

## Electronic Supplementary information

### New heterocyclic systems to afford microsecond green-light isomerisable azo dyes and their use as fast molecular photochromic switches

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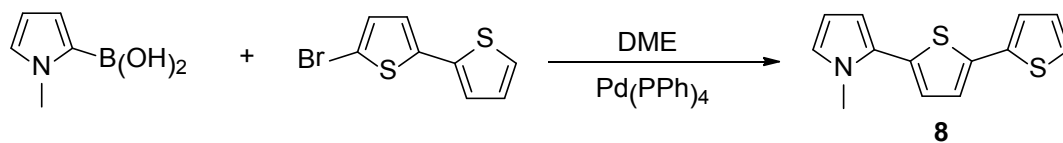
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### Synthesis of azoderivatives 1-7

**Materials:** 2-Aminobenzothiazole, 5-aminobenzothiazole, 4-nitroaniline, 2-bromo-2,2'-bithiophene and 1-methyl-1*H*-pyrrole-2-boronic used as precursors for the synthesis of azo dyes **5-7** were purchased from Aldrich and Fluka and used as received. TLC analyses were carried out on 0.25 mm thick precoated silica plates (Merck Fertigplatten Kieselgel 60F<sub>254</sub>) and spots were visualised under UV light. Chromatography on silica gel was carried out on Merck Kieselgel (230-240 mesh).

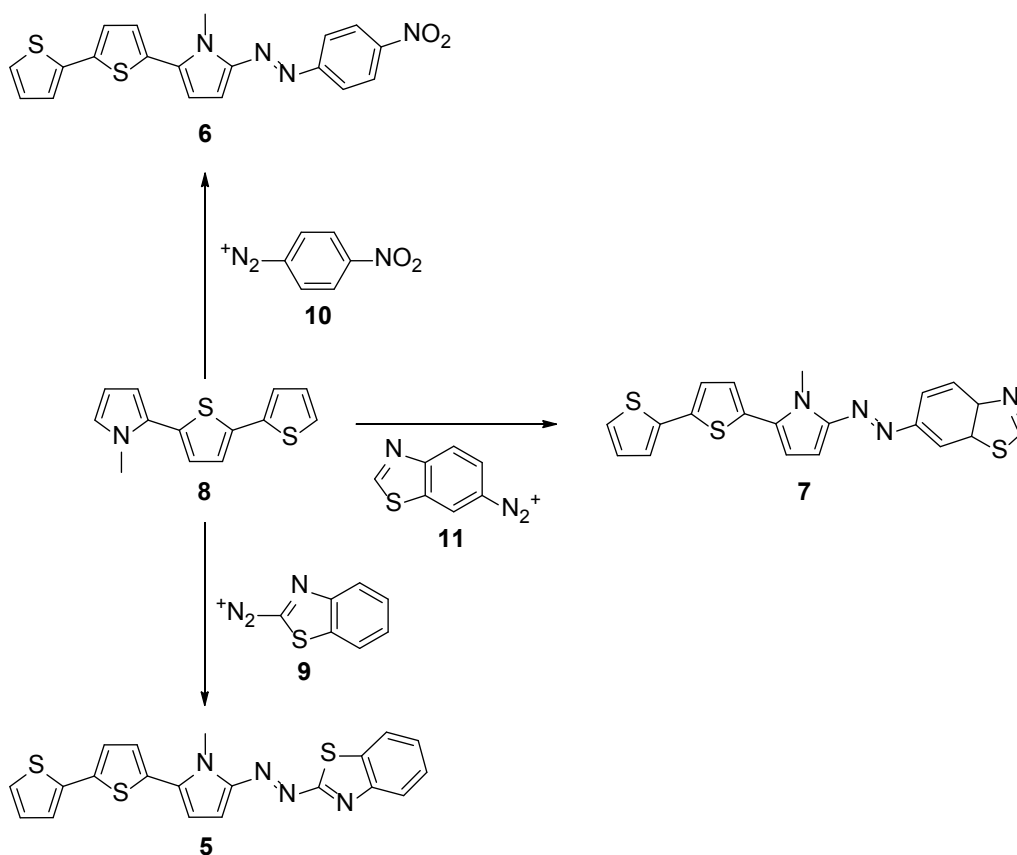
**Characterization:** NMR spectra were obtained on a Varian Unity Plus Spectrometer at an operating frequency of 300 MHz for <sup>1</sup>H NMR and 75.4 MHz for <sup>13</sup>C NMR or a Bruker Avance III 400 at an operating frequency of 400 MHz for <sup>1</sup>H NMR and 100.6 MHz for <sup>13</sup>C NMR using the solvent peak as an internal reference at 25 °C. All chemical shifts are given in ppm using  $\delta_{\text{H}}$  Me<sub>4</sub>Si = 0 ppm as a reference and *J* values are given in Hz. Assignments were made by comparison of chemical shifts, peak multiplicities and *J* values and were supported by spin decoupling-double resonance and bidimensional heteronuclear HMBC and HMQC correlation techniques. IR spectra were recorded on a BOMEM MB 104 spectrophotometer using KBr discs. Mass spectrometry analyses were performed at the "C.A.C.T.I.-Unidad de Espectrometria de Masas" at the University of Vigo (Spain). All melting points were measured on a Gallenkamp melting point apparatus and are uncorrected. UV-visible absorption spectra (200–800 nm) were obtained using a Shimadzu UV/2501PC spectrophotometer or a Varian Cary 500 UV-vis spectrophotometer.

**Synthesis of the azocompounds:** Bithienylpyrrole **8** was synthesized through a Suzuki cross-coupling reaction<sup>[4]</sup> between the commercially available 1-methyl-1*H*-pyrrole-2-yl-2-boronic acid and 5-bromo-2,2'-bithiophene. (Scheme S1).



**Scheme S1.** Synthetic concept for bithienylpyrrole **8**.

Precursor **8** was used as a coupling component together with benzothiazolyl- (**9** and **11**) and aryl- (**10**) diazonium salts in order to prepare bithienylpyrrole azo dyes **5-7** (Scheme S2).



**Scheme S2.** Synthesis of the heterocyclic azoderivatives **5-7**.

All new compounds were completely characterized by <sup>1</sup>H and <sup>13</sup>C NMR, IR, MS and HRMS and the data obtained were in full agreement with the proposed formulation.

**Synthesis of 1-methyl-2-(5-(thiophen-2-yl)thiophen-2-yl)-1*H*-pyrrole (**8**).** 2-Bromo-2,2'-bithiophene (1 equiv.) was coupled to 1-methyl-1*H*-pyrrole-2-boronic acid (1.2 equiv) in 1,2-dimethoxyethane (10 cm<sup>3</sup>) and a 2M Na<sub>2</sub>CO<sub>3</sub> aqueous solution (1 cm<sup>3</sup>) using Pd(PPh<sub>3</sub>)<sub>4</sub> as a

catalyst (3 mol %) at 80 °C under nitrogen. The reaction was monitored by TLC. After 32 hours, the reaction mixture was cooled down to room temperature. Next, a saturated solution of NaCl was added (20 cm<sup>3</sup>) and the product was extracted with chloroform (3×20 cm<sup>3</sup>). The organic layer was washed first with water (3×10 cm<sup>3</sup>) and, after, with an aqueous NaOH solution (10%, 1×10 cm<sup>3</sup>). The combined organic extract was dried over anhydrous MgSO<sub>4</sub>, filtered, and the solvent was removed. The crude was purified by means of column chromatography using mixtures of chloroform and light petroleum of increasing polarity to afford the coupled product **8** as a dark green oil (126 mg, 85 %). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 3.80 (s, 3H, CH<sub>3</sub>), 6.27-6.29 (m, 1H, 4-H), 6.47-6.48 (m, 1H, 3-H), 6.78 (m, 1H, 5-H), 7.00 (d, 1H, *J*=3.8 Hz, 4'-H), 7.09 (dd, 1H, *J*=3.6 and *J*=1.2 Hz, 4''-H), 7.21 (d, 1H, *J*=3.8 Hz, 3'-H), 7.25-7.28 (m, 2H, 3''- and 5''-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 35.2, 107.9, 109.9, 123.3, 123.9, 124.1, 124.3, 124.9, 126.7, 127.7, 133.9, 135.9, 137.2. λ<sub>max</sub> (Dioxane)/nm 352 (ε/dm<sup>3</sup> mol<sup>-1</sup> cm<sup>-1</sup> 17,880). IR (CHCl<sub>3</sub>): ν 3102, 3071, 2945, 1692, 1503, 1453, 1416, 1318, 1294, 1237, 1200, 1090, 1055 cm<sup>-1</sup>. MS (EI) *m/z* (%) = 245 ([M]<sup>+</sup>, 100), 244 (22), 230 (13). HRMS: *m/z* (EI) for C<sub>13</sub>H<sub>11</sub>NS<sub>2</sub>; calcd 245.0333; found: 245.0336.

***Procedure for the azo coupling of bithienylpyrrole 8 with the diazonium salt 9 to afford azo dye 5.***

***Diazotation of 2-aminobenzothiazole.*** 2-Aminobenzothiazole (1.0 mmol) was dissolved in concentrated H<sub>2</sub>SO<sub>4</sub> (1.2 mmol) at 0-5 °C. A mixture of NaNO<sub>2</sub> (1.2 mmol) in water (1 cm<sup>3</sup>) was slowly added to the well-stirred mixture of the amine derivative solution at 0-5 °C.

***Coupling reaction with bithienylpyrrole 8.*** The previously prepared solution of the diazonium salt **9** (1.0 mmol) was added dropwise to the solution of bithienylpyrrole **8** (1.2 mmol) in acetic anhydride (15 cm<sup>3</sup>): ice water (200 cm<sup>3</sup>). The combined solution was maintained at 0 °C for 1 h followed by stirring for 3 h at room temperature. Then, the precipitate was filtered, washed with warm water and *n*-hexane and dried over vacuum.

***2-(Benzo[d]thiazol-2-yl)-1-(1-methyl-5-(5-(thiophen-2-yl)thiophen-2-yl)-1H-pyrrol-2-yl)diazene (5).*** Dark purple solid (51 mg, 73 %). m.p. 164-165 °C. <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 4.11 (s, 3H, NCH<sub>3</sub>), 7.07 (d, 1H, *J*=4.6 Hz, 4'-H), 7.14-7.17 (m, 1H, 4'''-H), 7.18 (d, 1H, *J*=4.6 Hz, 3'-H), 7.41 (dt, 1H, *J*=8.4 and *J*=1.2 Hz, 5-H), 7.47-7.49 (m, 2H, 6- and 3'''-H), 7.51 (d, 1H, *J*=4 Hz, 4''-H), 7.62 (dd, 1H, *J*=4.8 and *J*=0.8 Hz, 5'''-H), 7.73 (d, 1H, *J*=4 Hz, 3''-H), 7.93-7.95 (m, 1H, 4-H), 7.99 (dd, 1H, *J*=7.6 and *J*=0.8 Hz, 7-H). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 30.7, 115.9, 122.4, 123.0, 125.2 (2C), 125.4, 126.1 (2C), 126.5, 126.8, 128.7, 130.0, 130.1, 133.0, 135.4, 138.6, 139.3, 152.6, 177.3. λ<sub>max</sub> (Dioxane)/nm 533 (ε/dm<sup>3</sup> mol<sup>-1</sup> cm<sup>-1</sup> 25,950). IR (Nujol): ν 1685, 1552, 1503, 1301, 1250, 1225, 1164, 1125, 1063 cm<sup>-1</sup>. MS (EI) *m/z* (%) = 406 ([M]<sup>+</sup>, 35),

380 (15), 378 (100), 260 (20), 258 (35), 257 (15), 245 (15), 206 (16), 204 (12), 203 (72), 190 (28), 179 (17), 96 (15). HRMS:  $m/z$  (EI) for  $C_{20}H_{14}N_4S_3$ ; calcd 406.0381; found: 406.0385.

**General procedure for the azo coupling of bithienylpyrrole 8 with diazonium salts 10 and 11 to afford azo dyes 6 and 7.**

*Diazotation of 5-aminobenzothiazole and 4-nitroaniline.* Heteroaromatic amines were dissolved in HCl 6 N (1 cm<sup>3</sup>) at 0-5 °C. A mixture of NaNO<sub>2</sub> (1.0 mmol) in water (2 cm<sup>3</sup>) was slowly added to the well-stirred mixture of the amine derivative solution at 0-5 °C. The reaction mixture was stirred for 10 min.

*Coupling reaction with bithienylpyrrole 8.* The previously prepared diazonium salt solution (1.0 mmol) was added dropwise to the solution of bithienylpyrrole **8** (0.52 mmol) in acetonitrile (10 cm<sup>3</sup>) and 2-3 drops of acetic acid. The resulting mixture was maintained at 0 °C for 1-2 h while stirred and then diluted with chloroform (20 cm<sup>3</sup>), washed with water and dried over anhydrous MgSO<sub>4</sub>. The dried solution was evaporated and the remaining azo dyes were purified by column chromatography using dichloromethane/*n*-hexane as eluent.

*1-(1-Methyl-5-(5-(thiophen-2-yl)thiophen-2-yl)-1H-pyrrol-2-yl)-2-(4-nitrophenyl)diazene (6).* Brown solid (72 mg, 60 %). m.p. 204-206 °C. <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 4.15 (s, 3H, NCH<sub>3</sub>), 6.86 (d, 1H,  $J=4.6$  Hz, 4'-H), 6.95 (d, 1H,  $J=4.6$  Hz, 3'-H), 7.12-7.15 (m, 1H, 4''-H), 7.43 (dd, 1H,  $J=3.6$  and  $J=1.2$  Hz, 3'''-H), 7.45 (d, 1H,  $J=4$  Hz, 4''-H), 7.58-7.59 (m, 2H, 3''- and 5'''-H), 7.96 (d, 2H,  $J=8.8$  Hz, 2- and 6-H), 8.34 (d, 2H,  $J=8.8$  Hz, 3- and 5-H). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 31.6, 113.7, 122 (2C), 124.8, 125.0 (2C), 125.2, 126.3, 128.5, 128.6, 130.8, 135.1, 135.6, 137.8, 146.4, 146.5, 148.1, 157.2.  $\lambda_{\max}$  (Dioxane)/nm 507 ( $\epsilon/\text{dm}^3 \text{ mol}^{-1}$  47,580). IR (Nujol):  $\nu$  3108, 2670, 1694, 1603, 1584, 1515, 1313, 1298, 1214, 1206, 1191, 1170, 1139, 1106, 1098, 1083, 1042 cm<sup>-1</sup>. MS (EI)  $m/z$  (%) = 394 ([M]<sup>+</sup>, 62), 365 (16), 364 (73), 257 (12), 246 (10), 245 (31), 244 (65), 217 (10), 203 (100), 190 (10), 179 (42). HRMS:  $m/z$  (EI) for  $C_{19}H_{14}N_4O_2S_2$ ; calcd 394.0558; found: 394.0575.

*2-(Benzo[d]thiazol-6-yl)-1-(1-methyl-5-(5-(thiophen-2-yl)thiophen-2-yl)-1H-pyrrol-2-yl)diazene (7).* Brown solid (33 mg, 40 %). m.p. 198-199 °C <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ 4.16 (s, 3H, NCH<sub>3</sub>), 6.73 (d, 1H,  $J=4.5$  Hz, 4'-H), 6.80 (d, 1H,  $J=4.5$  Hz, 3'-H), 7.11-7.14 (m, 1H, 4''-H), 7.39-7.40 (m, 1H, 3'''-H), 7.41 (d, 1H,  $J=3.8$  Hz, 4''-H), 7.48 (d, 1H,  $J=3.8$  Hz, 3''-H), 7.56 (dd, 1H,  $J=5.4$  and  $J=0.6$  Hz, 5'''-H), 8.0 (dd, 1H,  $J=8.9$  and  $J=2.1$  Hz, 5-H), 8.2 (d, 1H,  $J=8.9$  Hz, 4-H), 8.61 (d, 1H,  $J=2.1$  Hz, 7-H). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 31.5, 100.7, 112.3, 116.5, 120.1, 123.5, 124.5, 125.0, 126.0, 127.5, 128.5, 131.5, 132.4, 134.9, 135.8, 136.8, 147.4, 150.8, 153.6, 158.0.  $\lambda_{\max}$  (Dioxane)/nm 489 ( $\epsilon/\text{dm}^3 \text{ mol}^{-1}$  45,790). IR (Nujol):  $\nu$  2671, 1305, 1168, 1074,

1040, 965  $\text{cm}^{-1}$ . MS (EI)  $m/z$  (%) = 406 ( $[\text{M}]^+$ , 100), 378 (11), 363 (15), 244 (44), 204 (11), 203 (82), 179 (30), 134 (22). HRMS:  $m/z$  (EI) for  $\text{C}_{20}\text{H}_{14}\text{N}_4\text{S}_3$ ; calcd 406.0381; found: 406.0394.

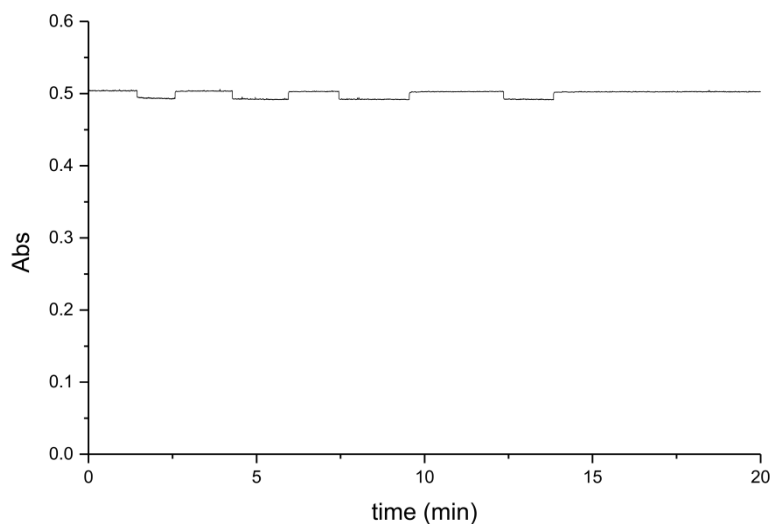
## **Spectroscopic and kinetic measurements**

**Time-resolved UV-vis spectroscopy:** For measurements in acetone, 20  $\mu\text{M}$  solutions were used. Irradiation experiments were made using a CARY 50 Varian spectrophotometer coupled to a 150W ozone free xenon lamp (6255 Oriel Instruments). The light from the UV-vis lamp was filtered using a water filter (61945 Oriel Instruments) and a long-pass filter (Schott GG 420) at 20  $^\circ\text{C}$  and carried to the spectrophotometer holder, perpendicular to the monitoring beam using a fibre-optic system (77654 Oriel Instruments).

**Nanosecond laser flash-photolysis setup:** In ethanol, a population of *cis* isomers was created by pulsed-laser irradiation of the *trans* isomer at 532 nm employing a Continuum Surelite I-10 Q-switched Nd-YAG laser (5 ns pulse width, *ca.* 10 mJ per pulse). The concomitant absorbance changes were monitored at  $90^\circ$  by a white-light analysing beam produced by a Xe lamp (PTI, 75 W) in combination with a dual-grating monochromator (PTI 101) coupled to a Hamamatsu R928 photomultiplier for detection.

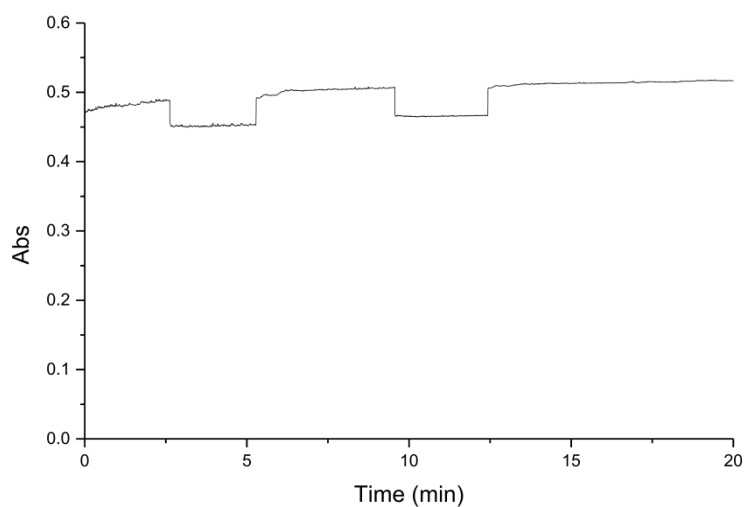
## **Photo and thermal isomerisation for azo dyes 5-7 in acetone**

### **Compound 5**



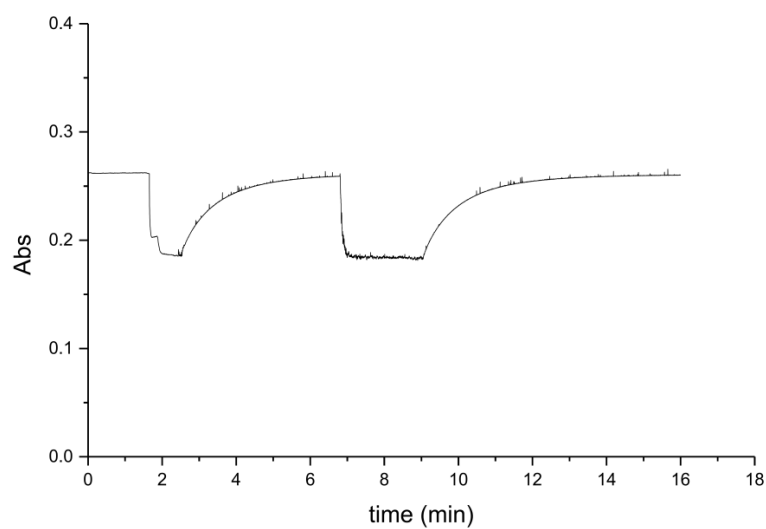
**Figure S1.** Visible light irradiation / dark cycles for azo compound **5** ( $[\mathbf{5}] = 20 \mu\text{M}$ ) in acetone at 298 K.

### Compound 6



**Figure S2.** Visible light irradiation / dark cycles for azo compound **6** ( $[6] = 20 \mu\text{M}$ ) in acetone at 298 K.

### Compound 7



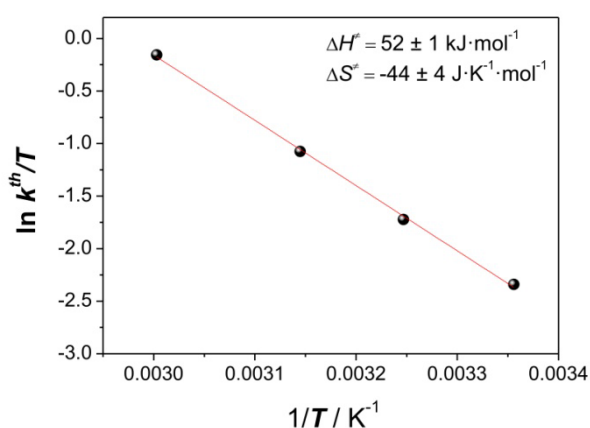
**Figure S3.** Visible light irradiation / dark cycles for azo compound **7** ( $[7] = 20 \mu\text{M}$ ) in acetone at 298 K.

## Kinetic and thermal activation parameters for the thermal *cis*-to-*trans* isomerisation kinetics for azo dyes 1-7 in ethanol

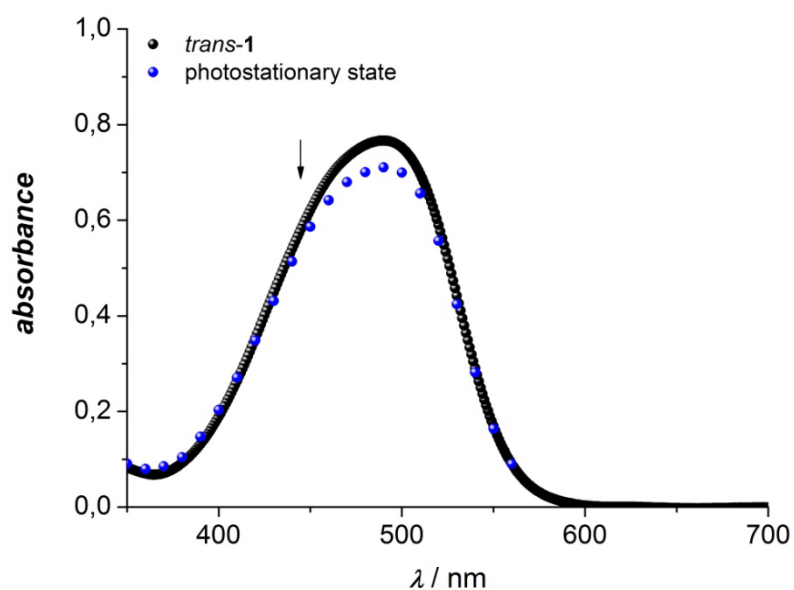
### Compound 1

|                      |    |    |     |     |
|----------------------|----|----|-----|-----|
| $T / ^\circ\text{C}$ | 25 | 35 | 45  | 60  |
| $\tau / \text{ms}$   | 35 | 19 | 9.8 | 3.5 |

**Table S1.** Relaxation time for *cis*-1 in ethanol at different temperatures.



**Figure S4.** Eyring plot for the thermal back reaction of *cis*-1.

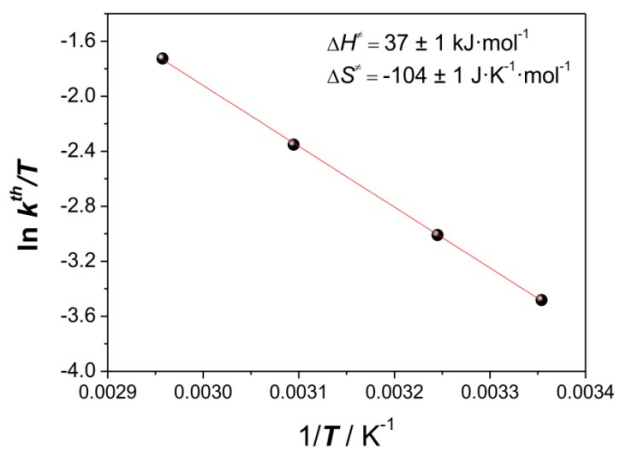


**Figure S5.** Spectral change for compound 1 after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532 \text{ nm}$ , 5 ns pulse width, *ca.* 10 mJ per pulse,  $[1] = 20 \mu\text{M}$ ).

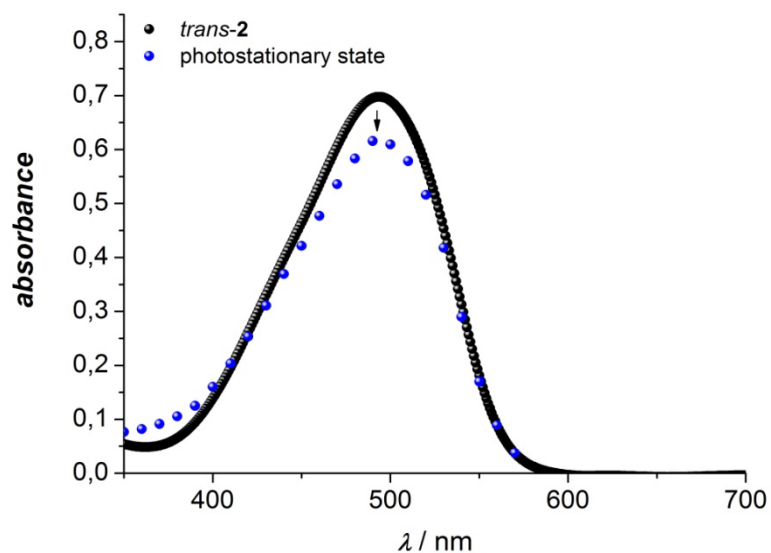
## Compound 2

|                      |     |    |    |    |
|----------------------|-----|----|----|----|
| $T / ^\circ\text{C}$ | 25  | 35 | 50 | 65 |
| $\tau / \text{ms}$   | 109 | 66 | 32 | 17 |

**Table S2.** Relaxation time for *cis*-2 in ethanol at different temperatures.



**Figure S6.** Eyring plot for the thermal back reaction of *cis*-2.

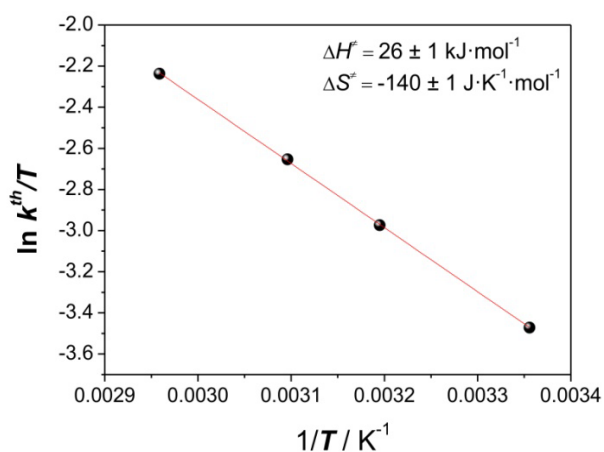


**Figure S7.** Spectral change for compound **2** after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532 \text{ nm}$ , 5 ns pulse width, *ca.* 10 mJ per pulse,  $[\mathbf{2}] = 20 \mu\text{M}$ ).

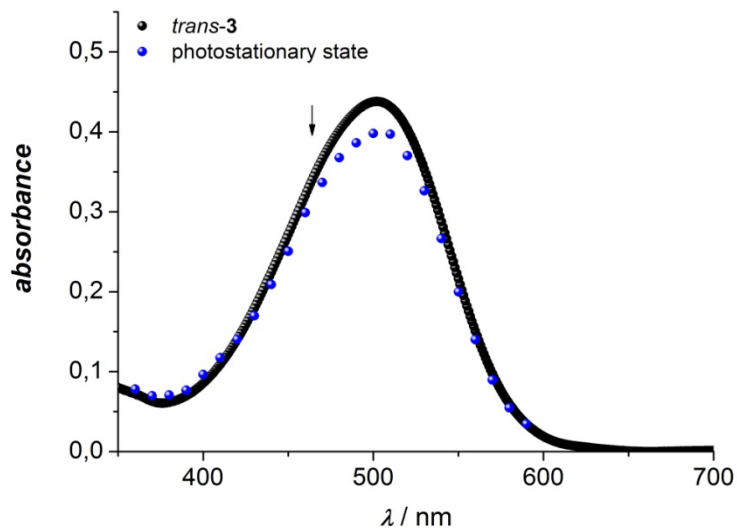
### Compound 3

|                      |     |    |    |    |
|----------------------|-----|----|----|----|
| $T / ^\circ\text{C}$ | 25  | 40 | 50 | 65 |
| $\tau / \text{ms}$   | 108 | 62 | 44 | 28 |

**Table S3.** Relaxation time for *cis*-3 in ethanol at different temperatures.



**Figure S8.** Eyring plot for the thermal back reaction of *cis*-3.

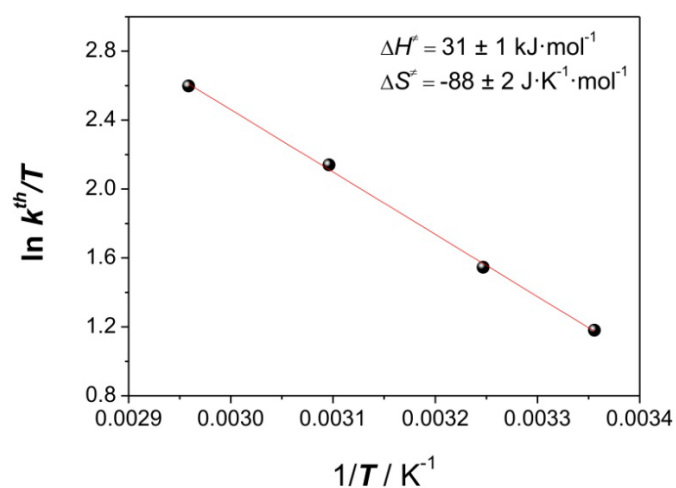


**Figure S9.** Spectral change for compound 3 after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532 \text{ nm}$ , 5 ns pulse width, *ca.* 10 mJ per pulse,  $[3] = 20 \mu\text{M}$ ).

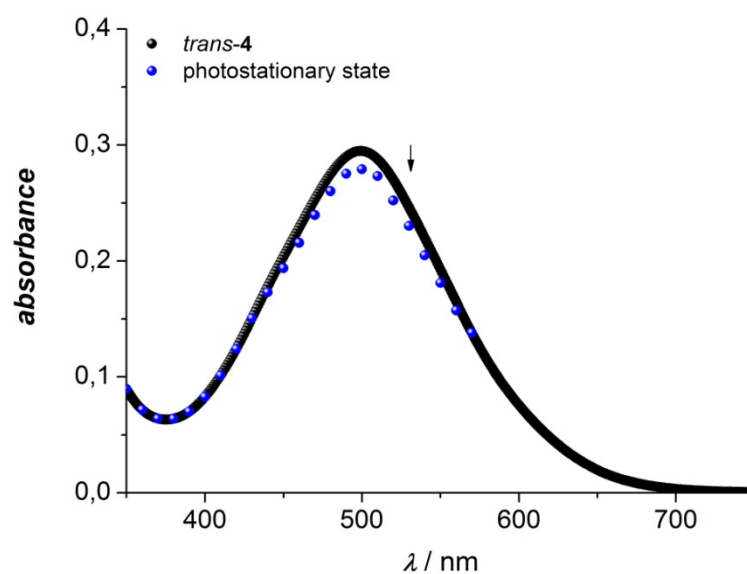
# Compound 4

|                      |     |      |      |      |
|----------------------|-----|------|------|------|
| $T / ^\circ\text{C}$ | 25  | 35   | 50   | 65   |
| $\tau / \text{ms}$   | 1.0 | 0.66 | 0.35 | 0.21 |

**Table S4.** Relaxation time for *cis*-4 in ethanol at different temperatures.



**Figure S10.** Eyring plot for the thermal back reaction of *cis*-4.

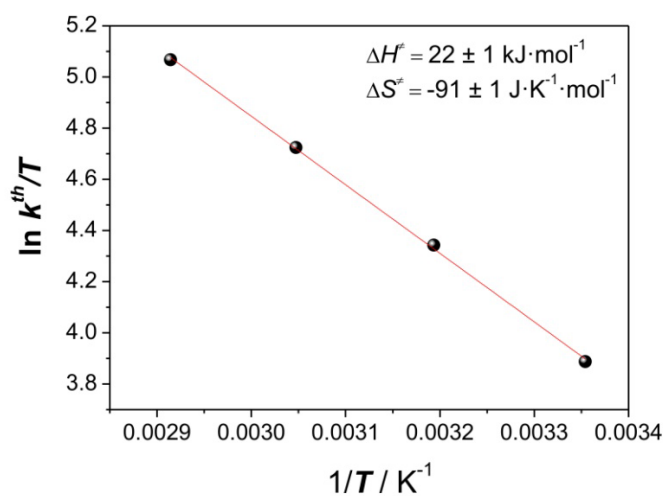


**Figure S11.** Spectral change for compound **4** after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532$  nm, 5 ns pulse width, *ca.* 10 mJ per pulse,  $[\mathbf{4}] = 20$   $\mu\text{M}$ ).

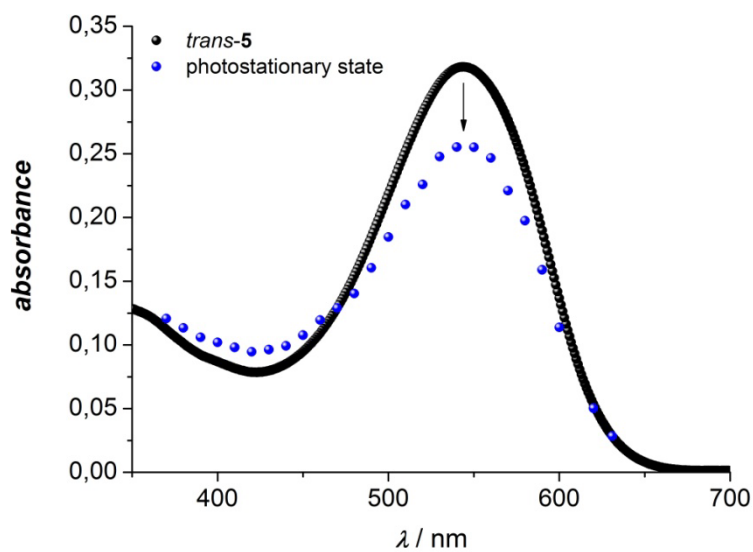
### Compound 5

|                      |    |    |    |    |
|----------------------|----|----|----|----|
| $T / ^\circ\text{C}$ | 25 | 40 | 55 | 70 |
| $\tau / \mu\text{s}$ | 69 | 42 | 27 | 18 |

**Table S5.** Relaxation time for *cis*-**5** in ethanol at different temperatures.



**Figure S12.** Eyring plot for the thermal back reaction of *cis*-**5**.

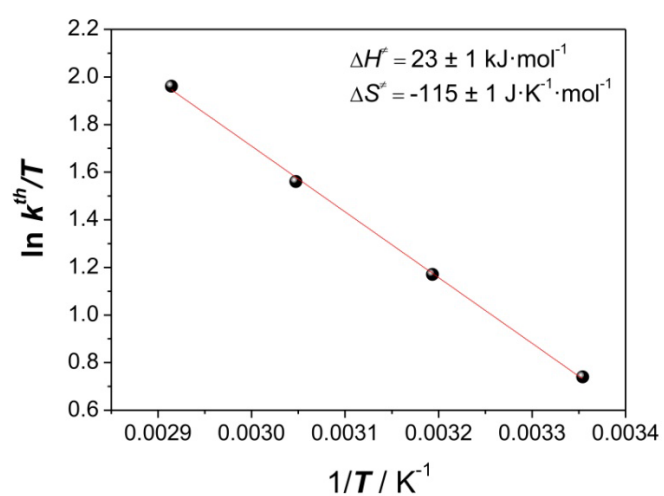


**Figure S13.** Spectral change for compound **5** after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532$  nm, 5 ns pulse width, *ca.* 10 mJ per pulse,  $[\mathbf{5}] = 20$   $\mu\text{M}$ ).

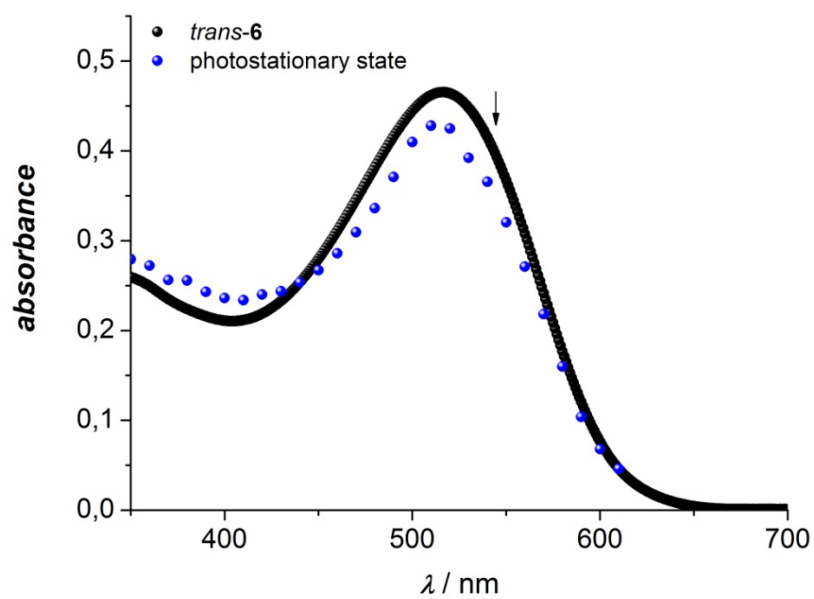
### Compound 6

| $T / ^\circ\text{C}$ | 25   | 40   | 55   | 70   |
|----------------------|------|------|------|------|
| $\tau / \text{ms}$   | 1.60 | 0.99 | 0.64 | 0.41 |

**Table S6.** Relaxation time for *cis*-**6** in ethanol at different temperatures.



**Figure S14.** Eyring plot for the thermal back reaction of *cis*-**6**.



**Figure S15.** Spectral change for compound **6** after *trans*-to-*cis* photoisomerisation with a green-light laser pulse in ethanol at 298 K ( $\lambda_{\text{irrad}} = 532$  nm, 5 ns pulse width, *ca.* 10 mJ per pulse,  $[\mathbf{6}] = 20$   $\mu\text{M}$ ).