

Electronic Supplementary Information

White-light emitting boronate microparticles for potential use as reusable bright chemosensors in water

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Materials

Boronate microparticles (**BP**) composed of polymeric 2,4,8,10-tetraoxa-3,9-diborospiro[5.5]undecane, that provide an available surface for dye grafting were prepared from benzene-1,4-diboronic acid and pentaerythritol according to previously reported procedures (Y. Matsushima, R. Nishiyabu, N. Takanashi, M. Haruta, H. Kimura and Y. Kubo, *J. Mater. Chem.*, 2012, **22**, 24124–24131), and their final diameters were $2.4 \pm 0.5 \mu\text{m}$ (Fig. a).

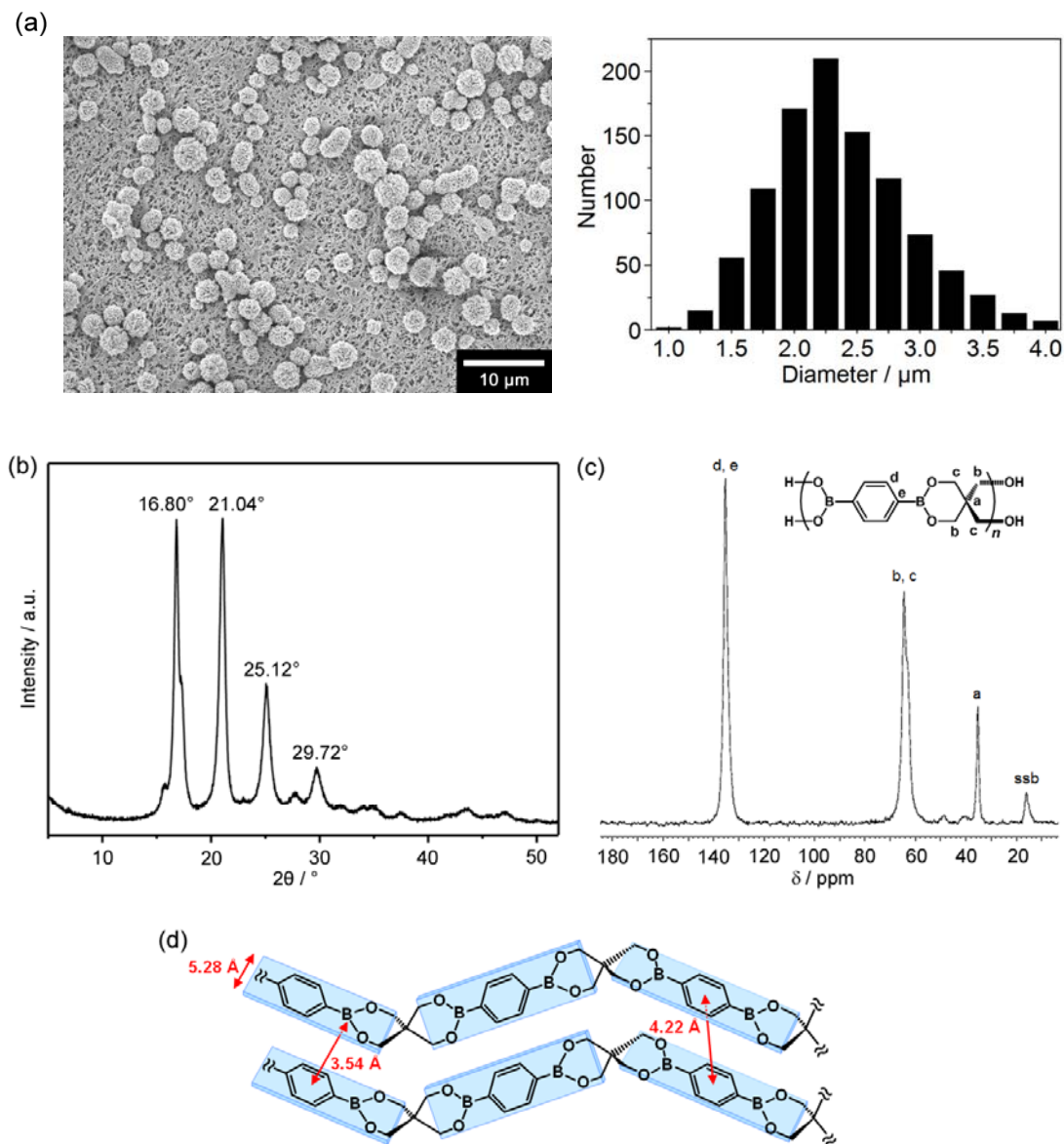


Fig. (a) The SEM image and the size distribution of boronate microparticles (**BP**), (b) PXRD pattern of **BP**, (c) ^{13}C CP-MAS NMR spectrum of **BP**, where ssb represents spinning side band and (d) plausible stacking structure of polymeric 2,4,8,10-tetraoxa-3,9-diborospiro[5.5]undecane in **BP**.

1-Pyreneboronic acid **1**, pyrene, lissaminerhodamine B sulfonyl chloride and 3-aminophenylboronic acid monohydrate were purchased from Tokyo Chemical Industry, Sigma-Aldrich, Acros Organics and Wako Chemicals, respectively. 3-[(*N*-Dansylamino)ethylamino]ethylamino)methyl]phenylboronic acid **2** was prepared previously; see, R. Nishiyabu, H. Kobayashi and Y. Kubo, *RSC Adv.*, 2012, **2**, 6555–6561.

General Experimental Procedure

Solution-state NMR spectra were recorded on a Bruker AVANCE-500 spectrometer using tetramethylsilane (TMS) as an internal standard (0 ppm) for ^1H and ^{13}C NMR analysis and $\text{BF}_3 \cdot \text{OEt}_2$ as an external standard (0 ppm) for ^{11}B NMR analysis. All solution-state NMR spectra were recorded at 298 K. Solid-state ^{13}C cross-polarization magic angle spinning (CP-MAS) NMR spectrum was measured by a JEOL JNM-ECA 500 spectrometer. The observation frequency was 125 MHz. The spectrometer is equipped with a 4 mm MAS probehead capable of producing an MAS speed of 15 kHz. Spectra were obtained by using a ^1H - ^{13}C CP contact time of 2 ms, an acquisition time of 40.7 ms, a recycle delay of 5 s between scans. The ^{13}C chemical shifts were calibrated by using adamantane ($\delta = 29.5$ ppm) as an external standard relative to tetramethylsilane (TMS; $\delta = 0$ ppm). The solid-state NMR spectra was recorded at 291 K. Fast atom bombardment (FAB) mass spectrum was obtained on a JEOL JMS-700 spectrometer where 3-nitrobenzyl alcohol was used as a matrix. Elemental analysis was performed on a Yanaco CHNcoder MT-5 analyzer. Field-emission scanning electron microscopy (FE-SEM) was performed on a JEOL JSM-7500F (acceleration voltage of 5 kV). For FE-SEM measurement, particles were coated with Au on an EIKO IB3ION COATER. Nitrogen gas adsorption isotherm was recorded by a Micrometrics Trister. The sample (20.8 mg) was degassed at 150 °C under vacuum for 5 h and the measurement was performed at 77 K. The surface area was calculated by BET method. Zeta potential measurement was performed using an OTSUKA ELECTRONICS ELSZ-2 instrument. UV-vis absorption spectra were measured on a Shimadzu UV-3600 spectrophotometer. Fluorescence spectra and quantum yields of dye-grafted **BP** were taken on using a JASCO FP-8500 spectrofluorometer with a substandard light source (JASCO ESC-846) and an integrating sphere. In addition, changes in fluorescence spectra upon adding metal ions were monitored by JASCO FP-6300Q spectrofluorometer. Quartz cell with 1-cm path length was used for UV-vis and fluorescence measurements. Fluorescence microscopic images were taken using a TS100 LED-F (λ_{ex} : 330–380 nm, λ_{em} : 420 nm~) equipped with a Plan APO 60X Oil objective lens. Photographed images of aqueous dispersions of particles were taken by a PENTAX *K-m* digital camera with a PENTAX DA 18-55mm lens (shutter speed: 0.25 s, f-number: 6.7) under UV irradiation using a handy lamp (365 nm).

Synthesis of 2-(3-diethylamino-6-diethylazaniumylidene-xanthen-9-yl)-5-[[3-(dihydroxyborylphenyl)amino]sulfonyl]-benzenesulfonate (**3**)

To lissaminerhodamine B sulfonyl chloride (150 mg, 0.26 mmol) in dry pyridine (5 mL) was added 3-aminophenylboronic acid monohydrate (40 mg, 0.26 mmol) under a N_2 atmosphere at room temperature. The mixture was stirred at room temperature for 24 h and then the solvent was removed *in vacuo*. The crude product was chromatographed on silica gel (wako gel C-300) using MeOH (3% (v/v)) in CHCl_3 as an eluent. In this way, 30 mg of **3** was obtained as a purple solid (17% yield). UV-vis absorption: $\lambda_{\text{max}} = 563$ nm ($\varepsilon = 1.1 \times 10^5 \text{ M}^{-1} \text{ cm}^{-1}$) in DMSO; Fluorescence: $\lambda_{\text{em}} = 589$ nm ($\lambda_{\text{ex}} = 520$ nm) in DMSO; ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 10.33 (*s*, 1H, NH), 8.44 (*d*, $J = 1.8$ Hz, 1H, Ar-H), 8.07 (*s*, 2H, B(OH) $_2$), 7.73 (*dd*, $J = 8.0$ Hz, 1.9 Hz, 1H, Ar-H), 7.54 (*d*, $J = 6.9$ Hz, 1H, Ar-H), 7.48 (*s*, 1H, Ar-H), 7.36 (*d*, $J = 8.0$ Hz, 1H, Ar-H), 7.28 (*dd*, $J = 7.6$ Hz, 7.6 Hz, 1H, Ar-H), 7.21 (*d*, $J = 8.2$ Hz, 1H, Ar-H), 7.02 (*dd*, $J = 9.6$ Hz, 2.4 Hz, 2H, Ar-H), 6.91 (*d*, $J = 2.4$ Hz, 2H, Ar-H), 6.85 (*d*, $J = 9.6$ Hz, 2H, Ar-H), 3.64 (*q*, $J = 8.0$ Hz, 8H, CH_2), 1.20 (*t*, $J = 7.0$ Hz, 12H, CH_3); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ 157.2, 157.1, 155.0, 148.0, 140.7, 136.6, 135.6, 133.4, 132.5, 130.6, 130.4, 128.2, 127.6, 126.7, 125.9, 123.4, 113.7, 113.4, 95.4, 45.3, 12.5; ^{11}B NMR (160 MHz, $\text{DMSO}-d_6$): δ 30.2; FABMS: m/z 947 ($[\text{M} + 2(3\text{-nitrobenzyl alcohol}) - 2\text{H}_2\text{O}]^+$); elemental analysis: anal. calcd for $\text{C}_{33}\text{H}_{36}\text{BN}_3\text{O}_8\text{S}_2 \cdot 0.1\text{CHCl}_3$: C 57.66; H 5.28; N 6.09, found: C 57.67; H 5.03; N 5.72%.

Surface functionalization of BP with fluorescent dyes

Methanol solution of fluorescent dye (1.0×10^{-5} M, 10 mL) was added to a vial where **BP** (10.0 mg) was present and was allowed to stand for 24 h at room temperature (25 °C). The resultant solid was collected by filtration and rinsed with methanol (10 mL), then dried *in vacuo* for 1 h.

Quantitative evaluation of dye grafting onto the surface of BP using Langmuir isotherm model

Methanol solutions of dye **1** at varied concentration ($0 - 2.5 \times 10^{-3}$ M, 5 mL) were added to vials where **BP** (5.0 mg) was present. The resultant suspensions were allowed to stand for 24 h at room temperature (25 °C) and then centrifuged with 4500 rpm. UV-vis absorption spectra of the supernatant solutions were measured to determine equilibrium concentrations and amounts of dye **1** grafted. The obtained data were analyzed by the Langmuir equation expressed in the following equation.

$$q = \frac{q_{\max} \times K \times C}{1 + K \times C}$$

where q , q_{\max} , K and C are adsorbed amount, maximum adsorption capacity, Langmuir adsorption constant and equilibrium concentration, respectively. The values of q_{\max} and K were evaluated by a nonlinear least-squares method.

Preparation of white-light emissive BP

A methanol solution (10 mL) of **1** (3.8×10^{-6} M), **2** (6.0×10^{-6} M) and **3** (2.0×10^{-7} M) was added to a vial where **BP** (10.0 mg) was present. The resultant suspension was allowed to stand for 24 h at room temperature (25 °C). The solid was collected by filtration and rinsed with methanol (10 mL). The obtained solid was dried *in vacuo* for 1 h and was used for further experiments.

Fluorescence measurements for the detection of Cu²⁺

For the measurements, nitrate salts of metal ions (Na⁺, K⁺, Mg²⁺, Ca²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Ag⁺, Zn²⁺, Cd²⁺, Hg²⁺, Al³⁺, and Pb²⁺) were used. Each aqueous solution of the metal ion was added to aqueous dispersions of **BP** ($1.0 \text{ mg} \cdot \text{mL}^{-1}$, 5 mM HEPES buffer, pH 7.0) at room temperature (25 °C). The resultant suspensions were measured by fluorometry. Association constant (K_a) for complexation of dansyldiethylenetriamine moiety with Cu²⁺ was determined from the titration curve of normalized changes in fluorescence intensities ($I - I_0$) / I_0 of **Sen-BP** at 510 nm upon the addition of incremental amounts of Cu²⁺. The normalized change in fluorescence intensity was expressed in the following equations.

$$(I - I_0) / I_0 = \frac{(\Delta I_{\text{lim}} / I_0) \times (A - \sqrt{A^2 - 4 \times K_a^2 \times S_0 \times M_0})}{2 \times K_a \times S_0}$$

$$A = K_a \times M_0 + K_a \times S_0 + 1$$

where I , I_0 and ΔI_{lim} are the observed fluorescence intensity, the fluorescence intensity in the absence of metal ion, and the saturation value of change in fluorescence intensity of **Sen-BP**, respectively. In the equation, K , S_0 and M_0 are the binding constant, the initial concentrations of dansyldiethylenetriamine moiety and Cu²⁺ added, respectively. The values of K_a , S_0 and $\Delta I_{\text{lim}} / I_0$ were determined to be $(5.9 \pm 0.6) \times 10^6 \text{ M}^{-1}$, $(9.6 \pm 0.4) \times 10^{-7} \text{ M}$ and -0.73 ± 0.01 by a nonlinear least-squares method, respectively.

Determination of the detection limit for Cu^{2+}

The detection limit for Cu^{2+} was determined from the point of intersection between the regression line obtained from fluorescence intensities (I) of **Sen-BP** at 510 nm in dynamic range of the titration curve and the line given by following equation.

$$Y = I_{\text{avg}} - 3 \times \sigma$$

where I_{avg} and σ are the average value and the standard deviation of fluorescence intensity of **Sen-BP** at 510 nm in the absence of metal ion measured by five times, respectively.

Reusability test of **Sen-BP** for Cu^{2+} detection

Initially, fluorescence spectrum of aqueous dispersion of **Sen-BP** ($1 \text{ mg} \cdot \text{mL}^{-1}$) in the presence of Cu^{2+} ($3.0 \times 10^{-5} \text{ M}$) was measured in 5 mM HEPES buffer (pH 7.0). After that aqueous EDTA ($5.0 \times 10^{-2} \text{ M}$) in 5 mM HEPES buffer (pH 7.0) was added to the dispersion at room temperature ($25 \text{ }^\circ\text{C}$). The resultant dispersion containing $1.0 \times 10^{-3} \text{ M}$ EDTA was allowed to stand for 5 min at room temperature ($25 \text{ }^\circ\text{C}$). Then, **Sen-BP** was collected by filtration, rinsed with water and dried *in vacuo* for 1 h. Such are cycled **Sen-BP** was again dispersed in 5 mM HEPES buffer (pH 7.0).

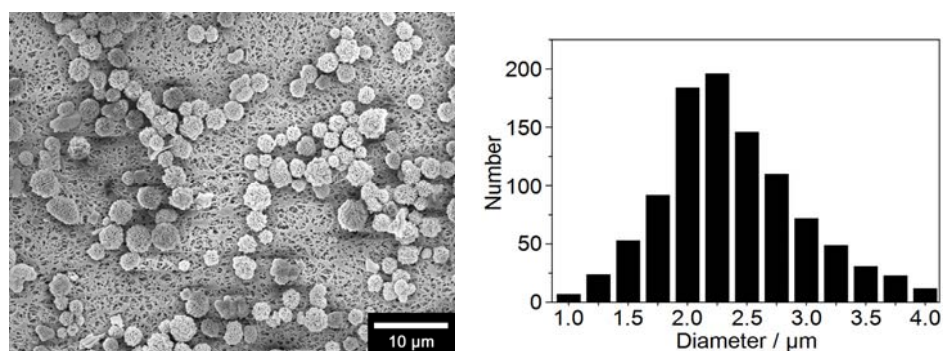


Fig. S1 SEM image and the size distribution of **BP** immersed in a MeOH solution of dye **1**.

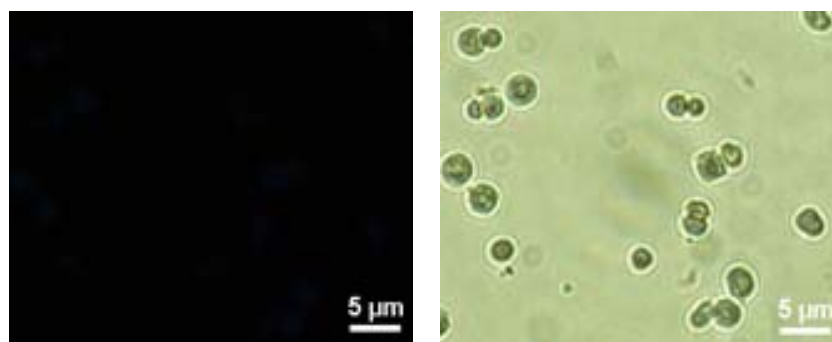


Fig. S2 Fluorescence microscopic image (left) and the corresponding optical microscopic image (right) of aqueous dispersion of **BP** (5 mM HEPES buffer, pH 7.0) after immersion in a MeOH solution of pyrene.

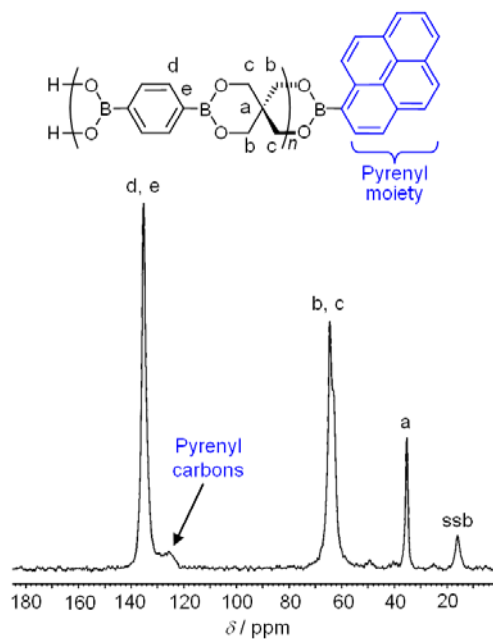


Fig. S3 ¹³C-CP-MAS NMR spectrum of **1-BP**, where ssb represents spinning side band.

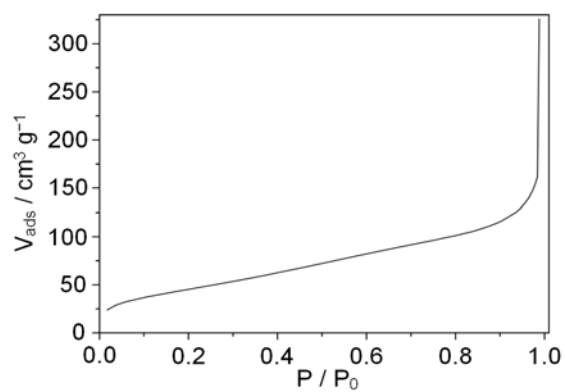


Fig. S4 Nitrogen adsorption isotherm of **BP**.

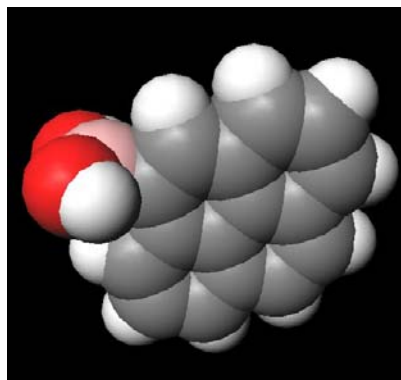


Fig. S5 Optimized structure of **1** using DFT calculation (B3LYP/6-31G*).

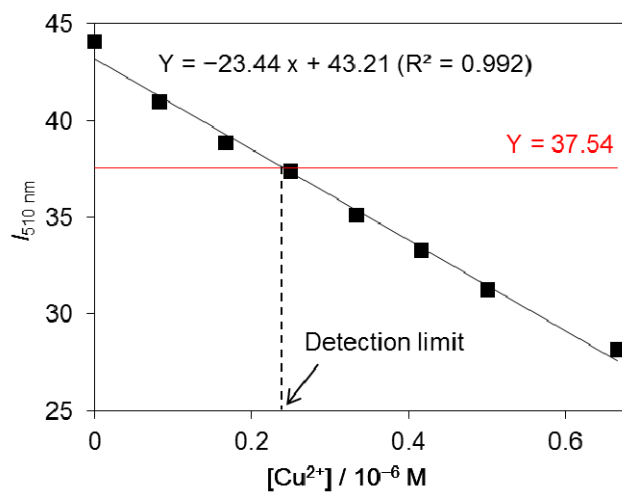


Fig. S6 Determination of the detection limit for Cu²⁺.