1	Supporting Information
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3	Self-Assembly of Diphenylalanine Peptide into Microtubes
4	with "Turn on" Fluorescence by an Aggregation-Induced
5	Emission Molecule
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10 Experimental Section

Reagents. All chemicals were of analytical or higher grade. All solutions were 11 prepared using ultrapure water from a Millipore Simplicity 185 water purification 12 (Millipore, Milford, MA, USA). The lyophilized diphenylalanine system 13 (NH₂-Phe-Phe-COOH, FF, ≥98%), 1,1,1,3,3,3-hexafluoro-2-propanol (HFP) were 14 purchased from Sigma-Aldrich Co. (USA). FF was characteristed by NMR: ¹H NMR 15 (CD₃OD, 400 MHz): δ 7.35~7.24 (m,10H), 4.52(dd, $J_1 = 8.1$ Hz, $J_2 = 4.8$ Hz, 1H), 16 $3.99 (dd, J_1 = 9.0 Hz, J_2 = 4.8 Hz, 1H), 3.28 \sim 3.22 (m, 2H), 3.02 (dd, J_1 = 13.9 Hz, J_2)$ 17 = 8.1 Hz, 1H), 2.92 (dd, J_1 = 14.4 Hz, J_2 = 9.0 Hz, 1H). The electrospray mass 18 19 spectra of FF is shown in Figure S1-A, which demonstrated the characteristic ions of FF in the positive mode including $[M+H]^+$ (*m/z* 313), $[M+Na]^+$ (*m/z* 335), $[2M+H]^+$ 20 (m/z 625) and $[2M+Na]^+ (m/z 647)$. 21

9, 10-Bis[4-(3-sulfonatopropoxyl)-styryl] anthracene (BSPSA) was synthesized 22 according to our previous works.¹⁵ Briefly, 9,10-Bis(4-methoxystyryl)anthracene was 23 first synthesized by dissolving 9,10-dibromoanthracene, 4-methoxystryrene, K₃PO₄ 24 and Pd(OAc)₂ in DMAC, and then stirred at 110 °C for 24 h. Then, the obtained 25 9,10-Bis(4-methoxystyryl)anthracene and dry CH₂Cl₂ were mixted and cooled to 26 -78 °C, after which boron tribromide/CH₂Cl₂ (1.51 g/10 mL) was added. After 27 stirring overnight, water was added to generate 9,10-Bis(4-hydroxystyryl)anthracene. 28 At last, 9,10-Bis(4-hydroxystyryl)anthracene was added into anhydrous under 29 nitrogen, after which NaOEt/anhydrous ethanol was slowly added. Then, with the 30 addition of 1,3-propanesultone/ethanol, the mixture was vigorously stirred overnight 31

32	and a yellow product of BSPSA was precipitated. ¹ H NMR (DMSO-d6, 400 MHz): δ
33	8.40 (d, J = 3.3 Hz, 2H), 8.38 (d, J = 3.3 Hz, 2H), 7.96 (d, J = 16.5 Hz, 2H), 7.74 (d, J
34	= 8.7 Hz, 4H), 7.55 (d, J = 3.2 Hz, 2H), 7.53 (d, J = 3.2 Hz, 2H), 7.01 (d, J = 8.7 Hz,
35	4H), 6.86 (d, = 16.5 Hz, 2H), 4.13 (t, J = 6.5 Hz, 4H), 2.58 (t, J = 7.2 Hz, 4H), 2.03 (m,
36	4H). The electrospray mass spectra of BSPSA are shown in Figure S1-B, which
37	shows the characteristic ion of $[M-2Na]^{2-}$ (<i>m/z</i> 328) in the negative mode.

Self-assembly of FF. For the self-assembly, 10 mg of lyophilized FF was 38 dissolved in 100 µL HFP to disrupt any possible aggregation in the peptide sample. 39 Then, 20 µL of FF/HFP (100 mg/mL) solution was transfered to a 1.5 ml centrifuge 40 tube, and allowed to dry by evaporation under ambient conditions. After FF/HFP 41 solution was dried completely by evaporation at ambient conditions, FF was 42 re-solubilized in ultrapure water by addition of 1 mL water ²⁰ or a certain 43 concentration of BSPSA agoeous solution and subsequent ultrasonic treated at 65 ° C 44 for 30 min. The FF aqueous solution was then annealed at room temperature and 45 allowed to age for one day. The obtained assemblage was utilized for further 46 characterization. 47

Characterization Methods. Fluorescence microscope images were recorded on an Olympus IX71 fluorescence microscope (Olympus, Japan). 20 µL 0.5 mg/ml FF aqueous solution was pipetted onto a glass slide to dry by evaporation under ambient conditions. Pictures were then taken after the assembly procedures. Scanning electron microscope (SEM) photographs were taken on a Hitachi S-4800 scanning electron microscope (Hitachi, Japan) by dripping 30 µL 0.5 mg/ml FF aqueous solution on

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54	glass slide to dry. Before SEM imaging, a thin layer of Pt was coated on the film
55	surface by an Auto Fine Coater (JEOL) at a current of 20 mA for 150 s. Fluorescence
56	spectra were recorded by a SHIMADZU RF-5301PC spectrophotometer (Shimadzu,
57	Japan) at 25 °C (5-nm slit width, 1-cm path-length cuvettes,). For polarization
58	experiments, the obtained microtubes after self-assembly of Phe-Phe in BSPSA
59	solution were directly subjected to the test in aqueous solution. UV-Vis absorption
60	spectra were obtained with a TU-1901 UV-Vis spectrophotometer (Beijing, China).
61	Circular dichroism (CD) spectra were recorded on a Pistar π -180 spectrophotometer
62	(Applied Photophysics Ltd., UK) at room temperature using 1 mm quartz cells. Scans
63	were obtained in a range between 190 and 350 nm by taking points at 1 nm (time per
64	point: 2 s, Bandwidth: 4 nm). Three spectra were averaged to improve the
65	signal-to-noise ratio.
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phenylalanine



9,10-bis [4-(3-sulfonatopropoxyl)-styryl] anthracene $(BSPSA)\ sodium$

- 75 **Figure S2.** Molecular structure of phenylalanine and BSPSA.
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Figure S3. (A) Fluorescence microscope image of microtubes obtained by self-assembly of 2.0 mg/mL FF in water and then incubated with 0.01 mg/mL BSPSA solution. (B) Fluorescence spectra of BSPSA, microtubes of incubating formed non-fluorescence microtubes with BSPSA, and microtubes obtained by FF self-assembly in BSPSA solution. BSPSA concentration: 0.01 mg/mL.







- 87 and the emission spectrum (red line). BSPSA concentration: 0.01 mg/mL.
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Figure S5. Fluorescence spectra of different concentration of FF assemblage in
BSPSA solution. FF concentration: 0.12, 0.25, 0.5, 1.0, 2.0, 4.0 mg/mL, respectively.
BSPSA concentration: 0.01 mg/mL. Excitation wavelength: 404 nm.





Figure S6. Polarized emissions from microtubes obtained by the self-assembly of
different concentrations of FF in BSPSA solutions. FF concentration: 0.12, 0.25, 0.5,
1.0, 2.0 mg/mL, respectively. BSPSA concentration: 0.01 mg/mL. Excitation
wavelength: 404 nm.





Figure S7. UV-Vis absorption spectra of microtubes by self-assembly of FF in
BSPSA aqueous solution. (A) 2.0 mg/mL FF in different concentration of BSPSA

107 solution. (B) Different concentration of FF in 0.01 mg/mL BSPSA.



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112 **Figure S8.** (A) FL spectra of 0.2 mg/mL FF solutions in different concentration of

BSPSA aqueous solution (0.005 to 0.07 mg/mL). Excitation: 260 nm. (B) CD spectra

of 0.16 mg/mL BSPSA solution, microtubes obtained by self-assembly of 2.0 mg/mL

115 FF in water, 0.01, 0.04 and 0.16 mg/mL BSPSA aqueous solution. The samples were

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116 four times diluted for the test.
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