1 Electronic Supplementary Information (ESI) for Chemical Communications

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- <sup>3</sup> In situ generated AgBr-enhanced ZnO
- <sup>4</sup> nanorod-based photoelectrochemical
- <sup>5</sup> aptasensing via layer-by-layer assembly
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# 1 Experimental Section

2	Materials and Reagents. Indium tin oxide (resistivity 10 $\Omega$ /sq.) is obtained from
3	Zhuhai Kaivo Electronic Components Co. Ltd., China. Zn $(NO_3)_2 \cdot 6H_2O \ge 99.0\%$ ),
4	Mercury (II) chloride (HgCl <sub>2</sub> ), Poly dimethyl diallyl ammonium chloride (PDDA;
5	20 wt% in water, $M_w$ (200,000-350,000)) and tris-(hydroxymethyl)-aminomethane
6	were bought from Sigma-Aldrich (St. Louis, MO). KNO3, AgNO3, NaBr and other
7	chemicals were purchased from Sinopharm Chemical Co., Ltd. (Shanghai, China).
8	Oligonucleotides were obtained from Sangon Biotechnology Co., Ltd. (Shanghai,
9	China) and had the sequences as: 5'-CCC CCC CCC CCC CCC CCC-3' (poly (C)).
10	The stock solution of poly (C) was 100 $\mu$ mol L <sup>-1</sup> in 10 mmol L <sup>-1</sup> TE buffer solution
11	(pH 8.0) and kept frozen. 10 mmol L <sup>-1</sup> Tris-HCl (pH 7.4) was used to dilute the
12	stock solution when needed. The stock solution of $Ag^+$ was prepared from $AgNO_3$
13	with the concentration of 1 mmol $L^{-1}$ . In this work, 0.1 mol $L^{-1}$ phosphate-buffered
14	saline (PBS) was always employed as the supporting electrolyte and the pH value of
15	PBS was 7.0 unless indicated otherwise. Millipore ultrapure water (resistivity $\geq 18.2$
16	$M\Omega$ ·cm) was used throughout the experiment and all chemicals were of analytical
17	reagent grade. AgBr was obtained in the characterizations of UV-vis spectroscopy
18	and XRD as following: Equal volume of 10 mmol $L^{-1}$ AgNO <sub>3</sub> and 0.1 mol $L^{-1}$ NaBr
19	solutions were mixed and shaken for a few minutes, and then a yellow precipitate
20	emerged. The above mixture was further shaken to get a homogeneous emulsion,
21	and then 100 uL of the emulsion was dropped onto the ITO electrode (1 cm $\times$ 1 cm)
22	and dried at room temperature in the dark.

1	Apparatus. Digital Lux Meter (MS6612, Shenzhen Huayi Instrument Co. Ltd.,
2	China). Field Emission Scanning Electron Microscopy (FESEM) and Energy
3	dispersive spectrum (EDS) were performed with a JSM-5610LV/VANTAGE $\mathbf{IV}$
4	(JEOL, Japan and Thermo Electron, USA). X-ray diffraction (XRD) patterns of ZnO
5	nanorods, AgBr and the composite film of ZnO-PDDA-poly (C)-AgBr (shortened as
6	ZnO-AgBr) on ITO electrode (1 cm $\times$ 1 cm) were measured on the D/max
7	2500VL/PC diffractometer (RIGAKU, Japan) equipped with graphite
8	monochromatized Cu K $\alpha$ radiation ( $\lambda = 1.54056$ Å) in 2 $\theta$ ranging from 20° to 80° with
9	a scan rate of 1.2° min <sup>-1</sup> at 40 kV and 100 mA, respectively. UV-visible diffuse
10	reflectance spectra were carried out in a quartz cuvette on a Cary 5000 UV-Vis-NIR
11	spectrophotometer (Varian, USA). The controlled experiment for $Ag^+$ was
12	performed using an Agilent 7500a ICP-MS (PaloAlto, CA, USA). PEC
13	measurements were performed with a home-built PEC system. Photocurrent was
14	measured by the current-time curve experimental technique on a CHI660D
15	electrochemical workstation (CH Instruments, Shanghai, China) at a bias voltage of
16	0.2 V following a 250 W tungsten halogen light excitation. The distance between the
17	light source and the electrode was fixed at 10 cm. All experiments were carried out
18	at room temperature in the dark using a conventional three electrode system with a
19	modified ITO (5.0 mm in diameter) as the working electrode, a platinum wire as the
20	auxiliary electrode, and a standard Ag/AgCl in 3 mol L <sup>-1</sup> KCl as the reference
21	electrode. The error bars in the respective calibration curves were derived from a set
22	of N=3 experiments.

1	Sensor Preparation and Detection Procedure. ZnO nanorod array was facilely
2	prepared by electrodeposition according to the reference with some modifications. <sup>S1</sup>
3	In detail, before electrodeposition, an ITO electrode (1 cm $\times$ 4 cm) was cleaned with
4	NaOH (1 mol $L^{-1}$ ) and H <sub>2</sub> O <sub>2</sub> (30%), washed with acetone and ultrapure water,
5	respectively. The ZnO nanorods were electrodeposited for 2 h at 80 $^{\circ}$ C and -1.0 V vs
6	a Ag/AgCl electrode in 3 mol L <sup>-1</sup> KCl from a hybrid solution of 0.5 mmol
7	$L^{-1}$ Zn(NO <sub>3</sub> ) <sub>2</sub> ·6H <sub>2</sub> O and 0.1 mol $L^{-1}$ KNO <sub>3</sub> on ITO substrates (1 cm × 1 cm) in a
8	three-electrode system. And then, the ZnO nanorods modified ITO electrodes were
9	annealed to ensure particles adhesion to the substrate at 110 °C for 1 h in air. Next,
10	10 $\mu$ L of 2.0 % PDDA containing 0.5 mol L <sup>-1</sup> KNO <sub>3</sub> was dropped onto the ZnO
11	nanorod array modified ITO electrode for 10 min. Following, 10 $\mu$ L of 10 $\mu$ mol L <sup>-1</sup>
12	poly (C) was assembled onto the modified electrode for 20 min via electrostatic
13	interaction. After each step in the above assembly, the modified ITO electrode was
14	washed with ultrapure water, and dried at room temperature. And thus, $\mathrm{Ag}^{\!\scriptscriptstyle +}$ was
15	selectively detected according to the C-Ag <sup>+</sup> -C coordination chemistry <sup>S2</sup> and then in
16	situ generated AgBr-enhanced ZnO nanorod-based photocurrent. Typically, 10 $\mu$ L of
17	AgNO <sub>3</sub> solution was dropped onto the modified electrode for 30 min, followed by
18	thoroughly rinsing with ultrapure water and dried at room temperature in the dark.
19	Finally, 10 $\mu$ L of 0. 1 mol L <sup>-1</sup> NaBr solution was dropped and incubated for 30 min
20	at room temperature in the dark. After rinsed with copious water, the fabricated PEC
21	sensor was applied to detect Ag <sup>+</sup> in 0.1 mol L <sup>-1</sup> PBS (pH 7.0) by current-time curve
22	experimental technique at the bias voltage of 0.2 V following the 250 W tungsten

halogen light excitation. The controlled experiments were operated according to the same procedure except for the metal ions were other ones (Li<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>,  $Hg^{2+}$ , Zn<sup>2+</sup>, Pb<sup>2+</sup>, Cd<sup>2+</sup> and Cu<sup>2+</sup>) and common anions (F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, PO<sub>4</sub><sup>3-</sup>,  $CO_3^{2-}$ ). All the analyte concentrations mentioned in this work referred to the final concentration in the detected samples. Except the characterization, the annealed ZnO nanorods modified ITO electrode was masked off with an effective working area of a circle that had a diameter of 5 mm for further assembly and experiment.

## Characterization of the ZnO Nanorod Array and the Modified Electrode. Fig. 8 S1 showed the top view observations of ZnO nanorod array and the composite film 9 10 of ZnO-PDDA-poly (C)-AgBr (ZnO-AgBr). The inset in Fig. S1A showed the top view of the ZnO nanorod array in a large magnification. As could be seen clearly in 11 KCl from a hybrid solution of 0.5 mmol L<sup>-1</sup> Zn(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O and 0.1 mol L<sup>-1</sup> KNO<sub>3</sub>. 12 And the synthesis yielded ZnO nanorods with diameters of 200-300 nm and lengths 13 of 0.5-1.0 µm. It could be indexed to wurtzite structure of hexagonal ZnO, indicating 14 its growth direction along [0001].<sup>S3,S4</sup> Fig. S1B showed the top view of ZnO-AgBr. 15 16 It showed that a thin polymer film covered most of the ZnO nanorods, especially for those not vertical to the substrate. AgBr in the composite could not be seen in (B) 17 owing to its very small size and the little quantity. But the mapping results based on 18 the EDS pattern indicated that the composite film concluded Zn (C), O (D), Ag (E) 19 20 and Br (F) (N, Cl and C not shown here). Quantitative analysis showed that mean atomic ratio of Zn/O in the ZnO nanorods is nearly stoichiometric. But the quantity 21 of Br (F) was obviously more than that of Ag (E). This is owing to the much excess 22

NaBr used in the assembly and the insufficient washing in the last step. However, the excess NaBr did no harm to the detection, actually it helped to enhance the photocurrent by digesting the holes left like that in photocatalysis.<sup>S5</sup> In summary, these data confirmed the high purity of the ZnO-nanorod and the successful assembly of ZnO-AgBr.

Fig. S2 showed the XRD patterns of (a) ZnO, (b) ZnO-AgBr, (c) AgBr. All the 6 peaks showed the existence of ZnO (JCPDS card No.36-1451) and AgBr (JCPDS 7 card No.06-0438), respectively. And it could be suitably indexed to wurtzite 8 (hexagonal) zinc oxide with a lattice constant of a=b=3.249 Å and c=5.206 Å, which 9 agreed well with the FESEM (inset in Fig. S1A) and also the JCPDS. The phase of 10 AgBr was cubic, and the lattice constant of a=b=c=5.774 Å was in agreement with 11 the literature.<sup>S6</sup> No peaks for other impurities were found in the spectra, revealing the 12 high purity of both products. The sharp and narrow peaks indicated that the 13 nanostructures were highly crystallized. As expected, a substantially higher intensity 14 was observed for the (002) diffraction peak in the XRD patterns of the ZnO nanorod 15 arrays (a), indicating that the ZnO nanorods were highly oriented with their c-axes 16 being perpendicular to the ITO glass substrate,<sup>S3</sup> which was well consistent with the 17 FESEM result in Fig. S1A. 18

The UV-vis diffuse reflectance spectra of ZnO (black), ZnO-AgBr (red), and AgBr (green) are presented in Fig. S3. Although the reflectance was observed in the whole visible wavelength range, the most pronounced enhancement occurred at 395 nm, corresponding to the optical bandgap of about 3.2 eV. This agreed well with the

literatures under similar deposition conditions.<sup>S4</sup> Compared with the ZnO single 1 crystal which has the optical band gap of 3.37 eV and the band edge of 376 nm,  $^{87}$  the 2 band edge here was a little red shift. The red shift band edge and the reflectance in 3 the whole visible region were due to the larger diameter of ZnO nanorods and the 4 more strong light scattering effect here.<sup>S8</sup> This scattering effect enhanced absorption 5 6 due to long optical path-lengths and photonic effects which could persist in periodic vertically-oriented nanorod arrays.<sup>89</sup> It also helped enhancing the photocurrent 7 through multiple absorption. AgBr had a wide absorption peak in the region of 8 400-500 nm, which accorded well with the literatures.<sup>\$5,\$10</sup> Consequently, 9 10 ZnO-AgBr exhibited a higher extinction intensity in the visible region than ZnO, which was beneficial for enhancing the photoresponse. Herein, both the light 11 12 scattering effect of ZnO nanorod array and the AgBr sensitization helped the enhancement of the total light absorption for the modified electrode in the visible 13 light region. 14

**Optimization of Detection Conditions.** The incubation time of Ag<sup>+</sup> and Br<sup>-</sup> were 15 important influencing factors for the sensitivity of the PEC sensing. Fig. S4A and 16 S4B showed the incubation time effect of AgNO<sub>3</sub> and NaBr on the PEC sensing, 17 respectively. As shown in Fig. S4A, after 30 min incubation of Ag<sup>+</sup>, the photocurrent 18 response arrived at a plateau. The incubation of largely excess Br<sup>-</sup> is the same case as 19 Ag<sup>+</sup> (Fig. S4B). Consequently, the incubation time for both Br<sup>-</sup> and Ag<sup>+</sup> was selected 20 as 30 min. In addition, pH effect on the response of the sensing system to  $Ag^+$  was 21 also evaluated as shown in Fig. S5. In the effective phosphate-buffered range of the 22

1	pH value of 5.3-8.04, pH value has no significant effect on the sensing system to
2	Ag <sup>+</sup> . This result can be explained as follows: Firstly, there is no proton taking part in
3	the reaction in the present sensing system. Secondly, as a kind of amphoteric
4	compound, ZnO is relative inert in the presence of weak acid or base. Thirdly,
5	PDDA can help keeping the stability of the ZnO in the presence of weak acid or
6	base. Considering the pH of real samples, pH 7.0 was chosen for the experiments.
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Method	Materials	DLR <sup>a</sup>	LOD <sup>a</sup>	Ref.	
Colorimetric <sup>d5</sup>	Hemin/G-quadruplex DNAzyme	5-100 nM <sup>b</sup>	2.5 nM <sup>d3</sup>	S11	
Colorimetric	Au NP-Citrate-Tween 20	400-1000 nM	100 nM <sup>d3</sup>	S12	
UV-vis	Au nanorod	1 nM-100 μM <sup>b</sup>	10 nM <sup>d4</sup>	S13	
Fluorescence	Al <sub>2</sub> O <sub>3</sub> NP @Au nanodot	3-300 nM	1.5 nM <sup>d4</sup>	S14	
Fluorescence	Graphene quantum dot	0-100 nM	3.5 nM <sup>d2</sup>	S15	
Fluorescence	fluorophore probe	0.1-5.0 µM	$0.13 \; \mu M^{d1}$	S16	
Fluorescence <sup>d5</sup>	the pyrrolo-dC mismatched ds-DNA	0-100 nM	9.2 nM <sup>d2</sup>	S17	
Fluorescence <sup>d5</sup>	CdSe/ZnS QD-labeled DNA	/	$1 \ \mu M^{d4}$	S18	
Elsonogo en o d <sup>5</sup>	SYER Green 1 as the DNA	50-700 nM	32 nM <sup>d1</sup>	S19	
Fluorescence	intercalator				
Fluorescence <sup>d5</sup>	FAM-labeled DNA-SWCNT	0-150 nM	1 nM <sup>d2</sup>	S20	
Fluorescence <sup>d5</sup>	FAM-labeled DNA-GO	/	$5 \text{ nM}^{d1}$	S21	
Fluorescence <sup>d5</sup>	Ru(bpy) <sub>2</sub> dppz <sup>2+</sup> -DNA-CdTe QDs	2 μM-10 μM	0.1 µM	S22	
Fluorescence	Fluorophore Probe	/	10.2 nM	S23	
Fluorescence <sup>d5</sup>	FAM-labeled DNA-nano C <sub>60</sub>	/	1 nM	S24	
CC <sup>c,d5</sup>	Au NPs and Cleavage NEase	1 pM <sup>b</sup> -100 nM	0.47 pM <sup>d1</sup>	S25	
ISE <sup>c</sup>	Ionophore-doped sensing membranes	0.1-1 nM	38 pM <sup>d4</sup>	S26	
DE C <sup>c.d5</sup>	ZnO nanorod array-PDDA-poly	04120 14	150 pM <sup>d2</sup>	TT1 1	
PEC	(C)-AgBr	0.4-12.8 nM	0.4-12.8 nM		I his work

### Table S1. The Comparison of Different Silver (I) Assay Methods

2 a DLR means dynamic linear range. LOD means limit of detection.

3 b nM means nmol  $L^{-1}$ ,  $\mu$ M means  $\mu$ mol  $L^{-1}$ , pM means pmol  $L^{-1}$ .

4 c CC means chronocoulometry. ISE means ion-selective electrode. PEC means

5 photoelectrochemical.

6 d1 LOD was defined at 3S/N.

7 d2 LOD was defined at  $3\sigma$ /slope.

8 d3 LOD was defined as the lowest value that could be detected.

9 d4 LOD was not told.

10 d5 Aptamer-based sensor for  $Ag^+$ .

1	Interference and Application in Real Samples. The selectivity of this system for
2	$Ag^+$ at 7.5 nmol L <sup>-1</sup> was evaluated over other common metal ions and anions such as
3	1000-fold K <sup>+</sup> , Na <sup>+</sup> , Li <sup>+</sup> , F <sup>-</sup> , Cl <sup>-</sup> , SO <sub>4</sub> <sup>2-</sup> , PO <sub>4</sub> <sup>3-</sup> , CO <sub>3</sub> <sup>2-</sup> ; 200-fold Ca <sup>2+</sup> , Mg <sup>2+</sup> ; 150-fold
4	Br <sup>-</sup> ; 30-fold I <sup>-</sup> ; 20-fold Cu <sup>2+</sup> , Cd <sup>2+</sup> , Zn <sup>2+</sup> and 10-fold Pb <sup>2+</sup> , Hg <sup>2+</sup> . As shown in Fig.
5	S6A, only the solution containing $Ag^+$ was obviously "Signal On", whereas the
6	others remained nearly unchanged (the deviation $<5.0\%$ ). Neither Hg <sup>2+</sup> nor Pb <sup>2+</sup>
7	disturbed Ag <sup>+</sup> detection, which was different from other detection methods. <sup>S12-14,S18</sup>
8	This remarkably high selectivity was better than or at least comparable to other
9	aptamer-based or the fluorescent iodide-based silver ion detection methods, \$11,\$16-\$21
10	let alone those nanomaterial-based methods. <sup>S12-S15</sup> This can be explained by both the
11	highly specific interaction of C-Ag <sup>+</sup> -C coordination effect <sup>S2</sup> and the specific ionic
12	interaction between $Ag^+$ and $Br^-$ , which is similar to that of $Ag^+$ and iodide. <sup>S16</sup>
13	Moreover, the in situ generated AgBr could largely enhance the photocurrent by
14	photosensitization. The dual function of AgBr here helps improving the analytical
15	performances to a large extent. Additionally, the influences of common anions on
16	$Ag^+$ assay were also performed. The results (Fig. S6B) showed that 1000-fold F <sup>-</sup> , Cl <sup>-</sup> ,
17	$SO_4^{2-}$ , $PO_4^{3-}$ , $CO_3^{2-}$ ; 150-fold Br <sup>-</sup> and 30-fold I <sup>-</sup> could coexit with Ag <sup>+</sup> . Except for F <sup>-</sup> ,
18	the concentration of any anion above is less than the critical value according to its
19	corresponding $K_{sp}$ constant value and the known concentration of $Ag^+$ added.
20	Therefore, their influences on the capture and detection of $Ag^+$ could be ignored. The
21	selectivity and real sample detection have confirmed this phenomena. This result
22	could be explained by their difficulty to assembly onto the anion oligonucleotides

owing to the electrostatic interaction and particularly the lack of specificity with poly
 (C).

On the bases of the above results, in order to evaluate the feasibility of the novel 3 PEC sensor, we further challenged the performance of the sensor in real samples, i.e. 4 5 the localized lake water by a standard addition method. After a minor pretreatment, 6 consisting of successive filtration through Whatman filter No.1 and Minisart (0.45  $\mu$ m), the real samples (5 mL) were added with 20 uL of 1  $\mu$ mol L<sup>-1</sup> AgNO<sub>3</sub>. And 7 then 10 uL of the mixed solution was dropped onto the modified ITO electrode prior 8 to thoroughly rinsing with water and then incubated with 0.1 mol L<sup>-1</sup> NaBr for 30 9 10 min before the PEC detection. The average recovery values were 95.0% and 107.4% (N=5) obtained by the PEC sensor and ICP-MS, respectively, indicating the potential 11 practicality of the PEC sensor for environmental samples with little interferences. 12 Additionally, this assay was inert toward the blank sample, suggesting the 13 interference of materials in lake water, such as bacteria and pathogenic, can not be 14 captured by the electrode and have little disturbance on the  $Ag^+$  assay. 15

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S11



2 Scheme S1. The structure segments of  $C-Ag^+-C$  for  $Ag^+$ -mediated DNA molecular

3 wire.



Fig. S1. FESEM images of (A) ZnO nanorod array, (B) ZnO-AgBr, Mapping of (C)
Zn, (D) O, (E) Ag, (F) Br in the ZnO-AgBr composite film. The inset in (A) showed
the top view of ZnO nanorod arrays in a large magnification.



Fig. S2. The XRD patterns of (a) ZnO, (b) ZnO-AgBr, (c) AgBr





8 AgBr (green).



Fig. S4. Effects of the incubation time for (A) 7.5 nmol L<sup>-1</sup> Ag<sup>+</sup> and (B) 0.1 mol
L<sup>-1</sup> Br<sup>-</sup> on the response of the modified ITO electrodes in 0.1 mol L<sup>-1</sup> PBS (pH 7.0) at
the bias voltage of 0.2 V following the 250 W tungsten halogen light irradiation.

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Fig. S5. The pH effects on the photocurrent responses of the ZnO-PDDA-poly (C)-AgBr modified ITO electrodes in 0.1 mol L<sup>-1</sup> PBS in the presence of 7.5 nmol L<sup>-1</sup> silver ions at the bias voltage of 0.2 V and following the 250 W tungsten halogen light (simulated sunlight) irradiation.



Fig. S6. Effects of common metal ions and anions on the detection of 7.5 nmol L<sup>-1</sup>
Ag<sup>+</sup> in 0.1 mol L<sup>-1</sup> PBS (pH 7.0) at the bias voltage of 0.2 V following the 250 W
tungsten halogen irradiation. The bars represent 7.5 nmol L<sup>-1</sup> Ag<sup>+</sup> and a mixture of
7.5 nmol L<sup>-1</sup> Ag<sup>+</sup> with one-tenth of the concentration of other ions mentioned as
following: 1000-fold K<sup>+</sup>, Na<sup>+</sup>, Li<sup>+</sup>, F<sup>-</sup>, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, PO<sub>4</sub><sup>3-</sup>, CO<sub>3</sub><sup>2-</sup>; 200-fold Ca<sup>2+</sup>, Mg<sup>2+</sup>;
150-fold Br<sup>-</sup>; 30-fold Γ; 20-fold Cu<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup> and 10-fold Pb<sup>2+</sup>, Hg<sup>2+</sup>. Error bars
represent the standard deviation of three replicates.

#### 9 Supplementary References

10 [S1] J. F. Qiu, M. Guo, Y. J. Feng and X. D. Wang, *Electrochim. Acta*, 2011, 56,

11 5776.

- [S2] A. Ono, S. Cao, H. Togashi, M. Tashiro, T. Fujimoto, T. Machinami, S. Oda, Y.
  Miyake, I. Okamoto and Y. Tanaka, *Chem. Commun.*, 2008, 4825.
- [S3] L. E. Greene, M. Law, J. Goldberger, F. Kim, J. C. Johnson, Y. F. Zhang, R. J.
  Saykally and P. D. Yang, *Angew. Chem. Int. Ed.*, 2003, 42, 3031.
- 16 [S4] F. Xu, Y. Lu, Y. Xie and Y. Liu, J. Solid State Electrochem., 2010, 14, 63.
- 17 [S5] P. Wang, B. Huang, X. Zhang, X. Qin, H. Jin, Y. Dai, Z. Wang, J. Wei, J. Zhan,
- 18 S. Wang, J. Wang and M. H. Whangbo, *Chem. Eur. J.*, 2009, **15**, 1821.
- [S6] Q. Zhu, W. S. Wang, L. Lin, G. Q. Gao, H. L. Guo, H. Du and A. W. Xu, *J. Phys. Chem. C*, 2013, 117, 5894.
- [S7] W. I. Park, Y. H. Jun, S. W. Jung and Y. Gyu-Chul, *Appl. Phys. Lett.*, 2003, 82, 964.
- 23 [S8] R. Tena-Zaera, J. Elias and C. Levy-Clement, Appl. Phys. Lett., 2008, 93,

1	233119.
2	[S9] (a) M. D. Kelzenberg, S. W. Boettcher, J. A. Petykiewicz, D. B. Turner-Evans,
3	M. C. Putnam, E. L. Warren, J. M. Spurgeon, R. M. Briggs, N. S. Lewis and H.
4	A. Atwater, Nat. Mater., 2010, 9, 239; (b) M. Law, L. E. Greene, J. C. Johnson,
5	R. Saykally and P. D. Yang, Nat. Mater., 2005, 4, 455.
6	[S10] (a) P. Wang, B. B. Huang, X. Y. Qin, X. Y. Zhang, Y. Dai and M. H.
7	Whangbo, Inorg. Chem., 2009, 48, 10697; (b) L. Q. Ye, J. Y. Liu, C. Q. Gong,
8	L. H. Tian, T. Y. Peng and L. Zan, ACS Catal., 2012, 2, 1677.
9	[S11] T. Li, L. L. Shi, E. K. Wang and S. J. Dong, Chem. Eur. J., 2009, 15, 3347.
10	[S12] C. Y. Lin, C. J. Yu, Y. H. Lin and W. L. Tseng, Anal. Chem., 2010, 82, 6830.
11	[S13] H. W. Huang, S. N. Chen, F. Liu, Q. Zhao, B. Liao, S. J. Yi and Y. L. Zeng,
12	Anal. Chem., 2013, 85, 2312.
13	[S14] P. C. Chen, T. Y. Yeh, C. M. Ou, C. C. Shih and H. T. Chang, Nanoscale, 2013,
14	<b>5</b> , 4691.
15	[S15] X. Ran, H. J. Sun, F. Pu, J. S. Ren and X. G. Qu, Chem. Commun., 2013, 49,
16	1079.
17	[S16] A. Chatterjee, M. Santra, N. Won, S. Kim, J. K. Kim, S. B. Kim and K. H.
18	Ahn, J. Am. Chem. Soc., 2009, 131, 2040.
19	[S17] K. S. Park, J. Y. Lee and H. G. Park, Chem. Commun., 2012, 48, 4549.
20	[S18] R. Freeman, T. Finder and I. Willner, Angew. Chem. Int. Ed., 2009, 48, 7818.
21	[S19] Y. H. Lin and W. L. Tseng, Chem. Commun., 2009, 6619.
22	[S20] C. Zhao, K. G. Qu, Y. J. Song, C. Xu, J. S. Ren and X. G. Qu, Chem. Eur. J.,
23	2010, <b>16</b> , 8147.
24	[S21] Y. Q. Wen, F. F. Xing, S. J. He, S. P. Song, L. H. Wang, Y. T. Long, D. Li
25	and C. H. Fan, Chem. Commun., 2010, 46, 2596.
26	[S22] W. Sun, J. Yao, T. Yao and S. Shi, Analyst, 2013, 138, 421.
27	[S23] J. Hatai, S. Pal and S. Bandyopadhyay, RSC Advances, 2012, 2, 10941.
28	[S24] H. Li, J. Zhai and X. Sun, Analyst, 2011, 136, 2040.
29	[S25] P. Miao, L. Ning and X. Li, Anal. Chem., 2013, 85, 7966.
30	[S26] C. Z. Lai, M. A. Fierke, R. C. da Costa, J. A. Gladysz, A. Stein and P.

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Bühlmann, Anal. Chem., 2010, 82, 7634.