## **Supporting Information**

# Probing the reactivity of *o*-phthalaldehydic acid/methyl ester: Synthesis of *N*-isoindolinones and 3-arylaminophthalides

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### General

Reagents and anhydrous solvents (THF, dichloromethane, and acetonitrile) were used as received. Triphenylphosphine, polymer-bound (100-200 mesh, 1.6 mmol/g) was purchased from Sigma-Aldrich. Reaction progress was monitored by TLC using Merck silica gel 60 F-254 with UV detection. Silica gel 60 (Merck 40–63  $\mu$ m) was used for column chromatography. The following stain solutions have been used in addition to UV light with fluorescent TLC plates: phosphomolybdic acid, anisaldehyde/EtOH. Reactions requiring anhydrous conditions were performed under nitrogen. NMR data were collected and calibrated in *d4*-MeOH or CDCl<sub>3</sub> at 298K on a Varian Unity 400 MHz Hz spectrometer. HPLC and routine mass spectra were acquired on an Agilent Technologies 1200 Series instrument, fitted with a G1316A UV-Vis detector, 1200 Series ELSD and 6110 quadrupole ESI-MS. High resolution mass spectrometry (HRMS) was performed on the Bruker MicroTOF mass spectrometer.

### **Experimental procedures**



#### Preparation of *o*-phthalaldehydic acid methyl ester derivatives<sup>1</sup>

Scheme 1. i)  $SOCl_2$ , reflux, 3 h; ii)  $Et_2NH$ ,  $CH_2Cl_2$ , 0 °C - RT, 12 h; iii) s-BuLi/TMEDA, -78 °C, DMF; iv) AcOH/HCl, reflux, 12 h; v) DBU-MeI, MeCN, RT.

#### **Preparation of aromatic azides**<sup>2</sup>

A solution of aromatic amines (4.0 mmol) in CH<sub>3</sub>CN (8 mL) was cooled to 0°C in an ice bath. To this stirred mixture was added *t*-BuONO (6.0 mmol) followed by TMSN<sub>3</sub> (6.0 mmol) drop wise. The resulting solution was stirred at room temperature for 1 h. The reaction mixture was concentrated under vacuum and the crude product was purified by silica gel chromatography (hexane) to give the azides **2** in quantitative yields.

#### Preparation of benzylic and aliphatic azides

To a solution of benzylic/aliphatic halides (4.0 mmol) in THF (10 mL) was added NaN<sub>3</sub> (8.0 mmol) in water (1.0 ml). The resulting solution was stirred at 80  $^{\circ}$ C for 3 h. The reaction mixture diluted with EtOAc (20 mL) and washed with water (20 ml) and brine (20 mL). The organic layer was separated dried (MgSO<sub>4</sub>) and concentrated under vacuum to give the azides **2** in quantitative yields.

# General procedure for the synthesis of *N*-isoindolinones through aza-Wittig/cyclisation reaction

#### Method A: using triphenylphosphine

To a stirred solution of *o*-phthalaldehydic acid methyl ester **1** (0.2 mmol) and azides **2** (0.3 mmol) in dry THF (5.0 mL) was added triphenylphosphine (0.4 mmol). The mixture was stirred at room temperature for 6 h. NaBH<sub>3</sub>CN (0.4 mmol) was added and the reaction mixture was stirred at room temperature under nitrogen. The solvent was removed under reduced pressure and the residue was dissolved in EtOAc, washed with water and brine. The organic layer was dried over MgSO<sub>4</sub> and concentrated. Purification by flash column chromatography (EtOAc/hexanes) gave the products.

#### Method B: using polymer-bound triphenylphosphine

To a stirred solution of *o*-phthalaldehydic acid methylester **1** (0.2 mmol) and azides **2** (0.4 mmol) in dry THF (5.0 mL) was added polymer-bound triphenylphosphine (0.4 mmol). The mixture was stirred at room temperature for 3 h. The resin was filtered, NaBH<sub>3</sub>CN (0.4 mmol) was added and the reaction mixture was stirred at reflux temperature under nitrogen. The solvent was removed under reduced pressure and the residue was dissolved in EtOAc, washed with water and brine. The organic layer was dried over MgSO<sub>4</sub> and concentrated. Purification by flash column chromatography (EtOAc/hexanes) gave the products.

# Synthesis of 3-arylaminophthalides through aza-Wittig/cyclisation reaction using polymer-bound triphenylphosphine

To a stirred solution of o-phthalaldehydic acid **5** (0.2 mmol) and azides **2** (0.4 mmol) in dry MeOH (5.0 mL) was added polymer-bound triphenylphosphine (0.4 mmol). The mixture was stirred at reflux temperature for 5 h. The resin was filtered and the solvent was evaporated to dryness. The crude residue was recrystallised using dry MeOH to obtain the 3-arylamino phthalides **6**.

# Synthesis of 3-arylaminophthalides using known procedure for the comparison of spectral data<sup>3</sup>

A solution of *o*-phthalaldehydic acid **5** (0.2 mmol) and aromatic amines (0.2 mmol) in dry MeOH (5.0 mL) was refluxed for 30 min. The solvent was evaporated to dryness and crude residue was recrystallised using dry MeOH to obtain the 3-arylamino phthalides.

### **Characterisation of compounds**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.44 (dd (merged), J = 8.0 Hz, 1H), 7.05 – 7.37 (m, 5H), 6.93 (d, J = 8.0 Hz, 1H), 6.89 (d, J = 8.0 Hz, 1H), 4.75 (s, 2H), 4.20 (s, 2H), 3.98 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 167.3, 157.5, 144.0, 137.3, 133.0, 128.7, 128.2, 127.5, 114.8, 110.0, 55.8, 48.9, 46.1; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 254.1176, found 254.1186.

#### Compound 4b



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.43 (dd (merged), J = 8.0 Hz, 1H), 7.21 (d, J = 8.0 Hz, 2H), 7.12 (d, J = 8.0 Hz, 2H), 6.92 (d, J = 8.0 Hz, 1H), 6.88 (d, J = 8.0 Hz, 1H), 4.71 (s, 2H), 4.19 (s, 2H), 3.98 (s, 3H), 2.32 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.3, 157.5, 144.0, 137.2, 134.3, 133.0, 132.9, 132.8, 129.4, 128.3, 120.0, 114.9, 110.1, 55.8, 48.9, 45.8, 21.1; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>18</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 268.1332, found 268.1344.

**Compound 4c** 



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.70 – 7.83 (m, 4H), 7.35 – 7. 49 (m, 4H), 6.84 – 6.92 (m, 2H), 4.88 (s, 2H), 4.19 (s, 2H), 3.97 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.4, 157.5, 144.0, 134.8, 133.3, 133.0, 132.8, 128.6, 127.7, 126.9, 126.8, 126.2, 125.9, 119.8, 114.9, 110.1, 55.8, 49.0, 46.3; HRMS (ESI) calcd. for C<sub>20</sub>H<sub>18</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 304.1332, found 304.1336.

Compound 4d OMe O



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.54 (d, J = 4.0 Hz, 1H), 7.63 (dd, J = 8.0 Hz, 1H), 7.46 (dd (merged), J = 8.0 Hz, 1H), 7.36 (d, J = 8.0 Hz, 1H), 7.18 (dd, J = 8.0, 4.0 Hz, 1H), 6.97 (d, J = 8.0 Hz, 1H), 6.89 (d, J = 8.0 Hz, 1H), 4.88 (s, 2H), 4.40 (s, 2H), 3.98 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 167.4, 157.6, 157.3, 149.1, 144.3, 137.1, 133.1, 122.8, 122.5, 119.7, 114.9, 110.1, 55.8, 49.8, 48.1; HRMS (ESI) calcd. for C<sub>20</sub>H<sub>25</sub>N<sub>2</sub>O<sub>4</sub> [M+H]<sup>+</sup> 255.1128, found 255.1141.

#### **Compound 4e**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.49 (dd (merged), J = 8.0 Hz, 1H), 7.04 (d, J = 8.0 Hz, 1H), 6.92 (d, J = 8.0 Hz, 1H), 4.36 (s, 2H), 4.04 (q, J = 8.0 Hz, 2H), 3.98 (s, 3H), 3.60 (t, J = 8.0 Hz, 2H), 2.37 (t, J = 8.0 Hz, 2H), 1.70 – 2.06 (m, 2H), 1.20 (t, J = 8.0 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 173.3, 168.1, 157.0, 143.9, 133.3, 119.7, 115.3, 110.3, 60.6, 56.0, 49.7, 41.6, 31.4, 23.6, 14.1; ESI-MS: m/z 278.1 [(M+H)<sup>+</sup>, 100); HRMS (ESI) calcd. for C<sub>15</sub>H<sub>20</sub>NO<sub>4</sub> [M+H]<sup>+</sup> 278.1387, found 278.1434.

#### **Compound 4f**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (d, *J* = 8.0 Hz, 2H), 7.52 (dd (merged), *J* = 8.0 Hz, 1H), 7.40 (dd, *J* = 8.0 Hz, 2H), 7.15 (dd, *J* = 8.0 Hz, 1H), 7.07 (d, *J* = 8.0 Hz, 1H), 6.93 (d, *J* = 8.0 Hz, 1H), 4.80 (s, 2H), 4.00 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.2, 157.9, 142.8, 139.7, 133.7, 129.0, 124.0, 119.2, 114.6, 110.3, 55.9, 50.1; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>14</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 240.1019, found 240.1027.

Compound 4g

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.74 (d, *J* = 8.0 Hz, 2H), 7.50 (dd (merged), *J* = 8.0 Hz, 1H), 7.05 (d, *J* = 8.0 Hz, 1H), 6.95 (d, *J* = 8.0 Hz, 2H), 6.92 (d, *J* = 8.0 Hz, 1H), 4.75 (s, 2H), 3.99 (s, 3H), 3.82 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.0, 157.8, 156.2, 142.8, 133.4, 132.9, 121.1, 121.1, 114.6, 114.2, 110.3, 55.9, 55.5, 50.5; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 270.1125, found 270.1136.

#### **Compound 4h**

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.88 (d, *J* = 8.0 Hz 2H), 7.48 (dd (merged), *J* = 8.0 Hz, 1H), 7.26 – 7.37 (m, 2H), 6.98 – 7.07 (m, 5H), 6.85 – 6.96 (m, 2H), 4.71 (s, 2H), 3.96 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.1, 157.8, 157.4, 153.34, 142.8, 135.2, 133.7, 129.7, 123.1, 120.9, 119.5, 118.5, 114.6, 110.4, 55.9, 50.3; HRMS (ESI) calcd. for C<sub>21</sub>H<sub>18</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 332.1281, found 332.1264.

#### **Compound 4i**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.84 (dd, *J* = 8.0, 4.0 Hz 2H), 7.52 (dd (merged), *J* = 8.0 Hz, 1H), 7.02 - 7.14 (m, 3H), 6.93 (d, *J* = 8.0 Hz, 1H), 4.76 (s, 2H), 4.00 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.1, 160.4, 157.9, 142.6, 133.8, 120.9, 115.8, 115.5, 114.6, 110.4, 55.9, 50.3; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>13</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 258.0925, found 258.0936.

#### **Compound 4**j



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.86 (s, 1H), 7.53 (dd (merged), J = 8.0 Hz, 1H), 7.29 (dd, J = 8.0 Hz, 1H), 7.20 (dd, J = 8.2, 2.2 Hz, 1H), 7.06 (d, J = 8.0 Hz, 1H), 6.93 (d, J = 8.0 Hz, 1H), 6.71 (dd, J = 8.2, 2.2 Hz, 1H), 4.78 (s, 2H), 4.01 (s, 3H), 3.85 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 166.4, 160.2, 157.9, 142.7, 141.0, 133.8, 129.5, 114.6, 110.5, 110.4, 110.3, 104.6, 104.6, 55.9, 55.3, 50.2; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 270.1125, found 270.1133.

**Compound 4k** 



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 (s, 1H), 7.81 (d, *J* = 8.0 Hz, 1H), 7.54 (dd (merged), *J* = 8.0 Hz, 1H), 7.32 (dd, *J* = 8.0 Hz, 1H), 7.12 (d, *J* = 8.0, Hz, 1H), 7.07 (d, *J* = 8.0 Hz, 1H), 6.94 (d, *J* = 8.0 Hz, 1H), 4.78 (s, 2H), 4.01 (s, 3H); <sup>13</sup>C NMR (101 MHz, cdcl<sub>3</sub>)  $\delta$ 166.1, 157.8, 142.4, 140.6, 134.6, 133.9, 129.8, 123.7, 119.8, 118.8, 116.6, 114.4, 110.3, 55.7, 49.8; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>13</sub>ClNO<sub>2</sub> [M+H]<sup>+</sup> 274.0629, found 274.0632.

#### **Compound 4l**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.73 (s, 1H), 7.62 (d, J = 8.0 Hz, 1H), 7.51 (dd (merged), J = 8.0 Hz, 1H), 7.29 (d, J = 8.0 Hz, 1H), 7.05 (d, J = 8.0 Hz, 1H), 6.97 (d, J = 8.0 Hz, 1H), 6.92 (d, J = 8.0 Hz, 1H), 4.78 (s, 2H), 4.00 (s, 3H), 2.39 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 166.2, 157.9, 142.9, 139.6, 138.8, 133.7, 128.8, 124.9, 120.5, 120.1, 116.3, 114.6, 110.3, 55.9, 50.2, 21.7; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 254.1176, found 254.1188.

Compound 3m



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (dd (merged), *J* = 8.0 Hz, 1H), 7.09 (dd, *J* = 8.0 Hz, 1H), 7.08 (d, *J* = 8.0 Hz, 1H), 7.03 (d, *J* = 8.0 Hz, 1H), 6.88 (d, *J* = 8.0 Hz, 1H), 6.72 (dd, *J* = 8.0 Hz, 1H), 6.61 (d, *J* = 8.0 Hz, 1H), 4.34 (s, 2H), 3.85 (s, 3H), 3.84 (s, 3H), 2.50 (q, *J* = 7.5 Hz, 2H), 1.24 (t, *J* = 7.5 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  168.3, 156.7, 145.2, 138.4, 130.9, 127.8, 127.8, 126.9, 122.8, 120.6, 117.5, 110.3, 110.2, 56.0, 55.9, 52.4, 52.3, 46.3, 23.7, 12.9; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>22</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 300.1594, found 300.1601.



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.32 (dd (merged), J = 8.0 Hz, 1H), 6.96 – 7.06 (m, 3H), 6.86 (d, J = 8.0 Hz, 1H), 6.68 – 6.78 (m, 1H), 6.60 (dd, J = 8.0, 1.4 Hz, 1H), 5.14 (br s, 1H), 4.33 (d, J = 4.4 Hz, 2H), 3.85 (s, 6H), 3.75 – 3.90 (m, 4H), 2.86 – 2.95 (m, 4H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  168.0, 156.6, 142.7, 138.6, 130.6, 125.2, 124.7, 122.3, 119.9, 119.5, 118.5, 117.1, 115.1, 110.4, 110.0, 67.5, 55.9, 51.7, 51.3, 46.0; HRMS (ESI) calcd. for C<sub>20</sub>H<sub>25</sub>N<sub>2</sub>O<sub>4</sub> [M+H]<sup>+</sup> 357.1809, found 357.1814.

#### **Compound 4ab**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.19 (d, *J* = 8.0 Hz, 2H), 7.11 (d, *J* = 8.0 Hz, 2H), 6.42 (s, 1H), 6.41 (s, 2H), 4.67 (s, 2H), 4.13 (s, 2H), 3.93 (s, 3H), 3.82 (s, 3H), 2.31 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.3, 164.2, 158.4, 145.9, 137.1, 134.5, 129.3, 128.2, 113.3, 99.1, 99.0, 98.2, 98.1, 55.9, 55.7, 49.0, 45.7, 21.1; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>20</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 298.1438, found 268.1381.



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.19 (d, J = 8.0 Hz, 2H), 7.11 (d, J = 8.0 Hz, 2H), 6.72 (s, 1H), 6.68 (s, 1H), 4.68 (s, 2H), 4.13 (s, 2H), 3.95 (s, 3H), 2.39 (s, 3H), 2.31 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 167.4, 157.2, 144.3, 143.9, 137.1, 134.4, 129.3, 128.2, 117.5, 115.5, 115.5, 111.0, 111.0, 55.8, 48.8, 45.8, 22.2, 21.1; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>20</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 282.1489, found 282.1480.

**Compound 4cb** OMe 0 Me

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.18 (d, *J* = 8.0 Hz, 2H), 7.12 (d, *J* = 8.0 Hz, 2H), 6.91 (s, 1H), 6.87 (s, 1H), 4.68 (s, 2H), 4.16 (s, 2H), 3.96 (s, 3H), 2.32 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.3, 157.7, 145.1, 138.7, 137.3, 133.9, 129.4, 128.2, 118.7, 115.3, 111.1, 56.1, 48.6, 45.8, 21.1; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>17</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 302.0942, found 302.0869.

#### **Compound 6f**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.68 (dd (merged), J = 8.0 Hz, 1H), 7.28 (dd, J = 8.0 Hz, 2H), 7.18 (d, J = 8.0 Hz, 1H), 7.04 (d, J = 8.0 Hz, 1H), 6.88 – 6.98 (m, 3H), 6.71 (d, J = 12.0 Hz, 1H), 4.64 (d, J = 12.0 Hz, 1H), 4.02 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.1, 158.2, 148.0, 143.6, 136.4, 129.3, 120.7, 114.9, 114.9, 114.7, 112.3, 85.8, 55.9; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>14</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 256.0968, found 256.0986.

Compound 6g

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.65 (dd (merged), J = 8.0 Hz, 1H), 7.18 (d, J = 8.0 Hz, 1H), 7.02 (d, J = 8.0 Hz, 1H), 6.91 (d, J = 8.0 Hz, 2H), 6.83 (d, J = 8.0 Hz, 2H), 6.61 (d, J = 12.0 Hz, 1H), 4.46 (d, J = 12.0 Hz, 1H), 3.99 (s, 3H), 3.77 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.3, 158.3, 154.5, 148.2, 137.3, 136.4, 117.3, 115.0, 114.8, 112.3, 87.4, 56.1, 55.6; ESI-MS: m/z 286.0 [(M+H)<sup>+</sup>, 100]; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>4</sub> [M+H]<sup>+</sup> 286.1074, found 286.1127.



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.67 (dd (merged), J = 8.0 Hz, 1H), 7.11 – 7.23 (m, 2H), 7.04 (d, J = 8.0 Hz, 1H), 6.70 (d, J = 12.0 Hz, 1H), 6.43 – 6.57 (m, 3H), 4.64 (d, J = 12.0 Hz, 1H), 4.01 (s, 3H), 3.80 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 167.2, 160.8, 158.4, 148.0, 145.1, 136.5, 130.2, 115.0, 114.8, 112.5, 107.6, 106.2, 101.2, 85.8, 56.1, 55.2; ESI-MS: m/z 286.0 [(M+H)<sup>+</sup>, 100]; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>16</sub>NO<sub>4</sub> [M+H]<sup>+</sup> 286.1074, found 286.1122.

#### **Compound 6m**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.69 (dd (merged), J = 8.0 Hz, 1H), 7.21 (d, J = 8.0 Hz, 2H), 7.12 – 7.19 (m, 2H), 7.06 (d, J = 8.0 Hz, 1H), 6.89 – 7.00 (m, 1H), 6.72 (d, J = 12.0 Hz, 1H), 4.49 (d, J = 12.0 Hz, 1H), 4.03 (s, 3H), 2.43 – 2.60 (m, 2H), 1.24 (t, J = 7.5 Hz, 3H).; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.2, 158.4, 148.5, 141.4, 136.6, 129.9, 128.4, 127.2, 121.1, 115.1, 114.8, 114.1, 112.5, 86.4, 56.2, 23.9, 13.1; ESI-MS: m/z 284.1 [(M+H)<sup>+</sup>, 100]; 286.1074 HRMS (ESI) calcd. for C<sub>17</sub>H<sub>18</sub>NO<sub>4</sub> [M+H]<sup>+</sup> 284.1281, found 284.1352.

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![](_page_15_Figure_1.jpeg)

![](_page_15_Figure_2.jpeg)

![](_page_16_Figure_1.jpeg)

![](_page_17_Figure_1.jpeg)

# Copies of 1H NMR and 13C NMR spectra

![](_page_18_Figure_1.jpeg)

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![](_page_21_Figure_1.jpeg)

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![](_page_38_Figure_1.jpeg)

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![](_page_40_Figure_1.jpeg)

![](_page_41_Figure_1.jpeg)

![](_page_42_Figure_1.jpeg)