Supplementary Information

Selective monomethylation of primary amines with simple electrophiles

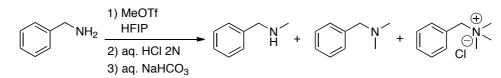
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General remarks

¹H NMR spectra were obtained on a Bruker 300. In all measurements CD₃OD was used as solvent unless otherwise noted. Chemical shifts δ are given in ppm relative to TMS as internal standard. Microflow reactions were performed with Harvard Apparatus syringe pumps (Pump 11 Elite) equipped with Hamilton gastight syringes. Peek (P-885) T-shaped micromixers with swept volume of 29 nL were manufactured by IDEX Health & Science. Peek (1532) microtubes with inner diameter of 500 µm and peek fittings were also purchased from IDEX Health & Science. All chemicals were used as provided without further purification. The conversion of amine into products was measured by ¹H NMR spectra directly from the crude product for benzylamine derivatives and for the other ones was added 3-bromotoluene as an internal standard.

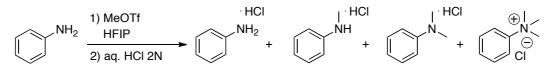
Experimental procedures

Typical procedure for the methylation of benzylamine

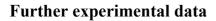


To a solution of HFIP (1.0 mL, 10 mmol) and benzylamine (109 μ L, 1 mmol) was added MeOTf (164 μ L, 1.5 mmol). The mixture was stirred for 1 h at room temperature and then quenched by a solution of HCl (2N, 1 mL). Volatiles were evaporated under reduced pressure, and the resulting mixture was neutralised with a saturated aqueous solution of NaHCO₃ and extracted with CH₂Cl₂ (3×5 mL). Combined organic phases were dried over MgSO₄, filtered and solvent was removed under reduced pressure. The crude product was analysed by ¹H NMR.

Typical procedure for the methylation of aniline



To a solution of HFIP (1.0 mL, 10 mmol) and aniline (91 μ L, 1 mmol) was added MeOTf (164 μ L, 1.5 mmol). The mixture was stirred for 1 h at room temperature and then quenched by a solution of HCl (2N, 1 mL). Volatiles were evaporated under reduced pressure. 3-Bromotoluene (123 μ L, 1 mmol) was added to the crude product and a ¹H NMR analysis was performed.



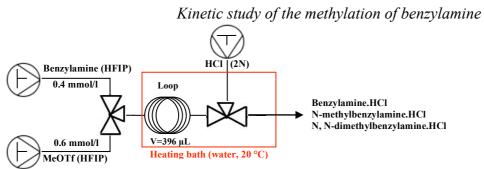


Figure S1: A flow microreactor for the methylation of benzylamine

Kinetic study was performed in a flow microreactor (Figure 1). The first syringe (3 mL) was filled with a mixture of benzylamine (330 μ L, 3.03 mmol) in HFIP (1.67 mL). The second syringe (3 mL) contained a mixture of MeOTf (495 μ L, 4.52 mmol) and HFIP (1.51 mL, 14.33 mmol). The reaction was quenched *in-situ* by a solution of aq. HCl (2N). Results of this study are shown below (Table S1 and Fig S2).

Table S1: Methylation of benzylamine with 1.5 equivalent of MeOTf in a flow microreactor

Time (min)	Benzylamine	N-methylbenzylamine	N, N-dimethylbenzylamine
0.5	70%	27%	3%
3	57%	41%	3%
7	41%	54%	5%
15	32%	62%	7%

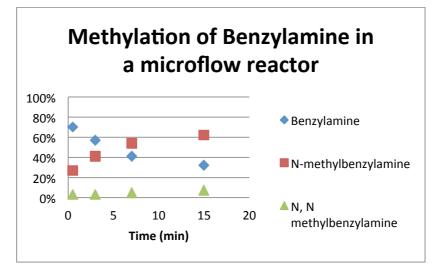


Figure S2: Methylation of benzylamine with 1.5 equivalent of MeOTf

Influence of HFIP on the methylating reagent

This study was performed to show the influence of HFIP on the methylating reagent. The variation of δ_{OH} (HFIP) in ¹H NMR according to the number of equivalents of methylating reagent was monitored.

Typical procedure

$$H_{O} \xrightarrow{CF_{3}}_{CF_{3}} + MeX \xrightarrow{XMe CF_{3}}_{H_{O}} \xrightarrow{K}_{CF_{3}}$$

HFIP (50 μ L, 475 μ mol) and 600 μ L of CDCl₃ were added into a NMR tube and a ¹H NMR spectrum was taken. The amount of methylating reagent was increased step by step (0, 0.5, 1, 1.5 and 2 eq) and each time a ¹H NMR spectrum was recorded (Fig S3).

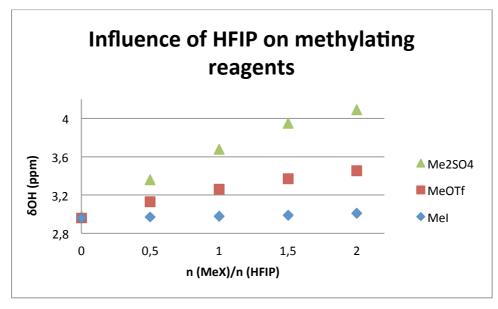


Figure S3 Δδ_{OH} according to MeX (MeOTf, Me₂SO₄ or MeI)

¹H NMR spectra

Amines are under hydrochloride salt form and the corresponding ¹H NMR spectra were recorded in CD₃OD, except where indicated.

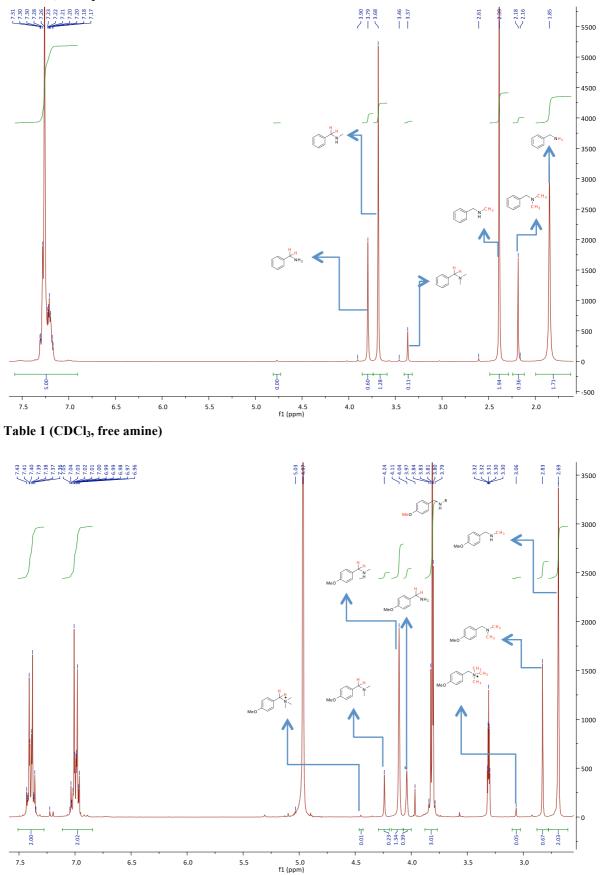


Table 2, entry 1

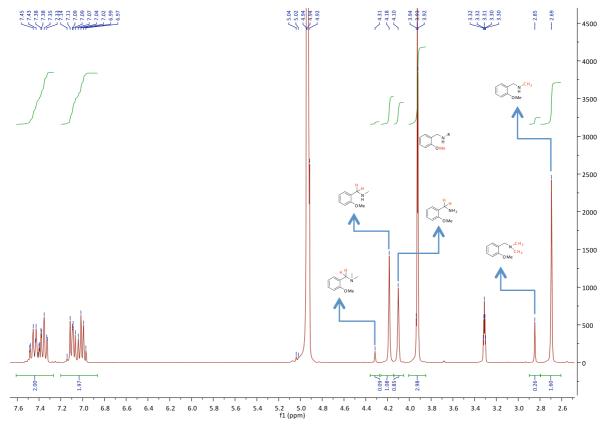


Table 2, entry 2

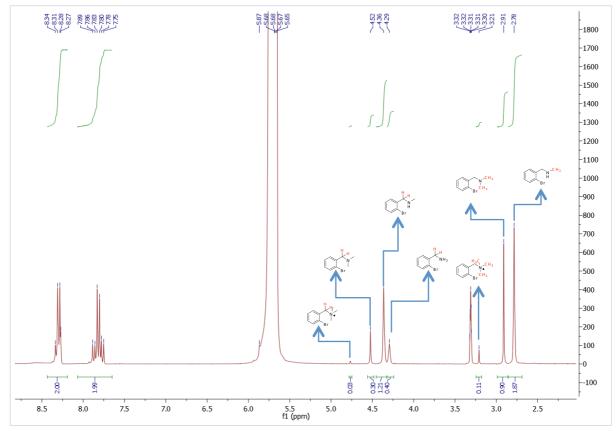
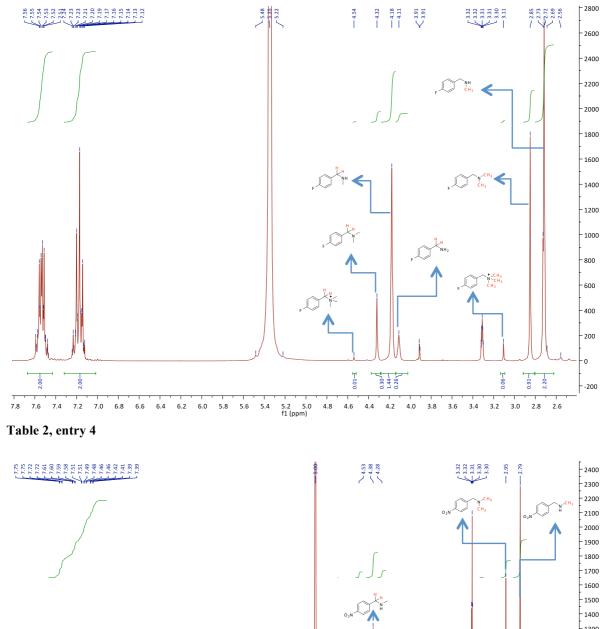


Table 2, entry 3



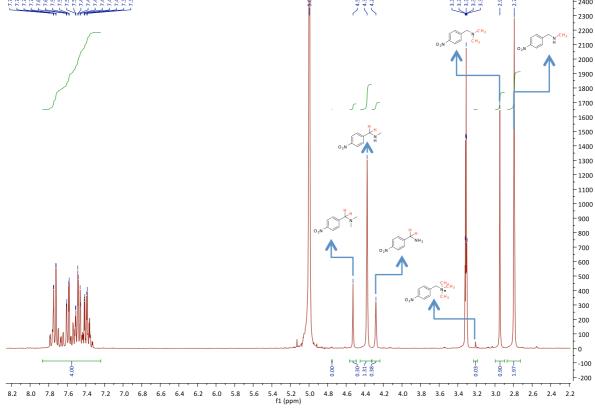


Table 2, entry 5

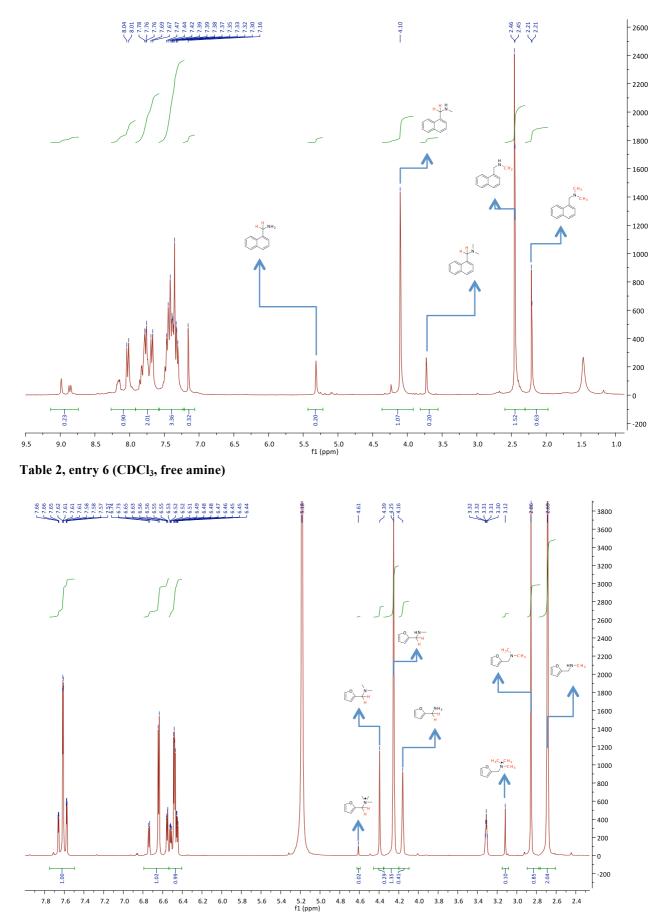
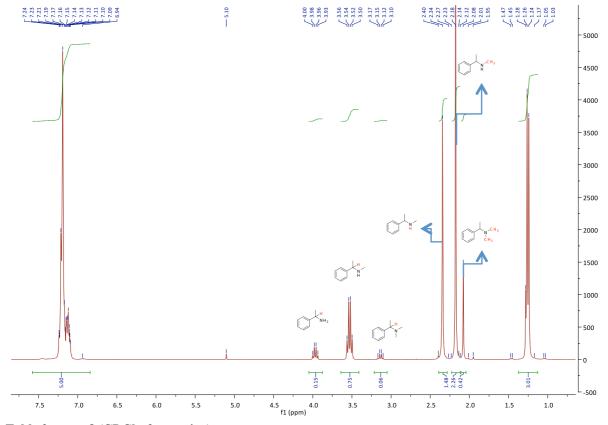


Table 2, entry 7

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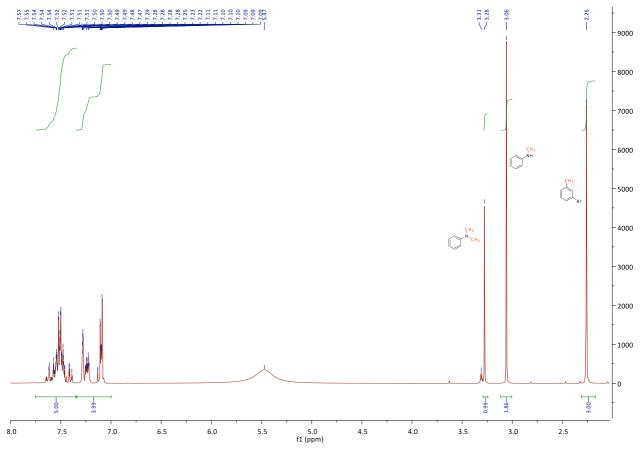


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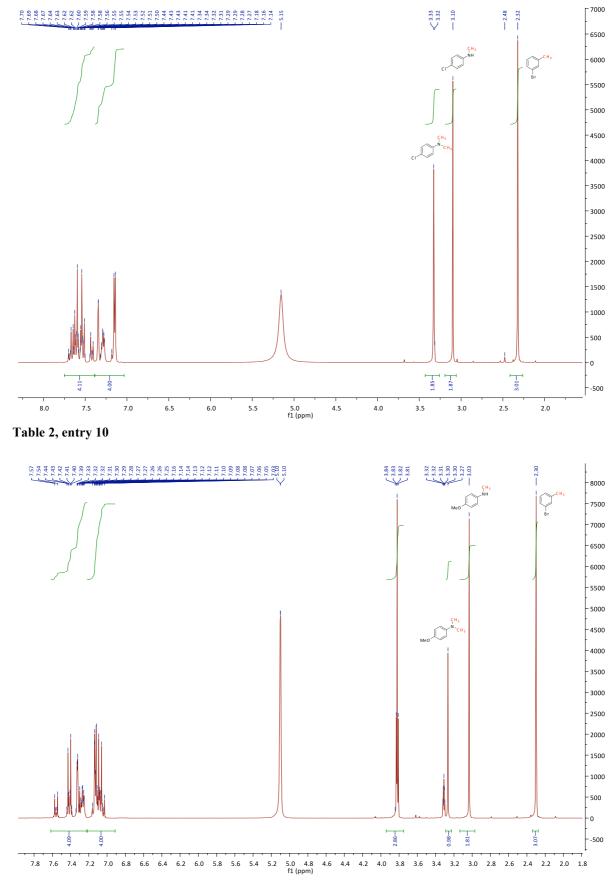


Table 2, entry 11

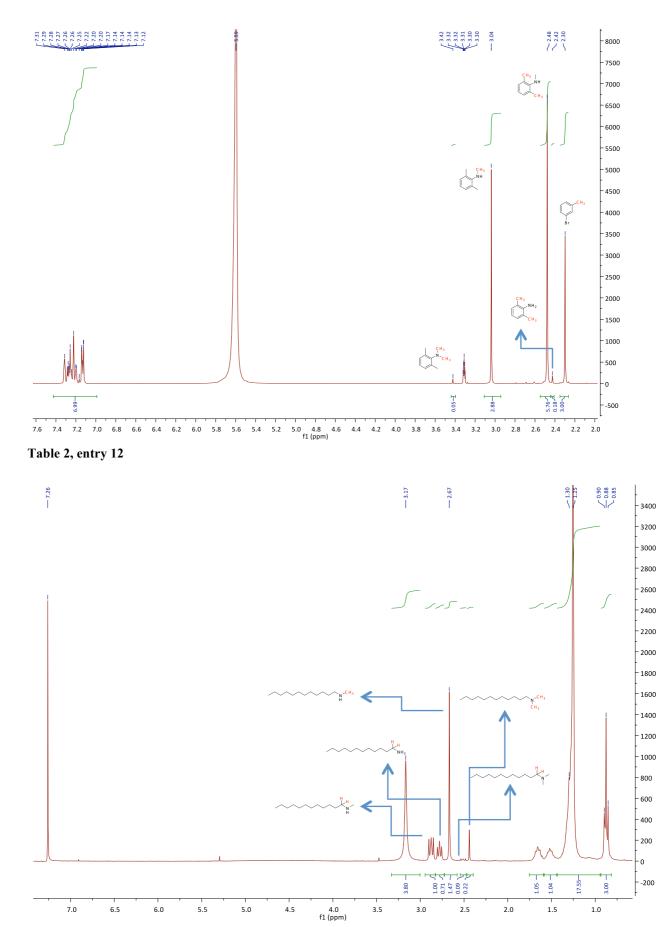


Table 2, entry 13 (CDCl₃, free amine)

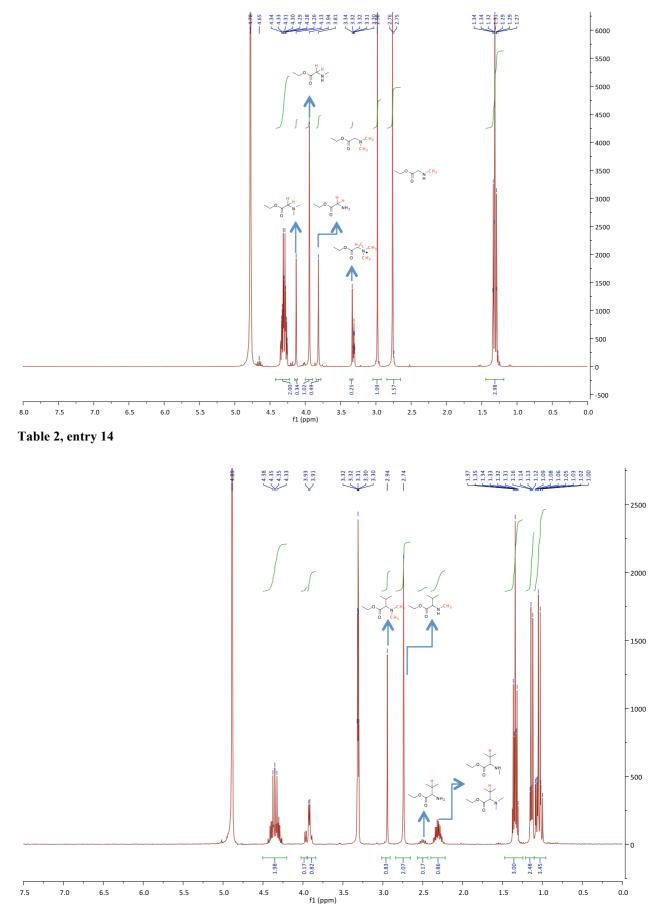


Table 2, entry 15

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