Supporting information figures

Table S1. Sequences of winner peptides.

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Figure S1. Outline of the TAT Hitchhiker in vivo selection system.



Figure S2. Analytical ultracentrifugation (AUC) of equimolar mixtures of HBZ-ZIP, JWH-ZIP and related peptides. A 1:1 mixture of 50 μ M peptides or a single peptide of 50 μ M was run at 42,000 rpm at 10°C in 10 mM HEPES containing 500mM NaCl (pH 7.4) and absorbance scans were carried out at 240 nm. AUC data for the HBZ-ZIP/JWH-ZIP mixture are best fit to a single-dispersed model with an apparent weight-average molecular weight (MW_{app}) of 10.1×10^4 , which corresponds well to the expected MW_{app} of the heterodimer (9.8×10⁴). JWH-ZIP, having a high sequence similarity with cJun-ZIP, was also suggested to form a homodimer (observed in AUC: 8.9×10³; expected as a dimer: 9.6×10³), as in the case of cJun-ZIP (observed in AUC: 9.0×10³; expected as a dimer: 9.6×10³). On the other hand, MW_{app} (5.2×10³) of HBZ-ZIP suggested that the peptide exists as a monomer (theoretical molecular weight of the monomer, 4.9×10³). Other expected molecular weight: cFos-ZIP/JWH-ZIP heterodimer, 9.7×10³; HBZ-ZIP/cJun-ZIP heterodimer, 9.7×10³; cJun-ZIP homodimer, 9.5×10³; cFos-ZIP homodimer, 9.6×10³; cFos-ZIP monomer, 4.8×10³.



Figure S3. Size-exclusion chromatography with on-line light-scattering (SEC-LS) analysis of (a) an equimolar mixtures of HBZ-ZIP and JWH-ZIP, (b) JWH-ZIP, (c) HBZ-ZIP, (d) an equimolar mixtures of cFos-ZIP and JWH-ZIP. Expected molecular masses; HBZ-ZIP/JWH-ZIP heterodimer, 9.8 kDa; JWH homodimer, 9.6 kDa; JWH-ZIP monomer, 4.8 kDa; HBZ-ZIP homodimer, 9.9 kDa; HBZ-ZIP monomer, 4.9 kDa; cFos-ZIP/JWH-ZIP heterodimer, 9.7 kDa. Total peptide concentration: 1 mg/mL (150 μL); eluent: 25 mM HEPES containing 150 mM NaCl (pH 7.4); temperature: 23°C.



Figure S4. CD spectra and denaturation curves. (a) Complex formation of JWH-ZIP with HBZ-ZIP. CD spectrum of the 1:1 mixture of HBZ-ZIP and JWH-ZIP (solid line, purple) was suggestive of a significantly more stabilized helical structure than the added curves of JWH-ZIP (25 μ M) and HBZ-ZIP (25 μ M) (dashed line, black). (b) Thermal denaturation curves for HBZ-ZIP (Tm=-1°C, squares, blue), cFos-ZIP (Tm=7°C, triangles, green), cJun-ZIP (Tm=34°C, open circles, black), and JWH-ZIP (Tm=35°C, closed circles, red). (c) CD spectra of cJun-ZIP alone (dashed line, black) and 1:1: mixtures of cJun-ZIP with HBZ-ZIP (solid line, red). (d) CD spectra of cFos-ZIP (dashed line, black), mixture of cFos-ZIP/cJun-ZIP (dashed dotted line, blue), and mixture of cFos-ZIP/JWH-ZIP (solid line, red). For (b), (c) and (d) total peptide concentration is 50 μ M.



Figure S5. ITC measurements. (a) HBZ-ZIP titrated into JWH-ZIP; (b) HBZ-ZIP titrated into JWH-ZIP; (c) cFos-ZIP titrated into JWH-ZIP; and (d) cFos-ZIP titrated into cJun-ZIP.



Figure S6. Co-localization of HA-JHW (red) and Myc-HBZ (green).



Figure S7. Expression check of the HA-cJun, Myc-HBZ, and HA-JWH. *protein from control vector pCAG-HA.