

Environmentally Sensitive Fluorescent Purine Nucleoside That Changes Emission Wavelength upon Hybridization

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Experimental

General: $^1\text{H-NMR}$ spectra (400 MHz) and $^{13}\text{C-NMR}$ spectra (100 MHz) were measured with Bruker Avance 400F spectrometer. Coupling constant (J value) are reported in hertz. The chemical shifts are shown in ppm downfield from tetramethylsilane, using residual dimethyl sulfoxide (δ 2.50 in $^1\text{H-NMR}$, δ 39.5 in $^{13}\text{C-NMR}$) and chloroform (δ 7.26 in $^1\text{H-NMR}$, δ 77.0 in $^{13}\text{C-NMR}$) as an internal standard. ESI-TOF masses were recorded on a JMS-T100LC "AccuTOF", Applied DATUM Solution Business Operations.

Oligonucleotides synthesis and characterization: The reagents for DNA synthesis were purchased from Glen Research. Mass spectra of oligodeoxynucleotides were determined with a MALDI-TOF MS (Shimadzu AXIMA-LNR, positive mode) with 2',3',4'-trihydroxyacetophenone as a matrix. Calf intestinal alkaline phosphatase (Promega), Crotalus adamanteus venom phosphodiesterase I (USB), and *Penicillium citrinum* nuclease P1 (Roche) were used for the enzymatic digestion of ODNs. All aqueous solutions utilized purified water (Millipore, Milli-Q sp UF). Reversed-phase HPLC was performed on CHEMCOBOND 5-ODS-H columns (10 \times 150 mm, 4.6 \times 150 mm) with a JASCO Chromatograph, Model PU-2080, using a UV detector, Model UV-2075 plus at 260 nm.

Procedure for the synthesis of modified nucleosides

6-Amino-1-(2-deoxy- β -D-erythro-pentofuranosyl)-1,5-dihydro-3-(2-naphthylethynyl)-4H-pyrazolo[3,4-*d*]pyrimidin-4-one (1) A mixture of 7-iodo-8-aza-7-deaza-2'-deoxyguanosine **2** (120 mg, 0.31 mmol), Pd(PPh₃)₄ (18.0 mg, 0.02 mmol), CuI (6.0 mg, 0.03 mmol) and 2-ethynynaphthalene (60.0 mg, 0.39 mmol) in DMF (5 ml) and Et₃N (0.1 ml) was stirred at 50 °C at under argon atmosphere for 30 min. After allowing to cool to room temperature, The reaction solution was concentrated in *vacuo*. The residue was purified by silica gel column chromatography with chloroform-methanol mixture (12 : 1 to 8 : 1) to give **1** as a colorless solid (87.2 mg, 68 %); $^1\text{H-NMR}$ (DMSO-*d*₆, 400 MHz) δ 2.20 (ddd, J = 4.1, 6.6, 13.2 Hz, 1H), 2.72 (m, 1H), 3.40 (m, 1H), 3.51 (m, 1H), 3.79 (m, 1H), 4.40 (m, 1H), 4.77 (m, 1H), 5.27 (d, J = 4.3 Hz, 1H), 6.34 (m, 1H), 6.80 (br, 2H), 7.59-7.63 (complex, 3H), 7.97-8.01 (complex, 3H), 8.23 (m, 1H), 10.84 (s, 1H); $^{13}\text{C-NMR}$ (DMSO-*d*₆, 100 MHz) δ 37.7, 62.4, 70.9, 82.2, 83.3, 87.6, 91.9, 100.4, 118.9, 127.0, 127.4, 127.8, 127.8, 127.9, 128.5, 129.7, 131.6, 132.5, 132.7, 155.4, 155.7, 157.1; HRMS (ESI) m/z 440.1335 calcd for C₂₂H₁₉N₅O₄Na [M + Na]⁺, found 440.1362.

6-[[Dimethylamino)methylidene]amino]-1-(2-deoxy- β -D-erythro-pentofuranosyl)-1,5-dihydro-3-(2-naphthylethynyl)-4H-pyrazolo[3,4-*d*]pyrimidin-4-one (3) A solution of **1** (75.0 mg, 0.18 mmol) in DMF (5 ml) was stirred with *N,N*-dimethylformamide diethyl acetal (45 μ l, 0.27 mmol) at 50 °C for 30 min. After completion of the reaction, the solvent was evaporated, and the residue was purified by silica gel column chromatography with chloroform-methanol mixture (9 : 1) to give compound **3** as a colorless solid (43.4 mg, 51 %); $^1\text{H-NMR}$ (DMSO-*d*₆, 400 MHz) δ 2.20 (ddd, J = 4.0, 6.7, 13.2 Hz, 1H), 2.75 (m, 1H), 3.08 (s, 3H), 3.21 (s, 3H), 3.42 (m, 1H), 3.53 (m, 1H), 3.82 (m, 1H), 4.44 (m, 1H), 4.78 (m, 1H), 5.30 (d, J = 4.2 Hz, 1H), 6.51 (m, 1H), 7.58-7.64 (complex, 3H), 7.97-8.01 (complex, 3H), 8.24 (m, 1H), 8.75 (s, 1H), 11.45 (s, 1H); $^{13}\text{C-NMR}$ (DMSO-*d*₆, 100

MHz) δ 34.9, 37.9, 40.8, 62.4, 71.0, 82.2, 83.4, 87.7, 92.0, 102.8, 118.9, 127.0, 127.4, 127.8, 127.8, 127.9, 128.5, 129.6, 131.6, 132.5, 132.7, 154.8, 157.9, 158.7, 159.6; HRMS (ESI) m/z 495.1757 calcd for $C_{25}H_{24}N_6O_4Na$ [$M + Na$]⁺, found 495.1786.

6-[[*(Dimethylamino)methylidene*]amino]-1-[2-deoxy-5-*O*-(4,4'-dimethoxytriphenylmethyl)- β -D-erythro-pentofuranosyl]-1,5-dihydro-3-(2-naphthylethynyl)-4*H*-pyrazolo[3,4-*d*]pyrimidin-4-one (4) To a solution of **3** (36.0 mg, 0.08 mmol) in anhydrous pyridine (3 ml) was added 4,4'-dimethoxytrityl chloride (39.0 mg, 0.11 mmol), and the stirring continued at room temperature for 1 h. After completion of the reaction, the solvent was evaporated, and the residue was diluted with ethyl acetate, washed with saturated $NaHCO_3$ and brine. The combined organic layer was dried with anhydrous Na_2SO_4 , filtered and concentrated. The purification by silica gel column chromatography with $CHCl_3$ -MeOH- Et_3N mixture (30 : 1 : 0.5) yields compound **4** as a colorless solid (42.8 mg, 72 %); 1H -NMR ($CDCl_3$, 400 MHz) δ 2.37 (m, 1H), 3.02-3.09 (complex, 4H), 3.16 (s, 3H), 3.31 (m, 1H), 3.38 (m, 1H), 3.70 (s, 6H), 4.05 (m, 1H), 4.83 (m, 1H), 6.63 (m, 1H), 6.78 (m, 4H), 7.16-7.33 (complex, 7H), 7.42 (m, 2H), 7.47-7.52 (complex, 2H), 7.67 (m, 1H), 7.79-7.81 (complex, 3H), 8.17 (m, 1H), 8.70 (s, 1H), 8.72 (s, 1H); ^{13}C -NMR ($CDCl_3$, 100 MHz) δ 35.2, 38.0, 41.5, 55.1 ($\times 2$), 64.5, 73.4, 81.5, 83.5, 85.6, 86.4, 93.3, 103.7, 113.1 ($\times 4$), 119.9, 126.5, 126.8, 126.8, 127.8, 127.8 ($\times 2$), 127.9, 128.0, 128.2 ($\times 2$), 128.6, 129.1, 130.0 ($\times 2$), 130.1 ($\times 2$), 132.4, 132.9, 133.1, 136.0, 136.0, 144.8, 155.2, 158.0, 158.4, 158.4, 158.6, 158.7; HRMS (ESI) m/z 797.3064 calcd for $C_{46}H_{42}N_6O_6Na$ [$M + Na$]⁺, found 797.3043.

6-[[*(Dimethylamino)methylidene*]amino]-1-[2-deoxy-5-*O*-(4,4'-dimethoxytriphenylmethyl)- β -D-erythro-pentofuranosyl]-1,5-dihydro-3-(2-naphthylethynyl)-4*H*-pyrazolo[3,4-*d*]pyrimidin-4-one-3'-[(2-cyanoethyl)-*N,N*-(diisopropyl)]phosphoramidite (5) To a solution of **4** (60.0 mg, 0.08 mmol) in anhydrous acetonitrile (1 ml) was added 2-cyanoethyl-diisopropyl-chlorophosphoramidite (70 μ l) in the presence of Et_3N (1 ml), and stirred at room temperature under an argon atmosphere for 1 h. After completion of the reaction, the solution was diluted with ethyl acetate, washed with saturated $NaHCO_3$ and brine. The combined organic layer was dried with anhydrous Na_2SO_4 , filtered and concentrated. The residue was incorporated into oligonucleotides without further purification.

Fluorescence spectra: The fluorescence spectra were obtained with a Shimadzu RF-5300PC spectrofluorophotometer at 25 °C using a cell with a 1 cm path length. The excitation and emission bandwidths were 1.5 nm. The fluorescence quantum yields (Φ_f) were determined using 9,10-diphenylanthracene as a reference with the known Φ_f (0.95) in ethanol.¹

UV absorption measurements: Absorption spectra were obtained using a Shimadzu UV-2550 spectrophotometer at room temperature using 1 cm length cell.

Melting temperature (T_m) measurements: All T_m s of the ODNs (2.5 μ M, final concentration) were measured in 50 mM sodium phosphate buffers (pH 7.0) containing 100 mM sodium chloride. Absorbance vs temperature profiles were measured at 260 nm using a Shimadzu UV-2550 spectrophotometer equipped with a Peltier temperature controller using 1 cm path length cell. The absorbance of the samples was monitored at 260 nm from 4 to 90 °C with a heating rate of 1 °C/min. From these profiles, first derivatives were calculated to determine T_m values.

Circular dichroism (CD) measurements: CD spectra were recorded with a JASCO J-805 CD spectrophotometer. CD spectra of oligonucleotides solutions (2.5 μ M ODNs in 50 mM sodium phosphate buffers (pH 7.0) containing 100 mM sodium chloride at 20 °C) were measured using 2 mm path length cell.

Reference

1. J. V. Morris, M. A. Mahaney, J. R. Huber, J. Phys. Chem., 1976, 80, 969-974.

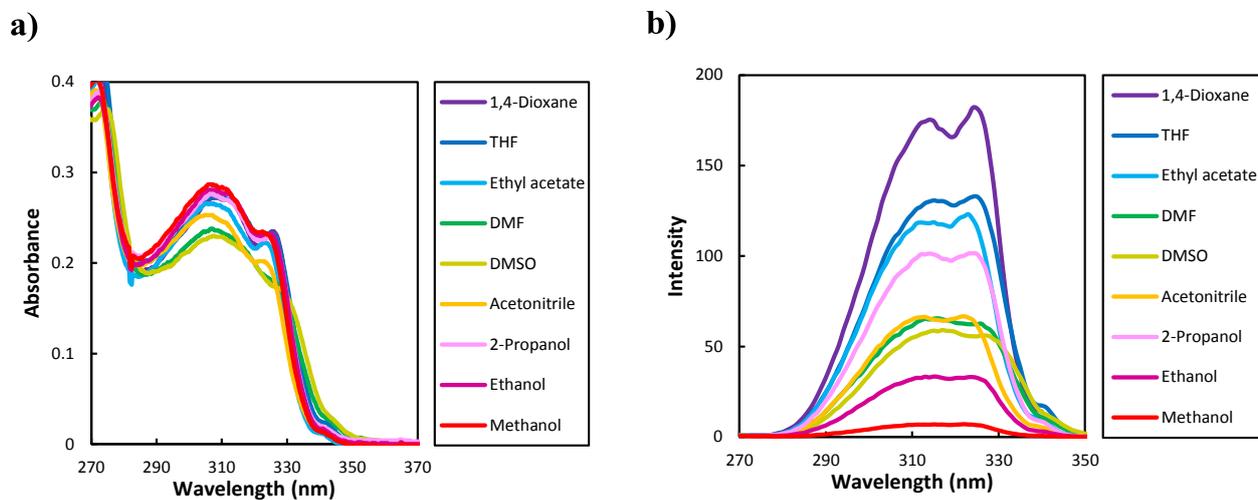


Fig. S1 UV absorption (a) and excitation (b) spectra of ^{na}G (1) (10 μM) in various solvents.^a

^a Solvent is used in the presence of 1 % v/v of DMF because of its low solubility.

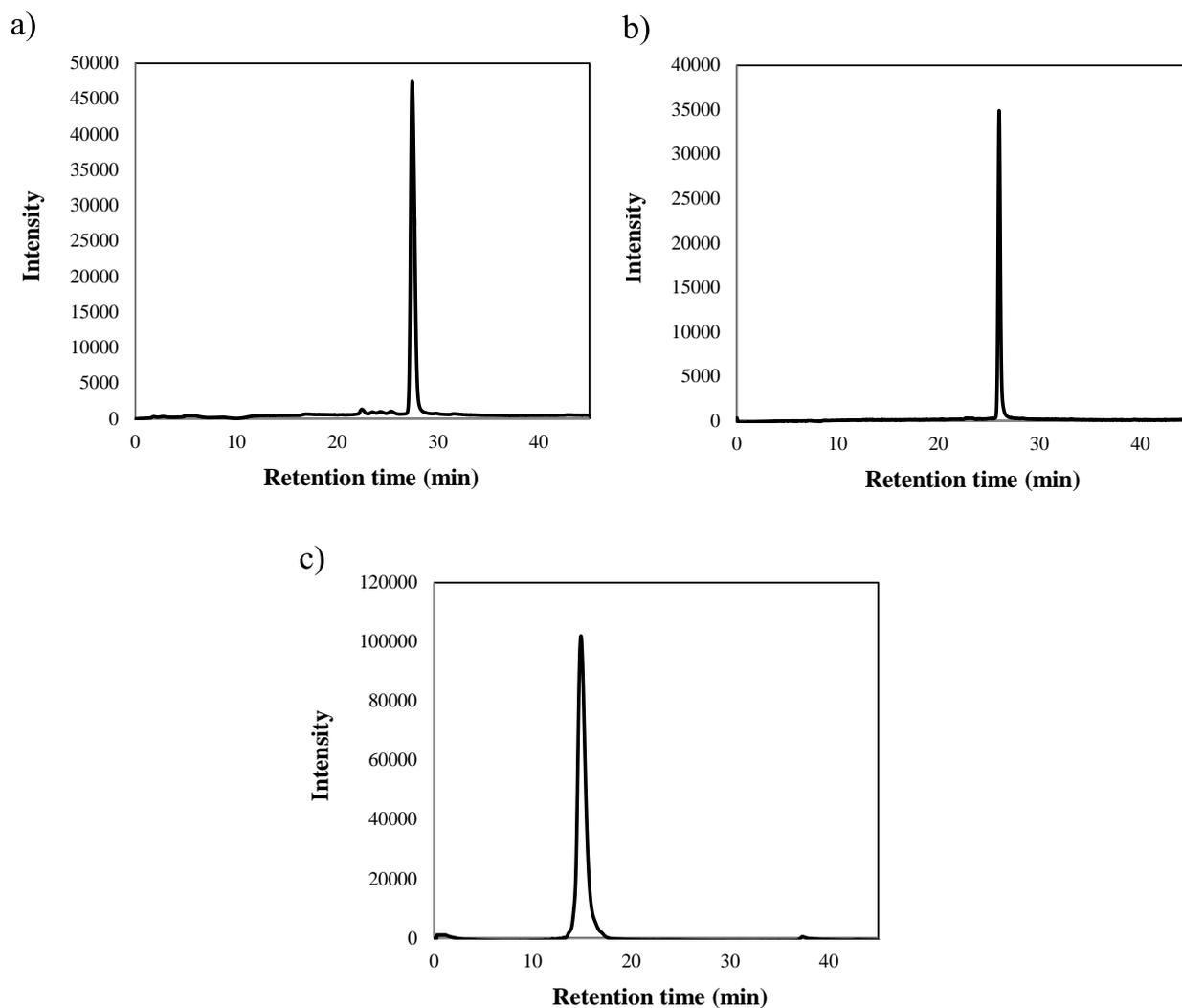
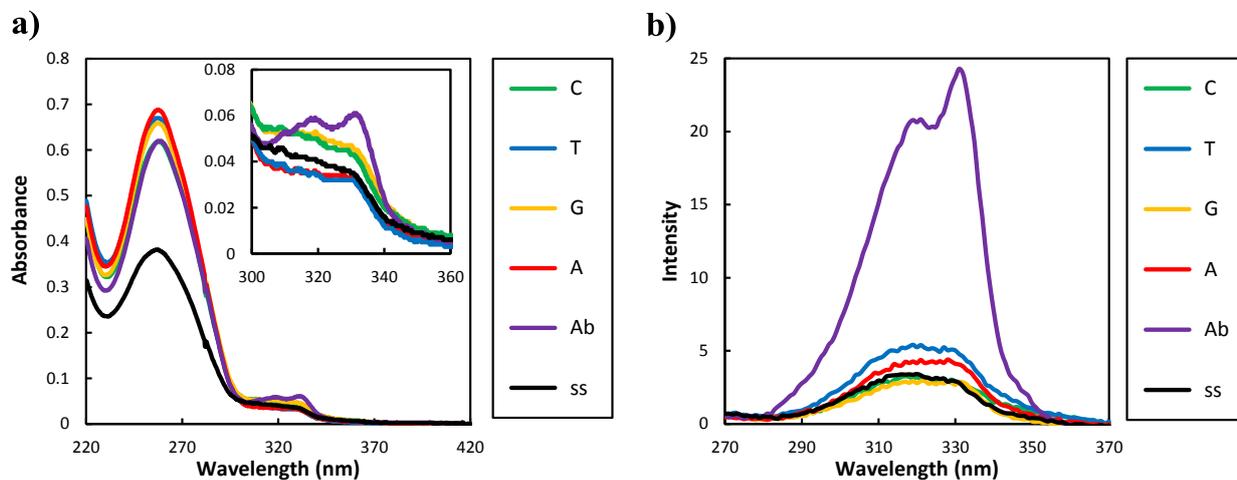


Fig. S2 HPLC profiles of single-stranded (a) ODN 1: 5'-CGCAAT^{na}GTAACGC-3', (b) ODN 3: 5'-CGCAAT^{na}GAAACGC-3', and (c) MB 1: 5'-CCACATGTGAAGGGCTTTT^{na}GAACTCTGCATGTGG-3'. HPLC analysis was carried out on a CHEMCOBOND 5-ODS-H column (10 × 150 mm) eluted with 0.05 M ammonium formate buffer containing acetonitrile. Gradient: from 3 to 20 % acetonitrile for (a) and (b), from 3 to 40 % acetonitrile for (c) at a flow rate 2.0 ml/min over 45 min.

Table S1 MALDI-TOF mass spectral data for the ODNs

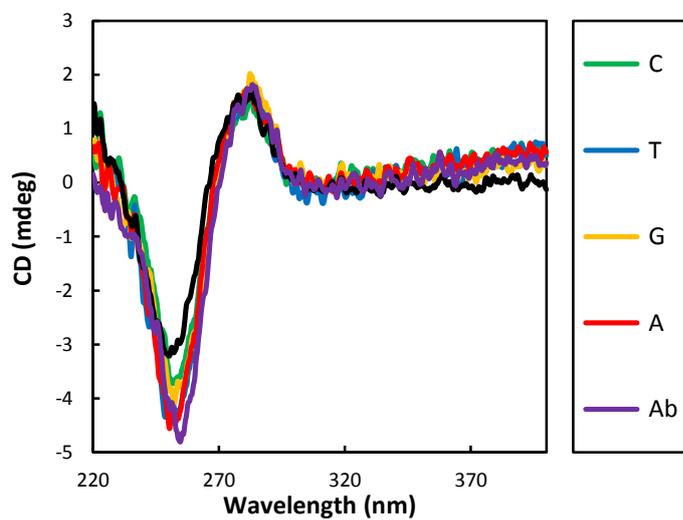
ODNs	MALDI-TOF mass	
	calcd. [M + H] ⁺	found [M + H] ⁺
ODN 1: 5'-CGCAAT ^{na} GTAACGC-3'	4093.83	4094.09
ODN 3: 5'-CGCAAT ^{na} GAAACGC-3'	4102.84	4102.60
MB 1: 5'-CCACATGTGAAGGGCTTTT ^{na} GAACTCTGCATGTGG-3'	10639.08	10640.58



ODN 1: 5'-CGCAAT ^{na}G TAACGC-3'

ODN 2: 3'-GCGTTA N ATTGCG-5' (N = C, T, G, A, Abasic site)

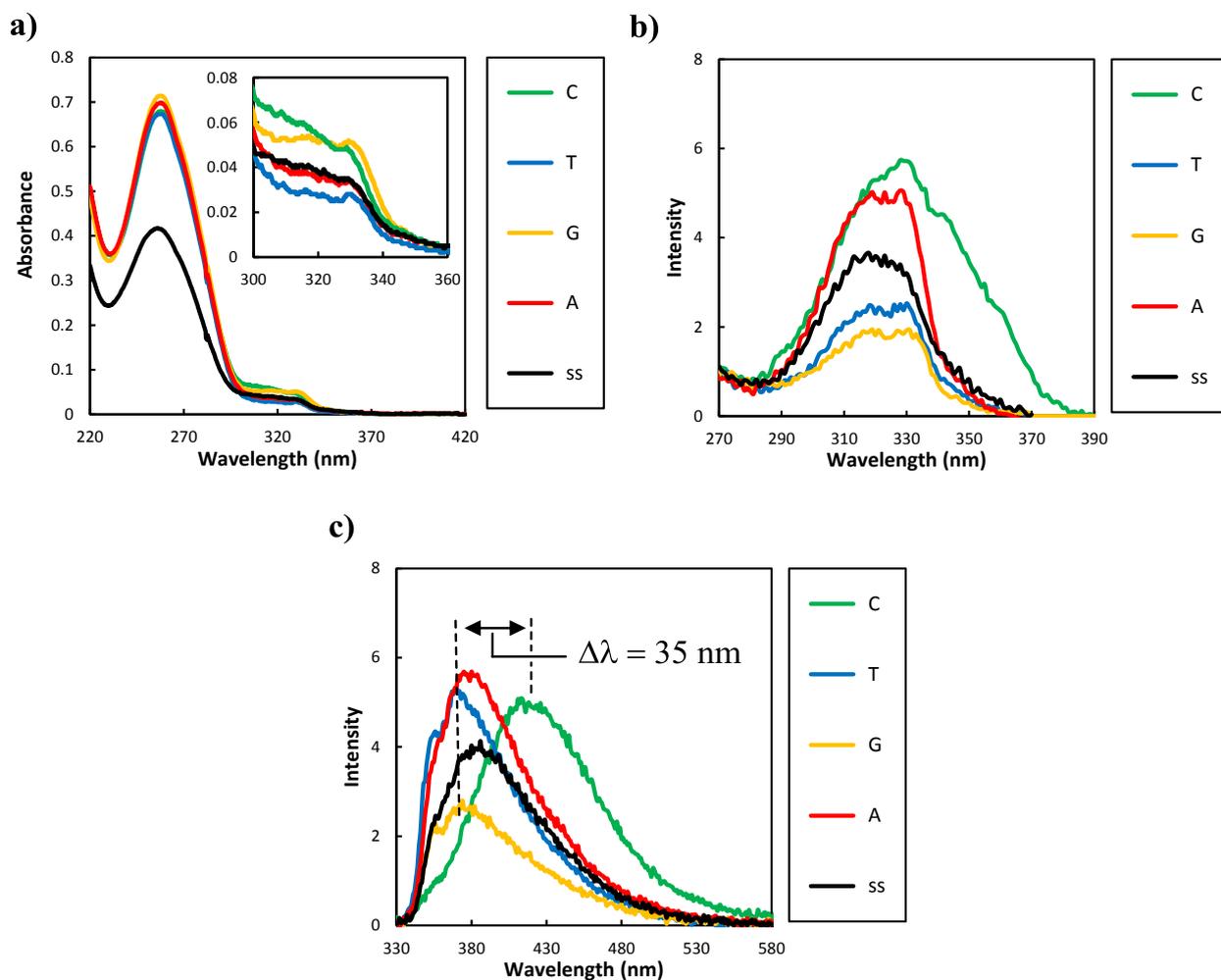
Fig. S3 UV absorption (a) and excitation (b) spectra of ODN 1 hybridized with ODN 2 (N = C, T, G, A, Ab). "Ab" denotes abasic site and "ss" denotes a single-strand ODN 1 (2.5 μ M ODNs, 0.1 M sodium chloride, 50 mM sodium phosphate buffer, pH 7.0, rt).



ODN 1: 5'-CGCAAT **X** TAACGC-3' (**X** = ^{na}G or G)

ODN 2: 3'-GCGTTA **N** ATTGCG-5' (**N** = C, T, G, A, Abasic site)

Fig. S4 CD spectra of ODN 1 (**X** = G) hybridized with ODN 2 (**N** = C) (black line), and of ODN 1 (**X** = ^{na}G) hybridized with ODN 2 (**N** = C, T, G, A, Ab). "Ab" denotes abasic site (2.5 uM ODNs, 50 mM sodium phosphate, 0.1 M sodium chloride, pH 7.0, rt).



ODN 3: 5'-CGCAAT ^{na}G AAACGC-3'

ODN 4: 3'-GCGTTA N TTTGCG-5' (N = C, T, G, A)

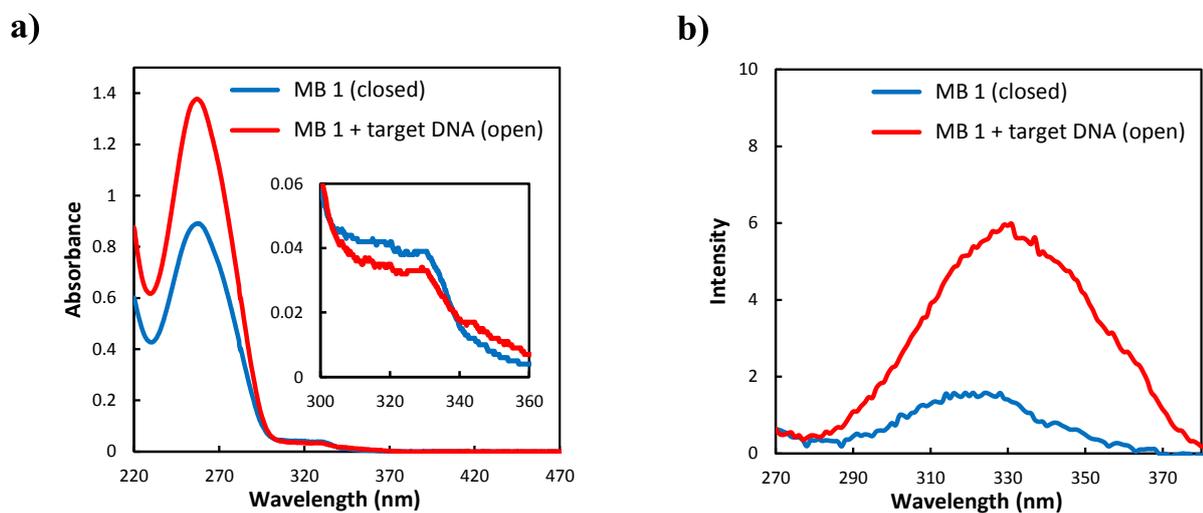
Fig. S5 UV absorption (a), excitation (b) and fluorescence (c) spectra of ODN 3 hybridized with ODN 4 (N = C, T, G, A). "ss" denotes a single-strand ODN 3 (2.5 μ M ODNs, 0.1 M sodium chloride, 50 mM sodium phosphate buffer, pH 7.0, rt).

Table S2 Thermal melting temperatures (T_m) and fluorescence maximum of duplexes ODN3/ODN4.

ODN 3 / ODN 4(N)	T_m (°C)	$\lambda_{\max}^{\text{fl}}$ (nm)
ODN 3 / ODN 4 (C)	55.3	410
ODN 3 / ODN 4 (T)	42.4	378
ODN 3 / ODN 4 (G)	43.7	375
ODN 3 / ODN 4 (A)	41.3	379
ODN 3	-	386

ODN 3: 5'-CGCAAT ^{na}G AAACGC-3'

ODN 4: 3'-GCGTTA N TTTGCG-5' (N = C, T, G, A)



MB 1: 5'-CCACATGTGAAGGGCTTTT ^{na}G AACTCTGCATGTGG-3'
target DNA(ODN_{bcr/abl}): 3'-ACTTCCCGAAA ACTTGAGAC-5'

Fig. S6 UV absorption (a) and excitation (b) spectra of hairpin MB 1 and the duplex formed by hybridization with target DNA (2.5 μ M ODNs, 0.1 M sodium chloride, 50 mM sodium phosphate buffer, pH 7.0, rt).