

Copper-mediated Tandem Reaction through Isocyanides N-H Bond Insertion: Efficient Access to Unsymmetrical Tetrasubstituted Ureas

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Supporting Information

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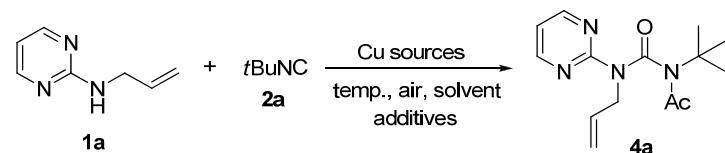
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1. General Information

All reagents and metal catalysts were obtained from commercial sources without further purification, and commercially available solvents were purified before use. All new compounds were fully characterized. All melting points were taken on a WRS-1A or a WRS-1B Digital Melting Point Apparatus without correction. Infrared spectra were obtained using an AVATAR 370 FT-IR spectrometer. ^1H , ^{13}C , and ^{19}F NMR spectra were recorded with a Bruker AV-500 spectrometer operating at 500 MHz and 125 MHz, respectively, with chemical shift values being reported in ppm relative to chloroform ($\delta = 7.26$ ppm), dimethyl sulfoxide ($\delta = 2.50$ ppm), acetone ($\delta = 2.05$ ppm) or TMS ($\delta = 0.00$ ppm) for ^1H NMR; chloroform ($\delta = 77.16$ ppm), acetone ($\delta = 29.84$ and 206.26 ppm) or dimethyl sulfoxide ($\delta = 39.52$ ppm) for ^{13}C NMR; and C_6F_6 ($\delta = -164.9$ ppm) for ^{19}F NMR. Mass spectra and high resolution mass spectra were recorded with an Agilent 5975N using an Electron impact (EI) or Electrospray ionization (ESI) techniques. Elemental analyses were carried out on an Elementar Vario EL elemental analyzer. Silica gel plate GF254 were used for thin layer chromatography (TLC) and silica gel H or 300-400 mesh were used for flash column chromatography. Yields refer to chromatographically and spectroscopically pure compounds, unless otherwise indicated.

2. Screening of the Reaction Conditions

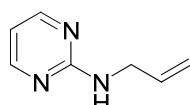
Table S1. Optimization of reaction conditions^a



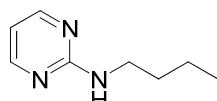
Entry	Cu sources	Solvent	Time [h]	Temp. [°C]	Yield [%] ^b
1	CuCl	toluene	36	110	n.d.
2	Cu(OAc) ₂	toluene	58	110	30 ^c
3	Cu(OAc) ₂	toluene	35	110	67
4	Cu(OAc) ₂ ·H ₂ O	toluene	27	110	65
5	Cu(OAc) ₂ ·H ₂ O	toluene	47	110	51 ^d
6	Cu(OAc) ₂ ·H ₂ O	PhCl	24	110	32
7	Cu(OAc) ₂ ·H ₂ O	dioxane	35	110	trace
8	Cu(OAc) ₂ ·H ₂ O	DMF	35	110	trace
9	Cu(OAc) ₂ ·H ₂ O	toluene	59	100	59
10	Cu(OAc) ₂ ·H ₂ O	toluene	36	120	48
11	Cu(OAc) ₂ ·H ₂ O	toluene	24	110	19 ^e
12	Cu(OAc) ₂ ·H ₂ O	toluene	40	110	16 ^f
13	Cu(OAc) ₂ ·H ₂ O	toluene	39	110	trace ^g
14	--	toluene	24	110	n.d.
15	--	toluene	12	110	n.d. ^h

^a Reaction conditions: **1a** (0.3 mmol), **2a** (0.9 mmol), Cu source (0.6 mmol), 110 °C, air, solvent (1.5 mL), dried through a calcium chloride tube. n.d. = not detected. ^b Isolated yield. ^c Cu(OAc)₂·H₂O (1.5 equiv). ^d Cu(OAc)₂·H₂O (2.5 equiv). ^e N₂. ^f O₂. ^g Cu(OAc)₂·H₂O (0.2 equiv), NaOAc (2.0 equiv). ^h Pd(OAc)₂ (5 mol %), NaOAc (3.0 equiv).

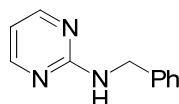
3. Synthesis and characterization for starting materials



Allyl-pyrimidin-2-yl-amine (1a)^[1]: To an oven-dried flask containing 2-chloro-pyrimidine (1.145 g, 10 mmol) was added allylamine (2.5 mL). The reaction mixture was stirred under reflux for 3 h and monitored by TLC. Upon completion, the mixture was washed with 10% NaOH aqueous solution and extracted with Et₂O (3×30 mL). The combined organic phase was washed with brine and dried over Na₂SO₄. After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **1a** as yellow liquid (1.318 g, 97%). IR (KBr, cm⁻¹): 3264, 1592, 1411, 1362, 919, 801; ¹H NMR (CDCl₃, 500 MHz): δ 8.25 (d, *J* = 5.0 Hz, 2H), 6.50 (t, *J* = 5.0 Hz, 1H), 5.98-5.91 (m, 2H), 5.26-5.22 (m, 1H), 5.13-5.10 (m, 1H), 4.06-4.04 (m, 2H); ¹³C NMR (CDCl₃, 125 MHz): δ 162.1, 158.0, 134.9, 115.7, 110.5, 43.7; LC-MS (ESI) m/z 136.1 [M⁺H].

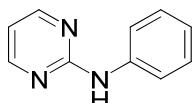


Butyl-pyrimidin-2-yl-amine (1b)^[2]: To an oven-dried flask containing 2-chloro-pyrimidine (2.2 g, 19.2 mmol), butylamine (1.4 g, 19.2 mmol) and triethylamine (6.0 mL) were added in *n*-butanol (40 mL) and the mixture was stirred at 110 °C for 18 h and monitored by TLC. Upon completion, the reaction mixture was extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with brine and dried over Na₂SO₄. After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **1b** as yellow liquid (2.3 g, 79%). IR (KBr, cm⁻¹): 3444, 3275, 2871, 1592, 801, 640; ¹H NMR (CDCl₃, 500 MHz): δ 8.26 (s, 2H), 6.51 (t, *J* = 5.0 Hz, 1H), 5.46 (s, 1H), 3.42-3.38 (m, 2H), 1.62-1.56 (m, 2H), 1.45-1.37 (m, 2H), 0.94 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 162.3, 157.9, 110.2, 41.2, 31.7, 20.1, 13.8; LC-MS (ESI) m/z 152.1 [M⁺H].

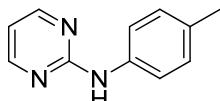


Benzyl-pyrimidin-2-yl-amine (1c)^[2]: To an oven-dried flask containing 2-chloro-pyrimidine (230.0 mg, 2.0 mmol), benzylamine (107.0 mg, 1.0 mmol) and triethylamine (101.0 mg, 1.0 mmol) were added in ethanol (2 mL) and the mixture was stirred at 78 °C for 24 h and monitored by TLC. Upon completion, the reaction mixture was extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with brine and dried over Na₂SO₄. After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **1c** as yellow solid (140.6 mg, 76%). M.p. 72-74 °C; IR (KBr, cm⁻¹): 3241, 2874, 1592, 1261, 667, 537; ¹H NMR (CDCl₃, 500 MHz): δ 8.28 (s, 2H), 7.37-7.32 (m, 4H), 7.29-7.26 (m, 1H), 6.58 (t, *J* = 5.0 Hz, 1H), 6.11 (s, 1H), 4.66 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (CDCl₃, 125 MHz): δ 162.1, 158.0, 139.0, 128.6,

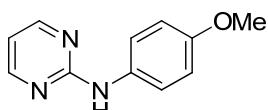
127.6, 127.3, 110.6, 45.5; LC-MS (ESI) m/z 186.1 [M⁺H].



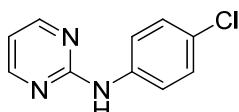
Phenyl-pyrimidin-2-yl-amine (1d)^[3]: To an oven-dried flask containing aniline (6.975 g, 75 mmol), 2-chloro-pyrimidine (5.725 g, 50 mmol) and acetic acid (50 mL) in 1,4-dioxane (130 mL) was added. The reaction mixture was stirred at 110 °C for 23 h and monitored by TLC. Upon completion, the mixture was extracted with CH₂Cl₂ (3×100 mL). The combined organic phase was washed with brine and dried over Na₂SO₄. After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel and then recrystallization to give **1d** as white solid (7.006 g, 82%). M.p. 106-108 °C; IR (KBr, cm⁻¹): 3441, 3258, 1613, 1252, 751, 640; ¹H NMR (CDCl₃, 500 MHz): δ 8.43 (d, J = 4.8 Hz, 2H), 7.71 (br, 1H), 7.62 (d, 2H), 7.35 (t, J = 8.2 Hz, 2H), 7.07 (t, J = 7.4 Hz, 1H), 6.73 (t, J = 4.8 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 160.2, 158.0, 139.4, 129.0, 122.9, 119.8, 112.4; EI-MS m/z (%): 171 (46) [M⁺], 170 (100).



Pyrimidin-2-yl-p-tolyl-amine (1e)^[4]: Following the same procedure as for **1d** with 4-methylaniline (321.0 mg, 3 mmol), 2-chloro-pyrimidine (520.0 mg, 4.5 mmol), and acetic acid (270 mg, 4.5 mmol) in 1,4-dioxane (6.0 mL) for 9 h to give **1e** as yellow solid (433.4 mg, 78%). M.p. 124-126 °C; IR (KBr, cm⁻¹): 3443, 3259, 1617, 1252, 794, 640, 507; ¹H NMR (CDCl₃, 500 MHz): δ 8.40 (d, J = 4.5 Hz, 2H), 7.95 (s, 1H), 7.49 (d, J = 8.5 Hz, 2H), 7.16 (d, J = 8.0 Hz, 2H), 6.67 (t, J = 4.5 Hz, 1H), 2.33 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 160.4, 158.0, 136.7, 132.6, 129.5, 120.3, 112.1, 20.9; EI-MS m/z (%): 185 (58) [M⁺], 184 (100).

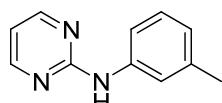


(4-Methoxy-phenyl)-pyrimidin-2-yl-amine (1f)^[4]: Following the same procedure as for **1d** with 4-methoxyaniline (370.0 mg, 3.0 mmol), 2-chloro-pyrimidine (520.0 mg, 4.5 mmol), and acetic acid (270 mg, 1.5 mmol) in 1,4-dioxane (6.0 mL) for 18 h to give **1f** as orange-red solid (509.0 mg, 84%). M.p. 123-124 °C; IR (KBr, cm⁻¹): 3442, 3258, 1617, 1424, 1243, 560; ¹H NMR (CDCl₃, 500 MHz): δ 8.37 (d, J = 4.5 Hz, 2H), 7.95 (br, 1H), 7.48 (d, J = 8.4 Hz, 2H), 7.16 (d, J = 8.2 Hz, 2H), 6.68 (t, J = 4.8 Hz, 1H), 3.80 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 160.6, 158.0, 155.9, 132.3, 122.4, 114.2, 111.9, 55.6; LC-MS (ESI) m/z 202.1 [M⁺H].

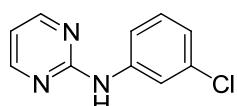


(4-Chloro-phenyl)-pyrimidin-2-yl-amine (1g)^[4]: Following the same procedure as for **1d** with *p*-chloroaniline (765.4 mg, 6.0 mmol), 2-chloro-pyrimidine (572.5 mg, 5.0 mmol), and acetic acid (300.3 mg, 5.0 mmol) in 1,4-dioxane (12.0 mL) for 29 h to give **1g** as yellow solid (849.3 mg, 83%). M.p. 172-174 °C; IR (KBr, cm⁻¹): 3455, 3259, 1616, 1424, 790, 464; ¹H NMR (CDCl₃, 500

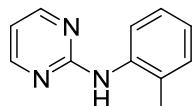
MHz): δ 8.43 (d, J = 4.8 Hz, 2H), 7.62 (br, 1H), 7.58 (d, J = 8.7 Hz, 2H), 7.30 (d, J = 8.7 Hz, 2H), 6.76 (t, J = 4.8 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.7, 158.0, 137.9, 128.9, 127.6, 120.7, 112.8; EI-MS m/z (%): 207 (18) [$\text{M}^+ (^{37}\text{Cl})$], 205 (58) [$\text{M}^+ (^{35}\text{Cl})$], 204 (100).



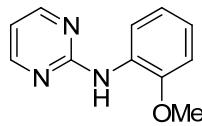
Pyrimidin-2-yl-*m*-tolyl-amine (1h)^[4]: Following the same procedure as for **1d** with 3-methylaniline (385.7 mg, 3.6 mmol), 2-chloro-pyrimidine (343.5 mg, 3.0 mmol), and acetic acid (180.0 mg, 3.0 mmol) in 1,4-dioxane (7.2 mL) for 18 h to give **1h** as yellow solid (245.0 mg, 44%). M.p. 80-82 °C; IR (KBr, cm^{-1}): 3314, 1614, 1254, 798, 519, 443; ^1H NMR (CDCl_3 , 500 MHz): δ 8.43 (d, J = 5.0 Hz, 2H), 7.49 (br, 1H), 7.44-7.42 (m, 2H), 7.23 (t, J = 8.0 Hz, 1H), 6.89 (d, J = 8.0 Hz, 1H), 6.72 (t, J = 5.0 Hz, 1H), 2.37 (s, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 160.3, 158.0, 139.3, 138.8, 128.8, 123.8, 120.4, 116.9, 112.3, 21.7; LC-MS (ESI) m/z 186.1 [M^+H].



(3-Chloro-phenyl)-pyrimidin-2-yl-amine (1i)^[4]: Following the same procedure as for **1d** with *m*-chloroaniline (765.0 mg, 6.0 mmol), 2-chloro-pyrimidine (572.5 mg, 5.0 mmol), and acetic acid (300.3 mg, 5.0 mmol) in 1,4-dioxane (12 mL) for 24 h to give **1i** as yellow solid (906.5 mg, 88%). M.p. 106-108 °C; IR (KBr, cm^{-1}): 3445, 1617, 1432, 995, 793, 640; ^1H NMR (CDCl_3 , 500 MHz): δ 8.45 (d, J = 4.5 Hz, 2H), 7.86 (s, 1H), 7.73 (br, 1H), 7.39 (d, J = 8.0 Hz, 1H), 7.25 (t, J = 7.5 Hz, 1H), 7.02 (d, J = 8.0 Hz, 1H), 6.79 (t, J = 4.5 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.8, 158.0, 140.8, 134.6, 129.9, 122.5, 119.4, 117.4, 113.0; LC-MS (ESI) m/z 208.1 [$\text{M}^+\text{H} (^{37}\text{Cl})$], 206.1 [$\text{M}^+\text{H} (^{35}\text{Cl})$].

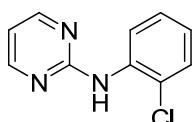


Pyrimidin-2-yl-*o*-tolyl-amine (1j)^[3]: Following the same procedure as for **1d** with *o*-methylaniline (642.9 mg, 6.0 mmol), 2-chloro-pyrimidine (572.5 mg, 5.0 mmol), and acetic acid (300.3 mg, 5.0 mmol) in 1,4-dioxane (12 mL) for 25 h to give **1j** as white solid (493.3 mg, 53%). M.p. 88-90 °C; IR (KBr, cm^{-1}): 3444, 3230, 1583, 1257, 752, 639; ^1H NMR (CDCl_3 , 500 MHz): δ 8.40 (d, J = 5.0 Hz, 2H), 7.89 (d, J = 8.0 Hz, 1H), 7.22-7.26 (m, 3H), 7.07 (t, J = 7.4 Hz, 1H), 6.72 (t, J = 5.0 Hz, 1H), 2.33 (s, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 160.6, 158.1, 137.1, 130.6, 129.8, 126.7, 124.3, 122.7, 122.2, 18.2; LC-MS (ESI) m/z 186.1 [M^+H].

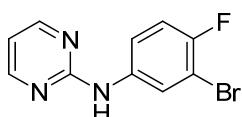


(2-Methoxy-phenyl)-pyrimidin-2-yl-amine (1k)^[3]: Following the same procedure as for **1d** with *o*-methoxyaniline (738.9 mg, 6.0 mmol), 2-chloro-pyrimidine (572.5 mg, 5.0 mmol), and acetic acid (300.3 mg, 5.0 mmol) in 1,4-dioxane (12 mL) for 18 h to give **1k** as yellow solid (382.7 mg, 38%). M.p. 54-56 °C; IR (KBr, cm^{-1}): 3385, 2998, 1581, 1245, 749, 583; ^1H NMR (CDCl_3 , 500 MHz): δ 8.42-8.45 (m, 3H), 8.05 (br, 1H), 7.00-7.01 (m, 2H), 6.90-6.92 (m, 1H), 6.73 (t, J = 5.0,

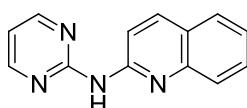
1H), 3.91 (s, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.9, 157.9, 148.0, 129.0, 122.0, 120.9, 118.9, 112.3, 110.0, 55.7; LC-MS (ESI) m/z 202.1 [M^+H].



(2-Chloro-phenyl)-pyrimidin-2-yl-amine (1l)^[4]: To an oven-dried flask containing 2-phenyl-pyrimidine (872.2 mg, 5.0 mmol), *N*-chlorosuccinimide (815.8 mg, 6.0 mg), $\text{Pd}(\text{OAc})_2$ (59.9 mg, 0.25 mmol) were added in HOAc (50 mL). The reaction mixture was stirred at 110 °C for 17 h and monitored by TLC. Upon completion, the reaction mixture was extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with brine and dried over Na_2SO_4 . After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel and then recrystallization to give **1l** as white solid (404.4 mg, 39%). M.p. 94-96 °C; IR (KBr, cm^{-1}): 3445, 3226, 1596, 1289, 798, 600; ^1H NMR (CDCl_3 , 500 MHz): δ 8.47-8.46 (m, 3H), 7.85 (br, 1H), 7.40 (d, J = 7.0 Hz, 1H), 7.30 (t, J = 8.0 Hz, 1H), 7.00 (t, J = 5.0 Hz, 1H), 6.81 (t, J = 5.0 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.7, 158.0, 136.0, 129.2, 127.4, 122.9, 122.7, 120.5, 113.3; LC-MS (ESI) m/z 208.1 [M^+H (^{37}Cl)], 206.1 [M^+H (^{35}Cl)].

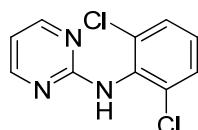


(3-Bromo-4-fluoro-phenyl)-pyrimidin-2-yl-amine (1m): Following the same procedure as for **1d** with 3-bromo-4-fluoro-phenylamine (912.1 mg, 4.8 mmol), 2-chloro-pyrimidine (458.0 mg, 4.0 mmol), and acetic acid (24.2 mg, 4.0 mmol) in 1,4-dioxane (9.6 mL) for 23 h to give **1m** as white solid (701.9 mg, 65%). M.p. 156-158 °C; IR (KBr, cm^{-1}): 3440, 3263, 1614, 1253, 798, 556 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz): δ 8.44 (d, J = 4.5 Hz, 2H), 7.98 (dd, $^4J_{\text{F}-\text{H}} = 6.0$ Hz, $^4J_{\text{H}-\text{H}} = 2.5$ Hz, 1H), 7.74 (br, 1H), 7.45-7.42 (m, 1H), 7.08 (t, J = 8.5 Hz, 1H), 6.78 (t, J = 5.0, 1H); ^{19}F NMR (CDCl_3 , 470 MHz): δ -115.0 (m, Ar-F); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.5, 158.0, 155.0 (d, $^1J_{\text{C}-\text{F}} = 237.5$ Hz), 136.2 (d, $^4J_{\text{C}-\text{F}} = 3.8$ Hz), 124.4, 120.1 (d, $^3J_{\text{C}-\text{F}} = 6.3$ Hz), 116.3 (d, $^2J_{\text{C}-\text{F}} = 23.8$ Hz), 112.9, 108.9 (d, $^2J_{\text{C}-\text{F}} = 21.3$ Hz); EI-MS m/z (%): 269 (62) [$\text{M}^+({}^{81}\text{Br})$], 267 (64) [$\text{M}^+({}^{79}\text{Br})$]; HRMS (EI) calcd for $\text{C}_{10}\text{H}_6\text{BrFN}_3$ [M-H]⁺ 265.9729, found 265.9728.

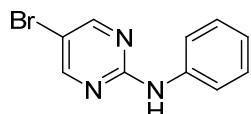


Pyrimidin-2-yl-quinolin-2-yl-amine (1n)^[5]: To an oven-dried flask containing 2-chloroquinoline (589.0 mg, 3.6 mmol), 2-aminopyrimidine (285.3 mg, 3.0 mg), $\text{Pd}_2(\text{dba})_3$ (41.2 mg, 0.045 mmol), DPPF (49.9 mg, 0.09 mmol) and *t*-BuOK (403.9 mg, 3.6 mmol) were added in toluene (30 mL). The reaction mixture was stirred at 80 °C for 27 h and monitored by TLC. Upon completion, the mixture was extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with brine and dried over Na_2SO_4 . After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel and then recrystallization to give **1n** as white solid (85 mg, 13%). M.p. 180-182 °C; IR (KBr, cm^{-1}): 3444, 3038, 1610, 1503, 1449, 1427, 1334, 817, 737; ^1H NMR (CDCl_3 , 500 MHz): δ 9.48 (br, 1H), 8.69-8.67 (m, 3H), 8.14 (d, J = 9.0 Hz, 1H), 7.87 (d, J = 8.0

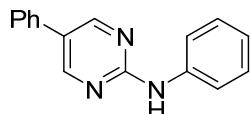
Hz, 1H), 7.75 (d, J = 8.0 Hz, 1H), 7.64 (td, J = 7.5, 1.5 Hz, 1H), 7.39 (td, J = 7.5, 0.5 Hz, 1H), 6.88 (t, J = 4.5 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 159.5, 158.2, 152.4, 138.0, 129.8, 127.5, 127.1, 125.4, 124.2, 114.1, 113.8; LC-MS (ESI) m/z 223.1 [M^+H].



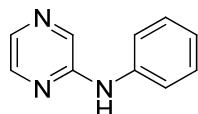
(2,6-Dichloro-phenyl)-pyrimidin-2-yl-amine (1o)^[6]: Following the same procedure as for **1l** with 2-phenyl-pyrimidine (872.2 mg, 5.0 mmol), *N*-chlorosuccinimide (815.8 mg, 6.0 mg), $\text{Pd}(\text{OAc})_2$ (59.9 mg, 0.25 mmol) in HOAc (50 mL) for 17 h to give **1o** as white solid (244.0 mg, 20%). M.p. 172-174 °C; IR (KBr, cm^{-1}): 3453, 3063, 1589, 1519, 1445, 1413, 1250, 784; ^1H NMR (CDCl_3 , 500 MHz): δ 8.37 (d, J = 5.0 Hz, 2H), 7.98 (br, 1H), 7.40 (d, J = 8.0 Hz, 2H), 7.16 (t, J = 8.5, 1H), 6.71 (t, J = 4.5 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 160.6, 158.2, 134.2, 133.9, 128.6, 127.8, 112.7; EI-MS m/z (%): 241 (1) [$\text{M}^+ ({}^{37}\text{Cl}, {}^{35}\text{Cl})$], 239 (3) [$\text{M}^+ (2 \times {}^{35}\text{Cl})$], 204 (100).



(5-Bromo-pyrimidin-2-yl)-phenyl-amine (1p)^[7]: To an oven-dried flask containing 5-bromo-2-chloro-pyrimidine (580.2 mg, 3.0 mmol), aniline (279.6 mg, 3.0 mmol) in *n*-butanol (3 mL) were added. The reaction mixture was stirred at 110 °C for 21 h and monitored by TLC. Upon completion, the mixture was filtered through a thin pad of celite, and the residue was purified by recrystallization to give **1p** as yellow solid (418.2 mg, 56%). M.p. 122-124 °C; IR (KBr, cm^{-1}): 3442, 3264, 1608, 1574, 1529, 1448, 1430, 753; ^1H NMR (d_6 -Acetone, 500 MHz): δ 8.89 (br, 1H), 8.53 (s, 2H), 7.81 (d, J = 7.5 Hz, 2H), 7.34 (t, J = 8.0 Hz, 2H), 7.03 (t, J = 7.5 Hz, 1H); ^{13}C NMR (d_6 -Acetone, 125 MHz): δ 158.8, 158.1, 140.0, 128.6, 122.2, 119.2, 107.8; LC-MS (ESI) m/z 252.0 [$\text{M}^+\text{H} ({}^{81}\text{Br})$], 250.0 [$\text{M}^+\text{H} ({}^{79}\text{Br})$].

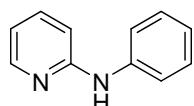


Phenyl-(5-phenyl-pyrimidin-2-yl)-amine (1q)^[8]: Following the same procedure as for **1p** with 2-chloro-5-phenyl-pyrimidine (190.6 mg, 1.0 mmol) and aniline (186.4 mg, 2.0 mmol) in *n*-butanol (1.0 mL) for 18 h to give **1q** as white solid (137.3 mg, 56%). M.p. 166-168 °C; IR (KBr, cm^{-1}): 3453, 3269, 3029, 1619, 1529, 1451, 756, 698; ^1H NMR (CDCl_3 , 500 MHz): δ 8.67 (s, 2H), 7.74 (br, 1H), 7.66 (d, J = 8.0 Hz, 2H), 7.53-7.47 (m, 4H), 7.41-7.36 (m, 3H), 7.09 (t, J = 7.0 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 158.8, 155.9, 139.0, 134.8, 129.3, 129.1, 127.9, 126.1, 125.7, 123.1, 119.7; EI-MS m/z (%): 247 (64) [M^+], 246 (100).

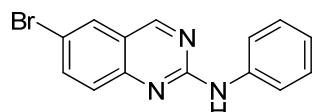


Phenyl-pyrazin-2-yl-amine (1r)^[9]: Following the same procedure as for **1p** with 2-chloropyrazine (343.5 mg, 3.0 mmol) and aniline (279.6 mg, 3.0 mmol) in *n*-butanol (3.0 mL) for 52 h to give **1r** as yellow solid (245.4 mg, 48%). M.p. 130-132 °C; IR (KBr, cm^{-1}): 3473, 3283,

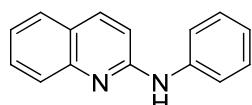
3101, 1626, 1584, 1521, 1498, 1352, 752; ^1H NMR (CDCl_3 , 500 MHz): δ 8.25 (s, 1H), 8.10 (d, J = 1.5 Hz, 1H), 7.96 (d, J = 3.0 Hz, 1H), 7.43 (d, J = 8.5 Hz, 2H), 7.35 (t, J = 8.0 Hz, 2H), 7.10 (m, 2H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 152.4, 141.9, 139.2, 134.7, 133.0, 129.4, 123.7, 120.4; EI-MS m/z (%): 171 (72) [M^+], 170 (100).



Phenyl-pyridin-2-yl-amine (1s)^[3]: To an oven-dried flask containing 2-bromopyridine (4.74 g, 30.0 mmol), aniline (2.794 g, 3.0 mmol) was added. The reaction mixture was stirred at 160 °C for 7 h and monitored by TLC. Upon completion, saturate NaHCO_3 was added, and the reaction mixture was extracted with ethyl acetate (3×30 mL). The combined organic phase was washed with brine and dried over Na_2SO_4 . After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **1s** as white solid (4.954 g, 97%). M.p. 106-108 °C; IR (KBr, cm^{-1}): 3450, 3224, 1590, 1574, 1464, 1444, 1327, 770, 670; ^1H NMR (CDCl_3 , 500 MHz): δ 8.17 (d, J = 4.0 Hz, 1H), 7.51 (td, J = 5.5, 2.0 Hz, 1H), 7.36-7.32 (m, 4H), 7.09-7.05 (m, 1H), 6.98 (s, 1H), 6.89 (d, J = 8.5 Hz, 1H), 6.75-6.73 (m, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 156.1, 148.2, 140.5, 137.9, 129.3, 122.9, 120.5, 114.9, 108.2; LC-MS (ESI) m/z 171.1 [M^+H].



(6-Bromo-quinazolin-2-yl)-phenyl-amine (1t)^[10]: Following the same procedure as for **1d** with aniline (139.7 mg, 1.5 mmol), 2,6-dibromoquinazoline (287.9 mg, 1.0 mmol), and acetic acid (60.1 mg, 1.0 mmol) in 1,4-dioxane (3.0 mL) for 22 h to give **1t** as yellow solid (208.0 mg, 70%). M.p. 154-156 °C; IR (KBr, cm^{-1}): 3442, 3279, 1601, 1583, 1543, 1446, 1353, 747; ^1H NMR (CDCl_3 , 500 MHz): δ 9.02 (s, 1H), 7.88 (d, J = 2.0 Hz, 1H), 7.82-7.79 (m, 3H), 7.63 (d, J = 9.0 Hz, 1H), 7.55 (br, 1H), 7.39 (t, J = 8.0 Hz, 2H), 7.10 (t, J = 7.0 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 160.9, 156.8, 150.1, 139.2, 137.7, 129.5, 129.0, 128.0, 123.0, 121.7, 119.3, 116.5; EI-MS m/z (%): 301 (1) [$\text{M}^+ ({}^{81}\text{Br})$], 299 (1) [$\text{M}^+ ({}^{79}\text{Br})$], 57 (100).



Phenyl-quinolin-2-yl-amine (1u)^[9]: To an oven-dried flask containing 2-chloroquinoline (327.2 mg, 2.0 mmol), aniline (186.2 mg, 2.0 mmol) were added in ethanol (10 mL). The reaction mixture was stirred under reflux for 10 h and monitored by TLC. Upon completion, the reaction mixture was extracted with ethyl acetate (3×10 mL). The combined organic phase was washed with brine and dried over Na_2SO_4 . After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **1u** as white solid (409.2 mg, 93%). M.p. 94-96 °C; IR (KBr, cm^{-1}): 3404, 3050, 1620, 1597, 1533, 1401, 823, 752; ^1H NMR (CDCl_3 , 500 MHz): δ 7.95 (d, J = 9.0 Hz, 1H), 7.79 (d, J = 8.5 Hz, 1H), 7.65 (d, J = 8.0 Hz, 1H), 7.61 (td, J = 8.0, 1.5 Hz, 1H), 7.55 (dd, J = 8.5, 1.0 Hz, 2H), 7.37 (t, J = 8.5 Hz, 2H), 7.31 (td, J = 8.0, 1.0 Hz, 1H), 7.12 (tt, J = 7.0, 1.5 Hz, 1H), 7.00 (d, J = 9.0 Hz, 1H), 2.50 (br, 1H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 154.5, 147.6,

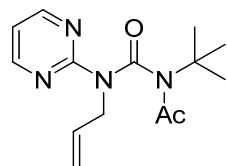
140.2, 137.8, 129.9, 129.3, 127.5, 126.7, 124.2, 123.2, 123.1, 120.6, 111.8; LC-MS (ESI) m/z 221.0 [M⁺H].

4. Synthesis of copper carboxylates

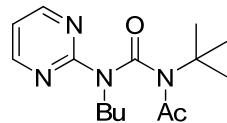
General procedure: To a round-bottom flask, sodium carboxylates was dissolved in water, and aqueous CuSO₄ was added. The reaction mixture was stirred at room temperature for 2 h. Upon completion, the reaction mixture filtered and washed with water and then dried in vacuum to give copper complex as copper (II) laurate monohydrate (**3b**),^[11] copper (II) phenylacetate (**3c**),^[12] copper (II) benzoate monohydrate (**3d**),^[13] copper (II) crotonate monohydrate (**3f**),^[14] and copper (II) phenylpropionate multihydrate (**3g**).^[15]

Copper (II) cinnamate monohydrate (3e**)**^[16]: To a round-bottom flask, A solution of CuCl₂·2H₂O in ethanol was added to a solution of cinnamic acid and Et₃N were added in ethanol. The reaction mixture was stirred at room temperature for 2 h. Upon completion, the reaction mixture filtered and washed with water and then dried in vacuum to give copper (II) cinnamate monohydrate (**3e**).

5. Synthesis and characterization of ureas

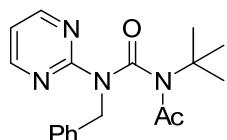


1-Acetyl-3-allyl-1-tert-butyl-3-pyrimidin-2-yl-urea (4a**):** To an oven-dried flask containing **1a** (67.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol) and *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) was added in toluene (3.0 mL). The reaction mixture was stirred at 110 °C for 36 h in the presence of air which was dried through a calcium chloride tube and monitored by TLC. Upon completion, the reaction mixture was washed with dilute ammonia and extracted with CH₂Cl₂ (3×10 mL). The combined organic phase was washed with brine and dried over Na₂SO₄. After that, the solid was filtered off through a thin pad of celite, and the filtrate was evaporated in vacuum to give the crude product which was purified by column chromatography on silica gel to give **4a** as white solid (89.8 mg, 65%). M.p. 50-52 °C; IR (KBr, cm⁻¹): 2979, 2964, 1675, 1568, 1415, 1315, 825; ¹H NMR (CDCl₃, 500 MHz): δ 8.69 (d, *J* = 4.5 Hz, 2H), 7.16 (t, *J* = 4.5 Hz, 1H), 5.96-5.88 (m, 1H), 5.24 (dd, *J* = 17.0, 1.0 Hz, 1H) 5.12 (dd, *J* = 10.0, 1.0 Hz, 1H), 4.68 (m, 2H), 2.18 (s, 3H), 1.23 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 169.9, 160.2, 158.3, 156.4, 132.0, 118.6, 118.4, 58.3, 50.8, 27.6, 25.3; LC-MS (ESI) m/z 299.1 [M⁺Na]; Anal. Calcd. For C₁₄H₂₀N₄O₂: C, 60.85; H, 7.30; N, 20.28. Found: C, 60.71; H, 7.29; N, 20.15.

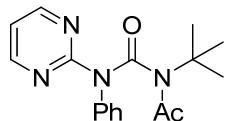


1-Acetyl-3-butyl-1-tert-butyl-3-pyrimidin-2-yl-urea (4b**):** Following the same procedure as for **4a** with **1b** (75.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 66 h to give **4b** as yellow liquid (97.6 mg, 67%). IR (KBr,

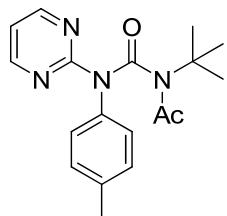
cm^{-1}): 2961, 2933, 1693, 1670, 1565, 1417, 803; ^1H NMR (CDCl_3 , 500 MHz): δ 8.70 (d, $J = 4.5$ Hz, 2H), 7.16 (t, $J = 5.0$ Hz, 1H), 4.06-4.03 (m, 2H), 2.19 (s, 3H), 1.68-1.54 (m, 2H), 1.36-1.28 (m, 2H), 1.21 (s, 9H), 0.88 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 169.9, 160.5, 158.3, 156.4, 118.6, 58.0, 48.5, 29.7, 27.5, 25.3, 20.1, 13.7; LC-MS (ESI) m/z 293.1 [M^+H]; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{24}\text{N}_4\text{NaO}_2$ [M^+Na] 315.1797, found 315.1778.



1-Acetyl-3-benzyl-1-tert-butyl-3-pyrimidin-2-yl-urea (4c)^[17]: Following the same procedure as for **4a** with **1c** (92.6 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 19 h to give **4c** as white solid (105.2 mg, 65%). M.p. 46-48 °C; IR (KBr, cm^{-1}): 2979, 2929, 1694, 1670, 1565, 1416, 1317, 700; ^1H NMR (CDCl_3 , 500 MHz): δ 8.66 (d, $J = 5.0$ Hz, 2H), 7.35-7.31 (m, 2H), 7.26-7.21 (m, 3H), 7.11 (t, $J = 5.0$ Hz, 1H), 5.37 (A of AB, $J = 14.5$ Hz, 1H), 5.26 (B of AB, $J = 15.0$ Hz, 1H), 2.16 (s, 3H), 1.24 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 169.9, 160.1, 158.3, 156.9, 136.3, 128.5, 128.3, 127.6, 118.6, 58.4, 51.6, 27.6, 25.4; LC-MS (ESI) m/z 349.1 [M^+Na]; Anal. Calcd. For $\text{C}_{18}\text{H}_{22}\text{N}_4\text{O}_2$: C, 66.24; H, 6.79; N, 17.17. Found: C, 66.20; H, 6.78; N, 17.28.

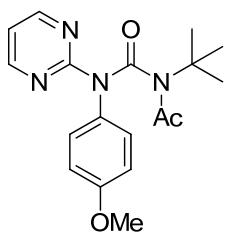


1-Acetyl-1-tert-butyl-3-phenyl-3-pyrimidin-2-yl-urea (4d): Following the same procedure as for **4a** with **1d** (85.5 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 25 h to give **4d** as white solid (140.4 mg, 90%). M.p. 82-83 °C; IR (KBr, cm^{-1}): 2979, 2923, 1704, 1668, 1566, 1512, 1413, 1297, 1021, 763, 734; ^1H NMR (CDCl_3 , 500 MHz): δ 8.73 (d, $J = 5.0$ Hz, 2H), 7.45-7.42 (m, 2H), 7.36-7.33 (m, 3H), 7.22 (t, $J = 5.0$ Hz, 1H), 2.39 (s, 3H), 1.33 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 170.1, 160.8, 158.9, 156.2, 140.4, 129.5, 128.0, 127.0, 119.4, 58.8, 27.7, 25.6; EI-MS m/z (%): 312 (1) [M^+], 170 (100); Anal. Calcd. For $\text{C}_{17}\text{H}_{20}\text{N}_4\text{O}_2$: C, 65.37; H, 6.45; N, 17.94. Found: C, 65.57; H, 6.47; N, 18.14.

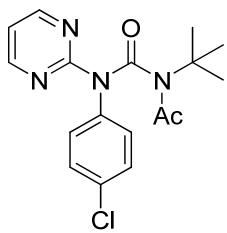


1-Acetyl-1-tert-butyl-3-pyrimidin-2-yl-3-p-tolyl-urea (4e)^[17]: Following the same procedure as for **4a** with **1e** (92.6 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 27 h to give **4e** as yellow solid (156.8 mg, 96%). M.p. 110-111 °C; IR (KBr, cm^{-1}): 3007, 2979, 2924, 1704, 1668, 1565, 1513, 1413, 1298, 763, 734; ^1H NMR (CDCl_3 , 500 MHz): δ 8.72 (d, $J = 5.0$ Hz, 2H), 7.23 (s, 4H), 7.20 (t, $J = 5.0$ Hz, 1H), 2.38 (s, 3H), 2.36 (s, 3H), 1.33 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 170.1, 160.9, 158.8, 156.3, 138.0, 137.8, 130.0, 126.8, 119.2, 58.7, 27.7, 25.6, 21.2; EI-MS m/z (%): 326 (4) [M^+], 184 (100); Anal.

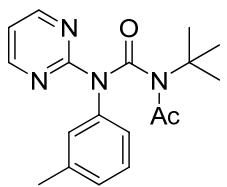
Calcd. For $C_{18}H_{22}N_4O_2$: C, 66.24; H, 6.79; N, 17.17. Found: C, 66.36; H, 6.73; N, 17.27.



1-Acetyl-1-tert-butyl-3-(4-methoxy-phenyl)-3-pyrimidin-2-yl-urea (4f)^[17]: Following the same procedure as for **4a** with **1f** (100.6 mg, 0.5 mmol), $Cu(OAc)_2 \cdot H_2O$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 25 h to give **4f** as white solid (145.2 mg, 85%). M.p. 124-126 °C; IR (KBr, cm^{-1}): 3010, 2977, 2933, 1699, 1672, 1565, 1510, 1411, 1295, 828; 1H NMR ($CDCl_3$, 500 MHz): δ 8.72 (d, J = 5.0 Hz, 2H), 7.28 (AA' of AA'BB', J = 9.5 Hz, 2H), 7.20 (t, J = 5.0 Hz, 1H), 6.93 (BB' of AA'BB', J = 9.0 Hz, 2H), 3.80 (s, 3H), 2.38 (s, 3H), 1.32 (s, 9H); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 170.1, 160.9, 159.0, 158.9, 156.4, 133.0, 128.3, 119.3, 114.7, 58.7, 55.5, 27.7, 25.6; LC-MS (ESI) m/z 365.2 [M^+Na]; Anal. Calcd. For $C_{18}H_{22}N_4O_3$: C, 63.14; H, 6.48; N, 16.36. Found: C, 63.26; H, 6.67; N, 16.23.

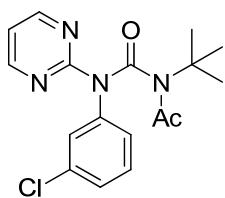


1-Acetyl-1-tert-butyl-3-(4-chloro-phenyl)-3-pyrimidin-2-yl-urea (4g): Following the same procedure as for **4a** with **1g** (102.8 mg, 0.5 mmol), $Cu(OAc)_2 \cdot H_2O$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 20 h to give **4g** as yellow solid (124.9 mg, 72%). M.p. 151-152 °C; IR (KBr, cm^{-1}): 3007, 2979, 2922, 1705, 1673, 1566, 1490, 1416, 1309, 835; 1H NMR ($CDCl_3$, 500 MHz): δ 8.72 (d, J = 5.0 Hz, 2H), 7.38 (AA' of AA'BB', J = 8.5 Hz, 2H), 7.27 (BB' of AA'BB', J = 8.5 Hz, 2H), 7.23 (t, J = 4.5 Hz, 1H), 2.36 (s, 3H), 1.31 (s, 9H); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 170.0, 160.5, 159.0, 156.2, 138.8, 133.8, 129.6, 128.4, 119.6, 58.9, 27.7, 25.6; LC-MS (ESI) m/z 371.1 [M^+Na (^{37}Cl)], 369.1 [M^+Na (^{35}Cl)]; Anal. Calcd. For $C_{17}H_{19}ClN_4O_2$: C, 58.87; H, 5.52; N, 16.15. Found: C, 58.84; H, 5.45; N, 15.85.

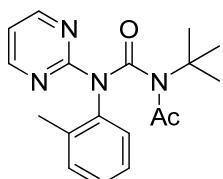


1-Acetyl-1-tert-butyl-3-pyrimidin-2-yl-3-m-tolyl-urea (4h)^[17]: Following the same procedure as for **4a** with **1h** (92.6 mg, 0.5 mmol), $Cu(OAc)_2 \cdot H_2O$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 22 h to give **4h** as white solid (142.3 mg, 87%). M.p. 76-78 °C; IR (KBr, cm^{-1}): 3009, 2968, 2922, 1706, 1673, 1564, 1409, 727; 1H NMR ($CDCl_3$, 500 MHz): δ 8.73 (d, J = 5.0 Hz, 2H), 7.31 (t, J = 7.5 Hz, 1H), 7.21 (t, J = 5.0 Hz, 1H), 7.16-7.12 (m, 3H), 2.39 (s, 3H), 2.36 (s, 3H), 1.33 (s, 9H); ^{13}C NMR ($CDCl_3$, 125 MHz): δ 170.1, 160.9, 158.9, 156.2, 140.2, 139.5, 129.3, 128.9, 127.5, 124.0, 119.4, 58.8, 27.7, 25.6, 21.4; LC-MS (ESI) m/z 349.1

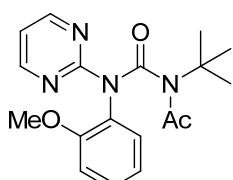
[M⁺Na]; Anal. Calcd. For C₁₈H₂₂N₄O₂: C, 66.24; H, 6.79; N, 17.17. Found: C, 66.50; H, 6.88; N, 17.35.



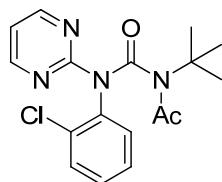
1-Acetyl-1-tert-butyl-3-(3-chloro-phenyl)-3-pyrimidin-2-yl-urea (4i)^[17]: Following the same procedure as for **4a** with **1i** (102.8 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 25 h to give **4i** as yellow solid (160.4 mg, 93%). M.p. 122-124 °C; IR (KBr, cm⁻¹): 2968, 2925, 1709, 1661, 1565, 1411, 1311, 785, 704; ¹H NMR (CDCl₃, 500 MHz): δ 8.74 (d, *J* = 4.5 Hz, 2H), 7.38-7.31 (m, 3H), 7.26-7.24 (m, 2H), 2.37 (s, 3H), 1.32 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.0, 160.4, 159.0, 156.2, 141.3, 134.9, 130.3, 128.2, 127.3, 125.3, 119.7, 58.9, 27.7, 25.6; LC-MS (ESI) m/z 371.1 [M⁺Na (³⁷Cl)], 369.1 [M⁺Na (³⁵Cl)]; Anal. Calcd. For C₁₇H₁₉ClN₄O₂: C, 58.87; H, 5.52; N, 16.15. Found: C, 58.81; H, 5.46; N, 16.01.



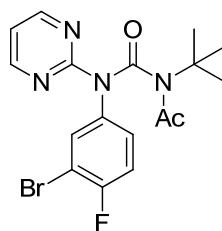
1-Acetyl-1-tert-butyl-3-pyrimidin-2-yl-3-o-tolyl-urea (4j): Following the same procedure as for **4a** with **1j** (92.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 23 h to give **4j** as white solid (129.8 mg, 80%). M.p. 102-104 °C; IR (KBr, cm⁻¹): 3007, 2969, 2925, 1700, 1667, 1566, 1409, 1310, 730; ¹H NMR (CDCl₃, 500 MHz): δ 8.70 (d, *J* = 5.0 Hz, 2H), 7.51 (s, 1H), 7.32-7.26 (m, 3H), 7.17 (t, *J* = 5.0 Hz, 1H), 2.41 (s, 3H), 2.22 (s, 3H), 1.35 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.2, 160.4, 158.6, 156.2, 139.1, 136.2, 131.4, 128.9, 127.0, 118.9, 58.8, 27.9, 25.7, 18.3; LC-MS (ESI) m/z 349.1 [M⁺Na]; Anal. Calcd. For C₁₈H₂₂N₄O₂: C, 66.24; H, 6.79; N, 17.17. Found: C, 66.12; H, 6.75; N, 17.04.



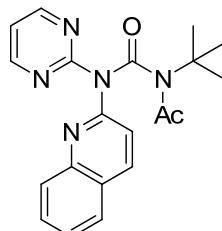
1-Acetyl-1-tert-butyl-3-(2-methoxy-phenyl)-3-pyrimidin-2-yl-urea (4k): Following the same procedure as for **4a** with **1k** (100.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 24 h to give **4k** as white solid (129.1 mg, 76%). M.p. 132-133 °C; IR (KBr, cm⁻¹): 2972, 2928, 1693, 1671, 1564, 1412, 1313, 750; ¹H NMR (CDCl₃, 500 MHz): δ 8.66 (d, *J* = 5.0 Hz, 2H), 7.39 (d, *J* = 8.0 Hz, 1H), 7.35 (t, *J* = 8 Hz, 1H), 7.12 (t, *J* = 4.5 Hz, 1H), 7.04 (t, *J* = 7.5 Hz, 1H), 6.98 (d, *J* = 8.0 Hz, 1H), 3.73 (s, 3H), 2.37 (s, 3H), 1.35 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.2, 160.7, 158.3, 155.9, 155.1, 129.8, 129.3, 129.2, 121.0, 118.5, 112.6, 58.6, 55.8, 27.8, 25.5; LC-MS (ESI) m/z 365.1 [M⁺Na]; Anal. Calcd. For C₁₈H₂₂N₄O₃: C, 63.14; H, 6.48; N, 16.36. Found: C, 63.09; H, 6.49; N, 16.29.



1-Acetyl-1-tert-butyl-3-(2-chloro-phenyl)-3-pyrimidin-2-yl-urea (4l): Following the same procedure as for **4a** with **1l** (102.8 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 21 h to give **4l** as white solid (135.9 mg, 78%). M.p. 132-134 °C; IR (KBr, cm⁻¹): 2980, 2915, 1703, 1679, 1567, 1411, 1304, 732; ¹H NMR (CDCl₃, 500 MHz): δ 8.68 (d, *J* = 5.0 Hz, 2H), 7.60 (d, *J* = 8.0 Hz, 1H), 7.47-7.41 (m, 2H), 7.36 (dt, *J* = 8.0, 1.5 Hz, 1H), 7.16 (t, *J* = 5.0 Hz, 1H), 2.39 (s, 3H), 1.37 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 159.8, 158.5, 158.4, 156.3, 137.8, 132.5, 130.9, 130.6, 129.8, 127.8, 118.7, 58.9, 27.9, 25.7; LC-MS (ESI) m/z 371.1 [M⁺Na (³⁷Cl)], 369.1 [M⁺Na (³⁵Cl)]; Anal. Calcd. For C₁₇H₁₉ClN₄O₂: C, 58.87; H, 5.52; N, 16.15. Found: C, 58.60; H, 5.52; N, 15.92.

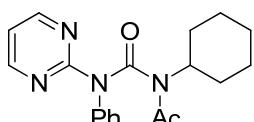


1-Acetyl-3-(3-bromo-4-fluoro-phenyl)-1-tert-butyl-3-pyrimidin-2-yl-urea (4m)^[17]: Following the same procedure as for **4a** with **1m** (142.6 mg, 0.53 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 22 h to give **4m** as white solid (198.0 mg, 91%). M.p. 88-90 °C; IR (KBr, cm⁻¹): 2981, 2925, 1707, 1676, 1566, 1412, 1309, 730; ¹H NMR (CDCl₃, 500 MHz): δ 8.74 (d, *J* = 4.5 Hz, 2H), 7.56 (dd, ⁴J_{F-H} = 6.0, ⁴J_{H-H} = 3.0 Hz, 1H), 7.33-7.30 (m, 1H), 7.26 (t, *J* = 5.0 Hz, 1H), 7.18 (t, *J* = 8.5 Hz, 1H), 2.36 (s, 3H), 1.31 (s, 9H); ¹⁹F NMR (CDCl₃, 470 MHz): δ -107.2 (m, Ar-F); ¹³C NMR (CDCl₃, 125 MHz): δ 169.9, 160.3, 159.0, 158.4 (d, ¹J_{C-F} = 247.5 Hz), 156.3, 136.8 (d, ³J_{C-F} = 3.8 Hz), 132.4, 128.0 (d, ³J_{C-F} = 7.5 Hz), 119.7, 117.0 (d, ²J_{C-F} = 23.8 Hz), 109.5 (d, ²J_{C-F} = 22.5 Hz), 58.9, 27.7, 25.6; LC-MS (ESI) m/z 433.0 [M⁺Na (⁸¹Br)], 431.0 [M⁺Na (⁷⁹Br)]; Anal. Calcd. For C₁₇H₁₈BrFN₄O₂: C, 49.89; H, 4.43; N, 13.69. Found: C, 50.09; H, 4.64; N, 13.77.

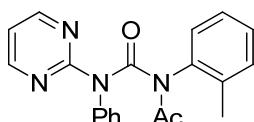


1-Acetyl-1-tert-butyl-3-pyrimidin-2-yl-3-quinolin-2-yl-urea (4n): Following the same procedure as for **4a** with **1n** (77.8 mg, 0.35 mmol), Cu(OAc)₂·H₂O (139.8 mg, 0.7 mmol), *tert*-butyl isocyanide (90 mg, 1.05 mmol) in *m*-xylene at 130 °C with O₂ balloon for 11 h to give **4n** as white solid (39.1 mg, 31%) together with recovered **1n** (34.1 mg, 56% conversion). M.p. 148-150 °C; IR (KBr, cm⁻¹): 2976, 1688, 1670, 1566, 1404, 765; ¹H NMR (CDCl₃, 500 MHz): δ 8.76 (d, *J* = 5.0 Hz, 2H), 8.25 (d, *J* = 8.5 Hz, 1H), 7.82 (d, *J* = 8.0 Hz, 2H), 7.70 (d, *J* = 9.0 Hz,

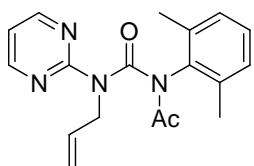
1H), 7.64 (td, $J = 7.0, 1.5$ Hz, 1H), 7.52 (td, $J = 7.5, 1.0$ Hz, 1H), 7.26 (t, $J = 4.5$ Hz, 1H), 2.46 (s, 3H), 1.39 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 170.2, 160.7, 158.8, 156.5, 152.9, 147.0, 138.4, 130.0, 129.0, 127.4, 127.0, 126.9, 119.4, 118.6, 59.4, 27.9, 25.9; LC-MS (ESI) m/z 364.0 [M^+H]; HRMS (ESI) m/z Calcd. for $\text{C}_{20}\text{H}_{22}\text{N}_5\text{O}_2$ [M^+H] 364.1773, found 364.1768.



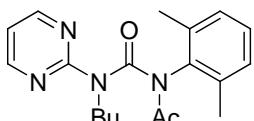
1-Acetyl-1-cyclohexyl-3-phenyl-3-pyrimidin-2-yl-urea (5a)^[17]: Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), cyclohexyl isonitrile (163.8 mg, 1.5 mmol) in toluene for 23 h to give **5a** as white solid (134.3 mg, 79%). M.p. 112-114 °C; IR (KBr, cm^{-1}): 2967, 2930, 1707, 1688, 1564, 1410, 1226, 702; ^1H NMR (CDCl_3 , 500 MHz): δ 8.65 (d, $J = 4.5$ Hz, 2H), 7.44 (t, $J = 7.5$ Hz, 2H), 7.34 (t, $J = 7.5$ Hz, 1H), 7.25 (d, $J = 7.5$ Hz, 2H), 7.12 (t, $J = 5.0$ Hz, 1H), 3.96-3.91 (m, 1H), 2.30 (s, 3H), 1.74-1.57 (m, 7H), 1.27-1.06 (m, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 170.9, 161.4, 158.6, 157.1, 140.8, 129.5, 127.8, 127.1, 118.2, 58.3, 30.1, 26.3, 25.4, 24.7; EI-MS m/z (%): 338 (3) [M^+]; HRMS (EI) m/s calcd for $\text{C}_{19}\text{H}_{22}\text{N}_4\text{O}_2$ 338.1743, found 338.1742.



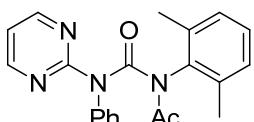
1-Acetyl-3-phenyl-3-pyrimidin-2-yl-1-o-tolyl-urea (5c): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), 2-methylisonitrile (175.7 mg, 1.5 mmol) in toluene for 19 h to give **5c** as yellow liquid (86.3 mg, 50%). IR (KBr, cm^{-1}): 3039, 1702, 1594, 1564, 1410, 1242, 724; ^1H NMR (CDCl_3 , 500 MHz): δ 8.55 (d, $J = 4.5$ Hz, 2H), 7.29-7.28 (m, 3H), 7.14 (t, $J = 7.0$ Hz, 1H), 7.07-7.04 (m, 3H), 6.99 (t, $J = 5.0$ Hz, 1H), 6.89 (d, $J = 4.5$ Hz, 2H), 2.45 (s, 3H), 1.90 (s, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 172.9, 161.4, 158.3, 157.1, 139.9, 137.0, 136.7, 130.8, 129.3, 128.3, 128.1, 127.7, 127.6, 126.4, 117.1, 25.0, 17.9; EI-MS m/z (%): 346 (5) [M^+], 170 (100); HRMS (EI): m/s Calcd. for $\text{C}_{20}\text{H}_{18}\text{N}_4\text{O}_2$: 346.1430, found: 346.1428.



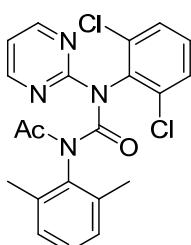
1-Acetyl-3-allyl-1-(2,6-dimethyl-phenyl)-3-pyrimidin-2-yl-urea (5d)^[17]: Following the same procedure as for **4a** with **1d** (72.4 mg, 0.54 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), 2,6-xylyl isocyanide (196.8 mg, 1.5 mmol) in toluene for 13 h to give **5d** as white solid (157.8 mg, 90%). M.p. 82-84 °C; IR (KBr, cm^{-1}): 2956, 2916, 1706, 1677, 1562, 1442, 1233, 808, 764; ^1H NMR (CDCl_3 , 500 MHz): δ 8.53 (d, $J = 4.5$ Hz, 2H), 7.13 (A of ABB', $J = 7.5$ Hz, 1H), 7.06 (BB' of ABB', $J = 7.5$ Hz, 2H), 6.93 (t, $J = 5.0$ Hz, 1H), 6.00-5.92 (m, 1H), 5.27 (dd, $J = 18.0, 1.5$ Hz, 1H), 5.09 (dd, $J = 10.5, 1.5$ Hz, 1H), 4.76 (d, $J = 5.5$ Hz, 2H), 2.31 (s, 6H), 1.88 (s, 3H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 172.6, 160.7, 157.6, 154.1, 137.7, 136.8, 133.5, 128.8, 128.5, 116.9, 116.4, 51.1, 24.8, 18.7; LC-MS (ESI) m/z 325.2 [M^+H]; Anal. Calcd. For $\text{C}_{18}\text{H}_{20}\text{N}_4\text{O}_2$: C, 66.65; H, 6.21; N, 17.27. Found: C, 66.65; H, 6.25; N, 16.97.



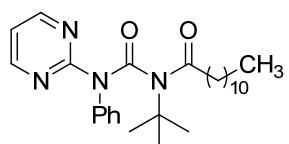
1-Acetyl-3-butyl-1-(2,6-dimethyl-phenyl)-3-pyrimidin-2-yl-urea (5e)^[17]: Following the same procedure as for **4a** with **1d** (80.6 mg, 0.53 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), 2,6-xylyl isocyanide (196.8 mg, 1.5 mmol) in toluene for 13 h to give **5e** as white solid (158.1 mg, 88%). M.p. 106-108 °C; IR (KBr, cm⁻¹): 2960, 2931, 1702, 1676, 1561, 1423, 1246, 806, 782; ¹H NMR (CDCl₃, 500 MHz): δ 8.51 (d, *J* = 5.0 Hz, 2H), 7.12 (t, *J* = 7.5 Hz, 1H), 7.04 (d, *J* = 7.5 Hz, 2H), 6.92 (t, *J* = 5.0 Hz, 1H), 4.12 (t, *J* = 7.5 Hz, 2H), 2.30 (s, 6H), 1.90 (s, 3H), 1.62 (quint, *J* = 7.5 Hz, 2H), 1.33 (sext, *J* = 7.5 Hz, 2H), 0.88 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 172.6, 160.9, 157.6, 154.5, 137.7, 136.8, 128.8, 128.4, 116.2, 49.1, 30.7, 24.8, 20.1, 18.7, 13.9; LC-MS (ESI) m/z 341.2 [M⁺H]; Anal. Calcd. For C₁₉H₂₄N₄O₂: C, 67.04; H, 7.11; N, 16.46. Found: C, 66.86; H, 7.08; N, 16.19.



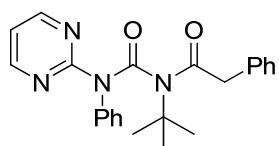
1-Acetyl-1-(2,6-dimethyl-phenyl)-3-phenyl-3-pyrimidin-2-yl-urea (5f): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), 2,6-xylyl isocyanide (196.8 mg, 1.5 mmol) in toluene for 13 h to give **5f** as white solid (167.2 mg, 93%). M.p. 148-150 °C; IR (KBr, cm⁻¹): 2920, 1718, 1684, 1578, 1563, 1419, 1238, 712; ¹H NMR (CDCl₃, 500 MHz): δ 8.52 (d, *J* = 4.5 Hz, 2H), 7.30-7.29 (m, 3H), 7.07 (t, *J* = 7.5 Hz, 1H), 6.97-6.92 (m, 5H), 2.38 (s, 3H), 2.09 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz): δ 173.3, 161.7, 158.0, 156.7, 139.8, 137.2, 136.3, 129.3, 128.5, 128.4, 128.3, 127.9, 116.7, 25.3, 18.5; EI-MS m/z (%): 360 (10) [M⁺], 170 (100); Anal. Calcd. For C₂₁H₂₀N₄O₂: C, 69.98; H, 5.59; N, 15.55. Found: C, 69.90; H, 5.63; N, 15.29.



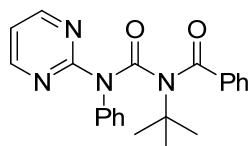
1-Acetyl-3-(2,6-dichloro-phenyl)-1-(2,6-dimethyl-phenyl)-3-pyrimidin-2-yl-urea (5g): Following the same procedure as for **4a** with **1d** (84.0 mg, 0.35 mmol), Cu(OAc)₂·H₂O (140.0 mg, 0.7 mmol), 2,6-xylyl isocyanide (137.7 mg, 1.1 mmol) in toluene for 14 h to give **5g** as white solid (114.6 mg, 76%). M.p. 196-198 °C; IR (KBr, cm⁻¹): 3033, 2917, 1724, 1691, 1565, 1417, 1311, 1234, 782, 772; ¹H NMR (CDCl₃, 500 MHz): δ 8.58 (d, *J* = 5.0 Hz, 2H), 7.29 (m, 2H), 7.19 (t, *J* = 8.0 Hz, 1H), 7.11 (t, *J* = 7.5 Hz, 1H), 7.02 (t, *J* = 4.5 Hz, 1H), 6.98 (d, *J* = 5.5 Hz, 2H), 2.24 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 172.8, 160.4, 158.0, 137.3, 136.3, 135.7, 129.3, 128.8, 128.7, 128.5, 117.0, 25.1, 18.5; EI-MS m/z (%): 430 [M⁺ (³⁷Cl, ³⁵Cl)], 428 [M⁺ (2×³⁵Cl)]; HRMS (EI) m/s calcd. for C₂₁H₁₈Cl₂N₄O₂ 428.0807, found 428.0804.



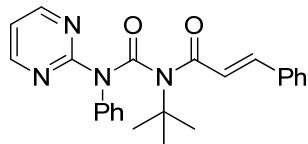
1-*tert*-Butyl-1-dodecanoyl-3-phenyl-3-pyrimidin-2-yl-urea (5i): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) laurate monohydrate (424.1 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 23 h to give **5i** as yellow liquid (144.8 mg, 64%) together with recovered **1d** (22.9 mg, 74% conversion). IR (KBr, cm⁻¹): 2925, 2854, 1701, 1675, 1565, 1409, 722; ¹H NMR (CDCl₃, 500 MHz): δ 8.70 (d, *J* = 5.0 Hz, 2H), 7.43 (t, *J* = 8.0 Hz, 2H), 7.35-7.32 (m, 3H), 7.20 (t, *J* = 5.0 Hz, 1H), 2.77-2.74 (m, 1H), 2.59-2.58 (m, 1H), 1.70-1.66 (m, 2H), 1.35-1.25 (m, 25H), 0.87 (t, *J* = 6.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 172.9, 160.9, 158.8, 156.1, 140.4, 129.4, 127.9, 127.0, 119.3, 58.7, 37.4, 31.9, 29.7, 29.6, 29.5, 29.4, 29.3, 27.8, 25.1, 22.7, 14.1; LC-MS (ESI) m/z 475.2 [M⁺Na]; HRMS (ESI) m/s calcd for C₂₇H₄₁N₄O₂ [M⁺H] 453.3230, found 453.3222.



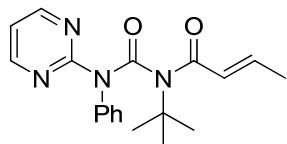
1-*tert*-Butyl-3-phenyl-1-phenylacetyl-3-pyrimidin-2-yl-urea (5j): Following the same procedure as for **4a** with **1d** (51.4 mg, 0.3 mmol), copper (II) phenylacetate monohydrate (250.4 mg, 0.75 mmol), *tert*-butyl isocyanide (74.8 mg, 0.9 mmol) in toluene at 90 °C for 41 h to give **5j** as white solid (66.2 mg, 57%) together with recovered **1d** (5.7 mg, 89% conversion). M.p. 128-130 °C; IR (KBr, cm⁻¹): 2977, 1704, 1674, 1564, 1409, 1320, 720; ¹H NMR (CDCl₃, 500 MHz): δ 8.75 (d, *J* = 5.0 Hz, 2H), 7.46 (t, *J* = 7.5 Hz, 2H), 7.41 (d, *J* = 8.5 Hz, 2H), 7.38-7.32 (m, 5H), 7.26 (d, *J* = 6.5 Hz, 1H), 7.23 (t, *J* = 5.0 Hz, 1H), 4.25 (d, *J* = 15.5 Hz, 1H), 3.96 (d, *J* = 16.0 Hz, 1H), 1.34 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.7, 160.9, 158.9, 156.1, 140.3, 135.5, 130.0, 129.5, 128.3, 128.1, 127.0, 126.6, 119.5, 59.1, 43.8, 27.8; LC-MS (ESI) m/z 389.1 [M⁺H]; Anal. Calcd. For C₂₃H₂₄N₄O₂: C, 71.11; H, 6.23; N, 14.42. Found: C, 71.18; H, 6.41; N, 14.44.



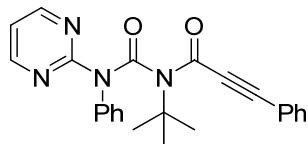
1-Benzoyl-1-*tert*-butyl-3-phenyl-3-pyrimidin-2-yl-urea (5k): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) benzoate monohydrate (323.8 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 33 h to give **5k** as white solid (165.2 mg, 88%). M.p. 110-111 °C; IR (KBr, cm⁻¹): 2983, 1702, 1648, 1565, 1405, 1314, 697; ¹H NMR (CDCl₃, 500 MHz): δ 8.56 (d, *J* = 4.5 Hz, 2H), 7.48-7.43 (m, 3H), 7.37 (t, *J* = 7.5 Hz, 2H), 7.28-7.21 (m, 3H), 7.04 (t, *J* = 4.5 Hz, 1H), 6.65 (d, *J* = 6.5 Hz, 2H), 1.70 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.0, 160.4, 158.1, 155.6, 140.3, 138.0, 131.0, 129.1, 128.2, 127.9, 127.7, 127.3, 117.6, 60.4, 28.7; LC-MS (ESI) m/z 375.1 [M⁺H]; Anal. Calcd. For C₂₂H₂₂N₄O₂: C, 70.57; H, 5.92; N, 14.96. Found: C, 70.83; H, 6.13; N, 14.68.



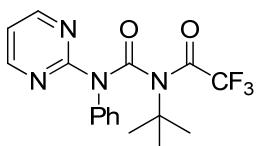
1-*tert*-Butyl-3-phenyl-1-(3-phenyl-acryloyl)-3-pyrimidin-2-yl-urea (5l): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) cinnamate monohydrate (375.9 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 33 h to give **5l** as white solid (157.0 mg, 78%). M.p. 146–148 °C; IR (KBr, cm^{−1}): 2974, 2930, 1693, 1664, 1567, 1410, 1193, 762, 703; ¹H NMR (CDCl₃, 500 MHz): δ 8.70 (d, *J* = 5.0 Hz, 2H), 7.62 (d, *J* = 15.0 Hz, 1H), 7.58 (d, *J* = 6.5 Hz, 2H), 7.42 (t, *J* = 8.0 Hz, 2H), 7.38–7.34 (m, 6H), 7.27 (d, *J* = 14.0 Hz, 1H), 7.16 (t, *J* = 5.0 Hz, 1H), 1.48 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 165.4, 161.0, 158.8, 156.0, 142.0, 140.3, 135.3, 129.7, 129.5, 128.8, 128.2, 128.1, 127.5, 121.7, 119.1, 59.3, 28.0; LC-MS (ESI) m/z 401.1 [M⁺H]; Anal. Calcd. For C₂₄H₂₄N₄O₂: C, 71.98; H, 6.04; N, 13.99. Found: C, 71.85; H, 6.17; N, 13.78.



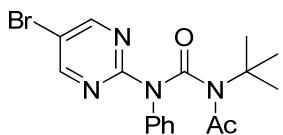
1-But-2-enoyl-1-*tert*-butyl-3-phenyl-3-pyrimidin-2-yl-urea (5m): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) crotonate monohydrate (233.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene at 90 °C for 26 h to give **5m** as white solid (78.7 mg, 47%). M.p. 116–118 °C; IR (KBr, cm^{−1}): 2973, 2925, 1689, 1672, 1565, 1407, 1330, 1191, 723; ¹H NMR (CDCl₃, 500 MHz): δ 8.69 (d, *J* = 5.0 Hz, 2H), 7.42 (t, *J* = 7.5 Hz, 2H), 7.35–7.33 (m, 3H), 7.16 (t, *J* = 4.5 Hz, 1H), 6.84 (dq, *J* = 15.0, 7.5 Hz, 1H), 6.53 (dq, *J* = 15.0, 2.0 Hz, 1H), 1.87 (dd, *J* = 7.0, 2.0 Hz, 3H), 1.40 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 165.5, 160.9, 158.7, 156.1, 141.1, 140.4, 129.4, 128.0, 127.3, 125.7, 119.1, 59.0, 27.9, 18.1; LC-MS (ESI) m/z 339.1 [M⁺H]; Anal. Calcd. For C₁₉H₂₂N₄O₂: C, 67.44; H, 6.55; N, 16.56. Found: C, 67.70; H, 6.42; N, 16.57.



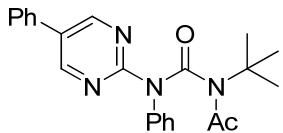
1-*tert*-Butyl-3-phenyl-1-(3-phenyl-propynoyl)-3-pyrimidin-2-yl-urea (5n): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) phenylpropionate multihydrate (851.7 mg, 2.0 mmol), *tert*-butyl isocyanide (249.4 mg, 3.0 mmol) in toluene at 90 °C for 22 h to give **5n** as white solid (93.5 mg, 47%). M.p. 37–39 °C; IR (KBr, cm^{−1}): 2981, 2927, 2211, 1703, 1646, 1565, 1407, 691; ¹H NMR (CDCl₃, 500 MHz): δ 8.67 (d, *J* = 5.0 Hz, 2H), 7.57 (d, *J* = 7.5 Hz, 2H), 7.40–7.30 (m, 8H), 7.14 (t, *J* = 4.5 Hz, 1H), 1.57 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 160.8, 158.4, 154.5, 151.8, 140.8, 132.9, 130.1, 129.3, 128.4, 128.0, 127.6, 120.4, 118.7, 88.3, 83.6, 60.1, 27.7; LC-MS (ESI) m/z 399.1 [M⁺H]; HRMS (ESI) m/s calcd for C₂₄H₂₃N₄O₂ [M⁺H] 399.1821, found 399.1807.



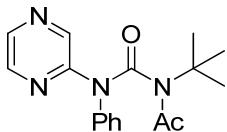
1-*tert*-Butyl-3-phenyl-3-pyrimidin-2-yl-1-(2,2,2-trifluoro-acetyl)-urea (5o): Following the same procedure as for **4a** with **1d** (85.6 mg, 0.5 mmol), copper (II) trifluoroacetate (289.6 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene at 90 °C for 45 h to give **5o** as white solid (58.2 mg, 32%) together with recovered **1d** (26.6 mg, 69% conversion). M.p. 118–119 °C; IR (KBr, cm⁻¹): 2977, 2934, 1721, 1695, 1570, 1409, 1325, 1134, 725; ¹H NMR (CDCl₃, 500 MHz): δ 8.68 (d, *J* = 5.0 Hz, 2H), 7.46 (t, *J* = 7.0 Hz, 2H), 7.39 (t, *J* = 7.5 Hz, 1H), 7.32 (d, *J* = 7.5 Hz, 2H), 7.18 (t, *J* = 5.0 Hz, 1H), 1.35 (s, 9H); ¹⁹F NMR (CDCl₃, 470 MHz): δ -70.74 (s); ¹³C NMR (CDCl₃, 125 MHz): δ 160.2, 158.5, 157.0 (q, ²J_{C-F} = 36.8 Hz), 151.9, 139.9, 129.6, 128.5, 127.5, 119.0, 116.0 (q, ¹J_{C-F} = 287.5 Hz), 61.5, 27.1; LC-MS (ESI) m/z 367.0 [M⁺H]; Anal. Calcd. For C₁₇H₁₇F₃N₄O₂: C, 55.74; H, 4.68; N, 15.29. Found: C, 55.88; H, 4.90; N, 15.20.



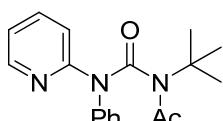
1-Acetyl-3-(5-bromo-pyrimidin-2-yl)-1-*tert*-butyl-3-phenyl-urea (6a)^[17]: Following the same procedure as for **4a** with **1p** (125.0 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 21 h to give **6a** as white solid (97.3 mg, 50%). M.p. 94–96 °C; IR (KBr, cm⁻¹): 2984, 2955, 1705, 1665, 1539, 1412, 1017, 731; ¹H NMR (CDCl₃, 500 MHz): δ 8.75 (s, 2H), 7.44 (t, *J* = 8.0 Hz, 2H), 7.36–7.31 (m, 3H), 2.35 (s, 3H), 1.36 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 169.9, 159.5, 159.1, 156.1, 140.0, 129.6, 128.2, 127.1, 117.9, 59.0, 27.8, 25.5; EI-MS m/z (%): 392 (0.6) [M⁺ (⁸¹Br)], 390 (0.6) [M⁺ (⁷⁹Br)], 251 (100); HRMS (EI) m/z Calcd. for C₁₇H₁₉BrN₄O₂ 390.0691, found 390.0695.



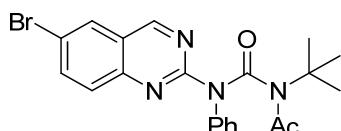
1-Acetyl-1-*tert*-butyl-3-phenyl-3-(5-phenyl-pyrimidin-2-yl)-urea (6b)^[17]: Following the same procedure as for **4a** with **1q** (86.6 mg, 0.35 mmol), Cu(OAc)₂·H₂O (139.8 mg, 0.7 mmol), *tert*-butyl isocyanide (91.9 mg, 1.1 mmol) in toluene for 12 h to give **6b** as yellow liquid (86.4 mg, 64%). IR (KBr, cm⁻¹): 2978, 1705, 1672, 1424, 1302, 1191, 758, 694; ¹H NMR (CDCl₃, 500 MHz): δ 8.93 (s, 2H), 7.56 (d, *J* = 7.5 Hz, 2H), 7.52 (t, *J* = 7.5 Hz, 2H), 7.49–7.44 (m, 3H), 7.40 (d, *J* = 7.5 Hz, 2H), 7.35 (t, *J* = 7.0 Hz, 1H), 2.44 (s, 3H), 1.38 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.1, 159.6, 156.6, 156.3, 140.4, 133.1, 132.3, 129.6, 129.5, 129.4, 128.0, 127.0, 126.9, 58.9, 27.8, 25.6; LC-MS (ESI) m/z 411.2 [M⁺Na]; HRMS (ESI) calcd for C₂₃H₂₄N₄NaO₂ [M⁺Na] 411.1797, found 411.1791.



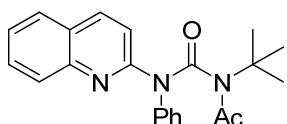
1-Acetyl-1-*tert*-butyl-3-phenyl-3-pyrazin-2-yl-urea (6c**)^[17]:** Following the same procedure as for **4a** with **1r** (85.6 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isonitrile (131.3 mg, 1.5 mmol) in toluene for 18 h to give **6c** as yellow solid (93.4 mg, 60%). M.p. 80-82 °C; IR (KBr, cm⁻¹): 2979, 1702, 1674, 1405, 1294, 1016, 728; ¹H NMR (CDCl₃, 500 MHz): δ 8.57 (s, 1H), 8.47 (s, 2H), 7.44 (t, *J* = 7.5 Hz, 2H), 7.35 (t, *J* = 7.5 Hz, 1H) 7.30 (d, *J* = 7.0 Hz, 2H), 2.33 (s, 3H), 1.31 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 169.6, 156.6, 143.2, 143.0, 142.5, 140.4, 129.8, 128.3, 127.2, 58.9, 27.8, 25.6; LC-MS (ESI) m/z 335.1 [M⁺Na]; HRMS (ESI) calcd for C₁₇H₂₀N₄NaO₂ [M⁺Na] 335.1484, found 335.1483.



1-Acetyl-1-*tert*-butyl-3-phenyl-3-pyridin-2-yl-urea (6d**)^[17]:** Following the same procedure as for **4a** with **1s** (85.1 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 26 h to give **6d** as white solid (110.5 mg, 71%). M.p. 92-94 °C; IR (KBr, cm⁻¹): 2961, 2925, 1695, 1665, 1469, 1318, 783, 711; ¹H NMR (CDCl₃, 500 MHz): δ 8.52-8.51(m, 1H), 7.72 (dt, *J* = 8.0, 1.5 Hz, 1H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.32-7.29 (m, 3H), 7.23-7.21 (m, 1H), 7.17 (d, *J* = 8.0 Hz, 1H), 2.37 (s, 3H), 1.31 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 169.8, 156.5, 154.9, 149.3, 141.4, 138.3, 129.5, 127.6, 126.9, 122.6, 121.6, 58.6, 27.7, 25.7; LC-MS (ESI) m/z 312.1 [M⁺H]; Anal. Calcd. For C₁₈H₂₁N₃O₂: C, 69.43; H, 6.80; N, 13.49. Found: C, 69.26; H, 6.78; N, 13.31.

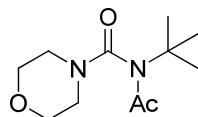


1-Acetyl-3-(6-bromo-quinazolin-2-yl)-1-*tert*-butyl-3-phenyl-urea (6e**):** Following the same procedure as for **4a** with **1t** (150.1 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 24 h to give **6e** as yellow liquid (90.9 mg, 41%). IR (KBr, cm⁻¹): 2977, 2927, 1700, 1669, 1569, 1420, 836; ¹H NMR (CDCl₃, 500 MHz): δ 9.28 (s, 1H), 8.09 (d, *J* = 2.0 Hz, 1H), 7.99 (dd, *J* = 9.0, 2.0 Hz, 1H), 7.86 (d, *J* = 9.0 Hz, 1H), 7.45-7.39 (m, 4H), 7.34 (t, *J* = 7.5 Hz, 1H), 2.47 (s, 3H), 1.31 (s, 9H); ¹³C NMR (CDCl₃, 125 MHz): δ 170.0, 161.4, 157.1, 156.3, 149.7, 140.3, 138.7, 129.9, 129.4, 129.3, 128.0, 127.1, 123.8, 122.3, 58.8, 27.8, 25.8; LC-MS (ESI) m/z 465.1 [M⁺Na (⁸¹Br)], 463.1 [M⁺Na (⁷⁹Br)]; HRMS (ESI) calcd for C₂₁H₂₁BrN₄NaO₂ [M⁺Na] 463.0746, found 463.0738.



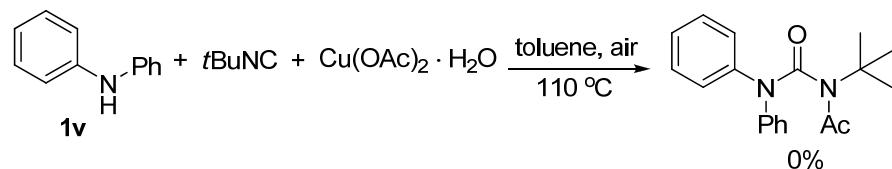
1-Acetyl-1-*tert*-butyl-3-phenyl-3-quinolin-2-yl-urea (6f**):** Following the same procedure as for **4a** with **1u** (110.1 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 18 h to give **6f** as yellow solid (122.6 mg, 68%). M.p. 90-92 °C; IR (KBr, cm⁻¹): 2978, 2928, 1698, 1669, 1593, 1501, 1296, 1193, 824, 756; ¹H NMR (CDCl₃, 500 MHz): δ 8.13 (d, *J* = 8.5 Hz, 1H), 8.02 (d, *J* = 8.0 Hz, 1H), 7.79 (d, *J* = 8.0 Hz, 1H), 7.74 (td, *J* = 7.0, 1.0 Hz, 1H), 7.57 (td, *J* = 8.0, 1.0 Hz, 1H), 7.42 (t, *J* = 7.0 Hz, 2H), 7.37 (d, *J* = 7.5 Hz,

2H), 7.32 (t, J = 8.0 Hz, 1H), 7.12 (d, J = 8.5 Hz, 1H), 2.47 (s, 3H), 1.31 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 169.7, 156.6, 153.1, 146.9, 141.0, 138.7, 130.5, 129.5, 129.2, 127.8, 127.4, 127.3, 127.0, 126.6, 119.1, 58.6, 27.9, 25.9; LC-MS (ESI) m/s 384.0 [M^+Na]; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{23}\text{N}_3\text{NaO}_2$ [M^+Na] 384.1688, found 384.1679.

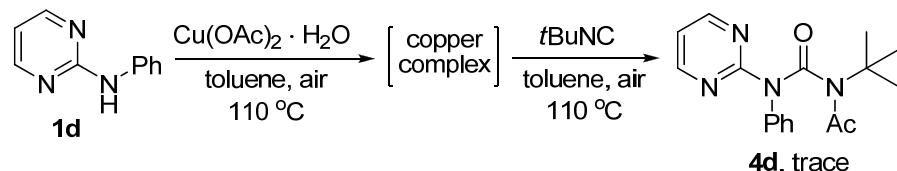


N-acetyl-N-tert-butylmorpholine-4-carboxamide (6g): Following the same procedure as for **4a** with morpholine (43.6 mg, 0.5 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol) in toluene for 22 h to give **6g** as pale yellow oil (79.7 mg, 70%). IR (KBr, cm^{-1}): 2967, 2924, 2856, 1681, 1426, 1364, 1317, 1273, 1220, 1196, 1117, 1027, 597; ^1H NMR (CDCl_3 , 500 MHz): δ 3.73-3.53 (m, 8H), 2.00 (s, 3H), 1.45 (s, 9H); ^{13}C NMR (CDCl_3 , 125 MHz): δ 168.7, 156.3, 66.7, 66.4, 57.9, 47.1, 43.8, 28.1, 24.0; LC-MS (ESI) m/s 251.1 [M^+Na]; HRMS (ESI) calcd for $\text{C}_{11}\text{H}_{20}\text{N}_2\text{NaO}_3$ [M^+Na] 251.1368, found 251.1366.

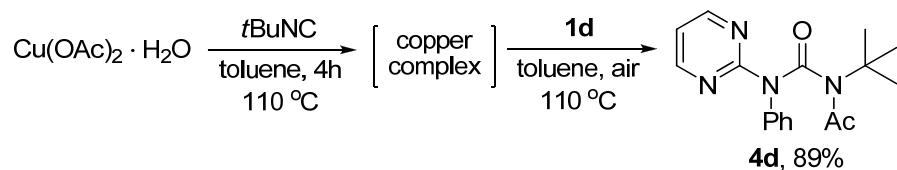
6. Mechanistic Studies



Following the same procedure as for **4a** with **1v** (16.9 mg, 0.1 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (40.0 mg, 0.2 mmol) and *tert*-butyl isocyanide (26 mg, 0.3 mmol) in toluene for 20 h, no desired urea product was detected by TLC and LC-MS.



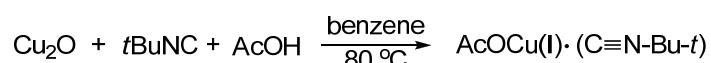
Following the same procedure as for **4a** with **1d** (342.4 mg, 2.0 mmol), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (798.6 mg, 4.0 mmol) in toluene for 1 h and monitored by TLC. Upon completion, the reaction mixture filtered and washed with water to give copper complex. Then the given copper complex and *tBuNC* was mixed in toluene and was stirred under 110 °C for 24 h, only trace amount of **4d** was detected which was monitored by TLC.



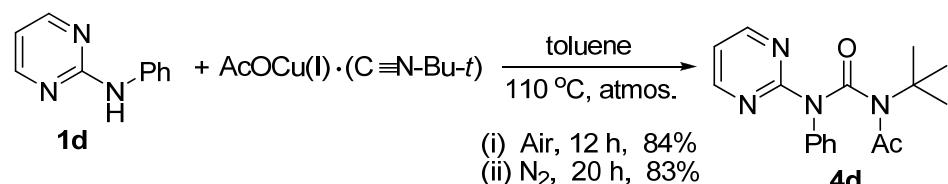
Synthesis of copper complex from $\text{Cu}(\text{OAc})_2$ and isocyanide. To an oven-dried flask containing $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (399.3 mg, 2.0 mmol), *tert*-butyl isocyanide (277.2 mg, 3.0 mmol) was added in

toluene (3.0 mL). The reaction mixture was stirred at 110 °C for 4 h in the presence of air which was dried through a calcium chloride tube. Upon completion, filtrated the reaction and the filtrate was cooled to give a crystal. Filtrated to give the Cu(II) complex, which was used directly for next step without further purification.

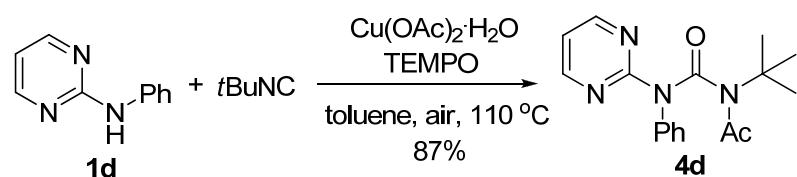
Following the same procedure used for **4a** with **1d** (34.2 mg, 0.2 mmol) and Cu(II) complex (113.1 mg, 0.4 mmol) in toluene (1.8 mL). After 22 h at 110 °C, purification by column chromatography on silica gel yielded **4d** (55.8 mg, 89%) as a white solid.



Synthesis of Cu(I) complex [AcOCu(I)•(C≡N-Bu-t)].^[18] Under nitrogen, a mixture of acetic acid (17 mmol), Cu₂O (8.5 mmol), and *t*-BuNC (17 mmol) was heated in 12 mL of benzene at 80 °C for 3 h. After filtration, the filtrate was subjected to evaporation in vacuo. Then benzene (10 mL) was added and recrystallization was carried out by warming the mixture up to 80 °C. The procedure was repeated for three times to give Cu(I) complex as a white solid. IR (KBr, cm⁻¹): 2174, 1581, 1562, 1410; ¹H NMR (*d*₆-DMSO, 500 MHz): δ 1.46 (s, 9H); ¹³C NMR (*d*₆-DMSO, 125 MHz): δ 57.1, 29.9.

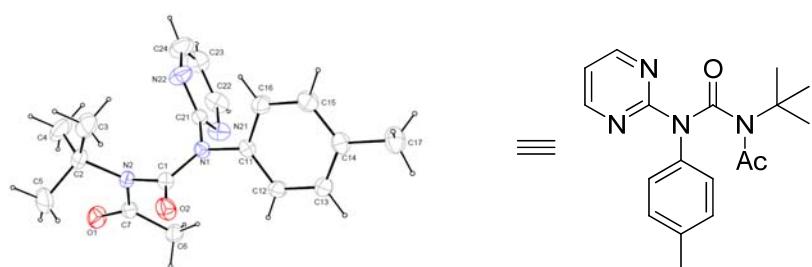


Following the same procedure used for **4a** with **1d** (34.2 mg, 0.2 mmol) and above prepared [AcOCu(I)•(C≡N-Bu-t)] (82.3 mg, 0.4 mmol) in toluene (1.2 mL) under air or N₂ atmosphere. After stirred at 110 °C, purification by column chromatography on silica gel yielded **4d** as a white solid in 84% and 83% yield, respectively.

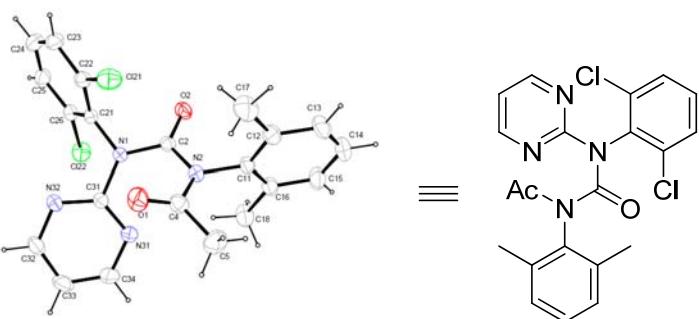


Following the same procedure as for **4a** with **1d** (85.5 mg, 0.5 mmol), Cu(OAc)₂·H₂O (199.7 mg, 1.0 mmol), *tert*-butyl isocyanide (131.3 mg, 1.5 mmol), TEMPO (78.2 mg, 0.5 mmol) in toluene for 16 h to give **4d** as white solid (135.6 mg, 87%).

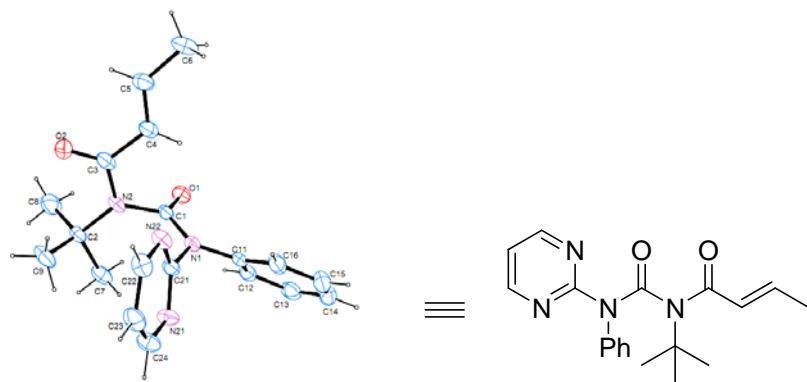
7. X-ray crystal structure for compounds **4e**, **5g** and **5m**



Crystallographic data for **4e**: C₁₈H₂₂N₄O₂, M = 326.40, monoclinic, P 21/n (No. 14), a = 10.438 (2) Å, b = 8.222 (1) Å, c = 20.993 (4) Å, β = 94.658(2)°, V = 1795.7 (5) Å³, Z = 4, Crystal size: 0.30 × 0.25 × 0.20 mm, T = 295 K, ρ_{calcd} = 1.207 g·cm⁻³, R₁ = 0.0528 (I>4σ(I)), wR₂ = 0.1700 (all data), GOF = 1.046, reflections collected/unique: 4120 / 2824 (Rint = 0.0261), Data: 2824, restraints: 0, parameters: 218. CCDC 882910 contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif



Crystallographic data for **5g**: C₂₁H₁₈Cl₂N₄O₂, M= 429.29, monoclinic, P 21/c (No. 14), a = 15.245 (5) Å, b = 8.417 (5) Å, c = 16.508 (5) Å, β = 104.637 (5)°, V = 2049.5 (15) Å³, Z = 4, Crystal size: 0.30 × 0.25 × 0.20 mm, T = 295 K, ρ_{calcd} = 1.391 g·cm⁻³, R₁ = 0.0421 (I>4σ(I)), wR₂ = 0.1278 (all data), GOF = 1.041, reflections collected/unique: 4697 / 3685 (Rint = 0.0200), Data: 3685, restraints: 0, parameters: 262. CCDC 883162 contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/date_request/cif.



Crystallographic data for **5m**: C₁₉H₂₂N₄O₂, M = 338.41, monoclinic, P21/n (No. 14), a = 10.47 (2) Å, b = 13.29 (3) Å, c = 14.71 (3) Å, β = 105.91 (3)°, V = 1969 (8) Å³, Z = 4, Crystal size: 0.28 × 0.25 × 0.21 mm, T = 295 K, ρ_{calcd} = 1.142 g·cm⁻³, R₁ = 0.1526 (I>4σ(I)), wR₂ = 0.4040 (all data), GOF = 1.223, reflections collected/unique: 8416 / 3226 (Rint = 0.0637), Data: 3226, restraints: 0, parameters: 227. CCDC 888379 contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/date_request/cif.

8. Reference

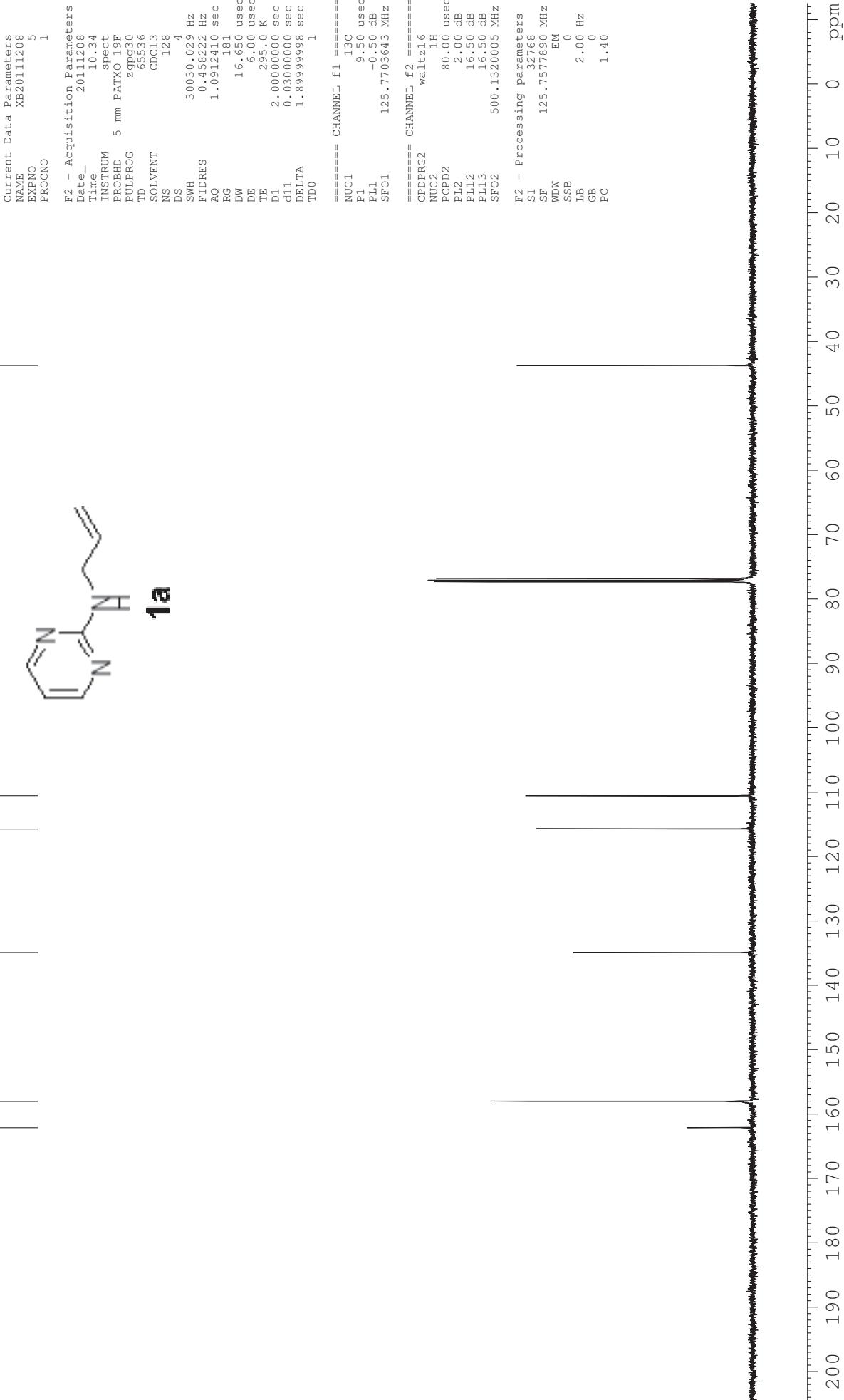
- [1] S. Jaime-Figueroa, Y. Liu, J. M. Muchowski and D. G. Putman, *Tetrahedron Lett.*, 1998, **39**, 1313.
- [2] H. P. L. Steenackers, *J. Med. Chem.*, 2010, **54**, 472.
- [3] L. Ackermann and A. V. Lygin, *Org. Lett.*, 2012, **14**, 764.
- [4] H. Takeuchi and K. Watanabe, *J. Phys. Org. Chem.*, 1998, 478.
- [5] R. Pierre, M. Florence, N. Romain, T. Jamal and G. Gilles, *Int. Patent*, WO 2010/143168A2, 2010.
- [6] E. Andre, *DE Patent*, DE 1978/2739659A1, 1978.
- [7] G. John, B. S. Armen, M. Allison, K. Tomas, F. K. Chiu, S. Jeongbeob, T. Hongqi, B. James and K. Kevin, *Int. Patent*, WO 2007/146824A2, 2007.
- [8] M. Colombo, M. Giglio and I. Peretto, *J. Heterocycl. Chem.*, 2008, **45**, 1077.
- [9] J. Chen, Q. Pang, Y. Sun and X. Li, *J. Org. Chem.*, 2011, **76**, 3523.
- [10] E. F. DiMauro, J. Newcomb, J. J. Nunes, J. E. Bemis, C. Boucher, J. L. Buchanan, W. H. Buckner, V. J. Cee, L. Chai, H. L. Deak, L. F. Epstein, T. Faust, P. Gallant, S. D. Geuns-Meyer, A. Gore, Y. Gu, B. Henkle, B. L. Hodous, F. Hsieh, X. Huang, J. L. Kim, J. H. Lee, M. W. Martin, C. E. Masse, D. C. McGowan, D. Metz, D. Mohn, K. A. Morgenstern, A. Oliveira-dos-Santos, V. F. Patel, D. Powers, P. E. Rose, S. Schneider, S. A. Tomlinson, Y.-Y. Tudor, S. M. Turci, A. A. Welcher, R. D. White, H. Zhao, L. Zhu and X. Zhu, *J. Med. Chem.*, 2006, **49**, 5671.
- [11] R. L. Martin and A. Whitley, *J. Chem. Soc.*, 1958, 1394.
- [12] M. Kondo and M. Kubo *J. Phys. Chem.*, 1958, **62**, 1558.
- [13] M. H. Borawska, P. Koczoń, J. Piekut, R. Świsłocka and W. Lewandowski, *J. Mol. Struct.*, 2009, **919**, 284.
- [14] M. Perec, R. Baggio, R. P. Sartoris, R. C. Santana, O. Peña and R. Calvo, *Inorg. Chem.*, 2009, **49**, 695.
- [15] M. T. Rogers, *J. Am. Chem. Soc.*, 1947, **69**, 1506.
- [16] M. D. B. Drew, A. P. Mullins and D. A. Rice, *Polyhedron*, 1994, **13**, 1631.
- [17] B. Xu, X. M. Huang, X. H. Hong, G. Y. Qian, T. Fang and X. C. Xu, *Faming Zhuanli Shengqing*, CN 102718719 A 20121010, 2012.
- [18] T. Saegusa, I. Murase and Y. Ito, *J. Org. Chem.*, 1973, **38**, 1753.

HXM-2-2
PROTON CDC13 I

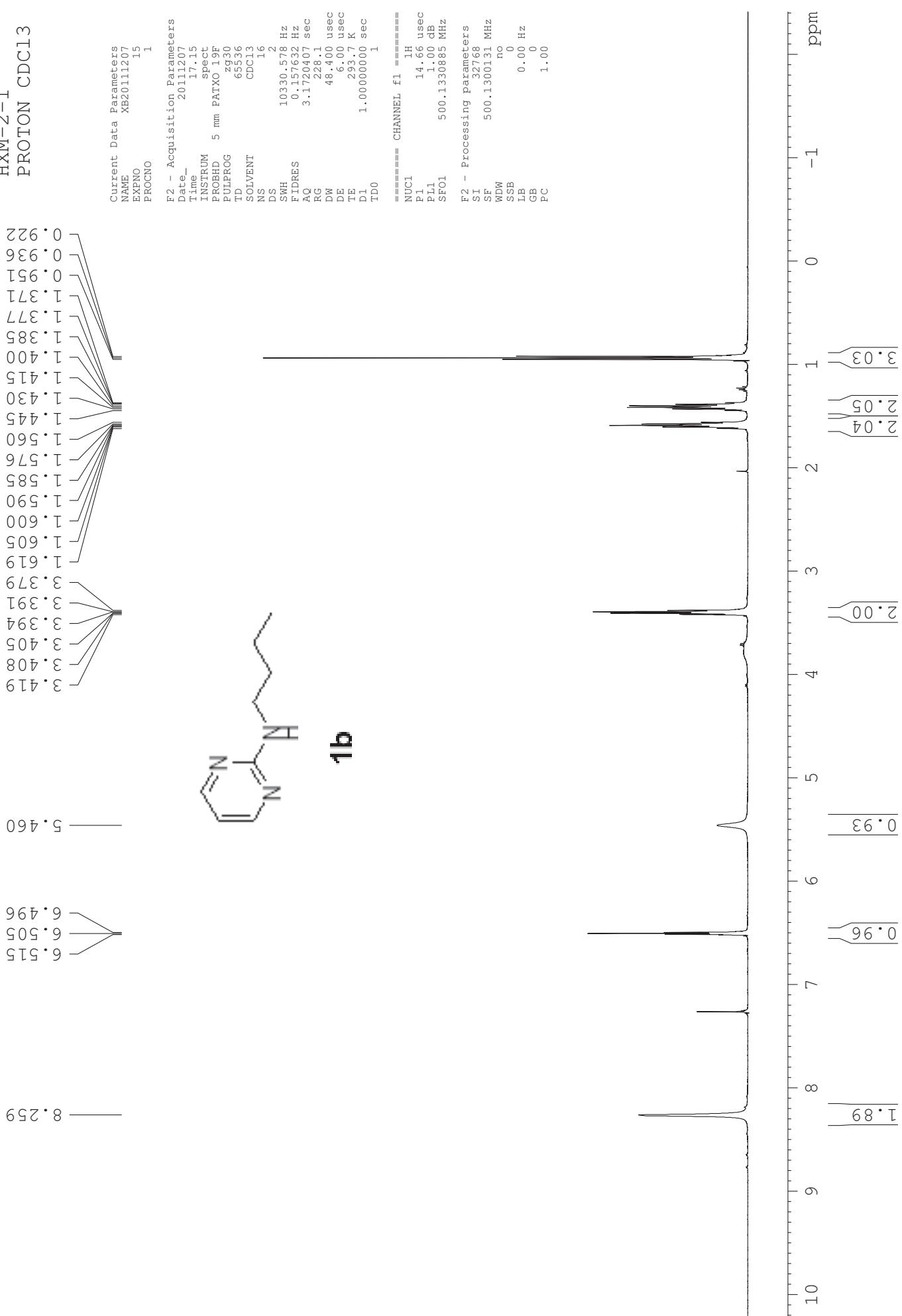
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RG 114
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DE 6.00 usec
TE 295.0 K
D1 1.0000000 sec
TD0 1
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P1 14.66 usec
PL1 1.00 dB
SF01 500.1330885 MHz
F2 - Processing parameters
SI 32768
SF 500.1300132 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

The figure shows the chemical structure of compound 1a, which is 1-(2-ethylbutyl)-4-methylimidazole. The structure features a five-membered imidazole ring with a methyl group at position 4 and a 2-ethylbutyl group at position 1. To the right of the structure is its ¹H NMR spectrum. The x-axis represents the chemical shift in ppm, ranging from 0 to 11.0. Key peaks are labeled with their corresponding chemical shifts: a sharp peak at 1.92 ppm (labeled 1.92), a peak at 2.00 ppm (labeled 2.00), a multiplet between 3.0 and 4.0 ppm, a peak at 4.99 ppm (labeled 4.99), a peak at 5.69 ppm (labeled 5.69), a peak at 6.98 ppm (labeled 6.98), a peak at 7.92 ppm (labeled 7.92), and a peak at 9.33 ppm (labeled 9.33). The spectrum also shows several aromatic peaks between 7.0 and 8.5 ppm.

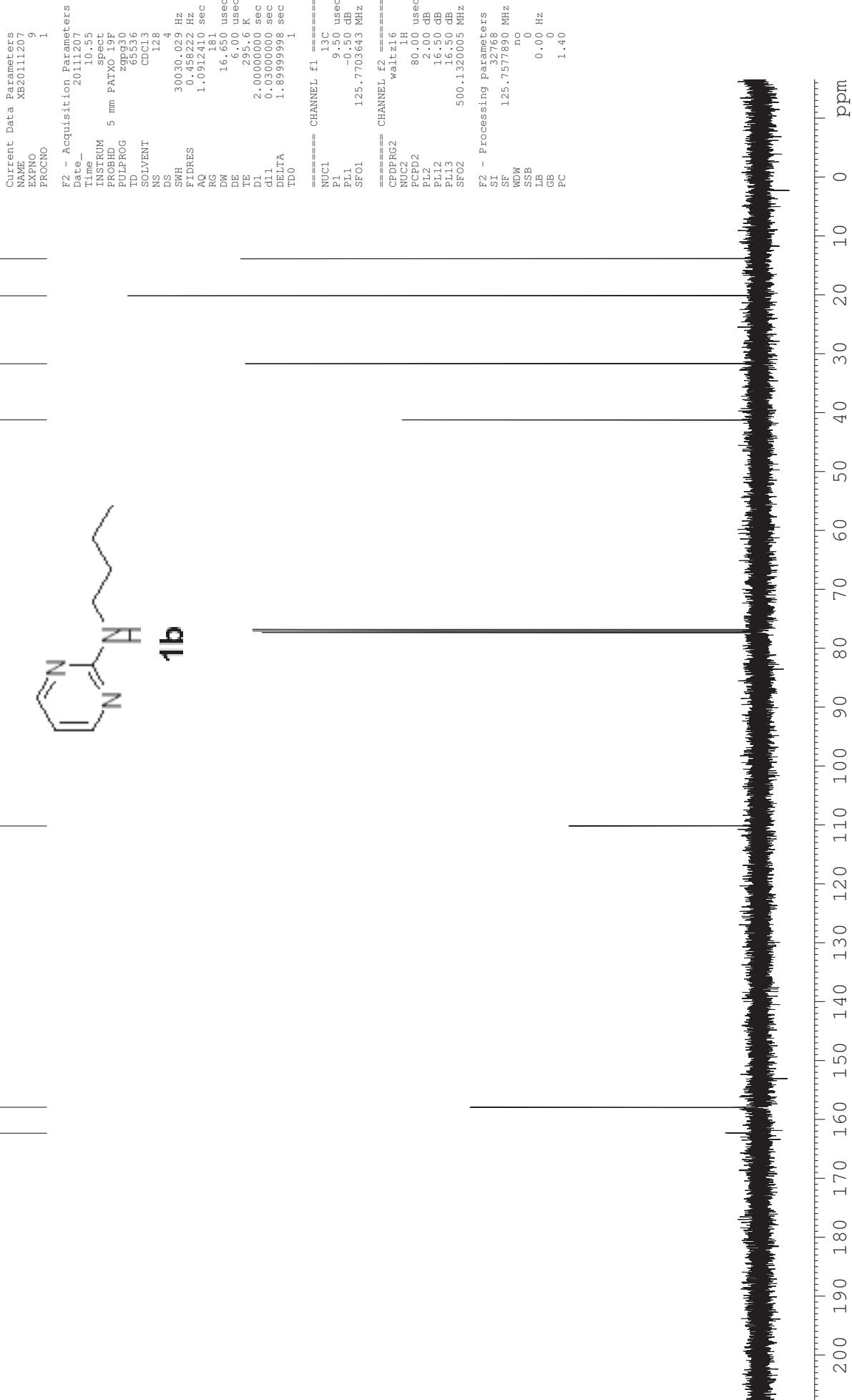
HXM-2-2
C13CPD CDC13



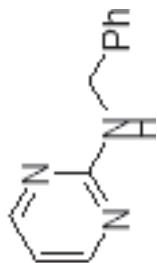
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PROTON CDCl₃



HXM-2-1
C13CPD CDC13



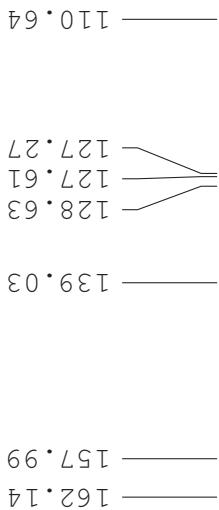
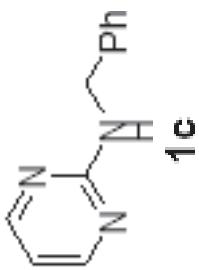
XSG-1-262
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1c

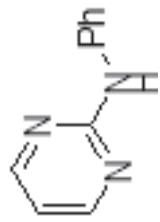


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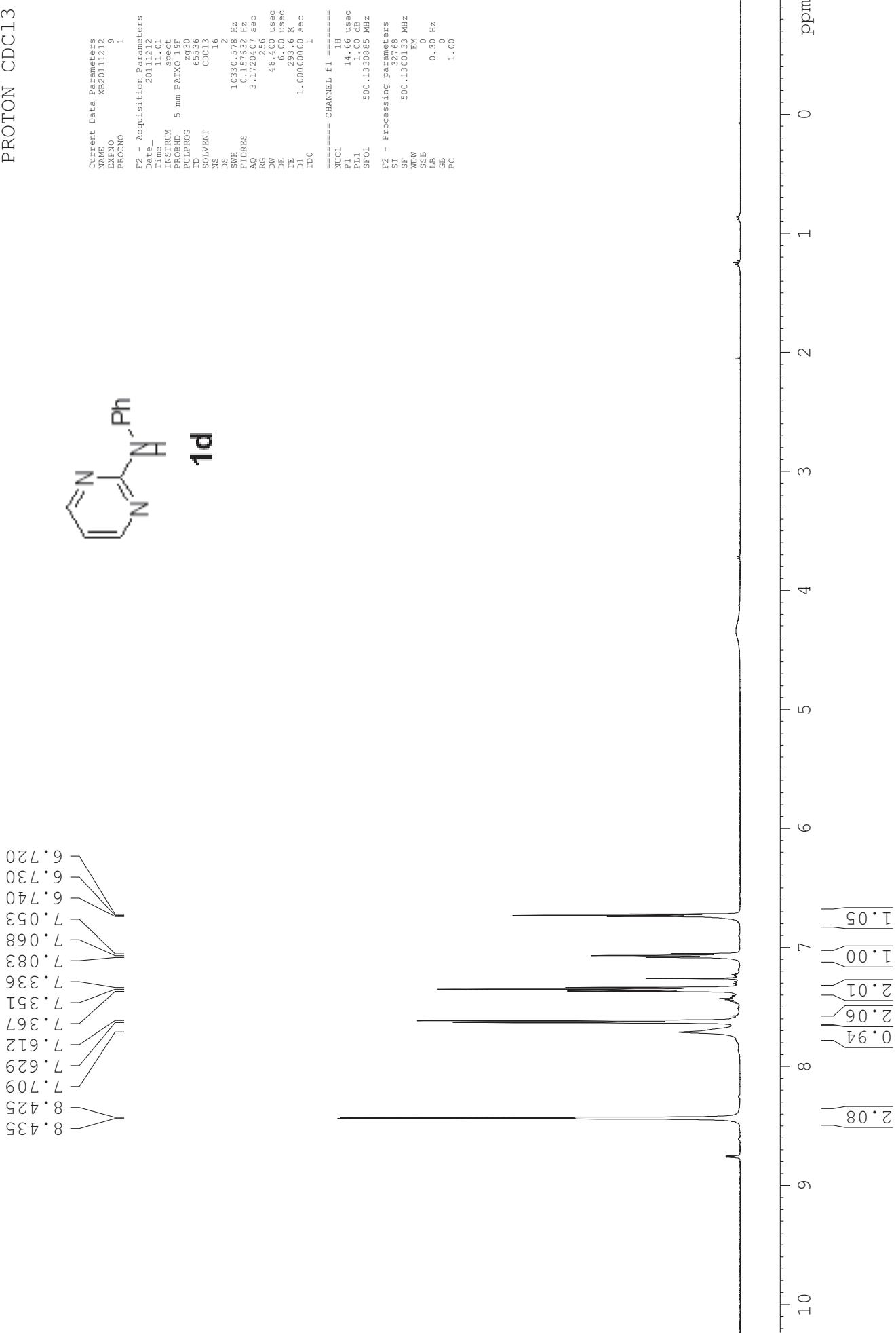


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SOLVENT CDCl₃
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FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 181
DW 16.650 usec
DE 6.00 usec
TE 295.2 K
D1 2.0000000 sec
d1 0.0300000 sec
DETA 1.8999998 sec
TDO 1
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NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 ======
CPDPG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
PL13 16.50 dB
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F2 - Processing parameters
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SSB 0
LB 0
GB 0
PC 1.40

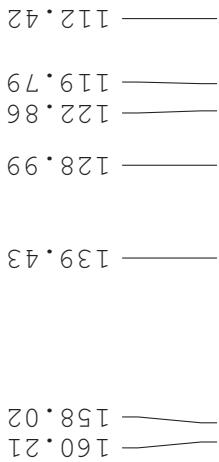
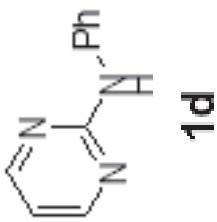
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7



QGY-1-31
C13CPD CDCL₃



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PULPROG zgpp30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 294.9 K
D1 2.0000000 sec
d1 0.0300000 sec
DETA 1.8999998 sec
TDO 1
===== CHANNEL f1 ======
NUC1 13C
P1 9.30 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 ======
CPDPFG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
F2 - Processing parameters
SI 32768
SF 125.757890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



HXM-2-5
PROTON CDC13

Current Data Parameters

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EXPNO	7
PROCNO	1

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TD	65536
SOLVENT	CDCl ₃
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DS	2
SWH	10330.578 Hz
ETR	0.157632 Hz
AQ	3.120407 sec
RG	114
DW	48.400 usec
DE	6.00 usec
TE	234.6 K
D1	1.0000000 sec
TD0	1

===== CHANNEL f1 =====

N1C1	1H
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P2J1	1.00 dB
SP01	500.1330685 MHz

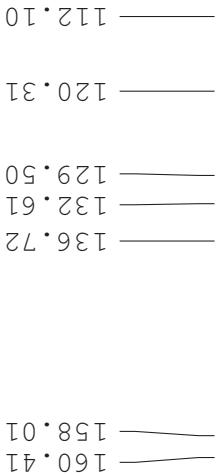
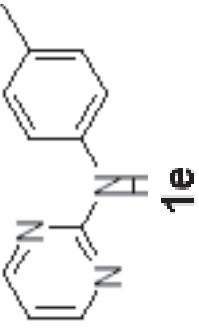
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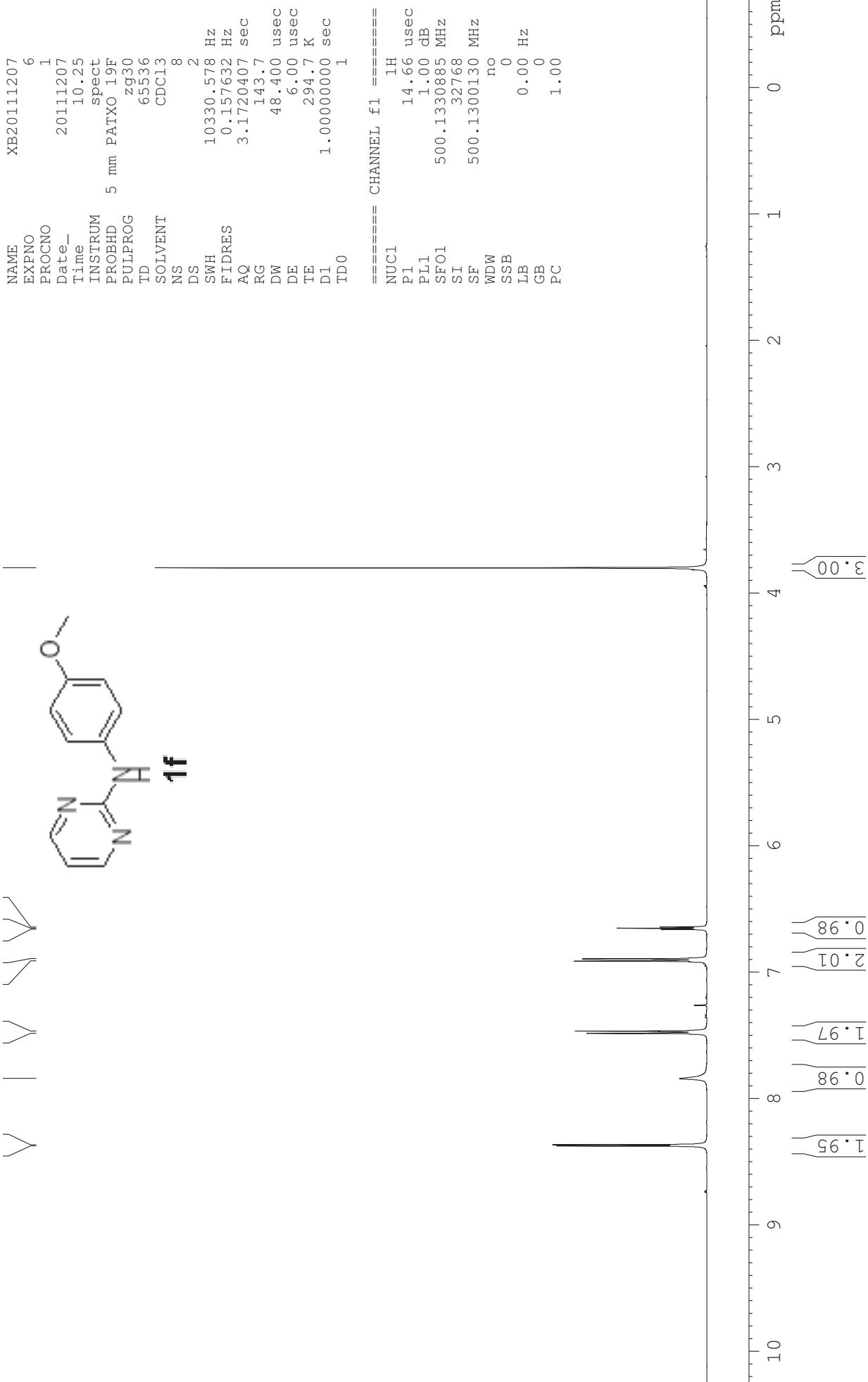
1e

HXM-2-5
C13CPD CDC13 I
20.87

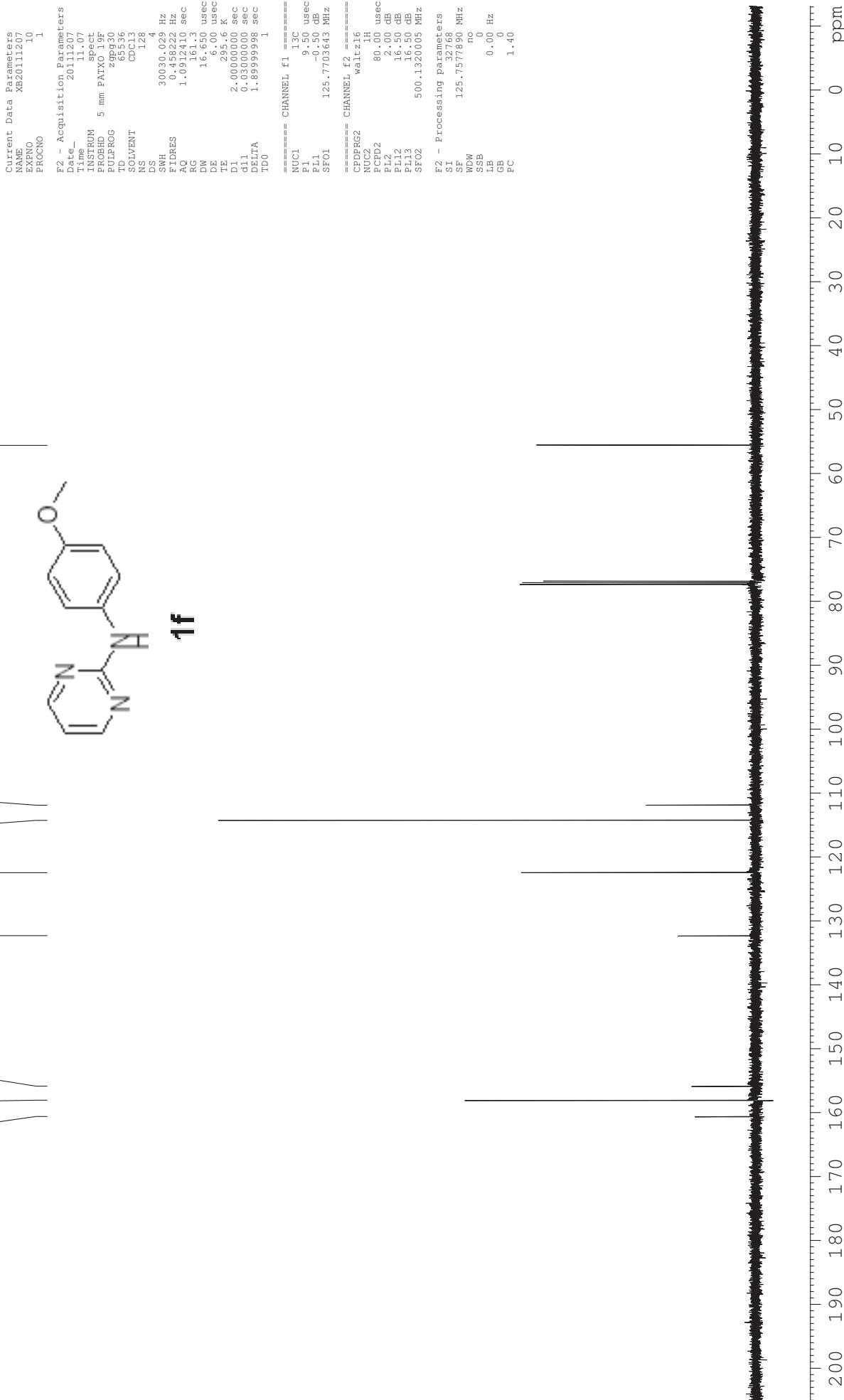
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SOLVENT CDCl3
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DS 4
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FIDRES 0.451222 Hz
AQ 1.0912410 sec
RG 181
DW 16.650 usec
DE 6.00 usec
TE 255.5 K
D1 2.0000000 sec
d11 0.0300000 sec
DETA 1.8939938 sec
TD0 1
===== CHANNEL f1 =====
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SF01 125.7701643 MHz
===== CHANNEL f2 =====
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NUC2 1H
PCPFG2 80.00 usec
PL2 2.00 dB
PL1,2 16.50 dB
PL1,3 16.50 dB
SFO2 500.1320005 MHz
F2 - Processing Parameters
SI 32768
SF 125.7577890 MHz
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HXM-2-3
PROTON CDC13



HXM-2-3
C13CPD CDC13



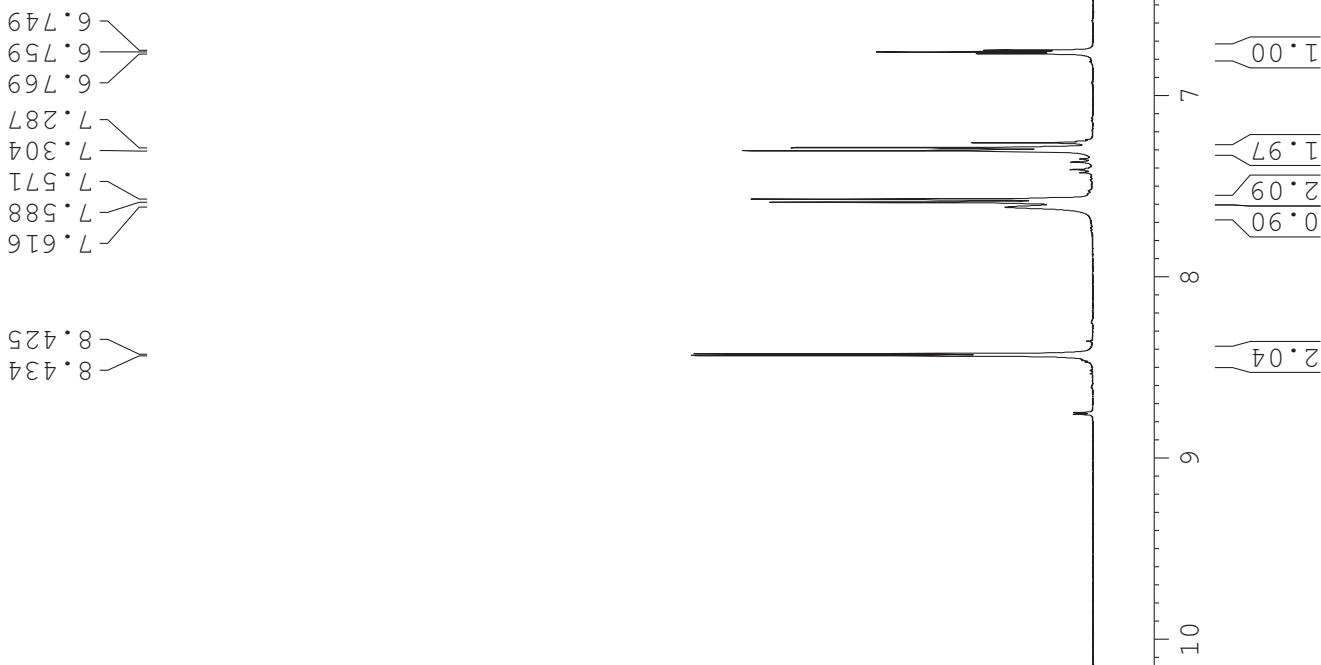
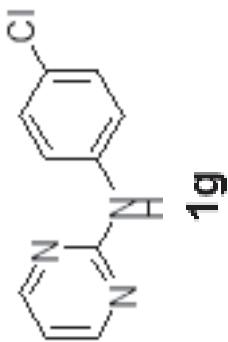
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PULPROG	PATXO 19F
TD	zg930
SOLVENT	65536
NS	CDC13
DS	16
SWH	2
FIDRES	10330.578 Hz
FAQ	0.157632 Hz
RG	3.1720407 sec
DW	287.4
DE	48.400 usec
TE	6.00 usec
DT	293.6 K
DDI	1.0000000 sec
DDFO	1

```

===== CHANNEL f1 =====
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PL1          1.00 dB
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SI           32768
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LB           0.00 Hz
GB           0
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```

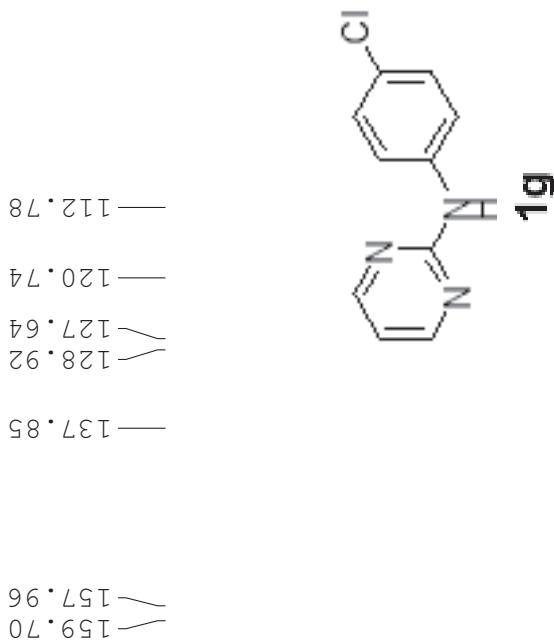


QGY-1-49
C13CPD CDC13

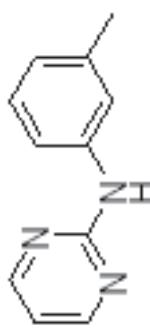
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SOLVENT CDCl3
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DS 4
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FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 295.1 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
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WDW EM
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GB 0

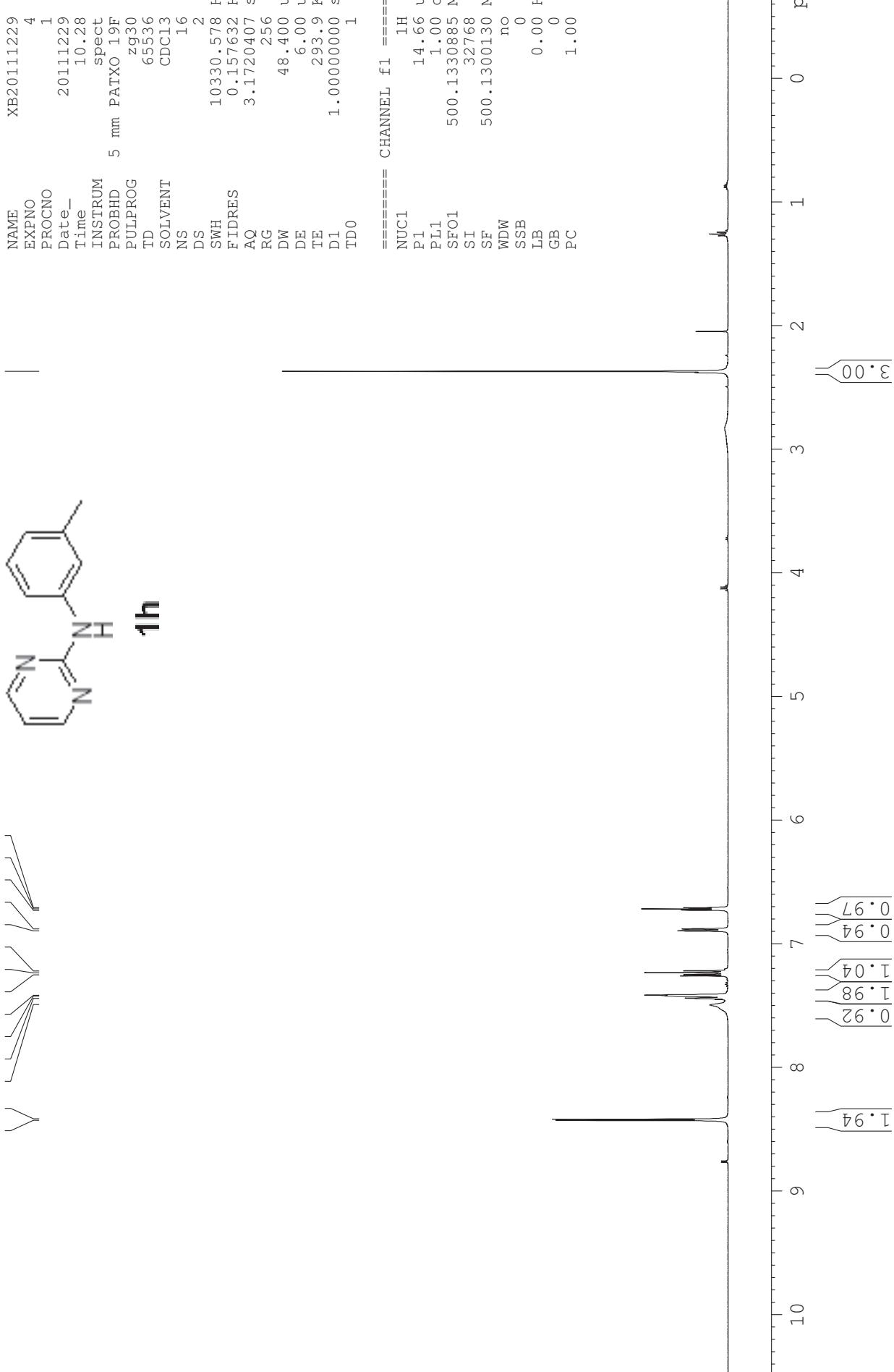


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PROTON CDCl₃

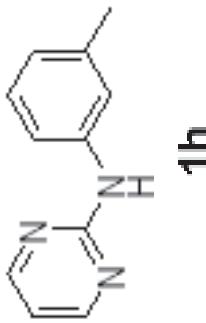


1h

8.426
8.416
7.492
7.439
7.422
7.415
7.250
7.234
7.219
6.896
6.880
6.728
6.718
6.708



FT-1-30
C13CPD CDC13



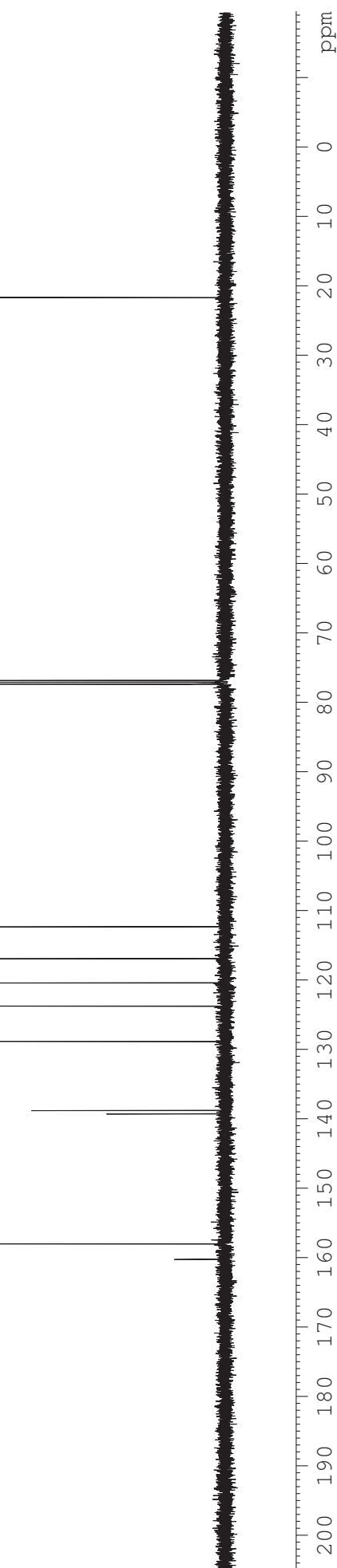
1h

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116.94
120.40
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128.82
138.80
139.29
158.00
160.25

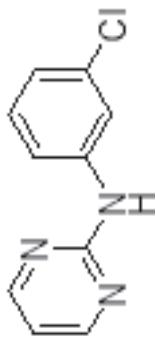
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SOLVENT CDCl3
NS 128
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FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 295.2 K
D1 2.0000000 sec
d11 0.0300000 sec
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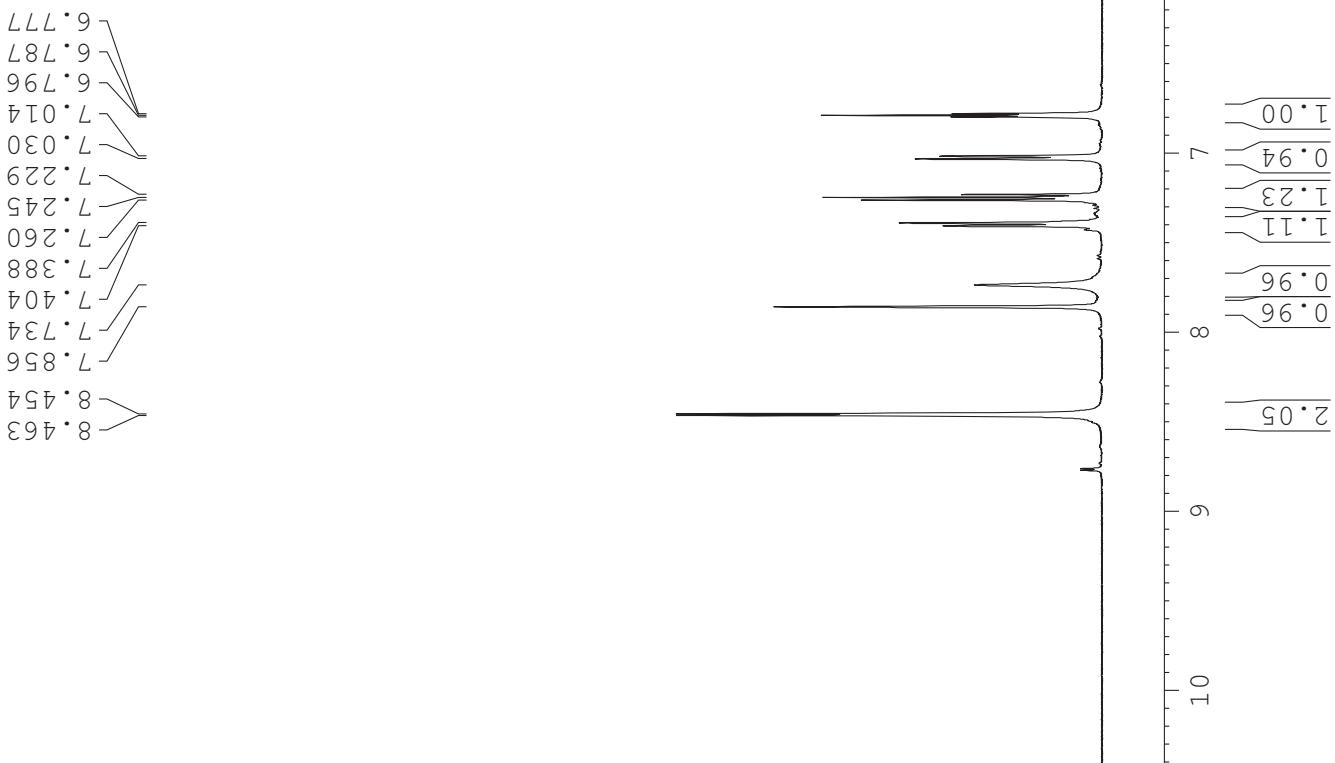
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PL13 16.50 dB
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SI 32768
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GB 1.0
PC 1.40



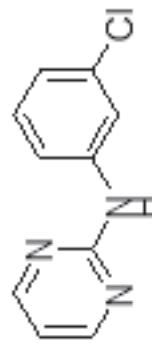
QGY-1-78-a
PROTON CDCl₃



1i



QGY-1-78-a
C13CPD CDC13



1i

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122.54
119.35
117.40
112.99

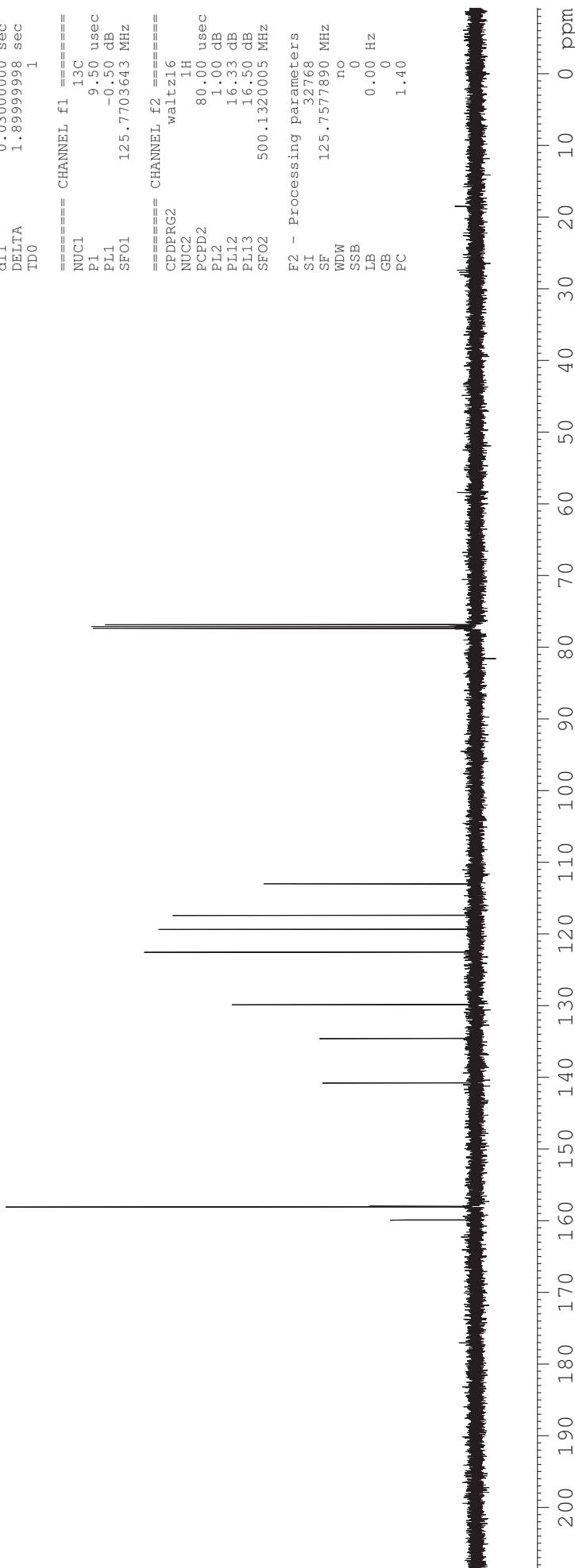
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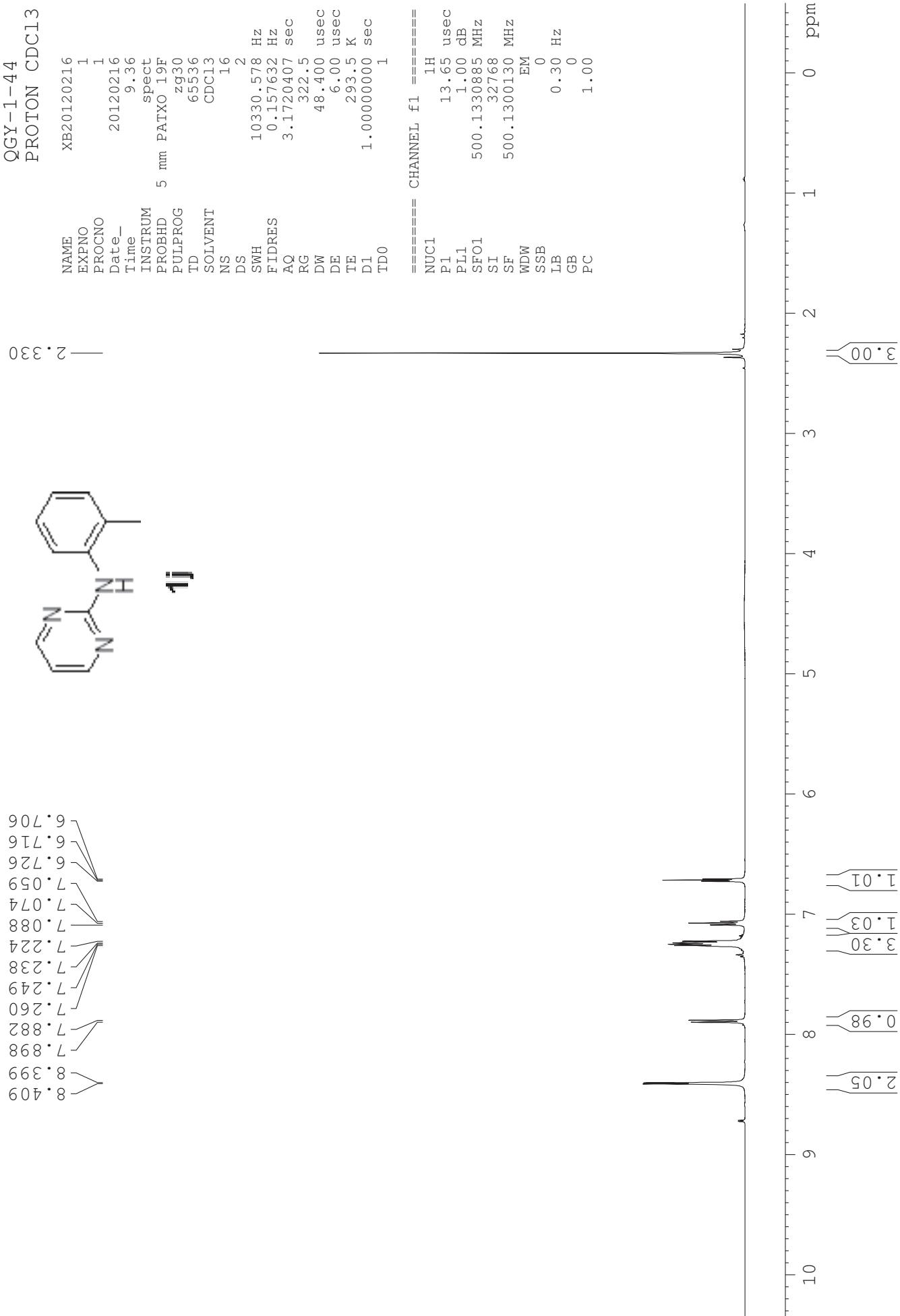
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FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 295.4 K
D1 2.0000000 sec
d11 0.0300000 sec
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SFO1 125.7703643 MHz

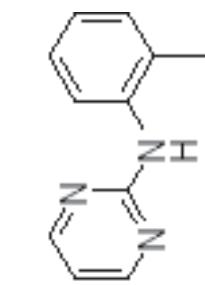
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PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.0 Hz
GB 1.0
PC 1.40





QGY-1-44
C13CPD CDC13



1j

— 112.20
— 122.69
— 124.33
— 126.66
— 129.77
— 130.64
— 137.14
— 158.11
— 160.61

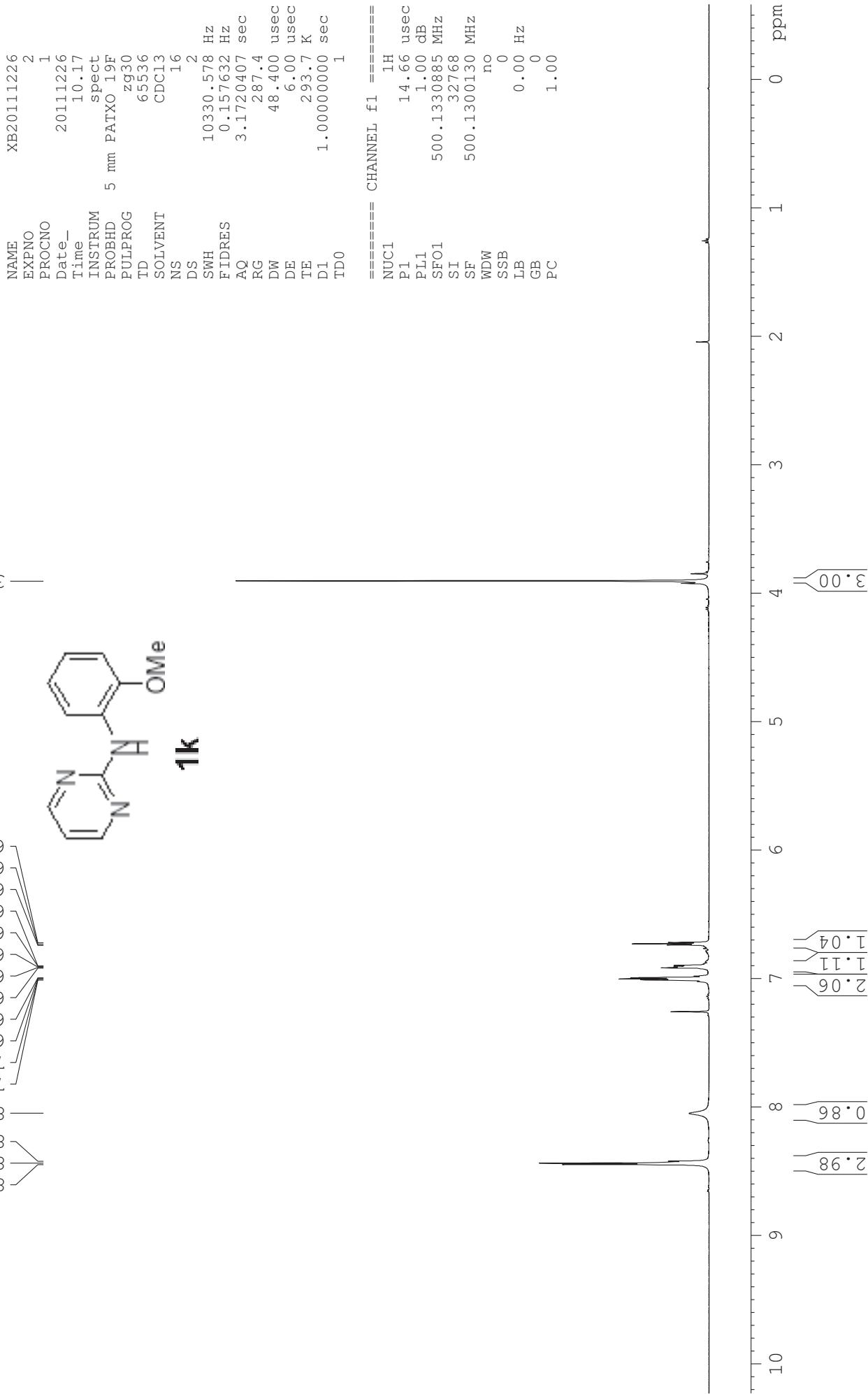
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FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 203.2
DW 16.650 usec
DE 6.00 usec
DEDE 295.4 K
TE 2.0000000 sec
D1 0.0300000 sec
d1 1
DELTA 1.8999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

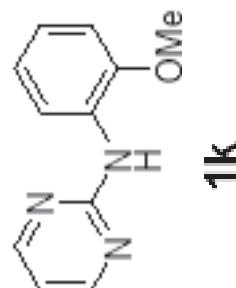
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.77 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40



QGY-1-45
PROTON CDC13



QGY-1-45
C13CPD CDC13



129.01
121.98
120.88
118.65
112.34
110.00
148.03
159.89
157.87

NAME XB20111231
EXPNO 8
PROCNO 1
Date_ 20111231
Time 11.03
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDC13
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 295.6 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPGR2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768 MHz
SF 125.7577890 MHz
WDW no
SSB 0 Hz
LB 0.00 Hz
GB 0

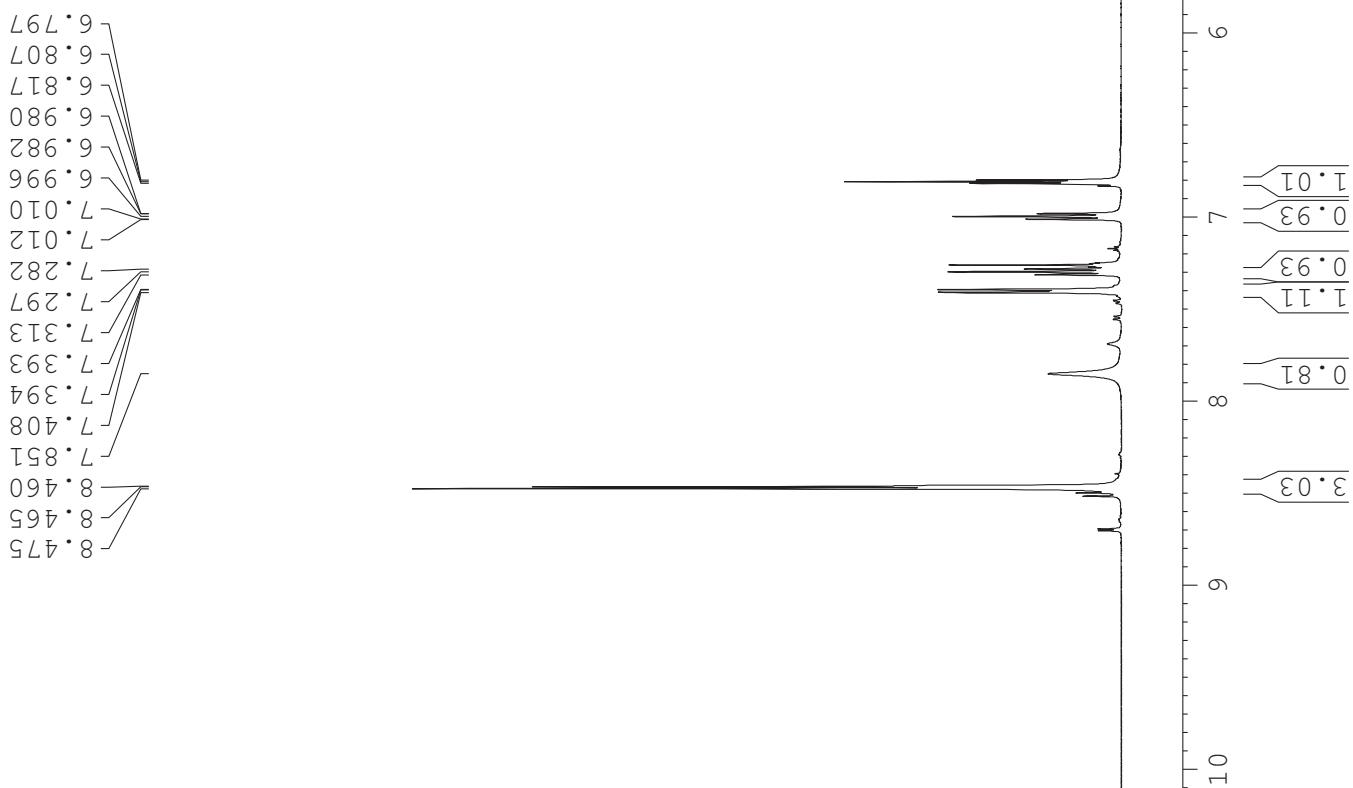
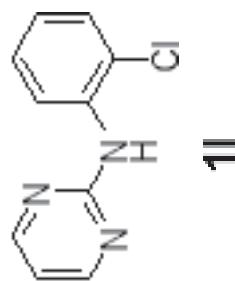


QGY-1-75
PROTON CDC13

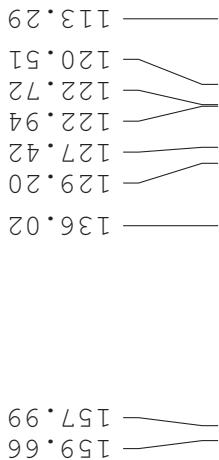
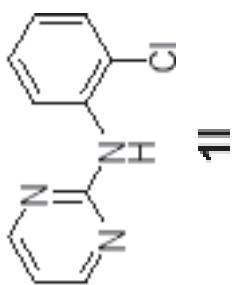
NAME XB20120227
EXPNO 3
PROCNO 1
Date 20120227
Time 10.41
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zg30
TD 65536
SOLVENT C6D13
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.1157632 Hz
AQ 3.1720407 sec
RG 362
DW 48.400 usec
DE 6.00 usec
TE 293.5 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====

NUC1 1H
P1 13.70 usec
PL1 1.00 dB
SFO1 500.1330885 MHz
SI 32768
SF 500.1300129 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



QGY-1-75
C13CPD CDC13



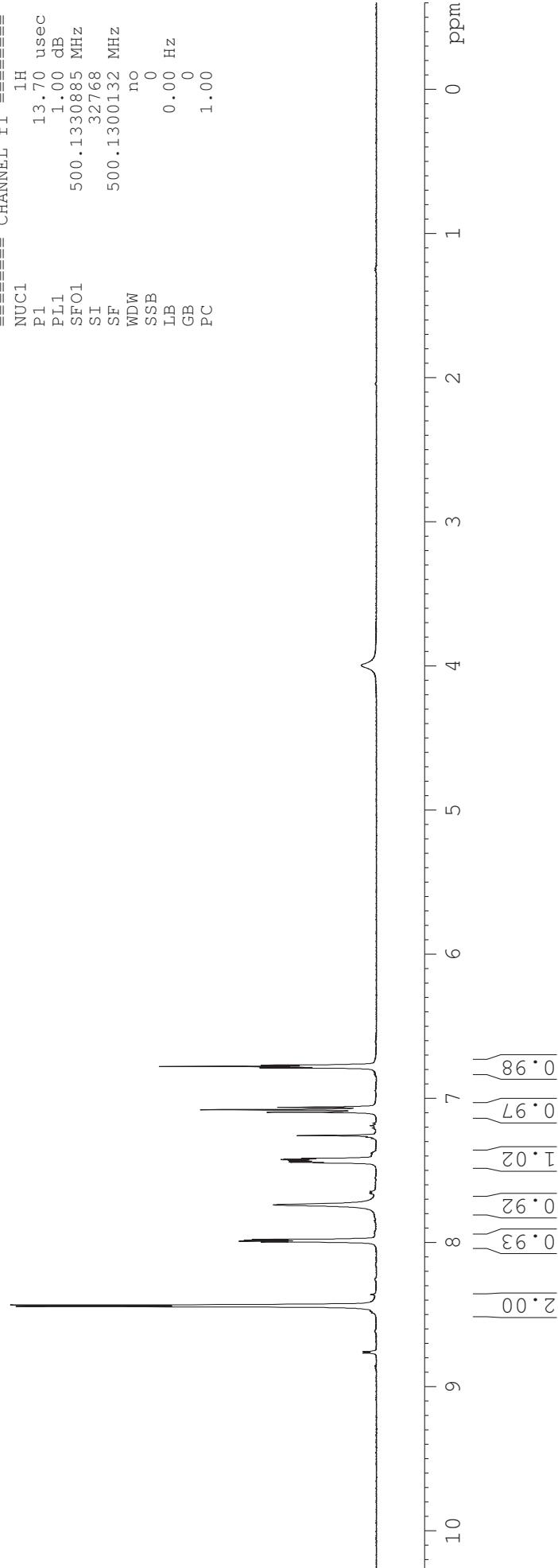
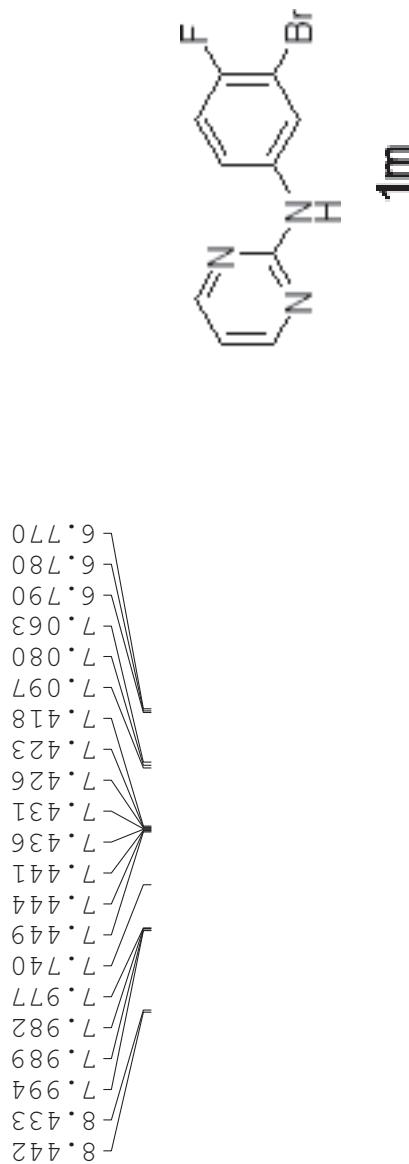
Current Data Parameters
NAME XB20120228
EXPNO 23
PROCNO 1
F2 - Acquisition Parameters
Date 20120228
Time 20.47
INSTRUM spect
PROBHD 5 mm PAXO 19F
PULPROG zgpp30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 101.6
DW 16.650 usec
DE 6.00 usec
TE 295.4 K
D1 2.0000000 sec
d1 0.0300000 sec
DELTA 1.8999998 sec
TD0 1
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 1.0
PC 1.40



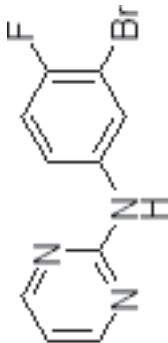
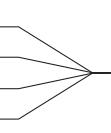
QGY-1-78-b
PROTON CDC13

NAME xb20120331
EXPNO 8
PROCNO 1
Date_ 20120331
Time 13.30
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.172047 sec
RG 362
DW 48.400 usec
DE 6.00 usec
TE 295.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.70 usec
PL1 1.00 dB
SFO1 500.1330885 MHz
SI 32768
SF 500.1300132 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

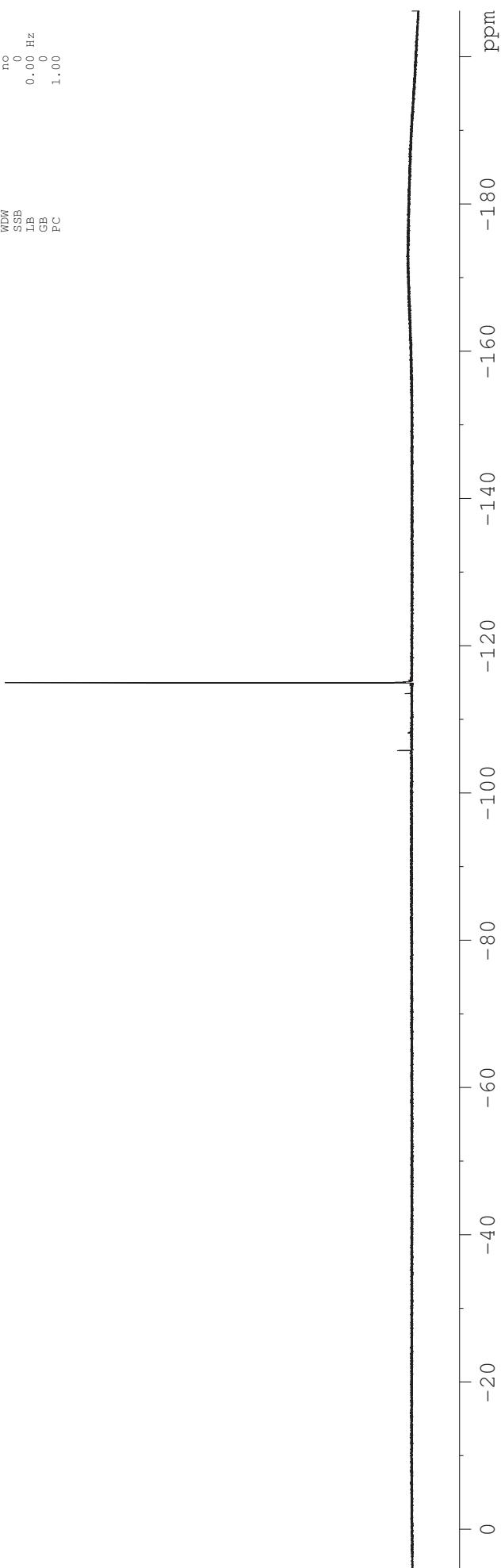


QGY-1-78-b
19F dft CDC13]

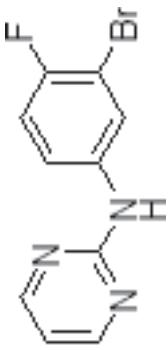


1m

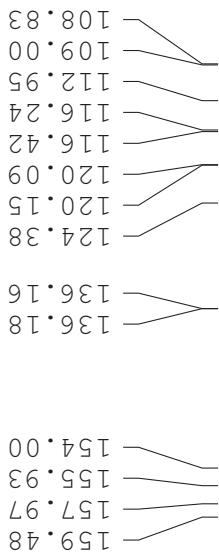
Current Data Parameters
NAME xb2012401
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20120401
Time 17.41
INSTRUM spect
PROBHD 5 mm PABX0 19F
PULPROG 131072
TD 131072
SOLVENT CDCl3
NS 8
DS 4
SWH 100000.000 Hz
FIDRES 0.762939 Hz
AQ 0.6554150 sec
RG 322.5
DW 5.000 usec
DE 6.00 usec
TE 294.4 K
D1 1.0000000 sec
TDO 1
===== CHANNEL f1 =====
NUC1 19F
P1 19.30 usec
PL1 4.00 dB
SFO1 470.545180 MHz
F2 - Processing parameters
SI 6536
SF 470.5923770 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



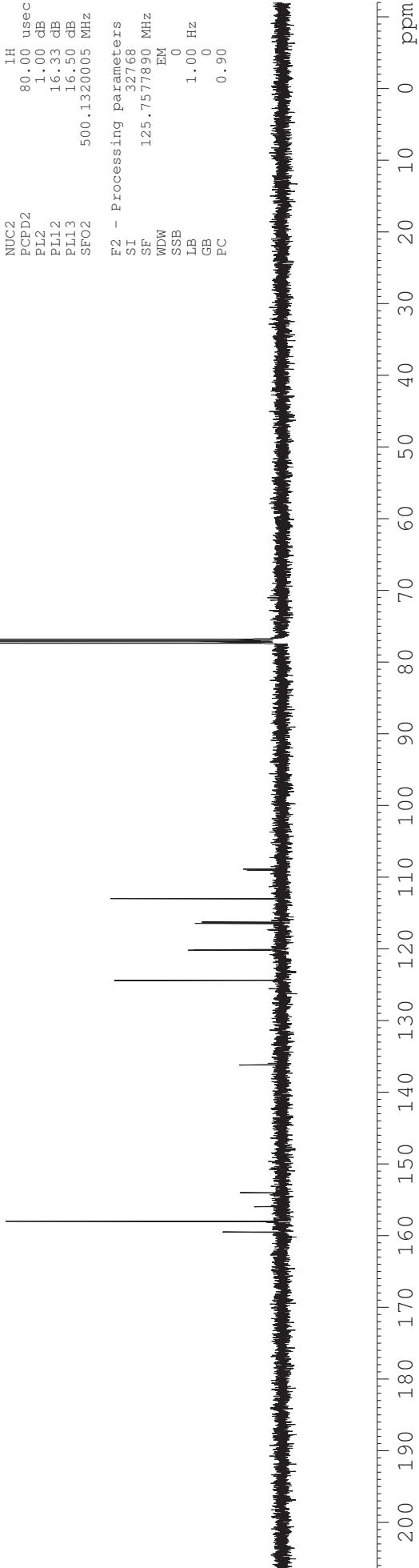
QGY-1-78-b
C13CPD CDC13



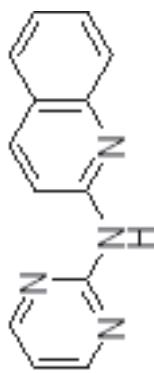
1m



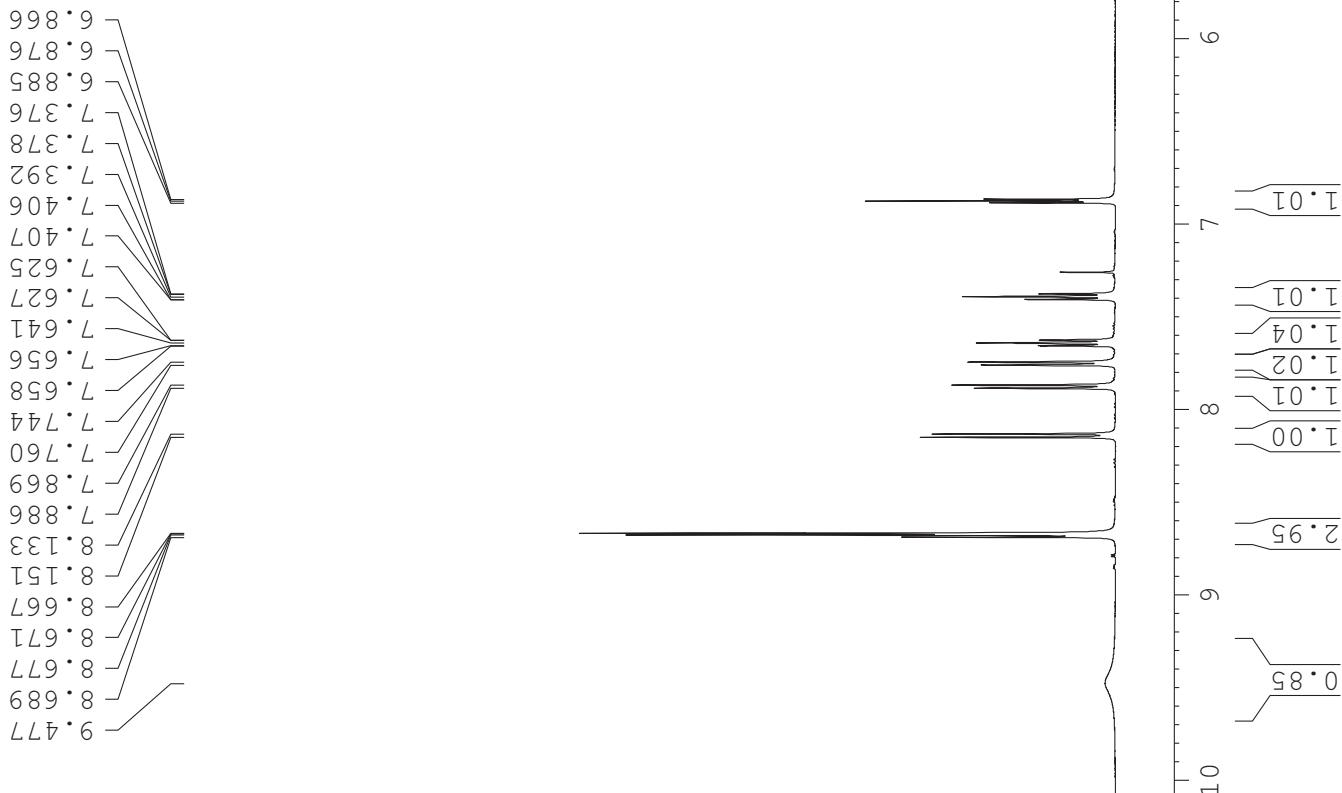
Current Data Parameters
NAME xb20120401
EXPNO 8
PROCNO 1
F2 - Acquisition Parameters
Date_ 20120401
Time 18.05
INSTRUM spect
PROBHD 5 mm PABX0 19F
PULPROG zgpp30
TD 65536
SOLVENT C6D13
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.459222 Hz
AQ 1.0912410 sec
RG 1.14
DW 16.650 usec
DE 6.00 usec
TE 295.7 K
D1 0.0300000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TDO 1
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 0.90



FT-1-96
PROTON CDC13



5



FT-1-96
C13CPD CDC13 I

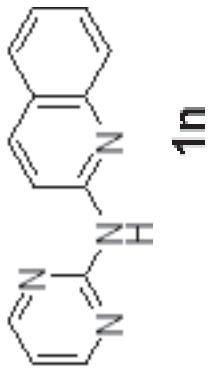
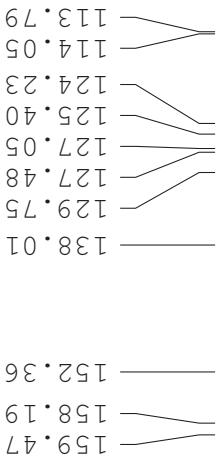
Current Data Parameters
NAME XB20120516
EXPNO 6
PROCNO 1

F2 - Acquisition Parameters
Date 20120516
Time 10.16
INSTRUM spect
PROBHD 5 mm PAXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDC13
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 128
DW 16.650 usec
DE 6.00 usec
TE 296.5 K
D1 2.0000000 sec
d1 0.3000000 sec
DELTA 1.8999998 sec
TD0 1

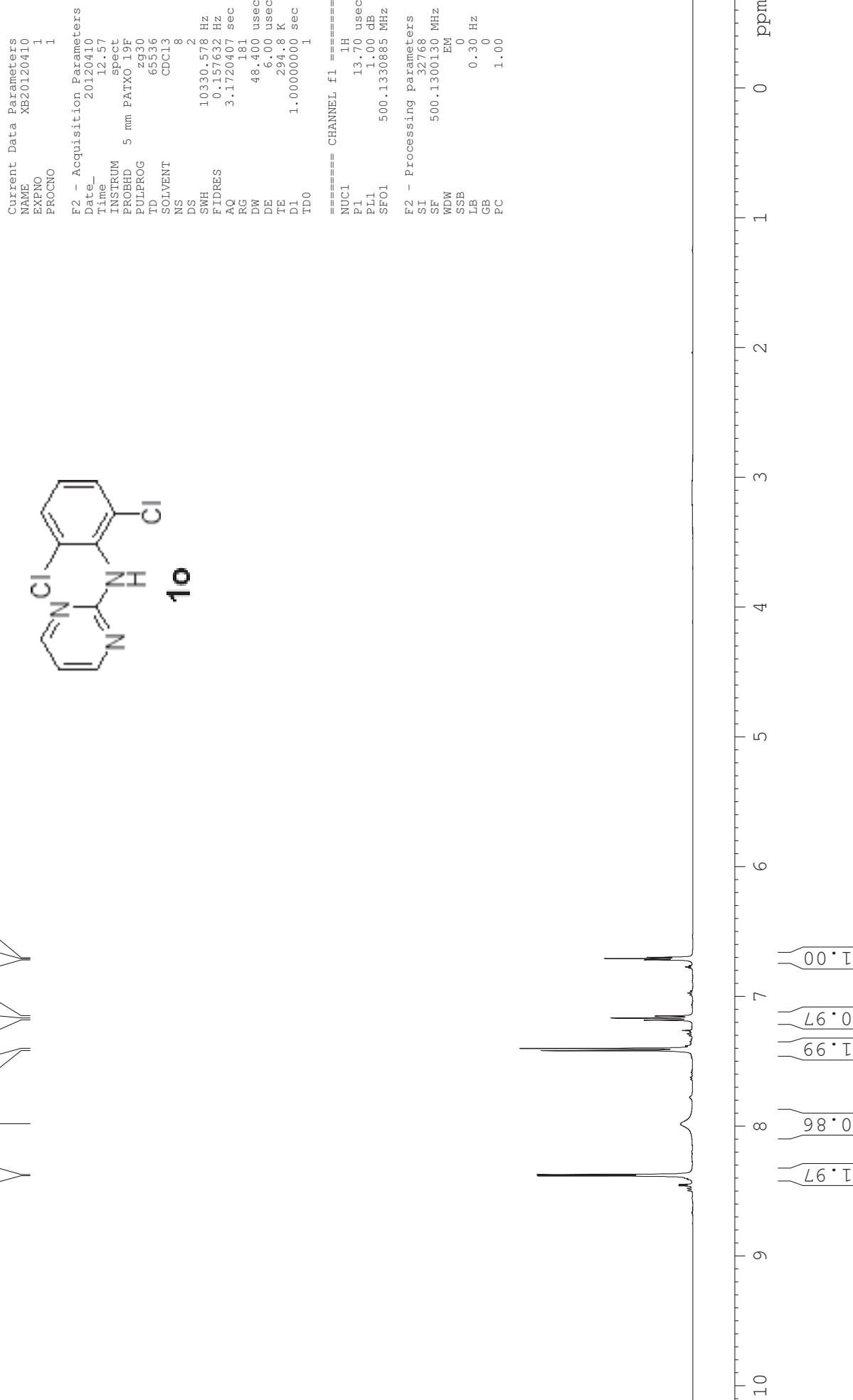
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SF01 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SF02 500.132005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0.0



QGY-1-102
PROTON CDCl₃



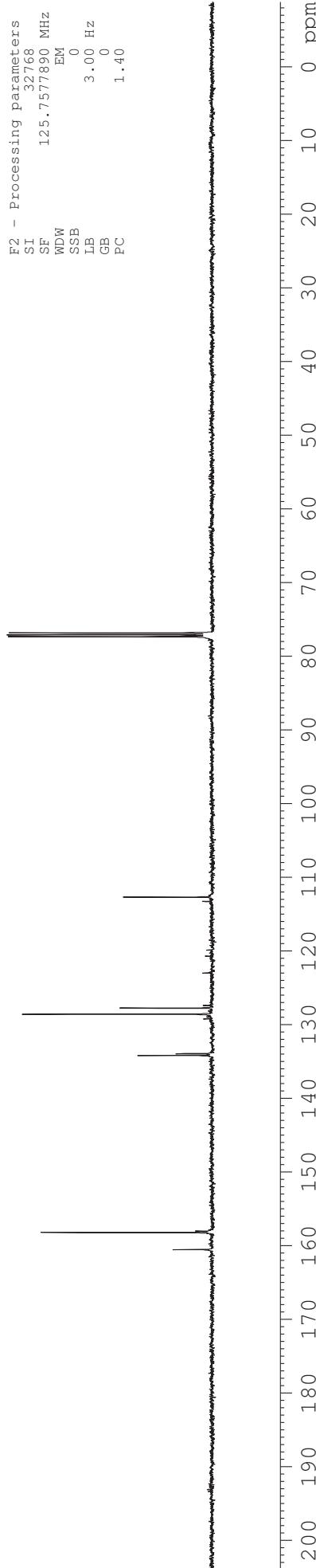
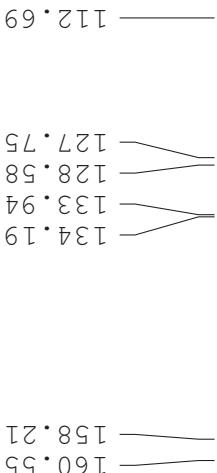
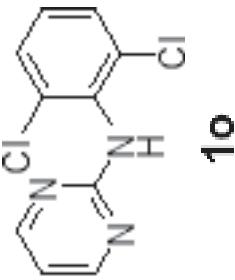
QGY-1-102
C13CPD CDCC13

Current Data Parameters
NAME XB20120410 4
EXPNO 1
PROCNO

F2 - Acquisition Parameters
Date_ 20120410
Time 13.21
INSTRUM spect
PROBHD 5 mm PABXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
E1DRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 296.2 K
D1 2.0000000 sec
Q1 0.0300000 sec
DELTA 1.89399998 sec
TDD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 3.00 Hz
GB 0
PC 1.40



FT-1-47
PROTON Acetone

NAME
EXPNO
PROCNO
Date
Time

20120223
17.37

INSTRUM
PROBHD
PULPROG
TD

5 mm PATXO 19F
zg30
65536

SOLVENT
NS
DS

Acetone
16
2
10330.578 Hz

FIDRES
AQ

0.1157632 Hz
3.1720407 sec

RG
DW

322.5
48.400 usec

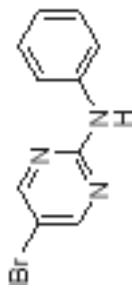
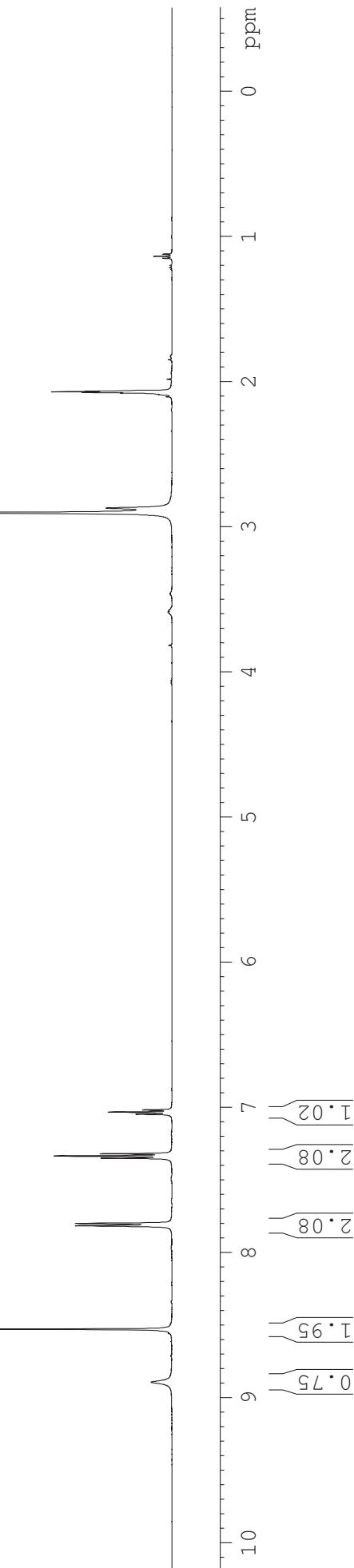
DE
TE

6.00 usec
2.93.8 K

D1
TDO

1.0000000 sec
1
1

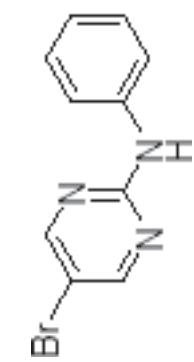
===== CHANNEL f1 =====
NUC1
P1
PL1
SFO1
SI
SF
WDW
SSB
LB
GB
PC



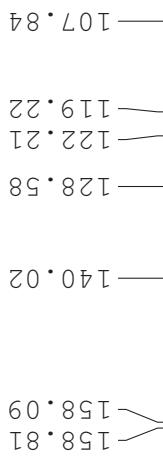
7.018
7.033
7.048
7.320
7.351
7.355
7.800
7.816

8.529
8.894

FT-1-47
C13CPD Acetone



1p

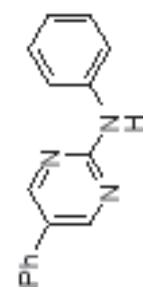


NAME XB20120228
EXPNO 25
PROCNO 1
Date_ 20120228
Time 21.01
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgppg30
TD 65536
SOLVENT Acetone
NS 128
DS 4
SWH 300030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 295.4 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 ======
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 ======
CPDPRG2
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



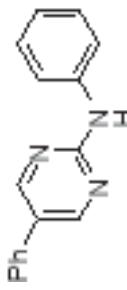
FT-1-70
PROTON CDCl₃



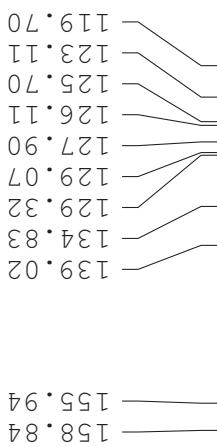
1q



FT-1-70
C13CPD CDC13



1q



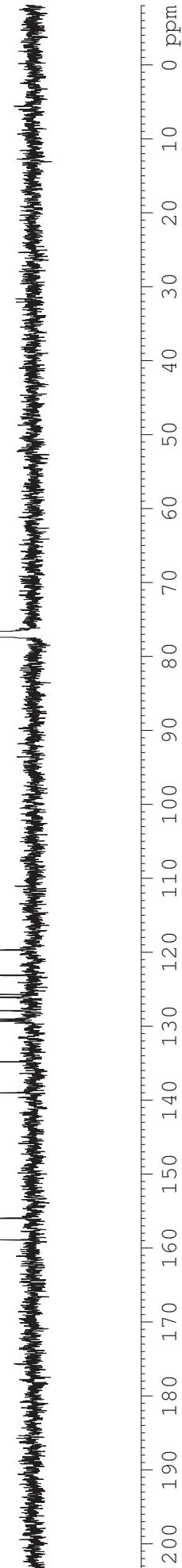
Current Data Parameters
NAME XB2120327
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
Date 2012027
Time 13:51
INSTRUM spect
PROBHD 5 mm PABTXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.45822 Hz
AQ 1.0912410 sec
RG 16.13
DW 16.550 usec
DE 6.00 usec
TE 296.2 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

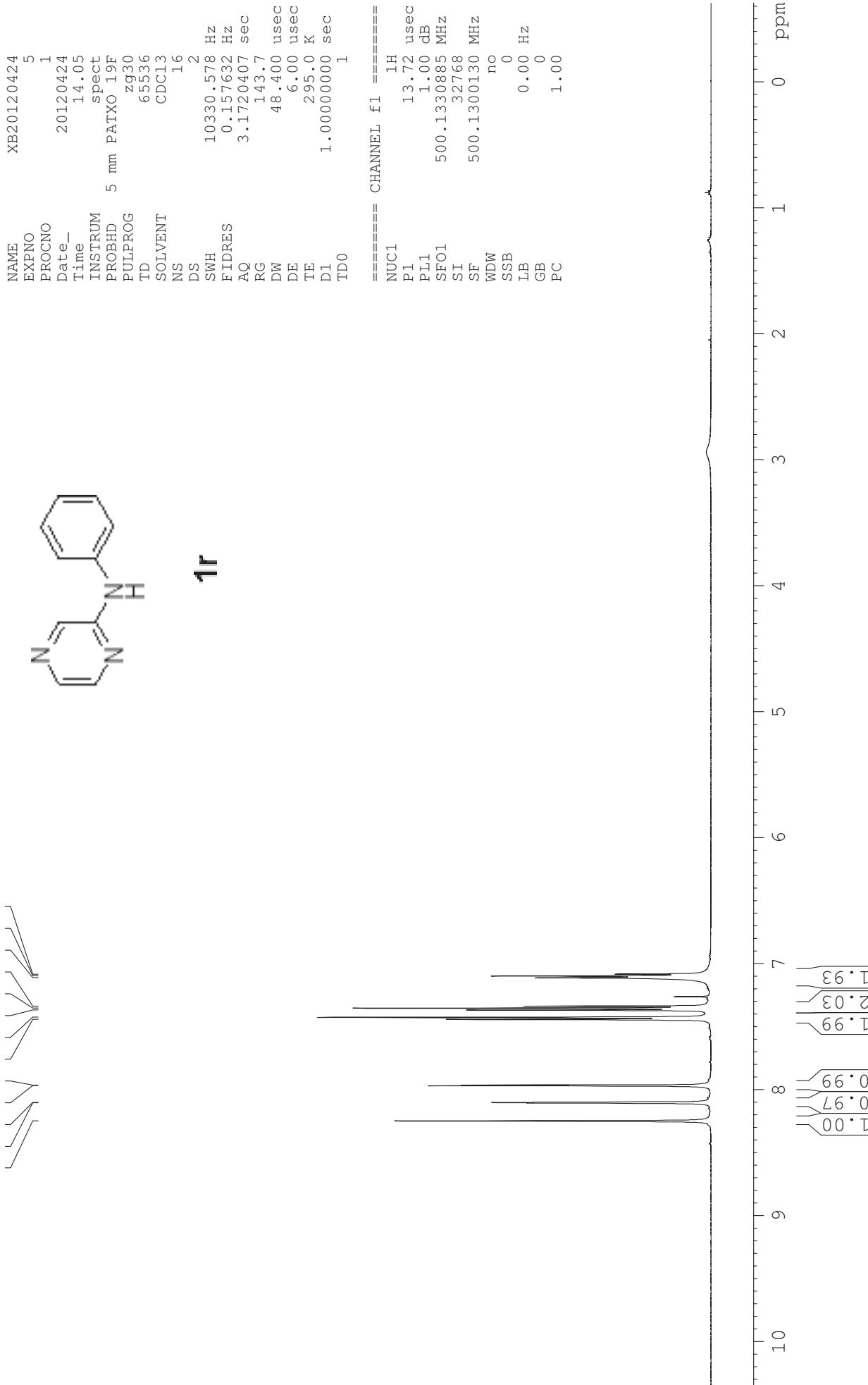
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 125.7703643 MHz
SFO1

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SF02 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.757790 MHz
WDW EM
SSB 3 0
LB 3.00 Hz
GB 0
PC 0.60



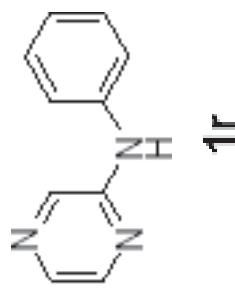
FT-1-72
PROTON CDCl₃



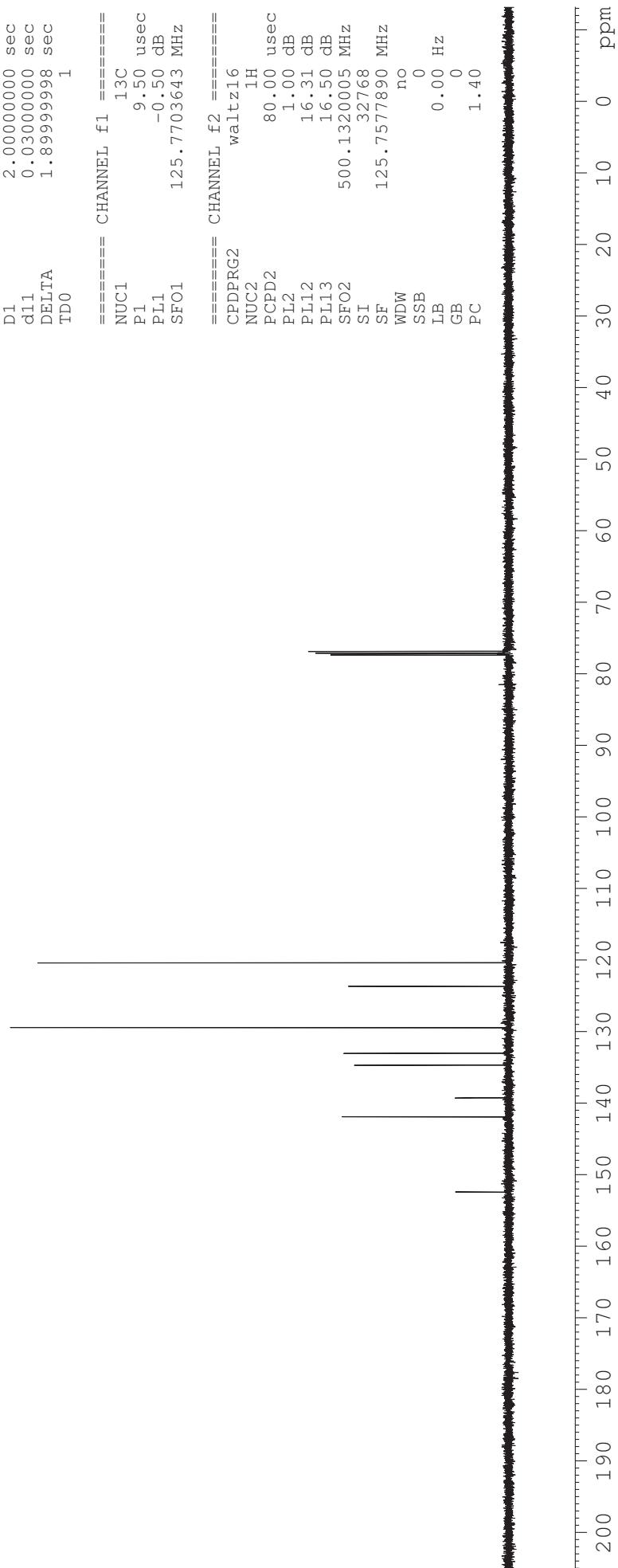
FT-1-72
C13CPD CDCl₃

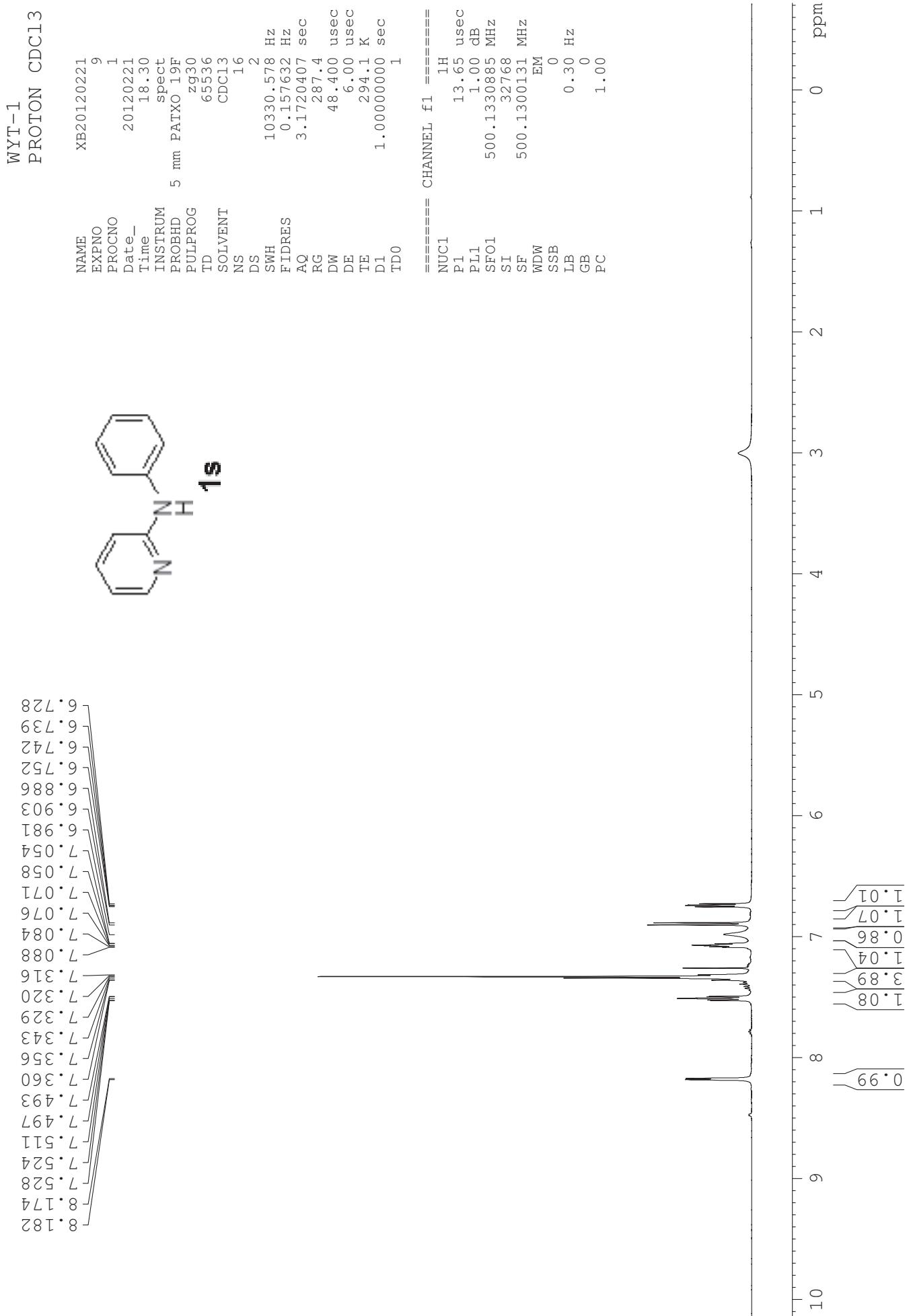
NAME XB20120424
EXPNO 7
PROCNO 1
Date 20120424
Time 14.16
INSTRUM spect
PROBHD 5 mm PABXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 362
DW 16.650 usec
DE 6.00 usec
DEDE 296.2 K
TE 2.00000000 sec
D1 0.03000000 sec
d11 1.89999998 sec
DELTA 1.89999998 sec
TDD 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768 MHz
SF 125.7577890 MHz
WDW no
SSB 0 0.00 Hz
LB 0 0
GB 0 0
PC 1.40

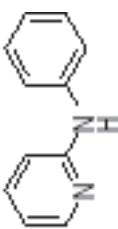


— 120.35
— 123.65
— 129.41
— 132.99
— 134.65
— 139.22
— 141.87
— 152.39





WYT-1
C13CPD CDC13

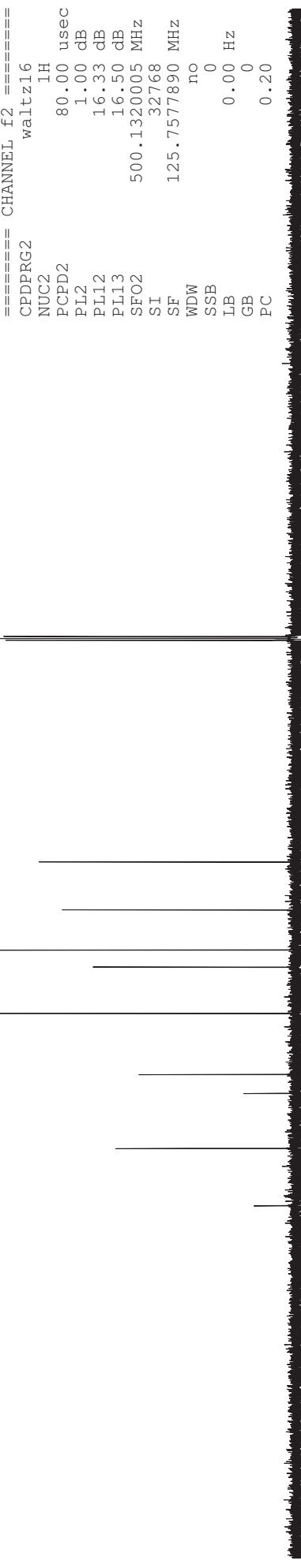


1s

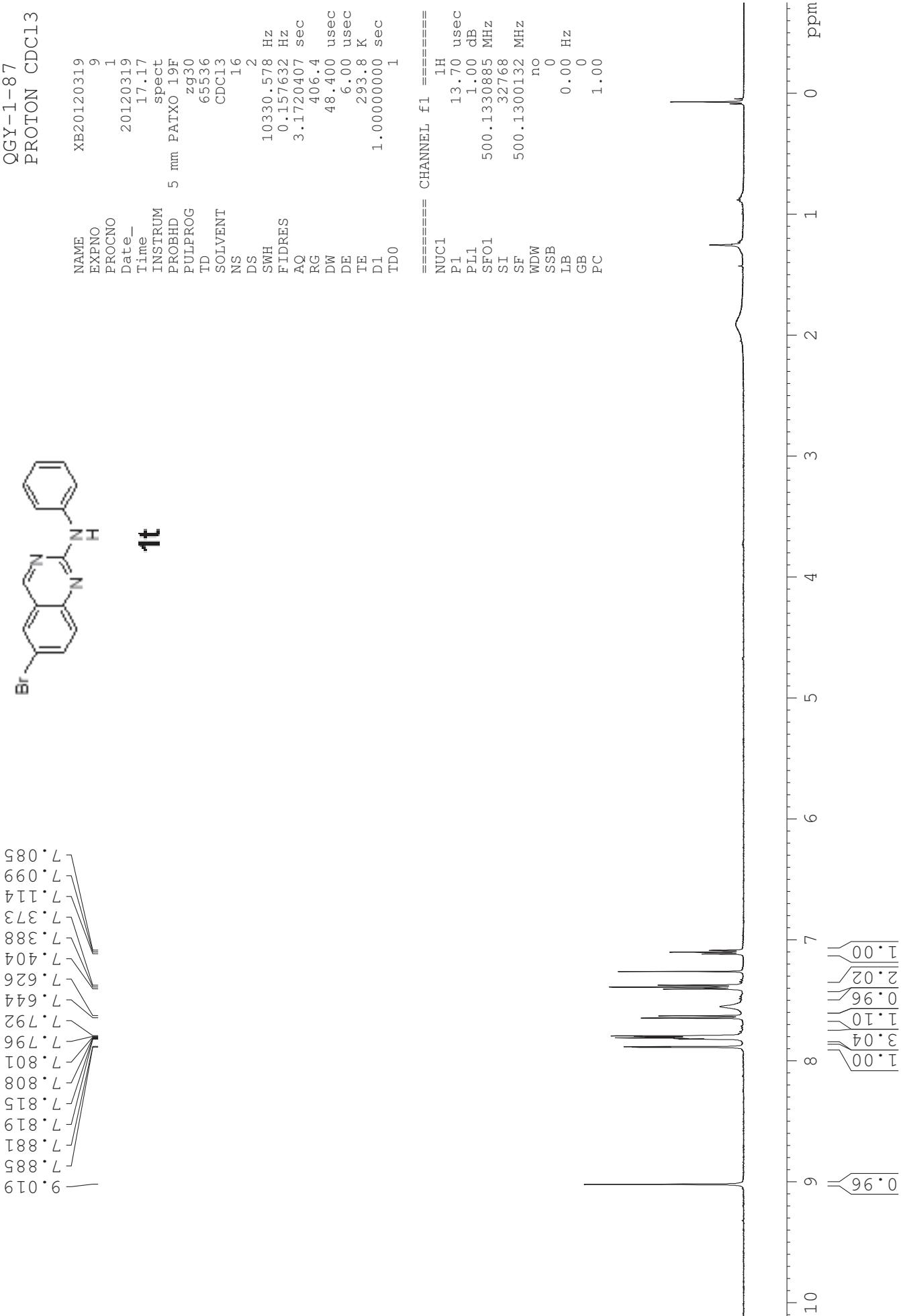
— 108.24
— 114.92
— 120.52
— 122.90
— 129.34
— 137.86
— 140.50
— 148.17
— 156.14

NAME XB20120227
EXPNO 7
PROCNO 1
Date_ 20120227
Time 11.38
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDC13
NS 110
DS 4
SWH 300030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 294.8 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768 MHz
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 0.20



200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm



QGY-1-87
C13CPD CDC13

NAME XB20120322

EXPNO 5

PROCNO 1

Date 20120322

Time 10.10

INSTRUM spect

PROBHD 5 mm PATXO 19F

PULPROG zppg30

TD 65536

SOLVENT CDCl3

NS 128

DS 4

SWH 300030.029 Hz

FIDRES 0.458222 Hz

AQ 1.0912410 sec

RG 114

DW 16.650 usec

DE 6.00 usec

TE 2.95.4 K

D1 2.0000000 sec

d11 0.0300000 sec

DELTA 1.8999998 sec

TDD0 1

===== CHANNEL f1 =====

NUC1 13C

P1 9.50 usec

PL1 -0.50 dB

SFO1 125.7703643 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16

NUC2 1H

PCPD2 80.00 usec

PL2 1.00 dB

PL12 16.33 dB

PL13 16.50 dB

SFO2 500.1320005 MHz

SI 32768

SF 125.7577890 MHz

WDW no

SSB 0

LB 0.00 Hz

GB 0

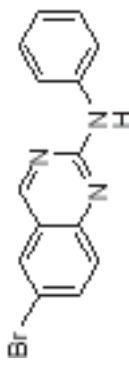
PC 0

PE 0

PO 0

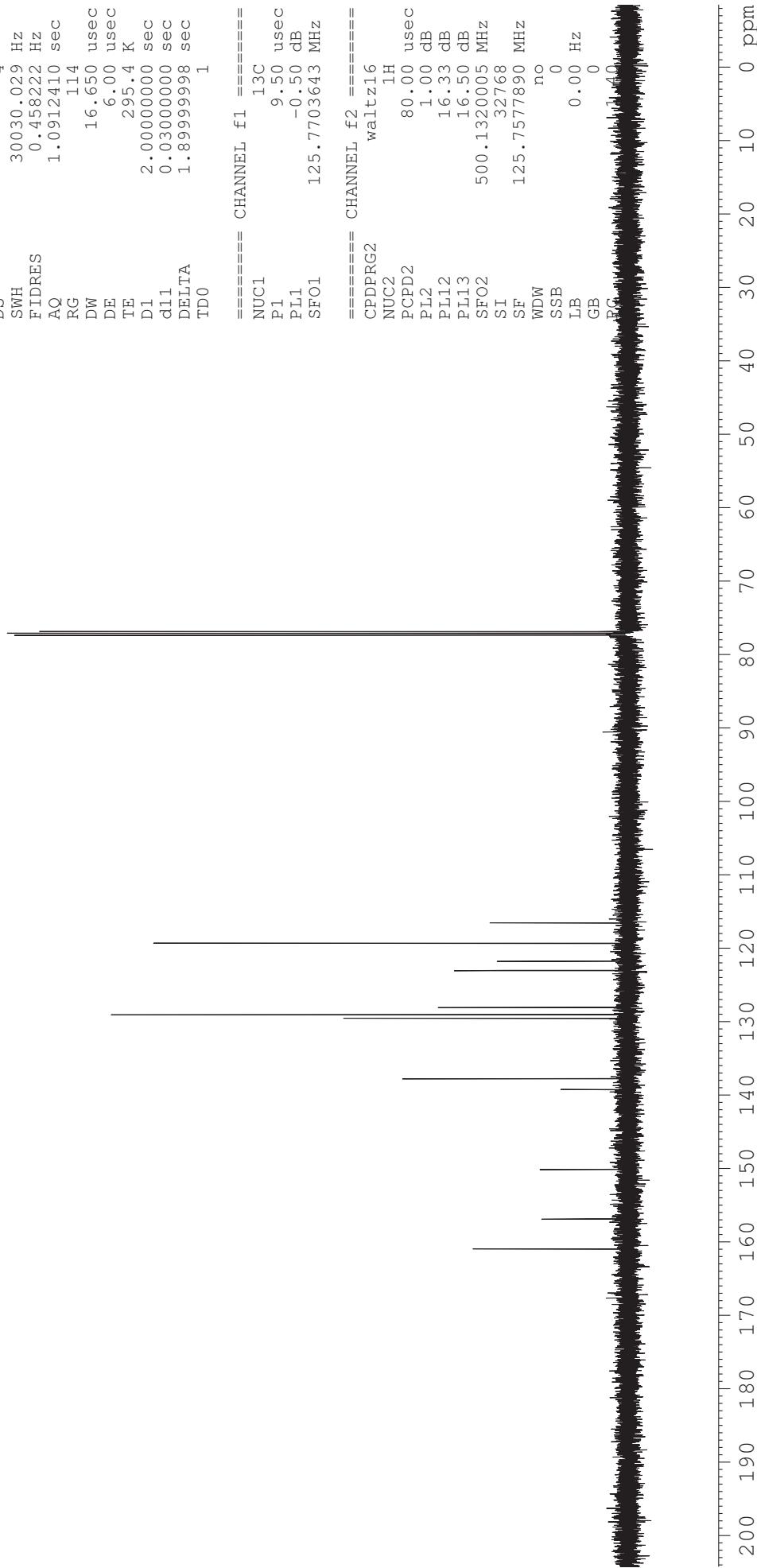
PD 0

PP 0



1t

116.52
119.28
121.72
122.99
128.03
129.03
129.49
137.73
139.20
150.09
156.84
160.91

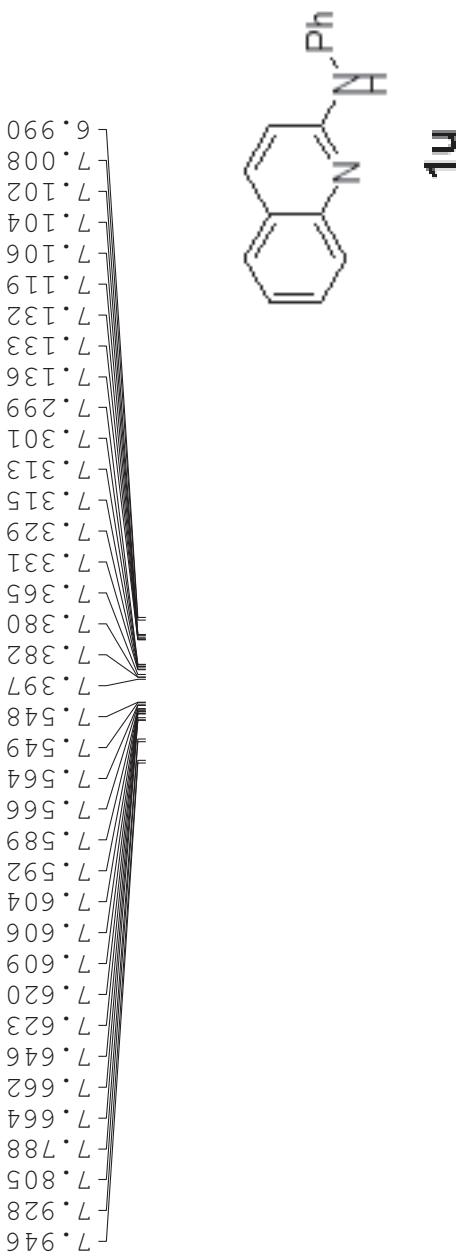


HXM-2-280
PROTON CDC13

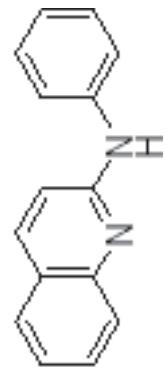
NAME XB20120504
EXPNO 5
PROCNO 1
Date 20120504
Time 15.30
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1720407 sec
RG 287.4
DW 48.400 usec
DE 6.00 usec
TE 294.8 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.72 usec
PL1 1.00 dB
SFO1 500.11330885 MHz
SI 32768
SF 500.11300129 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC

— 2.496 —



hxm-2-280
C13CPD CDCl₃



1u

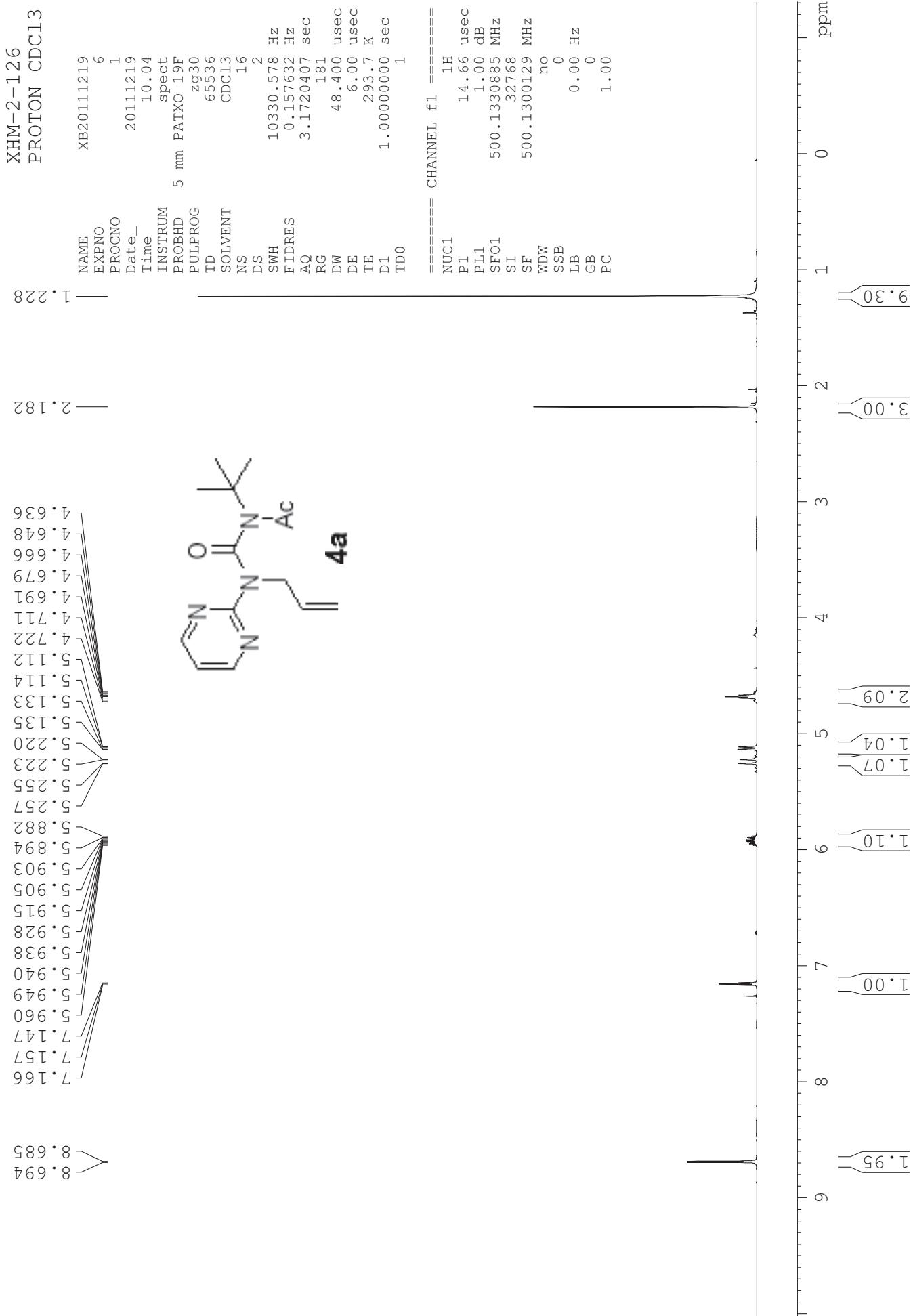
```
NAME          xb20120507
EXPNQ        4
PROCNO      1
Date_        20120507
Time         9.57
INSTRUM     spect
PROBHD      5 mm PATXO 19F
PULPROG     zgppg30
TD          65536
SOLVENT      CDCl3
NS          128
DS          4
SWH         30030.029 Hz
FIDRES      0.458222 Hz
AQ          1.0912410 sec
RG          143.7
DW          16.650 usec
DE          6.00
TE          296.2 K
D1          2.0000000 sec
d11         0.0300000 sec
DELTA       1.89999998 sec
TD0          1

===== CHANNEL f1 =====
NUC1         13C
P1          9.50 usec
PL1        -0.50 dB
SFO1       125.7703643 MHz

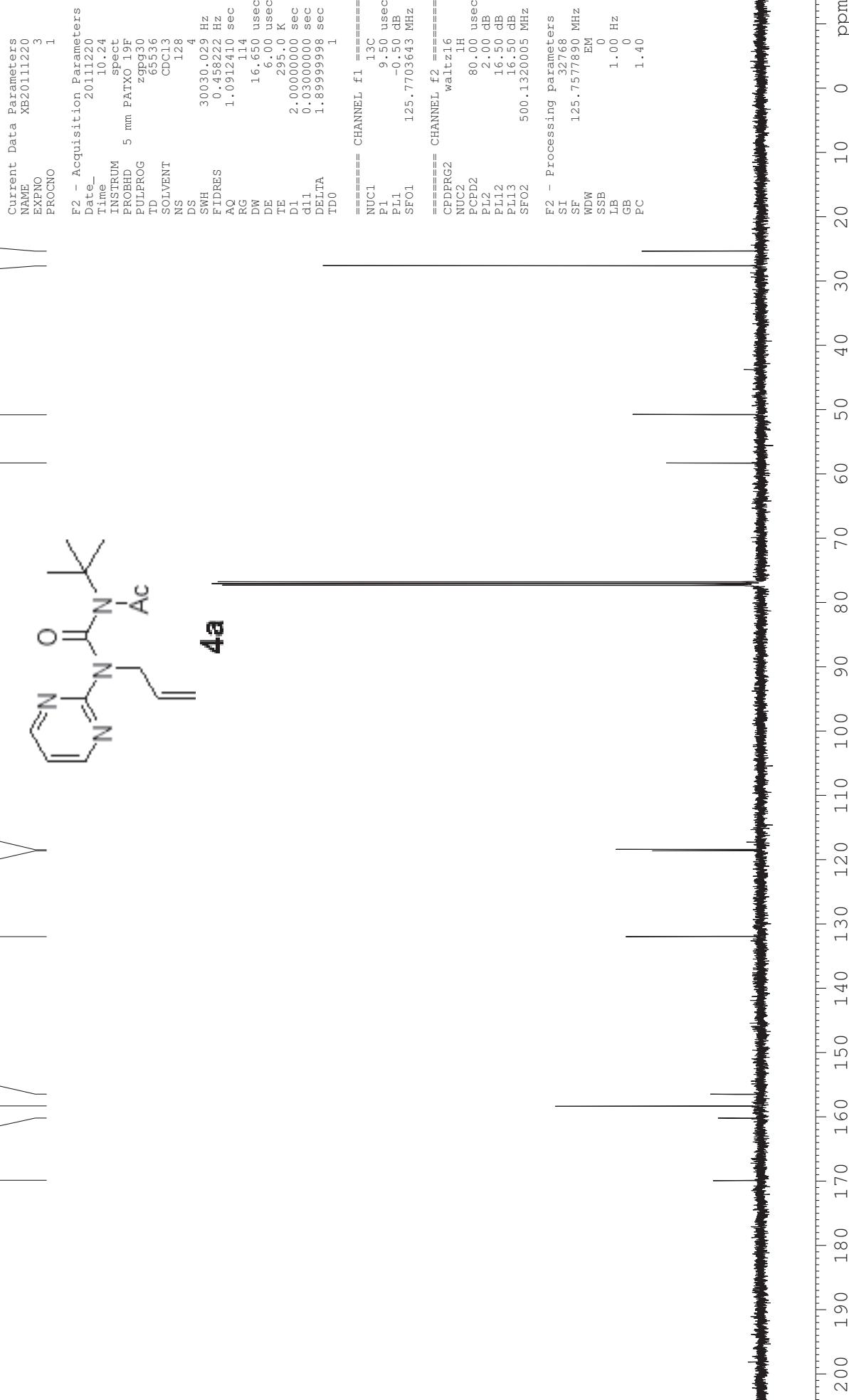
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2         1H
PCPD2       80.00 usec
PL2          1.00 dB
PL12        16.31 dB
PL13        16.50 dB
SFO2       500.1320005 MHz
SI          32768
SF       125.7577890 MHz
WDW         EM
SSB          0
LB          1.00 Hz
GB          0
PC         1.40
```



154.48
147.59
140.20
137.84
129.87
129.29
127.50
126.66
124.16
123.19
123.18
120.61
111.77

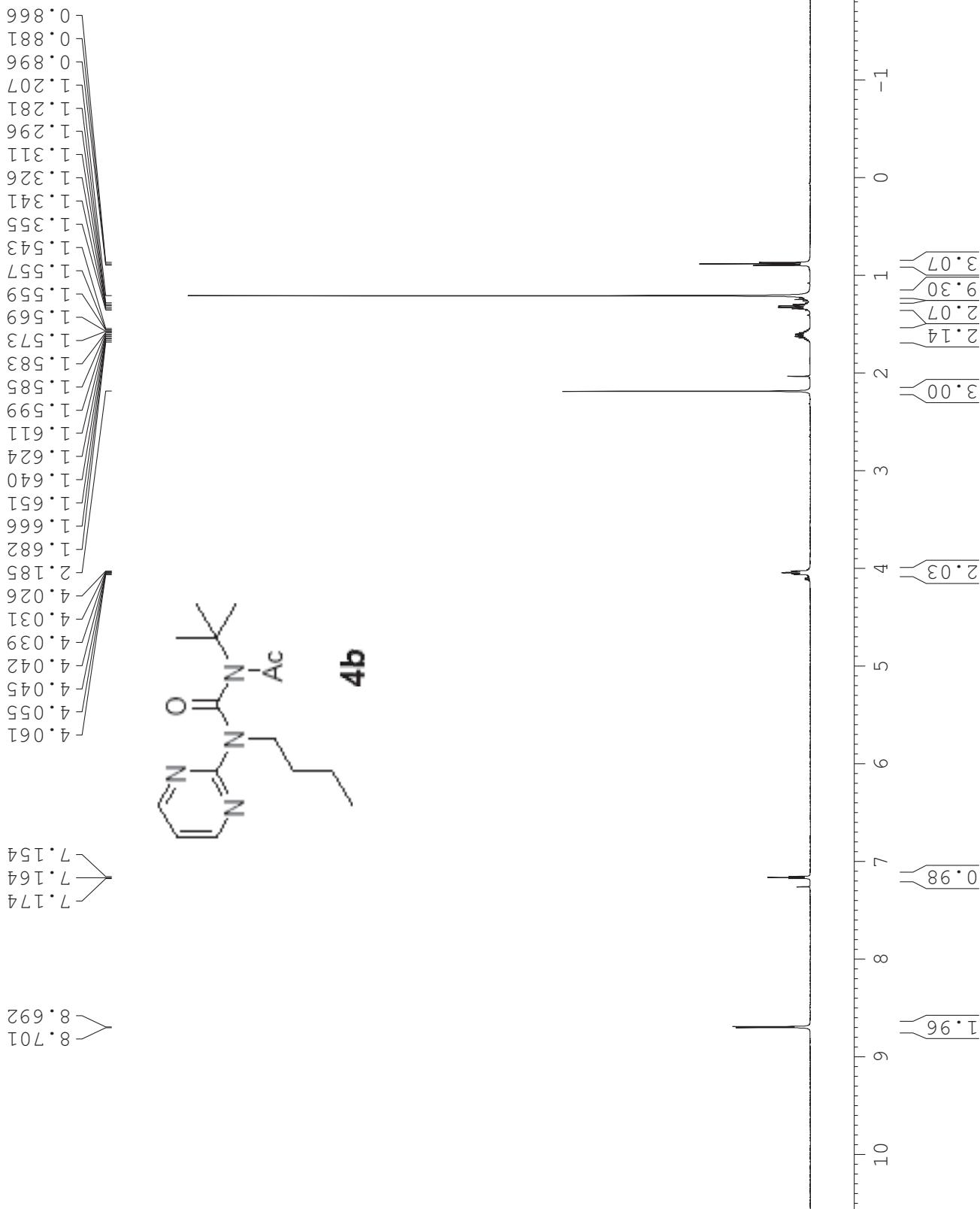


HXM-2-126
C13CPD CDC13

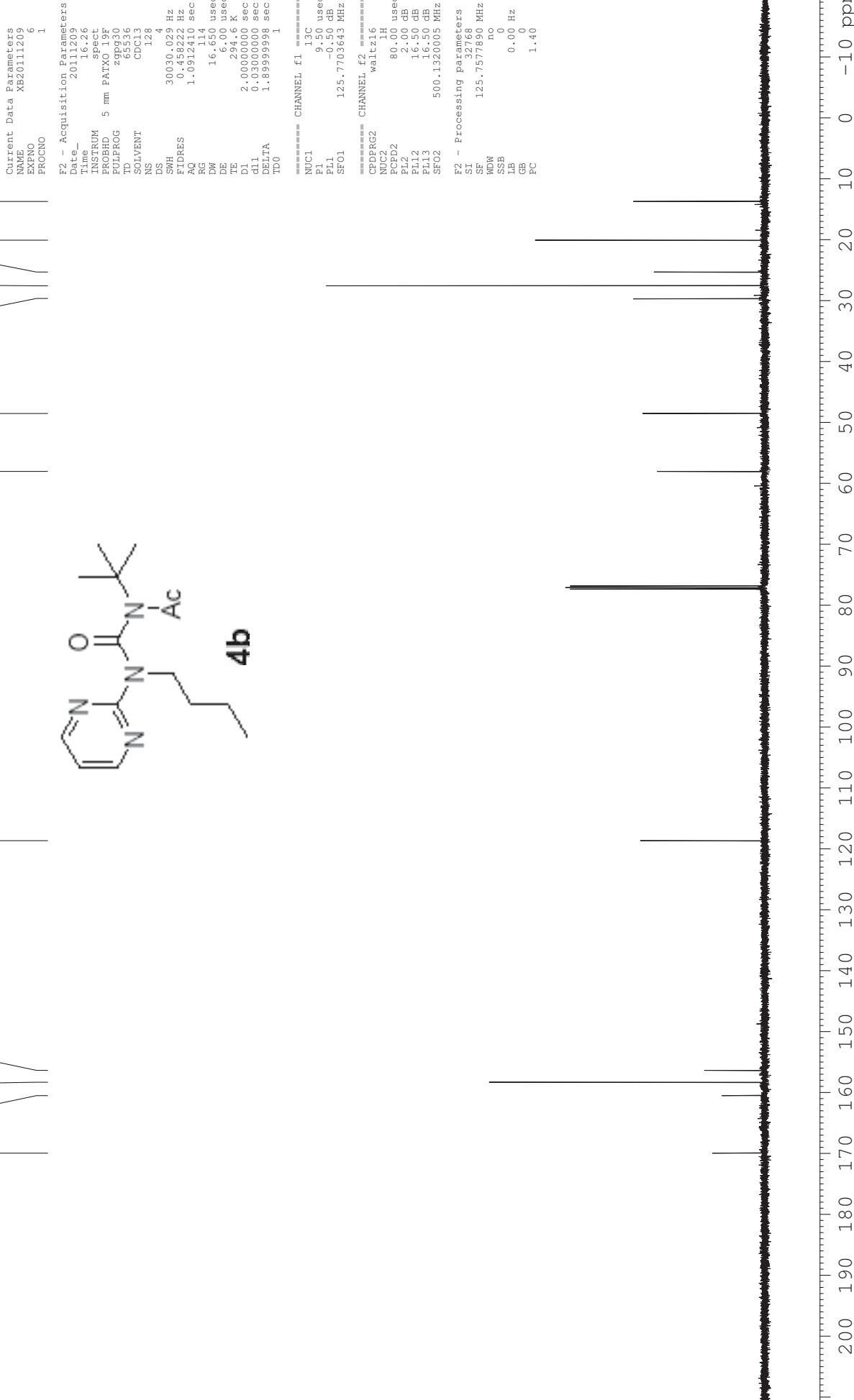


HXM--2-113
PRCTON CDC13

=====
NAME xb20111208
EXPNO 10
PROCNO 1
Date_ 20111208
Time 16.49
INSTRUM spect
PROBHD 5 mm PATRO 19F
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.172047 sec
RG 161.3
DW 48.400 usec
DE 6.000 usec
TE 293.8 K
D1 1.0000000 sec
TD0 1
=====



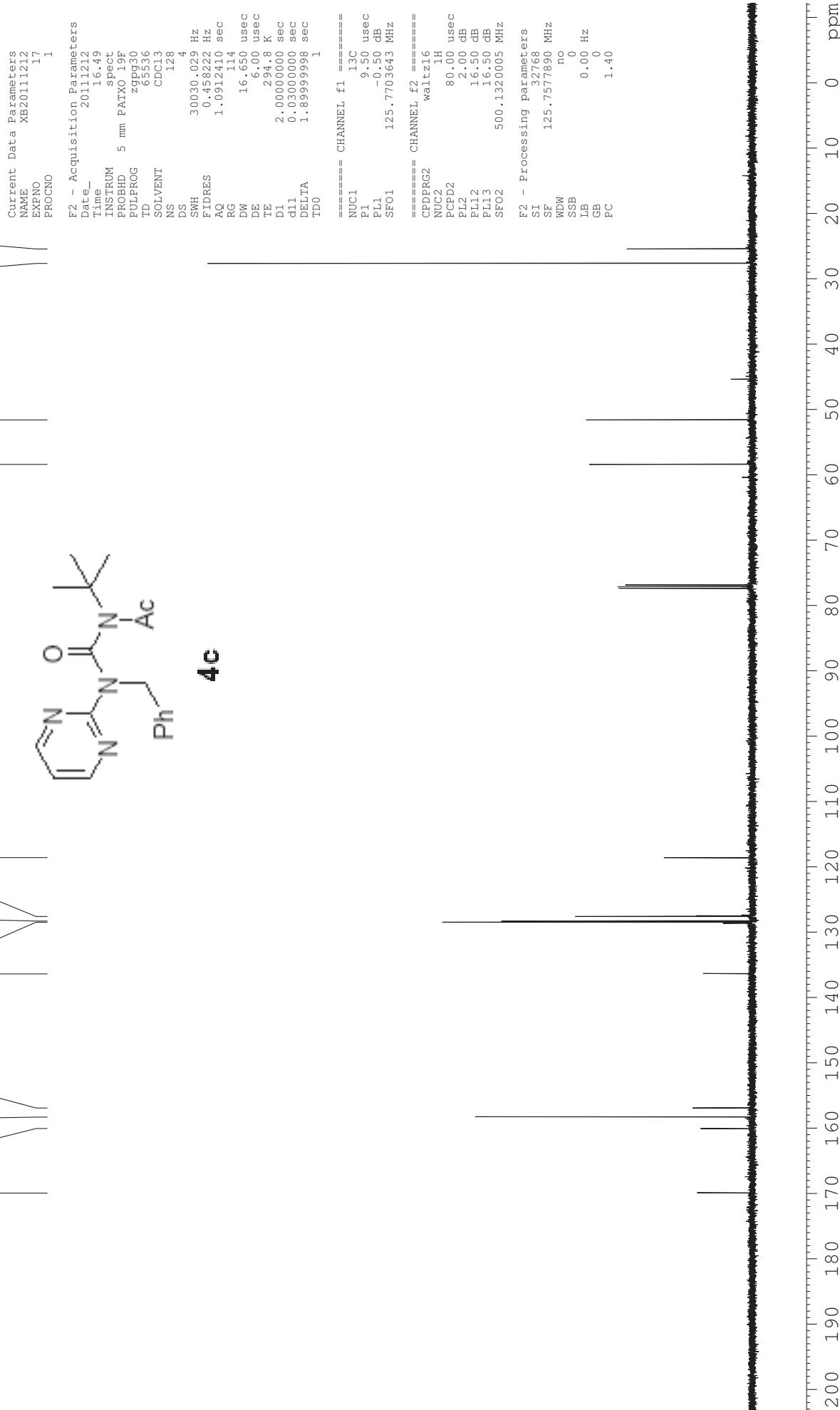
HXM-2-1113
C13CPD CDC13



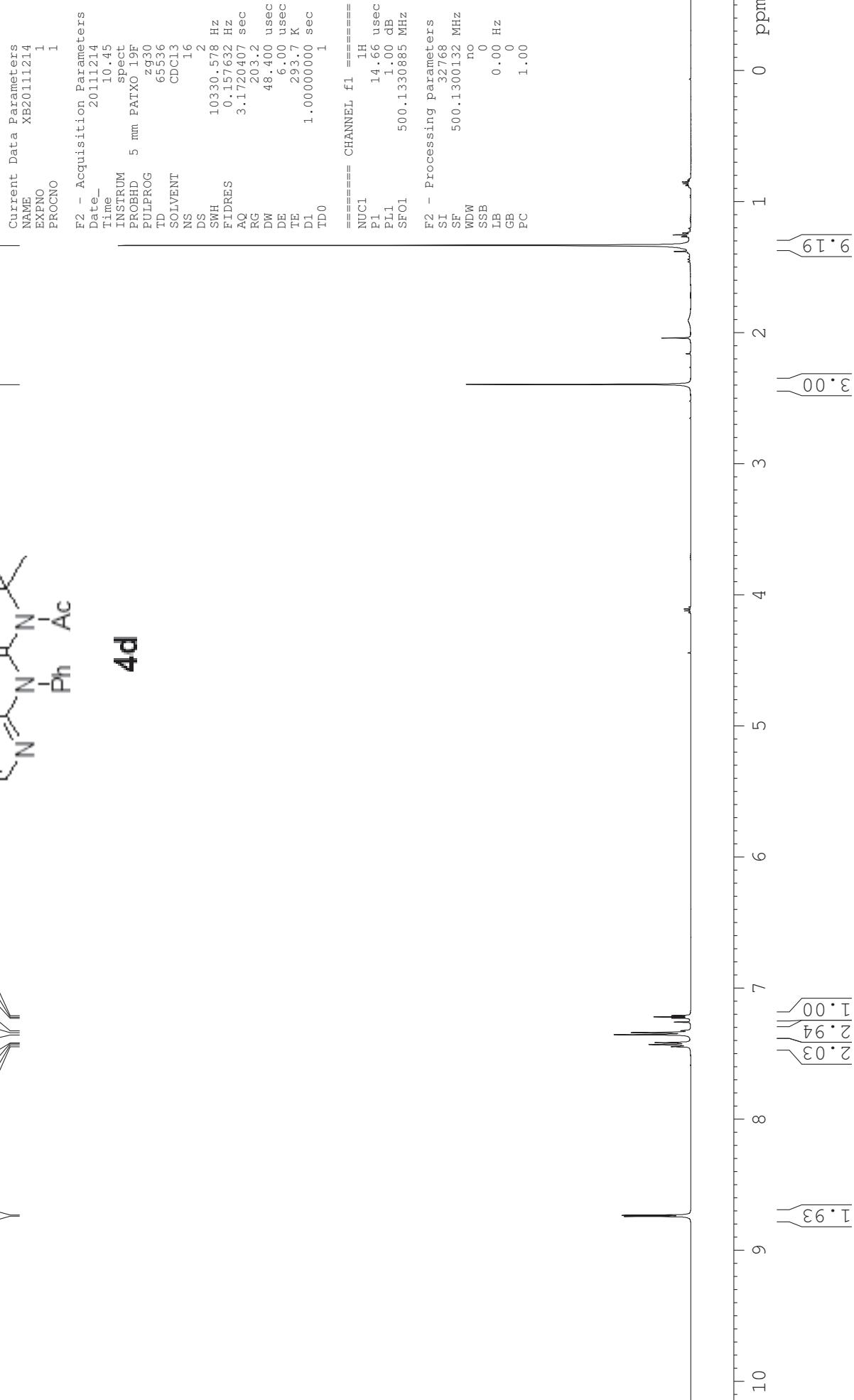
HXM-2-121
PROTON CDCl₃



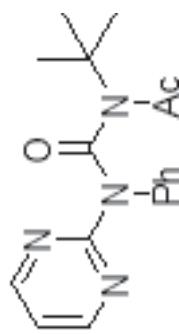
HXM-2-121
C13CPD CDC13



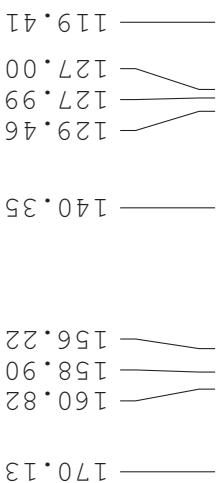
HXM-2-122
PROTON CDC13



HXM-2-122
C13CPD CDCL3



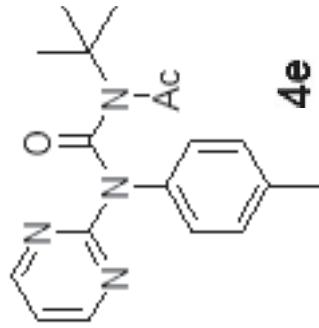
4d



Current Data Parameters
NAME XB2011214
EXPNO 3
PROCNO 1
F2 - Acquisition Parameters
Date_ 201114
Time_ 11.01
INSTRUM PROBHD
PROBHD 5 mm PATRO-19F
PULPROG zgpp930
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 295.0 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1
===== CHANNEL f1 =====
NUC1 13C
PL1 9.50 usec
PL1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
PL13 16.50 dB
SFO2 500.1120005 MHz
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40



HXM-2-114
PROTON CDC13 I



7.194
7.204
7.214
7.233

8.723
8.733

HXM-2-114
PROTON CDCl₃

Current Data Parameters
NAME XB02111208
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
Date_ 20111208
Time 10.19
INSTRUM spect
PROBHD 5 mm PABXO 19F
PULPROG 2930
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.15732 Hz
AQ 3.1720407 sec
RG 322.5
DW 48.400 usec
DE 6.00 usec
TE 293.7 K
D1 1.0000000 sec
TD0 1

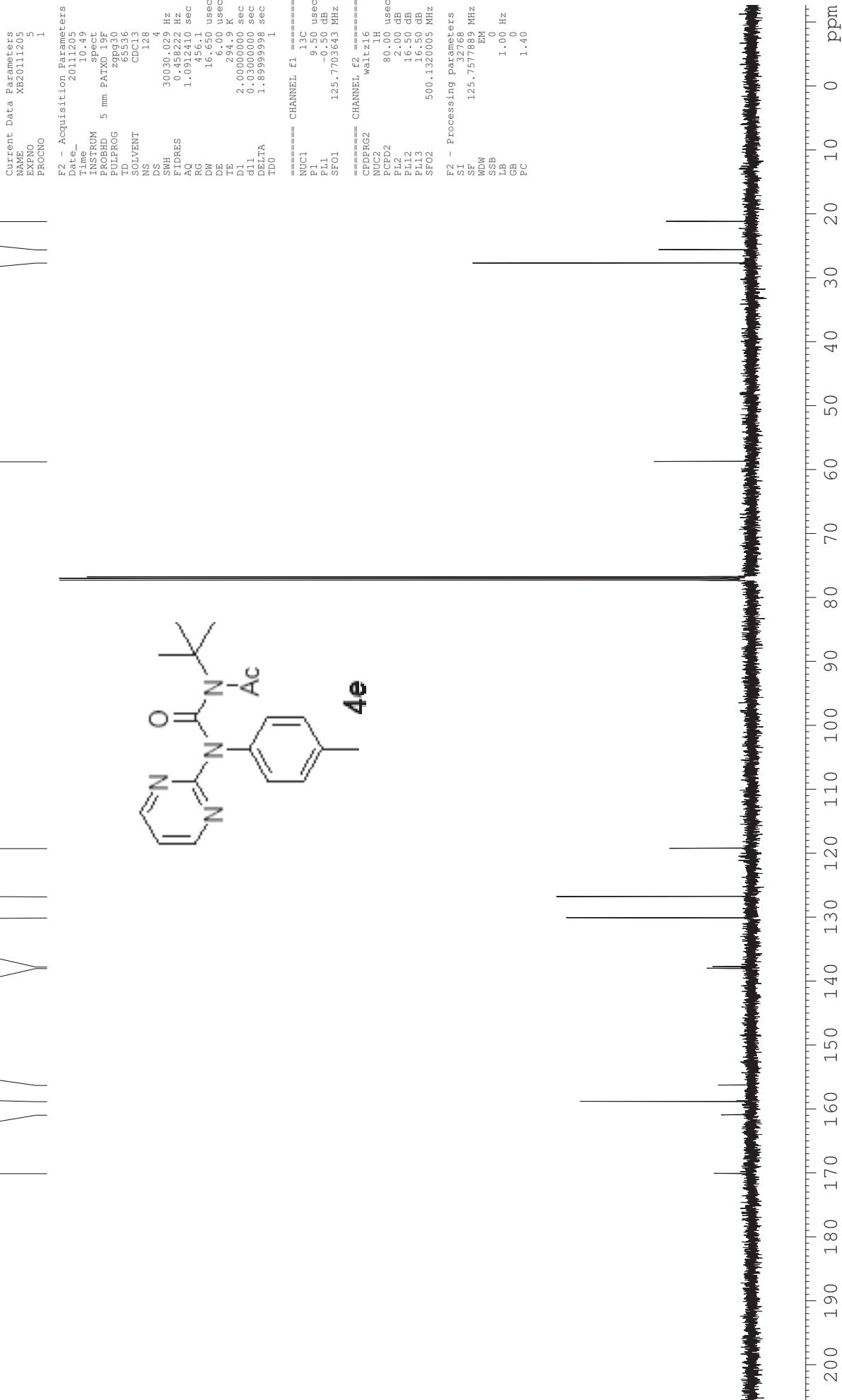
===== CHANNEL f1 =====

NUC1 1H
P1 14.66 usec
PL1 1.00 dB
SFO1 500.1330885 MHz

F2 - Processing parameters
SI 32768
SF 500.1300133 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

4e

HXM-2-104
C13CPD CDC13

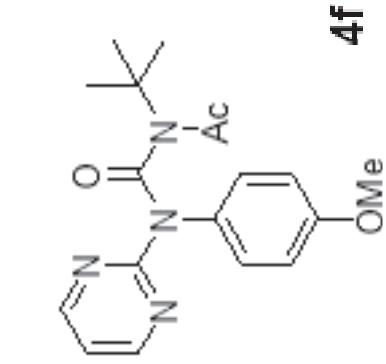


HXM-2-117
PROTON CDC13



HXM-2-1117
C13CPD CDC13

27.67
25.57
25.48
25.66

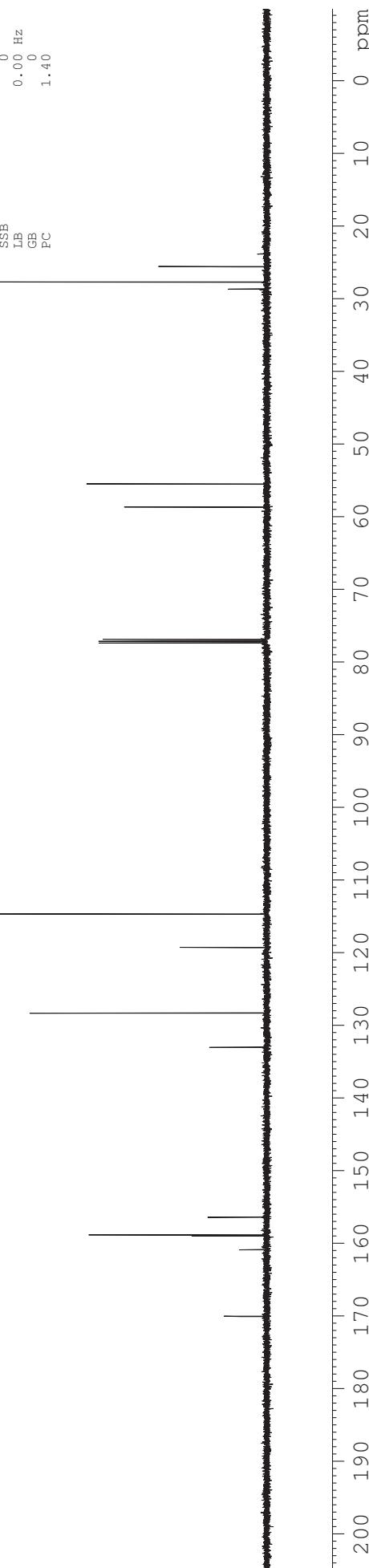


133.03
128.30
119.28
114.68

170.05
160.88
159.02
158.85
156.41

NAME XB20111213
EXPNO 3
PROCNO 1
Date 20111213
Time 10.59
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.45522 Hz
AQ 1.0912410 sec
RG 1114
DW 16.650 usec
DE 6.00 usec
TE 294.9 K
D1 2.0000000 sec
d1 0.0300000 sec
DETA 1.8999998 sec
TD0 1

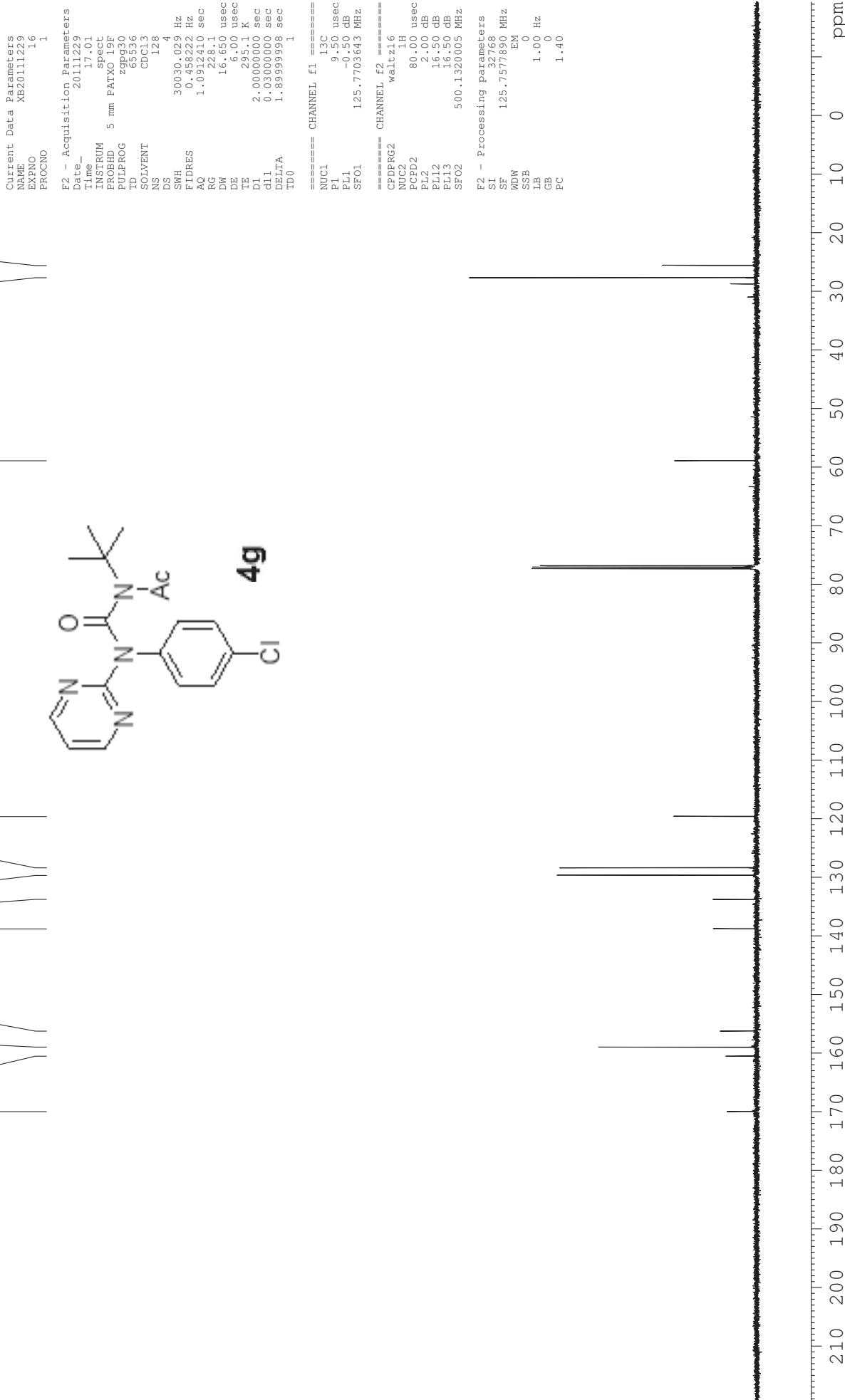
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SF01 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waitz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.50 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.75771890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40

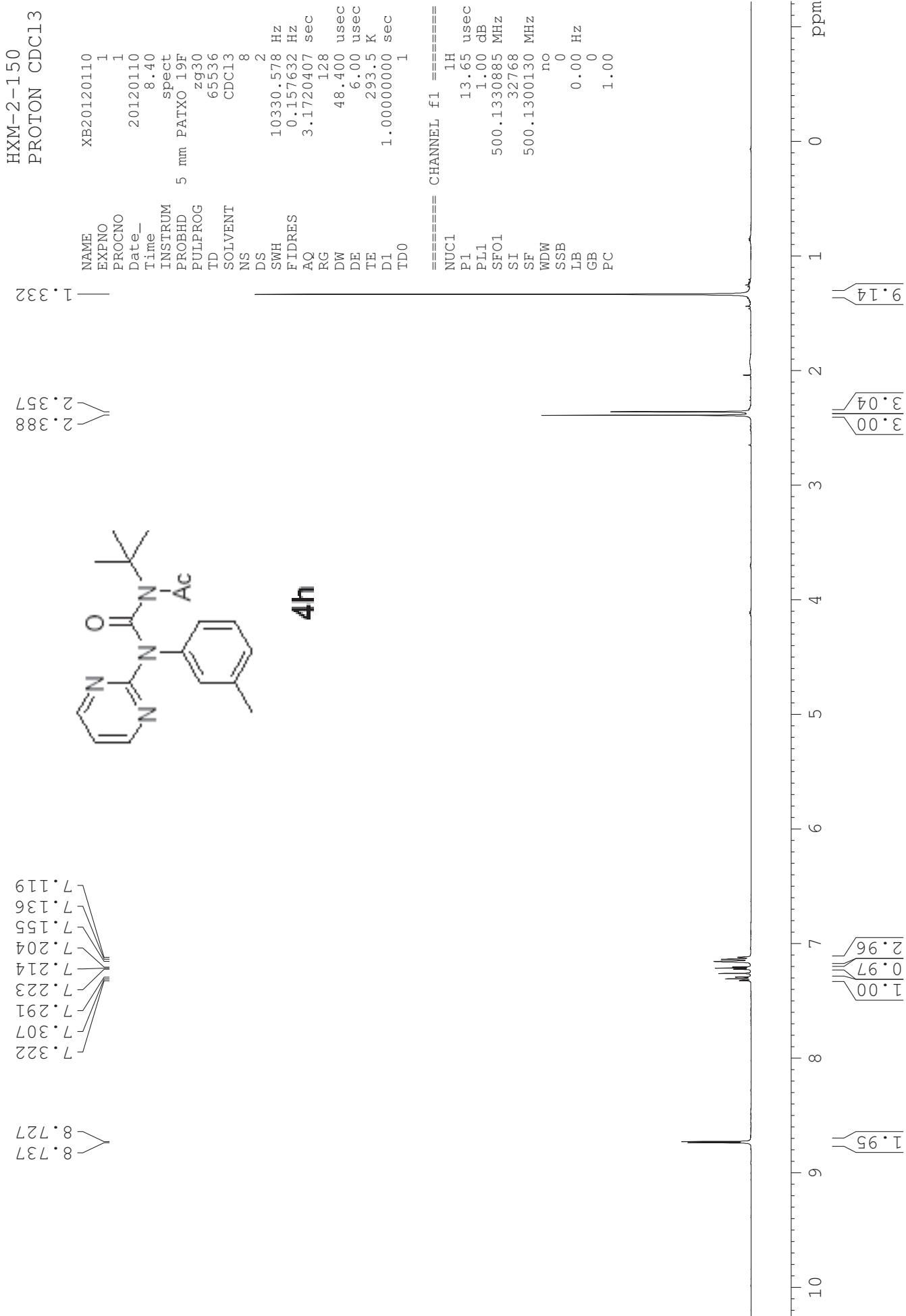


HXM-1-141
PROTON CDCl₃



HXM-1-141
C13CPD CDCL₃





HXM-2-150
C13CPD CDC13

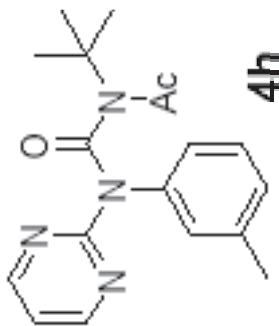
NAME XB20120110
EXPNO 6
PROCNO 1
Date_ 20120110
Time 9.20
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 294.9 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.77 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40



— 58.76 —

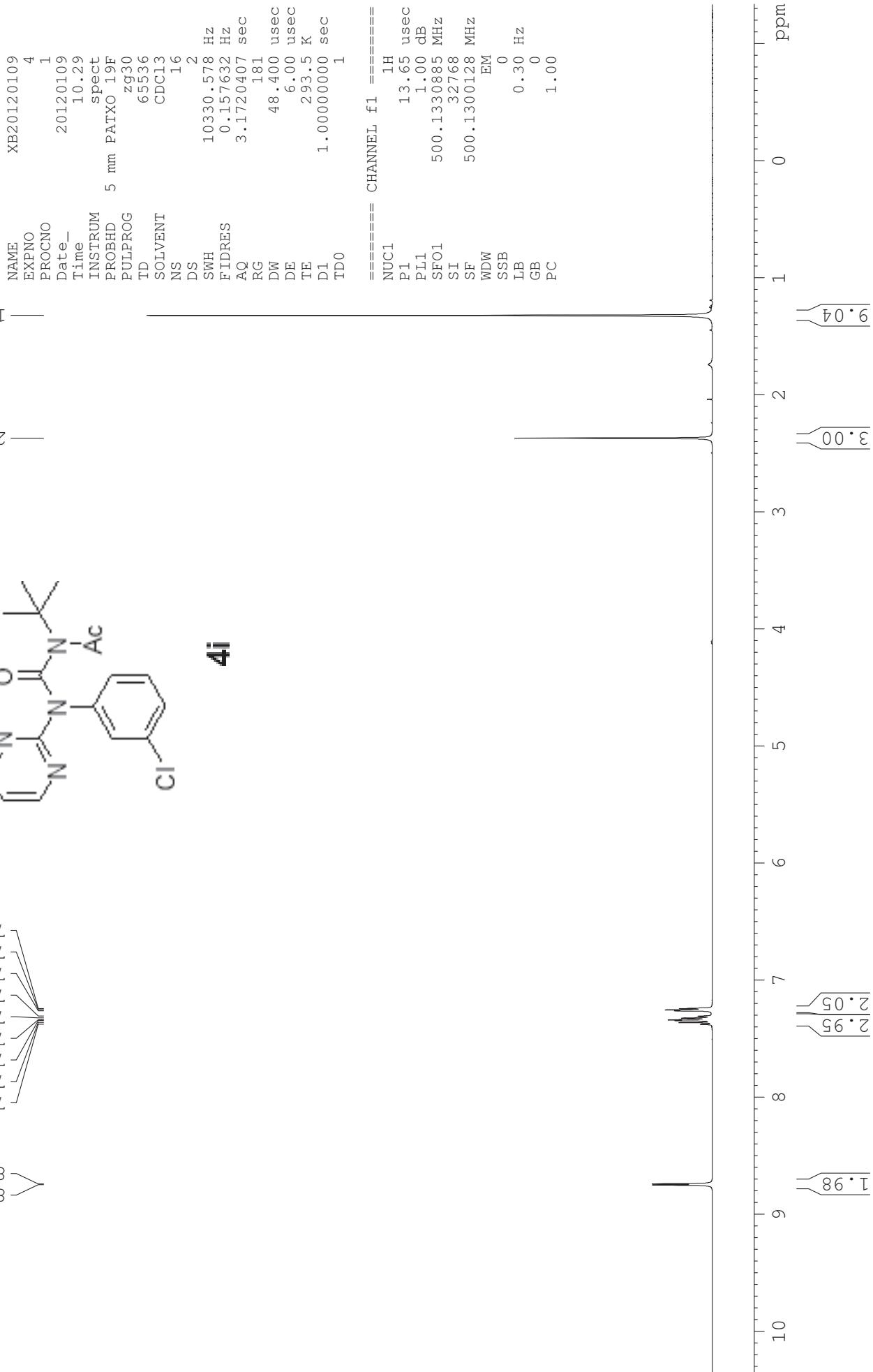


/ 119.36
/ 123.97
/ 127.49
/ 128.89
/ 129.25
/ 139.52
/ 140.22

/ 156.21
/ 158.89
/ 160.86

— 170.13 —

HXM-2-151
PROTON CDCl₃



HXM-2-151
C13CPD CDC13

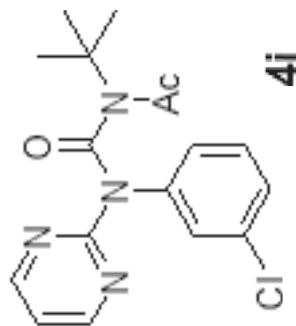
NAME XB20120109
EXPNO 11
PROCNO 1
Date_ 20120109
Time 11.13
INSTRUM spect
PROBHD 5 mm PAXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 322.5
DW 16.650 usec
DE 6.00 usec
TE 294.9 K
D1 2.0000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.77 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0.00 Hz
LB 0
GB 0
PC 1.40

25.59
27.68

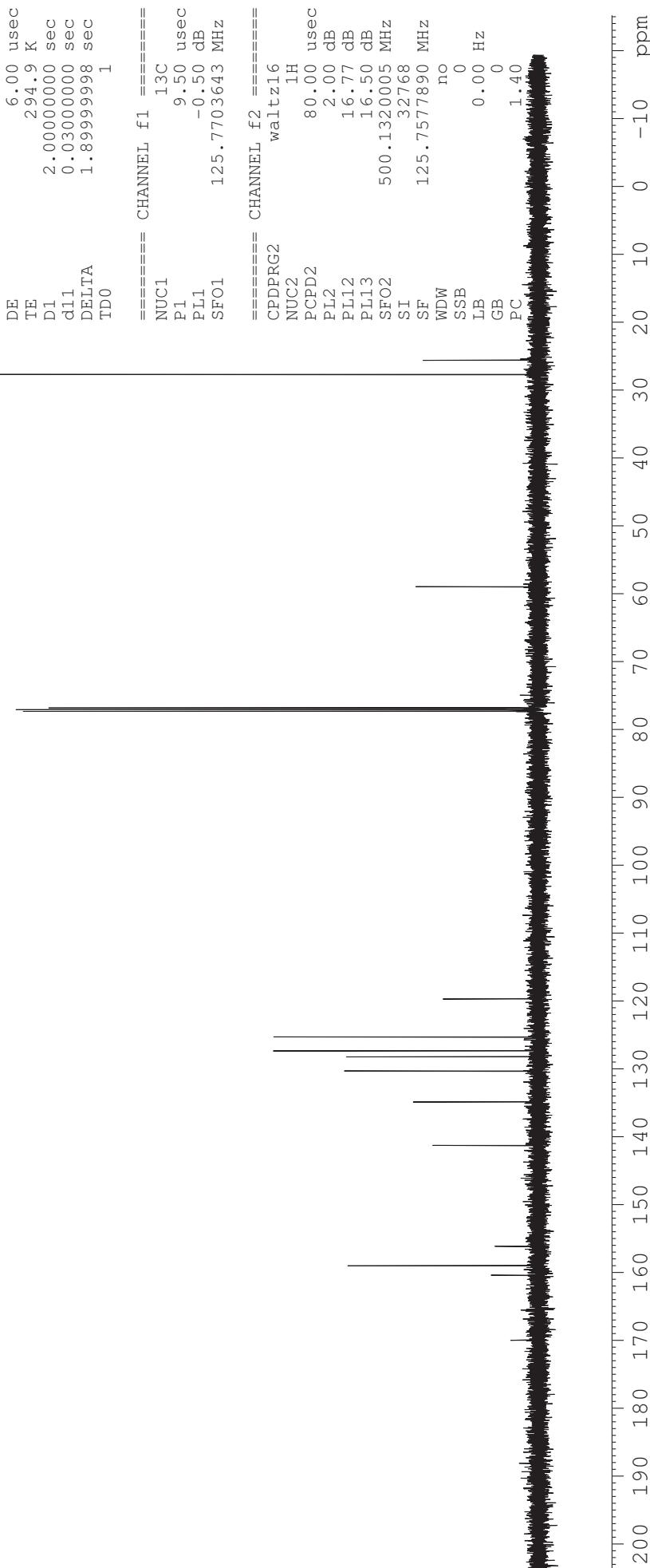
58.94

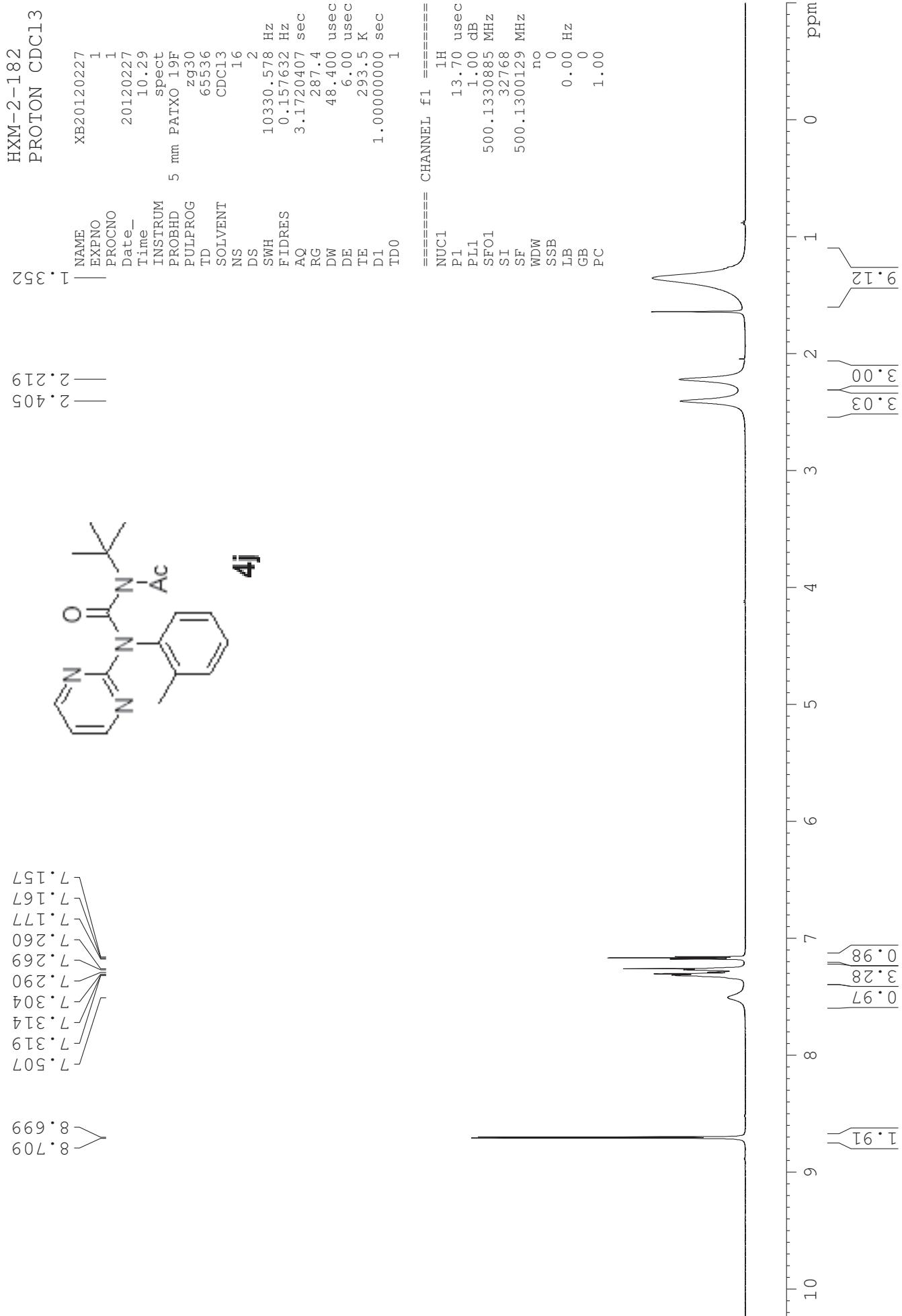


119.68
125.30
127.33
128.21
130.33
134.87
141.28

156.16
159.01
160.41

169.97





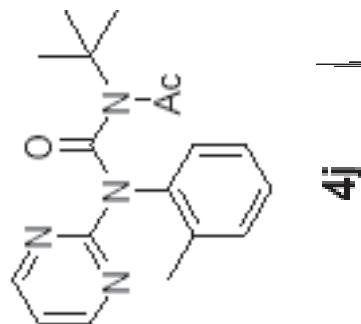
4j

HXM-2-182
C13CPD CDC13

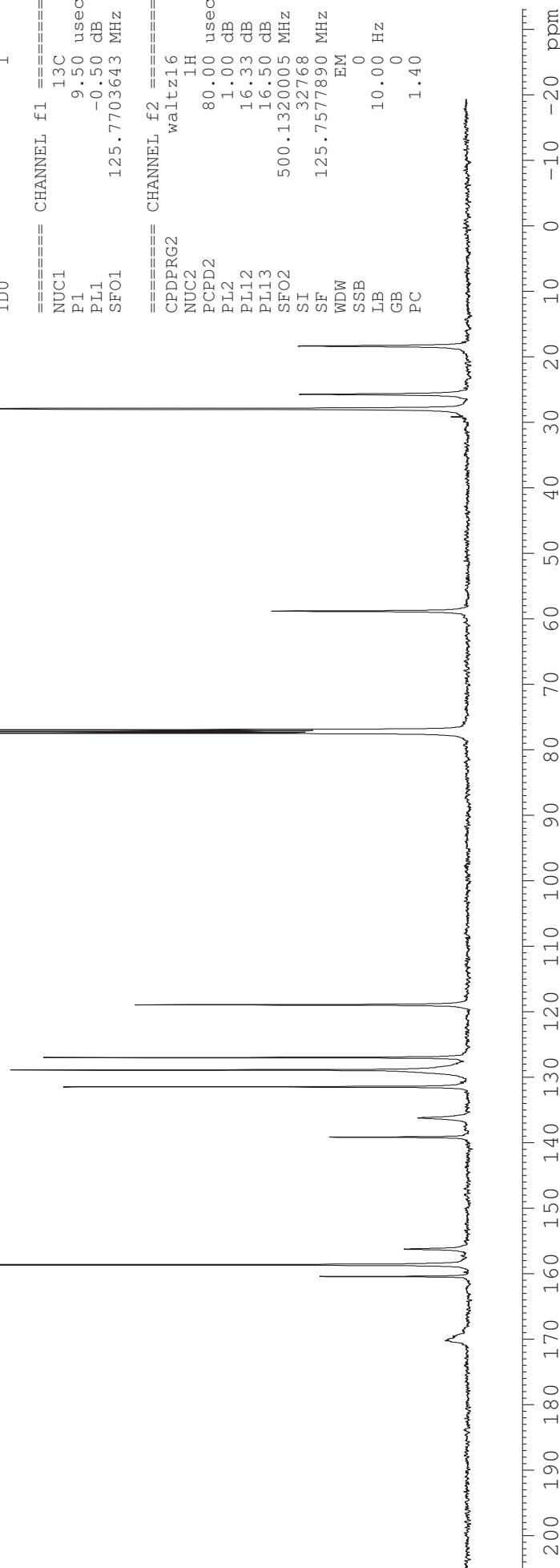
NAME XB20120316
EXPNO 12
PROCNO 1
Date 20120316
Time 18.11
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgppg30
TD 65536
SOLVENT CDC13
NS 2048
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 1114
DW 16.650 usec
DE 6.00 usec
TE 295.4 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TDO 1

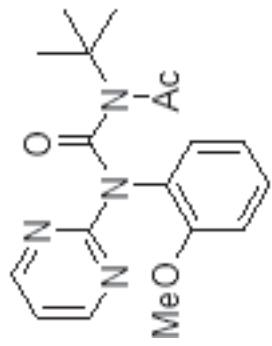
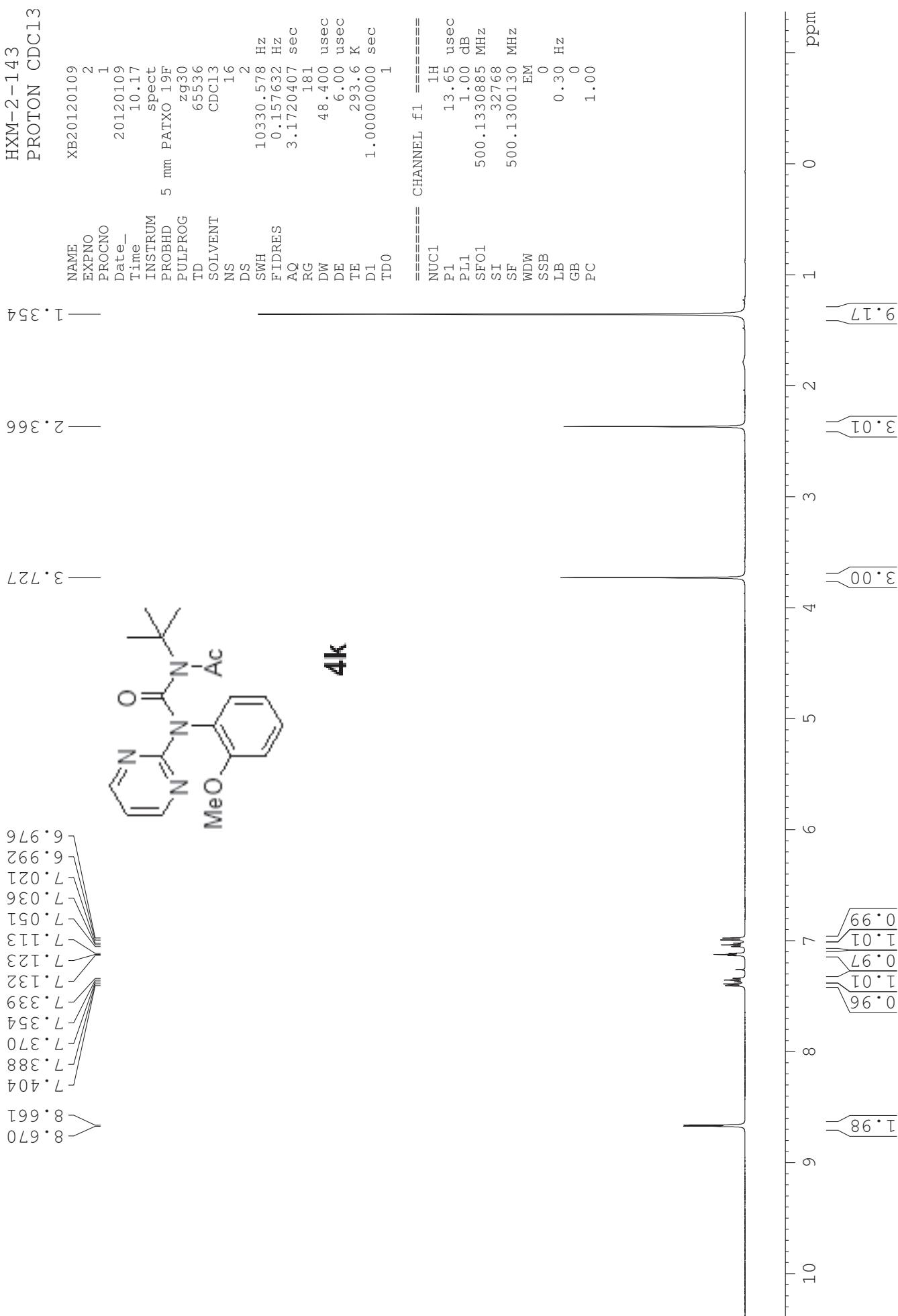
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 10.00 Hz
GB 0
PC 1.40

— 18.28
— 25.65
— 27.86
— 58.78



— 118.91
— 126.96
— 128.86
— 131.43
— 136.18
— 139.12
— 156.21
— 158.62
— 160.41
— 170.19





三

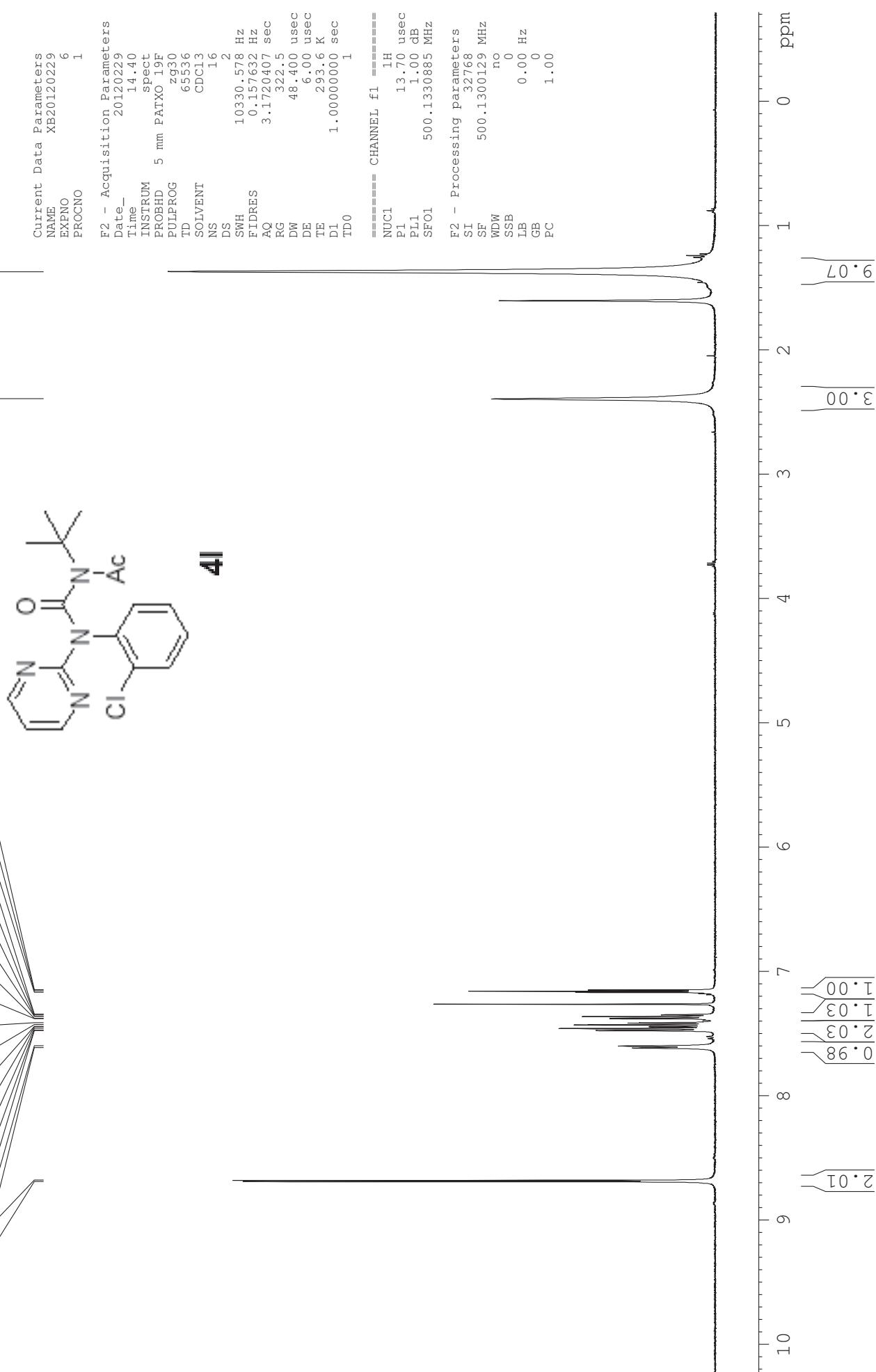
HXM-2-143
C13CPD CDC13

NAME XB20120109
EXPNO 10
PROCNO 1
Date 20120109
Time 11.01
INSTRUM spect
PROBHD 5 mm PAXCO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 181
DW 16.650 usec
DE 6.00 usec
TE 294.8 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TDD 1

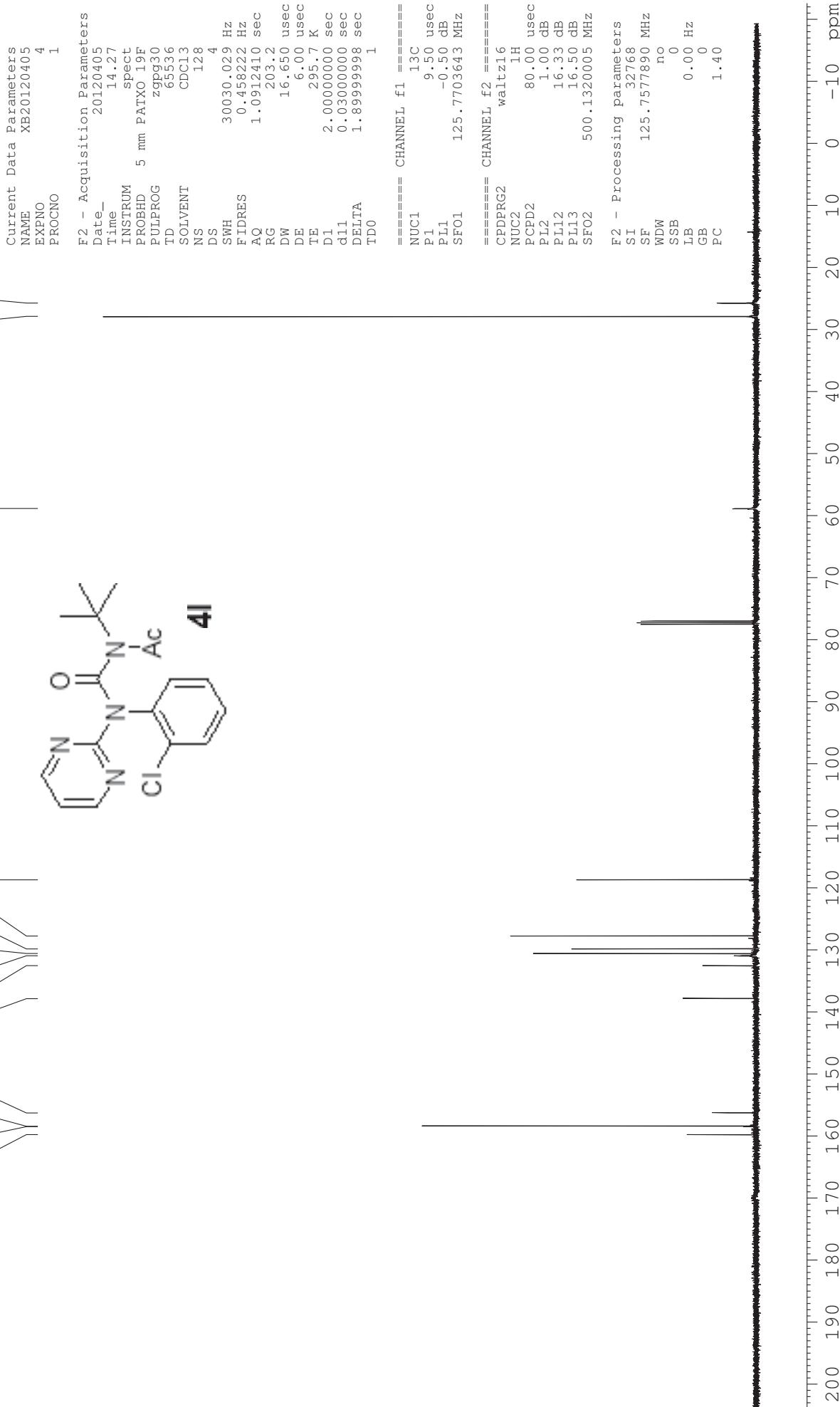
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.77 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40



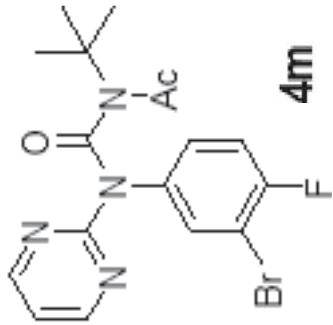
HXM-2-195
PROTON CDC13 I



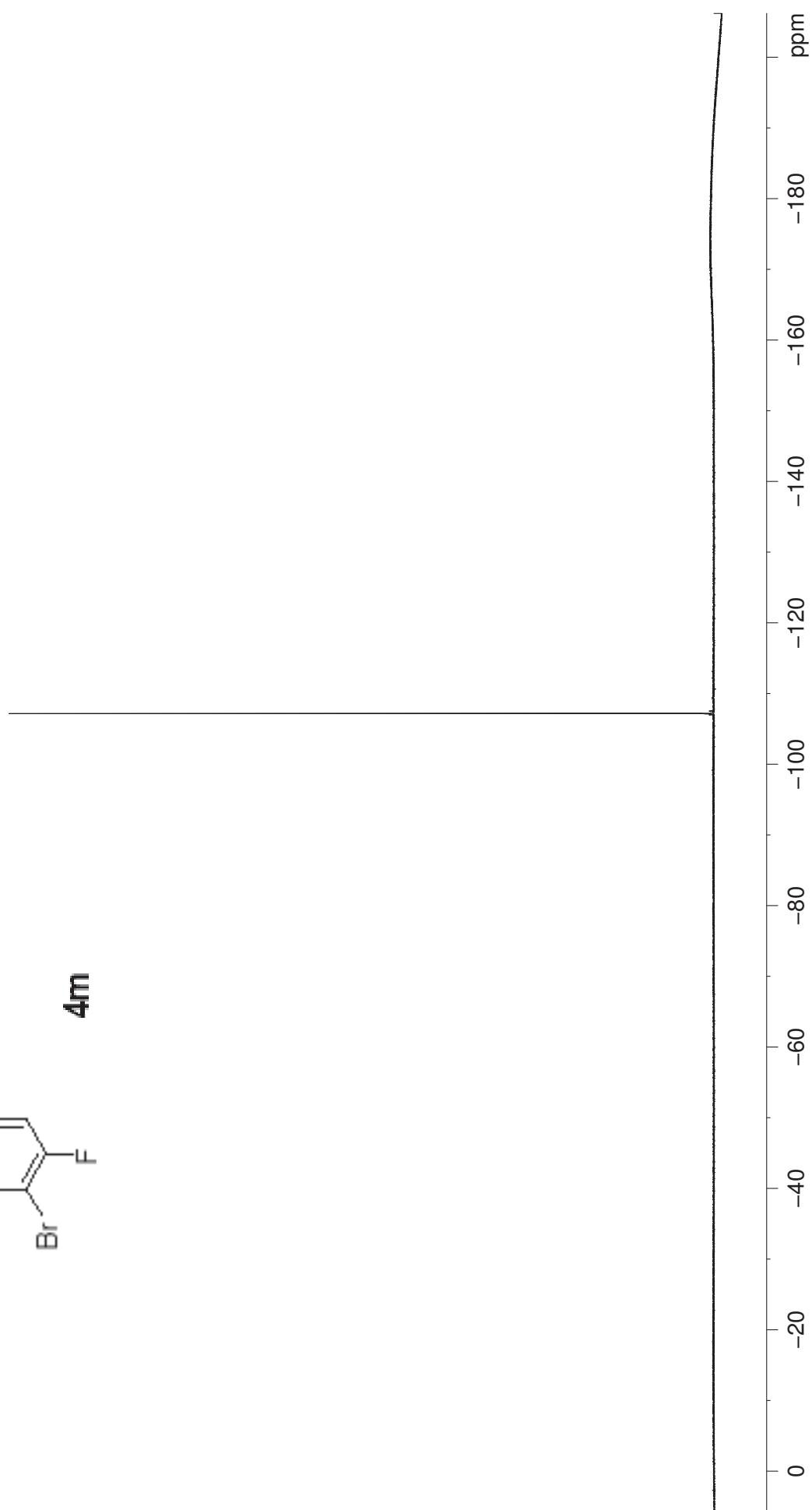
HXM-2-195
C13CPD CDC13



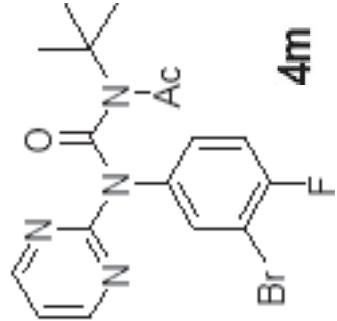
HXM-2-152
PROTON CDC13



HXM-2-152
19Fdeft CDCl₃ D:\deng 3C



HXM-2-152
C13CPD CDCl₃



27.68 25.56

58.93

109.67
116.91
117.09
119.72
128.03
128.09
132.41
136.82
136.85

156.33
159.03
159.43
160.27
169.85

Current Data Parameters
NAME XB20120112
EXPNO 4
PROCNO 1

F2 - Acquisition Parameters
Date 20120112
Time 13.41
INSTRUM spect
PROBHD 5 mm PAXO 19F
PULPROG zgpp30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 294.7 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 2.00 dB
PL12 16.77 dB
PL13 16.50 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 1.0
PC 1.40



HXM-2-364
PROTON CDCL:

```

=====
CHANNEL f1 =====

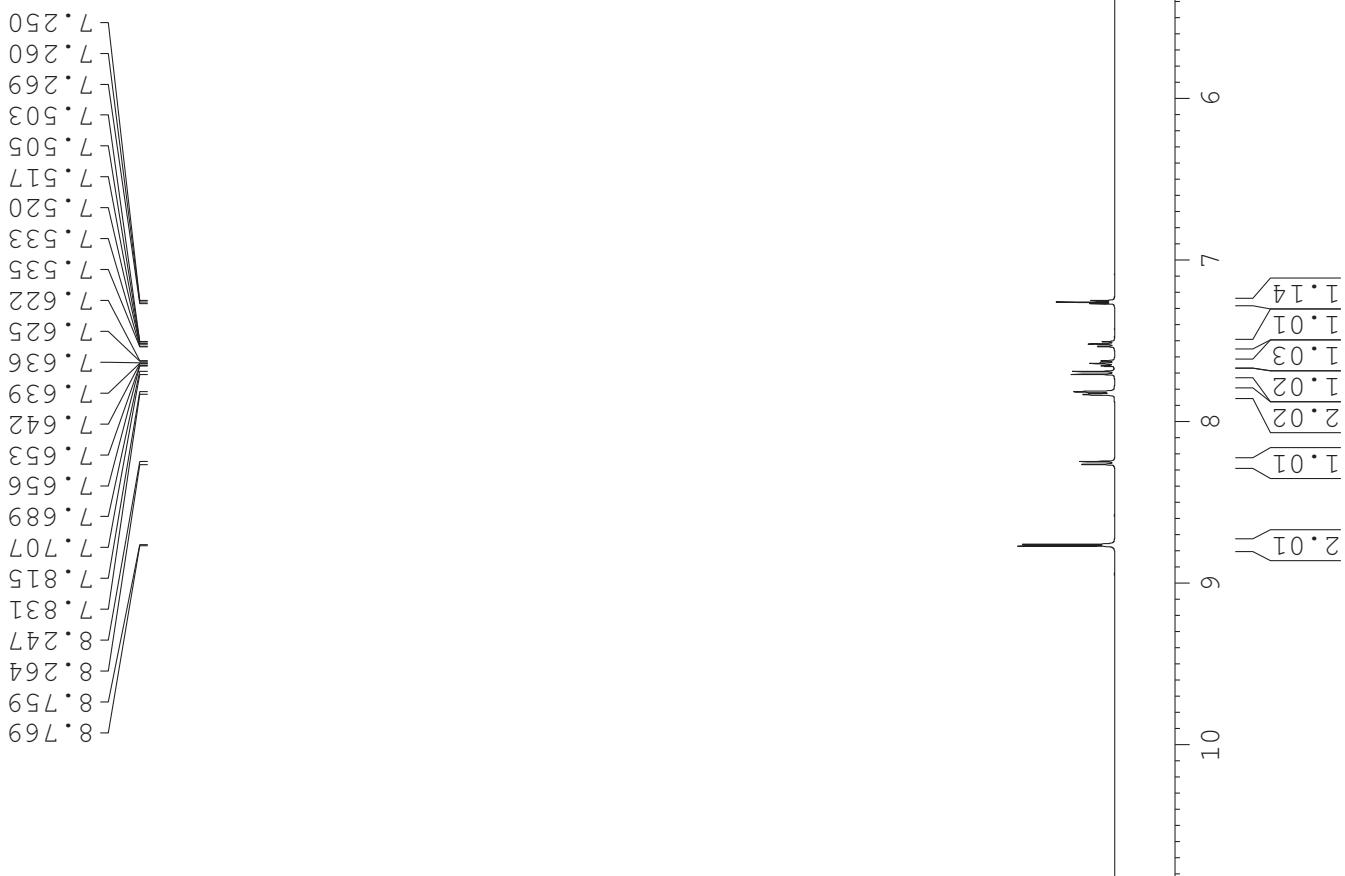
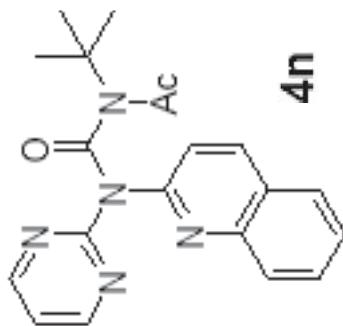
xb20120629 1
EXPNO 1
PROCNO 1
Date_ 20120629
Time_ 10.23
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG PULPROG
TD 65536
SOLVENT C6C13
NS 8
DS 2
SWH 10330.578 Hz
FDIDRES 0.157632 Hz
AQ 3.1720407 sec
RG 161.3
DW 48.400 usec
DE 6.00 usec
TE 295.8 K
D1 1.0000000 sec
TITD01 1

```

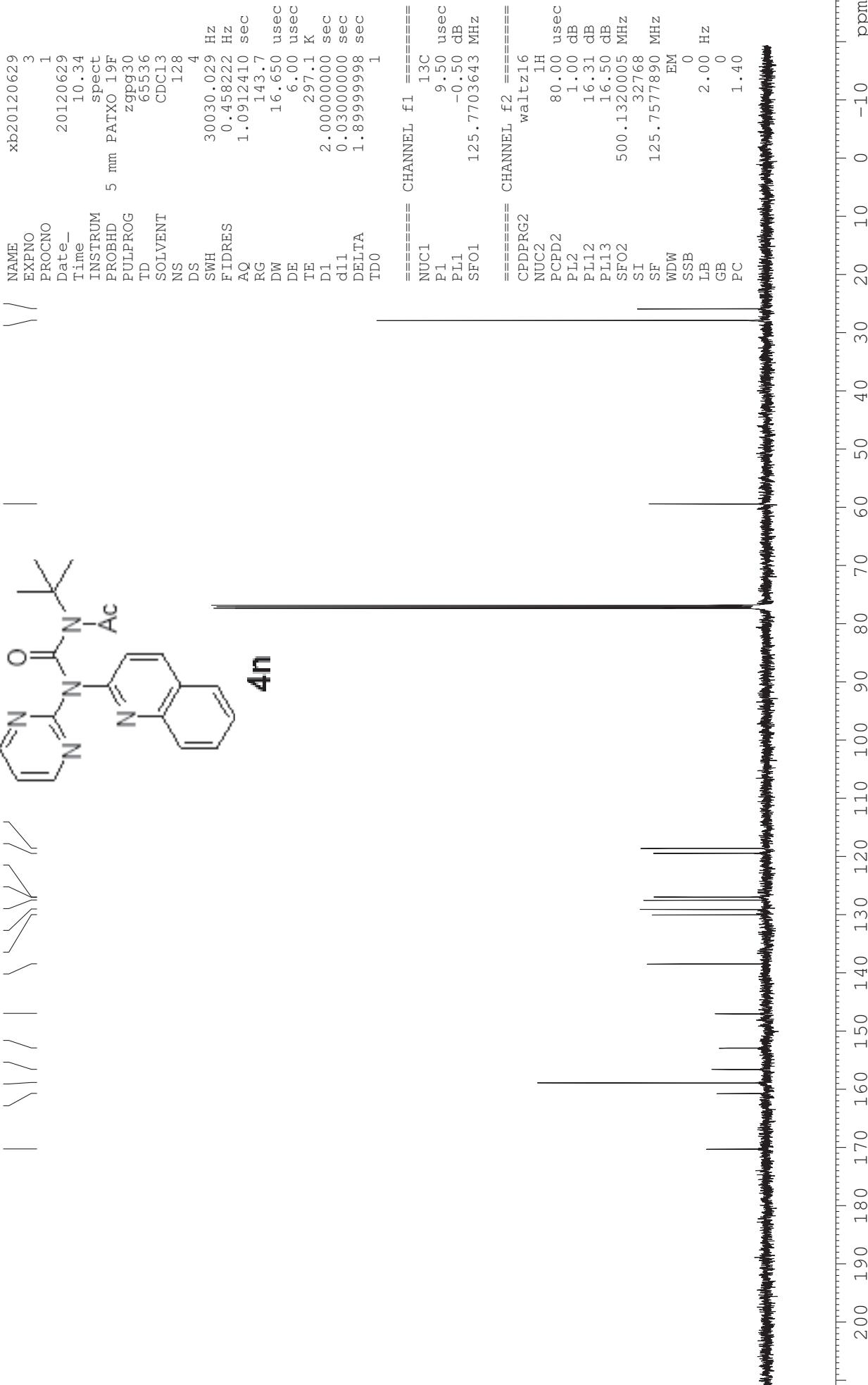
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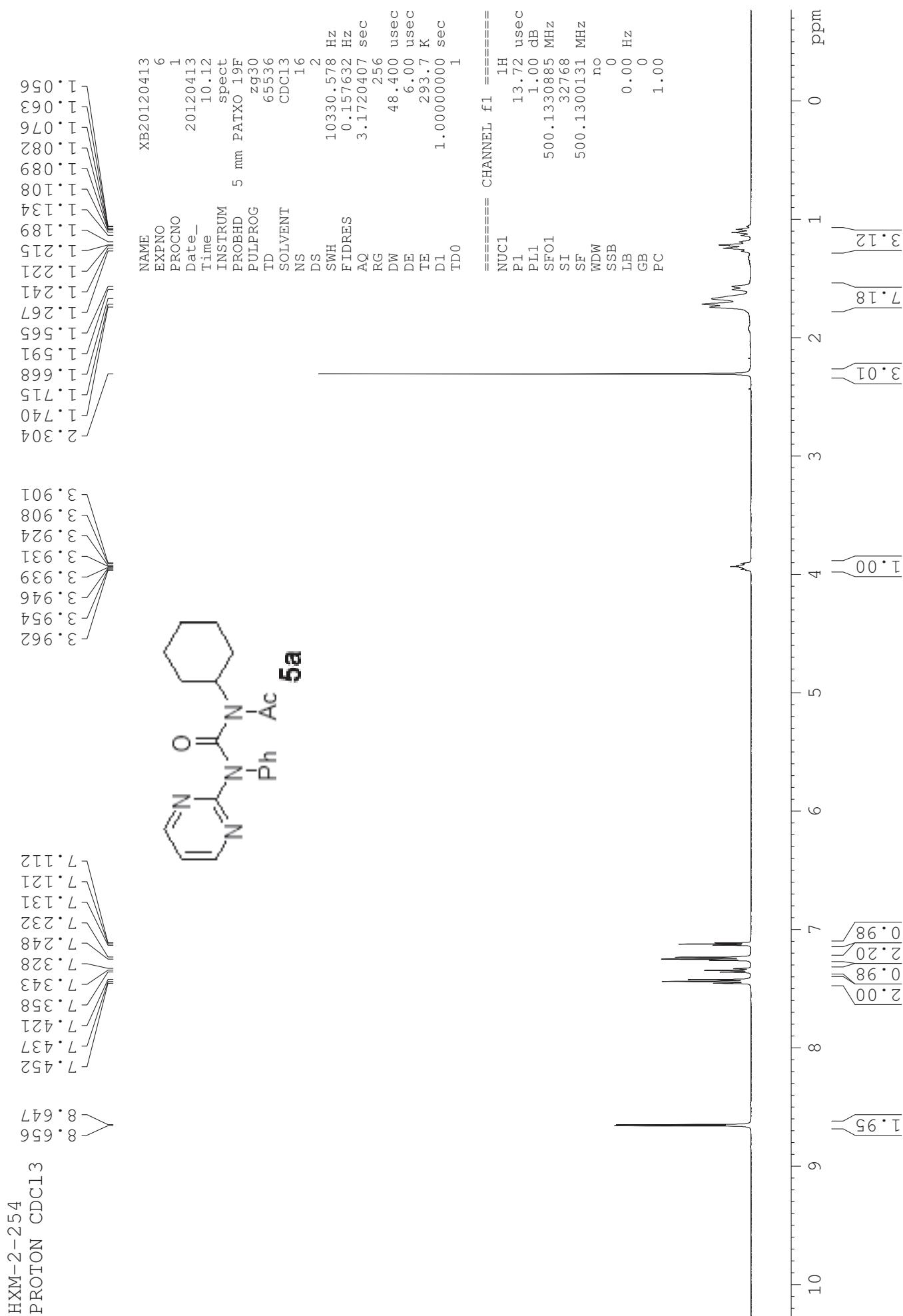
===== CHANNEL f1 =====
NUC1          1H
P1           13.72 usec
PPLI          1.00 dB
SSFO1         500.1330085 MHz
SI            327.68 MHz
SF            500.13000129 MHz
EM
WDW
SSSB          0
LLB          0.30 Hz
GB            0
PC          1.00

```



HXM-2-364
C13CPD CDCl₃



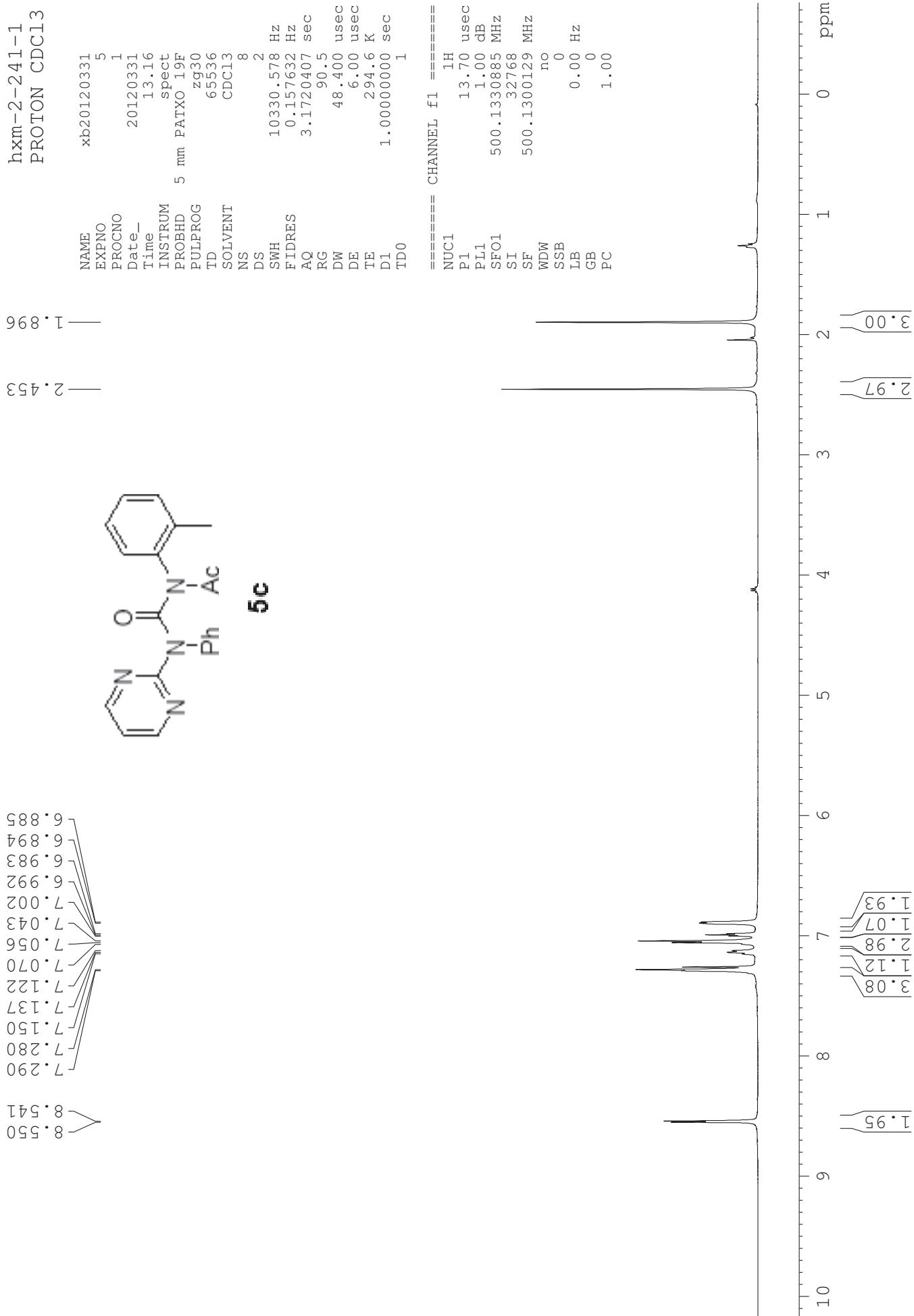


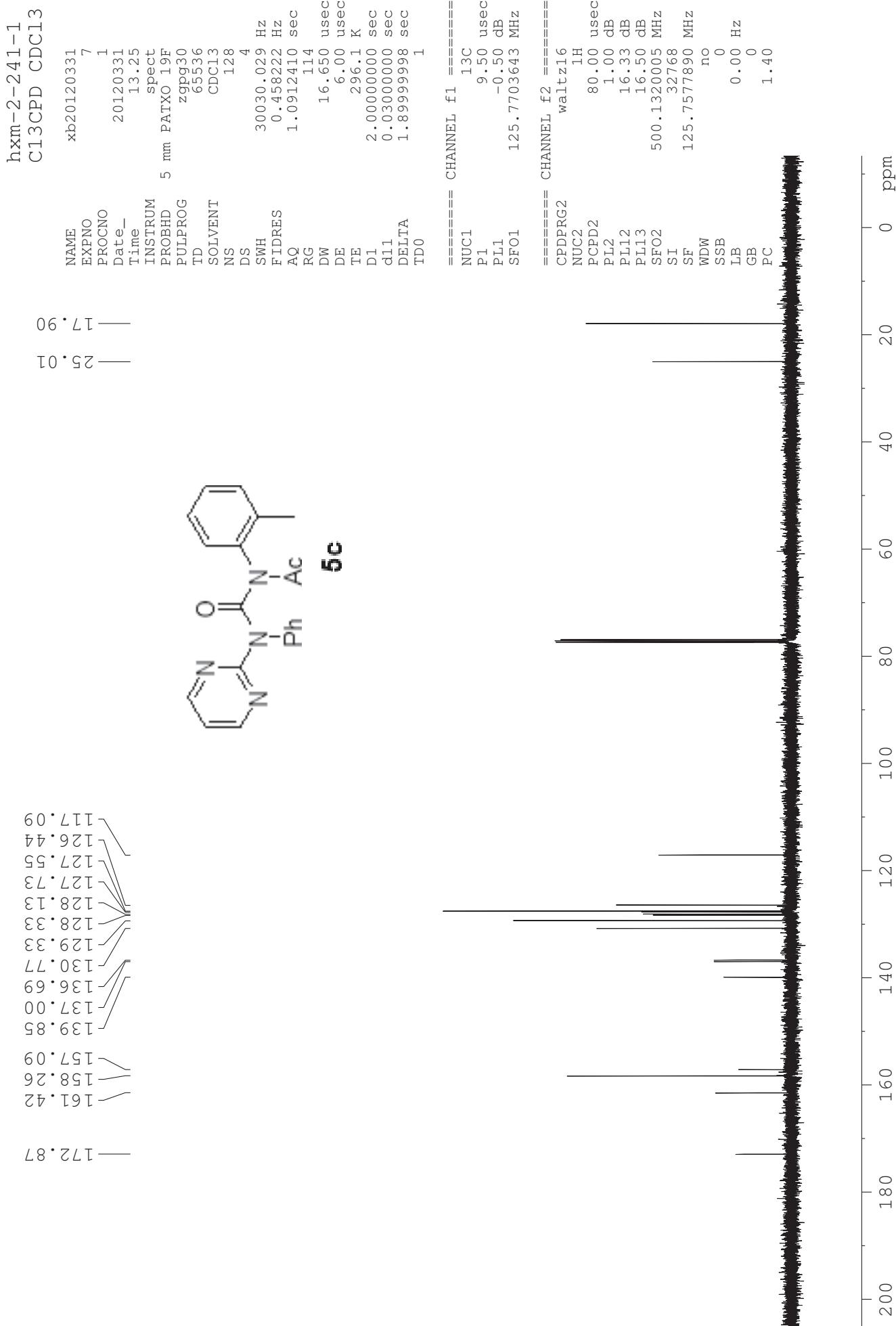
HXM-2-254
C13CPD CDC13

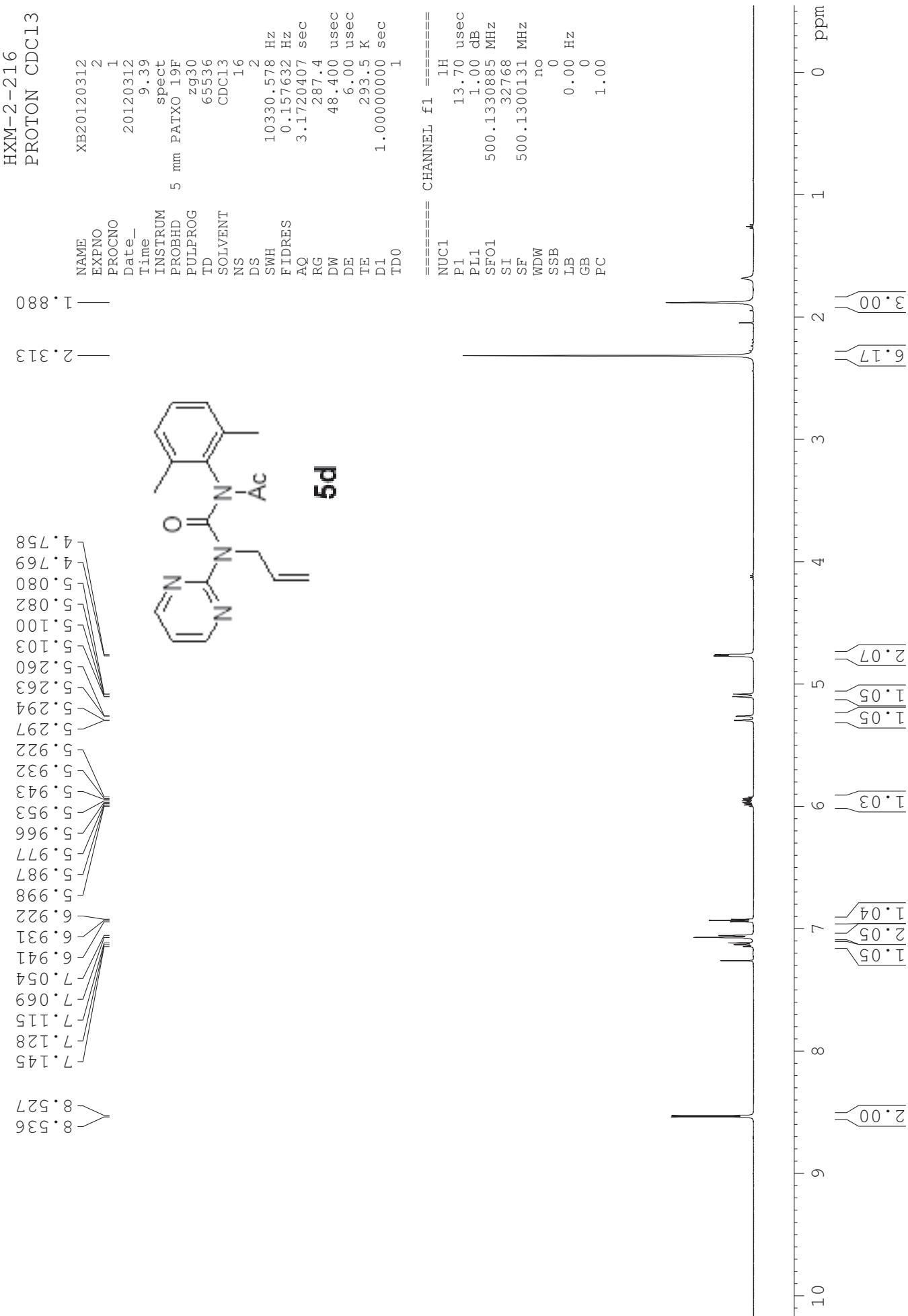
NAME XB20120411_5
EXPNO 5
PROCNO 1
Date 20120411
Time 10.34
INSTRUM spect
PROBHD 5 mm EATXO 19F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 296.9 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 327.68 MHz
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40









HXM-2-216
C13CPD CDC13

Current Data Parameters
NAME XB20120315
EXPNO 4
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120315
Time 16.24
INSTRUM spect
PROBID 5 mm PABX0 19F
PULPROG zgrg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
ETR 0.45222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 235.2 K
D1 2.0000000 sec
d1,1 0.0300000 sec
DELTA 1.8999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz

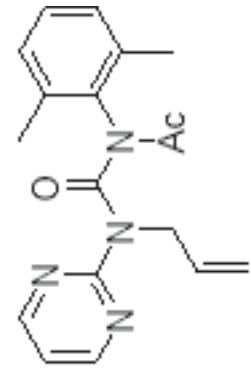
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40



51.13

24.78

18.71



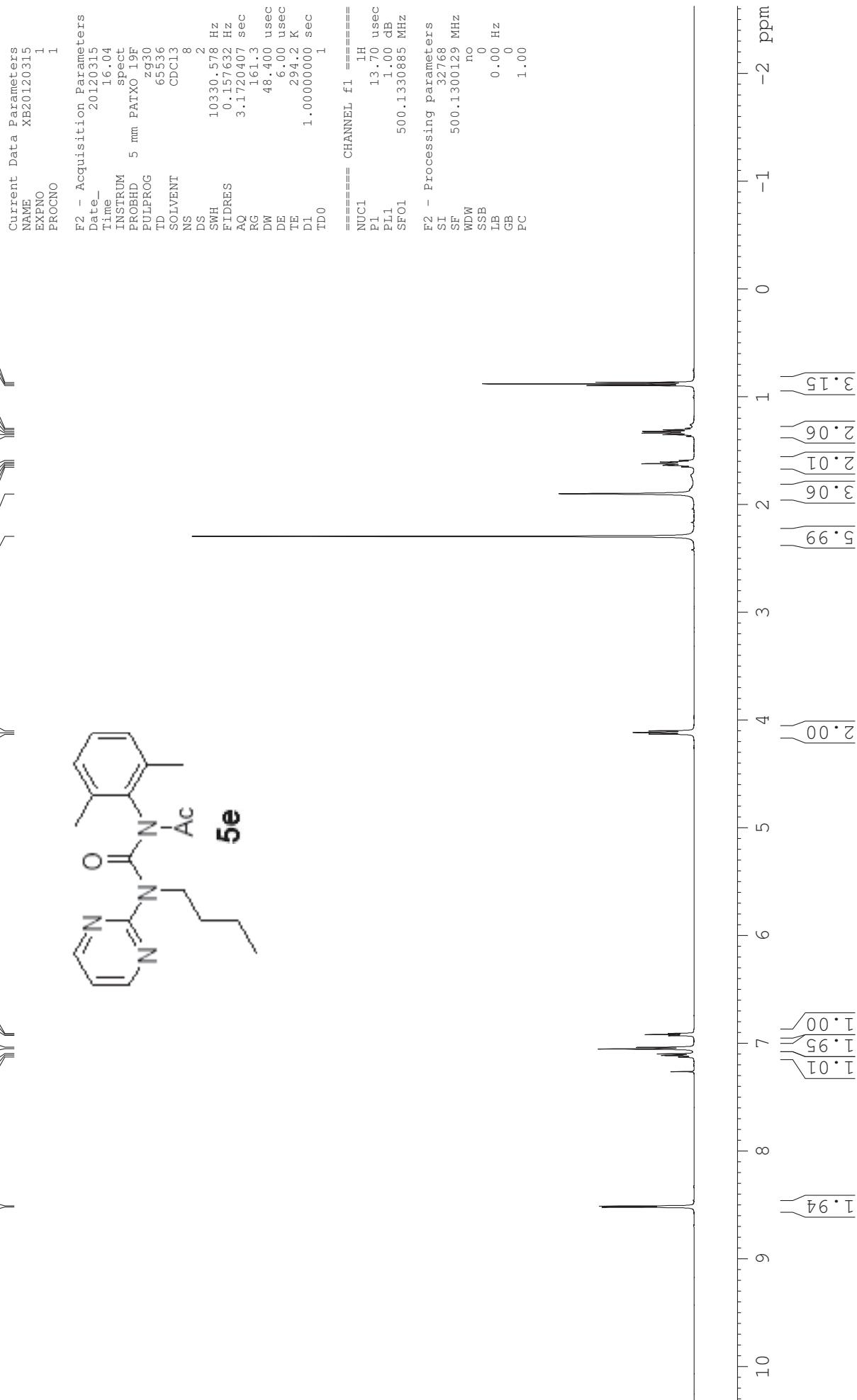
5d

116.35
116.94
128.48
128.80
133.47
136.83
137.72

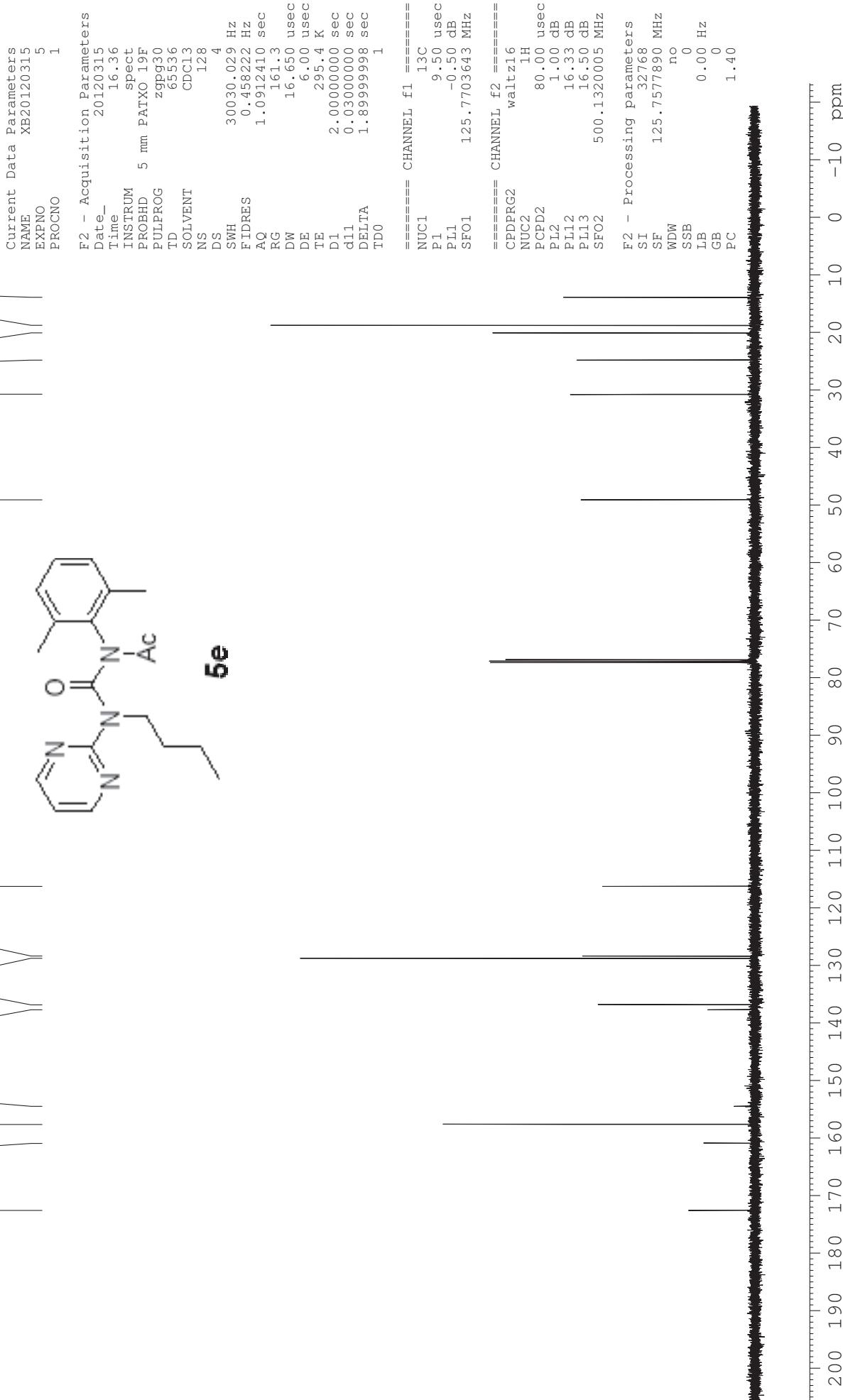
154.07
157.63
160.74

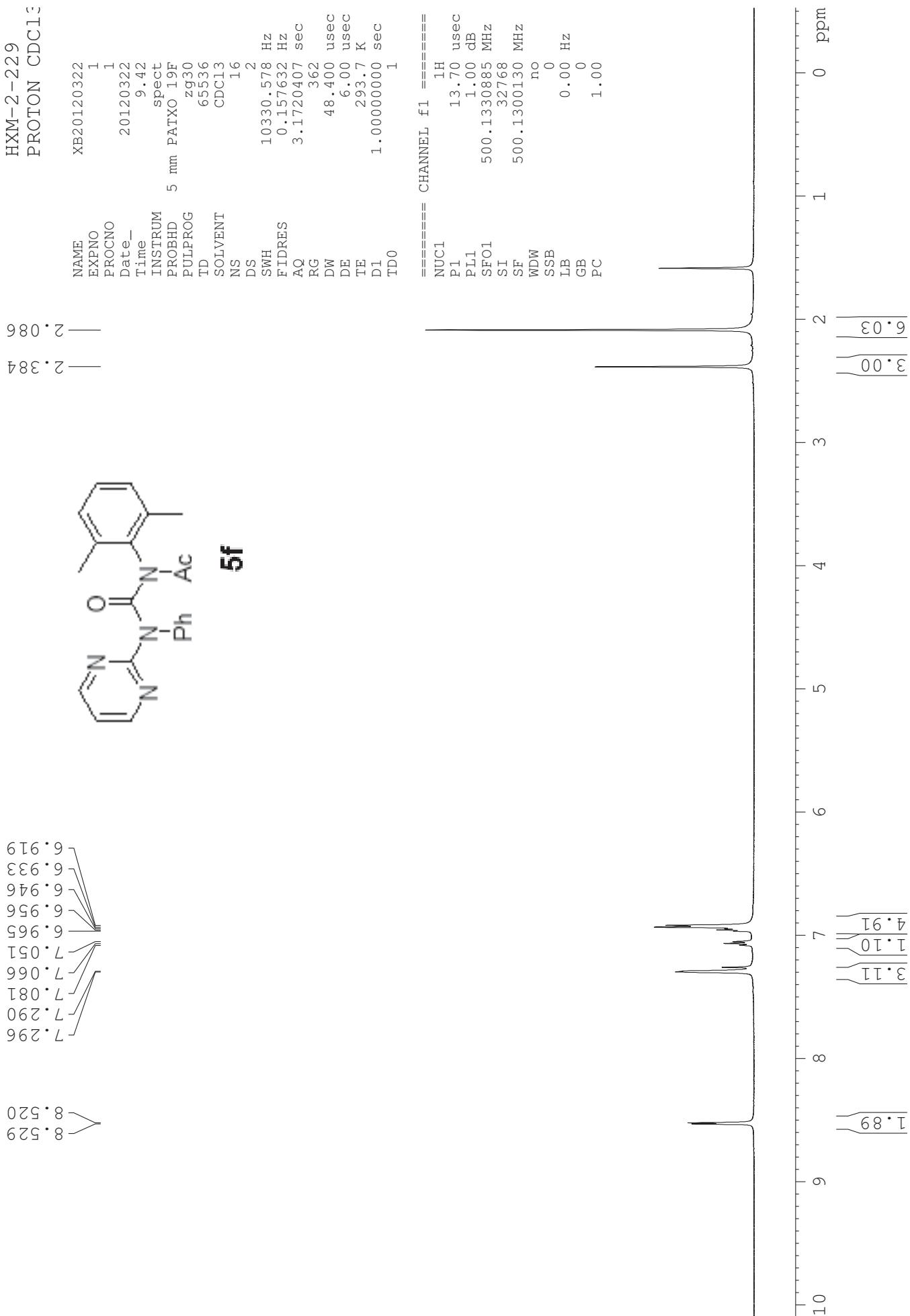
172.58

HXM-2-222
PROTON CDC13



HXM-2-222
C13CPD CDC13

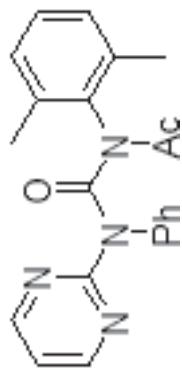




HXM-2-229
C13CPD CDCL₃

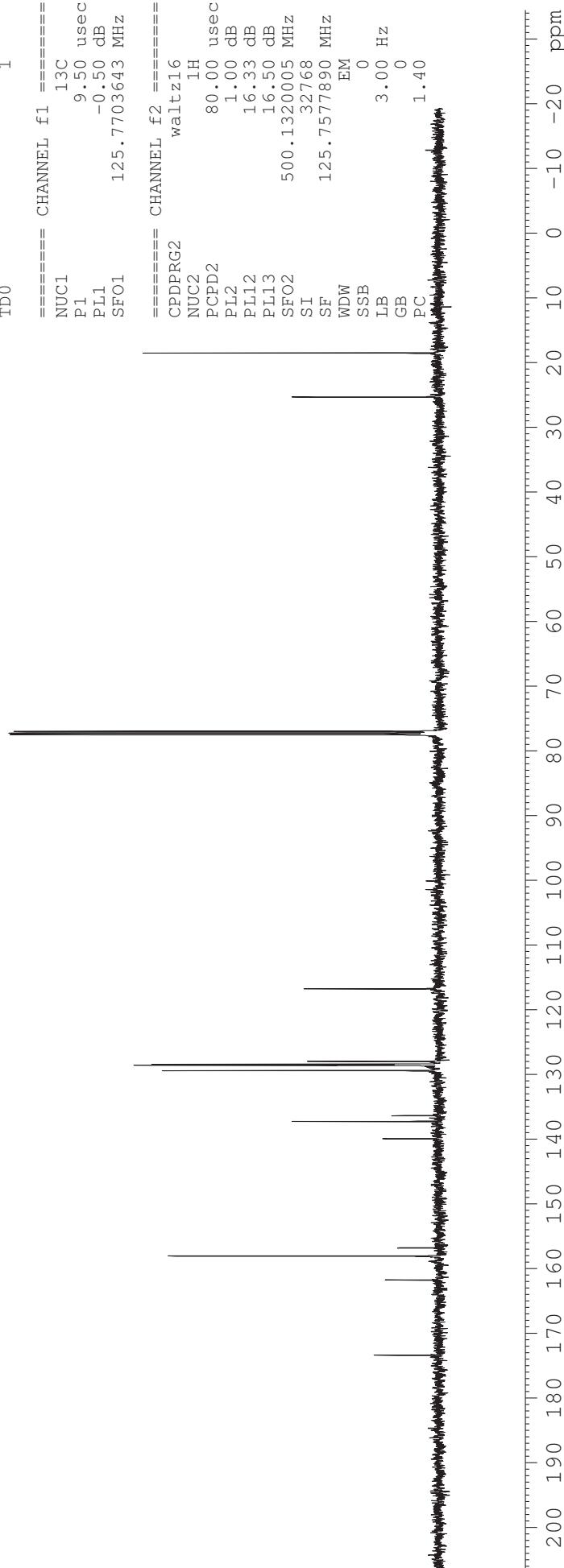
NAME XB20120322
EXPNO 8
PROCNO 1
Date 20120322
Time 10.28
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgppg30
TD 65536
SOLVENT CDCl₃
NS 59
DS 4
SWH 300030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 143.7
DW 16.650 usec
DE 6.00 usec
TE 294.8 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

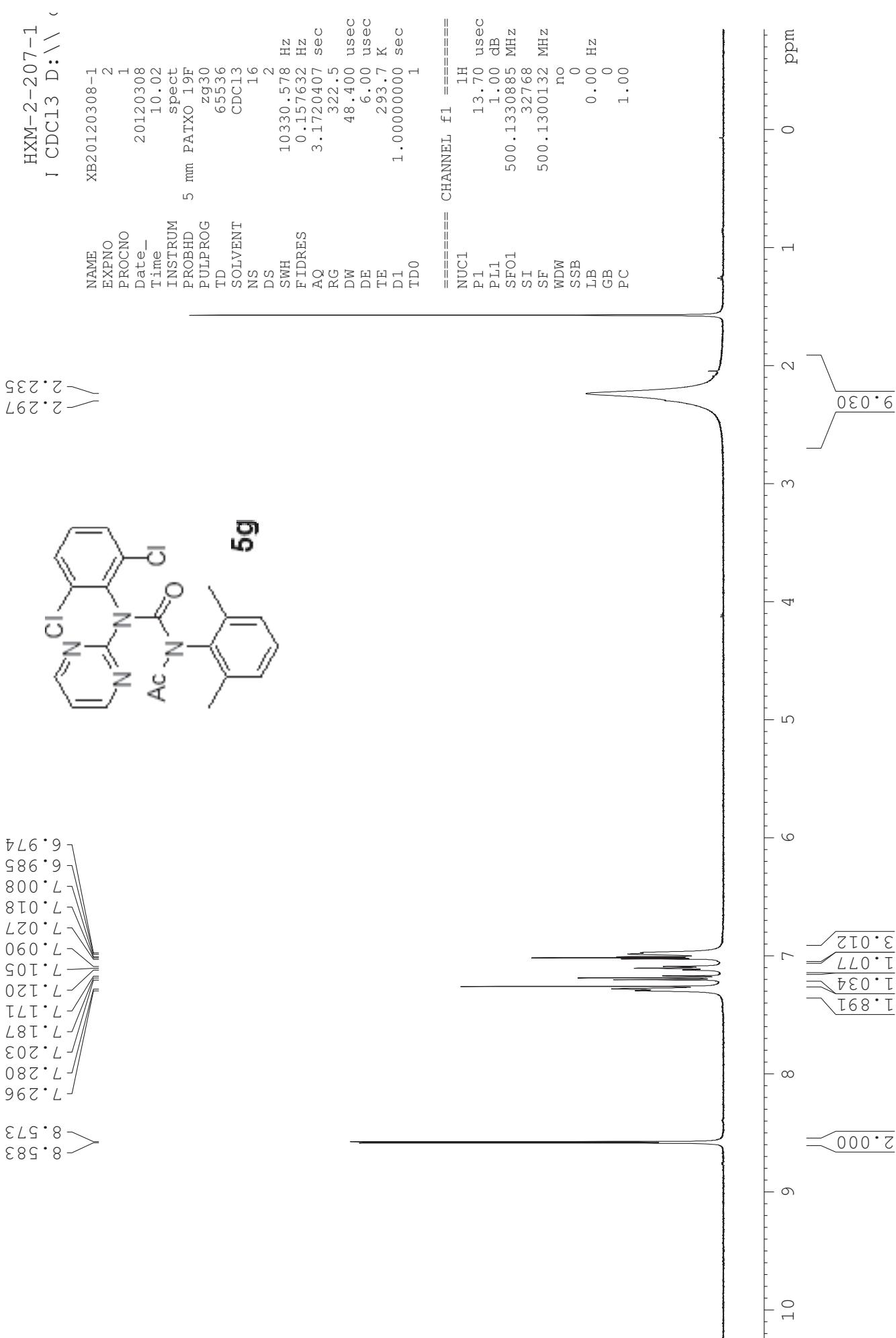
===== CHANNEL f1 =====
NUC1 ¹³C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 ¹H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 3.00 Hz
GB 0
PC 1.40

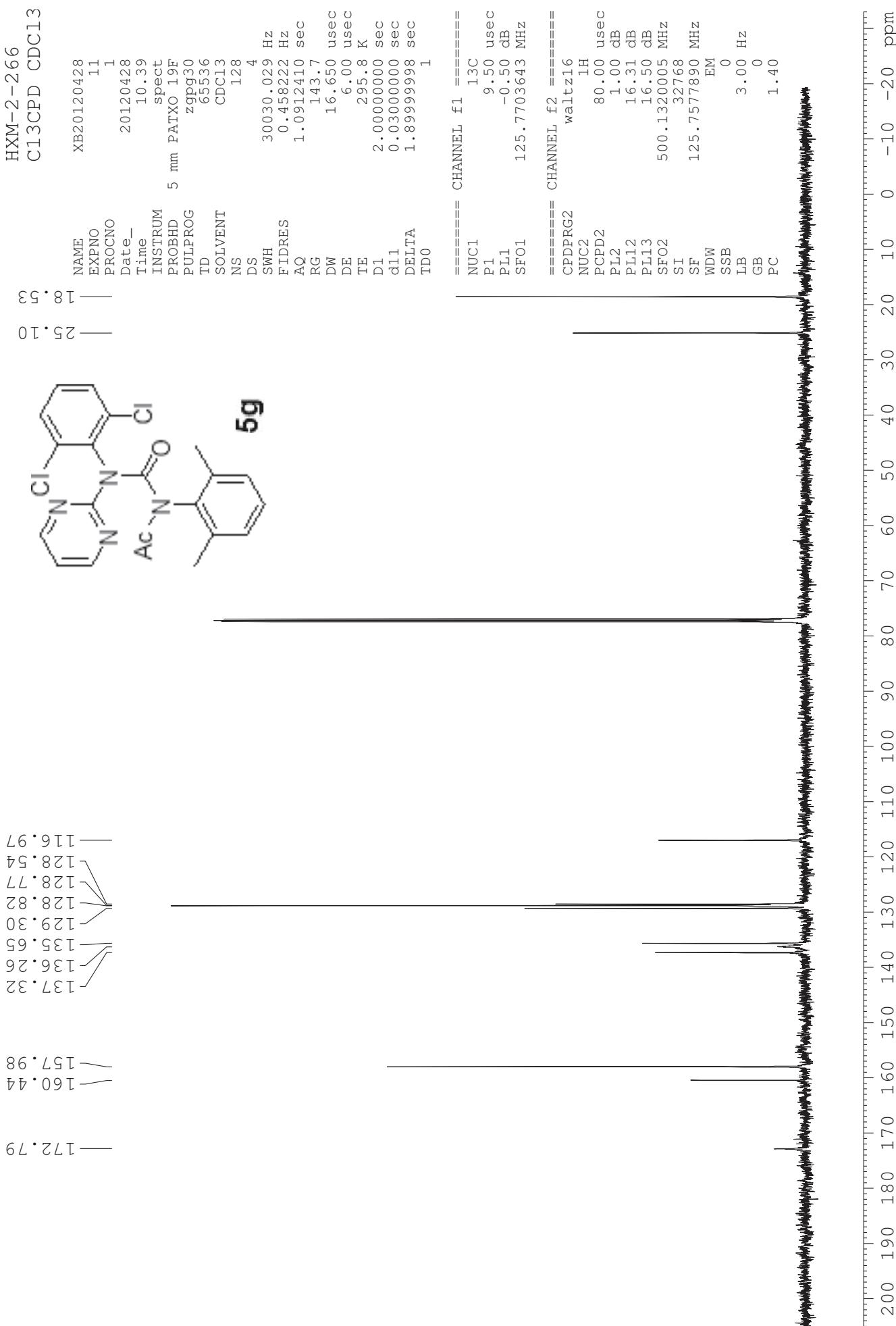


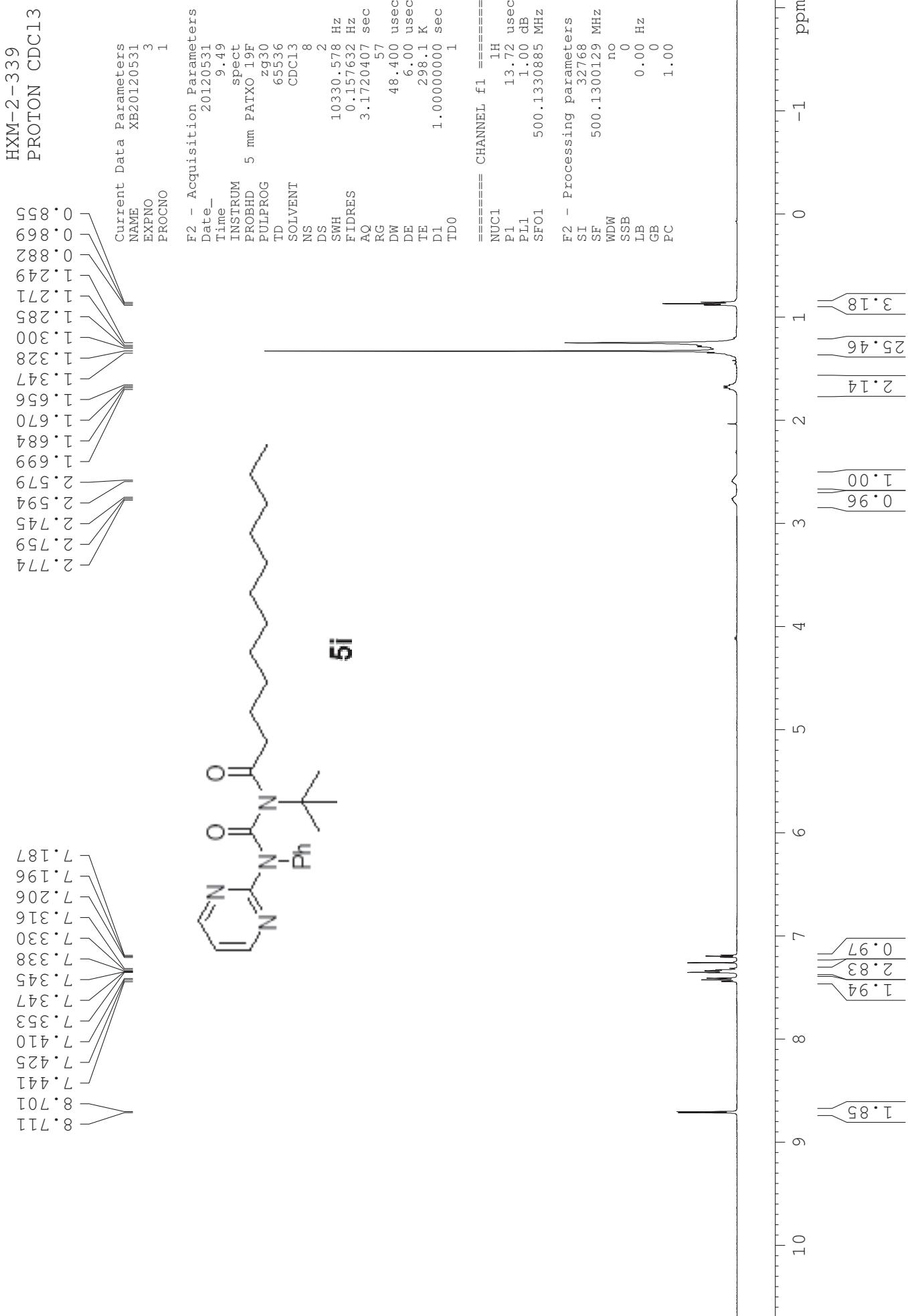
5f

116.66
127.88
128.32
128.42
128.48
129.30
129.36
137.16
139.84
156.70
157.97
161.66
173.30









HXM-2-339
C13CPD CDC13



Current Data Parameters
NAME XB20120531
EXPNO 12
PROCNO 1

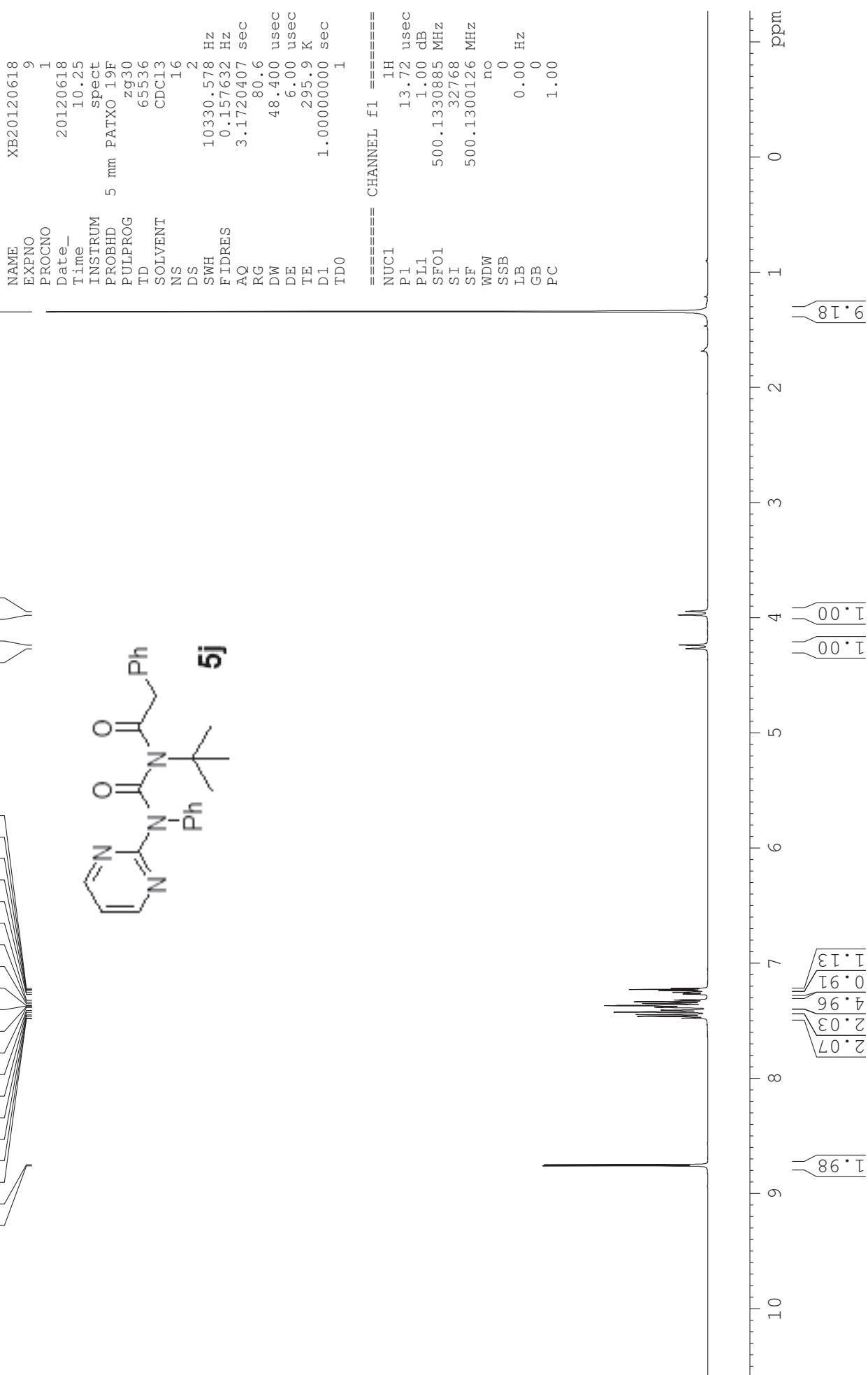
F2 - Acquisition Parameters
Date_ 20120531
Time_ 10.59
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zpgpg30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 128
DW 16.650 usec
DE 6.00 usec
TE 298.5 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 ======
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 ======
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 2.00 Hz
GB 0
TP 1.40



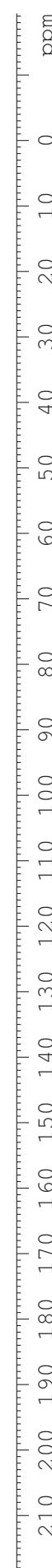
HXM-2-359
PROTON CDCl₃



HXM-2-359
C13CPD CDC13

NAME XB20120618
EXPNO 10
PROCNO 1
Date_ 20120618
Time 10.34
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 143.7
DW 16.650 usec
DE 6.00 usec
TE 297.0 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

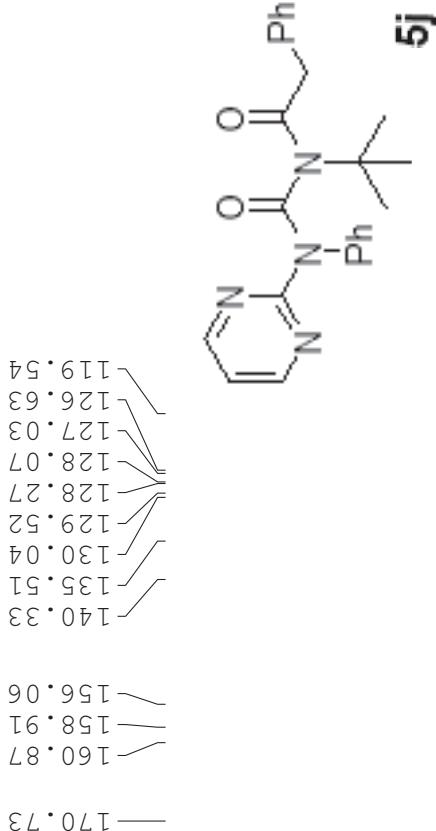
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768 MHz
SF 125.7577890 MHz
WDW EM
SSB 0
LB 2.00 Hz
GB 0
PC 1.40



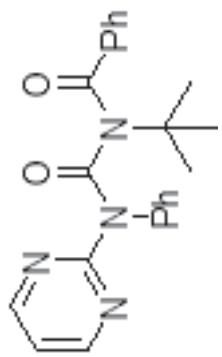
— 27.75 —

— 43.78 —

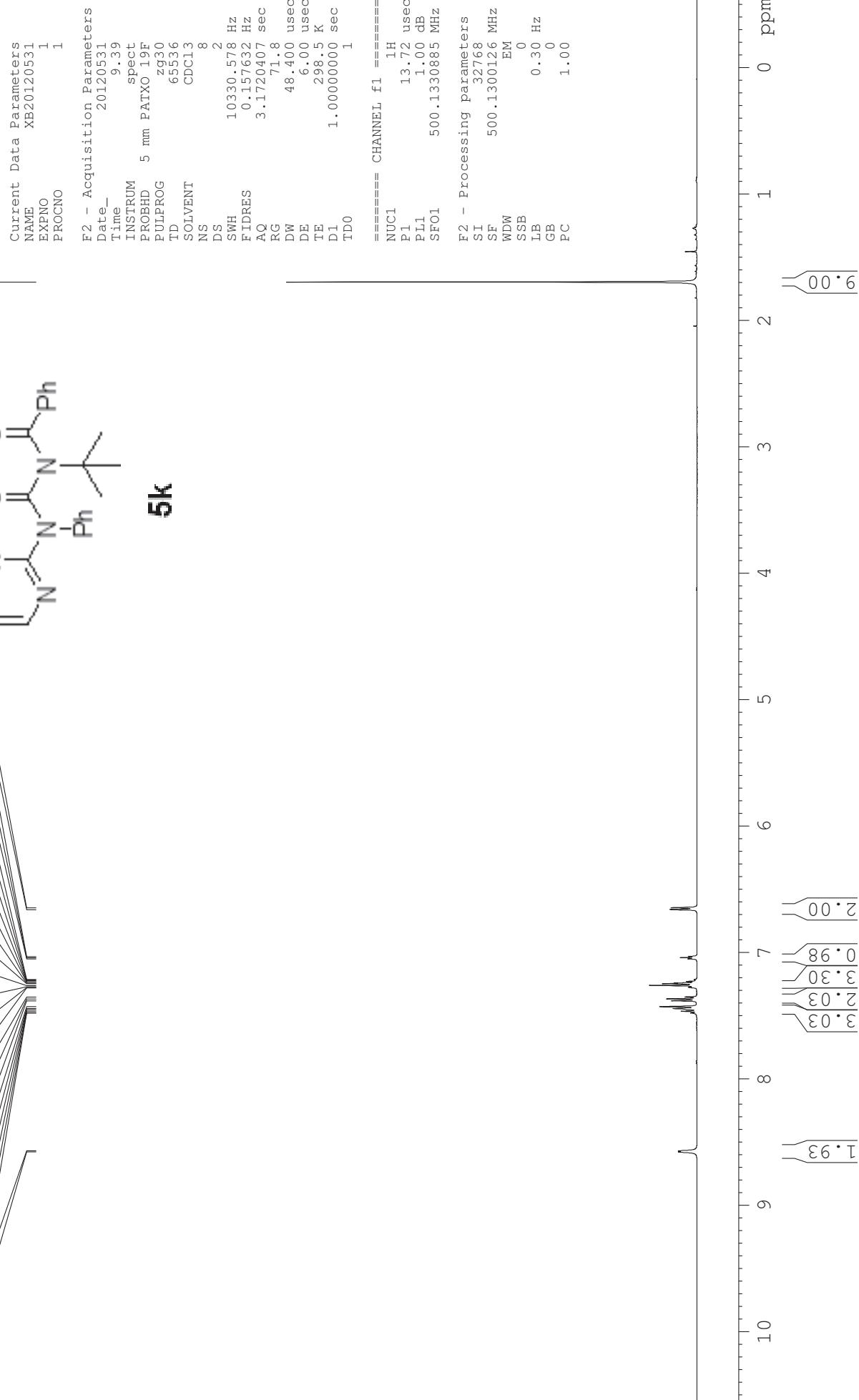
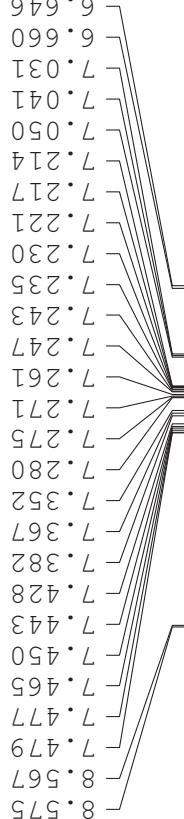
— 59.12 —



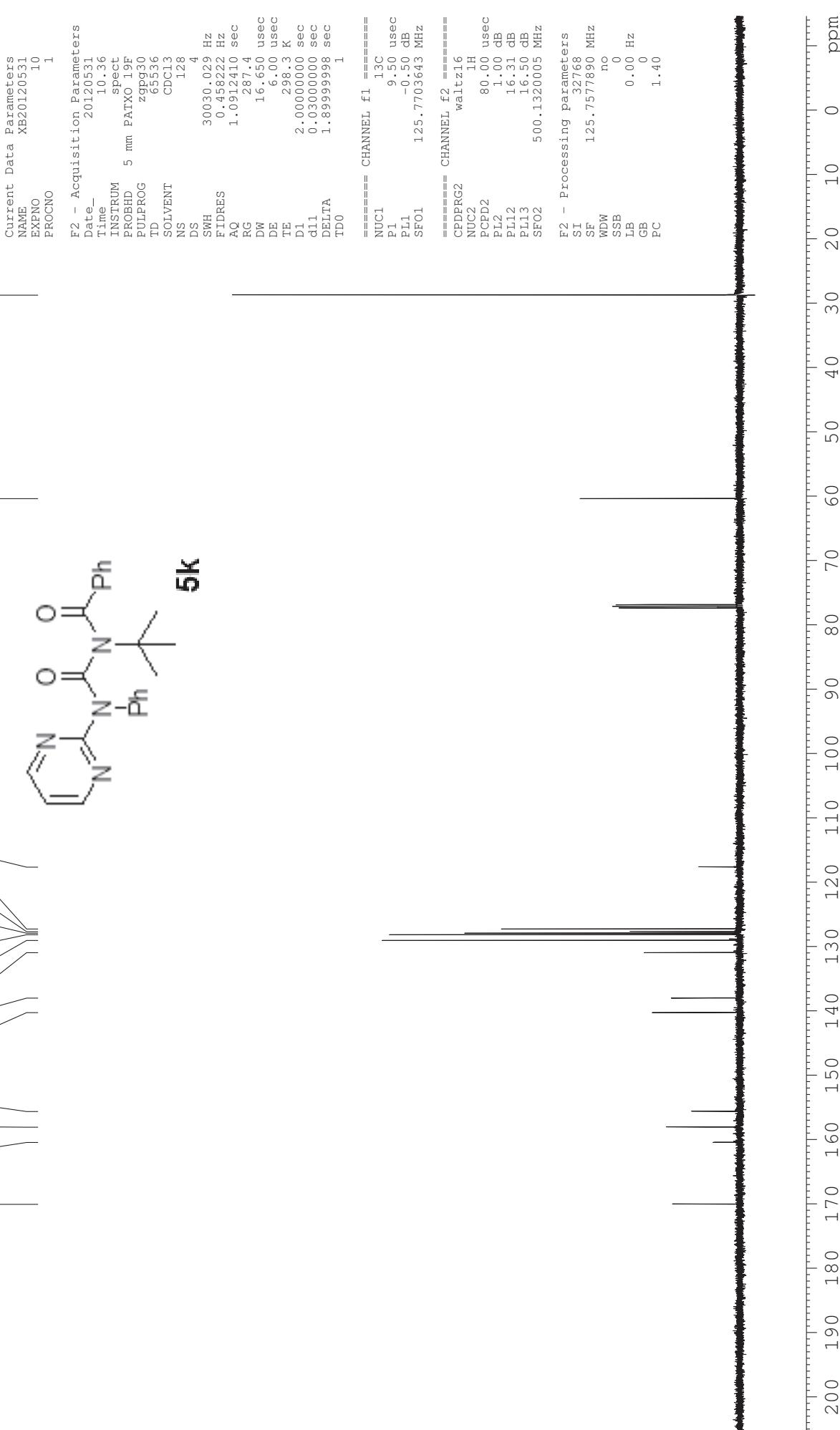
HXM-2-332
PROTON CDCl₃ I



5k



HXM-2-332
C13CPD CDCl₃ I
C13CPD CDCl₃ I



HXM-2-338
PROTON CDC13

Current Data Parameters
NAME XB20120531
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120531
Time 9.44
INSTRUM spect
PROBHD 5 mm PABRD 19F
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 8
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.170407 sec
RG 471.8
DW 48.400 usec
DE 6.00 usec
TE 298.4 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.72 usec
PL1 1.00 dB
SFO1 500.1330885 MHz

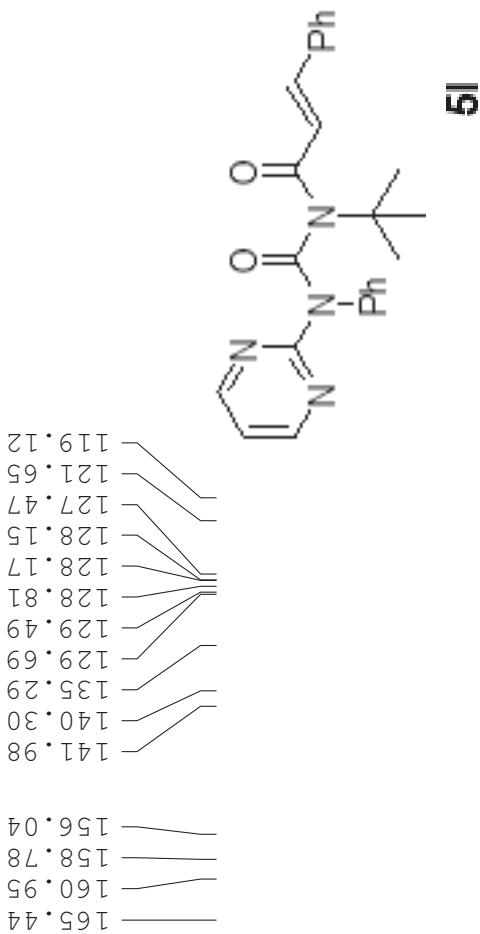
F2 - Processing parameters
SI 32768
SF 500.1300126 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

5l

HXM-2-338
C13CPD CDC13 1

28.03

59.29



Current Data Parameters
NAME XB20120531
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date 20120531
Time 10:49
INSTRUM spect
PROBHD 5 mm PAXO 19F
PULPROG zgpp30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 128
DW 16.650 usec
DE 6.00 usec
TE 298.5 K
D1 2.00000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz

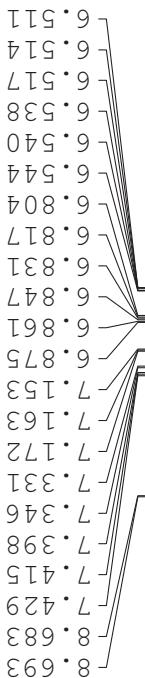
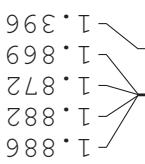
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0 Hz
GB 1.0
PC 1.40



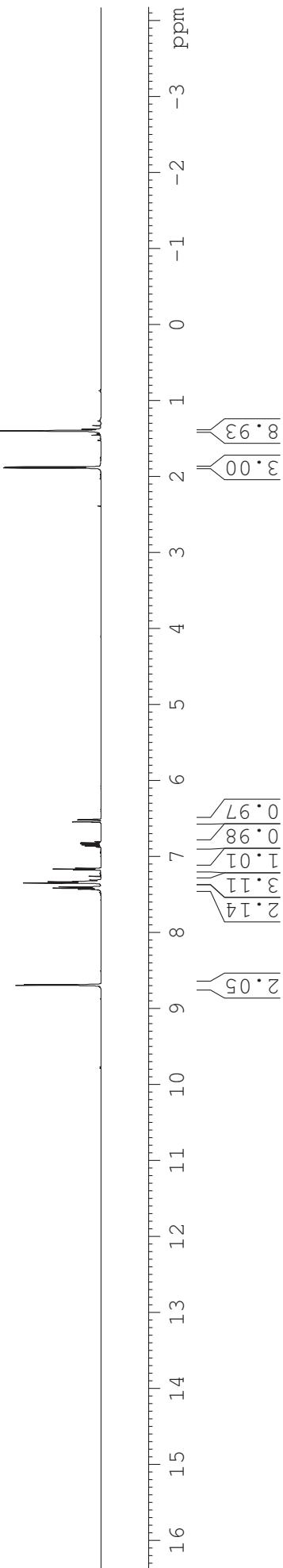
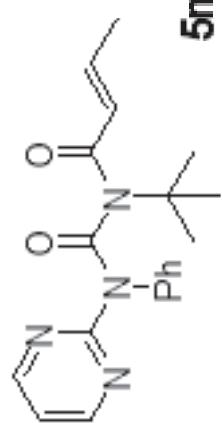
HXM-2-352
PROTON CDCl₃

NAME XB20120618
EXPNO 7
PROCNO 1
Date_ 20120618
Time 10.11
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 8
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1720407 sec
RG 80.6
DW 48.400 usec
DE 6.00 usec
TE 295.7 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.72 usec
PL1 1.00 dB
SFO1 500.1330885 MHz
SI 32768
SF 500.1300129 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



5m



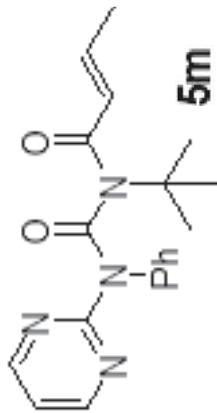
HXM-2-352
C13CPD CDC13

NAME XBB20120618
EXPNO 8
PROCNO 1
Date_ 20120618
Time 10.19
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 143.7
DW 16.650 usec
DE 6.00 usec
TE 296.8 K
D1 2.0000000 sec
d11 0.03000000 sec
DELTA 1.8999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768 MHz
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

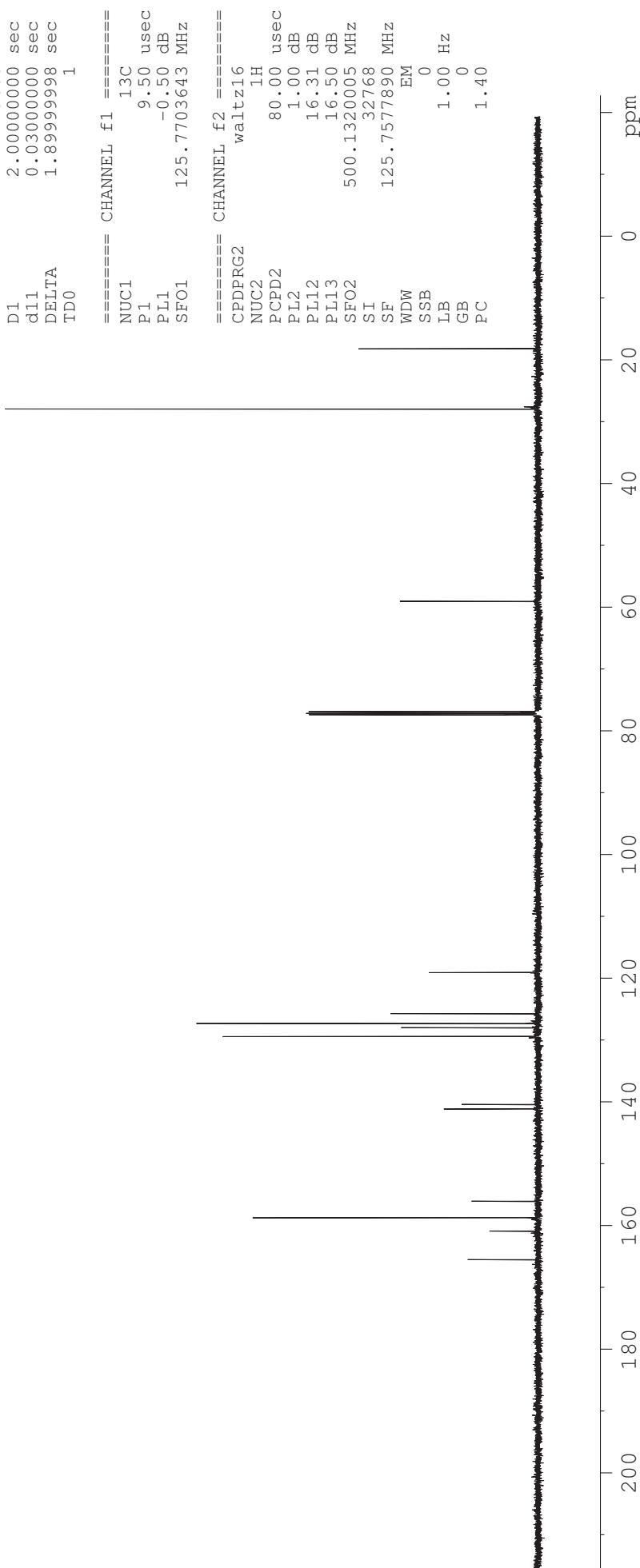
— 18.13 —
— 27.90 —

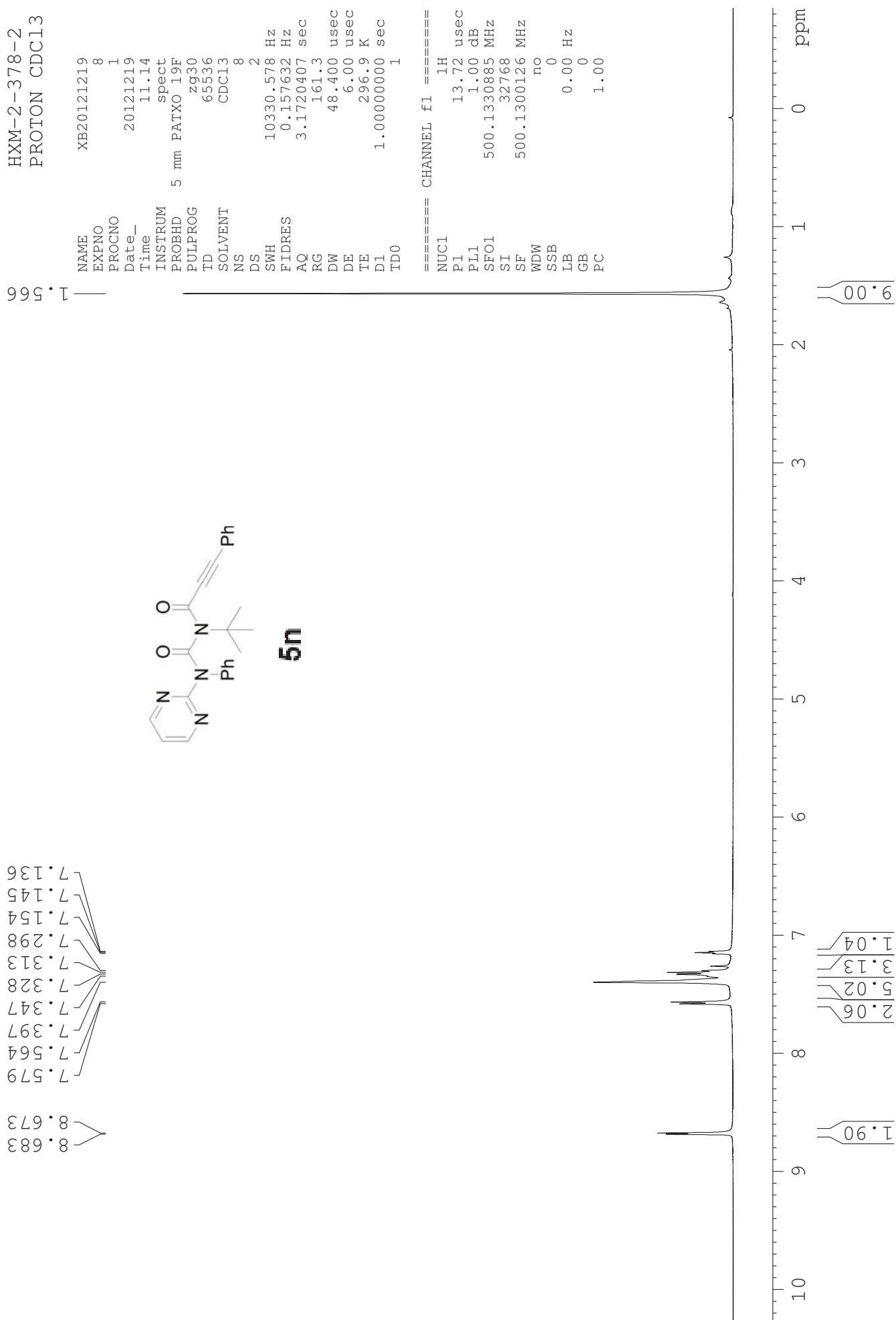
— 59.00 —



111.9.07
125.70
127.28
127.99
129.40
140.40
141.12
156.06
158.72
160.88
165.49

156.06
158.72
160.88
165.49





HXM-2-378-2
C13CPD CDC13

NAME XB20121219
EXPNO 21
PROCNO 1
Date_ 20121219
Time 17.41
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 300030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 161.3
DW 16.650 usec
DE 6.00 usec
TE 297.6 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

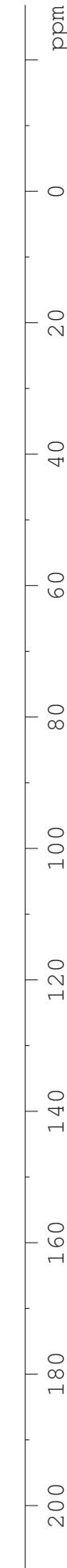
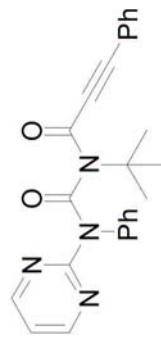
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

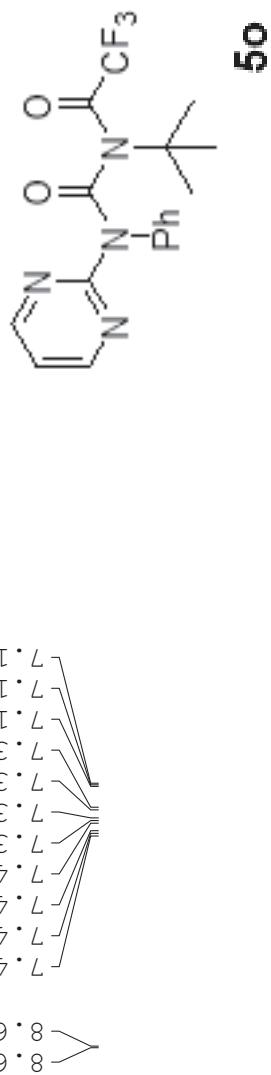
— 27.71 —

— 60.08 —

— 88.32 —
— 83.64 —

118.65
120.42
127.62
128.01
128.38
129.34
130.07
132.88
140.77
151.79
154.50
158.35
160.76



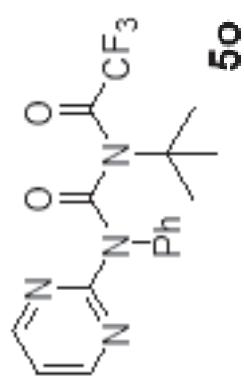


50

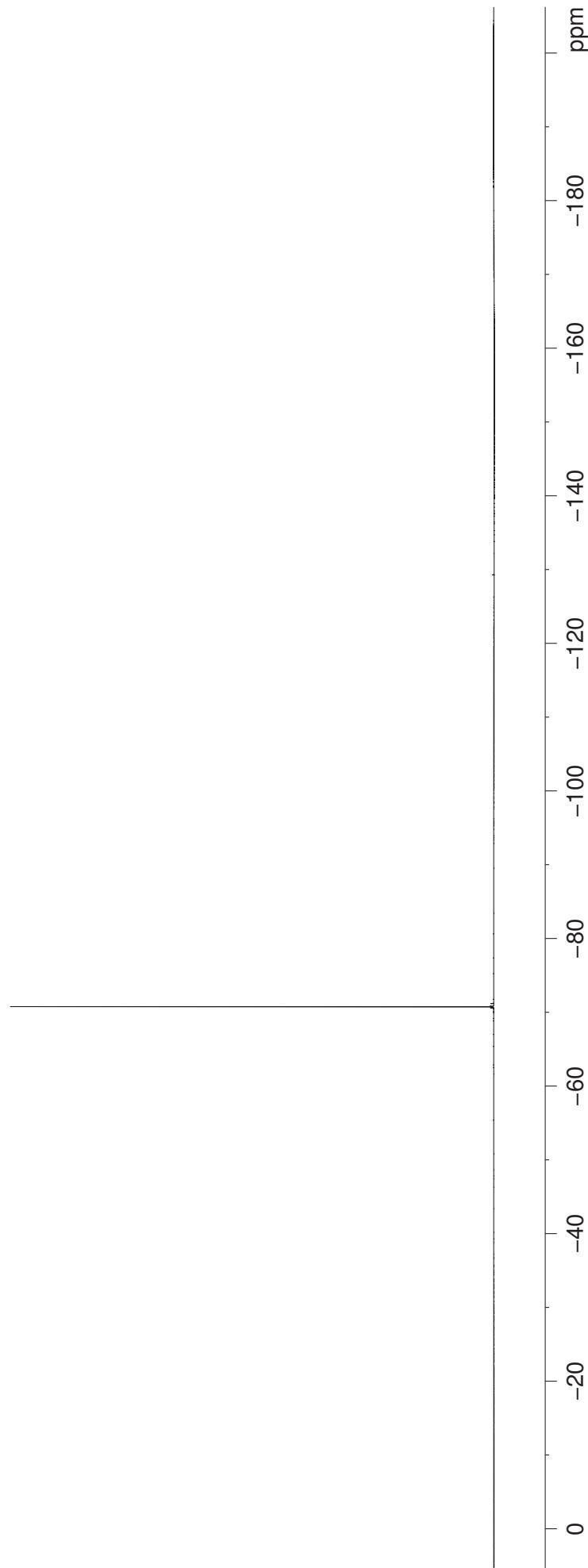
=====
EXPNO 14
PROCNO 1
Date 20120611
Time 14.33
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1720407 sec
RG 256
DW 48.400 usec
DE 6.000 usec
TE 296.8 K
D1 1.0000000 sec
TD0 1
=====
CHANNEL f1 ======
NUC1 ¹H
P1 13.72 usec
PL1 1.00 dB
SF01 500.133085 MHz
SI 32768
SF 500.1300129 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ppm

HXM-2-358
Jftt CDCl₃ D:\den



— -70.743 —



HXM-2-358
C13CPD CDC13

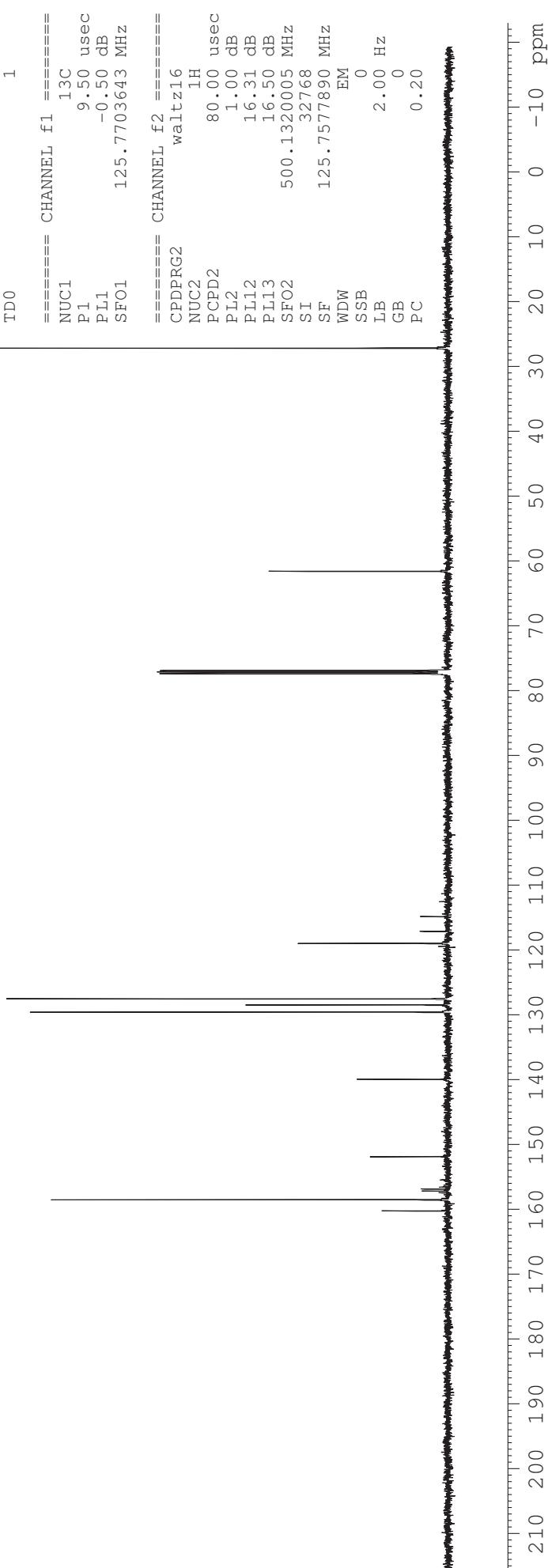
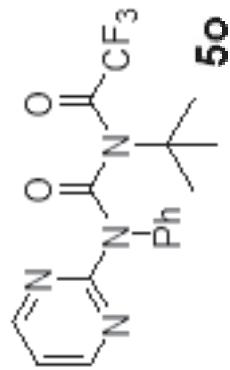
NAME XBX20120618
EXPNO 12
PROCNO 1
Date_ 20120618
Time 10.48
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgppg30
TD 65536
SOLVENT CDCl₃
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 1290.2
DW 16.650 usec
DE 6.00 usec
TE 297.0 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 2.00 Hz
GB 0
PC 0.20

27.08

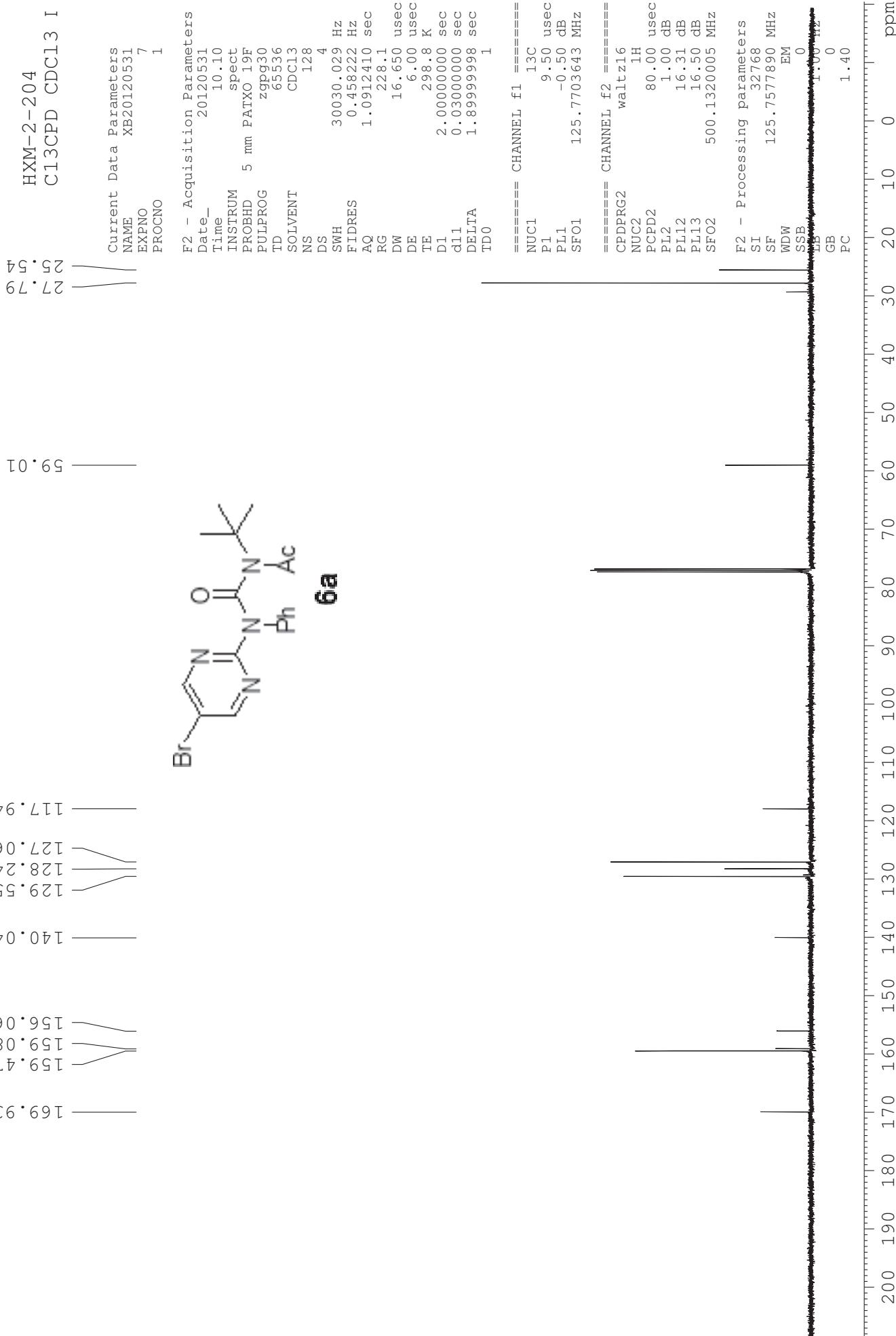
61.54

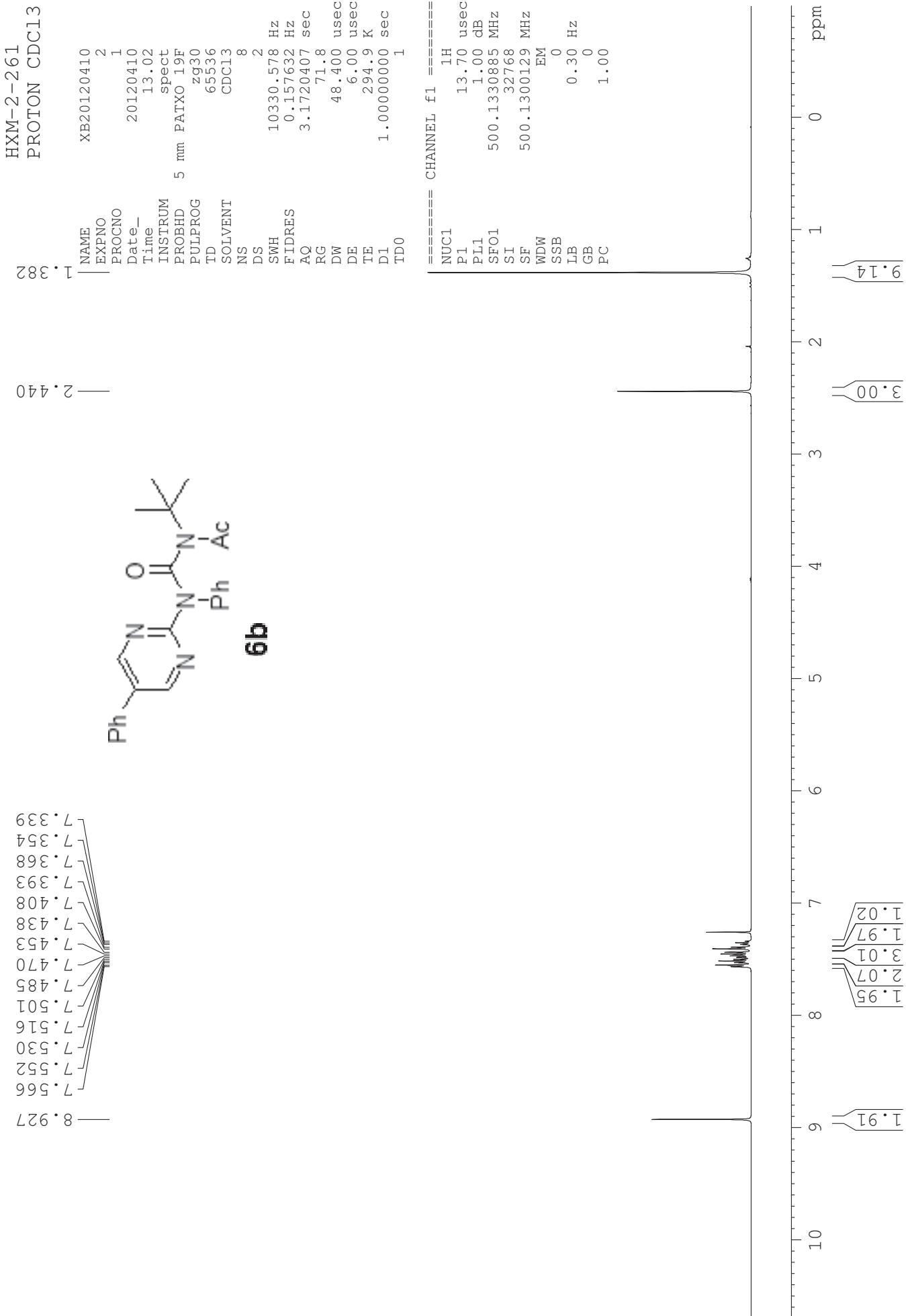
112.52
114.81
117.10
118.96
119.40
127.51
128.45
129.55
139.92
151.88
156.57
156.86
157.15
157.44
158.51
160.24



HXM-2-204
PROTON CDCl₃







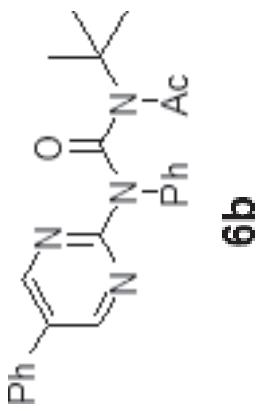
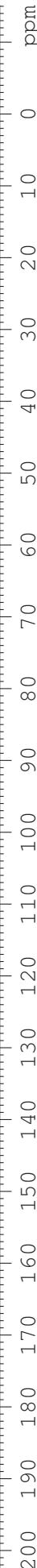
HXM-2-261
C13CPD CDC13

Current Data Parameters
NAME XB20120410
EXPNO 5
PROCNO 1

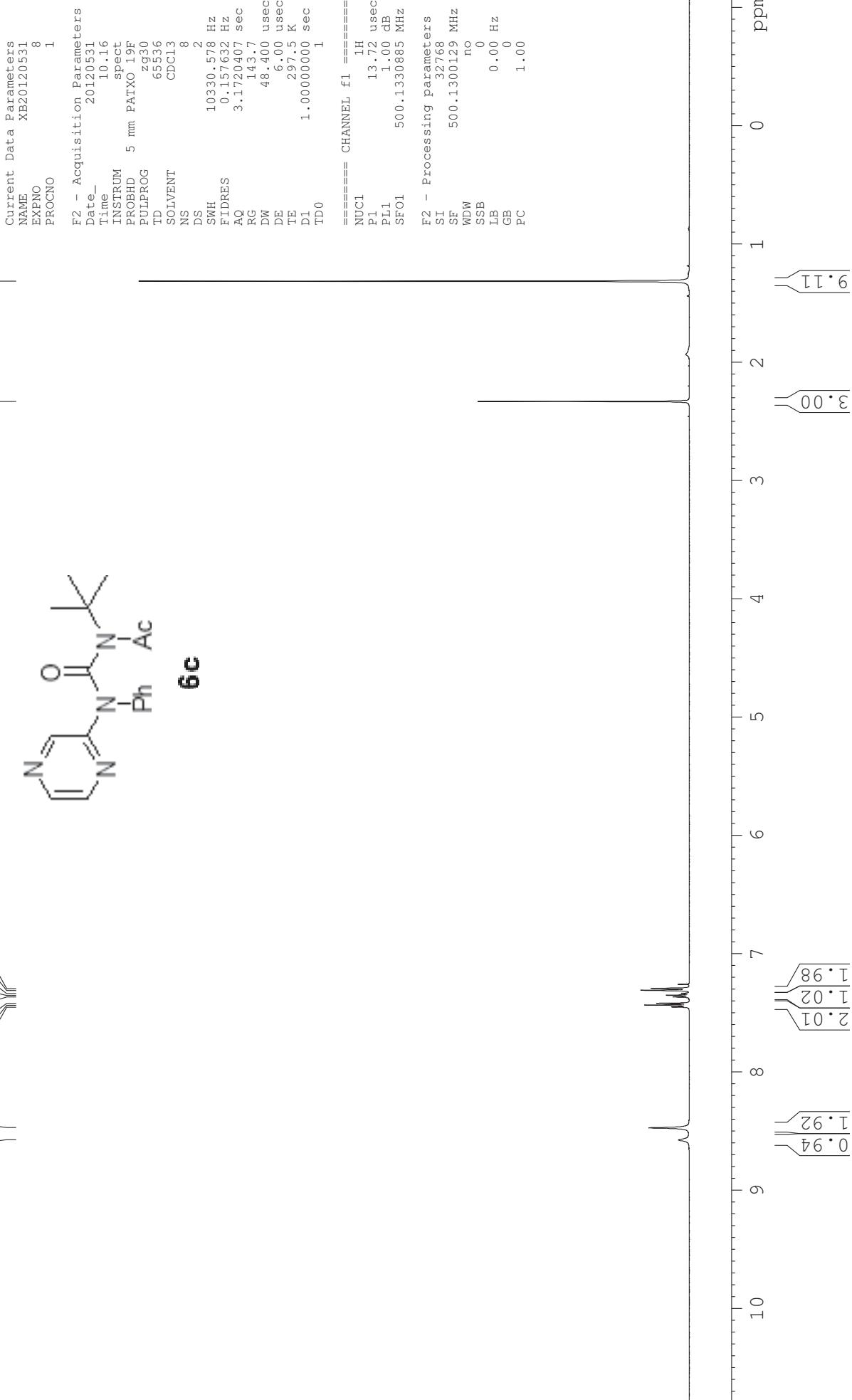
F2 - Acquisition Parameters
Date_ 20120410
Time 13.30
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgpp30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.45822 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 296.2 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1

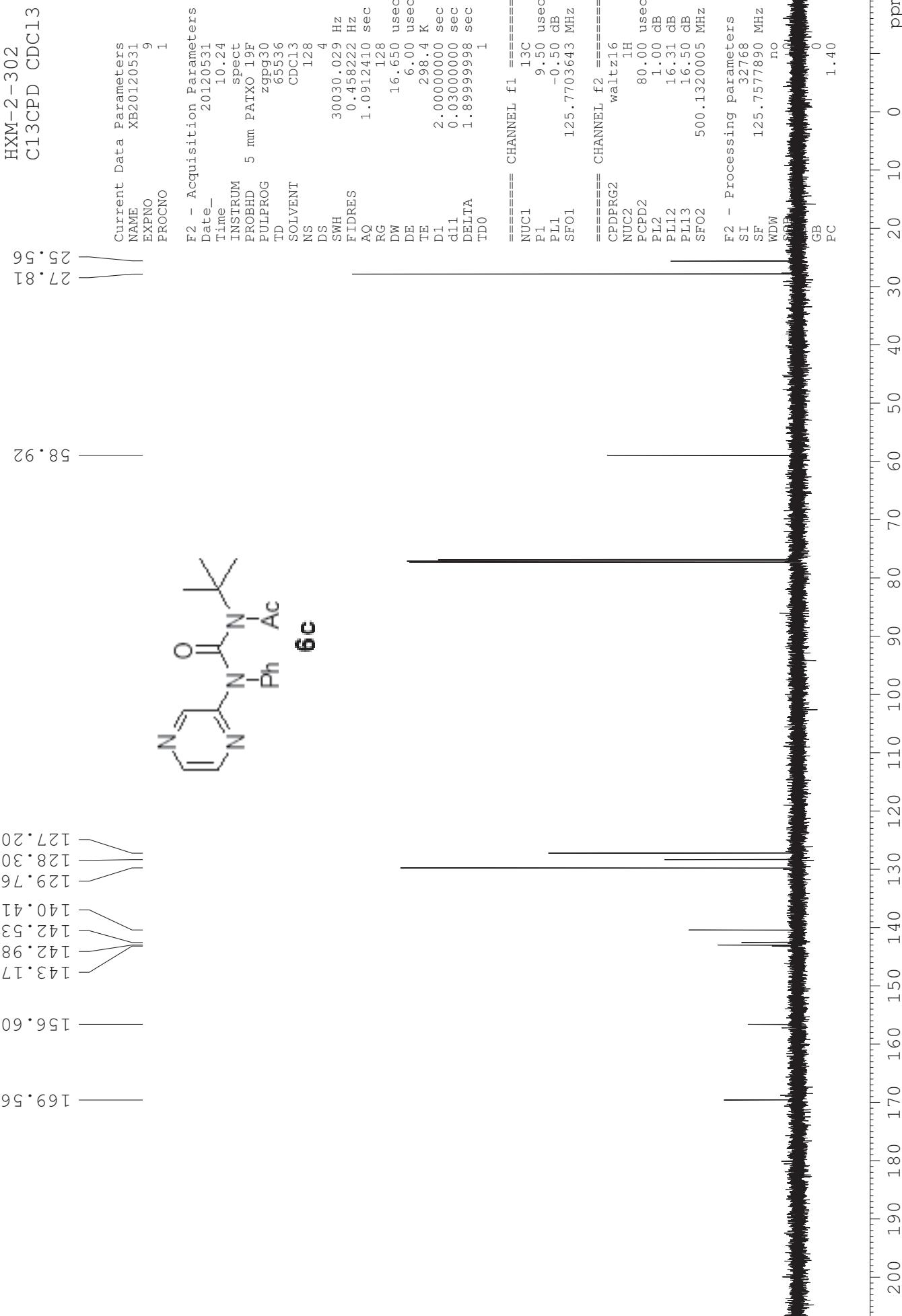
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SF01 125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2 w11z16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SF02 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0 0
LB 0.00 Hz
GB 0
PC 1.40



HXM-2-302
PROTON CDCl₃





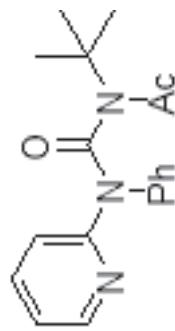
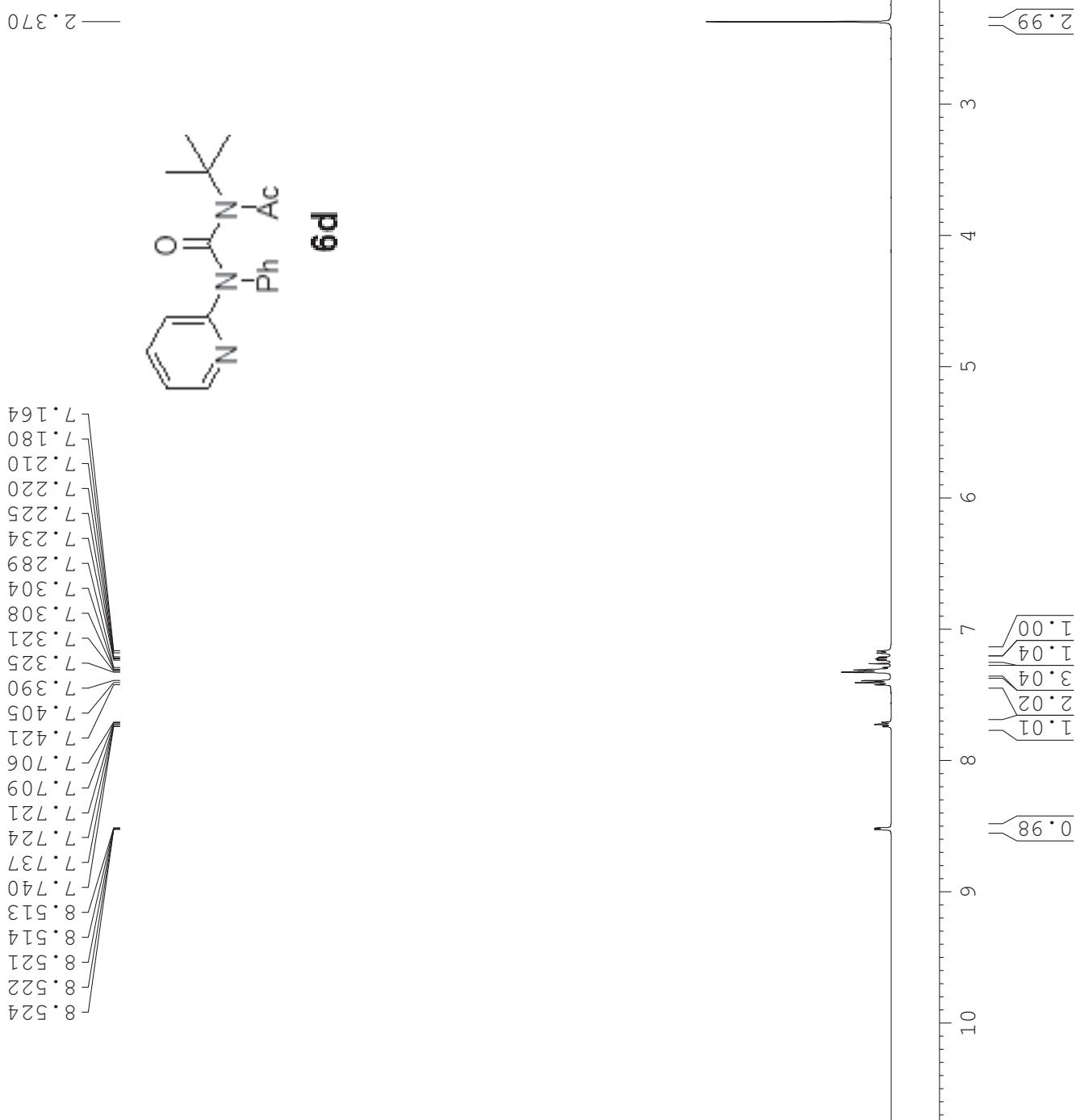
HXM-2-188
PROTON CDC13

NAME	EXPNO	PROCN0	DATE	TIME	INSTRUM	PROBHD	PULPROG	ID	SOLVENT	NS	DS	SWH	FIDRES	AQ	RG	DW	DE	TE	DC1	DDO
XB20120227	2		20120227	10.35	mm	PATXO	19F				2	10330.578	Hz							
					spec							0.157632	Hz							
												3.1720407	sec							
												322.5								
												48.400	usec							
												6.00	usec							
												293.6	K							
												1.00000000	sec							

```

===== CHANNEL f1 =====
NUC1          1H
?1           13..70 usec
?L1          1.00 dB
SF01         5000.11330885 MHz
?I           32768
?F           5000.13000132 MHz
WDW          EM
SSSB          0
?LB          0..30 Hz
?GB          0
?PC          1.00

```



20

HXM-2-188
C13CPD CDC13

Current Data Parameters
NAME XB20120228
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120228
Time 20:23
INSTRUM PROBHD spect
PROBHD 5 mm PATXO 19F
PULPROG zgppg30
TD 65536
SOLVENT C6D13
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.45822 Hz
AQ 1.0912410 sec
RG 114
DW 16.650 usec
DE 6.00 usec
TE 295.5 K
D1 2.0000000 sec
d11 0.03000000 sec
DELTA 1.8999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SF01 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 w11z16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SF02 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.757780 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

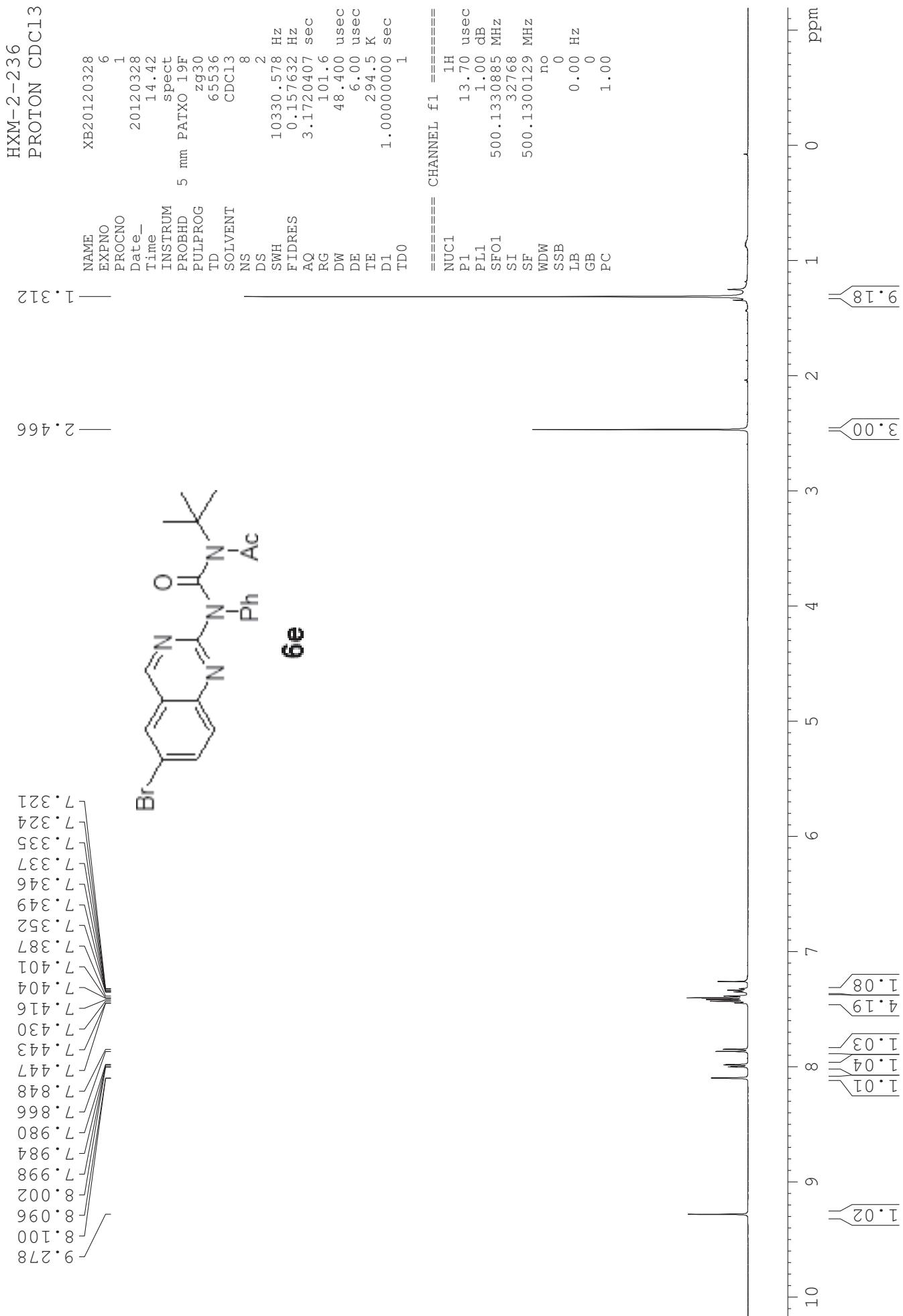


6d

58.63

121.62
122.60
126.88
127.62
129.45
138.34
141.40
149.29
154.90
156.52

169.79



HXM-2-236
C13CPD CDC13

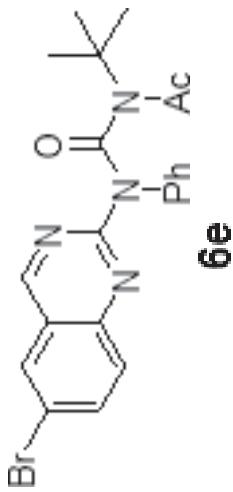
NAME XB20120328
EXPNO 8
PROCNO 1
Date 20120328
Time 14.52
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zgpp30
TD 65536
SOLVENT CDCl3
NS 128
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 322.5
DW 16.650 usec
DE 6.00 usec
TE 2.95.8 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.33 dB
PL13 16.50 dB
SFO2 500.11320005 MHz
SI 32768
SF 125.7577890 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.40

25 . 76
27 . 79

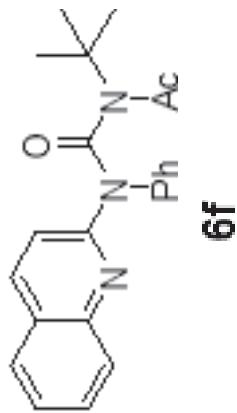
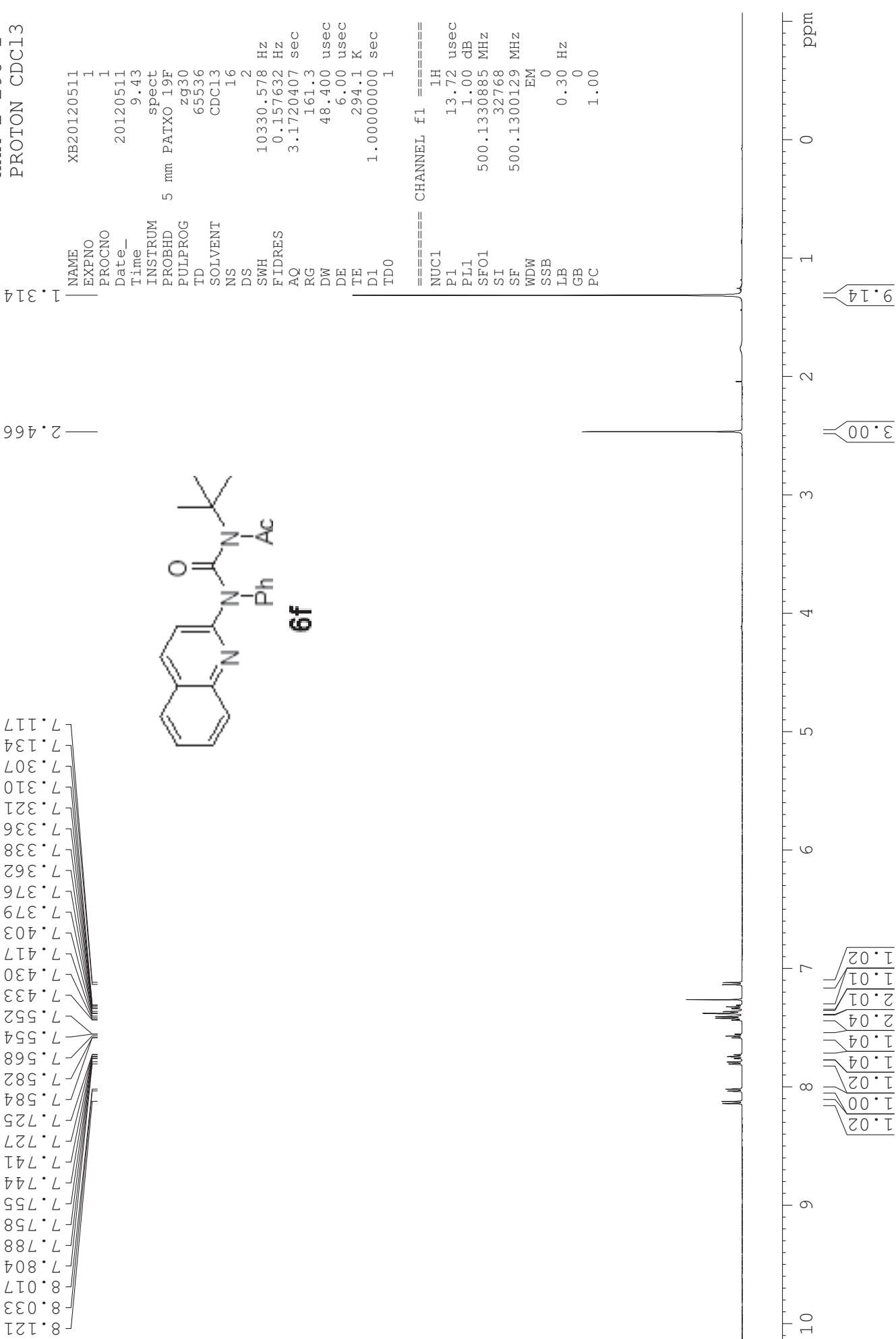
— 58 . 84 —



122.25
123.84
127.07
127.98
129.27
129.44
129.93
138.71
140.33
149.65
156.34
157.05
161.40
170.00



HXN-2-298-1
PROTON CDC13



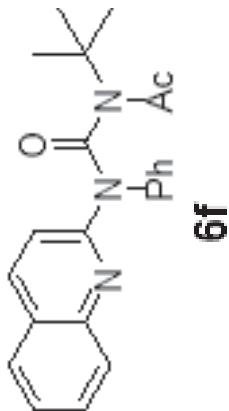
HXM-2-298
C13CPD CDC13

NAME xb20120514
EXPNO 9
PROCNO 1
Date 20120514
Time 16.51
INSTRUM spect
PROBHD 5 mm PATXO 19F
PULPROG zppg30
TD 65536
SOLVENT CDCl3
NS 39
DS 4
SWH 300030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 101.6
DW 16.650 usec
DE 6.00 usec
TE 295.6 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TD0 1
===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 125.7703643 MHz
SFO1 1
===== CHANNEL f2 =====
CPDPG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.31 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 327.68 MHz
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

25.87
27.85

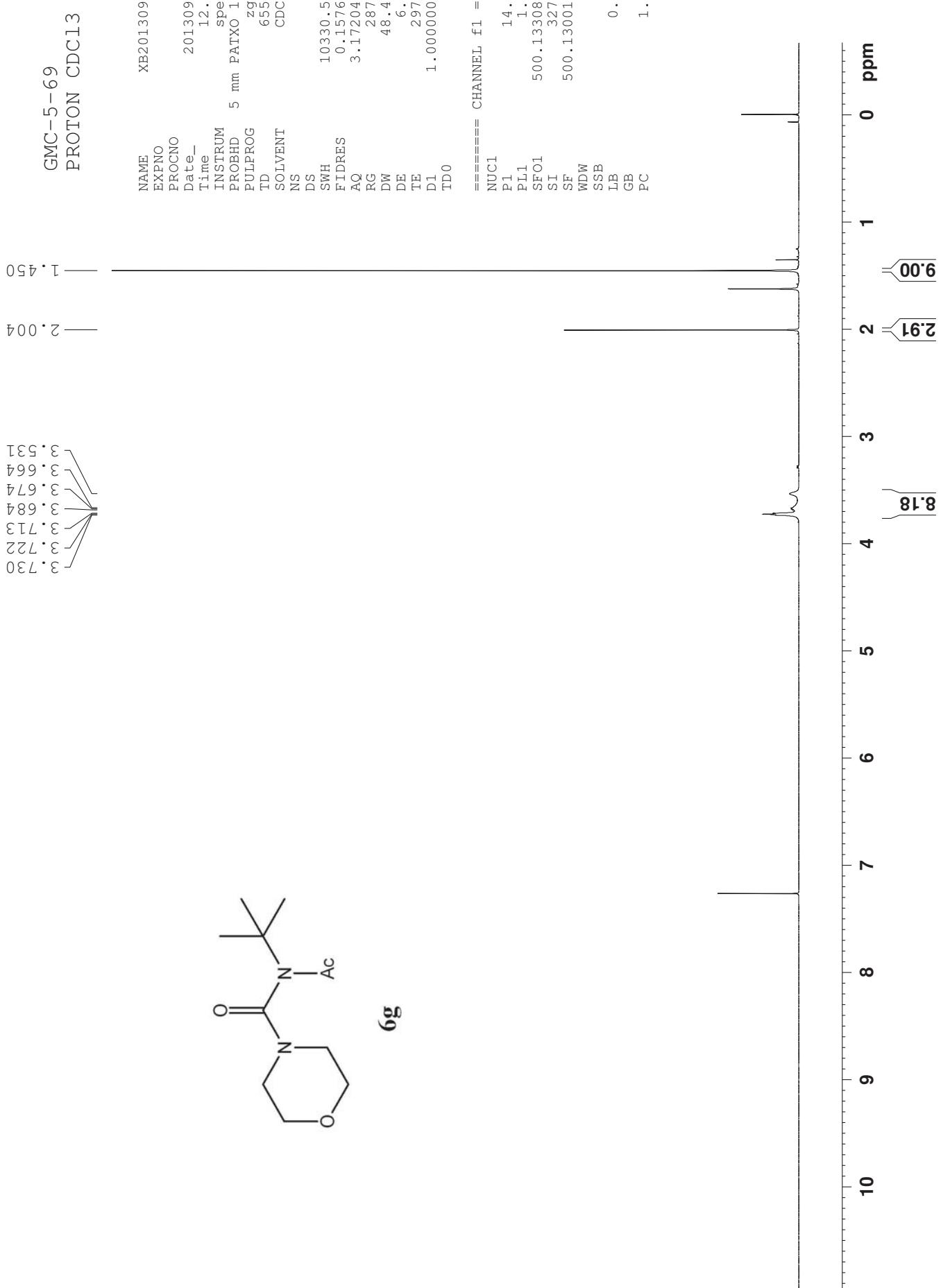
58.64

119.13
126.64
127.01
127.36
127.40
127.45
127.50
127.55
129.19
129.52
129.52
130.01
138.69
141.01
146.89
153.12
156.63



169.73





GMC-5-69
C13CPD CDC13

NAME XB20130912
EXPNO 2
PRCFCNO 1
Date 20130912
Time 9.53
INSTRUM spect
PROBHD 5 mm PAXO 1.9F
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0912410 sec
RG 362
DW 16.650 usec
DE 6.00 usec
TE 298.1 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -0.50 dB
SFO1 125.7703643 MHz

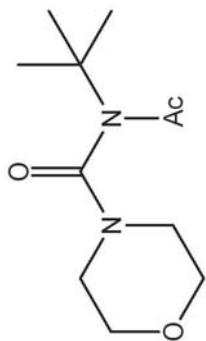
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 1.00 dB
PL12 16.05 dB
PL13 16.50 dB
SFO2 500.1320005 MHz
SI 32768
SF 125.7577709 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 0.20

23.98
28.10

43.79
47.12

57.88

66.40
66.68



6g

156.32

168.65

