

Supporting Information

For

Copper-catalyzed oxidation of arene-fused cyclic amines to cyclic imides

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Content

General Considerations	S2
Experimental Procedures	S3
Proposed mechanism	S7
References	S8
X-ray structure of 4a	S9
Copies of ¹ H and ¹³ C NMR Spectra	S10

General Considerations

All manipulations were conducted in sealed tube under an atmosphere of air. Unless otherwise noted, all starting materials were commercially available and were used without further purification. Isoindolines **1a-1k** were prepared by reaction of 1,2-bis(bromomethyl)benzene with amine in the presence of base.¹⁻³ Compound **II** was prepared by reaction of 1,8-bis(bromomethyl)naphthalene with methanamine. N,N-bispropargylamine was prepared by A³ coupling.⁴ ¹H NMR and ¹³C NMR spectra were recorded on 300 MHz and 400 MHz NMR spectrometer with TMS as internal standard. Mass spectra were obtained using a Bruker Esquire ion trap mass spectrometer in positive ion mode. Flash column chromatography was performed on silica gel and Al₂O₃ (200-300 mesh).

Experimental Procedures

General procedure for copper-catalyzed oxidation of isoindoline

To a 25 mL tube with teflon-coated magnetic stir bar and a rubber septum, was added isoindoline **1** (0.4 mmol), CuCl (0.04 mmol), CH₂Cl₂ 1 mL, TBHP (0.55 mL, 4 mmol, 70% in water). The tube was sealed and stirred for 24 h at 50 °C. The reaction mixture was quenched by water and extracted by ethyl acetate. The organic extract was dried over MgSO₄. Removing the solvent and subsequent purification by column chromatography on Al₂O₃ afforded **2**.

Procedure for copper-catalyzed oxidation of isoindoline on 5 mmol scale

To a 100 mL tube with teflon-coated magnetic stir bar and a rubber septum, was added isoindoline **1** (5 mmol), CuCl (0.5 mmol), CH₂Cl₂ 13 mL, TBHP (6.9 mL, 50 mmol, 70% in water). The tube was sealed and stirred for 24 h at 50 °C. The reaction mixture was quenched by water and extracted by ethyl acetate. The organic extract was dried over MgSO₄. Removing the solvent and subsequent purification by column chromatography on Al₂O₃ afforded **2a** (589 mg).

Procedure for copper-catalyzed oxidation of isoindoline on 10 mmol scale

To a 100 mL tube with teflon-coated magnetic stir bar and a rubber septum, was added isoindoline **1e** or **1g** (10 mmol), CuCl (1 mmol), CH₂Cl₂ 25 mL, TBHP (13.7 mL, 100 mmol, 70% in water). The tube was sealed and stirred for 24 h at 50 °C. The reaction mixture was quenched by water and extracted by ethyl acetate. The organic extract was dried over MgSO₄. Removing the solvent and subsequent purification by column chromatography on Al₂O₃ afforded **2e** (995 mg) or **2g** (825 mg).

2-Butylisoindoline-1,3-dione, 2a⁵

68% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.84-7.80 (m, 2H), 7.71-7.67 (m, 2H), 3.67 (t, *J* =, 2H), 1.68-1.60 (m, 2H), 1.40-1.31 (m, 2H), 0.93 (t, *J* =, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.5, 133.9, 132.3, 123.2, 37.9, 30.7, 20.2, 13.7. GC-MS: 203.

2-Octylisoindoline-1,3-dione, 2b⁶

73% isolated yield. ¹H NMR (300 MHz, CDCl₃) δ 7.84-7.80 (m, 2H), 7.72-7.68

(m, 2H), 3.66 (t, $J = 7.2$ Hz, 2H), 1.70-1.62 (m, 2H), 1.43-1.24 (m, 10H), 0.85 (t, $J = 6.4$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 168.5, 133.9, 132.3, 123.2, 38.2, 31.9, 29.2, 28.7, 27.0, 22.7, 14.2. GC-MS: 259.

2-(*Tert*-butyl)isoindoline-1,3-dione, 2c⁷

71% isolated yield. ^1H NMR (400 MHz, CDCl_3) δ 7.77-7.74 (m, 2H), 7.69-7.65 (m, 2H), 1.69 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 169.8, 133.7, 132.2, 122.7, 57.9, 29.2. GC-MS: 203.

2-Allylisoindoline-1,3-dione, 2d⁸

45% isolated yield. ^1H NMR (400 MHz, CDCl_3) δ 7.86-7.83 (m, 2H), 7.73-7.69 (m, 2H), 5.93-5.83 (m, 1H), 5.27-5.17 (m, 2H), 4.30-4.27 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.0, 134.0, 132.2, 131.6, 123.4, 117.8, 40.1. GC-MS: 187.

2-Benzylisoindoline-1,3-dione, 2e⁵

48% isolated yield. ^1H NMR (300 MHz, CDCl_3) δ 7.85-7.80 (m, 2H), 7.71-7.67 (m, 2H), 7.45-7.25 (m, 5H), 4.84 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 168.1, 136.5, 134.0, 132.2, 128.8, 128.7, 127.9, 123.4, 41.7. GC-MS: 237.

Methyl 2-(1,3-dioxoisoindolin-2-yl)acetate, 2f⁷

40% isolated yield. ^1H NMR (300 MHz, CDCl_3) δ 7.90-7.85 (m, 2H), 7.77-7.73 (m, 2H), 4.45 (s, 2H), 3.76 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 167.8, 167.5, 134.3, 132.1, 123.7, 52.8, 38.9. GC-MS: 219.

2-Phenylisoindoline-1,3-dione, 2g⁵

42% isolated yield. ^1H NMR (400 MHz, CDCl_3) δ 7.98-7.94 (m, 2H), 7.81-7.77 (m, 2H), 7.53-7.39 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 167.4, 134.5, 131.9, 131.8, 129.2, 128.2, 126.7, 123.8. GC-MS: 223.

2-(4-Chlorophenyl)isoindoline-1,3-dione, 2h⁵

47% isolated yield. ^1H NMR (300 MHz, CDCl_3) δ 7.98-7.93 (m, 2H), 7.83-7.78 (m, 2H), 7.52-7.46 (m, 2H), 7.44-7.38 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 167.1, 134.7, 133.9, 131.7, 130.3, 129.4, 127.8, 124.0. GC-MS: 257.

2-(4-Isopropylphenyl)isoindoline-1,3-dione, 2i

30% isolated yield. ^1H NMR (400 MHz, CDCl_3) δ 7.96-7.94 (m, 2H), 7.80-7.77 (m, 2H), 7.38-7.33 (m, 4H), 3.00-2.94 (m, 1H), 1.29 (d, $J = 6.9$ Hz, 6H); ^{13}C NMR

(100 MHz, CDCl₃) δ 167.5, 149.0, 134.4, 131.9, 129.3, 127.3, 126.5, 123.9, 34.0, 24.0. GC-MS: 265.

2-(Naphthalen-2-yl)isoindoline-1,3-dione 2j⁹

57% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.99-7.94(m, 4H), 7.90-7.88 (m, 2H), 7.82-7.79 (m, 5H), ¹³C NMR (100 MHz, CDCl₃) δ 167.5, 134.5, 133.4, 132.7, 129.2, 129.1, 128.3, 127.8, 126.8, 126.7, 125.6, 124.3, 123.9. GC-MS: 273.

2-Butyl-5-chloroisoindoline-1,3-dione, 2k

48% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 7.79-7.75 (m, 2H), 7.65 (q, *J* = 7.8 Hz, 1.8 Hz, 1H), 3.67 (t, *J* = 7.3 Hz, 2H), 1.68-1.60 (m, 2H), 1.40-1.30 (m, 2H), 0.93 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.5, 167.2, 140.6, 133.9, 130.3, 124.4, 123.7, 38.1, 30.6, 20.1, 13.6. GC-MS: 237.

2-Methyl-1H-benzo[de]isoquinoline-1,3(2H)-dione, 2l¹⁰

75% isolated yield. ¹H NMR (300 MHz, CDCl₃) δ 8.50 (d, *J* = 7.2 Hz, 2H), 8.13 (d, *J* = 8.3 Hz, 2H), 7.68 (dd, *J* = 8.3, 7.2 Hz, 2H), 3.50 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 164.4, 133.9, 131.5, 131.2, 128.0, 126.9, 122.5, 27.0. GC-MS: 211.

General procedure for preparation of TPD

To a solution of Cp₂ZrCl₂ (702 mg, 2 mmol) in 10 mL of THF, BuLi (1.6 M in hexane, 4.8 mmol), was added at -78 °C and the mixture was stirred for 1 h at the same temperature. To this solution, N,N-bis(3-phenylprop-2-yn-1-yl)butan-1-amine (2.0 mmol) was added and stirred for 3 h at room temperature. Then, S₂Cl₂ (3 mmol) was added and stirred overnight. The reaction mixture was quenched by water and extracted by ethyl acetate. The organic extract was dried over MgSO₄. Removing the solvent and subsequent purification by column chromatography on silica gel afforded **3a** in 64% isolated yield.¹¹ **3b** was prepared in 57% isolated yield by using similar procedure with N,N-bis(3-(thiophen-2-yl)prop-2-yn-1-yl)butan-1-amine as starting material.

To a 25 mL tube with teflon-coated magnetic stir bar and a rubber septum, was added **3** (0.4 mmol), CuCl (0.04 mol), CH₂Cl₂ 1 mL, TBHP (4 mmol, 70% in water). The tube was sealed and stirred for 24 h at 50°C. The reaction mixture was quenched by water and extracted by ethyl acetate. The organic extract was dried over MgSO₄.

Removing the solvent and subsequent purification by column chromatography on Al_2O_3 afforded **4**.

5-Butyl-1,3-diphenyl-4H-thieno[3,4-c]pyrrole-4,6(5H)-dione, 4a.

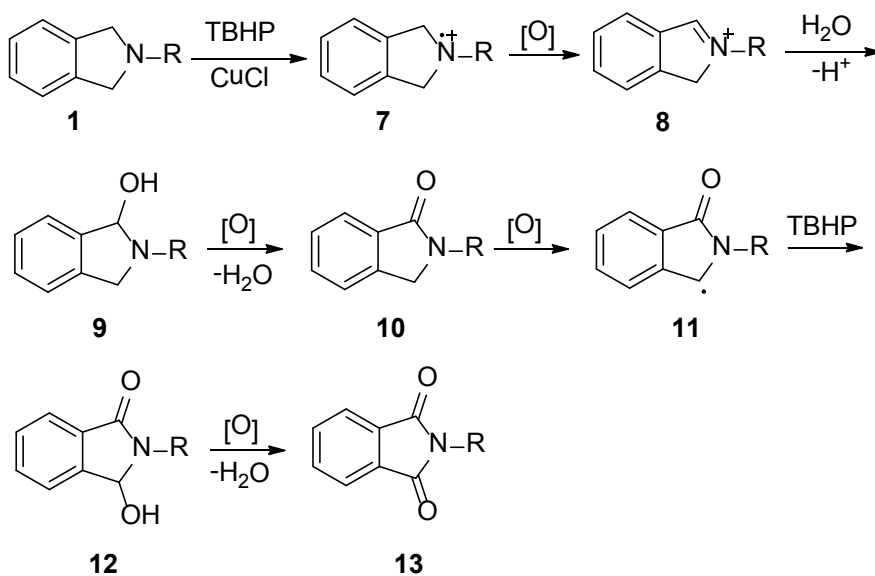
82% isolated yield. ^1H NMR (400 MHz, CDCl_3) δ 8.14-8.12 (m, 4H), 7.50-7.41 (m, 6H), 3.68 (t, $J = 7.3$ Hz, 2H), 1.71-1.63 (m, 2H), 1.42-1.36 (m, 2H), 0.95 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.1, 145.1, 130.7, 130.5, 130.2, 129.1, 128.2, 38.4, 30.6, 20.3, 13.8. HRMS calcd for $\text{C}_{22}\text{H}_{19}\text{NO}_2\text{S}$ 361.1136, found 361.1131.

5-Octyl-1,3-di(thiophen-2-yl)-4H-thieno[3,4-c]pyrrole-4,6(5H)-dione, 4b.

55% isolated yield. ^1H NMR (300 MHz, CDCl_3) δ 8.01 (d, $J = 3.8$ Hz, 2H), 7.43 (d, $J = 5.2$ Hz, 2H), 7.14-7.10 (m, 2H), 3.66 (t, $J = 7.2$ Hz, 2H), 1.72-1.63 (m, 2H), 1.42-1.20 (m, 10H), 0.87 (t, $J = 6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 162.7, 136.6, 132.5, 130.0, 128.7, 128.5, 38.7, 31.9, 29.3, 29.2, 28.6, 27.0, 22.7, 14.2. HRMS calcd for $\text{C}_{22}\text{H}_{23}\text{NO}_2\text{S}_3$ 429.0891, found 429.0893.

Proposed mechanism

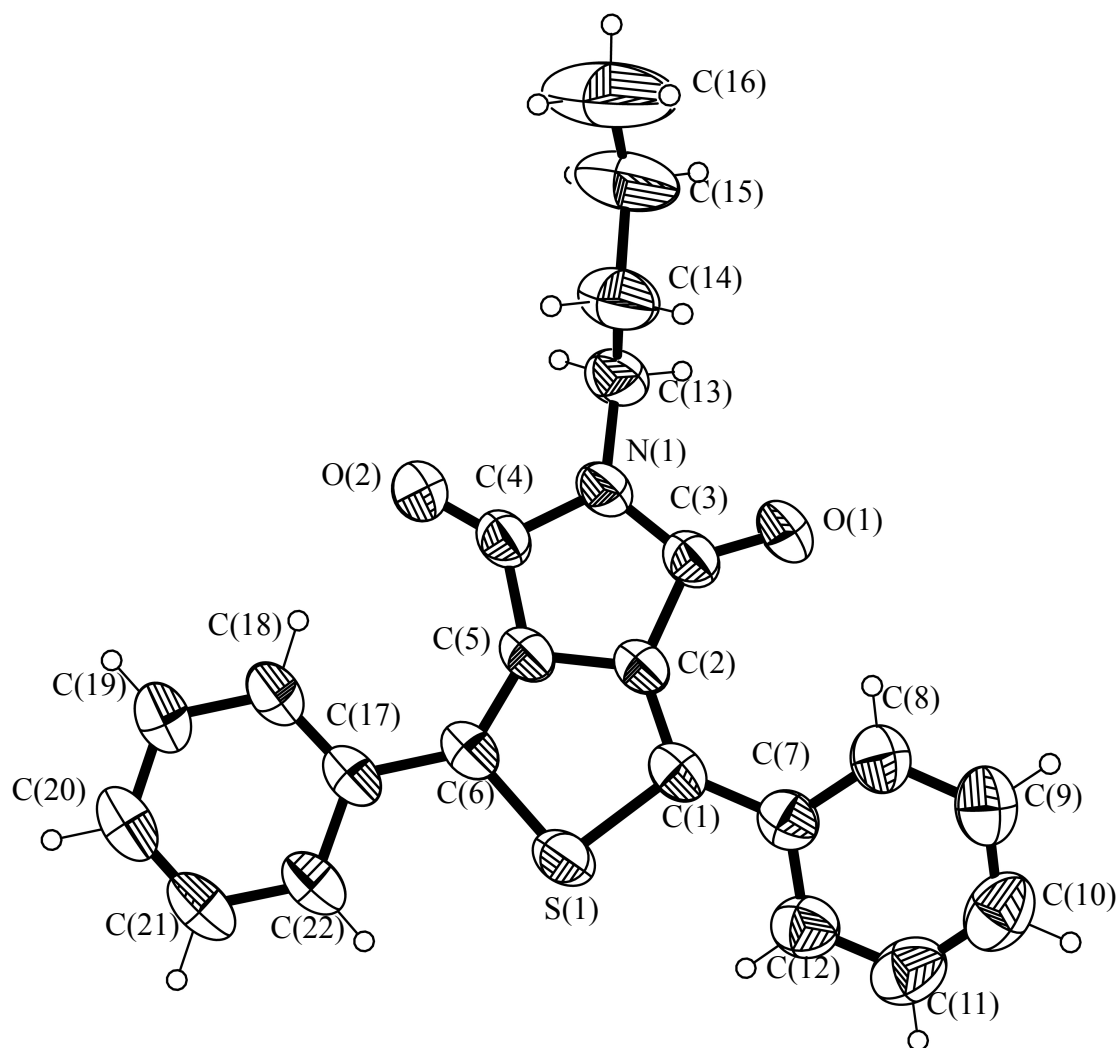
Based on the results, a plausible mechanism was proposed as follow. Firstly, amine **1** was oxidized to radical cation **7** in the present of TBHP and CuCl. **7** was further oxidized to iminium **8**. **8** reacted with water to yield compound **9**, which was then oxidized to amide **10**. **10** was further oxidized to form **11**, which reacted with TBHP to give **12**. Oxidation of **12** afforded imide **13**.



References

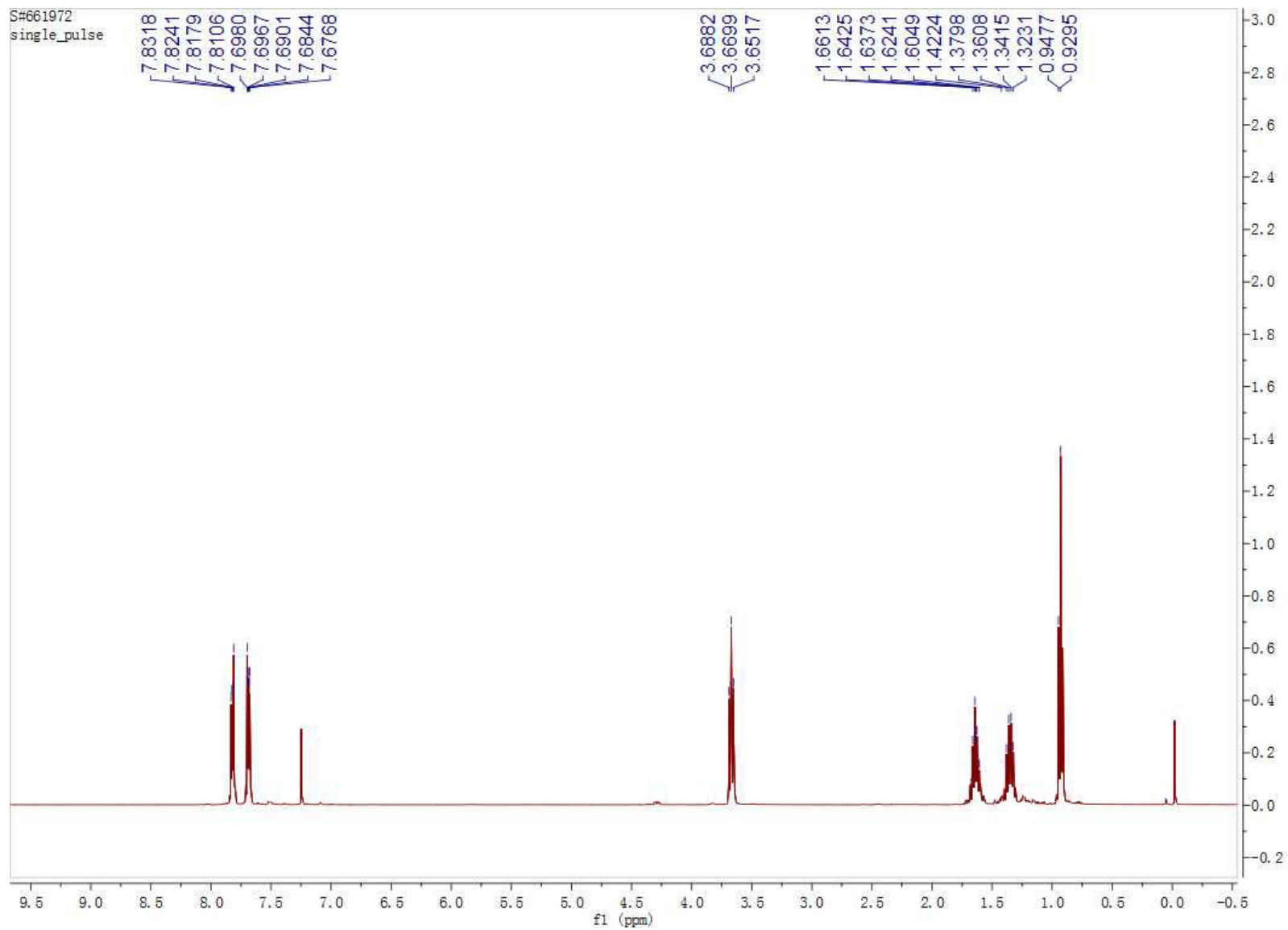
1. D. Hou, M. Wang, M. Chung, Y. Hsieh and H. Gavin, *J. Org. Chem.*, 2007, **72**, 9231.
2. F. N. Murigi, G. S. Nichol and E. A. Mash, *J. Org. Chem.*, 2010, **75**, 1293.
3. A. Subbarayappa and P. U. Patoliya, *Indian J. Chem. B*, 2009, **48B**, 545
4. N. Uhlig and C. Li, *Org. Lett.*, 2012, **14**, 3000.
5. J. Hsieh and C. Cheng, *Chem. Commun.*, 2005, 4554.
6. K. Asano and S. Matsubara, *Heterocycles*, 2010, **80**, 989.
7. D. Marosvoelgyi-Hasko, A. Petz, A. Takacs and L. Kollar, *Tetrahedron*, 2011, **67**, 9122.
8. V. Pace, P. Hoyos, M. Fernandez, J. V. Sinisterra and A. R. Alcantara, *Green Chem*, 2010, **12**, 1380.
9. H. J. Kim, J. Kim, S. H. Cho and S. Chang, *J. Am. Chem. Soc.*, 2011, **133**, 16382.
10. A. Takacs, A. Petz and L. Kollar, *Tetrahedron*, 2010, **66**, 4479.
11. P. J. Fagan and W. A. Nugent, *J. Am. Chem. Soc.*, 1988, **110**, 2310

X-ray structure of **4a**

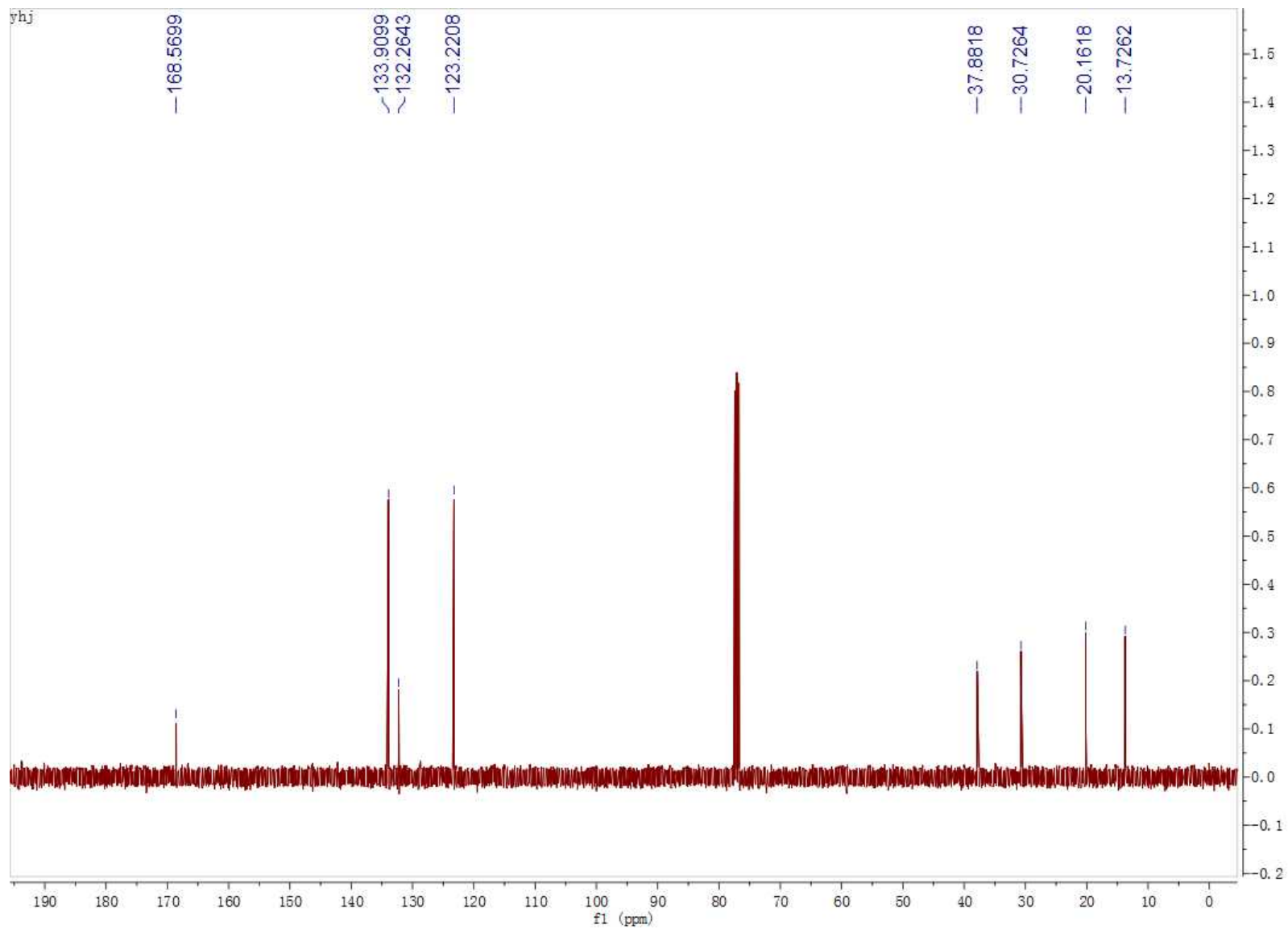


ORTEP drawing of $C_{22}H_{19}NO_2S$ with 35% probability ellipsoids, showing the atomic numbering scheme.

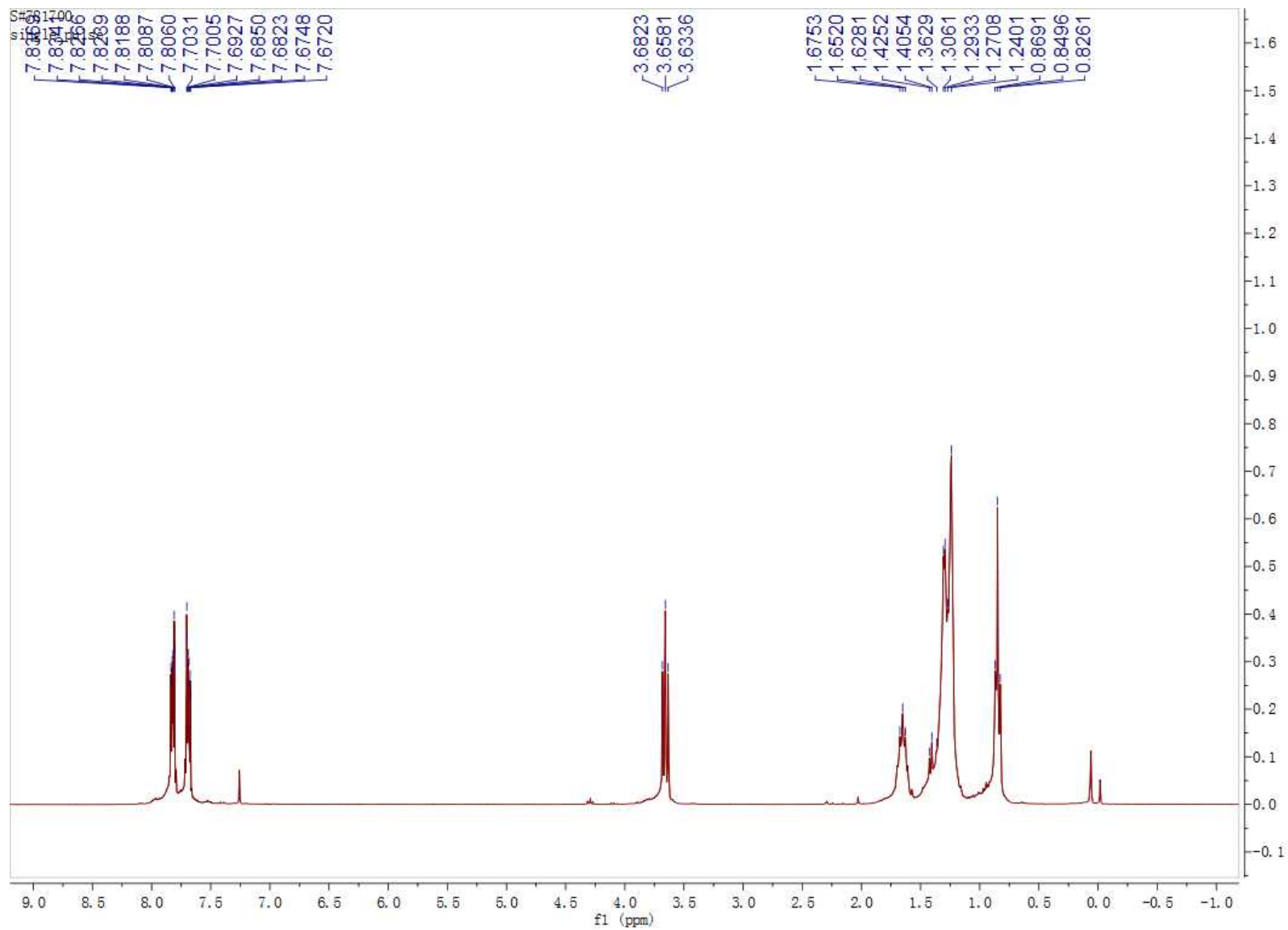
Copies of ^1H and ^{13}C NMR Spectra



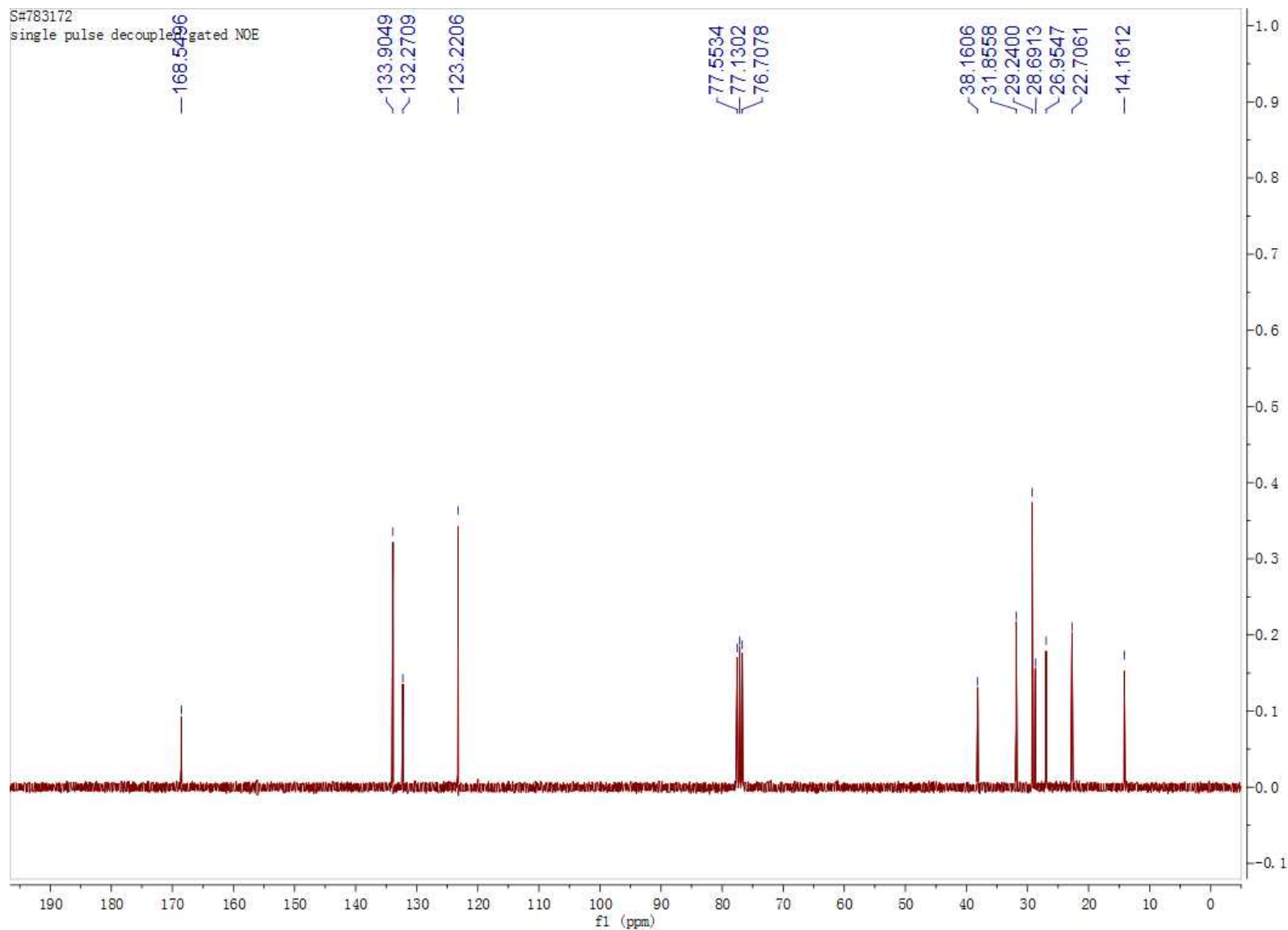
^1H NMR for compound **2a**



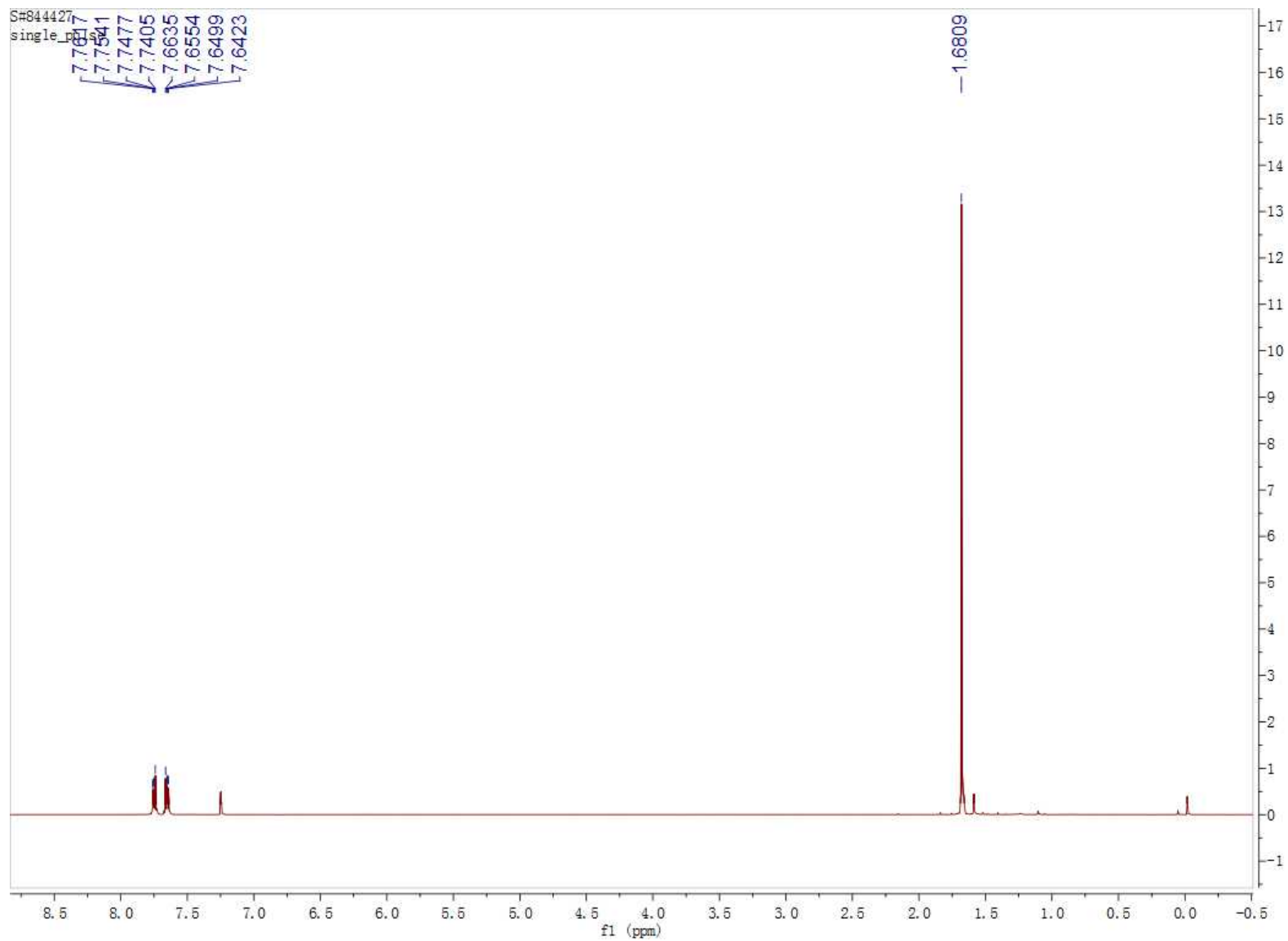
¹³C NMR for compound 2a



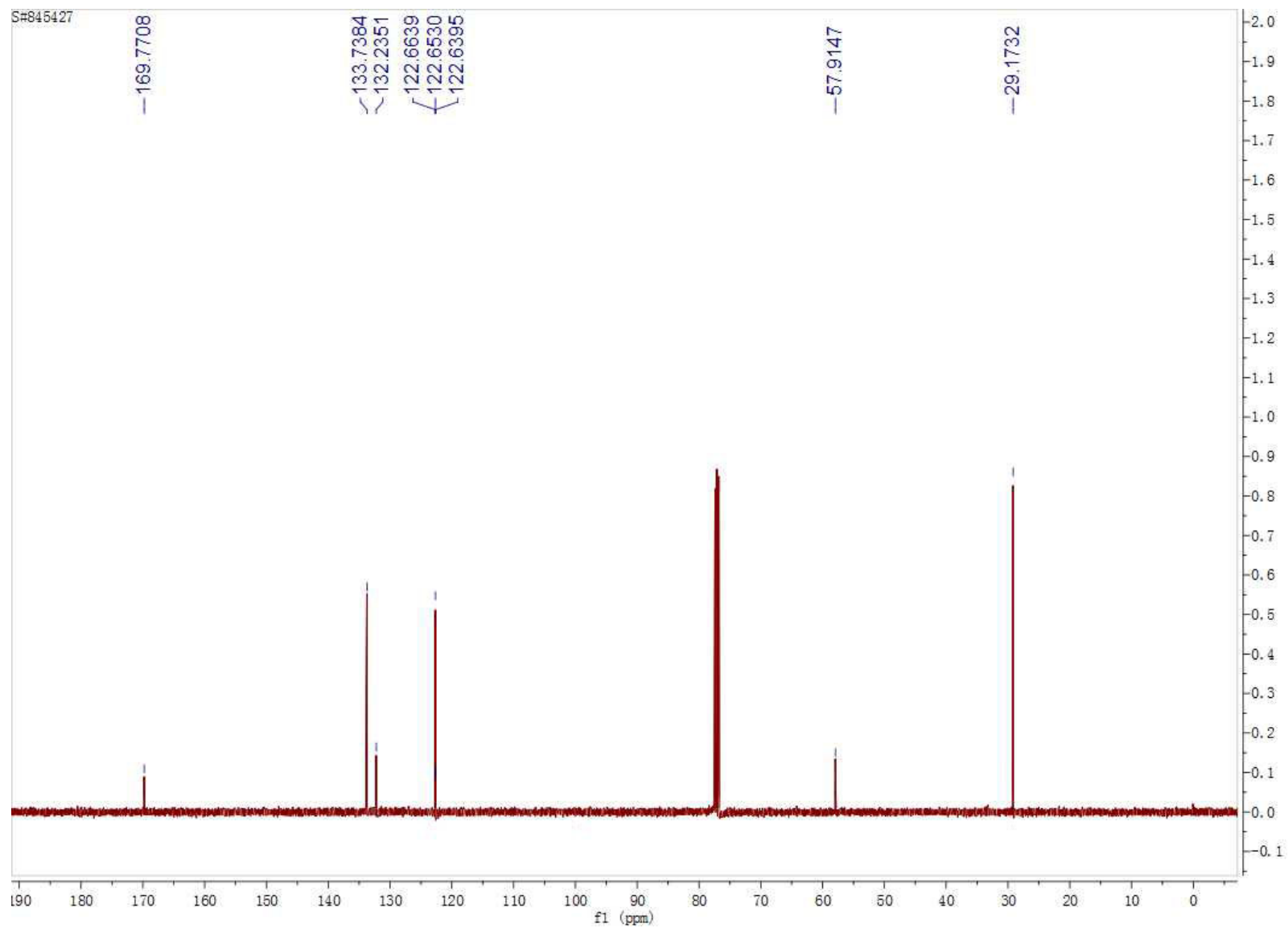
^1H NMR for compound **2b**



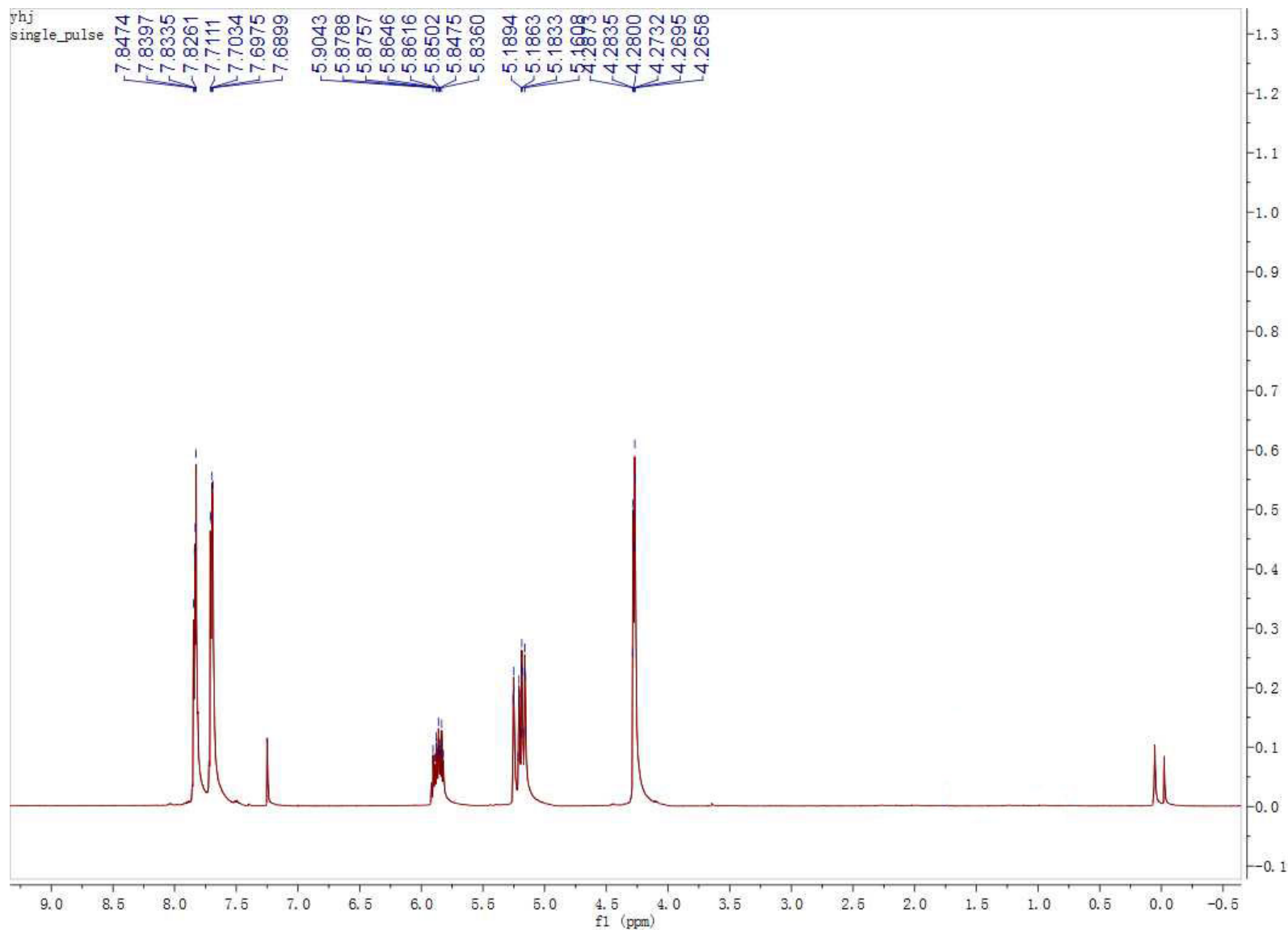
^{13}C NMR for compound **2b**



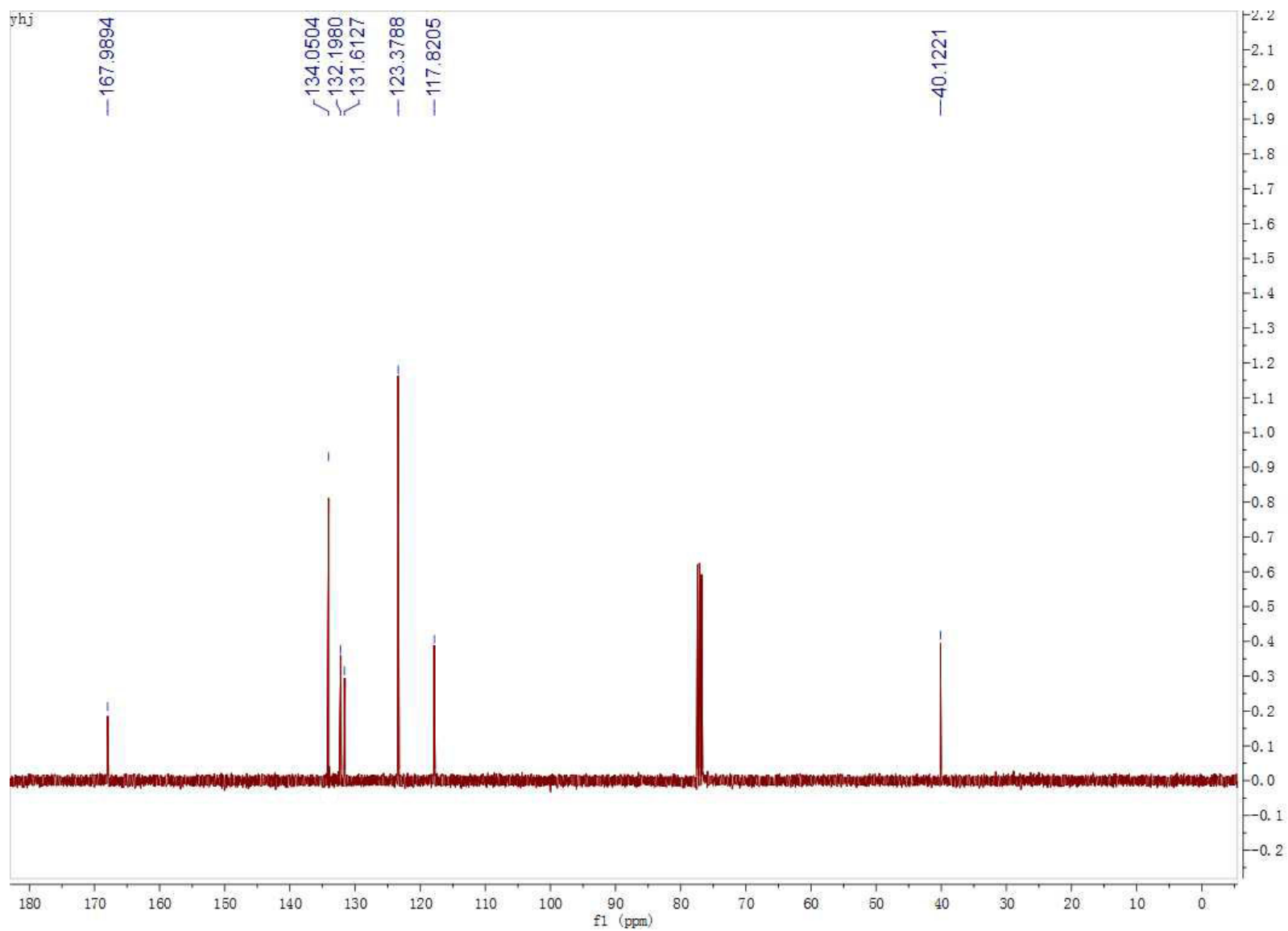
^1H NMR for compound **2c**



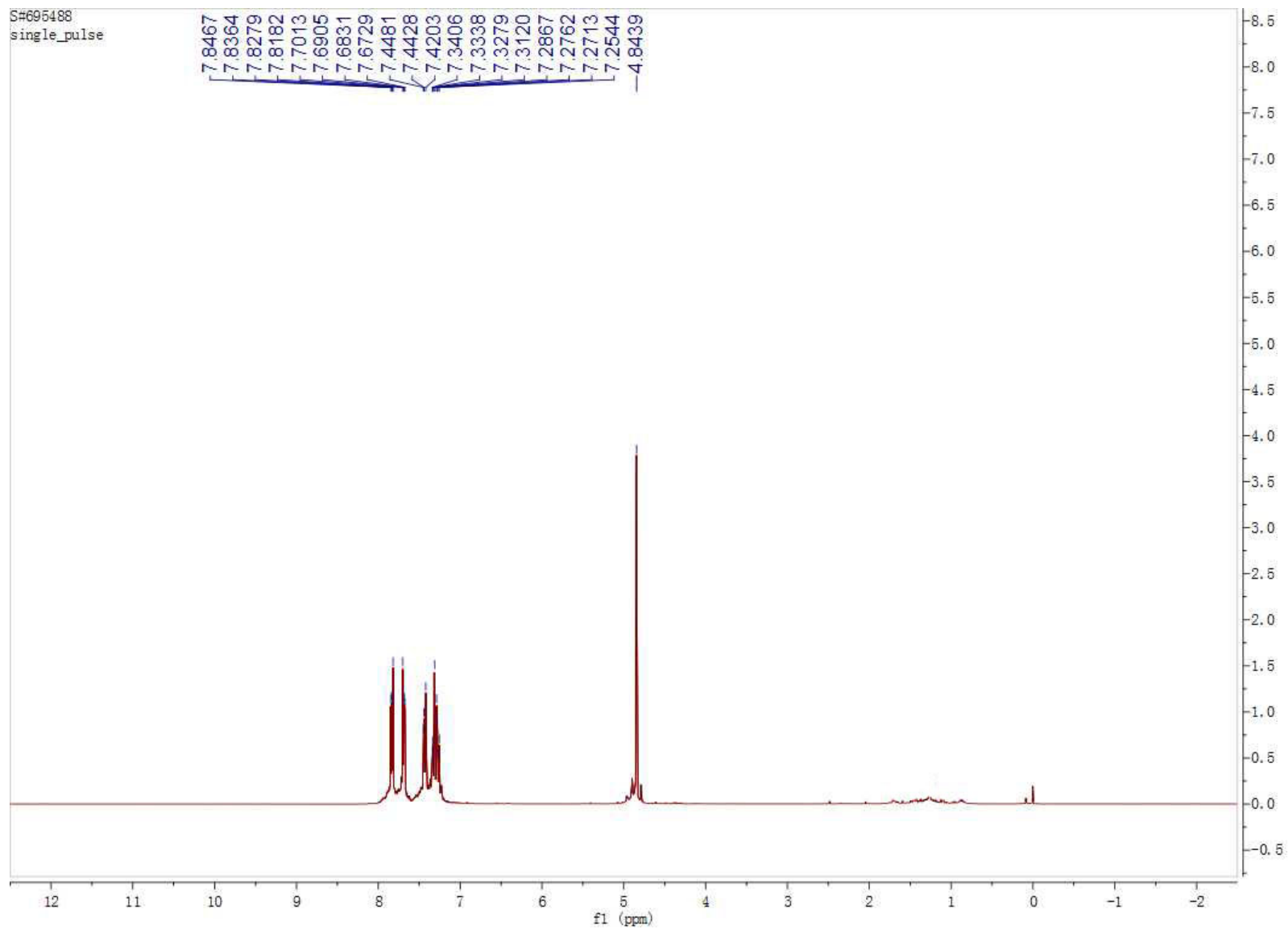
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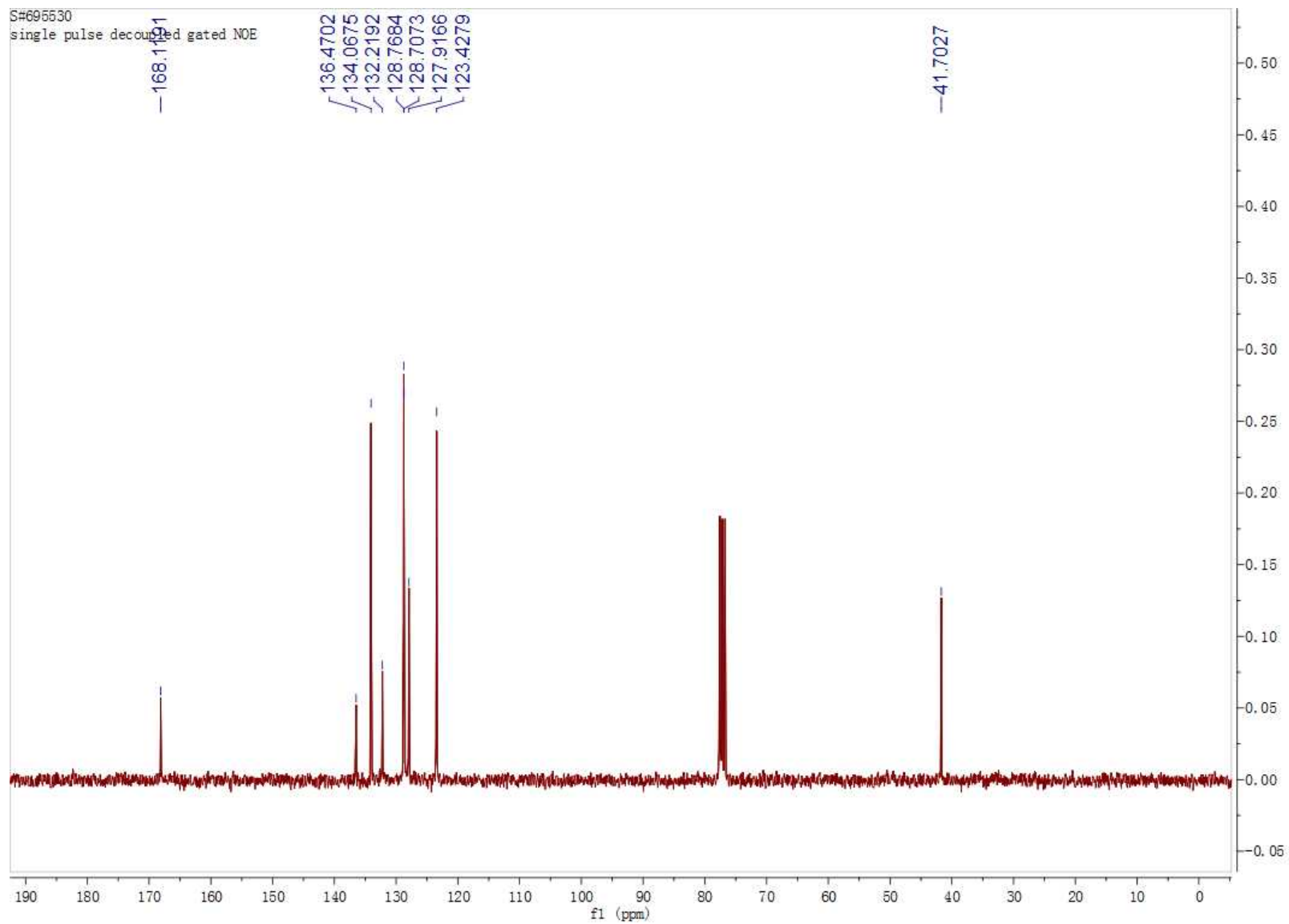
¹H NMR for compound 2d



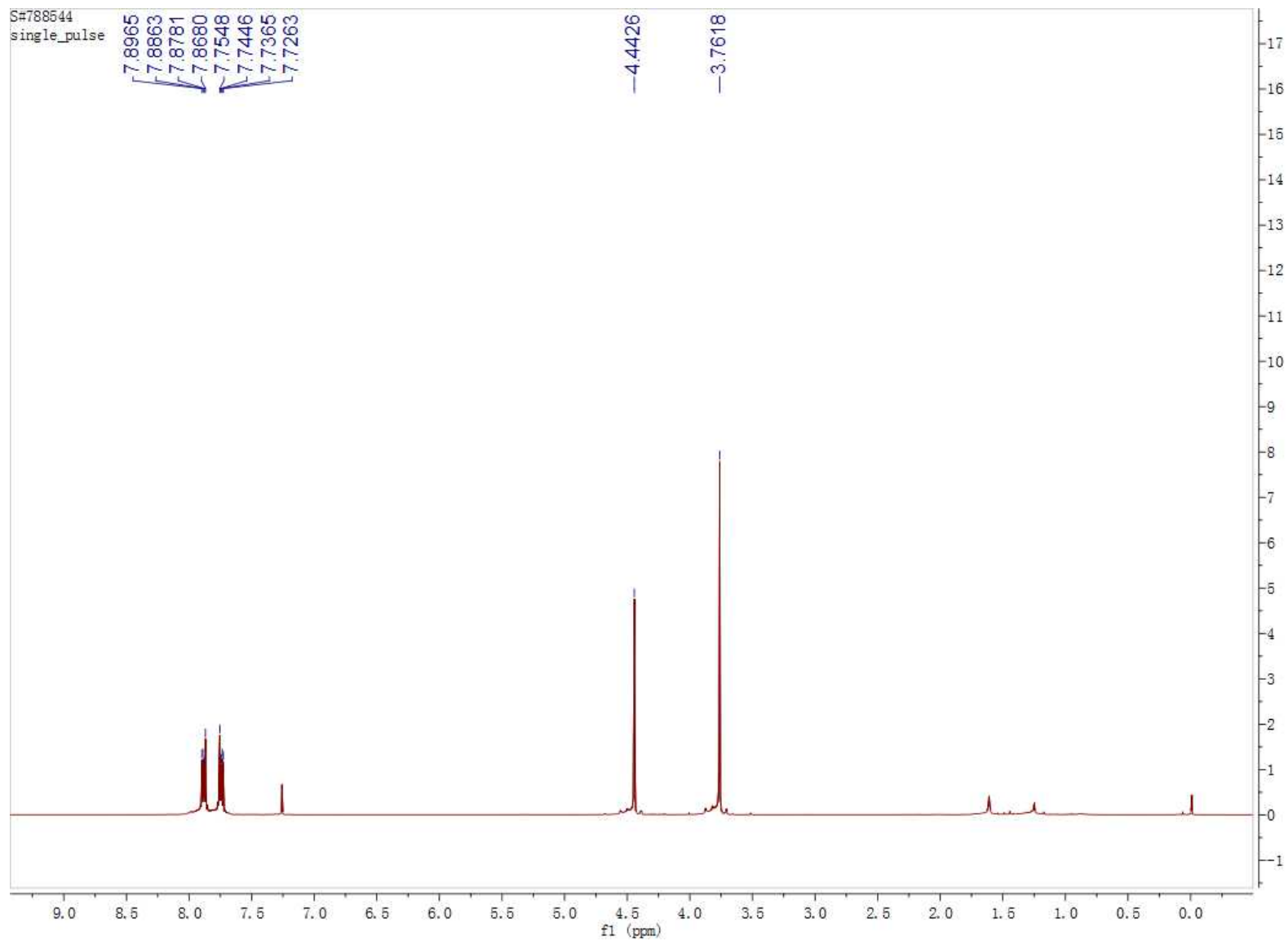
^{13}C NMR for compound **2d**



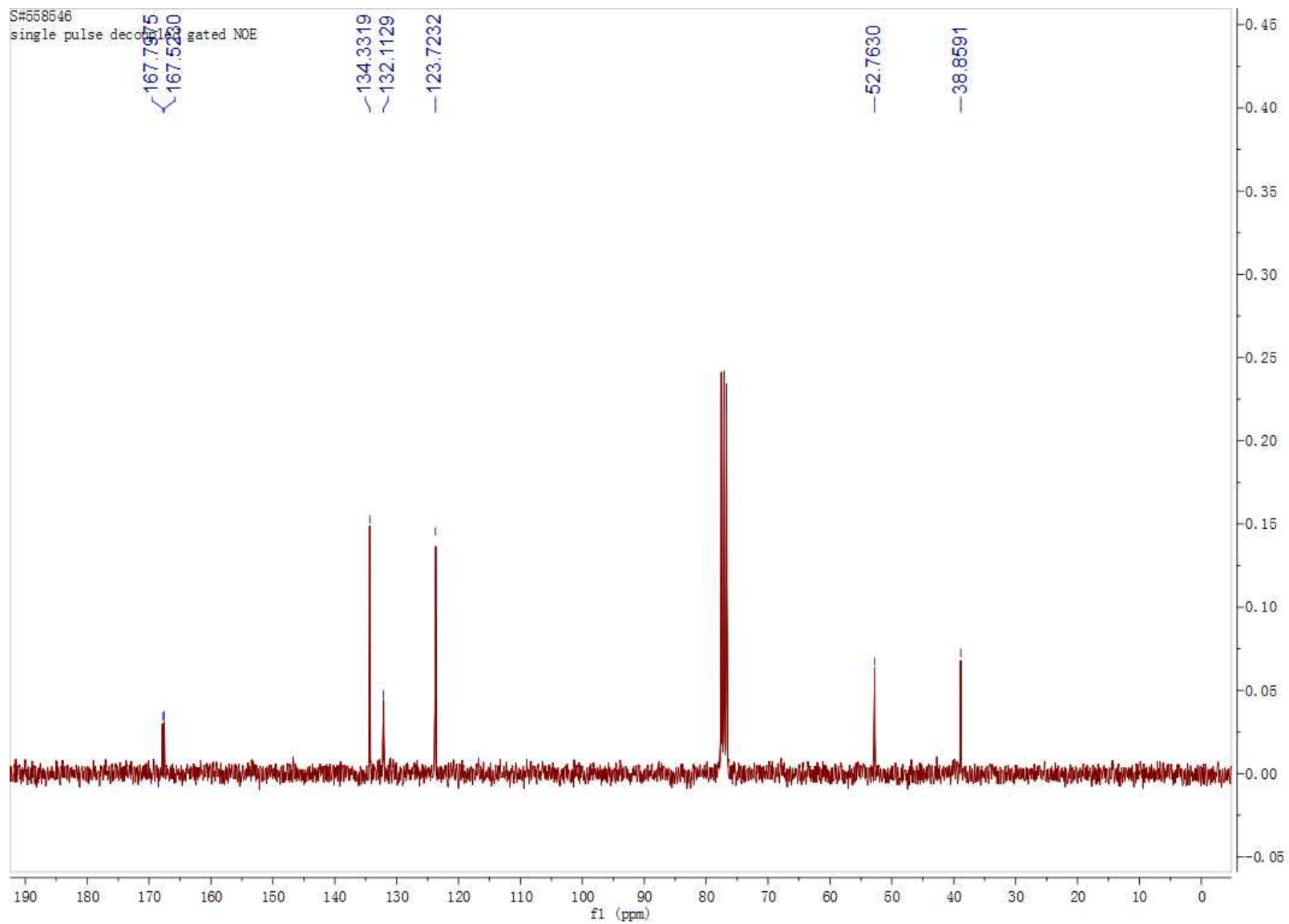
^1H NMR for compound **2e**



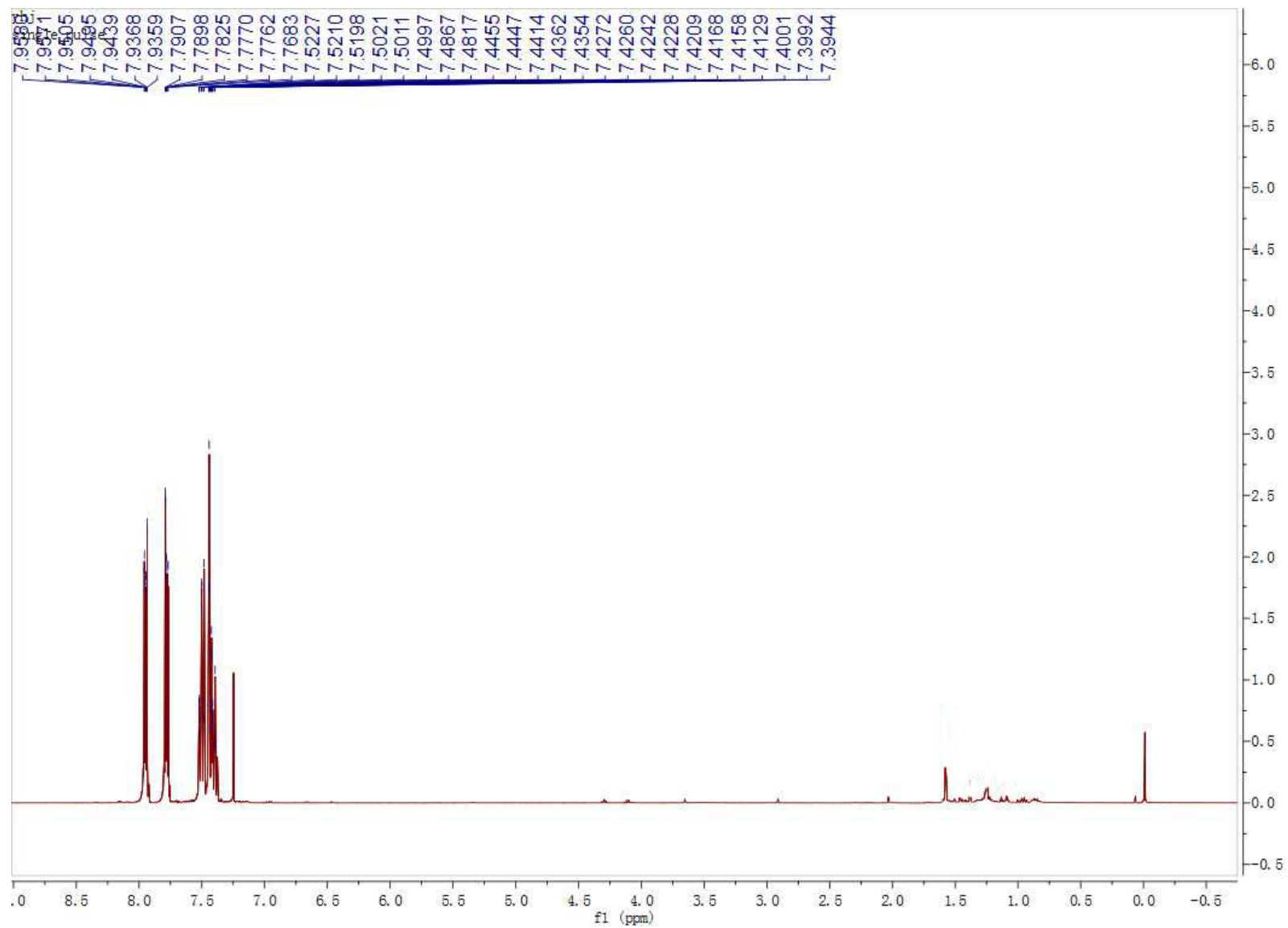
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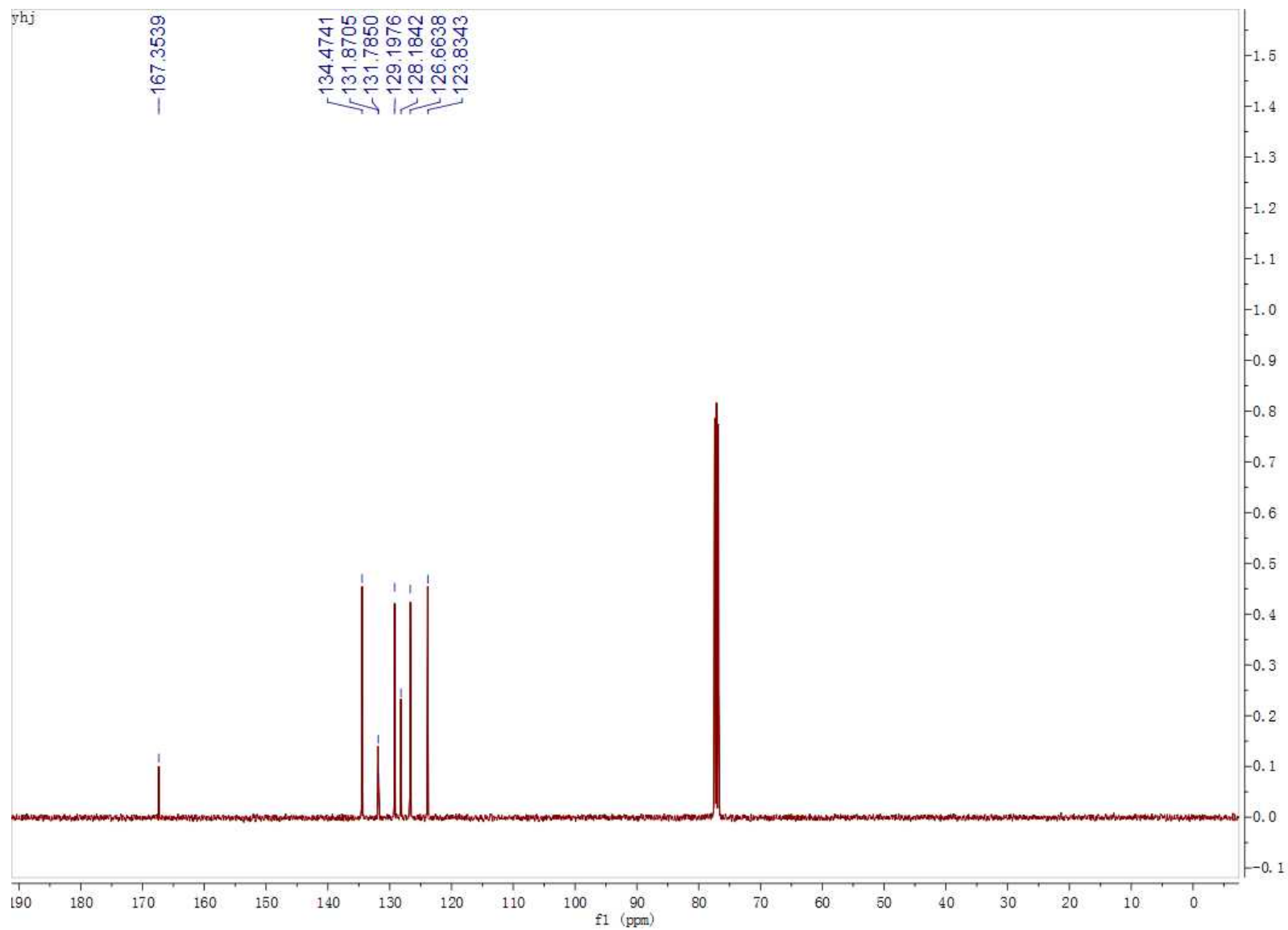
^1H NMR for compound **2f**



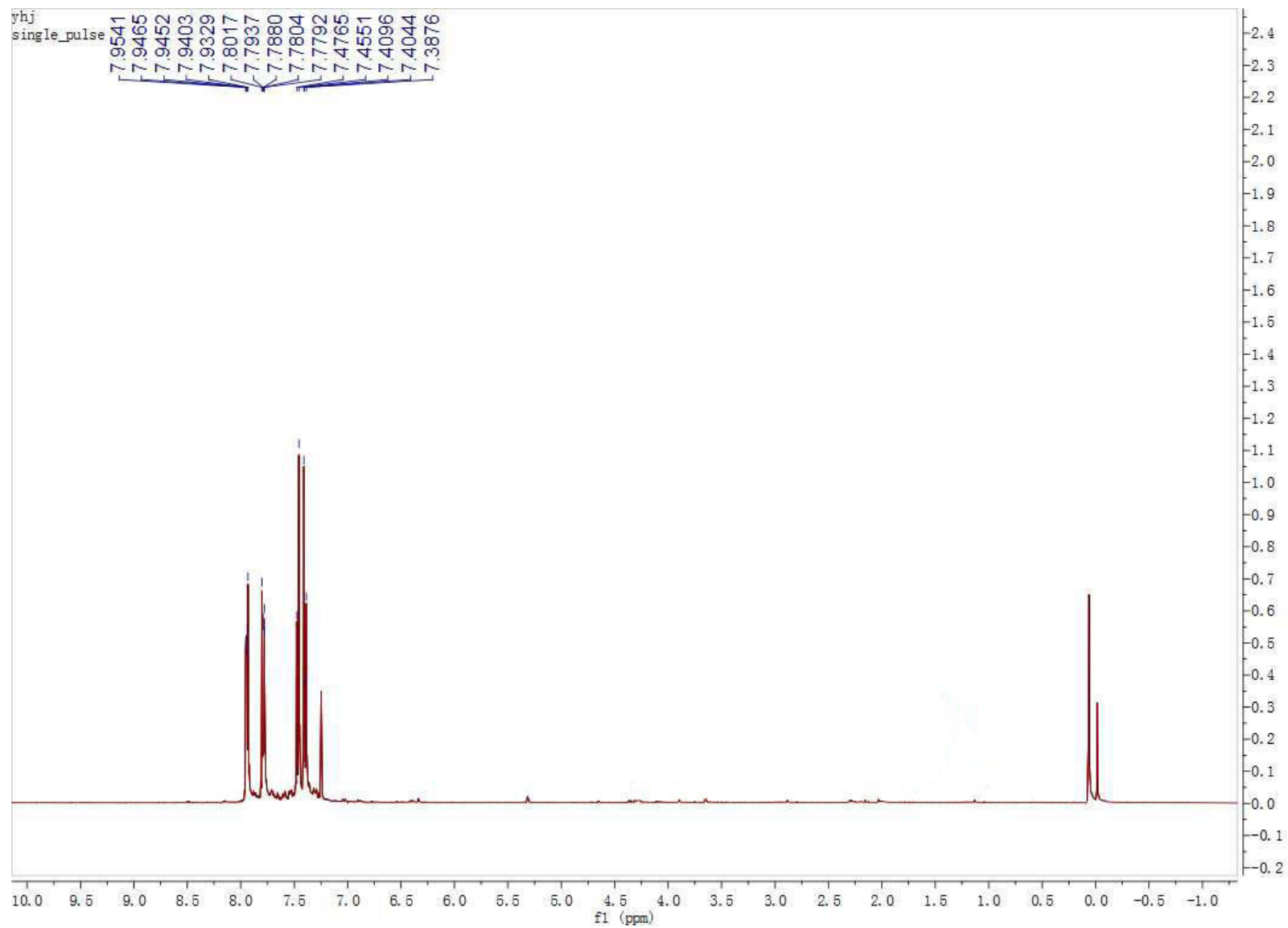
^{13}C NMR for compound **2f**



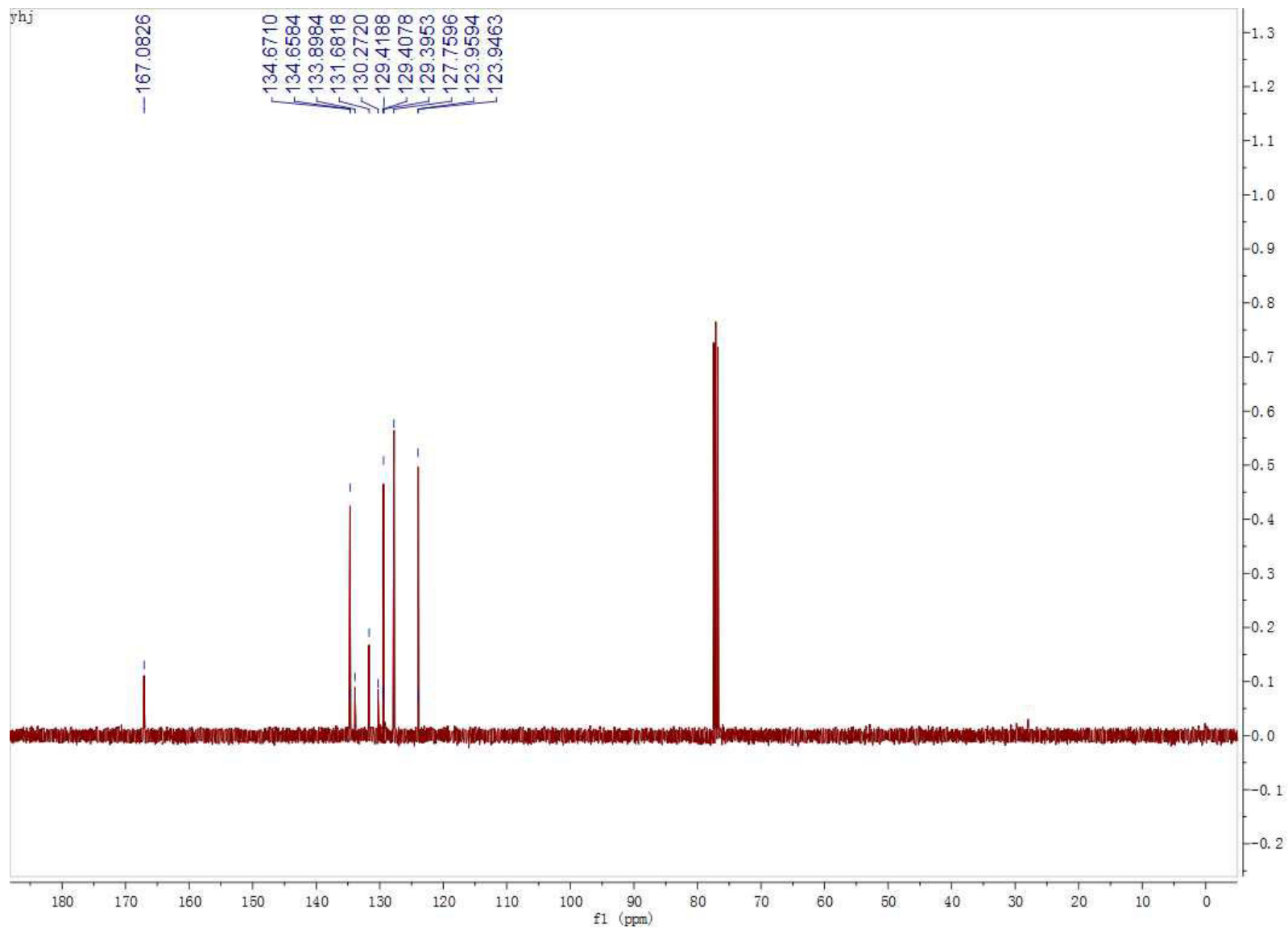
^1H NMR for compound **2g**



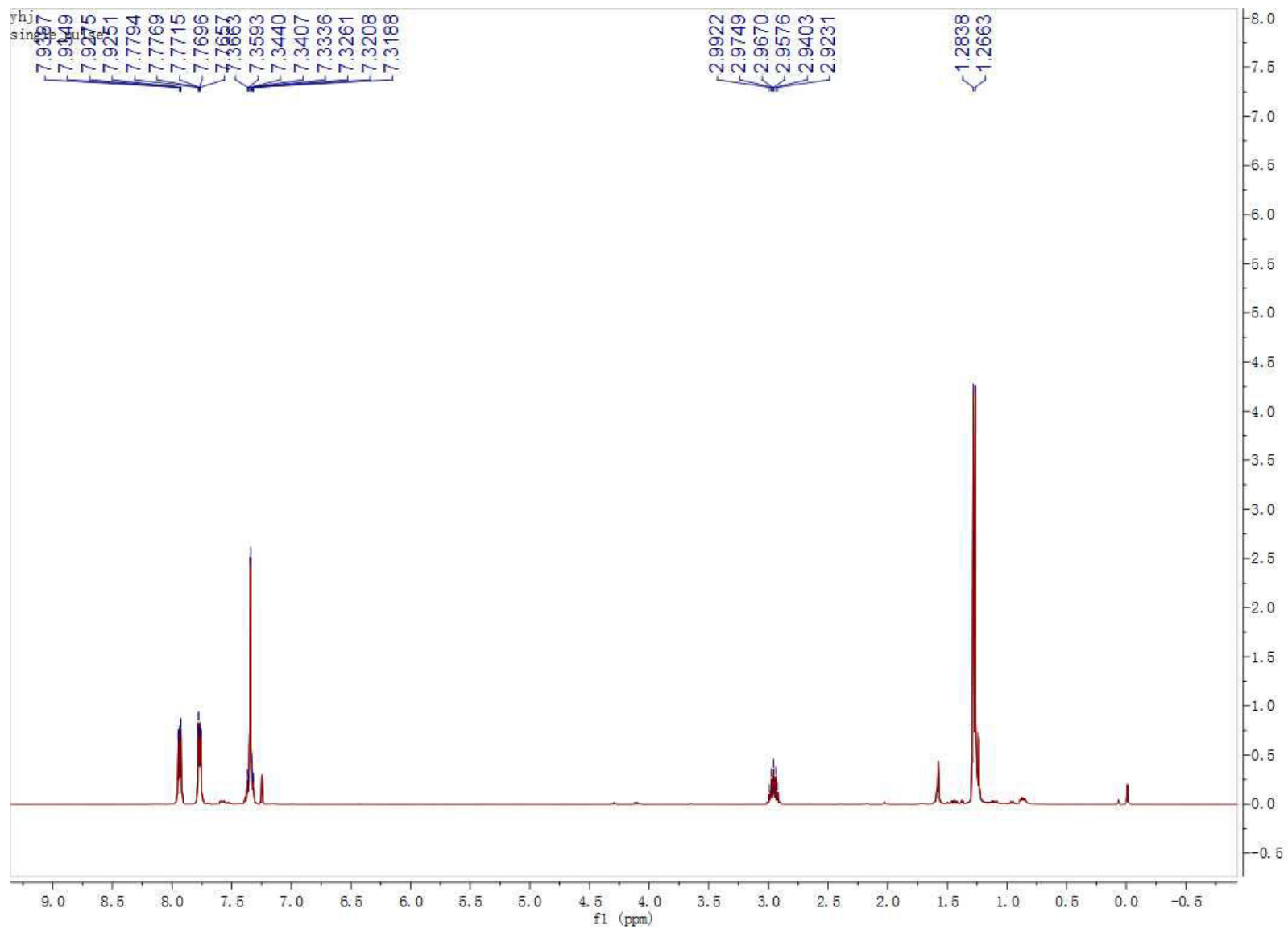
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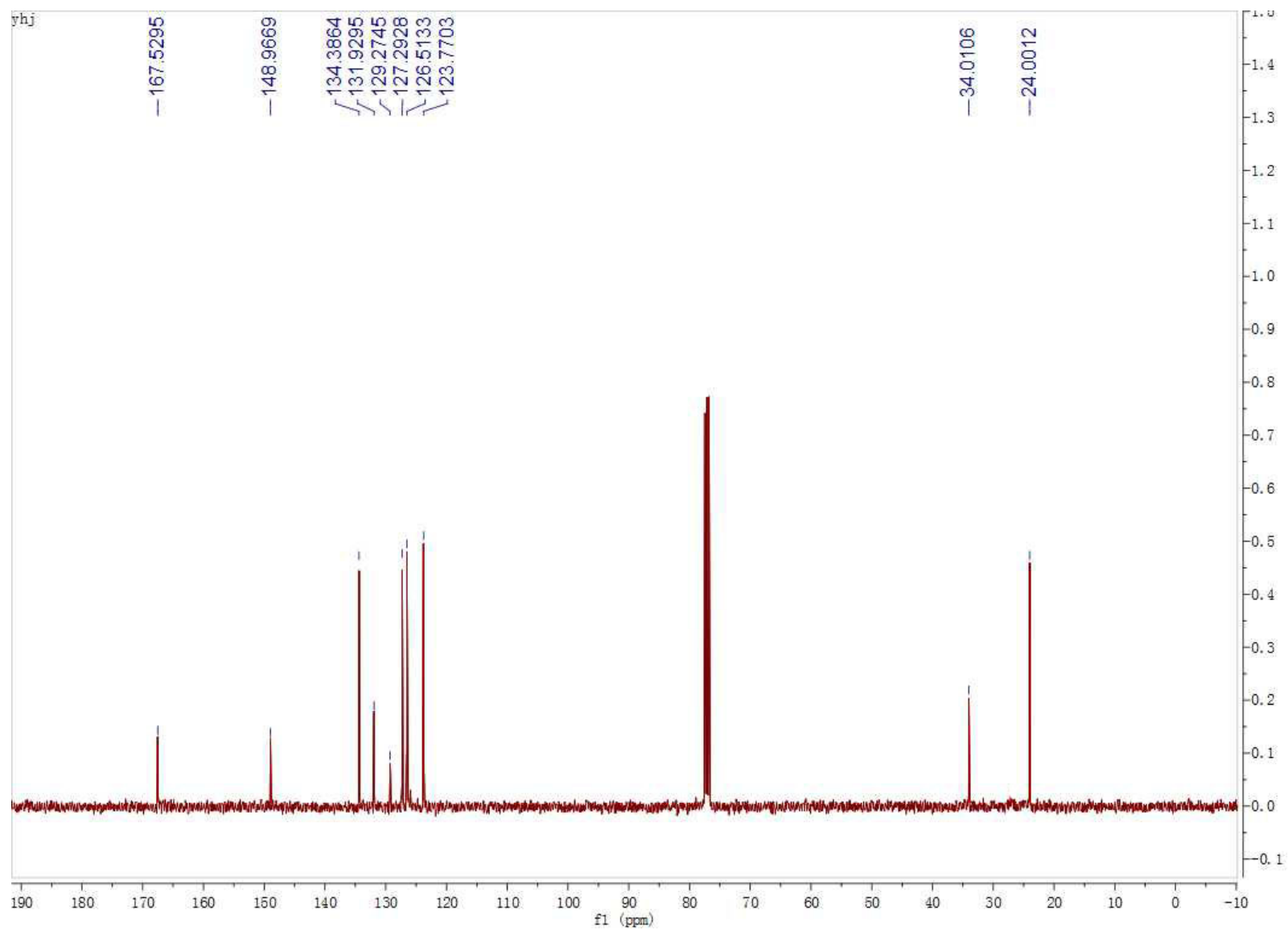
¹H NMR for compound **2h**



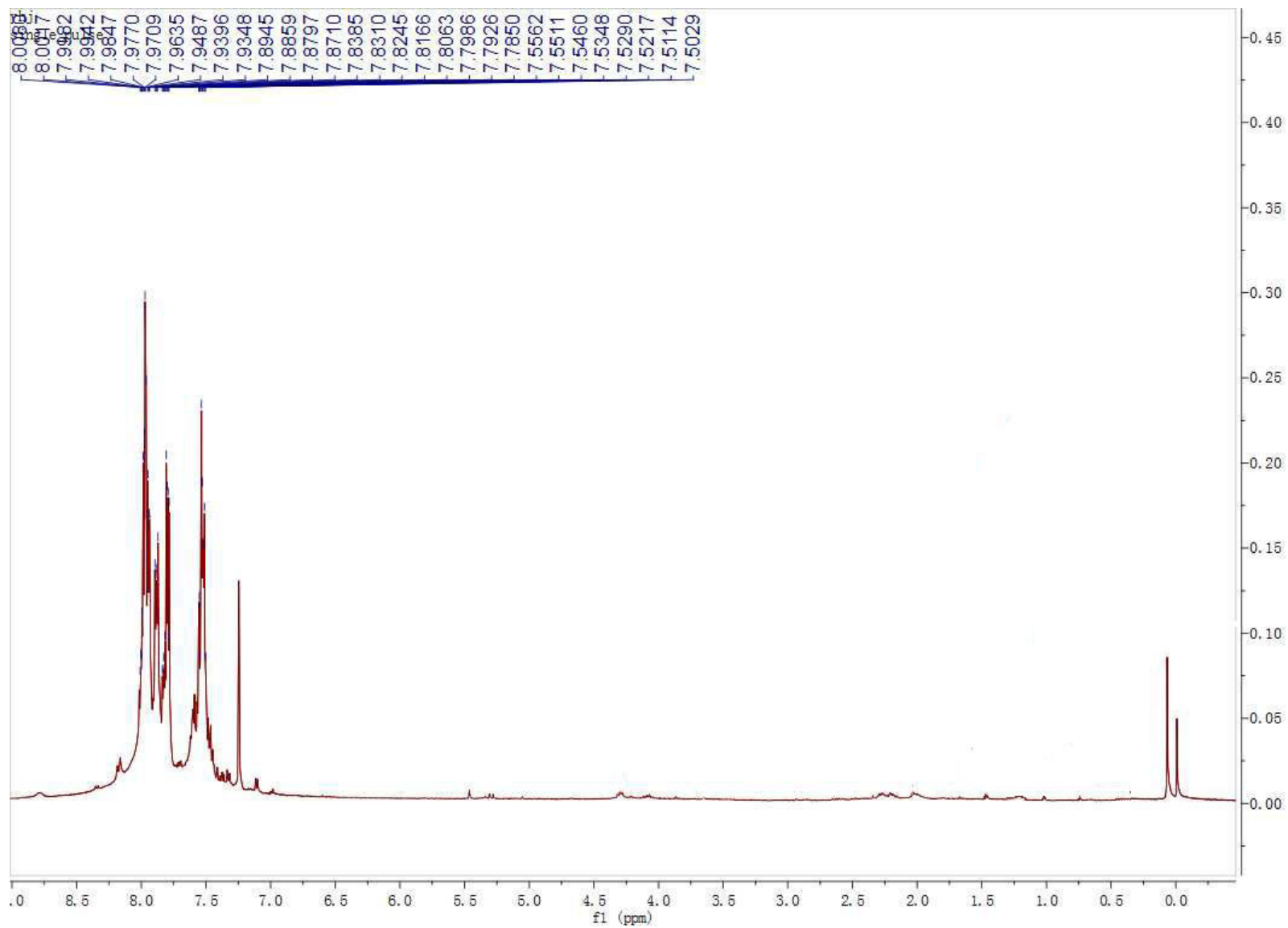
¹³C NMR for compound **2h**



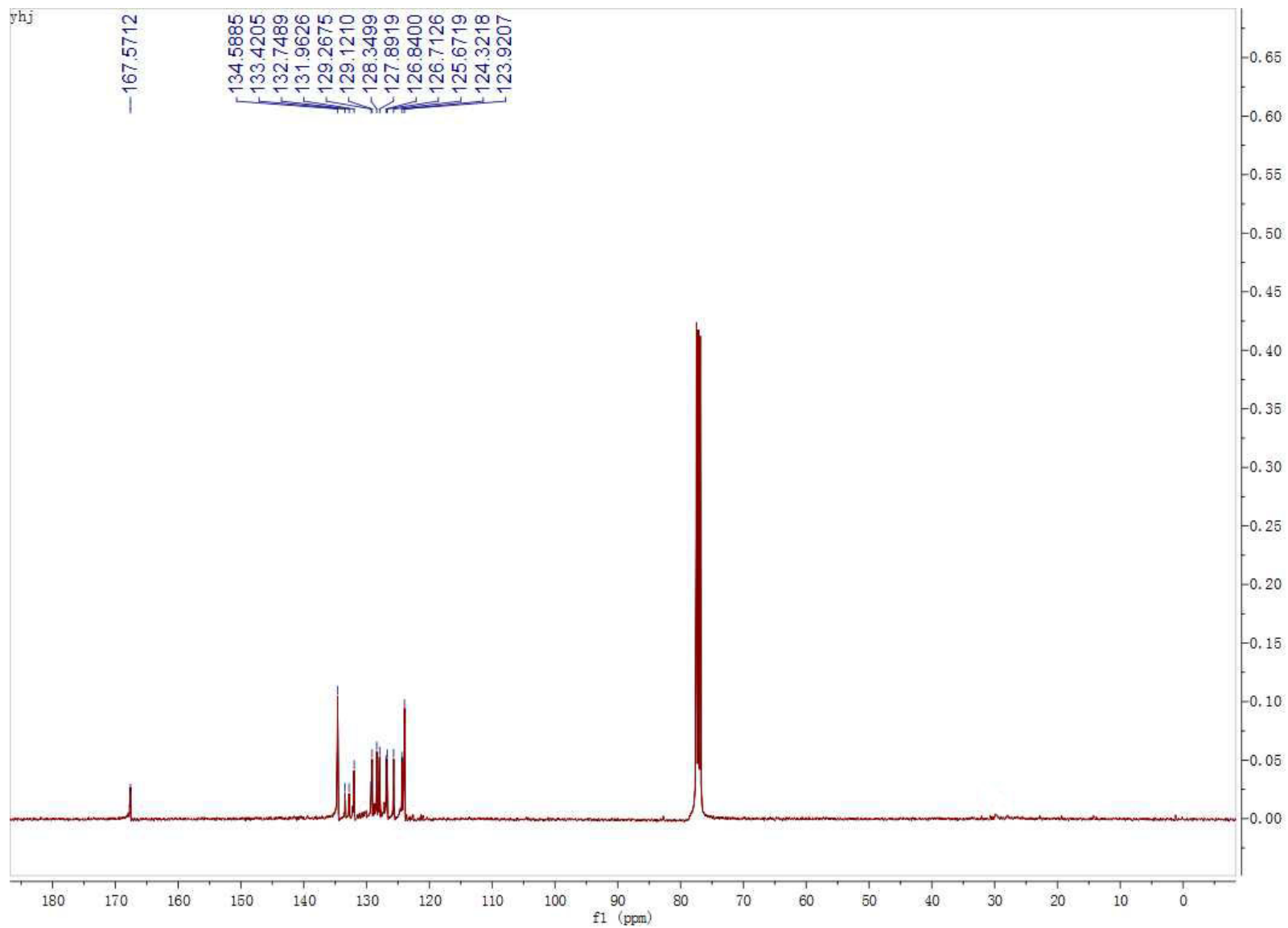
¹H NMR for compound 2i



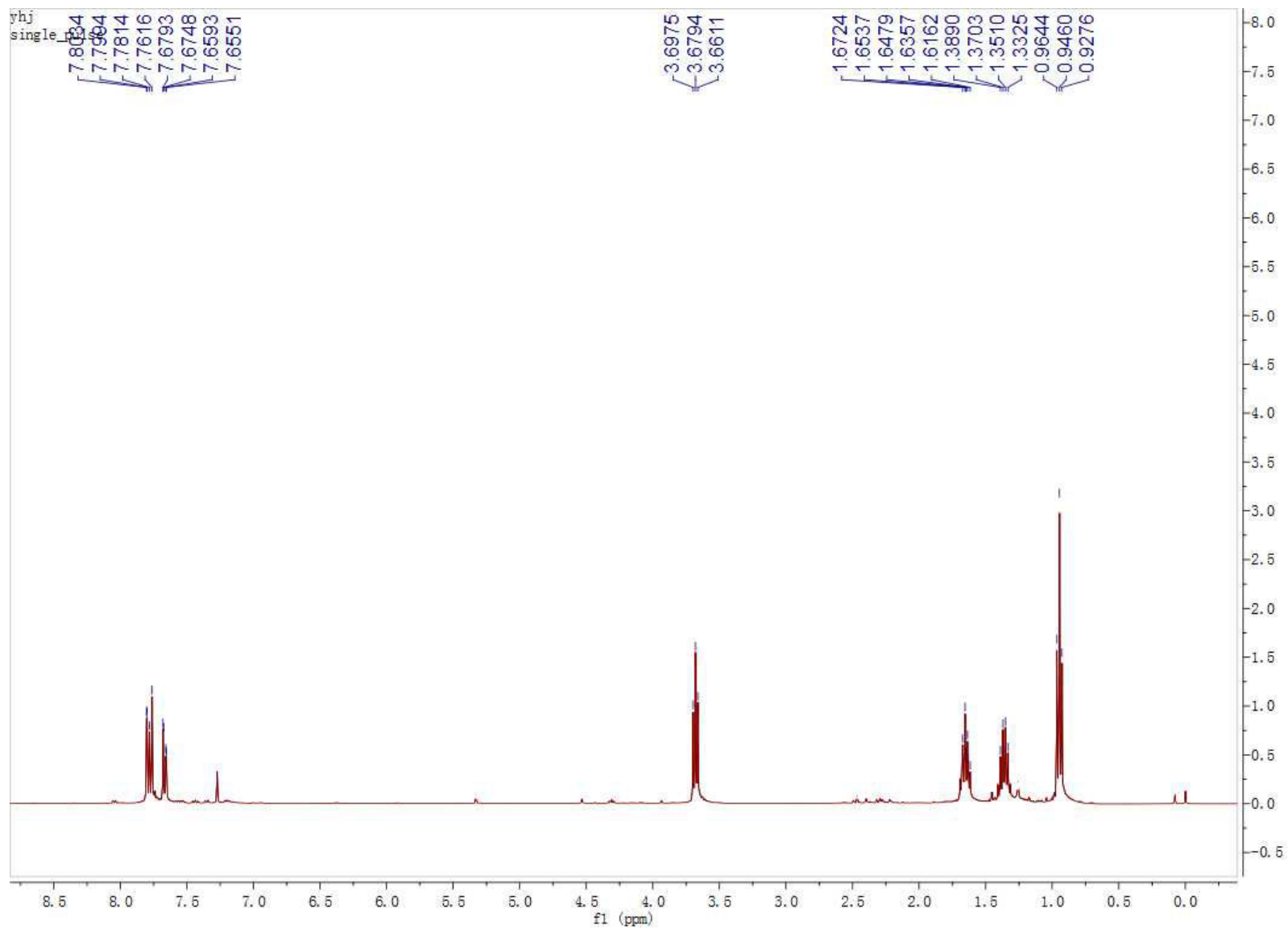
¹³C NMR for compound 2i



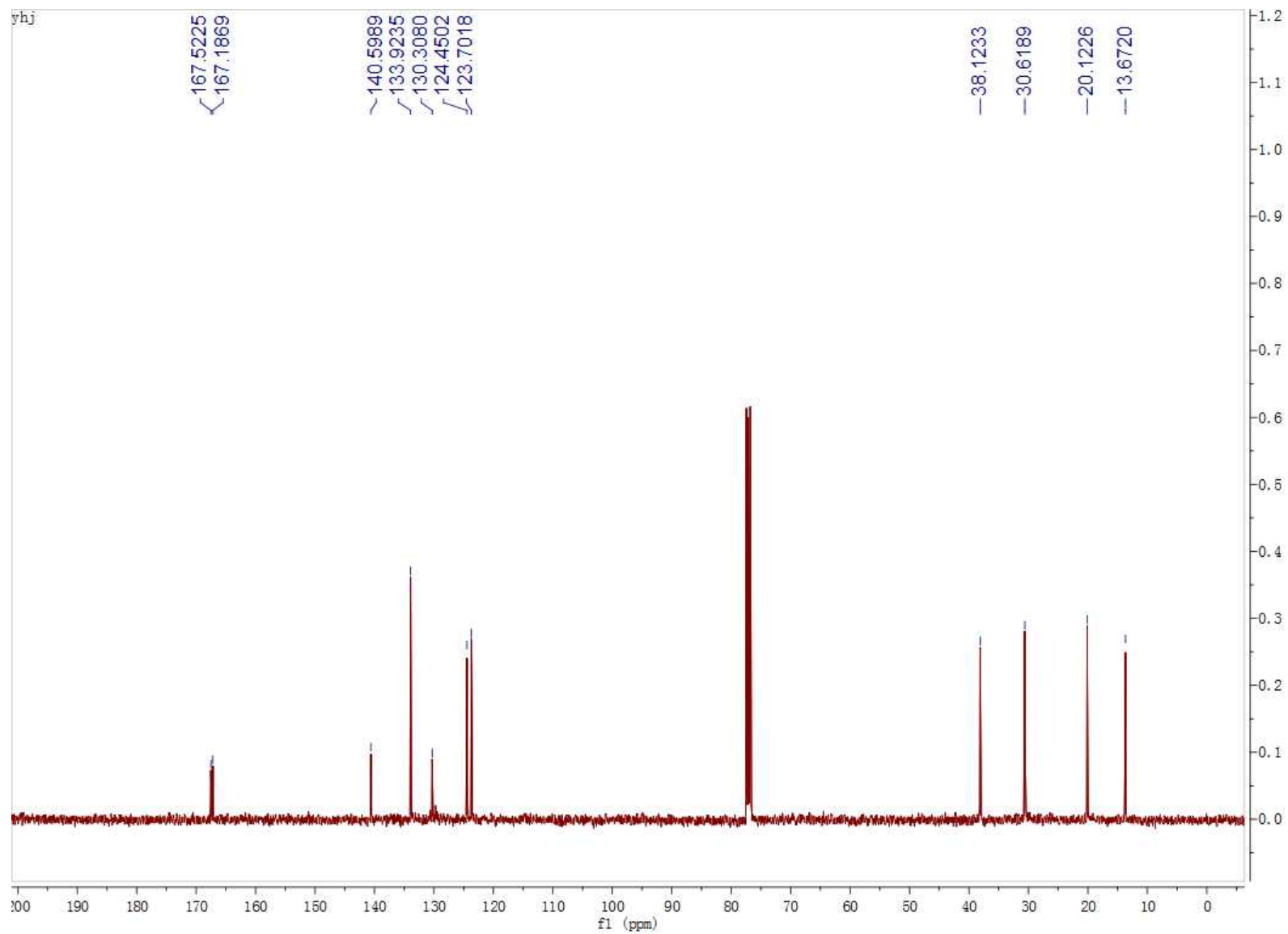
¹H NMR for compound 2j



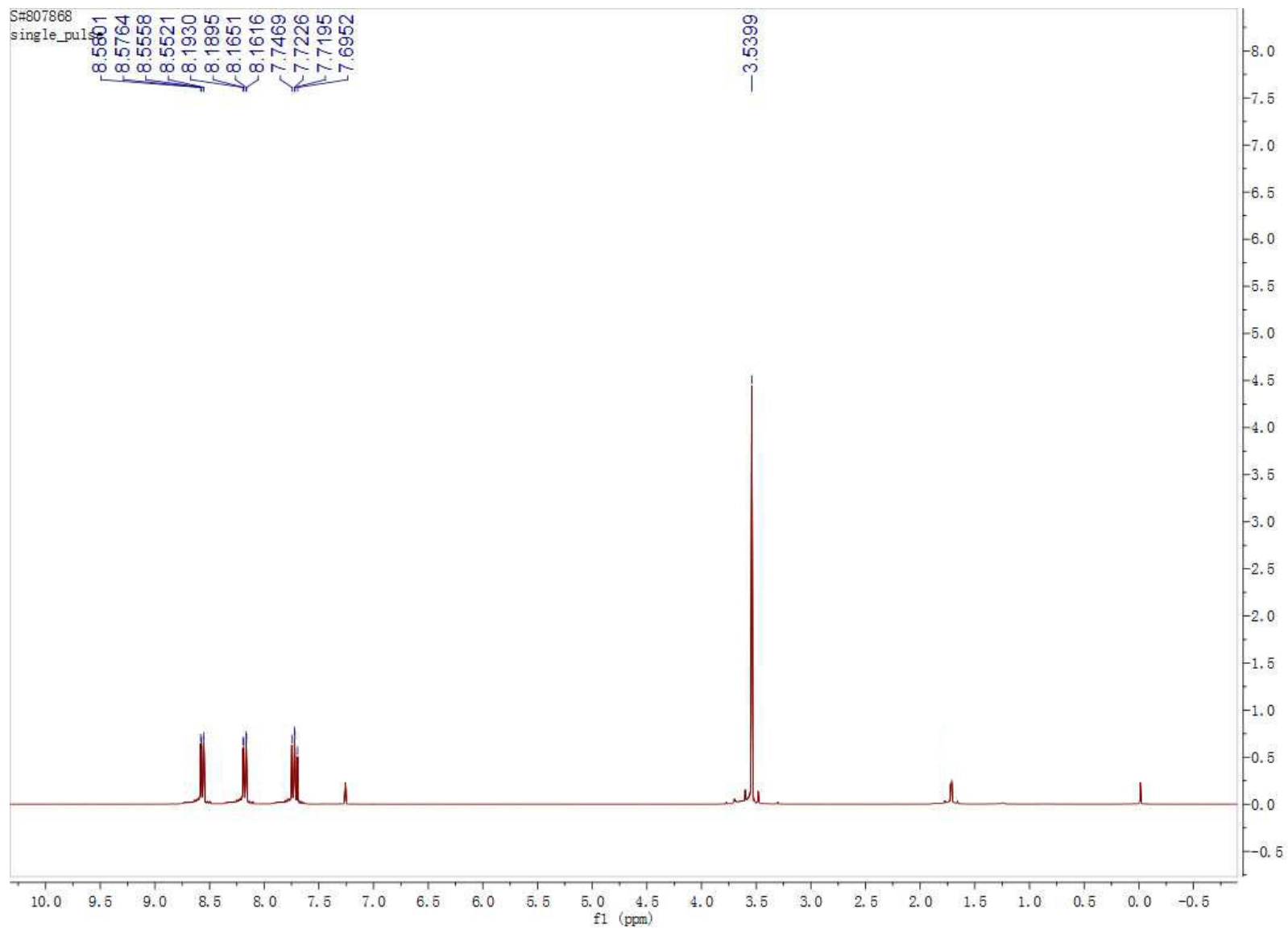
¹³C NMR for compound 2j



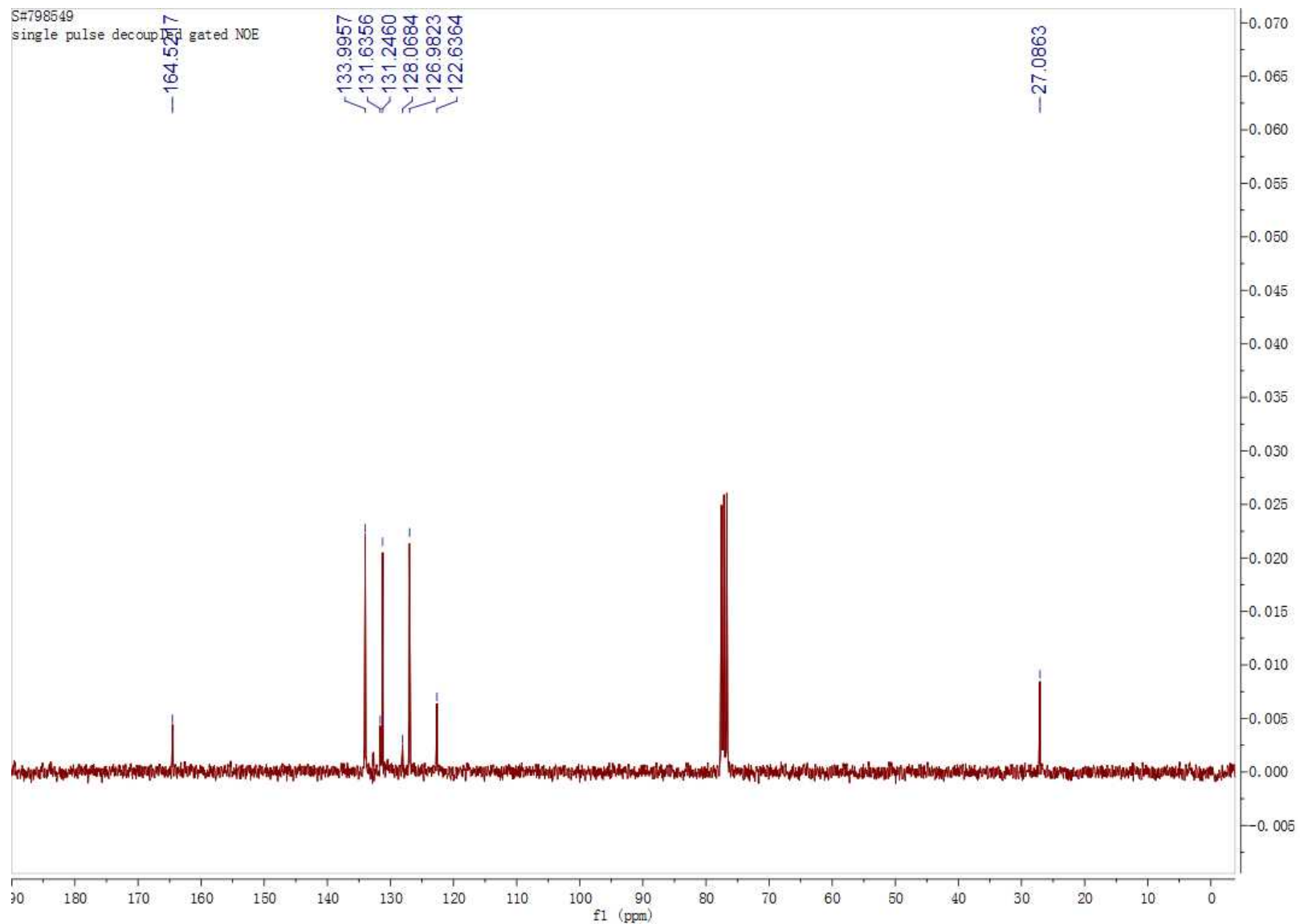
¹H NMR for compound 2k



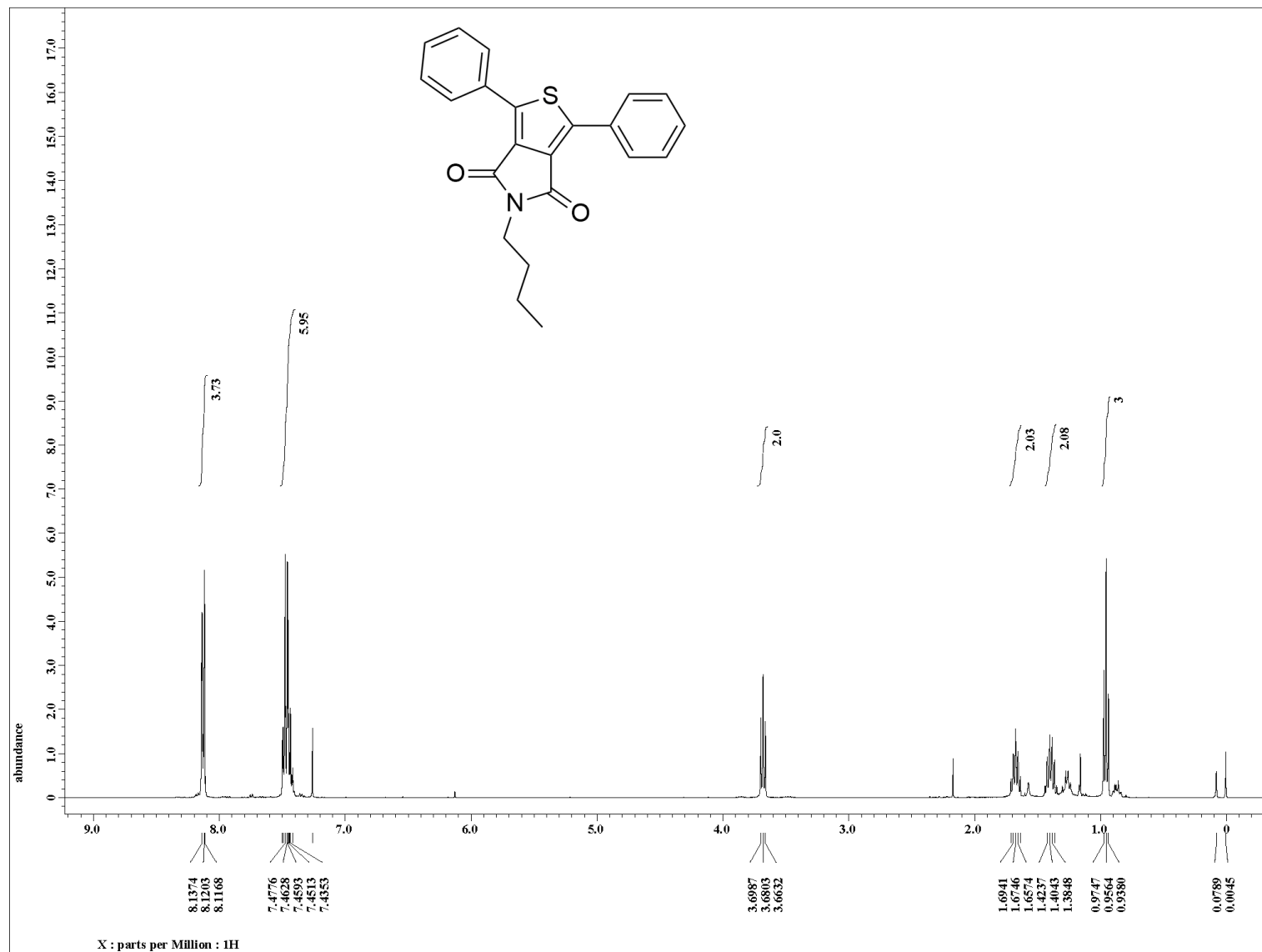
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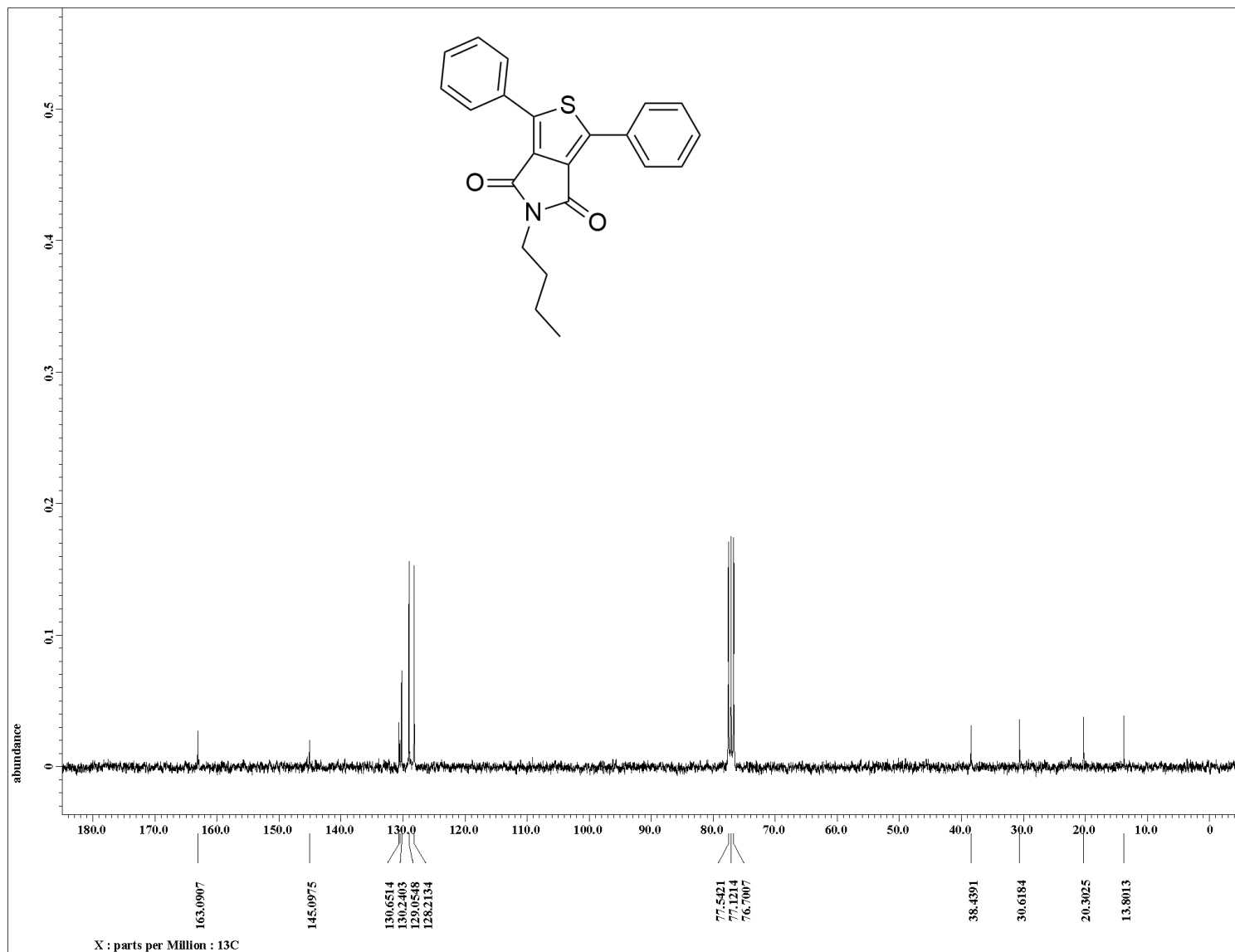
^1H NMR for compound **21**



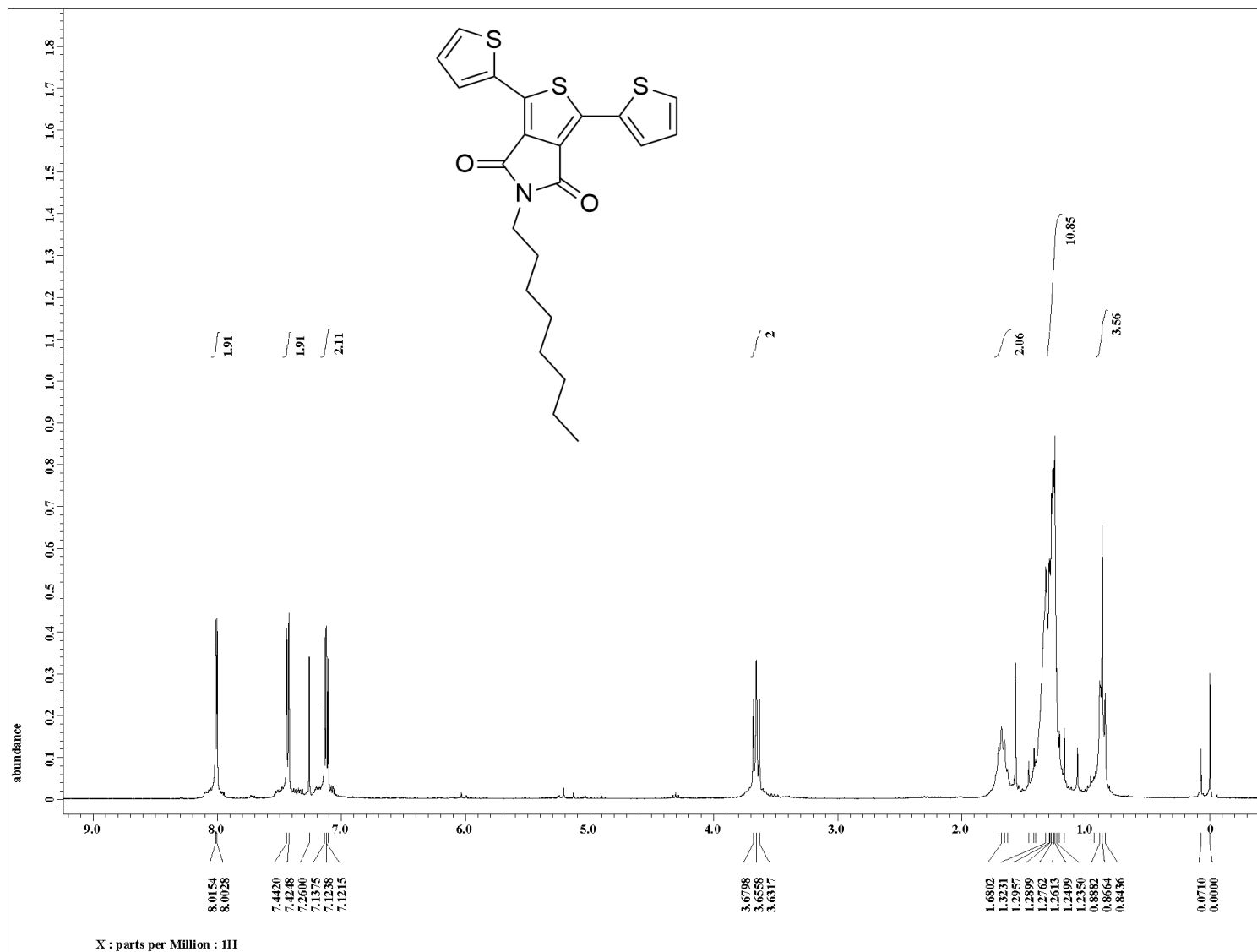
^{13}C NMR for compound **2I**



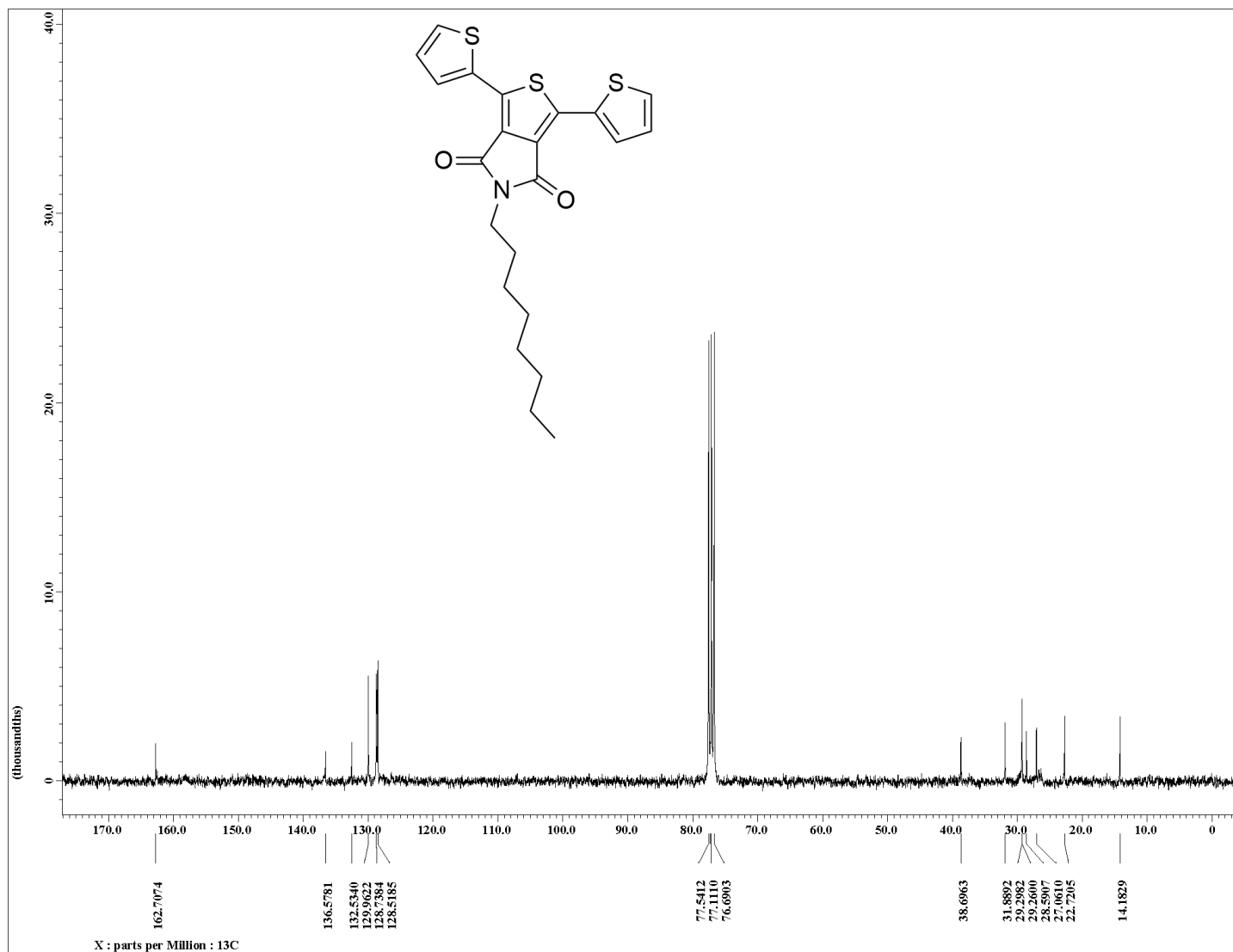
¹H NMR for compound 4a



^{13}C NMR for compound **4a**



¹H NMR for compound **4b**



^{13}C NMR for compound **4b**