

Supplementary Information for

Dendrimer-folate-copper conjugates as bioprobes for synchrotron x-ray fluorescence imaging

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Materials and methods

1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide HCl (EDC, 98%), polyamidoamine (PAMAM) dendrimer generation 5 (G5), triethylamine, Folic acid (FA) were purchased from Aldrich Co., Ltd; Acetic anhydride, dimethyl sulfoxide (DMSO, 99%), dimethylformamide (DMF, 99%), Dialysis membrane (MWCO=3500) were purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai); KB cells and A549 cell were purchased from Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences. ¹H-NMR spectrum was performed on Bruker AVANCE DRX 500Hz spectrometer in D₂O solution.

Preparation of dendrimer PAMAM generation 5 - folic acid conjugates (G5-FA)

4.90 mg of Folic Acid (MW = 441.4 g/mol) reacted with 29.69 mg of EDC·HCl (MW= 191.71 g/mol) in a mixture of 6 mL dry DMF and 2 mL dry DMSO for 1 h. The reaction mixture was added dropwise to the DI water solution (15 mL) of 1.39 μmol of G5 PAMAM and vigorously stirred for 3 days. After dialysis (using cellulose membrane with 3500 MWCO, PBS buffer and DI water 3 times with 1 L each time) and lyophilization, the yield was 95.4%. Further purification was carried out by membrane filtration with DI water. The obtained sample G5-FA was lyophilized and stored in a dry place before further modification. Yield: 92.1 %.

Preparation of acetylated dendrimer PAMAM generation 5 - folic acid conjugate

(Ac-G5-FA)

Acetylated PAMAM dendrimer generation 5(Ac-G5-FA) was prepared according to the reported procedure. Briefly acetic anhydride (150% of primary amine on the surface of PAMAM dendrimer G5) was slowly added to the dendrimer G5-FA solution (1.73 μmol dendrimer G5-Ac dissolved in 6 mL

methanol) in the presence of triethylamine (1.25 equivalent of acetic anhydride). The mixture was stirred under N₂ atmosphere at room temperature. After 18 h methanol was evaporated on rotary evaporator. The residue was dissolved in water and dialyzed (using cellulose membrane with 3500 MWCO) against PBS buffer and double distilled (DI) water for 3 days. The obtained sample Ac-G5-FA was lyophilized and stored in a dry place before further modification and characterization. Yield: 94.7%.

Measurements of metal ion binding to PAMAM dendrimers in aqueous solutions

Cu (II) was selected as a model cation to probe the binding of metal ions with affinity toward the amine groups inside the PAMAM dendrimers. Reagent grade CuSO₄ · 5H₂O was used as a source of Cu (II). Briefly, a CuSO₄ solution (2mM, 5 mL) was added into a G5-Ac-FA (or G5-Ac) dendrimer aqueous solution (0.1mM, 5mL) under vigorous stirring. The reaction mixture turned a deep-blue from wather blue within a few seconds.

Cell culture and cytotoxicity assay

KB cells (folate-receptor positive, FR+) and A549 cells (folate-receptor negative, FR-) were procured from the Stem Cell Bank of the Chinese Academy of Sciences. The KB cell line is a human oral epidermoid carcinoma that overexpresses FR, especially when grown in a low folic acid medium. The KB cells were grown continuously as a monolayer at 37 °C and 5% CO₂ in folic-acid-deficient RPMI 1640 medium containing 10% heat-inactivated fetal bovine serum (FBS), yielding a final folate concentration roughly that of normal human serum¹⁻². A549 Cell line is a human lung adenocarcinoma cells without folate-receptor, was grown continuously as a monolayer at 37 °C, and 5% CO₂ in RPMI 1640 medium containing 10% heat-inactivated fetal bovine serum (FBS).

We first tested the cytotoxicity of G5-Ac-FA-Cu²⁺ conjugates to KB cells and A549 cell . In brief, KB cells were respectively seeded in 96-well plates at a density of 5×10^3 cells per well and incubated for 24h. The medium in each well was replaced with 100 μ l of culture medium containing different concentration of treatments(0.1 μ M, 0.2 μ M, 0.4 μ M, 0.8 μ M,1.6 μ M,3.2 μ M) and cultured for another 24h. 10 μ L WST-1 solution was added to wells, absorbance values were taken using a 96-well Ophys Microplate Reader at 450 nm.³ Cell viability is expressed as a percentage of control. The cytotoxicity of G5-Ac-FA-Cu²⁺ conjugates to A549 cell have been performed according to the same procedure.

Preparation of samples for Synchrotron X-ray Fluorescence analysis

KB cells (folate-receptor positive, FR+) and A549 cells (folate-receptor negative, FR-) were procured from the Stem Cell Bank of the Chinese Academy of Sciences. These cells were grown continuously as a monolayer at 37 °C, and 5% CO₂ in RPMI 1640 medium containing 10% heat-inactivated fetal bovine serum (FBS). After trypsinization, cells were collected in a tube, centrifuged and resuspended in fresh RPMI 1640 medium to a final concentration of 1×10^5 cells/ml and seeded in 24 well plates with Mylar film. Copper uptake studies were performed in the same medium but supplemented with 0.2 μ M G5-Ac-FA-Cu²⁺ at 37°C for 4h, then washed with PBS for three times, fixed with paraformaldehyde in isotonic phosphate-buffered saline, and washed twice with ammonium acetate or deionized water to avoid formation of residual salt crystals that might increase background X-ray fluorescence. Cell samples were stored at 4°C before SXRF measurements.

Synchrotron radiation experiment

The experiment was performed at the Shanghai Synchrotron Radiation Facility in China. Scanning

X-ray fluorescence analysis was performed at XRF microprobe station. The X-ray light source comes from the BL15U1 beam line of Shanghai Synchrotron Radiation Facility (SSRF, Shanghai, China). The sample were placed onto a kinematic specimen holder suitable for both optical and X-ray fluorescence microscopy. The holder was then mounted on a light microscope (Leica DMXRE) and target cells imaged previously by standard fluorescence microscopy were located on the grid relative to a reference point using a high spatial resolution motorized x/y stage (Ludl Electronic Products, Hawthorne, NY). Coordinates were determined and used to precisely locate the target cell once the grid was transferred to the microprobe. A Fresnel zone plate was used to focus the monochromatic X-ray beam from an undulator source to a spot size of $2 \times 2 \mu\text{m}^2$ on the specimen. An incident photon energy of 7.9-8.2keV was chosen to ensure excitation of the K-line of Cu, and the sample was raster scanned through the beam at 298K under a helium atmosphere. The pixel step size was set to $1 \mu\text{m}$ and the entire X-ray spectrum was recorded at each pixel using an energy dispersive germanium detector. This beam line can provide multi-chromatic X-rays (white light), who's energy range from 3.5 – 22.5keV. The electron energy in the storage ring is 1-20 KeV, the spot size of the X-ray beam was $2.5 \times 2.6 \mu\text{m}^2$.

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2. Majoros, I. J.; Myc, A.; Thomas, T.; Mehta, C. B.; Baker, J. R., Jr. *Biomacromolecules* **2006**, 7, 572–579.
3. Worle-Knirsch, J. M.; Pulskamp, K.; Krug, H. F. *Nano Lett.* **2006**, 6, 1261.

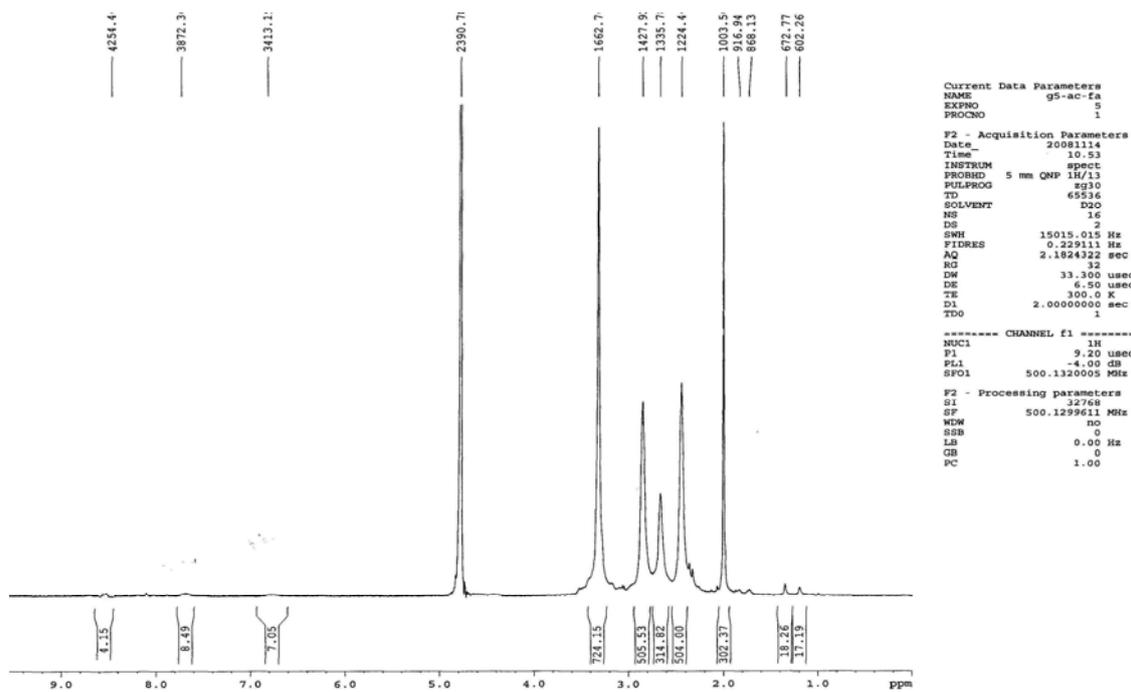


Fig. S1 The ¹H-NMR spectrum of G5-FA-Ac

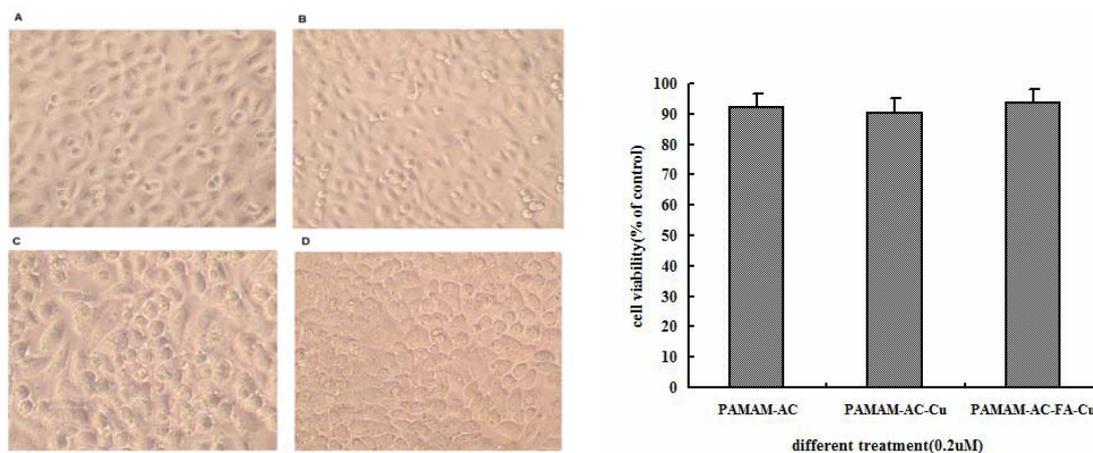


Fig. S2 The cytotoxicity assessments G5-Ac-FA-Cu²⁺ conjugates in concentration of 0.2 μM. A) KB cell, B) KB cell cultured with G5-Ac, C) KB cell cultured with G5-Ac-Cu, D) KB cell cultured with G5-Ac-FA-Cu

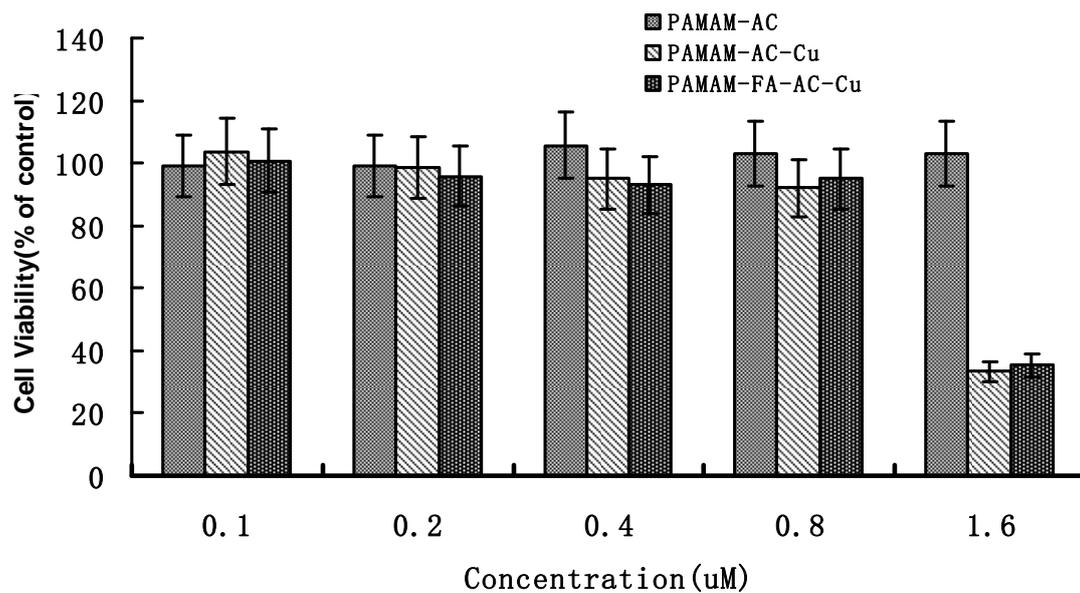


Fig. S3 The cytotoxicity assessments G5-Ac-FA-Cu²⁺ conjugates to A549 cells in different concentrations.

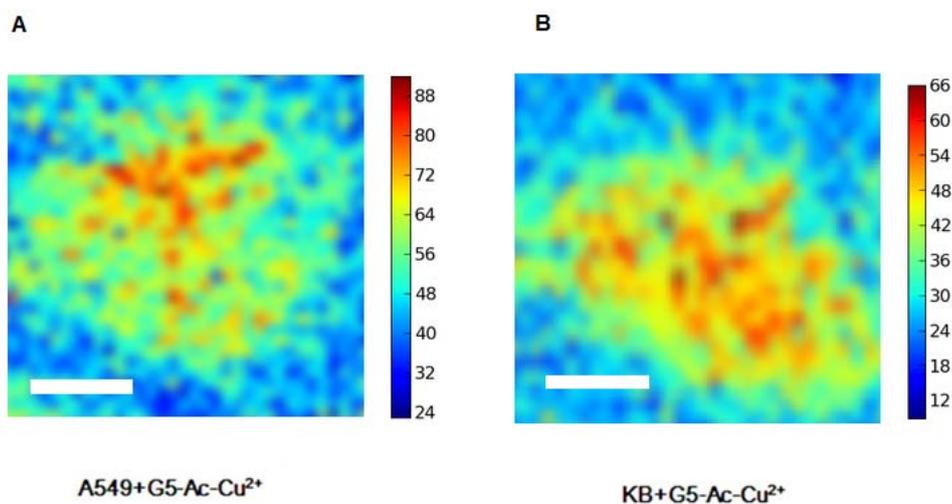


Fig. S4 The Synchrotron X-ray fluorescence analysis of KB cells and A549 cells cultured with G5-Ac-Cu²⁺ conjugates.

Scale bar: 5 μm.

A, A549 cell cultured with G5-Ac-Cu²⁺ conjugates B, KB cells cultured with G5-Ac-Cu²⁺ conjugates