

## S1: Mass spectrometer and DESI ion source parameters

Exactive mass spectrometer (Thermo Fisher Scientific Inc., Bremen, Germany)

Parameter	Setting
Injection time	1000 ms
Micro-scans	1
Mass Analyser	Fourier transform mass spectrometer
Mass resolution	100,000
Mass range	150-1000
Capillary temperature	250°C
Tube lens voltage	-150V
Skimmer Voltage	-40V

Geometric and electro-solvent parameters for home built DESI ion source

Parameter	Setting
Distance from sprayer tip to sample surface	2mm
Distance from sprayer tip to MS inlet capillary	14mm
Collection angle of MS inlet capillary	10°
Distance of MS inlet capillary from sample surface	<<1mm
Solvent constitution	Methanol:water (90:10 v/v)
Flow rate of solvent	1.5µl/min
Nebulising gas flow rate	7 Bar
Spray Voltage	5 kV

## S2: MSI data pre-processing

The MSI data was initially subjected to pre-processing which included peak detection, filtering of solvent/noise related peaks and variance stabilizing normalization with an in-house bio-informatics platform appointed on MATLAB (MathWorks). The molecular ion peaks within a  $m/z$  range smaller than the native accuracy of the mass spectrometer (<4ppm in this case) were assigned to the same molecular ion species uniformly for all pixels on a tissue section. The noise related  $m/z$  species were then found to be randomly located at <1% of pixels based on the distribution of molecular ion features across pixels. Additionally,  $m/z$  species were deemed to be of solvent-related origin if their mean peak intensity within tissue object pixels was less than their mean background intensity. All identified noise and solvent molecular ion peaks were removed from subsequent analysis.

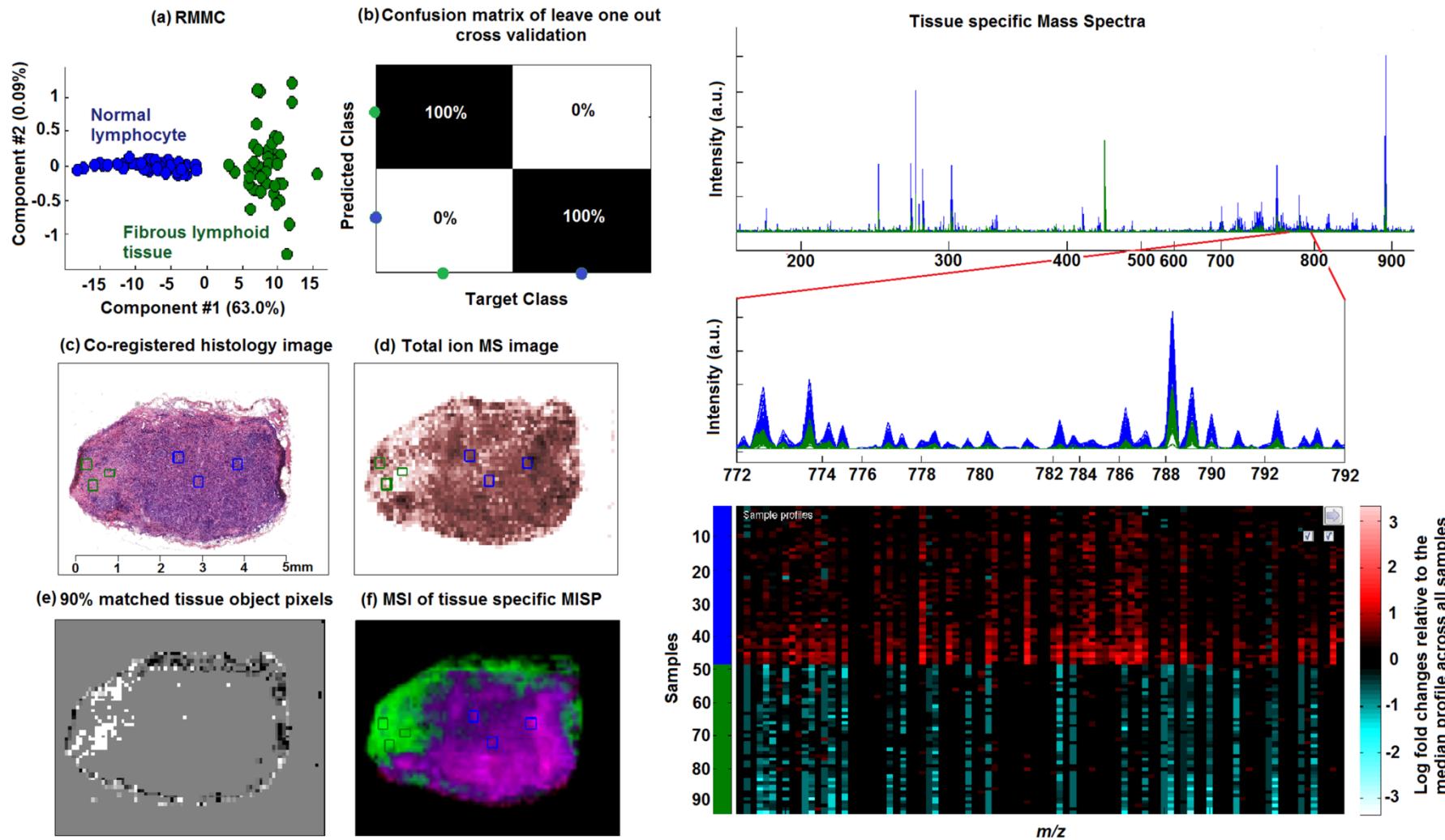
The combination of these processing steps resulted in a significant reduction of spectral data volume from >250,000 to approximately 1,000-5,000  $m/z$  features, thus enabling more efficient data handling/mining and pixel-wise tissue classification. Log-based transformation was subsequently applied to stabilise the increased variance of peak intensity as a function of increased signal intensity ("heteroscedastic noise structure").<sup>1</sup> This ensured that the data structure is consistent with the downstream application of multivariate statistical tools.

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<sup>1</sup>K. A. Veselkov, L. K. Vingara, P. Masson, S. L. Robinette, E. Want, J. V. Li, R. H. Barton, C. Boursier-Neyret, B. Walther, T. M. Ebbels, I. Pelczer, E. Holmes, J. C. Lindon, J. K. Nicholson, *Anal. Chem.*, 2011, 83(15), 5864-72.

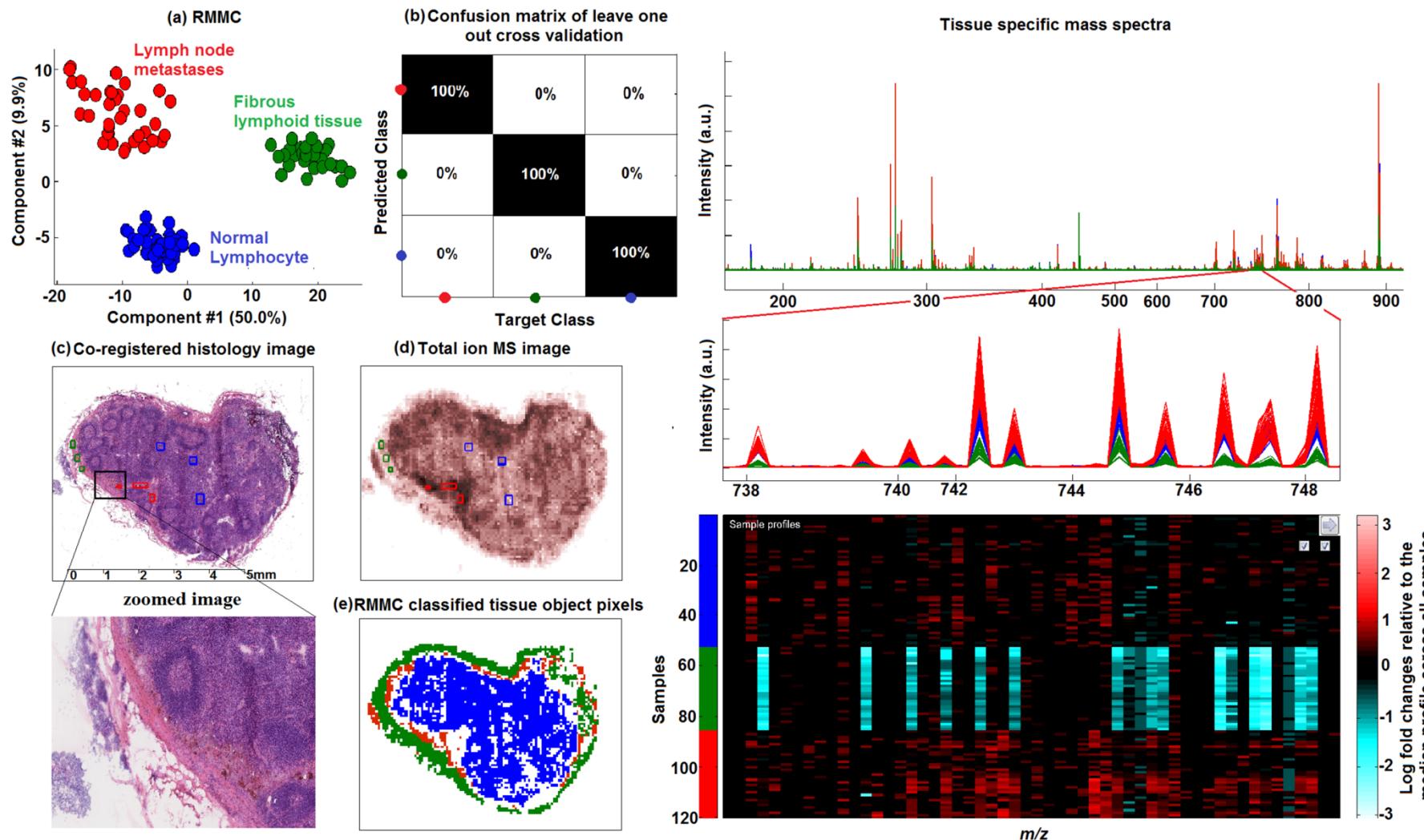
### S3: DESI-MSI data interrogation for a normal lymph node with no evidence of metastases

The blue and green regions highlighted in the histology image of the normal lymph node (Fig. c) represent normal lymphocyte and fibrous lymphoid tissue, respectively. This colour coding theme is representative of these tissue specific regions throughout the different sections of the whole figure. Figure 1 RMMC, Recursive Maximum Margin Criterium; MS, Mass Spectrometry; MSI, Mass Spectrometry Image; MISP, Molecular Ion Spectral Pattern.

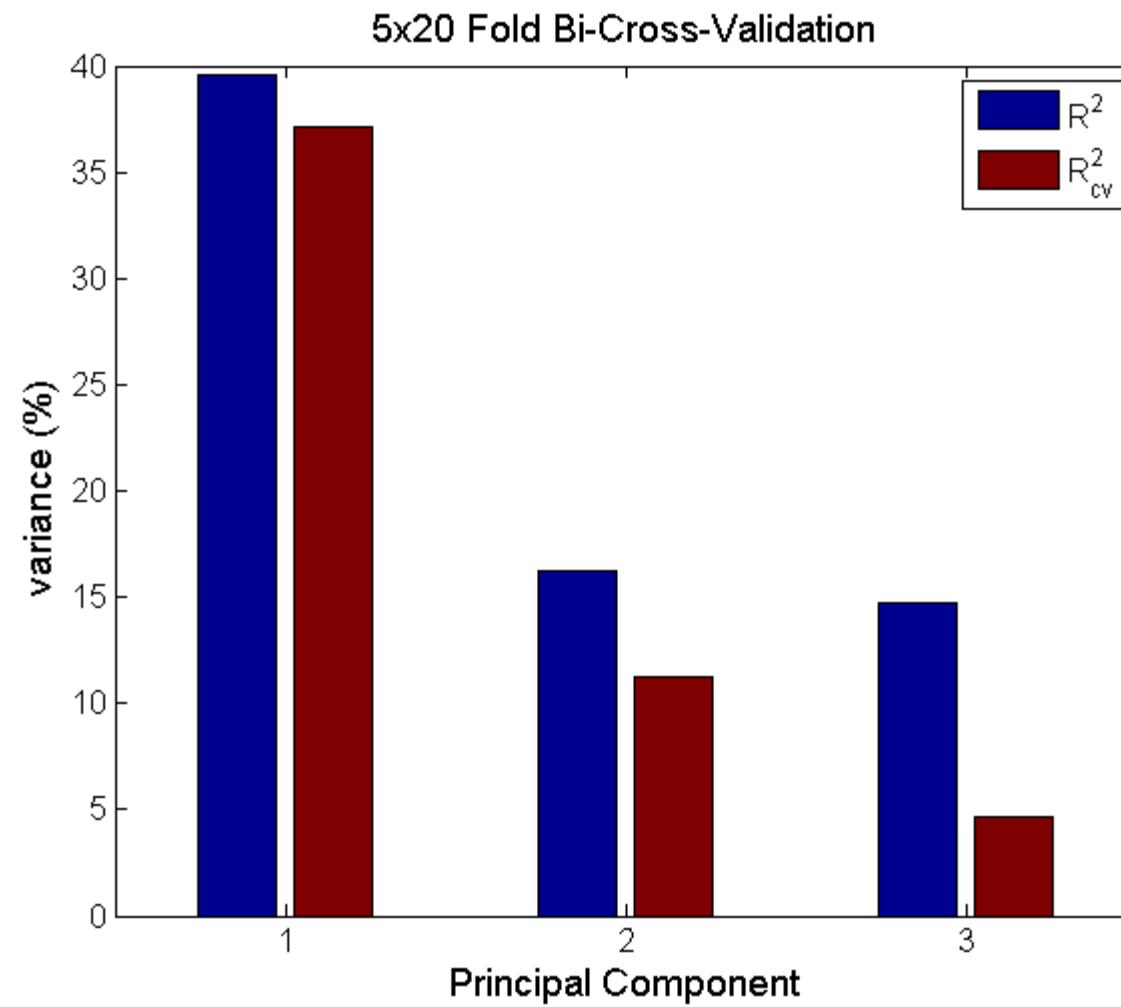


**S4 DESI-MSI data interrogation for a lymph node with evidence of micro-metastases**

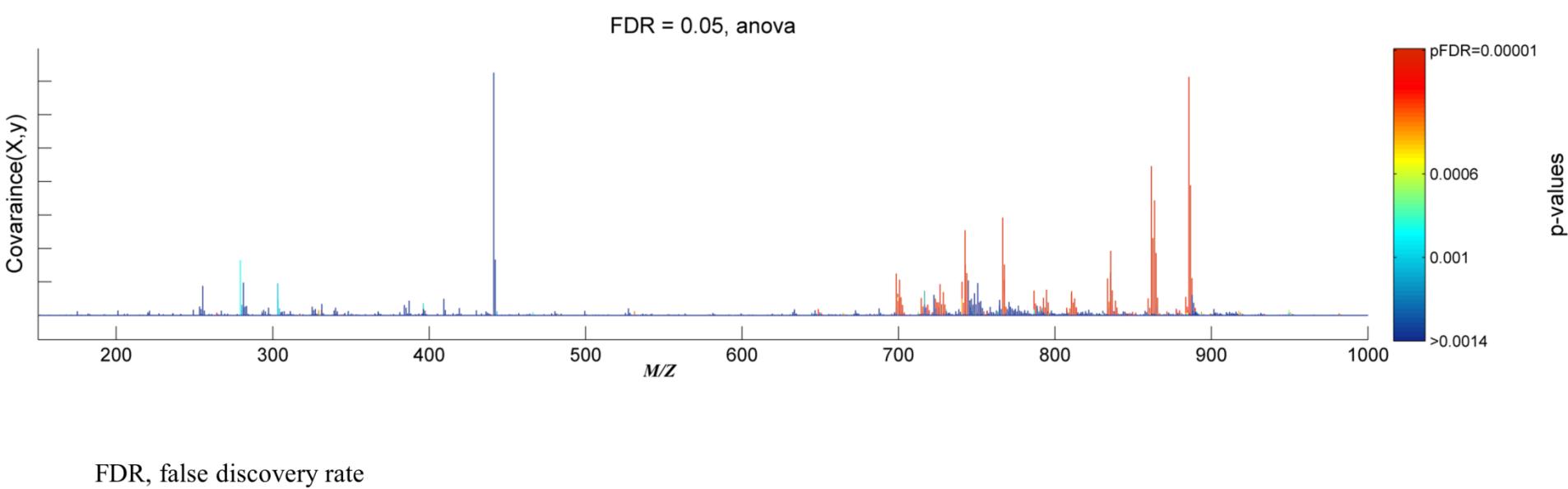
The blue, green and red regions highlighted in the histology image of the lymph node (Figure 3c), represent normal lymphocyte, fibrous lymphocyte tissue and lymph node micro-metastases, respectively. The colour coding theme is representative of these tissue specific regions throughout the different sections of the figure. The zoomed image shows the areas of micro-metastases highlighted by the brown staining of the AE1/AE1 anti-cytokeratin antibody. RMMC, Recursive Maximum Margin Criterium; MS, Mass Spectrometry; MSI, Mass Spectrometry Image; MISP, Molecular Ion Spectral Pattern.



S5: Bi-Cross Validation of PCA in Fig. 2

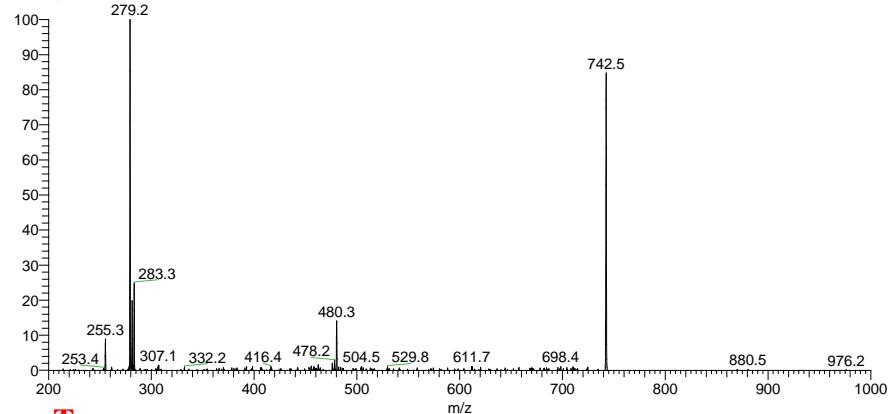


**S6: Discriminating features of comparative mass spectra of tissue specific regions (Figure 2c, main text) as a function of ANOVA based analysis of identified peaks**



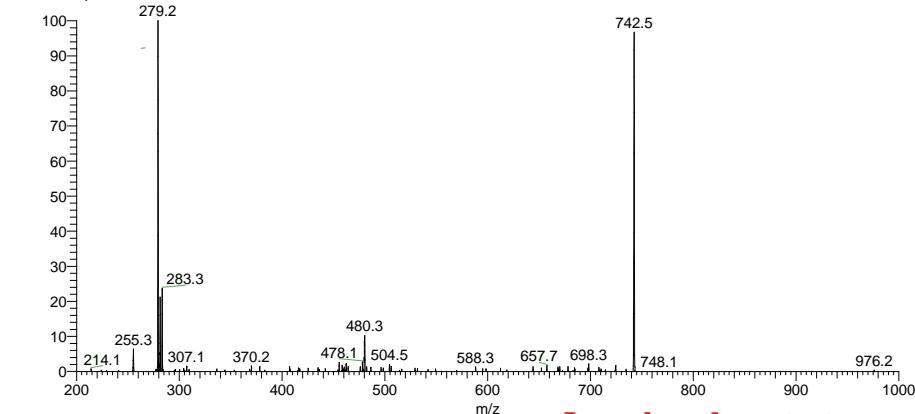
**Fig. S7a Direct tissue MS/MS spectra of *m/z* 742.54 [Phosphatidylethanolamine (18:2/18:0)-H] from tissue specific regions**

MSMS\_742p54\_Tumor\_131031122422 #22-149 RT: 0.43-3.03 AV: 128 NL: 8.01E-1  
T: ITMS - p NSI Full ms2 742.54@cid25.0



Tumour

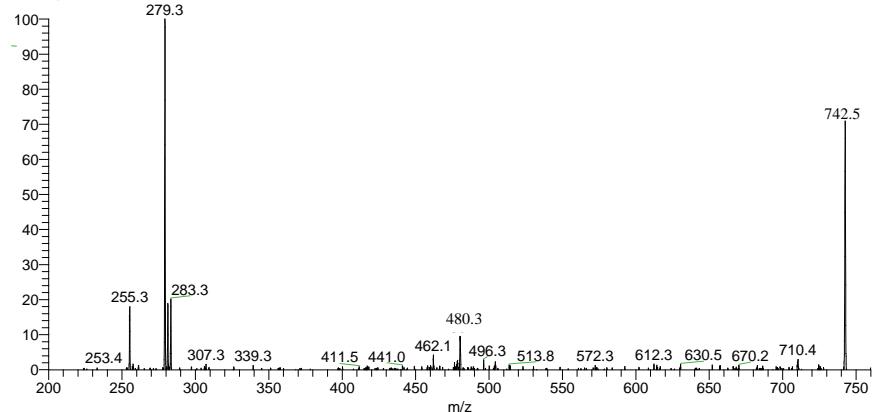
MSMS\_742p54\_LN3\_4\_131031122422 #25-75 RT: 0.49-1.52 AV: 51 NL: 9.30E-1  
T: ITMS - p NSI Full ms2 742.54@cid20.0



Lymph node metastases

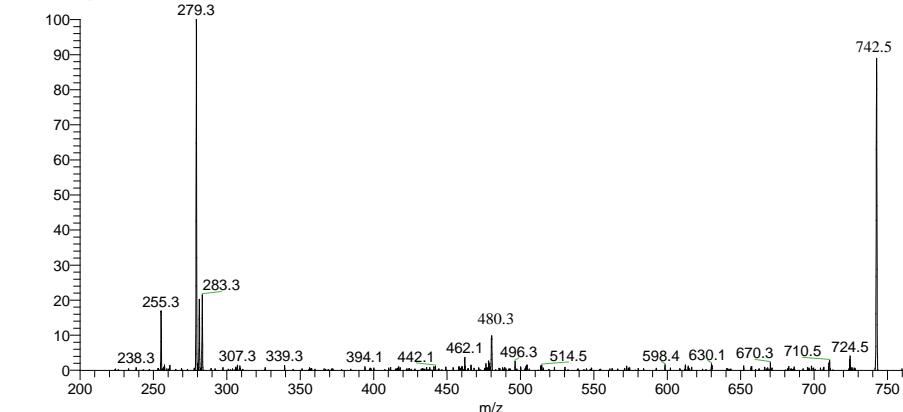
m/z ratio	Ion Description
742.54	Precursor ion [M-H] <sup>-</sup>
480.31	Loss of sn1 acyl chain as ketene (RCH=C=O) from [M-H] <sup>-</sup>
279.23	sn1 RCOO- ion
283.26	sn2 RCOO- ion

MSMS\_742p54\_nl\_131031122422 #28-50 RT: 0.53-0.97 AV: 23 NL: 6.28  
T: ITMS - p NSI Full ms2 742.53@cid20.0



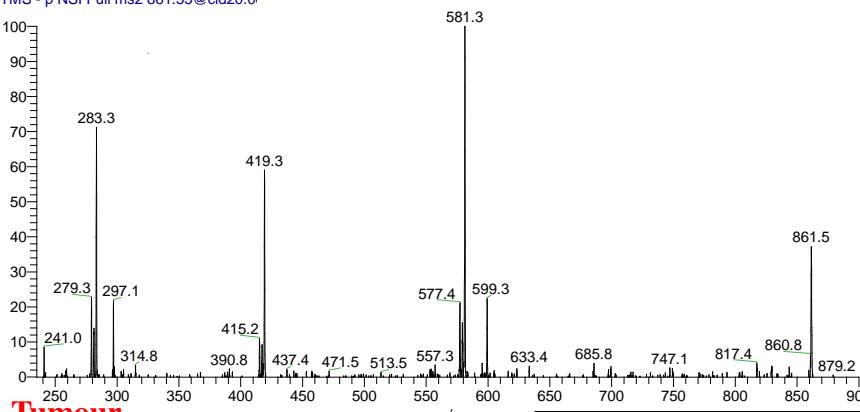
Normal lymphocytes

MSMS\_742p54\_fl\_131031122422 #29-66 RT: 0.55-1.28 AV: 38 NL: 4.10  
T: ITMS - p NSI Full ms2 742.53@cid20.0



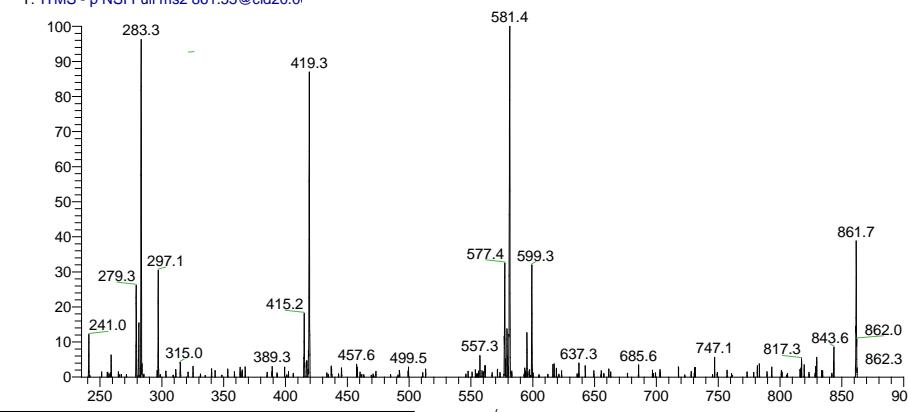
Fibrous lymphoid tissue

MSMS\_861p55\_131031122422 #49-119 RT: 0.96-2.36 AV: 71 NL: 1.28  
T: ITMS - p NSI Full ms2 861.55@cid20.0



Tumour

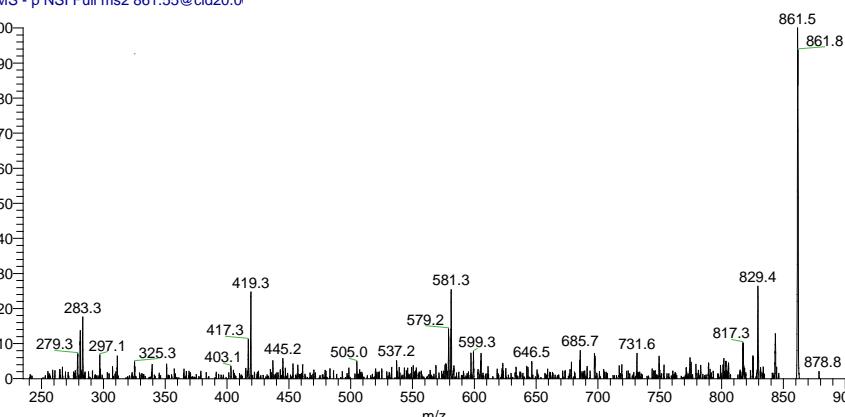
MSMS\_861p55\_LN3\_4\_131031122422 #91-139 RT: 1.80-2.76 AV: 49 NL: 8.61E-1  
T: ITMS - p NSI Full ms2 861.55@cid20.0



Lymph node metastases

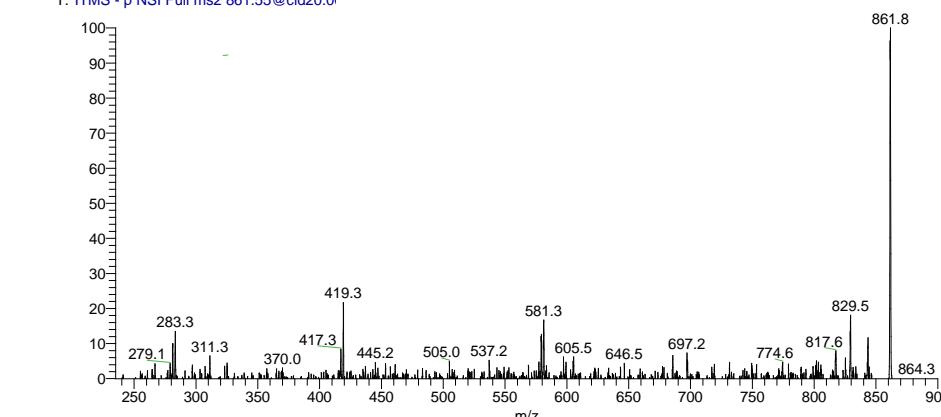
m/z ratio	Ion Description
861.55	Precursor ion [M-H] <sup>-</sup>
599.32	Loss of sn1 acyl chain as ketene (RCH=C=O) from [M-H] <sup>-</sup>
581.31	Neutral loss of sn1 RCOOH group from [M-H] <sup>-</sup>
577.28	Neutral loss of sn2 RCOOH group from [M-H] <sup>-</sup>
419.26	Neutral loss of sn1 RCOOH group and inositol from [M-H] <sup>-</sup>
415.23	Neutral loss of sn2 RCOOH group and inositol from [M-H] <sup>-</sup>
297.04	Glycerophosphoinositol -2H <sub>2</sub> O
283.26	sn2 RCOO- ion
279.23	sn1 RCOO- ion

MSMS\_861p55\_nl\_131113110640 #78-205 RT: 1.56-4.13 AV: 128 NL: 3.29E-1  
T: ITMS - p NSI Full ms2 861.55@cid20.0



Normal lymphocytes

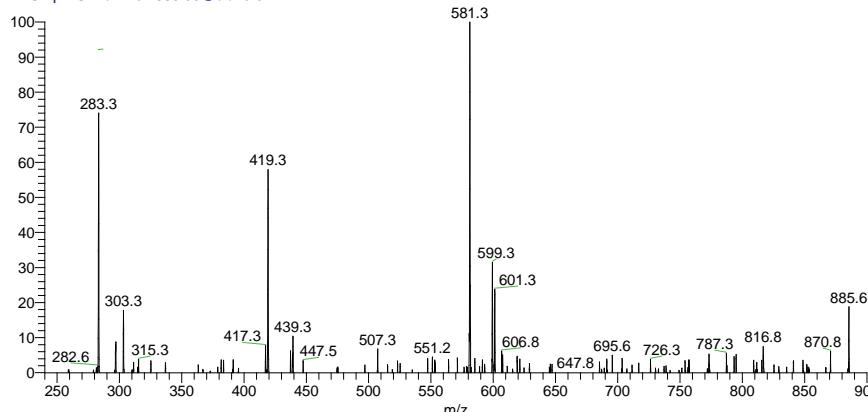
MSMS\_861p55\_fl\_131113110640 #69-171\_RT: 1.38-3.44 AV: 103 NL: 4.01E-1  
T: ITMS - p NSI Full ms2 861.55@cid20.0



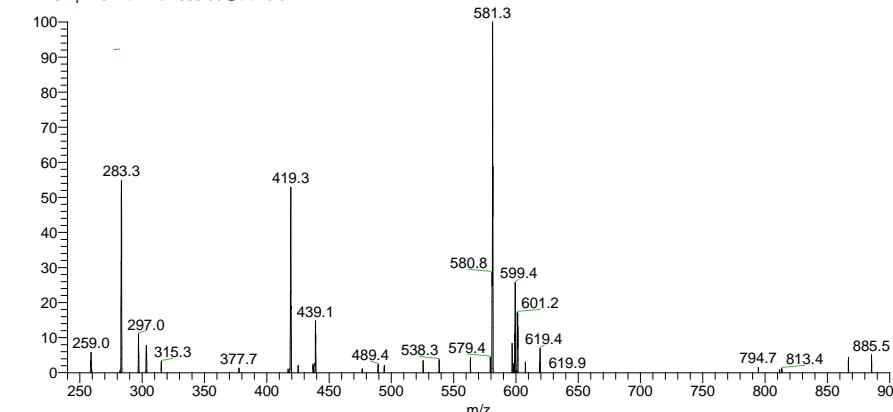
Fibrous lymphoid tissue

**Fig. S7c Direct Tissue DESI-MS/MS Spectra of  $m/z$  885.55 [Phosphatidylinositol (18:0/20:4)-H] from tissue specific regions**

MSMS885\_131024155248 #16-47 RT: 0.27-2.02 AV: 32 NL: 1.99E-1  
T: ITMS - p NSI Full ms2 885.55@cid20.0

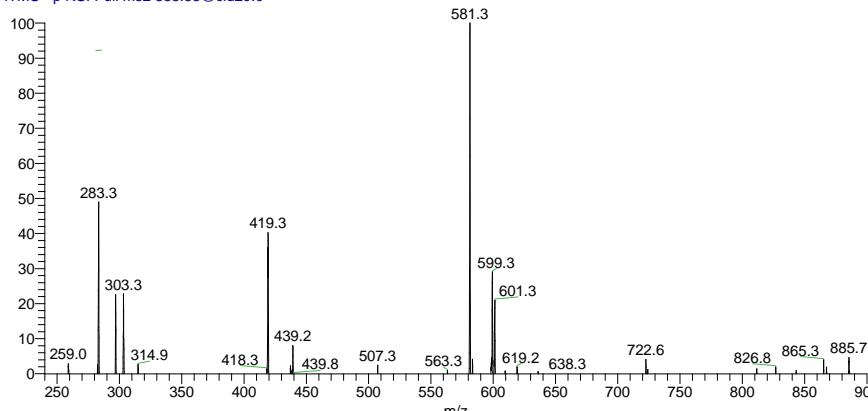
**Tumour**

LN3\_8nl\_885\_131024155248 #22-30 RT: 1.20-1.65 AV: 9 NL: 5.61E-1  
T: ITMS - p NSI Full ms2 885.55@cid20.0

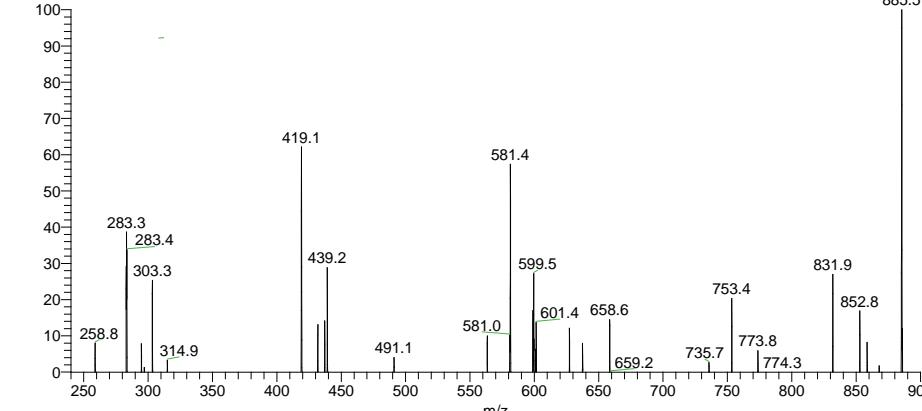
**Lymph node metastases**

m/z ratio	Ion Description
885.55	Precursor ion [M-H]-
599.32	Loss of sn2 acyl chain as ketene ( $\text{RCH}=\text{C=O}$ ) from [M-H]-
581.31	Neutral loss of sn2 RCOOH group from [M-H]-
419.26	Neutral loss of sn2 RCOOH group and inositol from [M-H]-
303.23	sn2 RCOO- ion
283.26	sn1 RCOO- ion

LN3\_8fl\_885\_131024155248 #7-16 RT: 0.34-0.86 AV: 10 NL: 7.20E-1  
T: ITMS - p NSI Full ms2 885.55@cid20.0

**Normal lymphocytes**

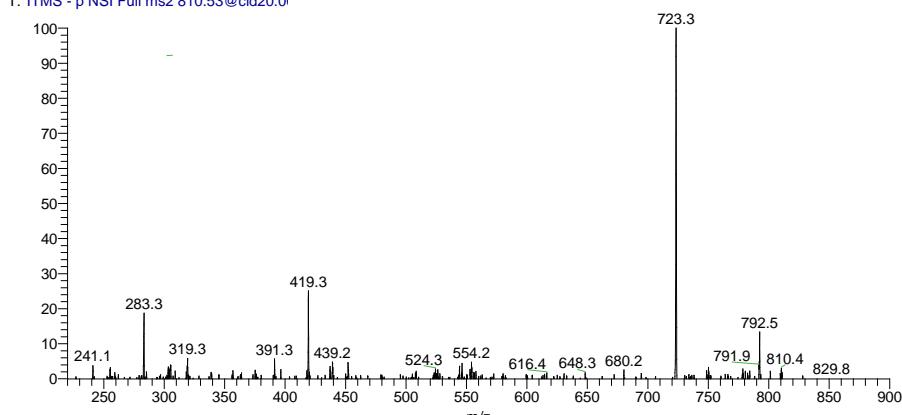
LN3\_8fl\_885\_131024155248 #27-39 RT: 0.50-0.77 AV: 13 NL: 2.11E-1  
T: ITMS - p NSI Full ms2 885.55@cid20.0

**Fibrous lymphoid tissue**

**Fig. S7d Direct tissue DESI-MS/MS Spectra of  $m/z$  810.53 [Phosphatidylserine (20:4/18:0)-H] from tissue specific regions**

MSMS810\_1D\_oldtumor\_131024155248 #9-40 RT: 0.42-2.06 AV: 32 NL: 3.19E-1  
 T: ITMS - p NSI Full ms2 810.53@cid20.0

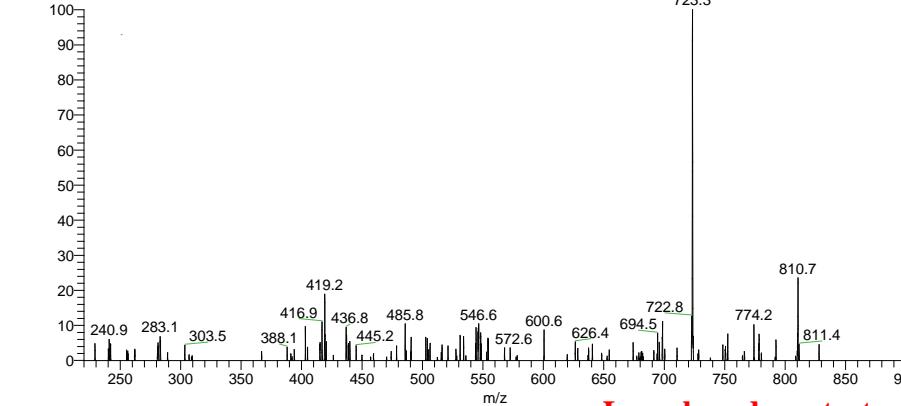
**PS - 20:4/18:0**



**Tumour**

LN3\_4\_810\_second\_131024155248 #18-64 RT: 0.96-3.57 AV: 47 NL: 9.64E-2  
 T: ITMS - p NSI Full ms2 810.55@cid20.0

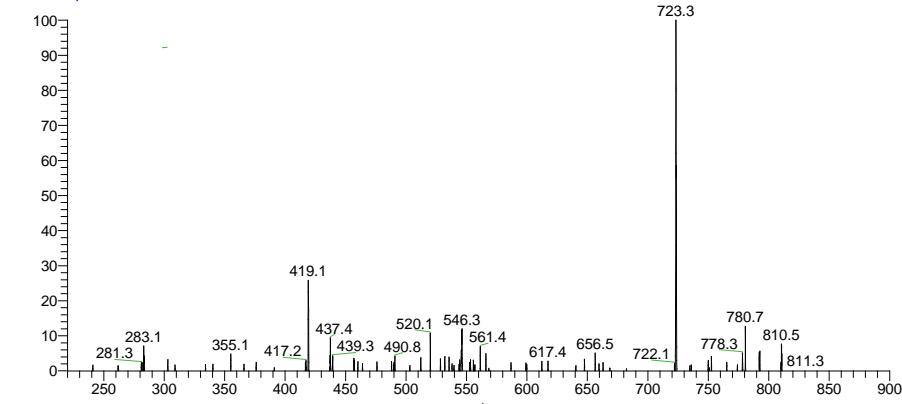
723.3



**Lymph node metastases**

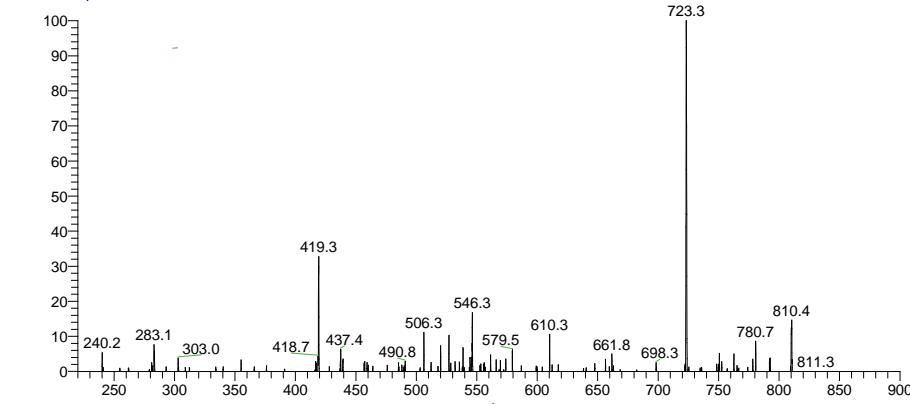
<b>m/z ratio</b>	<b>Ion Description</b>
810.53	Precursor ion [M-H] <sup>-</sup>
723.50	Loss of serine from precursor ion
419.26	Neutral loss of sn1 RCOOH group and serine from [M-H] <sup>-</sup>
283.26	sn2 RCOO- ion

LN3\_8nl\_810\_131024155248 #62-147 RT: 2.42-5.57 AV: 86 NL: 8.77E-2  
 T: ITMS - p NSI Full ms2 810.55@cid20.0



**Normal lymphocytes**

LN3\_8fl\_810\_1310241552482 #1-146 RT: 0.17-5.53 AV: 143 NL: 7.81E-2  
 T: ITMS - p NSI Full ms2 810.55@cid20.0



**Fibrous lymphoid tissue**