Regioselective synthesis of oxepinones and azepinones by goldcatalyzed cycloisomerizations of functionalyzed cyclopropyl alkynes

Jesús M. Fernández-García,^a Patricia García-García,^b Manuel A. Fernández-Rodríguez,^b Alexandra Pérez-Anes,^a Enrique Aguilar,*^a

^a Instituto Universitario de Química Organometálica "Enrique Moles", Departamento de Química Orgánica e Inorgánica, Universidad de Oviedo, C/Julián Clavería, 8. 33006, Oviedo, Spain. Fax: +34-985 103 446; E-mail: eah@uniovi.es

^b Área de Química Orgánica, Departamento de Química, Facultad de Ciencias, Universidad de Burgos, Pza. Misael Bañuelos, s/n, 09001, Burgos, Spain

Electronic Supplementary Information

TABLE OF CONTENTS

	Page
1. Optimization of the Reaction Conditions	ESI-3
2. Proposal to Explain the Lack of Reactivity of <i>trans</i> -DA Alkynylcyclopropanes	ESI-7
3. Experimental Part	ESI-8
• 3.1. General Considerations	ESI-8
• 3.2. Catalyst Lyophilisation Procedure	ESI-9
• 3.3. Synthetic Procedures and Compounds Characterization Data	ESI-10
4. Tables with the 2D-NMR Experiments for Selected Compounds	ESI-40
5. References	ESI-42
6. Copies of ¹ H- and ¹³ C-NMR Spectra	ESI-43

1. Optimization of the Reaction Conditions

A selected set of the tested conditions is displayed on table ESI-1.





^{*a*} Isolated yields. ^{*b*} Estimated by ¹H-NMR using hexamethylbenzene as internal standard. ^{*c*} The catalyst system was lyophilized prior to carrying out the reaction. ^{*d*} Only the silver salt was lyophilized prior to carrying out the reaction.

First essays using $Ph_3PAuCl/AgSbF_6$ as catalyst system did not lead to the expected oxepinone; instead they caused the opening of the cyclopropane ring leaving the triple bond unaltered and giving rise mainly to five-membered lactone **3**, with 4-oxo-6-phenyloct-5-ynoic acid **5** as byproduct (entry 1). Gratifyingly, when we switched to $Ph_3PAuNTf_2$ as catalyst¹ we obtained the desired seven-membered heterocycle **2a** as major product accompanied with the acyclic ester derivative **6** (entry 2). To determine whether this variation in the selectivity was caused by the change in the counterion or by the presence/absence of silver in the reaction mixture, we treated **1a** with catalytic amounts of AgSbF₆ and we observed complete and selective conversion to **5** (entry 3). This result shows how silver hexafluoroantimonate is able to catalyze the transformation of **1a** into **5** and, therefore, its presence should be avoided in order to get selective formation of oxepinone **2a**. As a consequence, preformed cationic gold catalysts were employed for evaluating the influence of different ligands. A considerable improvement in the isolated yield of the oxepin-2-one was achieved by modifying the

electronic properties of the ligand, by moving from an electron-deficient phosphine ligand to an electron-rich phosphine ligand and, then, to a NHC carbene ligand (entries 4-6). However, a five-membered lactone with concomitant hydration of the triple bond 4 as well as previously observed ester derivative 6 appeared as minor byproducts. A great advance was achieved when the catalyst was lyophilized (see below) prior to carrying out the reaction, which improved both the yield and the selectivity of the reaction (entries 6 vs 7). On the other hand, slightly lower yield of 2a and selectivity were reached when the Ph₃PAuCl/AgOTs system was tested (entry 8). A final survey revealed that the combination IPrAuCl/AgOTs led to a remarkable 90% isolated yield of 2a (entry 9). As suggested by one of the referees, considering that lyophilisation only removes water from the catalyst and gold complexes are usually not hygroscopic, it seems therefore essential to remove water from the hygroscopic silver salt by lyophilisation (otherwise the amount of wet silver salt contains less silver, not all the gold catalyst is activated, the conversion becomes slower and incomplete). A test carried out with lyophilised AgOTs and non-lyophilised gold complex led to a similar result, within the experimental error (86% isolated yield of 2a, entry 10). In these cases the presence of silver was not detrimental for the selectivity, as AgOTs turned out to be almost inert (entry 11).

During the catalyst optimization, the reactivity of carboxylic acids 1 was tested under very different conditions. Optimization of the reaction conditions is indeed a dynamic process where different reaction parameters are analysed, occasionally in a simultaneous manner. Thus, an extensive optimization of temperature, solvent and substrate concentration was carried out with lyophilized IPrAuNTf₂, which was the best catalyst system that we had found so far. The main results are listed in Table ESI-2. It is worth to notice that with such catalyst compound **2a** was isolated in 27% yield (entry 10) when the reaction was carried out at 0 °C and no by-products could be identified.

	O O O H 1a	−Ph IPrAuNTf ₂ solvent (conc.), [−] MS 4 Å	MeO T ► 2a	Ph O O A
Entry	Solvent ^a	Conc.	Т	Yield ^b
1	DCM	0.05 M	rt	69
2	DCE	0.05 M	rt	67
3	THF	0.05 M	rt	Traces
4	Hexane	0.05 M	rt	Traces
5	MeCN	0.05 M	rt	Traces
6	Toluene	0.05 M	rt	-
7	DMF	0.05 M	rt	-
8	DCM	0.025 M	rt	21
9	DCM	0.1 M	rt	26
10	DCE	0.05 M	0 °C	27
11	DCE	0.05 M	40 °C	42
12	DCE	0.05 M	60 °C	33
13	DCE	0.05 M	80 °C	31
1 DOM		alors are at hr		

Table ESI-2 Optimization of the reaction conditions (II)

^{*a*} DCM= CH₂Cl₂; DCE = ClCH₂CH₂Cl. ^{*b*} Isolated yield of 2a.

However, after establishing the system IPrAuCl/AgOTs as the best catalyst, we had not carried out lower temperature experiments. Therefore, attending at a request made by one of the referees, we decided to carry out two experiments at 0 °C and at -20 °C. Both reactions led to **2a** as major product (42-73% yield) and <u>no traces of products of type II</u> were observed neither in the crude nor after purification. In both cases, compound **4** was observed, but not isolated (10-25%, estimated yield).

On the other hand, it is also worth to remark that <u>molecular sieves are necessary</u>; during our initial tests for the optimization of the reaction conditions the formation of higher amounts of compounds **4** (which results from hydration of the triple bond), **5** or **6** (both of them come from hydrolysis of the quaternary cyclopropyl carbon) was observed. The amount of such by-products was reduced or suppressed by the employment of molecular sieves. A reaction carried out recently (at request of one referee) with model compound **1a** under the optimized reaction conditions but without the presence of molecular sieves resulted in 38% of **2a** and a 58% of **4**.

A commentary about the remarkable effect of the silver salt seems also required. There are precedents for counter-ion effects in Au/Ag catalysis² and it is usually explained on the basis of the different coordinating properties of the anion. Also, the long-overlooked "silver effect" in gold(I) catalysis has been recently confirmed as crucial and real³. Thus, we have observed that, depending on the reaction analysed, the effect of the counter-ion in the selectivity may be almost negligible,⁴ minor,⁵ or play a crucial role.⁶ On the other hand, counter-ion effects in silver catalysis are also known.⁷ In our case, an exhaustive study of the counter-ion effect has not been carried out and the fact that non-previously-lyophilized AgSbF₆ led exclusively to compound **5** (which was probably formed by hydrolysis of the acetal moiety in **3**), while the starting material is almost quantitatively recovered in the presence of AgOTs seems to indicate that the different hygroscopic character of both catalysts may have played also some role in the reaction outcome.

2. Proposal to Explain the Lack of Reactivity of trans-DA Alkynylcyclopropanes

As stated in the experimental part (see below), the cyclopropanation reaction, carried out to prepare alkynylcyclopropanecarboxylic esters **18** (which are later hydrolysed to the corresponding alkynylcyclopropanecarboxylic acids **1**, used as starting materials), takes place diastereoselectively leading to the desired *cis*-isomer as major component while the *trans*-isomer is the minor component.⁸

For compound **1a**, the separation of the diastereomers was carried at the ester stage (compound **18a**).

It was found that the *trans*-isomer **1a** remains inert under the optimized reaction conditions (listed in Table ESI-1, entry 9), being recovered once the reaction has ended. Two conclusions were reached from this observation:

1) A mechanistic conclusion: The lack of reactivity of the *trans*-isomers is probably due to the fact that the carboxylic group is not close enough to the triple bond because of the rigidity imposed by the cyclopropane ring, which inhibits the nucleophilic attack of the carboxylic group to the activated triple bond. This fact also suggests that, under the optimized conditions, the nucleophilic attack should take place prior to the cyclopropane ring opening, thus supporting *Via A* pathway; on the contrary, if the opening of the cyclopropane ring takes place prior to the nucleophilic attack (*Via B*), the loss of the rigidity should also allow the transformation of the *trans*-isomer into oxepinone 2a, which does not occur.



Figure S1. Proposal to explain the lack of reactivity of *trans*-DA alkynylcyclopropanecarboxylic acid 1a

2) A practical conclusion: Separation of the diastereomers was unnecessary, as only one of them reacted. Therefore, once the optimized conditions were reached, the reactions to establish the scope of the transformation were carried out with the mixture of both diastereomers. The reported yields in Table 1 and Scheme 3 were determined taking into account this lack of reactivity of the *trans*-isomers.

3. Experimental Part

3.1. General Considerations:

All reactions involving air sensitive compounds were carried out under inert atmosphere (Ar or N_2 , \geq 99.99%). All glassware was oven-dried (120 °C), evacuated and purged with nitrogen. All common reagents and solvents were obtained from commercial suppliers and used without any further purification unless otherwise indicated. Gold catalysts Ph3PAuNTf2,1 (p-CF₃C₆H₄)₃PAuNTf₂,¹ JohnphosAuNTf₂,⁹ IPrAuNTf₂,¹⁰ IPrAuCl¹¹ were prepared following previously reported literature procedures, and some of them were lyophilized prior to use (see Table ESI-1 for each case; see below for lyophilisation procedure). AgOTs was purchased from commercial supplies and, in some cases, lyophilized prior to use. Solvents were dried by standard methods.¹² Hexane (HxH), ethyl acetate and triethylamine were purchased as extra pure grade reagents and used as received. TLC was performed on aluminium-backed plates coated with silica gel 60 with F₂₅₄ indicator; the chromatograms were visualized under ultraviolet light and by staining with KMnO4 reagent and subsequent heating. Rf values are reported on silica gel. Column chromatography was carried out on silica gel 60, 230-240 mesh. Routine NMR measurements were recorded on Bruker AV-400 or DPX-300 spectrometers. The values of δ are expressed in ppm and referred to the residual signal of the solvent (CDCl₃). The coupling constants values J are expressed in Hz. ¹H NMR: splitting pattern abbreviations are: s, singlet; d, doublet; t, triplet; q, quartet; p, pentet; sext, sextet; at, apparent triplet; dd, double doublet; ddd, double doublet of doublet; m, multiplet. ¹³C NMR: multiplicities were determined by DEPT, abbreviations are: q, CH₃; t, CH₂; d, CH; s quaternary carbons, except for compounds bearing fluorine atoms 1g and 2g (see below). For these compounds, the abbreviation (q) regarding the carbon multiplicity refers to the F-C coupling and there will be no abbreviation if there is no F-C coupling; the number of hydrogen atoms linked to a determined carbon atom is indicated as C, CH, CH₂ or CH₃. In the cases where a mixture of two diastereomers was observed, the abbreviation "min" refers to the signals assigned to the minor diastereomer and the abbreviation "maj" to the signals belonging to the major one; in the cases where nothing is specified, either it hasn't been possible to assign the signal to any of the diastereomers or it belongs to both of them. COSY, HSQC, HMBC and NOESY experiments were carried out on a Bruker AV-400 spectrometer. Standard pulse sequences were employed for the DEPT experiments. FT-IR experiments were performed with a Varian 1000 FT-IR spectrometer using sodium chloride plates; samples were measured as films from dichloromethane. Mass spectra were determined by Universidad de Vigo (CACTI) with a VG AutoSpec M and Universidad de Burgos with a Micromass Autospec mass spectrometers respectively for high resolution mass spectra (HRMS); low resolution mass spectra were obtained on a gas chromatograph mass spectrometer Shimadzu QP2010 Plus with auto injector AOC-20i. Electron ionization (70 eV) or electrospray ionization (ESI) were employed.

3.2. Catalyst Lyophilisation Procedure.

Some catalysts were lyophilized prior to their use (see manuscript Table 1) by placing them into an Alpha 2-4 LSC (Christ) lyophilisation (freeze-drying) apparatus, and following a three-step procedure: (1) *manual freezing* + *warm up*, for 20 min.; (2) *main drying*, for 2 hours and (3) *final drying* for 12 hours.

The lyophilisation is essential to remove water from hygroscopic silver salts.

3.3. Synthetic Procedures and Compounds Characterization Data.

 Table ESI-3
 Synthesis of Alkynylcyclopropanecarboxylic Acids 1



^a Isolated yield. ^b Diastereomeric ratio: the major isomer has a *cis* relationship between alkynyl and ester substituents. ^c Isolated yield for the two-step sequence from Fischer carbene complexes 17.^d The TMS group is lost in the basic hydrolysis step (1n R = H).

General procedure for the synthesis of alkynylcyclopropanecarboxylic acids 1:

In a sealed tube methyl acrylate (15 equiv) was added to a solution of the corresponding Fischer alkoxy alkynylcarbene complex¹³ **17** (1 equiv, 5 mmol) in dry THF (100 mL). The mixture was stirred at 90° C until complete disappearance of the carbene complex was observed by TLC (~12 h). Solvent was removed under reduced pressure, and the residue was purified by flash chromatography (hexane/AcOEt); the corresponding alkynylcyclopropane ester **18** was isolated. Alkenes, resulting from the opening of the cyclopropane ring, were obtained as minor byproducts² and, in general, they could not be separated from alkynylcyclopropane esters **18**. In particular cases (for instance, for model compound **18a**), chromatographic separation of both

diastereomers was achieved at this stage. Isolated yields and diastereomeric ratios of alkynylcyclopropane esters are listed in previous table ESI-3.

LiOH·H₂O (6 equiv, 18 mmol, 755 mg) was added to a solution of the corresponding alkynylcyclopropanecarboxylic ester 18 (3 mmol) in MeOH/H₂O 1:1 mixture (50 mL) in a 250 mL flask. The mixture was stirred during 12 hr at room temperature, the final of the reaction was verified by TLC. The solution was cleaned with ether (2 x 25 mL), the resulting aqueous layer was neutralized with HCl 1N and was extracted employing ether (3 x 25 mL). In most cases the resulting alkynylcyclopropanecarboxylic acid 1 did not require further purification; needed, crude was purified by flash chromatography when the employing hexane/AcOEt/formic acid mixtures of variable composition. Epimerization was not observed during basic hydrolysis.



(1*S**,2*S**)-2-methoxy-2-(phenylethynyl)cyclopropanecarboxylic acid (1a). Yellow oil; isolated yield for the two-step sequence = 46 %; the major *cis*-diastereomer could be separated by column chromatography at the ester stage (after the first reaction of the synthesis). The following data correspond to the major *cis*-diastereomer: $R_f = 0.00$ (HxH/AcOEt, 5/1); ¹H NMR (300 MHz, CDCl₃) $\delta = 11.2$ (bs, 1 H), 7.47-7.41 (m, 2 H), 7.32- 7.26 (m, 3 H), 3.50 (s, 3 H), 2.18-2.09 (m, 1 H), 1.73 (dd, *J* = 7.2, 5.7 Hz, 1 H), 1.58 (dd, *J* = 9.3, 5.7 Hz, 1 H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 175.3$ (s), 132.2 (d, 2 CH), 128.8 (d), 128.7 (d, 2 CH), 122.8 (s), 86.9 (s), 84.0 (s), 60.0 (s), 56.4 (q), 30.6 (d), 22.9 (t); LRMS: (EI) *m/z* (%) = 216 ([M]⁺, 6), 171 (72), 141 (33), 129 (100), 115 (32); HRMS: (EI) calculated for C₁₃H₁₂O₃ [M]⁺: 216.0781, found: 216.0781.



 $(1S^*, 2S^*)$ -2-methoxy-2-[(4-methoxyphenyl)ethynyl]cyclopropanecarboxylic acid (1b). Yellow liquid; isolated yield for the two-step sequence = 46 %, of a 5:1 mixture of diasteroisomers (a 7 % of alkene carboxylic acid remains after purification); $R_f = 0.20$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) δ = 10.43 (bs, 2 H, min + maj), 7.38 (d, J = 8.9 Hz, 2 H, min), 7.35 (d, J = 8.8 Hz, 2 H, maj), 6.84 (d, J = 8.9 Hz, 2 H, min), 6.80 (d, J = 8.8 Hz, 2 H, maj), 3.81 (s, 3 H, min), 3.78 (s, 3 H, maj), 3.50 (s, 3 H, maj), 3.46 (s, 3 H, min), 2.22 (dd, J = 8.7, 7.4 Hz, 1 H, min), 2.17 (dd, J = 9.2, 7.4 Hz, 1 H, maj), 1.95 (dd, J= 7.4, 6.0 Hz, 1 H, min), 1.79 (dd, J = 7.4, 5.9 Hz, 1 H, maj), 1.68 (dd, J = 9.2, 5.9 Hz, 1 H, maj), 1.59 (dd, J = 8.7, 6.0 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.5$ (s, maj), 174.6 (s, min), 160.0 (s, min), 159.8 (s, maj), 133.4 (d, 2 CH, min), 133.3 (d, 2 CH, maj), 114.4 (s, min), 114.3 (s, maj), 114.0 (d, 2 CH min), 113.9 (d, 2 CH, maj), 86.7 (s, maj), 84.7 (s, min), 84.3 (s, min), 81.4 (s, maj), 60.2 (s, maj), 59.3 (s, min), 56.3 (q, min), 56.0 (q, maj), 55.3 (q, min), 55.2 (q, maj), 29.7 (d, min), 29.5 (d, maj), 22.7 (t, maj), 22.2 (t, min); LRMS: (EI) m/z $(\%) = 246 ([M]^+, 15), 201 (69), 173 (25), 159 (100), 144 (26); HRMS: (EI) calculated for$ C₁₄H₁₄O₄ [M]⁺: 246.0892, found: 246.0895.



(1S*,2S*)-2-methoxy-2-[(3-methoxy-4-methylphenyl)ethynyl]cyclopropanecarboxylic acid
(1c). Yellow liquid; isolated yield for the two-step sequence = 51 % of a 4:1 mixture of

diastereoisomers; $R_f = 0.25$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 11.12$ (bs, 2 H, min + maj), 7.08 (d, J = 7.8 Hz, 1 H, min), 7.03 (d, J = 7.7 Hz, 1 H, maj), 6.98 (d, J = 7.8 Hz, 1 H, min), 6.95 (d, J = 7.7 Hz, 1 H, maj), 6.90 (s, 1 H, min), 6.85 (s, 1 H, maj), 3.84 (s, 3 H, min), 3.81 (s, 3 H, maj), 3.52 (s, 3 H, maj), 3.47 (s, 3 H, min), 2.26 – 2.15 (m + s, 8 H, min + maj), 1.96 (at, J = 6.4 Hz, 1 H, min), 1.81 (at, J = 6.6 Hz, 1 H, maj), 1.70 (dd, J = 9.3, 5.8 Hz, 1 H, maj), 1.61 (dd, J = 8.7, 5.9 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.9$ (s, maj), 174.8 (s, min), 157.4 (s, min), 157.3 (s, maj), 130.6 (d, min), 130.4 (d, maj), 128.3 (s, min), 127.9 (s, maj), 124.2 (d, min), 124.0 (d, maj), 120.4 (s, maj), 120.1 (s, min), 113.0 (d, min), 112.9 (d, maj), 56.1 (q, maj), 55.3 (q, min), 55.2 (q, maj), 29.6 (d, maj), 28.1 (d, min), 22.8 (t, maj), 22.1 (t, min), 16.3 (q, min), 16.2 (q, maj); LRMS: (EI) m/z (%) = 260 ([M]⁺, 5), 215 (30), 173 (100), 158 (19); HRMS: (EI) calculated for C₁₅H₁₆O4 [M]⁺: 260.1049, found: 260.1056.



(1*S**, 2*S**)-2-methoxy-2-(*p*-tolylethynyl)cyclopropanecarboxylic acid (1d). Yellow liquid; isolated yield for the two-step sequence = 41 % of a 5:1 mixture of diasteroisomers (a 6 % of alkene carboxylic acid remains after purification); $R_f = 0.22$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 11.17$ (bs, 2 H, min + maj), 7.36 (d, *J* = 7.8 Hz, 2 H, min), 7.33 (d, *J* = 7.8 Hz, 2 H, maj), 7.14 (d, *J* = 7.8 Hz, 2 H, min), 7.09 (d, *J* = 7.8 Hz, 2 H, maj), 3.53 (s, 3 H, maj), 3.48 (s, 3 H, min), 2.37 (s, 3 H, min), 2.34 (s, 3 H, maj), 2.28 – 2.23 (m, 1 H, min), 2.23 – 2.17 (m, 1 H, maj), 2.00 – 1.97 (m, 1 H, min), 1.84 – 1.81 (m, 1 H, maj), 1.71 (dd, *J* = 9.1, 5.8 Hz, 1 H, maj), 1.61 (dd, *J* = 8.3, 6.1 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) δ = 175.7 (s, maj), 174.8 (s, min), 139.0 (s, min), 138.6 (s, maj), 131.8 (d, 2 CH, min), 131.7 (d, 2 CH, maj), 129.2 (d, 2 CH, min), 129.0 (d, 2 CH, maj), 119.2 (s, maj), 118.9 (s, min), 86.9 (s, maj), 85.4 (s, min), 84.5 (s, min), 82.2 (s, maj), 60.2 (s, maj), 59.3 (s, min), 56.4 (q, min), 56.1 (q, maj), 29.7 (d, min), 29.6 (d, maj), 22.8 (t, maj), 22.2 (t, min), 21.53 (q, min), 21.47 (q, maj); LRMS: (EI) m/z (%) = 230 ([M]⁺, 14), 197 (23), 185 (40), 169 (52), 143 (100), 114 (9); HRMS: (EI) calculated for C₁₄H₁₄O₃ [M]⁺: 230.0943, found: 230.0943.



(15*, 25*)-2-methoxy-2-(*o*-tolylethynyl)cyclopropanecarboxylic acid (1e). Yellow liquid; isolated yield for the two-step sequence = 54 % of a 6.7:1 mixture of diasteroisomers (a 4 % of alkene carboxylic acid remains after purification); $R_f = 0.24$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) δ = 11.52 (bs, 2 H, min + maj), 7.42 (dd, J = 7.6, 1.0 Hz, 1 H, maj + 1 H, min), 7.29 – 7.10 (m, 6 H, min + maj), 3.56 (s, 3 H, maj), 3.51 (s, 3 H, min), 2.46 (s, 3 H, min), 2.41 (s, 3 H, maj), 2.27 (dd, J = 8.6, 7.2 Hz, 1 H, min), 2.23 (dd, J = 9.3, 7.5 Hz, 1 H, maj), 2.02 (dd, J = 7.2, 5.9 Hz, 1 H, min), 1.85 (dd, J = 7.5, 5.8 Hz, 1 H, maj), 1.73 (dd, J = 9.3, 5.8 Hz, 1 H, maj), 1.63 (dd, J = 8.8, 5.9 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) δ = 175.8 (s, maj), 174.8 (s, min), 140.4 (s, min), 140.3 (s, maj), 122.1 (d, maj), 132.08 (d, min), 129.5 (d, min), 129.4 (d, maj), 128.8 (d, min), 128.5 (d, maj), 125.7 (d, min), 125.5 (d, maj), 122.1 (s, maj), 121.8 (s, min), 90.1 (s, min), 86.7 (s, maj), 85.7 (s, maj), 83.3 (s, min), 60.3 (s, maj), 59.4 (s, min), 56.4 (q, min), 56.1 (q, maj), 29.9 (d, min), 29.7 (d, maj), 22.9 (t, maj), 22.4 (t, min), 20.7 (q, min), 20.5 (q, maj); LRMS: (EI) m/z (%) = 230 ([M]⁺, 5), 185 (37), 169 (22), 143 (100), 115 (98); HRMS: (EI) calculated for C₁₄H₁₄O₃ [M]⁺: 230.0943, found: 230.0944.



(1*S**, 2*S**)-2-[(4-chlorophenyl)ethynyl]-2-methoxycyclopropanecarboxylic acid (1f). Yellow liquid; isolated yield for the two-step sequence = 45 % of a 4:1 mixture of diasteroisomers (a 3 % of alkene carboxylic acid remains after purification); $R_f = 0.27$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 10.46$ (bs, 2 H, min + maj), 7.37 (d, J = 8.3 Hz, 2 H, min), 7.32 (d, J = 8.3 Hz, 2 H, maj), 7.24 (d, J = 8.3, 2 H, maj + 2 H, min), 3.50 (s, 3 H, maj), 3.46 (s, 3 H, min), 2.28 – 2.20 (m, 1 H, min), 2.23 – 2.17 (m, 1 H, maj), 2.01 - 1.94 (m, 1 H, min), 1.86 - 1.79 (m, 1 H, maj), 1.71 (dd, J = 9.2, 5.9 Hz, 1 H, maj), 1.60 (dd, J = 8.7, 6.1 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.6$ (s, maj), 174.5 (s, min), 134.9 (s, min), 134.6 (s, maj), 133.1 (d, 2 CH, min), 132.9 (d, 2 CH, maj), 128.7 (d, 2 CH, min), 128.6 (d, 2 CH, maj), 120.7 (s, maj), 120.4 (s, min), 87.0 (s, min), 85.6 (s, maj), 84.0 (s, maj), 83.2 (s, min), 60.1 (s, maj), 59.0 (s, min), 56.5 (q, min), 56.2 (q, maj), 29.7 (d, min), 29.6 (d, maj), 22.8 (t, maj), 22.1 (t, min); LRMS: (EI) m/z (%) = 250 ([M]⁺, 1), 236 (4), 205 (23), 165 (30), 163 (100); HRMS: (EI) calculated for $C_{13}H_{11}ClO_3$ [M]⁺: 250.0397, found: 250.0406.



(1*S**, 2*S**)-2-methoxy-2-{[4-(trifluoromethyl)phenyl]ethynyl}cyclopropanecarboxylic acid (1g). Yellow liquid; isolated yield for the two-step sequence = 55 % of a 4.5:1 mixture of diasteroisomers (a 3 % of alkene carboxylic acid remains after purification); $R_f = 0.21$

(HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (401 MHz, CDCl₃) δ = 10.53 (bs, 2 H, min + maj), 7.61 – 7.45 (m, 8 H, maj + min), 3.50 (s, 3 H, maj), 3.46 (s, 3 H, min), 2.27 – 2.17 (m, 2 H, min + maj), 2.00 – 1.96 (m, 1 H, min), 1.86 – 1.80 (m, 1 H, maj), 1.71 (dd, J = 9.2, 6.0 Hz, 1 H, maj), 1.60 (dd, J = 8.6, 6.1 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) δ = 175.0 (C, maj), 173.8 (C, min), 132.0 (2 CH, min), 131.8 (2 CH, maj), 130.1 (q, ² J_{C-F} = 32.7 Hz, C, maj), 126.0 (C, maj), 125.8 (C, min), 125.3 (q, ³ J_{C-F} = 3.6 Hz, C, min), 125.1 (q, ³ J_{C-F} = 3.6 Hz, C, maj), 124.6 (q, ¹ J_{C-F} = 271.4 Hz, C, maj), 88.6 (C, min), 85.8 (C, maj), 85.2 (C, maj), 82.9 (C, min), 59.9 (C, maj), 58.8 (C, min), 56.4 (CH₃, min), 56.2 (CH₃, maj), 29.7 (CH, min), 29.6 (CH, maj), 22.7 (CH₂, maj), 21.9 (CH₂, min) [two signals corresponding to -CF₃ and to F₃C-<u>C</u>- of the minor isomer were not observed]; LRMS: (EI) *m/z* (%) = 284 ([M]⁺, 3), 239 (83), 209 (24), 197 (100); HRMS: (EI) calculated for C₁₄H₁₁O₃F₃ [M]⁺: 284.0660, found: 284.0664.



(1*S**, 2*S**)-2-methoxy-2-(thiophen-2-ylethynyl)cyclopropanecarboxylic acid (1h). Yellow liquid; isolated yield for the two-step sequence = 55 % of a 5:1 mixture of diasteroisomers; $R_f = 0.17$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 8.83$ (bs, 2 H, min + maj), 7.29 (dd, *J* = 5.1, 1.1 Hz, 1 H, min), 7.24 (dd, *J* = 5.1, 1.1 Hz, 1 H, maj + 1 H min), 7.20 (dd, *J* = 3.6, 1.1 Hz, 1 H, maj), 6.98 (dd, *J* = 5.1, 3.7 Hz, 1 H, min), 6.95 (dd, *J* = 5.1, 3.6 Hz, 1 H, maj), 3.49 (s, 3 H, maj), 3.46 (s, 3 H, min), 2.24 (dd, *J* = 8.9, 7.4 Hz, 1 H, min), 2.19 (dd, *J* = 9.3, 7.4 Hz, 1 H, maj), 1.96 (dd, *J* = 7.4, 6.0 Hz, 1 H, min), 1.80 (dd, *J* = 7.4, 5.9 Hz, 1 H, maj), 1.70 (dd, *J* = 9.3, 5.9 Hz, 1 H, maj), 1.60 (dd, *J* = 8.9, 6.0 Hz, 1 H, min); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.1$ (s, min), 175.0 (s, maj), 133.0 (d, min), 132.7 (d, maj), 127.9 (d, min), 127.6 (d, maj), 127.0 (d, min), 126.9 (d, maj), 122.3 (s, min), 122.2 (s, maj), 86.9 (s, maj), 80.0 (s, maj), 60.1 (s, maj), 59.4 (s, min), 56.5 (q, min), 56.2 (q, maj), 29.64 (d, min), 29.60 (d, maj),

22.9 (t, maj), 22.2 (t, min) [the signals corresponding to *sp* carbons belonging to the minor isomer were not observed]; LRMS: (ESI) m/z (%) = 223 ([M+1]⁺, 25), 209 (100), 191 (17); HRMS: (ESI) calculated for C₁₁H₁₁O₃S [M+1]⁺: 223.0423, found: 223.0434.



2S*)-(E)-2-methoxy-2-(3-methyl-4-phenylbut-3-en-1-yn-1-yl)cyclopropane-(1*S**, carboxylic acid (1i). Yellow liquid; isolated yield for the two-step sequence = 44 % of a 4:1 mixture of diasteroisomers (a 8 % of alkene carboxylic acid remains after purification); $R_f =$ 0.30 (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) δ = 9.60 (bs, 2 H, min + maj), 7.40 – 7.23 (m, 10 H, min + maj), 6.87 (s, 1 H, min), 6.84 (s, 1 H, maj), 3.49 (s, 3 H, maj), 3.47 (s, 3 H, min), 2.18 (dd, J = 9.2, 7.5 Hz, 1 H, maj + 1 H, min), 2.08 (s, 3 H, min), 2.05 (s, 3 H, maj), 1.95 (dd, J = 7.5, 5.9 Hz, 1 H, min), 1.79 (dd, J = 7.5, 5.8 Hz, 1 H, maj), 1.68 (dd, J = 9.2, 5.8 Hz, 1 H, maj), 1.58 (dd, J = 8.8, 5.9 Hz, 1 H, min); ¹³C NMR (100 MHz, $CDCl_3$) $\delta = 175.5$ (s, maj), 174.5 (s, min), 137.0 (d, min), 136.6 (d, maj), 129.1 (d, 2 CH, min), 129.0 (d, 2 CH, maj), 128.44 (s, min) 128.36 (d, 2 CH, min), 128.3 (d, 2 CH, maj), 128.2 (s, maj), 127.5 (d, min), 127.3 (d, maj), 118.9 (s, maj), 118.7 (s, min), 90.5 (s, maj), 88.1 (s, min), 85.0 (s, min), 81.9 (s, maj), 60.2 (s, maj), 59.3 (s, min), 56.3 (q, min), 56.0 (q, maj), 29.7 (d, maj), 25.6 (d, min), 22.8 (t, maj), 22.2 (t, min), 19.1 (q, min), 19.0 (q, maj). LRMS: (EI) m/z $(\%) = 256 ([M]^+, 3), 224 (12), 181 (20), 169 (43), 141 (76), 115 (100); HRMS: (EI) calculated$ for C₁₆H₁₆O₃ [M]⁺: 256.1099, found: 256.1093.



(1*S**,2*S**)-2-(hex-1-yn-1-yl)-2-methoxycyclopropanecarboxylic acid (1j). Yellow liquid; isolated yield for the two-step sequence = 37 % of a 4:1 mixture of diasteroisomers; $R_f = 0.28$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 11.36$ (bs, 2 H, min + maj), 3.39 (s, 3 H, maj), 3.38 (s, 3 H, min), 2.36 (t, *J* = 7.1 Hz, 2 H, min), 2.22 (t, *J* = 6.8 Hz, 2 H, maj), 2.10 – 1.97 (m, 2 H, maj + min), 1.89 – 1.76 (m, 1 H, min), 1.66 – 1.58 (m, 2 H, maj + min), 1.53 (dd, *J* = 9.1, 6.0 Hz, 1 H maj + 1 H min), 1.49 – 1.32 (m, 8 H, maj + min), 0.98 – 0.79 (m, 6 H, maj + min); ¹³C NMR (101 MHz, CDCl₃) $\delta = 175.9$ (s, maj), 174.9 (s, min), 87.7 (s, maj), 85.3 (s, min), 80.3 (s, min), 73.5 (s, maj), 60.0 (s, maj), 59.1 (s, min), 56.0 (q, min), 55.6 (q, maj), 30.5 (d, maj), 29.6 (t, min), 29.3 (d, min), 29.1 (t, maj), 22.3 (t, maj), 22.0 (t, min), 21.9 (t, min), 21.8 (t, maj), 18.4 (t, maj), 18.3 (t, min), 13.5 (q, maj), 13.4 (q, min); LRMS: (ESI) *m/z* (%) = 197 ([M+1]⁺, 32), 183 (71), 165 (100); HRMS: (ESI) calculated for C₁₁H₁₇O₃ [M+1]⁺: 197.1178, found: 197.1167.



(1*S**,2*S**)-2-(cyclopentylethynyl)-2-methoxycyclopropanecarboxylic acid (1k). Pale yellow liquid; isolated yield for the two-step sequence = 39 % of a 5:1 mixture of diasteroisomers; the following data correspond to the major diastereomer, which was separated by column chromatography. $R_f = 0.23$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 11.06$ (bs, 1 H), 3.41 (s, 3 H), 2.66 (p, *J* = 7.2 Hz, 1 H), 2.04 (dd, *J* = 9.2, 7.3 Hz, 1 H), 1.94 – 1.83 (m, 2 H), 1.73 - 1.69 (m, 2 H), 1.64 (dd, *J* = 7.3, 5.8 Hz, 1 H), 1.66 – 1.51 (m, 4 H), 1.55 (dd, *J* = 9.2, 5.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.9$ (s), 92.0 (s), 73.0 (s), 60.0 (s), 55.7 (q), 33.74 (t), 33.67 (t), 30.1 (d), 29.2 (d), 24.91 (t), 24.89 (t), 22.4 (t).



1*S**,2*S**)-2-(3,3-dimethylbut-1-yn-1-yl)-2-methoxycyclopropanecarboxylic acid (11). Colourless liquid; isolated yield for the two-step sequence = 57 % of a 5:1 mixture of diasteroisomers (a 5 % of alkene carboxylic acid remains after purification); $R_f = 0.34$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 10.81$ (bs, 2 H, min + maj), 3.34 (s, 3 H, maj), 3.32 (s, 3 H, min), 1.98 (dd, *J* = 9.2, 7.3 Hz, 1 H, maj + 1 H, min), 1.77 (dd, *J* = 7.2, 5.9 Hz, 1 H, min), 1.57 (dd, *J* = 7.3, 5.8 Hz, 1 H, maj), 1.48 (dd, *J* = 9.2, 5.8 Hz, 1 H, maj), 1.36 (dd, *J* = 8.7, 5.9 Hz, 1 H, min), 1.17 (s, 9 H, min), 1.15 (s, 9 H, maj); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.5$ (s, maj), 174.6 (s, min), 95.6 (s, maj), 93.3 (s, min), 75.5 (s, maj), 72.0 (s, maj), 59.8 (s, maj), 58.9 (s, min), 55.7 (q, min), 55.4 (q, maj), 30.73 (q, 3 CH₃, min), 30.69 (q, 3 CH₃, maj), 29.3 (d, min), 29.2 (d, maj), 27.4 (s, maj), 27.3 (s, min), 22.2 (t, maj), 22.0 (t, min); LRMS: (EI) *m/z* (%) = 196 ([M]⁺, 2), 181 (100), 151 (88), 109 (88), 93 (87), 91 (93), 79 (97); HRMS: (EI) Calculated for C₁₁H₁₆O₃ [M]⁺: 196.1099, found: 196.1093.



(1S*,2S*)-2-methoxy-2-[(*E*)-4-phenylbut-3-en-1-yn-1-yl]cyclopropanecarboxylic acid (1m). Yellow liquid; isolated yield for the two-step sequence = 45 % of a 3:1 mixture of diasteroisomers (a 10 % of alkene carboxylic acid remains after purification); $R_f = 0.23$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (401 MHz, CDCl₃) $\delta = 10.95$ (bs, 2 H, min + maj), 7.43 - 7.24 (m, 10 H, min + maj), 6.99 (d, *J* = 16.3 Hz, 1 H, min), 6.98 (d, *J* = 16.3 Hz, 1 H, maj), 6.31 (d, *J* = 16.3 Hz, 1 H, min), 6.18 (d, *J* = 16.3 Hz, 1 H, maj), 3.49 (s, 3 H, maj), 3.47 (s, 3 H, min), 2.24 - 2.20 (m, 1 H, min), 2.21 - 2.17 (m, 1 H, maj), 1.99 - 1.95 (m, 1 H, min), 1.81 - 1.78 (m, 1 H, maj), 1.69 (dd, J = 9.2, 5.9 Hz, 1 H, maj), 1.57 (dd, J = 8.8, 6.0 Hz, 1 H, min); ¹³C NMR (101 MHz, CDCl₃) $\delta = 175.7$ (s, maj), 174.7 (min), 142.5 (d, min), 142.3 (d, maj), 136.1 (s, maj), 135.9 (s, min), 129.1 (d, min), 129.0 (d, 2 CH, min), 128.8 (d, maj), 128.7 (d, 2 CH, maj), 126.5 (d, 2 CH, min), 126.4 (d, 2 CH, maj), 107.1 (d, maj), 106.8 (d, min), 88.2 (s, min), 86.1 (s, maj), 85.0 (s, maj), 83.7 (s, min), 60.3 (s, maj), 59.3 (s, min), 56.4 (q, min), 56.1 (q, maj), 29.8 (d, min), 29.7 (d, maj), 22.9 (t, maj), 22.2 (t, min); LRMS: (EI) *m/z* (%) = 242 ([M]⁺, 11), 209 (19), 197 (45), 181 (37), 165 (85), 153 (85), 127 (100); HRMS: (EI) calculated for C₁₅H₁₄O₃ [M]⁺: 242.0943, found: 242.0945.



(1*S**,2*S**)-2-ethynyl-2-methoxycyclopropanecarboxylic acid (1n). Obtained according to the general procedure by employing the corresponding silylated carbene complex 17n (R = Me₃Si). Desilylation takes place during basic hydrolysis. Colourless liquid; isolated yield for the two-step sequence = 58 % of a 10:1 mixture of diasteroisomers; R_f = 0.32 (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) δ = 10.66 (bs, 2 H, min + maj), 3.42 (s, 3 H, maj), 3.41 (s, 3 H, min), 2.56 (s, 1 H, maj), 2.50 (s, 1 H, min), 2.12 (dd, *J* = 8.8, 7.4 Hz, 1 H, min), 2.08 (dd, *J* = 9.3, 7.4 Hz, 1 H, maj), 1.85 (dd, *J* = 7.4, 6.0 Hz, 1 H, min), 1.68 (dd, *J* = 7.4, 5.9, 1 H, maj), 1.59 (dd, *J* = 9.3, 5.9 Hz, 1 H, maj), 1.49 (dd, *J* = 8.8, 6.0 Hz, 1 H, min); ¹³C NMR (101 MHz, CDCl₃) δ = 175.3 (s, maj), 174.3 (s, min), 80.7 (s, min), 77.4 (s, maj), 75.2 (d, maj), 74.5 (d, maj), 22.3 (t, maj), 21.6 (t, min). LRMS: (EI) *m*/*z* (%) = 140 ([M]⁺, 8), 125 (36), 115 (100), 95 (63), 83 (83); HRMS: (EI) calculated for C₇H₈O₃ [M]⁺: 140.0473, found: 140.0479.

General procedure for the synthesis of oxepin-2-ones 2:

A solution of freshly lyophilized IPrAuCl (3 mol%, 0.015 mmol, 9.3 mg) and AgOTs (3 mol%, 0.015 mmol, 4.2 mg) in CH₂Cl₂ (5 mL) was placed in a carrousel tube under Ar atmosphere and protected from light. The mixture was stirred during 30 min, and a solution of the corresponding alkynylcyclopropanecarboxylic acid 1 (0.5 mmol) in CH₂Cl₂ (5 mL) was then added; stirring was maintained for 12 h. End of reaction was monitored by TLC; solvents were removed under vacuum and the crude reaction mixture was purified by flash chromatography using hexane/AcOEt (5:1) as eluent to give the corresponding oxepinone 2 in the yield indicated in Scheme 1 or in Table 1.



5-methoxy-7-phenyloxepin-2(*3H*)**-one** (**2a**). Yellow liquid; yield = 90 %; $R_f = 0.65$ (HxH/AcOEt, 3/1); FT-IR (film, cm⁻¹): v = 1760, 1643; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.70$ -7.65 (m, 2 H), 7.41-7.37 (m, 3 H), 6.42 (s, 1 H), 4.85 (t, J = 6.8 Hz, 1 H), 3.65 (s, 3 H), 3.00 (d, J = 6.8 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 166.9$ (s), 155.8 (s), 150.3 (s), 133.5 (s), 129.5 (d), 128.5 (d, 2 CH), 125.4 (d, 2 CH), 106.2 (d), 89.7 (d), 55.5 (q), 32.0 (t). LRMS: (EI) m/z (%) = 216 ([M]⁺, 10), 187 (14), 147 (22), 111 (17), 105 (100), 77 (97); HRMS: (EI) calculated for C₁₃H₁₂O₃ [M]⁺: 216.0781, found: 216.0780.



5-methoxy-7-(4-methoxyphenyl)oxepin-2(3*H***)-one (2b).** Pale yellow liquid; yield = 78 %; R_f = 0.33 (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) δ = 7.61 (d, *J* = 8.8 Hz, 2 H), 6.90 (d, *J*

= 8.8 Hz, 2 H), 6.30 (s, 1 H), 4.81 (t, J = 7.0 Hz, 1 H), 3.83 (s, 3 H), 3.64 (s, 3 H), 2.99 (d, J = 7.0 Hz, 2 H); ¹³C NMR (101 MHz, CDCl₃) δ = 167.1 (s), 160.8 (s), 156.1 (s), 150.5 (s), 127.0 (d, 2 CH), 126.1 (s), 114.0 (d, 2 CH), 104.6 (d), 89.2 (d), 55.5 (q), 55.4 (q), 32.2 (t); LRMS: (EI) m/z (%)= 246 ([M]⁺, 11), 218 (9), 135 (100), 92 (9); HRMS: (EI) calculated for C₁₄H₁₄O₄ [M]⁺: 246.0892, found: 246.0894.



5-methoxy-7-(3-methoxy-4-methylphenyl)oxepin-2(3*H***)-one (2c). Pale yellow liquid; yield = 84 %; R_f = 0.27 (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) \delta = 7.27 - 7.11 (m, 3 H), 6.39 (s, 1 H), 4.84 (t,** *J* **= 7.0 Hz, 1 H), 3.87 (s, 3 H), 3.65 (s, 3 H), 3.00 (d,** *J* **= 7.0 Hz, 2 H), 2.23 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) \delta = 167.1 (s), 157.8 (s), 156.1 (s), 150.8 (s), 132.5 (s), 130.7 (d), 128.8 (s), 117.7 (d), 106.9 (d), 105.6 (d), 89.6 (d), 55.6 (q), 55.4 (q), 32.2 (t), 16.1 (q).**



5-methoxy-7-(*p***-tolyl)oxepin-2(3***H***)-one (2d).** Colourless liquid; yield = 92 %; $R_f = 0.35$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.57$ (d, J = 8.3 Hz, 2 H), 7.19 (d, J = 8.3 Hz, 2 H), 6.38 (s, 1 H), 4.83 (t, J = 7.0 Hz, 1 H), 3.63 (s, 3 H), 2.99 (d, J = 7.0 Hz, 2 H), 2.37 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 167.1$ (s), 156.1 (s), 150.7 (s), 139.8 (s), 130.8 (s), 129.4 (d, 2 CH), 125.5 (d, 2 CH), 105.5 (d), 89.6 (d), 55.5 (q), 32.1 (t), 21.2 (q).



5-methoxy-7-(*o***-tolyl)oxepin-2(3***H***)-one (2e).** Colourless liquid; yield = 72 %; $R_f = 0.35$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.40$ (d, J = 7.8 Hz, 1 H), 7.34 – 7.19 (m, 3 H), 6.01 (s, 1 H), 4.84 (t, J = 7.0 Hz, 1 H), 3.65 (s, 3 H), 3.08 (d, J = 7.0 Hz, 2 H), 2.41 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 166.9$ (s), 155.7 (s), 152.9 (s), 136.2 (s), 134.8 (s), 130.8 (d), 129.5 (d), 129.4 (d), 125.9 (d), 110.3 (d), 89.5 (d), 55.5 (q), 32.3 (t), 20.6 (q); LRMS: (EI) m/z (%)= 230 ([M]⁺, 8), 161 (84), 119 (100), 115 (18), 91 (40); HRMS: (EI) calculated for $C_{14}H_{14}O_3$ [M]⁺: 230.0943, found: 230.0948.



7-(4-chlorophenyl)-5-methoxyoxepin-2(3*H*)-one (2f). Colourless liquid; yield = 75 %; $R_f = 0.38$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.61$ (d, J = 8.7 Hz, 2 H), 7.37 (d, J = 8.7 Hz, 2 H), 6.39 (s, 1 H), 4.86 (t, J = 7.0 Hz, 1 H), 3.65 (s, 3 H), 3.00 (d, J = 7.0 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 166.7$ (s), 155.8 (s), 149.5 (s), 135.7 (s), 132.1 (s), 128.9 (d, 2 CH), 126.8 (d, 2 CH), 106.6 (d), 90.1 (d), 55.6 (q), 32.1 (t).



5-methoxy-7-[4-(trifluoromethyl)phenyl]oxepin-2(3*H***)-one (2g). Colourless liquid; yield = 86 %; R_f = 0.39 (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) \delta = 7.79 (d, J = 8.1 Hz, 2 H), 7.65 (d, J = 8.2 Hz, 2 H), 6.49 (s, 1 H), 4.90 (t, J = 6.9 Hz, 1 H), 3.66 (s, 3 H), 3.02 (d, J = 6.9 Hz, 2 H); for this compound, the abbreviation (q) regarding the carbon multiplicity refers to the F-C coupling and there will be no abbreviation if there is no F-C coupling; the number of hydrogen atoms linked to a determined carbon atom is indicated as C, CH, CH₂ or CH₃: ¹³C NMR (100 MHz, CDCl₃) \delta = 166.5 (C), 155.7 (C), 149.0 (C), 137.0 (C), 131.3 (q, ²***J***_{C-F} = 32.7 Hz, C), 125.8 (2 CH), 125.7 (q, ³***J***_{C-F} = 3.6 Hz, 2 CH), 123.8 (q, ¹***J***_{C-F} = 272.1 Hz, C), 108.1 (CH), 90.6 (CH), 55.6 (CH₃), 32.1 (CH₂).**



5-methoxy-7-(thiophen-2-yl)oxepin-2(3*H***)-one (2h).** Colourless liquid; yield = 62 %; $R_f = 0.43$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.38$ (dd, J = 3.7 and 1.1 Hz, 1 H), 7.32 (dd, J = 5.1 and 1.1 Hz, 1 H), 7.04 (dd, J = 5.1 and 3.7 Hz, 1 H), 6.31 (s, 1 H), 4.82 (t, J = 7.0 Hz, 1 H), 3.64 (s, 3 H), 3.03 (d, J = 7.0 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 166.6$ (s), 155.8 (s), 146.3 (s), 137.6 (s), 127.9 (d), 127.0 (d), 126.0 (d), 105.0 (d), 89.6 (d), 55.6 (q), 32.1 (t).



(*E*)-5-methoxy-7-(1-phenylprop-1-en-2-yl)oxepin-2(3*H*)-one (2i). Colourless liquid; yield = 82 %; $R_f = 0.50 (HxH/AcOEt, 5/1)$; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.38 - 7.26 (m, 6 H)$, 6.09

(s, 1 H), 4.84 (t, J = 6.9 Hz, 1 H), 3.63 (s, 3 H), 2.97 (d, J = 6.9 Hz, 2 H), 2.07 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 167.6$ (s), 156.0 (s), 152.3 (s), 136.9 (s), 130.3 (d), 129.5 (d, 2 CH), 129.3 (s), 128.2 (d, 2 CH), 127.3 (d), 107.2 (d), 90.2 (d), 55.5 (q), 32.0 (t), 14.7 (q); LRMS: (EI) m/z (%) = 256 ([M]⁺, 4), 187 (17), 159 (29), 145 (49), 117 (56), 115 (100); HRMS: (EI) calculated for C₁₆H₁₆O₃ [M]⁺: 256.1099, found: 256.1096.



7-butyl-5-methoxyoxepin-2(3*H*)-one (2j). Colourless liquid; yield = 89 %; $R_f = 0.55$ (HxH/AcOEt, 5/1); ¹H NMR (300 MHz, CDCl₃) $\delta = 5.64$ (s, 1 H), 4.68 (t, J = 7.0 Hz, 1 H), 3.57 (s, 3 H), 2.88 (d, J = 7.0 Hz, 2 H), 2.31 (t, J = 7.2 Hz, 2 H), 1.55 (p, J = 7.2 Hz, 2 H), 1.36 (sext, J = 7.2 Hz, 2 H), 0.92 (t, J = 7.2 Hz, 3 H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 167.5$ (s), 156.0 (s), 155.4 (s), 106.7 (d), 88.8 (d), 55.7 (q), 35.2 (t), 32.3 (t), 29.7 (t), 22.4 (t), 14.1 (q).



7-cyclopentyl-5-methoxyoxepin-2(*3H*)-one (2k). Colourless liquid; yield = 87 %; $R_f = 0.50$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 5.69$ (s, 1 H), 4.68 (t, J = 6.9 Hz, 1 H), 3.57 (s, 3 H), 2.86 (d, J = 6.9 Hz, 2 H), 2.71 (p, J = 8.2 Hz, 1 H); 1.91 – 1.88 (m, 2 H), 1.74 – 1.70 (m, 2 H), 1.65 – 1.60 (m, 4 H); ¹³C NMR (101 MHz, CDCl₃) $\delta = 167.3$ (s), 157.9 (s), 155.6 (s), 105.0 (d), 88.4 (d), 55.3 (q), 45.0 (d), 32.0 (t), 31.2 (t, 2 CH₂), 25.4 (t, 2 CH₂); LRMS: (EI) m/z (%) = 208 ([M]⁺, 2), 167 (100), 163 (61), 121 (70), 91 (86), 79 (65), 77 (56); HRMS: (EI) calculated for C₁₂H₁₆O₃ [M]⁺: 208.1099, found: 208.1096.



7-(*tert*-butyl)-5-methoxyoxepin-2(3*H*)-one (2l). Colourless liquid; yield = 74 %; $R_f = 0.53$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 5.74$ (s, 1 H), 4.70 (t, J = 6.9 Hz, 1 H), 3.58 (s, 3 H), 2.83 (d, J = 6.9 Hz, 2 H), 1.20 (s, 9 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 167.7$ (s), 162.5 (s), 155.6 (s), 103.7 (d), 88.7 (d), 55.3 (q), 36.7 (s), 31.8 (t), 28.2 (q, 3 CH₃).



(*E*)-5-methoxy-7-styryloxepin-2(3*H*)-one (2m). Colourless liquid; yield = 30 %; $R_f = 0.80$ (HxH/AcOEt, 1/1); ¹H NMR (300 MHz, CDCl₃) $\delta = 7.49 - 7.27$ (m, 5 H), 7.14 (d, *J* = 15.8 Hz, 1 H), 6.60 (d, *J* = 15.8 Hz, 1 H), 5.92 (s, 1 H), 4.83 (t, *J* = 7.0 Hz, 1 H), 3.64 (s, 3 H), 3.01 (d, *J* = 7.0 Hz, 2 H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 167.4$ (s), 156.4 (s), 150.2 (s), 136.2 (s), 132.7 (d), 129.2 (d, 2 CH), 129.1 (d), 127.5 (d, 2 CH), 122.4 (d), 110.4 (d), 90.5 (d), 55.9 (q), 32.5 (t); LRMS: (EI) *m/z* (%) = 242 ([M]⁺, 3), 173 (60), 131 (100), 103 (62); HRMS: (EI) calculated for $C_{15}H_{14}O_3$ [M]⁺: 242.0943, found: 242.0943



5-methoxyoxepin-2(3*H*)-one (2n). Not isolated. Data retrieved from a mixture of 2n:8 (54:46). Colourless liquid; combined yield (2n + 8) in the reaction carried out at rt, according to the general procedure = 30 %; combined yield (2n + 8) in the reaction carried out in refluxing dichloromethane = 71 %; R_f = 0.53 (HxH/AcOEt, 5/1. ¹H NMR (400 MHz, CDCl₃) δ = 6.57 (d, J = 7.2 Hz, 1 H), 5.79 (d, J = 7.2 Hz, 1 H), 4.77 (t, J= 7.0 Hz, 1 H), 3.60 (s, 3 H), 2.93 (d, J = 7.0 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ = 167.0 (s), 155.3 (s), 141.1 (d), 110.5 (d), 90.1 (d), 55.4 (q), 32.0 (t).

Procedure for the synthesis of oxepin-2-ones 2m or 2n in refluxing dichloromethane:

A solution of freshly lyophilized IPrAuCl (3 mol%, 0.015 mmol, 9.3 mg) and AgOTs (3 mol%, 0.015 mmol, 4.2 mg) in CH₂Cl₂ (5 mL) was placed in a carrousel tube under Ar atmosphere and protected from light. The mixture was stirred during 30 min, and a solution of the corresponding alkynylcyclopropanecarboxylic acid **1m** or **1n** (0.5 mmol) in CH₂Cl₂ (5 mL) was then added. The reaction tube was placed in an oil bath at 40 °C, and stirring was maintained for 12 h. End of reaction was monitored by TLC; solvents were removed under vacuum and the crude reaction mixture was purified by flash chromatography using hexane/AcOEt (5:1) as eluent to give the corresponding oxepinone **2m** or a mixture of oxepinone **2n** and (1*S**,5*S**)-5-methoxy-4-methylen-3-oxabicyclo[3.1.0]hexan-2-one **8** (ratio 52/48) in the yields indicated in Table 1 (entries 13 or 15).



5-Methoxy-5-(phenylethynyl)dihydrofuran-2(3*H***)-one (3). Obtained as a byproduct in some of the reactions during the optimization process (see Table ESI-1) and as major product when Ph_3PAuCl/AgSbF_6 was used as catalyst system. A solution of Ph_3PAuCl (3 mol%, 0.015 mmol, 7.4 mg) and AgSbF_6 (3 mol%, 0.015 mmol, 5.2 mg) in CH_2Cl_2 (5 mL) was put in a carrousel tube under Ar atmosphere and protected from light. The mixture was stirred during 30 min, and a solution of alkynylcyclopropanecarboxylic acid 1a** (0.5 mmol, 108 mg) in CH_2Cl_2 (5 mL)

was then added; stirring was maintained for 12 h. The end of the reaction was monitored by TLC, solvents were removed under vacuum and the crude reaction mixture was purified by flash chromatography using hexane/AcOEt (5:1) as eluent to give 5-methoxy-5-(phenylethynyl)dihydrofuran-2(3*H*)-one **3** in 54% yield (58 mg). Yellow liquid; $R_f = 0.55$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.50-7.48$ (m, 2 H), 7.39-7.33 (m, 3 H), 3.60 (s, 3 H), 2.79-2.53 (m, 4 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 175.1$ (s), 131.9 (d, 2 CH), 129.5 (d), 128.5 (d, 2 CH), 120.9 (s), 102.9 (s), 87.5 (s), 82.8 (s), 53.2 (q), 36.5 (t), 28.0 (t).



Dihydro-5-benzoylmethyl-5-methoxyfuran-2(3*H***)-one (4). Obtained as a byproduct in some of the reactions during the optimization process (see Table ESI-1). Isolated in 9% yield under the conditions reported in table ESI-1, entry 8. Yellow oil; R_f = 0.26 (HxH/AcOEt, 3/1); ¹H NMR (400 MHz, CDCl₃) \delta = 8.00 - 7.95 (m, 2 H), 7.67 - 7.60 (m, 1 H), 7.56 - 7.48 (m, 2 H), 3.73 (d, J = 17.1 Hz, 1 H), 3.61 (d, J = 17.1 Hz, 1 H), 3.42 (s, 3 H), 2.89 - 2.65 (m, 2 H), 2.63-2.43 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) \delta = 195.2 (s), 175.9 (s), 136.8 (s), 133.7 (d), 128.8 (d, 2 CH), 128.1 (d, 2 CH), 108.7 (s), 50.2 (q), 43.4 (t), 31.5 (t), 28.8 (t). LRMS: (EI)** *m/z* **(%) = 234 ([M]⁺, 5), 202 (23), 115 (81), 105 (100), 77 (97); HRMS: (EI) calculated for C₁₃H₁₄O₄ [M]⁺: 234.0892, found: 234.0895.**



4-Oxo-6-phenylhex-5-ynoic acid (5). Obtained as a byproduct in some of the reactions during the optimization process (see Table ESI-1) and as major product when $AgSbF_6$ was used as catalyst. A solution of $AgSbF_6$ (3 mol%, 0.015 mmol, 5.2 mg) in CH_2Cl_2 (5 mL) was put in a

carrousel tube under Ar atmosphere and protected from light. The mixture was stirred during 30 min, and a solution of alkynylcyclopropanecarboxylic acid **1a** (0.5 mmol, 108 mg) in CH₂Cl₂ (5 mL) was then added; stirring was maintained for 12 h. The end of the reaction was monitored by TLC, solvents were removed under vacuum and the crude reaction mixture was purified by flash chromatography using hexane/AcOEt (5:1) as eluent to give 4-oxo-6-phenylhex-5-ynoic acid **5** in 80% yield (162 mg). Yellow liquid; $R_f = 0.05$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.61 - 7.58$ (m, 2 H), 7.48 – 7.39 (m, 3 H), 3.04 (t, *J* = 6.6 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 185.1$ (s), 177.6 (s), 133.1 (d, 2 CH), 130.9 (d), 128.7 (d, 2 CH), 119.7 (s), 91.7 (s), 87.3 (s), 39.7 (t), 27.8 (t).



Methyl (Z)-6-methoxy-4-oxo-6-phenylhex-5-enoate (6). Obtained as a byproduct in some of the reactions during the optimization process (see Table ESI-1). Isolated in 15% yield under the conditions reported in Table ESI-1, entry 2. Yellow oil; $R_f = 0.10$ (HxH/AcOEt, 20/1); FT-IR (pure, cm⁻¹): v = 1732, 1651; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.92 - 7.88$ (m, 2 H), 7.55 - 7.42 (m, 3 H), 6.17 (s, 1 H), 3.78 (s, 3 H), 3.69 (s, 3 H), 3.20 (t, J = 7.6 Hz, 2 H), 2.65 (t, J = 7.6 Hz, 2 H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 189.9$ (s), 175.7 (s), 173.2 (s), 140.2 (s), 131.9 (d), 128.4 (d, 2 CH), 127.7 (d, 2 CH), 96.3 (d), 55.8 (q), 51.7 (q), 31.3 (t), 28.7 (t). LRMS: (EI) *m/z* (%) = 248 ([M]⁺, 1), 217 (4), 189 (9), 174 (100), 105 (83), 77 (26); HRMS: (EI) calculated for C₁₄H₁₆O₄ [M]⁺: 248.1043, found: 248.1044.



(1S*,6R*)-6-Methoxy-4-[(*E*)-styryl]-3-oxabicyclo[4.1.0]hept-4-en-2-one (7). Pale yellow liquid; isolated yield = 30 % (from alkynylcyclopropanecarboxylic acid 1m under the reaction conditions reported in the general procedure); R_f = 0.74 (HxH/AcOEt, 1/1); ¹H NMR (300 MHz, CDCl₃) δ = 7.44 – 7.27 (m, 5 H), 7.08 (d, *J* = 15.9 Hz, 1 H), 6.42 (d, *J* = 15.9 Hz, 1 H), 5.77 (bs, 1 H), 3.39 (s, 3 H), 2.36 (ddd, *J* = 11.2, 5.4, 1.4 Hz, 1 H), 2.01 (dd, *J* = 11.2, 5.4 Hz, 1 H), 1.17 (t, *J* = 5.4 Hz, 1 H); ¹³C NMR (75 MHz, CDCl₃) δ = 166.9 (s), 147.1 (s), 135.9 (s), 131.0 (d), 128.8 (d, 2 CH), 128.5 (d), 126.9 (d, 2 CH), 119.4 (d), 106.1 (d), 64.6 (s), 56.7 (q), 24.9 (d), 23.2 (t); LRMS: (EI) *m/z* (%)= 242 ([M]⁺, 1), 134 (22), 131 (100), 103 (61); HRMS: (EI) calculated for C₁₅H₁₄O₃ [M]⁺: 242.0943, found: 242.0954.



(1S*,5S*)-5-Methoxy-4-methylen-3-oxabicyclo[3.1.0]hexan-2-one (8). Not isolated. Data retrieved from a mixture of 2n:8 (54:46). Colourless liquid; combined yield (2n + 8) in the reaction carried out at rt, according to the general procedure = 30 %; combined yield (2n + 8) in the reaction carried out in refluxing dichloromethane = 71 %; R_f = 0.53 (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) δ = 4.90 (d, *J* = 2.5 Hz, 1 H), 4.64 (d, *J* = 2.5 Hz, 1 H), 3.45 (s, 3 H), 2.48 (dd, *J* = 10.0, 3.5 Hz, 1 H), 1.86 (dd, *J* = 10.0, 5.5 Hz, 1 H), 1.45 (dd, *J* = 5.5, 3.5 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.2 (s), 153.1 (s), 88.4 (t), 69.3 (s), 57.0 (q), 22.9 (d), 22.1 (t).



(1*S**,2*R**)-2-(phenylethynyl)cyclopropanecarboxylic acid (9). Jones reagent $(CrO_3/H_2SO_4/H_2O)$ dropwise added solution of $(1S^*, 2R^*)$ -2was to а (phenylethynyl)cyclopropylmethanol¹⁴ (1 mmol, 172 mg) at 0 °C till the orange colour persists in the solution. The reaction was stirred until disappearance of starting material was observed by TLC. Then, isopropanol was added till the orange colour vanished, and water (50 mL) was added. The reaction mixture was transferred to a separatory funnel and extracted with dichloromethane (2 x 20 mL). The combined organic layer was placed again in the separatory funnel and treated with 2% NaOH (2 x 20 mL). The combined basic aqueous layers are treated with concentrated HCl; cloudiness was observed. After dichloromethane extraction (2 x 15 mL), the combined organic layer was dried over sodium sulphate. Solvent was removed under vacuum. $(1S^*, 2R^*)$ -2-(phenylethynyl)cyclopropanecarboxylic acid 9 was isolated as a yellow liquid (182 mg, 98%), which did not require further purification. $R_f = 0.22$ (HxH/AcOEt/HCOOH, 5/1/0.01); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.39 - 7.36$ (m, 2 H), 7.27 -7.23 (m, 3 H), 2.16 - 2.03 (m, 2 H), 1.57 (td, J = 6.5, 4.8 Hz, 1 H), 1.38 (td, J = 8.3, 4.8 Hz, 1 H), the signal corresponding to the carboxylic acid proton was not observed. ¹³C NMR (75 MHz, CDCl3) $\delta = 176.7$ (s), 132.2 (d, 2 CH), 128.6 (d, 2 CH), 128.3 (d), 123.6 (s), 86.9 (s), 80.6 (s), 21.7 (d), 15.9 (t), 11.5 (d).



(1*R**,2*S**)-[2-methoxy-2-(phenylethynyl)cyclopropyl]methanol (10). Ester 18a [1 mmol, 230 mg, (used as a 6.7:1 mixture of diastereomers)] was dissolved in THF (10 mL) in a 50 mL round bottomed flask under inert atmosphere. Then LiBEt₃H (2 equiv) was added at -78 °C and the mixture was stirred for 12 hr. When end of reaction was observed by TLC, NaOH 1M (25

mL) was added. THF was removed by evaporation under reduced pressure, the crude was extracted with ether (2 x 25 mL) and the combined organic layer was dried with Na₂SO₄. The ether was removed by evaporation under reduced pressure leading to the final alcohol **10** as a colourless liquid (98 % yield, 198 mg of a 6.7:1 mixture of diastereomers), which was used without further purification. $R_f = 0.10$ (HxH/AcOEt, 5/1); ¹H NMR (300 MHz, CDCl₃) $\delta = 7.48$ – 7.44 (m, 4 H, min + maj), 7.37 – 7.31 (m, 6 H, min + maj), 4.04 – 3.95 (m, 1 H, min), 3.93 – 3.84 (m, 1 H, maj), 3.73 – 3.64 (m, 1 H, min), 3.58 (dd, J = 11.9, 9.0 Hz, 1 H, maj), 3.53 (s, 3 H, min), 3.48 (s, 3 H, maj), 1.93 (bs, 1 H, min), 1.83 (bs, 1 H, maj), 1.79 – 1.65 (m, 2 H, min + maj), 1.36 (dd, J = 10.0, 5.7 Hz, 1 H, maj), 1.21 (dd, J = 9.6, 5.6 Hz, 1 H, min), 1.02 (dd, J = 7.1, 5.6 Hz, 1 H, min), 0.97 (dd, J = 7.0, 5.7 Hz, 1 H, maj); ¹³C NMR (100 MHz, CDCl₃) $\delta = 131.73$ (d, 2 CH, maj), 131.70 (d, 2 CH, min), 128.8 (d, min), 128.5 (d, maj), 128.4 (d, 2 CH, maj), 128.3 (d, 2 CH, min), 122.7 (s, min), 122.4 (s, maj), 88.3 (s, min), 85.9 (s, maj), 85.7 (s, maj), 83.8 (s, min), 63.4 (t, maj), 62.6 (s, min), 61.4 (t, min), 57.1 (s, maj), 56.1 (q, min), 55.7 (q, maj), 29.9 (d, min), 29.0 (d, maj), 20.3 (t, maj), 18.7 (t, min).

of $(1S^*, 2R^*)$ -2-(phenylethynyl)cyclopropanecarboxylic acid Cycloisomerization 9: Formation of (1R*,6S*)-4-phenyl-3-oxabicyclo[4.1.0]hept-4-en-2-one (11)and (1S*,5R*,Z)-2-benzylidene-3-oxabicyclo[3.1.0]hexan-2-one (12). This cycloisomerization was carried out following the general procedure at 50 °C in 1,2-dichloroethane, by employing $(1S^*, 2R^*)$ -2-(phenylethynyl)cyclopropanecarboxylic acid 9 (0.5 mmol, 93 mg) as starting material. Once the reaction was concluded (checked by TLC), solvents were removed under vacuum give a crude reaction mixture formed by $(1R^{*}, 6S^{*})$ -4-phenyl-3to $(1S^*, 5R^*, Z)$ -2-benzylidene-3oxabicyclo[4.1.0]hept-4-en-2-one 11 and oxabicyclo[3.1.0]hexan-2-one 12, in 58/42 ratio. Separation of regiosiomers by column chromatography was not possible; a 54% yield of the mixture of 11 and 12 [containing some IPrAu(OH)] was recovered after column chromatography.



(1R*,6S*)-4-phenyl-3-oxabicyclo[4.1.0]hept-4-en-2-one (11). Not isolated; data retrieved form a mixture of 11/12 in 57/43 ratio, containing also some IPrAu(OH) (not quantified). R_f = 0.70 (HxH/AcOEt, 1/1); ¹H NMR (400 MHz, CDCl₃) δ = 7.58 – 7.56 (m, 2 H), 7.38 – 7.27 (m, 3 H), 6.03 (d, *J* = 5.1 Hz, 1 H), 2.21 – 2.17 (m, 1 H), 2.08 – 2.04 (m, 1 H), 1.71 – 1.55 (m, 1 H), 0.97 – 0.94 (m, 1 H). ¹³C NMR (100 MHz, CDCl₃) δ = 168.1 (s), 148.3 (s), 132.4 (s), 128.9 (d), 128.5 (d, 2 CH), 124.4 (d), 101.2 (d), 17.9 (d), 17.0 (d), 15.1 (t); LRMS: (EI) *m/z* (%) = 186 ([M]⁺, 9), 158 (100), 105 (66), 77 (61).



 $(1S^*, 5R^*, Z)$ -2-benzylidene-3-oxabicyclo[3.1.0]hexan-2-one (12). Not isolated; data retrieved form a mixture of 11/12 in 57/43 ratio, containing also some IPrAu(OH) (not quantified). R_f = 0.70 (HxH/AcOEt, 1/1); ¹H NMR (400 MHz, CDCl₃) δ = 7.54 – 7.51 (m, 2 H), 7.38 – 7.29 (m, 2 H), 7.22 – 7.17 (m, 1 H), 5.62 (s, 1H), 2.84 – 2.80 (m, 1 H), 2.44 – 2.40 (m, 1 H), 1.45 – 1.39 (m, 1 H), 1.20 – 1.17 (m, 1 H). ¹³C NMR (100 MHz, CDCl₃) δ = 172.6 (s), 147.7 (s), 133.7 (s), 128.5 (d, 4 CH), 126.8 (d), 104.1 (d), 22.4 (d), 18.3 (d), 15.0 (t); LRMS: (EI) *m/z* (%) = 186 ([M]⁺, 65), 158 (25), 129 (22), 118 (100), 90 (51).

Cycloisomerization of $(1R^*, 2S^*)$ -[2-methoxy-2-(phenylethynyl)cyclopropyl]methanol 10: Formation of $(1R^*, 6S^*)$ -6-methoxy-4-phenyl-3-oxabicyclo[4.1.0]hept-4-ene (13) and $(1S^*, 5R^*, Z)$ -2-benzylidene-1-methoxy-3-oxabicyclo[3.1.0]hexane (14). This cycloisomerization was carried out following the general procedure at room temperature, by employing alkynylcyclopropane alcohol **10** (0.5 mmol, 101 mg) as starting material. Once the reaction was concluded (as checked by TLC), solvents were removed under vacuum to give a crude reaction mixture formed by oxabicyclo[4.1.0]hept-4-ene **13** and oxabicyclo[3.1.0]hexane **14**, in 95% combined yield and 80/20 ratio.



(1*R**,6*S**)-6-methoxy-4-phenyl-3-oxabicyclo[4.1.0]hept-4-ene (13). Chromatographic purification using hexane/AcOEt (5:1) as eluent allowed the partial isolation of oxabicyclo[4.1.0]hept-4-ene 13 as colourless liquid. $R_f = 0.56$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.57$ (d, J = 7.2 Hz, 2 H), 7.47 – 7.21 (m, 3 H), 6.09 (s, 1 H), 4.28 (dd, J = 10.3, 0.9 Hz, 1 H), 3.99 (dd, J = 10.3, 1.8 Hz, 1 H), 3.41 (s, 3 H), 1.82 – 1.75 (m, 1 H), 1.33 (dd, J = 9.9, 5.1 Hz, 1 H), 1.10 (at, J = 5.7 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 149.4$ (s), 135.0 (s), 128.2 (d, 2 CH), 128.1 (d), 124.3 (d, 2 CH), 103.0 (d), 63.9 (t), 57.6 (s), 55.5 (q), 24.8 (t), 19.7 (d); HRMS: (ESI) Calculated for $C_{13}H_{15}O_2$ [M+1]⁺: 203.1067, found: 203.1070.



(1*S**,5*R**,*Z*)-2-benzylidene-1-methoxy-3-oxabicyclo[3.1.0]hexane (14). Colourless liquid; spectral data obtained from an enriched mixture containing 13 (ratio 14/13 = 3:2); R_f = 0.58 (Hexane/AcOEt, 5/1); ¹H NMR (401 MHz, CDCl₃) δ = 7.60 – 7.53 (m, 2 H), 7.35 – 7.26 (m, 2 H), 7.12 (d, *J* = 7.4 Hz, 1 H), 5.50 (s, 1 H), 4.41 (dd, *J* = 8.9, 4.3 Hz, 1 H), 4.07 (d, *J* = 8.9 Hz, 1 H), 3.46 (s, 3 H), 2.12 – 2.06 (m, 1 H), 1.60 (dd, *J* = 9.1, 5.3 Hz, 1 H), 0.96 (at, *J* = 5.3 Hz, 1 H); ¹³C NMR (101 MHz, CDCl₃) δ = 153.4 (s), 136.2 (s), 128.2 (d, 2 CH), 127.4 (d, 2 CH), 125.1 (d), 97.3 (d), 72.6 (t), 71.0 (s), 56.8 (q), 21.4 (d), 21.9 (t).



General procedure for the synthesis of alkynylcyclopropanecarboxamides 15:

A solution of alkynylcyclopropane acid 1 (1 mmol) in THF (15 mL) under inert atmosphere was cooled to -15 °C with stirring. Et₃N (1 equiv, 1 mmol, 0.14 mL) and ethyl chloroformate (1 equiv, 1 mmol, 0.96 mL) were sequentially added; a white precipitate was formed. Stirring was maintained for 1h at that temperature and, then, the corresponding amine (or ammonium hydroxide) was added. Stirring was continued for 15 min at -15 °C, before removing the cooling bath and allowing the mixture to reach room temperature. Solvent was evaporated under vacuum; ethyl acetate (25 mL) and 5% sodium bicarbonate solution (15 mL) were added. Following shaking and separation, the ethyl acetate layer was sequentially washed with water (15 mL), 1*N* hydrochloric acid (15 mL), and finally, water (15 mL). After drying over anhydrous sodium sulphate, evaporation of the ethyl acetate solution led to amides 15, which were used without further purification. No epimerization was observed under the reaction conditions.



(1*S**, 2*S**)-2-methoxy-2-(phenylethynyl)cyclopropanecarboxamide (15a). Colourless liquid; yield = 98 % (of a 5:1 mixture of diasteroisomers); $R_f = 0.05$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.43 - 7.39$ (m, 4 H, min + maj), 7.31 – 7.24 (m, 6 H, min + maj), 6.29 (bs, 4

H, min + maj), 3.48 (s, 3 H, min), 3.45 (s, 3 H, maj), 2.13 - 2.07 (m, 2 H, min + maj), 1.73 - 1.64 (m, 2 H, min + maj), 1.53 - 1.46 (m, 2 H, min + maj); ¹³C NMR (100 MHz, CDCl₃) $\delta = 171.1$ (s, min), 170.7 (s, maj), 131.9 (d, 2 CH, maj), 131.8 (d, 2 CH, min), 128.7 (d, min), 128.5 (d, maj), 128.4 (d, 2 CH, min), 128.2 (d, 2 CH, maj), 122.3 (s, maj), 122.0 (s, min), 86.7 (s, maj), 86.4 (s, min), 84.2 (s, min), 83.8 (s, maj), 58.6 (s, maj), 57.7 (s, min), 56.3 (q, min), 55.8 (q, maj), 31.1 (t, min), 31.0 (t, maj), 21.6 (d, min), 21.0 (d, maj).



 $(1S^{*},$ 2S*)-2-methoxy-N-methyl-2-(o-tolylethynyl)cyclopropanecarboxamide (15b). Colourless liquid; yield = 97% (of a 6.7:1 mixture of diasteroisomers); $R_f = 0.21$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.40 - 7.36$ (m, 2 H, min + maj), 7.23 - 7.07 (m, 6 H, min + maj), 6.25 (bs, 1 H, min), 6.11 (bs, 1 H, maj), 3.51 (s, 3 H, min), 3.48 (s, 3 H, maj), 2.82 (d, J = 4.8 Hz, 3 H, maj + 3 H min), 2.41 (s, 6 H, min + maj), 2.12 (dd, J = 9.8, 7.6 Hz, 1 H,min), 2.08 (dd, J = 9.7, 7.4 Hz, 1 H, maj), 1.72 (dd, J = 7.4, 5.7 Hz, 1 H, maj + 1 H min), 1.49 (dd, J = 9.7, 5.7 Hz, 1 H, maj + 1 H min); ¹³C NMR (100 MHz, CDCl₃) $\delta = 168.9$ (s, min), 168.4 (s, maj), 140.3 (s, maj), 140.2 (s, min), 132.1 (s, maj), 131.9 (d, min), 129.5 (d, min), 129.4 (d, maj), 128.6 (d, min), 128.4 (d, maj), 125.6 (d, min), 125.4 (d, maj), 122.3 (s, maj), 121.9 (s, min), 90.6 (s, min), 87.8 (s, maj), 85.5 (s, maj), 82.9 (s, min), 58.4 (s, maj), 57.4 (s, min) 56.3 (q, min), 55.7 (q, maj), 31.7 (t, min), 31.5 (t, maj), 26.6 (q, min), 26.5 (q, maj), 21.3 (d, min), 20.73 (d, maj), 20.66 (q, min), 20.60 (q, maj); LRMS: (EI) m/z (%) = 243 ([M]⁺, 19), 228 (33), 185 (99), 115 (100), 112 (57); HRMS: (EI) calculated for C₁₅H₁₇NO₂ [M]⁺: 243.1259, found: 243.1257.


(1S*,2S*)-2-methoxy-N-(4-methoxyphenyl)-2-(p-tolylethynyl)cyclopropanecarboxamide (15c). Colourless liquid; yield = 90% (of a 5:1 mixture of diasteroisomers); $R_f = 0.18$ (HxH/AcOEt, 5/1); ¹H NMR (401 MHz, CDCl₃) $\delta = 7.85$ (bs, 1 H, min), 7.57 (bs, 1 H, maj), 7.42 (d, J = 8.9 Hz, 2 H, min), 7.40 (d, J = 8.9 Hz, 2 H, maj), 7.34 (d, J = 8.0 Hz, 2 H, min), 7.30 (d, J = 7.9 Hz, 2 H, maj), 7.14 (d, J = 8.0 Hz, 2 H, min), 7.08 (d, J = 7.9, 2 H, maj), 6.85 $(d, J = 8.9 \text{ Hz}, 2 \text{ H}, \text{min}), 6.82 (d, J = 8.9 \text{ Hz}, 2 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{min}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{min}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{min}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{min}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{maj}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{maj}), 3.77 (s, 3 \text{ H}, \text{maj}), 3.79 (s, 3 \text{ H}, \text{maj}), 3.77 (s, 3 \text{ H}, \text$ 3.56 (s, 3 H, min), 3.51 (s, 3 H, maj), 2.36 (s, 3 H, min), 2.33 (s, 3 H, maj), 2.20 (dd, J = 9.2, 7.6 Hz, 1 H, maj + 1 H, min), 1.81 - 1.76 (m, 2 H, min + maj), 1.60 (dd, J = 9.4, 5.8 Hz, 1 H, maj + 1 H, min); ¹³C NMR (100 MHz, CDCl₃) δ = 166.3 (s, min), 166.2 (s, maj), 156.3 (s, min), 156.2 (s, maj), 138.9 (s, min), 138.6 (s, maj), 131.8 (d, 2 CH, maj), 131.7 (d, 2 CH, min), 131.3 (s, maj), 131.2 (s, min), 129.2 (d, 2 CH, min), 129.0 (d, 2 CH, maj), 121.9 (d, 2 CH, maj), 121.8 (d, 2 CH, min), 119.2 (s, maj), 119.0 (s, min), 114.2 (d, 2 CH, min), 114.0 (d, 2 CH, maj), 87.2 (s, maj), 85.8 (s, min), 84.4 (s, min), 83.1 (s, maj), 58.8 (s, maj), 57.8 (s, min), 56.4 (q, min), 55.8 (q, maj), 55.5 (q, min), 55.4 (q, maj), 32.2 (t, 2 CH₂, min + maj), 21.49 (d, min), 21.45 (d, maj), 21.3 (q, min), 20.9 (q, maj); LRMS: (EI) m/z (%) = 335 ([M]⁺, 34), 302 (51), 185 (100), 143 (52), 115 (39); HRMS: (EI) calculated for $C_{21}H_{21}NO_3$ [M]⁺: 335.1521, found: 335.1527.

General procedure for the synthesis of azepinones 16:

A solution of freshly lyophilized IPrAuCl (3 mol%, 0.015 mmol, 9.3 mg) and AgOTs (3 mol%, 0.015 mmol, 4.2 mg) in CH_2Cl_2 (5 mL) was placed in a sealed tube under Ar atmosphere and protected from light. The mixture was stirred during 30 min, and a solution of the

corresponding alkynylcyclopropanecarbamide **15** (0.5 mmol) in CH_2Cl_2 (5 mL) was then added; stirring was maintained for 12 h at 110 °C. End of reaction was monitored by TLC; solvents were removed under vacuum and the crude reaction mixture was purified by flash chromatography using hexane/AcOEt (5:1) as eluent to give the corresponding azepinone **16** in the yield indicated in Scheme 4.



5-methoxy-7-phenyl-1*H***-azepin-2(3***H***)-one (16a).** Colourless liquid; yield = 85 %; $R_f = 0.25$ (HxH/AcOEt, 5/1); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.56 - 7.53$ (m, 2 H), 7.44 - 7.38 (m, 3 H), 7.26 (bs, 1 H), 6.12 (s, 1 H), 4.76 (t, *J* = 7.1 Hz, 1 H), 3.63 (s, 3 H), 2.91 (d, *J* = 7.1 Hz, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 170.3$ (s), 156.1 (s), 138.9 (s), 136.9 (s), 129.6 (d), 129.0 (d, 2 CH), 126.6 (d, 2 CH), 109.8 (d), 90.8 (d), 55.6 (q), 32.9 (t). LRMS: (EI) *m/z* (%) = 215 ([M]⁺, 100), 201 (82), 186 (82), 146 (94), 104 (90); HRMS: (EI) calculated for C₁₃H₁₃NO₂ [M]⁺: 215.0946, found: 215.0948.



5-Methoxy-1-methyl-7-(*o*-tolyl)-1*H*-azepin-2(3*H*)-one (16b). Colourless liquid; yield = 76 %; $R_f = 0.36 (HxH/AcOEt, 5/1)$; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.41 - 7.17 (m, 4 H)$, 5.90 (s, 1 H), 4.83 (t, *J* = 7.0 Hz, 1 H), 3.62 (s, 3 H), 3.01 (bs, 2 H), 2.77 (s, 3 H), 2.23 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 169.6$ (s), 161.8 (s), 155.3 (s), 137.4 (s), 135.5 (s), 130.6 (d), 129.7 (d), 128.9 (d), 126.5 (d), 115.4 (d), 92.5 (d), 55.5 (q), 34.0 (q), 33.3 (d), 19.5 (q).



5-methoxy-1-(4-methoxyphenyl)-7-(*p***-tolyl)-1***H***-azepin-2(***3H***)-one (16c).** Colourless liquid; yield = 62 %; $R_f = 0.38$ (HxH/AcOEt, 5/1); ¹H NMR (401 MHz, CDCl₃) $\delta = 7.22$ (d, *J* = 7.9 Hz, 2 H), 7.06 (d, *J* = 8.9 Hz, 2 H), 6.99 (d, *J* = 7.9 Hz, 2 H), 6.70 (d, *J* = 8.9 Hz, 2 H), 6.27 (s, 1 H), 4.93 (t, *J* = 6.8 Hz, 1 H), 3.69 (s, 3 H), 3.67 (s, 3 H), 3.12 (bs, 2 H), 2.24 (s, 3 H); ¹³C NMR (1010 MHz, CDCl₃) $\delta = 169.1$ (s), 157.7 (s), 155.6 (s), 144.3 (s), 138.2 (s), 134.9 (s), 132.9 (s), 129.0 (d, 2 CH), 128.8 (d, 2 CH), 127.7 (d, 2 CH), 116.1 (d), 113.6 (d, 2 CH), 93.7 (d), 55.6 (q), 55.2 (q), 34.3 (t), 21.1 (q). LRMS: (EI) *m/z* (%) = 335 ([M]⁺, 63), 320 (15), 306 (100), 292 (14), 224 (32); HRMS: (EI) calculated for C₂₁H₂₁NO₃ [M]⁺: 335.1521, found: 335.1515.

4. Tables with the 2D-NMR (HSQC, COSY, HMBC and NOESY) Experiments for Selected Compounds [400 MHz (CDCl₃)]



Site	¹³ C RMN	DEPT	¹ H RMN	COSY	HMBC	NOESY
1	166.9	С				
2	32.0	CH_2	3.00 (d, J = 6.8 Hz, 2 H)	3	1, 3, 4, 6	3
3	89.7	СН	4.85 (t, $J = 6.8$ Hz, 1 H)	2	2, 5, 6, 1	2, 11
4	155.8	С				
5	106.2	СН	6.42 (s, 1 H)		3,4, 6, 7,	8,11
6	150.3	С				
7	133.5	С				
8	125.4	СН	7.70-7.65 (m, 2 H)	9, 10	6, 8, 10	5
9	128.5	СН	7.41-7.37 (m, 2 H)	8, 10	7, 9, 10	
10	129.5	СН	7.41-7.37 (m, 1 H)	8,9	8	
11	55.5	CH_3	3.65 (s, 3 H)		4	3, 5



Site	¹³ C RMN	DEPT	¹ H RMN	COSY	HMBC	NOESY
1	166.9	С				
2	24.9	СН	2.36 (ddd, <i>J</i> = 11.2, 5.4, 1.4 Hz, 1 H)	3, 5	3,4	
3	23.2	CH_2	1.17 (t, $J = 5.4$ Hz, 1 H),	2, 3	1, 3, 4, 5	
			2.01 (dd, <i>J</i> = 11.2, 5.4 Hz, 1 H)			
4	64.6	С				
5	106.1	СН	5.77 (s, 1 H)		3, 6, 7	7, 13
6	147.1	С				
7	119.4	СН	6.42 (d, <i>J</i> = 15.9 Hz, 1 H)	7	1, 5, 6, 8, 9,	5, 10
8	131.0	СН	7.08 (d, $J = 15.9$ Hz, 1 H)	8	5, 6, 7,9, 10	10
9	135.9	С				
10	126.9	СН	7.44-7.27 (m, 2 H)	11, 12	8, 10, 12	7, 8
11	128.8	СН	7.44-7.27 (m, 2 H)	10, 12	9, 11	
12	128.5	СН	7.44-7.27 (m, 1 H)	10, 11	10	
13	56.7	CH_3	3.39 (s, 3 H)		4	5

5. References and notes.

¹ N. Mézailles, L. Ricard, F. Gagosz, Org. Lett. 2005, 7, 4133-4136.

³ D. Wang, R. Cai, S. Sharma, J. Jirak, S. K. Thummanapelli, N. G. Akhmedov, H. Zhang, X. Liu, J. L. Petersen, X. Shi, *J. Am. Chem. Soc.* 2012, **134**, 9012–9019.

⁴ E. Álvarez, D. Miguel, P. García-García, M. A. Fernández-Rodríguez, F. Rodríguez, R. Sanz, *Beilstein J. Org. Chem.* 2011, **7**, 786–793.

⁵ E. Álvarez, P. García-García, M. A. Fernández-Rodríguez, R. Sanz, J. Org. Chem. 2013, dx.doi.org/10.1021/j0401388b.

⁶ P. García-García, M. A. Rashid, A. M. Sanjuán, M. A. Fernández-Rodríguez, R. Sanz, *Org. Lett.* 2012, **14**, 4778–4781.

⁷ P. Pale, J. Chuche, *Eur. J. Org. Chem.* 2010, 1019–1025.

⁸ In this notation, *cis* and *trans* prefixes denote the relative position of the reactive functional groups: the carboxylic group and the triple bond. For the synthesis of alkynylcyclopropanes see: J. Barluenga, M. A. Fernández-Rodríguez, P. García-García, E. Aguilar, I. Merino, *Chem. Eur. J.*, 2006, **12**, 303–313.

⁹ a) C. Nieto-Oberhuber, S. López, A. M. Echavarren, J. Am. Chem. Soc. 2005, **127**, 6178–6179. b) A. Buzas, F. Gagosz, J. Am. Chem. Soc. 2006, **128**, 12614–12615.

¹⁰ a) L. Ricard, F. Gagosz, *Organometallics* 2007, **26**, 4704–4707. (b) G. Li, L. Zhang, *Angew. Chem. Int. Ed.* 2007, **46**, 5156–5159.

¹¹ P. de Frémont, N. M. Scott, E. D. Stevens, S. P. Nolan, *Organometallics* 2005, **24**, 2411–2418.

¹² D. D. Perrin, W. L. F. Armarego, D. R. Perrin, *Purification of Laboratory Chemicals*, Pergamon Press., 3h Ed., 1998.

¹³ For the preparation of Fischer alkoxy alkynylcarbene complexes see ref. 8 and references cited therein.

¹⁴ B. De Carnné-Carnavalet, A. Archambeau, C. Meyer, J. Cossy, B. Folléas, J.-L. Brayer, J.-P. Demoute, *Org. Lett.* 2011, **13**, 956–959.

² See, for a recent example: S. Gupta, D. Koley, K. Ravikumar, B. Kundu, *J. Org. Chem.* 2013, **78**, 8624–8633.

6. Copies of ¹H- and ¹³C-NMR Spectra









Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013







Electronic Supplementary Material (ESI) for Chemical Communications This journal is The Royal Society of Chemistry 2013



Electronic Supplementary Material (ESI) for Chemical Communications This journal is © The Royal Society of Chemistry 2013





Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013





Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013





















Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013









Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013



```
Electronic Supplementary Material (ESI) for Chemical Communications This journal is \textcircled{C} The Royal Society of Chemistry 2013
```


Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013





Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013













Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013







Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013



Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013





Electronic Supplementary Material (ESI) for Chemical Communications This journal is O The Royal Society of Chemistry 2013



Electronic Supplementary Material (ESI) for Chemical Communications This journal is C The Royal Society of Chemistry 2013



