Supporting Information

Pd-catalyzed stereospecific allyl-aryl coupling of allylic alcohols with arylboronic acids

Jiang Ye, Jingming Zhao, Jing Xu, Yuxue Mao, Yong Jian Zhang*

School of Chemistry and Chemical Engineering, and Shanghai Key Laboratory of Electrical Insulation and Thermal Aging, Shanghai Jiao Tong University

Fax: (+) 86-21-54741297; E-mail: yjian@sjtu.edu.cn

Table of Contents

General Experimental Details General Procedure for Synthesis of Chiral Allylic Alcohols Spectroscopic Data for Allyl Alcohols General Procedure for Pd-Catalyzed Allyl-Aryl Coupling of Allyl Alcohols with Arylboronic Acids Spectroscopic Data for Substrates and Allyl-Aryl Coupling Products References HPLC Charts for Enantiomeric Enriched Substrates and Products

General Experimental Details

Analytical thin-layer chromatography (TLC) was carried out using 0.2-mm commercial silica gel plates (Yantai Jiangyou Silica Gel Development Co., Ltd., silica gel HSGF 254). Preparative column chromatography employing silica gel (Qingdao Shenghai Fine Silica Gel Chemical Co., Ltd., 200-300 mesh) was performed according to the method of Still.¹ Toluene, cyclohexane, dichloromethane, ether and pentane were distilled from CaH₂, THF was distilled from sodium/benzophenone. Methanol was treated with a small amount of sodium before distillation. Proton nuclear magnetic resonance (¹H NMR) spectra and Carbon-13 nuclear magnetic resonance (¹3C NMR) spectra were recorded with a Varian Mercuryplus 400 NMR spectrometer. Chemical shifts are reported in delta (δ) units, parts per million (ppm) downfield from tetramethylsilane or ppm relative to the center of the singlet at 7.26 ppm for deuteriochloroform. Coupling constants are reported in Hertz (Hz). Chemical shifts are reported in delta (δ) units, ppm relative to the center of the triplet at 77.0 ppm for deuteriochloroform. And the following abbreviations are used to indicate the multiplicity: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. High performance liquid chromatography (HPLC) was performed on Thermo Fisher Scientific Dionex Ultimate 3000 HPLC by using Daicel Chiracel OD-H or OJ-H columns with *i*-PrOH/hexane as the eluent. Optical rotations were measured on a SGW-1 polarimeter. The melting points were measured by Netzsch DSC-200 F3 differential scanning calorimeter.

 $Pd_2(dba)_3$ •CHCl₃ and all ligands were purchased from Sinocompound Co. and used as received. The arylboronic acids were purchased from Accela ChemBio Inc. and used after recrystallization with water. Racemic allylic alcohols were prepared by available methods: rac-1a,² rac-1b,³ rac-1c,⁴ rac-1f⁵; for rac-1d and rac-1e: the first step is to synthesis the corresponding unsaturated ketones,⁶ then reduce the ketones by using NaBH₄ and CeCl₃·7H₂O (MeOH as solvent, ice bath) under the monitoring of TLC. All other chemicals were used as received from commercial resources.

General Procedure for Synthesis of Chiral Allylic Alcohols⁷

$$R_{1} \xrightarrow{OH} R_{2} \xrightarrow{Ti(OiPr)_{4}, D-DIPT} OH \xrightarrow{OH} R_{1} \xrightarrow{OH} R_{2}$$

To a cooled (-20 °C) suspension of 4Å Molecular Sieves in dry DCM (0.2 M–0.25M), Ti(O*i*Pr)₄ (0.3 eq) and D-DIPT (0.3 eq) were added under N₂ atmosphere. After stirred for half an hour, allylic alcohol (1.0 eq) in DCM and TBHP (0.6–0.8 eq, approx 5.5 M solution in decane) was added subsequently. After keeping stirred for another 12 hours at the same temperature, the reaction was quenched by adding an aqueous solution of FeSO₄ 7H₂O (2.0 eq) and tartaric acid (1.3 eq). Then the mixture was continued to be stirred for 30 min at -20 °C and more than 3 hours in addition at room temperature. The mixture was extracted with DCM three times, dried over Na₂SO₄ and concentrated under reduced pressure. The crude compound was purified by flash column chromatography (EA:PE = 1:100–1:20, v/v) to afford enantioenriched allylic alcohols in 25–37% yields.

(S,E)-4-phenylbut-3-en-2-ol $(1a)^8$



White solid; m.p. 54 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.40–7.22 (m, 5H), 6.57 (d, J = 15.6 Hz, 1H), 6.26 (dd, J = 15.8, 6.4 Hz, 1H), 4.53–4.46 (m, 1H), 1.75 (br s, 1H), 1.38 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 136.6, 133.5, 129.3, 128.5, 127.6, 126.4, 68.9, 23.4; $[\alpha]^{25}{}_{D} = -26.0$ (c = 0.32, CHCl₃); 96% ee; [lit.⁸ $[\alpha]^{20}{}_{D} = -27.0$ (c = 1.0, CHCl₃), 98% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 13.2 min, t_{minor} = 8.7 min.

(S,E)-1-phenylhept-1-en-3-ol $(1b)^9$



Colourless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.40–7.22 (m, 5H), 6.57 (d, J = 16.4 Hz, 1H), 6.23 (dd, J = 15.6, 6.4 Hz, 1H), 4.28 (m, 1H), 1.71–1.56 (m, 3H), 1.46–1.32 (m, 4H), 0.91 (t, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 136.7, 132.6, 130.2, 128.6, 127.6, 126.4, 73.1, 37.1, 27.6, 22.6, 14.1; $[\alpha]^{22}_{D} = +1.3$ (c = 0.72, CHCl₃); 96% ee; [lit.⁹ $[\alpha]^{20}_{D} = +1.5$ (c = 0.68, CHCl₃), 98% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 12.2 min, t_{minor} = 7.9 min.

(*S*,*E*)-1,5-diphenylpent-1-en-3-ol (1c)¹⁰



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.40–7.18 (m, 10H), 6.59 (d, J = 15.6 Hz, 1H), 6.25 (dd, J = 16.0, 7.2 Hz, 1H), 4.34–4.28 (m, 1H), 2.83–2.70 (m, 2H), 2.04–1.89 (m, 2H), 1.61 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 141.7, 136.6, 132.1, 130.6, 128.6, 128.45, 128.38, 127.7, 126.4, 125.9, 72.3, 38.7, 31.7; $[\alpha]^{22}_{D} = +13.2$ (c = 0.68, CHCl₃); 98% ee; [lit.¹⁰ $[\alpha]^{20}_{D} = +20$ (c = 0.6, EtOH), 79% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 15/85, t_{major} = 16.0 min, t_{minor} = 14.1 min. (*S,E*)-4-(p-tolyl)but-3-en-2-ol (1d)^{8,11}



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.28 (d, *J* = 7.6 Hz, 2H), 7.13 (d, *J* = 8.0 Hz, 2H), 6.53 (d, *J* = 15.2, 1H), 6.21 (dd, *J* = 16.0, 6.8 Hz, 1H), 4.51–4.45 (m, 1H), 1.59 (bs, 1H), 2.34 (s, 3H), 1.37 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 137.4, 133.8, 132.5, 129.3, 129.2, 126.3, 69.0, 23.4, 21.2; $[\alpha]^{25}_{D}$ = -20.9 (*c* = 0.67, CHCl₃); 98% ee; [lit.⁸ $[\alpha]^{20}_{D}$ = -22.8 (*c* = 1.0, CHCl₃), >99% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 9.9 min, t_{minor} = 8.5 min. (*S,E*)-4-cvclohexvlbut-3-en-2-ol (1e)¹²

OH M

1e

Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 5.58 (dd, J = 15.2, 6.4 Hz, 1H), 5.46 (ddd, J = 15.2, 6.4, 0.8 Hz, 1H), 4.25 (m, 1H), 1.97–1.89 (m, 1H), 1.75–1.62 (m, 4H), 1.46 (d, J = 2.8 Hz, 1H, OH), 1.32–1.00 (m, 6H), 1.25 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 136.5, 131.5, 68.8, 40.0, 32.71, 32.69, 26.0, 25.9, 23.3; $[\alpha]^{22}{}_{D} = -16.4$ (c = 0.56, CHCl₃); 95% ee; [The *ee* value was determined after converted to *p*-nitrobenzoate. HPLC conditions: Chiralcel OJ-H column, 254 nm, flow rate: 0.8 ml/min, *i*-PrOH/hexane = 3/97, t_{major} = 7.32 min, t_{minor} = 8.76 min].

(*S*,*E*)-1-phenylhex-4-en-3-ol (1f)¹³



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.28 (t, *J* = 7.6 Hz, 2H), 7.21–7.16 (m, 3H), 5.68 (dq, *J* = 21.6, 6.8 Hz, 1H), 5.52 (ddd, *J* = 15.2, 6.8, 1.6 Hz, 1H), 4.09–4.04 (m, 1H), 2.75–2.62 (m, 2H), 1.71 (d, *J* = 6.4 Hz, 3H), 1.46 (bs, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 142.0, 134.0, 128.4, 128.3, 127.1, 125.7, 72.3, 38.7, 31.7, 17.7; $[\alpha]^{22}{}_{D} = -21.1$ (*c* = 0.56, CHCl₃); 98% ee; HPLC conditions: Chiralcel OD-H column, 220 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 6.8 min, t_{minor} = 8.9 min.

General Procedure for Pd-Catalyzed Allyl-Aryl Coupling of Allyl Alcohols with Arylboronic Acids

To a 10 mL Schlenck flask equipped with a stir bar, was added $Pd_2(dba)_3$ CHCl₃ (0.004 mmol), *rac*-BINAP (0.008 mmol), Allyl alcohol **1** (0.4 mmol), and Arylboronic acid **2** (0.6 mmol), after the system was degassed and recharged with N₂ three times, Toluene (0.8 mL) was injected *via* syringe. The mixture was stirred at 50 °C for 18 or 24 h, and then the solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (ethyl acetate: petroleum ether, 1:200–1000) to furnish corresponding allyl–aryl coupling products.

(R, E)-But-1-ene-1,3-diyldibenzene $(3a)^{12}$



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36–7.17 (m, 10H), 6.44–6.35 (m, 2H), 3.64 (dq, J = 6.8, 6.8 Hz, 1H), 1.46 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 145.6, 137.5, 135.2, 128.5, 127.3, 127.0, 126.2, 126.1, 42.5, 21.2; $[\alpha]^{25}{}_{D} = +34.7$ (c = 0.41, CHCl₃); 96% ee; [lit.¹² $[\alpha]^{20}{}_{D} = +37.4$ (c = 0.59, CHCl₃), 97% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexanes = 0.1/99.9, t_{minor}=10.8 min, t_{major} = 11.3 min. The absolute configurations were determined by comparing the sign of the optical rotation with that reported.¹²

(*R*, *E*)-1-methyl-2-(4-phenylbut-3-en-2-yl)benzene (3b)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.35–7.10 (m, 9H), 6.37–6.35 (m, 2H), 3.89–3.82 (m, 1H), 2.37 (s, 3H), 1.44 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 134.5, 137.6, 135.6, 134.8, 130.4, 128.5, 128.4, 127.0, 126.3, 126.2, 126.1, 126.0, 38.0, 20.4, 19.5; $[\alpha]^{25}_{D}$ = +32.7 (*c* = 1.47, CHCl₃); 94% ee; [lit.¹² $[\alpha]^{20}_{D}$ = +38.9 (*c* = 0.55, CHCl₃), 96% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 0.1/99.9, t_{major} = 8.9 min, t_{minor} = 7.9 min.

(*R*, *E*)-1-methoxy-3-(4-phenylbut-3-en-2-yl)benzene (3c)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36 – 7.17 (m, 6H), 6.88–6.74 (m, 3H), 6.44–6.34 (m, 2H), 3.80 (s, 3H), 3.61 (dq, *J* = 7.2, 6.8 Hz, 1H), 1.45 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 159.7, 147.3, 137.5, 135.0, 129.4, 128.5, 128.4, 127.0, 126.1, 119.7, 113.3, 111.2, 55.1, 42.6, 21.1; [α]²⁵_D = +8.5 (*c* = 0.45, CHCl₃);

95% ee; $[lit.^{12} [\alpha]_{D}^{20} = +10 \ (c = 0.28, CHCl_3), 92\%$ ee]; HPLC conditions: Chralcel OJ-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 8.2 min, t_{minor} = 9.5 min.

(R,E)-1-methyl-4-(4-phenylbut-3-en-2-yl)benzene (3d)¹⁴



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36–7.15 (m, 9H), 6.43–6.34 (m, 2H), 3.61 (dq, J = 6.8, 6.8 Hz, 1H), 2.33 (s, 3H), 1.46 (d, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 142.6, 137.6, 135.7, 135.4, 129.1, 128.4, 128.3, 127.2, 127.0, 126.1, 42.1, 21.25, 21.0; $[\alpha]^{20}_{D} = +39.8$ (c = 0.42, CHCl₃); 95% ee; HPLC conditions: Chralcel OJ-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 15/85, t_{major} = 6.4 min, t_{minor} = 5.9 min.

(*R*,*E*)-1-(tert-butyl)-4-(4-phenylbut-3-en-2-yl)benzene (3e)



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.38–7.17 (m, 9H), 6.45–6.34 (m, 2H), 3.62 (dq, J = 6.8, 6.8 Hz, 1H), 1.46 (d, J = 7.2 Hz, 3H), 1.31 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 148.9, 142.5, 137.6, 135.4, 128.4, 128.2, 126.9, 126.8, 126.1, 125.3, 42.1, 34.4, 31.4, 21.1; $[\alpha]^{15}_{D} = +20.2$ (c = 0.92, CHCl₃); 95% ee; HPLC conditions: Chralcel OD-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 0.1/99.9, t_{major} = 7.4 min, t_{minor} = 6.7 min.

(*R*,*E*)-1-methoxy-4-(4-phenylbut-3-en-2-yl)benzene(3f)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36–7.16 (m, 7H), 6.88–6.84 (m, 2H), 6.41–6.32 (m, 2H), 3.79 (s, 3H), 3.60 (dq, *J* = 7.2, 6.8 Hz, 1H), 1.44 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 137.7, 137.6, 135.6, 128.5, 128.2, 127.0, 126.1, 113.8, 55.3, 41.7, 21.3; $[\alpha]^{20}_{D} = +11.3$ (*c* = 0.51, CHCl₃); 94% ee; [lit.¹² $[\alpha]^{20}_{D} = +36.1$ (*c* = 0.29, CHCl₃), 94% ee]; HPLC conditions: Chralcel OJ-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 10.0 min, t_{minor} = 9.4 min.

(*R*, *E*)-1-chloro-4-(4-phenylbut-3-en-2-yl)benzene (3g)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36–7.19 (m, 9H), 6.42–6.30 (m, 2H), 3.62 (dq, J = 6.8, 6.8 Hz, 1H), 1.44 (d, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 144.0, 137.3, 134.5, 131.8, 128.8, 128.7, 128.5, 128.5, 127.2, 126.1, 41.9, 21.1; $[\alpha]^{25}_{D} = +23.4$ (c = 0.60, CHCl₃); 89% ee; [lit.¹² $[\alpha]^{20}_{D} = +14.2$ (c = 0.75, CHCl₃), 93% ee]; HPLC conditions: Chralcel OJ-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 1/99, t_{major} = 12.5 min, t_{minor} = 11.3 min.

(*R*, *E*)-2-(4-phenylbut-3-en-2-yl)naphthalene (3h)¹²



Off-white solid, m.p. 72 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.97–7.87 (m, 1H), 7.81–7.78 (m, 3H), 7.69 (s, 1H), 7.45–7.17 (m, 7H), 6.46–6.45 (m, 2H), 3.83–3.70 (m, 1H), 1.55 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 143.0, 137.5, 135.1, 133.6, 132.2, 128.8, 128.5, 128.0, 127.6, 127.6, 127.1, 126.3, 126.2, 125.9, 125.3, 125.2, 42.6, 21.1; $[\alpha]^{25}_{D} = +21.0$ (*c* = 0.40, CHCl₃); 94% ee; [lit.¹² $[\alpha]^{20}_{D} = +13.1$ (*c* = 0.17, CHCl₃), 93% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 0.1/99.9, t_{major} = 24.2 min, t_{minor} = 23.1 min.

(*R*, *E*)-hept-1-ene-1,3-diyldibenzene (3i)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.36–7.16 (m, 10H), 6.41–6.30 (m, 2H), 3.39 (dt, J = 7.2, 7.2 Hz), 1.83–1.76 (m, 2H), 1.37–1.20 (m, 4H), 0.87 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 134.5, 129.2, 128.4, 128.4, 127.6, 127.0, 126.1, 126.1, 49.2, 35.6, 29.8, 22.7, 14.0; $[\alpha]^{16}_{D}$ = +15.3 (c = 0.76, CHCl₃); 94% ee; [lit.¹² $[\alpha]^{20}_{D}$ = +17.4 (c = 0.46, CHCl₃), 95% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 0.1/99.9, t_{major} = 8.1 min, t_{minor} = 7.7 min.

(*R*, *E*)-pent-1-ene-1,3,5-triyltribenzene (3j)¹²



7 / 29

Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.35–7.15 (m, 15H), 6.43–6.31 (m, 2H), 3.45 (dt, J = 7.2, 7.6 Hz, 1H), 2.69–2.55 (m, 2H), 2.15 (dt, J = 8.4, 7.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 134.0, 129.7, 128.6, 128.4, 128.3, 127.7, 127.1, 126.3, 126.1, 125.8, 48.5, 37.3, 33.7; $[\alpha]^{20}{}_{D} = -9.0$ (c = 1.06, CHCl₃); 91% ee; [lit.¹² $[\alpha]^{20}{}_{D} = -4.5$ (c = 0.45, CHCl₃), 97% ee]; HPLC conditions: Chiralcel OD-H column, 254 nm, flow rate: 1.0 ml/min, *i*-PrOH/hexane = 0.5/99.5, t_{major} = 11.4 min, t_{minor} = 10.5 min.

(*R*,*E*)-1-methyl-4-(3-phenylbut-1-en-1-yl)benzene(3k)¹⁵



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.33–7.19 (m, 7H), 7.10–7.08 (m, 2H), 6.41–6.30 (m, 2H), 3.63 (dq, *J* = 7.2, 6.4 Hz, 1H), 2.32 (s, 3H), 1.46 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 145.8, 136.7, 134.7, 134.2, 129.2, 128.4, 128.3, 127.3, 126.1, 126.0, 42.5, 21.3, 21.1; $[\alpha]^{20}_{D}$ = +38.4 (*c* = 0.98, CHCl₃); 91% ee; HPLC conditions: Chralcel OJ-H column, 254 nm, flow rate: 1 ml/min, *i*-PrOH/hexane = 10/90, t_{major} = 11.6 min, t_{minor} = 14.3 min.

(*R*,*E*)-(4-cyclohexylbut-3-en-2-yl)benzene(31)¹²



Colorless oil; ¹H NMR (400 MHz, CDCl₃): δ 7.30–7.12 (m, 5H), 5.55 (ddd, J = 15.6, 6.8, 0.8 Hz, 1H), 5.41 (dd, J = 15.6, 6.0 Hz, 1H), 3.40 (dq, J = 7.2, 6.8 Hz, 1H), 1.96–1.89 (m, 1H), 1.74–1.67 (m, 4H), 1.32 (d, J = 7.2 Hz, 3H), 1.27–1.01 (m, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 135.2, 132.3, 128.3, 127.9, 127.1, 125.8, 42.2, 40.6, 33.18, 33.17, 26.2, 26.1, 21.6; $[\alpha]^{22}{}_{D} = +8.0$ (c = 0.50, CHCl₃); 93% ee; [lit.¹² $[\alpha]^{20}{}_{D} = +10.1$ (c = 0.45, CHCl₃), 95% ee]; HPLC conditions: Chiralcel OJ-H column, 254 nm, flow rate: 0.4 ml/min, *i*-PrOH/hexane = 0/100, $t_{major} = 11.7$ min, $t_{minor} = 12.8$ min.

(R,E)-hex-4-ene-1,3-diyldibenzene(3m) + (S,E)-hex-3-ene-1,5-diyldibenzene $(3n)^{12,16}$



Compouds **3m** and **3n** are inseparable mixture isolated in 91% yield with a ratio **3m**:**3n** = 42:58 by ¹H NMR analysis. Pure racemic **3m** and **3n** can be synthesized by the method described in the literature.¹⁶

For **3m**, ¹H NMR (400 MHz, CDCl₃) δ 7.32–7.14 (m, 10H), 5.63–5.59 (m, 1H), 5.47–5.40 (m, 1H), 3.22 (dt, J = 8.0, 7.2 Hz, 1H), 2.61–2.47 (m, 2H), 2.00 (dt, J = 8.4, 7.2, 2H), 1.68–1.66 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 145.0, 142.4, 134.9, 128.43, 128.41, 128.2, 127.5, 126.0, 125.6, 125.0, 48.4, 37.6, 33.8, 18.0.

For **3n**, ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.13 (m, 10H), 5.60–5.56 (m, 1H), 5.52–5.49 (m, 1H), 3.40 (dq, J = 7.2, 7.2 Hz, 1H), 2.70–2.63 (m, 2H), 2.36–2.31 (m, 2H), 1.31 (d, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 146.3, 142.0, 135.7, 128.5, 128.3, 128.2, 127.1, 125.9, 125.7, 42.2, 36.0, 34.4, 21.4.

HPLC conditions for **3m** and **3n**: Chiralcel OJ-H column, 220 nm, flow rate: 0.75 ml/min, *i*-PrOH/hexane = 0.3/99.7. t_{major} (**3m**) = 13.9 min, t_{minor} (**3m**) = 15.3 min, 96%ee; t_{major} (**3n**) = 19.3 min, t_{minor} (**3n**) = 18.0 min, 97% ee.

References

- 1. W. C. Still, M. Kahn and A. Mitra, J. Org. Chem., 1978, 43, 2923.
- 2. B. Martin-Matute, K. Bogar, M. Edin, F. B. Kaynak and J. E. Bäckvall, Chem. Eur. J., 2005, 11, 5832.
- 3. B. M. Trost and R. J. Kulawiec, J. Am. Chem. Soc., 1993, 115, 2027.
- 4. I. D. G. Watson, S. A. Styler and A. K. Yudin, J. Am. Chem. Soc., 2004, 126, 5086.
- 5. M. Dobmeier, J. M. Herrmann, D. Lenoir and B. König, Beilstein J. Org. Chem., 2012, 8, 330.
- 6. K. Zumbansen, A. Dohring and B. List, Adv. Synth. Catal., 2010, 352, 1135.
- (a) M. Sasaki, H. Ikemoto, M. Kawahata, K. Yamaguchi and K. Takeda, *Chem. Eur. J.*, 2009, **15**, 4663; (b) P. R. Carlier, W. S. Mungall, G. Sohröder and K. B. Sharpless, *J. Am. Chem. Soc.*, 1988, **110**, 2978.
- 8. C. Quirin and U. Kazmaier, Synthesis, 2009, 10, 1725.
- 9. M. Node, K. Nishide, Y. Shigeta, H. Shiraka and K. Obata, J. Am. Chem. Soc., 2000, 122, 1927.
- 10. Y. Liu, C.-S. Da, S.-L. Yu, X.-G. Yin, J.-R Wang, X.-Y Fan, W.-P Li and R. Wang, J. Org. Chem., 2010, 75, 6869.
- 11. S. R. Goudreau and A. B. Charette, J. Am. Chem. Soc., 2009, 131, 15633.
- (a) J. Zhao, J. Ye and Y-J. Zhang, *Adv. Synth. Catal.*, 2013, **355**, 491; (b) C. Li, J. Xing, J. Zhao, P. Huynh, W. Zhang, P. Jiang and Y. J. Zhang, *Org. Lett.*, 2012, **14**, 390.
- 13. S. Akai, R. Hanada, N. Fujiwara, Y. Kita and M. Egi, Org. Lett., 2010, 12, 4900.
- 14. R. Riveiros, R. Tato, J. P. Sestelo and L. A. Sarandeses, Eur. J. Org. Chem., 2012, 15, 3018.
- 15. H. Horibe, Y. Fukuda, K. Kondo, H. Okuno, Y. Murakami and T. Aoyama, Tetrahedron, 2004, 60, 10701.
- 16. H. Ohmiya, Y. Makida, T. Tanaka and M. Sawamura, J. Am. Chem. Soc., 2008, 130, 17276.

















Total:



1612.384

100.00

100.00





















Integration Results										
No.	Peak Name	Retention Time	Area	Height	Relative Area	Relative Height	Amount			
		min	mAU*min	mAU	%	%	n.a.			
1		6.040	118.731	992.652	49.89	52.34	n.a.			
2		6.570	119.240	903.736	50.11	47.66	n.a.			
Total:			237.971	1896.388	100.00	100.00				





























N	 Peak Name 	Retention Time	Area	Height	Relative Area	Relative Height	Amount
		min	mAU*min	mAU	%	%	n.a.
1		11.327	755.296	2103.646	49.11	60.59	n.a.
2		14.267	782.582	1368.177	50.89	39.41	n.a.
Т	otal:		1537.878	3471.823	100.00	100.00	











Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013

