Gate Effects in a Hexagonal Zinc-Imidazolate-4-amide-5-imidate Framework with Flexible Methoxy Substituent and CO₂ Selectivity

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Supporting Information

21 pages

Experimental Details

Synthesis of IFP-7 was performed in seal tube from Ace Glass. All reagents and solvents were used as purchased from great chemical suppliers without further purification if not stated otherwise.

The linker precursors 2-methoxy-4,5-dicyanoimidazole (1)^{1a} and IFP-1^{1b} were synthesized following published procedure.

Elemental analysis (C, H, N) was performed on Elementar Vario EL elemental analyzer.

Synthesis of IFP-7

In a sealed tube (Typ A, company: Ace) 2-methoxy-4,5-dicyanoimidazole (1) (0.1 g, 0.76 mmol) and Zn(NO₃)₂ ·4H₂O (0.19 g, 0.76 mmol) were solved in DMF (6 mL). The sealed tube was closed and the mixture was heated at 130 °C for 36 hours and was then allowed to cool down to room temperature with 5 °C per hour. Afterwards, large amount of powder of was obtained along with some by-product formed at the inner wall of sealed tube. By-product is bigger in size. IFP-7 was separated by sieving technique,² which is used extensively in the mining industry and for determining particle size. Minor by-product (~ 10 %) was trapped by a mesh while IFP-7 (~ 90 %) filtered through it. We could not able to define the structure of the by-product, till now. Powder material was collected by filtration, washed with DMF and EtOH and dried in air. The material named as IFP-7.

After several attempts, we could not find a suitable crystal of IFP-7 for single X-ray diffraction.

IFP-7: Yield: ~ 67 % based on Zn(NO₃)₂ ·4H₂O; ¹³C CP-MAS NMR: δ =170.1, 167.2, 157.1, 132.7, 126.9, 55.7 ppm; Elemental analysis of activated IFP-7: C₆H₆N₄O₃Zn; Calcd., C 29.11, H 2.44, N 22.64; Found: C 29.27, H 2.31, N 26.76; IR (KBr pellet): v_{max} = (3339 m, 3107 m, 1658 s, 1562 vs, 1478 m, 1285 m, 1252 m, 1220m, 1111 m, 795 m, 737 m) cm⁻¹.



Scheme S1. Synthesis of IFP-7.

Transformation of IFP-7 to the monomeric Zn-complex $[Zn(L2)_2(H_2O)]$ (2)

5 mL H₂O was added in a sealed tube (Typ A, company: Ace), containing IFP-7 (0.1 g). The sealed tube was closed and the mixture was heated at 100°C for 36 hours and was then allowed to cool down to room temperature. Crystals were formed. Crystals were washed with EtOH and dried in air.

Yield: ~ 51 %, Elemental analysis of 2: $C_{12}H_{20}N_8O_9Zn$: Calcd., C 29.67, H 4.15, N 23.07, Found: C 29.58, H 4.31, N 23.22; IR (KBr pellet): $v_{max} = (3443 \text{ m}, 3335 \text{ m}, 1651 \text{ s}, 1593 \text{ vs}, 1475 \text{ m}, 1272 \text{ m}, 1103 \text{ m}, 1011 \text{ m}, 723 \text{ m}) \text{ cm}^{-1}$.

IR spectra

IR spectra were recorded on FT-IR Nexus from Thermo Nicolet in the region of 4000 - 400 cm⁻¹ using KBr pellets as basis.

The fabrication of framework comes from IR stretch that no stretching bands related to C=N in the region of 2230–2240 cm⁻¹. Instead, new typical bands for amide and imidate groups observed between 3100-3350 cm⁻¹ and at 1560-1660 cm⁻¹.

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Fig. S1: IR-spectra of: **A**) 2-Methoxy-4,5-dicyanoimidazole (1); **B**) IFP-7 as-synthesized; **C**) Zn-complex [Zn(L2)₂(H₂O)] (2).

¹H and ¹³C CPMAS NMR Spectroscopy

¹H and ¹³C Cross Polarization experiments with Magic Angle Sample Spinning (¹³C CPMAS NMR) for IFP-7 was performed on a Bruker Avance 600 spectrometer (Bruker Biospin GmbH, Rheinstetten, Germany, $B_0 = 14.1$ T) operating at a frequency of 150.9 MHz using a double resonant 4 mm MAS (magic angle sample spinning) probe. The spinning frequency was 12.5 kHz. For CP, a ramped (50 % ramp) lock field was used on the ¹H channel with a CP contact time of 2 ms, a recycle delay of 3 s and 512 scans. The 90° ¹H pulse length was 3.3 µs. High power TPPM decoupling was applied (B₁ = 80 kHz). The ¹³C chemical shifts are referenced using the glycine carboxyl group signal (δ (¹³C) = 176.4 ppm).



Fig. S2: ¹H- **A**) and ¹³C- **B**) CP-MAS NMR spectra of IFP-7 (blue – as-synthesized, red – activated), activation procedure: 200° C and 10^{-3} mbar, 30 hrs.

Theoretical calculations of crystal structure of IFP-7

DFT calculations (PBE functional³) were performed by using the SIESTA program package.⁴ The DZP basis set used in the calculations was first tested on the structure of IMOF-3 (later, we named IMOF-3 as IFP-1) solved from single-crystal X-ray data,^{1b} and showed very good agreement with respect to unit cell parameters (compare a_{calcd} = 17.5313 Å and c_{calcd} = 18.6286 Å vs. a_{exptl} =17.9244 Å and c_{exptl} = 18.4454 Å), bond lengths and angles. Due to the large unit cell sizes, it was sufficient to include only the Γ -point of the Brillouin zone for the evaluation of integrals in the reciprocal space.

Powder X-ray-diffraction patterns

Powder X-ray diffraction (PXRD) patterns of all IFPs were measured on a Siemens

Diffractometer D5005 in Bragg-Brentano reflection geometry. The diffractometer was equipped with a copper tube, a scintillation counter, automatical incident- and diffracted-beam soller slits and with a graphite secondary monochromator. The generator was set to 40 kV and 40 mA. All measurements were performed with sample rotating. Data were collected digitally from 3° to 70° 2θ using a step size of $0.02^{\circ} 2\theta$ and a count time of 4 seconds per step. The simulated powder patterns for IFP-7 were calculated using single-crystal X-ray diffraction data and processed by the free Mercury v1.4.2 program provided by the Cambridge Crystallographic Data Centre.



Fig. S3: Powder X-ray diffraction patterns of IFP-7 (Color: black = simulated, blue = As-synthesized IFP-7, red = Activated).



Fig. S4: Powder X-ray diffraction profiles of IFP-7, collected during stability tests in refluxing methanol.



Fig. S5: Powder X-ray diffraction profiles of IFP-7, collected during stability tests in refluxing benzene.



Fig. S6: Powder X-ray diffraction profiles of IFP-7, collected during stability tests in refluxing water.

Single crystal X-ray structure determination of monomeric Zn-complex $[Zn(L2)_2(H_2O)]$ (2)

The crystal was embedded in perfluoropolyalkylether oil and mounted on a glass fibre. Intensity data were collected at 150 K using a STOE Imaging Plate Diffraction System IPDS-2 with graphite monochromatized $Mo_{K\alpha}$ radiation ($\lambda = 0.71073$ Å) at 50 kV and 40 mA (360 frames, $\Delta\omega=1^{\circ}$, 2 min exposure time per frame. The data were corrected for Lorentz polarization and absorption effects. The structure was resolved with direct methods using SHELXS-97⁵ and

refined with full-matrix least-squares on F^2 using the program SHELXL-97.⁶ All non-hydrogen atoms were refined anisotropically.

The hydrogen atoms of the methyl groups were calculated in their expected positions and refined as riding atoms with $U_{iso}(H) = 1.5 U_{eq}(C)$.

The other hydrogen atoms were located from the difference Fourier map and refined with $U_{iso}(H) = 1.5 U_{eq}(N, O)$.

Table S1. Selected crystallographic data and details of the structure refinements of monomeric Zn-
complex $[Zn(L2)_2(H_2O)]$ (2).Chemical formulaChemical formula

Chemical formula	$C_{12}H_{20}N_8O_9Zn$	
Formula Mass	485.73	
Crystal system	triclinic	
Space group	PĪ	
$a/ m \AA$	8.7253(6)	
$b/\text{\AA}$	9.1928(6)	
$c/{ m \AA}$	12.6864(8)	
$lpha/^{\circ}$	98.872(5)	
$\beta/^{\circ}$	100.538(5)	
$\gamma/^{\circ}$	109.715(5)	
Unit cell volume/Å ³	915.78(11)	
Temperature/K	150	
No. of formula units per unit cell, Z	2	
Radiation type	ΜοΚα	
Absorption coefficient, μ/mm^{-1}	1.411	
Reflections collected	11907	
No. of independent reflections	3230	
R _{int}	0.0184	
$R_1 / wR_2 [I > 2\sigma(I)]$	0.0210 / 0.0595	
R_1 / wR_2 (all data)	0.0222 / 0.0601	
Goodness of fit on F^2	1.064	



Fig. S7: Crystal structure of monomeric Zn-complex $[Zn(L2)_2(H_2O)]$ (2) showing the coordination environment of Zn^{II} and the coordination mode of the ligand 2-methoxyimidazolate-4,5-diamide (L2). Non-coordinating water molecules are omitted for clarity.

 Zn^{2+} ion is pentacoordinated by donor atoms of two ligands (4,5-diamide-2methoxyimidazolate). Ligand L1 in IFP-7 is transformed to 2-methoxyimidazolate-4,5-diamide (L2) in refluxing water. Two imidazolate N atoms (N1 and N5), two amide O atoms (O3 and O6), and one O atom (O7) from water molecule formed a distorted environment with a square pyramidal geometry. The packing is stabilized by a system of hydrogen bonds. For structural parameters as bond lengths , bond angles, and hydrogen bonds – see cif.

CCDC-925826 of $[Zn(L2)_2(H_2O)]$ (2) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Thermogravimetric (TG) analysis

The TG measurements were performed in a stationary air atmosphere (no purge) from room temperature up to 800 °C using a Linseis thermal analyzer (Linseis, Germany) working in the vertical mode. The heating rate was 10 °C/min. The samples were placed in cups of aluminium oxide.

Thermogravimetric analysis (TGA) trace for as-synthesized IFP-7 indicated a gradual weightloss of 12 % (25–250°C), corresponding to partial loss of guest species like water molecules and N,N'-dimethylformamide (DMF), followed by the decomposition of framework. Activated sample shows the thermal stability up to 300 °C.



Fig. S8: TGA curves of as-synthesized (color: blue) and activated compound (color : red) IFP-7.

Gas-sorption measurements

The sample was connected to the preparation port of the sorption analyzer and degassed under vacuum until the out gassing rate, i.e., the rate of pressure rise in the temporarily closed manifold with the connected sample tube, was less than 2 μ Torr/min at the specified temperature 160 °C for 24 h. After weighing, the sample tube was then transferred to the analysis port of the sorption analyzer. All used gases (H₂, He, N₂, CO₂, CH₄) were of ultra high purity (UHP, grade 5.0, 99.999%) and the STP volumes are given according to the NIST standards (293.15 K, 101.325 kPa). Helium gas was used for the determination of the cold and warm free space of the sample tubes. H₂ and N₂ sorption isotherms were measured at 77 K (liquid nitrogen bath), whereas CO₂ and CH₄ sorption isotherms were measured at 298±1 K (passive thermostating) 273.15 K (ice/deionized water bath) and 195.0 K (acetone/dry ice). The heat of adsorption values and the DFT calculations ('N₂ DFT slit pore' model) were done using the ASAP 2020 v3.05 software.

H₂ uptake

To check the hysteresis, we have measured the H_2 uptake capacity of a freshly prepare sample at 77 K by increasing the equilibrium time.

Table S2 summarizes the equilibrium time interval, measurement time and H_2 uptake for IFP-7. The H4 type hysteresis is mostly caused by irreversible uptake of molecules in pores (or through pore entrances).⁷ We have increased the equilibration interval up to 50 s which had taken the measurement time 68 h. If we further increased the equilibration interval, then it will be a problem to keep the temperature constant (77 K).

Condition		Curve or Experiment	Measurement time (h)	H_2 uptake $(cm^3/g)^a$
Absolute	Equilibration			
pressure	Interval ^b			
(mmHg)	(s)			
0-50	20	curve or experiment 1	IFP-7: 29 IFP-1: 8	35
50-500	15			
500-1000	10			
0-50	30	curve or experiment 2	38	37
50-500	30			
500-1000	30			
0-50	50	curve or experiment 3	68	50
50-500	50			
500-1000	50			

Table S2. H₂ uptake measurements at different equilibrium time interval.

^a H₂ uptake with a standard deviation of $\pm 15 \text{ cm}^3/\text{g}$ at 77 K and 1 bar.

^b Equilibration time allowed during both adsorption and desorption at each data point up in the stated pressure range



Fig. S9: H_2 gas sorption isotherms of IFP-7 under different equilibration interval. (left) and IFP-1 (right). Adsorption and desorption branches are indicated in closed and open symbols, respectively.

To compare the hysteresis of IFP-7, we measured the H_2 sorption at a similar condition (curve or exp.-1) for IFP-1 which is similar in structure to IFP-7 except for the methyl instead of a

methoxy group (IFP-1 contains a *methyl* group; IFP-7 contains a *methoxy* group, Fig. S10). In IFP-1, there is no hysteresis during the desorption (Fig. S9, right). Therefore, we confirmed that the equilibrium time interval is not a key issue for such broad hysteresis.



Fig. S10: Schematic structure of IFP-1 (left) and IFP-7 (right)

The broad desorption hysteresis for H_2 in IFP-7 does not change with the equilibrium time interval. This was also confirmed for the narrower desorption hysteresis of CO₂ at 273 K in IFP-7At 273 K, again we observed the similar hysteresis when the ad- and desorption equilibrium time interval was increased to 50 s over the whole pressure range (Fig.S14).



Fig. S11: CO_2 gas sorption isotherms of IFP-7 under different equilibration interval. Showing almost no change in hysteresis upon desorption.

Heat of adsorption

From two adsorption isotherms acquired at different temperatures T_1 and T_2 , the differential heat of adsorption $\Delta H_{ads,diff}$ can be calculated for any amount of adsorbed substance after determining the required relative pressures p_1 and p_2 . A modified form of the Clausius-Clapeyron equation is used (eq (1))⁸ $\Delta H_{ads,diff}$ was calculated over the whole adsorption range from the 273 K and 298 K isotherms for CO₂.

 $\Delta H_{ads,diff} = -Rln \left(\frac{p_2}{p_1}\right) \frac{T_1 T_2}{T_2 - T_1} \ (1)$



Fig. S12: Isosteric heats of CO₂ adsorption as a function of the adsorbent loading for IFP-7.



Fig. S13: The initial slope in the Henry region of the sorption isotherms of CO_2 (black) and CH_4 (red) and N_2 (blue) of IFP-7 at 298 K.



Fig. S14: The initial slope in the Henry region of the sorption isotherms of CO₂ (black) and CH₄ (red) and N₂ (blue) of IFP-7 at 273 K.



Fig. S15: The initial slope in the Henry region of the sorption isotherms of CO₂ (black) and CH₄ (red) and N₂ (blue) of IFP-7 at 195 K.

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