 (m, 3H), 3.4 (dd, *J* = 6.6 Hz, *J* = 5.4 Hz, 2H), 2.1 (m, 2H), 1.7 (m, 2H), 1.05 (s, 9H); ³¹P NMR (80 MHz, CDCl₃): δ 138.2, 137.6, 135.9, 135.8, 133.5, 133.2, 129.9, 129.4, 128.8, 128.4, 128.1, 127.9, 127.9, 126.3, 124.2, 115.1, 104.3, 79.6, 79.4, 78.8, 74.1, 69.8, 69.7, 62.6, 61.1, 30.5, 28.8, 26.9, 19.5.



5-*O*-dibenzylphosphoryl-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (21b): To a solution of dibenzylphosphoryl chloride (286.6 mg, 0.966 mmol) in anhydrous CH₂Cl₂ (3 mL) was added under a N₂ atmosphere and at –78 °C a solution of diol **16a** (100 mg, 0.322 mmol) and DMAP (118 mg, 0.966 mmol) in dry CH₂Cl₂ (5 mL). The reaction was allowed to warm to room temperature and then stirred overnight. The solvent was evaporated under vacuum and the mixture was purified by silica gel column chromatography using hexane/EtOAc 1:4 to afford C5'phosphorylated compound **22b** (97 mg, 53%). $[\alpha]_D^{25}$: + 60.0 (*c* 0.1, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 7.7–7.6 (m, 2H), 7.5–7.2 (m, 13H), 5.97 (d, J = 4.0 Hz, 1H), 5.9–5.7 (tqt, J = 7.0 Hz, J = 6.6 Hz, 1H), 5.1–4.9 (m, 6H), 4.78–4.73 (t, J = 4.4 Hz, 1H), 4.3–3.85 (m, 3H), 3.6–3.5 (m, 1H), 3.45–3.3 (m, 2H), 2.5 (d, J = 10.0Hz, 1H), 2.2–2.0 (q, J = 7.0Hz, 2H), 1.75–1.6 (m, 2H). ³¹P NMR (80 MHz, CDCl₃): δ 0.50; ¹³C NMR (80 MHz, CDCl₃):138.2, 129.6,

129.0, 128.8, 128.6, 128.4, 128.2, 126.3, 126.2, 115.2, 104.2, 79.5, 79.4, 71.4, 69.8, 69.7, 69.6, 65.6, 65.5, 62.9, 30.4, 28.8.



5-*O***-dibenzylphosphoryl-3-O-(9-fluorenylmethoxycarbonyl-L-valinoyl)-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (15g):** A solution of phosphorylated orthoester **22b** (130 mg, 0.228 mmol) in dry DMF was treated with Fmoc-valine (85.1 mg, 0.250 mmol), EDCI (65.6 mg, 0.342 mmol), and DMAP (14 mg, 0.114 mmol) and the resulting mixture was allowed to react for 12 h. The reaction crude was then washed with water and extracted with ethyl acetate (3 x 15 mL). The combined organic layer was concentrated and purified by silica gel column chromatography using hexane/EtOAc 1:1 to afford compound **11g** (122 mg, 61%). $[\alpha]_D^{25}$: + 60.0 (*c* 0.1, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 7.76 (d, *J* = 7.4 Hz, 1H), 7.60 (m, 3H), 7.4–7.1 (m, 18H), 6.0 (d, *J* = 4.0 Hz, 1H), 5.73 (m, 1H), 5.2–4.85 (m, 8H), 4.73 (dd, *J* = 5.4 Hz, 5.0 Hz, 1H), 4.5–4.13 (m, 5H), 3.98–3.83 (m, 2H), 3.42–3.25 (m, 2H), 2.1 (m, 3H), 1.6 (m, 2H), 1.0–0.78 (dd, *J* = 6.6 Hz, *J* = 7.0 Hz, 6H), ¹³C NMR (80 MHz, CDCl₃): δ 171.8, 143.95, 141.55, 138.16, 137.27, 129.51, 128.79, 128.39, 128.2, 128.1, 127.95, 127.29, 126.25, 125.23, 120.21, 115.73, 104.5, 77.65, 72.35, 69.66, 67.28, 64.96, 62.71, 59.13, 47.44, 31.54, 30.37, 28.73, 19.1, 17.7; ³¹P NMR (80 MHz, CDCl₃): δ 0.10.



3,5-di-*O*-chloroacetyl- β -D-ribofuranose-1,2-(pent-4-enyl orthochloroacetate (25a) To a cooled (0 °C) suspension of D- ribose (1 g, 6.66 mmol) in anhydrous methanol (24 ml), was added dropwise acetyl chloride (0.42 mL). The reaction mixture was allowed to warm to room temperature and stirred for 1 h. Pyridine (5 mL) was slowly added, the mixture was concentrated *in vacuo* and the residue co-evaporated with chloroform. The resulting methyl riboside (1.04 g) was dissolved in anhydrous DMF (10 mL), cooled to 0 °C, treated with sodium bicarbonate (2.52 g, 0.03 mol) and then with chloroacetyl chloride (5.21 g, 0.03 mol) dissolved in dry DMF (2 mL). The mixture was vigorously stirred overnight at rt. Water was added and after stirring for 30 min., it was extracted with diethyl ether (3 x 20 mL). The

combined organic layer was dried over MgSO₄, filtered, concentrated and the residue purified by silica gel column chromatography using hexane/EtOAc 4:1 to afford methyl tri-O-chloroacetyl Dribofuranoside (600 mg, 28%). This compound (500 mg, 1.27 mmol) was dissolved in acetic acid (2 mL) and HBr (2.5 mL, 45% in acetic acid) was added. The reaction vessel was tightly stoppered and stirred at rt for 1 hour. The reaction mixture was then diluted with CH₂Cl₂(25 mL) and ice-cold water was added (20 mL). The organic layer was washed with cold water (20 mL), cold saturated NaHCO₃ (2 x 10 mL) and dried. After evaporation of solvent, the corresponding glycosyl bromide was obtained in approximate yield 71 % (400 mg) and used immediately in the next step without any further purification. The crude glycosyl bromide (400 mg, 1.0 mol) was dissolved in dry CH₂Cl₂ (10 mL), and 4-penten-1-ol (0.208 mL, 2.0 mmol) and 2,6-lutidine (0.257 mL, 2.2 mol) were added followed by Bu₄NI in 3 lots (37.08 mg, 0.1 mol) in 30 min intervals at room temperature. The reaction mixture was stirred for 12 hours and then was washed with cold water (3 x 20 mL), brine (10 mL), dried (Na₂SO₄) and concentrated. The obtained crude was purified by flash column chromatography on silica gel using hexane/EtOAc 4:1 to afford orthoester **25a** (267 mg, 66.4%). ¹H NMR (200 MHz, CDCl₃): δ 6.08-6.0 (d, *J*=3.2 Hz, 1H), 5.9-5.7 (m, 1H), 5.1-4.92 (m, 3H), 4.84-4.76 (dddd, J=1.6 Hz, J=1.4 Hz, J=1.4 Hz, J=1.4 Hz, 1H), 4.6-4.48 (m, 2H), 4.3-4.23 (m, 1H), 4.19 (d, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 3.77-3.54 (t, J=1.4 Hz, 2H), 4.12-4.11 (d, J=1.6 Hz, 2H), 5.77-3.54 (t, J=1.4 Hz, 3H), 5.77-3.54 (t, J=1.4 Hz, 2H), 3.50-3.44 (t, J = 5.4 Hz 2H), 2.17-2.05 (m, 2H), 1.74-1.6 (m, 2H); ¹³C NMR (80 MHz, CDCl₃): δ 167.1, 166.8, 137.9, 122.9, 115.4, 104.9, 78.1, 73.7, 63.7, 62.4, 45.0, 40.7, 40.5, 30.2, 28.6.



3,5-di-*O*-benzoyl- β -D-ribofuranose-1,2-(pent-4-enyl orthochloroacetate (25b): To a well stirred solution of compound 25a (250 mg, 0.56 mmol) in MeOH (15 mL) was added NaOMe (0.5 ml, 1.0 mmol) at rt. The reaction mixture was then allowed to stir for 45 min and then concentrated under vacuo. The residue was purified by silicagel column chromatography using hexane/EtOAc 2:3 as eluent to give the corresponding diol (139 mg, 84.6%).

A stirred solution of diol (100 mg, 0.33 mmol) in anhydrous CH_2Cl_2 (5 mL) was cooled to 0 °C and treated with Et_3N (0.16 mL, 1.209 mmol), DMAP (78 mg, 0.64 mmol) and BzCl (0.10 ml, 0.88 mol). The mixture was allowed to stir for 15-20 min and then was washed with cold water (10 mL), extracted in CH_2Cl_2 (3 x 10 mL) and the combined organic layers were dried (Na₂SO₄), concentrated and the residue was purified by flash chromatography hexane/EtOAc 85:15 to afford orthoester **25b** (129 mg, 77%). ¹H

NMR (200 MHz, CDCl₃): δ 8.15-7.95 (m, 4H), 7.63-7.3 (m, 6H), 6.13-6.11 (d, *J* = 4.0 Hz, 1H), 5.88-5.67 (tqt, *J*=6.6 Hz , *J*=7.0 Hz, *J*=6.6 Hz, 1H), 5.2-5.15 (dd, *J*=4.0 Hz, *J*=4.2 Hz, 1H), 5.08-5.03 (m, 2H), 5.0-4.93 (m, 1H), 4.86-4.65 (m, 2H), 4.53-4.44 (dd, *J*=4.8 Hz, *J*=4.6 Hz, 1H), 3.8 (s, 2H), 3.51-3.44 (t, *J*=6.6 Hz, 2H), 2.15-2.04 (m, 2H), 1.72-1.58 (m, 2H); ¹³C NMR (80 MHz, CDCl₃): δ 166.3, 165.9, 137.9, 133.8, 133.4, 130.2, 129.9, 129.8, 129.1, 128.7, 128.6, 122.8, 115.4, 105.1, 78.8, 77.3, 73.4, 63.2. 62.3, 45.2, 30.2, 28.7.

1. Assembly of nucleobases with C3' and C5' differentiated ribo-NPOEs.



2-*O***-benzoyl-3,5-di-***O***-[9-fluorenylmethoxycarbonyl-L-valinoyl]-β-D-ribofuranosyl-uracil (16): The compound was prepared according to general procedure with uracil 7a (0.3 mmol, 34 mg) and orthoester 15b** (0.25 mmol, 241 mg) over a course of 3 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 3:1 to give the desired product **16** (205 mg, 83% yield). $[\alpha]_D^{25}$: + 23.7 (*c* 0.65, CHCl₃); ¹H NMR (200 MHz, CDCl₃): δ 8.83 (bs, 1H), 7.70 (d, *J* = 7.8 Hz, 2H), 7.75 (d, *J* = 7.4 Hz, 4H), 7.6–7.2 (m, 15H), 5.97 (s, 1H), 5.83 (d, *J* = 8.6 Hz, 1H), 5.60 (bs, 2H), 5.4 (d, *J* = 8.8 Hz, 1H), 5.23 (d, *J* = 8.8 Hz, 1H), 4.6 (d, *J* = 11.8 Hz, 1H), 4.5–4.0 (m, 10H), 2.1 (m, 2H), 1.0–0.6 (m, 12H); ¹³C NMR (80 MHz, CDCl₃): δ 172.1, 171.2, 165.6, 162.9, 156.6, 156.2, 150.2, 144.0, 143.9, 141.5, 140.8, 134.0, 130.1, 128.8, 128.5, 127.9, 127.3, 125.3, 125.2, 120.2, 103.8, 89.6, 80.4, 73.72, 71.4, 67.3, 63.8, 59.8, 59.2, 52, 47.4, 47.3, 31.4, 31.2, 29.9, 19.3, 19.1, 18.1, 17.8.



3,5-di-*O*-benzyl-2-*O*-benzoyl-5-O-*tert*-butyldiphenylsilyl- β -D-ribofuranosyl-uracil (17): The compound was prepared according to general procedure with uracil 7a (0.6 mmol, 34 mg) and orthoester **15c** (0.5 mmol, 301 mg) over a course of 2 h at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 4:1 to give the coupling product **17** (237 mg, 90%)

yield): $[\alpha]_D^{25}$: + 60.0 (*c* 1.0, CHCl₃); ¹H NMR (200 MHz, CDCl₃): δ 8.61 (bs, 1H), 8.1 (m, 2H), 7.8 (d, *J* = 8.0 Hz, 1H), 7.6 (m, 1H), 7.5–7.2 (m, 12H), 6.3 (d, *J* = 4.0 Hz, 1H), 5.5 (dd, *J* = 4.2 Hz, *J* = 4.6 Hz, 1H), 5.35 (dd, *J* = 2.2 Hz, *J* = 1.8 Hz, 1H), 4.65 (d, *J* = 11.8 Hz, 1H), 4.5–4.25 (m, 5H), 3.85 (dd, *J* = 2.2 Hz, *J* = 1.8 Hz, 1H), 3.6 (dd, *J* = 1.8 Hz, *J* = 2.2 Hz, 1H); ¹³C NMR (80 MHz, CDCl₃): δ 165.7, 164.0, 150.7, 140.5, 137.5, 133.8, 130.3, 129.4, 128.9, 128.8, 128.7, 128.5, 128.3, 128.2, 102.7, 88.12, 82.5, 76.1, 75.3, 73.9, 73.4, 68.9.



3-*O*-benzyl-2-*O*-benzoyl-5-O-*tert*-butyldiphenylsilyl-β-D-ribofuranosyl-uracil (18a): The compound was prepared according to general procedure with uracil 7a (0.3 mmol, 33 mg) and orthoester 15d (0.25 mmol, 162 mg) over a course of 2 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 3:1 to give the desired product 18a (126 mg, 75% yield) $[\alpha]_D^{25}$: +27.1 (*c* 0.5, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 8.9 (bs, 1H), 8.14–8.04 (m, 2H), 7.8 (d, *J* = 8.2 Hz, 1H), 7.71–7.11 (m, 18H), 6.37 (d, *J* = 4.4 Hz, 1H), 5.55 (t, *J* = 4.8 Hz, 1H), 5.4 (dd, *J* = 2.2 Hz, *J* = 2.2 Hz, 1H), 4.67 (d, *J* = 11.4 Hz, 1H), 4.48–4.38 (m, 2H), 4.26–4.02 (m, 2H), 3.8 (dd, *J* = 2.0 Hz, *J* = 2.2 Hz, 1H), 1.1 (s, 9H), ¹³C NMR (80 MHz, CDCl₃): δ 165.7, 163.2, 150.3, 139.9, 137.3, 135.9, 135.6, 133.7, 133.0, 132.4, 130.4, 130.3, 129.3, 128.7, 128.6, 128.2, 103.0, 87.4, 83.7, 75.8, 75.2, 73.6, 63.0, 27.3, 19.6.



2-*O***-benzoyl-5-O**-*tert*-butyldiphenylsilyl-3-*O*-(L-valinoyl)-β-D-ribofuranosyl-uracil (18b): The compound was prepared according to general procedure with uracil **7a** (0.3 mmol, 33 mg) and orthoester **15e** (0.25 mmol, 189 mg) over a course of 18 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 4:1 to give the Boc deprotected product **18b** (151 mg, 88% yield): $[\alpha]_D^{25}$: +5.897 (*c* 0.21, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 8.05–8.0 (m, 2H), 7.8–7.4 (m, 14H), 6.48 (d, *J* = 6.4 Hz, 1H), 5.72 (dd, *J* = 3.0 Hz, *J* = 3.2 Hz, 1H), 5.6 (m, 1H), 5.40 (d, *J* = 8.0 Hz,

1H), 4.22 (d, *J* = 3.0 Hz, 1H), 4.1–3.9 (dq, *J* = 11.8 Hz, *J* = 15.8 Hz, *J* = 1.8 Hz, 2H), 3.2 (d, *J* = 5.0 Hz, 1H), 1.9 (m, 1H), 1.5 (s, 9H), 0.85 (dd, *J* = 6.6 Hz, *J* = 6.6 Hz, 6H), ¹³C NMR (80 MHz, CDCl₃): δ 165.4, 163.0, 150.6, 139.5, 135.9, 135.5, 134.0, 132.9, 131.9, 130.5, 130.4, 130.1, 128.9, 128.8, 103.5, 85.8, 83.81, 74.2, 71.3, 63.7, 60.0, 32.1, 27.3, 19.6, 19.5, 17.4, FABMS: 686.3 (M+1).



3-*O***-dibenzylphosphoryl-2-***O***-benzoyl-5-O-***tert***-butyldiphenylsilyl-β-D-ribofuranosyl-uracil (18c): The compound was prepared according to general procedure with uracil 7a** (0.3 mmol, 33 mg) and orthoester **15f** (0.25 mmol, 205 mg) over a course of 1 hour at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 88:22 to give the product **18c** (180 mg, 85% yield) $[\alpha]_D^{25}$: -7.6 (*c* 0.21, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.27 (bs, 1H), 8.03 (d, *J* = 7.0 Hz, 1H), 7.70–7.62 (m, 5H), 7.61 (t, *J* = 7.5 Hz, 1H), 7.48–7.36 (m, 8H), 7.30–7.18 (m, 9H), 7.13–7.09 (m, 2H), 6.48 (d, *J* = 7.5 Hz, 1H), 5.52–5.46 (m, 1H), 5.36 (d, *J* = 6.0Hz, *J* = 8.0 Hz, 1H), 5.26–5.22 (m, 1H), 4.94–4.84 (m, 4H), 4.22 (bs, 1H), 3.98–3.76 (dddd, *J* = 1.5Hz, *J* = 12 Hz, *J* = 10 Hz, 2H), 1.12 (s, 9H), ³¹P NMR (80 MHz, CDCl₃): – 0.75, ¹³C NMR (80 MHz, CDCl₃): δ 165.4, 162.5, 150.3, 139.6, 135.9, 135.6, 133.9, 132.9, 131.9, 130.5, 130.5, 130.3, 128.9, 128.8, 128.8, 128.4, 128.3, 128.0, 103.4, 85.1, 84.9, 75.6, 75.5, 74.2, 74.1, 70.0, 69.9, 69.8, 63.9, 27.3, 19.6, HRMS (ESI): calculated for C₄₆H₄₇N₂O₁₀PSi + Na⁺ 869, found 869.2659.



5-O-dibenzylphosphoryl-2-O-benzoyl-3-O-(9-fluorenylmethoxycarbonyl-L-valinoyl)-β-D-

ribofuranosyl-uracil (19): The compound was prepared according to general procedure with uracil **7a** (0.3 mmol, 33 mg) and orthoester **15g** (0.25 mmol, 225 mg) over a course of 2 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 2:3 to give the

product **19** (139 mg, 60% yield): $[\alpha]_D^{25}$: -16.0 (*c* 0.1, CHCl₃), ¹H NMR (500 MHz, CDCl₃): δ 8.26 (bs, 1H), 7.95 (d, *J* = 7.5 Hz, 2H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.56–7.44 (m, 4H), 7.41–7.25 (m, 16H), 6.24 (d, *J* = 6.5 Hz, 1H), 5.54–5.38 (m, 2H), 5.18–5.03 (m, 5H), 4.42–4.14 (m, 6H), 2.16–2.04 (m, 1H), 0.92 (d, *J* = 7.0 Hz, 3H), 0.84 (d, *J* = 7.0 Hz, 3H), ¹³C NMR (80 MHz, CDCl₃): δ 170.8, 165.1, 162.15, 155.9, 149.9, 143.8, 143.6, 141.2, 139.4, 135.3, 133.8, 129.9, 128.8, 128.7, 126.6, 128.1, 127.7, 127.0, 124.9, 119.9, 103.4, 86.6, 77.4, 77.2, 72.9, 71.5, 69.9, 67.0, 66.3, 58.9, 41.1, 31.0, 29.6, 18.9, 17.4, ³¹P NMR (80 MHz, CDCl₃): δ 0.35, HRMS (ESI): calculated for C₅₀H₄₉N₃O₁₃P + H⁺930, found 930.2995.



2-*O***-benzoyl-5-***O***-***tert***butyldiphenylsilyl-β-D**-**ribofuranosyl-uracil (22):** The compound was prepared according to general procedure with uracil **7a** (0.3 mmol, 33 mg) and orthoester **21a** (0.25 mmol, 140 mg) over a course of 3 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 3:1 to give the product **22** (65 mg, 60.8% yield): $\alpha_{\rm lp}^{25}$: 14.5 (*c* 0.16, CHCl₃),¹H NMR (200 MHz, CDCl₃): δ 8.7 (bs, 1H), 8.04 to 8.08 (d,*J* = 8.0Hz, 2H), 7.8 to 7.85 (d, *J* = 8.0Hz, 1H), 7.3 to 7.76 (m, 13H), 6.36 (d, *J* = 4.8 Hz, 1H),5.4 to 5.49 (m, 2H), 4.68 (bs, 1H), 4.1 to 4.2 (m, 2H), 3.8 to 3.9 (d, *J* = 11.0 Hz, 1H), 2.6 (bs, 1H), 1.1 (s, 9H).¹³C NMR (80 MHz, CDCl₃): δ 166.0, 162.8, 150.2, 139.9, 135.9, 135.6, 134.0, 133.0, 132.3, 130.5, 130.4, 130.2, 128.9, 128.8, 128.3, 103.0, 86.7, 85.0, 76.9, 70.0, 63.4, 27.2, 19.6.



2-*O***-benzoyl-5-***O***-***tert***butyldiphenylsilyl β-D-ribofuranosyl-2,6-dichloropurine (23): The compound was prepared according to general procedure with 2,6-dichloropurine 9a** (0.3 mmol, 57 mg) and orthoester **21a** (0.25 mmol, 140 mg) over a course of 3 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 88:12 to give the product **23** (118 mg, 80.5% yield): ¹H NMR (200 MHz, CDCl₃): δ 8.43 (s, 1H), 8.1–8.0 (m, 2H), 7.7–7.3 (m, 13H), 6.44 (d, *J* = 5.0 Hz, 1H), 5.8 (t, *J* = 5.0 Hz, 1H), 4.9 (bs, 1H), 4.4–4.3 (m, 1H), 4.16–4.06 (dd, *J* = 2.4 Hz, *J* = 9.0 Hz, 1H), 3.96–3.86 (dd, *J* = 2.4 Hz, *J* = 10.0 Hz, 1 H), 2.3 (bs, 1H), 1.1 (s, 9H).



2,3,5-tri-*O***-chloroacetyl-β-D-ribofuranosyl-uracil (24):** The compound was prepared according to general procedure with uracil **7a** (0.3 mmol, 33 mg) and orthoester **25a** (0.25 mmol, 112 mg) over a course of 3 hours at -78°C. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 6:4 to give the product **24** (102 mg, 77% yield): ¹H NMR (200 MHz, CDCl₃): δ 12.0 (bs, 1H), 7.7 (d, *J*=3.4 Hz, 1H), 6.7 (s, 1H), 5.95 (d, *J*=3.4 Hz, 1H), 5.5-5.6 (m, 2H), 4.4-4.6 (m, 2H), 4.24-4.4 (m, 1H), 4.04-4.2 (m, 6H)..



3,5-di-*O*- **benzoyl-2**-*O*-**chloroacetyl-** β -**D**-**ribofuranosyl-uracil (26):** The compound was prepared according to general procedure with uracil **7a** (0.3 mmol, 33 mg) and orthoester **25b** (0.25 mmol, 125 mg) over a course of 3 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 1:1 to give the product 27 (99 mg, 69% yield): ¹H NMR (200 MHz, CDCl₃): δ 8.4 (bs. 1H), 7.95-8.1 (m, 4H), 7.3-7.7 (m, 7H), 6.2-6.1 (d, *J*=3.2 Hz, 1H), 5.82-5.5 (m, 3H), 4.84-4.5 (m, 3H), 4.05 (s, 2H).

6. Preparation and N-glycosylation of xylo n-pentenyl orthoesters.



5-O-tertbutyldiphenylsilyl- β -D-xylofuranose-1,2-(pent-4-enyl orthobenzoate) (27b): A well stirred solution of *n*-pentenyl orthoester 27a (3 g, 5.79 mmol) in methanol (50 mL) was treated with triethylamine (4.0 mL, 28.9 mmol) and water (0.5 mL). The reaction mixture was allowed to reflux

overnight after which it was determined to be complete by TLC. The solvent was removed under vacuum and the residue was purified by silica gel column chromatography using hexane/EtOAc 7:3 to afford the correspondig diol (1.51 g, 84 %) as a colorless gum. ¹H NMR (200 MHz, CDCl₃): δ 7.5–7.6 (m, 2H), 7.3– 7.4 (m, 3H), 6.2 (d, J = 4.0 Hz, 1H), 5.64–5.88 (m, 1H), 4.8–5.06 (m, 2H), 4.75 (d, J = 4.2 Hz, 1H), 4.20– 4.43 (m, 2H), 3.8–3.9 (m, 3H), 3.34–3.55 (m, 3H), 2.0–2.15 (m, 2H), 1.6–1.72 (m, 2H), ¹³C NMR (80 MHz, CDCl₃): 138.2, 137.2, 129.5, 128.5, 126.2, 123.0, 115.2, 105.2, 86.5, 79.9, 76.2, 62.9, 60.7, 30.4, 28.9. This material (1 g, 3.2 mmol) was pre-dried under vacuum in a flame-dried flask for 1 h and then dissolved in anhydrous CH₂Cl₂. The resulting solution was cooled to 0 °C, after which imidazole (435.7 mg, 6.4 mmol), DMAP (10 mol-%) and TBDPSCl (0.92 mL, 3.54 mmol) were added. The reaction mixture was allowed to warm to room temperature (rt) and stirred for another 3 h. The reaction mixture was washed with water (50 mL) and extracted with CH₂Cl₂ (3 x 40 mL). The combined organic layer was dried over sodium sulfate, concentrated and the residue was purified by flash chromatography using hexane/EtOAc 85:15 to afford **27b** (1.2 g, 68%) as colorless syrup. ¹H NMR (200 MHz, CDCl₃): δ 7.6 to 7.8 (m, 6H), 7.3 to 7.5 (m, 9H), 6.12 to 6.14 (d, J = 4.0 Hz, 1H), 5.73 to 5.9 (m, 1H), 5.0 to 5.1 (m, 2H), 4.86 (t, J = 4.6 Hz, 1H), 4.11 to 4.22 (m, 1H), 3.74 to 4.0 (m, 2H), 3.4 to 3.56 (m, 3H), 2.1 to 2.2 (m, 3H), 1.65 to 1.8 (pentet, J = 6.6 Hz, J = 7.0 Hz, 2H), 1.1 (s, 9H), ¹³C NMR (80 MHz, CDCl₃): 138.2, 137.4, 135.9, 135.8, 133.5, 133.4, 130.0, 129.6, 128.6, 128.0, 126.3, 123.8, 115.2, 104.5, 82.0, 79.8, 71.5, 62.8, 62.3, 30.5, 28.9, 27.1, 19.6.



5-*O-tert*butyldiphenylsilyl-3-azido-3-deoxy-β-D-ribofuranose-1,2-(pent-4-enyl orthobenzoate) (28) (27): To a stirred solution of 27b (300 mg, 0.547 mmol) in CH₂Cl₂ was added anhydrous pyridine (0.176 mL, 2.188 mmol) at 0 °C and allowed to stir for 10–20 min and added trifluoromethanesulfonic anhydride (0.110 mL, 0.656 mmol). The mixture was stirred until TLC (hexane/EtOAc 8:2) showed the complete formation of a single faster moving spot. The mixture was washed with water (20 mL) and extracted with CH₂Cl₂ (3 x 20 mL), dried over Na₂SO₄, concentrated under vacuum. The obtained crude mixture was co-evaporated with toluene (2x10 mL) and immediately proceeded for next reaction without any purification. Sodium azide (93.45 mg, 1.437 mmol) was added to a solution of compound **27c** (150 mg, 0.239 mmol) in dry DMF (12 mL) and then heated at 70 °C for 2 h. The reaction mixture was cooled to rt and poured into water (50 mL). The aqueous phase was extracted with diethyl ether (3 x 15 mL) and the combined organic layers were dried over sodium sulfate and concentrated under vacuum. Silica gel chromatography

of crude reaction mixture afforded azido compound **28** (108 mg, 79%). ¹H NMR (200 MHz, CDCl₃): δ 7.69–7.62 (m, 6H), 7.43–7.34 (m, 9H), 6.11–6.09 (d, J = 10.5Hz, 1H), 5.90–5.69 (tqt, J = 6.6 Hz, J = 6.8 Hz, 1H), 5.07–4.94 (m, 3H), 3.47–3.40 (t, J = 6.6 Hz, 2H), 2.18–2.07 (q, J = 7.2 Hz, 2H), 1.75–1.61 (pent, J = 8.2 Hz, J = 6.6 Hz, 2H), ¹³C NMR (80 MHz, CDCl₃): δ 138.2, 136.9, 135.8, 135.7, 133.3, 133.1, 130.0, 129.5, 128.4, 128.0, 127.9, 126.3, 124.1, 115.1, 104.4, 80.7, 79.1, 62.9, 61.7, 60.6, 30.4, 28.9, 27.0, 19.6.



5-*O*-*tert***butyldiphenylsilyl-3-deoxy-3-azido**-β-D-ribofuranosyl-uracil (29): The compound was prepared according to general procedure with uracil (0.3 mmol, 33 mg) and orthoester **28** (0.25 mmol, 146 mg) over a course of 2 hours at rt. The crude material was purified by column chromatography on silica gel using hexane/EtOAc 2:3 to give the product **29** (137 mg, 88% yield): $[\alpha]_D^{25}$: +54.0 (*c* 0.15, CHCl₃), ¹H NMR (200 MHz, CDCl₃): δ 8.71 (bs, 1H), 8.11–8.07 (m, 2H), 7.76–7.37 (m, 14H), 6.29–6.26 (d, *J* = 5.2 Hz, 1H), 5.68–5.62 (t, *J* = 5.4 Hz, 1H), 5.48–5.44 (d, *J* = 8.2Hz, 1H), 4.51–4.46 (t, *J* = 5.6 Hz, 1H), 4.13–4.07 (m, 2H), 3.87–3.80 (dd, *J* = 2.6 Hz, 1H), 1.17 (s, 9H).¹³C NMR (80 MHz, CDCl₃): δ 165.7, 163.1, 150.3, 139.8, 135.9, 135.6, 134.1, 132.7, 132.2, 130.6, 130.5, 130.4, 128.9, 128.6, 128.4, 128.3, 103.3, 87.2, 83.2, 75.9, 63.4, 60.5, 27.3, 19.6. LCMS: (M+Na) 634.

References

- [1] C. V. S. Ramamurty, P. Ganney, C. S. Rao and B. Fraser-Reid, J. Org. Chem. 2011, 76, 2245-2247.
- [2] P. J. Kocienski, Protecting Groups, 3rd Edn., 2005, Georg Thieme Verlag, Stuttgart.
- [3] I. Nowak, and M. J. Robins, Org. Lett. 2005, 7, 4903–4905.
- [4] Q. Zhang, J. Sun, Y. Zhu, F. Zhang and B. Yu, Angew. Chem. Int. Ed., 2011, 50, 4933–4936.

[5] a) D. H. Rammler and H. G. Khorana, J. Am. Chem. Soc. 1962, 84, 3112–3122. b) N. Shinomura, T. Matsutani and T. Mukaiyama, Bull. Chem. Soc. Jpn. 1994, 67, 3100–3106.

- [6] M. Hocek, A. Holy and H. Dvorakova, Collect. Czech. Chem. Commun. 2002, 67, 325–335.
- [7] I. Nowak, M. Conda-Sheridan and M. J. Robins, J. Org. Chem., 2005, 70, 7455-7458.



S19

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STANDARD 1H OBSERVE

Pulse Sequence: s2pul Solvent: COC13 Ambient temperature Mercury-200 "mercvx"

Relax. delay 1.000 sec Puise 32.9 degrees Acq. time 1.994 sec Width 3003.0 Hz 37 repetitions DBSERVE H1, 200.0803665 MHz DATA PROCESSING FT size 16384 Total time 7 min, 28 sec









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SURVEY PHOSPHORUS PARAMETERS

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "mercvx"

Pulse 45.3 degrees Acq. time 1.600 sec Width 10010.0 Hz 40 repetitions OBSERVE P31, 80.9939267 DECOUPLE H1, 200.0813150 Power 45 dB continuously on WALT2-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 32768 Total time 4 min, 36 sec BnO OBn



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SURVEY PHOSPHORUS PARAMETERS

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "mercvx"

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Pulse 45.3 degrees Acg. time 1.600 sec Width 10010.0 Hz S5 repetitions DBSERVE P31, 80.9939267 NHz DECOUPLE H1, 200.0813150 HHz Power 45 dB continuously on WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 32768 Total time 7 min, 12 sec



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STANDARD 1H OBSERVE

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "mercvx"



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GP-II

Sample: GP-II Sample ID: s_20100413_05 File: 0074.fid

Pulse Sequence: s2pul Solvent: cdc13 Temp. 26.0 C / 299.1 K Operator: walkup File: 0074 VMMRS-500 "NMRS00"

Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 2.049 sec Vidth 8012.8 Hz 8 repetitions OBSERVE H1, 499.7316295 MHz DATA PROCESSING Resol. enhancement -0.0 Hz FT size 55356 Total time 0 min, 31 sec







SURVEY PHOSPHORUS PARAMETERS

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "mercvx"

Pulse 45.3 degrees Acg. time 1.600 sec Width 10010.0 Hz DBSERVE P31, 60.3939267 MHz DECOUPLE H1, 200.0813150 MHz Power 45 dB Continuously on WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 32768 Total time 4 min, 36 sec







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SURVEY PHOSPHORUS PARAMITERS

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "mercvx"

Pulse 45.3 degrees Acq. time 1.600 sec Width 10010.0 Hz OSERVE P31, 80.9339267 MHz DECOUPLE H1, 200.0813150 MHz Power 45 dß continuously on WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 32768 Total time 0 min, 0 sec













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STANDARD 1H OBSERVE

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Nercury-200 "MercVX"

Relax. delay 1.000 sec Pulse 36.4 degrees Acq. time 1.994 sec Width 3003.0 Hz 200 repetitions OBSERVE H1, 200.0785680 MHz DATA PROCESSING FT size 16384 Total time 11 min, 40 sec

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21 - S - 41 13C OBSERVE Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature File: R-C1-Ac-De-13C Nercury-200 "MercVX" Pulse 45.0 degrees Acq. time 1.498 sec Vidth 12484.4 Hz 50000 repetitions OBSERVE C13, 50.3097842 MHz DECOUPLE H1, 200.0795755 MHz Power 45 dB continuously on WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 65536 Total time 0 min, 0 sec .847 77.211 76.583 5 C Ο 0 CI CI 25a 78.120 30.191 -63.731 62.391 73.667 40.710 860 049 115.439 50 រអ្ន .870 167.064 166.776 137. 122.329

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STANDARD 1H OBSERVE

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "MercVX"

Relax. delay 1.000 sec Pulse 38.4 degrees Acq. time 1.954 sec Width 3003.0 Hz 200 repetitions OBSERVE H1, 200.0785690 MHz DATA PROCESSING FT size 16354 Total time 0 min, 0 sec



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STANDARD 1H OBSERVE

Pulse Sequence: s2pul

Solvent: CDC13 Ambient temperature Mercury-200 "MercVX"

Relax. delay 1.000 sec Pulse 38.4 degrees Acq. time 1.394 sec Width 3003.0 Hz 200 repetitions OBSERVE H1, 200.0785590 MHZ DATA PROCESSING FT size 16384 Total time 11 min, 40 sec









STANDARD 1H OBSERVE

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "MercVX"

Relax. delay 1.000 sec Pulse 38.4 degrees Acq. time 1.994 sec Width 3003.0 Hz 200 repetitions OBSERVE H1, 200.0785690 MHz DATA PROCESSING FT size 16384 Total time 11 min, 40 sec












S74



STANDARD 1H OBSERVE

Pulse Sequence: s2pul

Solvent: CDC13 Ambient temperature Mercury-200 "MercVX"



13C OBSERVE

Pulse Sequence: s2pul Solvent: CDC13 Ambient temperature Mercury-200 "MercVX"

Pulse 45.0 degrees Acq. time 1.498 sec Width 12484.4 Hz 2864 repetitions OBSERVE C13, 50.3097842 MHz DECOUPLE H1, 200.0795755 MHz Power 45 dB continuously on WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz FT size 65536 Total time 4 hr, 40 min, 26 sec





