

Palladium-catalyzed asymmetric 1,6-addition of diarylphosphines to $\alpha,\beta,\gamma,\delta$ -unsaturated sulfonic esters: controlling regioselectivity by rational selection of electron-withdrawing groups

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General Methods

All air- and moisture-sensitive manipulations were carried out with standard Schlenk techniques under nitrogen or in a glove box under nitrogen. ^1H , ^{13}C and ^{31}P NMR spectra were recorded on a Varian instrument (300 MHz, 75 MHz and 121Hz; 400 MHz, 100 MHz and 162 MHz, respectively). ^1H , ^{13}C NMR chemical shifts are reported vs tetramethylsilane signal or residual protio solvent signals.

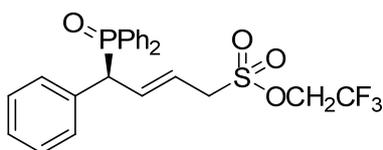
Toluene and THF were distilled over sodium benzophenone ketyl under nitrogen. Dichloromethane and tert-amyl alcohol was distilled over CaH_2 under nitrogen.

The pincer-PdOAc catalysts¹, diarylphosphine² and α,β -unsaturated sulfonic esters³ were synthesized following the literature procedures. All other chemicals and solvents were purchased from commercial company and used as received.

Experimental Details and Characterization Data

General Experimental Procedures for Table 2.

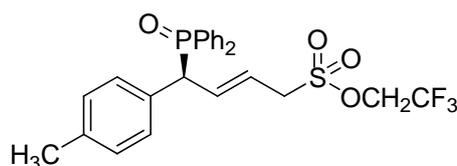
Diarylphosphine (0.20 mmol) was added to a solution of (*S,S*)-Pd catalyst (6.7 mg, 10 μmol Pd) and $\alpha,\beta,\gamma,\delta$ -unsaturated sulfonic ester **1** (0.21 mmol) in toluene (4.0 mL) at $-60\text{ }^\circ\text{C}$ (realized with a refrigerated bath circulator), then the resulting solution was stirred for 24 h at the same temperature. The reaction was quenched with 30% H_2O_2 aqueous solution (0.1 mL) and concentrated under vacuum. The residue was purified by silica gel chromatography with $\text{CH}_2\text{Cl}_2/\text{MeOH} = 100/1$ to afford the 1,6-adduct.



Entry 1. White solid. (80.2 mg; 81% yield). The ee was determined on a Daicel Chiralcel OD column with hexane/2-propanol = 70/30, flow = 0.7 mL/min. Retention times: 12.2 min [(*R*)-enantiomer], 14.1 min [(*S*)-enantiomer]. 94% ee. $[\alpha]_D^{20} = -12.1$

(c 0.500, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

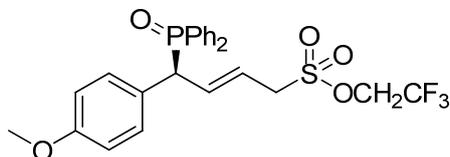
¹H NMR (400 MHz, CDCl₃): δ 7.89-7.83 (m, 2H, Ar), 7.61-7.45 (m, 5H, Ar), 7.40-7.35 (m, 1H, Ar), 7.29-7.18 (m, 7H, Ar), 6.36-6.16 (m, 1H, olefin), 5.58-5.48 (m, 1H, olefin), 4.33-4.21 (m, 3H), 3.89-3.76 (m, 2H, CH₂S). ¹³C NMR (100 MHz, CDCl₃): δ 136.7 (d, *J*_{CP} = 6.4 Hz), 134.5 (d, *J*_{CP} = 5.7 Hz), 132.3 (d, *J*_{CP} = 2.7 Hz), 131.8 (d, *J*_{CP} = 3.0 Hz), 131.6 (d, *J*_{CP} = 8.7 Hz), 131.19 (d, *J*_{CP} = 8.8 Hz), 131.16 (d, *J*_{CP} = 96.1 Hz), 131.0 (d, *J*_{CP} = 97.2 Hz), 129.4 (d, *J*_{CP} = 5.7 Hz), 128.9 (d, *J*_{CP} = 10.3 Hz), 128.85, 128.4 (d, *J*_{CP} = 11.8 Hz), 127.7 (d, *J*_{CP} = 2.3 Hz), 121.9 (q, *J*_{CF} = 277.8 Hz), 119.8 (d, *J*_{CP} = 11.0 Hz), 65.1 (q, *J*_{CF} = 37.8 Hz), 55.0 (d, *J*_{CP} = 1.9 Hz), 51.6 (d, *J*_{CP} = 64.5 Hz). ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 28.2 (s). HRMS (ESI) calcd for C₂₄H₂₃F₃O₄PS (M+H)⁺: 495.1001, found: 495.1002.



Entry 2. White solid. (79.1 mg; 78% yield). The ee was determined on a Daicel Chiralpak AD column with hexane/2-propanol = 90/10, flow = 1.0 mL/min. Retention times: 86.7 min [(*R*)-enantiomer], 98.5 min [(*S*)-enantiomer]. 94% ee. [α]_D²⁰ = -5.2 (c 0.50, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

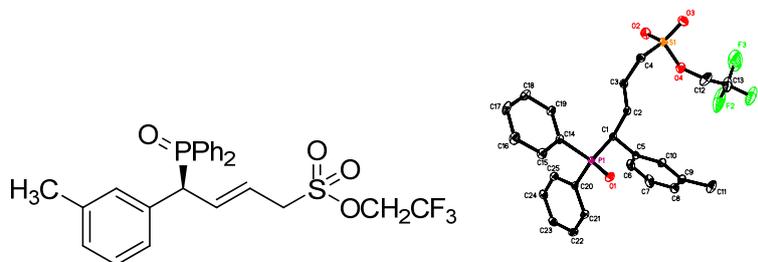
¹H NMR (400 MHz, CDCl₃): δ 7.86-7.82 (m, 2H, Ar), 7.56-7.48 (m, 5H, Ar), 7.40-7.35 (m, 1H, Ar), 7.31-7.25 (m, 2H, Ar), 7.15 (d, *J* = 7.6 Hz, 2H, Ar), 7.02 (d, *J* = 7.6 Hz, 2H, Ar), 6.36-6.16 (m, 1H, olefin), 5.55-5.46 (m, 1H, olefin), 4.33-4.15 (m, 3H), 3.89-3.75 (m, 2H, CH₂S), 2.26 (s, 3H, Me). ¹³C NMR (100 MHz, CDCl₃): δ 137.5 (d, *J*_{CP} = 2.3 Hz), 137.0 (d, *J*_{CP} = 6.0 Hz), 132.2 (d, *J*_{CP} = 3.0 Hz), 131.8 (d, *J*_{CP} = 2.7 Hz), 131.6 (d, *J*_{CP} = 8.8 Hz), 131.4 (d, *J*_{CP} = 6.1 Hz), 131.31 (d, *J*_{CP} = 89.0 Hz), 131.29 (d, *J*_{CP} = 9.1 Hz), 130.21 (d, *J*_{CP} = 96.9 Hz), 129.6 (d, *J*_{CP} = 1.5 Hz), 129.3 (d, *J*_{CP} = 5.7 Hz), 128.9 (d, *J*_{CP} = 11.4 Hz), 128.4 (d, *J*_{CP} = 11.8 Hz), 121.9 (q, *J*_{CF} = 277.7 Hz), 119.6 (d, *J*_{CP} = 11.0 Hz), 65.1 (q, *J*_{CF} = 38.2 Hz), 55.0 (d, *J*_{CP} = 1.9 Hz), 51.2 (d,

$J_{CP} = 64.8$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 28.2 (s). HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{25}\text{F}_3\text{O}_4\text{PS}$ ($\text{M}+\text{H}$) $^+$: 509.1158, found: 509.1156.



Entry 3. White solid. (66.3 mg; 63% yield). The ee was determined on a Daicel Chiralcel OD column with hexane/2-propanol = 85/15, flow = 1.0 mL/min. Retention times: 15.4 min [(*R*)-enantiomer], 26.6 min [(*S*)-enantiomer]. 96% ee. $[\alpha]_D^{20} = -18.4$ (c 0.50, CH_2Cl_2). The absolute configuration was assigned by analogy with Table 2, entry 4.

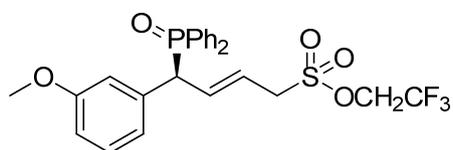
^1H NMR (400 MHz, CDCl_3): δ 7.86-7.82 (m, 2H, Ar), 7.56-7.46 (m, 5H, Ar), 7.40-7.35 (m, 1H, Ar), 7.31-7.25 (m, 2H, Ar), 7.15 (dd, $J = 6.8$ and 1.6 Hz, 2H, Ar), 6.75 (d, $J = 8.8$ Hz, 2H, Ar), 6.32-6.22 (m, 1H, olefin), 5.55-5.46 (m, 1H, olefin), 4.31-4.15 (m, 3H), 3.87-3.75 (m, 2H, CH_2S), 3.75 (s, 3H, OMe). ^{13}C NMR (100 MHz, CDCl_3): δ 159.0 (d, $J_{CP} = 2.3$ Hz), 136.8 (d, $J_{CP} = 6.1$ Hz), 132.2 (d, $J_{CP} = 2.7$ Hz), 131.8 (d, $J_{CP} = 3.0$ Hz), 131.6 (d, $J_{CP} = 8.3$ Hz), 131.21 (d, $J_{CP} = 9.1$ Hz), 131.18 (d, $J_{CP} = 99.1$ Hz), 131.0 (d, $J_{CP} = 96.5$ Hz), 130.5 (d, $J_{CP} = 5.7$ Hz), 128.9 (d, $J_{CP} = 11.3$ Hz), 128.4 (d, $J_{CP} = 11.7$ Hz), 126.5 (d, $J_{CP} = 6.1$ Hz), 121.9 (q, $J_{CF} = 278.1$ Hz), 119.6 (d, $J_{CP} = 11.0$ Hz), 114.3 (d, $J_{CP} = 1.9$ Hz), 65.1 (q, $J_{CF} = 37.8$ Hz), 55.2, 54.9 (d, $J_{CP} = 1.2$ Hz), 50.6 (d, $J_{CP} = 65.2$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 28.2 (s). HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{25}\text{F}_3\text{O}_5\text{PS}$ ($\text{M}+\text{H}$) $^+$: 525.1107, found: 525.1106.



Entry 4. White solid. (88.3 mg; 87% yield). The ee was determined on a Daicel Chiralpak AD column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention

times: 18.7 min [(*R*)-enantiomer], 20.8 min [(*S*)-enantiomer]. 93% ee. $[\alpha]_D^{20} = -11.9$ (c 0.500, CH₂Cl₂). The absolute configuration was determined as *S* according to the X-ray crystal diffraction analysis of the product.

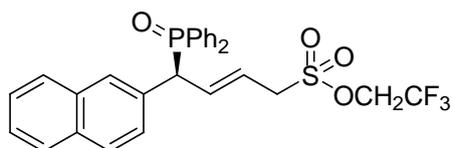
¹H NMR (400 MHz, CDCl₃): δ 7.87-7.82 (m, 2H, Ar), 7.55-7.46 (m, 5H, Ar), 7.40-7.35 (m, 1H, Ar), 7.31-7.25 (m, 2H, Ar), 7.10-7.00 (m, 3H, Ar), 7.00 (d, *J* = 7.2 Hz, 1H, Ar), 6.35-6.28 (m, 1H, olefin), 5.57-5.48 (m, 1H, olefin), 4.32-4.15 (m, 3H), 3.88-3.75 (m, 2H, CH₂S), 2.23 (s, 3H, Me). ¹³C NMR (100 MHz, CDCl₃): δ 138.5 (d, *J*_{CP} = 1.9 Hz), 136.7 (d, *J*_{CP} = 6.3 Hz), 134.3 (d, *J*_{CP} = 5.9 Hz), 132.2 (d, *J*_{CP} = 3.0 Hz), 131.8 (d, *J*_{CP} = 3.0 Hz), 131.6 (d, *J*_{CP} = 8.2 Hz), 131.3 (d, *J*_{CP} = 9.0 Hz), 131.2 (d, *J*_{CP} = 99.1 Hz), 131.0 (d, *J*_{CP} = 96.9 Hz), 130.0 (d, *J*_{CP} = 5.6 Hz), 128.9 (d, *J*_{CP} = 11.5 Hz), 128.7 (d, *J*_{CP} = 1.5 Hz), 128.4 (d, *J*_{CF} = 2.3 Hz), 128.3 (d, *J*_{CP} = 11.9 Hz), 126.4 (d, *J*_{CP} = 6.0 Hz), 121.9 (q, *J*_{CF} = 278.3 Hz), 119.7 (d, *J*_{CP} = 11.2 Hz), 65.1 (q, *J*_{CF} = 38.1 Hz), 55.0 (d, *J*_{CP} = 1.5 Hz), 51.6 (d, *J*_{CP} = 64.3 Hz), 21.4. ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 28.1 (s). HRMS (ESI) calcd for C₂₅H₂₅F₃O₄PS (M+H)⁺: 509.1158, found: 509.1162.



Entry 5. White solid. (82.1 mg; 78% yield). The ee was determined on a Daicel Chiralpak AD column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 24.8 min [(*R*)-enantiomer], 30.7 min [(*S*)-enantiomer]. 95% ee. The absolute configuration was assigned by analogy with Table 2, entry 4.

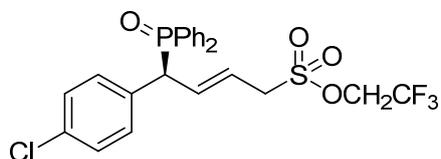
¹H NMR (400 MHz, CDCl₃): δ 7.88-7.82 (m, 2H, Ar), 7.55-7.46 (m, 5H, Ar), 7.40-7.35 (m, 1H, Ar), 7.31-7.25 (m, 2H, Ar), 7.13 (t, *J* = 8.0 Hz, 1H, Ar), 6.86-6.79 (m, 2H, Ar), 6.73 (d, *J* = 8.4 Hz, 1H, Ar), 6.35-6.28 (m, 1H, olefin), 5.55-5.48 (m, 1H, olefin), 4.32-4.15 (m, 3H), 3.88-3.75 (m, 2H, CH₂S), 3.68 (s, 3H, OMe). ¹³C NMR (100 MHz, CDCl₃): δ 159.8 (d, *J*_{CP} = 1.6 Hz), 136.6 (d, *J*_{CP} = 6.5 Hz), 136.0 (d, *J*_{CP} = 5.6 Hz), 132.3 (d, *J*_{CP} = 2.6 Hz), 131.9 (d, *J*_{CP} = 3.1 Hz), 131.6 (d, *J*_{CP} = 8.4 Hz), 131.3 (d, *J*_{CP} = 9.1 Hz), 131.2 (d, *J*_{CP} = 99.2 Hz), 131.0 (d, *J*_{CP} = 96.9 Hz), 129.8 (d,

$J_{CP} = 1.6$ Hz), 128.9 (d, $J_{CP} = 11.7$ Hz), 128.4 (d, $J_{CP} = 11.7$ Hz), 121.9 (q, $J_{CF} = 278.1$ Hz), 121.8 (d, $J_{CP} = 6.0$ Hz), 119.8 (d, $J_{CP} = 11.0$ Hz), 114.7 (d, $J_{CP} = 5.7$ Hz), 113.6 (d, $J_{CP} = 2.3$ Hz), 65.1 (q, $J_{CF} = 38.3$ Hz), 55.3, 55.0 (d, $J_{CP} = 1.9$ Hz), 51.7 (d, $J_{CP} = 64.5$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 28.2 (s). HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{25}\text{F}_3\text{O}_5\text{PS}$ ($\text{M}+\text{H}$) $^+$: 525.1107, found: 525.1107.



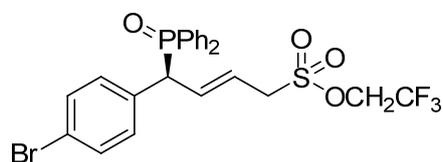
Entry 6. White solid. (95.0 mg; 87% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 19.1 min [(*R*)-enantiomer], 25.2 min [(*S*)-enantiomer]. 90% ee. $[\alpha]_{\text{D}}^{20} = -6.4$ (c 0.50, CH_2Cl_2). The absolute configuration was assigned by analogy with Table 2, entry 4.

^1H NMR (400 MHz, CDCl_3): δ 7.91-7.83 (m, 2H, Ar), 7.75-7.68 (m, 4H, Ar), 7.60-7.35 (m, 5H, Ar), 7.45-7.37 (m, 3H, Ar), 7.31 (t, $J = 7.6$ Hz, 1H, Ar), 7.22 (dd, $J = 7.6$ and 2.8 Hz, 2H, Ar), 6.45-6.37 (m, 1H, olefin), 5.60-5.52 (m, 1H, olefin), 4.50 (t, $J = 9.2$ Hz, 1H, PCH), 4.32-4.17 (m, 2H, OCH_2), 3.87-3.75 (m, 2H, CH_2S). ^{13}C NMR (100 MHz, CDCl_3): δ 136.6 (d, $J_{CP} = 6.3$ Hz), 133.4 (d, $J_{CP} = 1.9$ Hz), 132.6 (d, $J_{CP} = 1.5$ Hz), 132.3 (d, $J_{CP} = 2.6$ Hz), 132.1 (d, $J_{CP} = 6.0$ Hz), 131.9 (d, $J_{CP} = 2.6$ Hz), 131.6 (d, $J_{CP} = 8.5$ Hz), 131.2 (d, $J_{CP} = 8.9$ Hz), 131.1 (d, $J_{CP} = 98.4$ Hz), 131.05 (d, $J_{CP} = 97.3$ Hz), 128.9 (d, $J_{CP} = 11.6$ Hz), 128.62 (d, $J_{CP} = 8.2$ Hz), 128.60, 128.4 (d, $J_{CP} = 11.9$ Hz), 127.9, 127.7, 127.1 (d, $J_{CP} = 4.8$ Hz), 126.4, 126.2, 121.9 (q, $J_{CF} = 277.9$ Hz), 120.0 (d, $J_{CP} = 11.2$ Hz), 65.0 (q, $J_{CF} = 38.1$ Hz), 55.9 (d, $J_{CP} = 1.4$ Hz), 51.7 (d, $J_{CP} = 64.4$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 32.1 (s). HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{25}\text{F}_3\text{O}_4\text{PS}$ ($\text{M}+\text{H}$) $^+$: 545.1158, found: 545.1161.



Entry 7. White solid. (89.7 mg; 85% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 13.2 min [(*R*)-enantiomer], 17.0 min [(*S*)-enantiomer]. 88% ee. $[\alpha]_D^{20} = -20.7$ (c 0.500, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

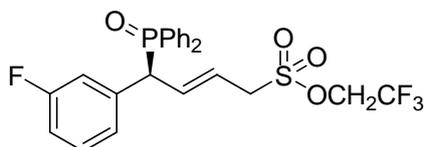
¹H NMR (400 MHz, CDCl₃): δ 7.87-7.81 (m, 2H, Ar), 7.58-7.45 (m, 5H, Ar), 7.41-7.35 (m, 1H, Ar), 7.35-7.28 (m, 2H, Ar), 7.24-7.17 (m, 4H, Ar), 6.32-6.21 (m, 1H, olefin), 5.56-5.46 (m, 1H, olefin), 4.30-4.15 (m, 3H), 3.87-3.75 (m, 2H, CH₂S). ¹³C NMR (100 MHz, CDCl₃): δ 136.2 (d, *J*_{CP} = 6.4 Hz), 133.7 (d, *J*_{CP} = 2.6 Hz), 133.1 (d, *J*_{CP} = 6.1 Hz), 132.4 (d, *J*_{CP} = 2.6 Hz), 132.1 (d, *J*_{CP} = 2.7 Hz), 131.6 (d, *J*_{CP} = 8.8 Hz), 131.1 (d, *J*_{CP} = 9.1 Hz), 130.9 (d, *J*_{CP} = 99.2 Hz), 130.7 (d, *J*_{CP} = 5.3 Hz), 130.67 (d, *J*_{CP} = 97.6 Hz), 129.04 (d, *J*_{CP} = 3.7 Hz), 128.99 (d, *J*_{CP} = 9.4 Hz), 121.9 (q, *J*_{CF} = 278.1 Hz), 120.2 (d, *J*_{CP} = 11.0 Hz), 64.9 (q, *J*_{CF} = 37.7 Hz), 54.9 (d, *J*_{CP} = 1.9 Hz), 51.9 (d, *J*_{CP} = 64.1 Hz). ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 27.9 (s). HRMS (ESI) calcd for C₂₄H₂₂ClF₃O₄PS (M+H)⁺: 529.0612, found: 529.0616.



Entry 8. White solid. (99.5 mg; 87% yield). The ee was determined on a Daicel Chiralpak AD column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 40.3 min [(*S*)-enantiomer], 53.1 min [(*R*)-enantiomer]. 93% ee. $[\alpha]_D^{20} = -25.9$ (c 0.500, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

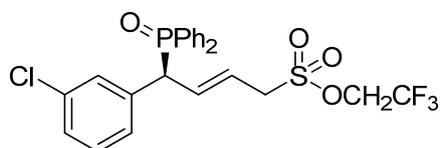
¹H NMR (400 MHz, CDCl₃): δ 7.89-7.83 (m, 2H, Ar), 7.61-7.48 (m, 5H, Ar), 7.40-7.38 (m, 1H, Ar), 7.35-7.30 (m, 4H, Ar), 7.15 (d, *J* = 6.8 Hz, 2H, Ar), 6.30-6.22 (m, 1H, olefin), 5.58-5.46 (m, 1H, olefin), 4.33-4.24 (m, 3H), 3.89-3.76 (m, 2H, CH₂S). ¹³C NMR (100 MHz, CDCl₃): δ 136.5 (d, *J*_{CP} = 6.8 Hz), 132.0 (d, *J*_{CP} = 2.7 Hz), 131.9 (d, *J*_{CP} = 2.6 Hz), 131.4 (d, *J*_{CP} = 97.7 Hz), 131.1 (d, *J*_{CP} = 8.7 Hz), 131.07 (d, *J*_{CP} = 8.8 Hz), 128.8 (d, *J*_{CP} = 13.3 Hz), 128.7 (d, *J*_{CP} = 12.9 Hz), 121.9 (q, *J*_{CF} =

277.7 Hz), 120.3 (d, $J_{CP} = 2.1$ Hz), 64.6 (q, $J_{CF} = 37.8$ Hz), 54.7, 45.9 (d, $J_{CP} = 68.0$ Hz), 20.7 (d, $J_{CP} = 2.6$ Hz), 12.4 (d, $J_{CP} = 13.3$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 27.9 (s). HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{22}\text{BrF}_3\text{O}_4\text{PS}$ ($\text{M}+\text{H}$) $^+$: 573.0106, found: 573.0114.



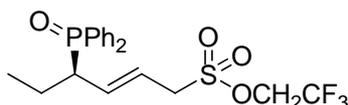
Entry 9. White solid. (86.9 mg; 86% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 12.1 min [(*R*)-enantiomer], 14.5 min [(*S*)-enantiomer]. 92% ee. $[\alpha]_D^{20} = -15.4$ (c 0.500, CH_2Cl_2). The absolute configuration was assigned by analogy with Table 2, entry 4.

^1H NMR (400 MHz, CDCl_3): δ 7.89-7.83 (m, 2H, Ar), 7.60-7.46 (m, 5H, Ar), 7.40-7.38 (m, 1H, Ar), 7.35-7.28 (m, 2H, Ar), 7.19-7.15 (m, 1H, Ar), 7.04 (d, $J = 8.0$ Hz, 1H, Ar), 7.15 (dd, $J = 9.2$ and 1.6 Hz, 1H, Ar), 6.89 (t, $J = 8.0$ Hz, 1H, Ar), 6.30-6.22 (m, 1H, olefin), 5.54-5.46 (m, 1H, olefin), 4.33-4.21 (m, 3H), 3.89-3.76 (m, 2H, CH_2S). ^{13}C NMR (100 MHz, CDCl_3): δ 162.7 (dd, $J_{CF} = 246.9$ Hz, $J_{CP} = 1.9$ Hz), 137.0 (dd, $J = 7.4$ and 5.9 Hz), 136.0 (d, $J_{CP} = 6.3$ Hz), 132.4 (d, $J_{CP} = 2.6$ Hz), 132.0 (d, $J_{CP} = 2.6$ Hz), 131.5 (d, $J_{CP} = 8.6$ Hz), 131.1 (d, $J_{CP} = 9.0$ Hz), 130.8 (d, $J_{CP} = 99.5$ Hz), 130.6 (d, $J_{CP} = 96.9$ Hz), 130.3 (dd, $J = 9.3$ and 2.6 Hz), 129.0 (d, $J_{CP} = 11.5$ Hz), 128.5 (d, $J_{CP} = 11.9$ Hz), 125.1 (dd, $J = 6.0$ and 3.0 Hz), 121.9 (q, $J_{CF} = 277.9$ Hz), 120.2 (d, $J = 10.7$ Hz), 116.4 (dd, $J = 12.3$ and 5.9 Hz), 114.6 (dd, $J = 21.2$ and 1.2 Hz), 64.9 (hept, $J_{CF} = 38.2$ Hz), 54.8 (d, $J_{CP} = 1.4$ Hz), 51.3 (dd, $J_{CP} = 63.9$ Hz, $J_{CF} = 1.8$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 27.9 (s). HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{22}\text{F}_4\text{O}_4\text{PS}$ ($\text{M}+\text{H}$) $^+$: 513.0907, found: 513.0911.



Entry 10. White solid. (86.5 mg; 82% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 12.0 min [(*R*)-enantiomer], 14.1 min [(*S*)-enantiomer]. 89% ee. $[\alpha]_D^{20} = -20.7$ (c 0.500, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

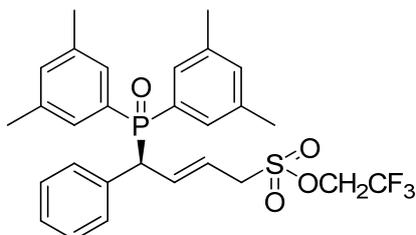
¹H NMR (400 MHz, CDCl₃): δ 7.88-7.83 (m, 2H, Ar), 7.61-7.46 (m, 5H, Ar), 7.43-7.38 (m, 1H, Ar), 7.34-7.28 (m, 2H, Ar), 7.23-7.12 (m, 4H, Ar), 6.32-6.22 (m, 1H, olefin), 5.58-5.48 (m, 1H, olefin), 4.35-4.24 (m, 3H), 3.89-3.77 (m, 2H, CH₂S).
¹³C NMR (100 MHz, CDCl₃): δ 136.6 (d, *J*_{CP} = 5.9 Hz), 135.9 (d, *J* = 6.3 Hz), 134.5 (d, *J*_{CP} = 1.9 Hz), 132.4 (d, *J*_{CP} = 2.9 Hz), 132.1 (d, *J*_{CP} = 3.0 Hz), 131.6 (d, *J*_{CP} = 8.6 Hz), 131.1 (d, *J*_{CP} = 8.9 Hz), 130.8 (d, *J*_{CP} = 99.5 Hz), 130.5 (d, *J*_{CP} = 98.4 Hz), 130.0 (d, *J* = 1.5 Hz), 129.5 (d, *J*_{CP} = 5.5 Hz), 129.0 (d, *J*_{CP} = 11.9 Hz), 128.5 (d, *J* = 11.9 Hz), 127.8 (d, *J* = 2.3 Hz), 127.5 (d, *J* = 5.6 Hz), 121.9 (q, *J*_{CF} = 277.9 Hz), 120.3 (d, *J* = 10.8 Hz), 64.9 (hept, *J*_{CF} = 37.8 Hz), 54.8 (d, *J*_{CP} = 1.1 Hz), 51.3 (d, *J*_{CP} = 63.6 Hz). ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 28.0 (s). HRMS (ESI) calcd for C₂₄H₂₂ClF₃O₄PS (M+H)⁺: 529.0612, found: 529.0615.



Entry 11. White solid. (74.8 mg; 84% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 18.2 min [(*R*)-enantiomer], 19.9 min [(*S*)-enantiomer]. 71% ee. $[\alpha]_D^{20} = -12.7$ (c 0.500, CH₂Cl₂), The absolute configuration was assigned by analogy with Table 2, entry 4.

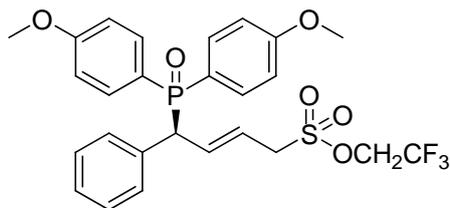
¹H NMR (400 MHz, CDCl₃): δ 7.82-7.50 (m, 4H, Ar), 7.52-7.45 (m, 6H, Ar), 5.89-5.82 (m, 1H, olefin), 5.56-5.48 (m, 1H, olefin), 4.40-4.33 (m, 2H, OCH₂), 3.91-3.77 (m, 2H, CH₂S), 3.08-2.95 (m, 1H, PCH), 1.79-1.65 (m, 2H), 0.92 (t, *J*_{CH} = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 136.6 (d, *J*_{CP} = 5.9 Hz), 135.9 (d, *J* = 6.3 Hz), 134.5 (d, *J*_{CP} = 1.9 Hz), 132.4 (d, *J*_{CP} = 2.9 Hz), 132.1 (d, *J*_{CP} = 3.0 Hz), 131.6 (d, *J*_{CP} = 8.6 Hz), 131.1 (d, *J*_{CP} = 8.9 Hz), 130.8 (d, *J*_{CP} = 99.5 Hz), 130.5 (d, *J*_{CP} = 98.4

Hz), 130.0 (d, $J = 1.5$ Hz), 129.5 (d, $J_{CP} = 5.5$ Hz), 129.0 (d, $J_{CP} = 11.9$ Hz), 128.5 (d, $J = 11.9$ Hz), 127.8 (d, $J = 2.3$ Hz), 127.5 (d, $J = 5.6$ Hz), 121.9 (q, $J_{CF} = 277.9$ Hz), 120.3 (d, $J = 10.8$ Hz), 64.9 (hept, $J_{CF} = 37.8$ Hz), 54.8 (d, $J_{CP} = 1.1$ Hz), 51.3 (d, $J_{CP} = 63.6$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 28.0 (s). HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{23}\text{F}_3\text{O}_4\text{PS}$ (M+H) $^+$: 447.1001, found: 447.0999.



Entry 12. White solid. (95.5 mg; 87% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 85/15, flow = 1.0 mL/min. Retention times: 12.7 min [(*R*)-enantiomer], 16.6 min [(*S*)-enantiomer]. 86% ee. $[\alpha]_{\text{D}}^{20} = -14.5$ (c 0.50, CH_2Cl_2). The absolute configuration was assigned by analogy with Table 2, entry 4.

^1H NMR (400 MHz, CDCl_3): δ 7.45 (d, $J = 11.2$ Hz, 2H, Ar), 7.28-7.18 (m, 6H, Ar), 7.04 (d, $J = 11.2$ Hz, 2H, Ar), 6.97 (s, 1H, Ar), 6.34-6.26 (m, 1H, olefin), 5.58-5.47 (m, 1H, olefin), 4.32-4.15 (m, 3H), 3.87-3.77 (m, 2H, CH_2S), 2.73 (s, 6H, Me), 2.17 (s, 6H, Me). ^{13}C NMR (100 MHz, CDCl_3): δ 138.6 (d, $J_{CP} = 11.9$ Hz), 137.9 (d, $J = 12.3$ Hz), 137.0 (d, $J_{CP} = 6.3$ Hz), 134.8 (d, $J_{CP} = 6.0$ Hz), 133.9 (d, $J_{CP} = 3.0$ Hz), 133.4 (d, $J_{CP} = 3.0$ Hz), 131.1 (d, $J_{CP} = 98.8$ Hz), 130.8 (d, $J_{CP} = 96.2$ Hz), 129.5 (d, $J_{CP} = 5.6$ Hz), 129.1 (d, $J = 8.2$ Hz), 128.8 (d, $J_{CP} = 8.9$ Hz), 128.7 (d, $J_{CP} = 2.8$ Hz), 127.5 (d, $J_{CP} = 2.2$ Hz), 121.9 (q, $J_{CF} = 277.9$ Hz), 119.6 (d, $J = 11.1$ Hz), 65.2 (q, $J_{CF} = 38.1$ Hz), 55.0 (d, $J_{CP} = 1.1$ Hz), 51.4 (d, $J_{CP} = 63.6$ Hz), 21.4 (d, $J_{CP} = 0.8$ Hz), 21.2 (d, $J_{CP} = 0.7$ Hz). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 28.8 (s). HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{31}\text{F}_3\text{O}_4\text{PS}$ (M+H) $^+$: 551.1627, found: 551.1626.



Entry 13. White solid. (95.0 mg; 86% yield). The ee was determined on a Daicel Chiralpak AS column with hexane/2-propanol = 85/15, flow = 1.0 mL/min. Retention times: 41.8 min [(*R*)-enantiomer], 54.8 min [(*S*)-enantiomer]. 76% ee. $[\alpha]_D^{20} = -8.6$ (c 0.50, CH₂Cl₂). The absolute configuration was assigned by analogy with Table 2, entry 4.

¹H NMR (400 MHz, CDCl₃): δ 7.73 (t, *J* = 8.8 Hz, 2H, Ar), 7.36 (dd, *J* = 10.4 and 8.8 Hz, 2H, Ar), 7.26-7.20 (m, 5H, Ar), 7.02 (d, *J* = 6.8 Hz, 2H, Ar), 6.78 (d, *J* = 6.8 Hz, 2H, Ar), 6.36-6.28 (m, 1H, olefin), 5.55-5.47 (m, 1H, olefin), 4.30-4.19 (m, 3H), 3.86 (s, 3H, OMe), 3.84-3.78 (m, 2H, CH₂S), 3.76 (s, 3H, OMe). ¹³C NMR (100 MHz, CDCl₃): δ 162.6 (d, *J*_{CP} = 2.6 Hz), 162.2 (d, *J* = 3.0 Hz), 137.2 (d, *J*_{CP} = 6.3 Hz), 135.0 (d, *J*_{CP} = 6.0 Hz), 133.4 (d, *J*_{CP} = 10.0 Hz), 133.1 (d, *J*_{CP} = 10.0 Hz), 129.5 (d, *J*_{CP} = 5.6 Hz), 128.8 (d, *J*_{CP} = 1.8 Hz), 127.5 (d, *J*_{CP} = 2.3 Hz), 122.7 (d, *J* = 105.4 Hz), 122.1 (d, *J*_{CP} = 104.3 Hz), 121.9 (q, *J*_{CF} = 277.9 Hz), 119.4 (d, *J* = 10.4 Hz), 114.4 (d, *J* = 12.2 Hz), 113.9 (d, *J* = 12.6 Hz), 65.1 (q, *J*_{CF} = 38.1 Hz), 55.4, 55.3, 55.0 (d, *J*_{CP} = 1.1 Hz), 52.3 (d, *J*_{CP} = 65.1 Hz). ³¹P {¹H} NMR (161 MHz, CDCl₃): δ 28.6 (s). HRMS (ESI) calcd for C₂₆H₂₇F₃O₆PS (M+H)⁺: 555.1213, found: 555.1212.

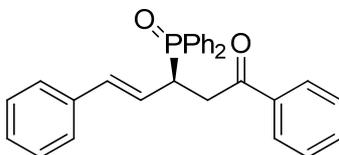


Table 1, Entry 1. White solid. (81.0 mg; 93% yield). The ee was determined on a Daicel Chiralpak IC column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 14.3 min [(*S*)-enantiomer], 17.2 min [(*R*)-enantiomer]. 93% ee. The absolute configuration was assigned by analogy with the literature example (Pincer Pd-catalyzed asymmetric 1,4-addition of diarylphosphines to α,β -unsaturated enones).¹

^1H NMR (400 MHz, CDCl_3): δ 7.93-7.88 (m, 4H, Ar), 7.83-7.76 (m, 2H, Ar), 7.56-7.38 (m, 9H, Ar), 7.23-7.12 (m, 5H, Ar), 6.43 (dd, $J = 16.0$ and 4.0 Hz, 1H, olefin), 6.15 (ddd, $J = 16.0$, 9.2 and 6.8 Hz, 1H, olefin), 4.22-4.10 (m, 1H), 3.69 (ddd, $J = 18.0$, 10.4 and 4.4 Hz, 1H), 3.30 (ddd, $J = 18.0$, 11.6 and 2.4 Hz, 1H). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 34.6 (s). HRMS (ESI) calcd for $\text{C}_{29}\text{H}_{26}\text{O}_2\text{P}$ ($\text{M}+\text{H}$) $^+$: 437.1665, found: 437.1666.

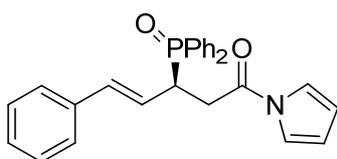


Table 1, Entry 2. (CAS: 1334387-70-0). White solid. (76.2 mg; 90% yield). The ee was determined on a Daicel Chiralpak AD-H column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 25.3 min [(*S*)-enantiomer], 33.1 min [(*R*)-enantiomer]. 96% ee. The absolute configuration was assigned by analogy with the literature example (Pincer Pd-catalyzed asymmetric 1,4-addition of diarylphosphines to α,β -unsaturated *N*-acylpyrroles).⁴

^1H NMR (400 MHz, CDCl_3): δ 7.91 (t, $J_{\text{HH}} = 8.8$ Hz, 2H, Ar), 7.79 (dd, $J = 10.0$ and 8.0 Hz, 2H, Ar), 7.57-7.16 (m, 13H), 6.45 (dd, $J = 15.6$ and 3.6 Hz, 1H, olefin), 6.25 (s, 2H), 6.13 (ddd, $J = 15.6$, 8.8 and 2.4 Hz, 1H, olefin), 4.06 (q, $J = 8.8$ Hz, 1H), 3.41 (ddd, $J_{\text{HH}} = 17.2$ and 10.0 Hz, $J_{\text{HP}} = 4.4$ Hz, 1H), 3.21 (ddd, $J_{\text{HH}} = 16.8$ and 2.0 Hz, $J_{\text{HP}} = 10.8$ Hz, 1H). $^{31}\text{P}\{^1\text{H}\}$ NMR (161 MHz, CDCl_3): δ 33.7 (s).

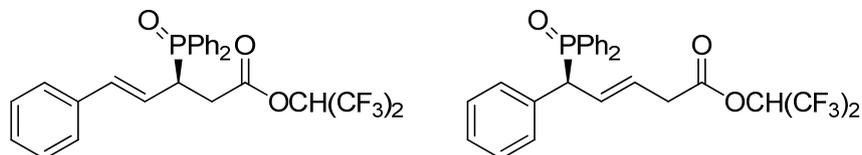


Table 1, Entry 3. White solid. (64.1 mg; 61% combined yield). 1,4-adduct : 1,6-adduct = 1 : 2.

The ee of 1,4-adduct was determined on a Daicel Chiralpak AD-H column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 6.6 min [(*S*)-enantiomer], 11.0 min [(*R*)-enantiomer]. 61% ee. The absolute configuration was

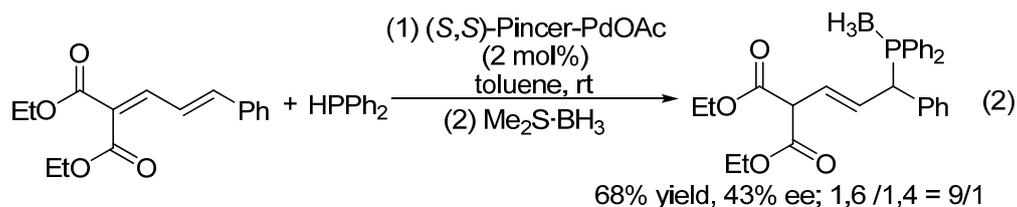
assigned by analogy with the literature examples (Pincer Pd-catalyzed asymmetric 1,4-addition of diarylphosphines to α,β -unsaturated carboxylic esters).⁵

¹H NMR (400 MHz, CDCl₃) for 1,4-adduct: δ 6.35 (dd, $J = 16.0$ and 4.4 Hz, 1H, olefin), 6.02 (ddd, $J = 16.0$, 8.8 and 6.4 Hz, 1H, olefin), 3.83-3.74 (m, 1H), 3.07 (ddd, $J = 16.4$ and 8.8 and 3.6 Hz, 1H), 2.94 (ddd, $J = 16.4$, 11.2 and 5.2 Hz, 1H). ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 33.5 (s). HRMS (ESI) calcd for C₂₆H₂₂F₆O₃P (M+H)⁺: 527.1205, found: 527.1201.

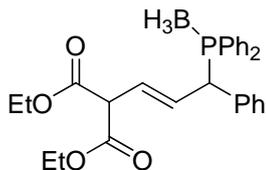
The ee of 1,6-adduct was determined on a Daicel Chiralpak AD-H column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 8.6 min [(*R*)-enantiomer], 9.6 min [(*S*)-enantiomer]. 71% ee. The absolute configuration was assigned by analogy with Table 2, entry 4.

¹H NMR (400 MHz, CDCl₃) for 1,6-adduct: δ 6.15-6.06 (m, 1H, olefin), 5.71-5.63 (m, 1H), 5.60-5.48 (m, 1H), 4.26 (t, $J = 9.6$ Hz, 1H), 3.24-3.10 (m, 2H). ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 33.0 (s). HRMS (ESI) calcd for C₂₆H₂₂F₆O₃P (M+H)⁺: 527.1205, found: 527.1201.

Experimental Procedures for the 1,6-Addition Reaction in Eq. 2



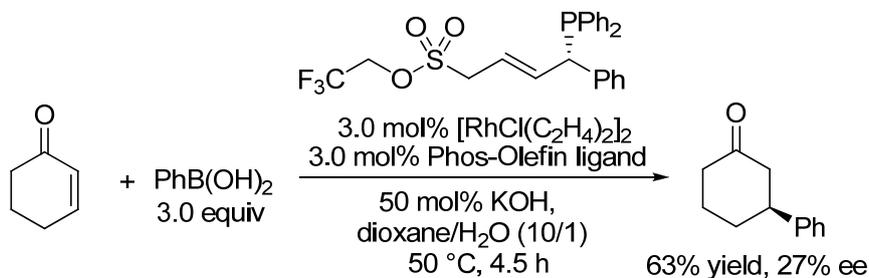
Diphenylphosphine (18.6 mg, 0.10 mmol) was added to a solution of (*S,S*)-Pd catalyst (1.4 mg, 2 μ mol Pd) and [(2*E*)-3-phenyl-2-propenylidene]-diethyl ester (32.9 mg, 0.12 mmol) in toluene (2.0 mL) at room temperature, then the resulting solution was stirred for 17 h. The reaction was quenched with Me₂S·BH₃ (0.15 mL; 2 M in THF) and concentrated under vacuum. The residue was purified by silica gel chromatography with hexane/EtOAc = 10/1 to afford the 1,6-adduct as a white solid.



White solid. (32.0 mg; 68% yield). The ee was determined on a Daicel Chiralpak AD column with hexane/2-propanol = 80/20, flow = 1.0 mL/min. Retention times: 7.8 min [major enantiomer], 8.7 min [minor enantiomer]. 43% ee. $[\alpha]_D^{20} = 7.3$ (c 1.00, CH₂Cl₂),

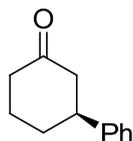
¹H NMR (400 MHz, CDCl₃): δ 7.83 (t, *J* = 8.8 Hz, 2H, Ar), 7.52-7.32 (m, 6H, Ar), 7.30-7.26 (m, 2H, Ar), 7.20-7.10 (m, 5H, Ar), 6.17 (ddd, *J* = 15.2, 9.6 and 6.4 Hz, 1H, olefin), 5.71 (ddd, *J* = 15.2, 9.6 and 3.2 Hz, 1H, olefin), 4.44 (dd, *J* = 15.2 and 9.6 Hz, 1H), 4.12 (q, *J* = 7.2 Hz, 2H), 4.10-4.00 (m, 2H), 3.96 (d, *J* = 9.2 Hz, 1H), 1.19 (t, *J* = 7.2 Hz, 3H), 1.18 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 167.5 (d, *J*_{CP} = 1.9 Hz), 167.3 (d, *J* = 2.6 Hz), 135.3 (d, *J*_{CP} = 1.5 Hz), 133.5 (d, *J*_{CP} = 8.4 Hz), 132.9 (d, *J*_{CP} = 8.4 Hz), 131.4 (d, *J*_{CP} = 2.2 Hz), 131.3, 131.0 (d, *J*_{CP} = 2.6 Hz), 129.4 (d, *J*_{CP} = 4.9 Hz), 128.6 (d, *J*_{CP} = 9.8 Hz), 128.3 (d, *J* = 9.8 Hz), 128.1 (d, *J*_{CP} = 2.3 Hz), 127.8 (d, *J*_{CP} = 52.6 Hz), 127.3 (d, *J*_{CP} = 2.7 Hz), 127.1 (q, *J*_{CF} = 52.7 Hz), 126.1 (d, *J* = 11.0 Hz), 61.7, 52.2 (q, *J*_{CF} = 1.5 Hz), 48.0 (d, *J*_{CP} = 29.9 Hz), 13.94, 13.93. ³¹P{¹H} NMR (161 MHz, CDCl₃): δ 24.7 (m). HRMS (ESI) calcd for C₂₈H₃₆BNO₄P (M+NH₄)⁺: 491.2506, found: 491.2511.

Experimental Procedure for Rhodium-Catalyzed 1,4-Addition of Phenylboronic Acid to Cyclohexenone



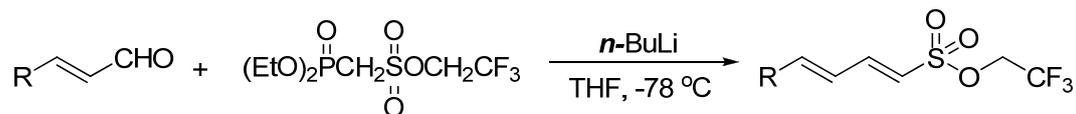
A mixture of [RhCl(C₂H₄)₂]₂ (1.2 mg, 6 μmol Rh) and the allylic phosphine (2.9 mg, 6 μmol; Table 2, Entry 1) in dioxane (2 mL) was stirred at room temperature for 20 min. Then, 0.50 M aqueous KOH (0.20 mL, 0.10 mmol) was added into it, followed by the

addition of cyclohexenone (119.2 mg, 0.20 mmol) and PhB(OH)₂ (73.2 mg, 0.60 mmol). The solution was stirred for 4.5 h at 50 °C, and was then passed through a pad of silica gel with EtOAc, and the solvent was removed under vacuum. The residue was chromatographed on silica gel with EtOAc/hexane to afford the 1,4-adduct .

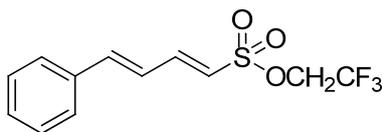


(CAS 57344-86-2). (22.0 mg; 63% yield). ¹H NMR data of the product are identical with the reported data of the product in literature.⁶ The ee was determined on a Daicel Chiralcel OD column with hexane/2-propanol = 98/2, flow = 1.0 mL/min. Retention times: 12.8 min [(*S*)-enantiomer], 13.6 min [(*R*)-enantiomer]. 27% ee.

General Procedures for the Preparation of $\alpha,\beta,\gamma,\delta$ -Unsaturated Sulfonic Esters

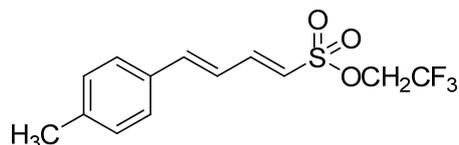


n-BuLi (1.92 mmol; 2.5 M in hexane) was added to a solution of 2,2,2-trifluoroethyl (diethoxyphosphoryl)methanesulfonate (0.6 g, 1.92 mmol) in THF at -78 °C and stirred for 20 min. Aldehyde (1.6 mmol) was added to it, and the resulting solution was allowed to warm to room temperature slowly and stirred at room temperature for 20 h. After quenching with AcOH (0.2 mL), the mixture was extracted with EtOAc, dried over MgSO₄, filtered and concentrated under vacuum. The residue was purified by silica gel chromatography with hexane/EtOAc = 30/1 to afford product as a white solid.

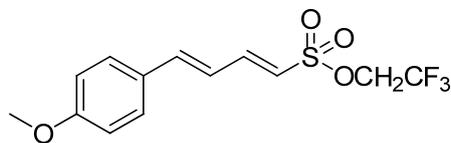


(0.42 g; 89% yield). ¹H NMR (300MHz, CDCl₃): δ 7.50-7.46 (m, 2H), 7.42-7.36 (m, 4H), 7.03 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.87-6.78 (m, 1H, olefin), 6.35 (d, $J_{\text{HH}} = 15.0$

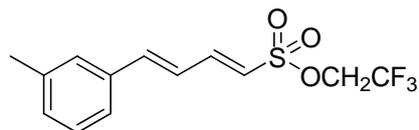
Hz, 1H, olefin), 4.42 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH₂). ¹³C NMR (75 MHz, CDCl₃): δ 146.7, 145.0, 135.0, 130.3, 129.1, 127.8, 122.7, 122.1 (q, $J_{\text{CF}} = 277.8$ Hz), 120.3, 64.6 (q, $J_{\text{CF}} = 38.0$ Hz). HRMS (ESI) calcd for C₁₂H₁₅F₃NO₃S (M+NH₄)⁺: 310.0719, found: 310.0726.



(0.38 g; 77% yield). ¹H NMR (300MHz, CDCl₃): δ 7.36-7.28 (m, 3H), 7.12-7.09 (m, 2H), 6.92 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.74-6.65 (m, 1H, olefin), 6.22 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.32 (q, $J_{\text{HF}} = 8.1$ Hz, 2H, OCH₂), 2.28 (s, 3H, Me). ¹³C NMR (75 MHz, CDCl₃): δ 147.0, 145.1, 140.8, 132.3, 129.8, 127.8, 122.1 (q, $J_{\text{CF}} = 277.8$ Hz), 121.8, 120.5, 64.6 (q, $J_{\text{CF}} = 38.0$ Hz), 21.5. HRMS (ESI) calcd for C₁₃H₁₃F₃NaO₃S (M+Na)⁺: 329.0430, found: 329.0417.

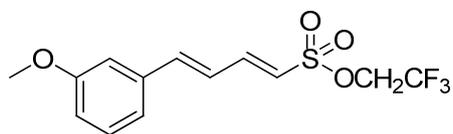


(0.39 g; 76% yield). ¹H NMR (300MHz, CDCl₃): δ 7.46-7.37 (m, 3H), 7.01 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.93-6.90 (m, 2H, Ar), 6.77-6.68 (m, 1H, olefin), 6.28 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.41 (q, $J_{\text{HF}} = 8.1$ Hz, 2H, OCH₂), 3.85 (s, 3H, OMe). ¹³C NMR (75 MHz, CDCl₃): δ 161.5, 147.3, 144.9, 129.5, 127.8, 122.3 (q, $J_{\text{CF}} = 277.8$ Hz), 120.6, 119.5, 114.6, 64.6 (q, $J_{\text{CF}} = 38.0$ Hz), 55.5. HRMS (ESI) calcd for C₁₃H₁₇F₃NO₄S (M+NH₄)⁺: 340.0825, found: 340.0817.

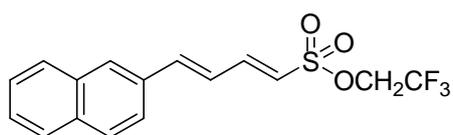


(0.37 g; 76% yield). ¹H NMR (300MHz, CDCl₃): δ 7.46-7.38 (m, 1H), 7.30-7.25 (m, 3H), 7.20-7.18 (m, 1H), 7.02 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.88-6.79 (m, 1H, olefin), 6.33 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.42 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH₂), 2.37 (s, 3H, Me).

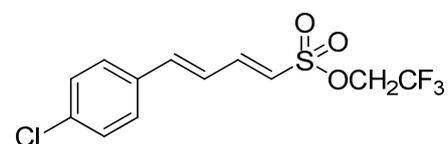
^{13}C NMR (75 MHz, CDCl_3): δ 146.8, 145.3, 138.9, 135.0, 131.2, 129.0, 128.5, 125.1, 122.6, 122.1 (q, $J_{\text{CF}} = 277.8$ Hz), 121.0, 64.6 (q, $J_{\text{CF}} = 37.5$ Hz), 21.4. HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{17}\text{F}_3\text{NO}_3\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 324.0876, found: 324.0879.



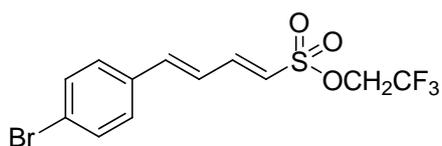
(0.38 g; 76% yield). ^1H NMR (300MHz, CDCl_3): δ 7.45-7.29 (m, 2H), 7.09-6.78 (m, 5H), 6.35 (d, $J_{\text{HH}} = 14.7$ Hz, 1H, olefin), 4.42 (q, $J_{\text{HF}} = 7.2$ Hz, 2H, OCH_2), 3.83 (s, 3H, OMe). ^{13}C NMR (75 MHz, CDCl_3): δ 160.1, 146.5, 144.9, 136.4, 130.1, 123.1, 122.3 (q, $J_{\text{CF}} = 277.8$ Hz), 121.4, 120.5, 116.0, 112.9, 64.6 (q, $J_{\text{CF}} = 38.6$ Hz), 55.4. HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{17}\text{F}_3\text{NO}_4\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 340.0825, found: 340.0829.



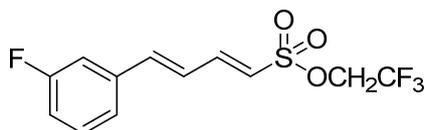
(0.27 g; 50% yield). ^1H NMR (300MHz, CDCl_3): δ 7.74-7.71 (m, 4H), 7.51-7.31 (m, 4H), 7.03 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.84-6.75 (m, 1H, olefin), 6.25 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.34 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH_2). ^{13}C NMR (75 MHz, CDCl_3): δ 146.7, 145.1, 134.2, 133.4, 132.5, 129.6, 128.9, 128.6, 127.9, 127.5, 127.0, 123.3, 123.0, 122.2, 122.3 (q, $J_{\text{CF}} = 277.8$ Hz), 64.6 (q, $J_{\text{CF}} = 37.4$ Hz). HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{17}\text{F}_3\text{NO}_3\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 360.0876, found: 360.0879.



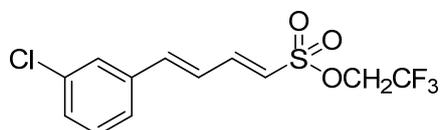
(0.38 g; 72% yield). ^1H NMR (300MHz, CDCl_3): δ 7.35-7.46 (m, 5H), 7.01 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.78-6.86 (m, 1H, olefin), 6.38 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.43 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH_2). ^{13}C NMR (75 MHz, CDCl_3): δ 146.2, 143.5, 136.2, 133.5, 129.4, 129.0, 123.3, 122.1 (q, $J_{\text{CF}} = 277.9$ Hz), 121.9, 64.7 (q, $J_{\text{CF}} = 38.0$ Hz). HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{ClF}_3\text{NO}_3\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 344.0330, found: 344.0331.



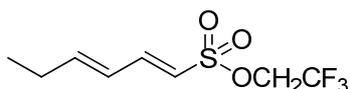
(0.36 g; 61% yield). ^1H NMR (300MHz, CDCl_3): δ 7.54-7.51 (m, 2H), 7.45-7.34 (m, 3H), 6.99 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.88-6.79 (m, 1H, olefin), 6.38 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.43 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH_2). ^{13}C NMR (75 MHz, CDCl_3): δ 146.1, 143.5, 133.9, 132.4, 129.2, 124.6, 123.4, 122.1 (q, $J_{\text{CF}} = 277.9$ Hz), 122.1, 64.7 (q, $J_{\text{CF}} = 38.0$ Hz). HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{10}\text{BrF}_3\text{NaO}_3\text{S}$ ($\text{M}+\text{Na}$) $^+$: 392.9378, found: 392.9362.



(0.35 g; 77% yield). ^1H NMR (300MHz, CDCl_3): δ 7.46-7.33 (m, 2H), 7.27-7.17 (m, 2H), 7.10-6.98 (m, 2H), 6.88-6.79 (m, 1H, olefin), 6.40 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.44 (q, $J_{\text{HF}} = 7.8$ Hz, 2H, OCH_2). ^{13}C NMR (75 MHz, CDCl_3): δ 163.1 (d, $J_{\text{CF}} = 247.1$ Hz), 145.9, 143.4 (d, $J_{\text{CF}} = 2.9$ Hz), 137.2 (d, $J_{\text{CF}} = 8.0$ Hz), 130.7 (d, $J_{\text{CF}} = 8.4$ Hz), 124.0, 123.8 (d, $J_{\text{CF}} = 2.8$ Hz), 122.4, 122.1 (q, $J_{\text{CF}} = 278.3$ Hz), 117.1 (d, $J_{\text{CF}} = 21.2$ Hz), 114.0 (d, $J_{\text{CF}} = 22.4$ Hz), 64.7 (q, $J_{\text{CF}} = 38.0$ Hz). HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{F}_4\text{NO}_3\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 328.0625, found: 328.0631.

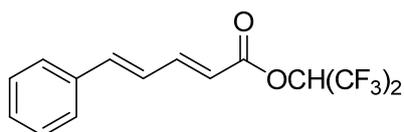


(0.6 g; 69% yield). ^1H NMR (300MHz, CDCl_3): δ 7.47-7.34 (m, 5H), 6.98 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 6.88-6.80 (m, 1H, olefin), 6.40 (d, $J_{\text{HH}} = 15.0$ Hz, 1H, olefin), 4.40 (q, $J_{\text{HF}} = 8.1$ Hz, 2H, OCH_2). ^{13}C NMR (75 MHz, CDCl_3): δ 145.9, 143.1, 136.8, 135.1, 130.4, 130.1, 127.5, 126.0, 124.1, 122.5, 122.1 (q, $J_{\text{CF}} = 277.9$ Hz), 64.7 (q, $J_{\text{CF}} = 38.0$ Hz). HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{ClF}_3\text{NO}_3\text{S}$ ($\text{M}+\text{NH}_4$) $^+$: 344.0330, found: 344.0333.



(0.23 g; 60% yield). ¹H NMR (300MHz, CDCl₃): δ 7.30-7.22 (m, 1H, olefin), 6.45-6.35 (m, 1H, olefin), 6.24-6.16 (m, 2H, olefin), 4.39 (q, *J*_{HF} = 7.8 Hz, 2H, OCH₂), 2.31-2.22 (m, 2H, =CCH₂), 1.08 (t, *J*_{HH} = 7.5 Hz, 3H, Me). ¹³C NMR (75 MHz, CDCl₃): δ 151.4, 147.3, 124.5, 120.8, 122.1 (q, *J*_{CF} = 208.5 Hz), 64.4 (q, *J*_{CF} = 28.3 Hz), 26.1, 12.3. HRMS (ESI) calcd for C₈H₁₂F₃O₃S (M+H)⁺: 245.0454, found: 245.0452.

The substrate **1f** was prepared from the corresponding acyl chloride with hexafluoroisopropanol in the presence of Et₃N following the literature procedure.

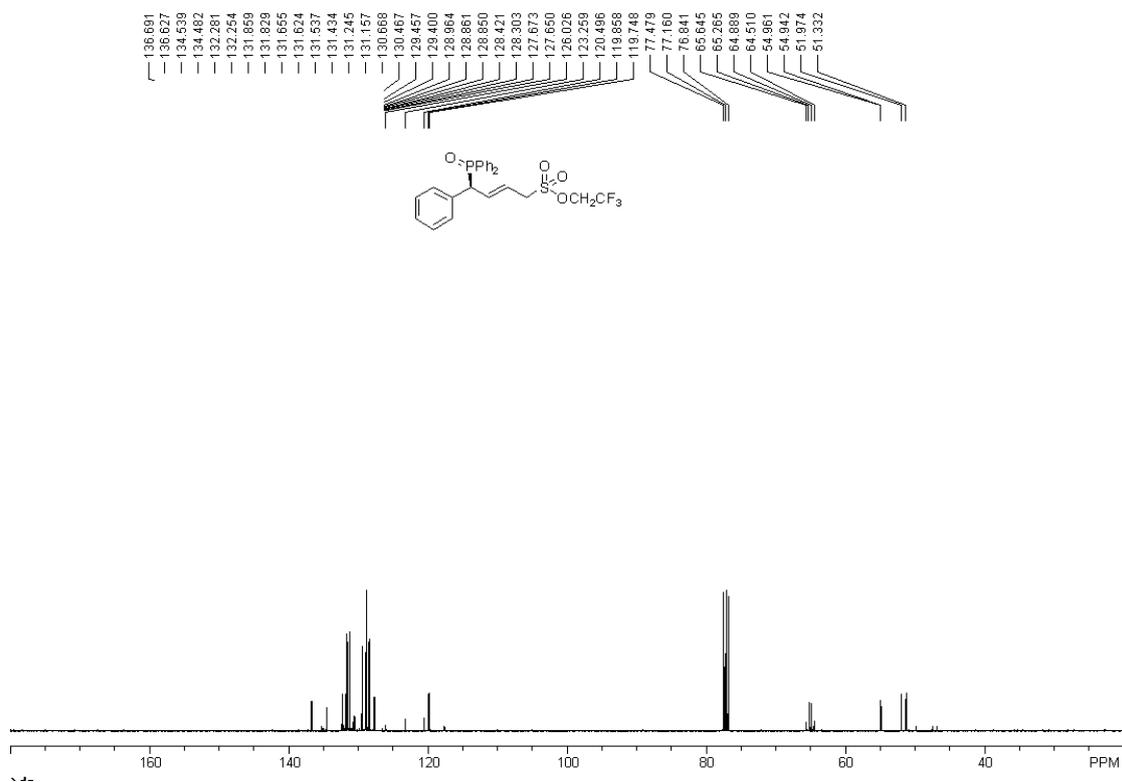
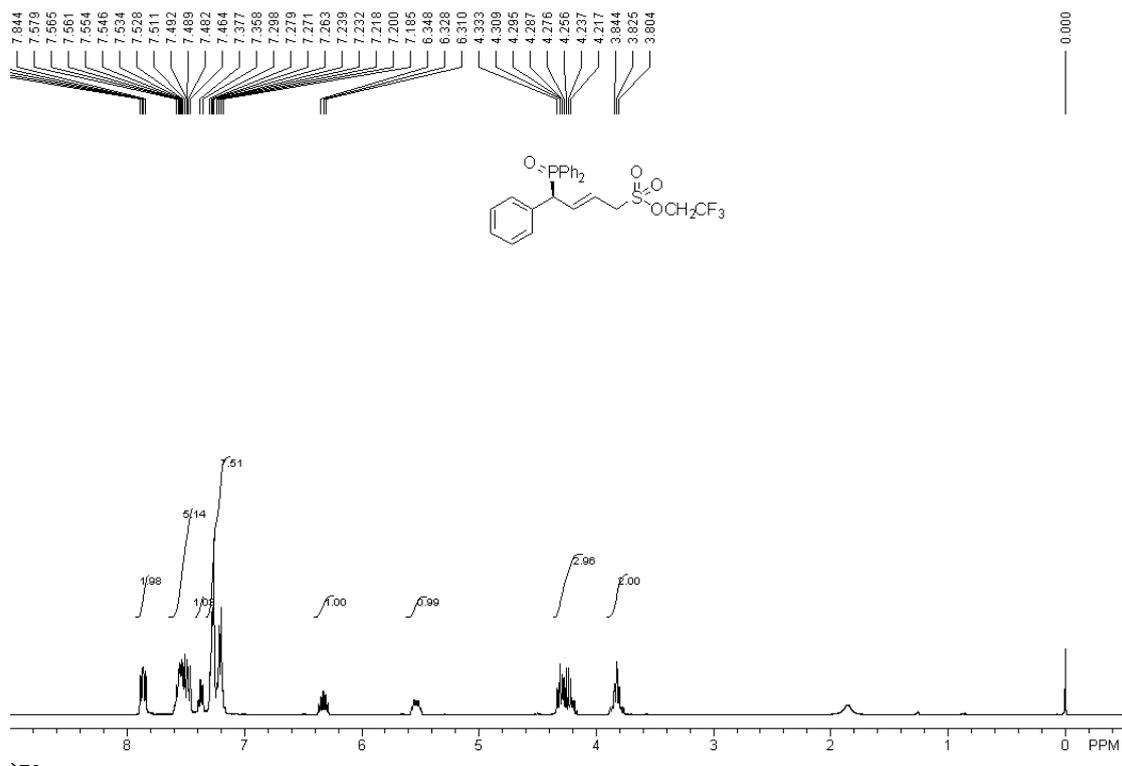


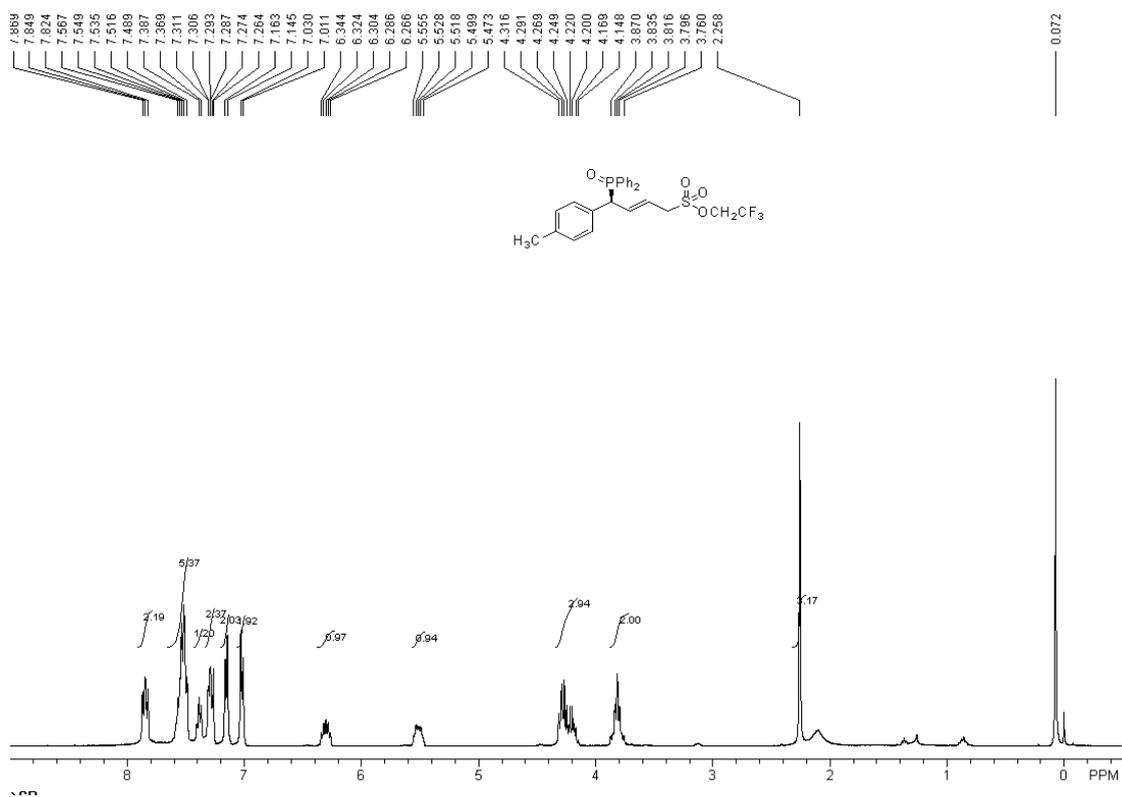
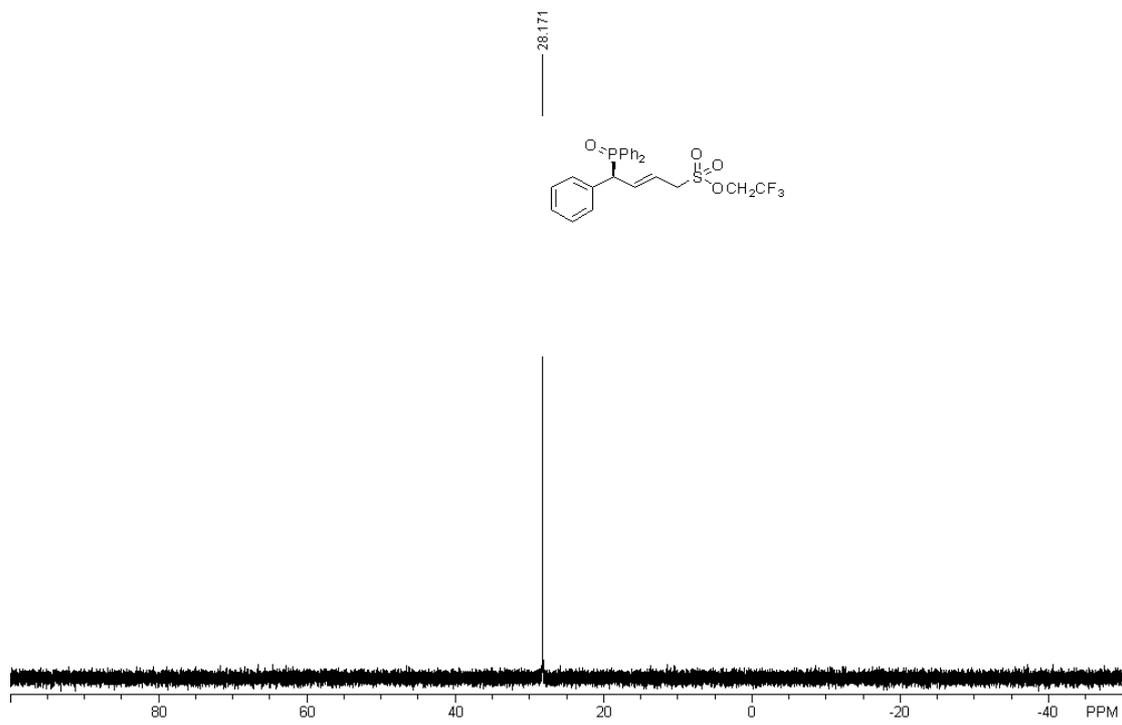
71% yield. ¹H NMR (300MHz, CDCl₃): δ 7.66 (dd, *J*_{HH} = 15.3 and 10.2 Hz, 1H, olefin), 7.53-7.50 (m, 2H, Ar), 7.46-7.40 (m, 3H, Ar), 7.02-6.85 (m, 2H, olefin), 6.06 (q, *J*_{HH} = 15.3 Hz, 1H, olefin), 6.08-5.98 (m, 1H, OCH). ¹³C NMR (75 MHz, CDCl₃): δ 163.5, 149.1, 143.4, 135.7, 129.9, 129.0, 127.7, 125.5, 122.7, 120.9 (q, *J*_{CF} = 282.9 Hz), 116.9, 66.6 (sept, *J*_{CF} = 34.7 Hz). MS (ESI): 324.8 (M⁺).

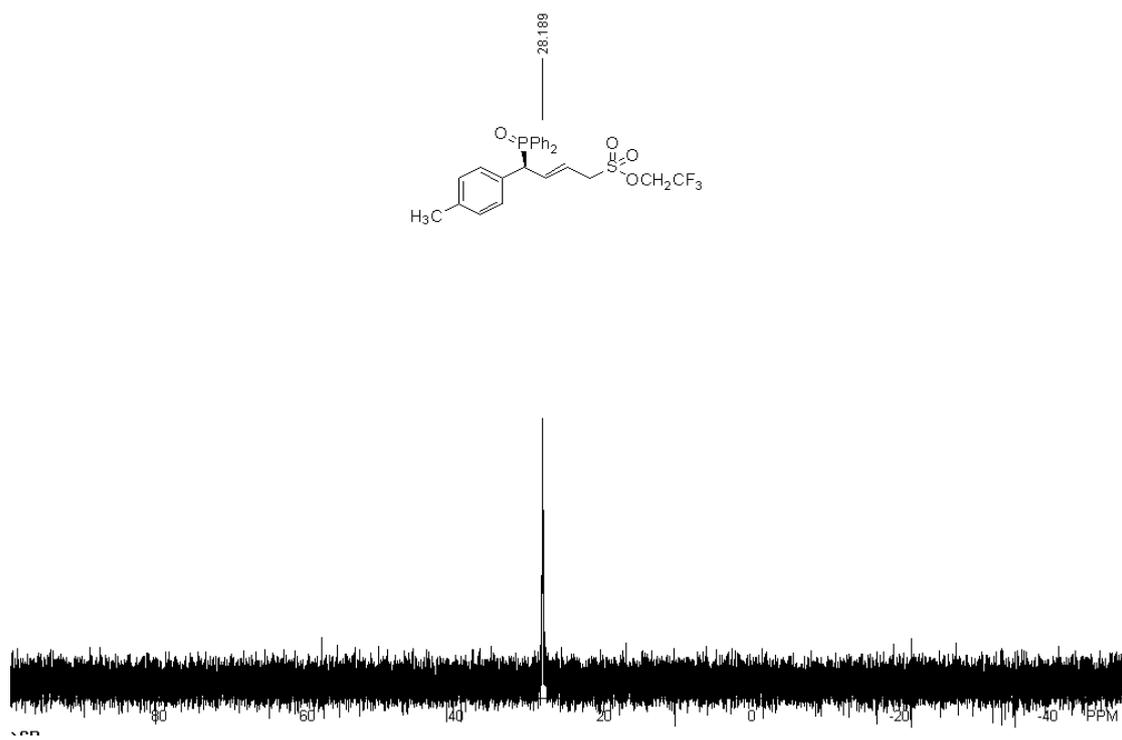
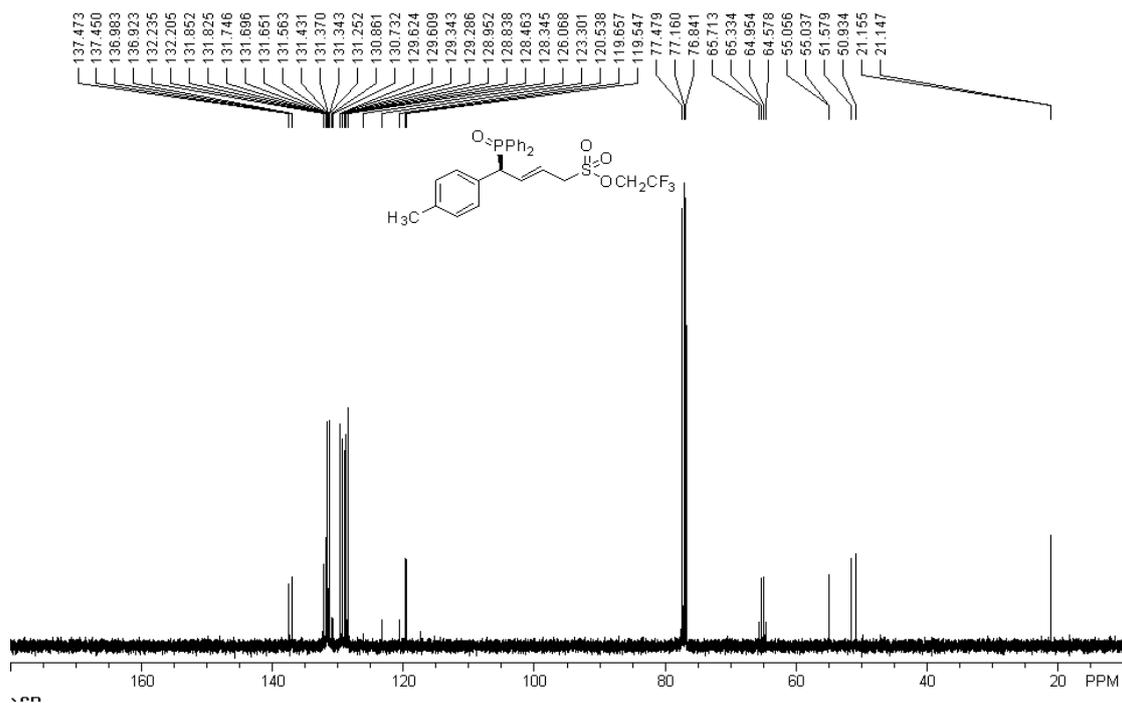
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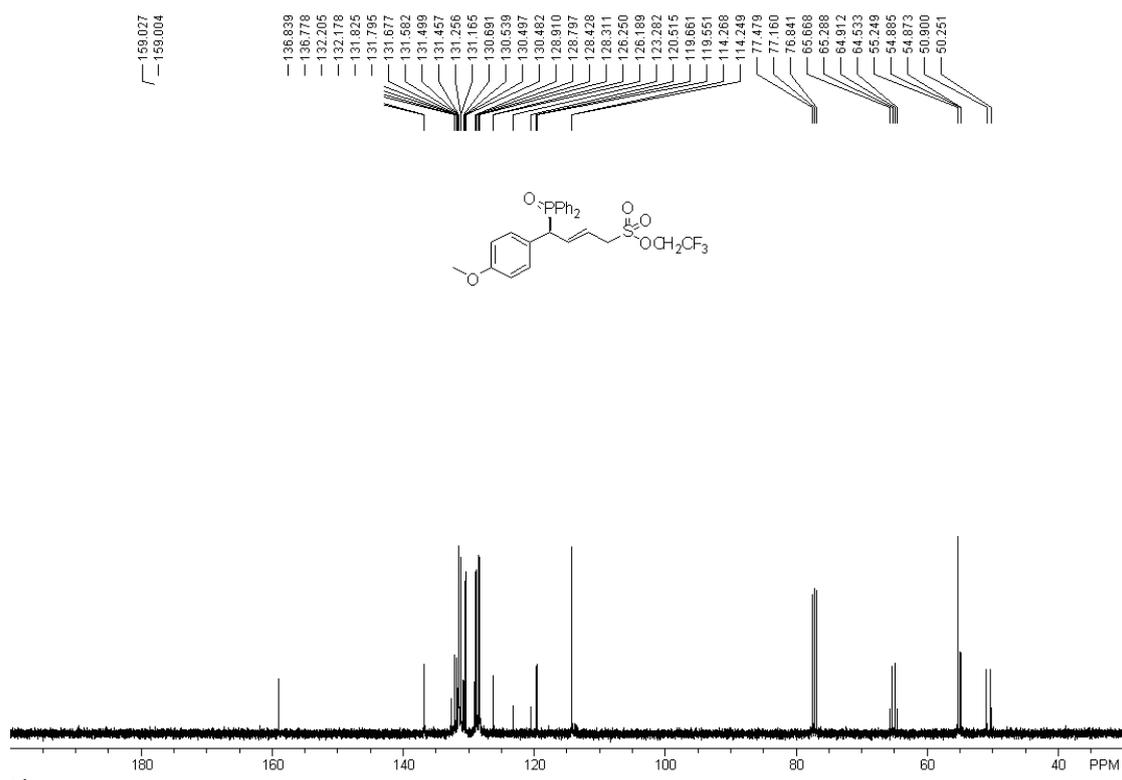
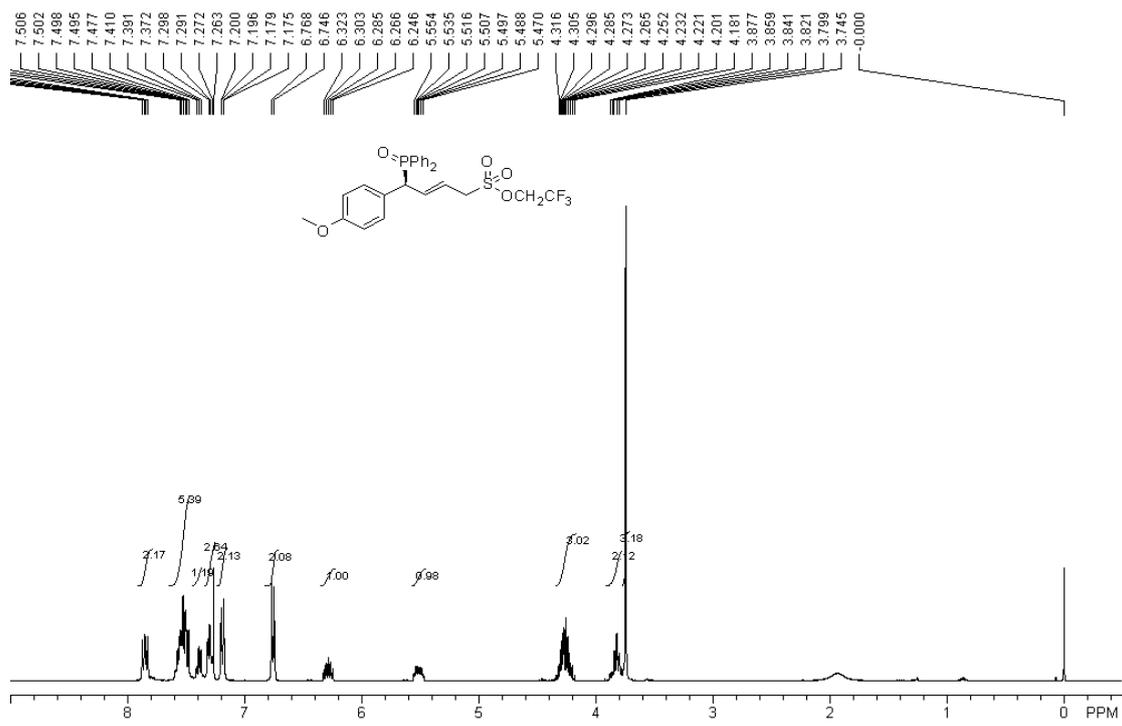
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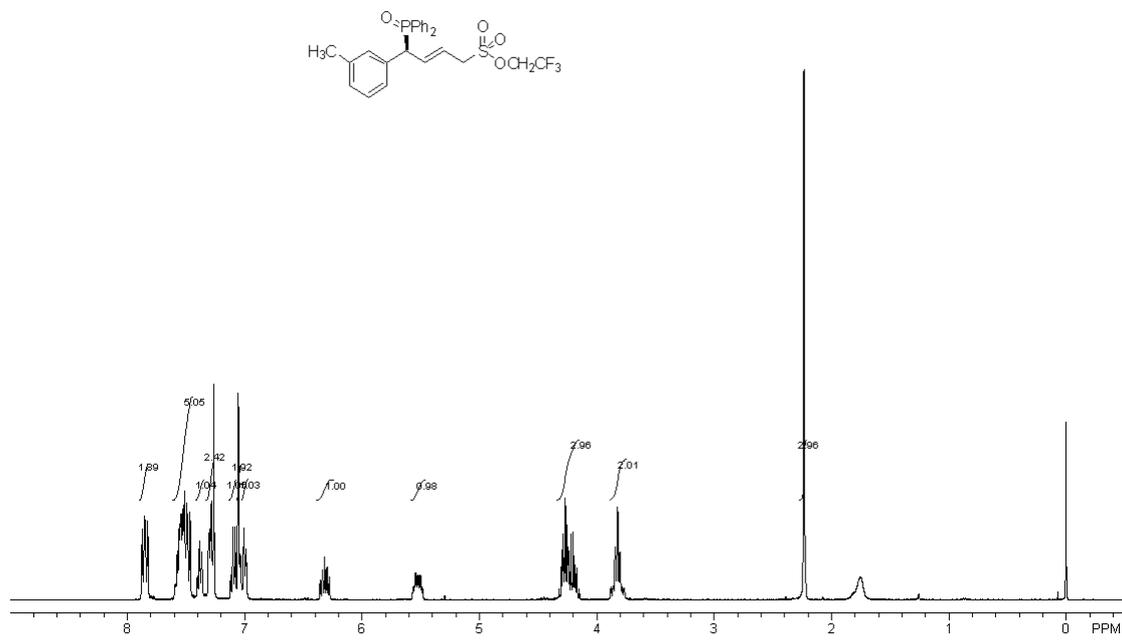
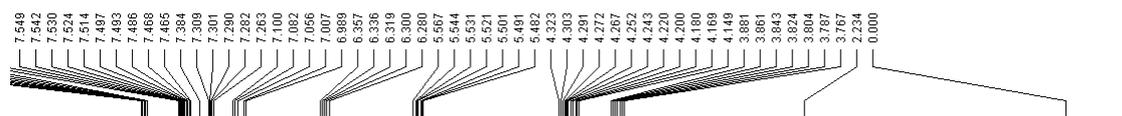
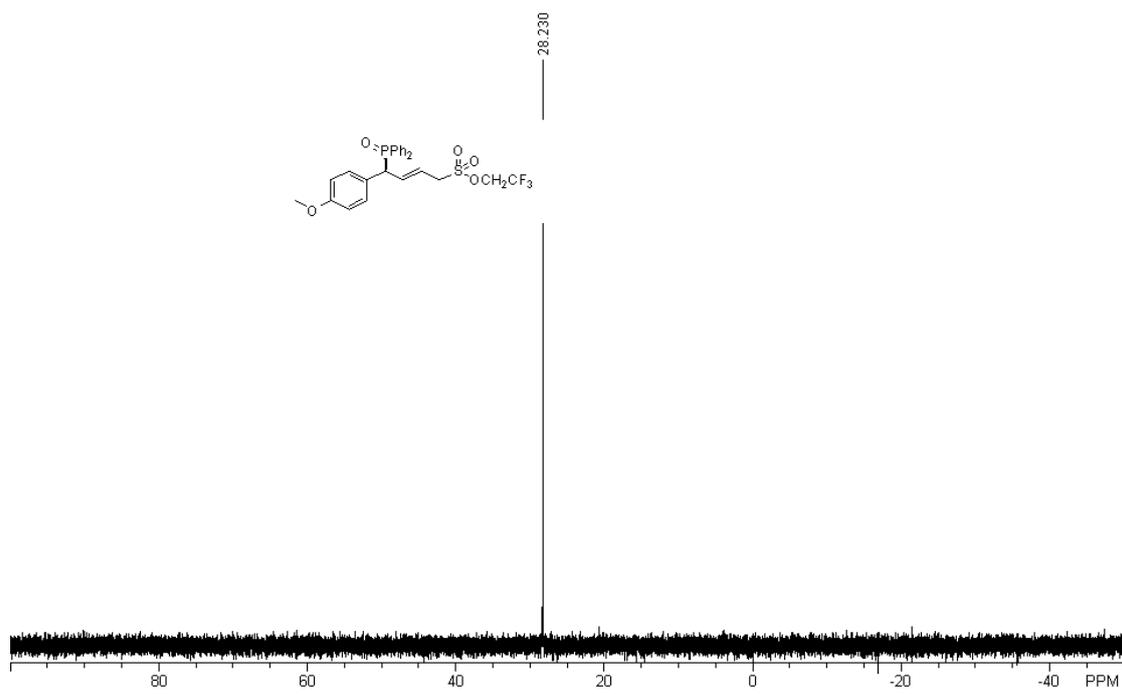
NMR Spectra

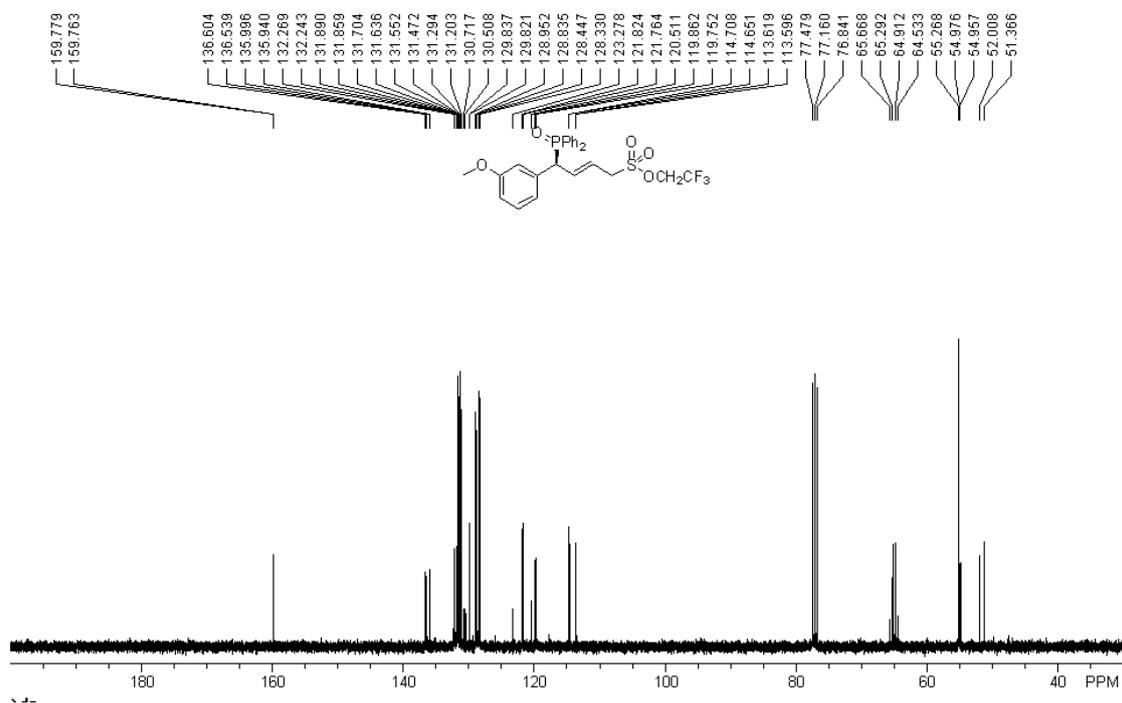
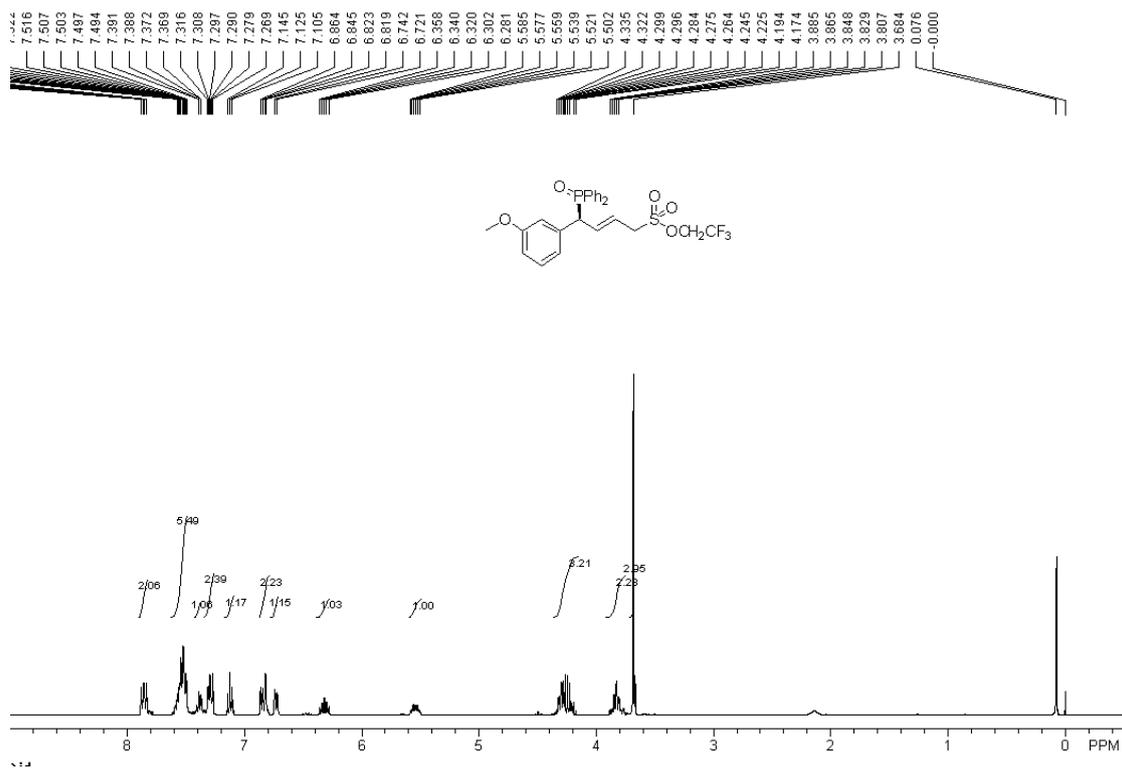


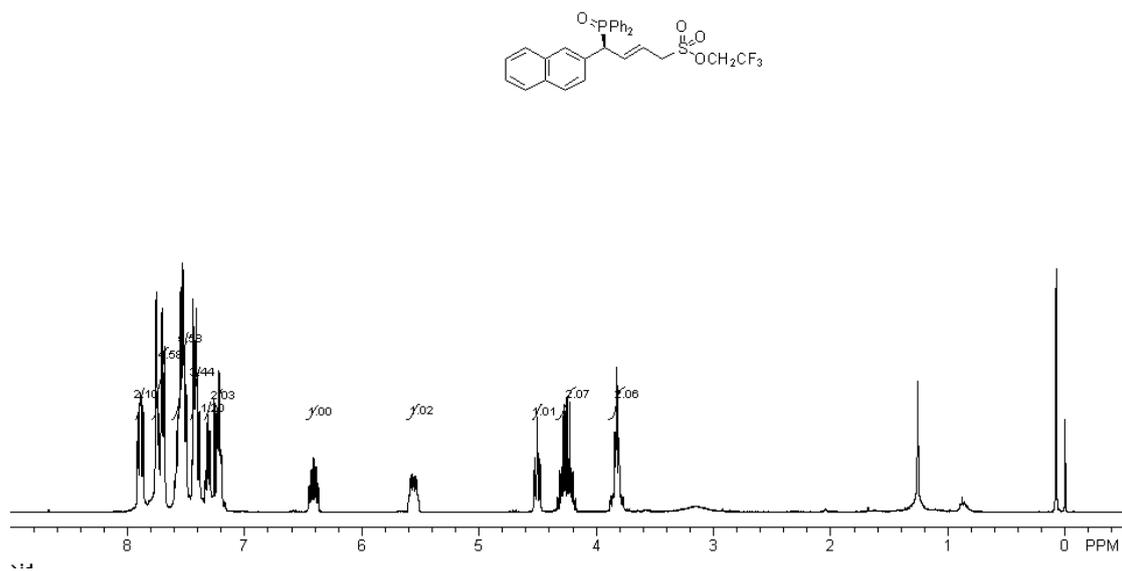
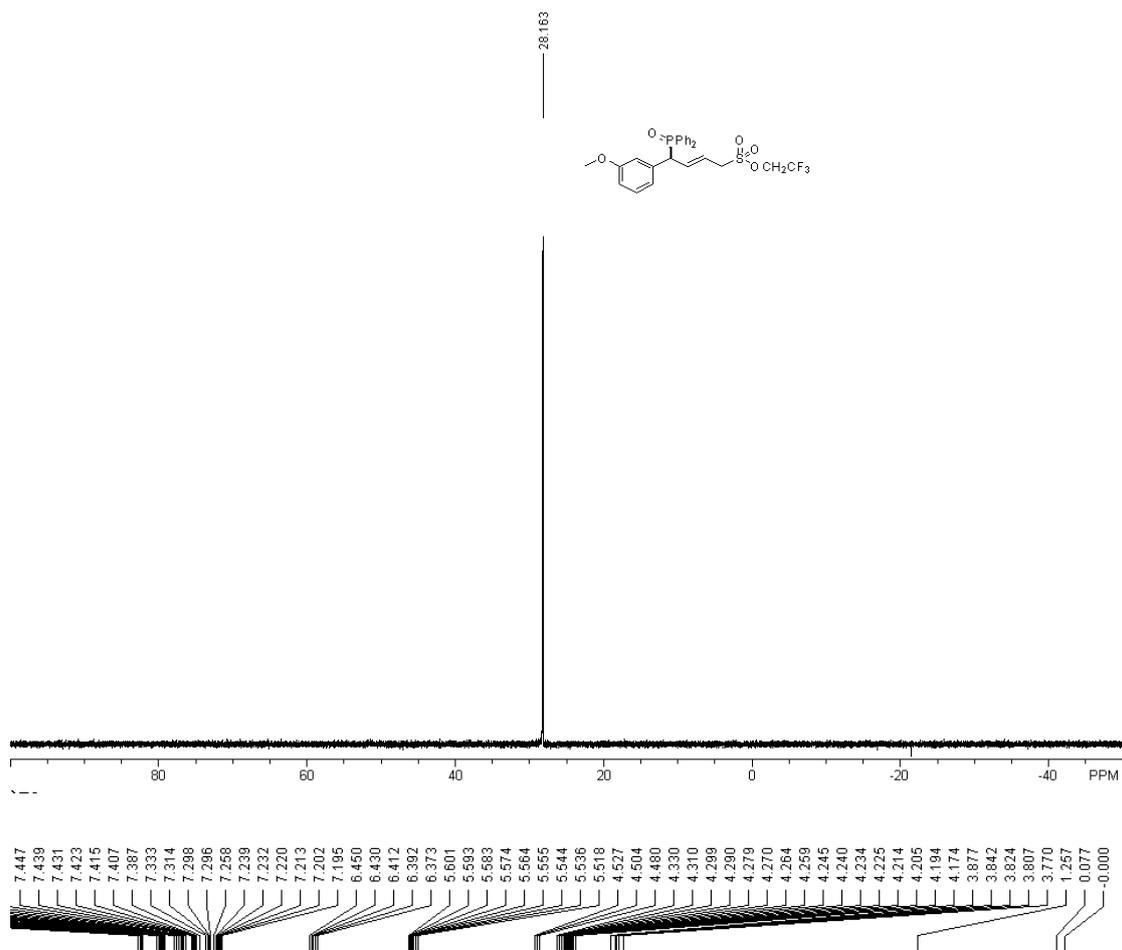


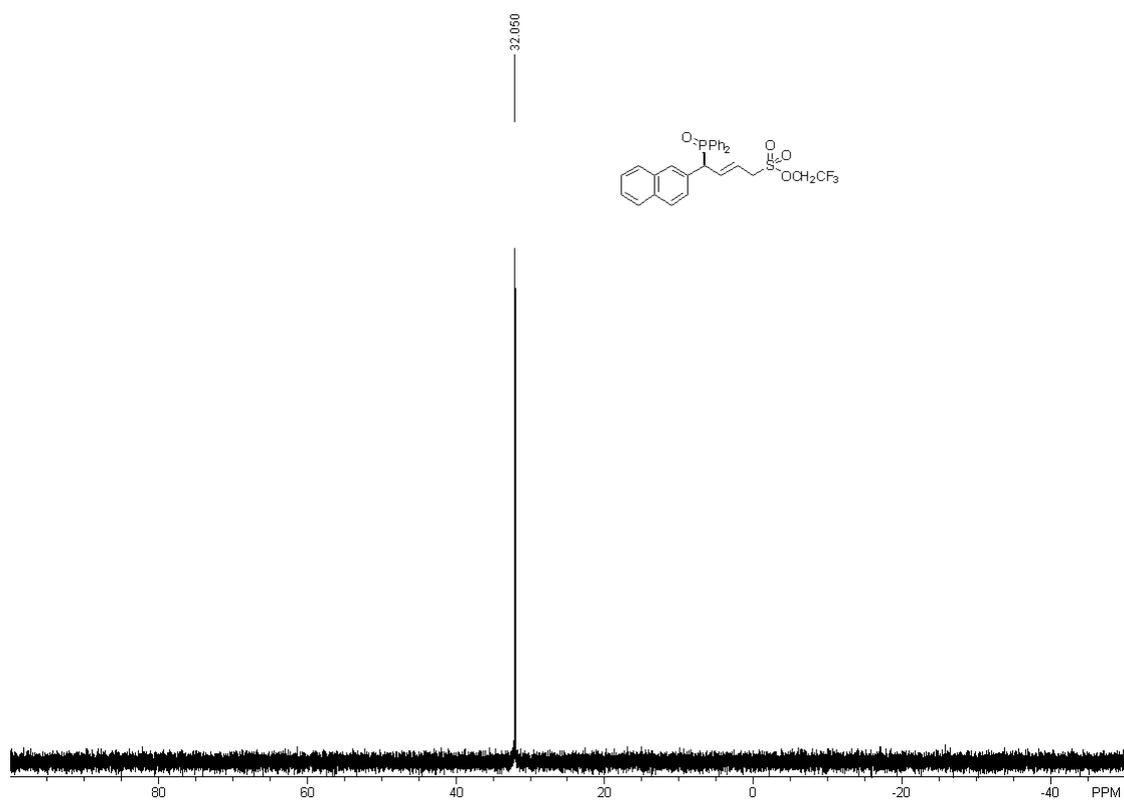
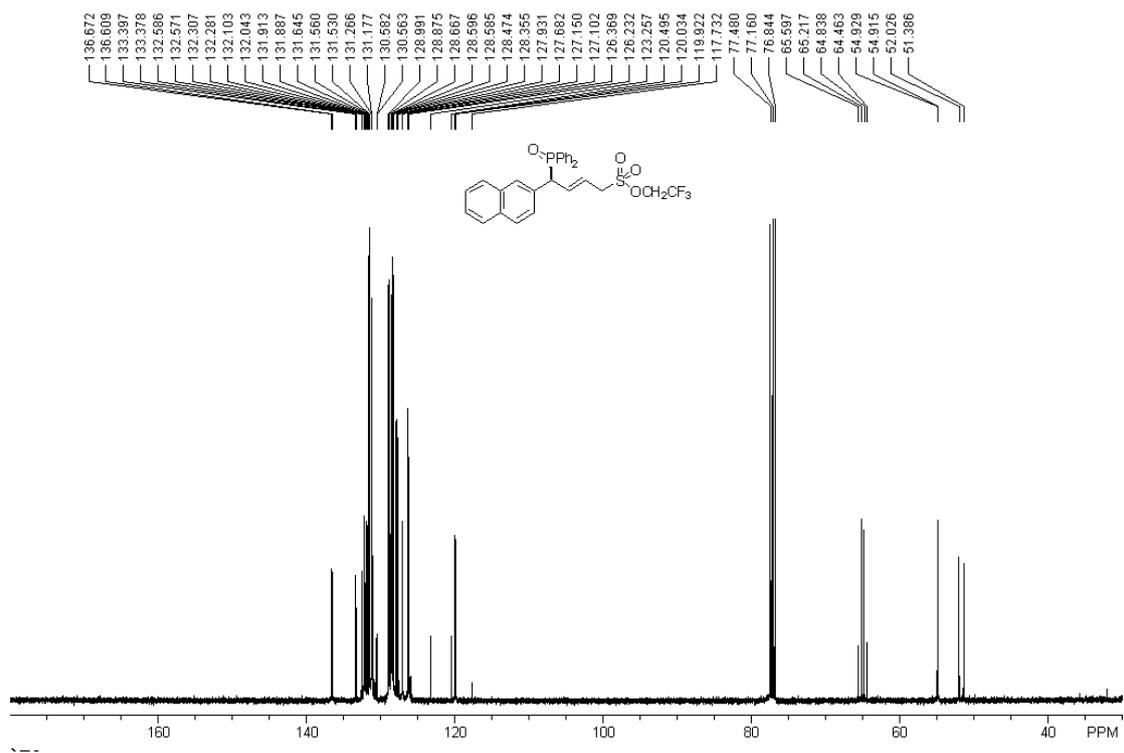


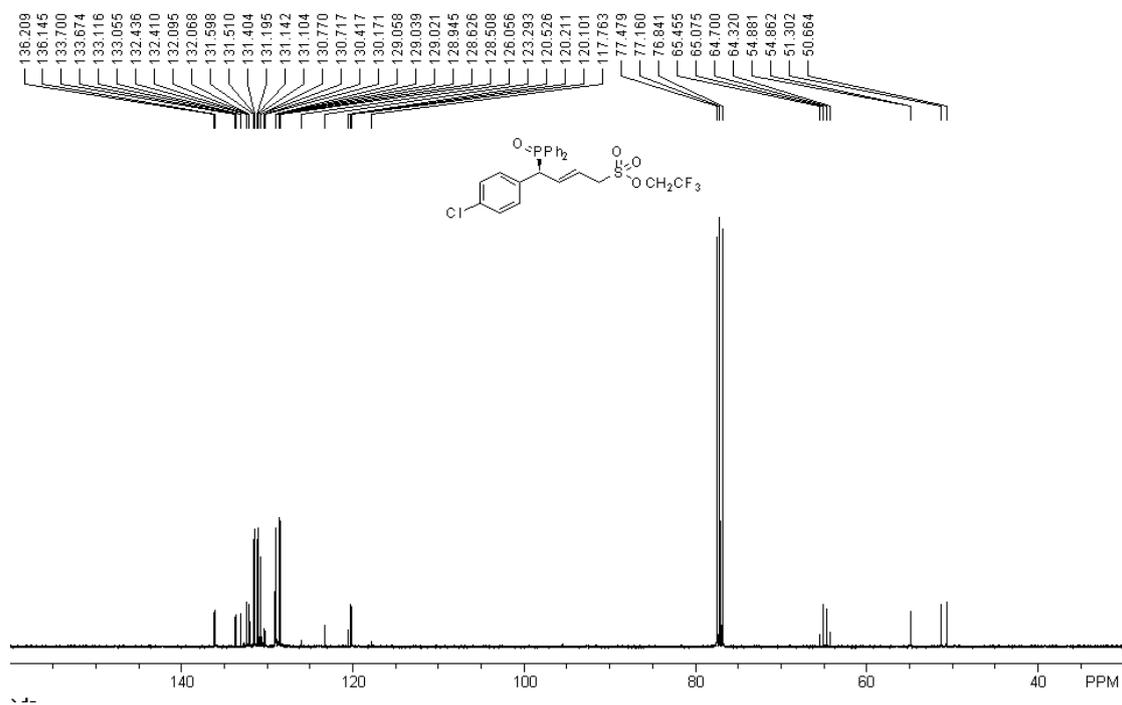
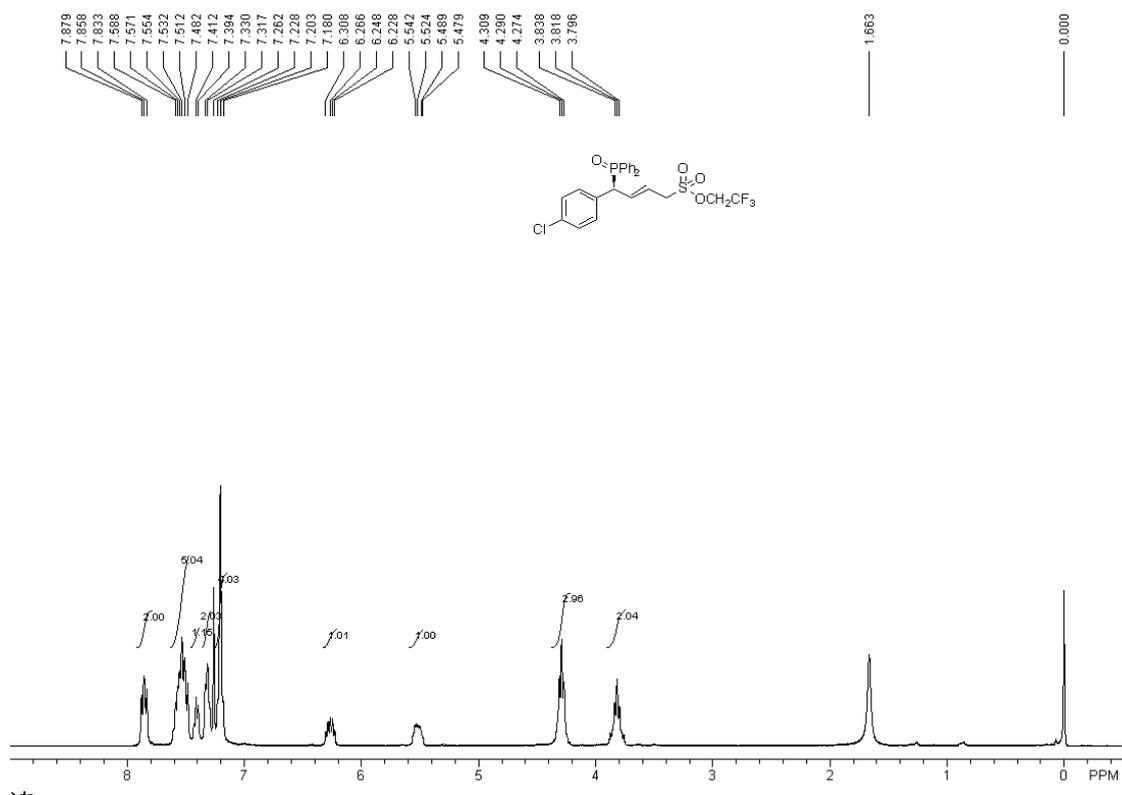


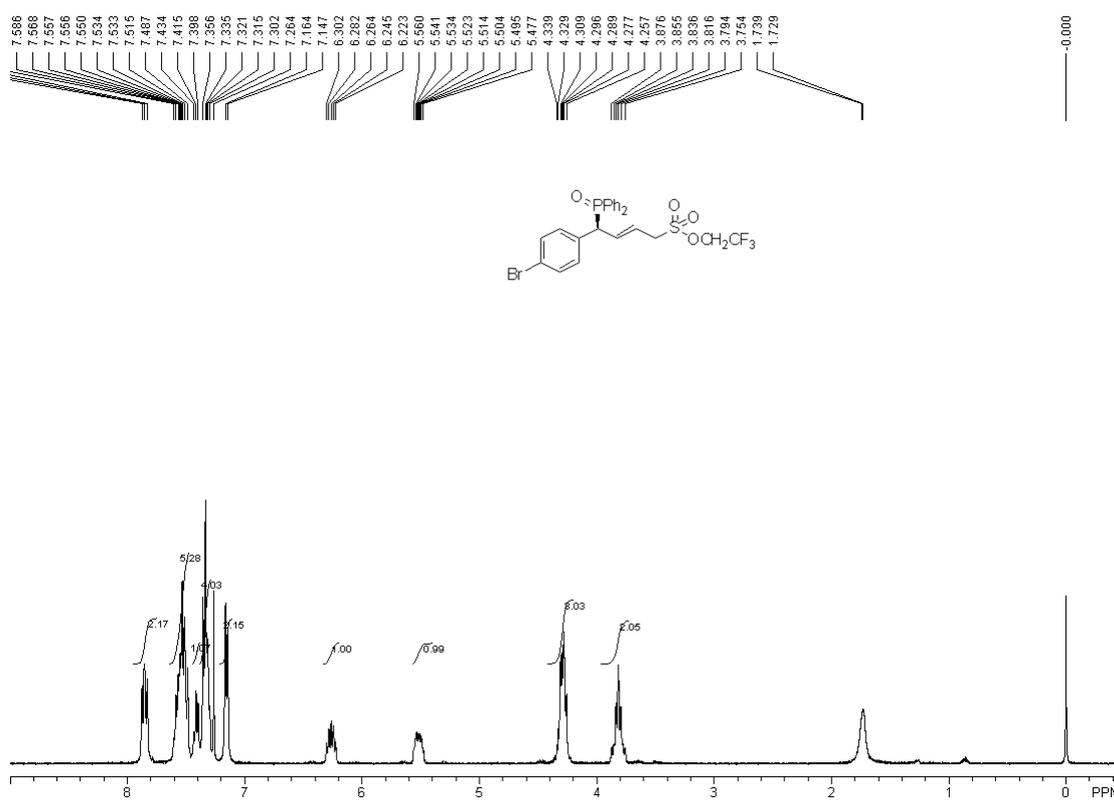
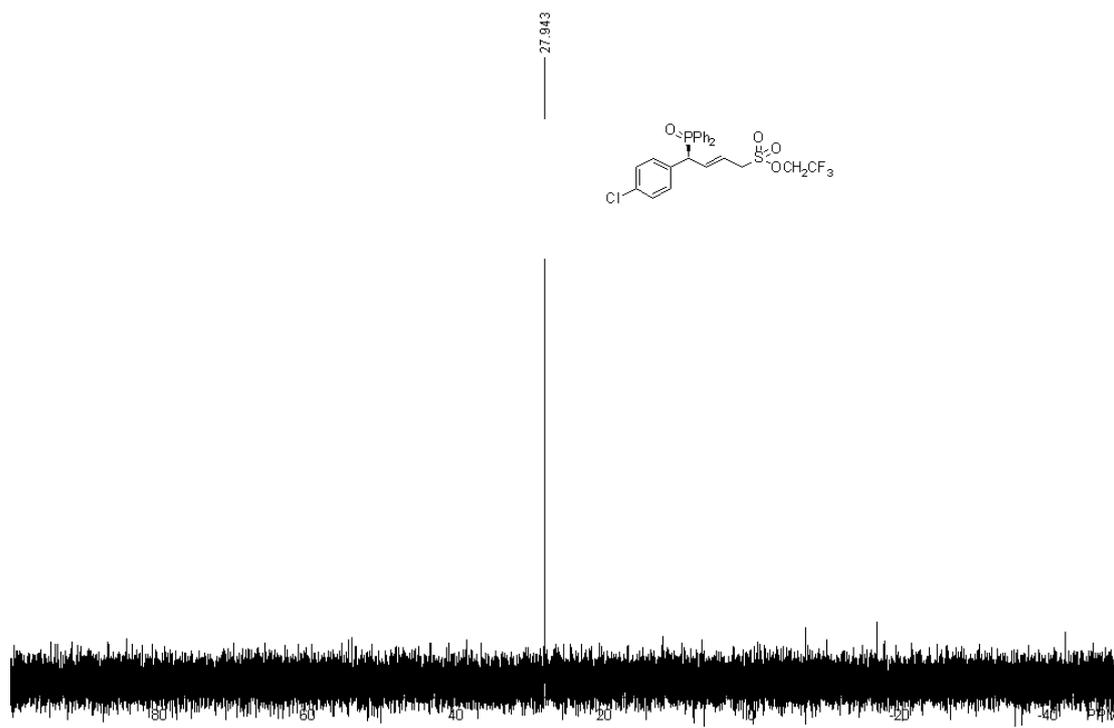


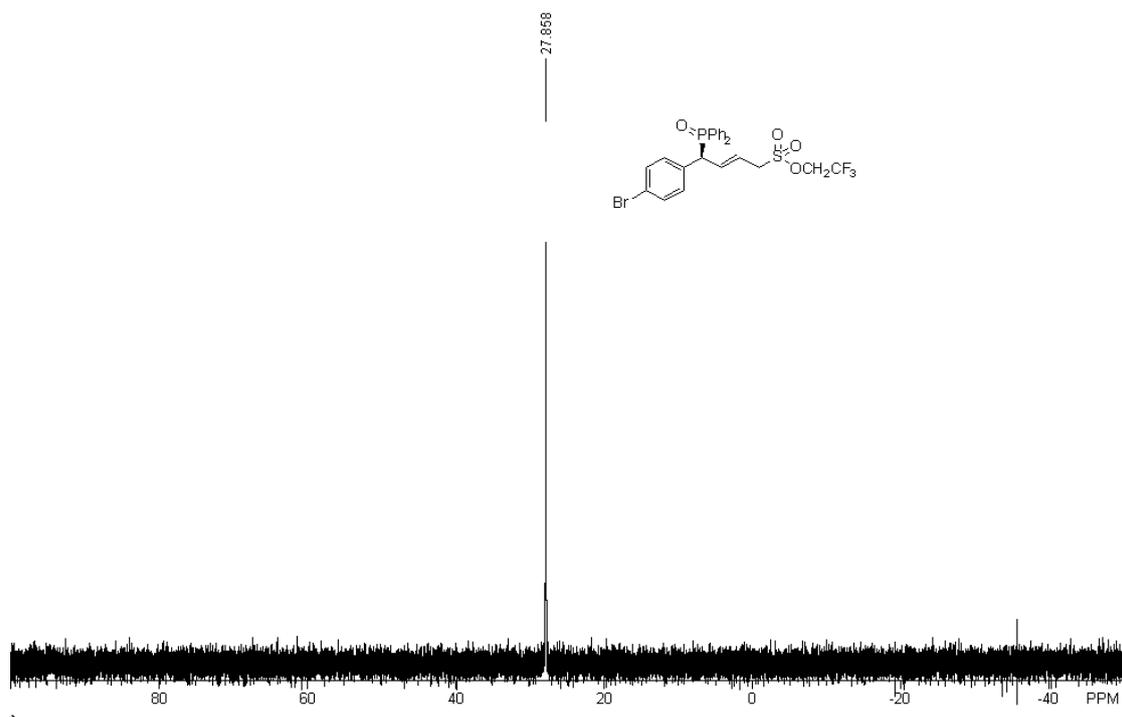
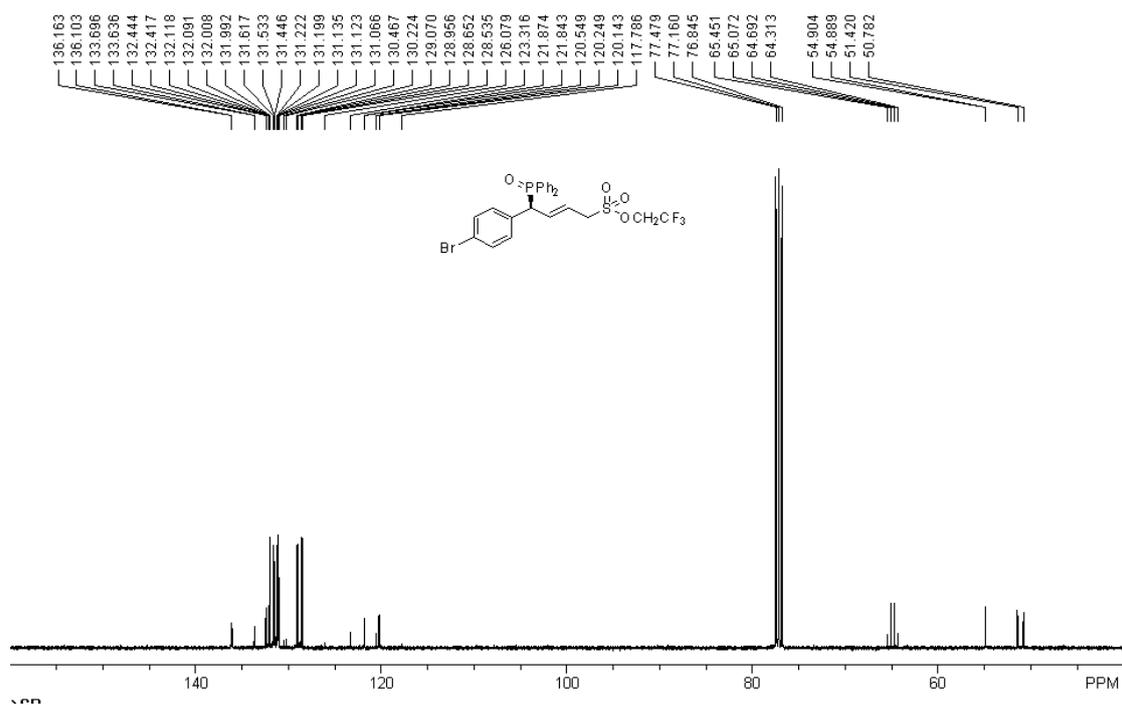


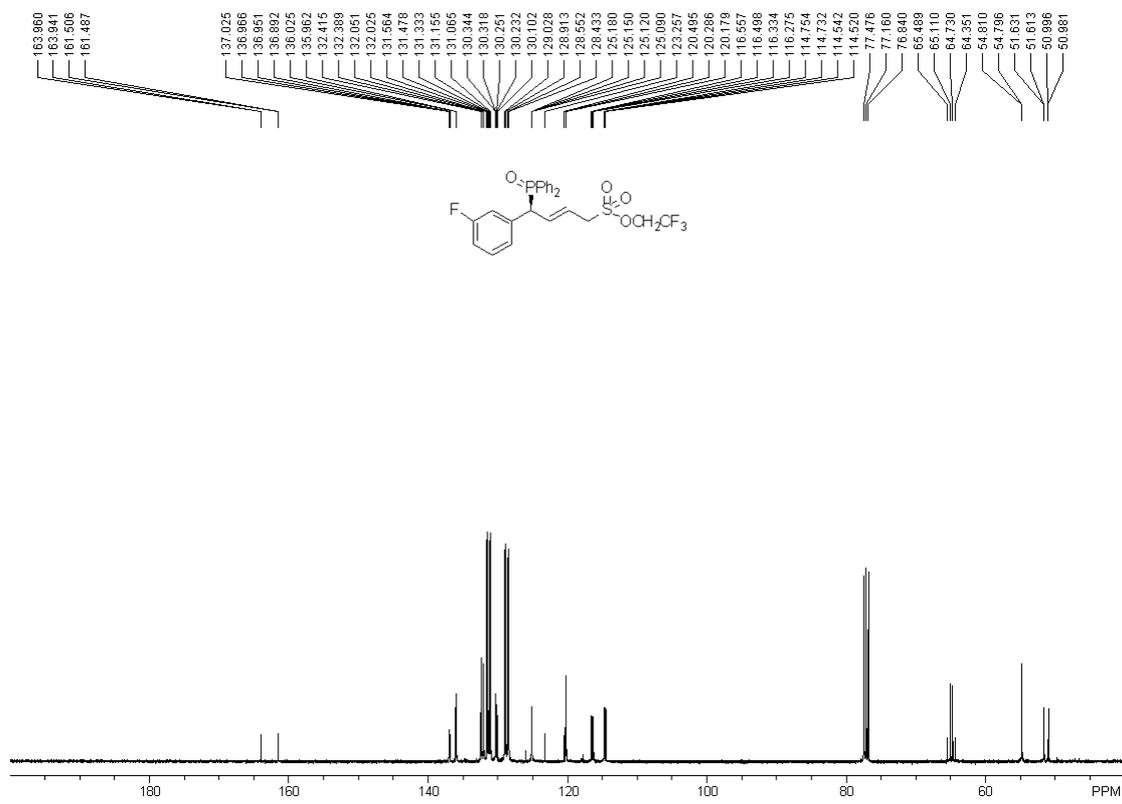
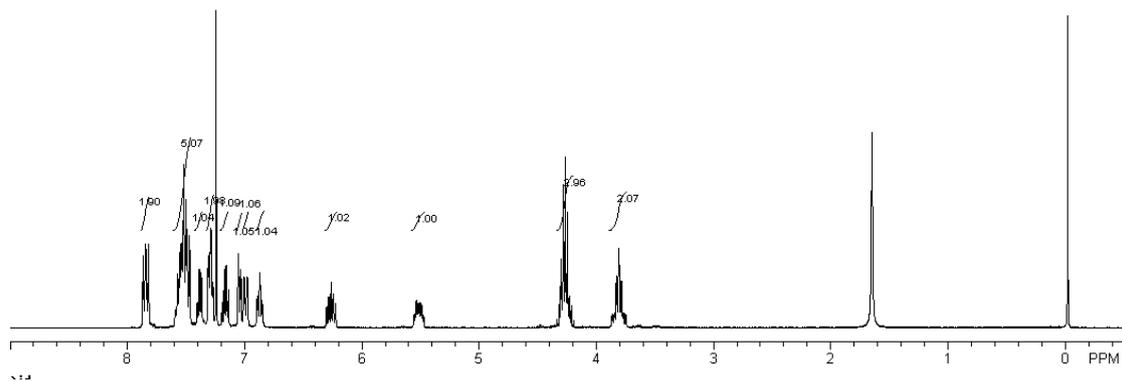
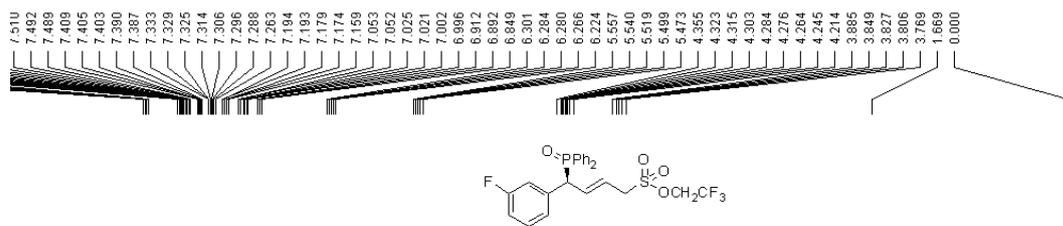


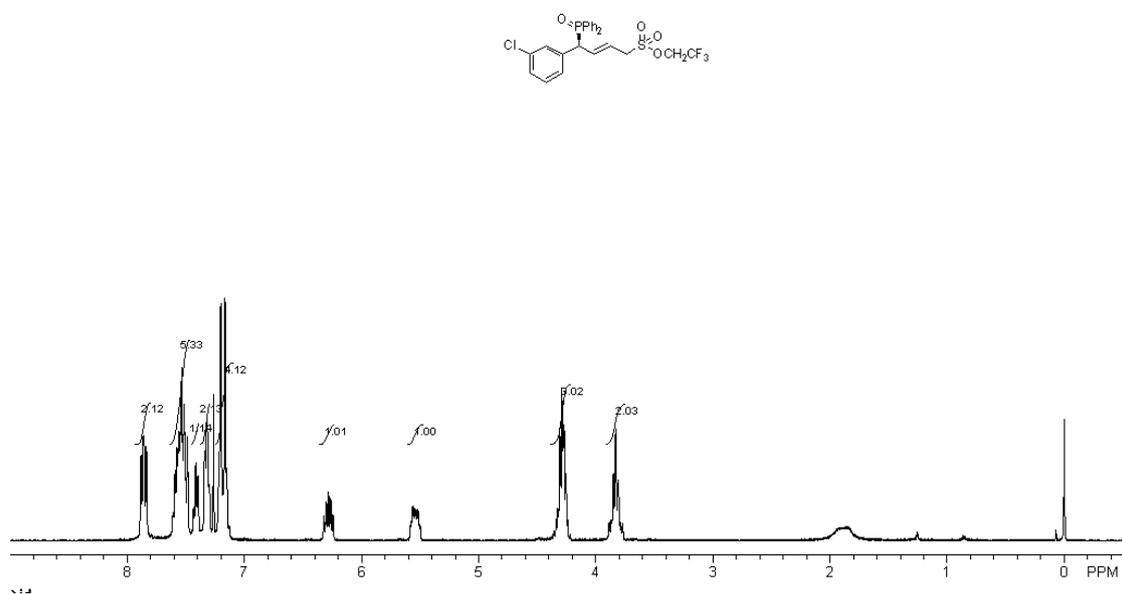
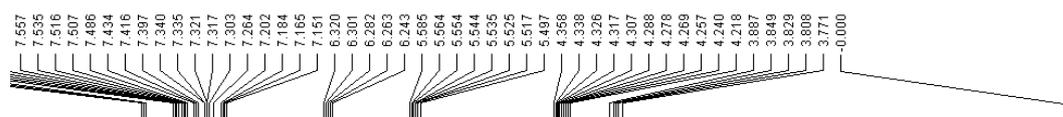
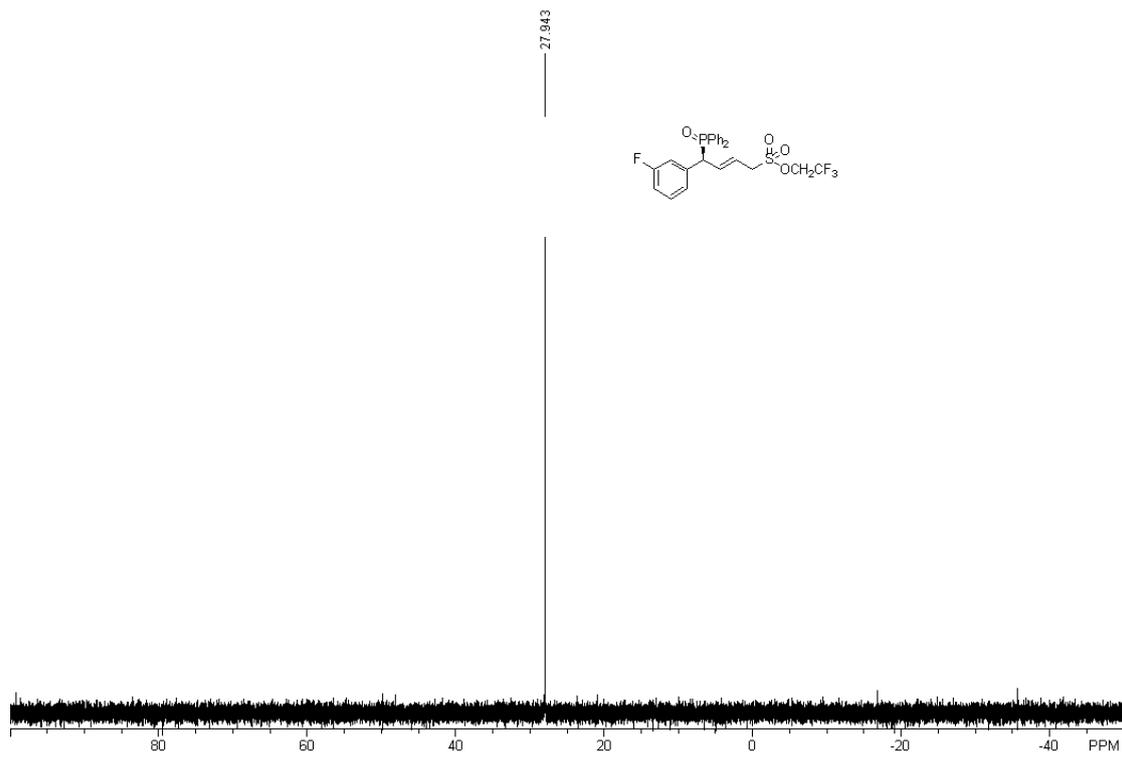


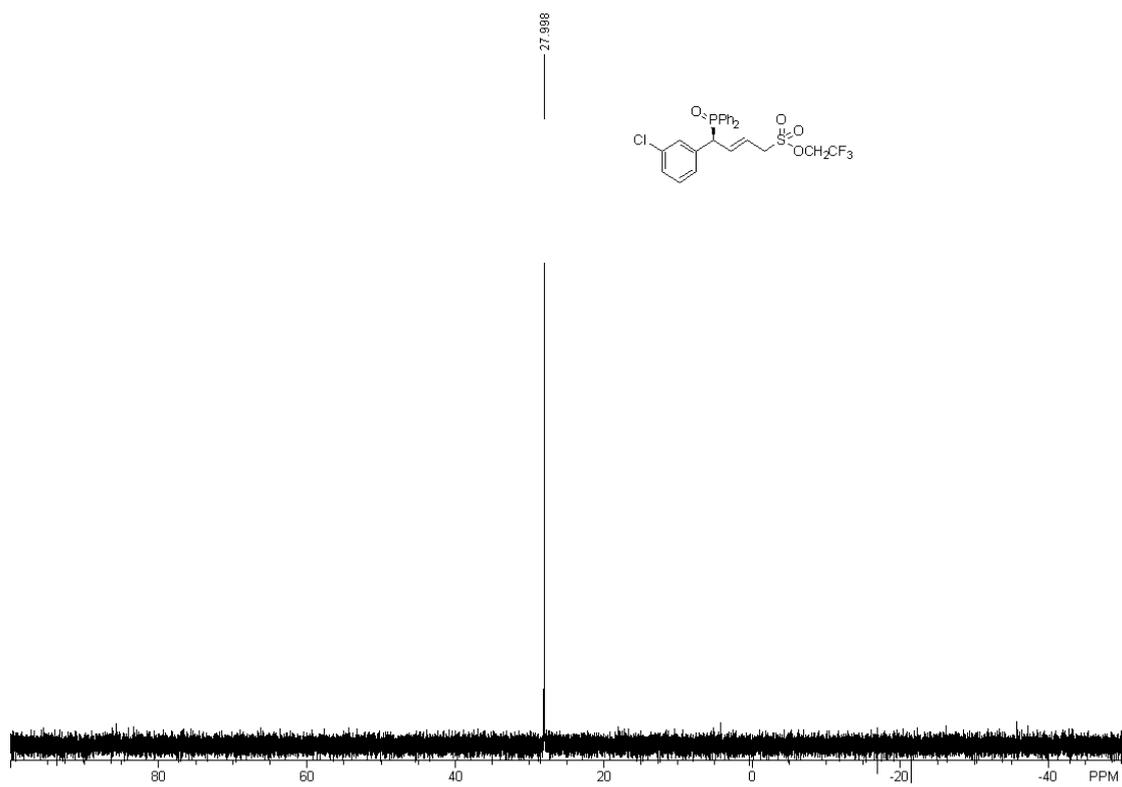
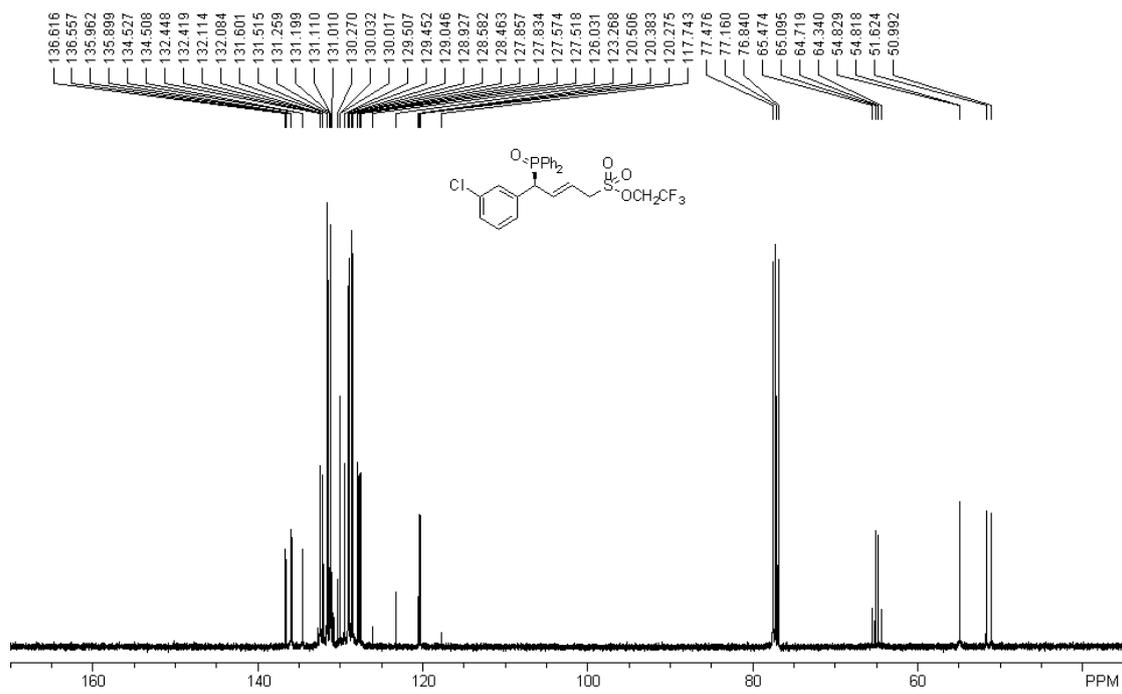


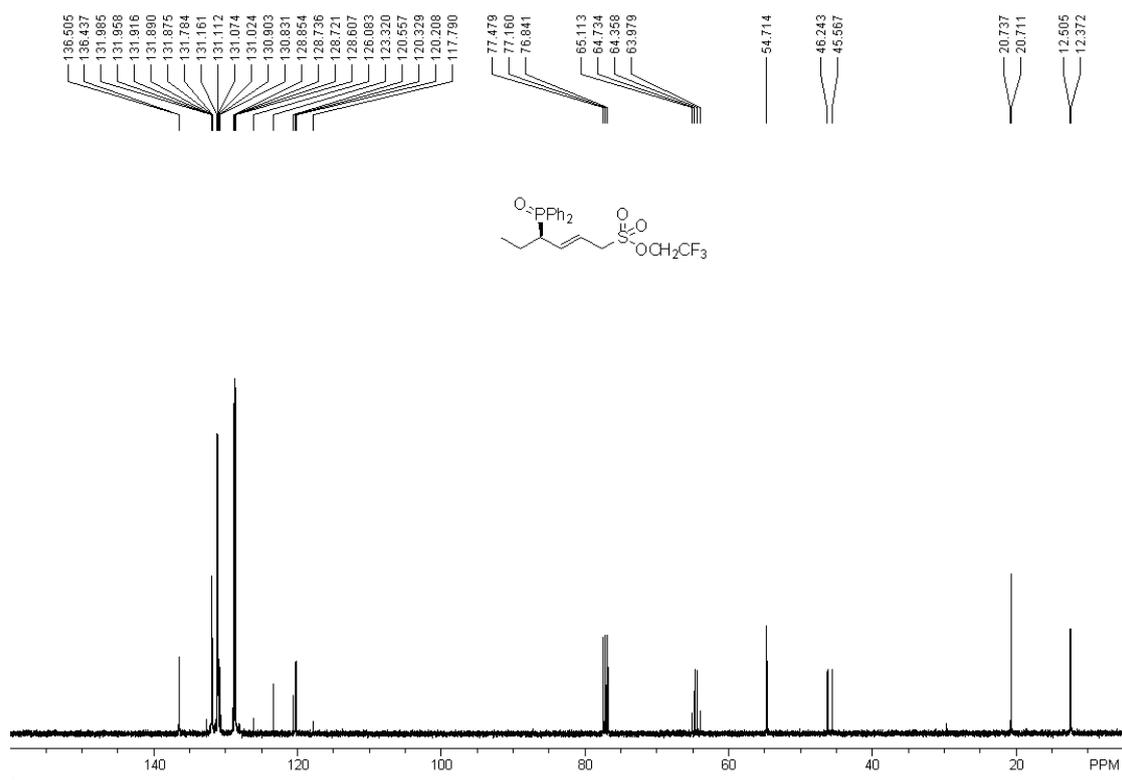
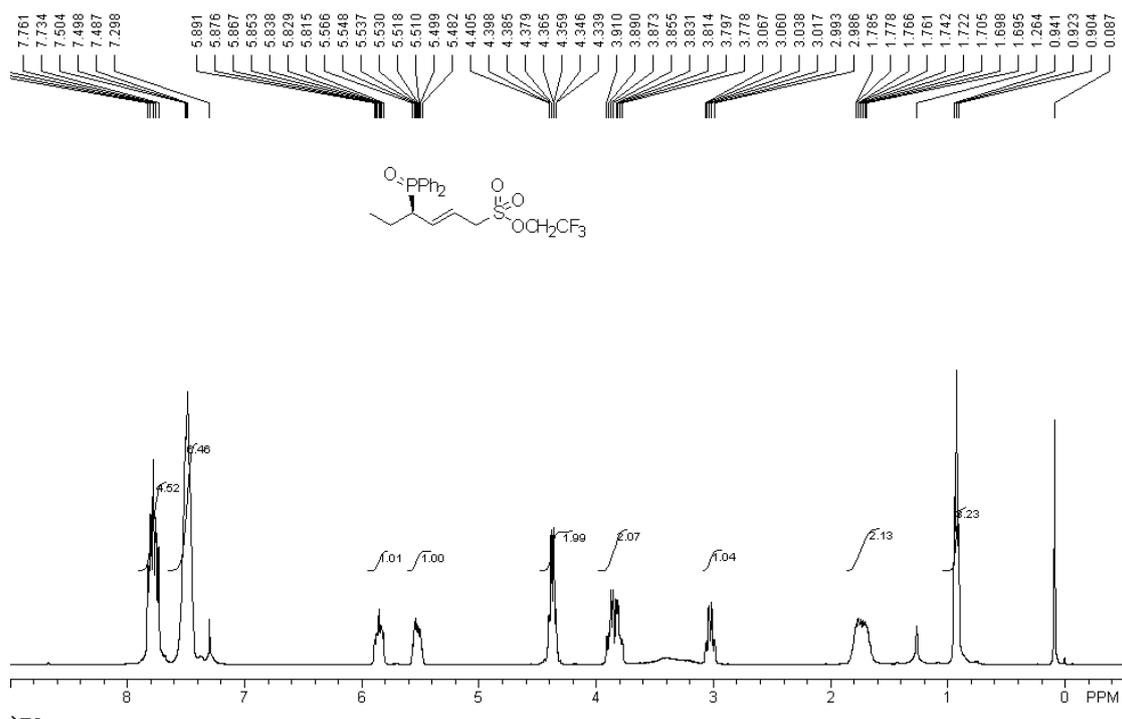


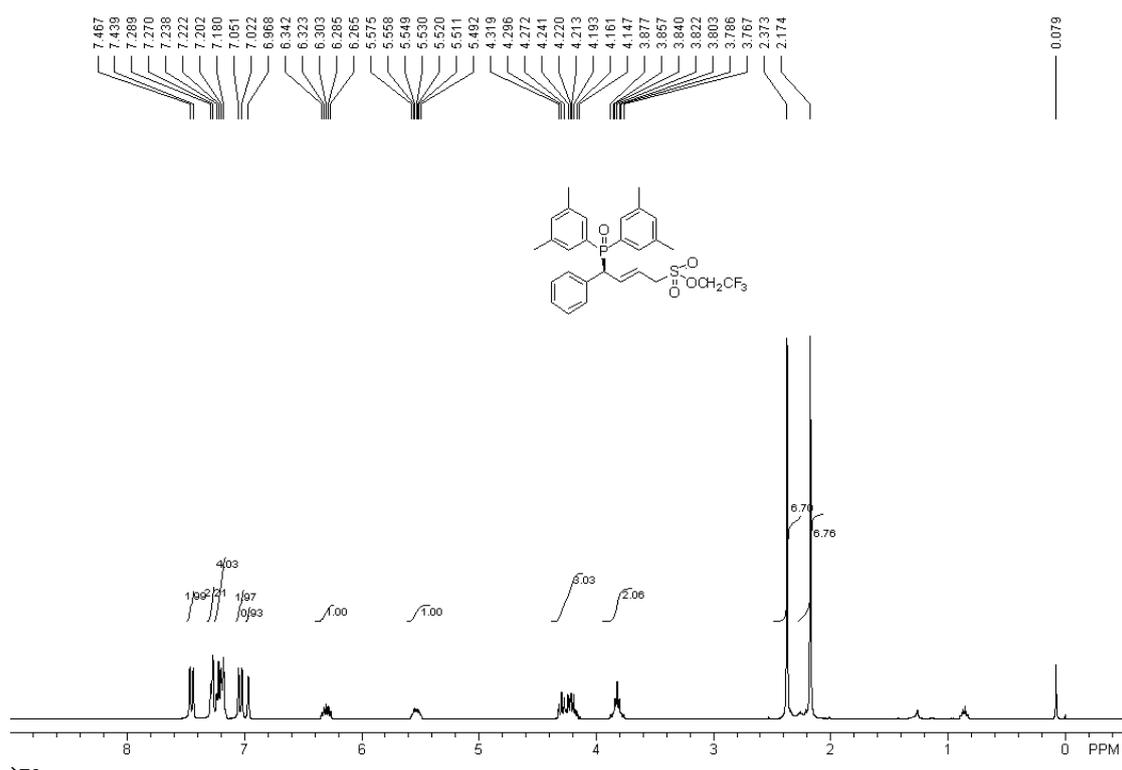
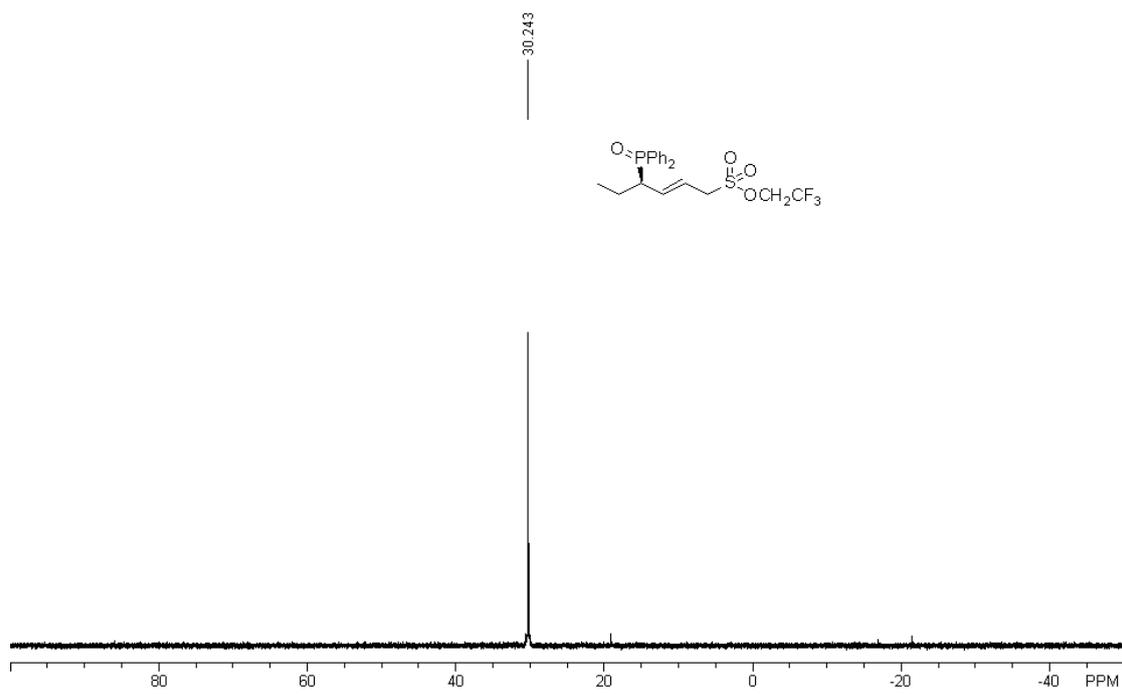


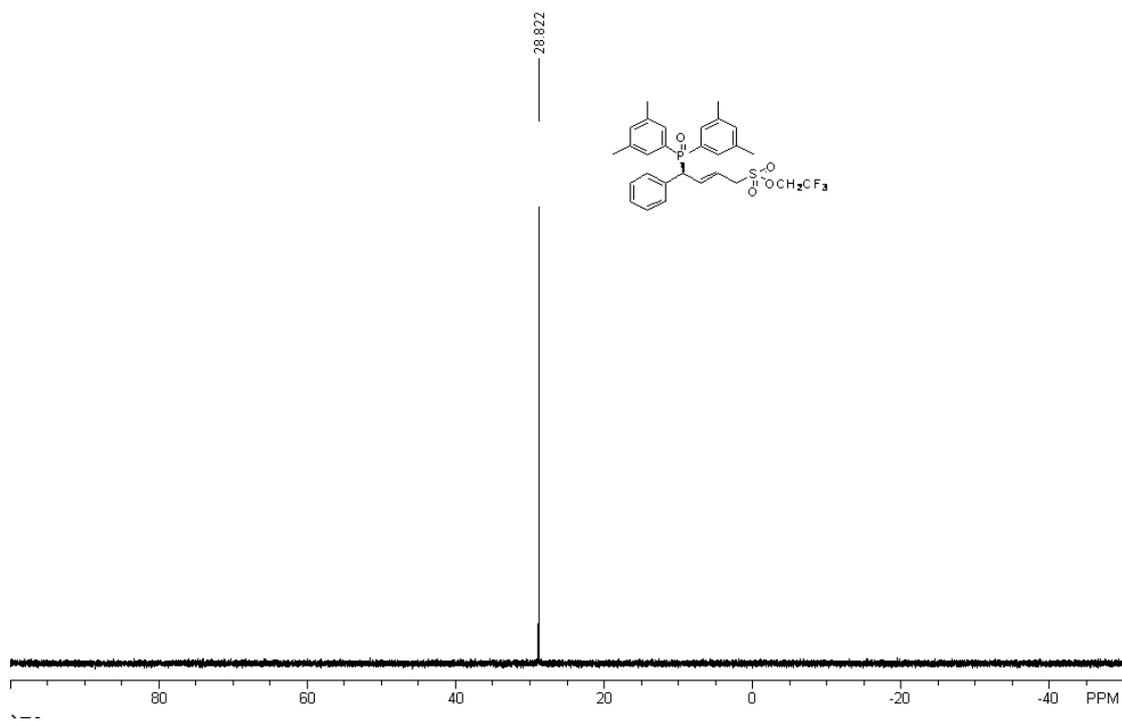
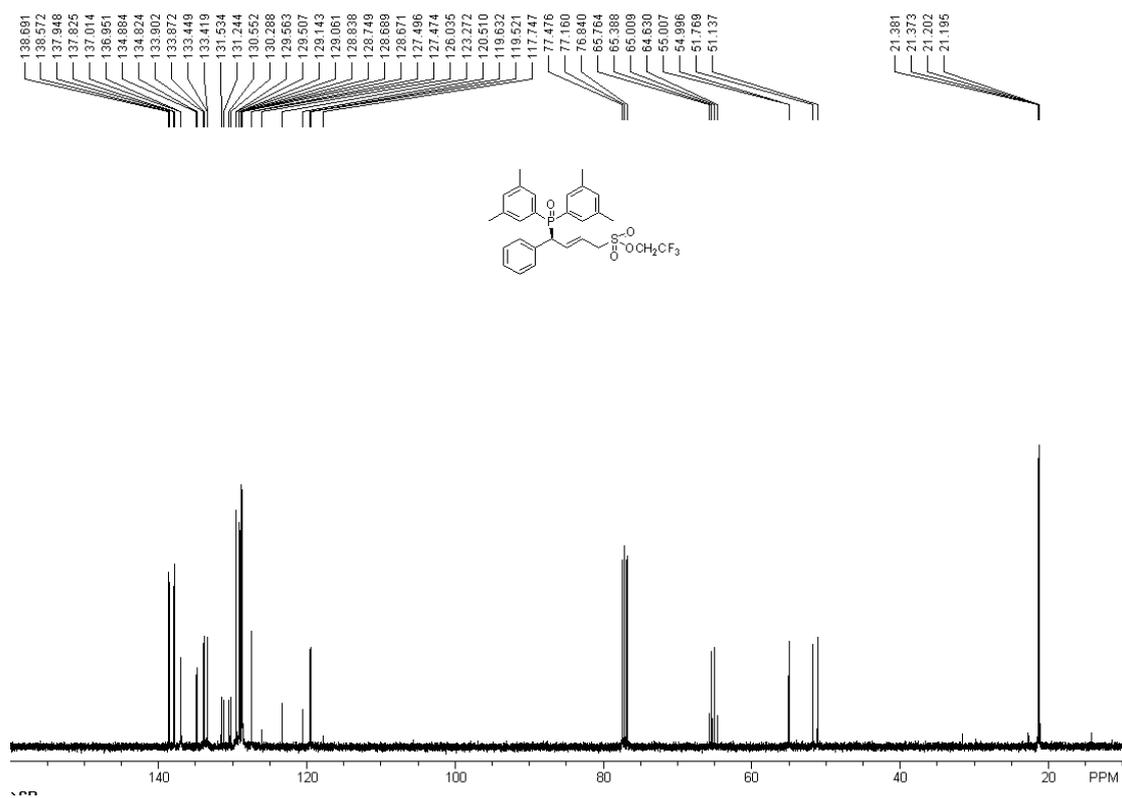


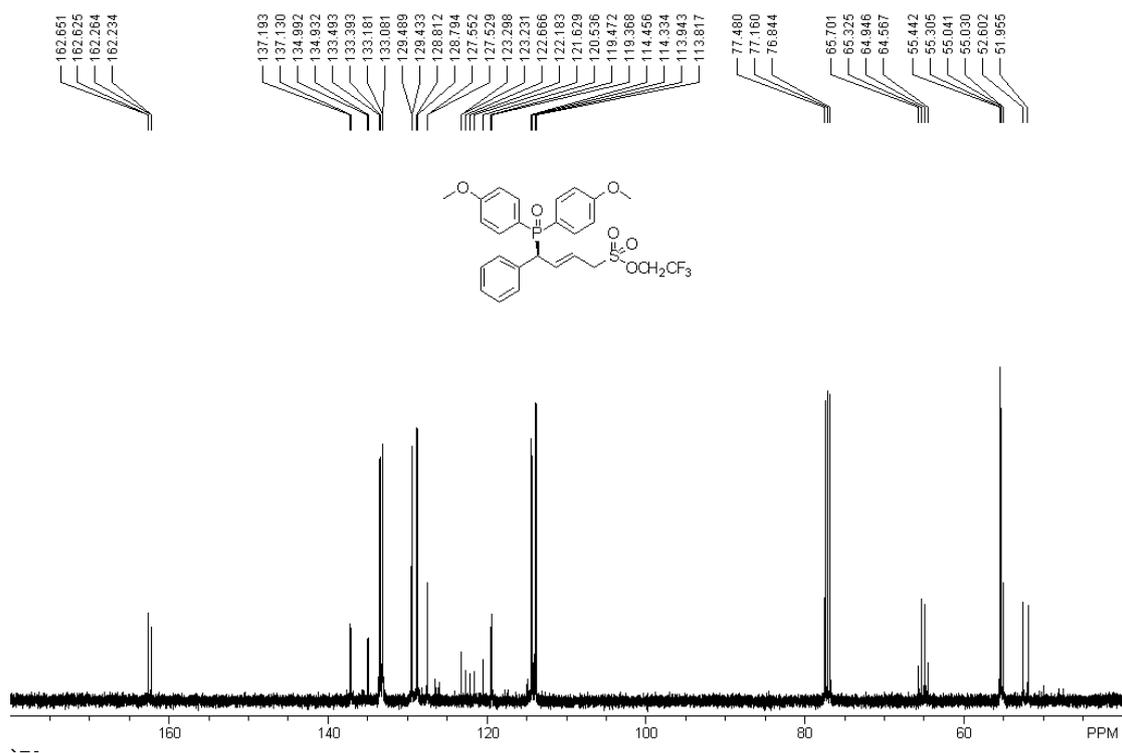
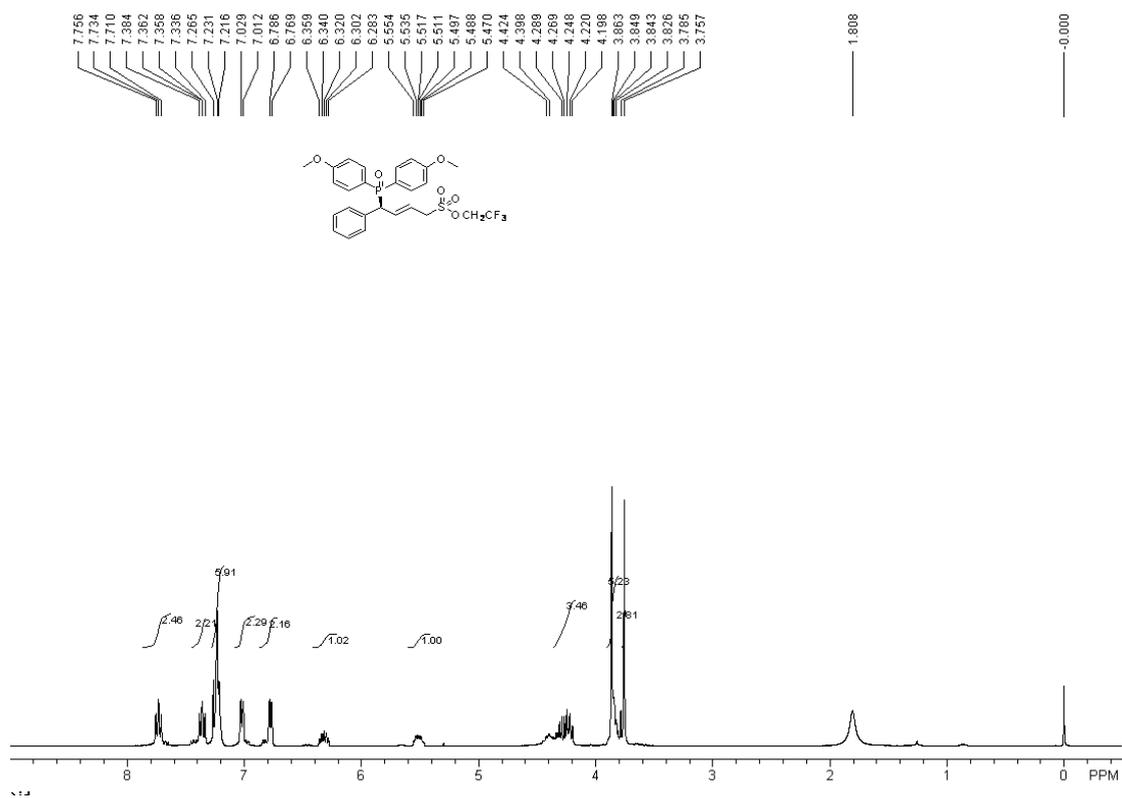


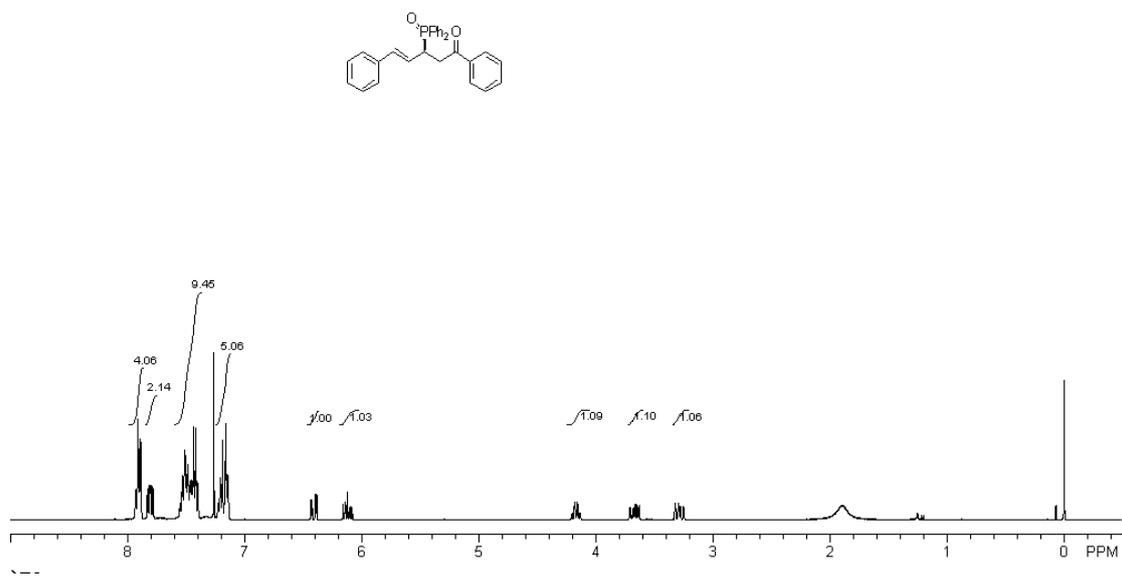
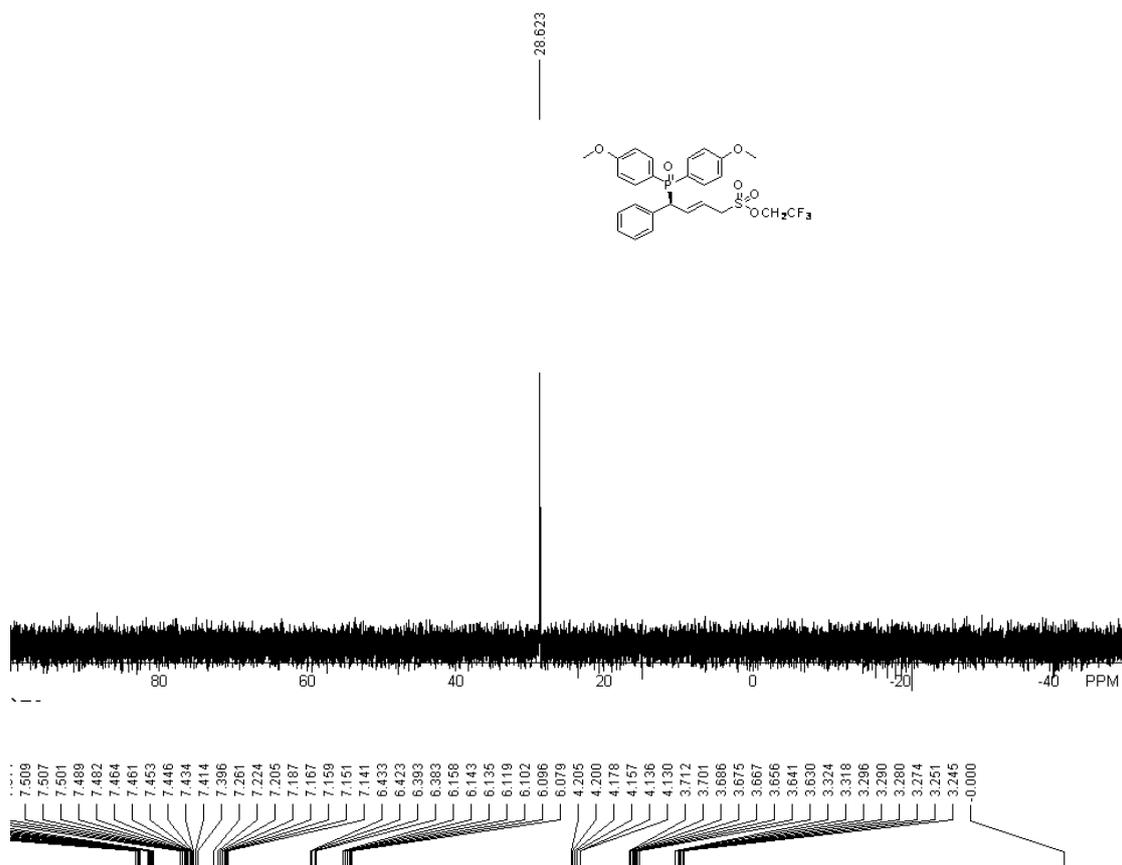


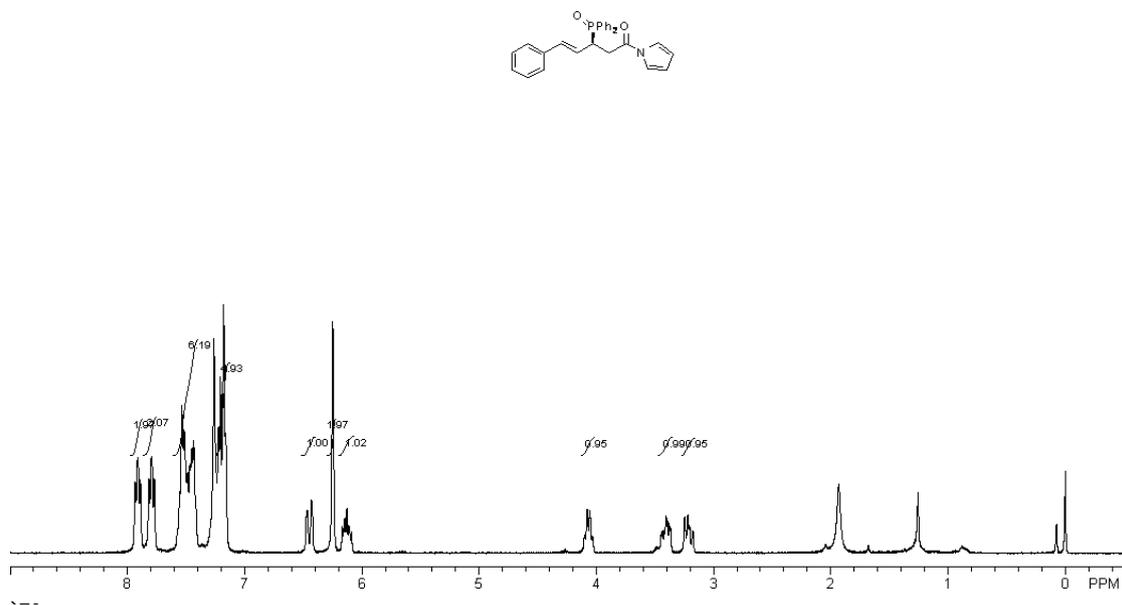
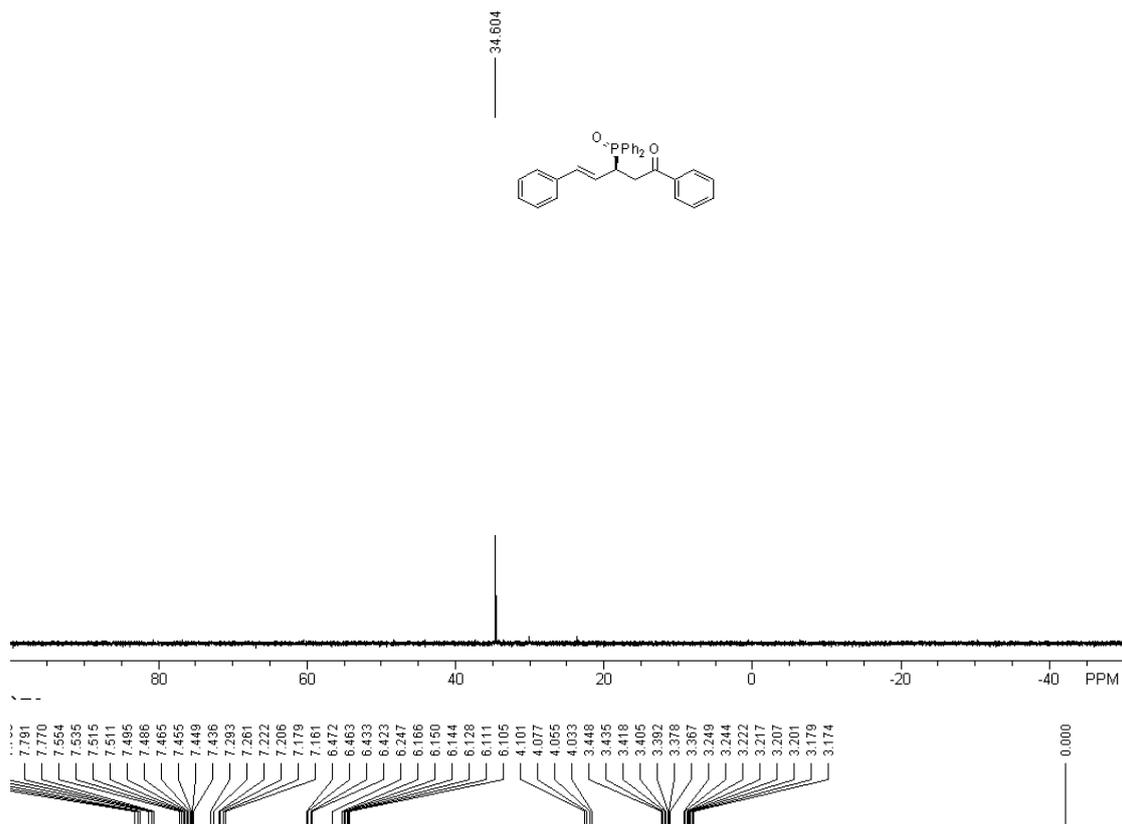


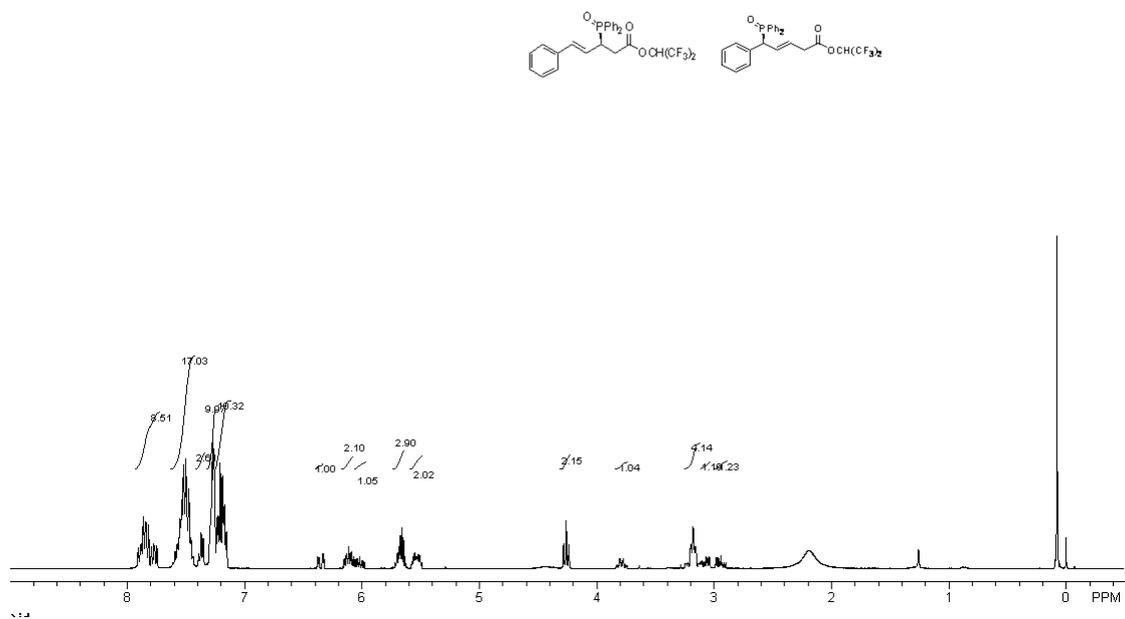
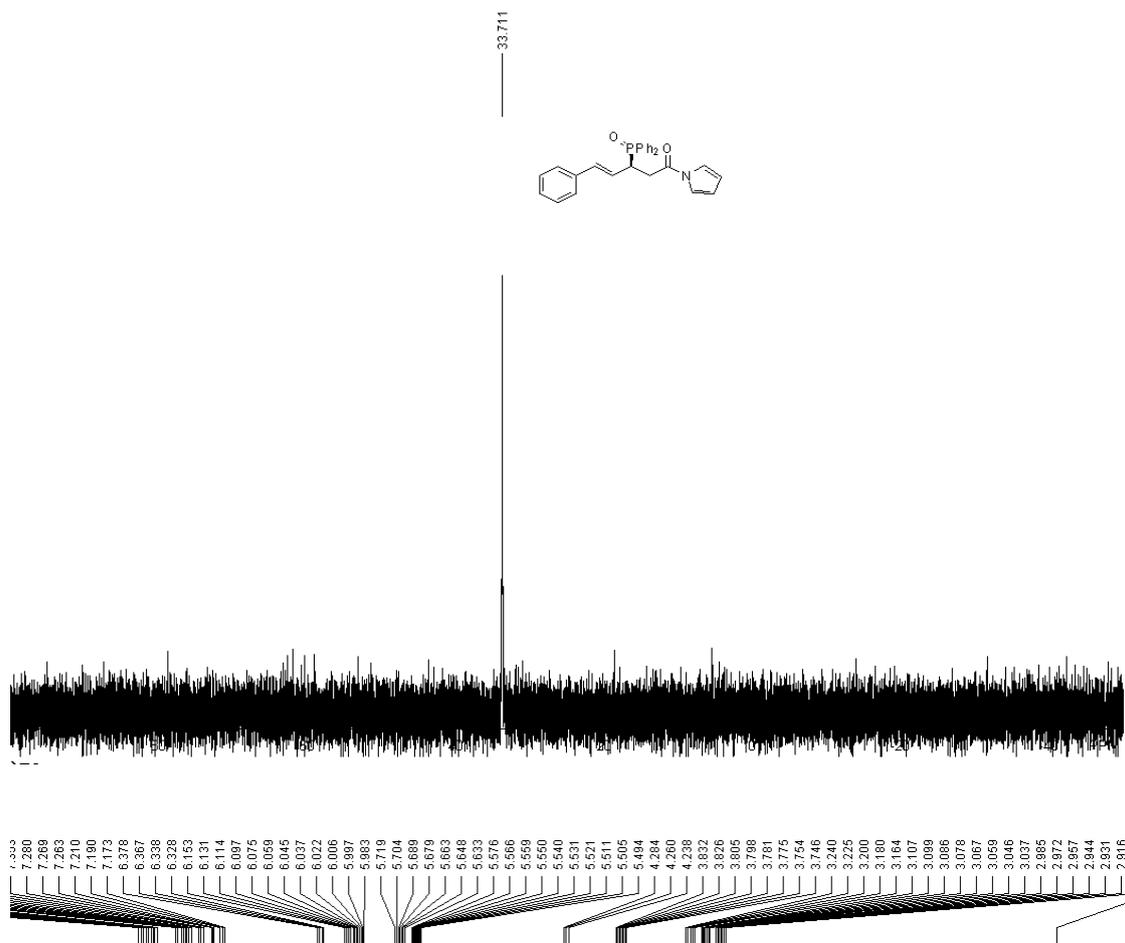


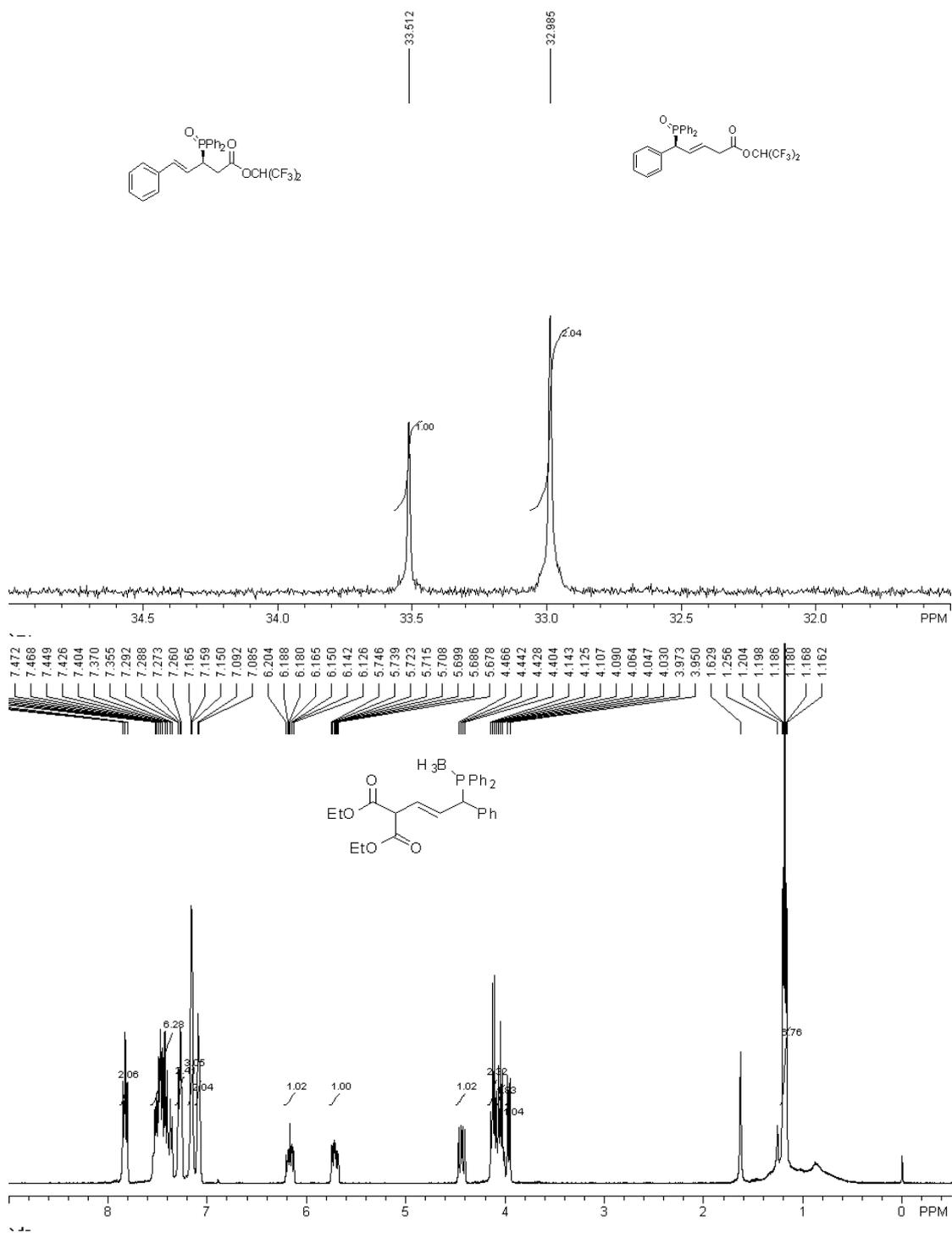


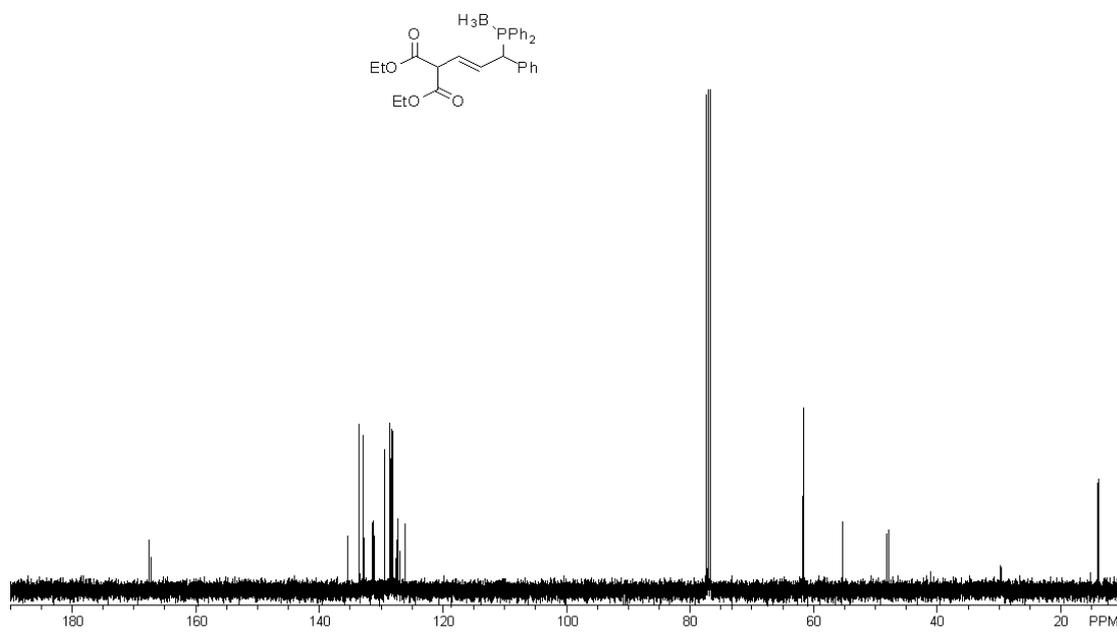
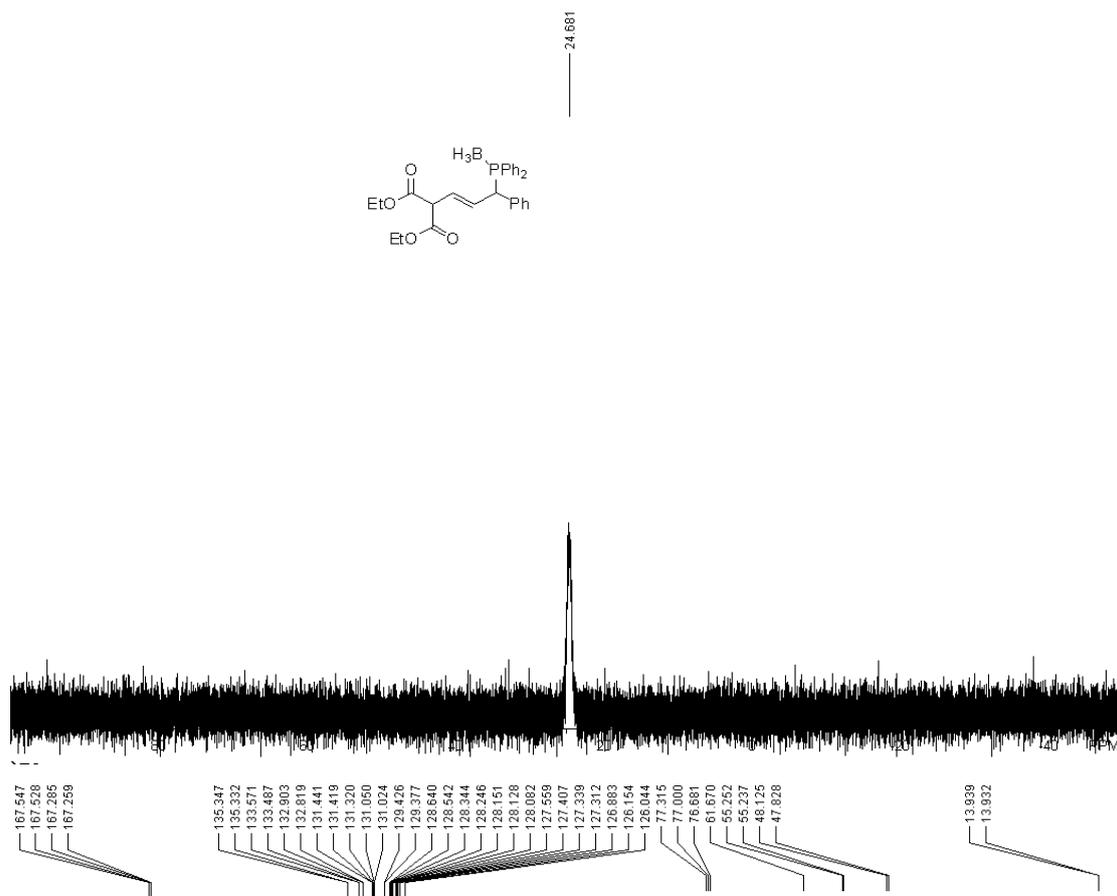


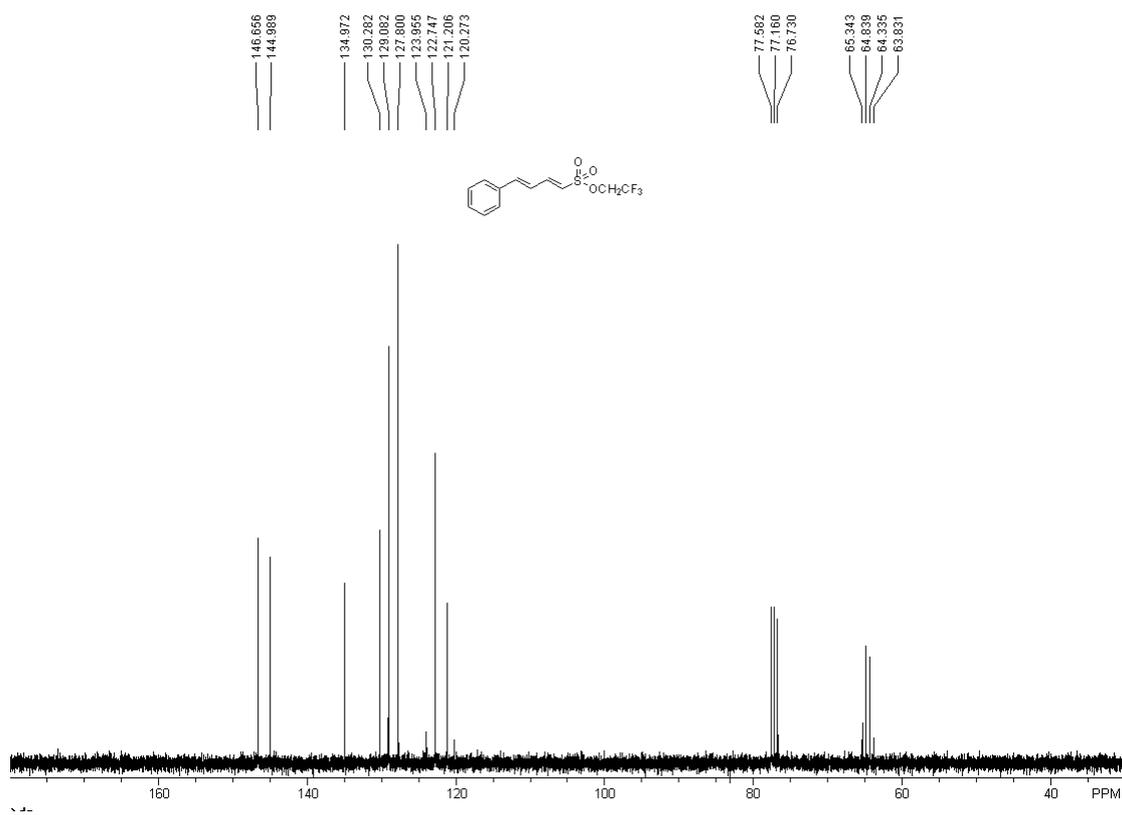
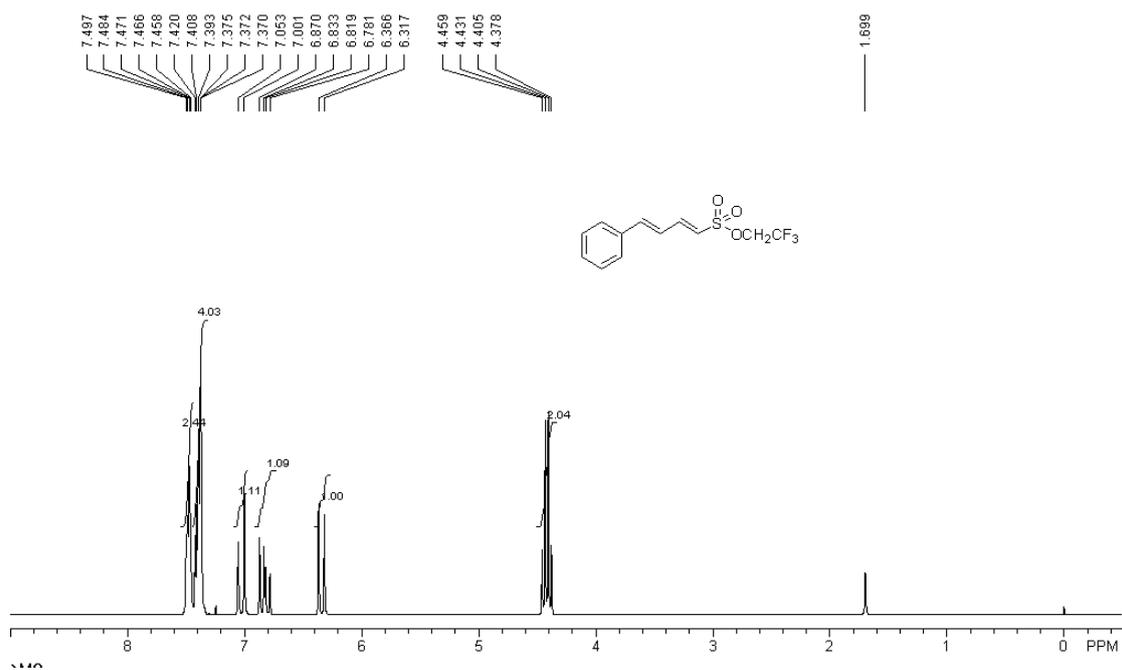


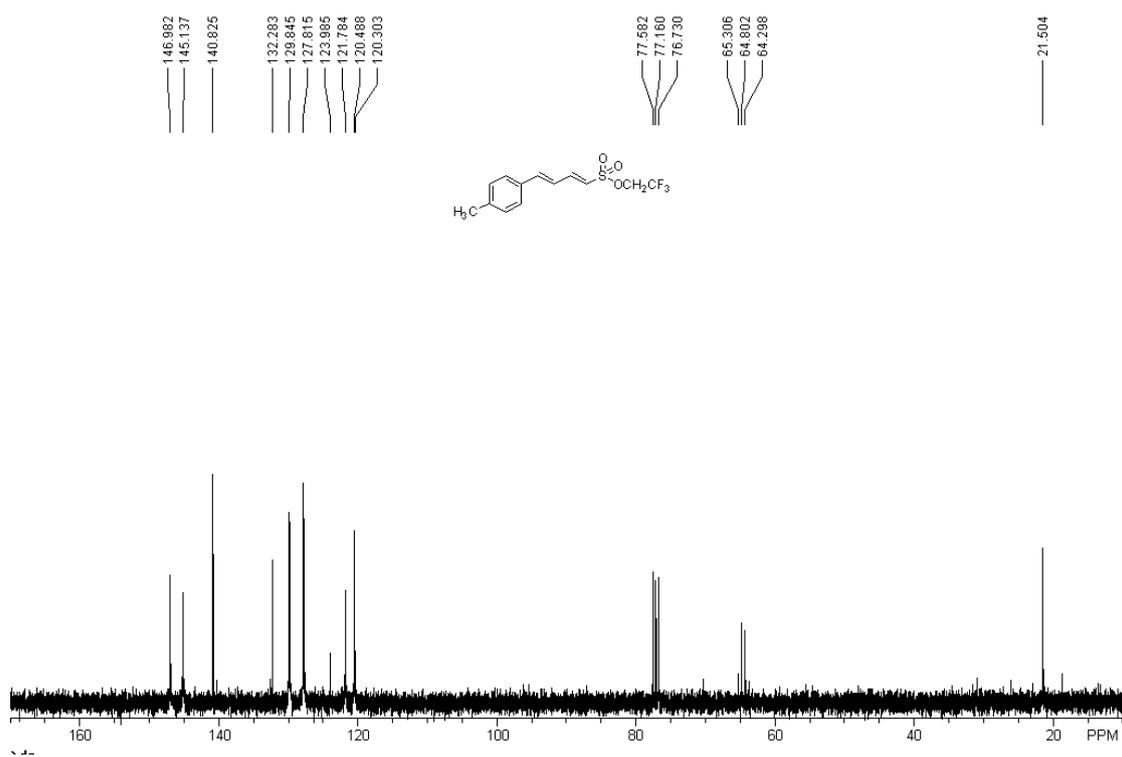
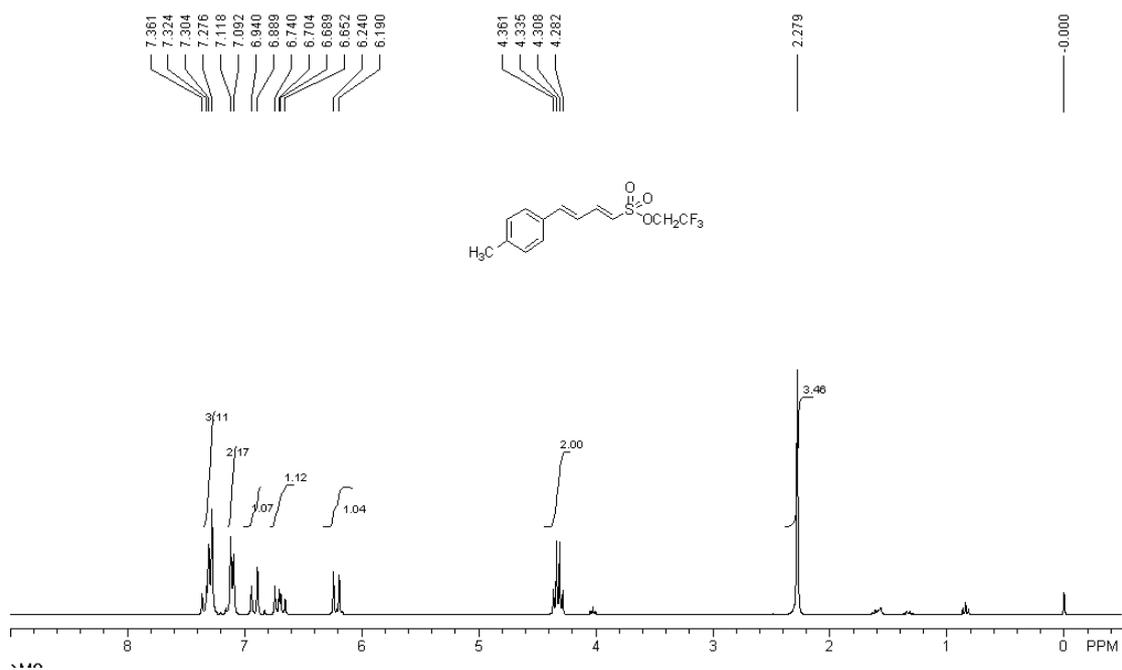


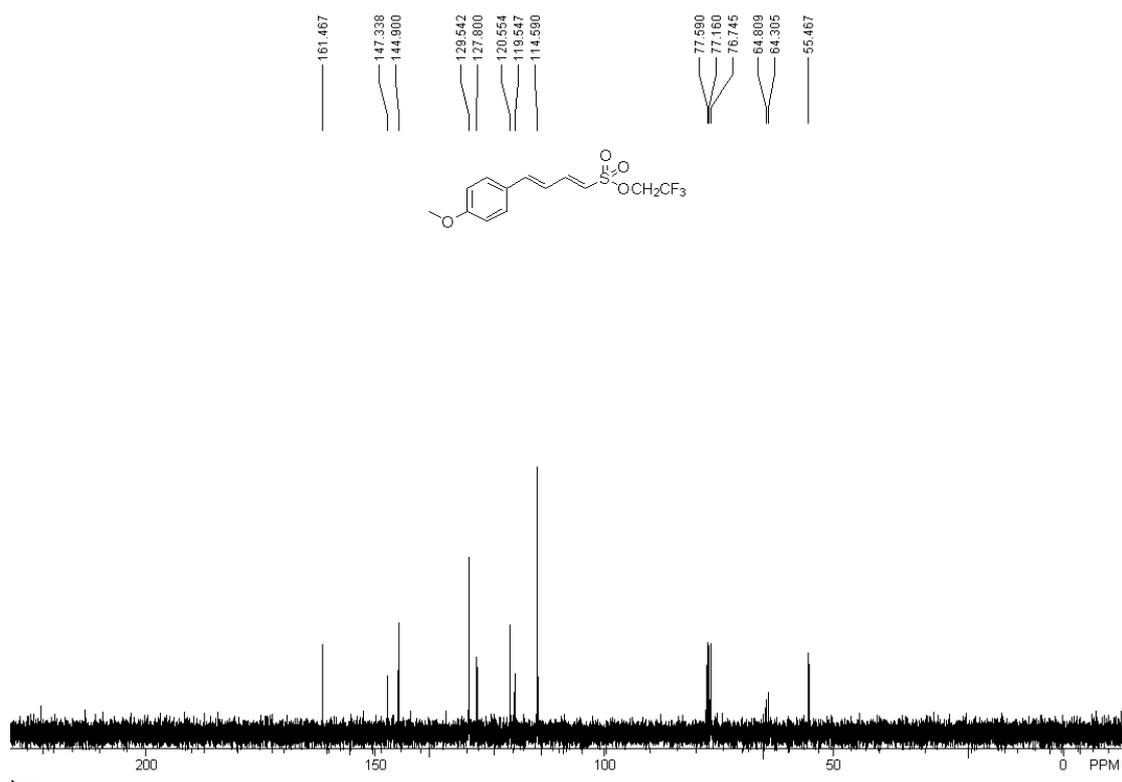
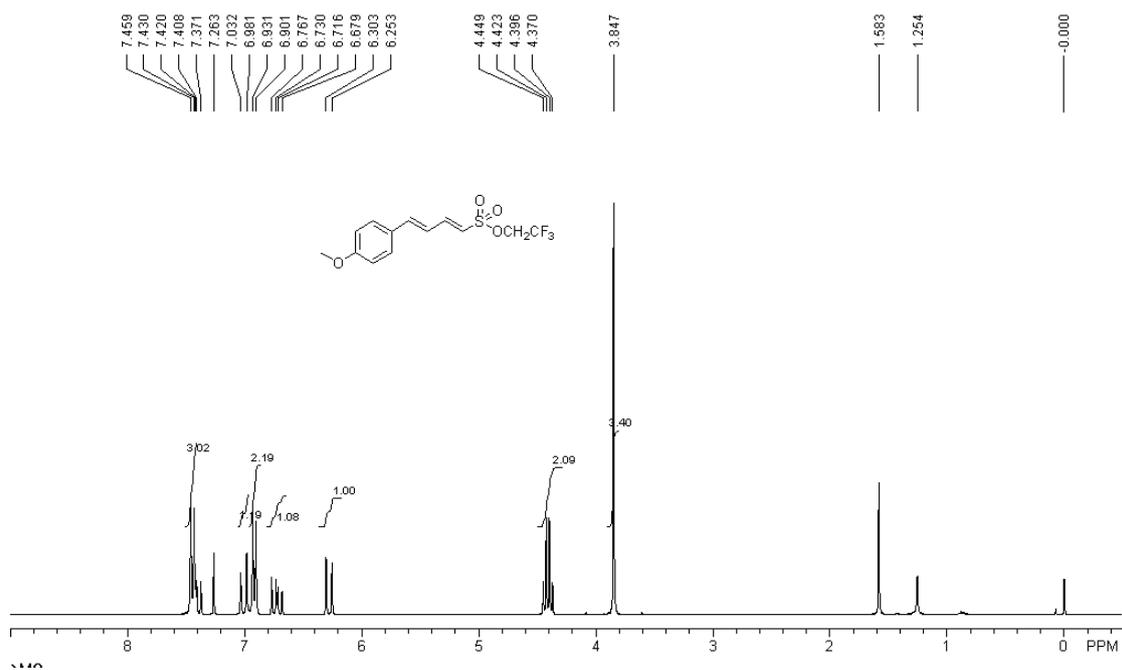


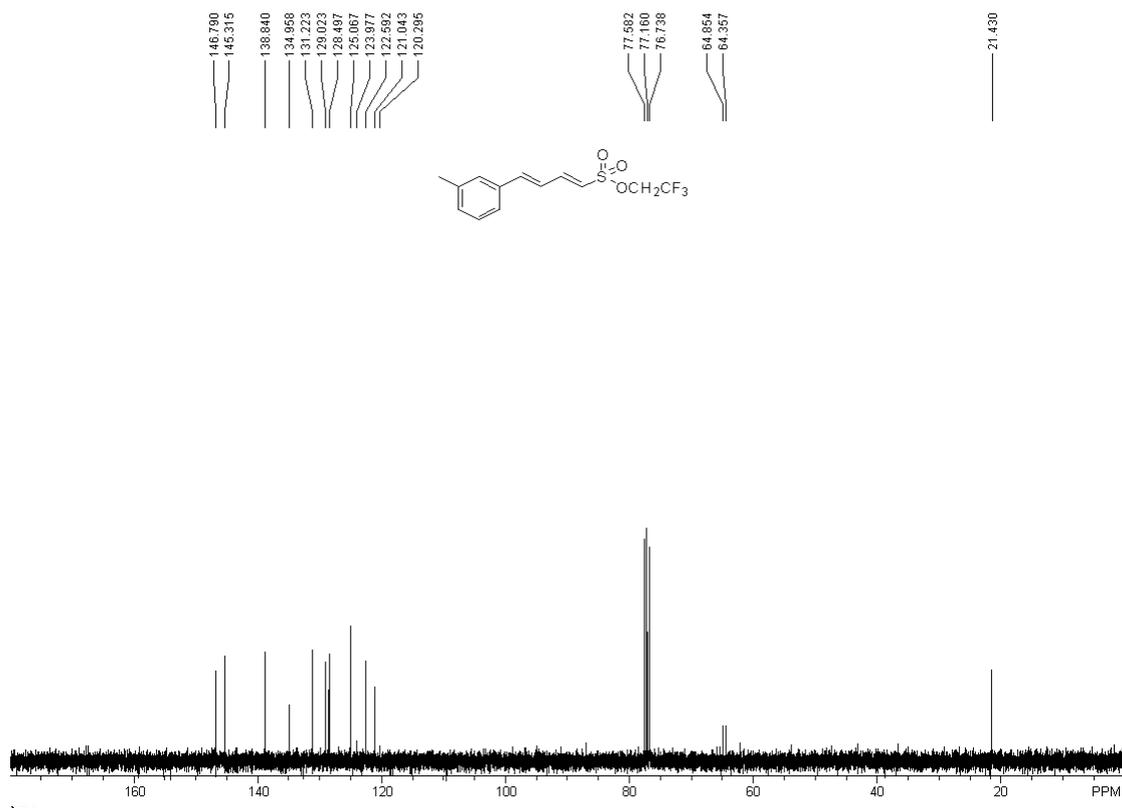
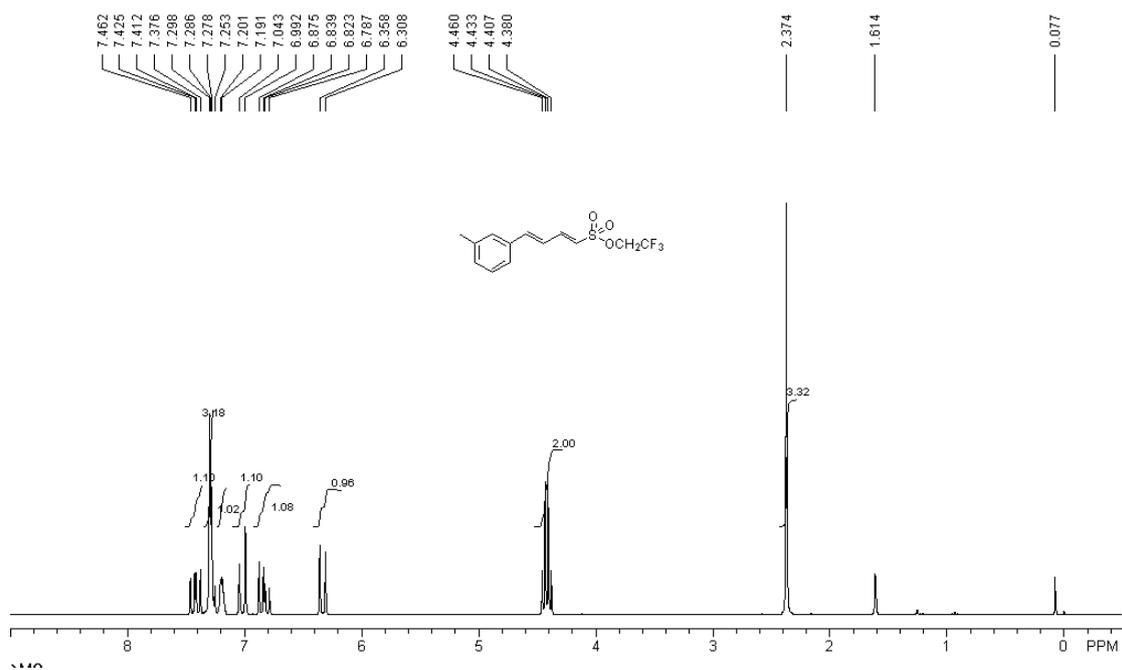


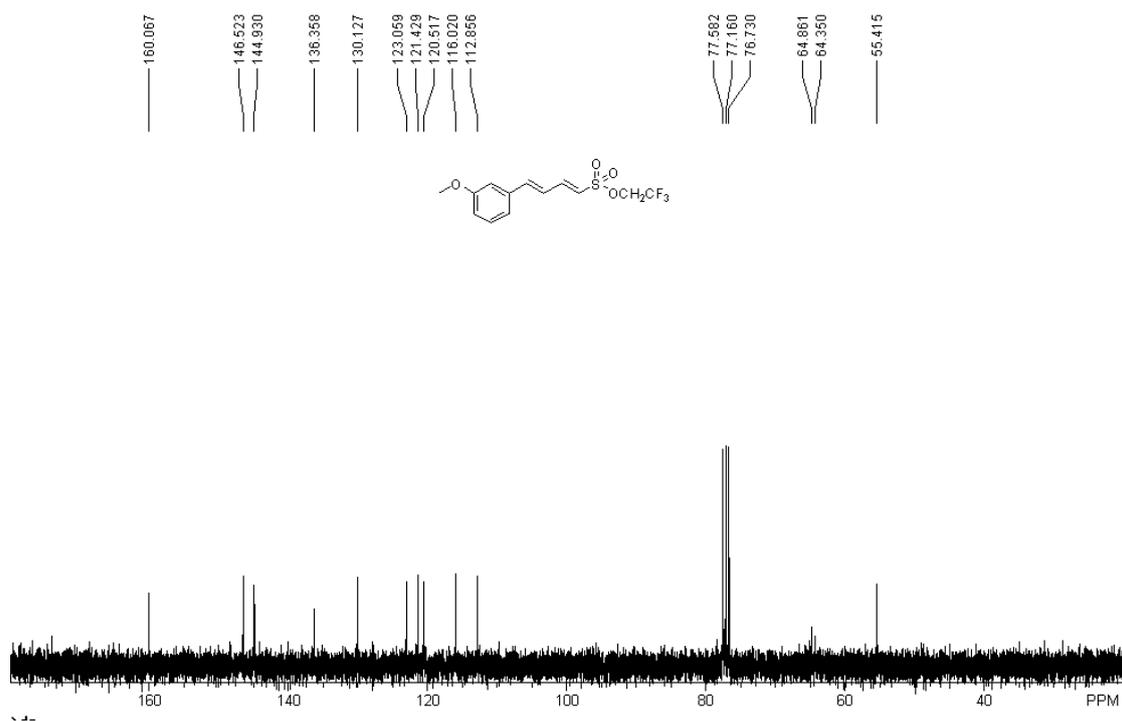
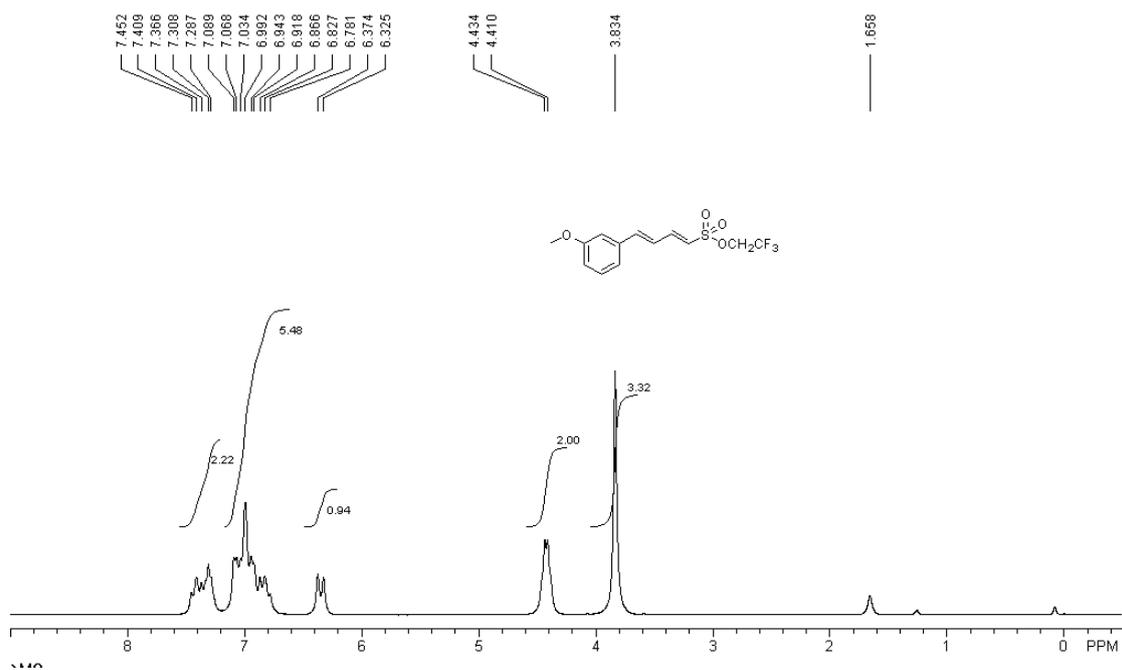


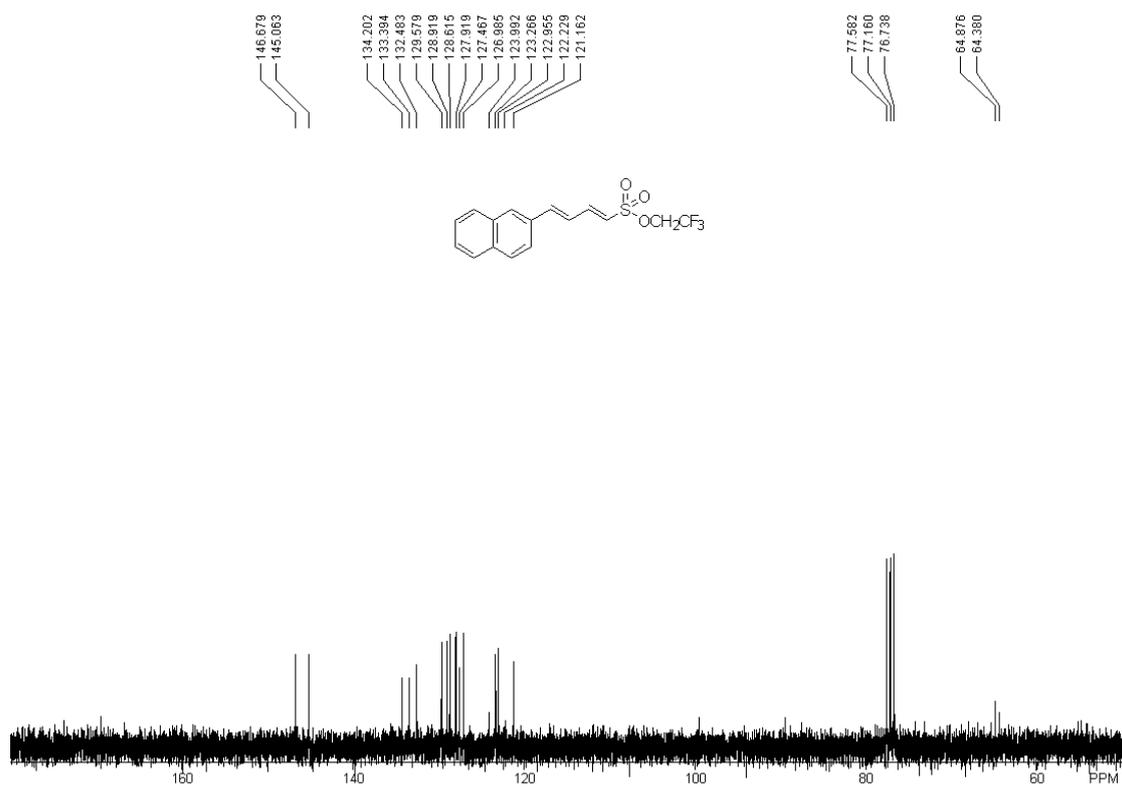
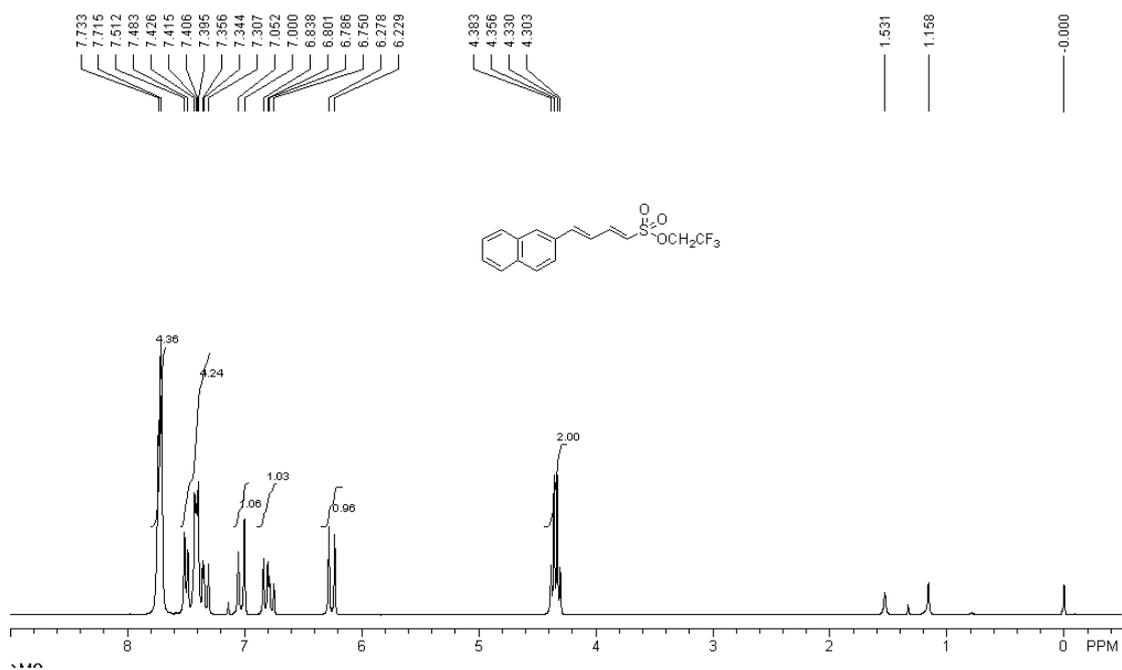


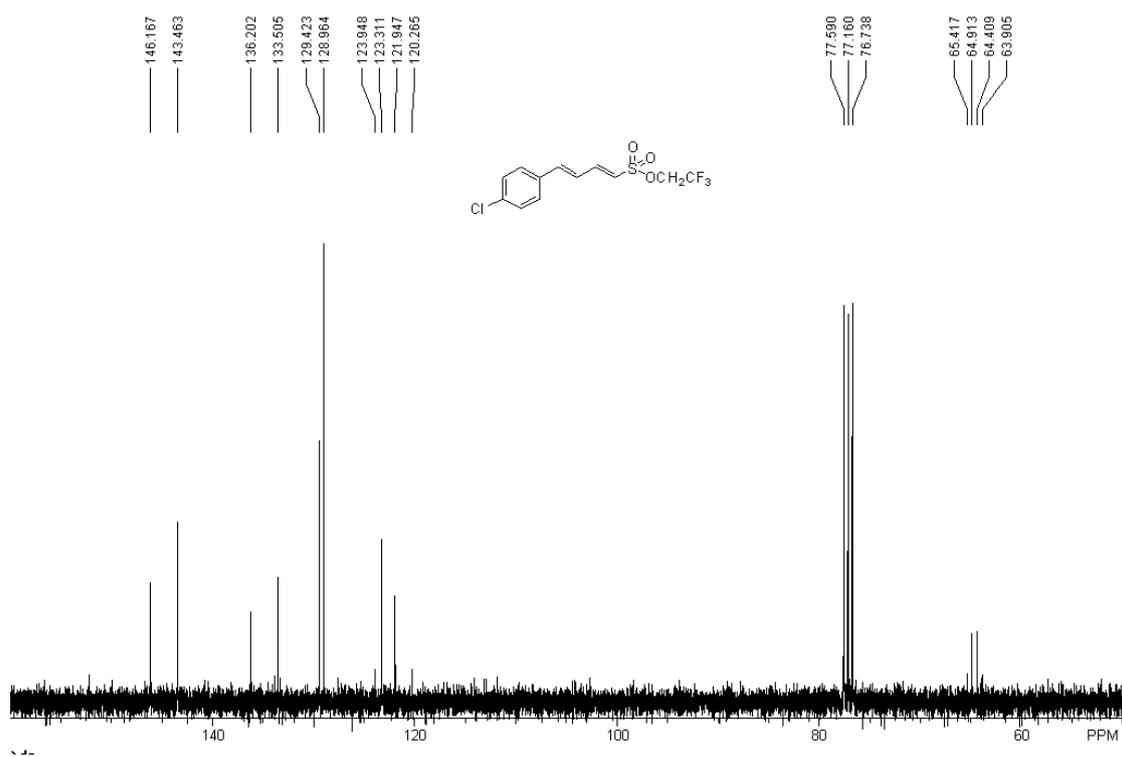
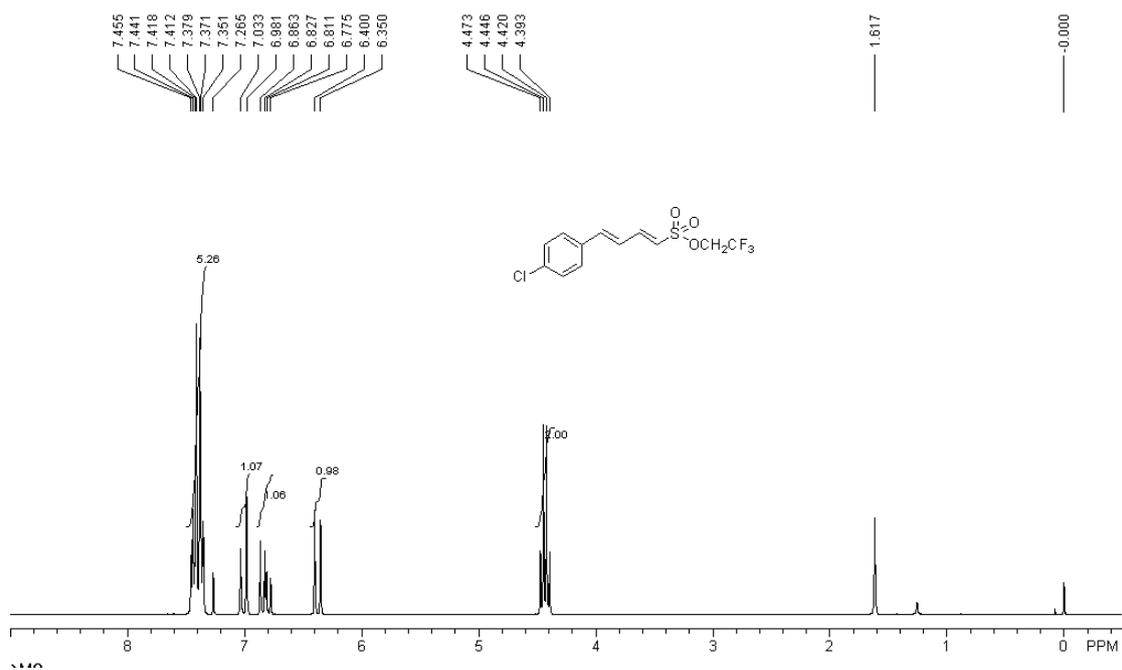


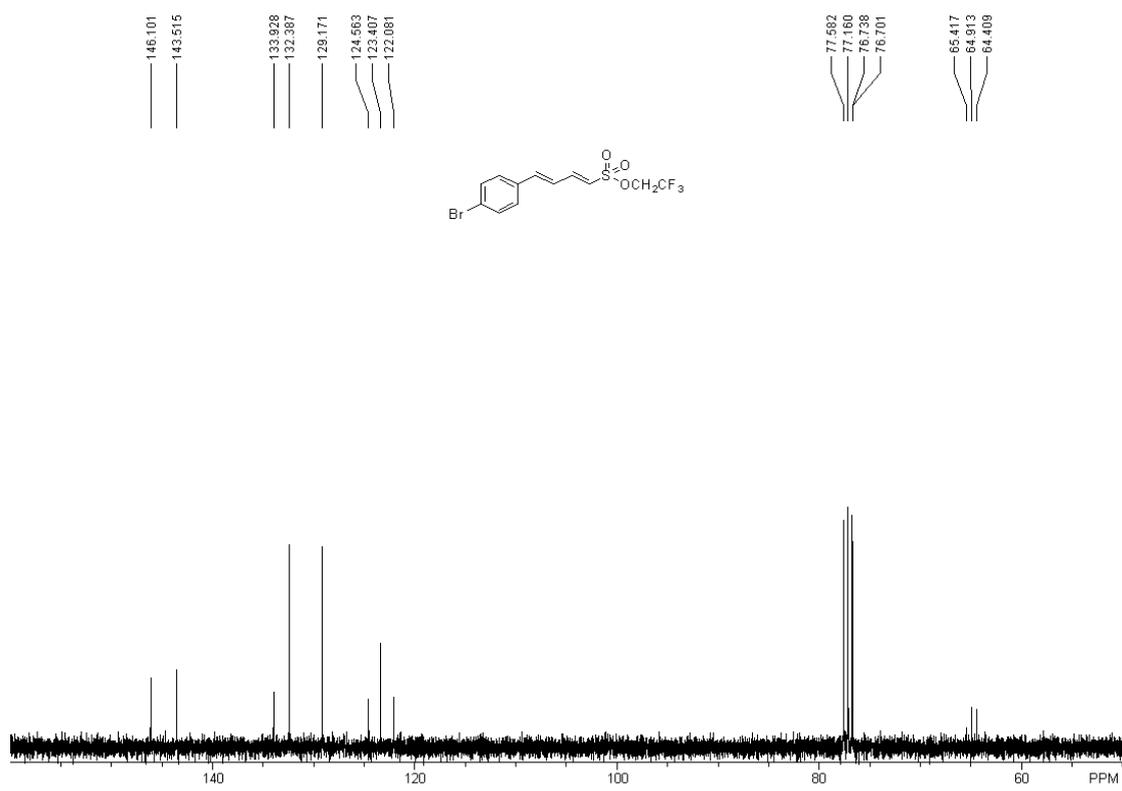
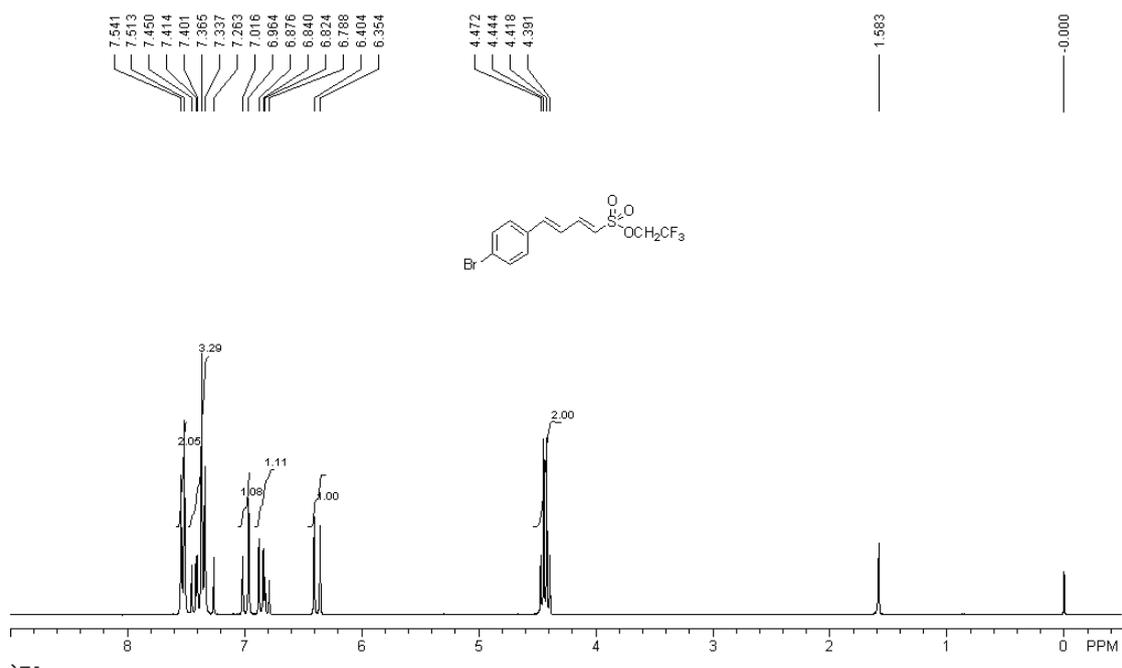


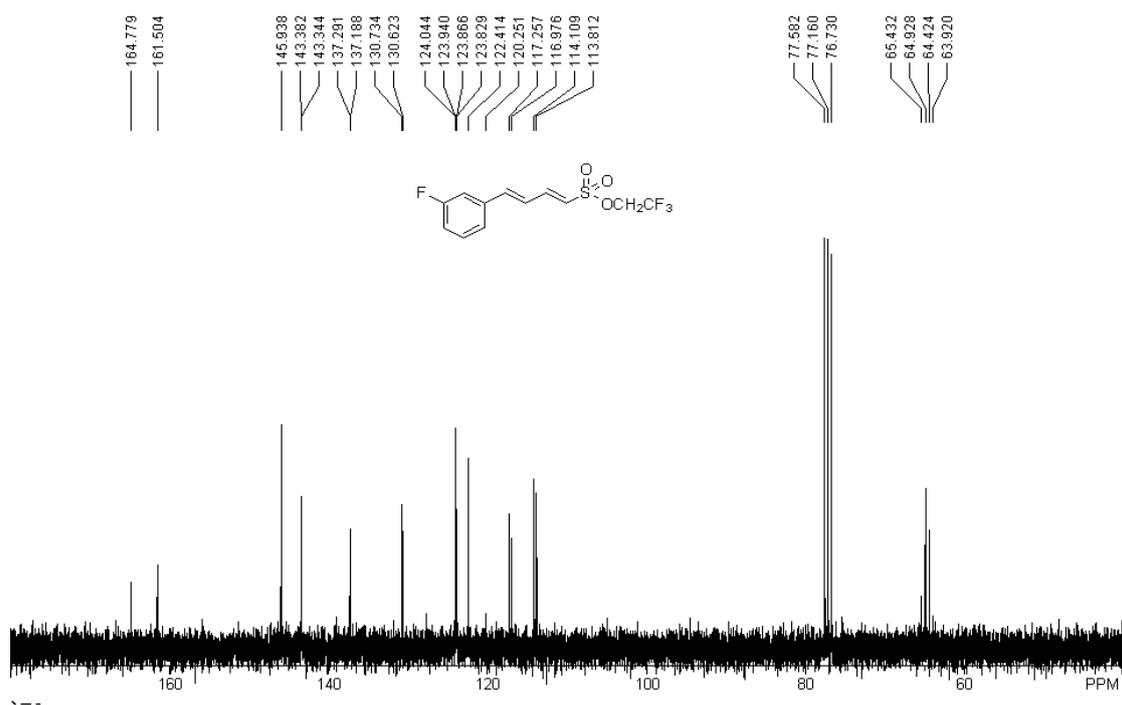
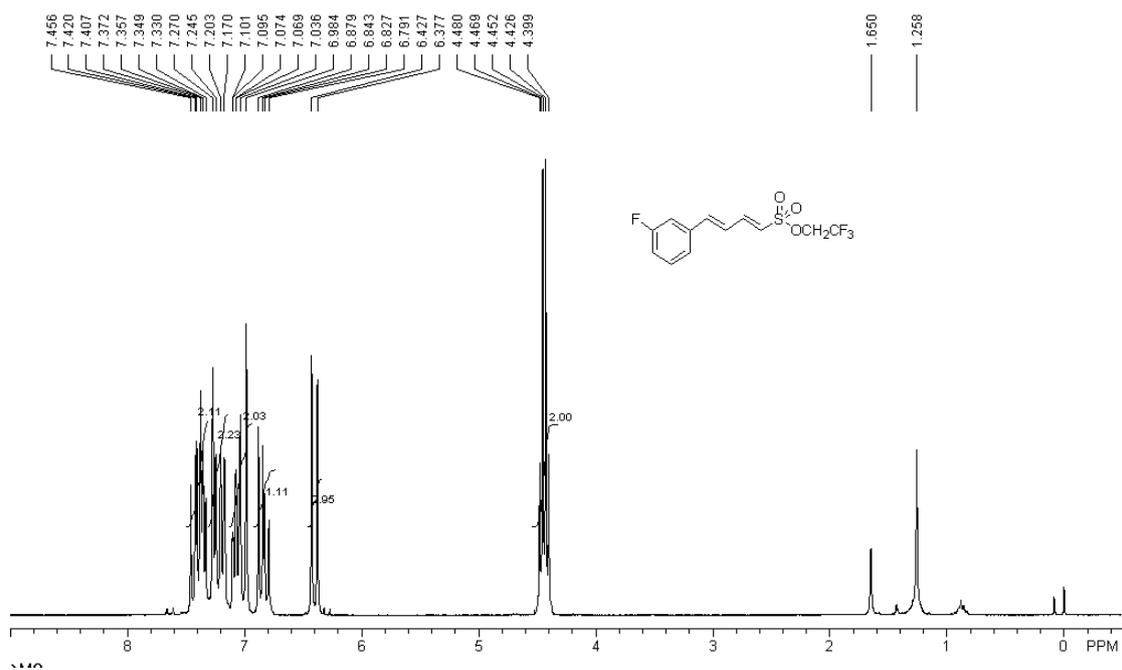


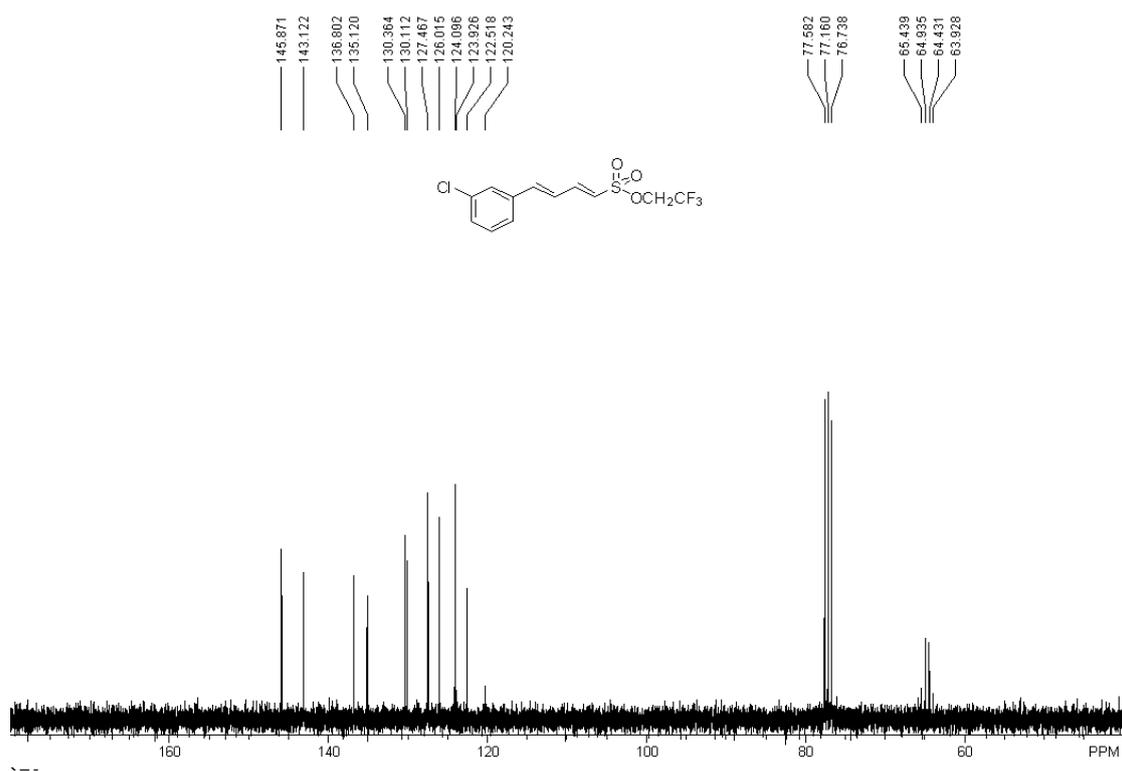
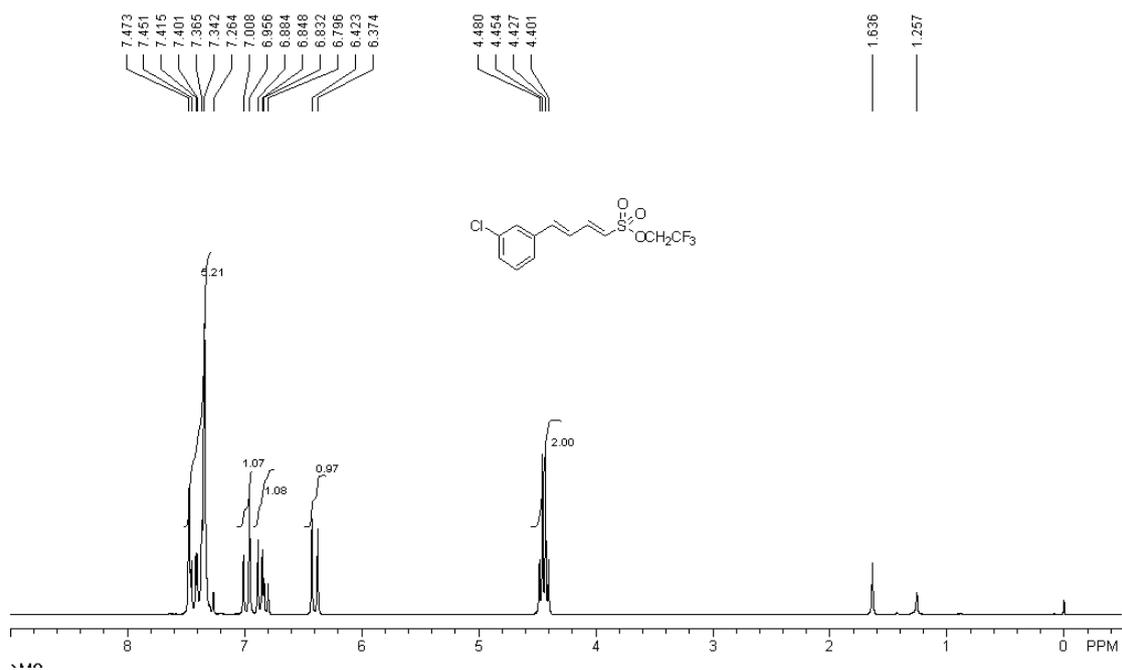


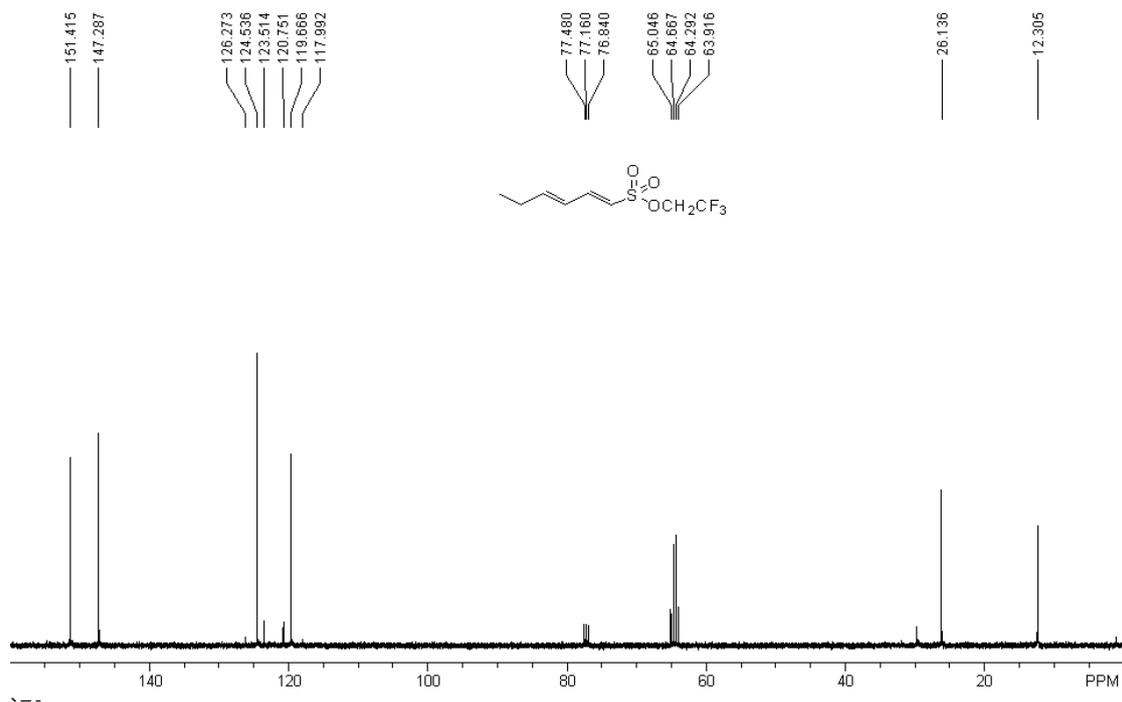
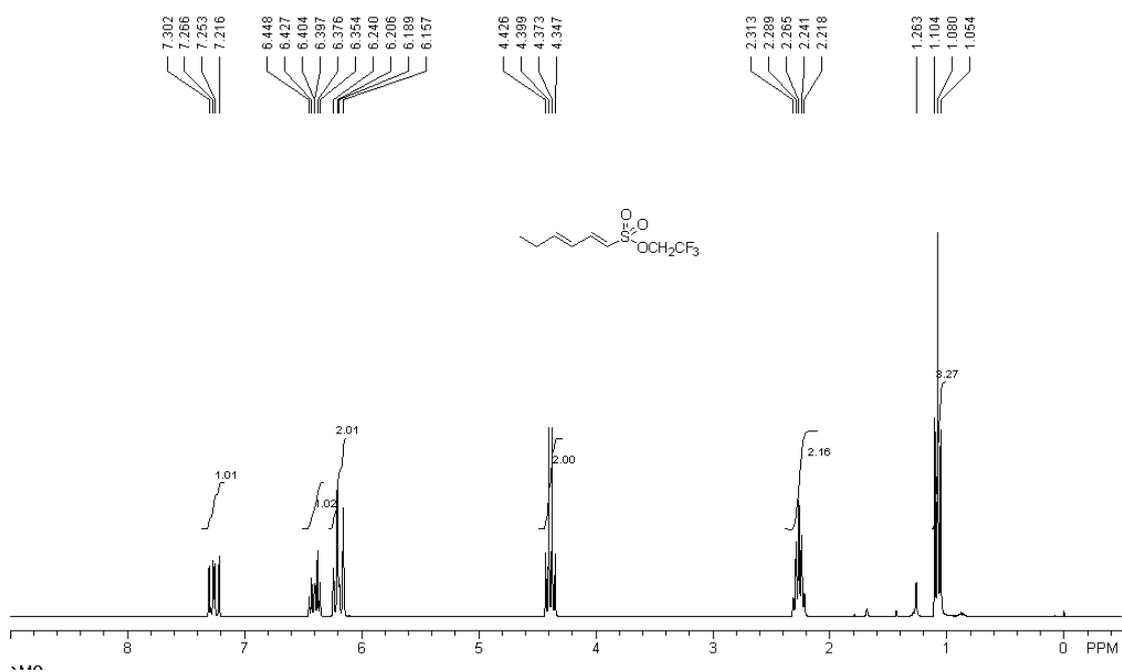


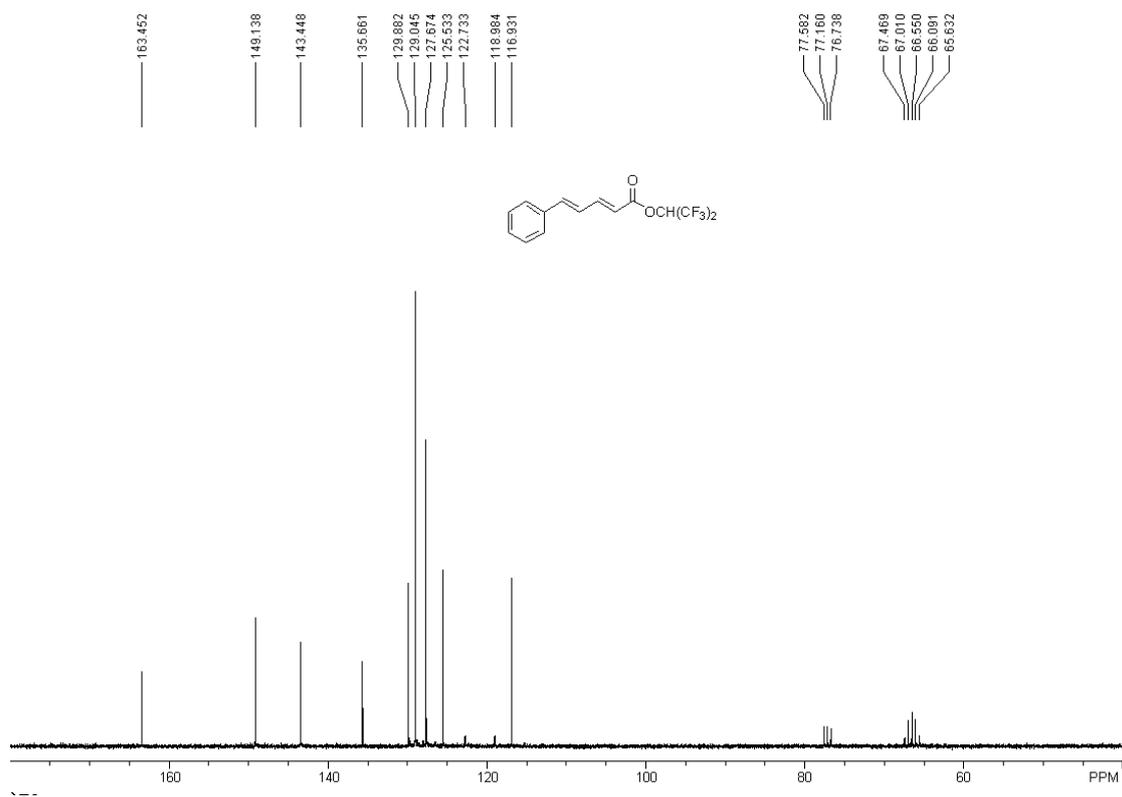
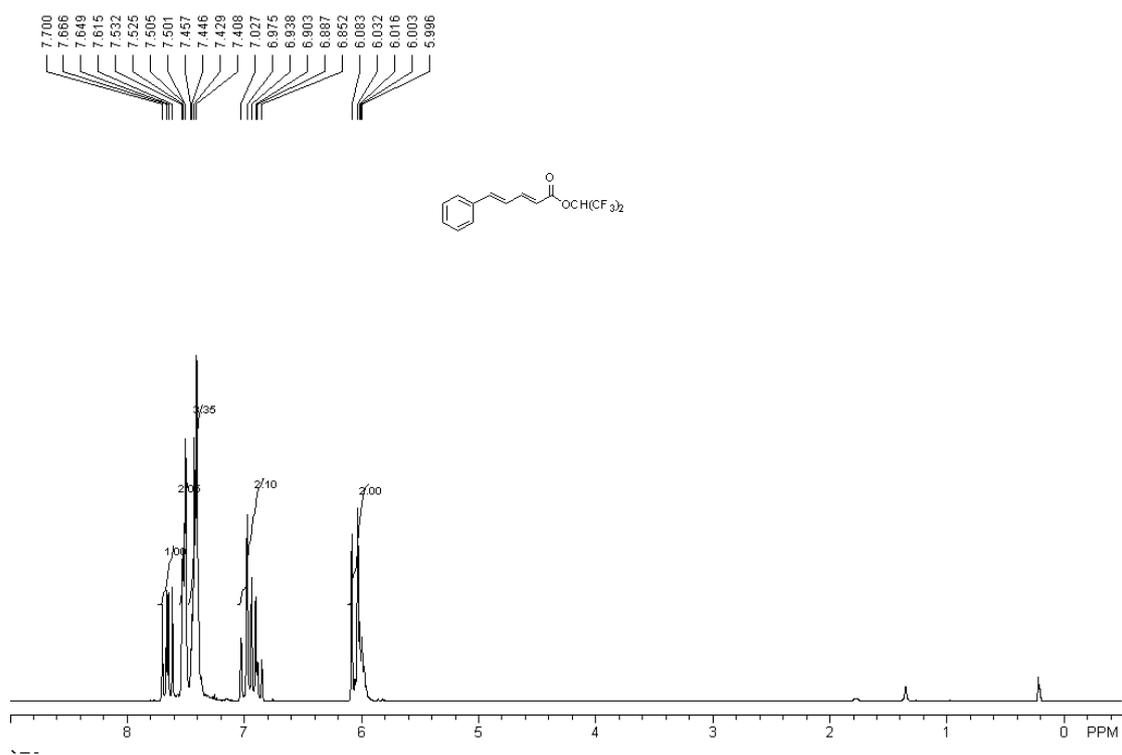






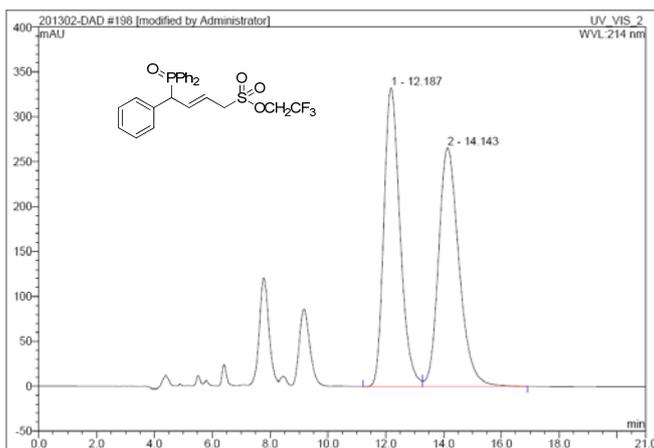






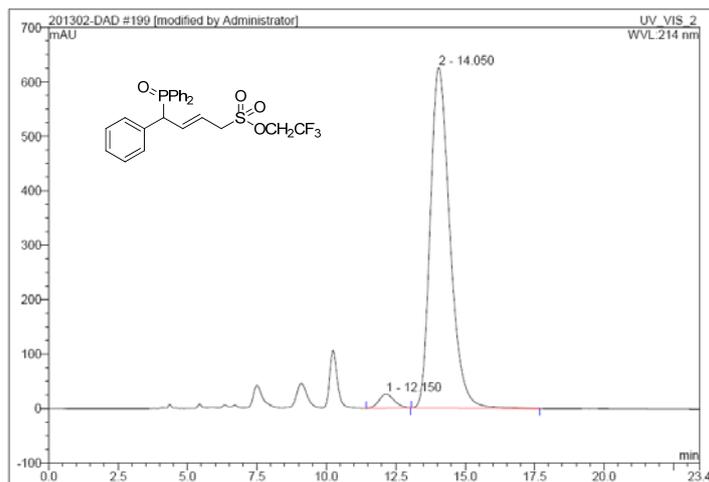
HPLC Chart

198 CJZ-5-6+- OD-H 73 214 0.7			
Sample Name:	CJZ-5-6+- OD-H 73 214 0.7	Injection Volume:	8.0
Vial Number:	RA2	Channel:	UV_VIS_2
Sample Type:	unknown	Wavelength:	214.0
Control Program:	test-dad	Bandwidth:	4
Quantif. Method:	WXL	Dilution Factor:	1.0000
Recording Time:	2013-3-25 10:13	Sample Weight:	1.0000
Run Time (min):	21.00	Sample Amount:	1.0000



No.	Ret.Time min	Peak Name	Height mAU	Area mAU*min	Rel.Area %	Amount	Type
1	12.19	n.a.	333.231	216.146	49.71	n.a.	BM
2	14.14	n.a.	265.955	218.711	50.29	n.a.	MB
Total:			599.185	434.856	100.00	0.000	

199 CJZ-6-9-2 OD-H 73 214 0.7			
Sample Name:	CJZ-6-9-2 OD-H 73 214 0.7	Injection Volume:	4.0
Vial Number:	RA1	Channel:	UV_VIS_2
Sample Type:	unknown	Wavelength:	214.0
Control Program:	test-dad	Bandwidth:	4
Quantif. Method:	WXL	Dilution Factor:	1.0000
Recording Time:	2013-3-25 10:36	Sample Weight:	1.0000
Run Time (min):	23.44	Sample Amount:	1.0000



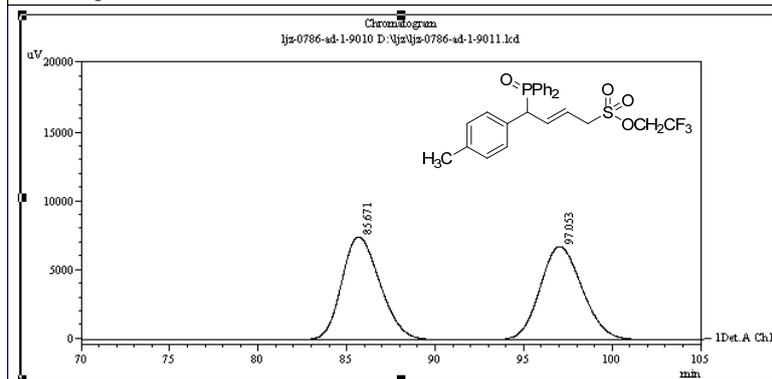
No.	Ret.Time min	Peak Name	Height mAU	Area mAU*min	Rel.Area %	Amount	Type
1	12.15	n.a.	25.745	16.234	3.08	n.a.	BMB*
2	14.05	n.a.	625.605	510.931	96.92	n.a.	BMB*
Total:			651.350	527.165	100.00	0.000	

==== Shimadzu LCsolution Analysis Report ====

D:\Vjz\ljjz-0786-ad-1-9011.lcd

Sample Name : ljjz-0786-ad-1-9010 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

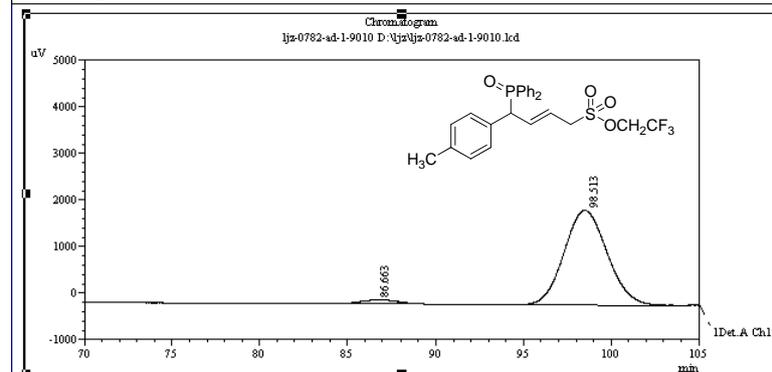
Peak#	Ret. Time	Area	Height	Area %
1	85.671	1121298	7476	49.948
2	97.053	1123618	6766	50.052
Total		2244915	14241	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\Vjz\ljjz-0782-ad-1-9010.lcd

Sample Name : ljjz-0782-ad-1-9010 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

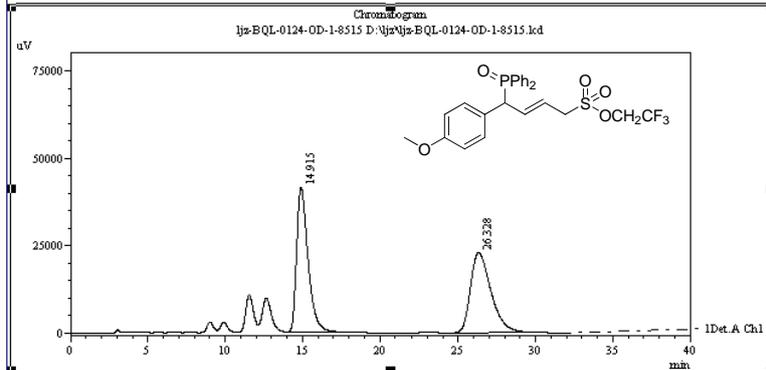
Peak#	Ret. Time	Area	Height	Area %
1	86.663	11584	89	3.217
2	98.513	348468	2036	96.783
Total		360051	2125	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\jz\jz-BQL-0124-OD-1-8515.lcd

Sample Name : ljz-BQL-0124-OD-1-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

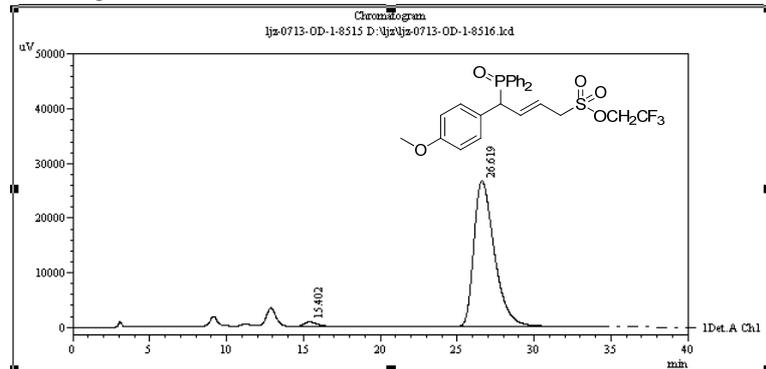
Peak#	Ret. Time	Area	Height	Area %
1	14.915	2116386	41234	50.126
2	26.328	2105977	22883	49.874
Total		4222363	64116	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\jz\jz-0713-OD-1-8516.lcd

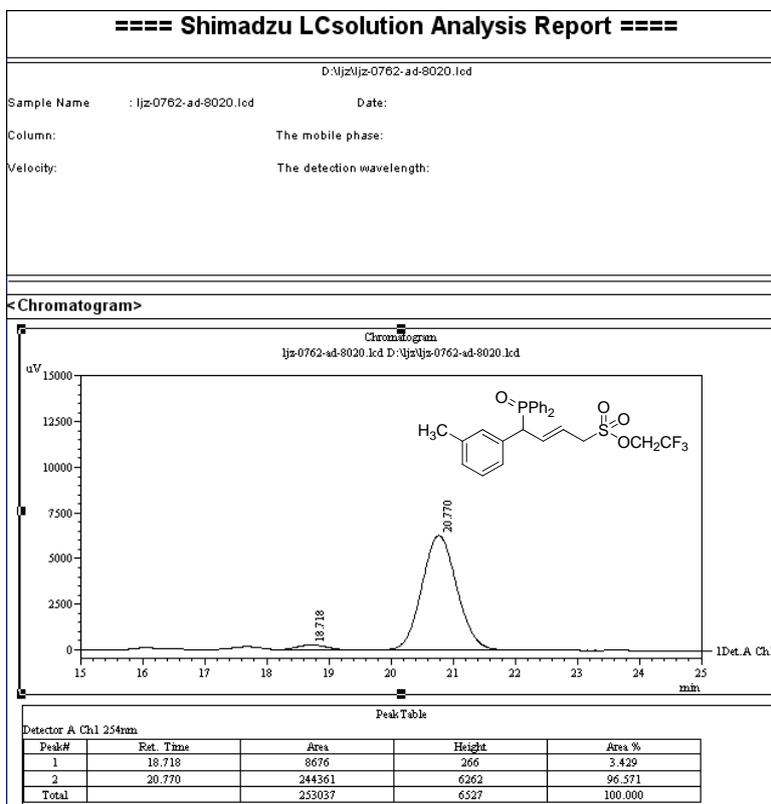
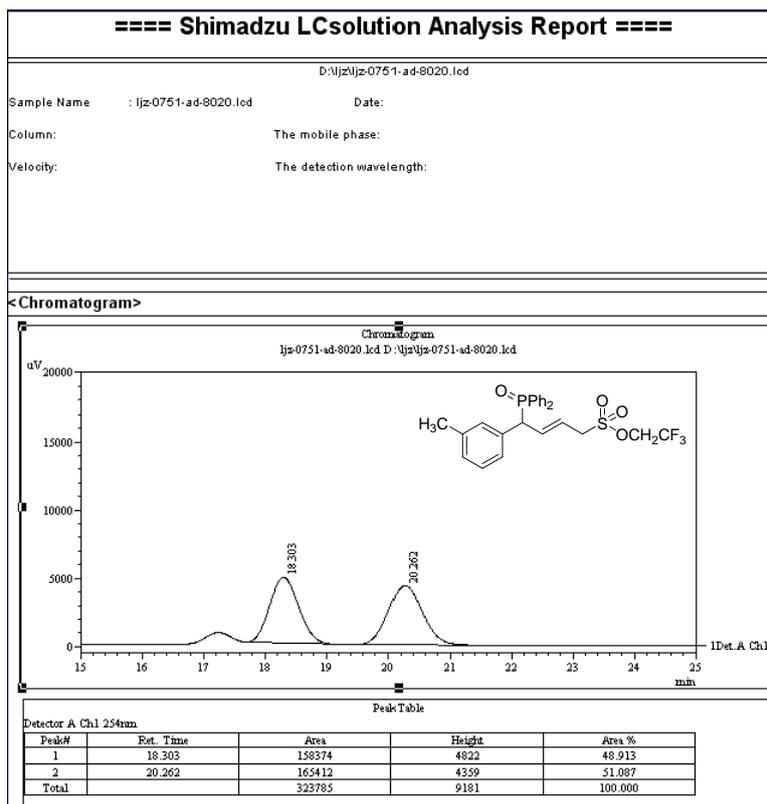
Sample Name : ljz-0713-OD-1-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	15.402	51452	870	2.042
2	26.619	2468763	26684	97.958
Total		2520215	27553	100.000

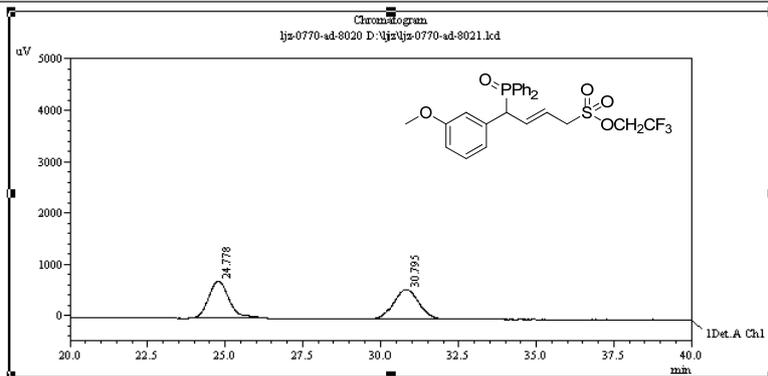


==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0770-ad-8021.lcd

Sample Name : ljz-0770-ad-8020 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

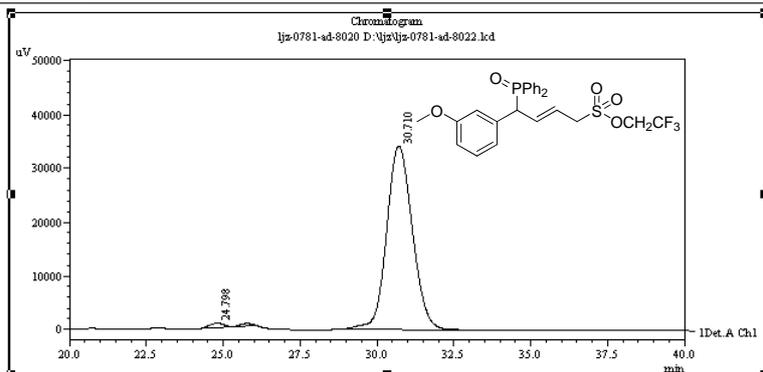
Peak#	Ret. Time	Area	Height	Area %
1	24.778	31716	714	49.636
2	30.795	32182	578	50.364
Total		63898	1292	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0781-ad-8022.lcd

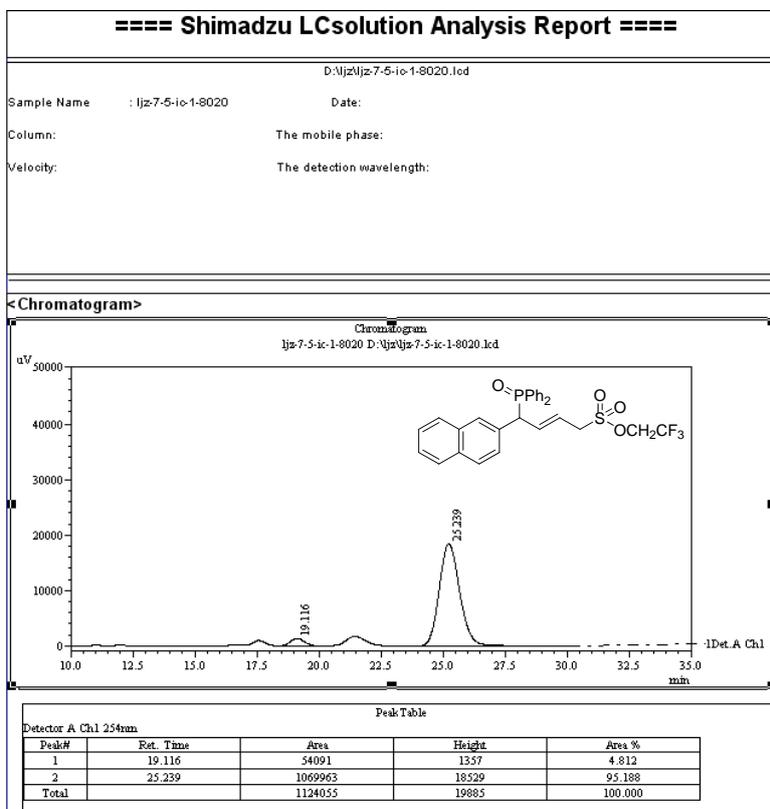
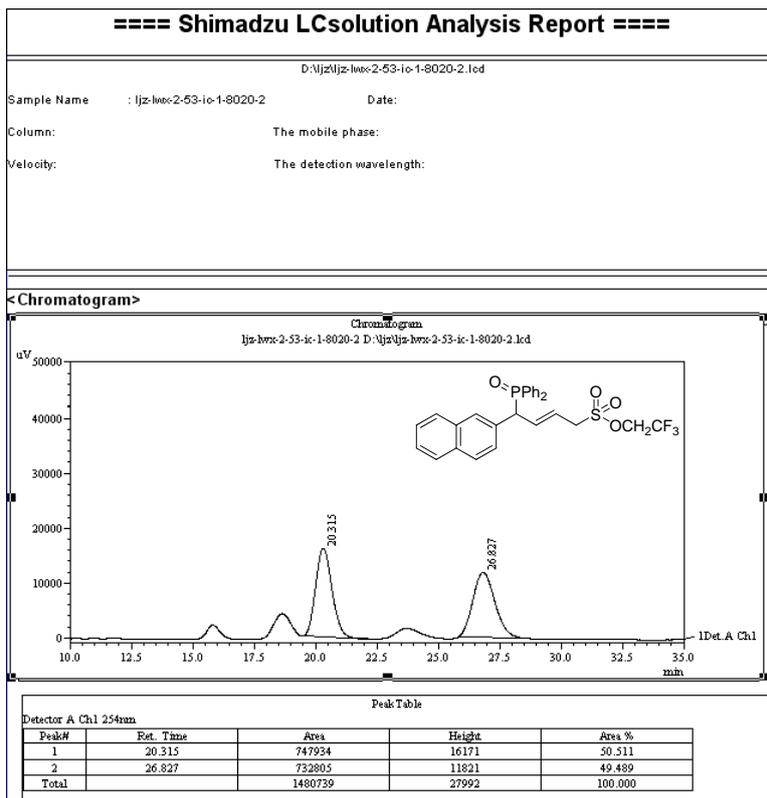
Sample Name : ljz-0781-ad-8020 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	24.798	48131	882	2.353
2	30.710	1997251	34210	97.647
Total		2045382	35092	100.000



==== Shimadzu LCsolution Analysis Report ====

D:\jz\jz-0690-ic-1-8020.lcd

Sample Name : ljz-0690-ic-1-8020

Date:

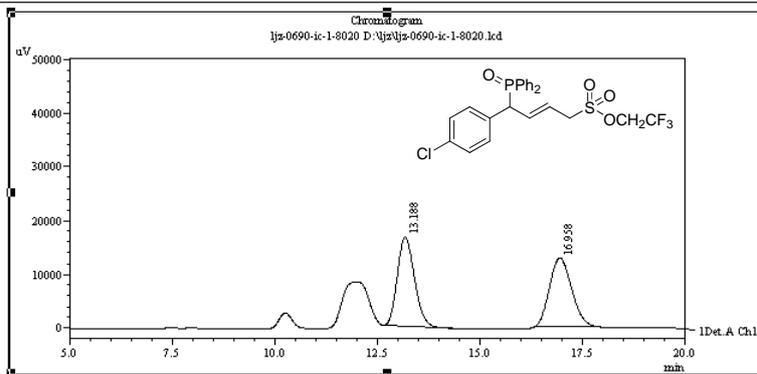
Column:

The mobile phase:

Velocity:

The detection wavelength:

<Chromatogram>



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	13.188	482165	16568	50.532
2	16.958	472009	12722	49.468
Total		954175	29290	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\jz\jz-0710-ic-1-8020.lcd

Sample Name : ljz-0710-ic-1-8020

Date:

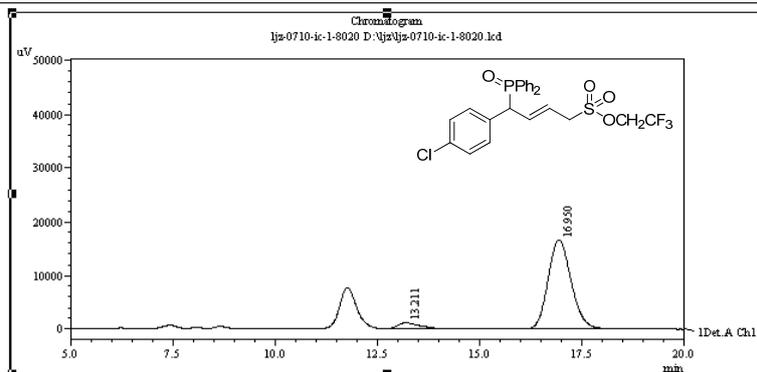
Column:

The mobile phase:

Velocity:

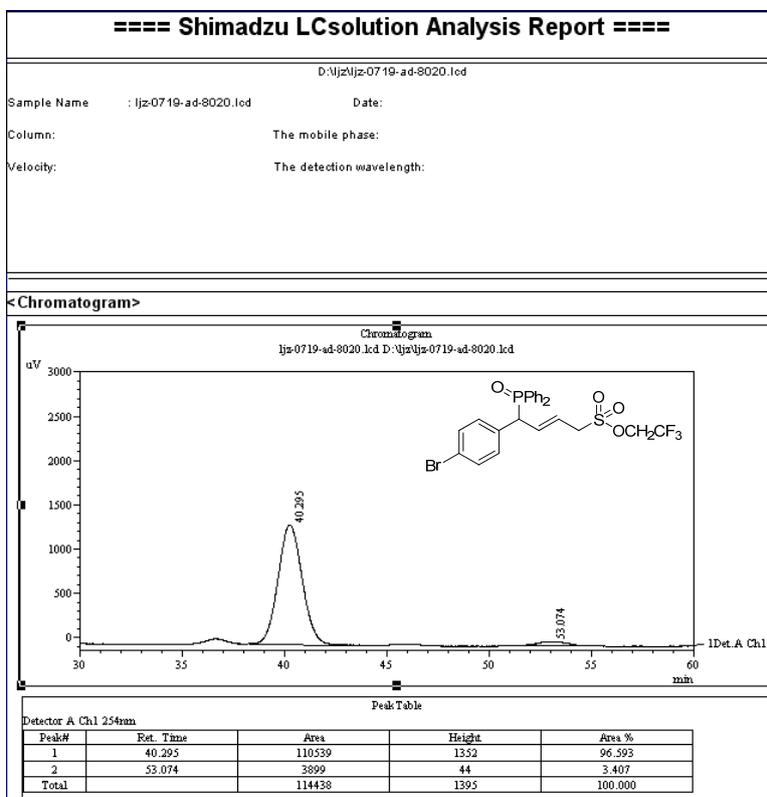
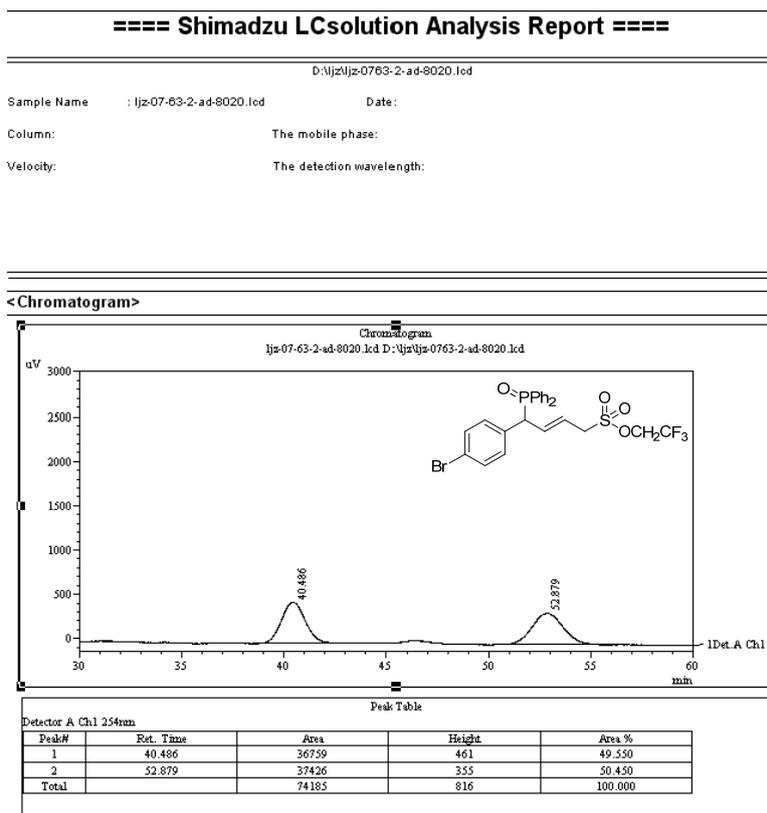
The detection wavelength:

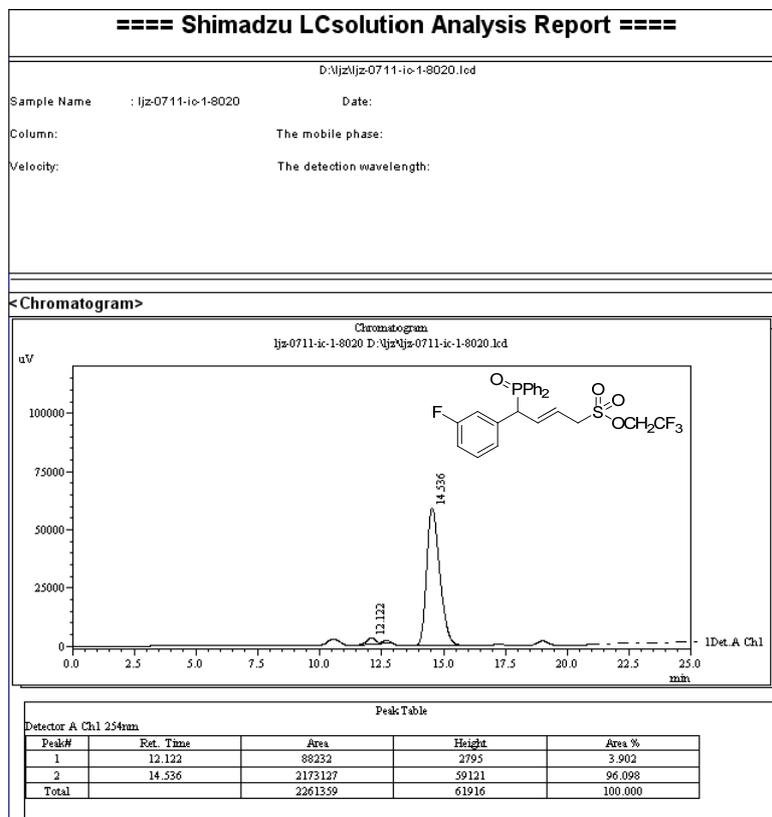
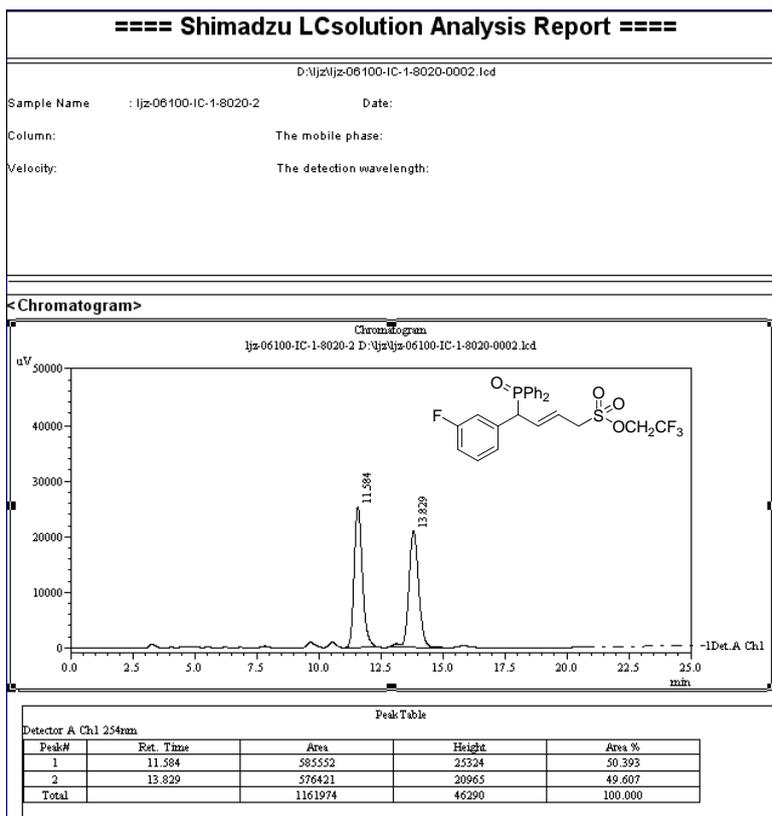
<Chromatogram>



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	13.211	40916	1100	5.872
2	16.950	655904	16654	94.128
Total		696820	17754	100.000



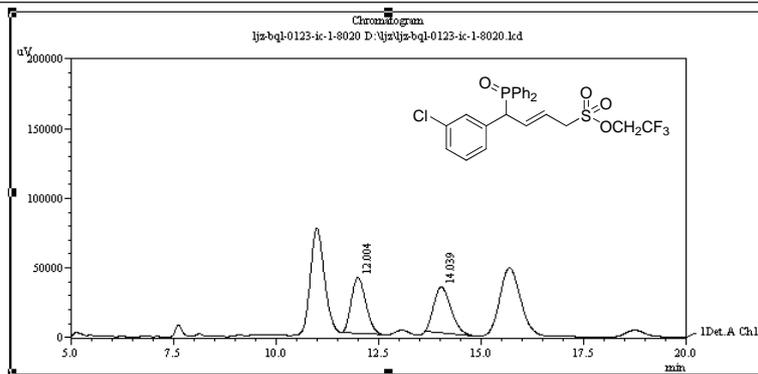


==== Shimadzu LCsolution Analysis Report ====

D:\jz\l\jz-bql-0123-ic-1-8020.lcd

Sample Name : ljz-bql-0123-ic-1-8020 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

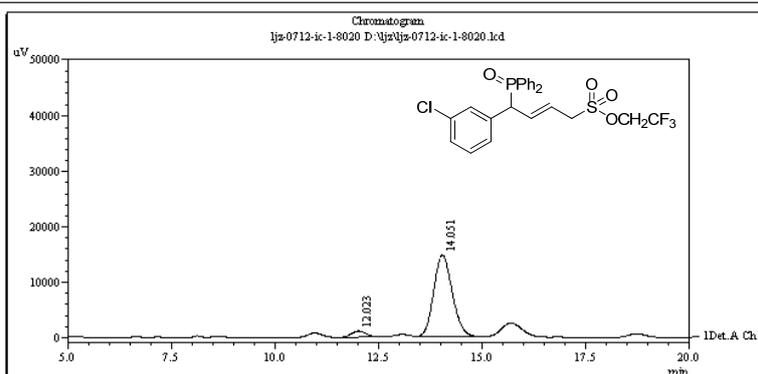
Peak#	Ret. Time	Area	Height	Area %
1	12.004	972842	39924	50.056
2	14.039	970650	33031	49.944
Total		1943492	72955	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\jz\l\jz-0712-ic-1-8020.lcd

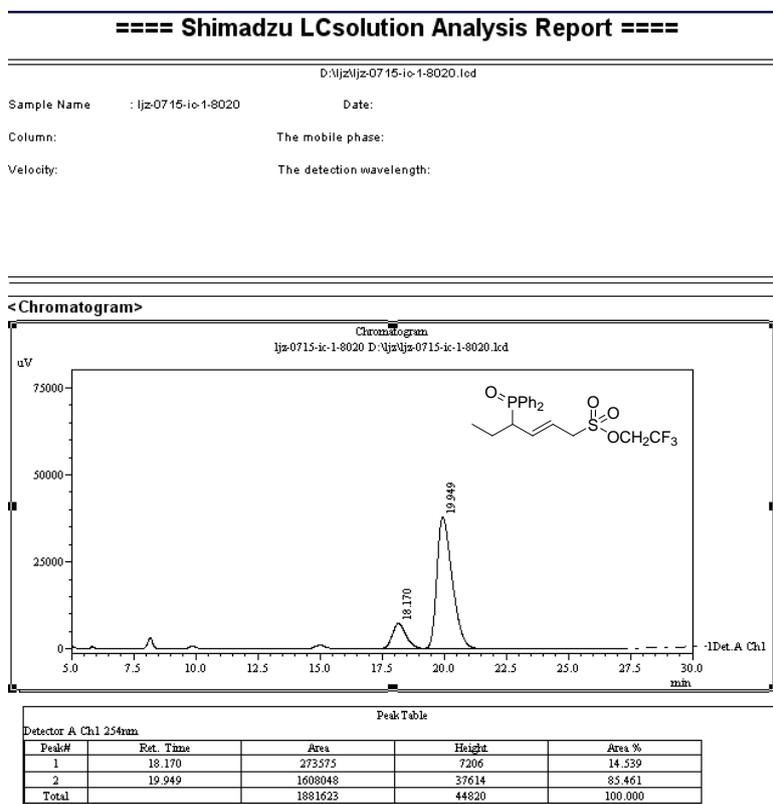
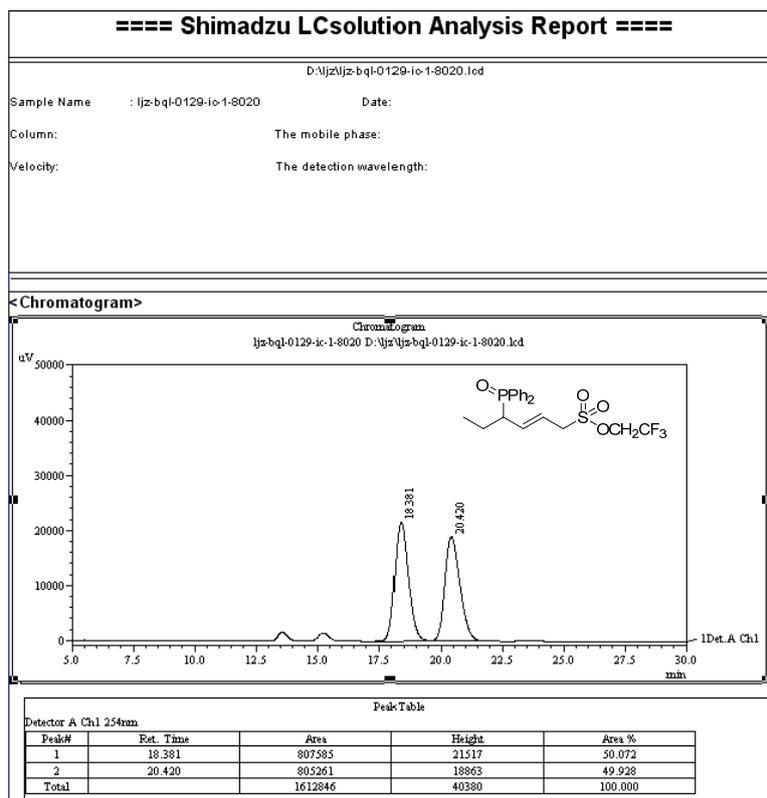
Sample Name : ljz-0712-ic-1-8020 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

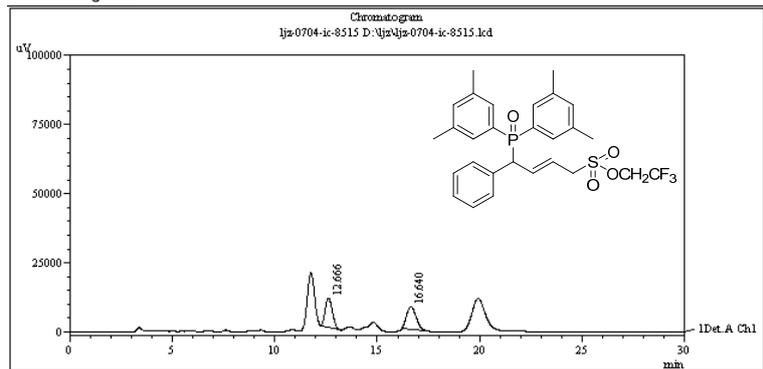
Peak#	Ret. Time	Area	Height	Area %
1	12.023	27862	1110	5.582
2	14.051	457746	14729	94.418
Total		484809	15840	100.000



==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0704-ic-8515.lcd
 Sample Name : ljz-0704-ic-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



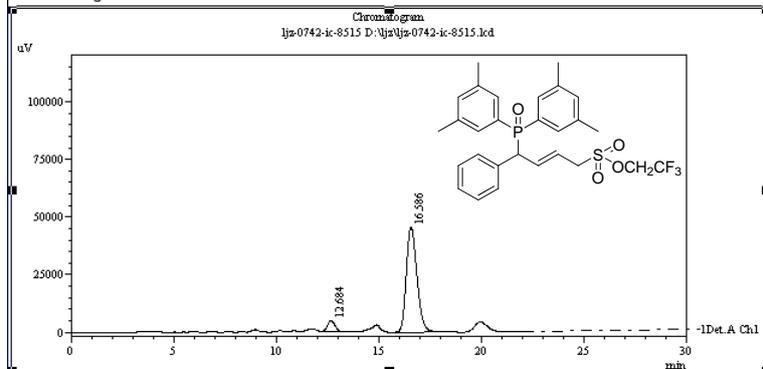
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	12.666	252616	10469	49.140
2	16.640	261458	8112	50.860
Total		514074	18581	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0742-ic-8515.lcd
 Sample Name : ljz-0742-ic-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

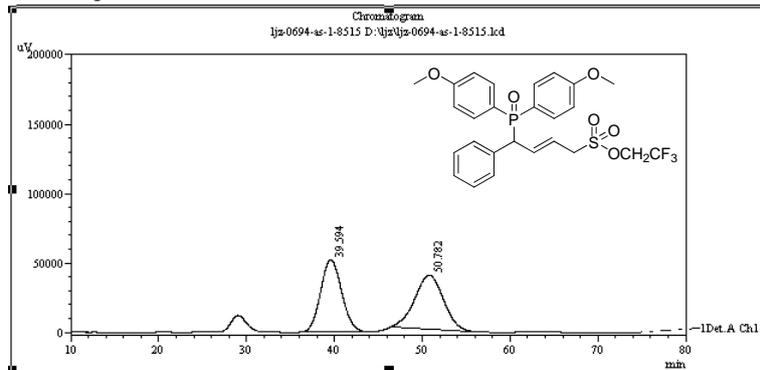
Peak#	Ret. Time	Area	Height	Area %
1	12.684	121656	4847	6.861
2	16.586	1651367	45608	93.139
Total		1773023	50455	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0694-as-1-8515.lcd

Sample Name : ljz-0694-as-1-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



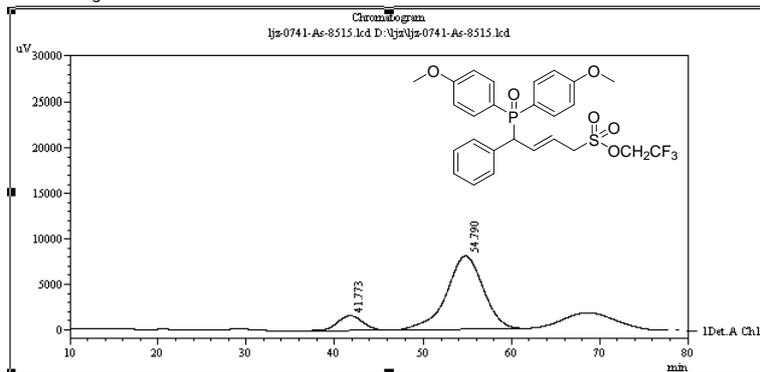
Peak Table				
Peak#	Ret. Time	Area	Height	Area %
1	39.594	8701891	52084	49.564
2	50.782	8855048	38892	50.436
Total		17556940	90976	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0741-As-8515.lcd

Sample Name : ljz-0741-As-8515 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table				
Peak#	Ret. Time	Area	Height	Area %
1	41.773	322026	1628	12.075
2	54.790	2344904	8065	87.925
Total		2666929	9693	100.000

==== Shimadzu LCsolution Analysis Report ====

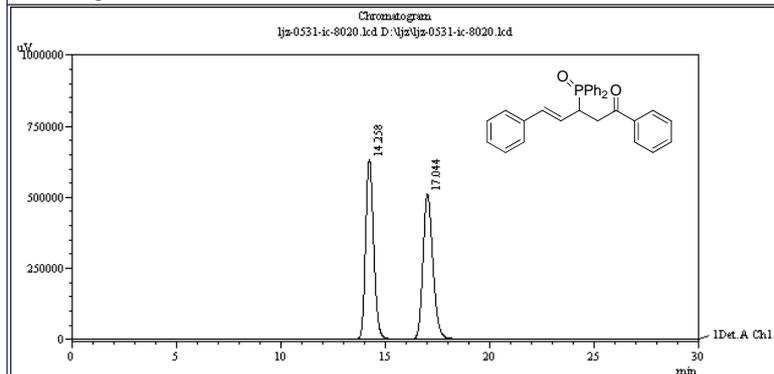
D:\jz\l\jz-0531-ic-8020.lcd

Sample Name : l\jz-0531-ic-8020.lcd Date:

Column: The mobile phase:

Velocity: The detection wavelength:

<Chromatogram>



Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	14.238	16826752	632342	49.889
2	17.044	16901672	509331	50.111
Total		33728425	1141673	100.000

==== Shimadzu LCsolution Analysis Report ====

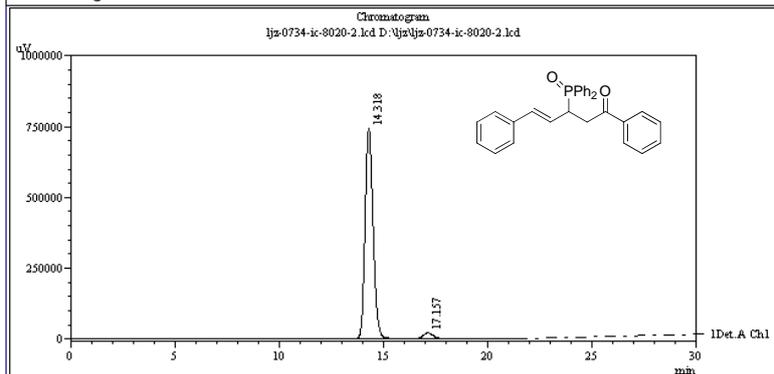
D:\jz\l\jz-0734-ic-8020-2.lcd

Sample Name : l\jz-0734-ic-8020-2.lcd Date:

Column: The mobile phase:

Velocity: The detection wavelength:

<Chromatogram>



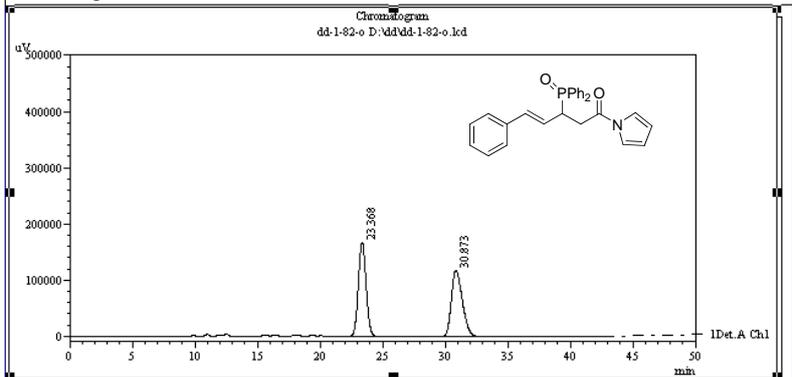
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	14.318	19746403	742746	96.592
2	17.157	696782	20714	3.408
Total		20443185	763461	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\dd\dd-1-82-o.lcd
 Sample Name : dd-1-82-o Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



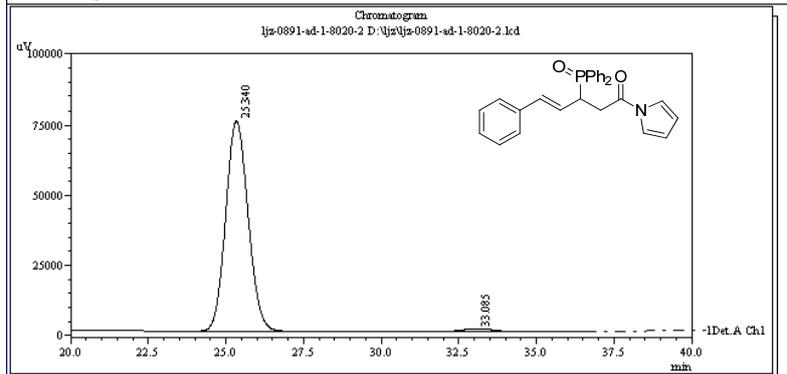
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	23.368	7239253	167694	50.109
2	30.873	7207838	118112	49.891
Total		14447111	285806	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0891-ad-1-8020-2.lcd
 Sample Name : ljz-0891-ad-1-8020-2 Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

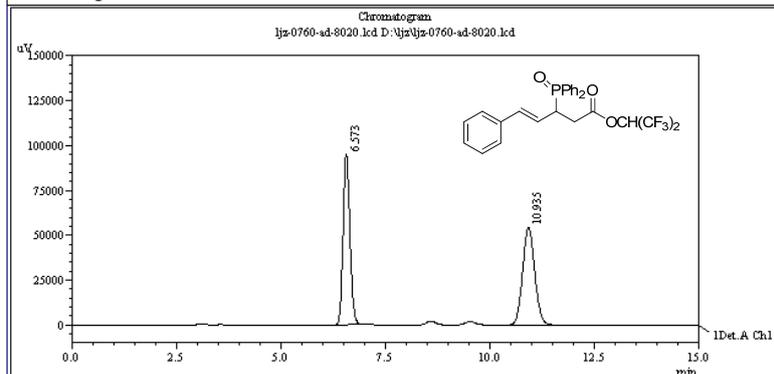
Peak#	Ret. Time	Area	Height	Area %
1	25.340	3784791	79085	97.958
2	33.085	78891	1145	2.042
Total		3863682	76230	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0760-ad-8020.lcd

Sample Name : ljz-0760-ad-8020.lcd Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



Peak Table

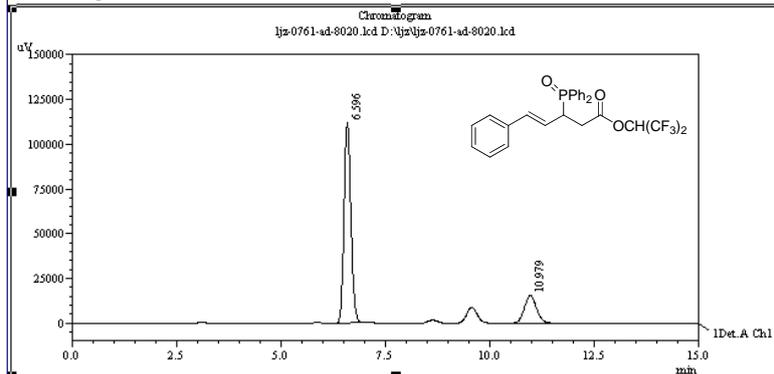
Peak#	Ret. Time	Area	Height	Area %
1	6.573	1090506	95029	49.861
2	10.935	1096586	54173	50.139
Total		2187092	149202	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\ljz\ljz-0761-ad-8020.lcd

Sample Name : ljz-0761-ad-8020.lcd Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



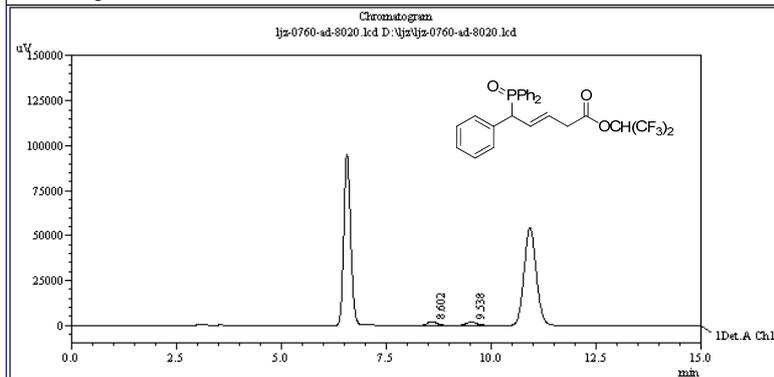
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	6.596	1288561	112030	80.476
2	10.979	312612	15386	19.524
Total		1601173	127416	100.000

==== Shimadzu LCsolution Analysis Report ====

D:\jz\jz-0760-ad-8020.lcd
 Sample Name : jz-0760-ad-8020.lcd Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

<Chromatogram>



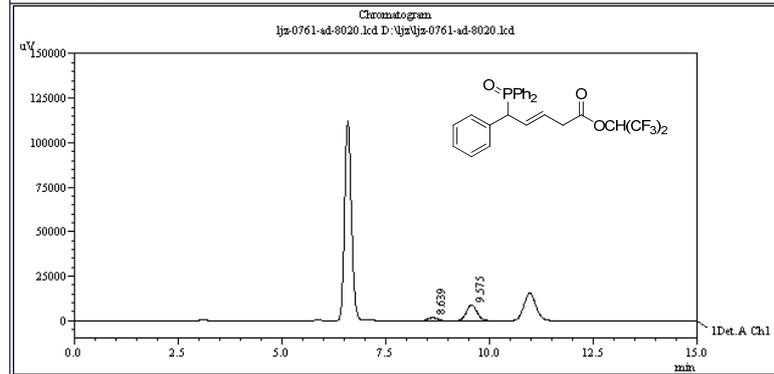
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	8.602	33674	2150	50.553
2	9.538	32937	1835	49.447
Total		66611	3985	100.000

==== Shimadzu LCsolution Analysis Report ====

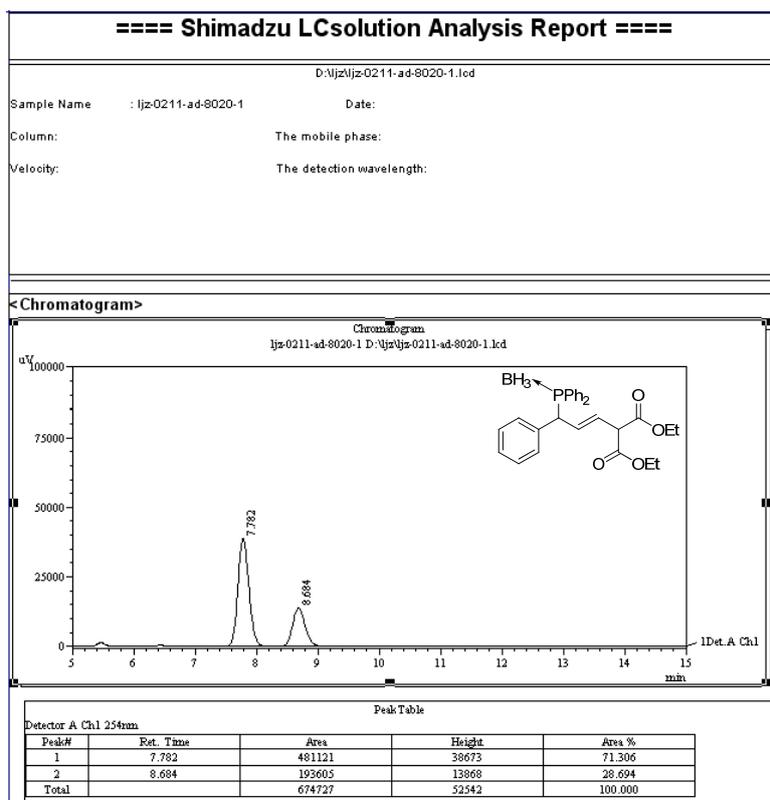
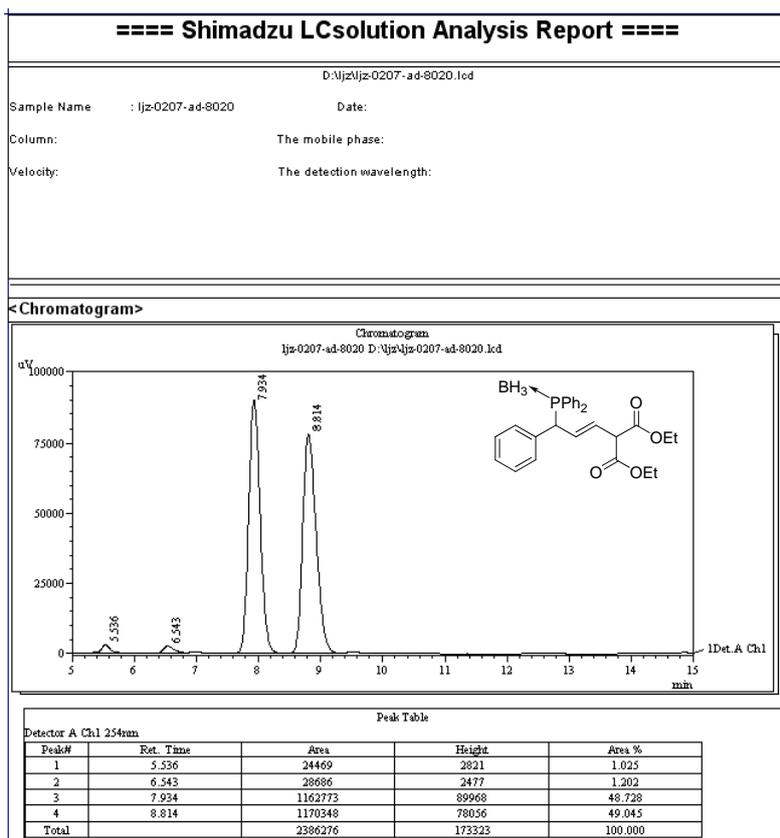
D:\jz\jz-0761-ad-8020.lcd
 Sample Name : jz-0761-ad-8020.lcd Date:
 Column: The mobile phase:
 Velocity: The detection wavelength:

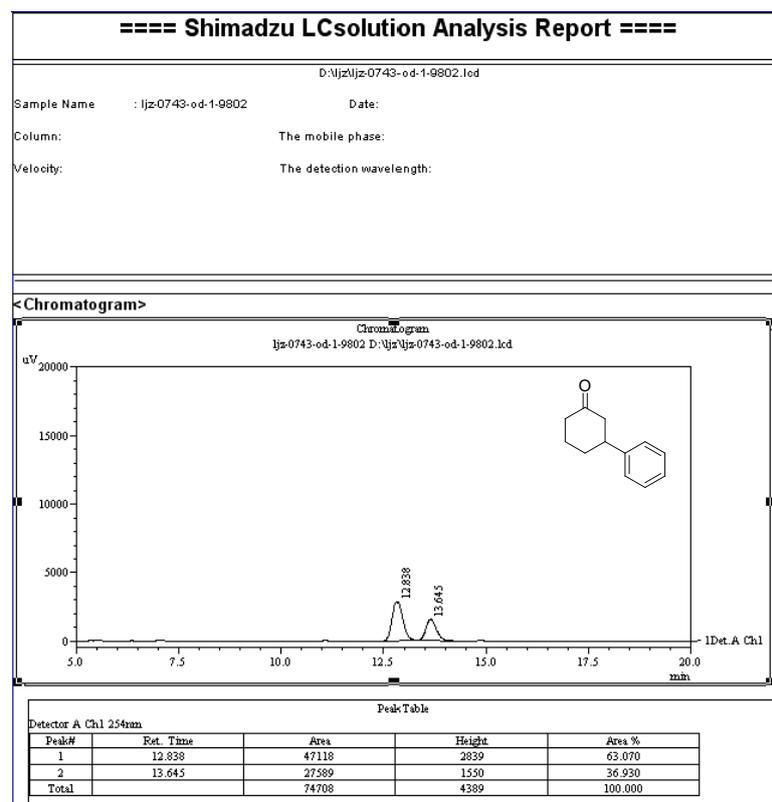
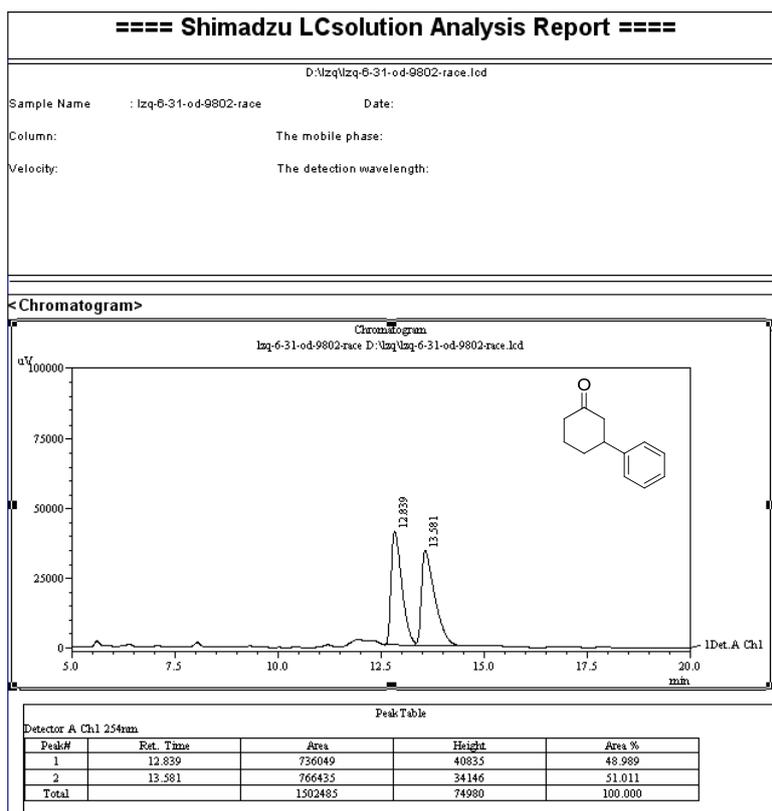
<Chromatogram>



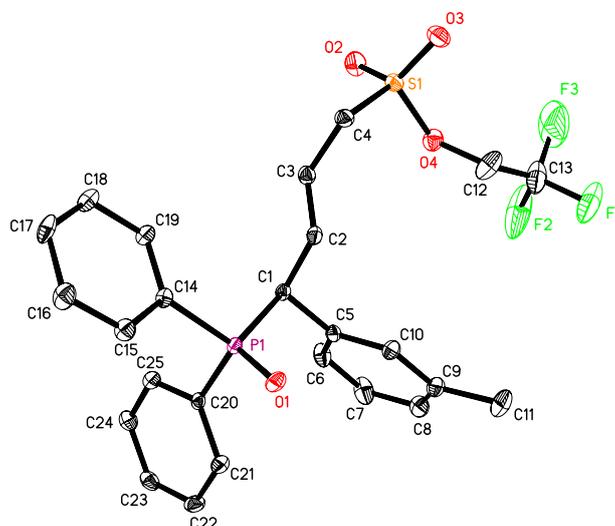
Peak Table

Peak#	Ret. Time	Area	Height	Area %
1	8.639	27189	1740	14.624
2	9.575	158753	8940	85.376
Total		185942	10680	100.000





X-ray Data



1.6-adduct in Table 2, Entry 4

Table 1. Crystal data and structure refinement for mo_dm13397_0m.

Identification code	mo_dm13397_0m	
Empirical formula	C ₂₅ H ₂₄ F ₃ O ₄ P S	
Formula weight	508.47	
Temperature	140(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P 21	
Unit cell dimensions	a = 5.6851(12) Å	$\alpha = 90^\circ$.
	b = 16.633(4) Å	$\beta = 99.518(4)^\circ$.
	c = 13.304(3) Å	$\gamma = 90^\circ$.
Volume	1240.7(4) Å ³	
Z	2	
Density (calculated)	1.361 Mg/m ³	
Absorption coefficient	0.246 mm ⁻¹	
F(000)	528	
Crystal size	0.390 x 0.150 x 0.100 mm ³	
Theta range for data collection	1.552 to 30.606°.	
Index ranges	-8 ≤ h ≤ 8, -23 ≤ k ≤ 23, -19 ≤ l ≤ 17	
Reflections collected	12282	
Independent reflections	6964 [R(int) = 0.0457]	
Completeness to theta = 25.242°	100.0 %	

Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.7461 and 0.6580
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	6964 / 19 / 308
Goodness-of-fit on F ²	1.029
Final R indices [I > 2σ(I)]	R1 = 0.0513, wR2 = 0.1135
R indices (all data)	R1 = 0.0691, wR2 = 0.1236
Absolute structure parameter	-0.06(6)
Extinction coefficient	n/a
Largest diff. peak and hole	0.455 and -0.275 e.Å