SUPPORTING INFORMATION

Efficient Asymmetric Synthesis of Lamivudine via Enzymatic Dynamic Kinetic Resolution

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Table of contents

General	S 2
Immobilization of subtilisin Carlsberg	S2
DKR of $((2R)$ -5-acetoxy-1,3-oxathiolan-2-yl)methyl benzoate 9a	S2
Synthesis of ((2 <i>R</i> ,5 <i>S</i>)-5-(4-acetamido-2-oxopyrimidin-1(2 <i>H</i>)-yl)-1,3-oxathiolan-2-yl)methyl benzoate 10	S2
Synthesis of 4-amino-1-((2 <i>R</i> ,5 <i>S</i>)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl)pyrimidin-2(1 <i>H</i>)- one 1a	S2
Synthesis of ((2S)-5-acetoxy-1,3-oxathiolan-2-yl)methyl acetate 7a	S 3
Synthesis of ((2 <i>R</i>)-5-acetoxy-1,3-oxathiolan-2-yl)methyl acetate 7b	S 3
Synthesis of 4-amino-1-((2 <i>S</i> ,5 <i>R</i>)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl)pyrimidin-2(1 <i>H</i>)- one 1b	S 3
Chiral HPLC of 9a	S 3
Chiral HPLC of 9b	S 4
Chiral HPLC of racemic compound 9	S 4
Chiral HPLC of 7a	S 4
Chiral HPLC of 7b	S4
Chiral HPLC of racemic compound 7	S4
¹ H NMR and ¹³ C NMR of 9	S5
¹ H NMR and ¹³ C NMR of 10	S6
¹ H NMR and ¹³ C NMR of 1a	S 7
¹ H NMR and ¹³ C NMR of 7	S 8
¹ H NMR and ¹³ C NMR of 1b	S 9
References	S 9

General: All commercially available starting materials were of reagent grade and used as received. Subtilisin Carlsberg was purchased from Sigma-Aldrich (Art. No: P5380). ¹H NMR and ¹³C NMR data were recorded on a Bruker Avance DMX 500 at 500 MHz. Chemical shifts are reported as δ values (ppm) with CDCl₃ (¹H NMR δ 7.26, ¹³C NMR δ 77.16) as internal standard. *J* values are given in Hertz (Hz). Thin layer chromatography (TLC) was performed on precoated Cromatofolios AL Silica gel 60 F₂₅₄ (Merck, Darmstadt, Germany), visualized with UV-detection. Flash column chromatography was performed on silica gel 60, 0.040-0.063 mm (SDS). Analytical high performance liquid chromatography (HPLC) with chiral stationary phases was performed using an HP-Agilent 1110 series controller, equipped with a Daicel chiralpak OJ (4.6x250 mm, 20 µm) column. Solvents for HPLC use were of spectrometric grade.

Immobilization of subtilisin Carlsberg: To a solution of octyl β -D-glucopyranoside (25 mg) and Brij 56 (25 mg) in phosphate buffer (0.5 M, pH 7.2, 10 mL) was added subtilisin Carlsberg (100 mg). The mixture was rapidly frozen with liquid nitrogen and lyophilized overnight.

((2R)-5-acetoxy-1,3-oxathiolan-2-yl)methyl benzoate (9a)²

Compound **8** (85.4 mg, 0.52 mmol), 1,4-dithiane-2,5-diol **6** (47.5 mg, 0.31 mmol), triethylamine (72.5 μ L, 0.52 mmol), and phenyl acetate (196 μ L, 1.56 mmol) in dry THF (4 mL) were added to a sealed-cap vial containing surfactant-treated subtilisin Carlsberg (80 mg) together with 4Å molecular sieves (120 mg). The reaction mixture was cooled to 4 °C. After 2 days, the mixture was filtered and washed with saturated NH₄Cl and brine, after which the mixture was dried over MgSO₄ and the solvent removed under vacuum. The crude product was purified by column chromatography (hexane:EtOAc = 8:1) affording compound **9** as a clear oil (130.5 mg, 0.46 mmol, 89%), 82% *ee*, determined by HPLC analysis (Chiral OJ, λ = 254 nm, Hex:¹PrOH = 9:1, 0.5 mL/min; Colorless oil; ¹H NMR (500 Hz, CDCl₃) δ = 8.05 (2H, d, *J* = 8.0 Hz), 7.58 (1H, t, *J* = 7.13 Hz), 7.45 (2H, t, *J* = 7.57 Hz), 6.72 (1H, d, *J* = 4.12 Hz), 5.68 (1H, dd, *J* = 4.96, 1.53 Hz), 4.53 (2H, m), 3.20 (2H, m), 2.12 (3H, s); ¹³C NMR (125 Hz, CDCl₃) δ = 169.2 166.0, 133.29, 129.8, 128.4, 99.2, 83.3, 66.1, 37.6, 21.1.

((2S)-5-acetoxy-1,3-oxathiolan-2-yl)methyl benzoate $(9b)^2$

Following the procedure for compound **9a**, the reaction of compound **8** (8.5 mg, 0.05 mmol) and **6** with TEA (7 μ L, 0.05 mmol), phenyl acetate (19 μ L, 0.15 mmol), and CAL B (20 mg) in toluene (0.5 mL) afforded compound **9b** (11.7 mg, 83%), 84% *ee*, determined by HPLC analysis (Chiral OJ, λ = 254 nm, Hex:ⁱPrOH = 9:1, 0.5 mL/min. The NMR data were the same as compound **9a**, which was the enantiomer of compound **9b**.

((2R,5S)-5-(4-acetamido-2-oxopyrimidin-1(2H)-yl)-1,3-oxathiolan-2-yl)methyl benzoate (10)²

Silylated N⁴-acetylcytosine was generated in situ by stirring N⁴-acetylcytosine (55.4 mg, 0.362 mmol) with hexamethyldisilazane (HMDS, 1.0 mL, 4.8 mmol) at reflux (135 °C) under argon until all solid had dissolved (approximately 3 h). The solution was slowly cooled to rt, and HMDS was removed under anhydrous conditions. The system was evacuated five times with argon, and dried under HV for 30 minutes. Meanwhile, compound 9 (63.2 mg, 0.224 mmol) was dissolved in anhydrous acetonitrile (6 mL) and stirred with pre-activated 3Å molecular sieves for 30 minutes under N₂. The solution was transferred via syringe to the dried silvlated N^4 -acetylcytosine, and the resulting solution was cooled to 0 °C. TMSI (70.0 µL, 0.504 mmol) was added dropwise over 15 minutes and the resulting yellow solution was stirred under argon at 0 °C for 1 h. The reaction was quenched by addition of a mixture of ethyl acetate (10 mL) and aqueous NaHCO₃ solution (5 wt%, 6 mL) and the resulting two-phase system was stirred at rt for 15 minutes. The mixture was further diluted with ethyl acetate (10 mL) and the organic phase was washed with saturated aqueous NaHCO₃ solution (15 mL), distilled H₂O (2 x 15 mL) and brine (2 x 15 mL). Drying over MgSO₄, filtration and concentration *in vacuo* yielded the crude product mixture as a vellow solid. Purification by column chromatography (EtOAc : methanol = 5:1) allowed for separation of the diastereoisomers to yield the *cis* (34.4 mg, 0.113 mmol, 51%) isomers (10) as white solids. ¹H NMR (CDCl₃) $\delta = 8.23$ (1H, d, J = 7.54 Hz), 8.07 (1H, d, J = 8.32 Hz), 7.64 (1H, t, J = 9.26 Hz), 7.52 (1H, t, J = 7.79 Hz), 7.32 (1H, d, J = 8.13 Hz), 6.36 (1H, dd, J = 8.12 Hz), 7.52 (1H, dd, J = 8.12 H 5.31, 2.76 Hz), 5.53 (1H, dd, J = 4.21, 2.98 Hz), 4.82 (2H, m), 3.67 (1H, dd, J = 12.71, 5.39 Hz), 3.26 (1H, dd, J = 12.69, 2.69 Hz), 2.25 (3H, s); ¹³C NMR δ = 170.3, 165.9, 162.6, 154.7, 144.7, 133.8, 129.7, 129.1, 128.7, 96.2, 97.8, 85.5, 63.5, 39.3, 29.7, 25.0.

4-amino-1-((2*R*,5*S*)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl)pyrimidin-2(1*H*)-one (Lamivudine 1a)

To a solution of compound **10** (19 mg, 0.05 mmol) in methanol (0.5 mL), K_2CO_3 (6.9 mg, 0.05 mmol) was added and the mixture stirred at rt under N₂ for 14 h. The mixture was diluted with methanol, filtered and concentrated. Purification by column chromatography (EtOAc:methanol = 4:1), yielded compound **1a** (10.2 mg, 0.045 mmol, 89%). ¹H NMR (500 Hz, D₂O) δ = 7.96 (1H, dd, *J* = 7.58 Hz), 6.29 (1H, dd, *J* = 5.54, 4.33 Hz), 5.99 (1H, d, *J* = 7.57 Hz), 5.32 (1H, dd, *J* = 4.44, 3.40 Hz), 3.91 (2H, m), 3.53 (1H, dd, *J* = 12.26, 5.65 Hz), 3.18 (1H, dd, *J* = 12.26, 4.23 Hz). ¹³C NMR (125 Hz, D₂O) δ = 164.8, 155.8, 140.3, 94.5, 85.8, 84.5, 60.7, 35.3.

((2S)-5-acetoxy-1,3-oxathiolan-2-yl)methyl acetate (7a)¹

Following the procedure for compound **9a**, the reaction of compound **5** (60.1 mg, 0.50 mmol) and **6** (45.7 mg, 0.30 mmol) with CAL B (20 mg), TEA (70 μ L, 0.50 mmol), phenyl acetate (188 μ L, 1.5 mmol) in toluene (4 mL) afforded compound **7a** (101.3 mg, 92%), 83% *ee*, determined by HPLC analysis (Chiral OJ, λ = 210 nm, Hex:¹PrOH = 9:1, 0.5 mL/min. ¹H NMR (500 Hz, CDCl₃) δ = 6.67 (1H, d, *J* = 4.15 Hz), 5.53 (1H, dd, *J* = 6.38, 3.97 Hz), 4.20-4.32 (2H, m), 3.10-3.34 (2H, m), 2.09 (3H, s). ¹³C NMR (125 Hz, CDCl₃) δ = 168.1, 167.3, 96.8, 80.7, 63.4, 35.1, 18.8, 18.4.

((2R)-5-acetoxy-1,3-oxathiolan-2-yl)methyl acetate $(7b)^1$

Following the procedure for compound **9a**, the reaction of compound **5** (6 mg, 0.05 mmol) and **6** (4.6 mg, 0.03 mmol) with STS (10 mg), TEA (7 μ L, 0.05 mmol), phenyl acetate (19 μ L, 0.15 mmol) in toluene (0.5 mL) afforded compound **7b** (9.6 mg, 87%), 64% *ee*, determined by HPLC analysis (Chiral OJ, λ = 210 nm, Hex:ⁱPrOH = 9:1, 0.5 mL/min. The NMR data were tha same as compound **7a**, which was the enantiomer of compound **7b**.

4-amino-1-((2S,5R)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl)pyrimidin-2(1H)-one (1b)

Silvlated N^4 -acetylcytosine was generated in situ by stirring N^4 -acetylcytosine (64.3 mg, 0.42 mmol) with hexamethyldisilazane (HMDS, 868 µL, 4.1 mmol) at reflux (135 °C) under argon until all solid had dissolved (approximately 3 h). The solution was slowly cooled to rt, and HMDS was removed under anhydrous conditions. The system was evacuated five times with argon, and dried under HV for 30 minutes. Meanwhile, compound 7a (62 mg, 0.28 mmol) was dissolved in anhydrous acetonitrile (6 mL) and stirred with pre-activated 3Å molecular sieves for 30 minutes under N₂. The solution was transferred via syringe to the dried silvlated N^4 -acetylcytosine, and the resulting solution was cooled to 0 °C. TMSI (70.0 µL, 0.504 mmol) was added dropwise over 15 minutes and the resulting yellow solution was stirred under argon at 0 °C for 1 h. The reaction was quenched by addition of a mixture of ethyl acetate (10 mL) and aqueous NaHCO₃ solution (5 wt%, 6 mL) and the resulting two-phase system was stirred at rt for 15 minutes. The mixture was further diluted with ethyl acetate (10 mL) and the organic phase was washed with saturated aqueous NaHCO₃ solution (15 mL), distilled H₂O (2 x 15 mL) and brine (2 x 15 mL). Drying over MgSO₄, filtration and concentration *in vacuo* yielded the crude product mixture as a yellow solid (65 mg). The solid was dissolved in methanol (2 mL), and K₂CO₃ (24 mg, 0.17 mmol) was added. The mixture was stirred at rt overnight. The mixture was diluted with methanol, filtered and concentrated. Purification by column chromatography (EtOAc:methanol = 4:1), yielded compound **1a** (26 mg, 40% for two steps). ¹H NMR (500 Hz, DMSO) δ = 7.82 (1H, d, J = 7.48 Hz), 7.21 (1H, d, J = 33.29 Hz), 6.21 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, d, J = 6.93 Hz), 5.32 (1H, t, J = 5.65 Hz), 5.73 (1H, t, J = 5.65 Hz), 5.75 (1H, t, J = 5.65 5.72 Hz), 5.17 (1H, t, J = 4.95 Hz), 3.69-3.75 (1H, m), 3.41 (1H, dd, J = 12.29, 4.02 Hz), 3.04 (1H, dd, J = 11.58, 4.73 Hz). ¹³C NMR (125 Hz, DMSO) δ = 165.6, 154.6, 140.9, 93.9, 86.5, 85.7, 62.8, 36.2.



Chiral HPLC of compound 9a

Chiral HPLC of compound 9b



Chiral HPLC of racemic compound 9



Chiral HPLC of compound 7a



Chiral HPLC of compound 7b



Chiral HPLC of racemic compound 7



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¹H NMR and ¹³C NMR of 9



Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013

¹H NMR and ¹³C NMR of 10



S6

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¹H NMR and ¹³C NMR of 1a





¹H NMR and ¹³C NMR of 7



¹H NMR and ¹³C NMR of 1b

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