Supporting information

Tryptoline-3-hydroxypyridinaldoxime conjugates as efficient

reactivators of phosphorylated human acetyl and

butyrylcholinesterases

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1. Chemistry

General. Column chromatography purifications were performed on Merck silica gel (40-63 μ m). Thin-layer chromatography (TLC) was carried out on Merck DC Kieselgel 60 F-254 aluminium sheets. Compounds were visualized by illumination with a short wavelength UV lamp ($\lambda = 254$ nm). All solvents were dried following standard procedures (CH₂Cl₂, CH₃CN: distillation over P₂O₅, DMF: distillation over BaO under reduced pressure, THF: distillation over Na/benzophenone). Triethylamine was distilled from CaH₂ and stored over BaO or KOH. ¹H and ¹³C NMR spectra were recorded on a Bruker DPX 300 spectrometer (Bruker, Wissembourg, France). Chemical shifts are expressed in parts per million (ppm) from CDCl₃ (δ_{H} = 7.26, δ_{C} = 77.16), MeOD (δ_{H} = 3.31, δ_{C} = 49.00) or DMSO (δ_{H} = 2.50, δ_{C} = 39.52).¹J values are expressed in Hz. Residual solvents contained in NMR sample were indicated on spectra. Mass spectra were obtained with a Finnigan LCQ Advantage MAX (ion trap) apparatus equipped with an electrospray source. All analyses were performed in the positive mode.

All final oximes were confirmed to be \geq 95% purity based on HPLC analysis. Analytical HPLC was performed on a Thermo Electron Surveyor instrument equipped with a PDA detector under the following conditions: Thermo Hypersil GOLD C18 column (5 µm, 4.6 x 100 mm) with MeOH and 0.1% aq. trifluoroacetic acid (TFA) as eluents [0.1% aq. TFA/MeOH (90/10) (5 min), followed by linear gradient from 10% to 100% of MeOH (45 min)] at a flow rate of 1.0 mL/min and UV detection Max Plot 220-360 nm.

Methyl 3-benzyloxy-6-bromopicolinate 6



 H_2SO_4 (1.8 mL, 31.5 mmol, 3 equiv.) was added dropwise to a suspension of 3hydroxypicolinic acid (1.5 g, 10.5 mmol) in MeOH (24 mL). The mixture was refluxed for 24 h. Then, the mixture was neutralized with an aqueous solution of K_2CO_3 (pH 8.5). The aqueous layer was extracted with EtOAc (thrice). The organic layer was dried over MgSO₄ and concentrated under reduced pressure to give methyl 3-hydroxypicolinate (1.28 g, 80%) as a white solid. ¹H and ¹³C NMR data *were* in agreement with *those* given in the *literature*.¹ At 0°C, Br₂ (335 µL, 6.5 mmol) was added portionwise (4 x 84 µL in 2 h) to a suspension of previous methyl ester (1 g, 6.5 mmol) in water (40 mL). The mixture was stirred at 0 °C for 2 h then 15 h at rt. The solution was extracted with dichloromethane (thrice). The organic layer was washed with brine, dried over MgSO₄ and concentrated under reduced pressure. Purification by chromatography on silica gel (cylohexane/EtOAc 8/2, v/v) afforded methyl 6-bromo-3-hydroxypicolinate (796 mg, 53%) as a white powder. R*f* = 0.45 (cylohexane/EtOAc 8/2, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 4.07 (s, 3H), 7.29 (d, *J* = 8.7 Hz, 1H), 7.58 (dd, *J* = 0.3, 8.7 Hz, 1H), 10.72 (br s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 53.5 (CH₃), 129.5 (CH), 130.0 (C), 130.7 (C), 134.5 (CH), 158.5 (C), 169.1 (C). MS (ESI+): *m/z* (%): 234 (85) and 232 (100) [M+H]⁺.

Benzyl bromide (770 µL, 6.3 mmol, 3 equiv.) was slowly added to a mixture of previous compound (500 mg, 2.1 mmol) and K₂CO₃ (1.3 g, 10.8 mmol, 4.5 equiv.) in acetone (30 mL). The solution was refluxed for 15 h. The resulting mixture was filtered and concentrated under reduced pressure. Purification by chromatography on silica gel (cyclohexane/EtOAc 9/1, v/v) gave **6** (631 mg, 93%) as a white solid. R*f* = 0.3 (cyclohexane/EtOAc 8/2, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 3.98 (s, 3H), 5.22 (s, 2H), 7.25 (d, *J* = 8.7 Hz, 1H), 7.46-7.34 (m, 5H), 7.51 (d, *J* = 8.7 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 52.9 (CH₃), 71.2 (CH₂), 125.0 (CH), 126.9 (2 x CH), 128.4 (CH), 128.8 (2 x CH), 131.2 (C), 131.4 (CH), 135.2 (C), 139.8 (C), 154.0 (C), 164.0 (C). MS (ESI+): *m/z* (%): 324 (85) and 322 (100) [M+H]⁺.

Methyl 3-(benzyloxy)-6-(4-hydroxybut-1-ynyl)picolinate 9



To a solution of methyl 3-(benzyloxy)-6-bromopicolinate **6** (800 mg, 2.48 mmol) in CH_2Cl_2 (25 mL) and Et_3N (13 mL) was added 3-butyn-1-ol (174 mg, 2.48 mmol). The resulting mixture was degassed for 15 min. CuI (47 mg, 0.25 mmol, 0.1 equiv.) and Pd(PPh₃)₄ (143 mg, 0.12 mmol, 0.05 equiv.) were then poured and the solution was stirred under argon at rt overnight. The reaction mixture was concentrated under reduced pressure. Purification by chromatography on silica gel (cyclohexane/EtOAc 4/6 to 3/7, v/v) afforded the desired

¹ Louise-Leriche, L.; Paunescu, E.; Saint-André, G.; Baati, R.; Romieu, A.; Wagner, A.; Renard, P.-Y., *Chem. Eur. J.* **2010**, *16*, 3510-3523.

product **9** as a yellow solid (770 mg, 99%). R*f* = 0.15 (cyclohexane/EtOAc 4/6, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 2.66 (t, J = 6.6 Hz, 2H), 3.00 (br s, 1H), 3.82 (t, J = 6.6 Hz, 2H), 3.95 (s, 3H), 4.29-4.35 (m, 1H), 5.18 (s, 2H), 7.28-7.44 (m, 7H). ¹³C NMR (75 MHz, CDCl₃) δ 23.8 (CH₂), 52.7 (CH₃), 60.7 (CH₂), 70.9 (CH₂), 80.6 (C), 87.4 (C), 121.9 (CH), 126.9 (2 x CH), 128.3 (CH), 128.8 (2 x CH), 130.1 (CH), 135.0 (C), 135.5 (C), 139.9 (C), 153.2 (C), 164.8 (C). MS (ESI+): *m/z* (%): 312 (100) [M+H]⁺.

Methyl 3-(benzyloxy)-6-(5-hydroxypent-1-ynyl)picolinate 10



To a solution of methyl 3-(benzyloxy)-6-bromopicolinate **6** (800 mg, 2.48 mmol) in CH₂Cl₂ (25 mL) and Et₃N (13 mL) was added 4-pentyn-1-ol (208 mg, 2.48 mmol). The resulting mixture was degassed for 15 min. CuI (47 mg, 0.25 mmol, 0.1 equiv.) and Pd(PPh₃)₄ (143 mg, 0.12 mmol, 0.05 equiv.) were then poured and the solution was stirred under argon at rt overnight. The reaction mixture was concentrated under reduced pressure. Purification by flash chromatography (cyclohexane/EtOAc 5/5 to 4/6, v/v) afforded the desired product **10** as a yellow solid (800 mg, 99%). R*f* = 0.16 (cyclohexane/EtOAc 4/6, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.86 (qt, J = 6.4 Hz, 2H), 2.38 (br s, 1H), 2.54 (t, J = 7.0 Hz, 2H), 3.78 (t, J = 6.2 Hz, 2H), 3.96 (s, 3H), 5.20 (s, 2H), 7.30-7.45 (m, 7H). ¹³C NMR (75 MHz, CDCl₃) δ 15.9 (CH₂), 31.0 (CH₂), 52.7 (CH₃), 61.4 (CH₂), 70.9 (CH₂), 79.6 (C), 89.8 (C), 121.9 (CH), 127.0 (2 x CH), 128.8 (2 x CH), 130.1 (CH), 135.4 (C), 135.6 (C), 140.0 (C), 153.0 (C), 164.9 (C). MS (ESI+): *m/z* (%): 326 (100) [M+H]⁺, 348 (95) [M+Na]⁺ and 673 (100) [2M + Na]⁺.

Methyl 3-(benzyloxy)-6-(4-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)but-1ynyl)picolinate 11



To a mixture of methyl 3-(benzyloxy)-6-(4-hydroxybut-1-ynyl)picolinate **9** (771 mg, 2.5 mmol) in dry CH_2Cl_2 (25 mL) was added Et_3N (1 mL, 7.5 mmol, 3 equiv.) and MsCl (290 μ L, 3.8 mmol, 1.5 equiv.). The mixture was stirred at reflux for 4h. The resulting solution was cooled to rt and filtrated under Celite.

The crude product was dissolved in CH₃CN (30 mL) with 2,3,4,9-tetrahydro-1*H*-pyrido[3,4*b*]indole **5** (430 mg, 2.5 mmol, 1 equiv.) and K₂CO₃ (1 g, 7.5 mmol, 3 equiv.). The solution was heated under reflux for 24 h and then cooled at rt. Salts were filtrated. Concentration under reduced pressure and purification by flash chromatography (cyclohexane/EtOAc 1/9, v/v) afforded the desired product **11** as a brown solid (389 mg, 34%). R*f* = 0.21 (cyclohexane/EtOAc 1/9, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 2.71 (t, J = 14.7 Hz, 2H), 2.81-2.84 (m, 2H), 2.91-2.95 (m, 4H), 3.82 (s, 2H), 3.97 (s, 3H), 5.20 (s, 2H), 7.05-7.15 (m, 2H), 7.25-7.48 (m, 9H), 8.00 (br s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 18.6 (CH₂), 21.2 (CH₂), 50.0 (CH₂), 51.1 (CH₂), 52.8 (CH₃), 55.6 (CH₂), 71.0 (CH₂), 80.3 (C), 88.7 (C), 108.2 (C), 110.9 (CH), 118.0 (CH), 119.3 (CH), 121.4 (CH), 121.9 (CH), 127.0 (2 x CH), 127.3 (C), 128.4 (CH), 128.9 (2 x CH), 130.1 (CH), 131.7 (C), 135.4 (C), 135.6 (C), 136.2 (C), 140.0 (C), 153.2 (C), 165.0 (C). MS (ESI+): *m/z* (%): 466 (100) [M+H]⁺, 931 (33) [2M + H]⁺.

Methyl 3-(benzyloxy)-6-(5-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)pent-1-ynyl)picolinate 12



To a mixture of methyl 3-(benzyloxy)-6-(5-hydroxypent-1-ynyl)picolinate **10** (800 mg, 2.5 mmol) in dry CH_2Cl_2 (25 mL) was added Et_3N (1 mL, 7.5 mmol, 3 equiv.) and MsCl (290 μ L, 3.8 mmol, 1.5 equiv.). The mixture was stirred at reflux for 4 h. The resulting solution was cooled to rt and filtrated under Celite.

The crude product was placed in CH₃CN (30 mL) with 2,3,4,9-tetrahydro-1*H*-pyrido[3,4*b*]indole **5** (430 mg, 2.5 mmol, 1 equiv.) and K₂CO₃ (1 g, 7.5 mmol, 3 equiv.). The solution was heated under reflux for 24 h and then cooled at rt. Salts were removed by filtration. Concentration under reduced pressure and purification by flash chromatography (cyclohexane/EtOAc 1/9, v/v to 100% EtOAc) afforded the desired product **12** as orange oil (458 mg, 38%). R*f* = 0.14 (cyclohexane/EtOAc 1/9, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.81 (qt, J = 6.9 Hz, 2H), 2.45 (t, J = 7.2 Hz, 2H), 2.63 (t, J = 6.9 Hz, 2H), 2.78-2.81 (m, 4H), 3.49 (s, 2H), 3.93 (s, 3H), 5.13 (s, 2H), 7.05-7.10 (m, 2H), 7.16-7.43 (m, 9H), 8.62 (br s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 17.4 (CH₂), 21.4 (CH₂), 25.9 (CH₂), 50.1 (CH₂), 51.2 (CH₂), 52.8 (CH₃), 56.7 (CH₂), 70.7 (CH₂), 77.6 (C), 90.3 (C), 107.9 (C), 110.9 (CH), 117.9 (CH), 118.6 (CH), 119.0 (CH), 121.1 (CH), 121.9 (CH), 126.9 (2 x CH), 127.2 (C), 128.3 (CH), 128.8 (2 x CH), 130.2 (CH), 132.2 (C), 135.4 (C), 135.5 (C), 136.2 (C), 139.8 (C), 153.0 (C), 165.0 (C). MS (ESI+): *m/z* (%): 480 (100) [M+H]⁺.

6-(4-(3,4-dihydro-1*H*-pyrido[3,4-b]indol-2(9*H*)-yl)butyl)-3-hydroxypicolinaldehyde 13



Methyl 3-(benzyloxy)-6-(4-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)but-1ynyl)picolinate **11** (389 mg, 0.84 mmol) was dissolved in degassed mixture of MeOH (40 mL) and EtOAc (20 mL). Pearlman's catalyst (480 mg, 0.19 mmol, 0.4 equiv., 20% Pd, moisture 50%) was added and the solution was bubbled with H₂ and the reaction was stirred at rt under H₂ *atmosphere* (1 atm) for 24 h. The mixture was filtrated through Celite and concentrated under reduced pressure.

To a solution of crude product in dry CH_2Cl_2 (6 mL) were successively added 2,6-lutidine (209 µL, 1.8 mmol, 3 equiv.) and TBDMSOTF (413 µL, 1.8 mmol, 3 equiv.). The mixture was stirred at rt for 4 h under argon atmosphere. The mixture was washed with NaCl sat., dried over MgSO₄ and concentrated under reduced pressure.

To a solution of the resulting residue in dry CH_2Cl_2 (6 mL) was added dropwise DIBAL-H (1.8 mL, 1M in CH_2Cl_2 , 1.8 mmol, 3 equiv.) at -78°C. Then, the reaction mixture was stirred

at this temperature for 12 min. The reaction was quenched with MeOH (1.8 mL) and the mixture was allowed to warm at rt. The organic layer was washed with an aqueous solution of NaOH (1M), dried over MgSO₄ and concentrated under reduced pressure.

Then, TBAF (650 µL, 1M in THF, 0.65 mmol, 1.1 equiv.) was added at 0 °C to the residue in dry THF (6 mL) and the mixture was stirred for 2 h at this temperature. After concentration under reduced pressure, a chromatography on silica gel (EtOAc/MeOH 9/1, v/v) afforded **13** as a yellow solid (60 mg, 29%). R*f* = 0.58 (EtOAc/MeOH 9/1, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.61-1.66 (m, 2H), 1.75-1.80 (m, 2H), 2.57 (t, J = 7.2 Hz, 2H), 2.78-2.87 (m, 6H), 3.53 (s, 2H), 7.03-7.12 (m, 2H), 7.22-7.26 (s, 3H), 7.44 (d, J = 7.5 Hz, 1H), 8.34 (br s, 1H), 10.02 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 21.2 (CH₂), 26.9 (CH₂), 27.6 (CH₂), 37.1 (CH₂), 50.3 (CH₂), 51.0 (CH₂), 57.4 (CH₂), 108.2 (C), 110.8 (CH), 118.0 (CH), 119.3 (CH), 121.3 (CH), 126.5 (CH), 127.2 (C), 129.8 (CH), 131.7 (C), 135.7 (C), 136.2 (C), 154.7 (C), 157.1 (C), 198.7 (C). MS (ESI+): *m/z* (%): 350 (100) [M+H]⁺.

6-(5-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)pentyl)-3-hydroxypicolinaldehyde 14



Methyl 3-(benzyloxy)-6-(5-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)pent-1ynyl)picolinate **12** (458 mg, 0.96 mmol) was dissolved in degassed mixture of MeOH (50 mL) and EtOAc (25 mL). Pearlman's catalyst (270 mg, 0.19 mmol, 0.2 equiv., 20% Pd, moisture 50%) was added and the solution was bubbled with H_2 and the reaction was stirred at rt under H_2 *atmosphere* (1 atm) for 2 h. The mixture was filtrated through Celite and concentrated under reduced pressure.

To a solution of crude product in dry CH_2Cl_2 (10 mL) were successively added 2,6-lutidine (334 μ L, 2.9 mmol, 3 equiv.) and TBDMSOTf (666 μ L, 2.9 mmol, 3 equiv.). The mixture was stirred at rt for 4 h under argon atmosphere. The mixture was washed with NaCl sat., dried over MgSO₄ and concentrated under reduced pressure.

To a solution of the resulting residue in dry CH_2Cl_2 (10 mL) was added dropwise DIBAL-H (2.9 mL, 1M in CH_2Cl_2 , 2.9 mmol, 3 equiv.) at -78°C. Then, the reaction mixture was stirred at this temperature for 12 min. The reaction was quenched with MeOH (2.9 mL) and the mixture was allowed to warm at room temperature. The organic layer was washed with an

aqueous solution of NaOH (1 M), dried over MgSO₄ and concentrated under reduced pressure. Then, TBAF (1.1 mL, 1M in THF, 1.1 mmol, 1.1 equiv.) was added at 0 °C to the residue in dry THF (10 mL) and the mixture was stirred for 16 h at this temperature. After concentration under reduced pressure, a chromatography on silica gel (EtOAc/MeOH 95/5, v/v) afforded access to **14** as a yellow solid (119 mg, 34%). R*f* = 0.68 (EtOAc/MeOH 8/2, v/v). ¹H NMR (300 MHz, CDCl₃) δ (ppm) 1.35-1.41 (m, 2H), 1.58-1.63 (m, 2H), 1.70-1.77 (m, 2H), 2.48 (t, J = 7.2 Hz, 2H), 2.76-2.83 (m, 6H), 3.48 (s, 2H), 7.04-7.12 (m, 2H), 7.20-7.26 (m 3H), 7.42-7.45 (m, 1H), 8.43 (br s, 1H), 10.04 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 21.2 (CH₂), 22.7 (CH₂), 27.0 (CH₂), 27.2 (CH₂), 29.7 (CH₂), 37.3 (CH₂), 50.3 (CH₂), 50.9 (CH₂), 57.6 (CH₂), 108.1 (C), 110.8 (CH), 117.9 (CH), 119.2 (CH), 121.2 (CH), 126.5 (CH), 127.2 (C), 129.8 (CH), 131.7 (C), 135.7 (C), 136.1 (C), 154.9 (C), 157.1 (C), 198.7 (C). MS (ESI+): *m/z* (%): 364 (100) [M+H]⁺.

6-(4-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)butyl)-3-hydroxypicolinaldehyde oxime 3



То 6-(4-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)butyl)-3а solution of hydroxypicolinaldehyde 13 (60 mg, 0.17 mmol) in dry EtOH (5 mL) were successively added NH₂OH.HCl (15 mg, 0.21 mmol, 1.2 equiv.) and NaOAc (19 mg, 0.22 mmol, 1.3 equiv.). The mixture was stirred at rt for 3 h under argon atmosphere. After concentration under reduced pressure, a chromatography on silica gel (EtOAc/MeOH 95/5, v/v) afforded 3 as a white solid (14 mg, 22%). Rf = 0.49 (EtOAc/MeOH 9/1, v/v). ¹H NMR (300 MHz, DMSO) δ (ppm) 1.55-1.59 (m, 2H), 1.64-1.76 (m, 2H), 2.51-2.57 (m, 2H), 2.65-2.72 (m, 6H), 3.56 (s, 2H), 6.90-7.02 (m, 2H), 7.15 (d, J = 8.4 Hz, 2H), 7.26 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 7.6 Hz, 1H), 8.28 (s, 1H), 10.11 (br s, 1H), 11.82 (br s, 1H). ¹³C NMR (75 MHz, DMSO) δ 21.2 (CH₂), 26.4 (CH₂), 27.3 (CH₂), 36.4 (CH₂), 50.1 (CH₂), 50.9 (CH₂), 57.1 (CH₂), 106.5 (C), 110.8 (CH), 117.3 (CH), 118.2 (CH), 120.2 (CH), 123.8 (CH), 124.1 (CH), 126.7 (C), 132.9 (C), 135.4 (C), 135.8 (C), 151.1 (C), 151.4 (C), 152.9 (CH). MS (ESI+): *m/z* (%): 365 (100) [M+H]⁺. HPLC: $t_R = 23.02 \text{ min}$ (purity = 97.4%). HRMS (ESI+): m/z calcd for $C_{21}H_{25}N_4O_2$ 365.1978; found: 365.1970.

6-(5-(3,4-dihydro-1*H*-pyrido[3,4-b]indol-2(9*H*)-yl)pentyl)-3-hydroxypicolinaldehyde oxime 4



To a solution of 6-(5-(3,4-dihydro-1*H*-pyrido[3,4-*b*]indol-2(9*H*)-yl)pentyl)-3hydroxypicolinaldehyde **14** (119 mg, 0.33 mmol) in dry EtOH (5 mL) were successively added NH₂OH.HCl (27 mg, 0.39 mmol, 1.2 equiv.) and NaOAc (35 mg, 0.43 mmol, 1.3 equiv.). The mixture was stirred at rt for 1 h under argon atmosphere. After concentration under reduced pressure, chromatography on silica gel (EtOAc/MeOH 9/1, v/v) afforded **4** as a white solid (31 mg, 25%). R*f* = 0.68 (EtOAc/MeOH 9/1 v/v). ¹H NMR (300 MHz, MeOD) δ (ppm) 1.43 (dt, J = 7.4, 15.0 Hz, 2H), 1.67-1.82 (m, 4H), 2.71-2.85 (m, 4H), 2.91 (t, J = 5.8 Hz, 2H), 3.10 (t, J = 5.8 Hz, 2H), 3.92 (s, 2H), 6.95-7.10 (m, 2H), 7.17 (d, J = 8.5 Hz, 1H), 7.28 (dd, J = 3.1, 8.2 Hz, 2H), 7.40 (d, J = 7.6 Hz, 1H), 8.30 (s, 1H).

¹³C NMR (75 MHz, MeOD) δ 21.4 (CH₂), 26.9 (CH₂), 27.9 (CH₂), 31.0 (CH₂), 37.7 (CH₂), 51.0 (CH₂), 52.2 (CH₂), 58.4 (CH₂), 107.6 (C), 112.0 (CH), 118.6 (CH), 119.9 (CH), 122.3 (CH), 125.4 (CH), 126.1 (CH), 127.9 (C), 130.8 (C), 136.2 (C), 138.0 (C), 152.7 (2 x C), 154.5 (CH). MS (ESI+): *m/z* (%): 379 (100) [M+H]⁺. HPLC: *t*_R = 24.88 min (purity = 96.0%). HRMS (ESI+): *m/z* calcd for C₂₂H₂₆N₄O₂ 379.2134; found: 379.2133.

2. Biological assays

Inhibition of *hAChE* and *h*BChE by OPNAs. Recombinant *h*AChE and *h*BChE were produced and purified as previously described.^{2,3} VX and tabun were from DGA maîtrise NRBC (Vert le Petit, France). Paraoxon-ethyl was purchased from Sigma-Aldrich. HI-6 was from Pharmacie Centrale des Armées (Orléans, France). All other chemicals including paraoxon were from Sigma. Stock solution of VX and tabun were 5 mM in isopropanol. The inhibition of 120 μ M *h*AChE or 100 μ M of *h*BChE is realized with a 5-fold excess of OPNAs

² Carletti, E.; Li, H.; Li, B.; Ekström, F.; Nicolet, Y.; Loiodice, M.; Gillon, E.; Froment, M. T.; Lockridge, O.; Schopfer, L. M.; Masson, P.; Nachon, F., *J. Am. Chem. Soc.* **2008**, *130*, 16011-16020.

³ Brazzolotto, X.; Wandhammer, M.; Ronco, C.; Trovaslet, M.; Jean, L.; Lockridge, O.; Renard, P.-Y.; Nachon, F., *FEBS J.* **2012**, *279*, 2905-2916.

and was performed in tris buffer (20 mM, pH 7.4, 0.1% BSA) at 25 °C. After a 20-minute incubation, inhibited *h*AChE or *h*BChE was desalted on PD-10 column (GE Healthcare).

Reactivation of hAChE and hBChE inhibited by OPNAs. OPNA-inhibited hAChE was incubated at 37 °C with different concentrations of oxime in 0.1 M phosphate buffer, pH 7.4, 0.1% BSA, 5% methanol (See table below). Methanol was used for complete dissolution of the oximes. 50-µl aliquots of mix was transferred to 1-mL cuvettes at time intervals ranging from 1 to 10 minutes depending on the reactivation rate, for measurement of hAChE activity (1 mM acetylthiocholine) or hBChE activity (1 mM butyrylthiocholine), in Ellman's buffer (phosphate 0.1 M, pH 7.4, 0.1% BSA, 0.5 mM DTNB, 25 °C). The increase in absorbance at 412 nm was followed on a Uvikon 943 spectrophotometer.

The enzyme activity in the control remained constant during the experiment. The percentage of reactivated enzyme (% E_{react}) was calculated as the ratio of the recovered enzyme activity and activity in the control. The apparent reactivation rate k_{obs} for each oxime concentration, the dissociation constant K_D of inhibited enzyme-oxime complex (E-POx) and the maximal reactivation rate constant k_r , were calculated by non-linear fit using the standard oxime concentration-dependent reactivation derived from the following scheme:

E-P + Ox
$$\xleftarrow{K_D}$$
 E-POx $\xleftarrow{k_r}$ E + POx
%E_{react} = 100. (1-e^{k_{obs}.t}) and $k_{obs} = \frac{k_r[Ox]}{K_D + [Ox]}$

Concentration of oximes 2-4 (μ M) used to determine the concentration dependence of the reactivation rate k_{obs} for reactivation of VX-*h*BChE and VX-, tabun- and paraoxon-inhibited *h*AChE.

	VX-hBChE	VX-hAChE	tabun-hAChE	paraoxon-hAChE
2	10-25-50-75 100-150	-	-	-
3	10-25-50-75 100-150	10-25-50-75-100 120-150-200-250	15-35-50-75-100 150-200	10-25-50-75-100 150-200
4	10-30-50-70 100	10-25-35-50-100 150-200-300	20-50-70-100	10-25-50-80-100 120-150



Reactivation of VX-inhibited hBChE ; Plot of k_{obs} vs [2] or [3] or [4]



Reactivation of VX-inhibited hAChE ; Plot of $k_{obs} \mbox{ vs } [\textbf{3}] \mbox{ or } [\textbf{4}]$



Reactivation of tabun-inhibited hAChE ; Plot of $k_{obs} \mbox{ vs } [\textbf{3}] \mbox{ or } [\textbf{4}]$



Reactivation of Paraoxon-inhibited hAChE ; Plot of $k_{obs} \, vs \, \textbf{[3]} \, or \, \textbf{[4]}$























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