Supplementary Information

'Click cyclic ADP-ribose': a neutral second messenger mimic

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Potter*.

Contents:

Supplementary Figures	S2
Computational modelling	S 3
Experimental details for synthesis of compounds	S4
Enzymatic Assay for cADPR Hydrolysis	S8
Assay for Calcium release in SUH	S9
¹ H-NMR and ¹³ C-NMR for all new compounds	S10
HPLC traces for 3 and 4	S22

Supplementary Figures



Fig. S1 Compound **3** docked pose in the co-crystal structure of CD38 and cIDPR a) southern ribose pulled out of plane b) overlap of triazole and pyrophosphate groups.



Fig. S2 H-bonds to the pyrophosphate in the crystal structure of cIDPR bound to CD38.



Fig. S3 8-NH₂ group of docked 4 is predicted to H-bond to Asp-155.

Modelling—Compounds **3** and **4** were built and minimised using the Schrödinger software. The 2PGJ structure was passed through the Protein Preparation Wizard of the Schrödinger software after the structure of the ligand in the crystal structure was corrected. Ligands were docked using GOLD with the binding site defined as a sphere of 5Å radius centred on the centroid of the ligand in the binding site. Each ligand was docked 25 times.

Synthesis of Compounds

General Experimental

All reagents and solvents were of commercial quality and were used without further purification, unless described otherwise. Unless otherwise stated, all reactions were carried out under an inert atmosphere of argon. ¹H- and ¹³C chemical shifts (δ) were internally referenced to the residual solvent peak. ¹H- and ¹³C-NMR assignments are based on *g*COSY, *g*HMBC, *g*HSQC, and DEPT-135 experiments. Abbreviations for splitting patterns are as follows: br, broad; s, singlet; d, doublet; t, triplet; m, multiplet etc. Coupling constants are given in Hertz (Hz).

5'-Azido-5'-deoxy-2',3'-*O***-isopropylideneadenosine 6** - Diphenylphosphoryl azide (2.1 mL, 9.77 mmol) and DBU (2.2 mL, 16.64 mmol) were added dropwise to a suspension of 2',3'-isopropylidene adenosine (1.5 g, 4.88 mmol) in dioxane (25 mL) and stirring was continued for 3 h. NaN₃ (1.6 g, 24.4 mmol), TBAI (0.18 g, 0.488 mmol) and 15-crown-5 (96 µL, 0.488 mmol) were then added and the solution was refluxed for 4 h. The solvent was removed under reduced pressure and the residue was purified on an Isco chromatographic system (PE:EtOAc, 1:0 \rightarrow 0:1 v/v) to yield the *title compound* (1.62 g, 100 %) as a pale yellow foam; R_f = 0.47 (DCM:Acetone 1:1 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.27 (s, 1H, H-8), 7.82 (s, 1H, H-2), 6.02 (d, 1H, J = 2.2, H-1'), 5.81 (bs, 2H, NH₂), 5.38 (dd, 1H, J = 6.3, 2.2, H-2'), 4.97 (dd, 1H, J = 6.3, 3.4, H-3'), 4.29 (dt, 1H, J = 3.4, 5.5, H-4'), 3.53-3.44 (m, 2H, H-5'), 1.53 (s, 3H, CH₃) and 1.30 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) δ 155.6, 153.2, 149.1, 139.8, 120.3, 114.7, 90.6, 85.6, 84.0, 82.0, 52.3, 27.0 and 23.7 ppm; HRMS (ES⁺) calcd for C₁₃H₁₇N₈O₃ 333.1418, found *m/z* [M+H]⁺ 333.1386.

5'-Azido-5'-deoxy-2',3'-O-isopropylidene-8-bromoadenosine 7 - Adenosine **6** (2.16 g, 6.50 mmol) was taken up in dioxane:NaOAc (0.5 M, pH 4, aq.) (1:1 v/v, 60 mL). Br₂ (0.67 mL, 13.0 mmol) was added and the resulting solution stirred in the dark for 16 h. Na₂S₂O₃ (sat. aq.), NaHCO₃ (sat. aq.) and DCM were added and the solution stirred vigorously for 30 min. The organic layer was washed with NaCl (sat. aq.), dried (Na₂SO₄) and all solvents evaporated. The residue was purified by column chromatography on silica gel eluting with PE:EtOAc (1:0 \rightarrow 1:0 v/v) to afford the *title compound* (2.63 mg, 99 %) as a pale yellow foam; $R_{\rm f} = 0.67$ (PE:EtOAc, 1:3 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.28 (s, 1H, H-2), 6.21 (d, 1H, *J* = 1.8, H-1'), 5.96 (bs, 2H, NH₂), 5.68 (dd, 1H, *J* = 6.3, 1.8, H-2'), 5.15 (dd, 1H, *J* = 6.3, 3.7, H-3'), 4.34 (ddd, 1H, *J* = 7.3, 5.8, 3.7, H-4'), 3.51 (dd, 1H, *J* = 12.8, 7.3, H-5'a), 3.45 (dd, 1H, *J* = 12.8, 5.8, H-5'b), 1.61 (s, 3H, CH₃), 1.39 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 154.4, 153.0, 150.3, 127.5, 120.2, 114.5, 91.2, 86.4, 83.4, 82.4, 52.1, 27.1, 25.3 ppm; HRMS (ESI⁺) found *m/z* [M+H]⁺ 411.0532, 413.0521; C₁₃H₁₆N₈O₃⁷⁹Br requires 411.0523, C₁₃H₁₆N₈O₃⁸¹Br requires 413.0503.

5'-Azido-5'-deoxy-2',3'-O-isopropylidene-8-bromoinosine 8 - Bromoadenosine **7** (2.23 g, 5.42 mmol) was taken up in acetic acid-water (46 mL, 20:3 v/v). NaNO₂ (4.49 g, 65.08 mmol) was added in one portion and the resulting solution stirred for 16 h. All solvents were evaporated and the residue taken up in EtOH and evaporated. The residue was partitioned between CHCl₃ and H₂O, and the organic layer washed with

NaHCO₃ (sat. aq.), then brine, dried (Na₂SO₄) and evaporated to dryness. The residue was purified by column chromatography on silica gel eluting with PE:EtOAc (1:0 \rightarrow 1:0 v/v) to afford the *title compound* (2.06 mg, 92 %) as an orange foam; $R_{\rm f} = 0.24$ (PE:EtOAc, 1:3 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 13.08 (bs, 1H, NH), 8.46 (s, 1H, H-2), 6.19 (d, 1H, J = 2.0, H-1'), 5.57 (dd, 1H, J = 6.4, 2.0, H-2'), 5.08 (dd, 1H, J = 6.4, 4.1, H-3'), 4.30 (dd, 1H, J = 6.1, 4.2, H-4'), 3.50 (d, 2H, J = 6.1, both H-5'), 1.60 (s, 3H, CH₃), 1.39 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 157.7, 149.5, 146.2, 126.5, 125.5, 114.9, 91.1, 86.1, 83.5, 81.9, 52.0, 27.1, 25.3 ppm; HRMS (ESI⁻) found *m*/*z* [M-H]⁻ 410.0227, 412.0194; C₁₃H₁₃N₇O₄⁷⁹Br requires 410.0218, C₁₃H₁₃N₇O₄⁸¹Br requires 412.0197.

*N*1-(2",3",5"-Tri-*O*-acetyl-β-D-ribofuranosyl)-5'-azido-5'-deoxy-2',3'-*O*-isopropylidene-8-bromoinosine

9 - Bromoinosine **8** (250 mg, 0.61 mmol) was taken up in DCM (2.5 mL) and DBU (272 µL, 1.82 mmol) added. After 30 minutes, 1,2,3,5-tetra-*O*-acetyl-β-D-ribofuranose (213 mg, 0.67 mmol) was added and the solution cooled to -78 °C. Trimethylsilyl trifluoromethanesulfonate (439 µL, 2.43 mmol) was added dropwise and the solution stirred for a further 45 min before warming to rt. After 1 h, NaHCO₃ (sat. aq.) was added and the crude material extracted into DCM (× 3). The combined organic fractions were dried (Na₂SO₄), and solvent evaporated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with PE:EtOAc (1:0 → 0:1 v/v) to afford the *title compound* (369 mg, 91 %) as a colorless glass; $R_f = 0.53$ (EtOAc:PE 3:1 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.18 (s, 1H, H-2), 6.32 (d, 1H, *J* = 4.3, H-1″), 6.15 (d, 1H, *J* = 2.2, H-1′), 5.48 (dd, 1H, *J* = 6.4, 2.2, H-2′), 5.43-5.42 (m, 2H, H-2″ and H-3″), 5.03 (dd, 1H, *J* = 6.4, 4.4, H-3′), 4.44-4.36 (m, 3H, H-4″ and 2 × H-5″), 4.26 (dt, 1H, *J* = 6.4, 5.9, H-4′), 3.49 (d, 2H, *J* = 5.9, both H-5′), 2.16, 2.11, 2.07 (each s, 3H, 3 × OAc), 1.60 (s, 3H, CH₃), 1.37 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 170.1, 169.6, 169.5, 154.4, 147.7, 144.4, 126.2, 124.9, 115.1, 90.8, 87.6, 85.5, 83.5, 81.6, 80.4, 74.2, 70.2, 63.0, 51.8, 27.2, 25.3, 20.7, 20.4, 20.3 ppm; HRMS (ESI⁺) found *m*/*z* [M+Na]⁺ 692.0891, 694.0869; C₂₄H₂₈N₇O₁₁⁷⁹BrNa requires 692.0922, C₂₄H₂₈N₇O₁₁⁸¹BrNa requires 694.0902.

*N*1-(β-D-ribofuranosyl)-5'-azido-5'-deoxy-2',3'-O-isopropylidene-8-bromoinosine 10 - Triacetyl 9 (1.45 g, 2.16 mmol) was taken up in MeOH (10 mL) in a pressure tube. The solution was saturated with NH₃ (g) at 0 °C, then stirred at rt for 12 h. The solvent was evaporated and the residue purified by column chromatography on silica gel eluting with DCM:Acetone (1:0 → 0:1 v/v) to afford the *title compound* (962 mg, 81 %) as a clear oil; $R_f = 0.38$ (DCM:Acetone 1:1 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.63 (s, 1H, H-2), 6.11 (d, 1H, J = 2.1, H-1'), 6.03 (d, 1H, J = 3.8, H-1"), 5.48 (dd, 1H, J = 6.4, 2.1, H-2'), 5.14 (bd, 1H, J = 3.9, 2'-OH), 5.02 (dd, 1H, J = 6.4, 4.2, H-3'), 4.65 (bd, 1H, J = 4.6, 3'-OH), 4.47 (m, 1H, H-2"), 4.40 (dd, 1H, J = 6.2, 4.7, H-3"), 4.24 (dt, 1H, J = 6.2, 4.2, H-4'), 4.18-4.16 (m, 1H, H-4"), 4.02-3.99 (m, 1H, 5'-OH), 3.95-3.86 (m, 2H, both H-5"), 3.45 (d, 2H, J = 6.2, both H-5'), 1.57 (s, 3H, CH₃), 1.34 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 155.7, 148.4, 145.9, 126.5, 124.5, 115.0, 92.2, 91.0, 85.8 (2C), 83.4, 81.7, 75.3, 70.1, 61.5, 51.9, 27.1, 25.3 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 566.0618, 568.0596; C₁₈H₂₂N₇O₈⁷⁹BrNa requires 566.0605, C₁₈H₂₂N₇O₈⁸¹BrNa requires 568.0585.

N1-(2",3"-O-isopropylidene-β-D-ribofuranosyl)-5'-azido-5'-deoxy-2',3'-O-isopropylidene-8-

bromoinosine 11 - To **10** (75 mg, 0.14 mmol) in acetone-2,2-dimethoxypropane (2.5 mL, 4:1 v/v) was added *p*-TsOH (26 mg, 0.14 mmol). After 1 h, DCM and NaHCO₃ (sat. aq.) were added, and the organic layer dried (Na₂SO₄) and evaporated to dryness. The residue was taken up in MeOH (5 mL) and DOWEX H⁺ resin (25 mg) added. After 30 min the resin was removed by filtration under gravity and the solvent evaporated to obtain the *title compound* (75 mg, 93 %) as a colourless glass; $R_f = 0.50$ (PE:EtOAc 1:3 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.10 (s, 1H, H-2), 6.15 (d, 1H, J = 2.2, H-1'), 5.78 (d, 1H, J = 2.9, H-1"), 5.47 (dd, 1H, J = 6.4, 2.2, H-2'), 5.22 (dd, 1H, J = 6.4, 2.9, H-2"), 5.09 (dd, 1H, J = 6.4, 3.4, H-3"), 5.03 (dd, 1H, J = 6.4, 4.2, H-3'), 4.37 (ddd, 1H, J = 3.7, 3.4, 2.5, H-4"), 4.27 (dt, 1H, J = 5.9, 4.2, H-4'), 3.93 (dd, 1H, J = 12.2, 2.5, H-5"a), 3.83 (dd, 1H, J = 12.2, 3.7, H-5"b), 3.49 (d, 2H, J = 5.9, both H-5'), 1.60 (s, 3H, CH₃), 1.58 (s, 3H, CH₃), 1.37 (s, 3H, CH₃), 1.35 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 155.0, 148.1, 146.6, 126.5, 125.6, 115.1, 114.3, 96.6, 90.9, 87.9, 85.6, 83.6, 83.5, 81.7, 80.6, 62.7, 51.9, 27.3, 27.1, 25.3, 25.2 ppm; HRMS (ESI⁺) found m/z [M+H]⁺ 584.1096, 586.1091; C₂₁H₂₇N₇O₈⁷⁹Br requires 584.1099, C₂₁H₂₇N₇O₈⁸¹Br requires 586.1079.

N1-(2",3"-O-isopropylidene-5"-O-propargyl-β-D-ribofuranosyl)-5'-azido-5'-deoxy-2',3'-O-

isopropylidene-8-bromoinosine 12 - To **11** (145 mg, 0.25 mmol) at 0 °C in toluene-dioxane (1.5 mL, 2:1 v/v) was added powdered KOH (28 mg, 0.50 mmol). After 15 min, propargyl bromide (80 %, 55 µL, 0.50 mmol) and TBAI (92 mg, 0.25 mmol) were added, the solution was allowed to warm to rt and stirred for a further 16 h. The reaction mixture was diluted with DCM, washed with H₂O and the organic layer dried (Na₂SO₄) and all solvents evaporated. The residue was purified by column chromatography on silica gel eluting with PE:EtOAc (1:0 \rightarrow 0:1 v/v) to afford the *title compound* (60 mg, 39 %) as a colourless oil; $R_{\rm f}$ = 0.66 (EtOAc:PE 3:1 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 8.19 (s, 1H, H-2), 6.17 (d, 1H, J = 2.1, H-1'), 6.07 (d, 1H, J = 1.9, H-1''), 5.48 (dd, 1H, J = 6.4, 2.1, H-2'), 5.06 (dd, 1H, J = 6.4, 4.2, H-3'), 4.99 (dd, 1H, J = 6.3, 1.9, H-2''), 4.93 (dd, 1H, J = 6.3, 3.6, H-3''), 4.46 (ddd, 1H, J = 5.1, 3.6, 3.3, H-4''), 4.29 (dt, 1H, J = 5.8, 4.2, H-4'), 4.21 (dd, 1H, J = 15.8, 2.4, propargyl-Ha), 4.16 (dd, 1H, J = 15.8, 2.4, propargyl-Hb), 3.89 (dd, 1H, J = 10.3, 5.1, H-5''b), 3.50 (d, 2H, J = 5.8, both H-5'), 2.46 (t, 1H, J = 2.4, alkynyl-H) 1.61 (s, 3H, CH₃), 1.59 (s, 3H, CH₃), 1.37 (s, 3H, CH₃), 1.35 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 154.7, 148.1, 145.8, 126.1, 125.3, 115.0, 114.2, 94.2, 90.9, 87.1, 85.8, 85.7, 83.7, 81.9, 81.4, 78.8, 75.4, 69.9, 58.6, 52.0, 27.22, 27.20, 25.4, 25.3 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 644.1053, 646.1062; C₂₄H₂₈N₇O₈⁷⁹BrNa requires 644.1075, C₂₄H₂₈N₇O₈⁸¹BrNa requires 646.1054.

N1-Cyclic-2",3"-O-isopropylidene-8-bromoinosine-5'-deoxy-5'-(4-methyl-1,2,3-triazole)-2',3'-O-

isopropylideneribose 13 - To 12 (60 mg, 0.097 mmol) in THF (60 mL) was added DIPEA (50 μ L, 0.290 mmol). The solution was degassed with Argon (30 min), CuI (37 mg, 0.193 mmol) added and the solution refluxed at 70 °C for 16 h. All solvents were evaporated, and the residue was purified by column chromatography on silica gel eluting with PE:EtOAc (1:0 \rightarrow 0:1 v/v) to afford the *title compound* (45 mg, 75 %); $R_{\rm f} = 0.11$ (EtOAc:PE 3:1 v/v); ¹H-NMR (400 MHz, CDCl₃) δ 7.81 (s, 1H, H-2), 7.15 (s, 1H, triazole-

CH), 6.09 (d, 1H, J = 1.2, H-1'), 5.84 (d, 1H, J = 1.5, H-1"), 5.21 (t, 1H, J = 6.6, H-3'), 5.04 (dd, 1H, J = 6.6, 1.2, H-2'), 4.94 (dd, 1H, J = 15.1, 1.4, H-5'a), 4.89 (dd, 1H, J = 5.6, 1.5, H-2"), 4.82-4.79 (m, 2H, H-3", H-4"), 4.67 (d, 1H, J = 11.0, H-5"a), 4.49 (dd, 1H, J = 15.1, 3.2, H-5'b), 4.46 (d, 1H, J = 13.8, triazole-CH₂-Ha), 4.28-4.26 (m, 1H, H-4'), 4.25 (d, 1H, J = 13.8, triazole-CH₂-Hb), 3.68 (dd, 1H, J = 11.0, H-5"b), 1.62 (s, 3H, CH₃), 1.58 (s, 3H, CH₃), 1.39 (s, 3H, CH₃), 1.32 (s, 3H, CH₃) ppm; ¹³C-NMR (100 MHz, CDCl₃) 155.1, 148.5, 144.5, 143.7 (2C), 125.7, 124.6, 115.4, 112.6, 97.4, 88.7, 87.5, 87.3, 84.2, 83.6, 81.7, 78.1, 72.2, 63.5, 48.7, 27.5, 26.9, 25.5, 25.0 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 644.1014, 646.1006; $C_{24}H_{28}N_7O_8^{79}$ BrNa requires 644.1075, $C_{24}H_{28}N_7O_8^{81}$ BrNa requires 646.1054.

N1-Cyclic 8-bromoinosine 5'-deoxy-5'-(4-methyl-1,2,3-triazole) ribose 14 - Compound **13** (20 mg, 0.032) was taken up in aqueous TFA (2 mL, 50 %) at 0 °C and then stirred at rt for 3 h. All solvents were evaporated and the residue coevaporated with MeOH (×3). The residue was then purified by column chromatography on silica gel eluting with DCM:MeOH (1:0 → 7:3 v/v) to afford the *title compound* (15 mg, 88 %) as a white amorphous solid; $R_f = 0.42$ (DCM:MeOH 4:1 v/v); ¹H-NMR (500 MHz, d₄-MeOD) δ 8.68 (s, 1H, H-2), 7.88 (s, 1H, triazole-CH), 5.90 (d, 1H, J = 2.4, H-1'), 5.86 (s, 1H, H-1''), 4.81 (d, 2H, J = 8.5, both H-5'), 4.72 (d, 1H, J = 11.1, triazole-CH₂-Ha), 4.65-4.63 (m, 1H, H-3'), 4.61 (d, 1H, J = 11.1, triazole-CH₂-Hb), 4.54 (dd, 1H, J = 6.3, 2.4, H-2'), 4.34-4.29 (m, 3H, H-2'', H-3'', H-4''), 4.26 (d, 1H, J = 11.5, H-5''a), 4.26-4.24 (m, 1H, H-4'), 3.83 (d, 1H, J = 11.5, H-5''b) ppm; ¹³C-NMR (125 MHz, d₄-MeOD) 157.2, 150.5, 146.6 (2C), 127.5, 127.4, 125.1, 94.6, 93.5, 85.7, 82.3, 76.9, 74.1, 69.64, 69.58, 69.3, 65.5, 50.2 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 564.0467, 566.0448; C₁₈H₂₀N₇O₈⁷⁹BrNa requires 564.0449, C₁₈H₂₀N₇O₈⁸¹BrNa requires 566.0428.

N1-Cyclic inosine 5'-deoxy-5'-(4-methyl-1,2,3-triazole) ribose 3 - To **14** (8 mg, 0.015 mmol) in MeOH (2.5 mL) was added Pd/C (1 mg, 10 mol %) and NaHCO₃ (6 mg, 0.074 mmol). The resulting suspension was stirred under an atmosphere of H₂ for 8 h, filtered and all solvents evaporated. The residue was purified by semi-preparative reverse phase HPLC eluted at 5 mL/min with acetonitrile:0.1 M TEAB (1:19 → 13:7 v/v) over 25 min. Fractions were analysed by analytical RP-HPLC eluted at 1 mL/min with ion-pair buffer: 0.17% (*m/v*) cetrimide and 45% (*v/v*) phosphate buffer (pH 6.4) in MeOH. Appropriate fractions were collected and evaporated under vacuum to give the *title compound* (2.0 mg, 31 %); ¹H-NMR (500 MHz, D₂O) δ 8.53 (s, 1H, H-2), 8.05 (s, 1H, H-8), 7.95 (s, 1H, triazole-CH), 5.90 (s, 1H, H-1"), 5.88 (d, 1H, *J* = 1.6, H-1'), 4.87 (dd, 1H, *J* = 15.7, 2.9, H-5'a), 4.77-4.72 (m, 3H, triazole-CH₂-Ha, H-5'b, H-3'), 4.54 (d, 1H, *J* = 11.3, triazole-CH₂-Hb), 4.46 (dd, 1H, *J* = 6.1, 1.6, H-2'), 4.43-4.39 (m, 2H, H-2", H-3"), 4.33 (d, 1H, *J* = 7.1, H-4"), 4.30-4.27 (m, 1H, H-4'), 4.21 (dd, 1H, *J* = 12.2, 1.9, H-5"a), 3.84 (dd, 1H, *J* = 12.2, 1.1, H-5"b) ppm; ¹³C-NMR (125 MHz, D₂O) 157.8, 147.7, 144.5, 143.7, 142.0, 126.7, 123.4, 92.4, 91.1, 84.1, 79.6, 75.4, 73.4, 68.4, 68.1, 67.7, 63.9, 48.5 ppm; HRMS (ESI⁺) found *m*/z [M+Na]⁺ 486.1354; C₁₈H₂₁N₇O₈Na requires 486.1344.

*N*1-Cyclic 8-azidoinosine 5'-deoxy-5'-(4-methyl-1,2,3-triazole) ribose 15 - Compound 14 (9.0 mg, 0.016 mmol) was evaporated from dry DMF (4×2 mL), taken up in DMF (1 mL) and stirred under Argon. NaN₃

(11 mg, 0.17 mmol) was added and the resulting suspension stirred at 70 °C in the dark for 16 h, after which a second portion of NaN₃ (11 mg, 0.17 mmol) was added and the suspension stirred for a further 16 h, after which full conversion of the starting material to product was observed by HPLC ($\lambda = 255 \text{ nm} \rightarrow \lambda = 277 \text{ nm}$). All solvent was evaporated and the resulting residue co-evaporated with MilliQ (2 × 5 mL). The residue was then taken up in MilliQ (5 mL), filtered through cotton wool, and purified by semi-preparative HPLC (1.1 × 25 cm C₁₈ column) eluted with acetonitrile:MilliQ (1:0 \rightarrow 13:7 v/v) over 25 min. Fractions were analysed by analytical HPLC and appropriate fractions collected and evaporated under vacuum to give the *title compound* (6.0 mg, 71 %); ¹H-NMR (500 MHz, d₇-DMF) δ 8.95 (s, 1H, H-2), 8.22 (s, 1H, triazole-CH), 6.07 (s, 1H, H-1"), 5.76 (d, 1H, J = 3.1, H-1'), 5.02 (d, 2H, J = 2.3, both H-5'), 4.99 (d, 1H, J = 10.8, triazole-CH₂-Ha), 4.84 (d, 1H, J = 10.8, triazole-CH₂-Hb), 4.75 (dd, 1H, J = 7.2, 6.3, H-3'), 4.70 (dd, 1H, J = 6.3, 3.1, H-2'), 4.61 (dd, 1H, J = 8.2, 4.2, H-3"), 4.48-4.43 (m, 4H, H-2", H-4", H-5"a), 4.06 (d, 1H, J = 11.3, H-5"b) ppm; ¹³C-NMR (125 MHz, d₇-DMF) 155.3, 148.2, 145.0, 143.8, 143.7, 126.3, 121.9, 92.7, 89.5, 84.1, 81.5, 75.7, 73.1, 68.7, 68.1, 68.0, 65.5, 49.3 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 527.1381; C₁₈H₂₀N₁₀O₈Na requires 527.1358.

N1-Cyclic 8-aminoinosine 5'-deoxy-5'-(4-methyl-1,2,3-triazole) ribose 4 - Compound 14 (5.0 mg, 9.2 μ mol) was evaporated from dry DMF (4 \times 2 mL), taken up in DMF (1 mL) and stirred under Argon. NaN₃ (6 mg, 0.092 mmol) was added and the resulting suspension stirred at 70 °C in the dark for 16 h, after which a second portion of NaN₃ (6 mg, 0.092 mmol) was added and the suspension stirred for a further 16 h, after which full conversion of the starting material to product was observed by HPLC ($\lambda = 255 \text{ nm} \rightarrow \lambda = 277$ nm). All solvent was evaporated and the resulting residue co-evaporated with MilliO (2×5 mL). The residue was taken up in EtOH-MilliO (3 mL, 1:2 v/v), Pd/C (5 mg) added and stirred under an atmosphere of H₂ for 16 h. The suspension was filtered and all solvents evaporated. The residue was purified by semipreparative HPLC (1.1 × 25 cm C₁₈ column) eluted with acetonitrile:0.1 M TEAB (1:0 \rightarrow 13:7 v/v) over 25 min. Fractions were analysed by analytical HPLC and appropriate fractions collected and evaporated under vacuum to give the *title compound* (1.85 mg, 42 % over 2 steps); ¹H-NMR (500 MHz, D₂O) δ 8.33 (s, 1H, H-2), 7.92 (s, 1H, triazole-CH), 5.87 (s, 1H, H-1"), 5.70 (d, 1H, J = 1.9, H-1'), 4.83 (dd, 1H, J = 15.6, 2.6, H-5'a), 4.76-4.66 (m, 2H, H-5'b, triazole-CH₂-Ha), 4.60 (dd, 1H, J = 8.9, 5.2, H-3'), 4.55-4.50 (m, 2H, H-2', triazole-CH₂-Hb), 4.40-4.37 (m, 2H, H-2", H-3"), 4.32 (d, 1H, J = 6.5, H-4"), 4.24 (d, 1H, J = 8.9, H-4'), 4.20 (dd, 1H, J = 11.8, 1.6, H-5"a), 3.82 (d, 1H, J = 11.8, H-5"b) ppm; ¹³C-NMR (125 MHz, D₂O) 156.2, 147.5, 143.7, 141.8 (2C), 126.7, 120.1, 92.5, 88.8, 84.2, 79.5, 75.6, 72.9, 68.4, 67.9, 67.8, 64.0, 48.7 ppm; HRMS (ESI⁺) found m/z [M+Na]⁺ 501.1467; C₁₈H₂₂N₈O₈Na requires 501.1453.

Enzymatic Assay for cADPR Hydrolysis—The inhibition of cADPR hydrolysis by various concentrations of analogue (0–1 mM) was determined by incubating 1 μ M cADPR with 1 μ g/ml of CD38 for 10 min at 20 – 24 °C in 25 mM sodium acetate, pH 4.5. The reaction was stopped by the addition of 150 mM HCl. The precipitated protein was filtered, and the pH was neutralized with Tris base. After diluting the mixture 20-

fold, the concentration of the unhydrolyzed cADPR present in the diluted reaction mixture was assayed by the fluorimetric cycling assay as described previously.¹

Preparation of Sea urchin Egg Homogenate (SUH) and Assay for Calcium Release²

Eggs from the sea urchin *Lytechinus pictus* (Marinus Scientific, Long Beach, CA) were collected into artificial seawater (435 mM NaCl, 40 mM MgCl₂, 15 mM MgSO₄, 11 mM CaCl₂, 10 mM KCl, 2.5 mM NaHCO₃, 20 mM Tris and 1mM EDTA, pH 8), following intracoelomic injection of 0.5 M KCl, de-jellied by passing through 100 µm nylon net (Millipore, Watford), then washed to reduce extra-cellular calcium: twice in calcium-free artificial seawater with EGTA (470 mM NaCl, 27 mM MgCl₂, 28 mM MgSO₄, 10 mM KCl, 2.5 mM NaHCO₃, and 1mM EGTA, pH 8), then twice in calcium-free artificial seawater (470 mM NaCl, 27 mM MgCl₂, 28 mM MgSO₄, 10 mM KCl, 2.7 mM MgCl₂, 28 mM MgSO₄, 10 mM KCl, 27 mM MgCl₂, 28 mM MgSO₄, 10 mM KCl, and 2.5 mM NaHCO₃, pH 8), each time gently spinning down the eggs and discarding the supernatant, re-suspending the eggs at ten times their packed volume.

The eggs were then washed in an ice-cold intra-cellular-like medium (Glu-IM) (250 mM N-methyl glucamine, 250 mM potassium gluconate, 20 mM Hepes, 1 mM MgCl₂, pH 7.2), before being re-suspended in an equivalent volume of Glu-IM supplemented with 2 mM ATP, 20 U/ml creatine phosphokinase, 20 U/ml phosphocreatine and protease-inhibitor cocktail tablets (completeTM EDTA-free, Roche, Burgess Hill) one per 50 ml, and then homogenised using a Dounce homogeniser with a tight-fitting pestle. The homogenate was frozen and stored as aliquots at -80°C till required.

For use in the calcium-release assay an aliquot of SUH would be thawed and progressively diluted (approximately doubling the volume each time) over a 3.5 hour period kept at 17°C in Glu-IM supplemented with 1 mM ATP, 10 U/ml creatine phosphokinase, 10 U/ml phosphocreatine to give a final dilution of 1 in 20 (equivalent to 2.5% packed eggs). The fluorescent calcium reporter dye fluo-3 (Life Technologies, Paisley) was added in the final dilution to be at 3 μ M. The assays were performed using a Novostar plate reader (BMG Labtech, Aylesbury). The diluted SUEH was placed in wells of a micro plate, 100 μ l/well for 98-well plates, 50 or 20 μ l/well for 384-well plates and baseline fluorescence recorded. Additions of the compounds being studied were then made at 1%–2.5% volume of SUEH and the subsequent maximum fluorescence recorded, baseline subtracted to give a value for the rise in fluorescence and that value normalised to the rise evoked by 0.5 μ M cADPR (an amount that generates a near maximal rise) under the same conditions.

Data was manipulated and plots generated with "Prism" (GraphPad Software, La Jolla, CA).

^{1.} R. M. Graeff, H. C. Lee, *Biochem. J.*, 2002, **361**, 379-384.

^{2. (}a) D. L. Clapper, T. F. Walseth, P. J. Dargie, H. C. Lee, *J. Biol. Chem.*, 1987, **262**, 9561-9568; (b) A. J. Morgan, G. C. Churchill, R. Masgrau, M. Ruas, L. C. Davis, R. A. Billington, S. Patel, M. Yamasaki, J. M. Thomas, A. A. Genazzani, A. Galione, *Methods in cADPR and NAADP research*, CRC Press, Boca Raton, 2006.



S10





















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S21

HPLC trace for compound 3



HPLC trace for compound 4

