

Organocatalytic tandem Morita-Baylis-Hillman/Michael reaction for asymmetric synthesis of a drug-like oxa-spirocyclic indanone scaffold

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Supporting Information

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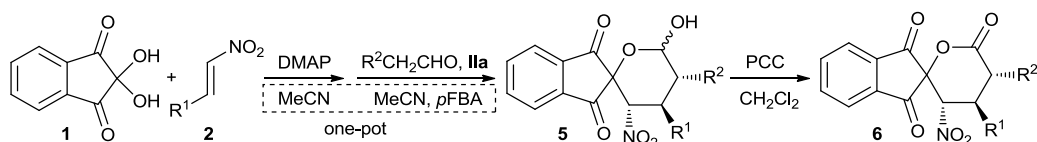
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1. General methods

NMR data was obtained for ^1H at 400 MHz, and for ^{13}C at 100 MHz. Chemical shifts were reported in ppm from tetramethylsilane with the solvent resonance as the internal standard in CDCl_3 solution. ESI HRMS was recorded on a Waters SYNAPT G2. In each case, enantiomeric ratio was determined by HPLC analysis on chiral column in comparison with authentic racemates, using a Daicel Chiralpak AD-H Column (250 x 4.6 mm), Daicel Chiralpak OD-H Column (250 x 4.6 mm) or Kromasil CelluCoat Column (250 x 4.6 mm). UV detection was monitored at 220 nm or 254 nm. Optical rotation data were examined in CH_2Cl_2 solution at 20 °C. Column chromatography was performed on silica gel (200-300 mesh) eluting with ethyl acetate and petroleum ether. TLC was performed on glass-backed silica plates. UV light and I_2 were used to visualize products. Melting points were determined on a Mel-Temp apparatus and are uncorrected. All chemicals were used without purification as commercially available unless otherwise noted. The catalysts **II** were synthesized according to the literature procedures.^[1]

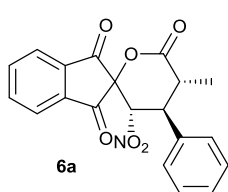
[1]. (a) Marigo, M.; Wabnitz, T. C.; Fielenbach, D.; Jørgensen, K. A. *Angew. Chem., Int. Ed.* **2005**, *44*, 794. (b) Hayashi, Y.; Gotoh, H.; Hayasi, T.; Shoji, M. *Angew. Chem., Int. Ed.* **2005**, *44*, 4212.

2. General procedure for the asymmetric synthesis of spiroindanone δ -lactone **6**



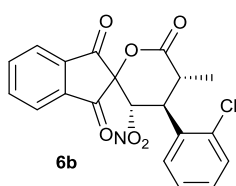
The reaction was carried out with ninhydrin **1** (89 mg, 0.5 mmol) and nitroolefin **2** (0.4 mmol) in the presence of DMAP (3.6 mg, 0.04 mmol) in MeCN (2.0 mL) at room temperature for 1 h to afford the MBH adduct **3**, after which aldehyde **4** (0.5 mmol), diphenyl prolinol TMS ether **IIa** (32.5 mg, 0.1 mmol) and 4-fluorobenzoic acid (14.0 mg, 0.1 mmol) were added in one-pot. The reaction mixture was stirred at room temperature for a specified reaction time (about 3h) until the reaction completed (monitored by TLC). Then the reaction mixture was concentrated and the residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate = 10:1) to give hemiacetal **5**.

Hemiacetal **5** were oxidized to the stable corresponding spiroindanone δ -lactone **6**. To a solution of **5** in methylene chloride (5 mL) was added PCC (107.8 mg, 0.5 mmol). The mixture was stirred for 2 h at 50 °C. The solid was removed by filtration through celite. The filtrate was evaporated under reduced pressure and the residual was purified by column chromatography (petroleum ether/ethyl acetate = 15:1) to give spiroindanone δ -lactone **6**. The dr values of **6** were calculated by ¹H NMR analysis of the crude reaction mixture and the major diastereomers of **6** could be isolated in pure form for NMR, HRMS and HPLC analysis.

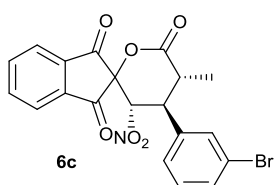


6a was obtained as a white solid in 76% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 93% by HPLC on Chiralpak OD-H column (30% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 11.05$ min, $t_{\text{major}} = 12.11$ min. m.p. 167-168 °C; $[\alpha]_{\text{D}}^{20} -129.4$ ($c = 0.12$ in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): $\delta = 8.12$ -8.05 (m, 2H), 8.01-7.94 (m, 2H), 7.40-7.34 (m, 3H), 7.27-7.26 (m, 2H), 5.64 (d, $J = 12.4$ Hz, 1H), 4.16 (t, $J = 12.0$ Hz, 1H), 3.10-3.02 (m, 1H), 1.39 (d, $J = 6.8$ Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 192.31$, 190.03, 169.62, 141.15, 140.18, 138.00, 137.48, 135.43, 129.51, 129.05, 127.77, 124.99, 124.72, 87.28, 46.58, 41.42, 14.74 ppm; ESI HRMS: calcd. For C₂₀H₁₅NO₆+Na 388.0797, found 388.0795.

For the minor diastereomer of **6a**, ¹H NMR (400 MHz, CDCl₃): $\delta = 8.05$ -7.84 (m, 4H), 7.33-7.18 (m, 5H), 5.56 (d, $J = 12.0$ Hz, 1H), 4.09 (t, $J = 11.6$ Hz, 1H), 3.02-2.94 (m, 1H), 1.31 (d, $J = 6.8$ Hz, 3H) ppm; ESI HRMS: calcd. For C₂₀H₁₅NO₆+Na 388.0797, found 388.0796.

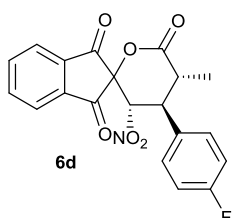


6b was obtained as a white solid in 76% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 99% by HPLC on Kromasil CelluCoat column (10% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 26.42$ min, $t_{\text{major}} = 28.33$ min. m.p. 167-168 °C; $[\alpha]_{\text{D}}^{20} -126.2$ ($c = 0.18$ in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): $\delta = 8.15$ -8.07 (m, 2H), 8.03-7.95 (m, 2H), 7.43-7.39 (m, 3H), 7.30-7.27 (m, 1H), 5.63 (d, $J = 12.4$ Hz, 1H), 5.19 (t, $J = 12.0$ Hz, 1H), 2.99-2.97 (m, 1H), 1.42 (d, $J = 6.4$ Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 192.07$, 190.04, 169.22, 141.15, 140.21, 137.90, 137.43, 135.66, 133.85, 130.44, 129.83, 128.10, 126.70, 124.99, 124.93, 86.23, 42.39, 40.71, 14.31 ppm; ESI HRMS: calcd. For C₂₀H₁₄ClNO₆+Na 422.0407, found 422.0409.



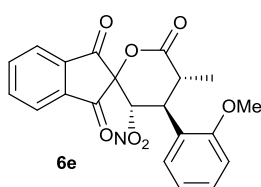
6c was obtained as a white solid in 78% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 98% by HPLC on Chiralpak OD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, $t_{\text{minor}} = 13.62$ min, $t_{\text{major}} = 14.97$ min. m.p. 168-169 °C; $[\alpha]_{\text{D}}^{20}$

-95.2 ($c = 0.12$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.13$ -8.07 (m, 2H), 8.03-7.98 (m, 2H), 7.50-7.43 (m, 2H), 7.30-7.20 (m, 2H), 5.60 (d, $J = 12.0$ Hz, 1H), 4.15 (t, $J = 11.6$ Hz, 1H), 3.03-2.99 (m, 1H), 1.40 (d, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.17$, 189.80, 169.08, 141.07, 140.21, 138.01, 137.82, 137.57, 132.32, 131.06, 130.90, 126.44, 125.06, 124.78, 123.53, 86.78, 46.17, 41.30, 14.78 ppm; ESI HRMS: calcd. For $\text{C}_{20}\text{H}_{14}\text{BrNO}_6 + \text{Na}$ 465.9902, found 465.9905.



6d was obtained as a white solid in 72% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Chiralpak OD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 11.74$ min, $t_{\text{major}} = 13.13$ min. m.p. 170-172 °C; $[\alpha]_{\text{D}}^{20}$

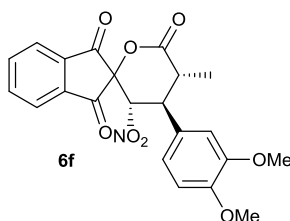
-107.6 ($c = 0.12$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.13$ -8.06 (m, 2H), 8.03-7.96 (m, 2H), 7.26-7.23 (m, 2H), 7.11-7.07 (m, 2H), 5.57 (d, $J = 12.4$ Hz, 1H), 4.18 (t, $J = 12.0$ Hz, 1H), 3.02-2.97 (m, 1H), 1.39 (d, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.25$, 189.90, 169.29, 162.84 (d, $J_{\text{CF}} = 248$ Hz), 141.11, 140.17, 138.04, 137.55, 131.20, 129.49 (d, $J_{\text{CF}} = 8$ Hz), 125.04, 124.76, 116.65 (d, $J_{\text{CF}} = 22$ Hz), 99.99, 87.21, 45.87, 41.54, 14.70 ppm; ESI HRMS: calcd. For $\text{C}_{20}\text{H}_{14}\text{FNO}_6 + \text{H}$ 384.0883, found 384.0886.



6e was obtained as a white solid in 70% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 99% by HPLC on Chiralpak OD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 15.64$ min, $t_{\text{major}} = 17.68$ min. m.p. 168-170 °C; $[\alpha]_{\text{D}}^{20}$

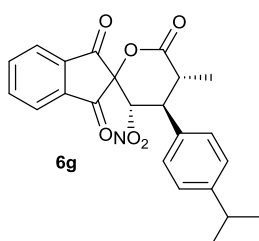
-125.8 ($c = 0.08$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.12$ -8.05 (m, 2H), 8.01-7.93 (m, 2H), 7.34-7.30 (m, 1H), 7.08-7.06 (m, 1H), 6.95-6.89 (m, 2H), 6.14 (br s, 1H), 4.24 (br s, 1H), 3.93 (s, 3H), 3.52 (br s, 1H), 1.39 (d, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.67$, 190.39, 170.38, 157.58, 141.32, 140.22, 137.72, 137.18, 130.31, 124.86, 124.61, 122.27, 122.21, 121.25, 111.62, 84.56, 55.59, 50.44, 38.60, 15.38 ppm; ESI HRMS: calcd. For $\text{C}_{21}\text{H}_{17}\text{NO}_6 + \text{Na}$

418.0903, found 418.0901.



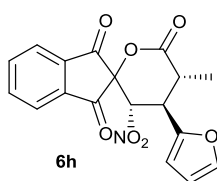
6f was obtained as a white solid in 65% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Chiralpak OD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, $t_{\text{minor}} = 27.63$ min, $t_{\text{major}} = 36.53$ min. m.p. 171-172 °C; $[\alpha]_{\text{D}}^{20}$

-139.9 ($c = 0.12$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.13$ -8.06 (m, 2H), 8.03-7.95 (m, 2H), 7.27 (s, 1H), 6.86-6.79 (m, 2H), 6.71-6.70 (m, 1H), 5.58 (d, $J = 12.4$ Hz, 1H), 4.09 (t, $J = 12.0$ Hz, 1H), 3.89 (s, 3H), 3.87 (s, 3H), 3.06-2.98 (m, 1H), 1.41 (d, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.36, 190.09, 169.64, 149.61, 149.43, 141.18, 140.17, 138.01, 137.47, 127.55, 124.99, 124.67, 120.08, 111.67, 110.36, 87.54, 56.07, 55.90, 46.35, 41.61, 14.77$ ppm; ESI HRMS: calcd. For $\text{C}_{22}\text{H}_{19}\text{NO}_8 + \text{Na}$ 448.1008, found 448.1011.



6g was obtained as a white solid in 72% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 98% by HPLC on Chiralpak OD-H column (10% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 16.32$ min, $t_{\text{major}} = 18.03$ min. m.p. 156-158 °C; $[\alpha]_{\text{D}}^{20}$

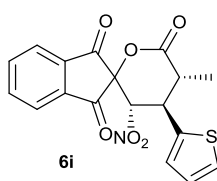
-117.2 ($c = 0.10$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.13$ -8.05 (m, 2H), 8.02-7.94 (m, 2H), 7.23-7.21 (m, 2H), 7.17-7.15 (m, 2H), 5.60 (d, $J = 12.4$ Hz, 1H), 4.11 (t, $J = 12.0$ Hz, 1H), 3.06-2.98 (m, 1H), 2.92-2.85 (m, 1H), 1.39 (d, $J = 6.8$ Hz, 3H), 1.23 (s, 3H), 1.22 (s, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.31, 190.08, 169.74, 149.73, 141.17, 140.20, 137.95, 137.43, 132.58, 127.63, 127.54, 124.98, 124.70, 87.36, 77.23, 77.10, 46.21, 41.52, 33.75, 23.81, 14.78$ ppm; ESI HRMS: calcd. For $\text{C}_{23}\text{H}_{21}\text{NO}_6 + \text{H}$ 408.1447, found 408.1450.



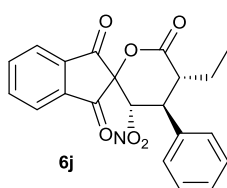
6h was obtained as a white solid in 66% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 95% by HPLC on Chiralpak OD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, $t_{\text{major}} = 8.85$ min, $t_{\text{minor}} = 18.05$ min. m.p. 152-154 °C; $[\alpha]_{\text{D}}^{20}$

-106.2 ($c = 0.09$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.13$ -8.06 (m, 2H), 8.02-7.95 (m, 2H), 7.45 (s, 1H), 6.33-6.28 (m, 2H), 5.69 (d, $J = 12.0$ Hz, 1H), 4.34 (t, $J = 11.6$ Hz, 1H), 3.28-3.20 (m, 1H), 1.45 (d, $J = 6.8$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 192.48, 189.77, 169.22, 147.41,$

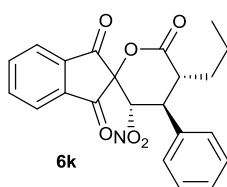
143.68, 141.09, 140.27, 137.88, 137.47, 125.03, 124.76, 110.77, 110.70, 84.91, 76.18, 40.69, 38.95, 14.94 ppm; ESI HRMS: calcd. For $C_{18}H_{13}NO_7+Na$ 378.0590, found 378.0593.



6i was obtained as a white solid in 69% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Chiralpak OD-H column (10% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 21.08$ min, $t_{\text{major}} = 27.83$ min. m.p. 156-158 °C; $[\alpha]_D^{20} -125.8$ ($c = 0.14$ in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): $\delta = 8.13-8.06$ (m, 2H), 8.01-7.94 (m, 2H), 7.33 (s, 1H), 6.97 (s, 2H), 5.54 (d, $J = 12.0$ Hz, 1H), 4.52 (t, $J = 11.6$ Hz, 1H), 3.07-3.05 (m, 1H), 1.49 (d, $J = 6.4$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$): $\delta = 192.26, 189.65, 168.82, 141.09, 140.20, 138.35, 138.02, 137.54, 127.78, 127.48, 126.32, 125.03, 124.73, 88.19, 76.78, 42.82, 42.36, 14.92$ ppm; ESI HRMS: calcd. For $C_{18}H_{13}NO_6S+Na$ 394.0361, found 394.0357.

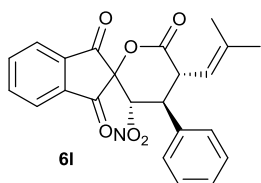


6j was obtained as a white solid in 78% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 99% by HPLC on Kromasil CelluCoat column (20% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 8.15$ min, $t_{\text{major}} = 13.39$ min. m.p. 169-171 °C; $[\alpha]_D^{20} -94.7$ ($c = 0.09$ in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): $\delta = 8.12-8.05$ (m, 2H), 8.01-7.93 (m, 2H), 7.40-7.32 (m, 3H), 7.28-7.26 (m, 2H), 5.61 (d, $J = 12.0$ Hz, 1H), 4.44 (t, $J = 12.0$ Hz, 1H), 3.12-3.08 (m, 1H), 2.08-2.02 (m, 1H), 1.56-1.48 (m, 1H), 1.12 (t, $J = 7.6$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$): $\delta = 192.19, 190.11, 168.56, 141.20, 140.11, 137.96, 137.41, 135.30, 129.52, 129.02, 127.69, 124.96, 124.67, 87.75, 77.23, 46.68, 42.77, 21.08, 9.71$ ppm; ESI HRMS: calcd. For $C_{21}H_{17}NO_6+Na$ 402.0954, found 402.0955.



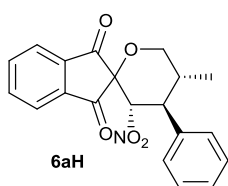
6k was obtained as a white solid in 75% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 99% by HPLC on Chiralpak AD-H column (10% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, $t_{\text{major}} = 11.22$ min, $t_{\text{minor}} = 12.98$ min. m.p. 155-156 °C; $[\alpha]_D^{20} -91.8$ ($c = 0.08$ in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): $\delta = 8.12-8.05$ (m, 2H), 8.02-7.94 (m, 2H), 7.41-7.33 (m, 3H), 7.27-7.26 (m, 2H), 5.59 (d, $J = 12.4$ Hz, 1H), 4.39 (t, $J = 12.0$ Hz, 1H), 3.13-3.08 (m, 1H), 1.92-1.85 (m, 1H), 1.69-1.46 (m, 3H), 0.87 (t, $J = 7.2$ Hz, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$):

δ = 191.23, 189.06, 167.79, 140.16, 139.10, 136.93, 136.39, 134.37, 128.47, 127.98, 126.69, 123.92, 123.63, 86.76, 76.21, 44.71, 42.57, 29.46, 17.86, 13.05 ppm; ESI HRMS: calcd. For $C_{22}H_{19}NO_6+Na$ 416.1110, found 416.1113.



6I was obtained as a white solid in 60% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 99% by HPLC on Chiralpak OD-H column (15% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, t_{minor} = 14.76 min, t_{major} = 15.86 min. m.p. 162-164 °C; $[\alpha]_D^{20}$

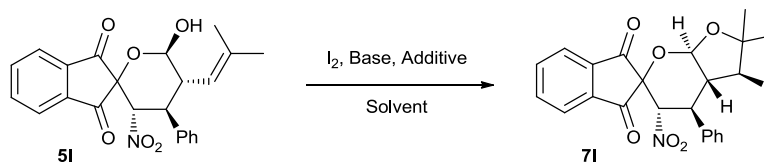
-125.2 (c = 0.10 in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): δ = 8.13-8.06 (m, 2H), 8.02-7.94 (m, 2H), 7.36-7.27 (m, 3H), 7.24-7.22 (m, 2H), 5.75 (d, J = 12.4 Hz, 1H), 5.36 (d, J = 9.2 Hz, 1H), 4.24 (t, J = 12.0 Hz, 1H), 3.81 (dd, J = 10.8 Hz, J = 9.2 Hz, 1H), 1.68 (s, 3H), 1.14 (s, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$): δ = 192.31, 189.98, 168.32, 141.18, 140.23, 139.57, 137.95, 137.47, 135.32, 129.13, 128.69, 127.97, 124.98, 124.72, 117.60, 86.67, 77.22, 46.55, 46.03, 25.53, 17.78 ppm; ESI HRMS: calcd. For $C_{23}H_{19}NO_6+Na$ 428.1110, found 428.1107.



6aH was obtained as a white solid in 75% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Chiralpak AD-H column (10% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, t_{major} = 12.75 min, t_{minor} = 23.03 min. m.p. 183-184 °C; $[\alpha]_D^{20}$ -145.8 (c

= 0.08 in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): δ = 8.06-8.01 (m, 2H), 7.93-7.86 (m, 2H), 7.37-7.28 (m, 5H), 5.41 (d, J = 12.0 Hz, 1H), 4.25 (t, J = 12.0 Hz, 1H), 3.93 (dd, J = 12.0 Hz, J = 5.2 Hz, 1H), 3.85 (t, J = 12.0 Hz, 1H), 2.35-2.27 (m, 1H), 0.78 (d, J = 6.4 Hz, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$): δ = 194.83, 193.66, 141.50, 140.09, 137.09, 137.04, 136.60, 128.94, 127.99, 124.49, 124.16, 88.45, 73.97, 69.77, 47.58, 36.19, 13.93 ppm; ESI HRMS: calcd. For $C_{20}H_{17}NO_5+Na$ 374.1004, found 374.1007.

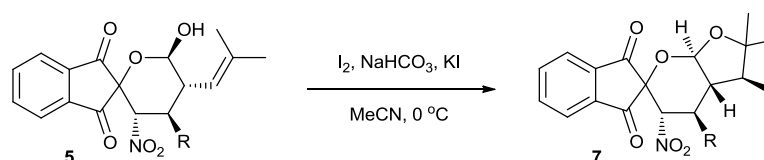
3. Screening studies to improve the yield of hexahydrofuro[2,3-*b*]pyran derivatives **7**^a



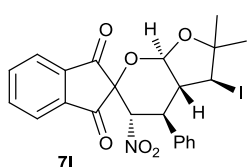
entry	Base	additive	Solvent ^b	Time (h)	Yield ^c (%)	ee ^d
1	Na ₂ CO ₃	-	THF	12	49	82
2	K ₂ CO ₃	-	THF	12	45	78
3	NaHCO ₃	-	THF	12	48	95
4	NaHCO ₃	KI	THF	6	53	96
5	NaHCO ₃	KI	Dioxane	8	31	95
6	NaHCO ₃	KI	DCM	8	38	96
7	NaHCO ₃	KI	MeCN	2	61	97
8	NaHCO₃	KI	MeCN^e	2	71	97

^a Unless noted otherwise, reactions were performed with allylic hemiacetal **5X** (0.4 mmol), I₂ (1.2 mmol), Base (1.2mmol), and additive (0.04 mmol) at 0 °C in 4 mL solvent for the times specified (TLC monitoring). ^b The commercially available solvent was used without distillation or drying. ^c Isolated yield. ^d By chiral HPLC analysis. ^e The anhydrous acetonitrile was used.

4. General procedure for the asymmetric synthesis of **7**

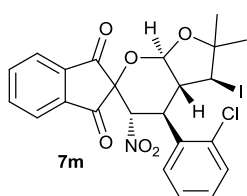


To a solution of allylic hemiacetal **5** (0.4 mmol) in 4 mL anhydrous MeCN were added I₂ (304.8 mg, 1.2 mmol), NaHCO₃ (100.8 mg, 1.2 mmol), and KI (6.6 mg, 0.04 mmol). The mixture was stirred for the times specified (TLC monitoring) at 0 °C. Ethyl acetate (20 mL) was added and the resulting mixture washed with saturated aqueous sodium thiosulfate (30 mL). The separated aqueous phase was extracted with ether (3×10 mL) and the combined organic solution dried and evaporated. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate = 20:1) to give tetracyclic hexahydrofuro[2,3-*b*]pyran-based compounds **7**. For the reason that the major and minor diastereomers of compounds **7** could not be isolated in pure form by column chromatography, the dr values of **7** were calculated by chiral HPLC analysis.



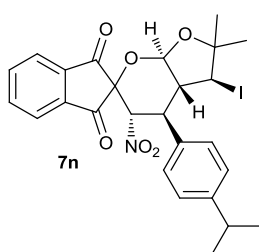
7l was obtained as a white solid in 71% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Chiralpak AD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{major}} = 10.09$ min, $t_{\text{minor}} = 12.14$ min. m.p. 194-196 °C; $[\alpha]_{\text{D}}^{20}$

-124.2 ($c = 0.08$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.09$ -8.01 (m, 2H), 7.92-7.90 (m, 2H), 7.44-7.31 (m, 5H), 5.66 (d, $J = 8.0$ Hz, 1H), 5.56 (d, $J = 10.8$ Hz, 1H), 4.18 (t, $J = 11.6$ Hz, 1H), 3.94 (d, $J = 12.4$ Hz, 1H), 2.53 (td, $J = 12.0$ Hz, $J = 8.0$ Hz, 1H), 1.51 (s, 3H), 1.36 (s, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.71, 171.25, 140.76, 140.41, 137.01, 136.99, 131.77, 129.03, 128.86, 128.66, 124.82, 124.60, 100.20, 85.95, 85.75, 52.08, 45.90, 30.61, 26.68, 25.22$ ppm; ESI HRMS: calcd. For $\text{C}_{23}\text{H}_{20}\text{INO}_6 + \text{Na}$ 556.0233, found 556.0231.



7m was obtained as a white solid in 69% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 94% by HPLC on Chiralpak AD-H column (20% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{major}} = 6.56$ min, $t_{\text{minor}} = 12.24$ min. m.p. 206-207 °C; $[\alpha]_{\text{D}}^{20}$

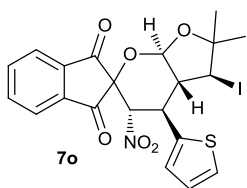
-149.5 ($c = 0.32$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.10$ -8.02 (m, 2H), 7.93-7.92 (m, 2H), 7.51-7.47 (m, 2H), 7.38-7.25 (m, 2H), 5.65-5.62 (m, 2H), 5.05 (t, $J = 11.6$ Hz, 1H), 4.07 (d, $J = 12.4$ Hz, 1H), 2.41 (td, $J = 12.0$ Hz, $J = 8.0$ Hz, 1H), 1.49 (s, 3H), 1.38 (s, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.44, 191.14, 140.73, 140.41, 136.98, 136.94, 136.21, 132.80, 130.14, 129.25, 127.31, 126.78, 124.88, 124.68, 100.26, 85.95, 84.30, 81.83, 53.04, 40.26, 30.59, 26.72, 24.99$ ppm; ESI HRMS: calcd. For $\text{C}_{23}\text{H}_{19}\text{ClINO}_5 + \text{Na}$ 589.9843, found 589.9841.



7n was obtained as a white solid in 65% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 98% by HPLC on Kromasil CelluCoat column (5% 2-propanol/n-hexane, 1 mL/min), UV 220 nm, $t_{\text{minor}} = 9.09$ min, $t_{\text{major}} = 10.13$ min. m.p. 171-173 °C; $[\alpha]_{\text{D}}^{20}$

-139.6 ($c = 0.18$ in CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3): $\delta = 8.09$ -8.01 (m, 2H), 7.93-7.90 (m, 2H), 7.34-7.24 (m, 4H), 5.66 (d, $J = 8.0$ Hz, 1H), 5.55 (d, $J = 11.2$ Hz, 1H), 4.15 (t, $J = 11.6$ Hz, 1H), 3.92 (d, $J = 12.4$ Hz, 1H), 2.94-2.87 (m, 1H), 2.50 (td, $J = 12.4$ Hz, $J = 8.0$ Hz, 1H), 1.51 (s, 3H), 1.36 (s, 3H), 1.24 (s, 3H), 1.22 (s, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.76, 191.37, 149.66, 140.78, 140.42, 137.00, 131.32, 126.88, 125.38, 124.81, 124.58, 100.21,$

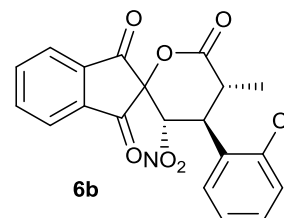
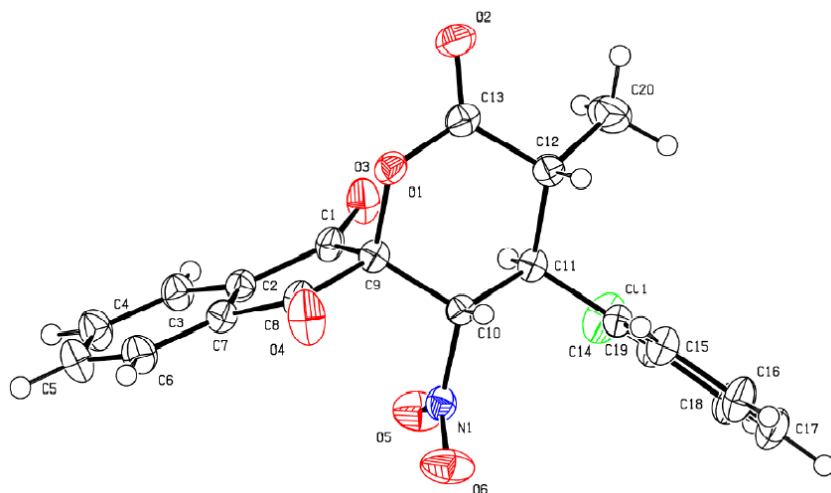
86.08, 85.73, 52.11, 45.53, 33.75, 30.60, 26.68, 25.20, 24.00 ppm; ESI HRMS: calcd. For $C_{26}H_{26}INO_6+H$ 576.0883, found 576.0886.



7o was obtained as a white solid in 61% yield for two steps after flash chromatography and the enantiomeric excess was determined to be 97% by HPLC on Kromasil CelluCoat column (5% 2-propanol/n-hexane, 1 mL/min), UV 254 nm, $t_{\text{minor}} = 14.13$ min, $t_{\text{major}} = 28.33$ min. m.p. 188-190 °C; $[\alpha]_D^{20}$

-157.6 ($c = 0.06$ in CH_2Cl_2); 1H NMR (400 MHz, $CDCl_3$): $\delta = 8.10$ -8.00 (m, 2H), 7.95-7.90 (m, 2H), 7.30 (d, $J = 5.2$ Hz, 1H), 7.13 (d, $J = 3.2$ Hz, 1H), 7.02 (dd, $J = 4.8$ Hz, $J = 3.6$ Hz, 1H), 5.65 (d, $J = 8.0$ Hz, 1H), 5.46 (d, $J = 10.8$ Hz, 1H), 4.51 (t, $J = 11.6$ Hz, 1H), 3.98 (d, $J = 12.4$ Hz, 1H), 2.50 (td, $J = 12.0$ Hz, $J = 8.0$ Hz, 1H), 1.53 (s, 3H), 1.38 (s, 3H) ppm; ^{13}C NMR (100 MHz, $CDCl_3$): $\delta = 193.61, 190.93, 140.71, 140.42, 137.13, 137.06, 127.18, 125.12, 124.86, 124.63, 100.06, 89.88, 87.07, 85.84, 53.34, 30.60, 26.71, 24.91$ ppm; ESI HRMS: calcd. For $C_{21}H_{18}INO_6S+Na$ 561.9797, found 561.9798.

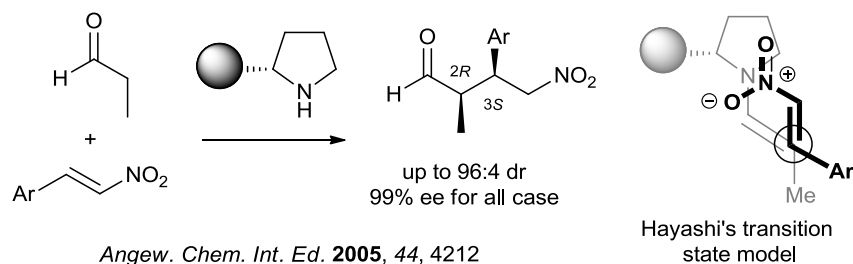
5. Crystal data of 6b



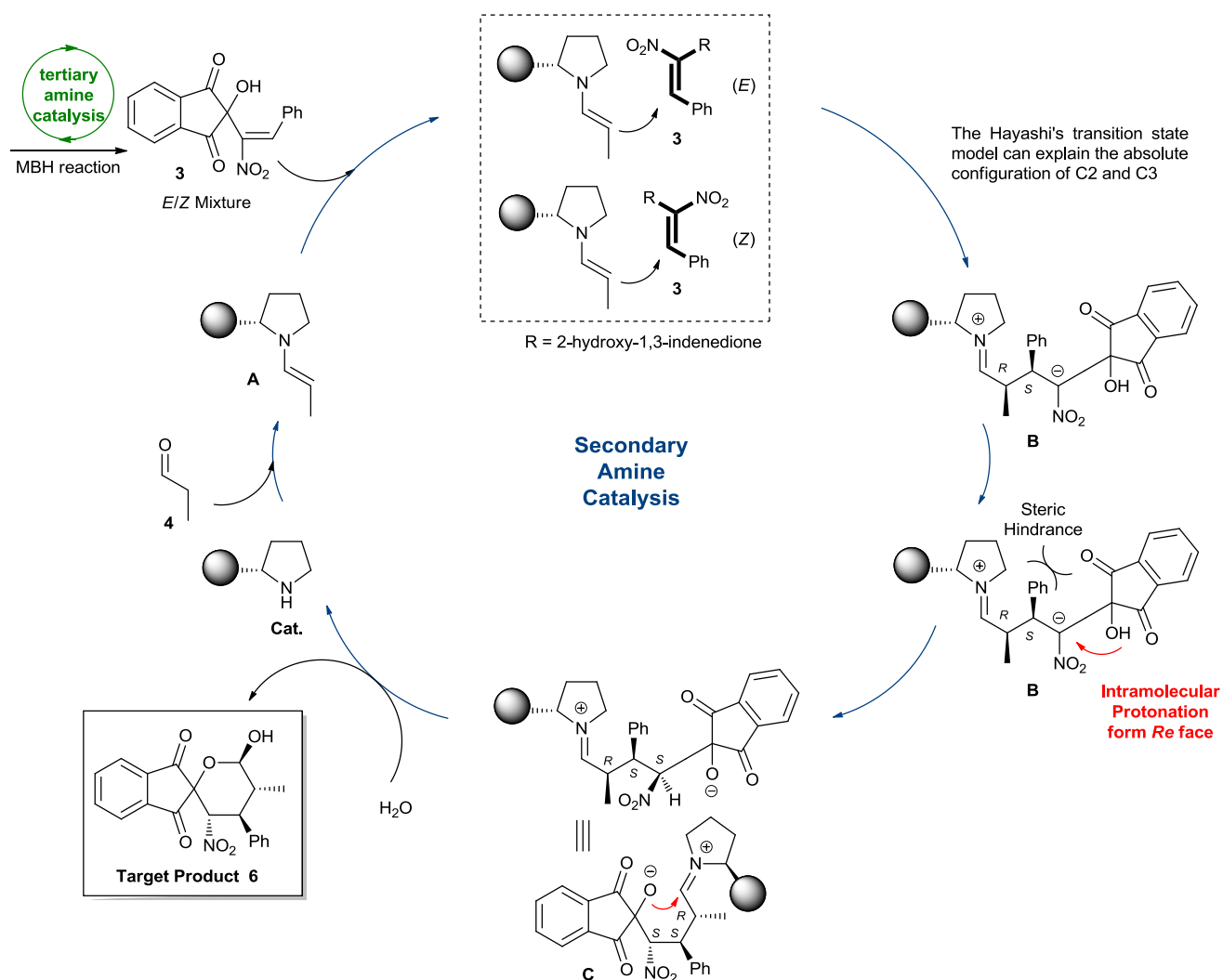
Empirical formula	C ₂₀ H ₁₄ ClNO ₆
Formula weight	399.77
Temperature/K	143(2)
Crystal system	orthorhombic
Space group	P212121
a/Å	8.1843(16)
b/Å	10.679(2)
c/Å	21.790(4)
α/°	90.00
β/°	90.00
γ/°	90.00
Volume/Å ³	1904.4(6)
Z	4
ρ _{calc} /mm ³	1.394
m/mm ⁻¹	0.238
F(000)	824.0
Crystal size/mm ³	0.4 × 0.38 × 0.37
2θ range for data collection	5.34 to 58.28 °
Index ranges	-11 ≤ h ≤ 11, -14 ≤ k ≤ 13, -29 ≤ l ≤ 29
Reflections collected	16915
Independent reflections	5110[R(int) = 0.0281]
Data/restraints/parameters	5110/0/254
Goodness-of-fit on F ²	1.000
Final R indexes [I ≥ 2σ (I)]	R1 = 0.0340, wR2 = 0.0802
Final R indexes [all data]	R1 = 0.0363, wR2 = 0.0819
Largest diff. peak/hole / e Å ⁻³	0.23/-0.30
Flack parameter	0.03(5)

6. Proposed catalytic cycles for the enantioselective transformation

✂ In 2005, Yujiro Hayashi demonstrated that diphenylprolinol silyl ethers were efficient chiral secondary amine catalyst in the asymmetric Michael reaction of aldehydes and nitroalkenes. A reasonable transition state model has been proposed to explain the high diastereoselectivities and excellent enantioselectivities of this reaction. The Hayashi's transition state model can explain the absolute configuration of C2 (α -position of CHO) and C3 (β -position of NO₂). (ACIE. 2005, 44, 4212)

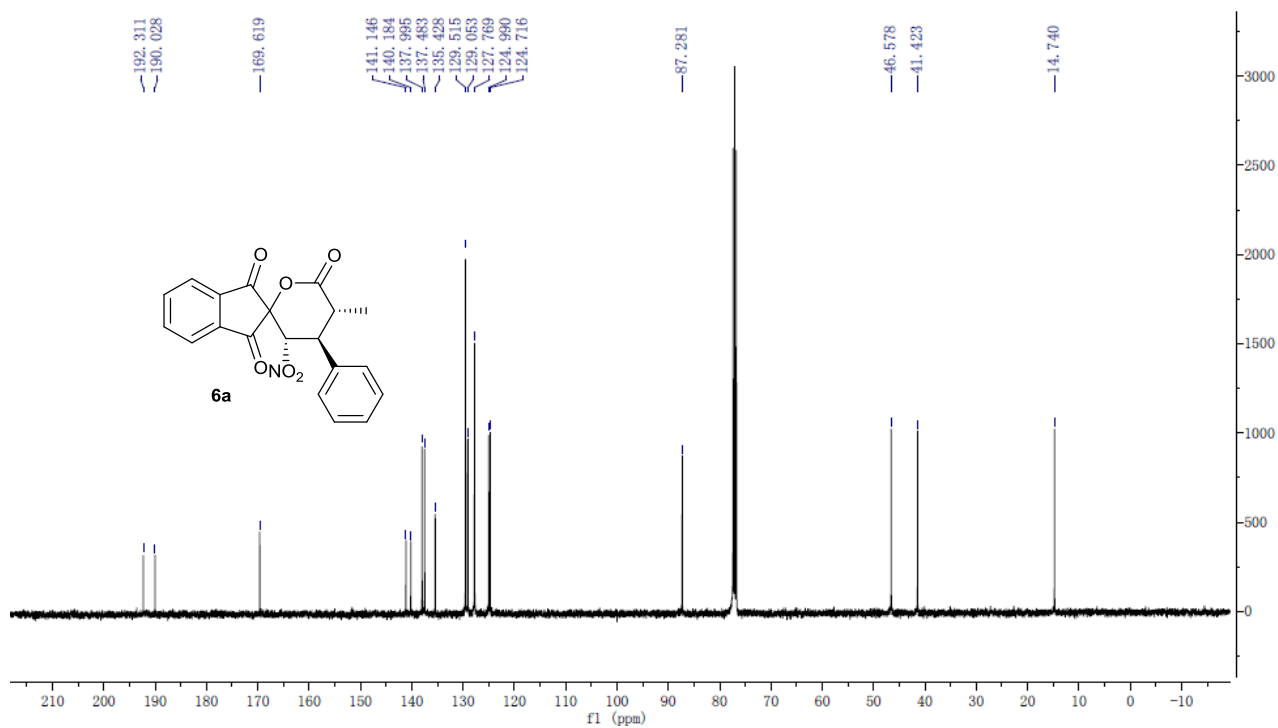
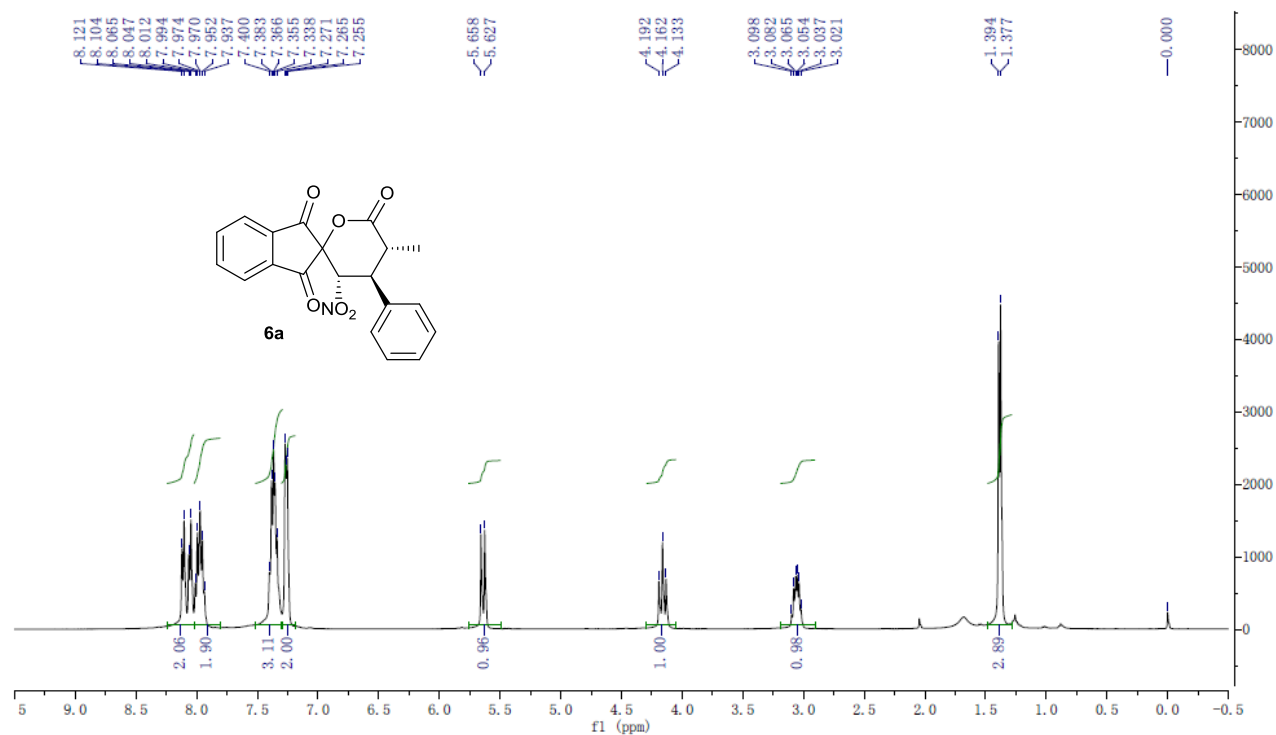


Based on Hayashi's transition state model, we proposed the plausible catalytic cycles and transition-state structures which were responsible for the enantioselective transformation:



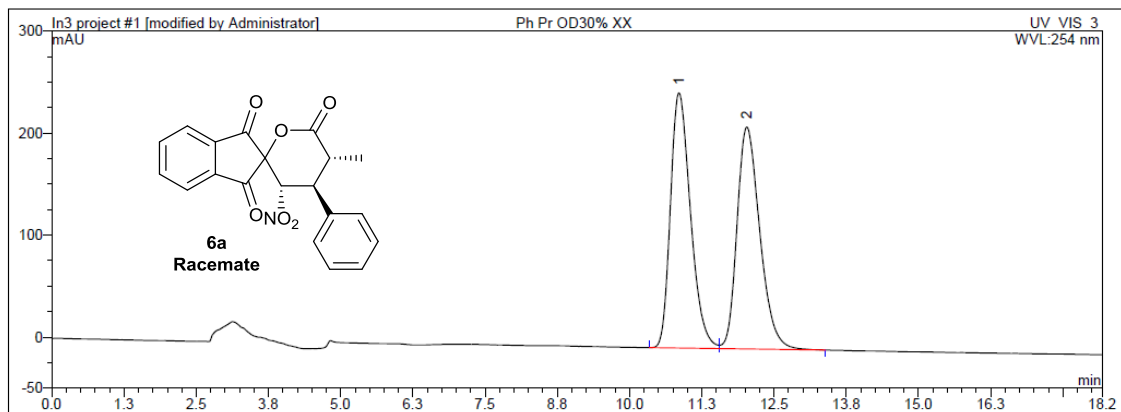
The plausible mechanism includes two catalytic cycles: tertiary amine catalysis and secondary amine catalysis. In the first catalytic cycle, the tertiary amine-catalyzed MBH reaction of conjugated nitroalkene with ninhydrin can provide tertiary alcohol intermediate **3** (E/Z mixtures), which then participates directly in the second catalytic cycle by serving as the receptor in an asymmetric Michael addition. The activated nucleophilic enamine specie **A** would be generated from aldehyde **4** and secondary amine catalyst. The transition state structure of asymmetric Michael addition between enamine **A** and nitroalkene derivative **3** is similar to Hyashi's model. Therefore, Zwitterion intermediate **B** could be obtained with good stereoselectivities at C2 (α -position of CHO) and C3 (β -position of NO₂). It is noteworthy that the E/Z configuration of MBH tertiary alcohol **3** will not affect the chirality of C4 (α -position of NO₂), owing to the generation of achiral carbanion at this position. Influenced by the steric hindrance effect of benzene ring at C3 position, intramolecular C4 protonation from *Re* face will afford Zwitterion intermediate **C** with high diastereoselectivity. Subsequent hydrolysis and hemi-acetalization provides the desired oxa-spirocyclic indanone **6** with excellent ee and good dr value, and the catalyst is regenerated.

7. NMR spectra and HPLC chromatograms



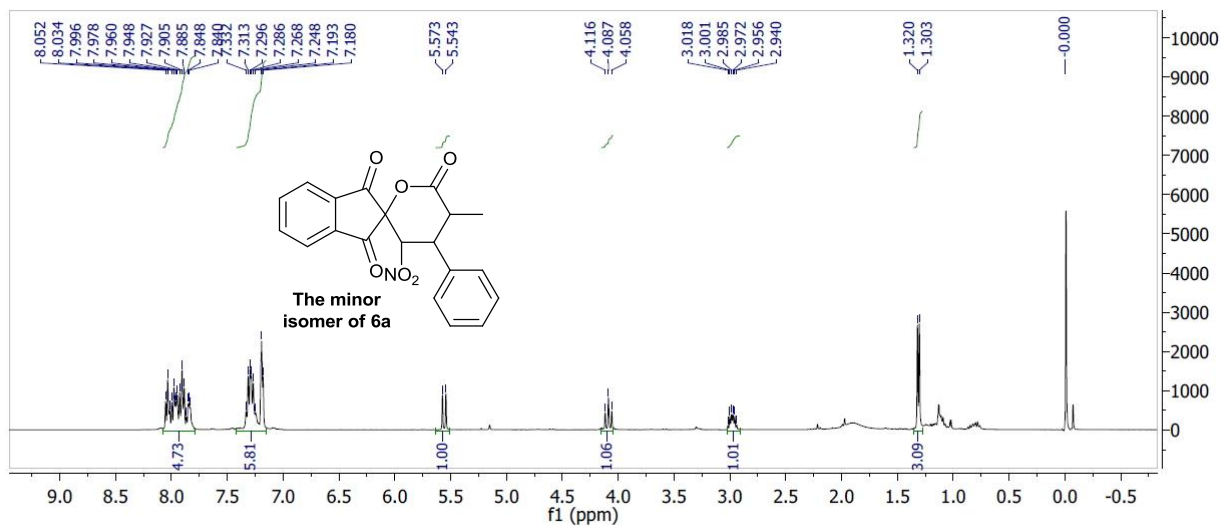
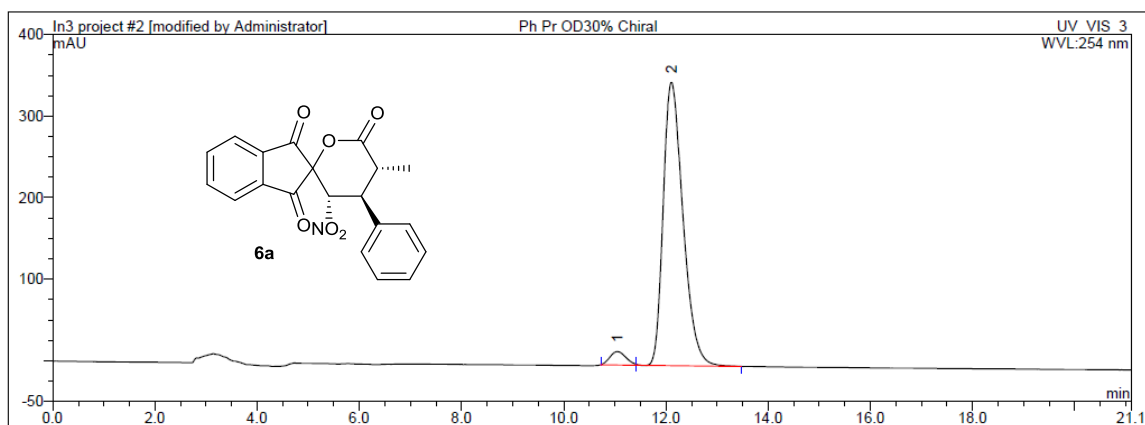
Peak Analysis Report

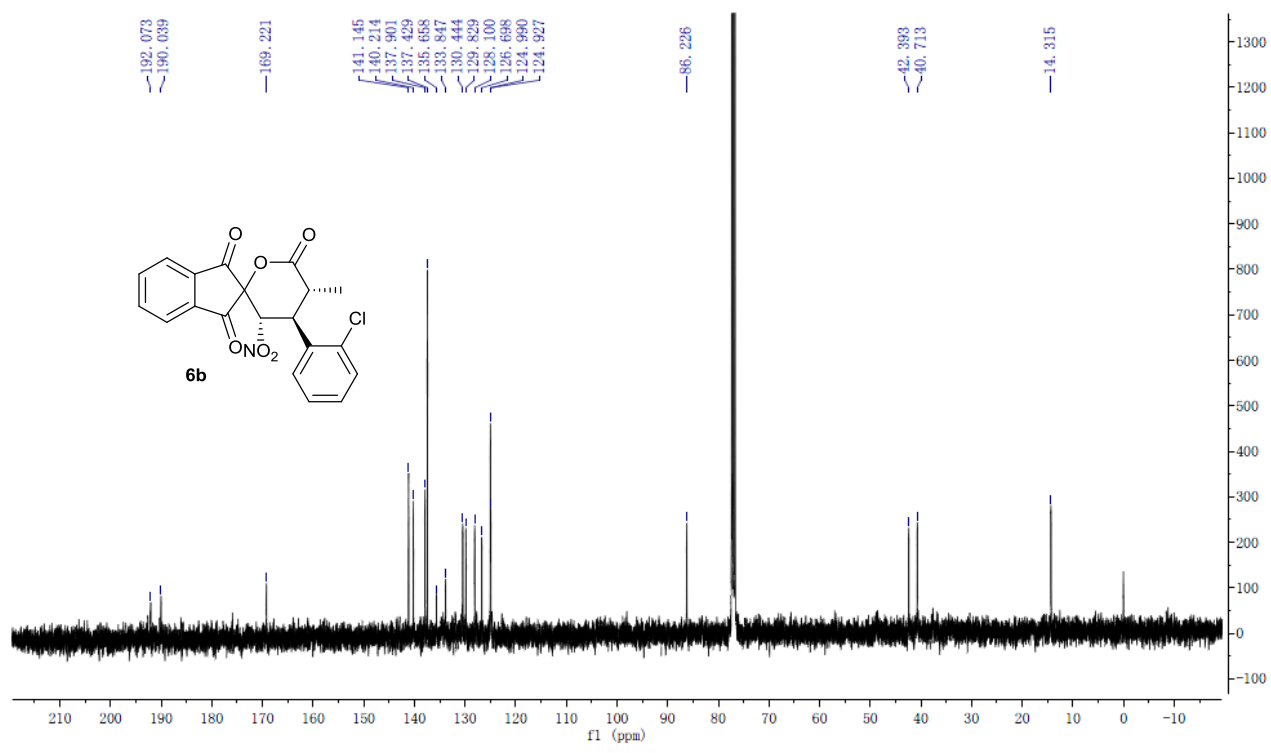
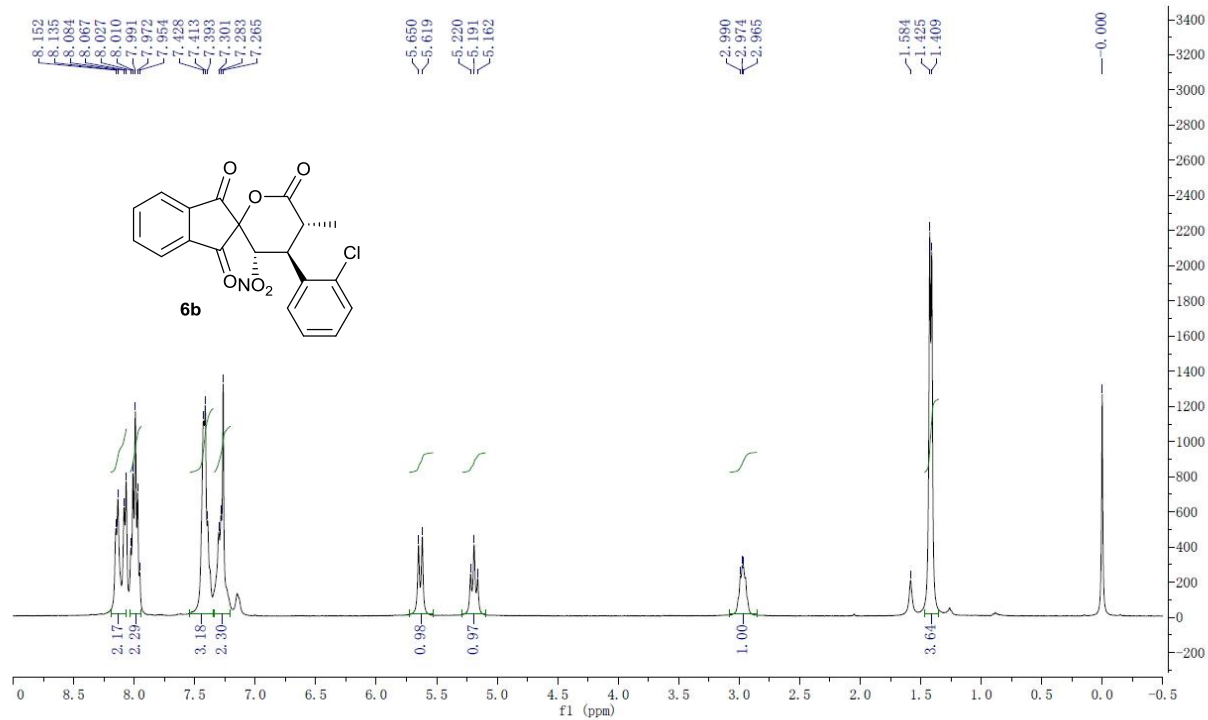
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	10.86	100.687	50.30	250.073	n.a.
2	n.a.	12.03	99.479	49.70	217.544	n.a.



Peak Analysis Report

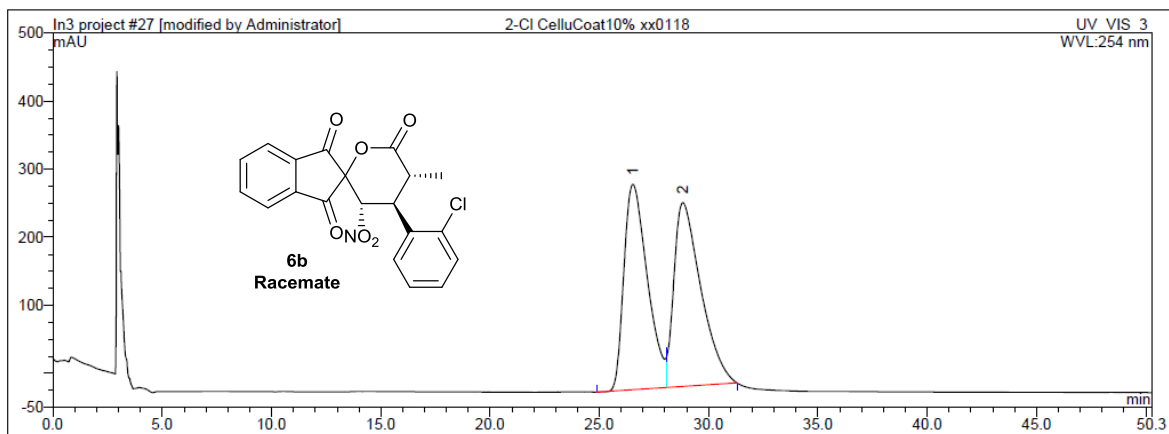
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	11.05	5.913	3.66	16.395	n.a.
2	n.a.	12.11	155.766	96.34	347.363	n.a.





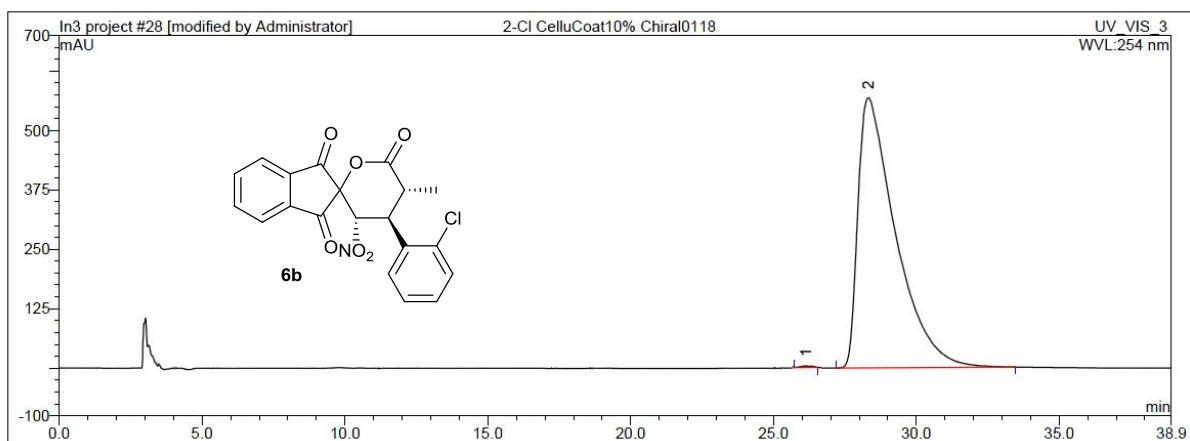
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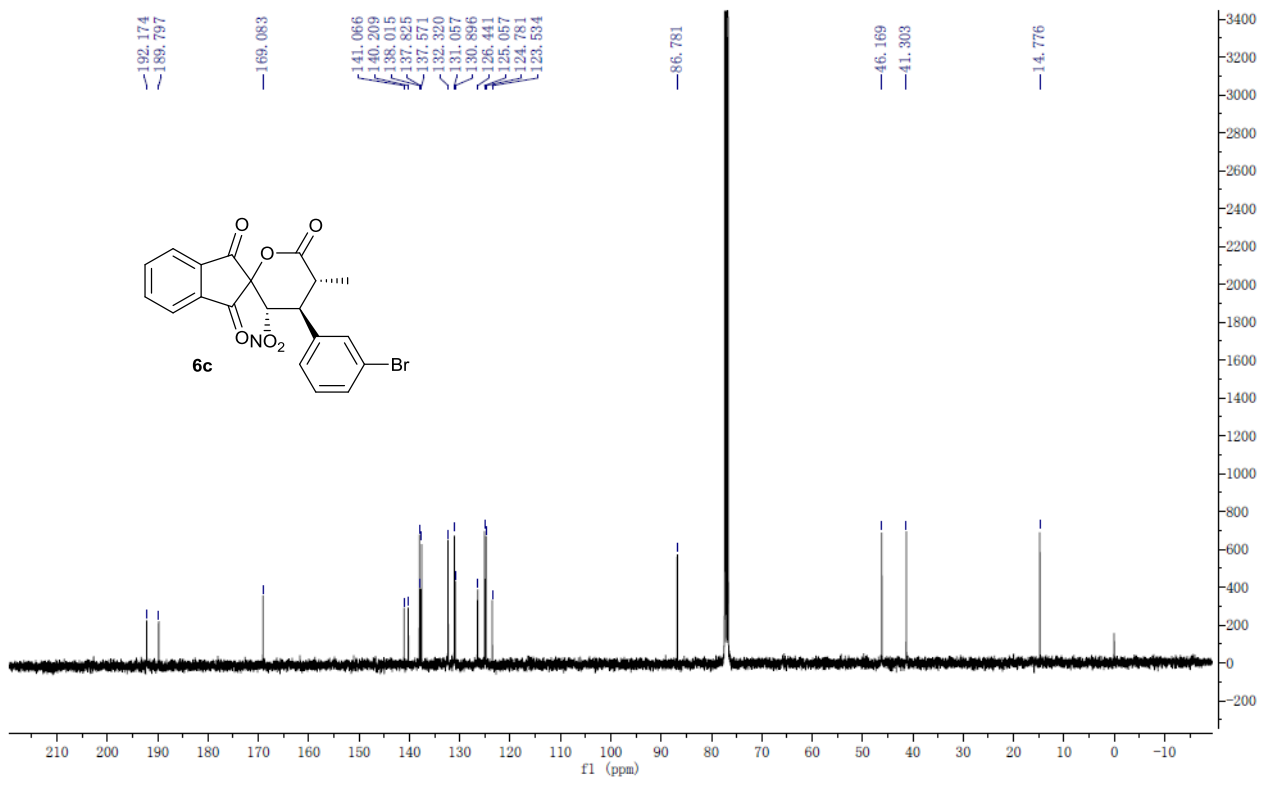
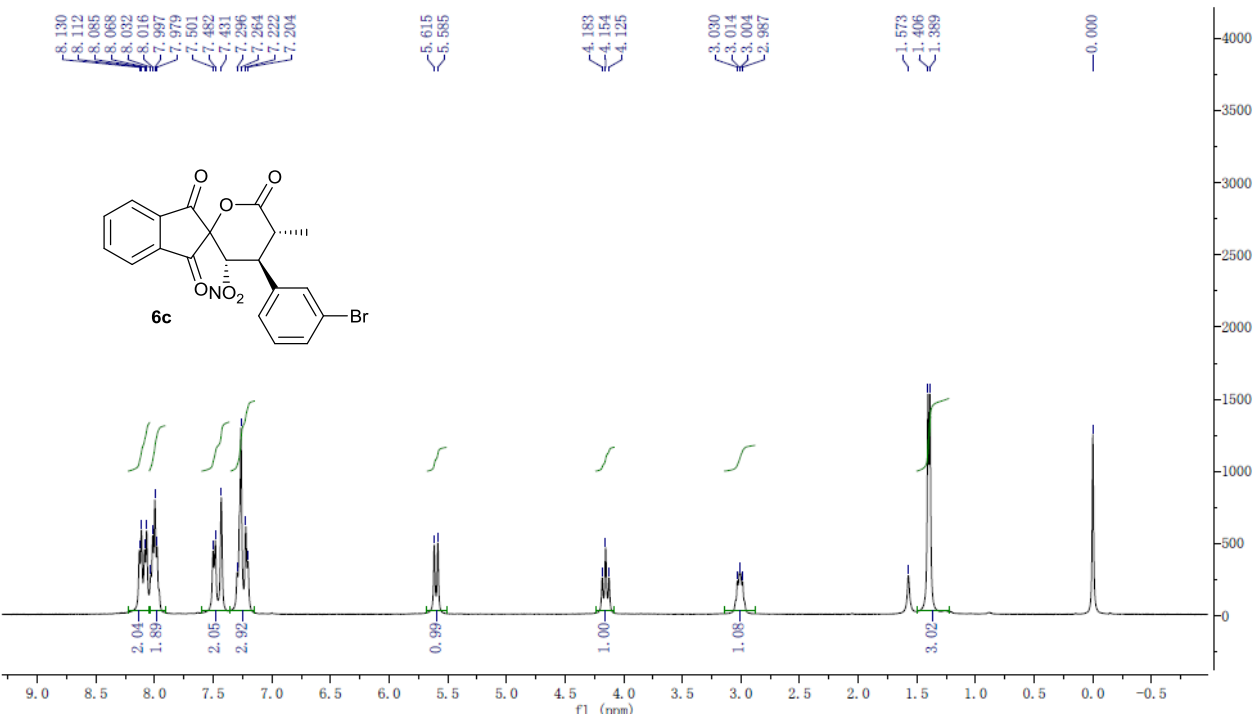
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	26.53	376.848	49.53	301.463	n.a.
2	n.a.	28.82	383.954	50.47	269.928	n.a.



Peak Analysis Report

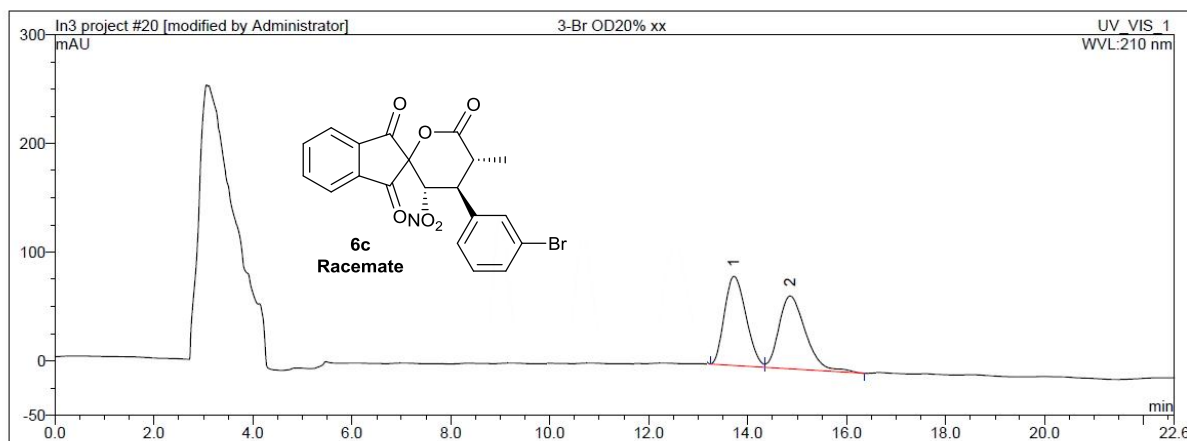
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	26.42	1.508	0.17	2.785	n.a.
2	n.a.	28.33	865.230	99.83	567.553	n.a.





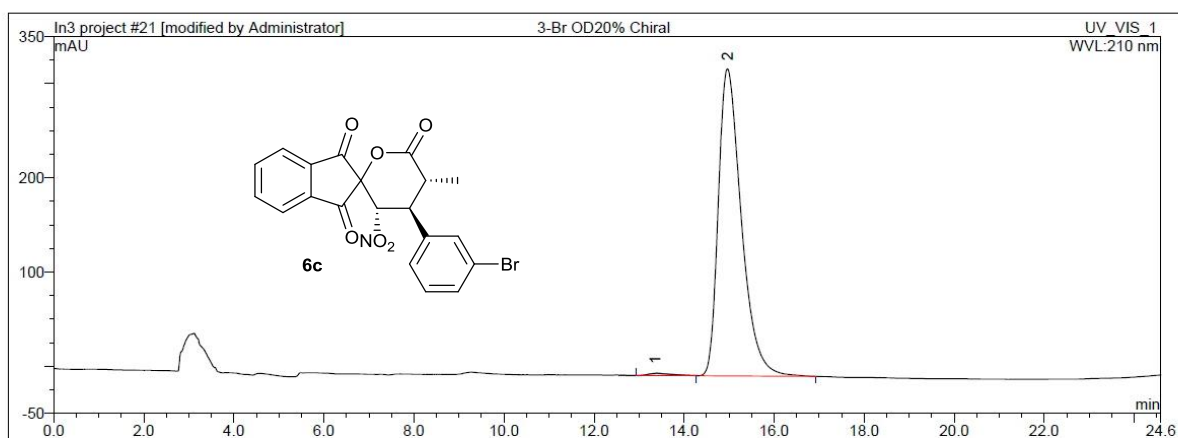
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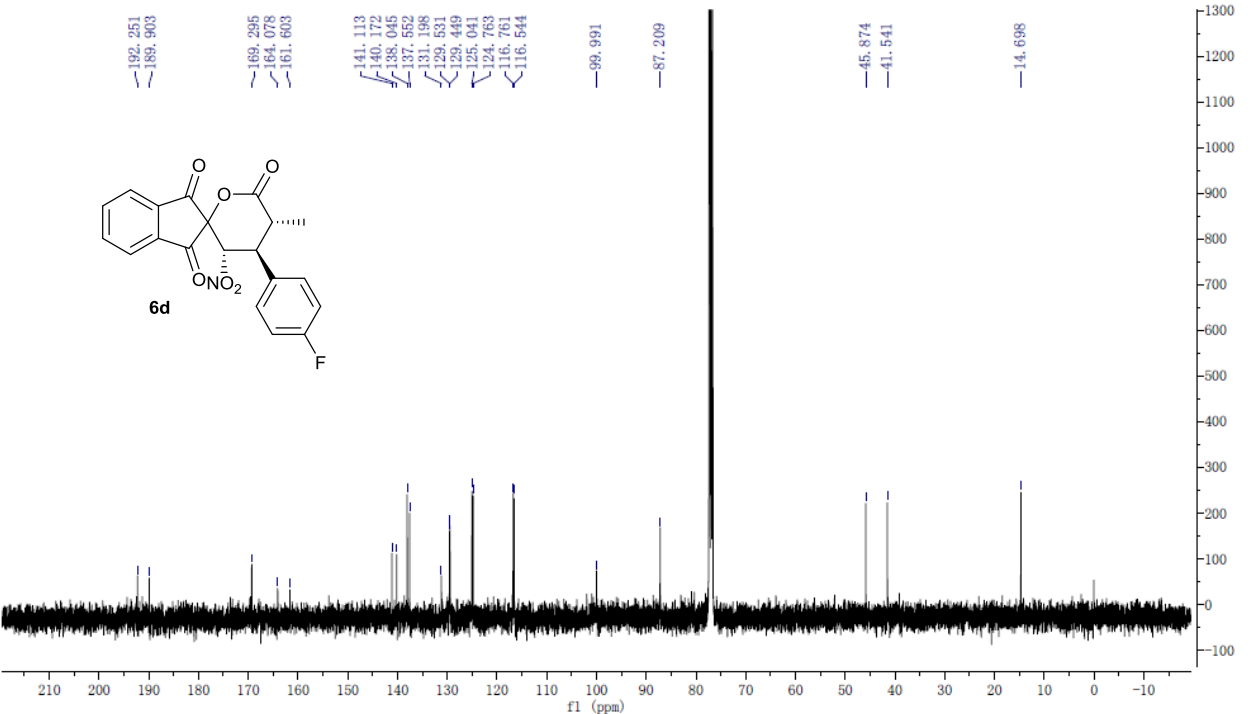
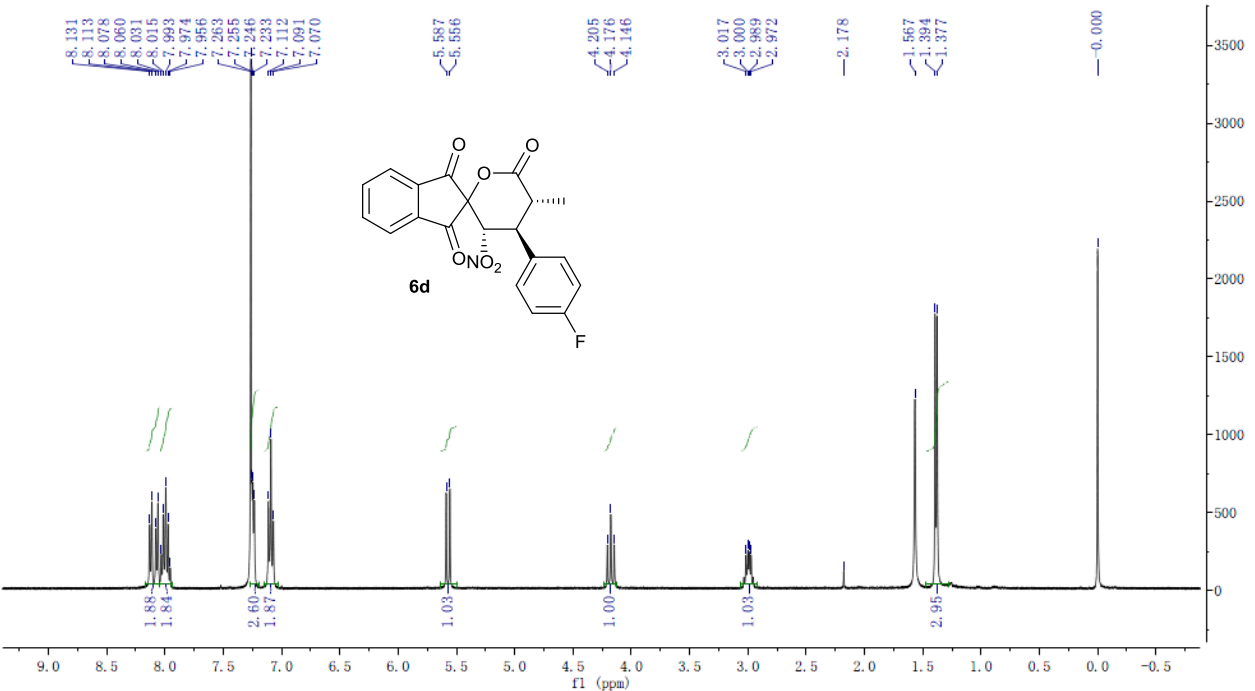
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	13.66	41.322	49.80	81.882	n.a.
2	n.a.	14.79	41.654	50.20	66.969	n.a.



Peak Analysis Report

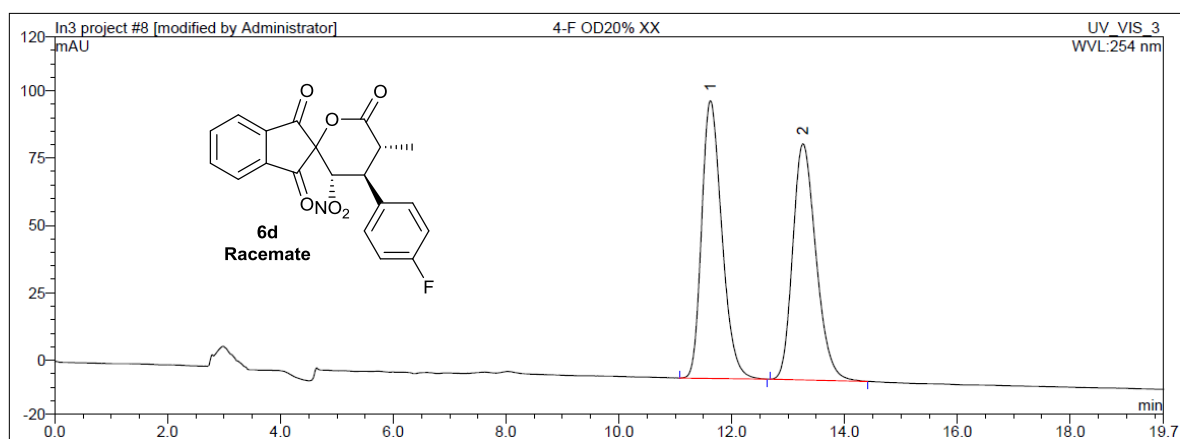
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	13.62	1.383	0.71	2.471	n.a.
2	n.a.	14.97	193.164	99.29	325.816	n.a.





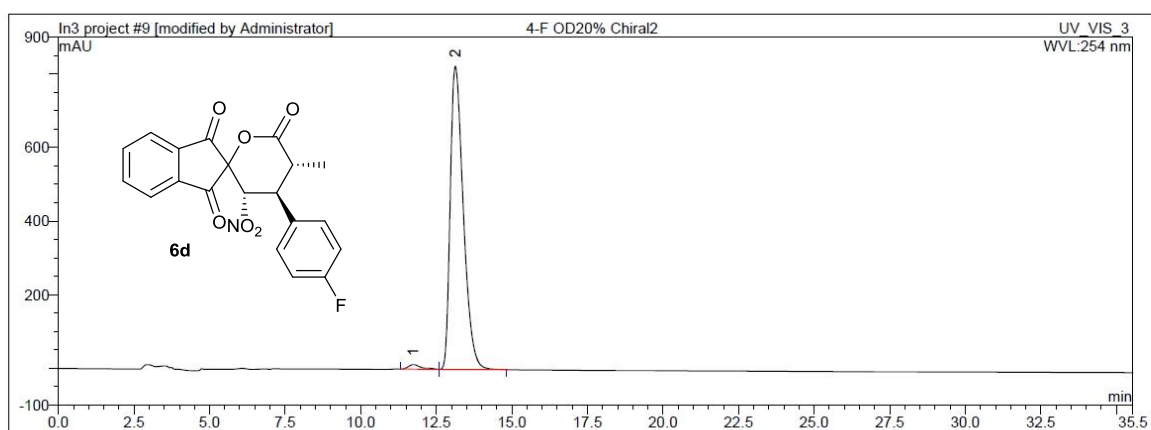
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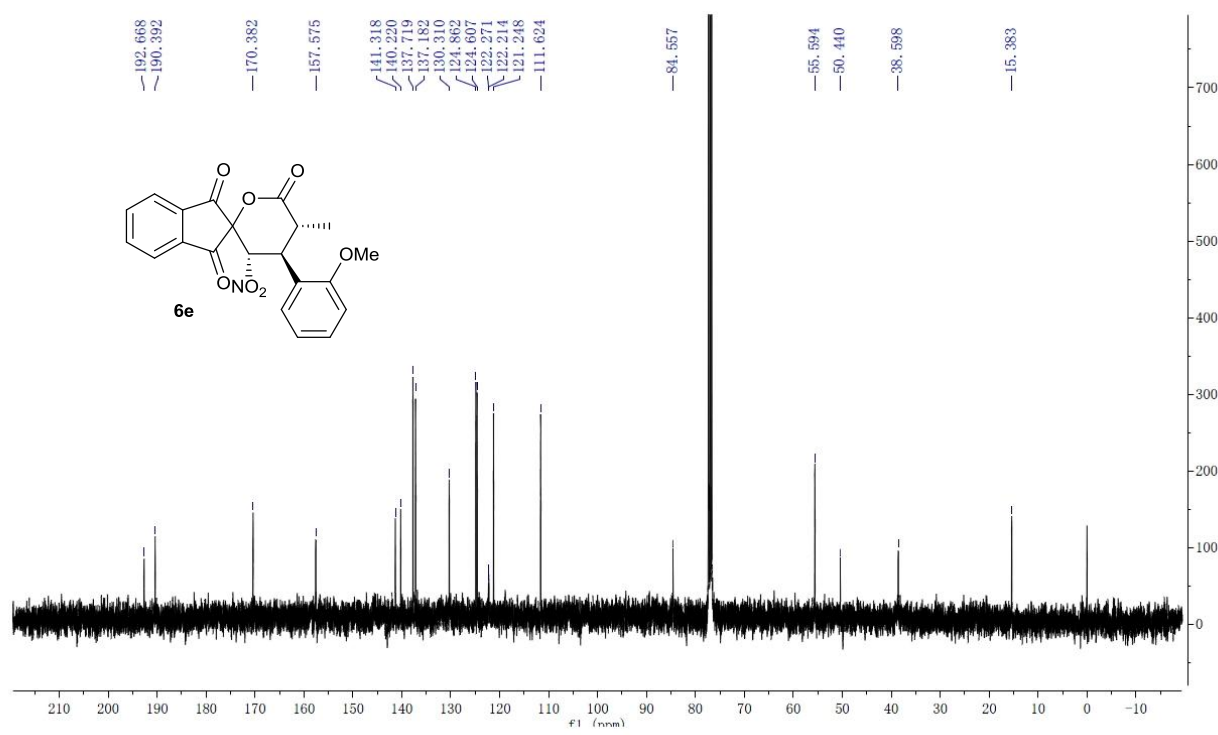
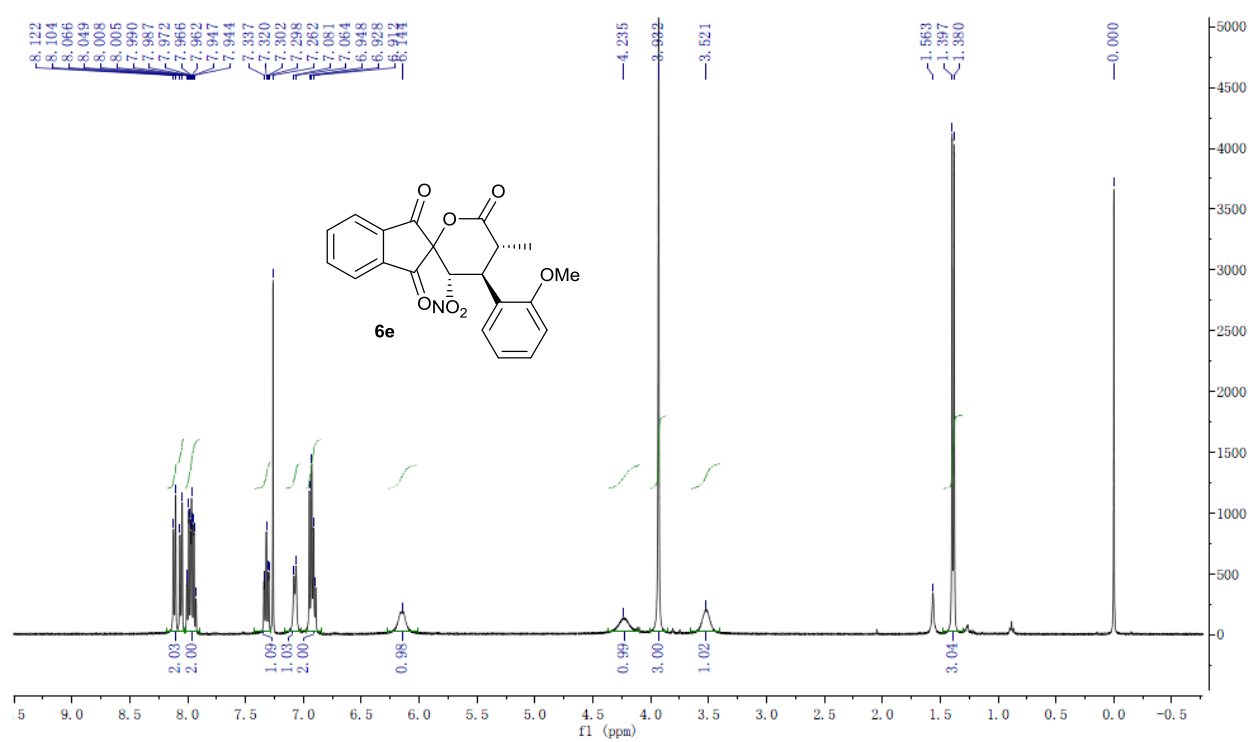
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	11.63	42.711	50.47	103.008	n.a.
2	n.a.	13.27	41.908	49.53	87.620	n.a.



Peak Analysis Report

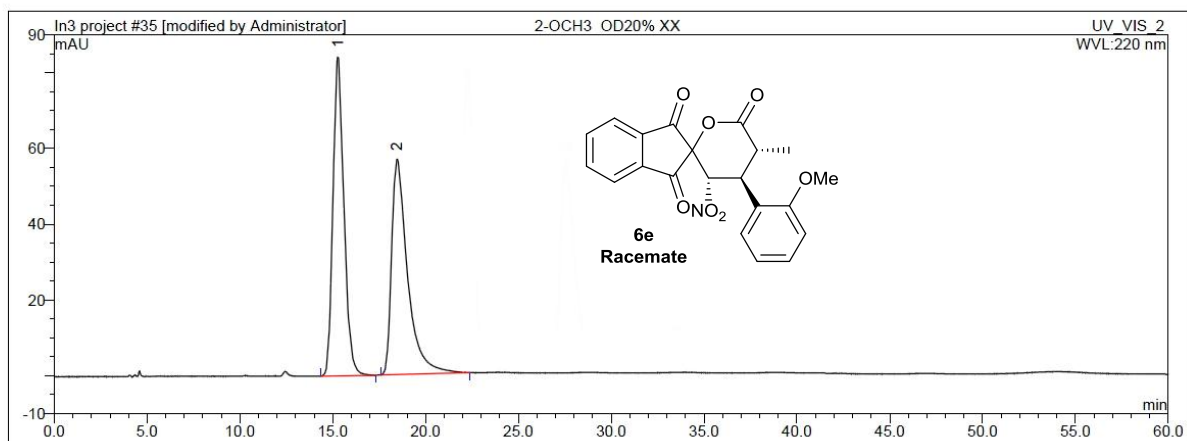
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	11.74	5.833	1.40	12.197	n.a.
2	n.a.	13.13	412.027	98.60	823.225	n.a.





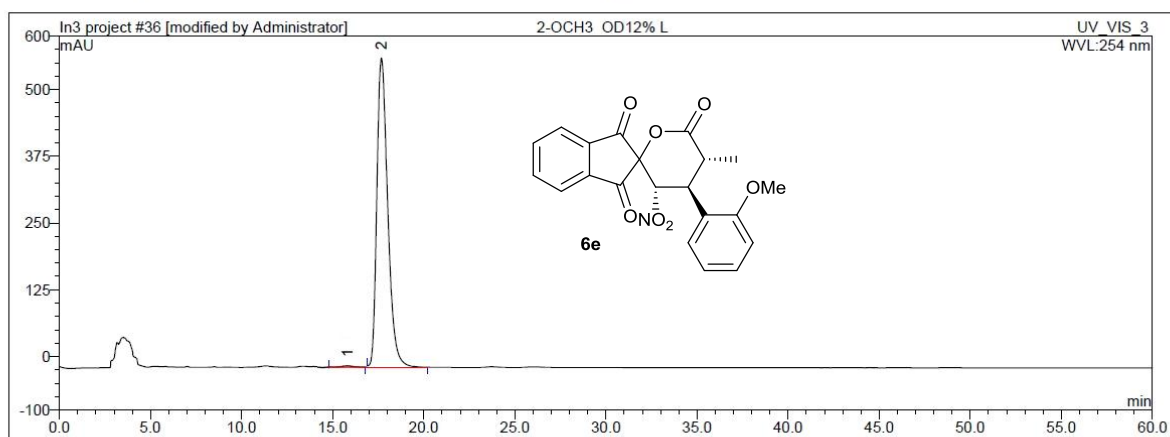
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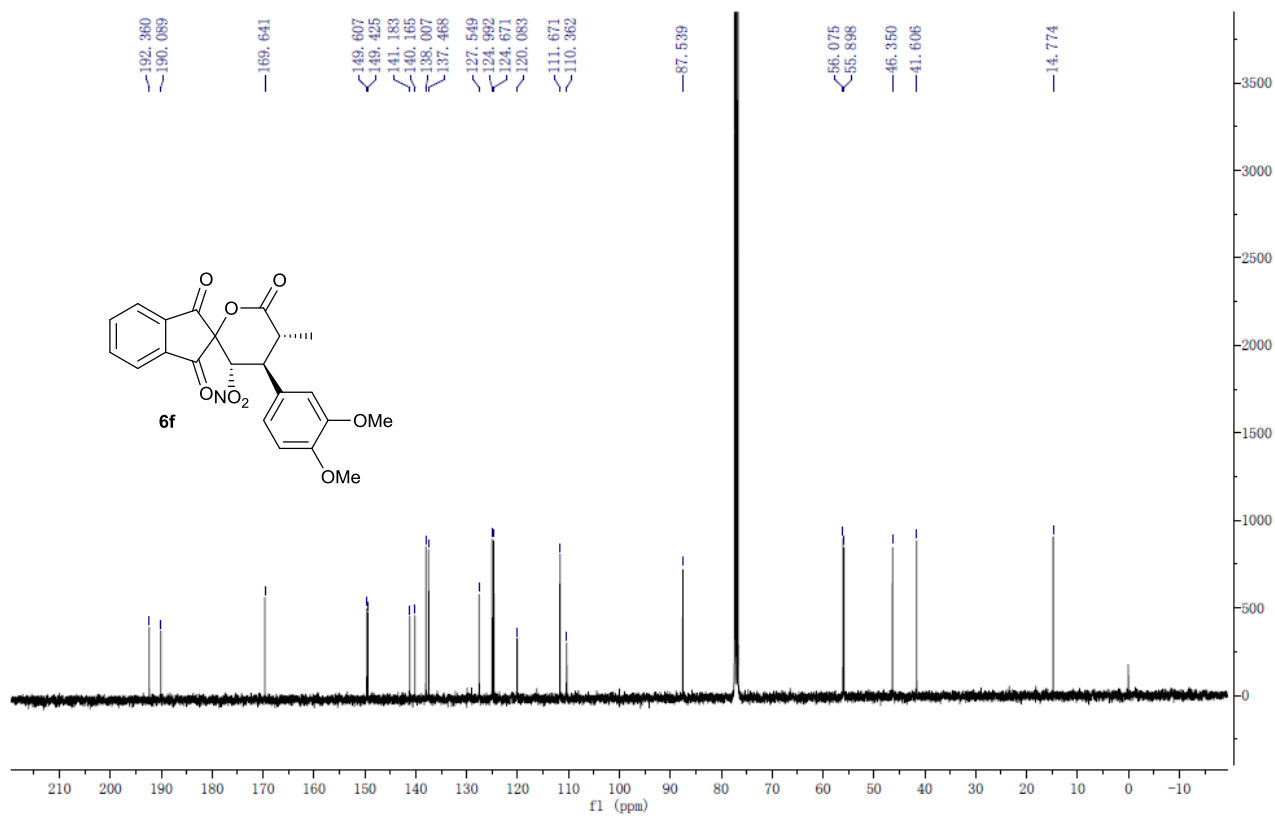
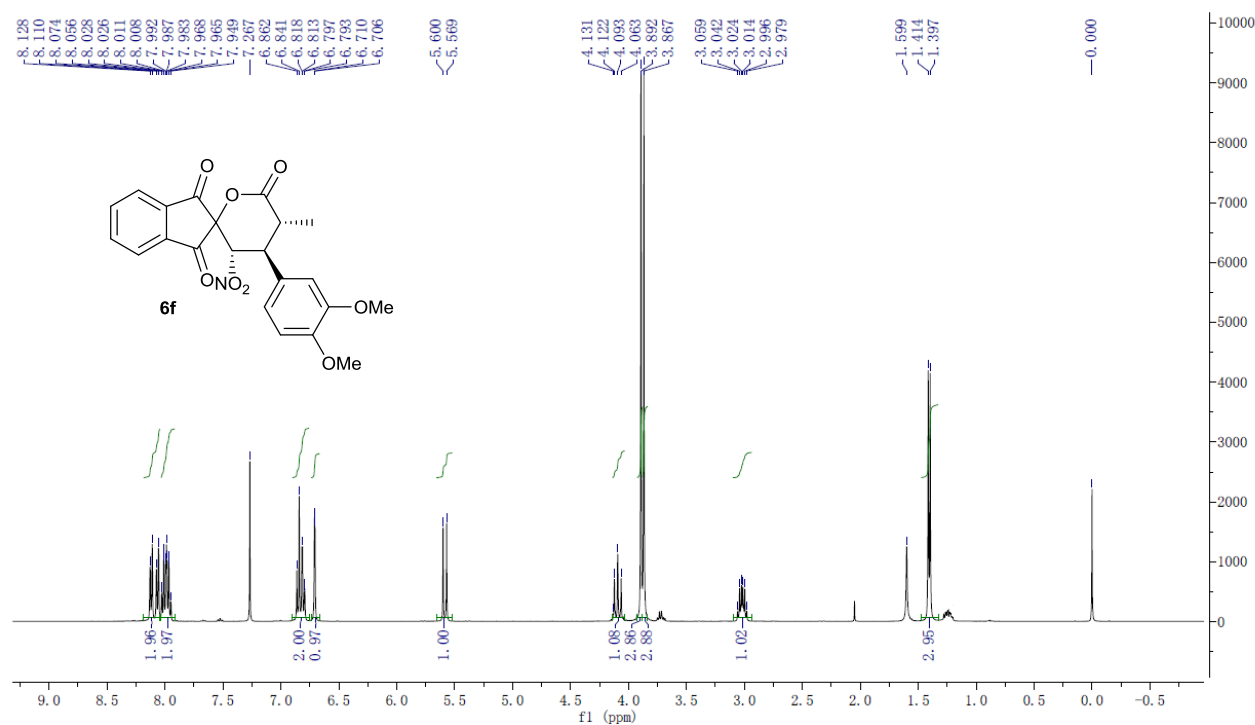
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	15.62	39.256	50.28	84.166	n.a.
2	n.a.	17.55	38.824	49.72	56.744	n.a.



Peak Analysis Report

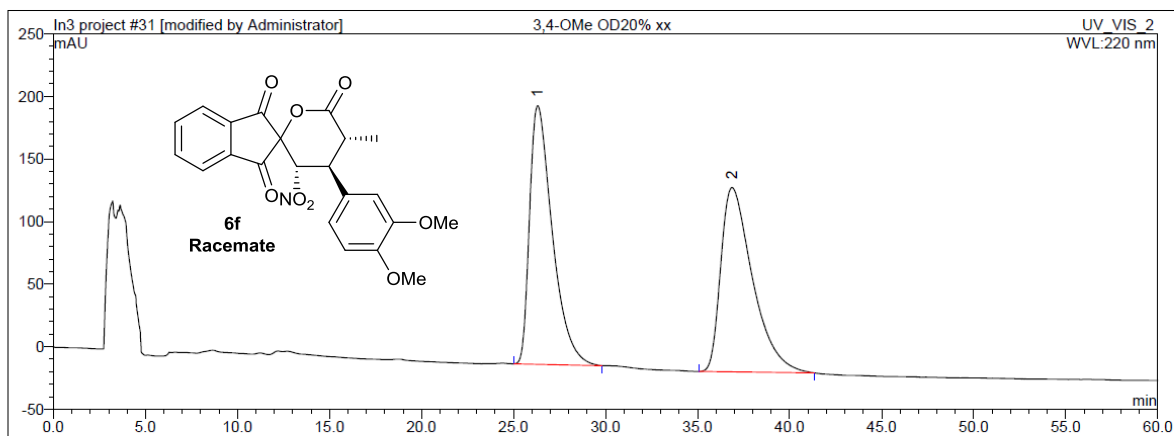
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1	n.a.	15.64	1.939	0.48	2.615	n.a.
2	n.a.	17.68	400.824	99.52	578.678	n.a.





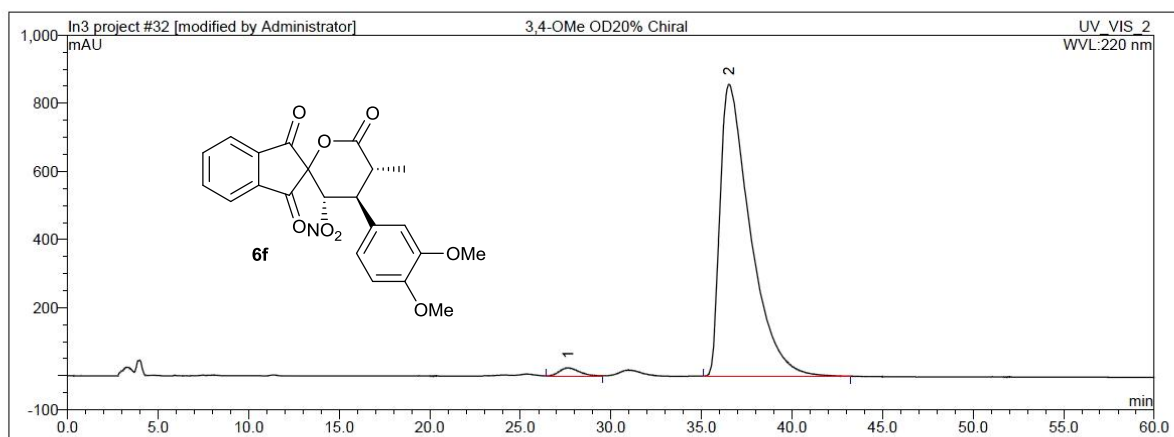
Peak Analysis Report

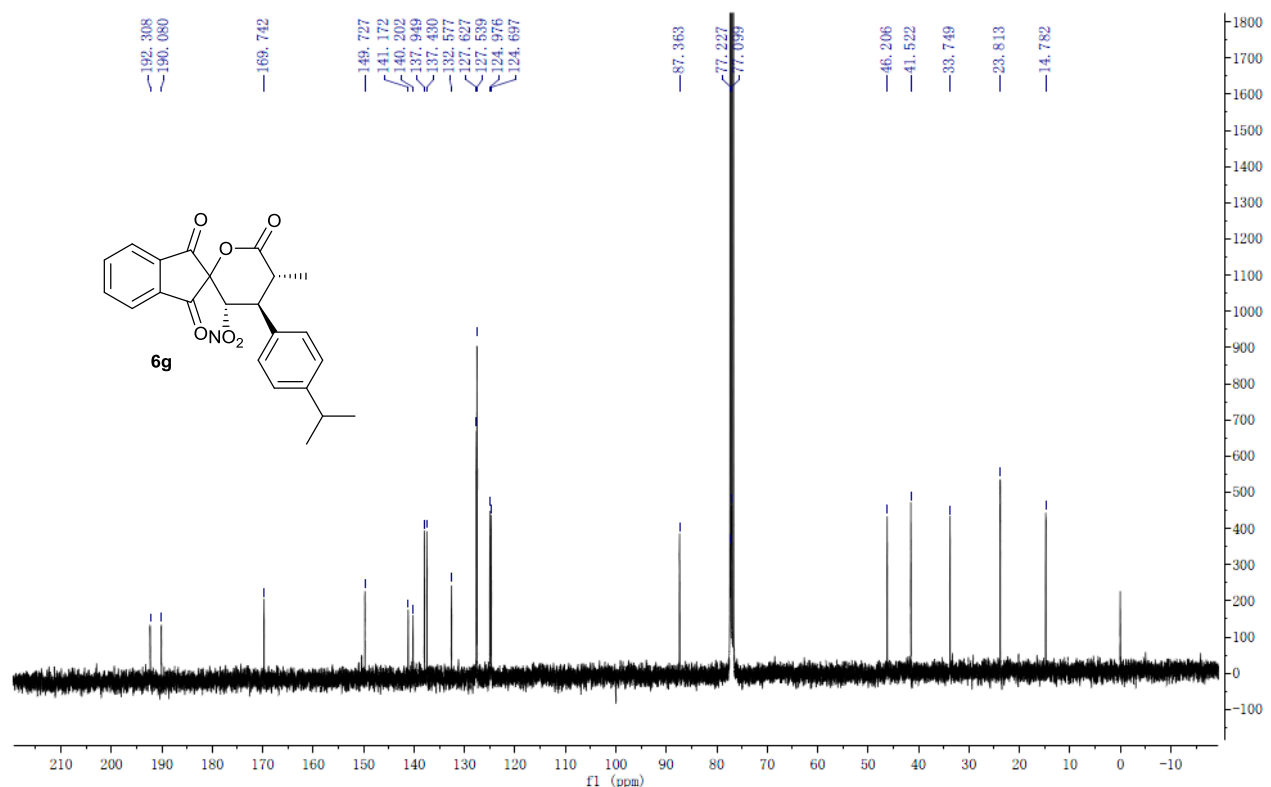
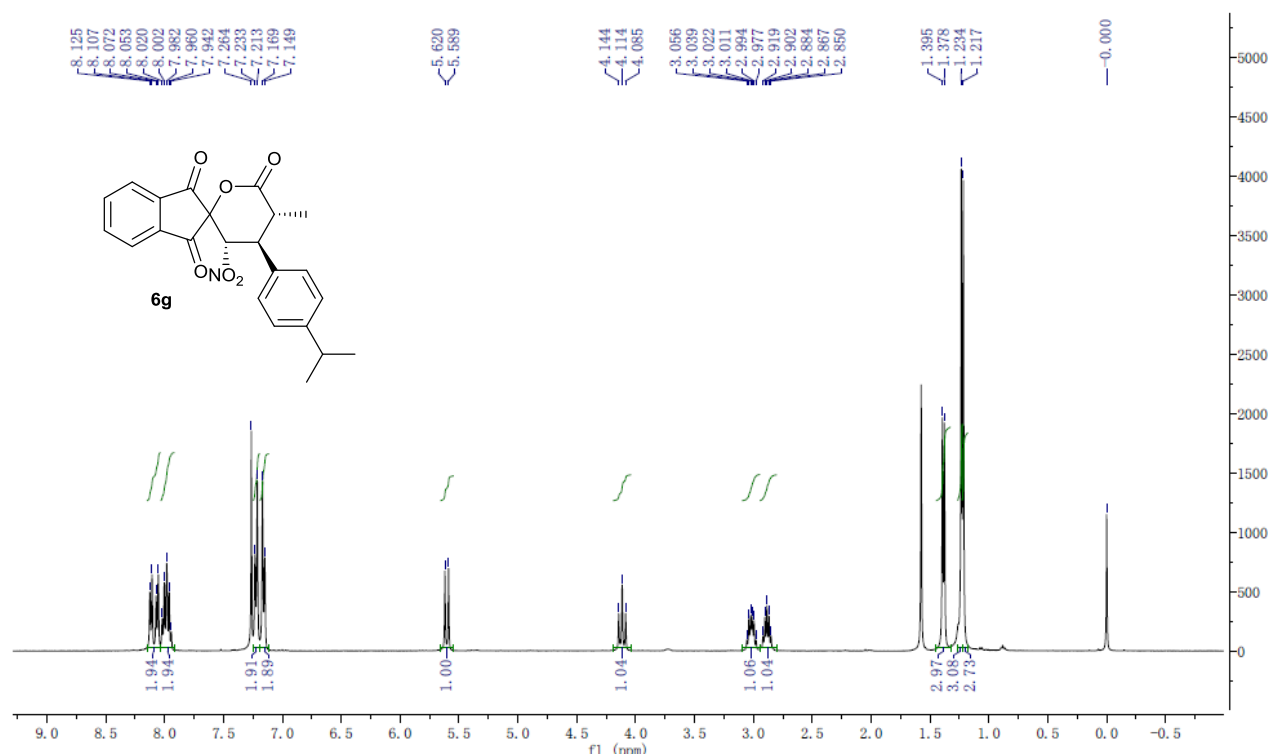
No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
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1	n.a.	26.31	295.694	50.12	206.502	n.a.
2	n.a.	36.88	294.240	49.88	147.219	n.a.



Peak Analysis Report

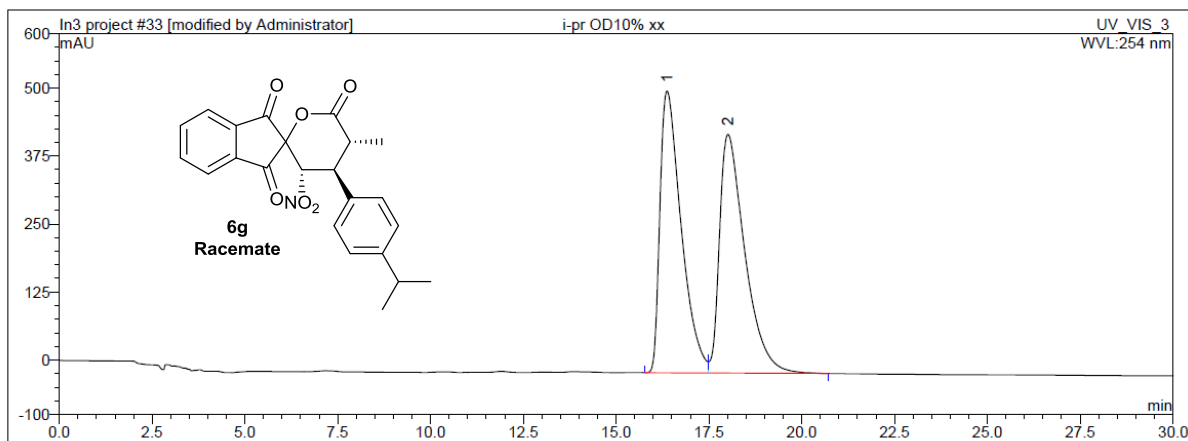
No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	27.63	28.594	1.70	23.184	n.a.
2	n.a.	36.53	1655.628	98.30	856.754	n.a.





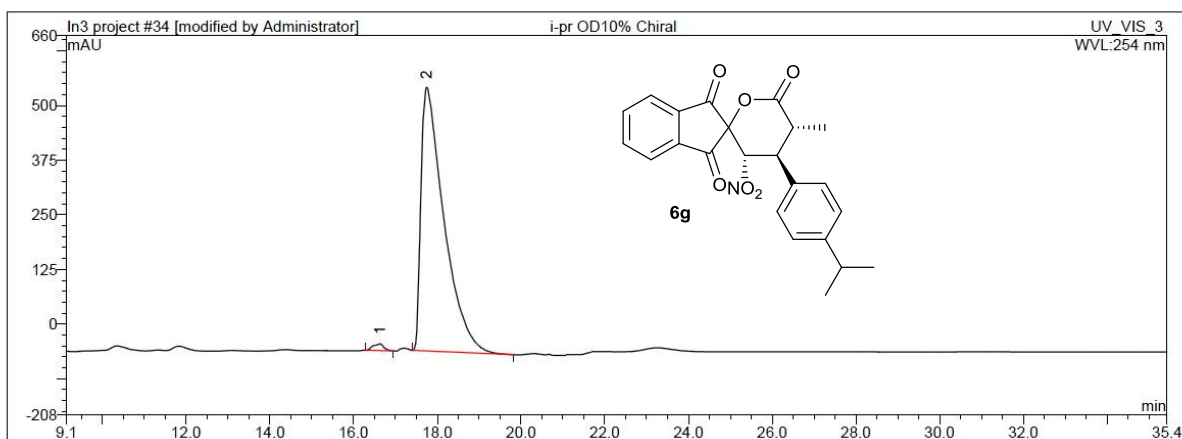
Peak Analysis Report

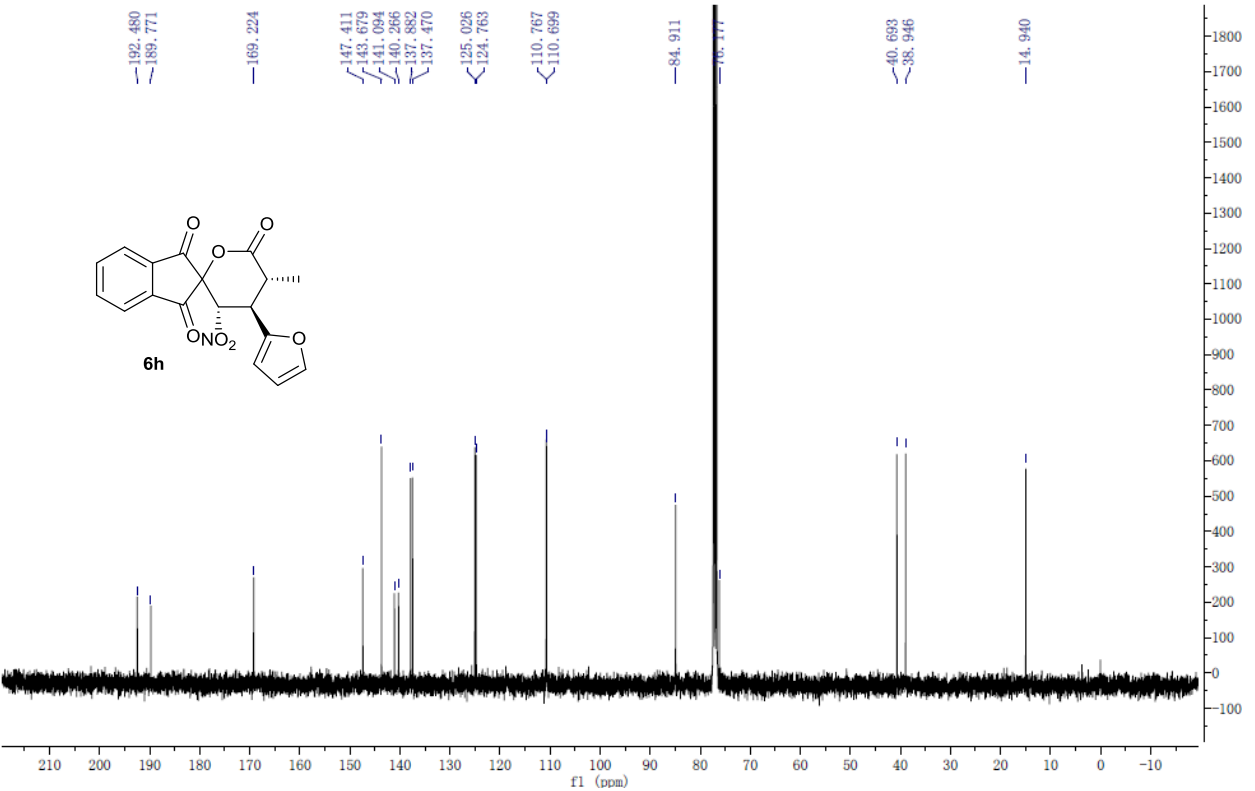
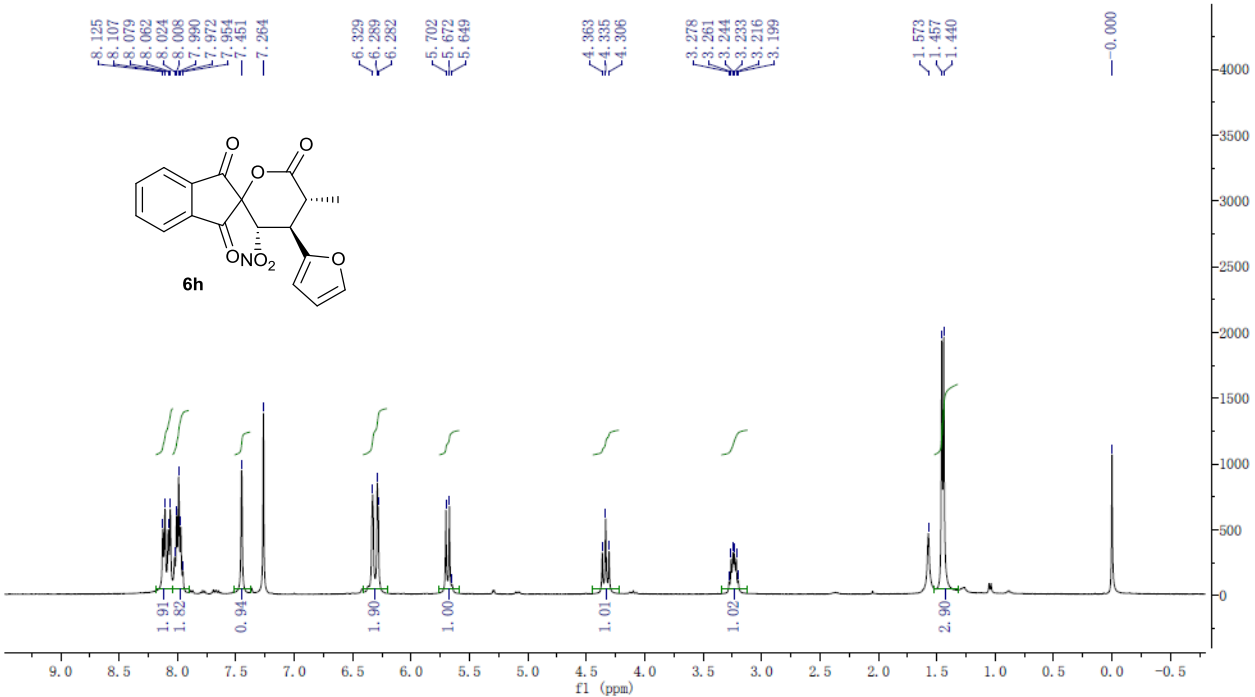
No.	Peak Name	Ret Time (detected)	Area	Rel Area	Height	Amount
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1	n.a.	16.38	342.639	49.63	517.674	n.a.
2	n.a.	18.02	347.684	50.37	438.691	n.a.



Peak Analysis Report

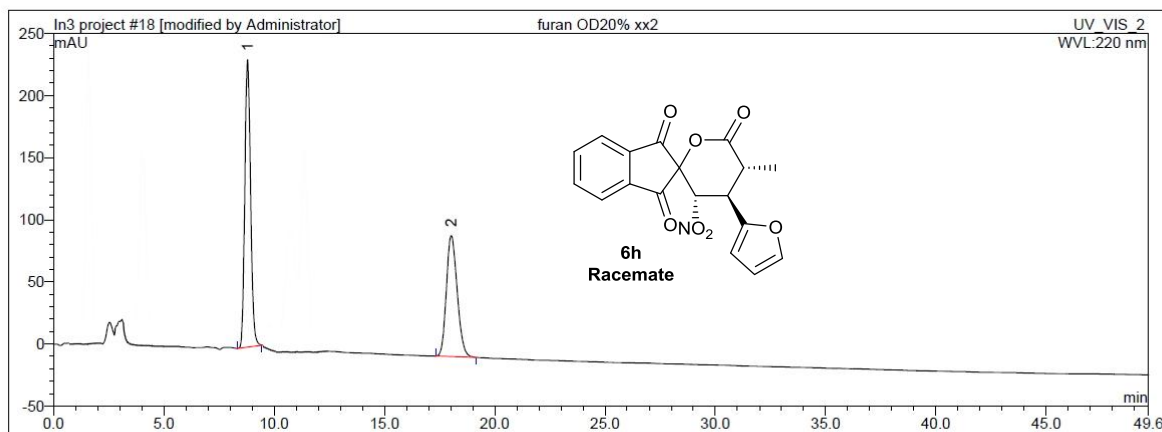
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		min	mAU*min	%	mAU	
1	n.a.	16.32	6.123	1.18	15.217	n.a.
2	n.a.	18.03	512.026	98.82	602.632	n.a.





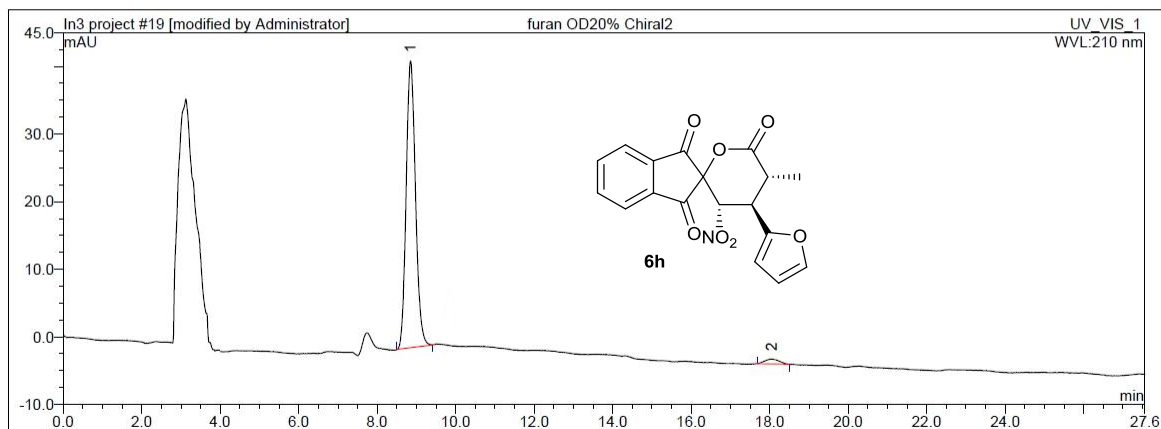
Peak Analysis Report

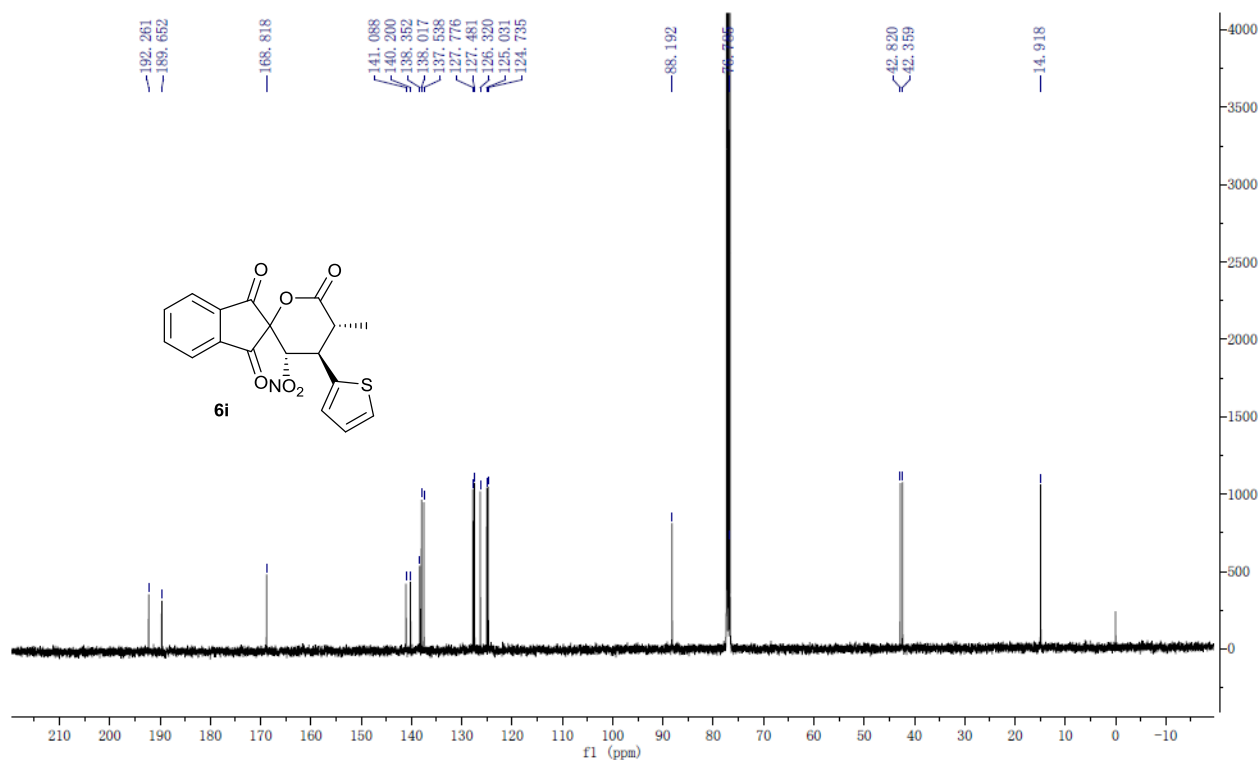
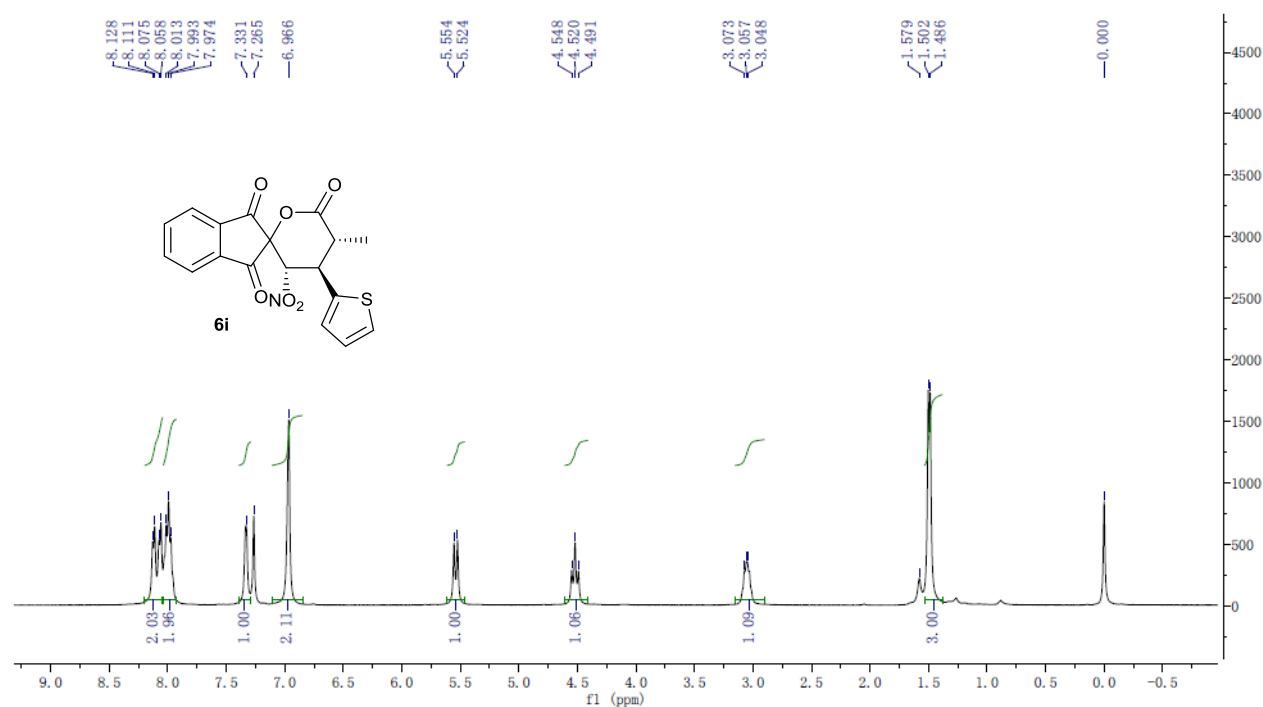
No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	8.79	77.142	50.91	231.274	n.a.
2	n.a.	18.03	74.372	49.09	97.405	n.a.

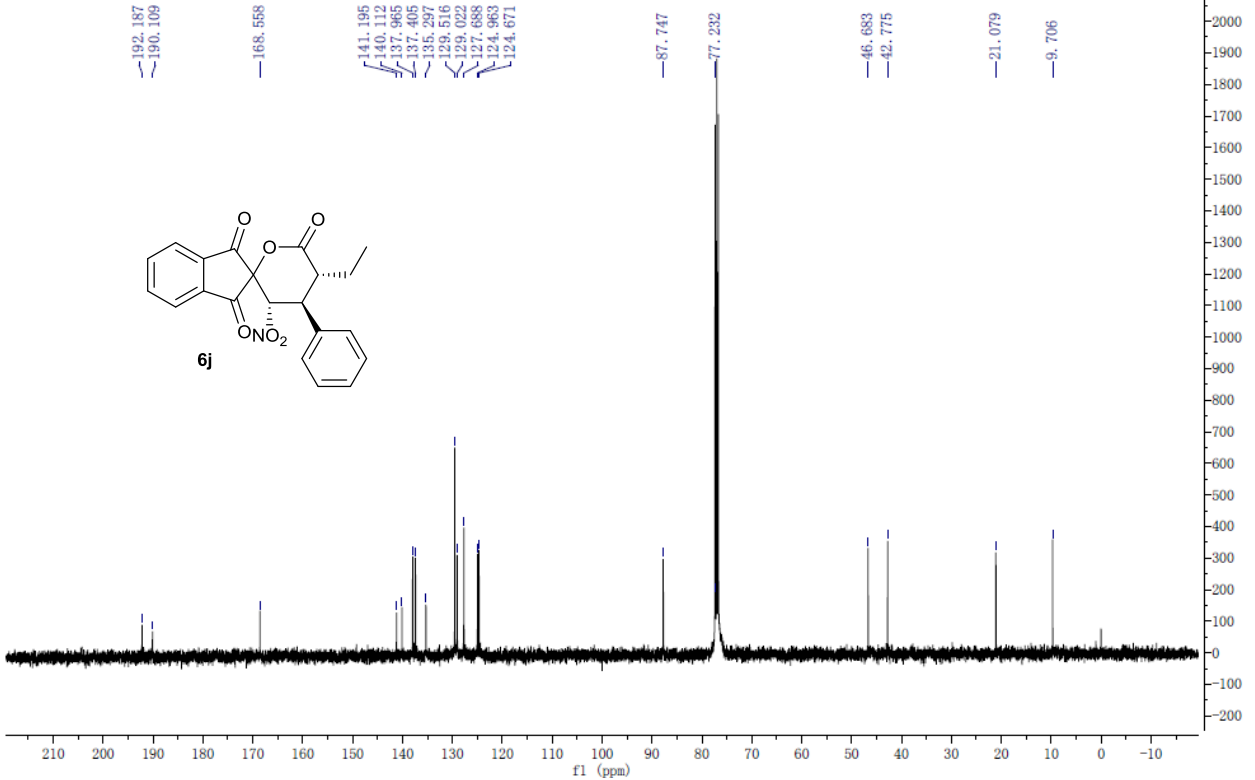
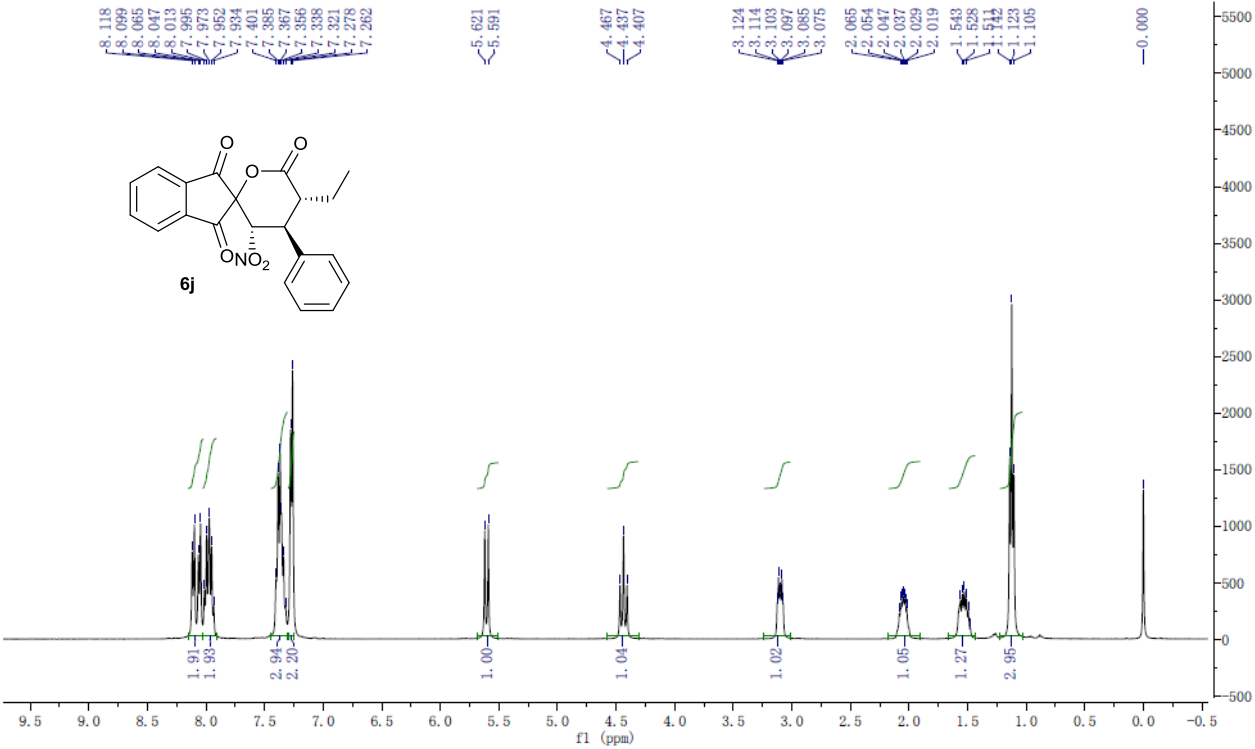


Peak Analysis Report

No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	8.85	11.711	97.50	42.418	n.a.
2	n.a.	18.05	0.301	2.50	0.716	n.a.

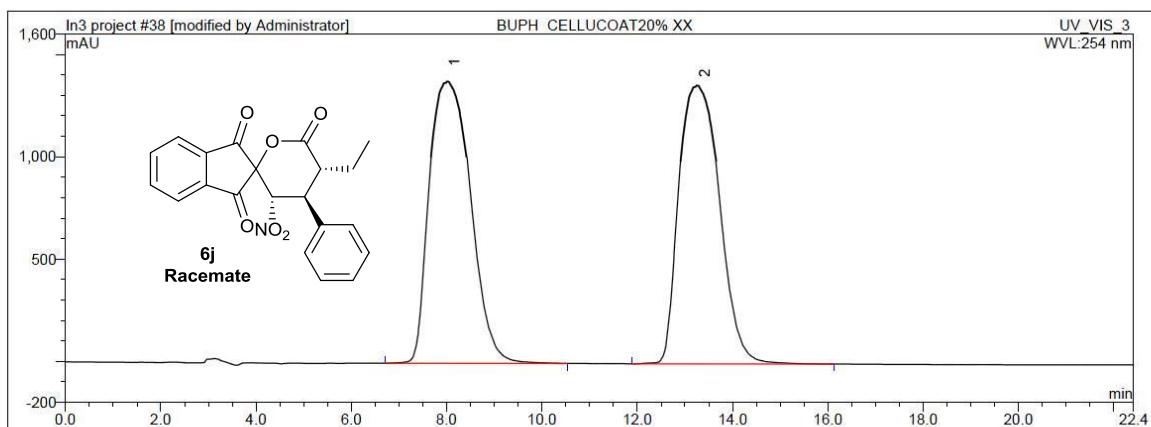






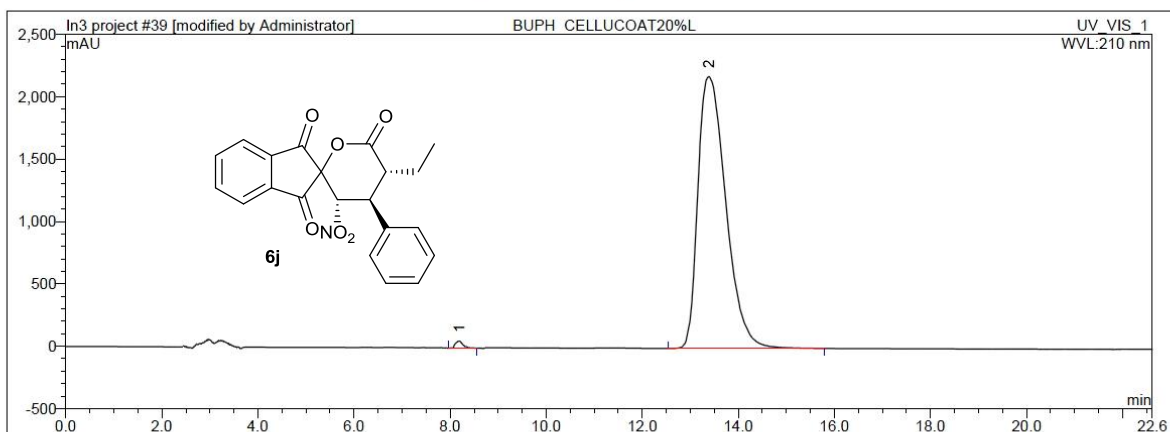
Peak Analysis Report

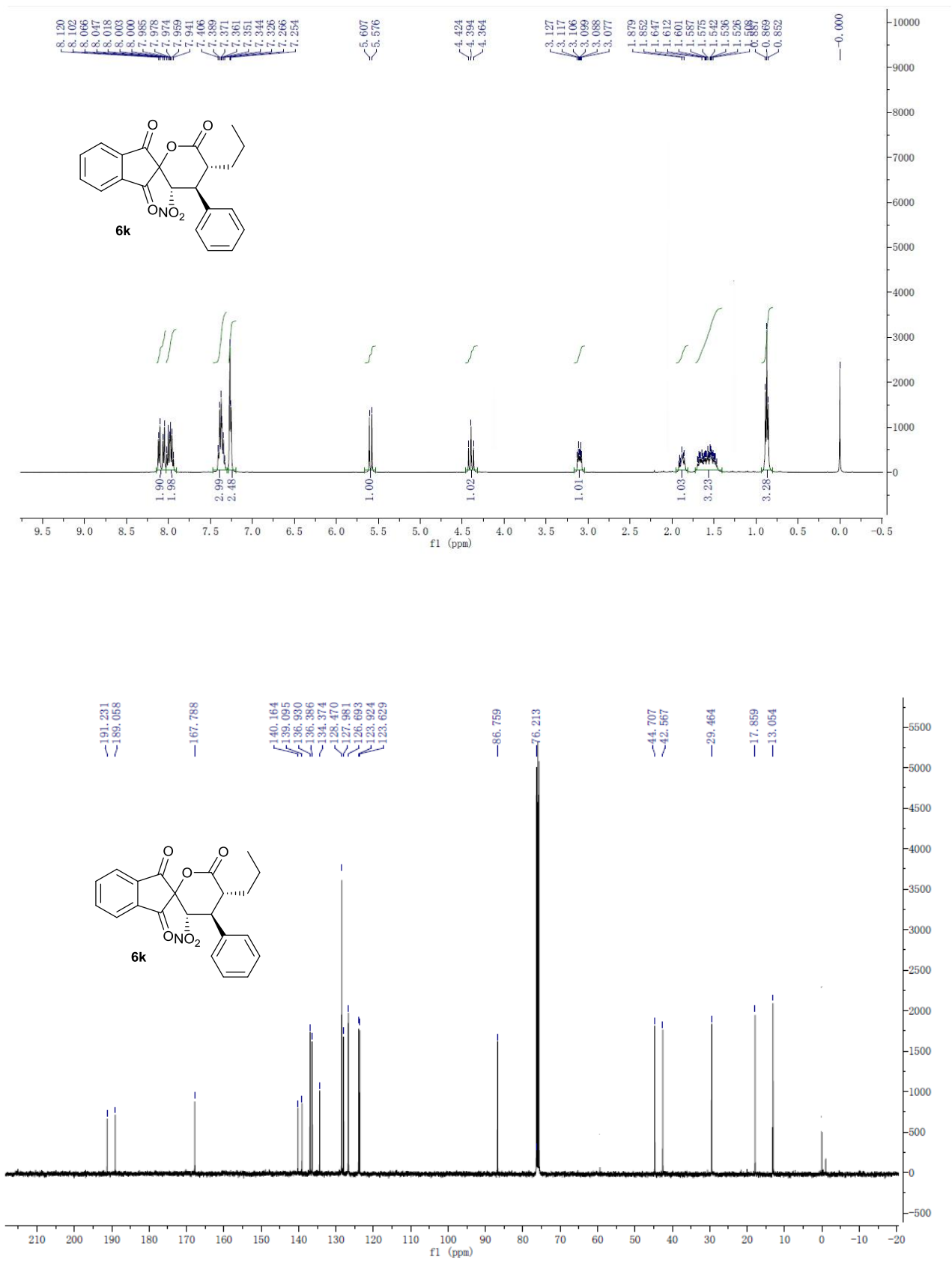
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	8.17	1380.660	51.10	1407.284	n.a.
2	n.a.	13.42	1321.286	48.90	1359.002	n.a.



Peak Analysis Report

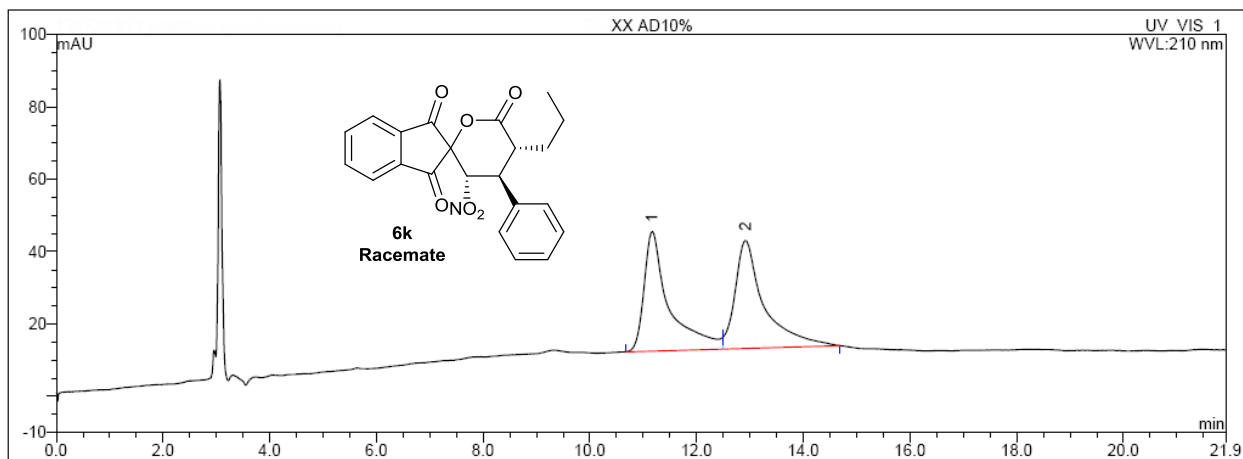
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	8.15	10.085	0.69	55.658	n.a.
2	n.a.	13.39	1462.010	99.31	2176.358	n.a.





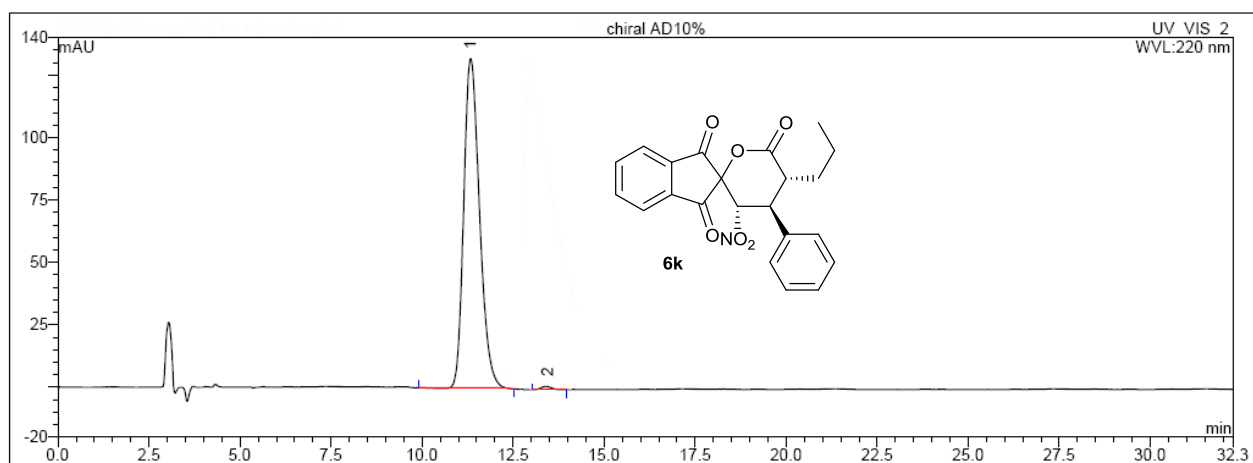
Peak Analysis Report

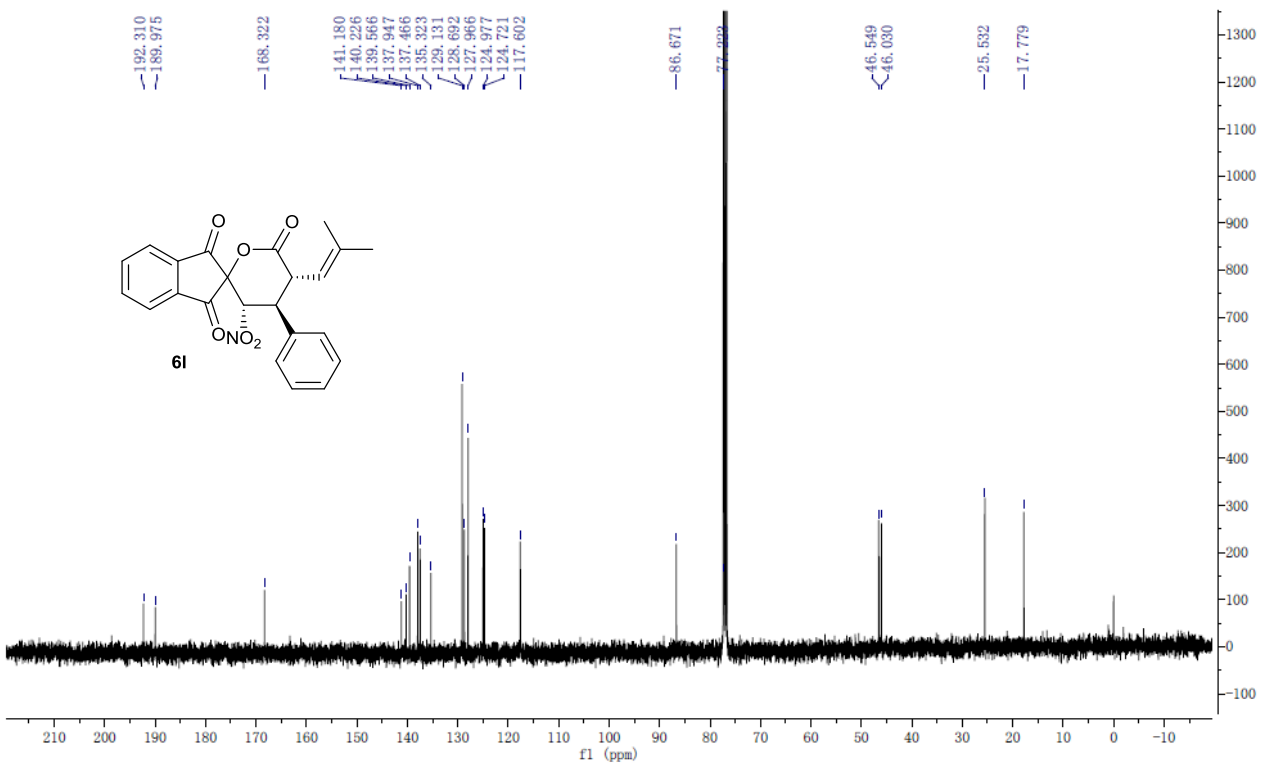
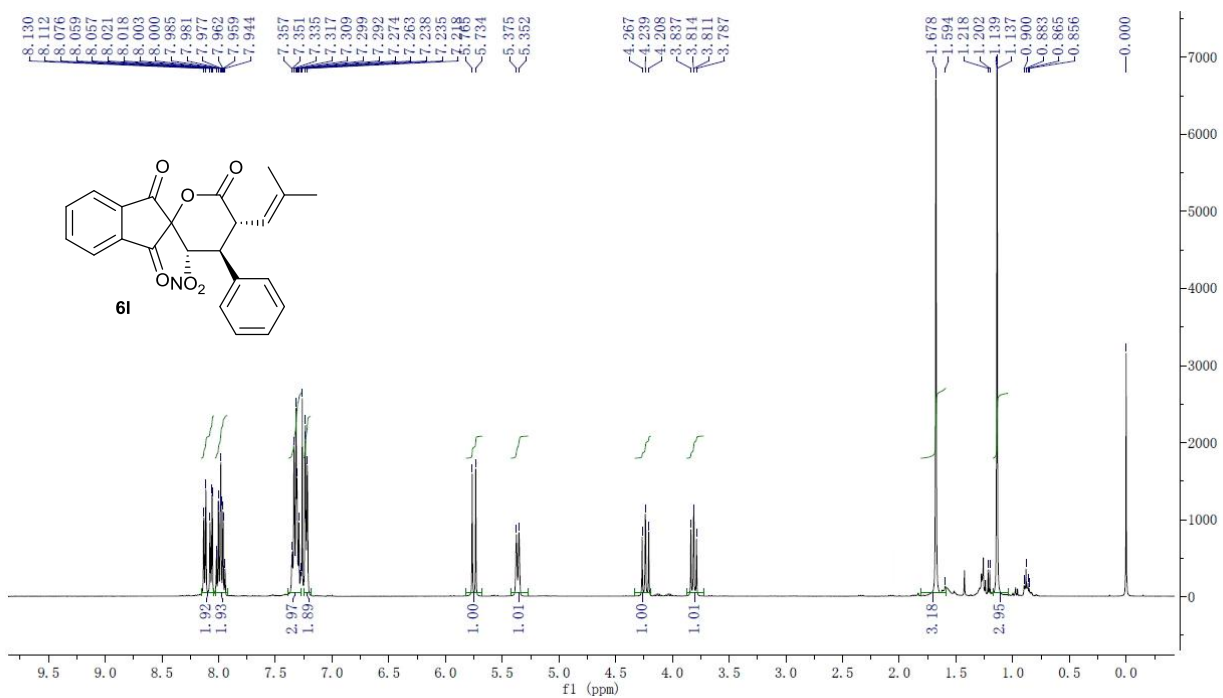
No.	Peak Name	Ret.Time (detected)	Area	Rel.Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	11.17	18.054	49.04	33.008	n.a.
2	n.a.	12.92	18.759	50.96	29.807	n.a.



Peak Analysis Report

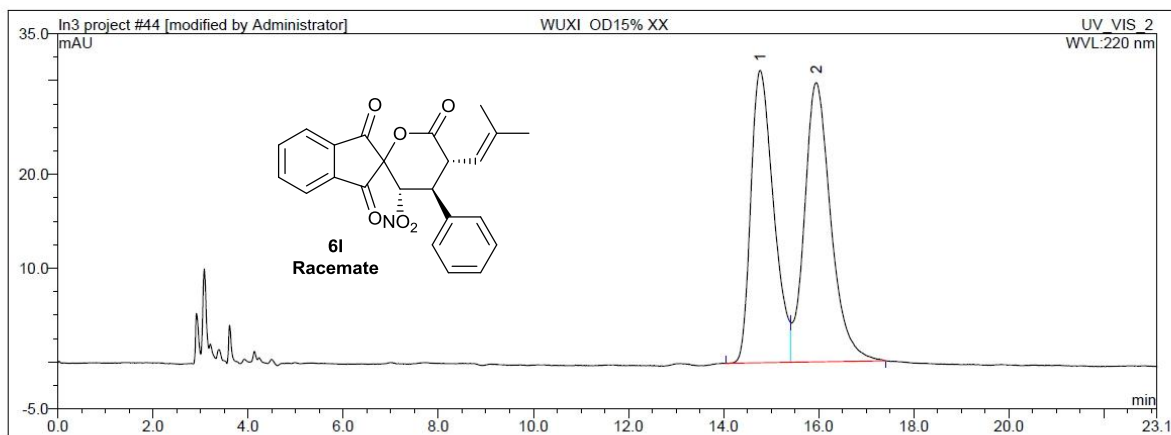
No.	Peak Name	Ret.Time (detected)	Area	Rel.Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	11.22	76.891	99.37	124.374	n.a.
2	n.a.	12.98	0.485	0.63	1.403	n.a.





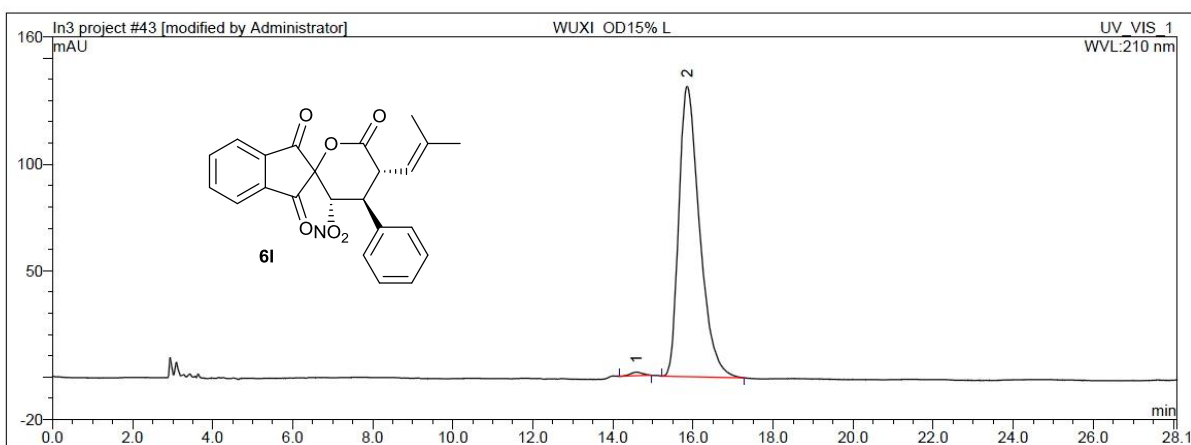
Peak Analysis Report

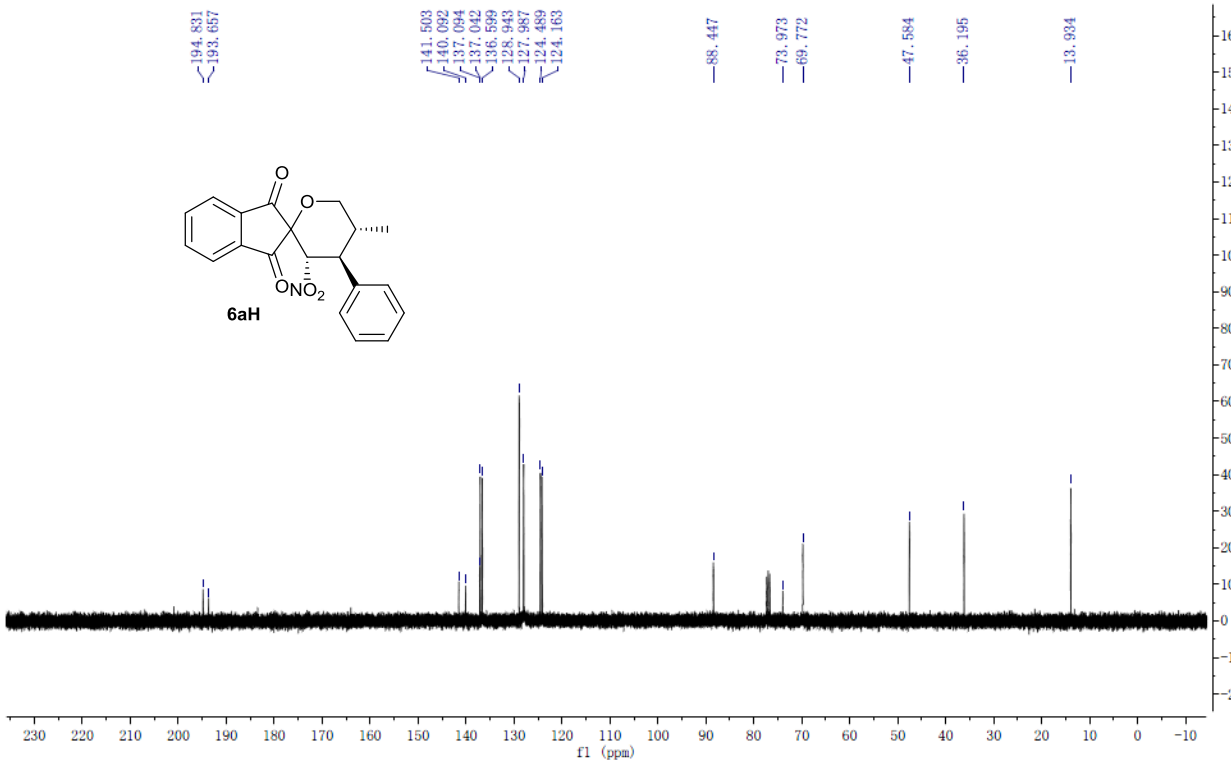
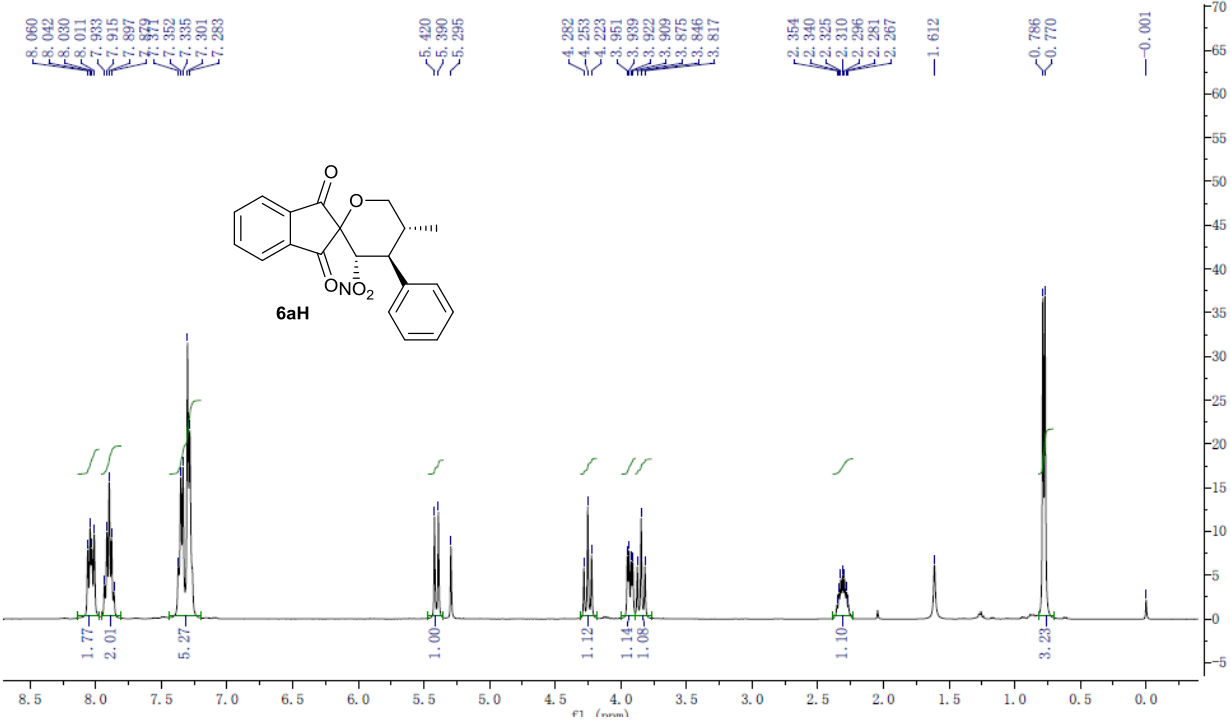
No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	14.78	16.762	49.06	30.809	n.a.
2	n.a.	15.95	17.403	50.94	28.831	n.a.



Peak Analysis Report

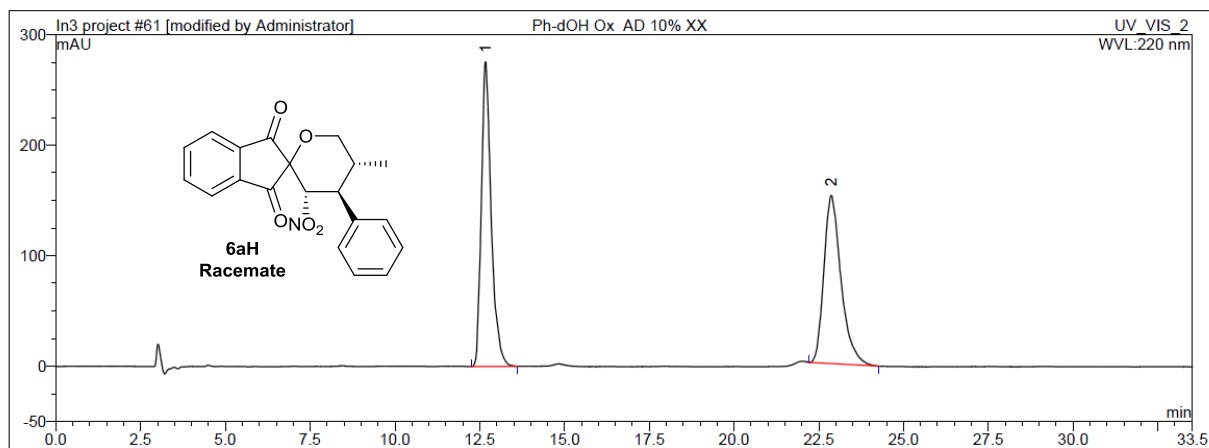
No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	14.76	0.570	0.70	1.620	n.a.
2	n.a.	15.86	81.387	99.30	136.373	n.a.





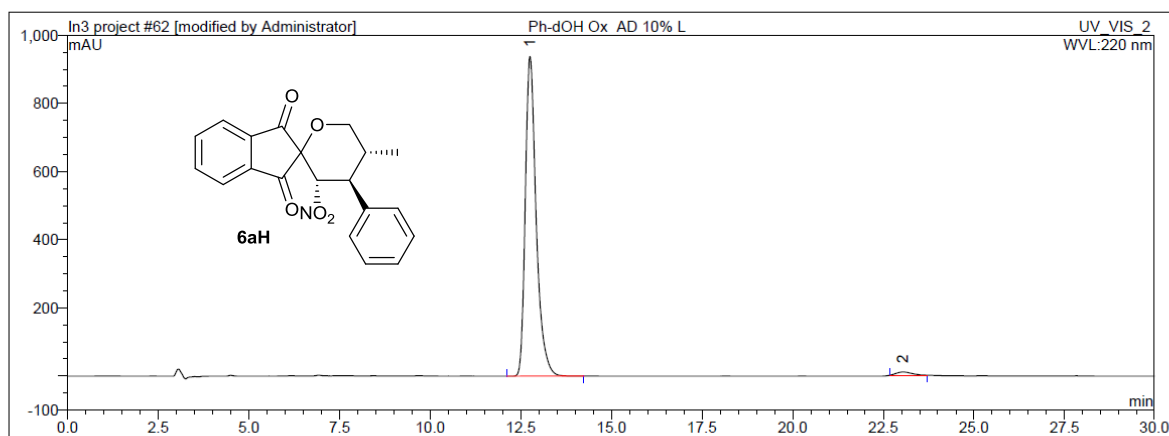
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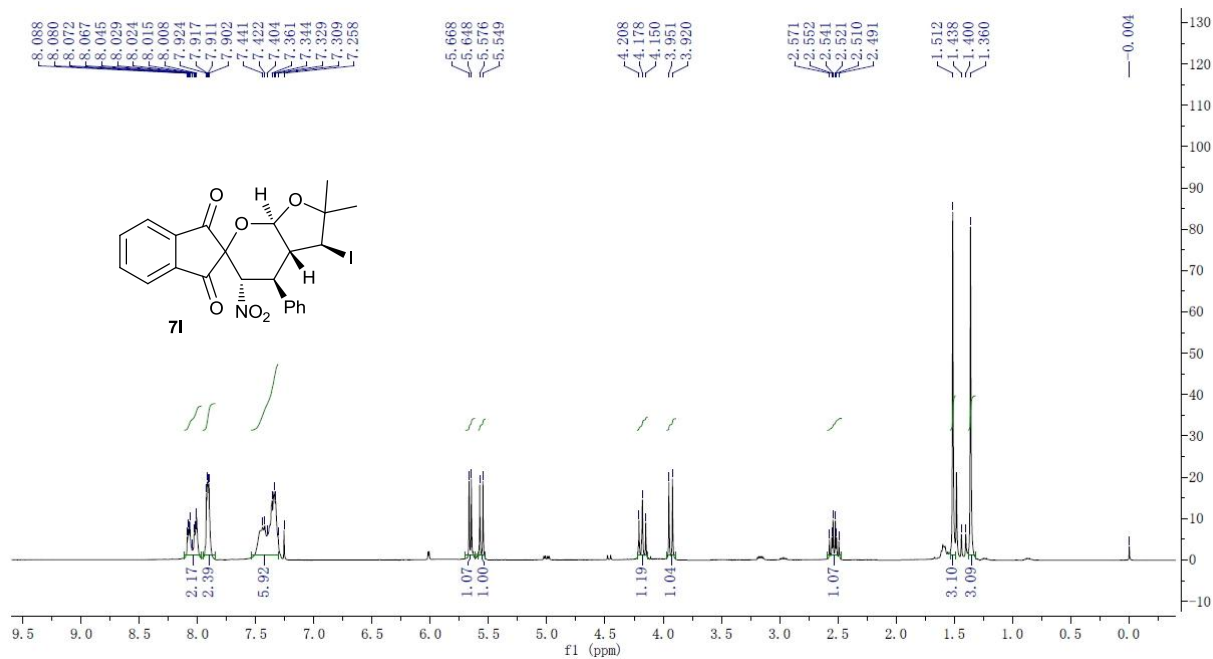
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	12.68	92.147	50.87	275.469	n.a.
2	n.a.	22.87	89.002	49.13	151.869	n.a.



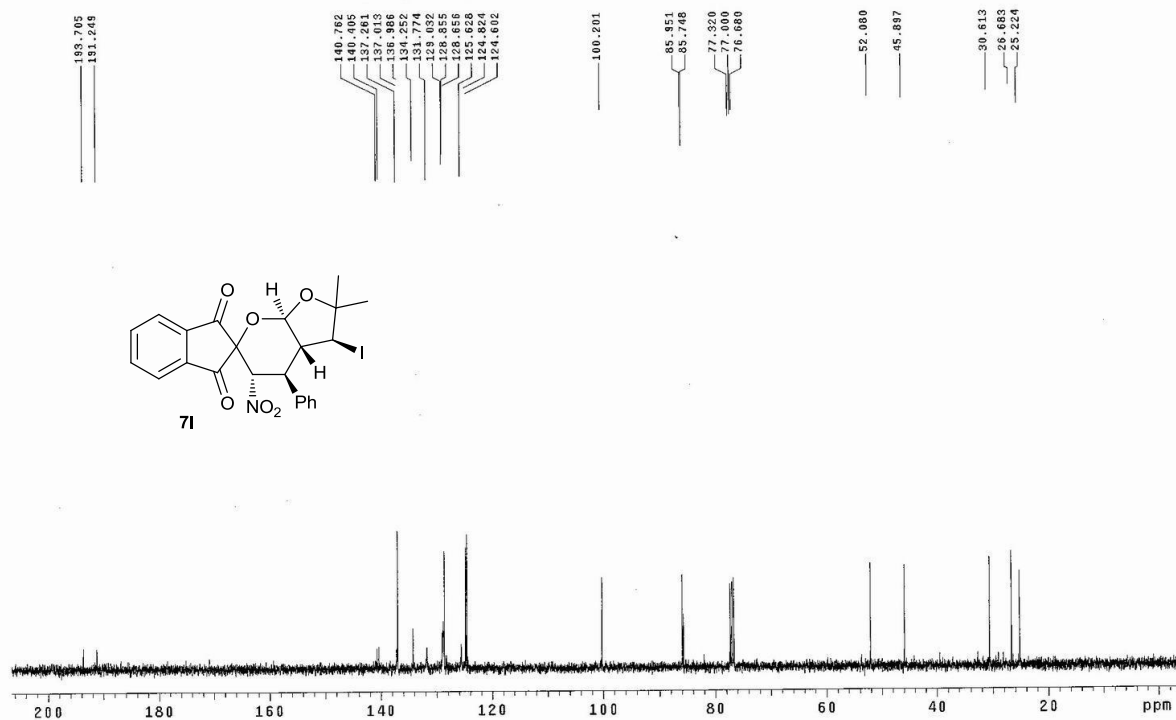
Peak Analysis Report

No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	12.75	326.697	98.55	938.310	n.a.
2	n.a.	23.03	4.814	1.45	9.684	n.a.



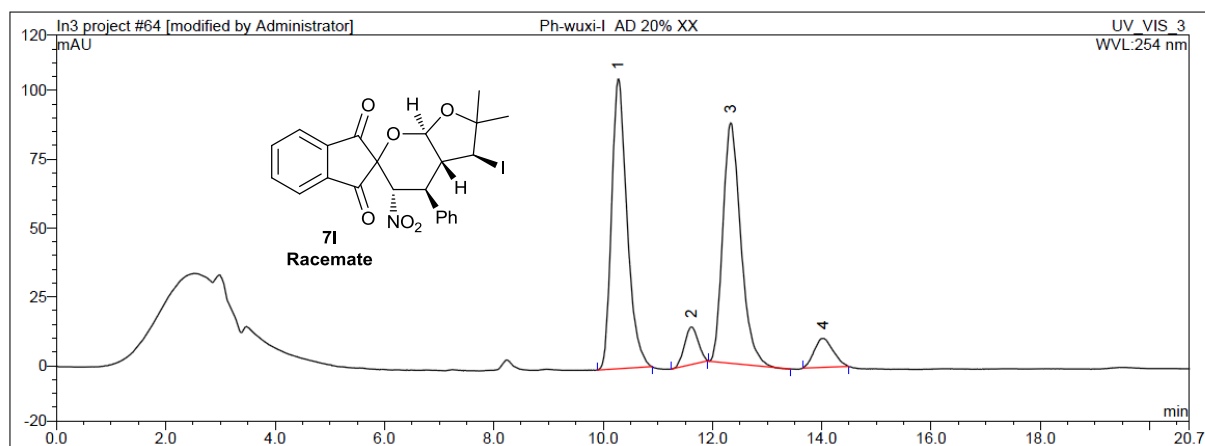


YL-0321-ph-CDCl₃-C13-2013-3-30
Pulse Sequence: s2pu1



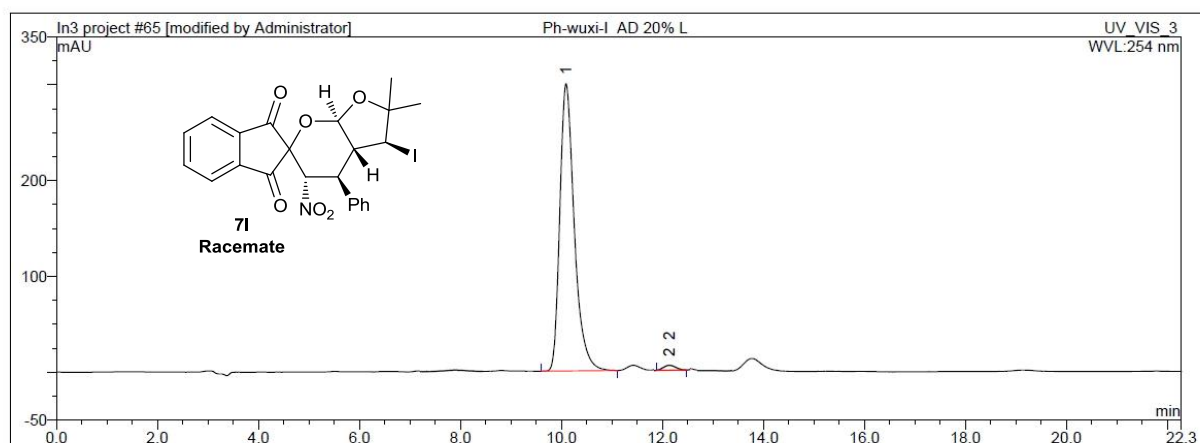
Peak Analysis Report

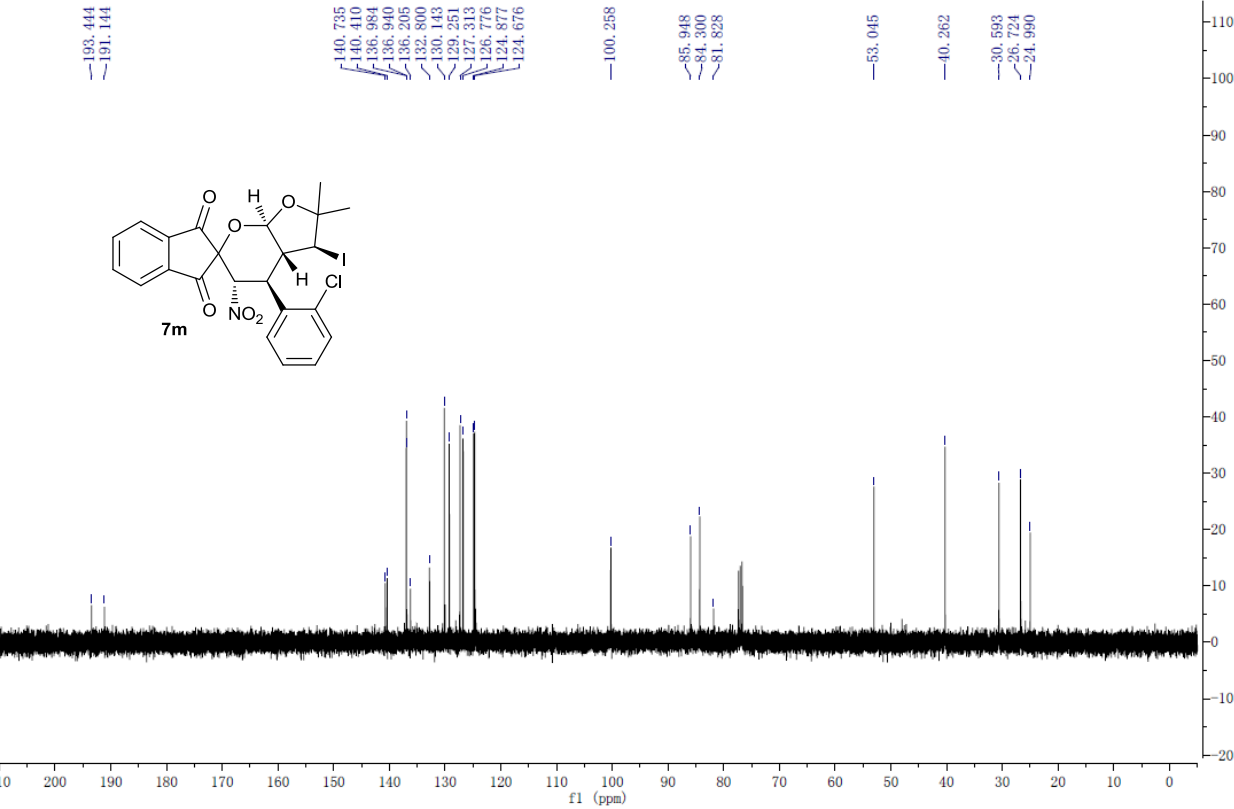
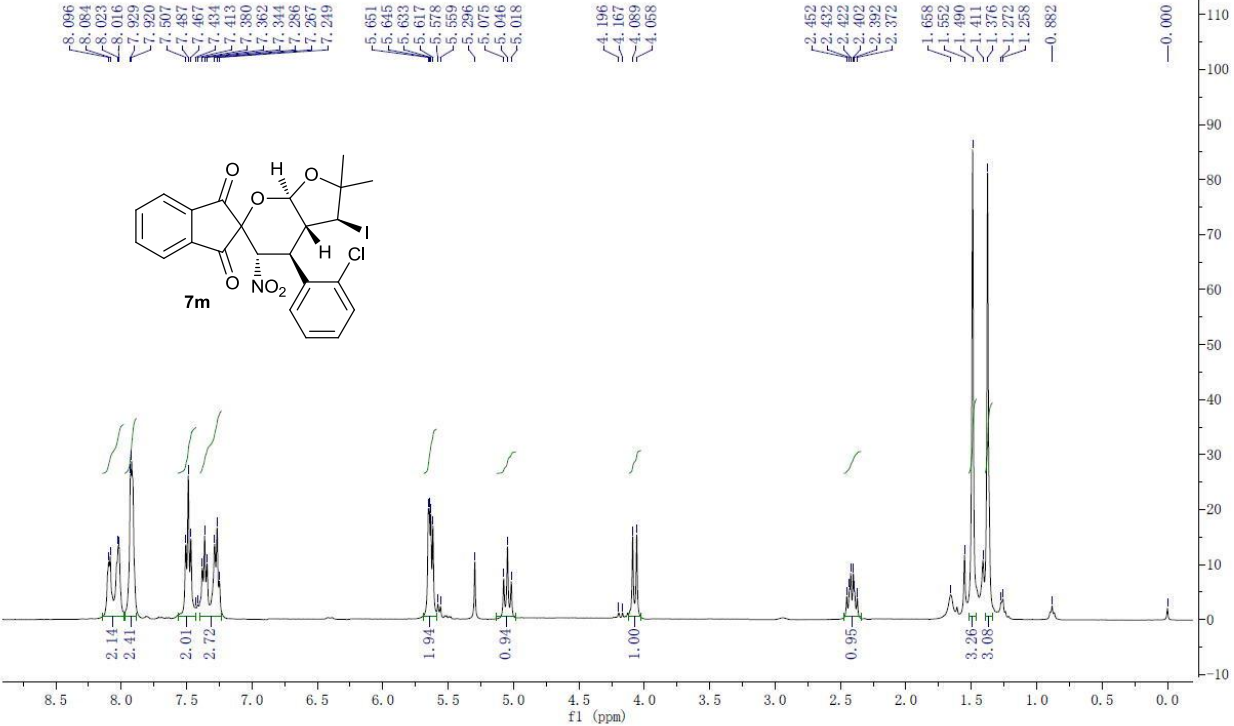
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	10.28	34.496	45.46	105.150	n.a.
2	n.a.	11.62	3.930	5.18	13.560	n.a.
3	n.a.	12.33	33.259	43.83	87.219	n.a.
4	n.a.	14.03	4.202	5.54	10.503	n.a.



Peak Analysis Report

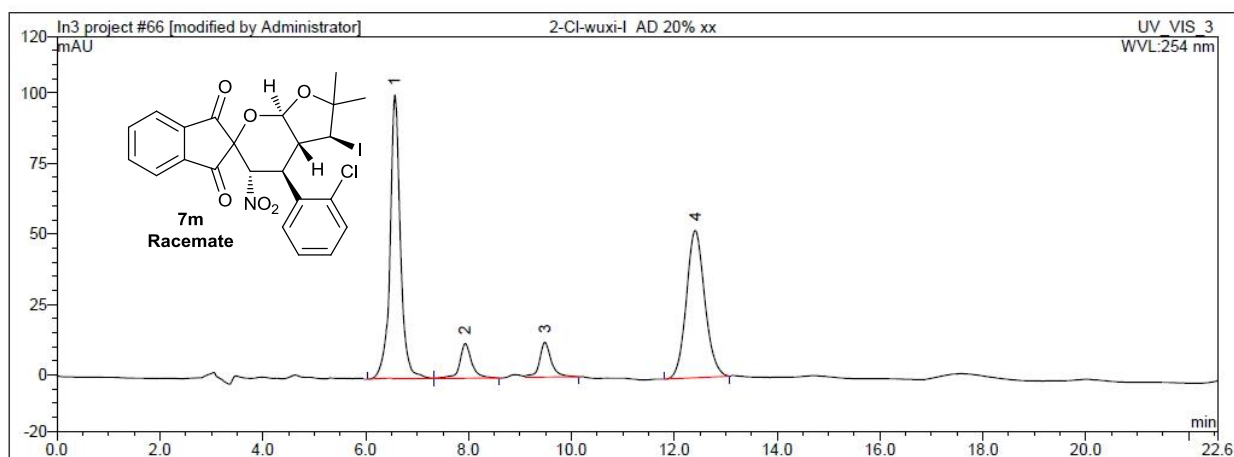
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	10.09	99.440	98.59	299.955	n.a.
2	n.a.	12.14	1.425	1.41	5.103	n.a.





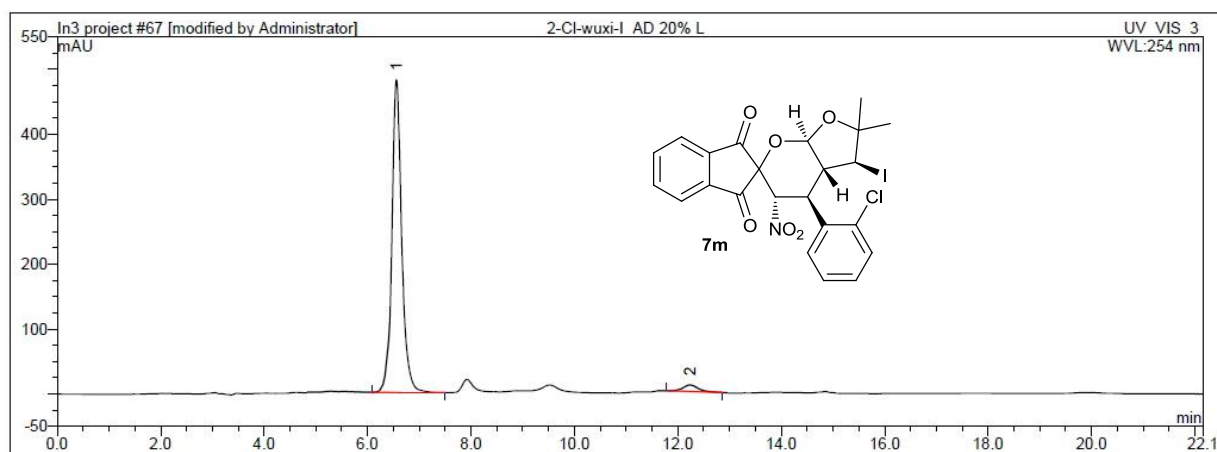
Peak Analysis Report

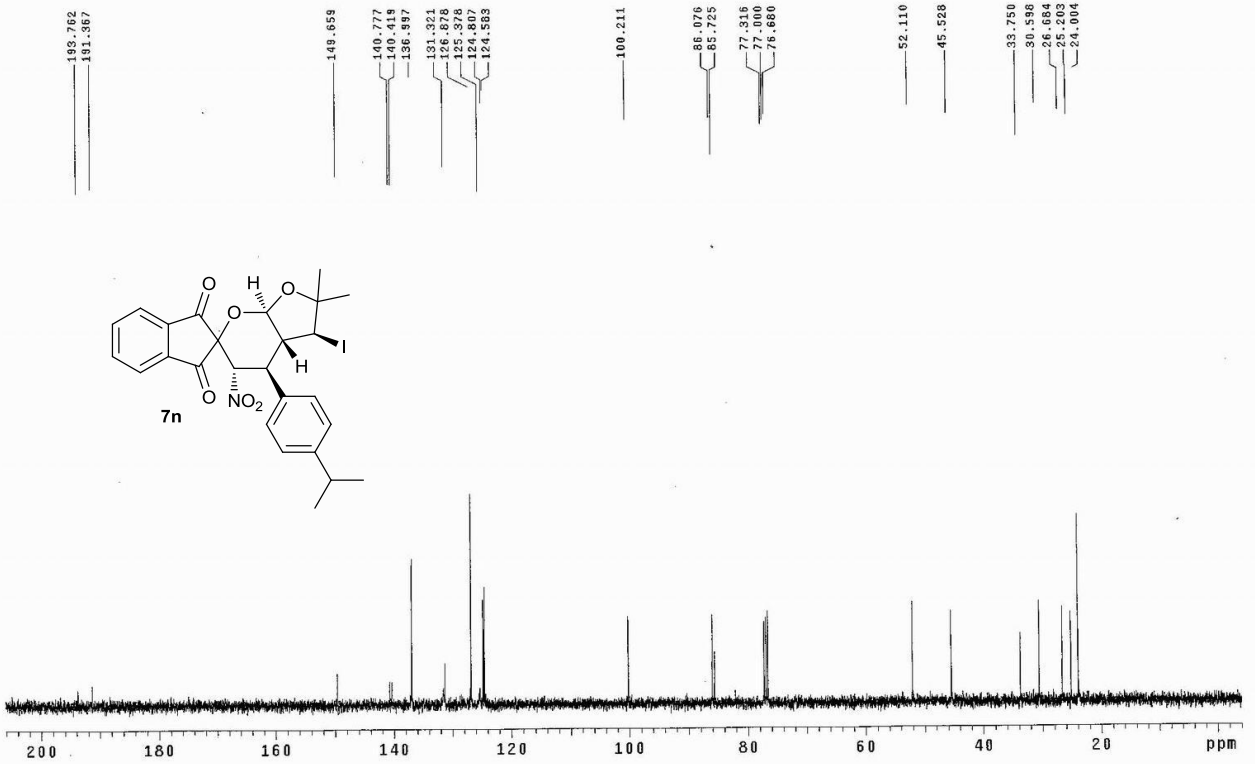
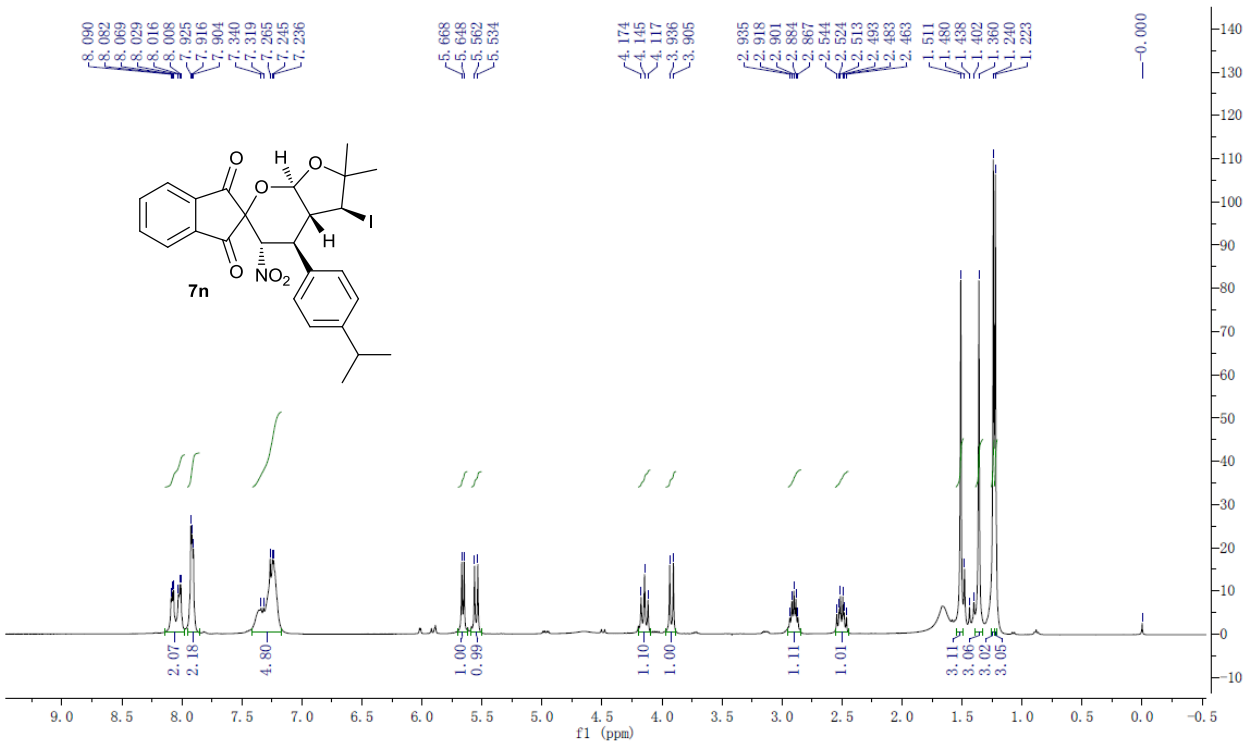
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	6.57	23.919	43.89	100.246	n.a.
2	n.a.	7.93	3.326	6.10	11.981	n.a.
3	n.a.	9.51	3.285	6.03	11.763	n.a.
4	n.a.	12.41	23.971	43.98	51.902	n.a.



Peak Analysis Report

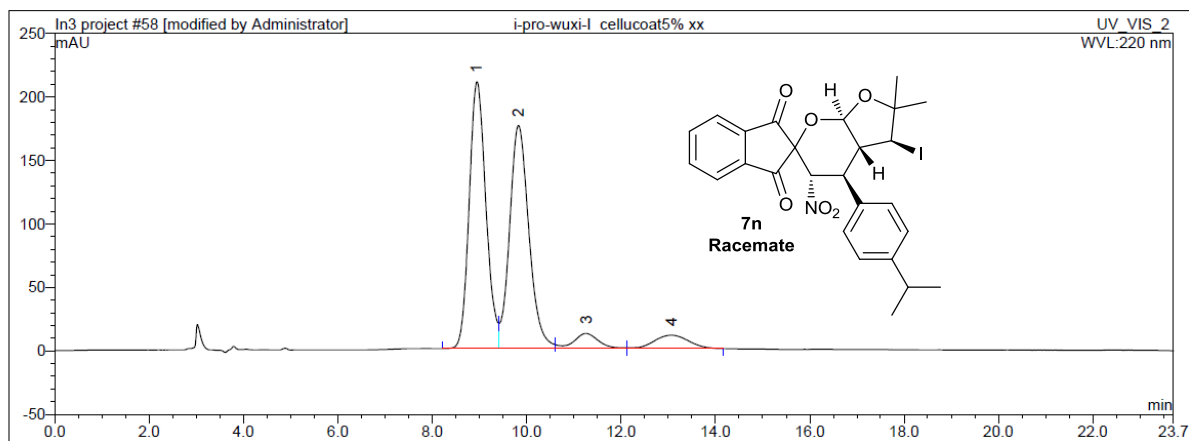
No.	Peak Name	Ret. Time (detected) min	Area mAU*min	Rel. Area %	Height mAU	Amount
1	n.a.	6.56	108.420	96.92	482.328	n.a.
2	n.a.	12.24	3.450	3.08	10.118	n.a.





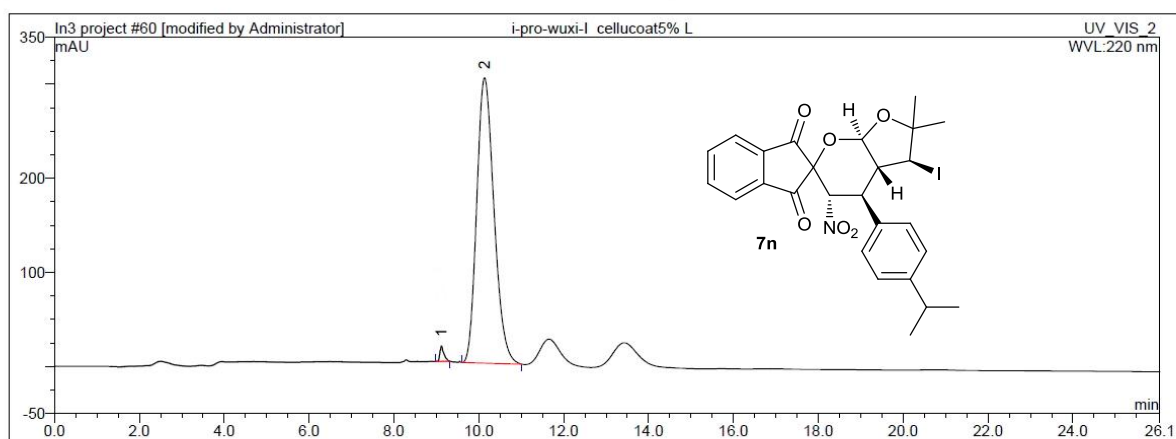
Peak Analysis Report

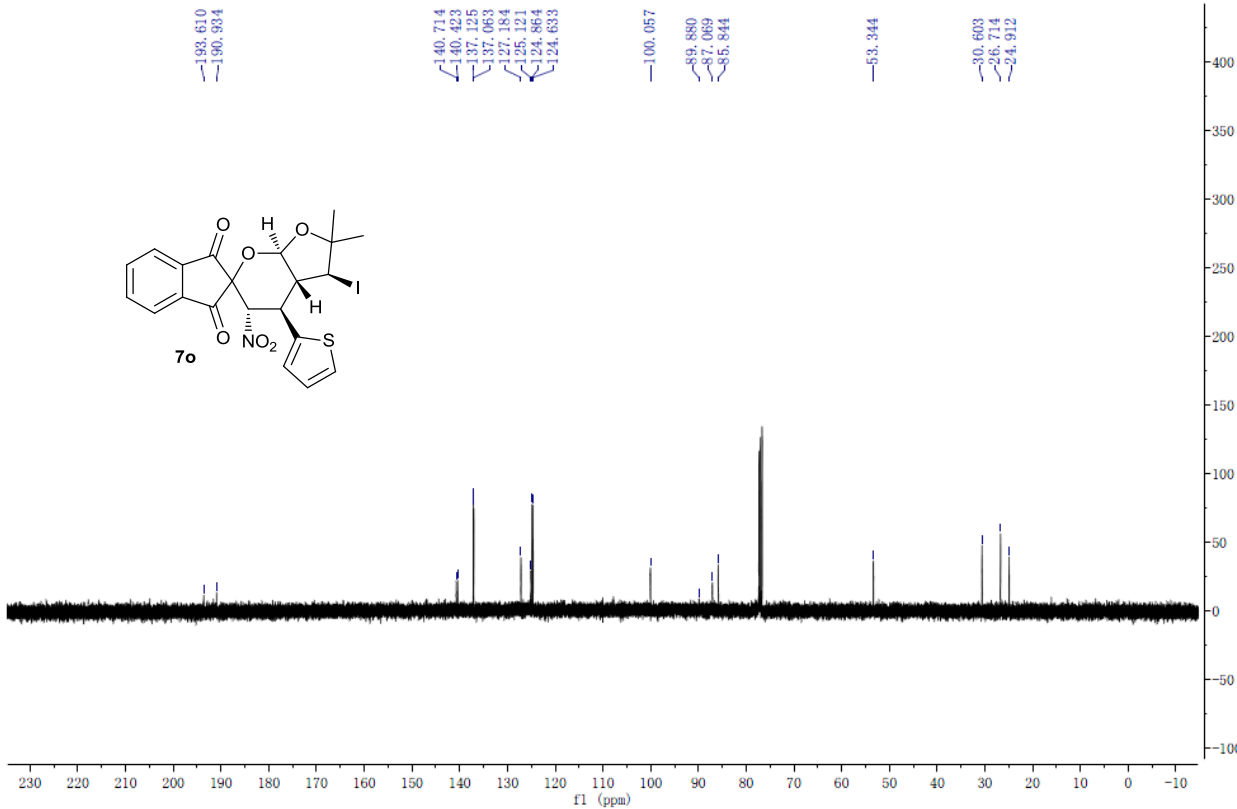
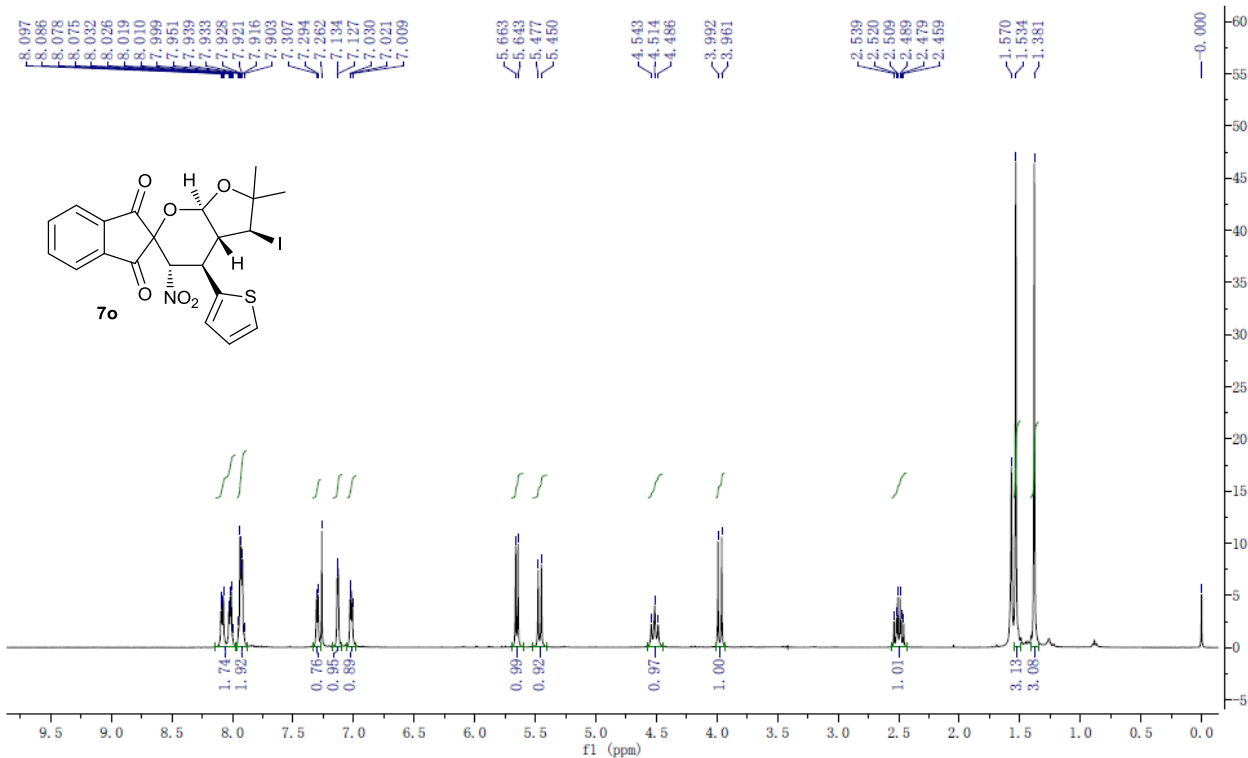
No.	Peak Name	Ret.Time (detected)	Area	Rel.Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	8.94	86.329	46.41	210.160	n.a.
2	n.a.	9.83	83.154	44.70	175.807	n.a.
3	n.a.	11.27	7.444	4.00	11.983	n.a.
4	n.a.	13.07	9.092	4.89	10.634	n.a.



Peak Analysis Report

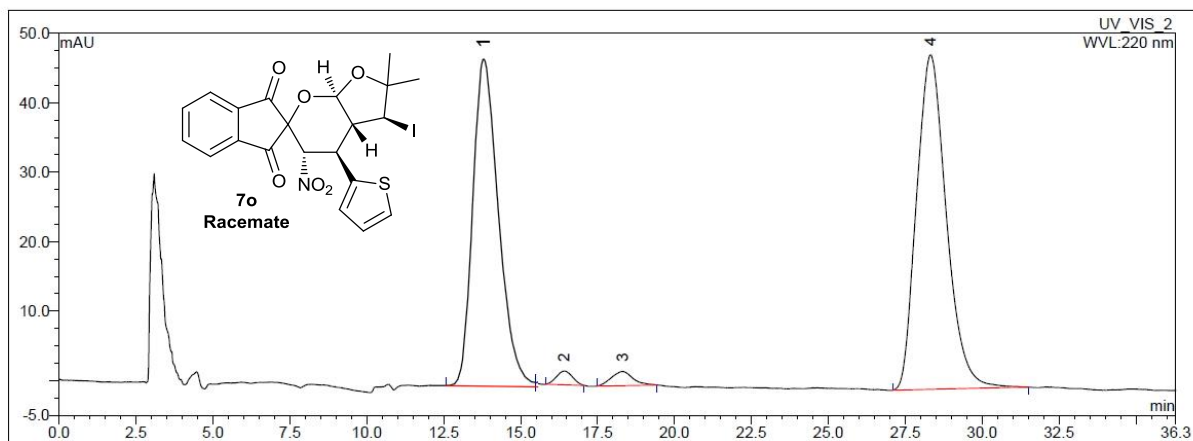
No.	Peak Name	Ret.Time (detected)	Area	Rel.Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	9.09	1.761	1.22	16.901	n.a.
2	n.a.	10.13	142.786	98.78	303.280	n.a.





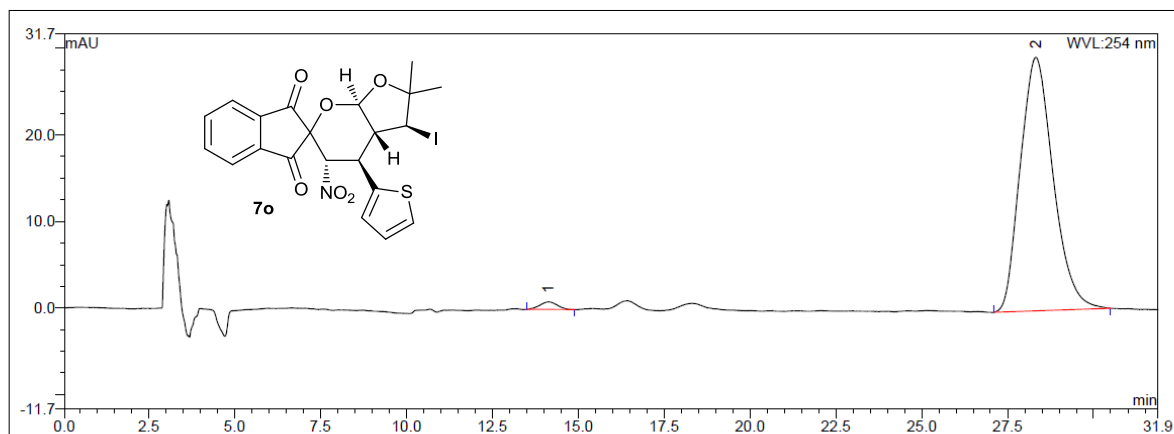
Peak Analysis Report

No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	14.13	13.453	46.07	46.062	n.a.
2	n.a.	16.41	1.187	4.06	1.972	n.a.
3	n.a.	18.32	1.206	4.13	1.787	n.a.
4	n.a.	28.32	13.353	45.73	48.365	n.a.



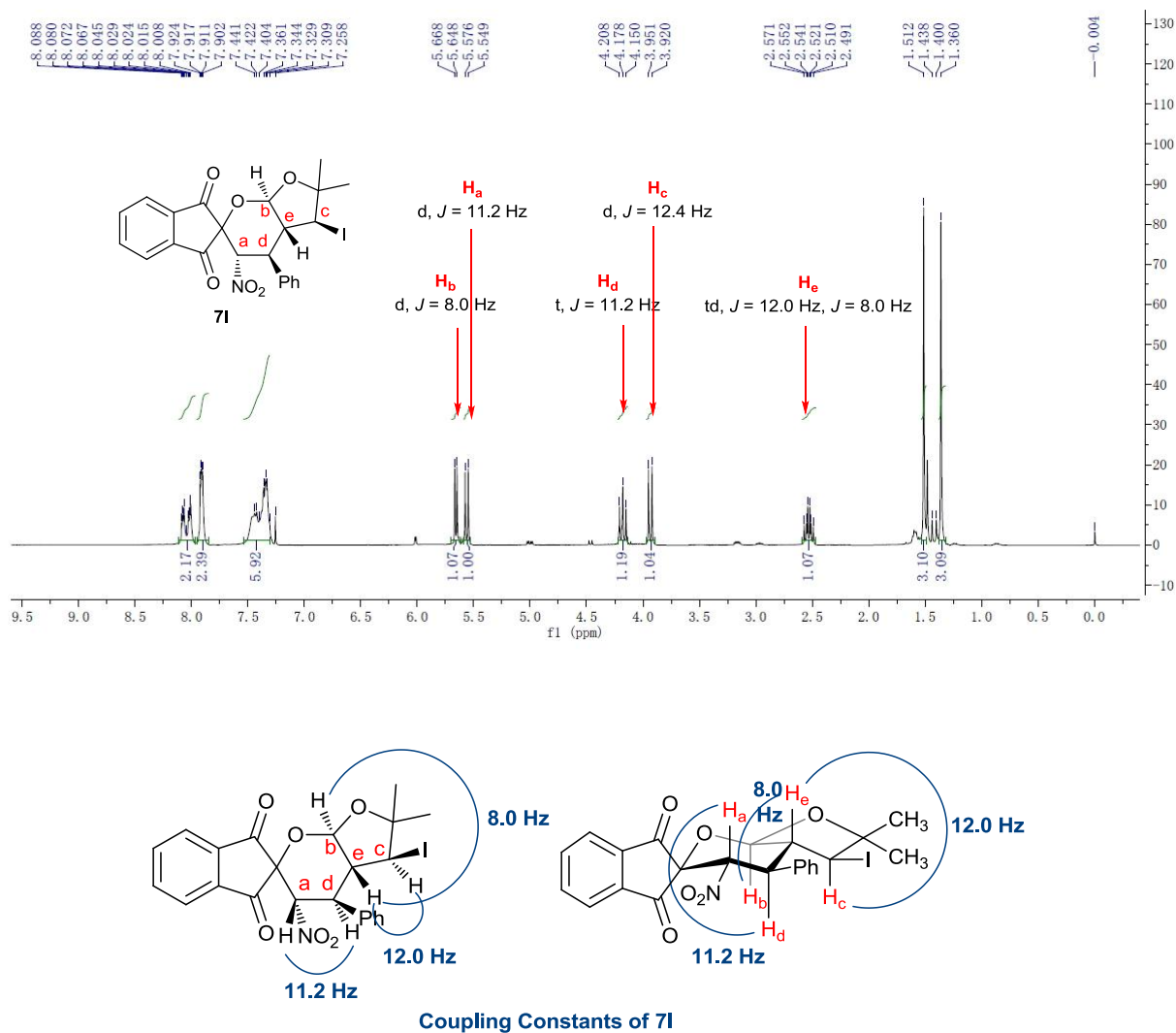
Peak Analysis Report

No.	Peak Name	Ret. Time (detected)	Area	Rel. Area	Height	Amount
		min	mAU*min	%	mAU	
1	n.a.	14.13	0.514	1.56	0.894	n.a.
2	n.a.	28.33	32.543	98.44	29.291	n.a.



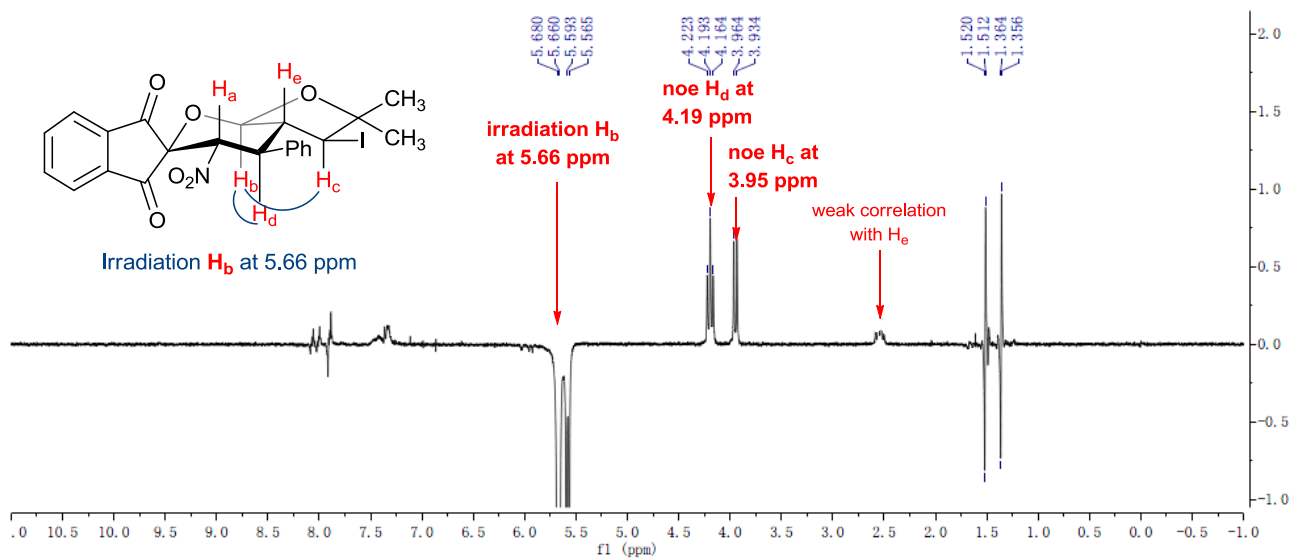
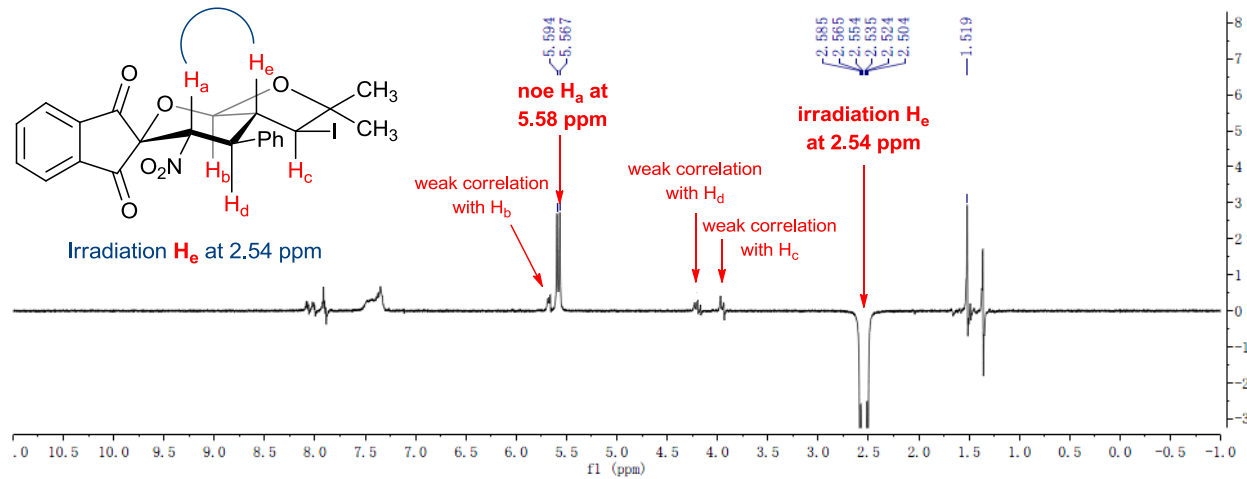
8. The confirmation of the absolute configuration of **7l**

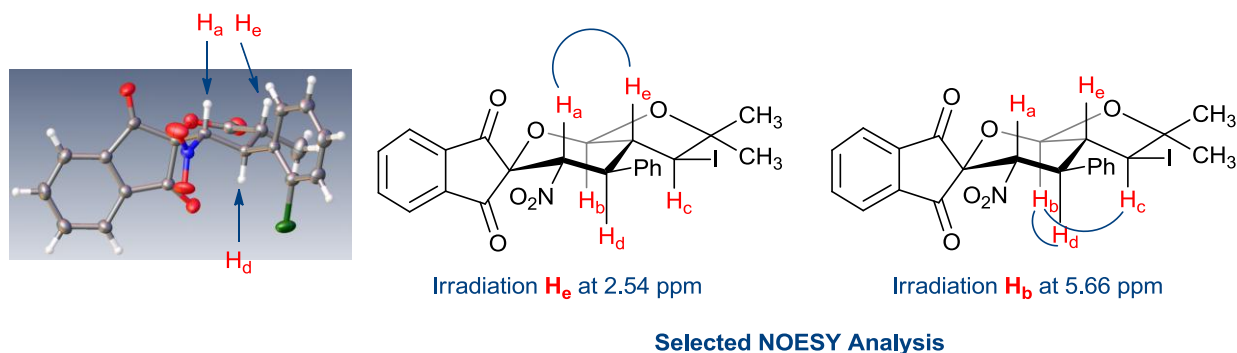
The absolute configuration of **7l** was determined by selected NOESY spectra and the products **7m-7o** were assigned by analogy.



◇ The ¹H NMR spectrum of **7l** exhibited five proton signals ascribable to **H_a**, **H_b**, **H_c**, **H_d** and **H_e** on the hexahydrofuro[2,3-*b*]pyran framework. Other signals observed in the ¹H NMR spectrum included nine aromatic protons and two methyl singlets (δ 1.51, 1.36 ppm).

◇ Three doublets could be assigned as **H_a**, **H_b** and **H_c** by chemical shifts and coupling constants. **H_d** appeared as a triplet at δ 4.18 (t, *J* = 11.2 Hz, 1H) and **H_e** appeared as a triple doublet at δ 2.53 (td, *J* = 12.0, 8.0 Hz, 1H).





- ◇ The absolute configuration of H_a , H_d and H_e could be confirmed by analogy from X-ray crystal structure of **6b** and the conformation of the three C-H bonds could be determined as axial.
- ◇ Irradiation **H_e** at 2.54 ppm: The strong NOESY correlation between H_e and H_a indicated that H_e and H_a were on the same side of the ring which was consistent with the X-ray crystal structure.
- ◇ Irradiation **H_b** at 5.66 ppm: The strong NOESY correlations of H_b with H_d and H_c indicated that H_b , H_d and H_c were on the same side of the ring.