

Supporting Information

Direct N^9 -arylation of purines with aryl halides

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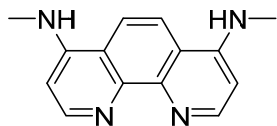
General Methods

All commercial solvents and reagents were used as obtained without further purification, except for CH_2Cl_2 which was distilled before use. Water was purified by a Milli-Q system and degassed by letting through nitrogen gas. All reactions were carried out under a nitrogen atmosphere. Semi-automatic flash chromatography was performed using Merck silica gel 60 (0.040-0.063 mm), on a Biotage HPFC SP4 Flash Purification System. ^1H and ^{13}C NMR spectra were recorded at 300 K on a Bruker Avance III 400 operating at 400 MHz and 101 MHz, respectively, with chemical shifts calibrated relative to solvent residual peaks (^1H NMR ($\text{DMSO}-d_6$): δ 2.50 ppm; ^1H NMR (CDCl_3): δ 7.26 ppm; ^{13}C NMR ($\text{DMSO}-d_6$): δ 39.52 ppm; ^{13}C NMR (CDCl_3): δ 77.16 ppm). Assignments are based on chemical shifts and/or COSY, HSQC and NOESY spectra. Mass spectra were recorded on a Bruker MicroTOF-Q II (ESI) spectrometer. Reactions were monitored by thin-layer chromatography (TLC) carried out on 0.25 mm E. Merck silica gel plates (60F-254) using UV-light and HPLC using a Dionex 120A C18 column (5μ , 4.6x150 mm) with gradient elution of MeCN and water, both solvents containing 0.05% trifluoroacetic acid (TFA), at a flow rate of 1 mL min^{-1} and UV detection at 230, 254 and 320 nm. HPLC purification was performed by using a Phenomenex Luna C18 5μ column (250x21.20 mm) with gradient elution of MeCN and water, both solvents containing 0.05% TFA, at a flow rate of 20 mL min^{-1} .

Synthesis of phenanthroline ligands

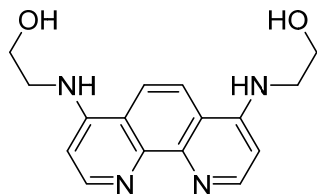
Ligands **L1**(DPPhen)¹ and **G**² were prepared as described previously.

*N*⁴,*N*⁷-Dimethyl-1,10-phenanthroline-4,7-diamine (Ligand **L2**)



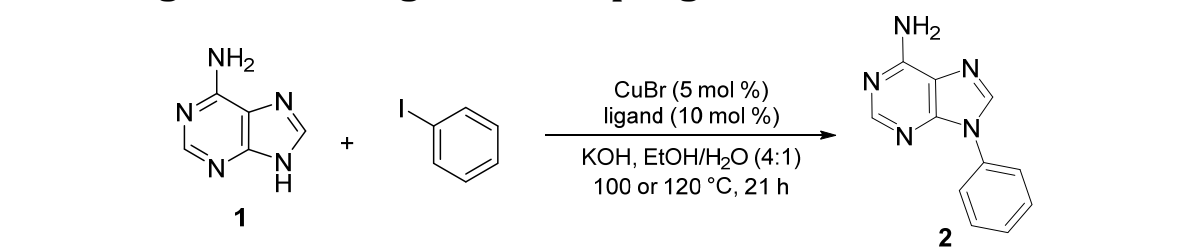
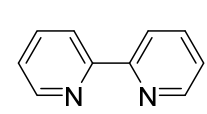
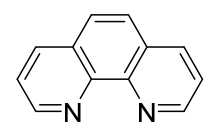
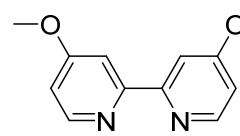
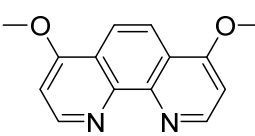
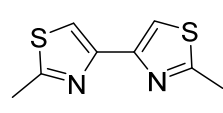
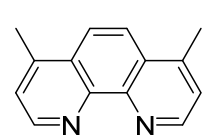
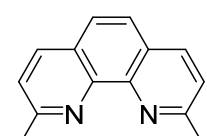
A sealed vial containing 4,7-dichloro-1,10-phenanthroline (300 mg, 1.2 mmol) and methylamine (5.0 mL, 33% in EtOH, 40 mmol) was heated in a microwave reactor (Emrys Creator) at 150 °C for 40 min to give an orange mixture with a precipitate. The precipitate was isolated by filtration, washed with water, ethanol and diethyl ether, and dried under vacuum to give the title compound as a pale yellow solid (255 mg, 89% yield). m.p. > 250 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.73 (s, 2H), 8.53 (d, J = 6.2 Hz, 2H), 8.36 (s, 2H), 6.84 (d, J = 6.3 Hz, 2H), 3.04 ppm (d, J = 4.4 Hz, 6H); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 153.6, 145.8, 137.3, 118.2, 117.0, 101.1, 29.7 ppm. HRMS-ESI: m/z calcd for $\text{C}_{14}\text{H}_{15}\text{N}_4^+$: 239.1291 [$M+\text{H}^+$], found: 239.1298.

*N*⁴,*N*⁷-Bis(2-hydroxyethyl)-1,10-phenanthroline-4,7-diamine (Ligand **L3**, BHPPhen)



A sealed vial containing 4,7-dichloro-1,10-phenanthroline (747 mg, 3.0 mmol), 2-aminoethanol (1.81 mL, 30.0 mmol) and anhydrous EtOH (15.0 mL) was heated in a microwave reactor (Biotage Initiator⁺ EU) at 140 °C for 3 hours to give a red mixture with a precipitate. The precipitate was isolated by filtration, washed sequentially with water, ethanol and diethyl ether, and dried under vacuum to give the title compound as a pale yellow solid (795 mg, 89% yield). m.p. > 250 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.48 (d, J = 5.3 Hz, 2H), 8.06 (s, 2H), 7.19–7.03 (m, 2H), 6.67 (d, J = 5.4 Hz, 2H), 4.91 (s, 2H), 3.70 (t, J = 5.7 Hz, 4H), 3.42–3.35 ppm (m, 4H); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 150.2, 149.4, 146.3, 117.4, 117.2, 100.7, 58.9, 45.3 ppm. HRMS-ESI: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{N}_4\text{O}_2^+$: 299.1503 [$M+\text{H}^+$], found: 299.1514.

Table S1: Ligand screening for the coupling of adenine and iodobenzene

				
<div style="display: flex; justify-content: space-around; align-items: flex-end;"> <div style="text-align: center;">  A </div> <div style="text-align: center;">  B </div> <div style="text-align: center;">  C </div> <div style="text-align: center;">  D </div> </div>				
<div style="display: flex; justify-content: space-around; align-items: flex-end;"> <div style="text-align: center;">  E </div> <div style="text-align: center;">  F </div> <div style="text-align: center;">  G </div> </div>				
entry	ligand	temp (°C)	conversion (%) ^b	yield (%) ^c
1	A	100	<1	ND
2	B	100	6	ND
3	C	100	2	ND
4	D	100	13	ND
5	E	100	<1	ND
6	L1	100	22	19
7	L2	100	31	20
8	F	120	32	30
9	G	120	<1	ND
10	L1	120	65	46
11	L2	120	ND	56

Reaction conditions: **1** (0.60 mmol), iodobenzene (0.50 mmol), ligand (10 mol %), CuBr (5 mol %), KOH (1.0 mmol), EtOH/H₂O (1.0 mL, 4:1), heated in a sealed vial to 100 °C or 120 °C for 18 h. ^b Determined from integration of the area under the signal of **2** (HPLC; UV absorbtion at 254 nm) as the percentage of the total area of the signals corresponding to **1** and **2**. ^c Yield of isolated product. ND: not determined.

Reaction optimisation

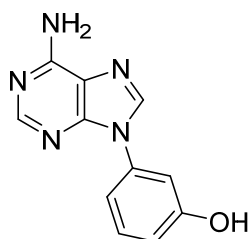
Entry 1: A reaction tube equipped with a magnetic stirring bar was charged with CuBr (3.6 mg, 0.025 mmol), ligand (**L1**, 15.9 mg, 0.05 mmol), adenine (**1**, 81 mg, 0.60 mmol) and KOH (56.1 mg, 1.0 mmol). The tube was sealed with a rubber septum, evacuated and back-filled with nitrogen three times. EtOH/H₂O (4:1 v/v, 1 mL) and iodobenzene (56 μ L, 0.50 mmol) was added by syringe. The rubber septum was exchanged with a cap designed to withstand moderate pressure under a stream of nitrogen and the reaction tube was placed in a preheated oil bath at 120 °C. After 21 hours, the reaction mixture was cooled to RT, silica (1.5 g) was added and the mixture was concentrated to dryness on a rotary evaporator. The dry residue was purified by semi-automated flash column chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% \rightarrow 5%)), to provide 49 mg (46%) of 9-phenyladenine (**2**).³

Entries 2-19 were performed by the same procedure, but with the modifications indicated in Table 1.

General procedure for *N*⁹-arylation of adenine (Table 2)

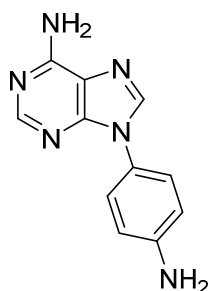
A reaction tube equipped with a magnetic stirring bar was charged with CuBr (3.6 mg, 0.025 mmol), ligand (**L3**, 14.9 mg, 0.05 mmol), sodium ascorbate (10 mg, 0.05 mmol), adenine (67.6 mg, 0.50 mmol), KOH (56.1 mg, 1.0 mmol) and aryl halide (if solid, 0.75 mmol). The reaction tube was sealed with a rubber septum, evacuated and back-filled with nitrogen three times. DMF/H₂O (4:1 v/v, 1 mL) and aryl halide (if liquid, 0.75 mmol) was added by syringe. The rubber septum was exchanged with a cap designed to withstand moderate pressure under a stream of nitrogen and the reaction tube was placed in a preheated oil bath at 120 °C. After 21 hours (for aryl bromides 48 hours), the reaction mixture was cooled to RT, silica (1.5 g) was added and the mixture was concentrated to dryness on a rotary evaporator. The dry residue was purified by semi-automated flash column chromatography. In some cases an inorganic iodide salt eluted with the product on the column, which was removed by recrystallisation from H₂O or MeOH. Known compounds exhibited ¹H and ¹³C NMR in agreement with previously reported data cited below and ESI-MS was in agreement with the predicted molecular mass.

3-(6-Aminopurin-9-yl)phenol (**3a**)



Following the general procedure using 3-iodophenol (165 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% \rightarrow 7%)), followed by a recrystallisation from water. Yield: 101 mg (89%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.89 (s, 1H), 8.54 (s, 1H), 8.22 (s, 1H), 7.45–7.29 (m, 4H), 7.29–7.24 (m, 1H), 6.88–6.78 ppm (m, 1H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 158.1, 156.3, 153.1, 149.1, 139.6, 136.1, 130.3, 119.4, 114.4, 113.2, 110.0 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₁₀N₅O⁺: 228.0880 [*M*+H⁺], found: 228.0884.

9-(4-Aminophenyl)purin-6-amine (3b)

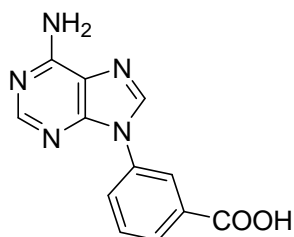


Using 4-iodoaniline: Following the general procedure using 4-iodoaniline (164 mg, 0.75 mmol) and heating for 21 hours provided the title compound as a pale tan solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (6% → 10%)), followed by a recrystallisation from water. Yield: 100 mg (88%).

Using 4-bromoaniline: Following the general procedure using 4-bromoaniline (129 mg, 0.75 mmol) and heating for 48 hours provided the title compound as a pale tan solid after semi-automated flash chromatography (Biotage SNAP 25 g, 10% MeOH in CH₂Cl₂). Yield: 86 mg (76%).

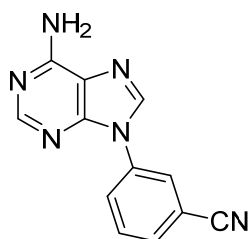
9-(4-Aminophenyl)purin-6-amine (3b): m.p. > 250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.32 (s, 1H), 8.14 (s, 1H), 7.38 (d, *J* = 8.6 Hz, 2H), 7.26 (s, 2H), 6.69 (d, *J* = 8.6 Hz, 2H), 5.36 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.1, 152.7, 149.3, 148.4, 140.0, 124.6, 123.5, 119.0, 113.8 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₁₁N₆⁺: 227.1040 [*M*+H⁺], found: 227.1042.

3-(6-Aminopurin-9-yl)benzoic acid (3c)



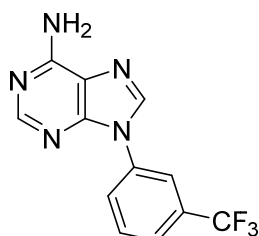
Following the general procedure using 3-iodobenzoic acid (186 mg, 0.75 mmol) provided the title compound as a tan solid after semi-automated flash chromatography (Biotage SNAP 25 g, MeOH/AcOH 2:1 in CH₂Cl₂ (6% → 12%)), followed by a recrystallisation from MeOH. Yield: 84 mg (66%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.32 (s, 1H), 8.68 (s, 1H), 8.51 (s, 1H), 8.32–8.20 (m, 1H), 8.19–8.11 (m, 1H), 8.07–7.94 (m, 1H), 7.81–7.68 (m, 1H), 7.42 ppm (s, 2H); HRMS-ESI: *m/z* calcd for C₁₂H₁₀N₅O₂⁺: 256.0829 [*M*+H⁺], found: 256.0831.

3-(6-Aminopurin-9-yl)benzonitrile (3d)



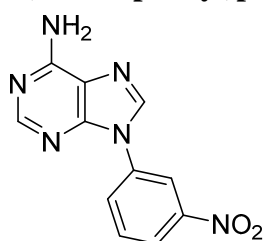
Following the general procedure using 3-iodobenzonitrile (172 mg, 0.75 mmol) provided the title compound as an off-white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 8%)), followed by a recrystallisation from H₂O. Yield: 91 mg (77%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.71 (s, 1H), 8.48 (t, *J* = 1.7 Hz, 1H), 8.40–8.32 (m, 1H), 8.25 (s, 1H), 7.95–7.87 (m, 1H), 7.81 (t, *J* = 8.0 Hz, 1H), 7.47 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.4, 153.4, 149.0, 139.3, 135.9, 130.8, 130.8, 127.2, 125.8, 119.3, 118.1, 112.3 ppm; HRMS-ESI: *m/z* calcd for C₁₂H₉N₆⁺: 237.0883 [*M*+H⁺], found: 237.0881.

9-(3-(Trifluoromethyl)phenyl)purin-6-amine (3e)



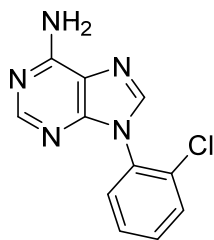
Following the general procedure using 1-iodo-3-(trifluoromethyl)benzene (204 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 6%)). Yield: 116.5 mg (83%). m.p. 226–228°C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.74 (s, 1H), 8.40 (s, 1H), 8.33–8.26 (m, 1H), 8.25 (s, 1H), 7.88–7.76 (m, 2H), 7.46 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.4, 153.4, 149.1, 139.5, 135.9, 130.8, 130.2 (q, *J* = 32.2 Hz), 126.4, 123.8 (q, *J* = 272.5 Hz), 123.8 (q, *J* = 3.6 Hz), 119.3, 119.2 ppm (q, *J* = 3.9 Hz); HRMS-ESI: *m/z* calcd for C₁₂H₉F₃N₅⁺: 280.0805 [*M*+H⁺], found: 280.0818.

9-(3-Nitrophenyl)purin-6-amine (3f)



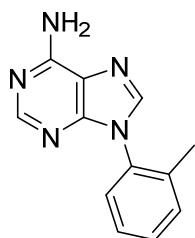
Following the general procedure using 1-iodo-3-nitrobenzene (187 mg, 0.75 mmol) provided the title compound as a yellow solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)), followed by a recrystallisation from water. Yield: 104 mg (81%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.96 (t, *J* = 2.2 Hz, 1H), 8.80 (s, 1H), 8.48–8.40 (m, 1H), 8.31–8.24 (m, 2H), 7.89 (t, *J* = 8.2 Hz, 1H), 7.49 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.4, 153.4, 149.1, 148.3, 139.4, 136.2, 131.0, 128.4, 121.7, 119.3, 117.1 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉N₆O₂⁺: 257.0781 [*M*+H⁺], found: 257.0794.

9-(2-Chlorophenyl)purin-6-amine (3g)



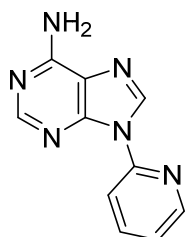
Following the general procedure using 1-chloro-2-iodobenzene (179 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)). Yield: 2.2 mg (2%). m.p. 233-234 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.33 (s, 1H), 8.11 (s, 1H), 7.79–7.73 (m, 1H), 7.69–7.64 (m, 1H), 7.64–7.53 (m, 2H), 7.39 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.2, 153.2, 150.2, 140.6, 132.1, 131.0, 130.7, 130.2, 130.1, 128.3, 118.2 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉ClN₅⁺: 246.0541 [*M*+H⁺], found: 246.0553.⁴

9-(*o*-Tolyl)purin-6-amine (3h)



Following the general procedure using 2-iodotoluene (164 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)). Yield: 7.5 mg (7%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.26 (s, 1H), 8.11 (s, 1H), 7.49–7.45 (m, 2H), 7.40–7.37 (m, 2H), 7.35 (s, 2H), 2.08 ppm (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.3, 153.0, 150.1, 140.8, 134.9, 133.7, 131.0, 129.2, 127.8, 126.8, 118.4, 17.5 ppm; HRMS-ESI: *m/z* calcd for C₁₂H₁₂N₅⁺: 226.1087 [*M*+H⁺], found: 226.1088.^{5*}

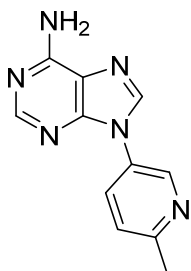
9-(Pyridin-2-yl)purin-6-amine (3i)



Following the general procedure using 2-bromopyridine (118 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 6%)). Yield: 87 mg (82%). m.p. 230-231 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.88 (s, 1H), 8.62–8.53 (m, 2H), 8.29 (s, 1H), 8.15–8.07 (m, 1H), 7.54–7.40 ppm (m, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.4, 153.3, 148.7, 148.7, 148.2, 139.5, 138.0, 122.7, 119.9, 115.1 ppm; HRMS-ESI: *m/z* calcd for C₁₀H₉N₆⁺: 213.0883 [*M*+H⁺], found: 213.0876.

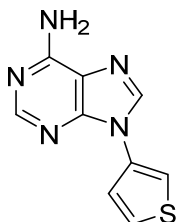
* ¹H NMR is not in agreement with that reported by Tao *et al.*⁵ No ¹³C NMR has been reported previously.

9-(6-Methylpyridin-3-yl)purin-6-amine (3j)



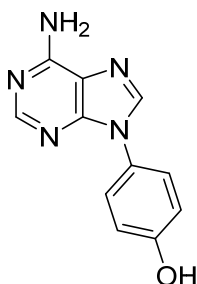
Following the general procedure using 5-bromo-2-methylpyridine (129 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 6%)). Yield: 92 mg (81%). m.p. 259-260 °C (dec.); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.95 (d, *J* = 2.4 Hz, 1H), 8.61 (s, 1H), 8.27 – 8.16 (m, 2H), 7.48 (d, *J* = 8.4 Hz, 1H), 7.43 (s, 2H), 2.55 ppm (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.0, 156.4, 153.3, 149.2, 143.1, 139.4, 130.9, 129.6, 123.4, 119.1, 23.7 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₁₁N₆⁺: 227.1040 [*M*+H⁺], found: 227.1029.

9-(Thiophen-3-yl)purin-6-amine (3k)



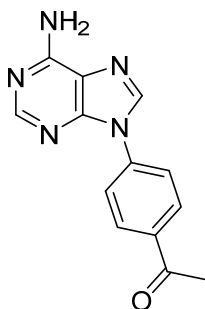
Following the general procedure using 3-bromothiophene (122 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 8%)). Yield: 83 mg (76%). m.p. 218-219 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.65 (s, 1H), 8.24 (s, 1H), 8.16 – 8.08 (m, 1H), 7.84 – 7.73 (m, 2H), 7.40 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.3, 153.3, 148.7, 139.3, 133.3, 127.1, 121.8, 119.0, 114.5 ppm; HRMS-ESI: *m/z* calcd for C₉H₈N₅S⁺: 218.0495 [*M*+H⁺], found: 218.0487.

4-(6-Aminopurin-9-yl)phenol (3l)



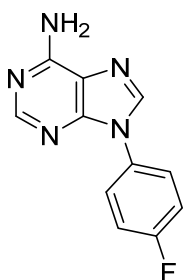
Following the general procedure using 4-bromophenol (130 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 25 g, 10% MeOH in CH₂Cl₂). Yield: 72 mg (63%). m.p. 303-305 °C (dec.); ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.79 (s, 1H), 8.41 (s, 1H), 8.17 (s, 1H), 7.59 (d, *J* = 8.8 Hz, 2H), 7.31 (s, 2H), 6.93 ppm (d, *J* = 8.8 Hz, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.8, 156.3, 153.0, 149.2, 139.9, 126.6, 124.9, 119.0, 115.8 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₁₀N₅O⁺: 228.0880 [*M*+H⁺], found: 228.0873.

1-(4-(6-Aminopurin-9-yl)phenyl)ethan-1-one (3m)



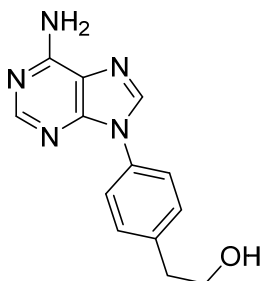
Following the general procedure using 4'-bromoacetophenone (149 mg, 0.75 mmol) provided the title compound as a yellow solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)). Yield: 98 mg (77%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.74 (s, 1H), 8.25 (s, 1H), 8.19–8.14 (m, 4H), 7.45 (s, 2H), 2.64 ppm (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 197.0, 156.4, 153.4, 149.1, 139.3, 139.0, 135.1, 129.6, 122.0, 119.5, 26.8 ppm; HRMS-ESI: *m/z* calcd for C₁₃H₁₂N₅O⁺: 254.1036 [*M*+H⁺], found: 254.1047.

9-(4-Fluorophenyl)purin-6-amine (3n)



Following the general procedure using 1-bromo-4-fluorobenzene (131 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)). Yield: 96 mg (84%). m.p. 255–256 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.56 (s, 1H), 8.20 (s, 1H), 7.96–7.90 (m, 2H), 7.47–7.42 (m, 2H), 7.39 ppm (s, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 160.9 (d, *J* = 244.4 Hz), 156.3, 153.2, 149.1, 139.7, 131.5 (d, *J* = 2.8 Hz), 125.2 (d, *J* = 8.6 Hz), 119.1, 116.3 ppm (d, *J* = 22.9 Hz); HRMS-ESI: *m/z* calcd for C₁₁H₉FN₅⁺: 230.0836 [*M*+H⁺], found: 230.0836.⁶

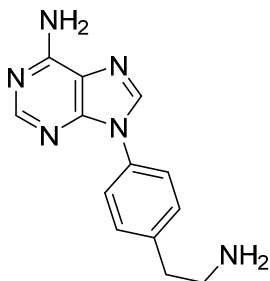
2-(4-(6-Aminopurin-9-yl)phenyl)ethan-1-ol (3o)



Following the general procedure using 4-bromophenethyl alcohol (151 mg, 0.75 mmol) provided the title compound as a white solid after semi-automated flash chromatography (Biotage SNAP 25 g, MeOH in CH₂Cl₂ (5% → 8%)), followed by recrystallisation from H₂O. Yield: 105 mg (82%). m.p. >250 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.53 (s, 1H), 8.20 (s, 1H), 7.80–7.73 (m, 2H), 7.44–7.40 (m, 2H), 7.37 (s, 2H), 4.70 (t, *J* = 5.2 Hz, 1H), 3.65 (td, *J* = 6.9, 5.3 Hz, 2H), 2.80 ppm (t, *J* = 6.9 Hz, 2H); ¹³C NMR (101 MHz, DMSO-

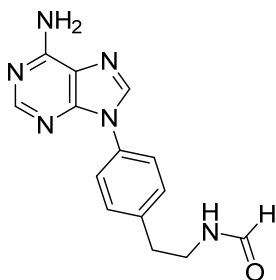
d_6) δ 156.3, 153.1, 149.2, 139.6, 139.1, 133.1, 129.9, 122.8, 119.2, 62.0, 38.5 ppm; HRMS-ESI: m/z calcd for $C_{13}H_{14}N_5O^+$: 256.1193 [$M+H^+$], found: 256.1204.

9-(4-(2-Aminoethyl)phenyl)purin-6-amine (3p)



Following the general procedure using 4-bromophenethyl amine (151 mg, 0.75 mmol) and EtOH/H₂O (1 mL, 4:1 v/v) as solvent provided the title compound as a pale yellow solid after semi-automated flash chromatography (Biotage SNAP 10 g, 24% NH_{3(aq)}/MeOH 1:9 in CH₂Cl₂ (5% → 8%)). Yield: 101 mg (79%). m.p. 219-220 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.53 (s, 1H), 8.19 (s, 1H), 7.82–7.74 (m, 2H), 7.42–7.38 (m, 2H), 7.37 (s, 2H), 2.81 (t, J = 6.9 Hz, 2H), 2.71 (t, J = 7.0 Hz, 2H), 1.47 ppm (s br, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.3, 153.01, 149.1, 139.9, 139.6, 133.0, 129.6, 122.9, 119.2, 43.6, 39.3 ppm (merged with DMSO); HRMS-ESI: m/z calcd for $C_{13}H_{15}N_6^+$: 255.1353 [$M+H^+$], found: 255.1355.

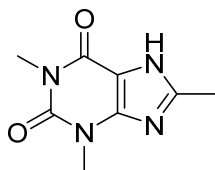
N-(4-(6-Aminopurin-9-yl)phenethyl)formamide



Following the general procedure using 4-bromophenethyl amine (151 mg, 0.75 mmol) and DMF/H₂O (1 mL, 4:1 v/v) as solvent provided *N*-(4-(6-amino-9*H*-purin-9-yl)phenethyl)formamide as a pale yellow solid after semi-automated flash chromatography (Biotage SNAP 10g, MeOH in CH₂Cl₂ (5% → 9%)). Yield: 112 mg (79%). m.p. 169-170 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.55 (s, 1H), 8.21 (s, 1H), 8.11 (s, 1H), 8.02 (d, J = 1.6 Hz, 1H), 7.81 (d, J = 8.5 Hz, 2H), 7.46–7.40 (m, 2H), 7.37 (s, 2H), 3.42–3.38 (m, 2H), 2.82 ppm (t, J = 7.1 Hz, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 161.1, 156.3, 153.1, 149.1, 139.6, 138.7, 133.3, 129.7, 122.9, 119.3, 38.6, 34.5 ppm. HRMS-ESI: m/z calcd for $C_{14}H_{15}N_6O^+$: 283.1302 [$M+H^+$], found: 283.1295.

Synthetic procedure for *N*-arylation of various purines (Table 3)

Preparation of 8-methyltheophylline (**4d**)

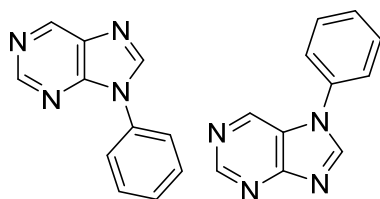


A sealed vial containing 5,6-diamino-1,3-dimethyluracil hydrate (565 mg, 3.0 mmol), acetic anhydride (0.31 mL, 3.3 mmol) and pyridine (6 mL) was heated in a microwave reactor (Emrys Creator) at 110 °C for 20 min to give a red mixture with a precipitate. The solvent was evaporated under vacuum to dryness. The resulting amide was converted to benzimidazole by treatment with *t*-BuOK (1.01 g, 9.0 mmol) in isopropyl alcohol (14 mL) at 60 °C. After 16 hours, the reaction mixture was cooled to RT, silica (5 g) was added and the mixture was concentrated to dryness on a rotary evaporator. The dry residue was purified by semi-automated flash column chromatography (Biotage SNAP 25 g, MeOH in CH₂Cl₂ (3% → 4.5%)), to provide 382 mg (66%) of 8-methyltheophylline (**4d**). m.p. 322-324 °C (lit. 325 °C⁷); ¹H NMR (400 MHz, DMSO) δ 3.39 (s, 3H), 3.21 (s, 3H), 2.36 ppm (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 153.9, 151.2, 150.4, 148.1, 106.1, 29.7, 27.6, 14.3 ppm. HRMS-ESI: *m/z* calcd for C₁₄H₁₅N₆O⁺: 283.1302 [*M*+H⁺], found: 283.1295.

General procedure A - purine, 2,6-diaminopurine, theophylline and 8-methyltheophylline (4a-d): A reaction tube equipped with a magnetic stirring bar was charged with CuBr (3.6 mg, 0.025 mmol), ligand (**L3**, 14.9 mg, 0.05 mmol), sodium ascorbate (10 mg, 0.05 mmol), the purine substrate (**4a**, **4b**, **4c** or **4d**, 0.50 mmol) and KOH (56.1 mg, 1.0 mmol). The tube was sealed with a rubber septum, evacuated and back-filled with nitrogen three times. Then DMF/H₂O (4:1 v/v, 1 mL) and aryl halide (0.75 mmol) was added by syringe. The rubber septum was exchanged with a cap designed to withstand moderate pressure under a stream of nitrogen and the reaction tube was placed in a preheated oil bath at 120 °C. After 21 hours or 48 hours, the reaction mixture was cooled to RT, silica (1.5 g) was added and the mixture was concentrated to dryness on a rotary evaporator. The dry residue was purified by semi-automated flash column chromatography.

General procedure B - guanine and hypoxanthine (4e and 4f): A reaction tube equipped with a magnetic stirring bar was charged with CuBr (3.6 mg, 0.025 mmol), ligand (**L3**, 14.9 mg, 0.05 mmol), the purine substrate (**4e** or **4f**, 0.50 mmol) and KOH (56.1 mg, 1.0 mmol). The tube was sealed with a rubber septum, evacuated and back-filled with nitrogen three times. Then DMSO/H₂O (4:1 v/v, 1 mL) and aryl halide (0.75 mmol) was added by syringe. The rubber septum was exchanged with a cap designed to withstand moderate pressure under a stream of nitrogen and the reaction tube was placed in a preheated oil bath at 120 °C. After 6-48 hours, the reaction mixture was cooled to RT, silica (1.5 g) was added and the mixture concentrated to dryness on a rotary evaporator at 60 °C. The dry residue was purified by semi-automated flash column chromatography.

9-Phenylpurine (5a) and 7-phenylpurine(6a)



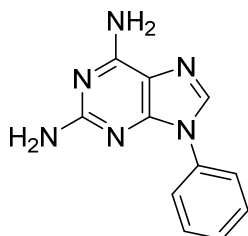
Using iodobenzene: Synthesised by general procedure A using iodobenzene (153 mg, 0.75 mmol) and heating for 21 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (2% → 3%)) gave 61 mg (62%) of 9-phenylpurine (**5a**) and 7 mg (7%) of 7-phenylpurine (**6a**), both products as pale brown solids.

Using bromobenzene: Synthesised by general procedure A using bromobenzene (118 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (2% → 3%)) gave 63 mg (64%) of 9-phenylpurine (**5a**) and 3 mg (3%) of 7-phenylpurine (**6a**), both products as pale brown solids.

9-phenylpurine (5a): ¹H NMR (400 MHz, CDCl₃) δ 9.24 (s, 1H), 9.05 (s, 1H), 8.38 (s, 1H), 7.78–7.67 (m, 2H), 7.66–7.55 (m, 2H), 7.53–7.44 ppm (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 153.5, 151.2, 149.4, 144.3, 134.7, 134.3, 130.2, 128.7, 123.6 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉N₄⁺: 197.0822 [*M*+H⁺], found: 197.0823.^{8†}

7-phenylpurine (6a): ¹H NMR (400 MHz, CDCl₃) δ 9.21 (s, 1H), 9.06 (s, 1H), 8.47 (s, 1H), 7.68–7.62 (m, 2H), 7.59–7.51 ppm (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 161.3, 154.0, 146.9, 140.8, 135.0, 130.8, 129.4, 125.3, 123.7 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉N₄⁺: 197.0822 [*M*+H⁺], found: 197.0826.^{9‡}

9-Phenylpurine-2,6-diamine (5b)



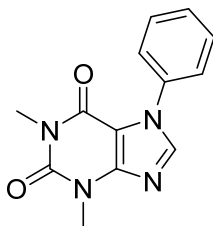
Using iodobenzene: Synthesised by general procedure A using iodobenzene (153 mg, 0.75 mmol) and heating for 21 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 5%)) gave 106 mg (94%) of the title compound as a white solid.¹

Using bromobenzene: Synthesised by general procedure A using bromobenzene (118 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (4% → 5%)) gave 68 mg (60%) of the title compound as a white solid.

[†] In accordance with the ¹H-NMR reported by Hayashi *et al.* No ¹³C-NMR has been reported previously.

[‡] ¹H-NMR reported by Laufer *et al.* used DMSO-*d*₆ as solvent. No ¹³C-NMR has been reported previously.

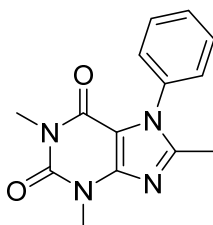
7-Phenyltheophylline (6c)



Using iodobenzene: Synthesised by general procedure A using iodobenzene (153 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, EtOAc in PE (50% → 100%)) gave 101 mg (79%) of the title compound as a white solid.¹⁰

Using bromobenzene: Synthesised by general procedure A using bromobenzene (118 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, EtOAc in PE (50% → 100%)) gave 100 mg (78%) of the title compound as a white solid.

8-methyl-7-phenyltheophylline (6d)

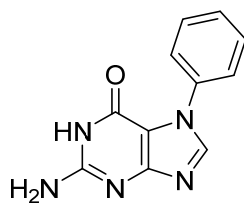
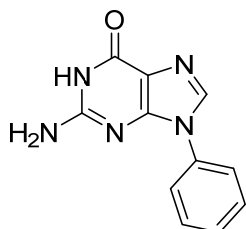


Using iodobenzene: Synthesised by general procedure A using iodobenzene (153 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, EtOAc in PE (60% → 100%)) gave 16 mg (12%) of the title compound as a white solid.

Using bromobenzene: Synthesised by general procedure A using bromobenzene (118 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, EtOAc in PE (60% → 100%)) gave 13 mg (10%) of the title compound as a white solid.

m.p. 230-231 °C (lit. 235 °C¹¹); ¹H NMR (400 MHz, CDCl₃) δ 7.57 – 7.49 (m, 3H), 7.38 – 7.28 (m, 2H), 3.63 (s, 3H), 3.34 (s, 3H), 2.36 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 154.3, 151.8, 151.2, 148.6, 135.0, 129.6, 129.5, 127.1, 108.3, 29.9, 28.1, 13.9 ppm; HRMS-ESI: *m/z* calcd for C₁₄H₁₅N₄O₂⁺: 271.1190 [*M*+H⁺], found: 271.1184.

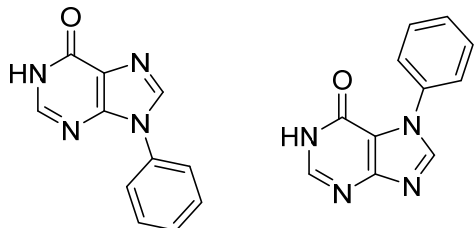
9-Phenylguanine (5e) and 7-phenylguanine (6e)



Using iodobenzene: Synthesised by general procedure B using iodobenzene (153 mg, 0.75 mmol) and heating for 21 hours. Purification by semi-automated flash chromatography (Biotage SNAP 25 g, MeOH in CH₂Cl₂ (4% → 8%)) gave a mixture of two isomers: 9-phenylguanine (**5e**) and 7-phenylguanine (**6e**) that were separated on HPLC (5-22% MeCN in H₂O (0-5 min), *R*_t(**6e**) = 3.9 min, *R*_t(**5e**) = 4.7 min) to give 68 mg (60%) of **5e** as a white solid and 24 mg (21%) of **6e** as a pale yellow solid.^{3,12}

Using bromobenzene: Synthesised by general procedure B using bromobenzene (118 mg, 0.75 mmol) and heating for 48 hours. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (5% → 10%)) gave a mixture of two isomers: 9-phenylguanine (**5e**) and 7-phenylguanine (**6e**) that were separated on HPLC (5-22% MeCN in H₂O (0-5 min), *R_t*(**6e**) = 3.9 min, *R_t*(**5e**) = 4.7 min) to give 67 mg (59%) of **5e** as a white solid and 15 mg (13%) of **6e** as a pale yellow solid.

9-Phenylhypoxanthine (**5f**) and 7-phenylhypoxanthine (**6f**)



Using iodobenzene: Synthesised by general procedure B using iodobenzene (153 mg, 0.75 mmol) and heating for 3 hours, after which the starting material was almost consumed, and diarylated products had started to form. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)) gave a mixture of two isomers: 9-phenylhypoxanthine (**5f**) and 7-phenylhypoxanthine (**6f**) that were separated on HPLC (10-21% MeCN in H₂O (0-6 min), *R_t*(**6f**) = 4.4 min, *R_t*(**5f**) = 5.3 min) to give 46 mg (43%) of **5f** and 15 mg (14%) of **6f**, both as white solids.

Using bromobenzene: Synthesised by general procedure B using bromobenzene (118 mg, 0.75 mmol) and heating for 32 hours, after which the starting material was almost consumed, and diarylated products had started to form. Purification by semi-automated flash chromatography (Biotage SNAP 10 g, MeOH in CH₂Cl₂ (3% → 5%)) gave a mixture of two isomers: 9-phenylhypoxanthine (**5f**) and 7-phenylhypoxanthine (**6f**) that were separated on HPLC (10-21% MeCN in H₂O (0-6 min), *R_t*(**6f**) = 4.4 min, *R_t*(**5f**) = 5.3 min) to give 51 mg (48%) of **5f** and 12 mg (11%) of **6f**, both as white solids.

9-phenylhypoxanthine (5f): m.p. 317-318 °C (lit. 316-318 °C¹³); ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.47 (s, 1H), 8.47 (s, 1H), 8.09 (s, 1H), 7.83–7.69 (m, 2H), 7.67–7.55 (m, 2H), 7.55–7.44 ppm (m, 1H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.7, 147.9, 146.1, 139.4, 134.5, 129.5, 128.1, 124.8, 123.8 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉N₄O⁺: 213.0771 [*M*+H⁺], found: 213.0761.¹³

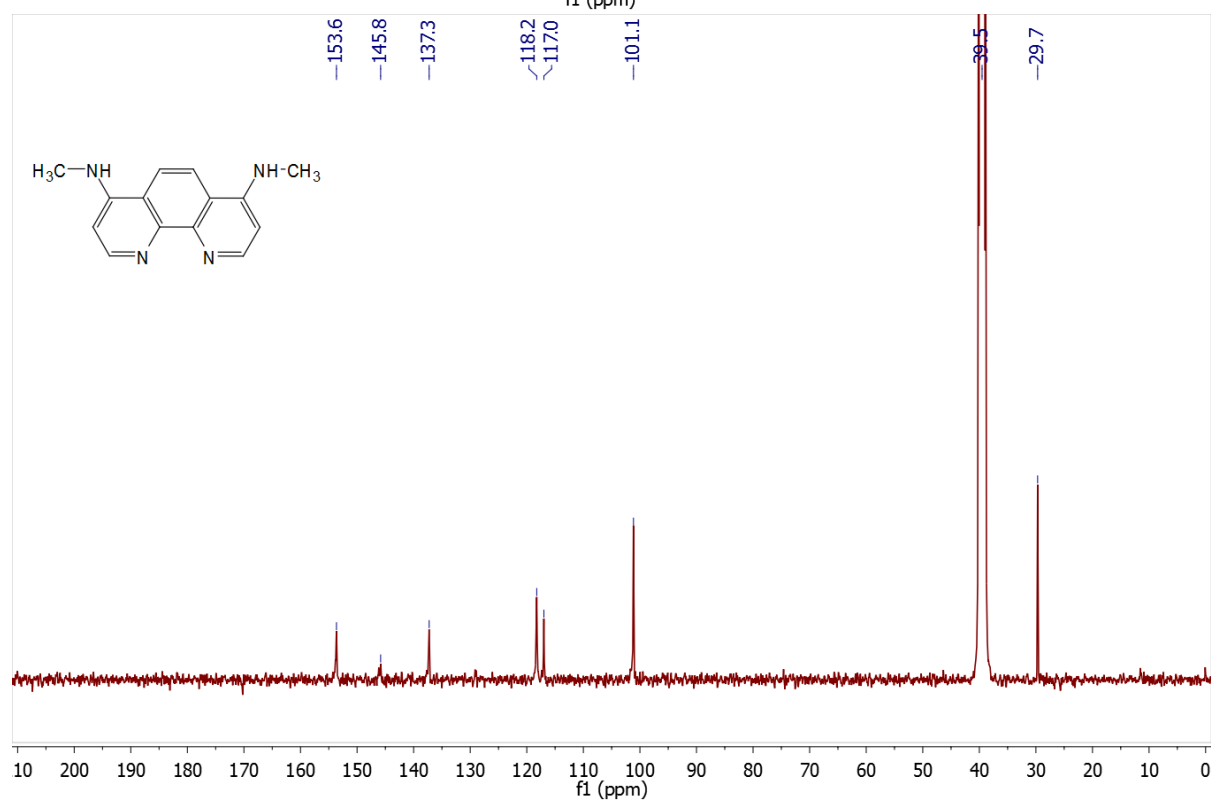
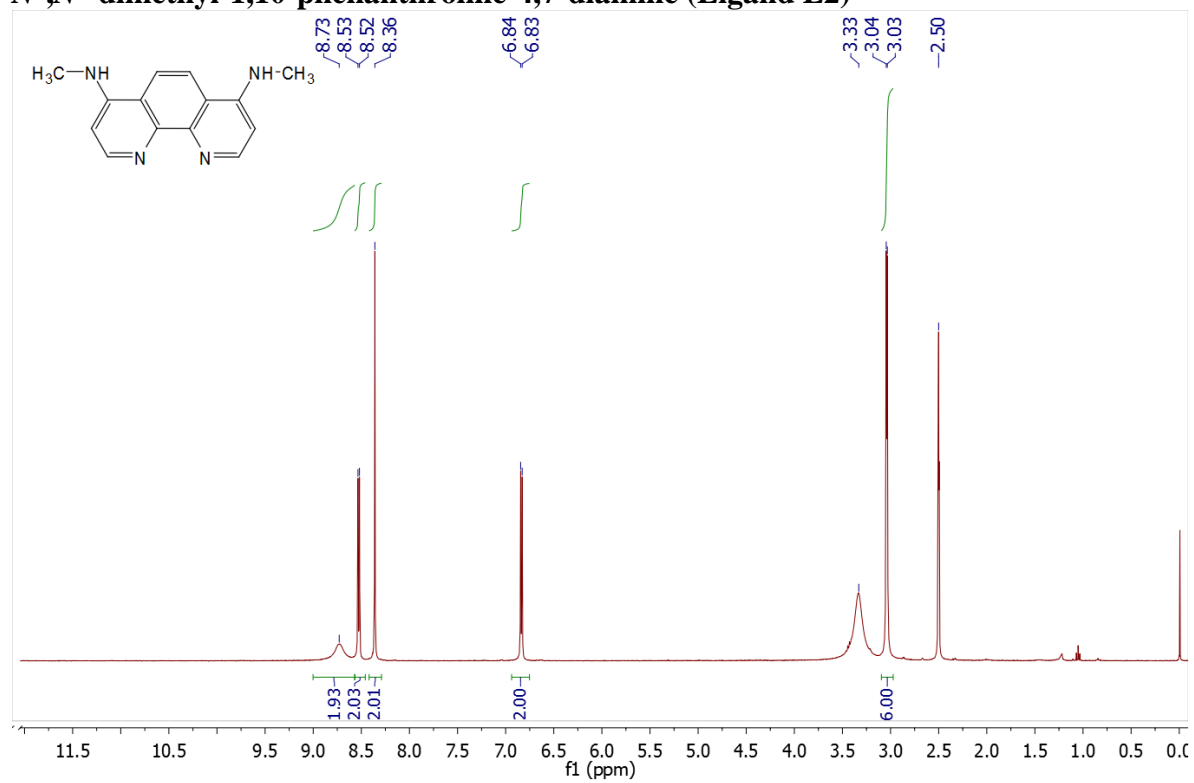
7-phenylhypoxanthine (6f): m.p. 295-296 °C (lit. 295-297 °C¹⁴); ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.43 (s, 1H), 8.51 (s, 1H), 8.06 (s, 1H), 7.68–7.59 (m, 2H), 7.57–7.51 (m, 2H), 7.51–7.44 ppm (m, 1H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 158.2, 153.6, 145.2, 144.0, 135.4, 128.9, 128.2, 125.2, 114.6 ppm; HRMS-ESI: *m/z* calcd for C₁₁H₉N₄⁺: 213.0771 [*M*+H⁺], found: 213.0761.¹⁴

References

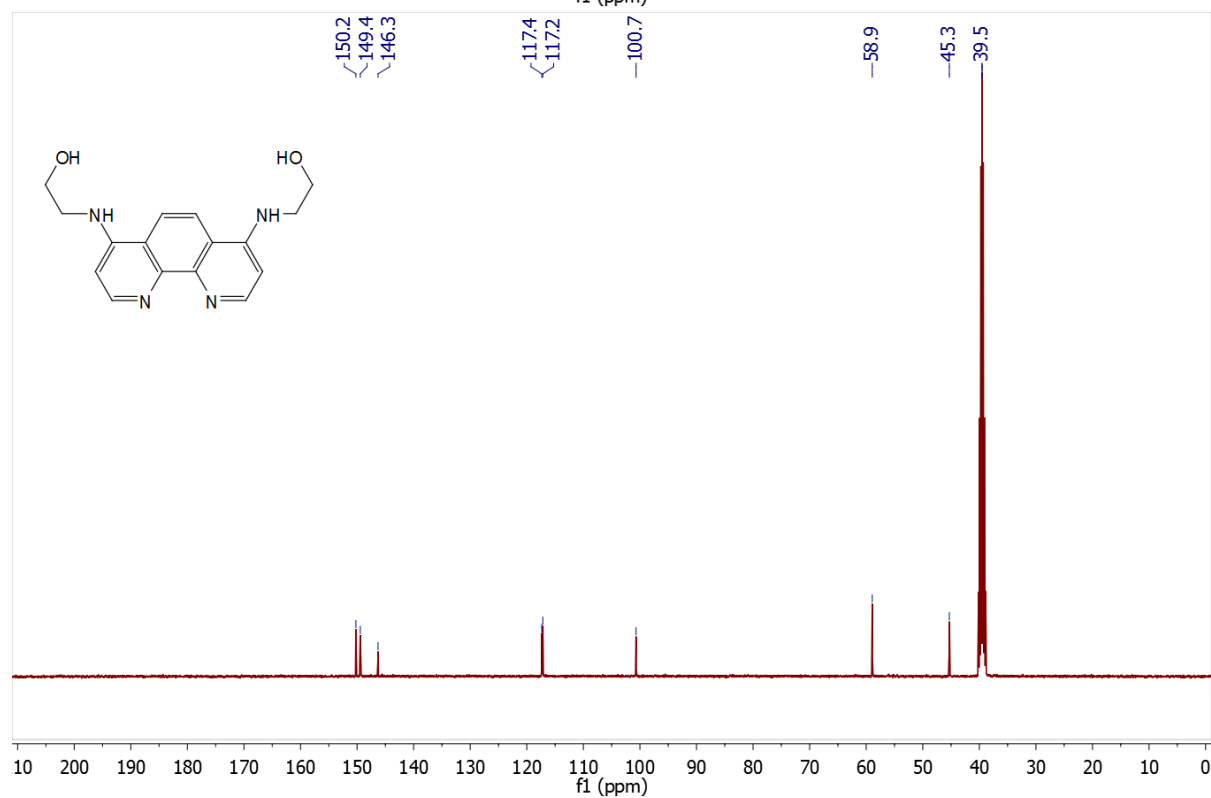
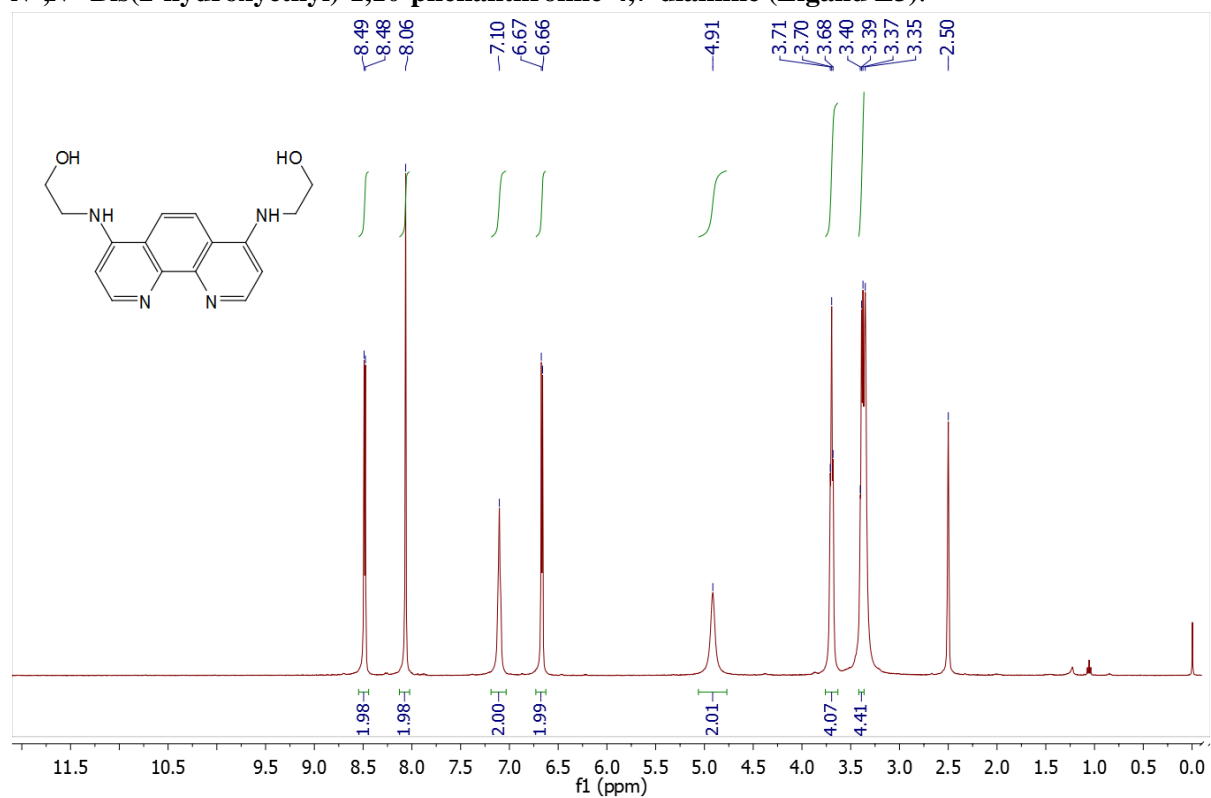
- (1) Engel-Andreasen, J.; Shimpukade, B.; Ulven, T. *Green Chem.* **2013**, *15*, 336.
- (2) Hashemi, M. M.; Asadollahi, H.; Mostaghim, R. *Russ. J. Org. Chem.* **2005**, *41*, 623.
- (3) Keder, R.; Dvořáková, H.; Dvořák, D. *Eur. J. Org. Chem.* **2009**, 2009, 1522.
- (4) Koppel, H. C.; O'Brien, D. E.; Robins, R. K. *J. Am. Chem. Soc.* **1959**, *81*, 3046.
- (5) Tao, L.; Yue, Y.; Zhang, J.; Chen, S.-Y.; Yu, X.-Q. *Helv. Chim. Acta* **2008**, *91*, 1008.
- (6) Yahyazadeh, A.; Habibi, F. *E-J. Chem.* **2007**, *4*, 372.
- (7) Speer, J. H.; Raymond, A. L. *J. Am. Chem. Soc.* **1953**, *75*, 114.
- (8) Hayashi, E.; Shimada, N.; Matsuoka, Y. *Yakugaku Zasshi* **1979**, *99*, 114.
- (9) Laufer, S. A.; Domeyer, D. M.; Scior, T. R. F.; Albrecht, W.; Hauser, D. R. *J. Med. Chem.* **2005**, *48*, 710.
- (10) Kim, D.; Lee, H.; Jun, H.; Hong, S.-S.; Hong, S. *Bioorg. Med. Chem.* **2011**, *19*, 2508.
- (11) Taylor, E. C.; Yoneda, F. *J. Org. Chem.* **1972**, *37*, 4464.
- (12) Jacobsen, M. F.; Knudsen, M. M.; Gothelf, K. V. *J. Org. Chem.* **2006**, *71*, 9183.
- (13) El-Bayouki, K. A. M.; El-Sayed, A. S.; Basyouni, W. M. *Gazz. Chim. Ital.* **1989**, *119*, 163.
- (14) Gewald, K.; Heinhold, G. *Monatsh. Chem.* **1976**, *107*, 1413.

NMR-spectra

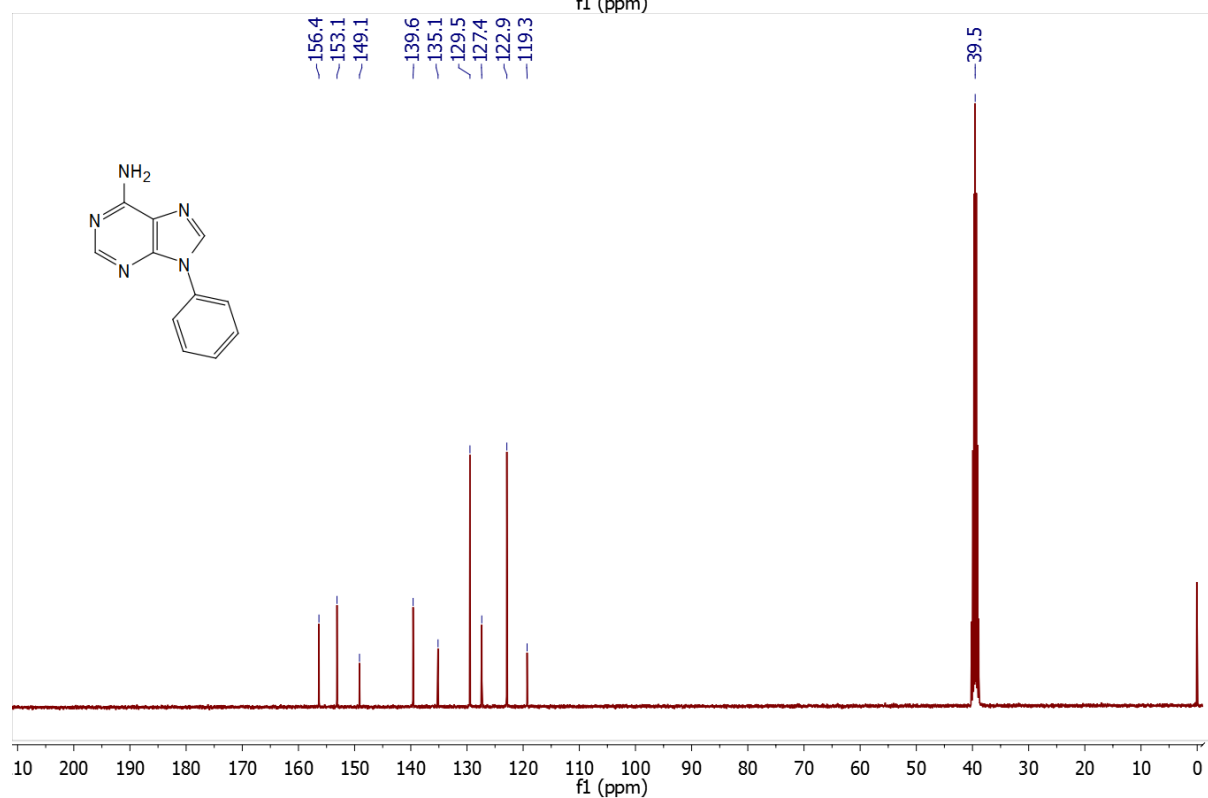
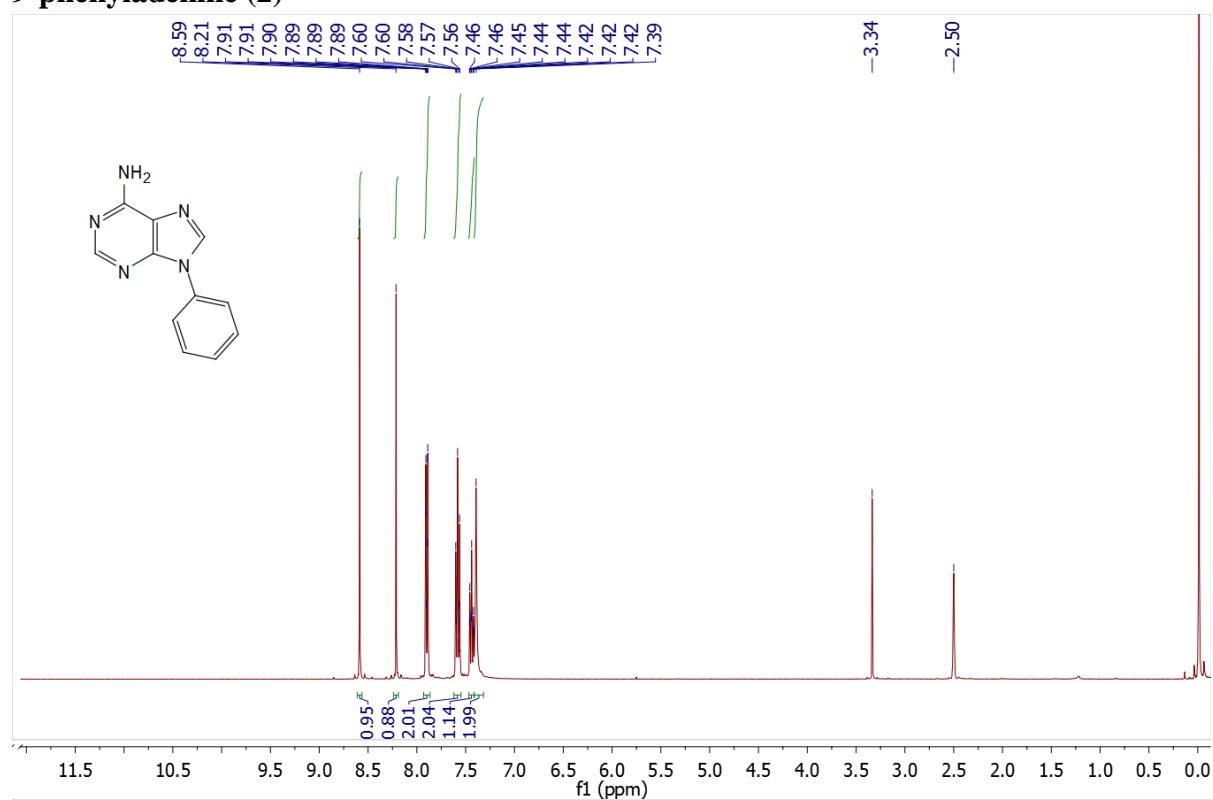
N^4,N^7 -dimethyl-1,10-phenanthroline (Ligand L2)



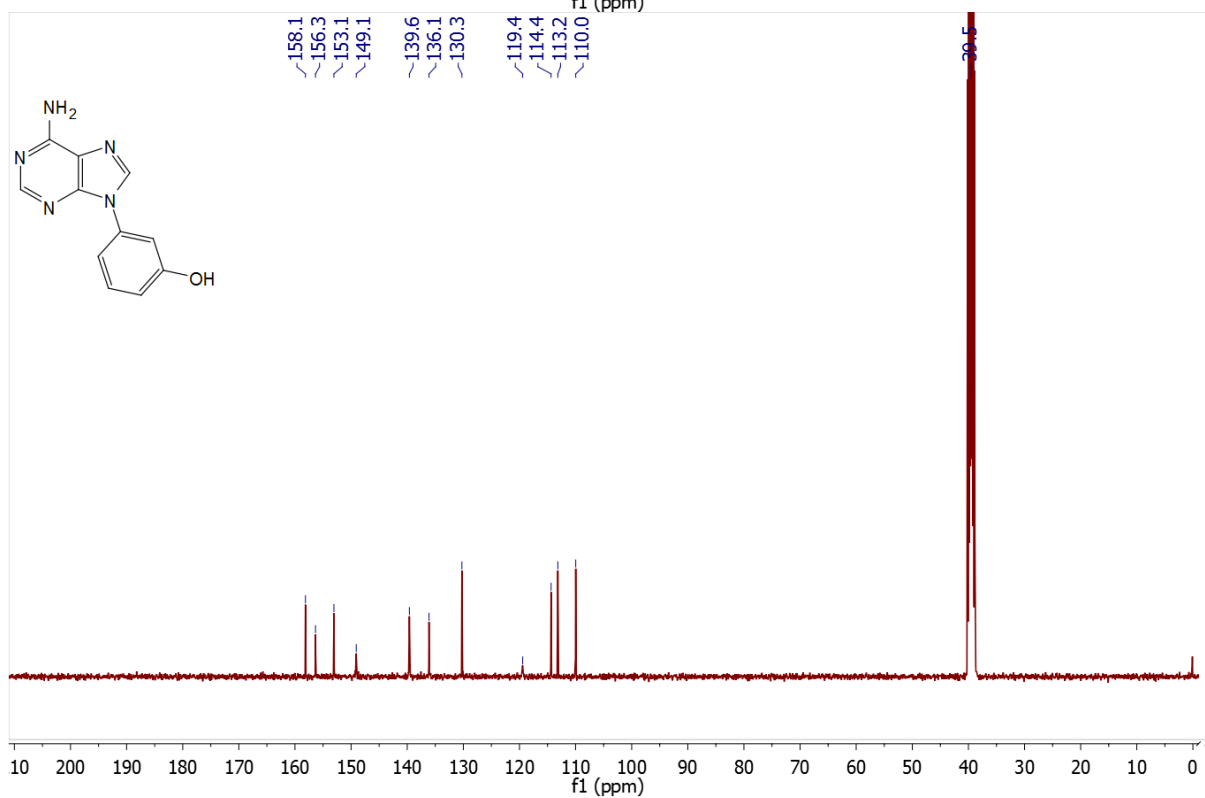
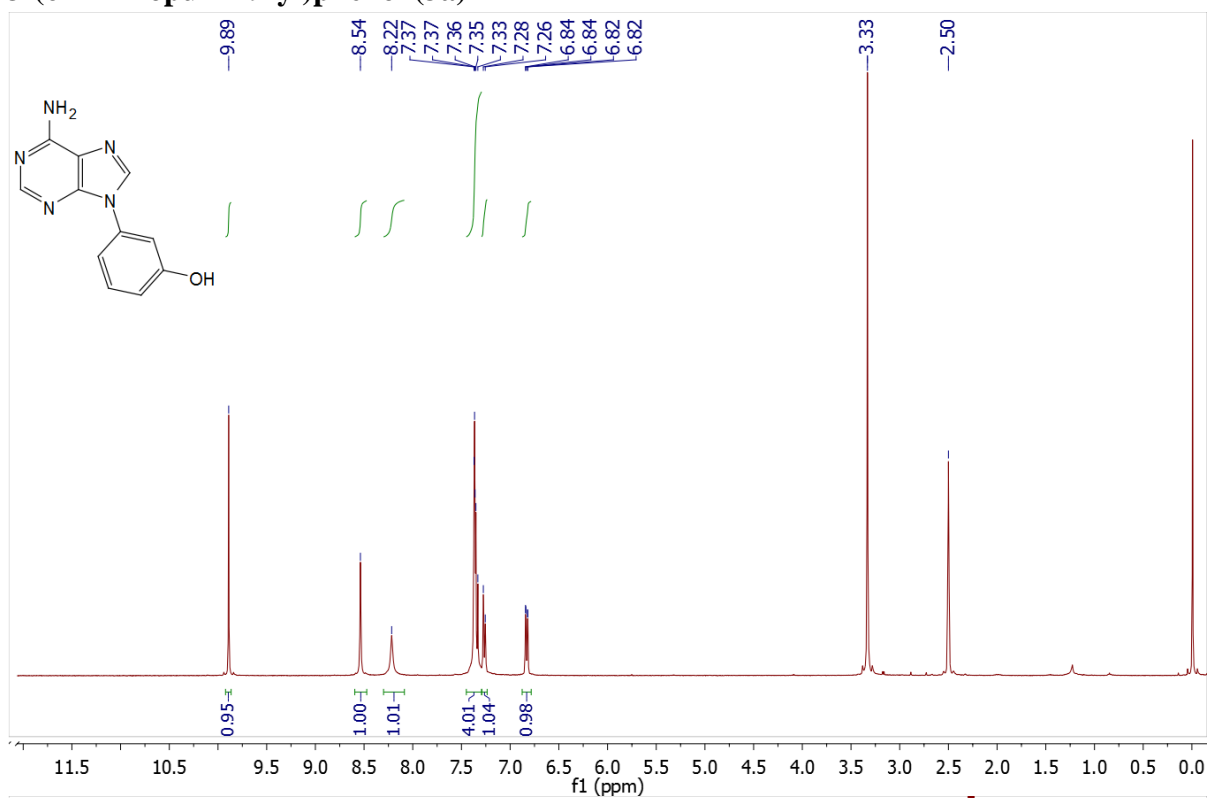
***N*⁴,*N*⁷-Bis(2-hydroxyethyl)-1,10-phenanthroline-4,7-diamine (Ligand L3).**



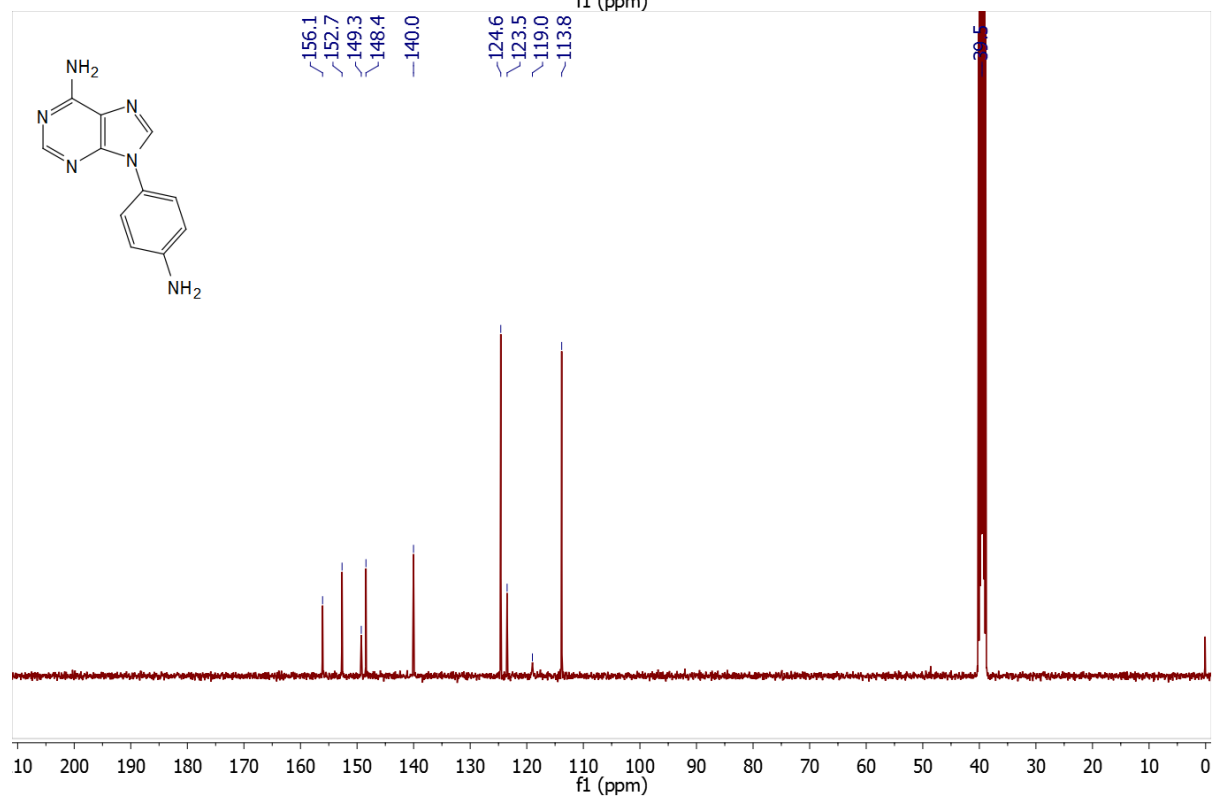
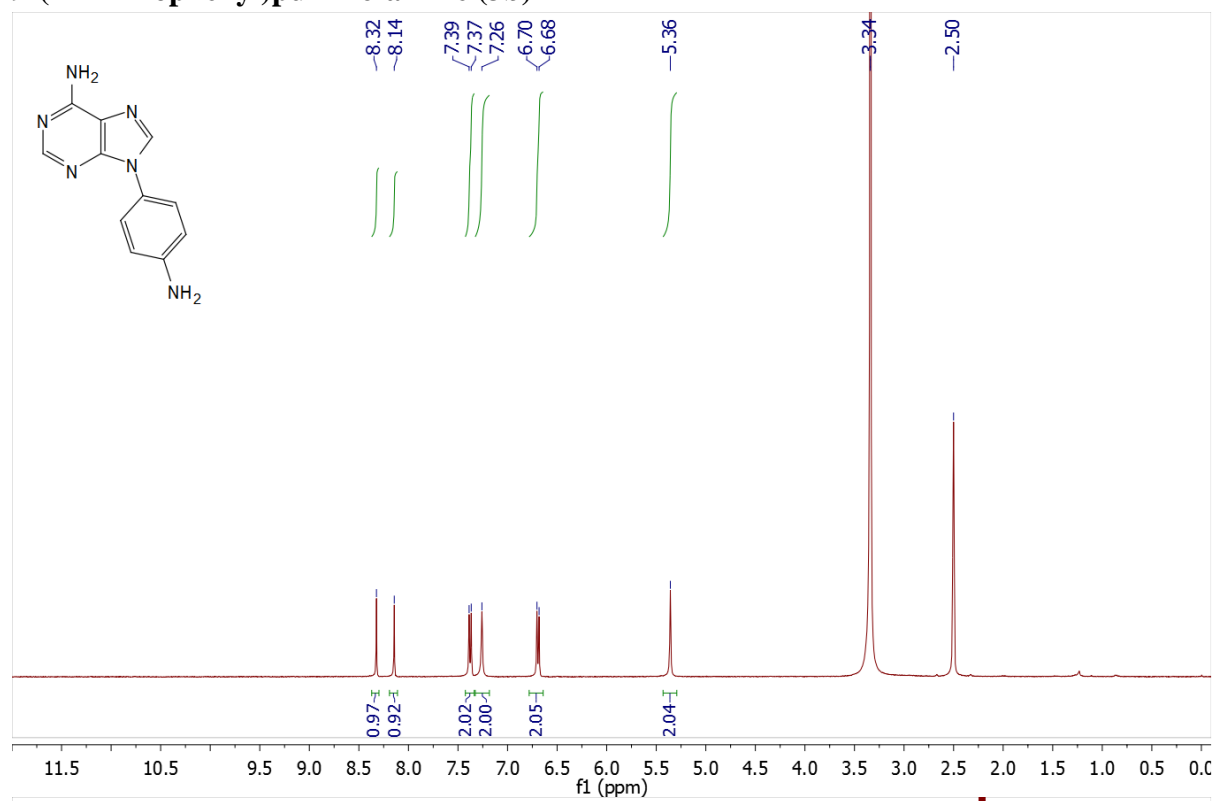
9-phenyladenine (2)



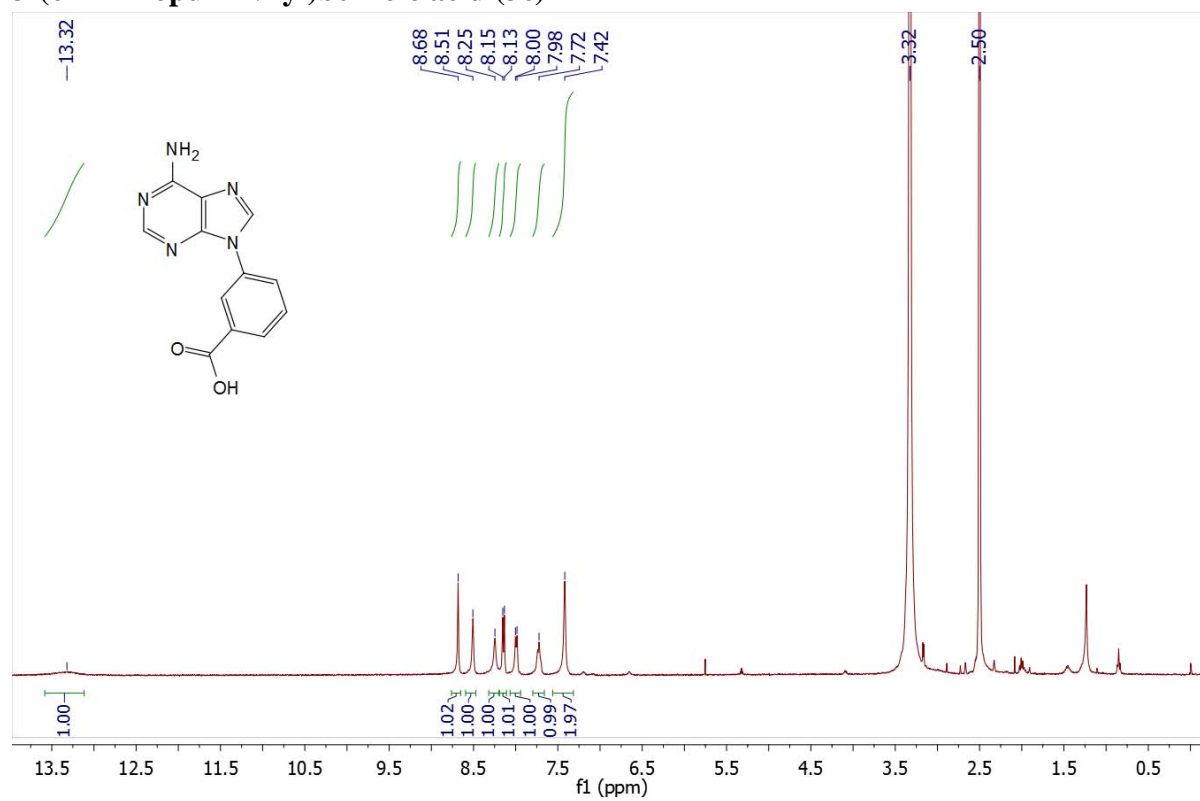
3-(6-Aminopurin-9-yl)phenol (3a)



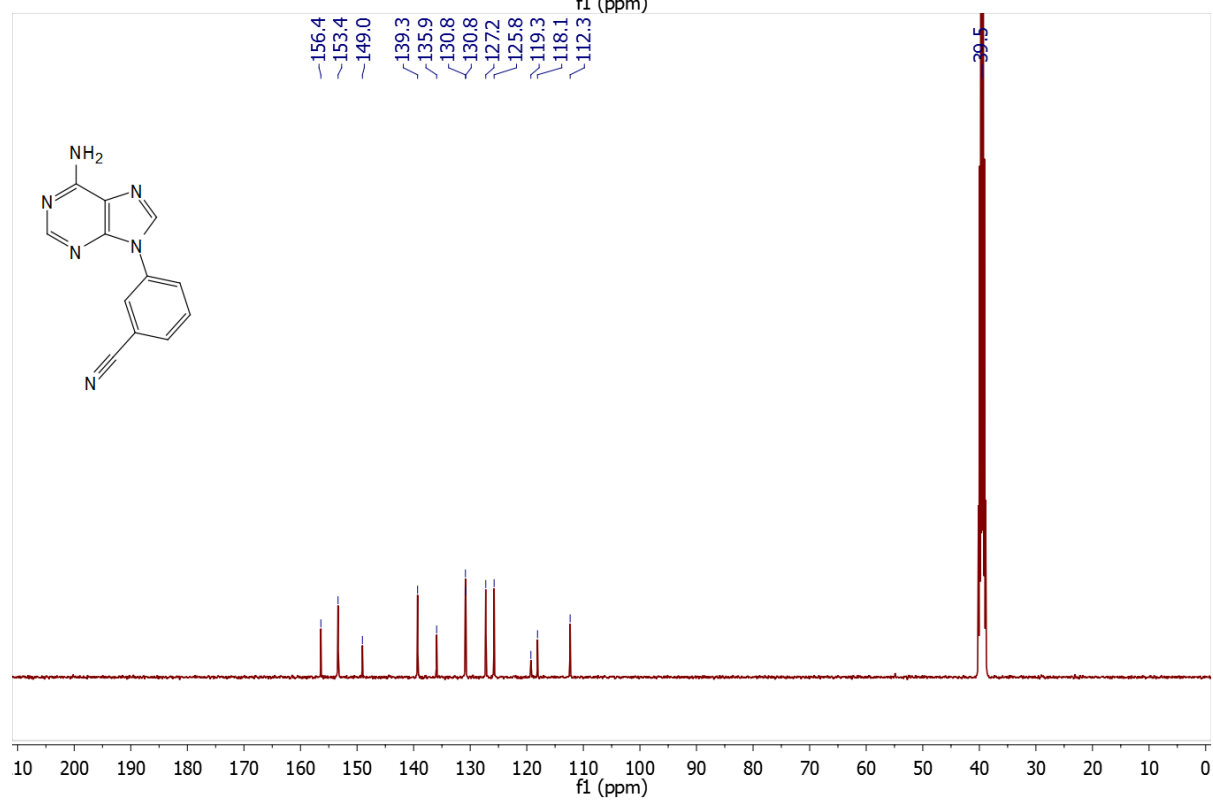
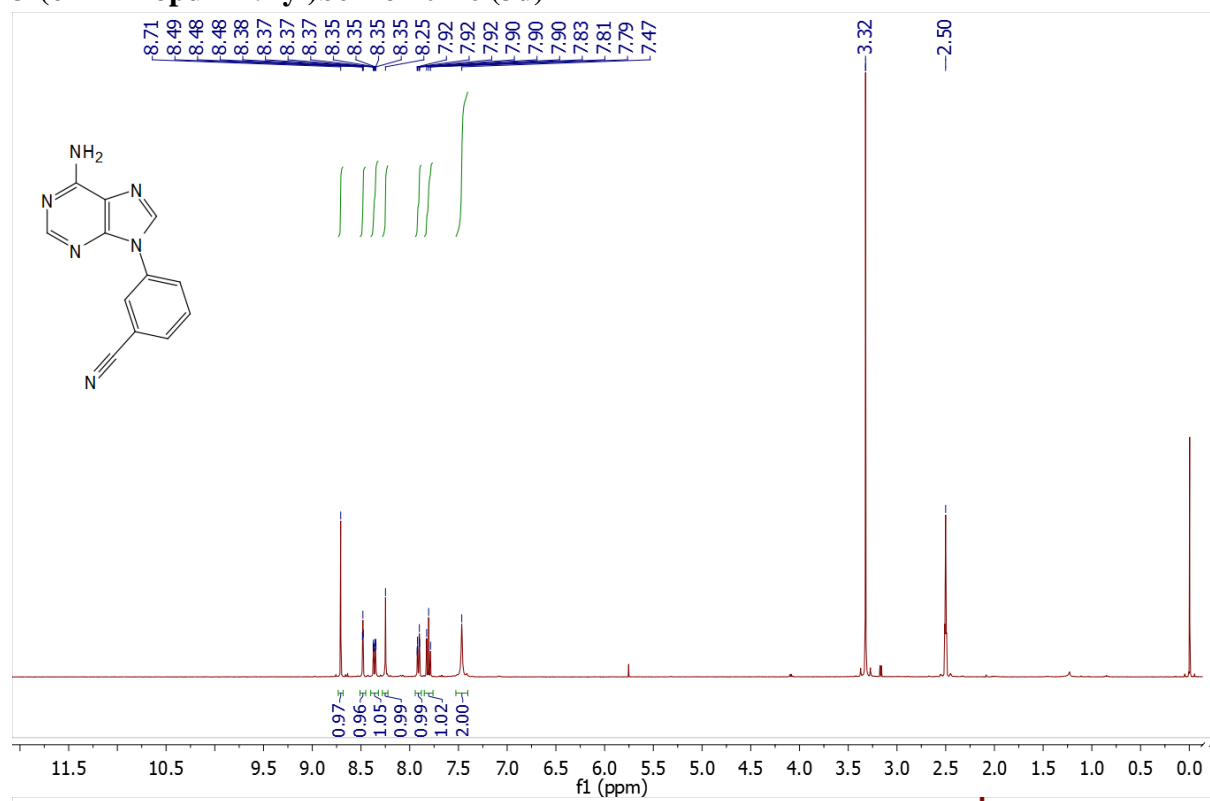
9-(4-Aminophenyl)purin-6-amine (3b)



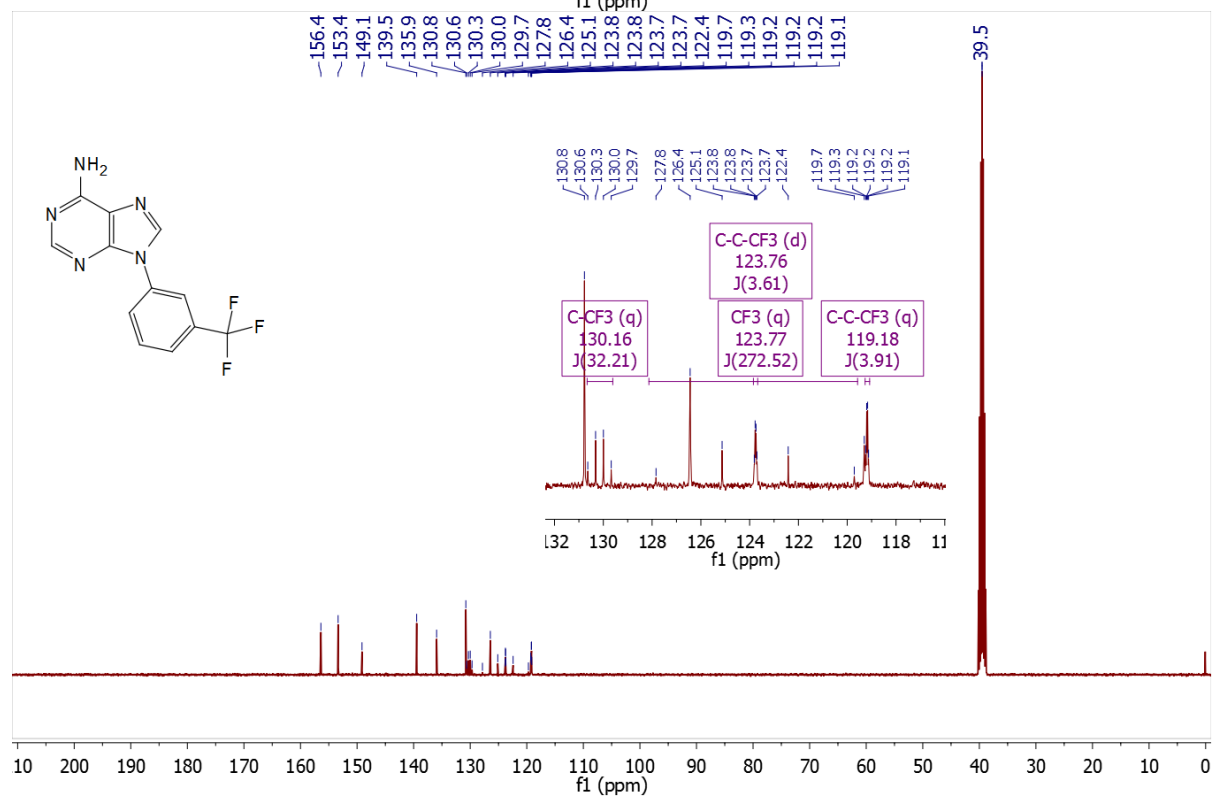
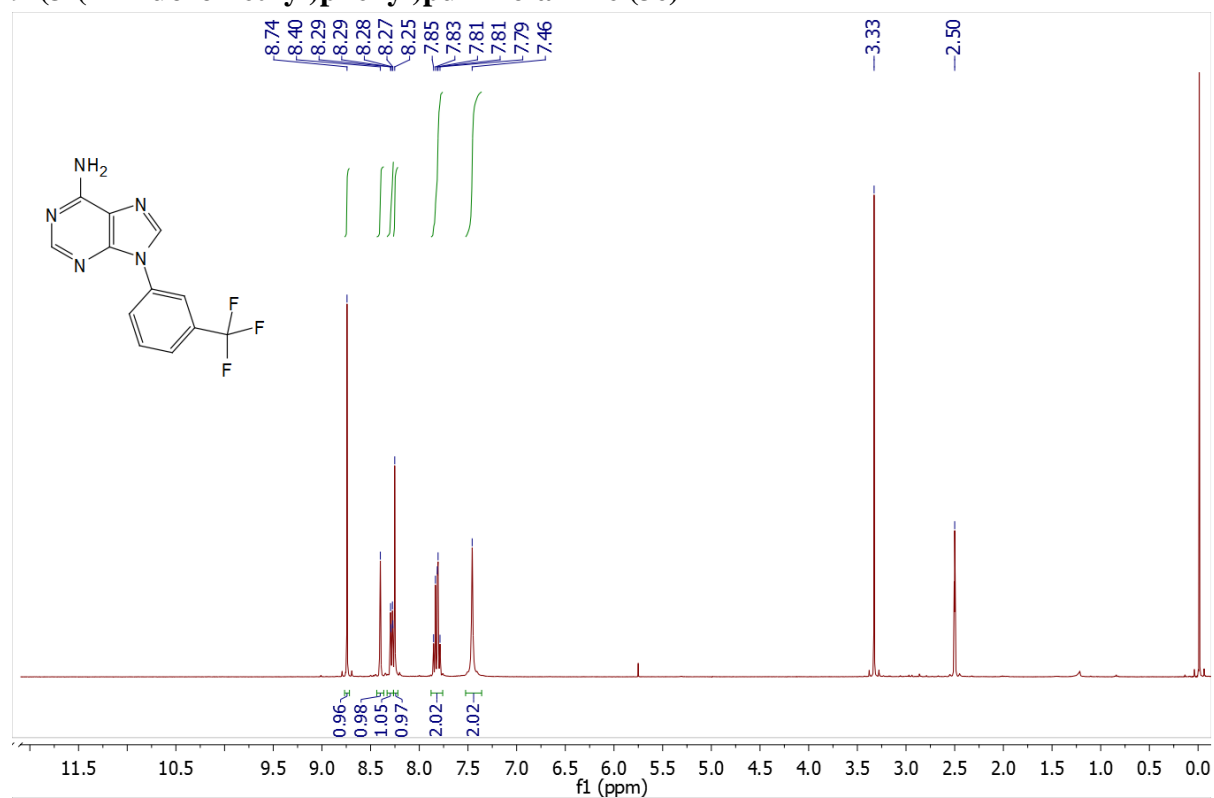
3-(6-Aminopurin-9-yl)benzoic acid (3c)



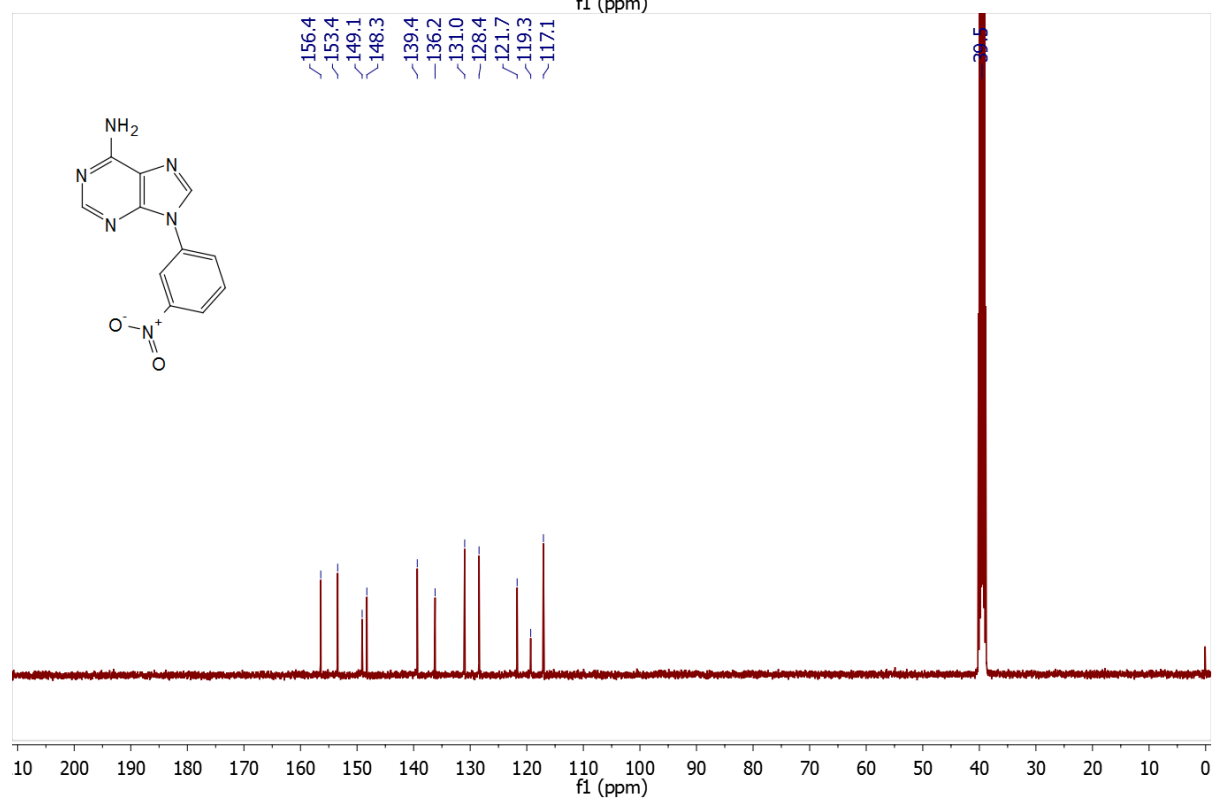
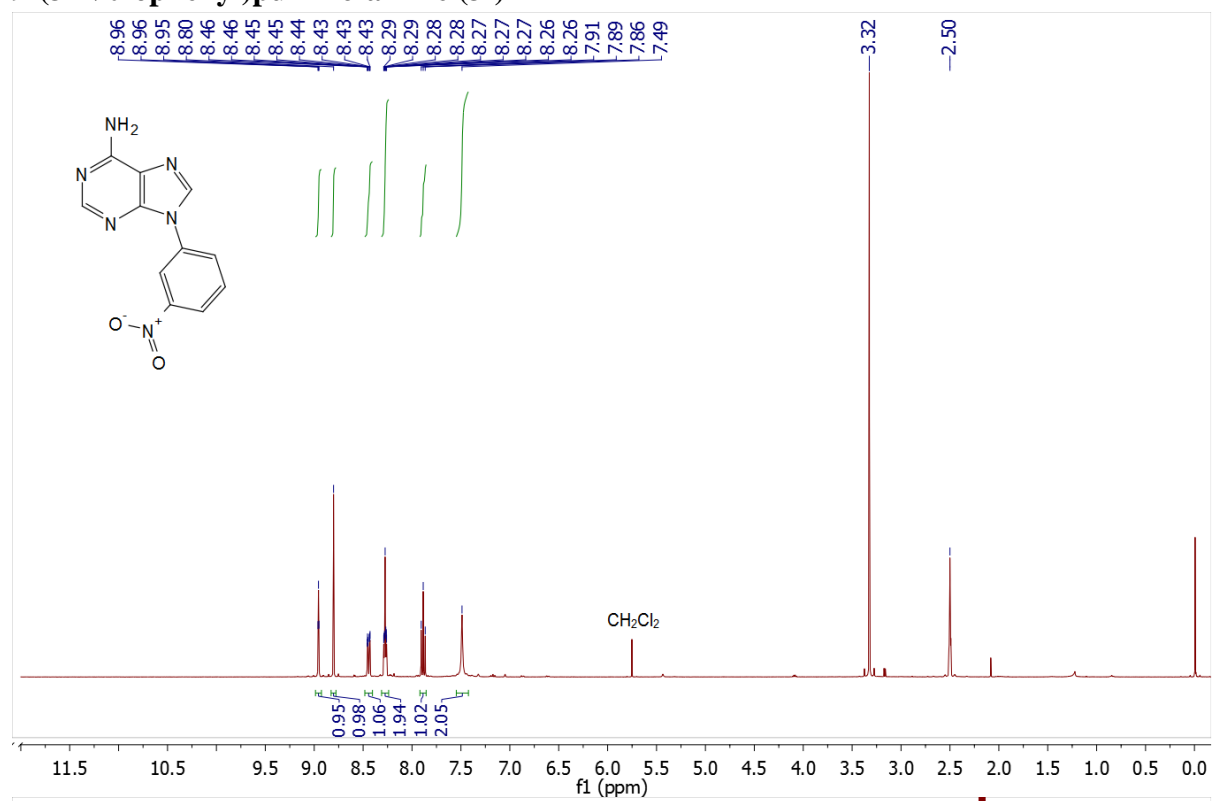
3-(6-Aminopurin-9-yl)benzonitrile (3d)



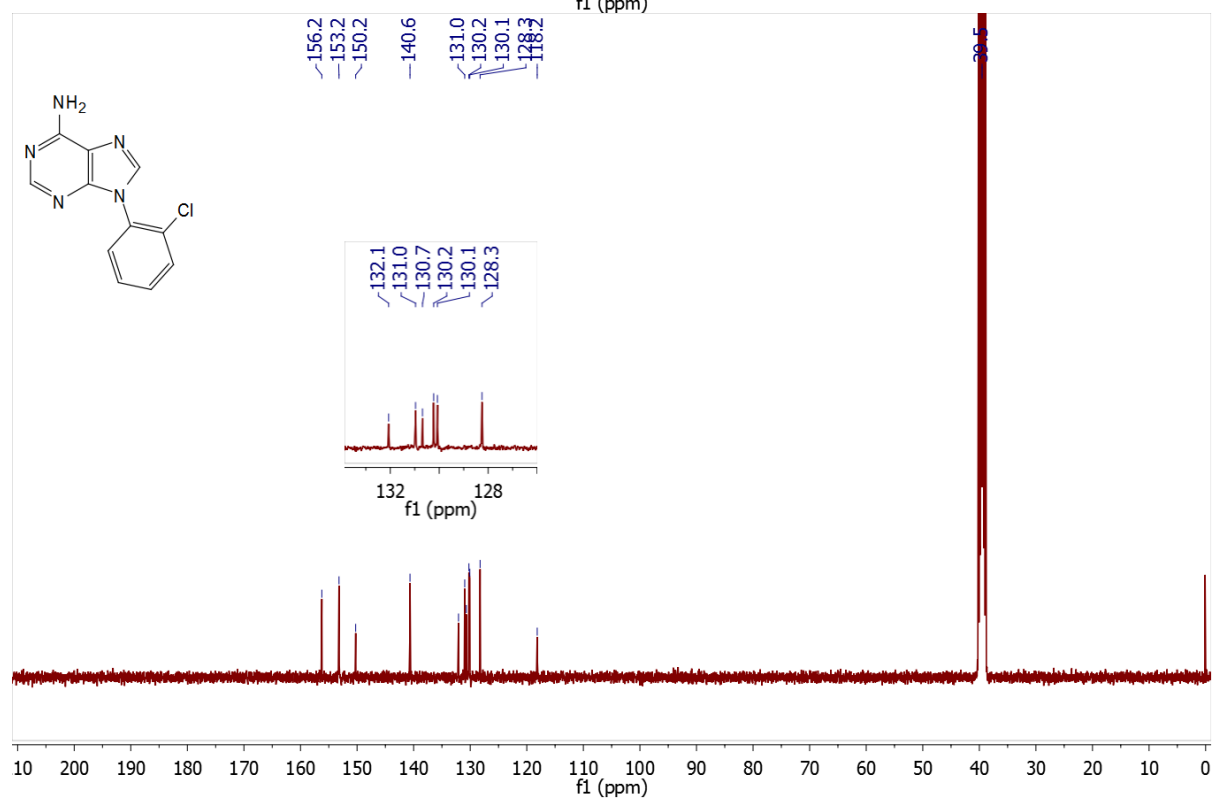
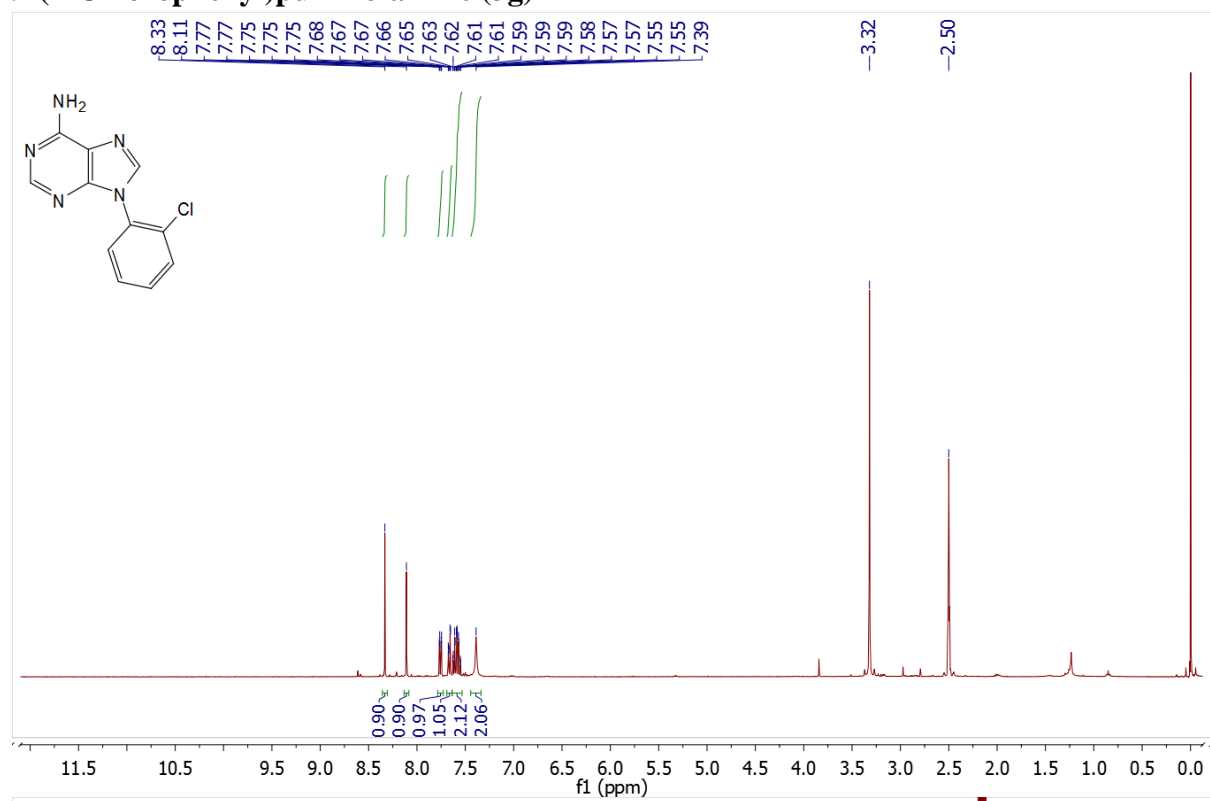
9-(3-(Trifluoromethyl)phenyl)purin-6-amine (3e)



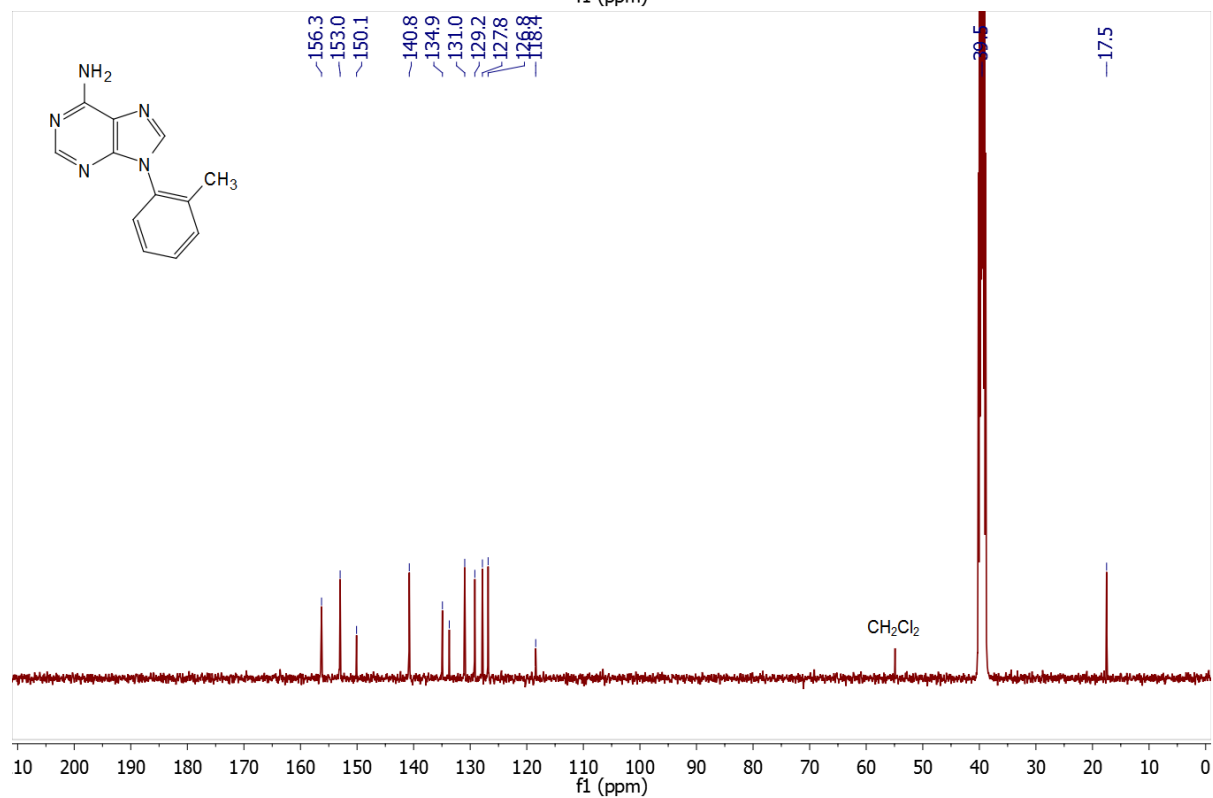
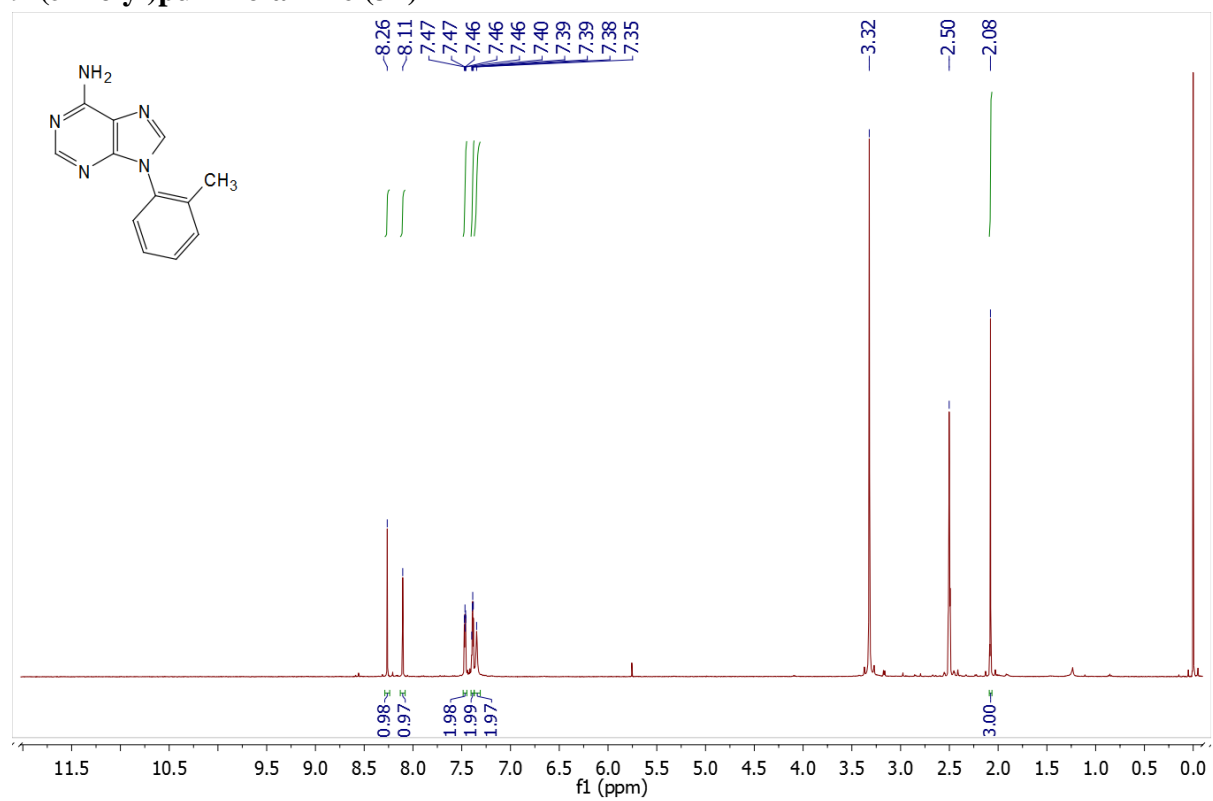
9-(3-Nitrophenyl)purin-6-amine (3f)



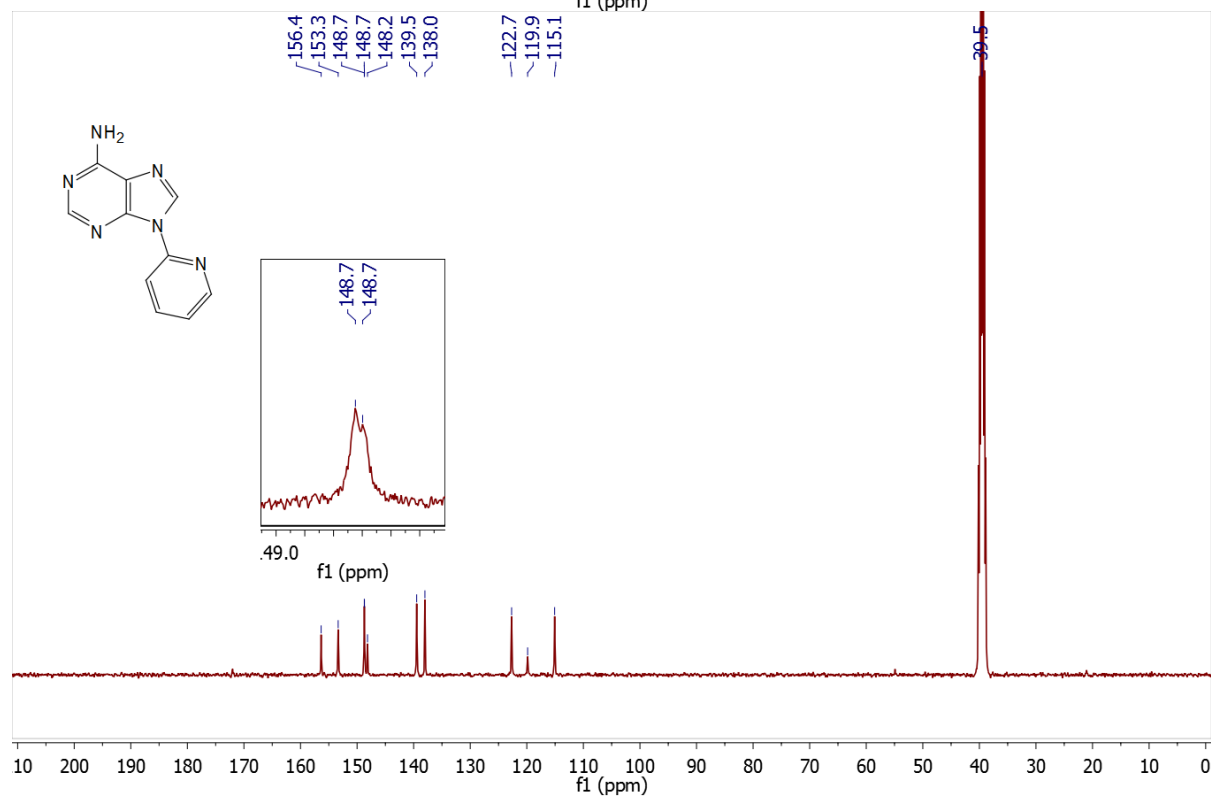
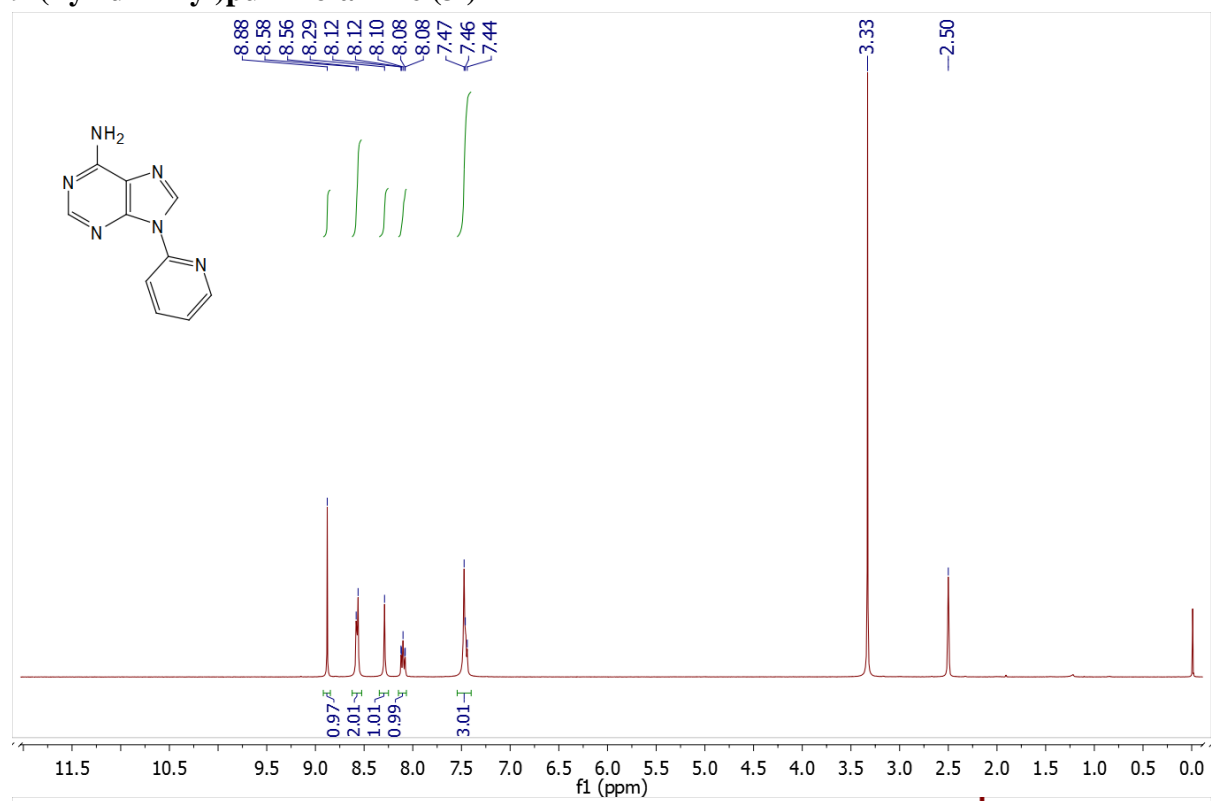
9-(2-Chlorophenyl)purin-6-amine (3g)



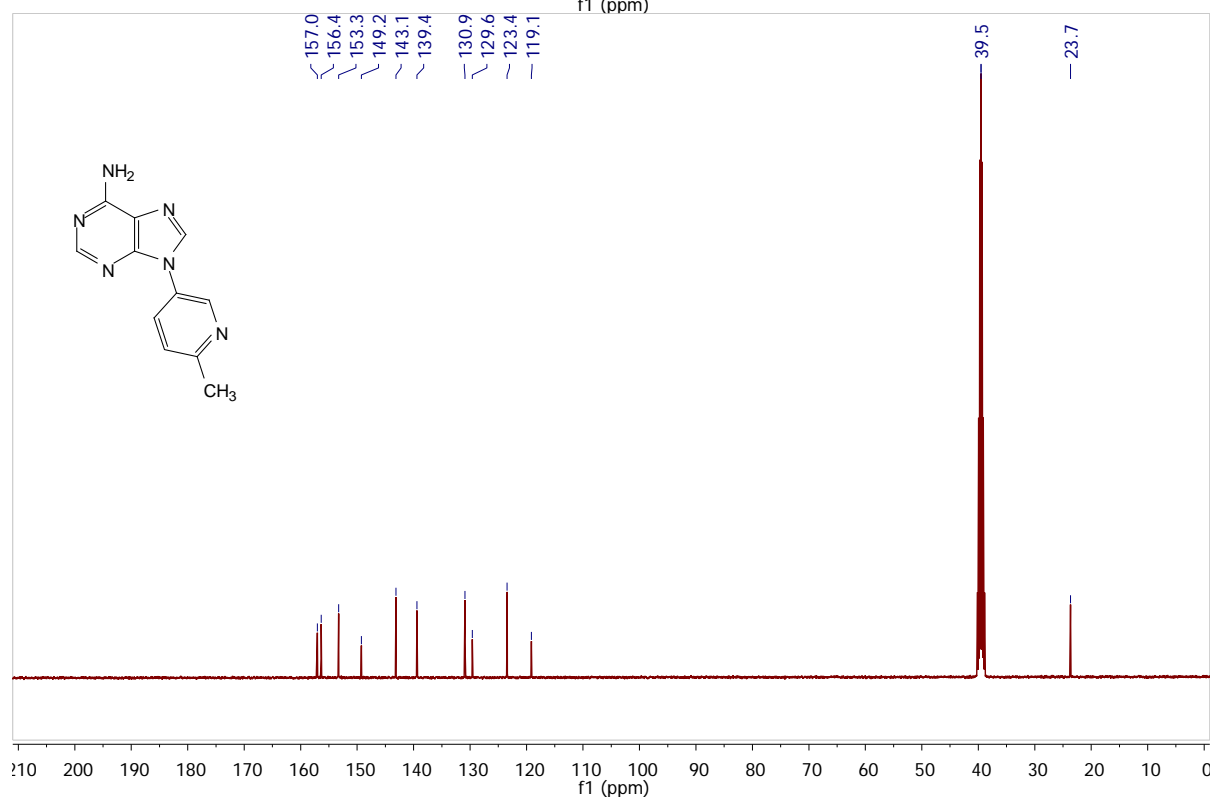
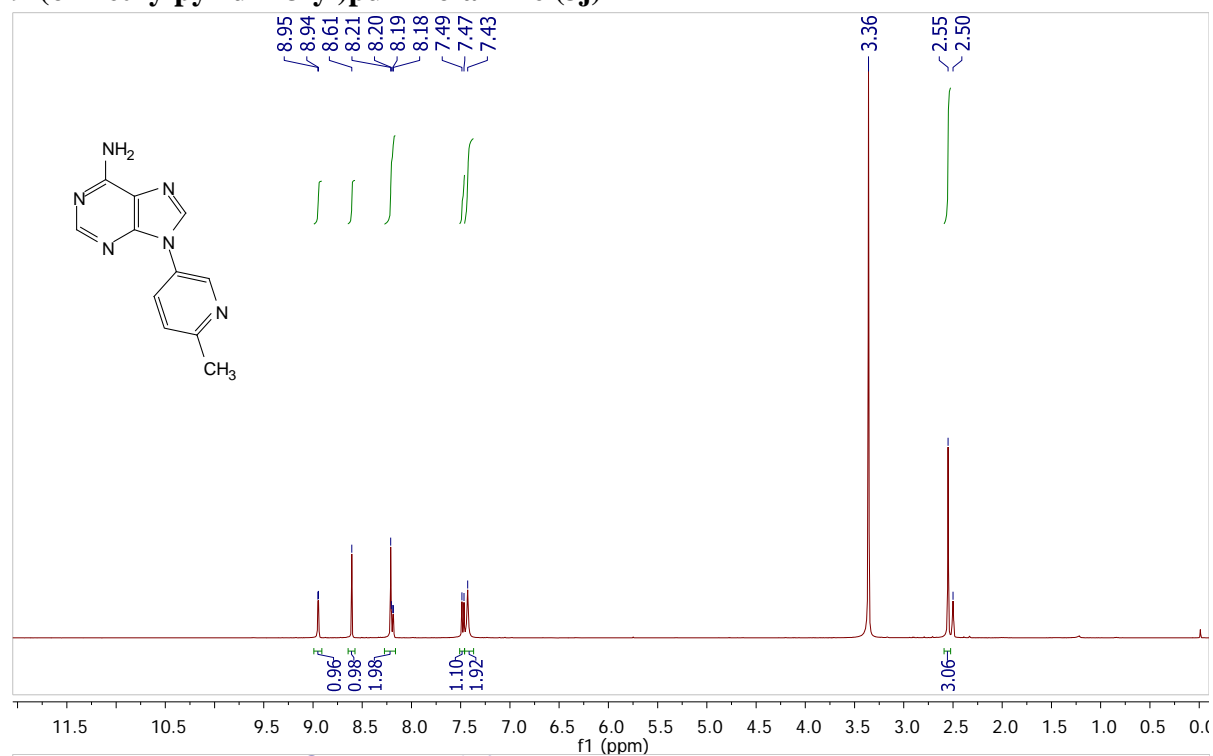
9-(*o*-Tolyl)purin-6-amine (3h)



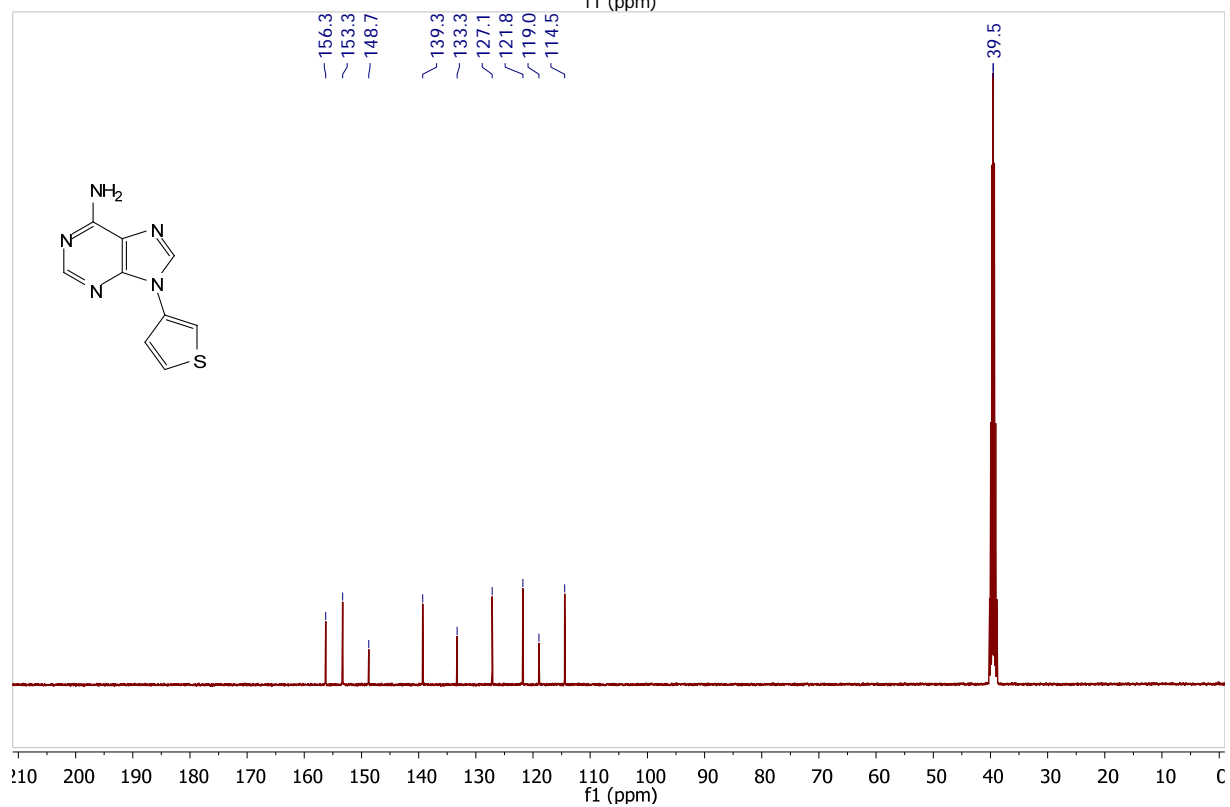
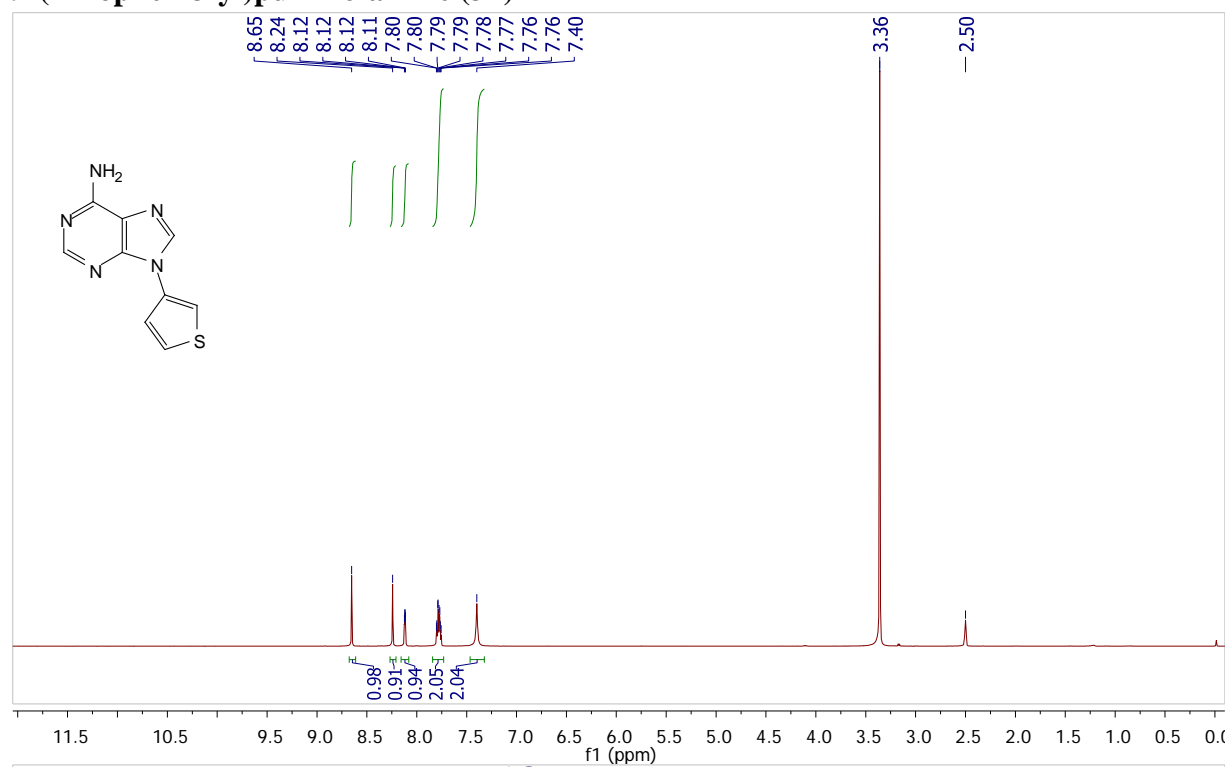
9-(Pyridin-2-yl)purin-6-amine (3i)



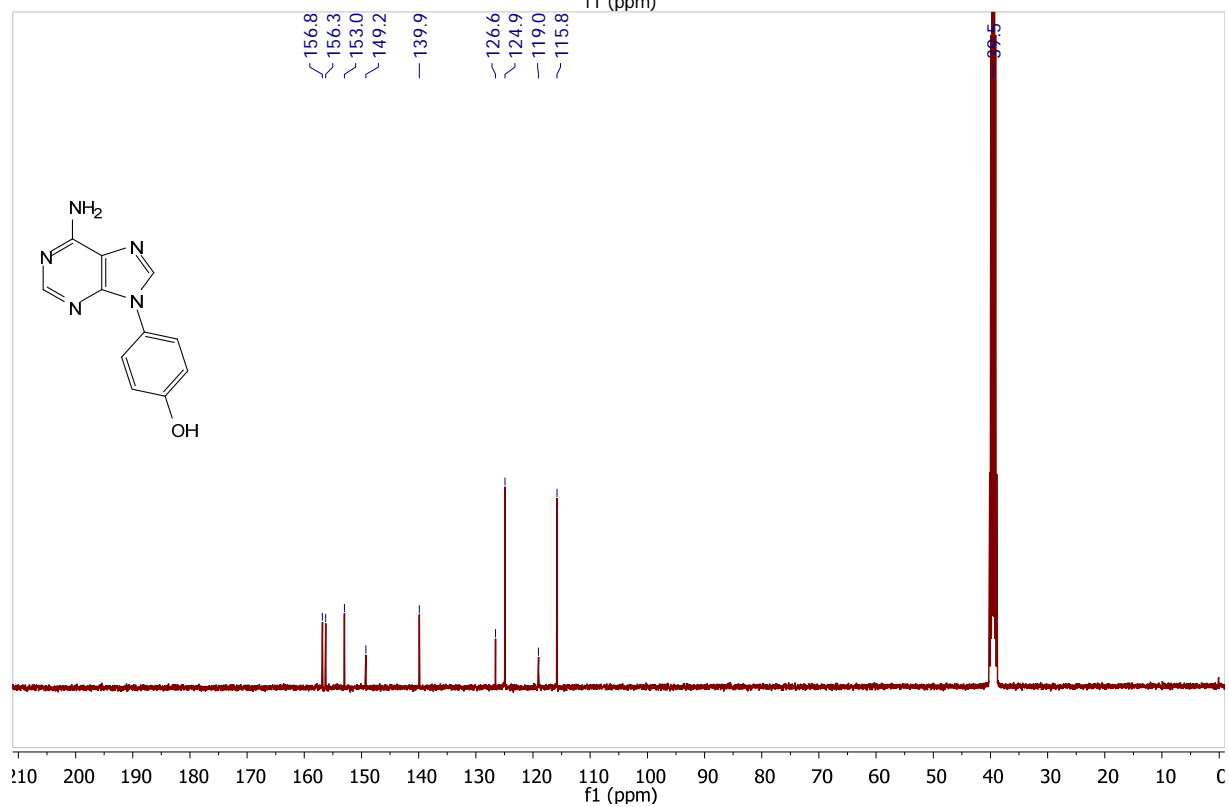
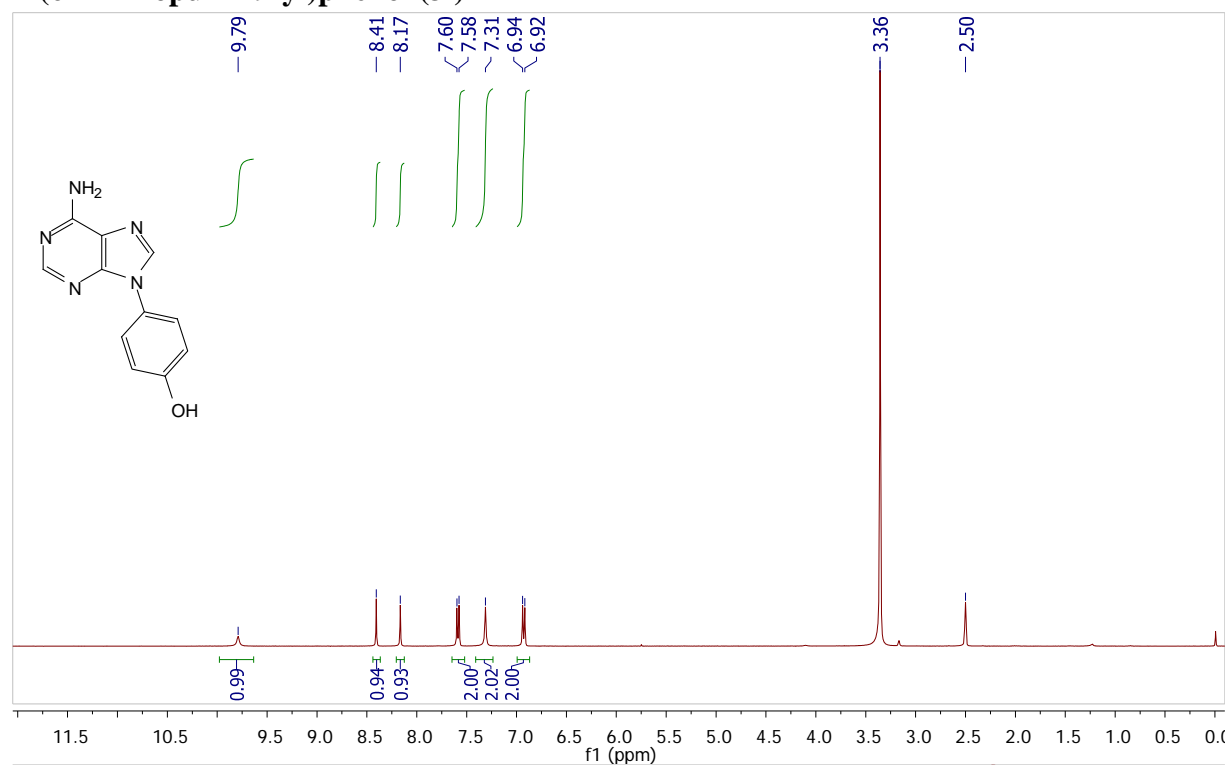
9-(6-Methylpyridin-3-yl)purin-6-amine (3j)



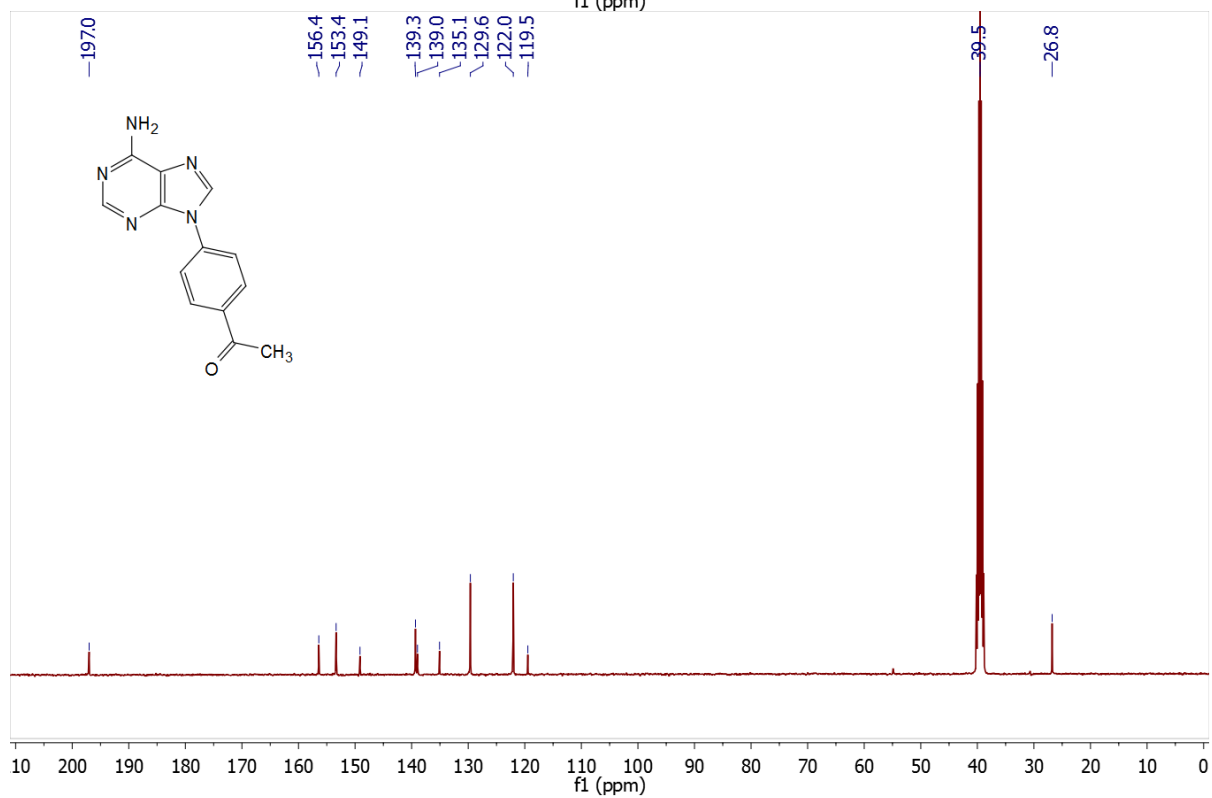
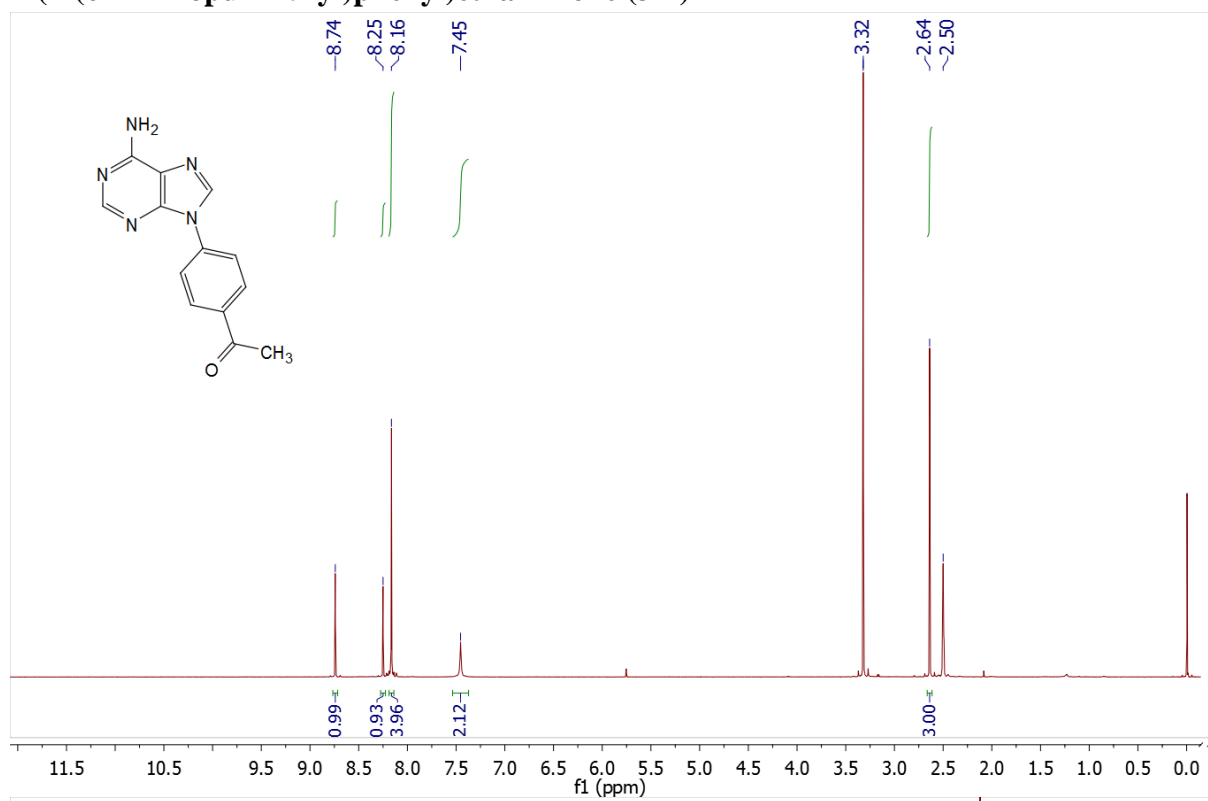
9-(Thiophen-3-yl)purin-6-amine (3k)



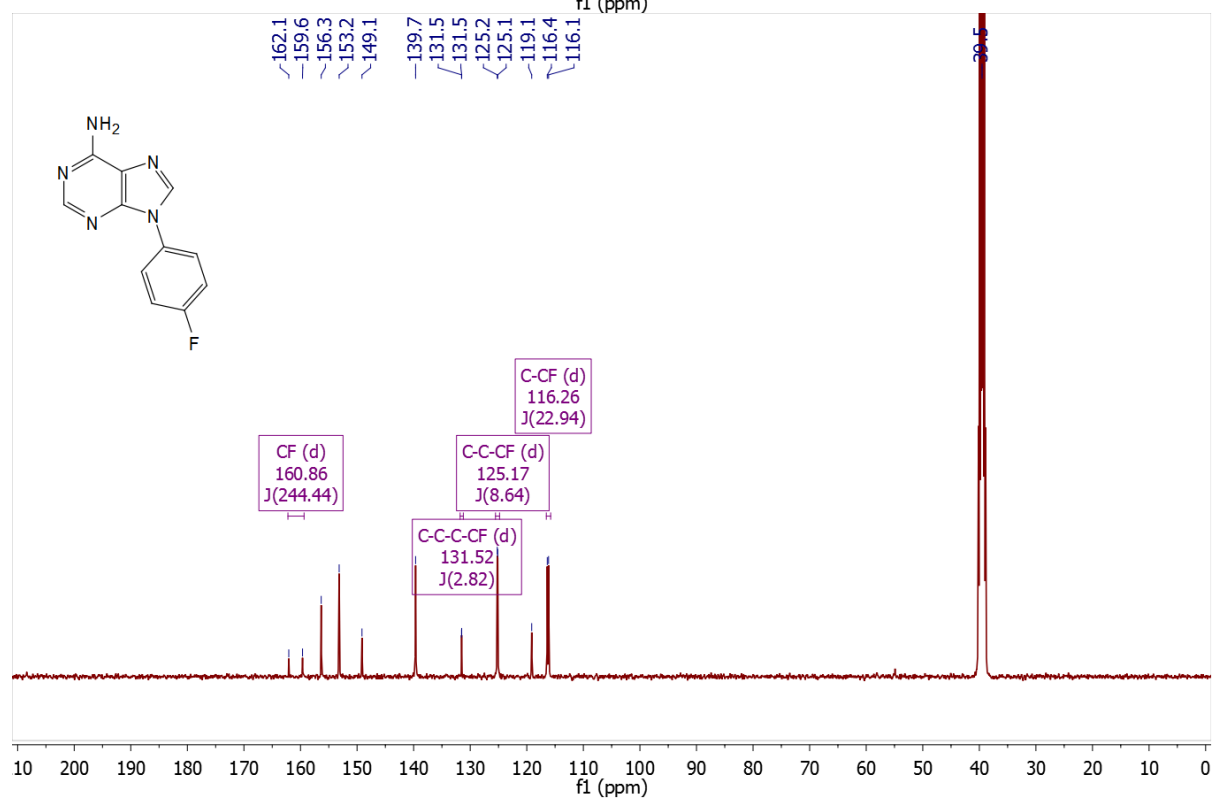
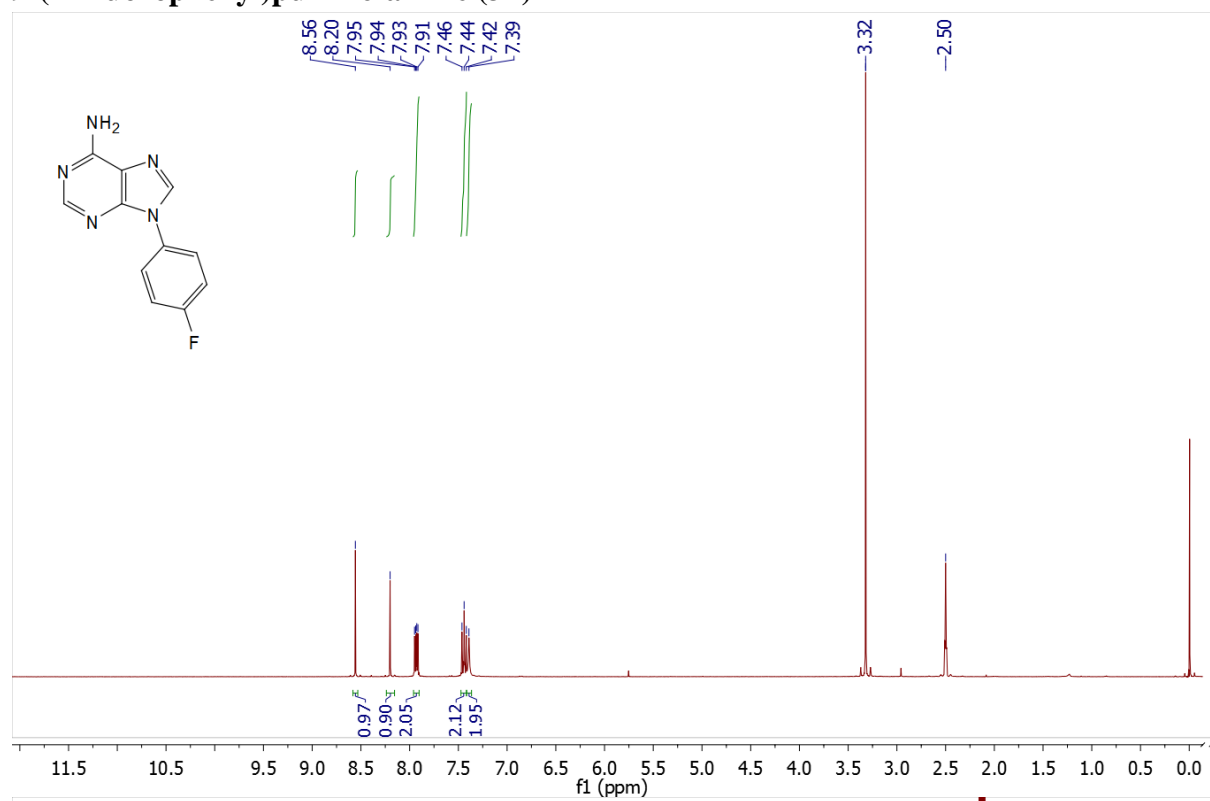
4-(6-Aminopurin-9-yl)phenol (3l)



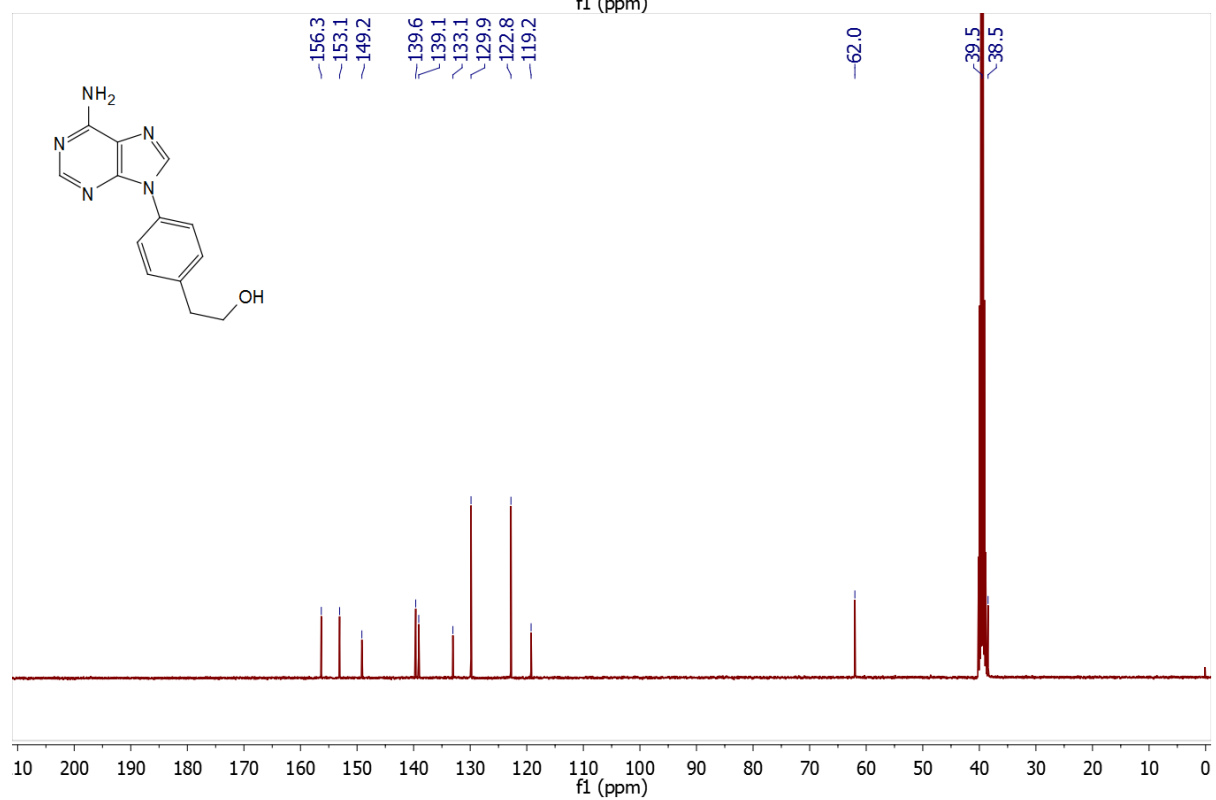
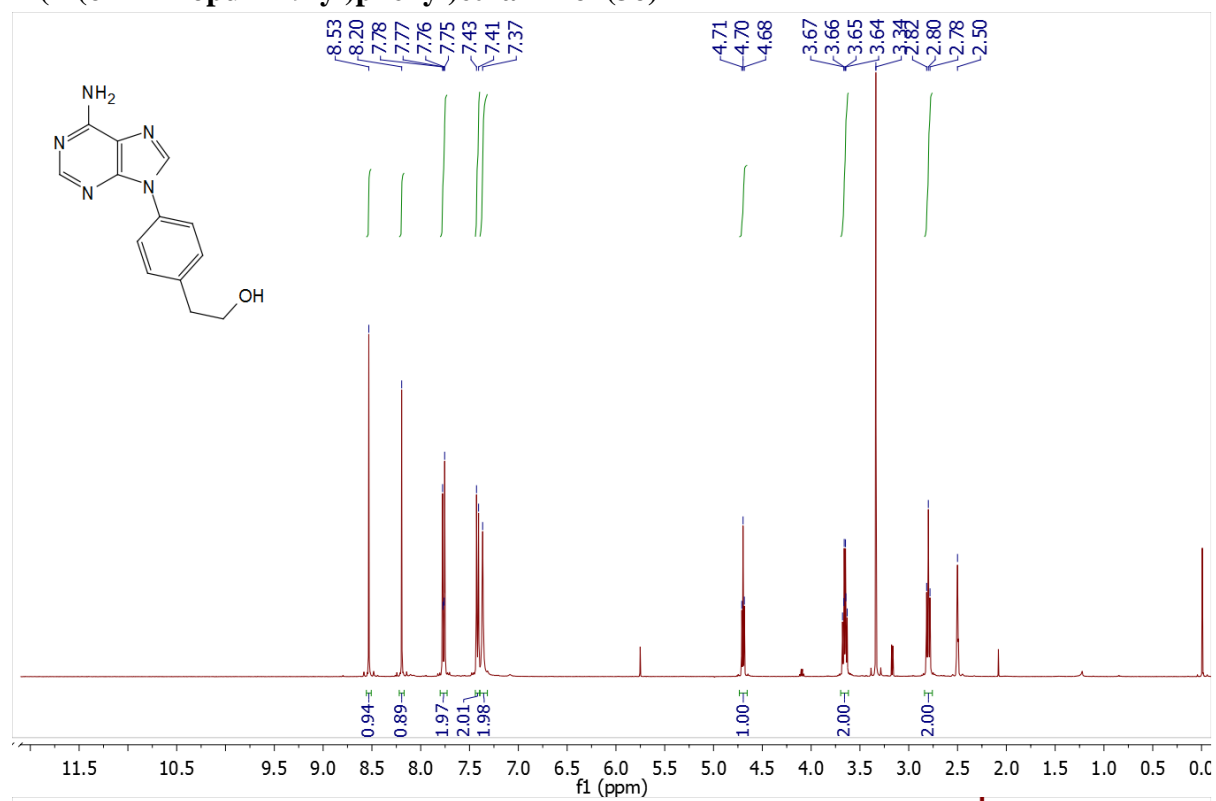
1-(4-(6-Aminopurin-9-yl)phenyl)ethan-1-one (3m)



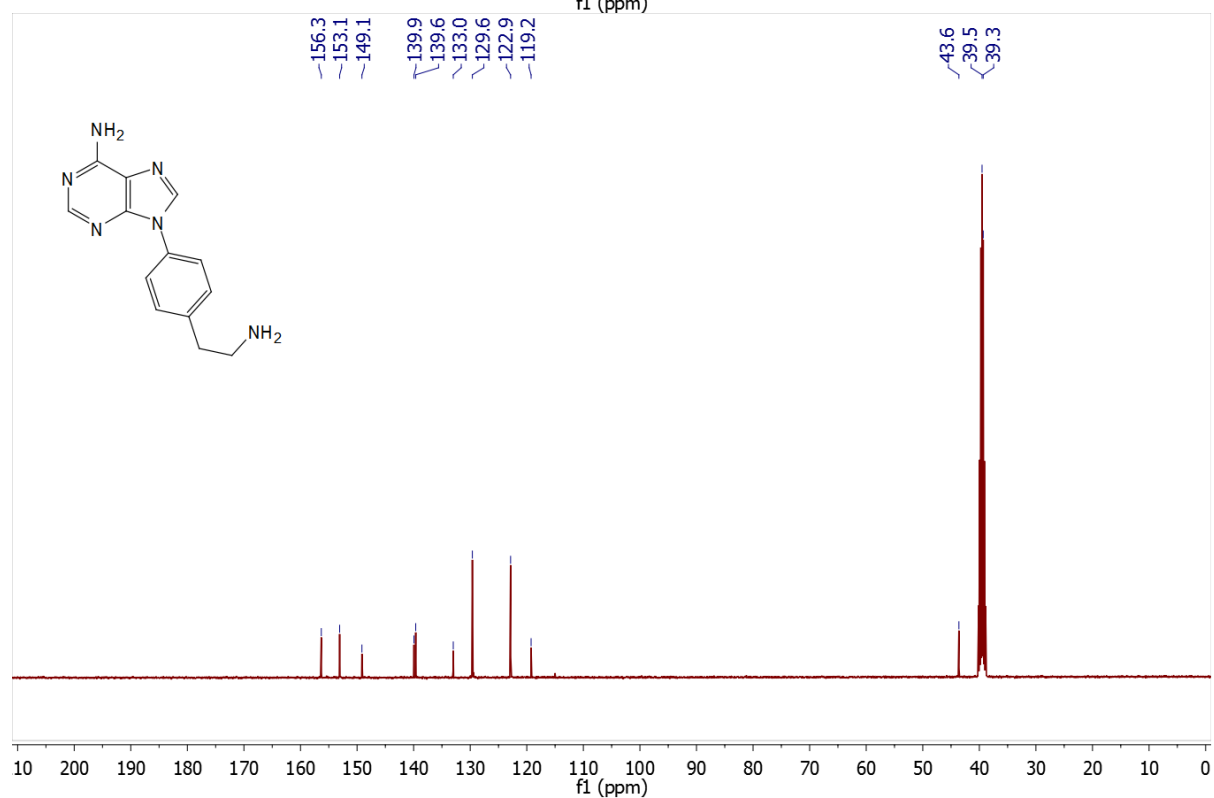
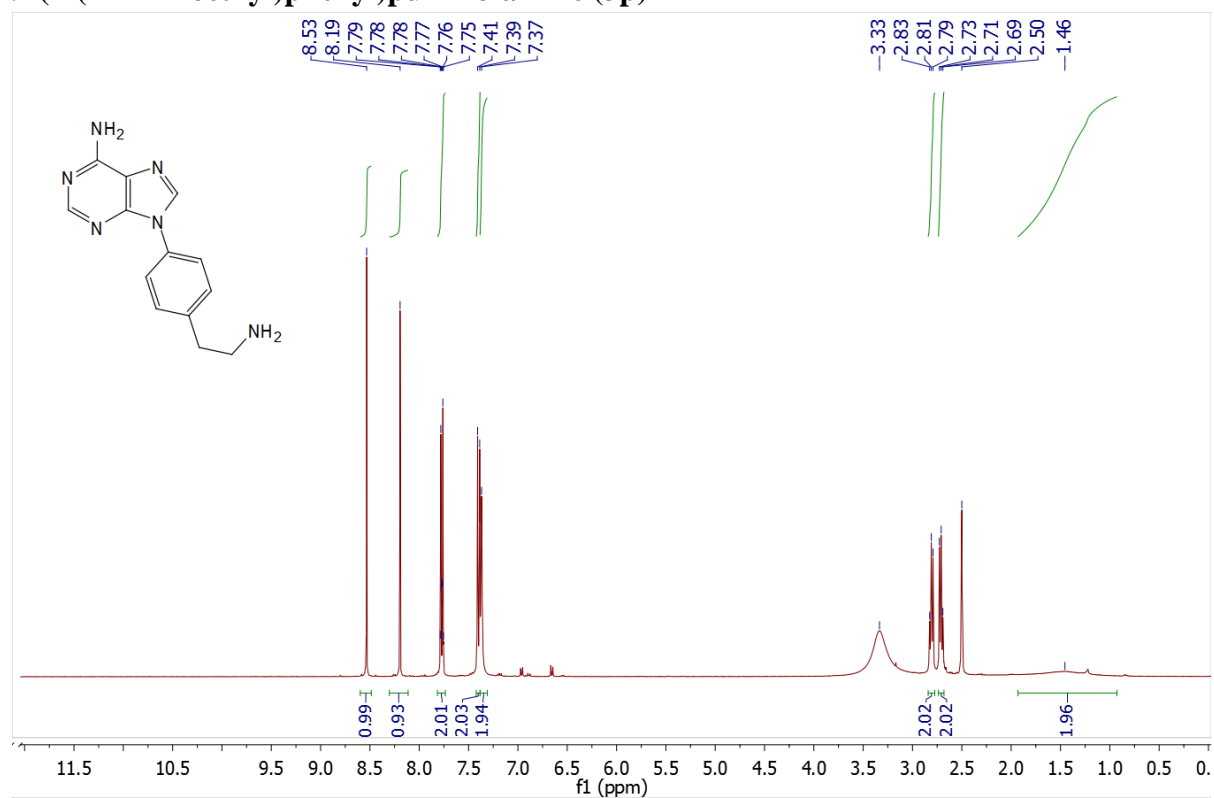
9-(4-Fluorophenyl)purin-6-amine (3n)



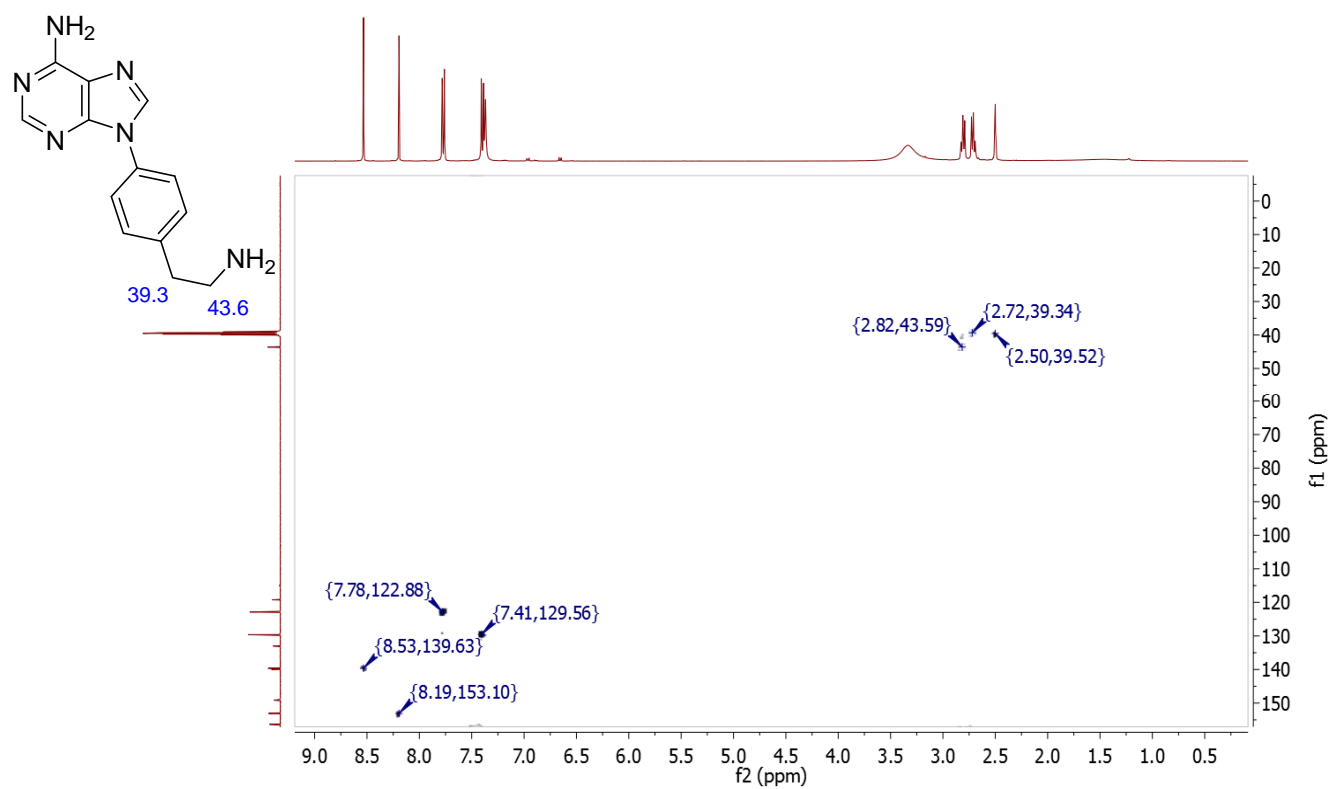
2-(4-(6-Aminopurin-9-yl)phenyl)ethan-1-ol (3o)



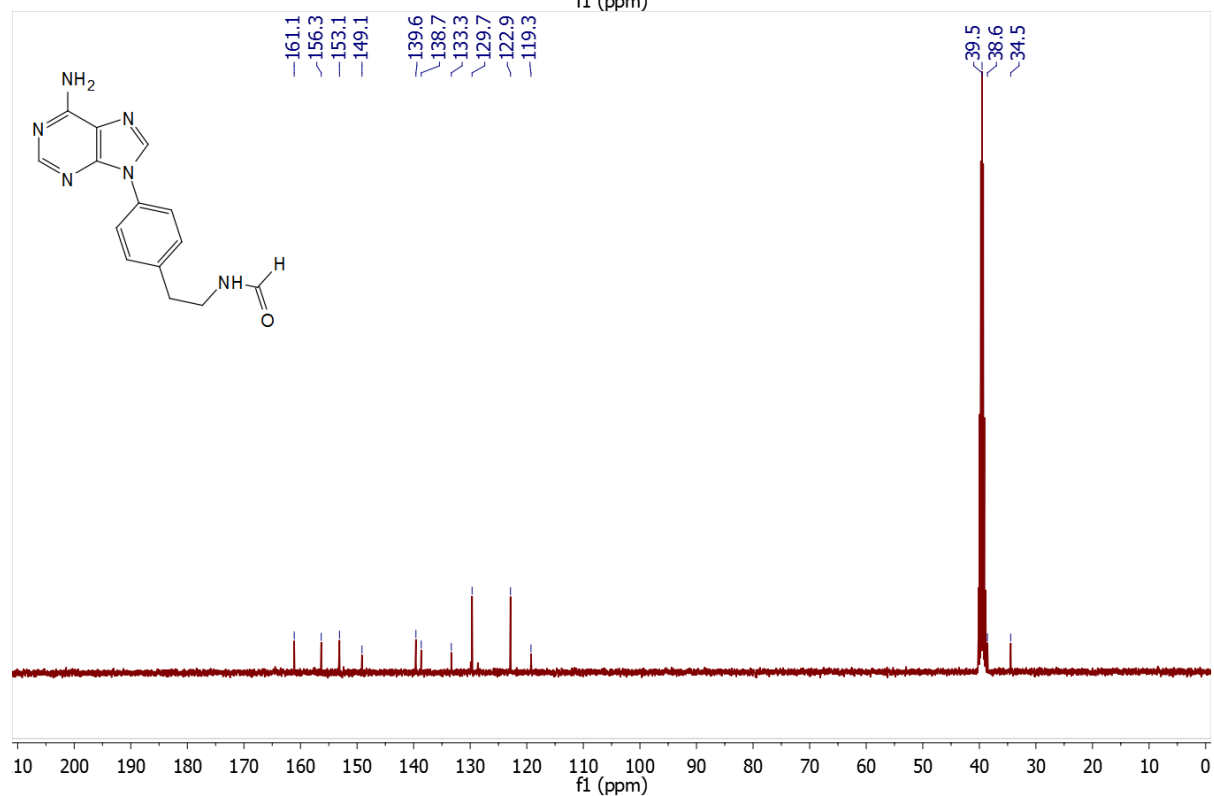
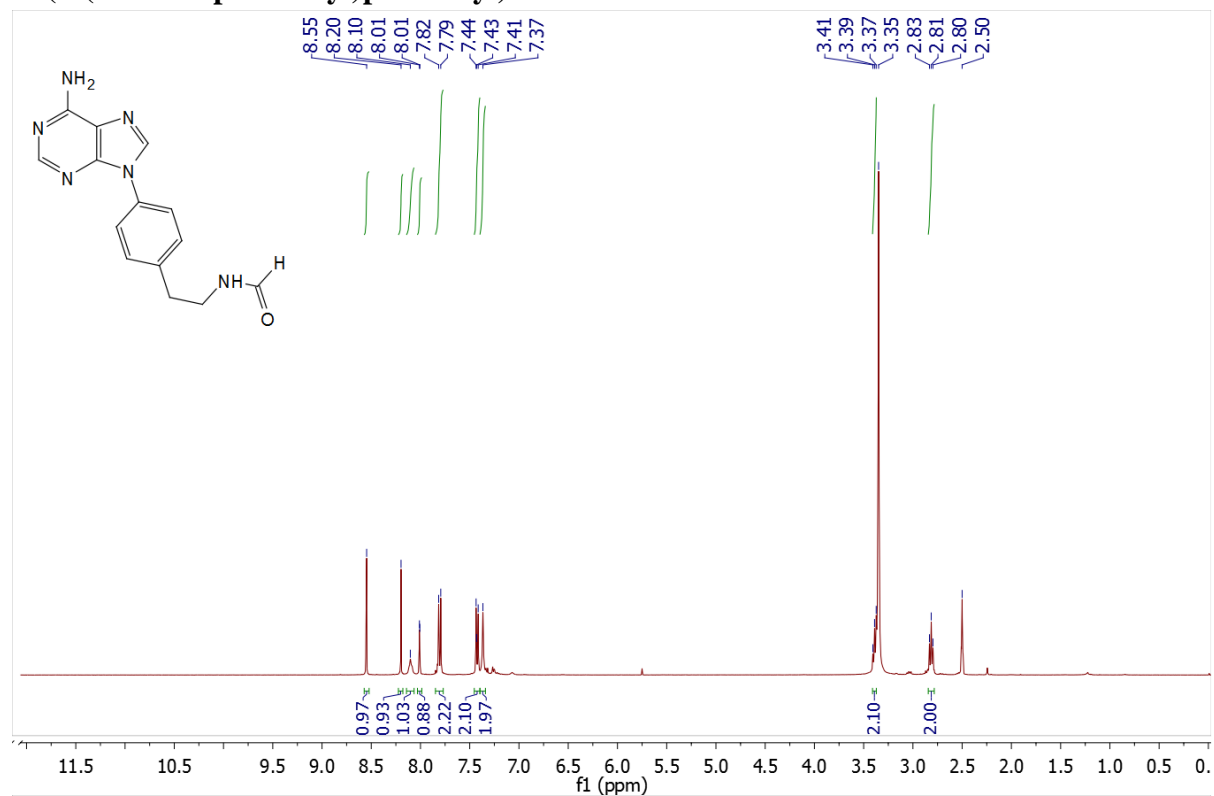
9-(4-(2-Aminoethyl)phenyl)purin-6-amine (3p)



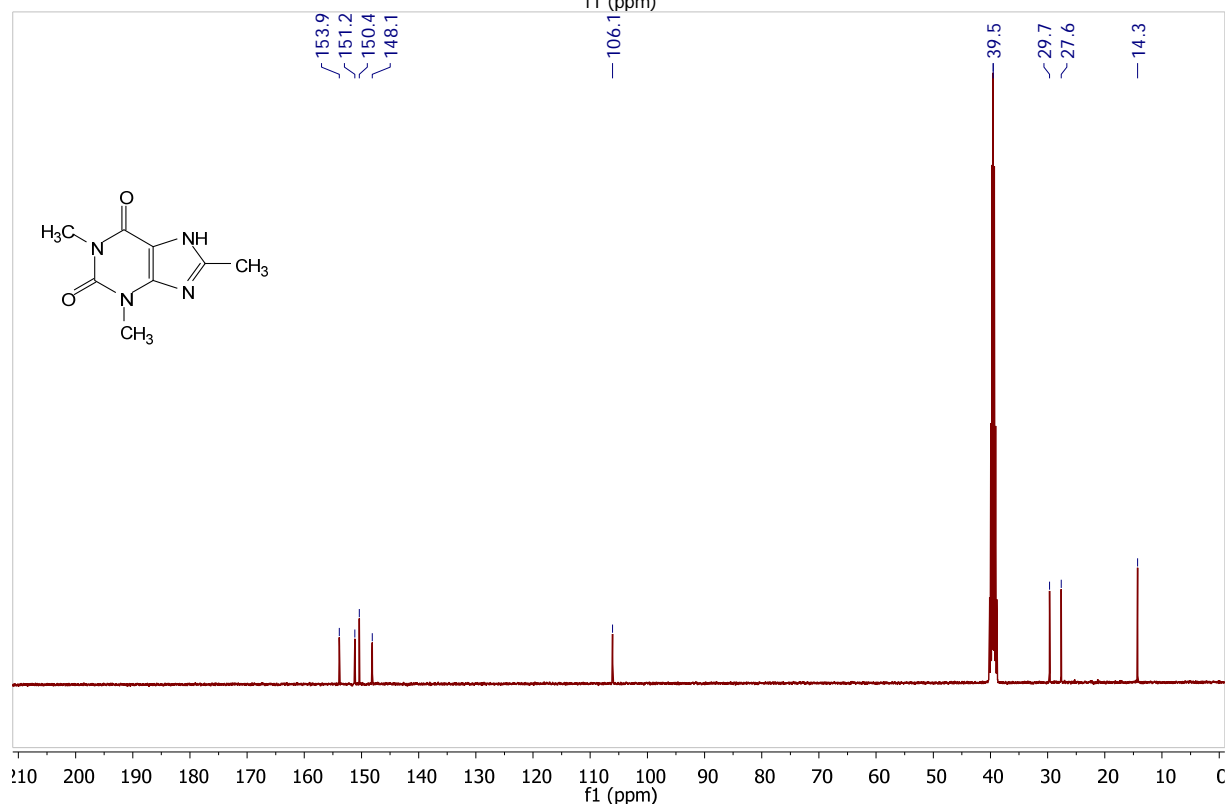
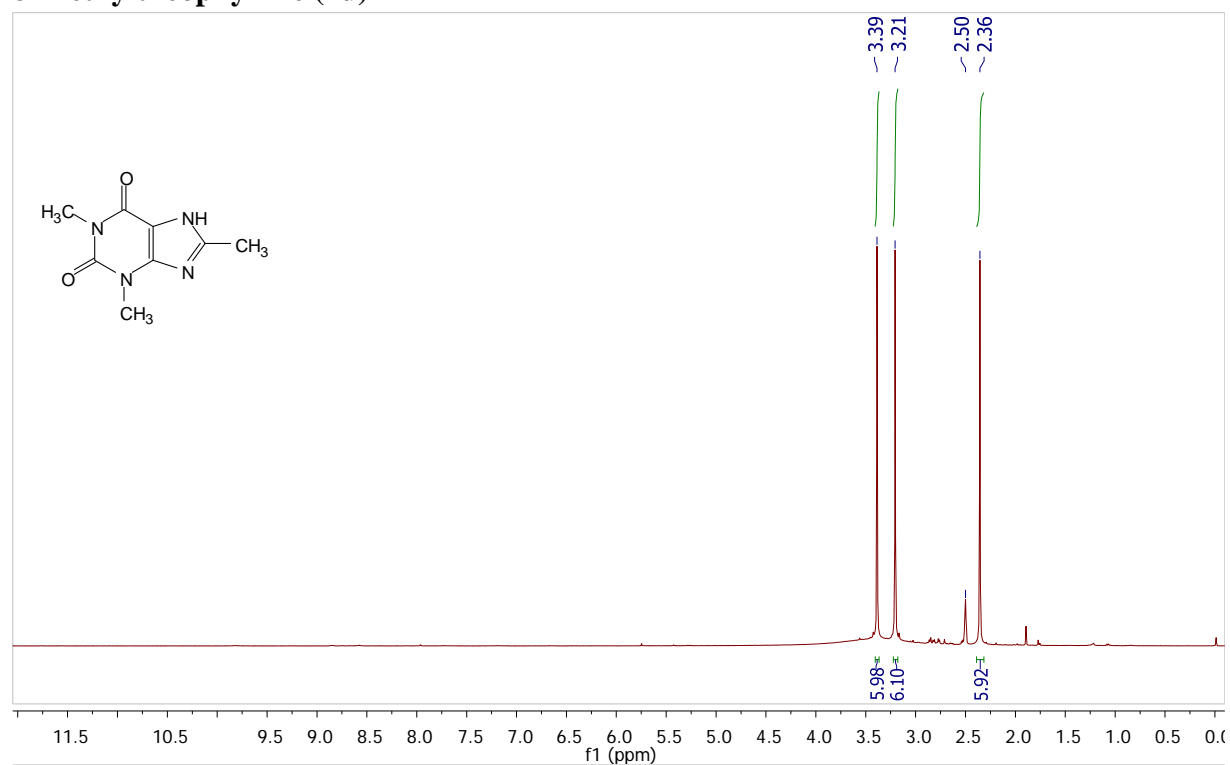
HSQC of **3p**:



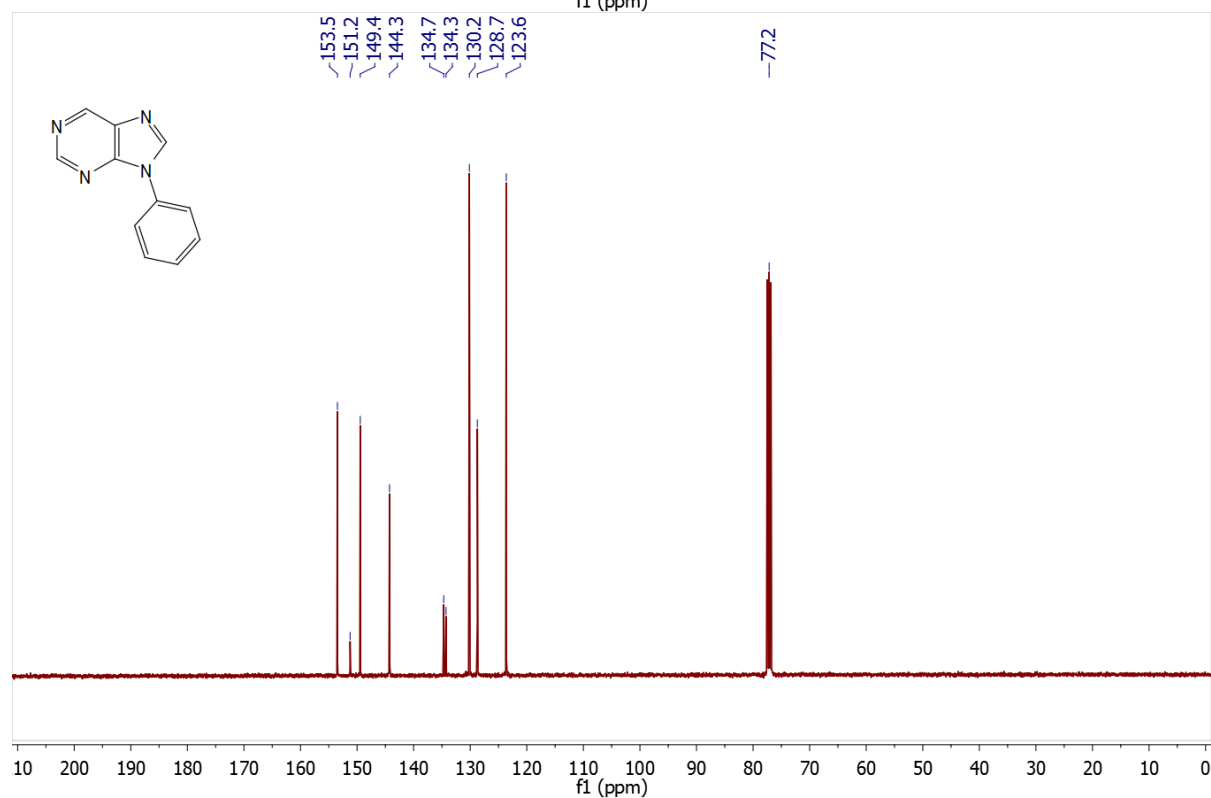
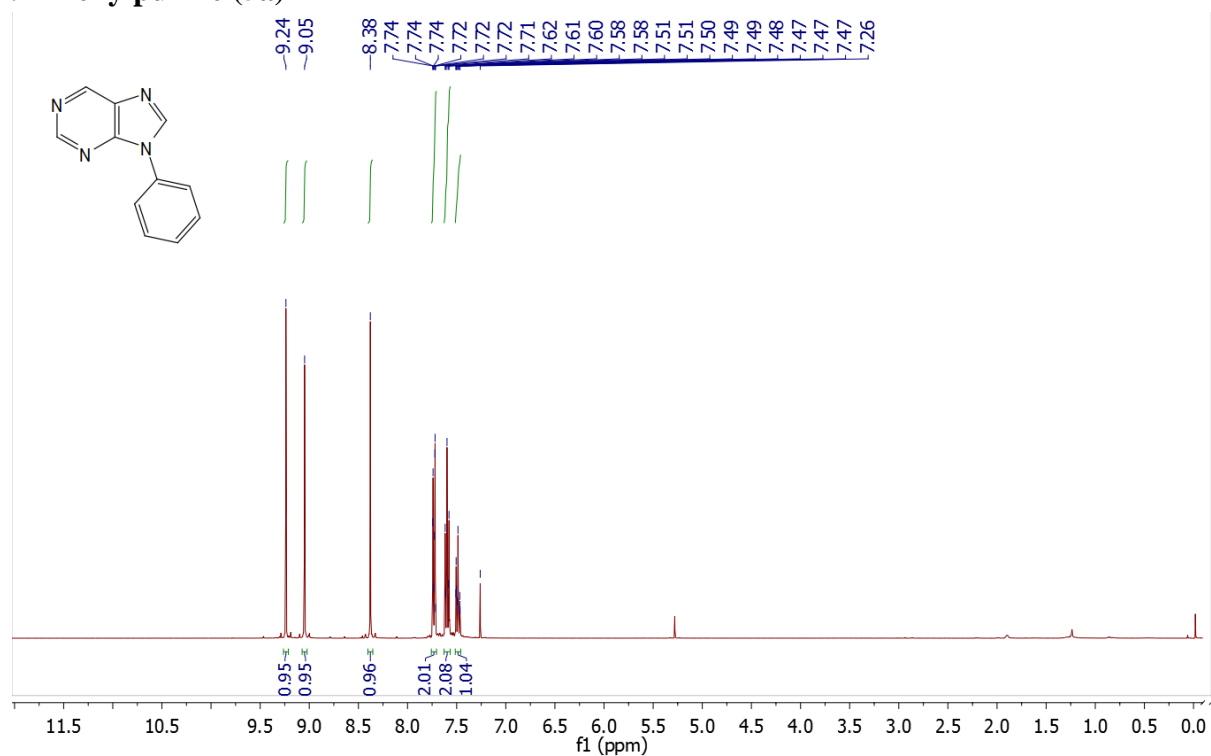
***N*-(4-(6-Aminopurin-9-yl)phenethyl)formamide**



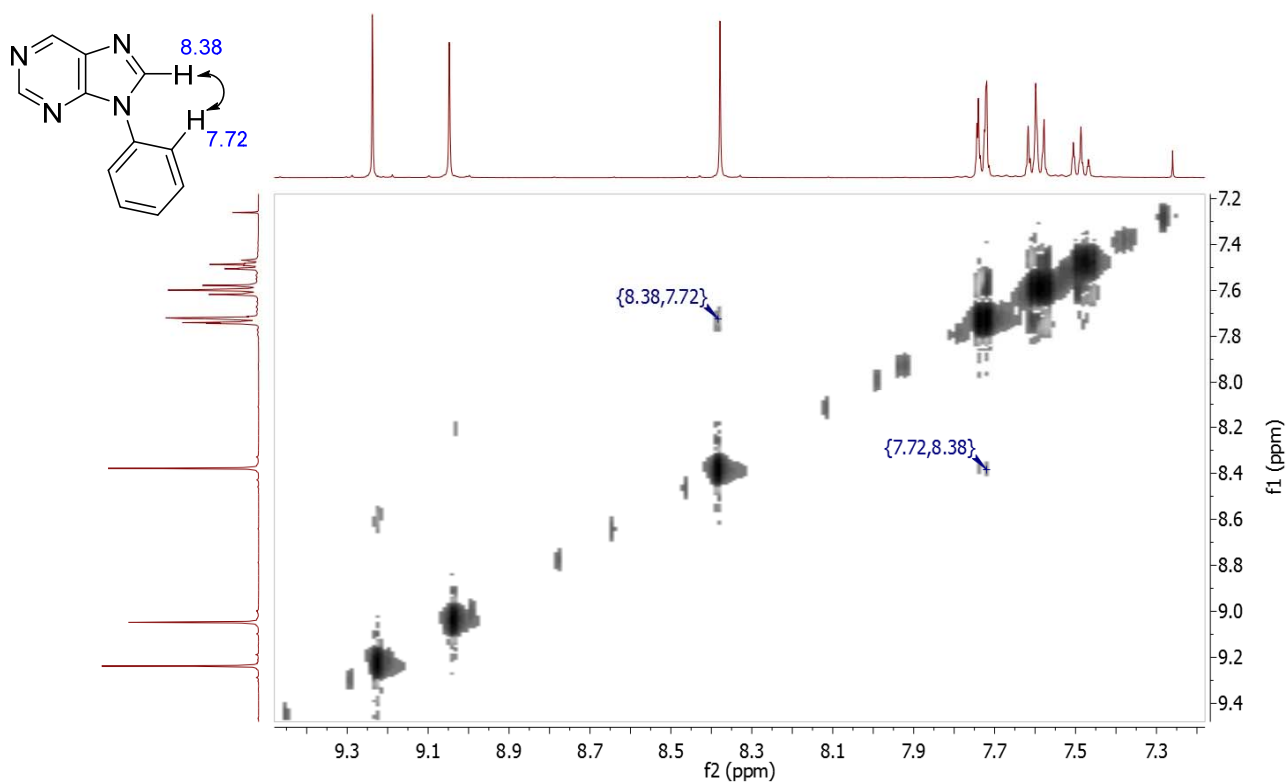
8-Methyltheophylline (4d)



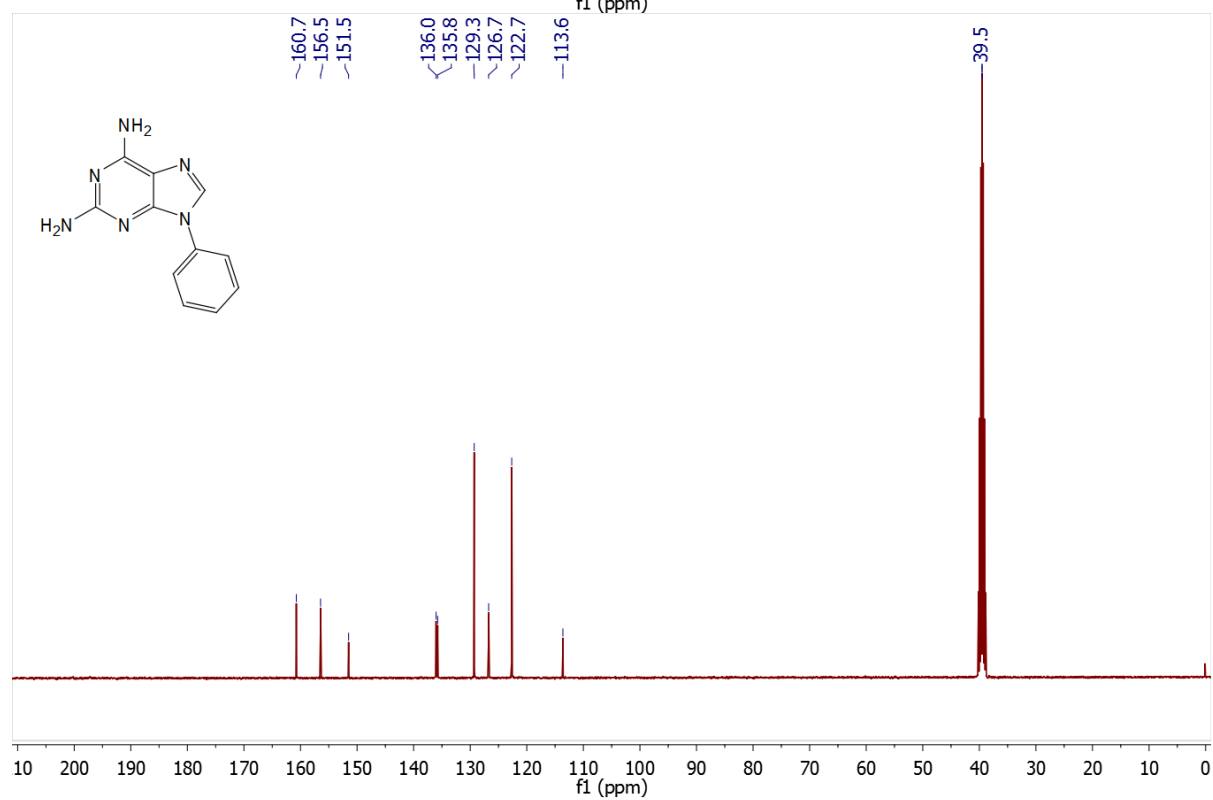
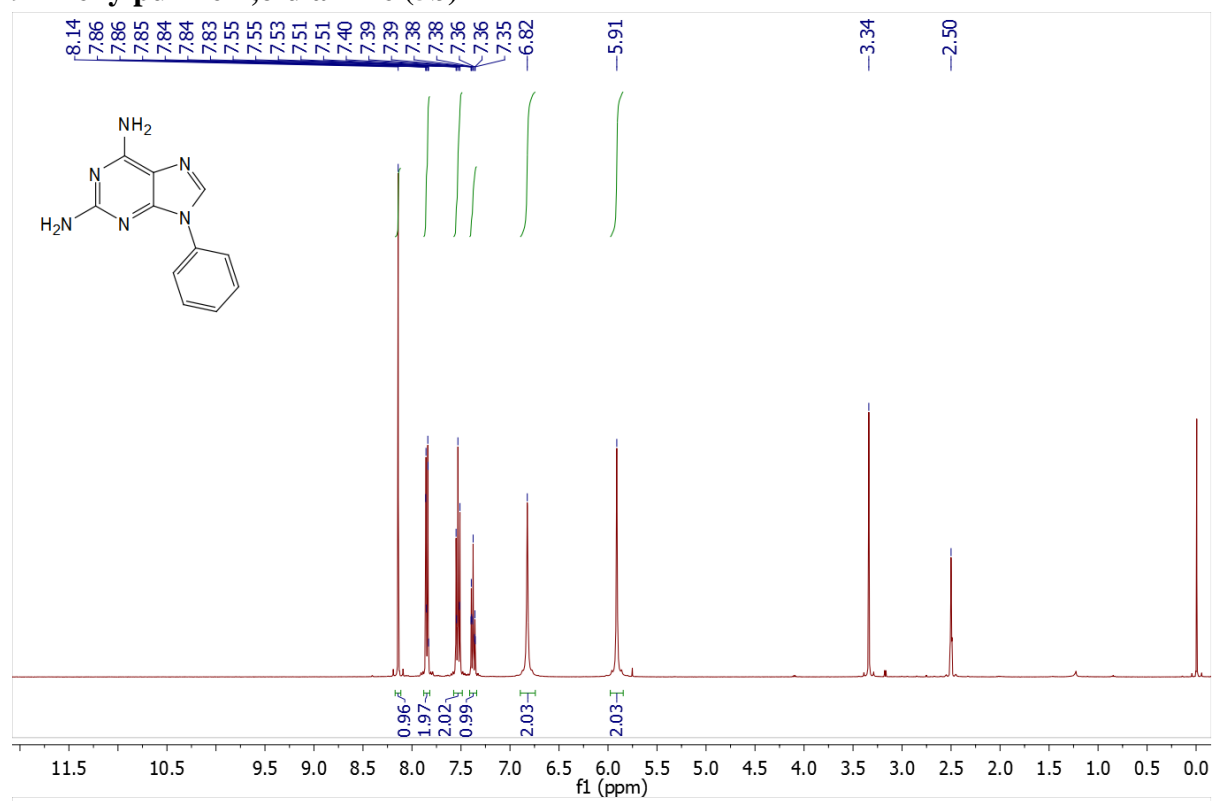
9-Phenylpurine (5a)



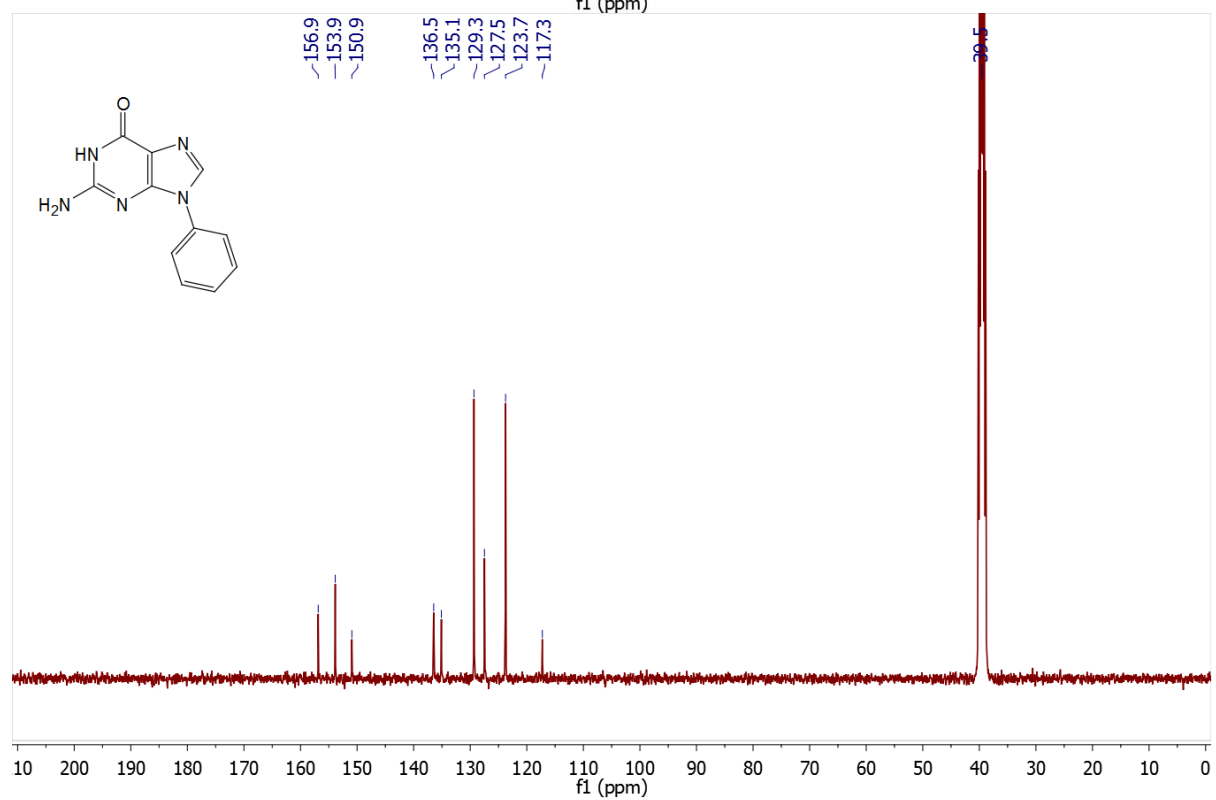
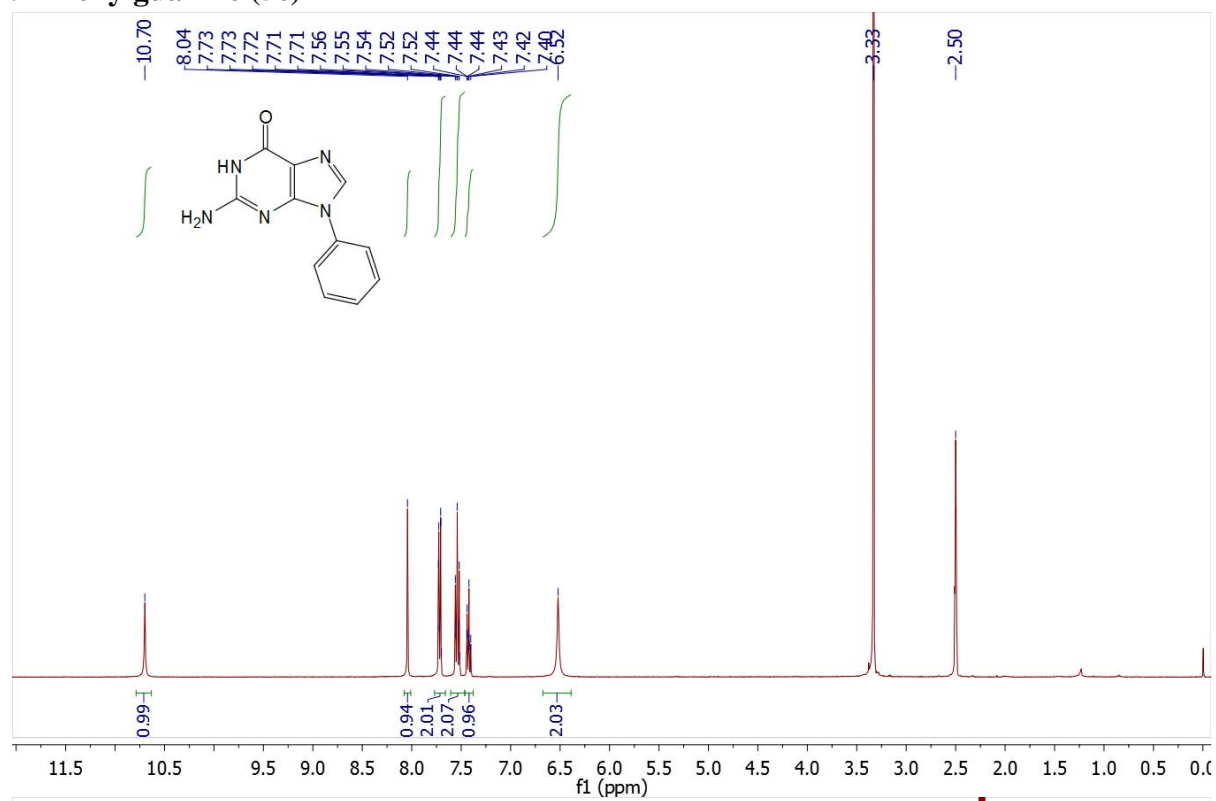
NOESY of **5a**



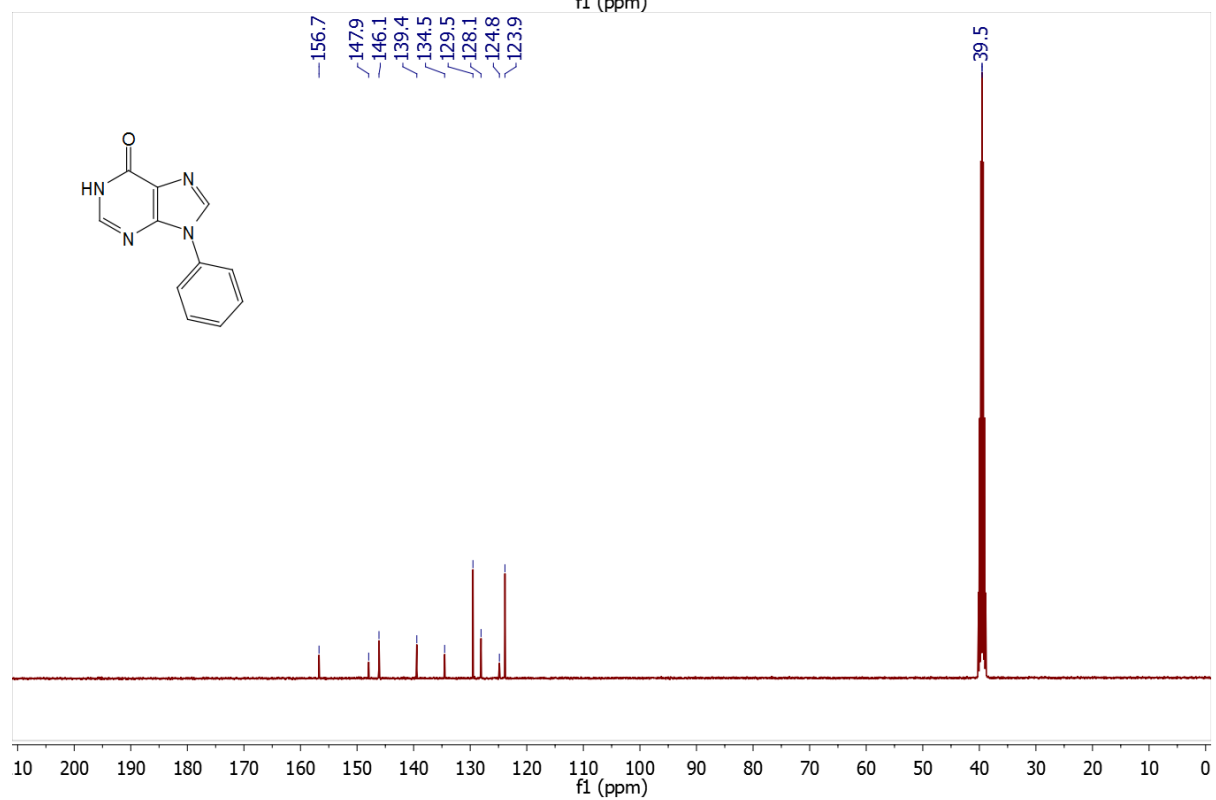
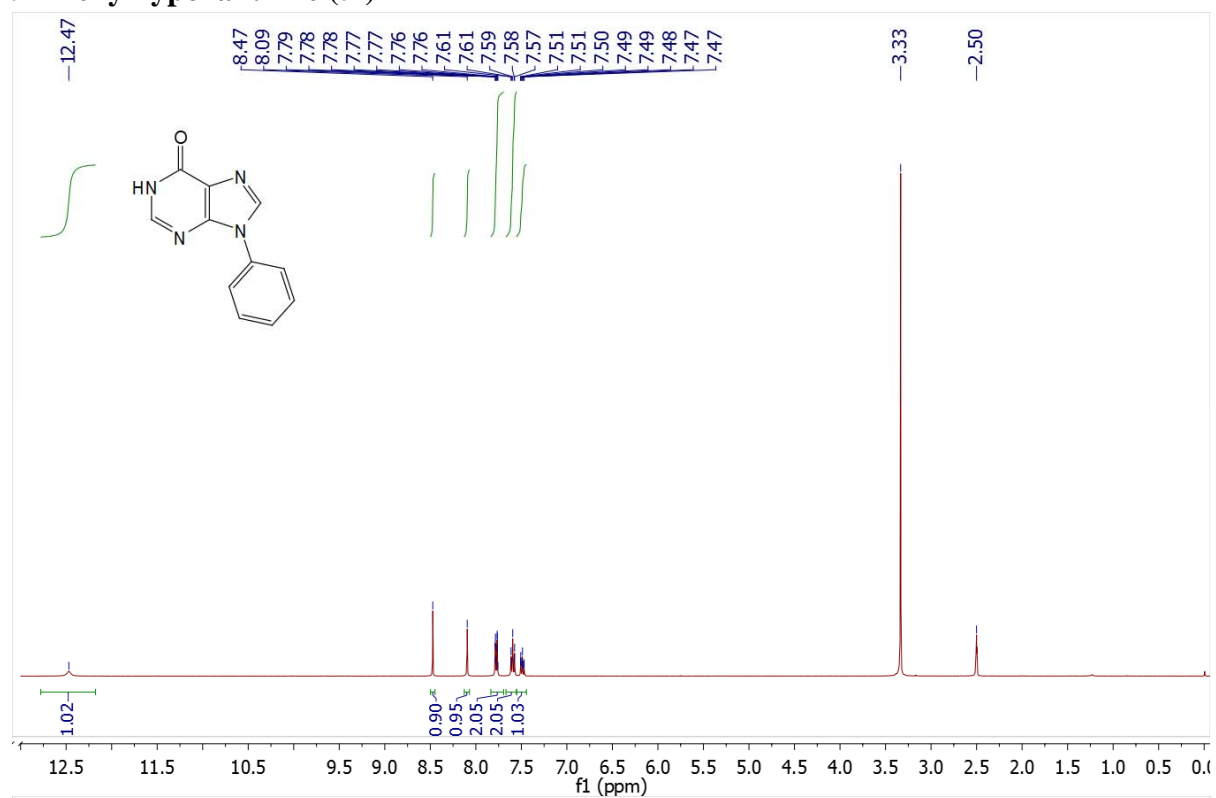
9-Phenylpurine-2,6-diamine (5b)



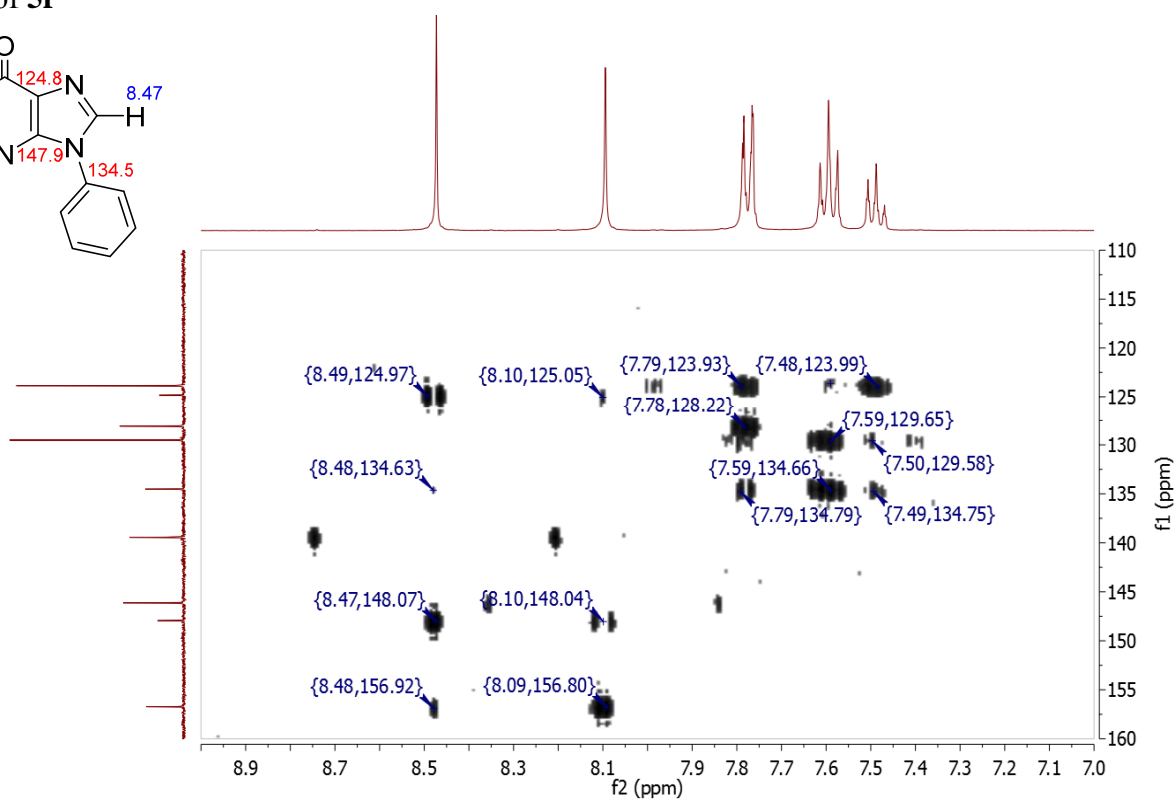
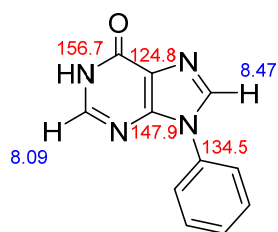
9-Phenylguanine (5e)



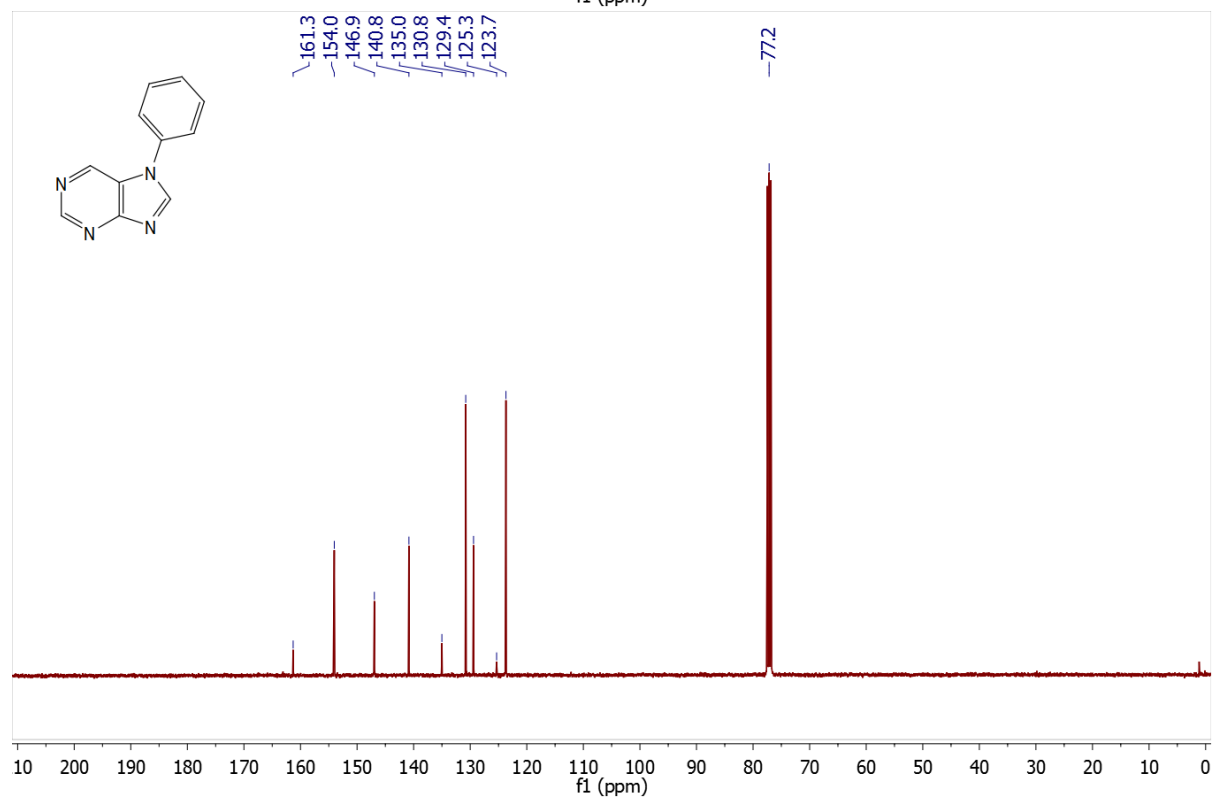
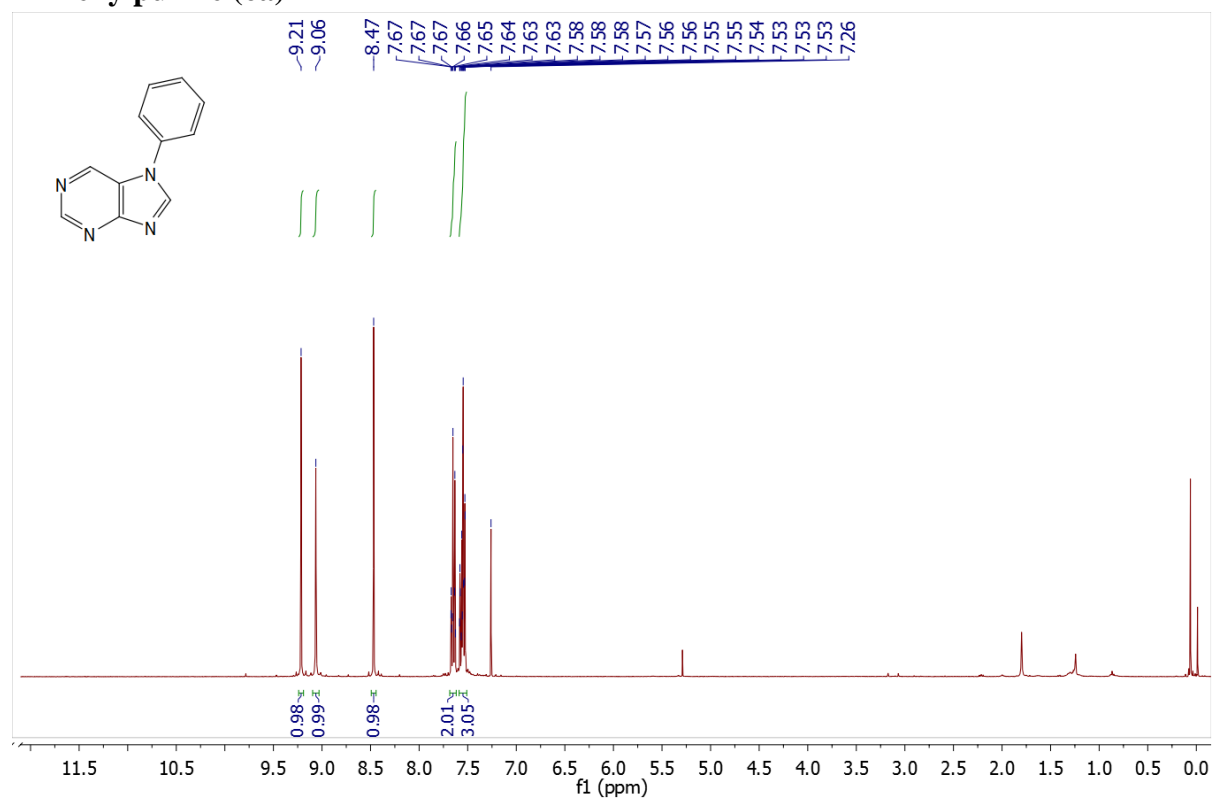
9-Phenylhypoxanthine (5f)



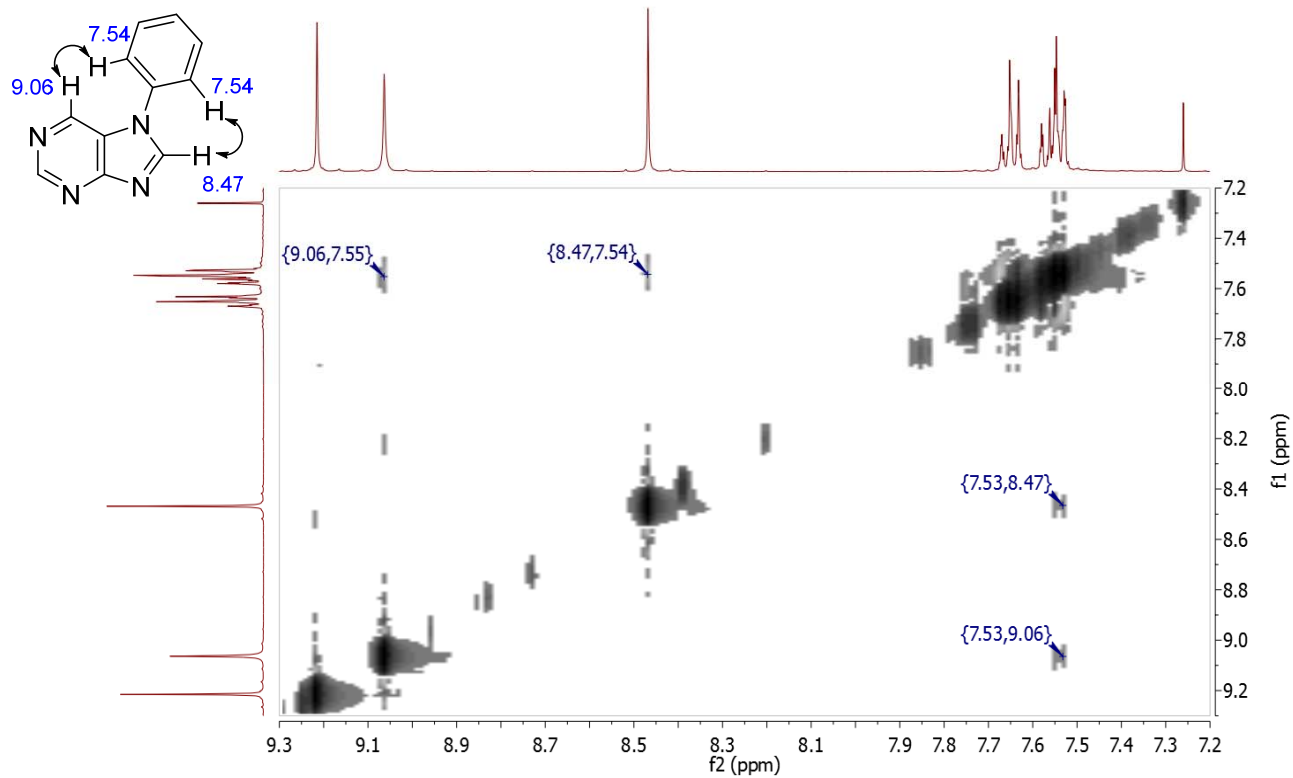
HMBC of **5f**



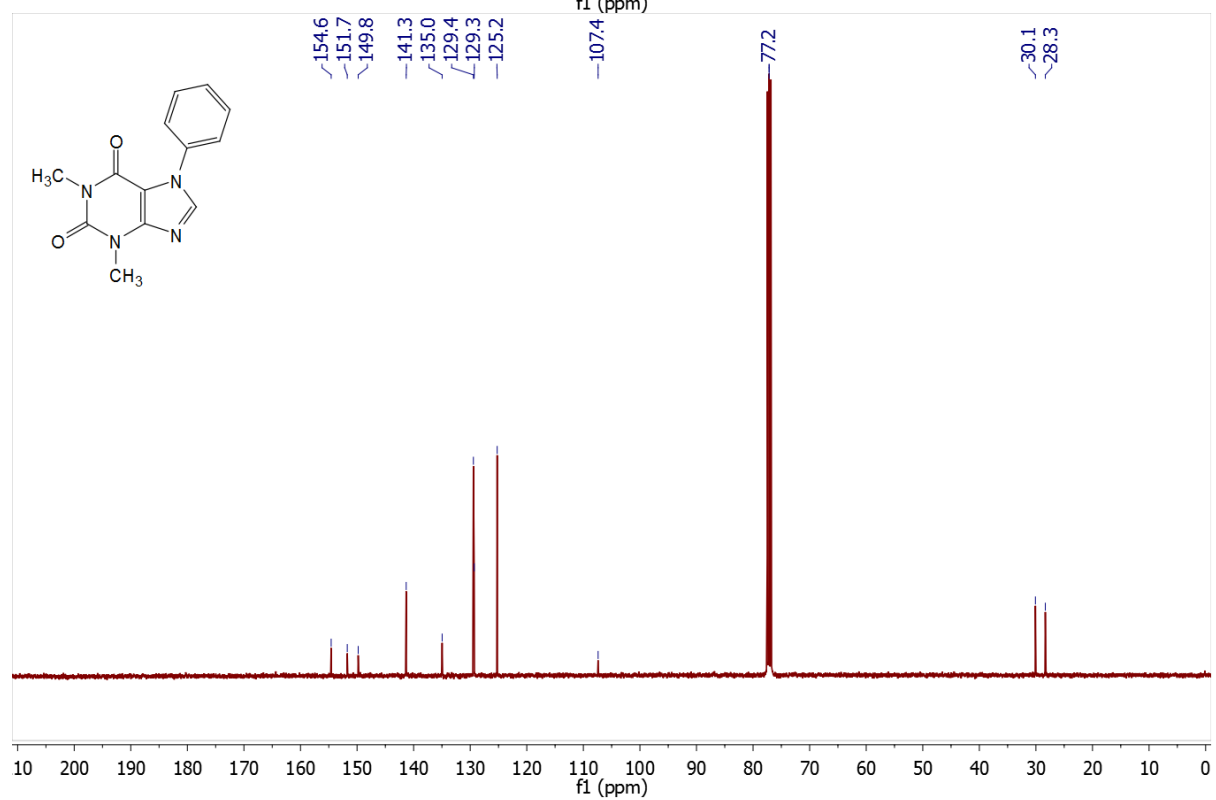
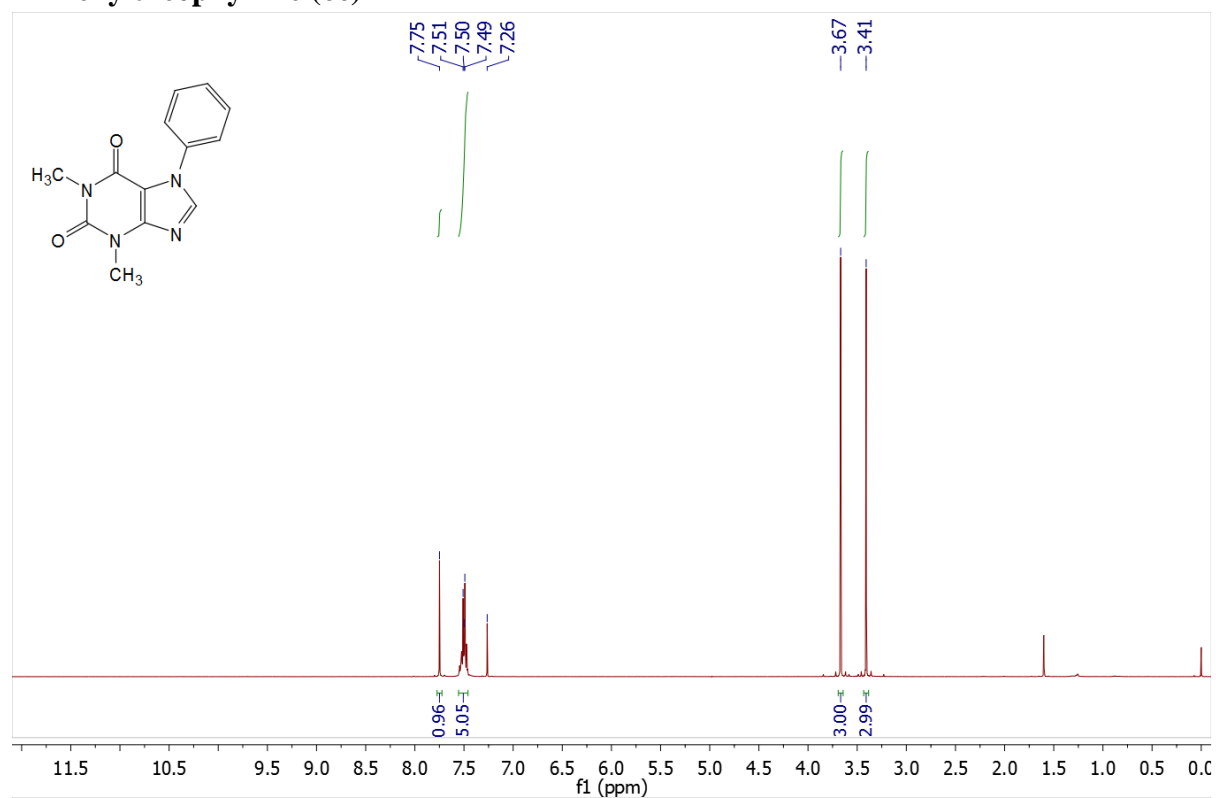
7-Phenylpurine (6a)



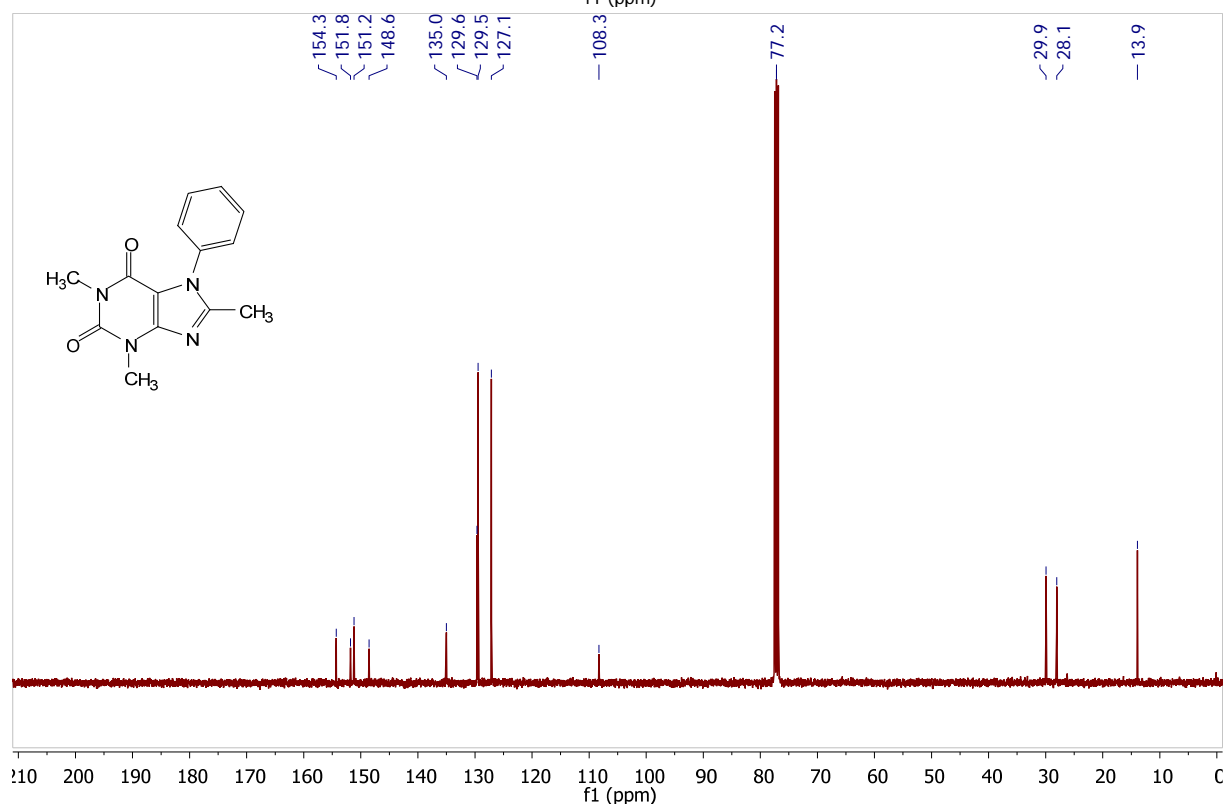
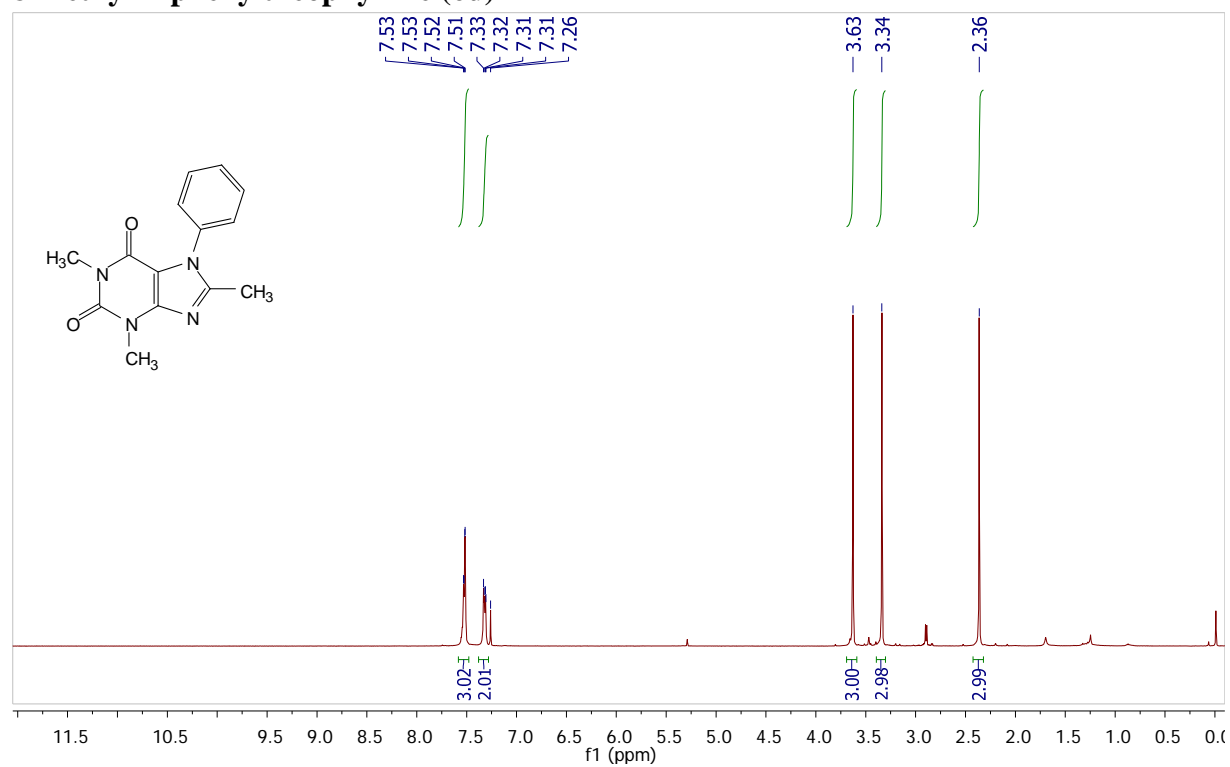
NOESY of **6a**



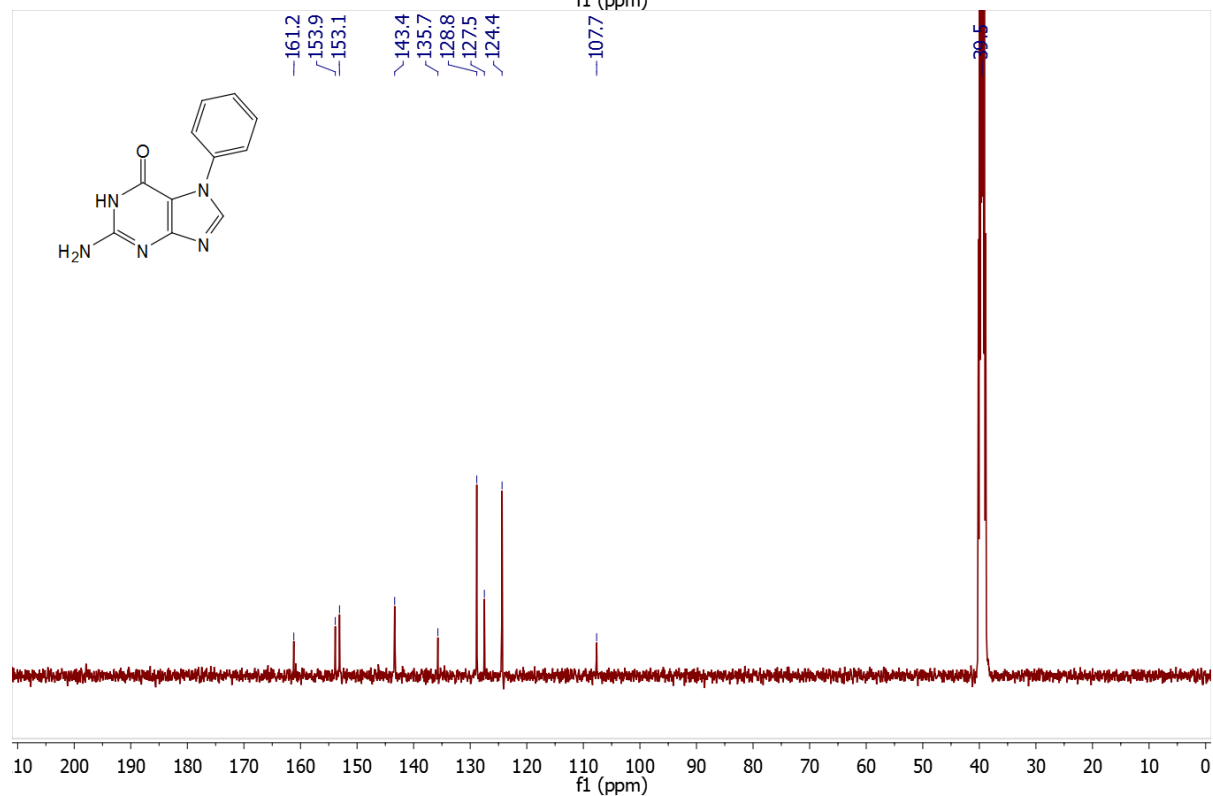
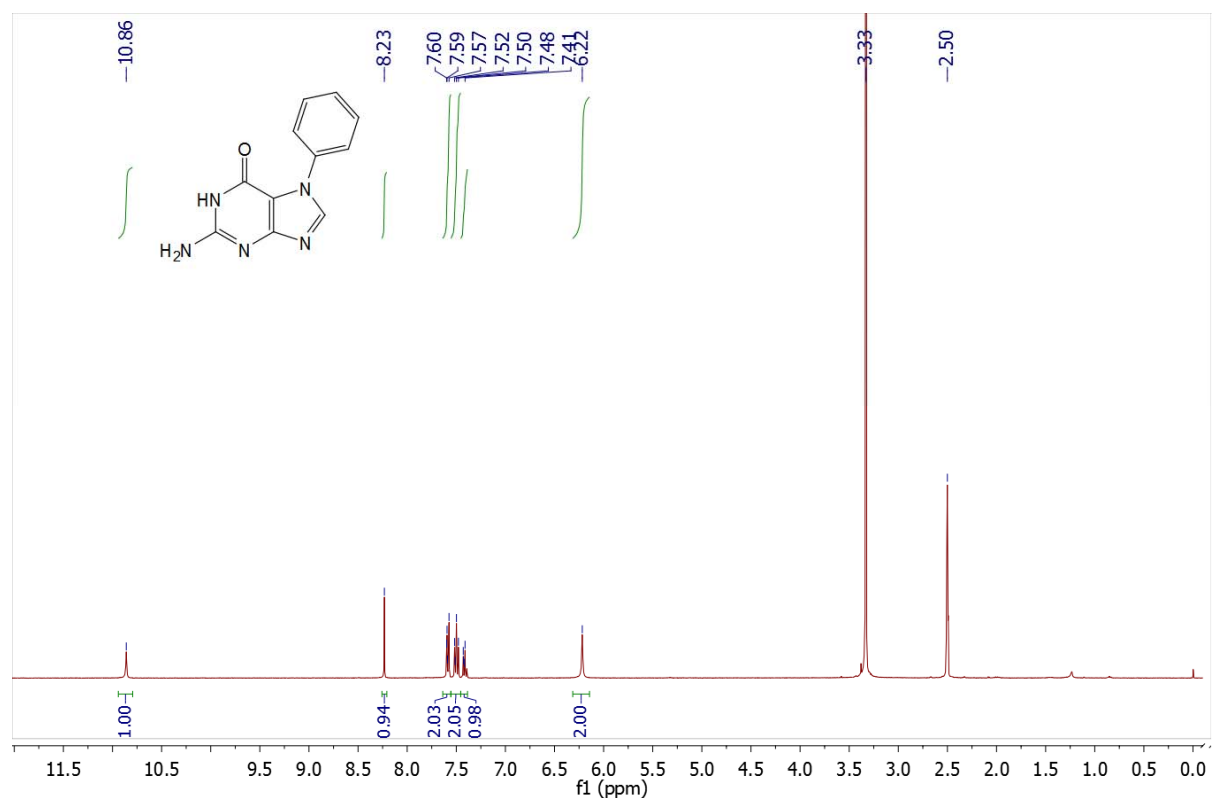
7-Phenyltheophylline (6c)



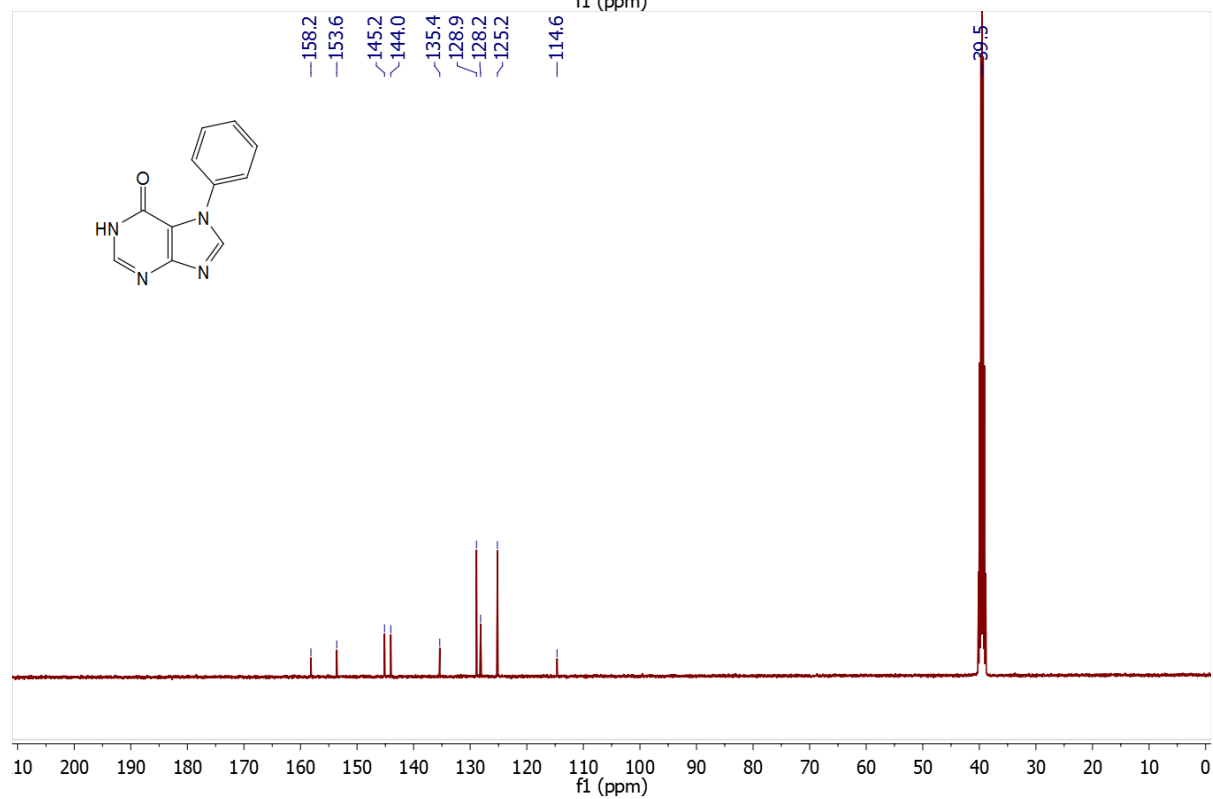
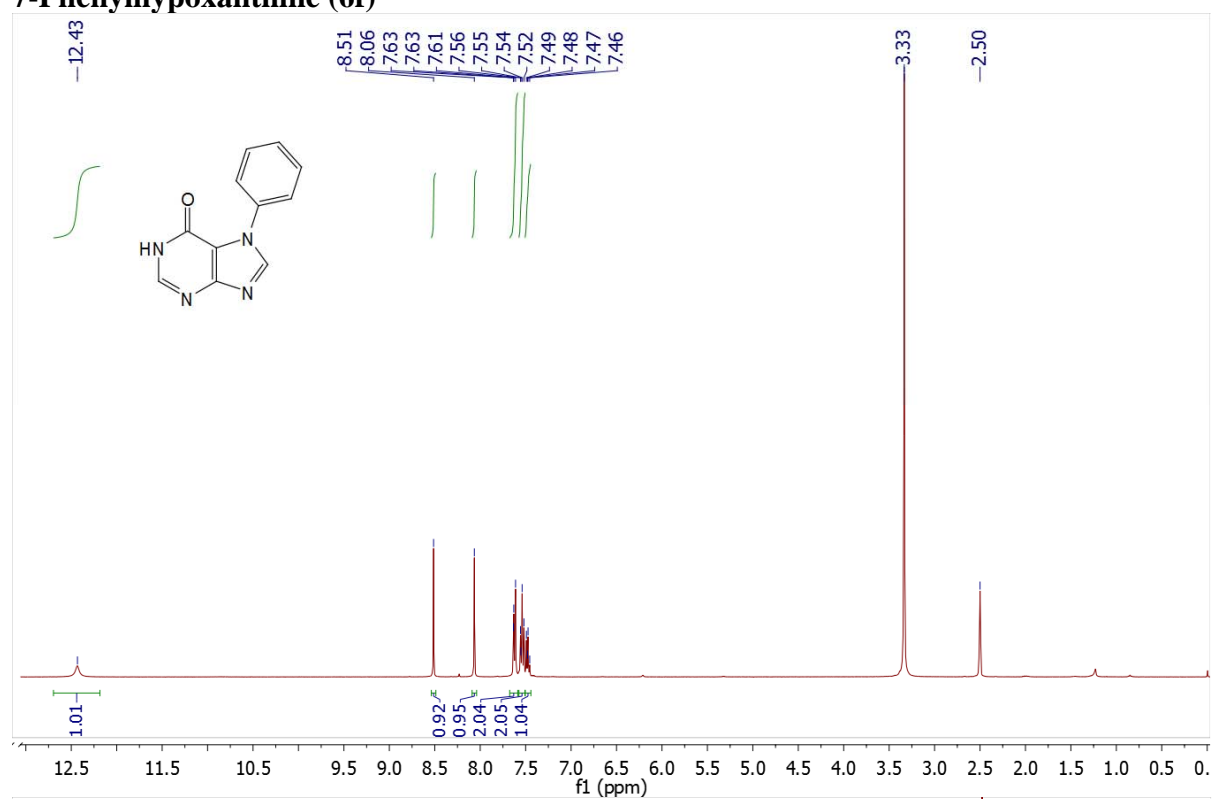
8-methyl-7-phenyltheophylline (6d)



7-phenylguanine (6e)



7-Phenylhypoxanthine (6f)



HMBC of **6f**

