Supporting Information

MCF-supported boronic acids as efficient catalysts for direct amide condensations of carboxylic acids and amines

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General Information: All anhydrous solvents were purchased from Sigma-Aldrich and used without further purification. All silanes including dialkyltetramethyldisilazanes were purchased from Gelest, Inc. ¹H and ¹³C NMR spectra were recorded on Bruker AV-400 (400 MHz) instrument. ¹³C CP-MAS and ²⁹Si CP-MAS NMR spectra were recorded on a 400 MHz Bruker spectrometer for the MCF-supported compounds. The mass spectra were recorded on Shimadzu GC-2010 (QP–2010 gas chromatograph). Reaction yields were determined by purification of crude products using preparative TLC or flash column chromatography. Amides **11a**,¹ **11d**,² **11e**,³ **11f**,¹ **11g**,⁴ **11h**,⁵ and **11i**⁶ were known and their NMR spectra were consistent with those reported in literature. The NMR data of boronic acid **2**, amides **11b** and **11c** has been not reported although they have been known in the literature.



General procedure for the synthesis of MCF-supported boronic acid

Scheme S1 Preparation of MCF-supported boronic acid catalysts

MCF (8.0 g) was degassed at 120 °C overnight under vacuum before reaction. HMDS (675 μ l, 3.2 mmol) was added to a suspension of MCF in anhydrous toluene (50 ml). The mixture was stirred at room temperature under argon for 3 h, and then heated at 60 °C overnight. The resulting suspension was filtered, washed with toluene and methanol, and then dried under vacuum. The resulting TMS-capped MCF (5.0 g) was degassed at 120 °C overnight under vacuum before reaction and then dispersed in anhydrous toluene (30 ml). 3-aminopropyltriethoxysilane (APTES) (232 mg, 1.05 mmol) was added to the mixture. The mixture was stirred at room temperature under argon for 3 h, and then

heated at 100 °C overnight. The mixture was cooled down, filtered and washed with toluene, CH₂Cl₂, and methanol. The collected particles were dried under vacuum. The ¹H NMR spectrum of the collected filtrate after solvent evaporation did not show any organic compounds confirming the immobilization of all APTES. The weight gain also confirmed the complete anchoring of all APTES. The particles were post-capped with TMS by the vapor-phase reaction with HMDS. The amine-functionalized MCF was used for the immobilization of boronic acid. This amine-functionalized MCF could be prepared in one pot without recovering the MCF particles at each step. 5-Carboxy-2fluorophenylboronic acid (232 mg, 1.26 mmol) in anhydrous N-methyl-2-pyrrolidone (NMP) was added to 5.0 g of the amine-functionalized MCF dispersed in anhydrous NMP. 2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate (TBTU) (405 mg, 1.26 mmol) in anhydrous NMP was added to the mixture and then N,Ndiisopropylethylamine (DIEA) (658 µL, 3.78 mmol) was added. The resulting mixture was stirred overnight at room temperature, filtered, and washed with NMP (4×15 ml), CH_2Cl_2 (4 × 15 ml) and methanol (4 × 15 ml). The final MCF-supported boronic acid was dried under vacuum. A Kaiser test confirmed that most of amine groups were coupled to the boronic acid. Considering overall weight increase by both post-capping and the coupling of the boronic acid, the loading of the boronic acid was calculated to give 0.2 mmol/g. For fluoroalkyl capping, bis(trifluoropropyl)tetramethyldisilazane (Gelest, Inc.) was used. For propyl and octyl capping, di-n-propyltetramethylsilazane and di-n-octylteteramethylsilazance were used, respectively. With these disilazane compounds, the pre-capping was done at 80 °C and the post-capping was done in anhydrous toluene at 90 °C overnight and then further post-capping with HMDS under the vapor-phase condition.



Fig. S1. Nitrogen adsorption –desorption isotherms for the bare support MCF (\Box), amine-functionalized MCF with fluoroalkyl capping (**=**) and MCF-supported boronic acid with fluoroalkyl capping **8a** (•)

Support materials	Surface area ^a (m ² /g)	Pore volume ^b (cm ³ /g)
Bare MCF	421.5	2.29
Amine-functionalized MCF	382.8	1.96
8 a	320.3	1.80

Table S1. Characteristics of bare MCF, amine-functionalized MCF and the catalyst 8a

^aBET surface area; ^bEstimated at the relative pressure of P/P₀=0.99.



Fig. S2. A Kaiser test of a) amine-functionalized MCF and b) MCF-supported boronic acid

General procedure for the direct amide condensation of carboxylic acids and amines (Table 1)

To a 10 ml headspace vial was added phenylacetic acid (68mg, 0.5 mmol), catalyst (5 mol%) and toluene (2 ml), then benzylamine (55 μ l, 0.5 mmol) was added. After the reaction vial was sealed with PTFE/silicone septa, it was placed in a heating block on a magnetic stirrer at 120 °C for 1-2 h. After reaction was completed, 5 ml of EtOAc was added to fully dissolve the crude products following with filtration through a fritted glass funnel. And then the catalyst was washed with EtOAc (3 \times 5 ml). The filtrate was concentrated to afford crude products. The crude products were purified with preparative TLC or flash column chromatography to give a pure amide product.





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4 : R₁ = (CH₂)₂CF₃, (Loading: 0.2 mmol/g)

5a :R₁ = CH₃, R₂ = H, X = F (Loading: 1.0 mmol/g) **5b** : R₁ = CH₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **5c** : R₁ = CH₃, R₂ = Me, X = F (Loading: 0.2 mmol/g) **6** : R₁ = (CH₂)₂CH₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **7** : R₁ = (CH₂)₇CH₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8a**: R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8b**: R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **7** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **7** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R₁ = (CH₂)₂CF₃, R₂ = H, X = F (Loading: 0.2 mmol/g) **8** : R_1 = (CH_2)_2CF₃, R_2 = H, X = F (Loading: 0.2 mmol/g) **8** : R_1 = (CH_2)_2CF₃, R_2 = H, X = F (Loading: 0.2 mmol/g) **8** : R_1 = (CH_2)_2CF₃, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X = F (Loading: 0.2 mmol/g) **9** : R_1 = (CH_2)_2CF_3, R_2 = H, X =

8c: $R_1 = (CH_2)_2CF_3$, $R_2 = H$, X = F (Loading: 0.5 mmol/g) **8d** : $R_1 = (CH_2)_2CF_3$, $R_2 = H$, X = H (Loading: 0.2 mmol/g)



10 : R₁ = (CH₂)₂CF₃, (Loading: 0.2 mmol/g)

Fig. S3. Catalysts tested. Silica: Silica gel 60 (0.040-0.063 mm), Merck KGaA



Fig. S4. Nitrogen adsorption–desorption isotherms for MCF (\blacksquare), SBA-16 (\bullet) and the commercial silica (\diamond)

Support materials	Surface area ^a (m ² /g)	Pore volume ^b (cm ³ /g)	Pore size ^c (nm)
MCF	421.5	2.29	23.8
SBA-16	527.4	0.38	3.7
Silica	448.7	0.76	4.7

Table S2. Characteristics of the support materials

^aBET surface area; ^bEstimated at the relative pressure of $P/P_0=0.99$; ^cThe pore size was derived from the N₂ adsorption branch using the BJH method.

Ph 🧹	OH O +	Ph NH ₂	Catalyst Foluene, 120 °C (sealed tube)	► Ph H O	Ph
	Entry	Catalyst (mol%)	Time (hr)	Yield (%) ^{b}	_
	1	bare MCF ^c	2	23	_
	2	1 (10)	2	74	
	3	2 (10)	2	78	
	4	3 ^d	2	20	
	5	$3^{d} + 1$ (10)	2	27	
	6	4 (5)	2	11 ^e	
	7	5a (10)	2	44	
	8	5b (10)	2	71	
	9	5c (10)	2	79	
	10	6 (10)	2	98	
	11	7 (10)	2	88	
	12	8a (10)	2	>98	
	13	8d (10)	2	84	
	14	6 (5)	1	55	
	15	8a (5)	1	82	
	16	8b (5)	1	26	
	17	8c (5)	1	40	
	18	9 (5)	1	28	
	19	10 (5)	1	42	

Table S3. Direct amide condensations of phenylacetic acid and benzylamine by various catalysts under sealed conditions^a

^{*a*}Conditions: catalyst (10 or 5 mol%), phenylacetic acid (0.5 mmol), benzylamine (0.5 mmol), toluene (2 ml), 120 °C, 1 or 2 hr; ^{*b*}Isolated yield; ^c250 mg, dried under high vacuum at 120 °C for 15 hr; ^d250 mg; ^eAzeotropic removal of water.

General procedure for the direct amide condensation of carboxylic acids and amines (Table 2)

To a 10 ml headspace vial was added carboxylic acid (75mg, 0.55 mmol), catalyst **8a** (5 mmol%, 125 mg) and toluene (or *o*-xylene) (2 ml), and then amine (0.5 mmol or other amounts as shown in Table 1) were added. After the reaction vial was sealed with PTFE/silicone septa, it was placed in a heating block on a stirrer for 2-16 h at 120 °C (or 155 °C). After the reaction was completed, 5 ml of EtOAc was added to fully dissolve the crude products, followed by filtration with celite. And then the catalyst was washed with EtOAc (3 × 5 ml). The filtrate was concentrated to afford crude products. The crude products were purified with preparative TLC afforded to give a pure product. For azeotropic removal of water, a Dean-Stark trap was used under reflux conditions.

Recycling of the catalyst 8a (Table 3)

To a 10 ml round-bottomed flask containing phenylacetic acid (340 mg, 2.5 mmol), MCF-supported catalyst **8a** (625 mg, 5 mol%) and toluene (5 ml), was added benzylamine (265 μ l, 2.5 mmol). After the reaction mixture was stirred under azeotropic removal of *in situ* generated water for 2 h (oil bath temperature: 130 °C), it was cooled to room temperature. 5 ml of EtOAc was added to fully dissolve the crude products, followed by filtration through a fritted glass funnel. And then the catalyst was thoroughly washed with EtOAc (3 × 5 ml). The filtrate was concentrated to afford crude products and the recovered catalyst was dried under high vacuum for the next round of run. The crude products were purified by flash column chromatography to give a pure amide product.

¹H and ¹³C NMR Data for 2, the supported catalysts and intermediates



¹H NMR (CDCl₃, 400 MHz): δ 8.44 (t, *J* = 5.6 Hz, 1H), 8.36 (s, 2H), 8.05 (dd, *J* = 2.4, 5.6 Hz, 1H), 7.87 (m, 1H), 7.16 (t, *J* = 8.8 Hz, 1H), 3.23 (dd, *J* = 6.8, 12.4 Hz, 2H), 1.48 (m, 2H), 1.31 (m, 2H), 0.89 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (CDCl₃, 100 MHz): δ 168.5, 166.1, 165.8, 135.2, 135.1, 131.3, 131.2, 130.7, 130.7, 115.3, 115.0, 100.0, 31.7, 20.1, 14.2. Mol. Wt.: 239.1 GC - MS *m/z* (%): 195 (6) (M-HBO2), 153 (16), 152 (12), 124 (7), 123 (100), 95 (33), 75 (11).

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 ^{13}C NMR (100 MHz): δ 0.9.



¹³C NMR (100 MHz): δ 23.5, 19.8, 1.0.



¹³C NMR (100 MHz): δ 36.4, 35.2, 32.4, 25.8, 20.4, 15.4, 0.8.



¹³C NMR (100 MHz): δ 31.8, 13.0, 0.9.



¹³C NMR (100 MHz): δ 59.9, 43.9, 25.6, 17.0, 10.5, 0.9.



¹³C NMR (100 MHz): δ 60.2, 54.8, 35.0, 23.5, 17.6, 11.0, 1.0.



¹³C NMR (100 MHz): δ 60.3, 44.1, 29.0, 17.2, 10.3, 0.9 . ²⁹Si NMR: δ 13.8, -57.7, -64.9, -100.5, -108.8.



¹³C NMR (100 MHz): δ 170.0, 135.9, 128.7, 60.7, 51.3, 43.8, 29.4, 24.5, 17.8, 10.8, 1.0. ²⁹Si NMR: δ 14.4, -55.0, -63.6, -99.8, -108.4.

Cat 5b



¹³C NMR (100 MHz): δ 169.5, 134.5, 114.6, 60.0, 43.1, 31.3, 23.9, 17.0, 10.5, 0.9. ²⁹Si NMR: δ 13.9, -56.4, -63.2, -100.0, -108.5.

Cat 5c



¹³C NMR (100 MHz): δ 170.6, 133.3, 114.3, 59.9, 51.3, 33.9, 21.2, 10.1, 0.9. ²⁹Si NMR: δ 13.7, -56.8, -65.3, -99.9, -108.6.

Cat 6



¹³C NMR (100 MHz): δ 169.9, 134.3, 115.1, 60.5, 51.5, 43.3, 21.5, 17.6, 11.0, 1.0. ²⁹Si NMR: δ 13.8, -56.5, -64.9, -100.5, -109.2.



¹³C NMR (100 MHz): δ 169.5, 133.8, 115.1, 60.2, 51.5, 42.8, 34.2, 32.9, 30.2, 23.7, 18.6, 12.8, 1.0.

²⁹Si NMR: δ 13.7, -56.8, -64.0, -100.8, -109.3.





¹³C NMR (100 MHz): δ 170.3, 130.4, 115.6, 60.9, 51.3, 43.7, 29.5, 24.5, 17.7, 10.8, 1.0. ²⁹Si NMR δ = 13.7, -56.9, -64.3, -100.9, -108.6.





¹³C NMR (100 MHz): δ 172.7, 151.9, 133.2, 118.51, 63.5, 52.7, 46.14, 31.5, 26.2, 19.6, 0.8.

²⁹Si NMR δ = 14.2, -54.9, -64.7, -99.9, -108.4.

Cat 8c



¹³C NMR (100 MHz): δ 169.6, 131.6, 115.4, 60.4, 50.3, 43.3, 29.1, 24.3, 17.6, 10.5, 0.9. ²⁹Si NMR δ = 15.0, -56.5, -63.8, -100.3, -107.4.

Cat 8d



¹³C NMR (100 MHz): δ 170.1, 135.7, 129.8, 127.0, 60.2, 50.9, 43.1, 28.8, 17.2, 10.2, 0.8. ²⁹Si NMR δ = 13.7, -56.3, -64.2, -100.1, -108.1.





¹³C NMR (100 MHz): δ 178.3, 130.5, 128.1, 60.8, 51.6, 44.2, 29.2, 18.8, 10.6, 0.7. ²⁹Si NMR δ = 15.0, -55.7, -61.8, -99.9, -107.1.



¹³C NMR (100 MHz): δ 169.4, 129.9, 127.2, 115.1, 60.3, 51.3, 42.7, 28.8, 17.58, 10.3, 0.8. ²⁹Si NMR δ = 15.2, -54.9, -62.2, -99.9, -108.2.

¹H and ¹³C NMR data for amides

11a

H N Bn

¹H NMR (CDCl₃, 400 MHz): δ 7.31-7.18 (m, 10H), 5.92 (brs, 1H), 4.41 (d, *J* = 5.8 Hz, 2H), 3.62 (s, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 171.0, 134.8, 129.4, 129.0, 128.7, 127.5, 127.4, 127.3, 43.8, 43.6. Mol. Wt.: 225.29 GC - MS *m/z* (%):225 (32), 91.5 (100).

11b

_H ∠N _{Bn}

¹H NMR (CDCl₃, 400 MHz): δ 7.32-7.24 (m, 5H), 7.05 (brs, 1H), 4.44 (t, *J* = 2.8 Hz, 2H), 4.39 (dd, *J* = 6 Hz, 8.4 Hz, 1H), 3.91-3.83 (m, 2H), 2.29 (m, 1H), 2.09 (m, 1H), 1.88 (m, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 173.2, 138.2, 128.7, 127.6, 127.5, 78.5, 69.4, 42.8, 30.3, 25.6. Mol. Wt.: 205.25 GC - MS *m/z* (%):205 (9), 177 (13), 106 (15), 91 (43), 71 (100).

11c

¹H NMR (CDCl₃, 400 MHz): δ 7.33-7.25 (m, 5H), 4.41 (d, *J* = 6.0 Hz, 2H), 3.19 (brs, 1H), 1.47 (s, 6H).

¹³C NMR (CDCl₃, 100 MHz): δ 176.6, 138.1, 128.7, 127.6, 127.5, 73.5, 43.2, 27.9. Mol. Wt.: 193.24. GC - MS *m/z* (%): 193 (16), 175 (22), 107 (24), 92 (25), 134 (33), 106 (53), 135 (63), 91 (100).

11d



¹H NMR (CDCl₃, 400 MHz): δ 7.67 (d, J = 15.6 Hz, 1H), 7.47 (t, J = 3.6 Hz, 2H), 7.36 (m, 8H), 6.48 (d, J = 15.6 Hz, 1H), 6.39 (brs, 1H), 4.55 (d, J = 6.0 Hz, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 166.0, 141.3, 138.2, 134.8, 129.7, 128.8, 128.7, 127.9, 127.8, 127.6, 120.5, 43.8. Mol. Wt.: 237.3. GC - MS *m/z* (%):237 (72), 131 (100).

 $\overset{11e}{\overbrace{O}}\overset{H}{\underset{O}{\overset{H}}}_{N_{Bn}}$

¹H NMR (CDCl₃, 400 MHz): δ 7.96 (s, 1H), 7.43 (s, 1H) 7.33(m, 5H), 6.65 (s, 1H), 6.36 (brs, 1H), 4.56 (d, *J* = 5.6 Hz, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 162.6, 144.9, 143.8, 143.8, 138.1, 138.1, 128.8, 127.9, 127.6, 122.4, 108.3, 43.5. Mol. Wt.: 201.22 GC - MS *m/z* (%):201 (29), 95 (100).

11f

.N Bn

¹H NMR (CDCl₃, 400 MHz): δ 7.82 (d, J = 0.8 Hz, 2H), 7.79 (m, 1H), 7.43 (m, 2H), 7.27 (m, 5H), 6.77 (brs, 1H), 4.62 (d, J = 7.0 Hz, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 167.5, 138.2, 134.3, 131.6, 128.8, 128.6, 127.9, 127.6, 127.0, 44.1. Mol. Wt.: 211.26 GC - MS *m/z* (%): 211 (66), 105 (100).

11g NC H NBn ¹H NMR (CDCl₃, 400 MHz): δ 7.89 (dd, *J* = 0.8 Hz, 8.4 Hz, 1H), 7.71 (dd, *J* = 0.4 Hz, 8.4 Hz, 1H), 7.35 (m, 5H), 6.78 (brs, 1H), 4.62 (d, *J* = 5.6 Hz, 2H).

¹³C NMR (CDCl₃, 100 MHz): δ 165.7, 138.2, 137.5, 132.4, 128.9, 127.9, 127.9, 127.8, 118.0, 115.1, 44.3. Mol. Wt.: 236.27 GC - MS *m/z* (%): 77 (17), 91 (25), 79 (28), 106 (46), 235 (50), 102 (66), 130 (94), 236 (100).

11h



¹H NMR (CDCl₃, 400 MHz): δ 7.36-7.25 (m, 5H), 5.68 (brs, 1H), 3.56 (s, 2H), 3.19 (m, 2H), 1.43 (m, 2H), 1.26 (m, 2H), 0.87 (t, *J* = 9.6 Hz, 3H).

¹³C NMR (CDCl₃, 100 MHz): δ 171.0, 135.1, 129.4, 128.9, 128.4, 127.4, 127.3, 43.8, 39.4, 31.5, 19.9, 13.7. Mol. Wt.: 191.27 GC - MS *m/z* (%):191 (9), 100 (13), 91 (58), 92 (100).

11i

Bn

¹H NMR (CDCl₃, 400 MHz), a mixture of two rotamers: δ 7.30-7.23 (m, 10H), 7.13 (m, 1H), 4.63 (s, 1.3H, major rotamer), 4.55 (s, 1H, minor rotamer), 3.81 (s, 1.3H, major rotamer), 3.78 (s, 1H, minor rotamer), 2.98 (s, 1.5H, minor rotamer), 2.92 (s, 2H, major rotamer).

¹³C NMR (CDCl₃, 100 MHz), a mixture of two rotamers: δ 171.6 (minor rotamer), 171.2 (major rotamer), 137.3, 136.5, 135.1, 135.0, 129.0, 128.9, 128.8, 128.7, 128.6, 128.1, 127.7, 127.4, 126.9, 126.8, 126.4, 53.7 (minor rotamer), 51.0 (major rotamer), 41.2 (major rotamer), 40.9 (minor rotamer), 35.3 (major rotamer), 34.1 (minor rotamer). Mol. Wt.: 239.13.

GC - MS *m/z* (%): 239 (57), 91 (100).

11j



¹H NMR (CDCl₃, 400 MHz): δ 7.39-7.27 (m, 9H), 7.05 (m, 1H), 3.71 (brs, 1H).

¹³C NMR (CDCl₃, 100 MHz): δ 169.3, 137.7, 134.5, 129.6, 129.3, 129.0, 127.7, 124.5, 120.0, 44.8. Mol. Wt.: 211.26 GC - MS *m/z* (%): 211 (35), 91 (59), 93 (100).

11k



¹H NMR (CDCl₃, 400 MHz): δ 9.91 (brs, 1H), 9.28 (brs, 1H), 7.21 (s, 2H), 7.11 (d, J = 8.4 Hz, 2H), 6.70 (dd, J = 1.6 Hz, 6.8 Hz, 2H), 6.66 (s, 1H), 3.46 (s, 2H) 2.21 (s, 6H).

¹³C NMR (CDCl₃, 100 MHz): δ 170.0, 156.5, 139.6, 138.1, 130.4, 126.7, 125.1, 117.3, 115.5, 43.1, 21.6. Mol. Wt.: 255.31 GC - MS *m/z* (%): 255 (53), 121 (100), 108 (56).

References

1. L. U. Nordstrøm, H. Vogt, R. Madsen J. Am. Chem.Soc., 2008, 130, 17672.

2. W. Ren and M. Yamane J. Org. Chem., 2010, 75, 3017.

3. N. Zanatta, D. Faoro, S. C. Silva, H. G. Bonacorso, M. A.P. Martins *Tetrahedron Lett.*, 2004, **45**, 5689.

4. T. Maki, K. Ishihara and H. Yamamoto, Org. Lett., 2005, 7, 5043.

5. G. Pelletier, W. S. Bechara, and A. B. Charette J. Am. Chem. Soc., 2010, 132, 121817.

6. Z.-W. Chen, H.-F. Jiang, X.-Y. Pan, Z.-J. He Tetrahedron, 2011, 61, 1431.



Fig. S5. ¹H and ¹³C NMR Spectra for immobilized catalysts and intermediates.





Fig. S7. ¹H and ¹³C NMR Spectra of 2 and amides

