

An Unusual Macrocyclization Reagent for Highly Selective One-Pot Synthesis of Strained Macrocyclic Aromatic Hexamers†

Haoliang Fu,^a Hu Chang,^b Jie Shen,^{c,d} Lin Yu,^a Bo Qin,^{*b} Kun Zhang,^{*a} and Huaqiang Zeng^{*c,d}

^a Faculty of Chemical Engineering and Light Industry, Guang Dong University of Technology, Guang Dong, China 510006, China

^b College of Chemistry and Chemical Engineering, Chongqing University, Chongqing, China 400030. E-mail: qinbo@cqu.edu.cn

^c Department of Chemistry, National University of Singapore, 3 Science Drive 3, Singapore 117543. E-mail: chmzh@nus.edu.sg; Tel: +65-6516-2683

^d Institute of Bioengineering and Nanotechnology, 31 Biopolis Way, The Nanos, Singapore 138669. E-mail: hqzeng@ibn.a-star.edu.sg

Supporting Information

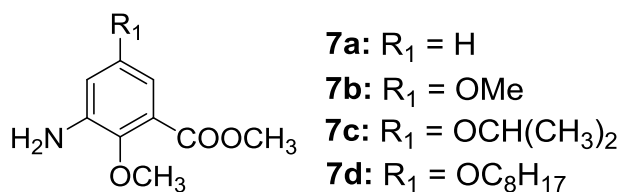
Table of contents

1. General remarks	S2
2. Synthesis of amino esters 7a-7d	S3
3. ¹ H NMR, ¹³ C NMR, and (HR)MS of hexamers 8a , 8c and 8d	S4
4. ¹ H NMR, ¹³ C NMR, and (HR)MS of intermediate oligomers P2-P5	S7

1. General remarks

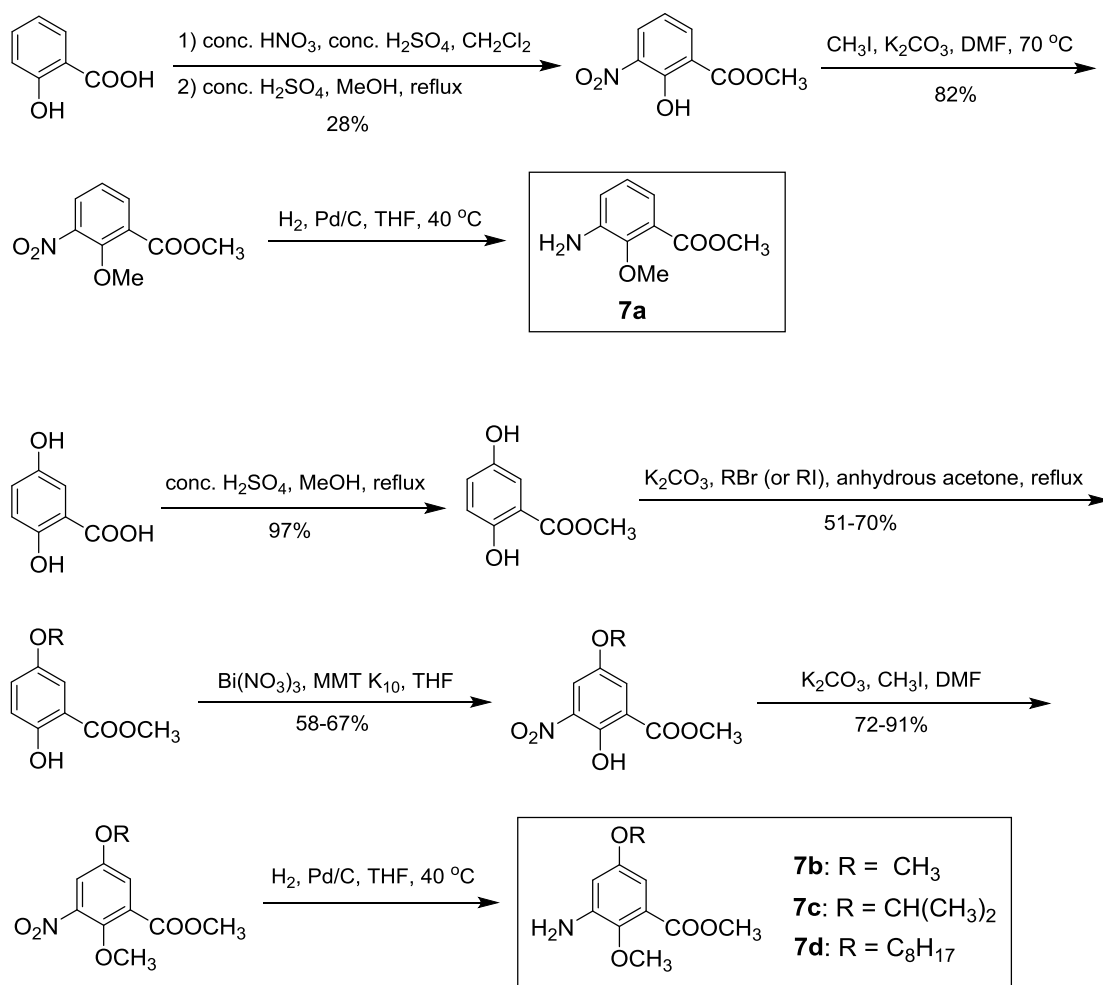
^1H NMR and ^{13}C NMR data were obtained on a Bruker AMX300 or 600 (300 or 600 MHz) nuclear resonance spectrometer with CDCl_3 as solvent and tetramethylsilane (TMS) as internal standard. Chemical shifts were reported in units (ppm) by assigning TMS resonance in the ^1H NMR spectra as 0.00 ppm (chloroform, 7.26 ppm). Data were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet and m = multiplet), coupling constant (J values) in Hz and integration. Chemical shifts for ^{13}C NMR spectra were recorded in ppm from tetramethylsilane using the central peak of CDCl_3 (77.0 ppm) as the internal standard. All reagents were purchased from Acros, Aldrich, and Alfa Aesar without further purification in advance before use.

2. Synthesis of monomeric amino esters 7a-7d have been reported in the following journal articles

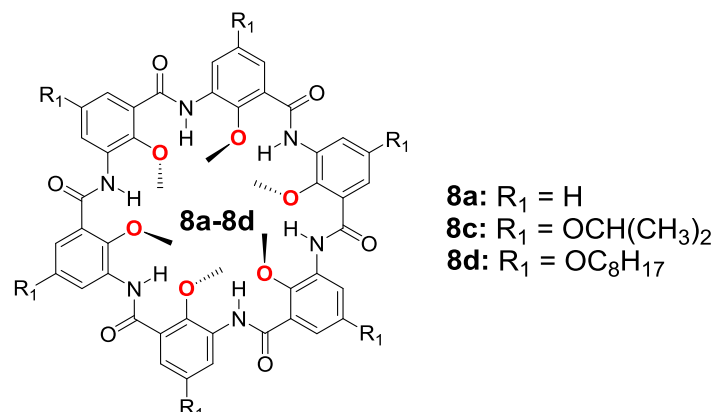


For synthesis of **7a**, see: B. Qin, X. Y. Chen, X. Fang, Y. Y. Shu, Y. K. Yip, Y. Yan, S. Y. Pan, W. Q. Ong, C. L. Ren, H. B. Su and H. Q. Zeng, *Org. Lett.* 2008, **10**, 5127.

For synthesis of **7b-7d**, see: Y. Yan, B. Qin, C. L. Ren, X. Y. Chen, Y. K. Yip, R. J. Ye, D. W. Zhang, H. B. Su and H. Q. Zeng, *J. Am. Chem. Soc.* 2010, **132**, 5869.



3. ^1H NMR, ^{13}C NMR, and (HR)MS of hexamers **8a**, **8c** and **8d**

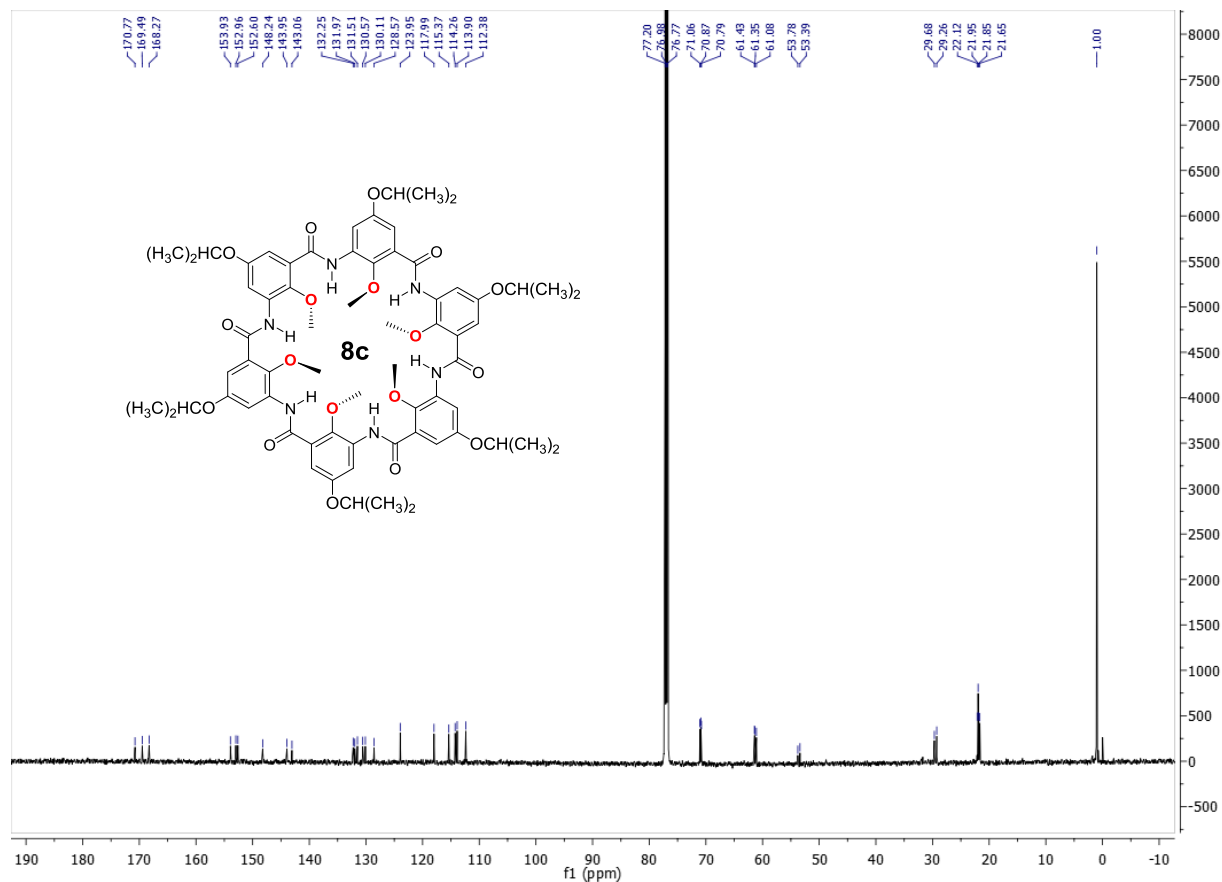
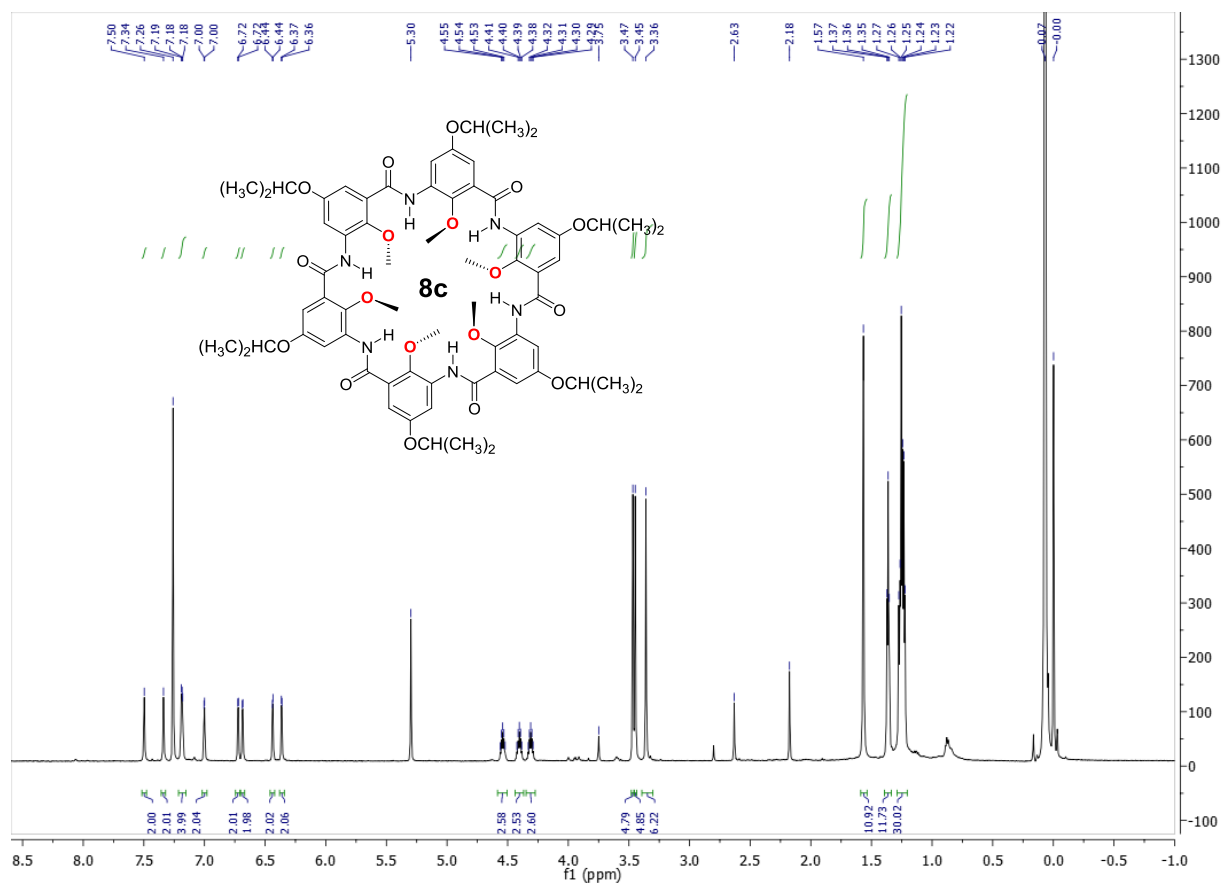


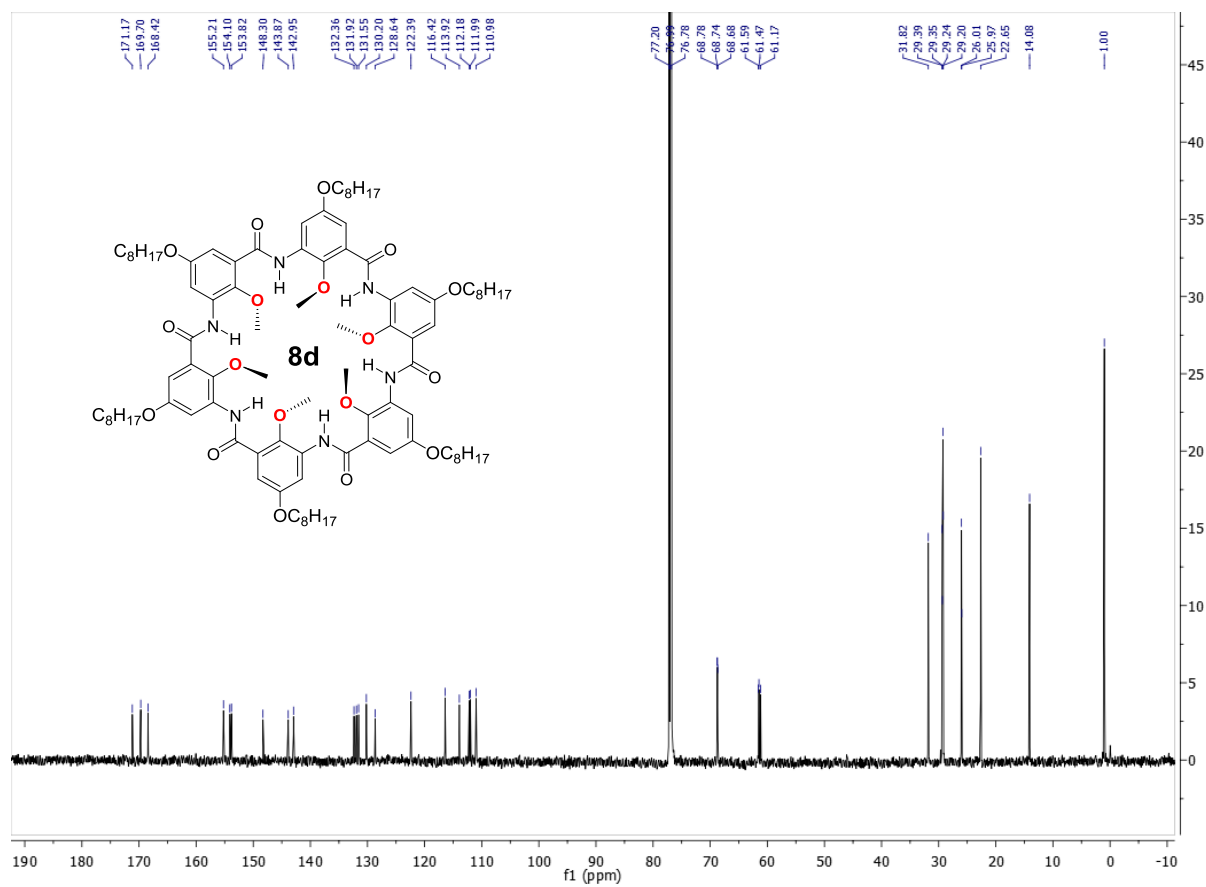
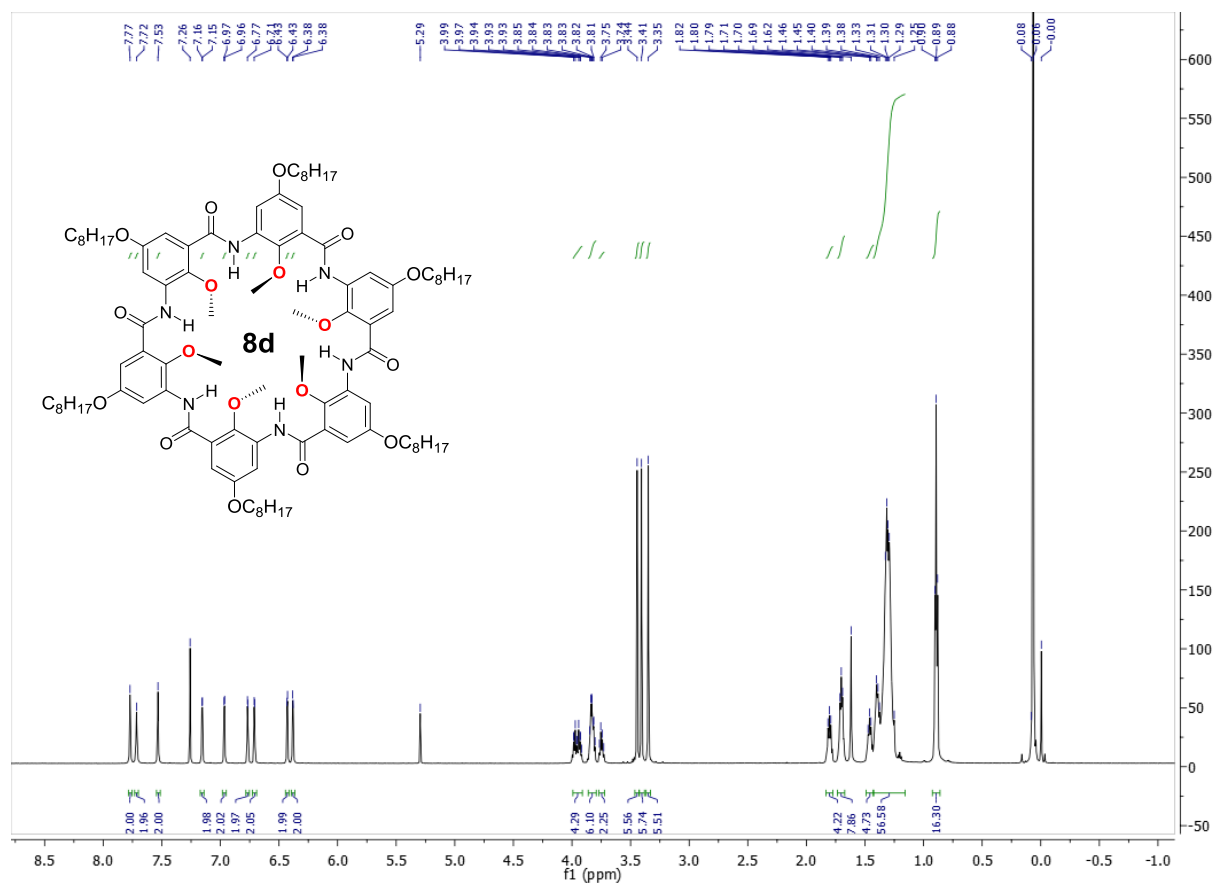
For ^1H NMR, ^{13}C NMR, and (HR)MS of hexamer **8a**, see Y. Liu, B. Qin and H. Q. Zeng, *Sci. China Chem.* 2012, **55**, 55.

Due to their highly distorted backbones as evidenced from the computationally determined structure of **8a** (Fig 2c of the text), the ^1H signals for the six aromatic units in both **8c** and **8d** split into three individual sets of signals. The same splitting is applicable to other proton signals from amide bonds and interior and exterior side chains as well as the ^{13}C NMR.

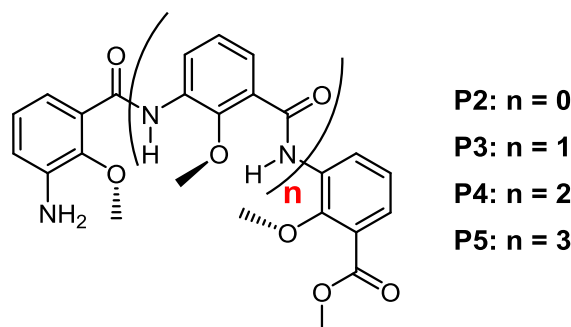
Hexamer 8c: ^1H NMR (600 MHz, CDCl_3) δ 7.50 (s, 2H), 7.34 (s, 2H), 7.21 – 7.15 (m, 4H), 7.00 (d, $J = 2.7$ Hz, 2H), 6.72 (d, $J = 2.6$ Hz, 2H), 6.69 (d, $J = 2.7$ Hz, 2H), 6.44 (d, $J = 2.6$ Hz, 2H), 6.36 (d, $J = 2.5$ Hz, 2H), 4.54 (dt, $J = 12.1, 6.0$ Hz, 3H), 4.40 (dt, $J = 11.9, 5.9$ Hz, 3H), 4.31 (dt, $J = 12.0, 5.9$ Hz, 3H), 3.47 (s, 5H), 3.45 (s, 5H), 3.36 (s, 6H), 1.57 (s, 11H), 1.36 (t, $J = 5.6$ Hz, 12H), 1.25 (td, $J = 12.0, 6.0$ Hz, 30H). ^{13}C NMR (151 MHz, CDCl_3) δ 170.77, 169.49, 168.27, 153.93, 152.96, 152.60, 148.24, 143.95, 143.06, 132.25, 131.97, 131.51, 130.57, 130.11, 128.57, 123.95, 117.99, 115.37, 114.26, 113.90, 112.38, 71.06, 70.87, 70.79, 61.43, 61.35, 61.08, 53.78, 53.39, 29.68, 29.26, 22.12, 21.95, 21.85, 21.65. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{66}\text{H}_{78}\text{N}_6\text{O}_{18}\text{K}$): m/z 1265.5270, found: m/z 1265.5075.

Hexamer 8d: ^1H NMR (600 MHz, CDCl_3) δ 7.77 (s, 2H), 7.72 (s, 2H), 7.53 (s, 2H), 7.16 (d, $J = 3.0$ Hz, 2H), 6.97 (d, $J = 2.9$ Hz, 2H), 6.77 (d, $J = 2.8$ Hz, 2H), 6.71 (d, $J = 2.9$ Hz, 2H), 6.43 (d, $J = 2.7$ Hz, 2H), 6.38 (d, $J = 2.7$ Hz, 2H), 3.95 (m, 4H), 3.83 (m, 6H), 3.75 (dd, $J = 15.4, 6.6$ Hz, 2H), 3.44 (s, 6H), 3.41 (s, 6H), 3.35 (s, 6H), 1.80 (m, 4H), 1.70 (m, 8H), 1.46 (m, 5H), 1.33 (m, 57H), 0.89 (t, $J = 6.7$ Hz, 16H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.17, 169.70, 168.42, 155.21, 154.10, 153.82, 148.30, 143.87, 142.95, 132.36, 131.92, 131.55, 130.20, 128.64, 122.39, 116.42, 113.92, 112.18, 111.99, 110.98, 68.78, 68.74, 68.68, 61.59, 61.47, 61.17, 31.82, 29.39, 29.35, 29.24, 29.20, 26.01, 25.97, 22.65, 14.08. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{96}\text{H}_{138}\text{N}_6\text{O}_{18}\text{K}$): m/z 1685.9965, found: m/z 1685.9752.





4. ^1H NMR, ^{13}C NMR, and (HR)MS of intermediate oligomers P2-P5



Dimer **P2**: ^1H NMR (300 MHz, CDCl_3) δ 10.62 (s, 1H), 8.86 (s, 1H), 8.84 (d, $J = 1.6$ Hz, 1H), 7.57 (ddd, $J = 7.5, 5.8, 1.6$ Hz, 2H), 7.20 (t, $J = 8.0$ Hz, 1H), 7.08 (t, $J = 7.8$ Hz, 1H), 6.95 (d, $J = 1.6$ Hz, 1H), 6.93 (d, $J = 1.6$ Hz, 1H), 3.95 (s, 6H), 3.91 (s, 3H). ^{13}C NMR (75.4 MHz, CDCl_3) δ 165.00, 162.56, 148.35, 144.18, 139.27, 132.51, 125.40, 124.75, 124.30, 123.72, 123.29, 122.47, 119.96, 118.83, 61.39, 59.79, 51.23. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_5\text{Na}$): m/z 353.1108, found: m/z 353.1125.

Trimer **P3**: ^1H NMR (300 MHz, CDCl_3) δ 10.54 (s, 1H), 10.38 (s, 1H), 8.88 – 8.81 (m, 2H), 7.89 (d, $J = 1.6$ Hz, 1H), 7.87 (d, $J = 1.6$ Hz, 1H), 7.60 (ddd, $J = 9.6, 7.9, 1.6$ Hz, 2H), 7.34 (t, $J = 8.0$ Hz, 1H), 7.23 (s, 1H), 7.11 (t, $J = 7.8$ Hz, 1H), 6.98 (d, $J = 1.6$ Hz, 1H), 6.95 (d, $J = 1.6$ Hz, 1H), 3.97 (s, 3H), 3.96 (s, 3H), 3.96 (s, 3H), 3.95 (s, 3H). ^{13}C NMR (75.4 MHz, CDCl_3) δ 164.87, 162.56, 162.03, 148.29, 146.27, 144.04, 139.25, 132.24, 131.74, 125.31, 125.29, 125.04, 124.91, 124.67, 124.50, 123.82, 123.68, 123.43, 122.44, 120.03, 119.03, 62.00, 61.42, 59.75, 51.28. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{25}\text{H}_{25}\text{N}_3\text{O}_7\text{Na}$): m/z 502.1585, found: m/z 502.1578.

Tetramer **P4**: ^1H NMR (300 MHz, CDCl_3) δ 10.44 (s, 1H), 10.23 (s, 1H), 10.20 (s, 1H), 8.86 (s, 1H), 8.84 (s, 1H), 7.94 – 7.86 (m, 2H), 7.65 (d, $J = 1.6$ Hz, 1H), 7.62 (d, $J = 1.6$ Hz, 1H), 7.59 (s, 1H), 7.56 (d, $J = 1.5$ Hz, 1H), 7.37 (t, $J = 8.0$ Hz, 2H), 7.23 (d, $J = 8.1$ Hz, 1H), 7.12 (t, $J = 7.8$ Hz, 1H), 6.99 (d, $J = 1.6$ Hz, 1H), 6.97 (d, $J = 1.6$ Hz, 1H), 4.01 (s, 3H), 3.99 (s, 3H), 3.96 (d, $J = 3.5$ Hz, 9H). ^{13}C NMR (75.4 MHz, CDCl_3) δ 164.83, 162.70, 162.12, 162.01, 148.36, 146.21, 144.03, 139.31, 132.15, 131.69, 131.46, 125.45, 125.36, 125.25, 125.19, 125.04, 124.82, 124.77, 124.55, 124.17, 123.92, 123.87, 123.50, 122.34, 119.98, 119.05, 62.06, 61.97, 61.44, 59.79, 51.31. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{33}\text{H}_{32}\text{N}_4\text{O}_9\text{Na}$): m/z 651.2062, found: m/z 651.2026.

Pentamer **P5**: ^1H NMR (300 MHz, CDCl_3) δ 10.41 (s, 1H), 10.17 (s, 1H), 10.04 (s, 1H), 10.03 (s, 1H), 8.85 (d, $J = 4.7$ Hz, 2H), 7.93 – 7.87 (m, 4H), 7.64 (d, $J = 1.6$ Hz, 1H), 7.62 (d, $J = 1.6$ Hz, 1H), 7.58 (d, $J = 1.6$ Hz, 1H), 7.56 (d, $J = 1.6$ Hz, 1H), 7.43 – 7.37 (m, 4H), 7.24 (d, $J = 8.1$ Hz, 1H), 7.12 (t, $J = 7.8$ Hz, 1H), 6.97 (dd, $J = 7.8, 1.6$ Hz, 1H), 4.04 (s, 6H), 3.99 (s, 3H), 3.98 (s, 3H), 3.94 (s, 3H), 3.89 (s, 3H). ^{13}C NMR (75.4 MHz, CDCl_3) δ 171.09, 165.77, 163.74, 163.32, 163.13, 162.99, 149.22, 147.25, 147.23, 147.18, 144.94, 140.35, 133.10, 132.72, 132.39, 132.34, 126.56, 126.47, 126.45, 126.28, 126.25, 126.16, 125.88, 125.87, 125.84, 125.77, 125.54, 125.15, 124.94, 124.78, 124.48, 123.47, 120.88, 120.01, 63.03, 62.42, 60.87, 60.36, 52.29, 21.02, 14.19. HRMS-EI: calculated for $[\text{M}+\text{Na}]^+$ ($\text{C}_{41}\text{H}_{39}\text{N}_5\text{O}_{11}\text{Na}$): m/z 800.2538, found: m/z 800.2582.

