

## **Supporting Information**

### **Enantioselective intramolecular propargylic amination using chiral copper-pybox catalyst**

Masashi Shibata, Kazunari Nakajima, and Yoshiaki Nishibayashi\*

*Institute of Engineering Innovation, School of Engineering, The University of Tokyo,*

*Yayoi, Bunkyo-ku, Tokyo 113-8656, Japan*

#### **List of Contents of Supporting Information**

1. General Methods	Page S2
2. General Procedure for the Preparation of Propargylic Acetates	Page S3
3. Spectroscopic Data of Other Propargylic Acetates	Page S5
4. Enantioselective Intramolecular Propargylic Amination of Propargylic Acetates	Page S8
5. Spectroscopic Data and Isolated Yields of Other Products	Page S9
6. Preparation of 1,1'-(1,2-phenylene)-bis(prop-2-yne-1,1-diyl) Diacetate ( <b>3</b> ).	Page S12
7. Double Propargylic Amination of Propargylic Diacetate with Amines	Page S13
8. Spectroscopic Data and Isolated Yield of Other Product	Page S14
9. X-ray Diffraction Study of <b>2d</b>	Page S15
10. References and Notes	Page S18
11. <sup>1</sup> H and <sup>13</sup> C NMR Spectra	Page S20
12. Charts of Propargylic Aminated Products by HPLC Analysis	Page S34

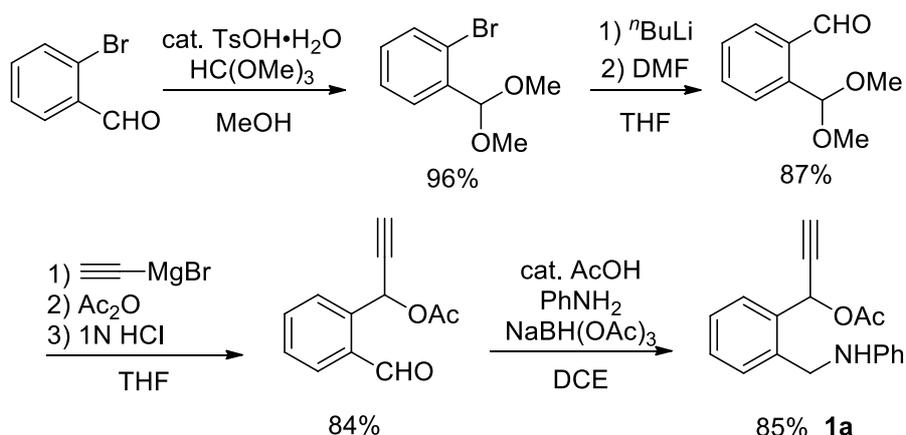
## General Methods.

$^1\text{H}$  NMR (270 MHz) and  $^{13}\text{C}$  NMR (67.8 MHz) spectra were measured on a JEOL Excalibur 270 spectrometer using  $\text{CDCl}_3$  as solvent. HPLC analyses were performed on Hitachi L-7100 and GL-7410 apparatuses equipped with a UV detector using 25 cm x 4.6 mm DAICEL Chiralpak AD, IA columns. Elemental analyses were performed at Microanalytical Center of The University of Tokyo. Mass spectra were measured on a JEOL JMS-700 mass spectrometer. Specific rotations were measured on a JASCO DIP-1000 polarimeter.

All reactions were carried out under a dry nitrogen atmosphere. Solvents were dried by the usual methods, then distilled under  $\text{N}_2$  and degassed before use. Optically pure diphosphines **L1-L3**, pybox ligands **L5**, **L9**,  $\text{CuOTf} \cdot 1/2\text{C}_6\text{H}_6$ , aniline, and *N,N'*-diphenylethylenediamine are commercially available reagents. Optically pure pybox ligands **L4**,<sup>S1</sup> **L6**,<sup>S2</sup> **L7**,<sup>S3</sup> **L8**,<sup>S2</sup> **L10**,<sup>S3</sup> bis(oxazoline) ligand **L11**,<sup>S4</sup> and propargylic acetate **3**<sup>S5</sup> were prepared according to literature procedures.

## General Procedure for the Preparation of Propargylic Acetates.

**Scheme S1.** Preparation of Propargylic Acetate **1a**



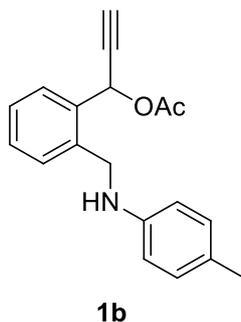
A typical experimental procedure for the preparation of 1-(2-((phenylamino)methyl)phenyl)prop-2-yn-1-yl acetate (**1a**) is described below. In a 50 mL Schlenk flask were placed 2-bromobenzaldehyde (3.80 g, 20.5 mmol) and anhydrous MeOH (20 mL) under  $\text{N}_2$ . Trimethyl orthoformate (8.75 mL, 80 mmol) and  $\text{TsOH}\cdot\text{H}_2\text{O}$  (37.6 mg, 0.20 mmol) were added to the solution and the mixture was stirred at room temperature for 2 h. The resulting mixture was passed through a short silicagel pad to give 1-bromo-2-(dimethoxymethyl)benzene as a colorless oil (4.54 g, 19.6 mmol, 96% isolated yield).

To a solution of 1-bromo-2-(dimethoxymethyl)benzene (4.54 g, 19.6 mmol) in anhydrous THF (60 mL) was added  $n\text{-BuLi}$  (1.65 M in hexane, 18.0 mL, 29.7 mmol) at  $-78\text{ }^\circ\text{C}$  and the mixture was stirred for 30 min. DMF (3.1 mL, 40.0 mmol) was added to the solution and the mixture was stirred at  $-78\text{ }^\circ\text{C}$  for 30 min. The solution was allowed to warm to room temperature and stirred for another 1 h. After the reaction mixture was quenched by addition of saturated  $\text{NaHCO}_3$  aq., the solution was extracted with hexane (20 mL x 3). The combined organic layers were dried over anhydrous  $\text{Na}_2\text{SO}_4$  and concentrated under reduced pressure. The residue was purified by column chromatography (triethylamine-pretreated  $\text{SiO}_2$ ) with hexane/ethyl acetate (90/10) to give 2-(dimethoxymethyl)benzaldehyde as a colorless oil (3.07 g, 17.0 mmol, 87% isolated yield).

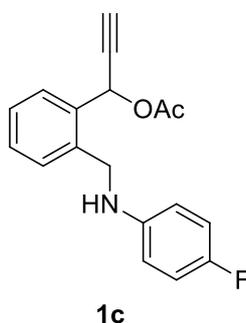
To a solution of 2-(dimethoxymethyl)benzaldehyde (1.46 g, 8.09 mmol) in anhydrous THF (45 mL) was added ethynylmagnesium bromide (0.5 M in THF, 24 mL, 12 mmol) at 0 °C and the mixture was stirred for 40 min. Acetic anhydride (1.50 mL, 15.9 mmol) was added to the solution and the mixture was allowed to warm to room temperature. After stirring for 1 h, the mixture was acidified by addition of 1N HCl aq. (30 mL) and stirred for 3 h. The solution was extracted with ethyl acetate (15 mL x 3). The combined organic layers were washed with brine and dried over anhydrous MgSO<sub>4</sub>. The solvent was removed under reduced pressure and the residue was purified by column chromatography (SiO<sub>2</sub>) with hexane/ethyl acetate (85/15) to give 1-(2-formylphenyl)prop-2-yn-1-yl acetate<sup>S6</sup> as a pale yellow oil (1.37 g, 6.7 mmol, 84% isolated yield).

In a 20 mL Schlenk flask were placed 1-(2-formylphenyl)prop-2-yn-1-yl acetate (208 mg, 1.03 mmol), aniline (0.19 mL, 2.1 mmol), and anhydrous dichloroethane (4 mL) under N<sub>2</sub>. To the solution was added acetic acid (6 µL, 0.1 mmol) at room temperature and the mixture was stirred for 1 h. Sodium triacetoxyborohydride (283.7 mg, 1.34 mmol) was added to the solution and the mixture was stirred for 4 h. After the reaction mixture was quenched by addition of saturated NaHCO<sub>3</sub> aq., the solution was extracted with dichloromethane (5 mL x 3). The combined organic layers were dried over anhydrous MgSO<sub>4</sub> and concentrated under reduced pressure. The residue was purified by column chromatography (SiO<sub>2</sub>) with hexane/ethyl acetate (85/15) to give 1-(2-((phenylamino)methyl)phenyl)prop-2-yn-1-yl acetate (**1a**) as a brown oil (244.6 mg, 0.876 mmol, 85% isolated yield). <sup>1</sup>H NMR δ 7.67-7.63 (m, 1H), 7.46-7.43 (m, 1H), 7.36-7.33 (m, 2H), 7.21-7.15 (m, 2H), 6.76-6.70 (m, 1H), 6.66-6.63 (m, 3H), 4.50 (d, *J* = 14.0 Hz, 1H), 4.43 (d, *J* = 14.0 Hz, 1H), 4.09 (br, 1H), 2.65 (d, *J* = 2.2 Hz, 1H), 2.09 (s, 3H). <sup>13</sup>C NMR δ 169.5, 147.9, 136.9, 135.0, 129.4, 129.3, 129.2, 128.3, 128.0, 117.8, 112.9, 80.2, 75.6, 63.0, 45.7, 20.9. HRMS (EI) Calcd. for C<sub>18</sub>H<sub>17</sub>NO<sub>2</sub> [M]: 279.1259. Found: 279.1249.

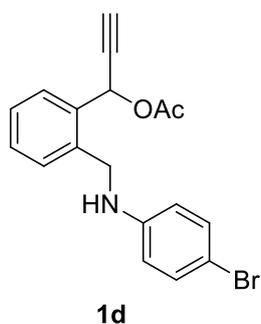
## Spectroscopic Data of Other Propargylic Acetates



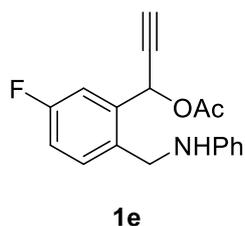
**1-(2-(((4-methylphenyl)amino)methyl)phenyl)prop-2-yn-1-yl acetate (1b):** A brown oil.  $^1\text{H}$  NMR  $\delta$  7.74-7.70 (m, 1H), 7.51-7.48 (m, 1H), 7.42-7.35 (m, 2H), 7.05 (d,  $J = 8.1$  Hz, 2H), 6.73 (d,  $J = 2.4$  Hz, 1H), 6.62 (d,  $J = 8.1$  Hz, 2H), 4.53 (d,  $J = 14.2$  Hz, 1H), 4.46 (d,  $J = 14.2$  Hz, 1H), 4.05 (br, 1H), 2.71 (d,  $J = 2.4$  Hz, 1H), 2.30 (s, 3H), 2.14 (s, 3H).  $^{13}\text{C}$  NMR  $\delta$  169.4, 145.6, 137.1, 134.8, 129.6, 129.3, 129.0, 128.2, 127.8, 126.7, 112.9, 80.1, 75.6, 62.9, 45.9, 20.8, 20.3. HRMS (EI) Calcd. for  $\text{C}_{19}\text{H}_{19}\text{NO}_2$  [M]: 293.1416. Found: 293.1420.



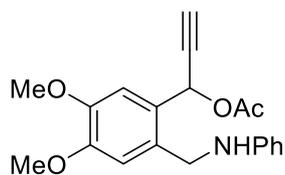
**1-(2-(((4-fluorophenyl)amino)methyl)phenyl)prop-2-yn-1-yl acetate (1c):** A brown solid, m.p. 76.1-78.0 °C.  $^1\text{H}$  NMR  $\delta$  7.68-7.62 (m, 1H), 7.41-7.28 (m, 3H), 6.90-6.81 (m, 2H), 6.65 (d,  $J = 2.4$  Hz, 1H), 6.58-6.50 (m, 2H), 4.42 (d,  $J = 14.0$  Hz, 1H), 4.36 (d,  $J = 14.0$  Hz, 1H), 4.01 (br, 1H), 2.64 (d,  $J = 2.4$  Hz, 1H), 2.05 (s, 3H).  $^{13}\text{C}$  NMR  $\delta$  169.4, 155.9 (d,  $^1J_{\text{C-F}} = 234.7$  Hz), 144.2 (d,  $^4J_{\text{C-F}} = 2.2$  Hz), 136.6, 134.9, 129.3, 129.0, 128.3, 128.0, 115.6 (d,  $^2J_{\text{C-F}} = 22.3$  Hz), 113.6 (d,  $^3J_{\text{C-F}} = 7.8$  Hz), 80.1, 75.7, 62.8, 46.2, 20.8. HRMS (EI) Calcd. for  $\text{C}_{18}\text{H}_{16}\text{FNO}_2$  [M]: 297.1165. Found: 297.1177.



**1-(2-(((4-bromophenyl)amino)methyl)phenyl)prop-2-yn-1-yl acetate (1d):** A pale yellow solid, m.p. 108.1-110.0 °C.  $^1\text{H}$  NMR  $\delta$  7.65-7.62 (m, 1H), 7.39-7.32 (m, 3H), 7.23 (d,  $J = 8.9$  Hz, 2H), 6.62 (d,  $J = 2.4$  Hz, 1H), 6.50 (d,  $J = 8.9$  Hz, 2H), 4.48-4.37 (m, 2H), 4.22 (br, 1H), 2.65 (d,  $J = 2.4$  Hz, 1H), 2.08 (s, 3H).  $^{13}\text{C}$  NMR  $\delta$  169.5, 146.8, 136.3, 135.0, 131.9, 129.4, 129.0, 128.4, 128.1, 114.5, 109.3, 80.1, 75.7, 62.9, 45.7, 20.9. HRMS (EI) Calcd. for  $\text{C}_{18}\text{H}_{16}\text{BrNO}_2$  [M]: 357.0364. Found: 357.0369.



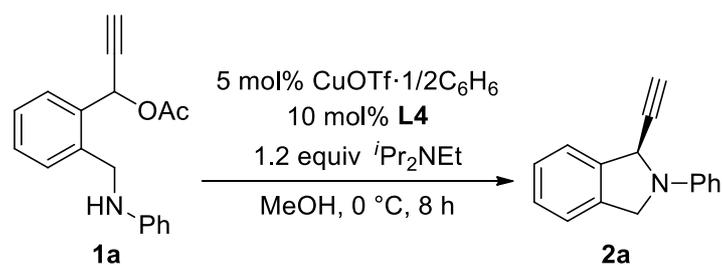
**1-(5-fluoro-2-((phenylamino)methyl)phenyl)prop-2-yn-1-yl acetate (1e):** A brown oil.  $^1\text{H}$  NMR  $\delta$  7.43-7.34 (m, 2H), 7.21-7.15 (m, 2H), 7.06-6.99 (m, 1H), 6.77-6.71 (m, 1H), 6.65-6.62 (m, 3H), 4.44 (d,  $J = 14.2$  Hz, 1H), 4.37 (d,  $J = 14.2$  Hz, 1H), 4.06 (br, 1H), 2.67 (d,  $J = 2.2$  Hz, 1H), 2.11 (s, 3H).  $^{13}\text{C}$  NMR  $\delta$  169.4, 162.2 (d,  $^1J_{\text{C-F}} = 245.9$  Hz), 147.7, 137.2 (d,  $^3J_{\text{C-F}} = 7.3$  Hz), 132.3 (d,  $^4J_{\text{C-F}} = 3.4$  Hz), 130.9 (d,  $^3J_{\text{C-F}} = 8.4$  Hz), 129.3, 117.9, 116.0 (d,  $^2J_{\text{C-F}} = 21.2$  Hz), 115.0 (d,  $^2J_{\text{C-F}} = 22.8$  Hz), 112.9, 79.6, 76.0, 62.2 (d,  $^4J_{\text{C-F}} = 1.7$  Hz), 45.2, 20.8. HRMS (EI) Calcd. for  $\text{C}_{18}\text{H}_{16}\text{NO}_2\text{F}$  [M]: 297.1165. Found: 297.1173.



**1f**

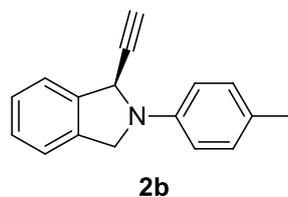
**1-(4,5-dimethoxy-2-((phenylamino)methyl)phenyl)prop-2-yn-1-yl acetate (1f):** A yellow oil.  $^1\text{H}$  NMR  $\delta$  7.22-7.16 (m, 3H), 6.94 (s, 1H), 6.76-6.65 (m, 3H), 6.61 (d,  $J = 2.2$  Hz, 1H), 4.41 (d,  $J = 13.4$  Hz, 1H), 4.32 (d,  $J = 13.4$  Hz, 1H), 4.02 (br, 1H), 3.93 (s, 3H), 3.84 (s, 3H), 2.66 (d,  $J = 2.2$  Hz, 1H), 2.08 (s, 3H).  $^{13}\text{C}$  NMR  $\delta$  169.5, 149.6, 148.4, 148.0, 129.6, 129.2, 127.0, 117.8, 112.9, 112.3, 111.5, 80.4, 75.5, 62.7, 56.0, 55.9, 45.6, 20.9. HRMS (EI) Calcd. for  $\text{C}_{20}\text{H}_{21}\text{NO}_4$  [M]: 339.1471. Found: 339.1454.

## Enantioselective Intramolecular Propargylic Amination of Propargylic Acetates.

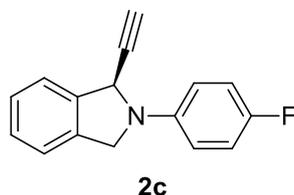


A typical experimental procedure for the reaction of 1-((phenylamino)methyl)phenylprop-2-yn-1-yl acetate (**1a**) is described below. In a 20 mL Schlenk flask were placed CuOTf·1/2C<sub>6</sub>H<sub>6</sub> (2.6 mg, 0.010 mmol) and (*S*)-Me-pybox (**L4**) (5.0 mg, 0.020 mmol) under N<sub>2</sub>. Anhydrous methanol (1.0 mL) was added, and then the mixture was magnetically stirred at 60 °C for 1 h. After the solution was cooled to 0 °C, **1a** (55.9 mg, 0.20 mmol) in anhydrous methanol (1.0 mL) and diisopropylethylamine (42 μL, 0.24 mmol) were added under N<sub>2</sub>, and the reaction was kept at 0 °C for 8 h. The solvent was concentrated under reduced pressure, and the residue was purified by the column chromatography (SiO<sub>2</sub>) with hexane/ethyl acetate (93/7) to give 1-ethynyl-2-phenylisoindoline (**2a**) as a white solid (38.2 mg, 0.173 mmol, 87% isolated yield), m.p. 110.1 °C (decomp.). <sup>1</sup>H NMR δ 7.51-7.48 (m, 1H), 7.39-7.31 (m, 5H), 6.93-6.90 (m, 2H), 6.86-6.80 (m, 1H), 5.61 (br, 1H), 4.82 (dd, *J* = 3.1 and 13.0 Hz, 1H), 4.60 (d, *J* = 13.0 Hz, 1H), 2.41 (d, *J* = 2.2 Hz, 1H). <sup>13</sup>C NMR δ 145.9, 138.5, 137.0, 129.2, 128.2, 127.7, 123.1, 122.6, 117.4, 112.9, 82.6, 72.3, 55.1, 53.7. Anal. Calcd. for C<sub>16</sub>H<sub>13</sub>N: C, 87.64; H, 5.98; N, 6.39. Found: C, 87.44; H, 6.25; N, 6.32. [α]<sub>D</sub><sup>25</sup> = -153.9 (*c* = 0.435, CHCl<sub>3</sub>). The enantiomeric excess of **2a** was determined by HPLC analysis; DAICEL Chiralpak AD, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min, λ = 254 nm, retention time: 9.4 min (minor) and 12.2 min (major), 93% *ee*.

## Spectroscopic Data and Isolated Yields of Other Products.

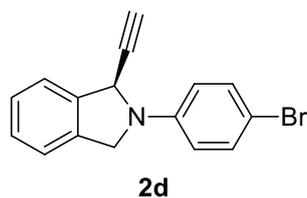


**1-ethynyl-2-(4-methylphenyl)isoindoline (2b):** Isolated yield 79% (with **L4**). A white solid, m.p. 121.4 °C (decomp.). <sup>1</sup>H NMR δ 7.43-7.40 (m, 1H), 7.30-7.26 (m, 3H), 7.08 (d, *J* = 8.4 Hz, 2H), 6.76 (d, *J* = 8.4 Hz, 2H), 5.50 (br, 1H), 4.72 (dd, *J* = 3.2 and 13.0 Hz, 1H), 4.49 (d, *J* = 13.0 Hz, 1H), 2.33 (d, *J* = 2.2 Hz, 1H), 2.23 (s, 3H). <sup>13</sup>C NMR δ 143.7, 138.6, 137.1, 129.8, 128.2, 127.7, 126.8, 123.0, 122.6, 113.1, 82.6, 72.4, 55.4, 54.0, 20.3. HRMS (EI) Calcd. for C<sub>17</sub>H<sub>15</sub>N [M]: 233.1204. Found: 233.1205. [α]<sub>D</sub><sup>25</sup> = - 149.0 (*c* = 0.490, CHCl<sub>3</sub>). The enantiomeric excess of **2b** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min, λ = 254 nm, retention time: 7.2 min (minor) and 11.5 min (major), 92% *ee*.

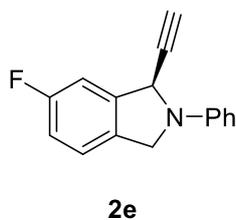


**1-ethynyl-2-(4-fluorophenyl)isoindoline (2c):** Isolated yield 79% (with **L4**). A white solid, m.p. 93.8 °C (decomp.). <sup>1</sup>H NMR δ 7.50-7.47 (m, 1H), 7.38-7.35 (m, 3H), 7.05 (m, 2H), 6.82 (dd, *J* = 4.3 and 9.2 Hz, 2H), 5.55 (br, 1H), 4.78 (dd, *J* = 3.1 and 12.9 Hz, 1H), 4.54 (d, *J* = 12.9 Hz, 1H), 2.42 (d, *J* = 1.9 Hz, 1H). <sup>13</sup>C NMR δ 156.0 (d, <sup>1</sup>*J*<sub>C-F</sub> = 235.3 Hz), 142.5 (d, <sup>4</sup>*J*<sub>C-F</sub> = 1.7 Hz), 138.5, 136.9, 128.3, 127.8, 123.1, 122.6, 115.7 (d, <sup>2</sup>*J*<sub>C-F</sub> = 22.3 Hz), 113.6 (d, <sup>3</sup>*J*<sub>C-P</sub> = 7.2 Hz), 82.4, 72.5, 55.6, 54.2. Anal. Calcd. for C<sub>16</sub>H<sub>12</sub>FN: C, 80.99; H, 5.10; N, 5.90. Found: C, 80.83; H, 5.22; N, 5.65. [α]<sub>D</sub><sup>25</sup> = - 150.9 (*c* = 0.470, CHCl<sub>3</sub>). The enantiomeric excess of **2c** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5,

flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, retention time: 7.6 min (minor) and 13.7 min (major), 95% *ee*.

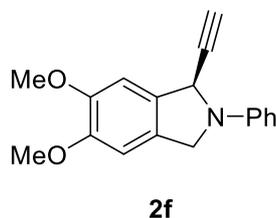


**2-(4-bromophenyl)-1-ethynylisoindoline (2d)**: Isolated yield 89% (with **L5**). A white solid, m.p. 163.2 °C (decomp.). <sup>1</sup>H NMR  $\delta$  7.49-7.46 (m, 1H), 7.42-7.34 (m, 5H), 6.76 (d,  $J$  = 9.2 Hz, 2H), 5.55 (br, 1H), 4.75 (dd,  $J$  = 3.1 and 12.8 Hz, 1H), 4.54 (d,  $J$  = 12.8 Hz, 1H), 2.41 (d,  $J$  = 2.2 Hz, 1H). <sup>13</sup>C NMR  $\delta$  144.8, 138.2, 136.6, 131.9, 128.3, 127.9, 123.1, 122.6, 114.5, 109.6, 82.0, 72.7, 55.2, 53.7. Anal. Calcd. for C<sub>16</sub>H<sub>12</sub>BrN: C, 64.45; H, 4.06; N, 4.70. Found: C, 64.23; H, 4.19; N, 4.46.  $[\alpha]^{25}_D = -146.2$  ( $c$  = 0.455, CHCl<sub>3</sub>). The enantiomeric excess of **2d** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, retention time: 8.4 min (minor) and 15.7 min (major), 96% *ee*.



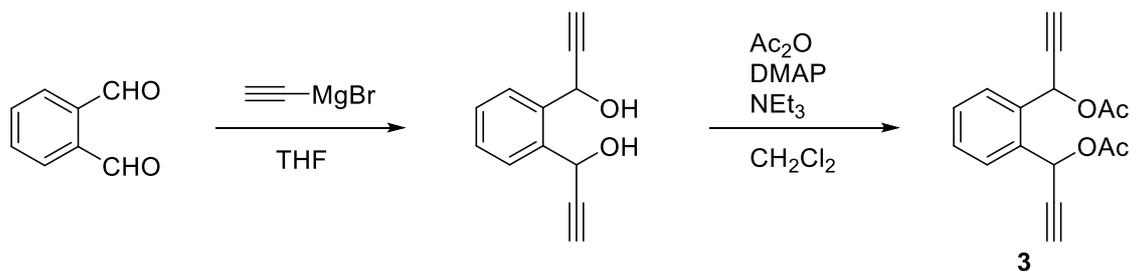
**1-ethynyl-6-fluoro-2-phenylisoindoline (2e)**: Isolated yield 81% (with **L4**). A white solid, m.p. 139.8 °C (decomp.). <sup>1</sup>H NMR  $\delta$  7.37-7.23 (m, 3H), 7.19-7.15 (m, 1H), 7.08-7.00 (m, 1H), 6.89-6.80 (m, 3H), 5.56 (br, 1H), 4.74 (dd,  $J$  = 3.2 and 12.6 Hz, 1H), 4.52 (d,  $J$  = 12.6 Hz, 1H), 2.42 (d,  $J$  = 1.9 Hz, 1H). <sup>13</sup>C NMR  $\delta$  162.7 (d, <sup>1</sup> $J_{C-F}$  = 244.8 Hz), 145.8, 140.4 (d, <sup>3</sup> $J_{C-F}$  = 8.9 Hz), 132.5 (d, <sup>4</sup> $J_{C-F}$  = 2.8 Hz), 129.3, 123.9 (d, <sup>3</sup> $J_{C-F}$  = 8.9 Hz), 117.6, 115.6 (d, <sup>2</sup> $J_{C-F}$  = 22.9 Hz), 112.9, 110.3 (d, <sup>2</sup> $J_{C-F}$  = 24.0 Hz), 81.9, 72.8, 55.1 (d, <sup>4</sup> $J_{C-F}$  = 2.8 Hz), 53.2. HRMS (EI) Calcd. for C<sub>16</sub>H<sub>12</sub>NF [M]: 237.0954. Found: 237.0950.  $[\alpha]^{25}_D = -156.8$  ( $c$  = 0.555, CHCl<sub>3</sub>).

The enantiomeric excess of **2e** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 97/3, flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, retention time: 9.5 min (major) and 11.6 min (minor), 98% *ee*.



**1-ethynyl-5,6-dimethoxy-2-phenylisoindoline (2f)**: Isolated yield 79% (with **L5**). A white solid, m.p. 149.3 °C (decomp.). <sup>1</sup>H NMR  $\delta$  7.34-7.30 (m, 2H), 6.96 (s, 1H), 6.88-6.78 (m, 4H), 5.52 (br, 1H), 4.73 (dd, *J* = 3.5 and 12.3 Hz, 1H), 4.50 (d, *J* = 12.3 Hz, 1H), 3.92 (s, 3H), 3.89 (s, 3H), 2.42 (d, *J* = 2.2 Hz, 1H). <sup>13</sup>C NMR  $\delta$  149.7, 149.3, 145.9, 130.0, 129.2, 128.6, 117.2, 112.7, 105.7, 105.3, 82.7, 72.3, 56.12, 56.10, 55.2, 53.8. HRMS (EI) Calcd. for C<sub>18</sub>H<sub>17</sub>NO<sub>2</sub> [M]: 279.1259. Found: 279.1246.  $[\alpha]_D^{25} = -120.2$  (*c* = 0.415, CHCl<sub>3</sub>). The enantiomeric excess of **2f** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, retention time: 27.0 min (major) and 29.3 min (minor), 96% *ee*.

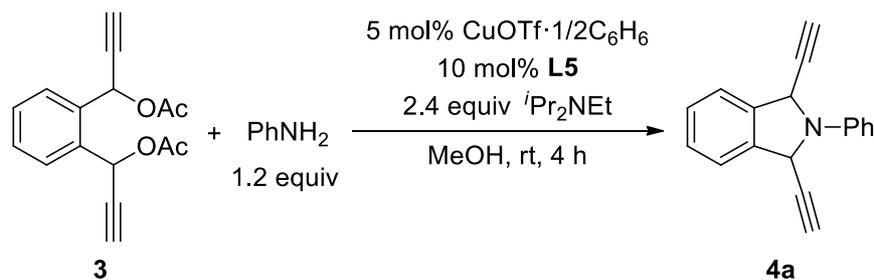
### Preparation of 1,1'-(1,2-phenylene)-bis(prop-2-yne-1,1-diyl) Diacetate (**3**).



To a flame-dried Schlenk flask was placed ethynylmagnesium bromide (0.5 M in THF, 24 mL, 12 mmol). *o*-Phthalaldehyde (405.5 mg, 3.02 mmol) was added at room temperature, and the resulting mixture was stirred at 60 °C for 2 h. After the reaction, water (50 mL) was added, and the resulting mixture was extracted with EtOAc (30 mL x 3). The combined organic layer was dried over anhydrous  $\text{MgSO}_4$ . After concentration *in vacuo*, the residue was passed through a short silicagel pad with hexane/EtOAc (7/3) to give 1,1'-(1,2-phenylene)-bis(prop-2-yne-1-ol) as a pale yellow oil (549.4 mg, 2.95 mmol).

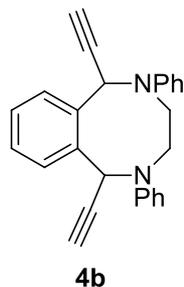
To a flame-dried Schlenk flask were placed  $\text{CH}_2\text{Cl}_2$  (6.0 mL), DMAP (36.9 mg, 0.302 mmol),  $\text{NEt}_3$  (625  $\mu\text{L}$ , 4.55 mmol), and 1,1'-(1,2-phenylene)-bis(prop-2-yne-1-ol) (549.4 mg, 2.95 mmol).  $\text{Ac}_2\text{O}$  (425  $\mu\text{L}$ , 4.53 mmol) dissolved in  $\text{CH}_2\text{Cl}_2$  (1.0 mL) was added dropwise, and the resulting mixture was stirred at room temperature for 5 h. After the reaction, water (50 mL) was added, and the resulting mixture was extracted with  $\text{CH}_2\text{Cl}_2$  (30 mL x 3). The combined organic layer was dried over anhydrous  $\text{MgSO}_4$ . After concentration *in vacuo*, the residue was purified by column chromatography ( $\text{SiO}_2$ ) with hexane/EtOAc (7/3) to give **3**<sup>S5</sup> as a white solid (716.4 mg, 2.65 mmol, 88% isolated yield).

### Double Propargylic Amination of Propargylic Diacetate with Amines.



A typical experimental procedure for the reaction of 1,1'-(1,2-phenylene)-bis(prop-2-ynyl)-1,1-diyl diacetate (**3**) with aniline is described below. In a 20 mL Schlenk flask were placed CuOTf·1/2C<sub>6</sub>H<sub>6</sub> (2.4 mg, 0.010 mmol) and (*S*)-Ph-pybox (**L5**) (7.3 mg, 0.020 mmol) under N<sub>2</sub>. Anhydrous methanol (1.0 mL) was added, and then the mixture was magnetically stirred at 60 °C for 1 h. After the solution was cooled to room temperature, **3** (53.0 mg, 0.20 mmol) in anhydrous methanol (1.0 mL), aniline (22 μL, 0.24 mmol), and diisopropylethylamine (84 μL, 0.48 mmol) were added under N<sub>2</sub>, and the reaction was kept at room temperature for 4 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography (SiO<sub>2</sub>) with hexane/ethyl acetate (93/7) to give 1,3-diethynyl-2-phenylisoindoline (**4a**) as a white solid (33.3 mg, 0.137 mmol, 70% isolated yield, *meso*-**4a**/*dl*-**4a** = 5.0/1). <sup>1</sup>H NMR (*meso*-isomer): δ 7.51-7.34 (m, 6H), 7.15-7.07 (m, 2H) 6.91-6.85 (m, 1H), 5.54 (br, 2H), 2.45 (br, 2H). <sup>1</sup>H NMR (*dl*-isomer): δ 5.76 (br, 2H), 2.36 (br, 2H). <sup>13</sup>C NMR (*meso*-isomer): δ 145.0, 137.6, 129.3, 128.8, 123.1, 118.3, 113.4, 82.4, 72.6, 55.4. <sup>13</sup>C NMR (*dl*-isomer): δ 143.7, 137.8, 123.2, 118.6, 115.4, 81.7, 73.3, 54.6. HRMS (EI) Calcd. for C<sub>18</sub>H<sub>13</sub>N [M]: 243.1048. Found: 243.1038. The enantiomeric excess of *dl*-**4a** was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min, λ = 254 nm, retention time: 13.4 min (minor) and 29.1 min (major), 75% *ee*.

## Spectroscopic Data and Isolated Yield of Other Product.



**1,6-diethynyl-2,5-diphenyl-1,2,3,4,5,6-hexahydrobenzo[f][1,4]diazocine (4b):** Isolated yield 68% (*meso-4b/dl-4b* = 7.7/1). A white solid.  $^1\text{H}$  NMR (*meso*-isomer):  $\delta$  7.76-7.73 (m, 2H), 7.36-7.33 (m, 2H) 7.15-7.06 (m, 4H), 6.76-6.65 (m, 6H), 5.70 (d,  $J$  = 2.2 Hz, 2H), 4.23-4.13 (m, 2H), 3.92-3.82 (m, 2H), 2.44 (d,  $J$  = 2.2 Hz, 2H).  $^1\text{H}$  NMR (*dl*-isomer):  $\delta$  8.01-7.97 (m, 2H), 5.57 (d,  $J$  = 2.3 Hz, 2H), 3.97 (br, 4H), 2.66 (d,  $J$  = 2.3 Hz, 2H).  $^{13}\text{C}$  NMR (*meso*-isomer):  $\delta$  145.5, 135.8, 130.0, 129.0, 128.2, 117.3, 113.1, 80.1, 77.4, 56.7, 46.8.  $^{13}\text{C}$  NMR (*dl*-isomer):  $\delta$  129.0, 128.1, 117.2, 112.5, 58.0, 49.1. HRMS (EI) Calcd. for  $\text{C}_{26}\text{H}_{22}\text{N}_2$  [M]: 362.1783. Found: 362.1781. The enantiomeric excess of *dl-4b* was determined by HPLC analysis; DAICEL Chiralpak IA, hexane/*i*PrOH = 95/5, flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, retention time: 6.8 min (minor) and 12.4 min (major), 66% *ee*.

## X-ray Diffraction Study of **2d**.

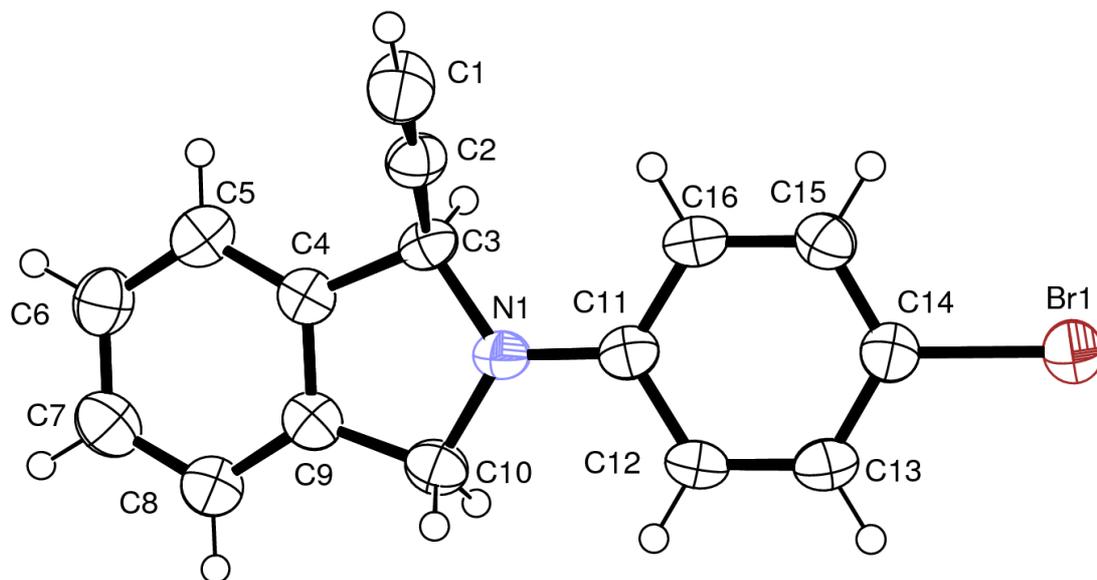
Diffraction data for (*R*)-2-(4-bromophenyl)-1-ethynylisoindoline (**2d**) were collected on a Rigaku R-AXIS RAPID imaging plate diffractometer with graphite-monochromated Mo-K $\alpha$  radiation ( $\lambda = 0.71075 \text{ \AA}$ ) with Varimax optics. Reflections were collected for the  $2\theta$  range of  $6^\circ$  to  $55^\circ$ . Intensity data were corrected for numerical absorptions (NUMABS)<sup>S7</sup>, and for Lorentz and polarization effects. A correction for secondary extinction<sup>S8</sup> was further applied (coefficient, 19(12)). The structure solution and refinements were carried out by using CrystalStructure package.<sup>S9</sup> The positions of all the non-hydrogen atoms were determined by direct methods (SIR97)<sup>S10</sup> and subsequent Fourier syntheses, and were refined on  $F_o^2$  with all the unique reflections by full-matrix least squares with anisotropic thermal parameters. All the hydrogen atoms were placed at the calculated positions with fixed isotropic parameters. Goodness of fit indicator  $[\sum w(|F_o| - |F_c|)^2 / (N_{\text{obs}} - N_{\text{params}})]^{1/2}$  were all refined to the value of 1.000. The atomic scattering factors were taken from reference S11, and anomalous dispersion effects were included.<sup>S12</sup> The values of  $\Delta f'$  and  $\Delta f''$  were taken from reference S13. The Flack parameter<sup>S14</sup> for **2d** was refined to the value of 0.010(14), which clearly suggests that the absolute configuration of the major isomer of **2d** is (*R*) as shown in Figure S1, where the ORTEP drawing of **2d** is depicted. Details of the crystals and data collection parameters of **2d** are summarized in Table S1.

CCDC 989441 (**2d**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).

**Table S1.** Crystallographic Data for **2d**.

	<b>2d</b>
chemical formula	C <sub>16</sub> H <sub>12</sub> BrN
formula weight	298.18
crystal size	0.42 × 0.11 × 0.08
color, habit	colorless, needle
temperature (°C)	−75
crystal system	orthorhombic
space group	<i>P</i> 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub> (#19)
<i>a</i> (Å)	4.08853(17)
<i>b</i> (Å)	11.1945(6)
<i>c</i> (Å)	27.8189(12)
<i>α</i> (deg)	90
<i>β</i> (deg)	90
<i>γ</i> (deg)	90
<i>V</i> (Å <sup>3</sup> )	1273.25(10)
<i>Z</i>	4
<i>d</i> <sub>calcd</sub> (g cm <sup>−3</sup> )	1.555
<i>F</i> (000)	600
<i>μ</i> (cm <sup>−1</sup> )	32.173
transmission factors range	0.440–0.773
measured reflections	10532
unique reflections	2885
<i>R</i> <sub>int</sub>	0.0594
refined parameters	177
<i>R</i> 1 ( <i>I</i> > 2σ( <i>I</i> )) <sup>a</sup>	0.0365
<i>wR</i> 2 (all data) <sup>b</sup>	0.0705
residual peaks (e Å <sup>−3</sup> )	+0.531/−0.657
CCDC number	989441

<sup>a</sup>  $R1 = \Sigma||F_o| - |F_c||/\Sigma|F_o|$ .    <sup>b</sup>  $wR2 = [\Sigma\{w(F_o^2 - F_c^2)^2\}/\Sigma w(F_o^2)^2]^{1/2}$ ,  $w = 4F_o^2/q\sigma(F_o^2)$ ,  $q = 1.847$ .



**Figure S1.** ORTEP drawing of **2d**. Thermal ellipsoids are given at the 50% probability level.

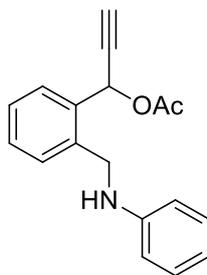
## References and Notes

- (S1) Tse, M. K.; Bhor, S.; Klawonn, M.; Anilkumar, G.; Jiao, H.; Döbler, C.; Spannenberg, A.; Mägerlein, W.; Hugl, H.; Beller, M. *Chem. Eur. J.* **2006**, *12*, 1855.
- (S2) Nishiyama, H.; Kondo, M.; Nakamura, T.; Itoh, K. *Organometallics* **1991**, *10*, 500.
- (S3) Meng, J.-C.; Fokin, V. V.; Finn, M. G. *Tetrahedron Lett.* **2005**, *46*, 4543.
- (S4) Ginotra, S. K.; Singh, V. K. *Org. Biomol. Chem.* **2007**, *5*, 3932.
- (S5) Sugimoto, Y.; Hanamoto, T.; Inanaga, J. *Appl. Organomet. Chem.* **1995**, *9*, 369.
- (S6) Teng, T.-M.; Das, A.; Huple, D. B.; Liu, R.-S. *J. Am. Chem. Soc.* **2010**, *132*, 12565.
- (S7) Higashi, T. *ABSCOR: empirical absorption correction based on Fourier series approximation*; Rigaku Corp.: Tokyo, Japan, 1995.
- (S8) Larson, A. C. In *Crystallographic Computing: Proceedings of an International Summer School organized by The Commission on Crystallographic Computing of the International Union of Crystallography and held in Ottawa, 4–11 August 1969*; Ahmed, F. R., Hall, S. R., Huber, C. P., Eds.; Munksgaard, Copenhagen, Denmark, 1970; pp. 291–294.
- (S9) (a) *CrystalStructure 4.0: Single Crystal Structure Analysis Software*; Rigaku Corp: Tokyo, Japan and MSC: The Woodlands, TX, 2010. (b) Carruthers, J. R.; Rollett, J. S.; Betteridge, P. W.; Kinna, D.; Pearce, L.; Larsen, A.; Gabe, E. *CRYSTALS Issue 11*; Chemical Crystallography Laboratory: Oxford, UK, 1999.
- (S10) Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G. L.; Giacovazzo, C.; Guagliardi, A.; Moliterni, A. G. G.; Polidori, G.; Spagna, R. *J. Appl. Crystallogr.* **1999**, *32*, 115.
- (S11) Cromer, D. T.; Waber, J. T. In *International Tables for X-ray Crystallography*; Ibers, J. A., Hamilton, W. C. Eds.; Kynoch Press: Birmingham, England, 1974; Vol. IV., Table 2.2 A.
- (S12) Ibers, J. A.; Hamilton, W. C. *Acta Crystallogr.* **1964**, *17*, 781.
- (S13) (a) Creagh, D. C.; McAuley, W. J. In *International Tables for X-ray Crystallography*; Wilson, A. J. C. Ed.; Kluwer Academic Publishers: Boston, MA, 1992; Vol. C. Table 4.2.6.8.

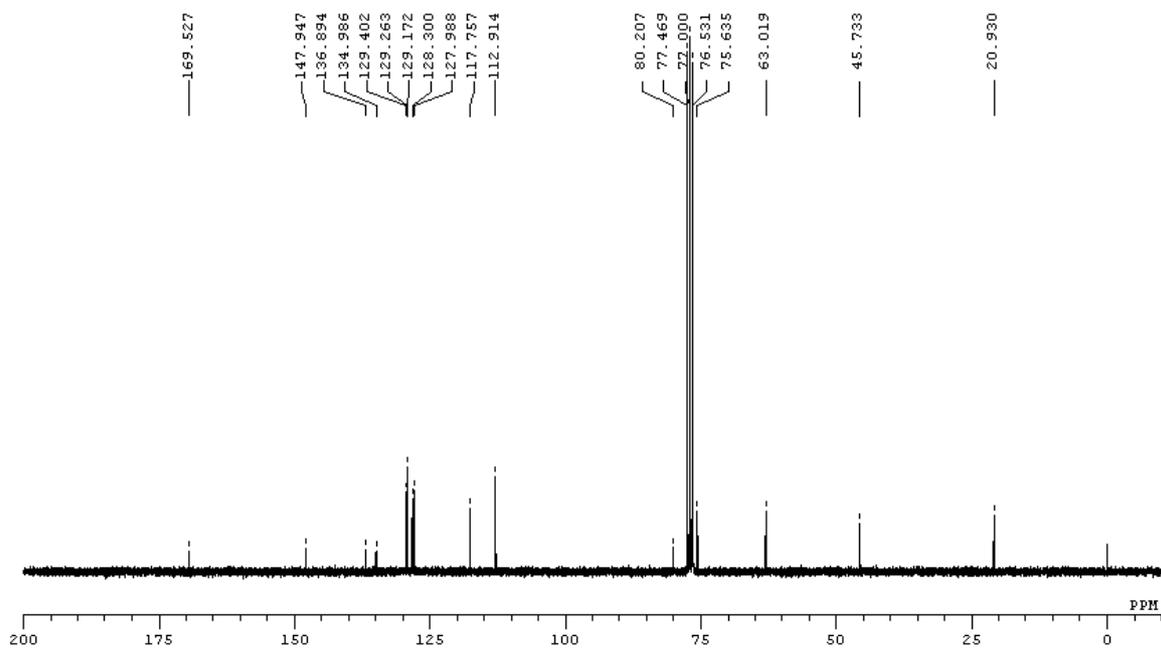
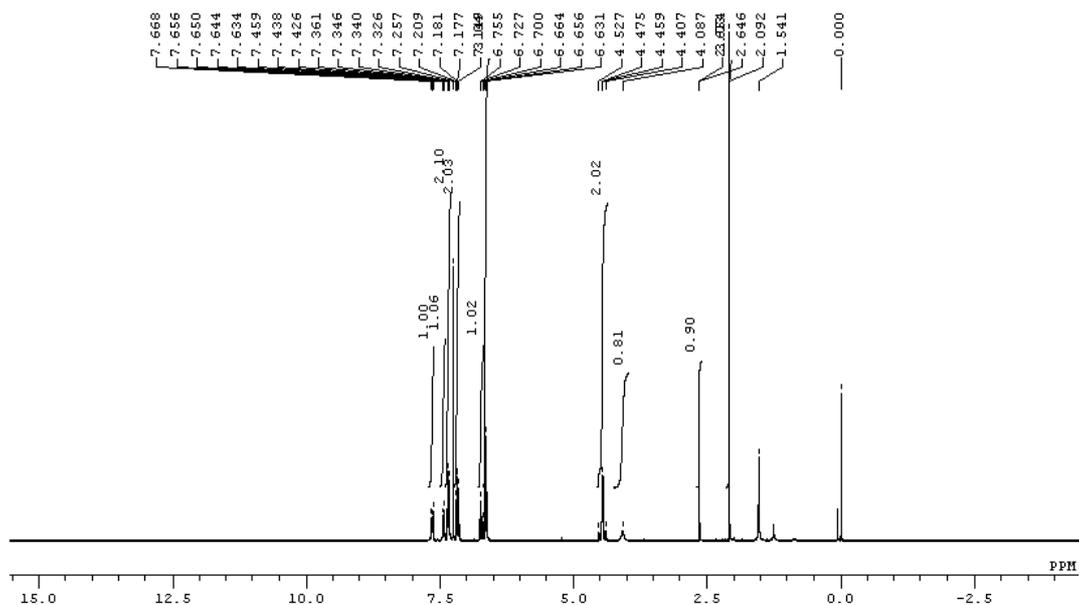
(b) Creagh, D. C.; Hubbell, J. H. In *International Tables for X-ray Crystallography*; Wilson, A. J. C. Ed.; Kluwer Academic Publishers: Boston, MA, 1992; Vol. C. Table 4.2.4.3.

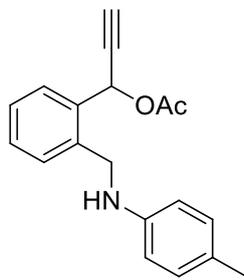
(S14) Flack, H. D. *Acta Crystallogr.* **1983**, A39, 876.

# <sup>1</sup>H and <sup>13</sup>C NMR Spectra

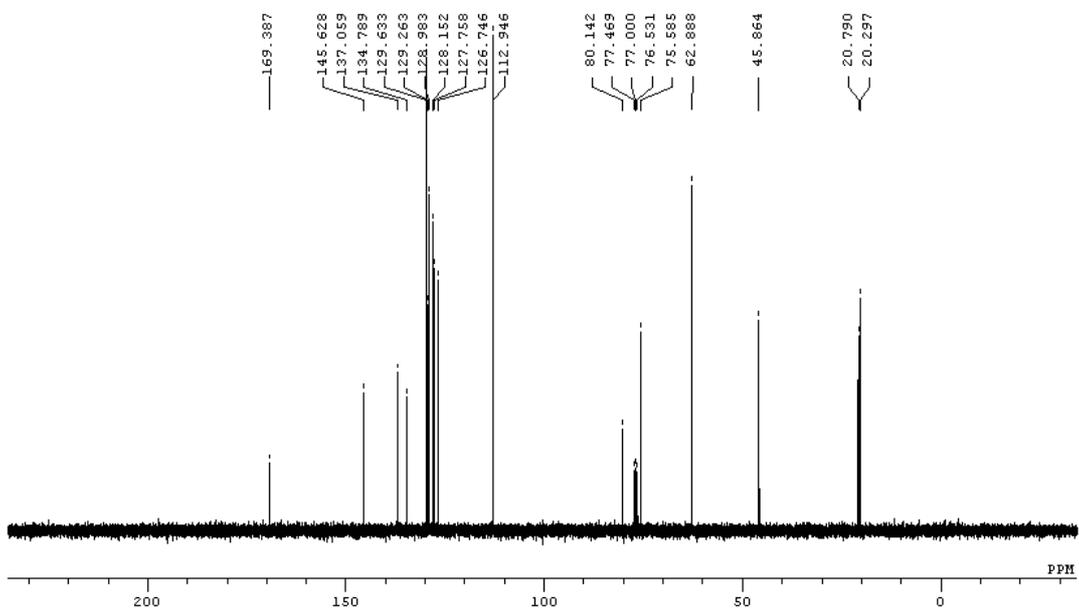
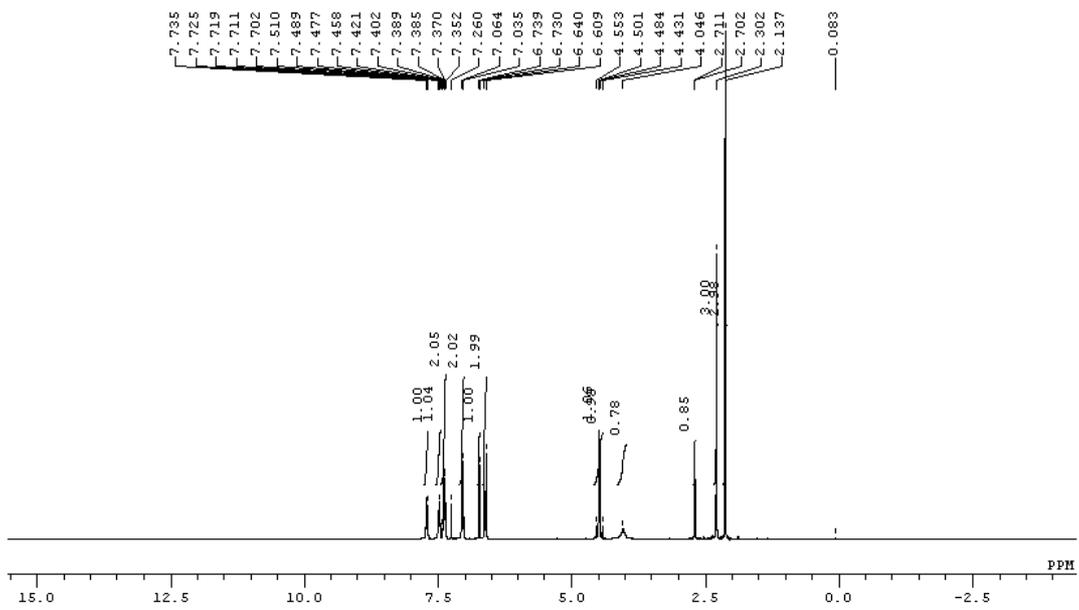


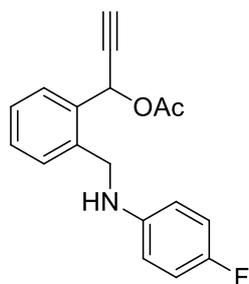
1a



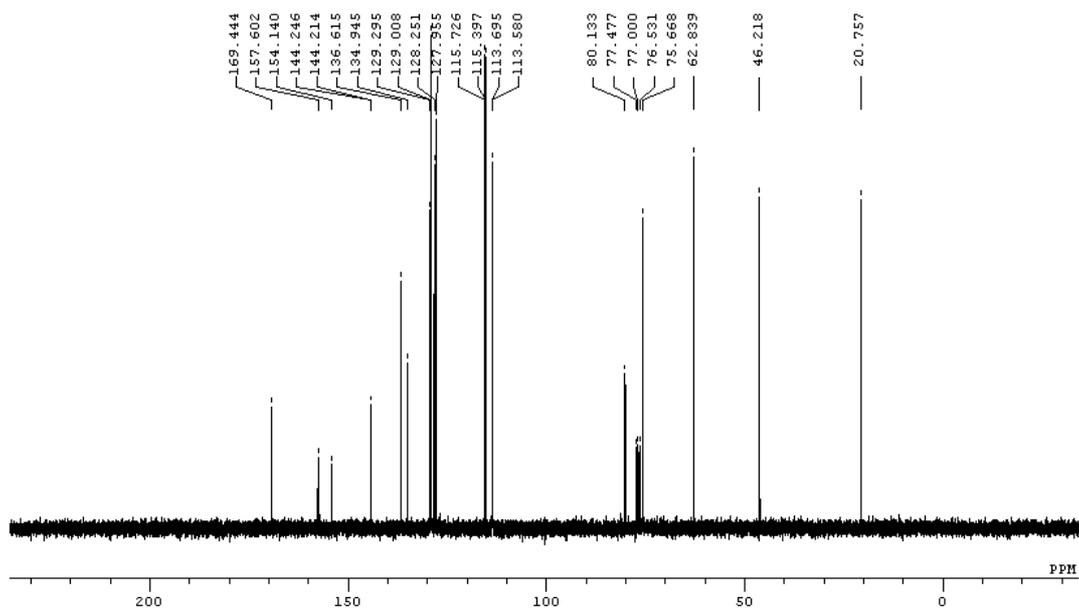
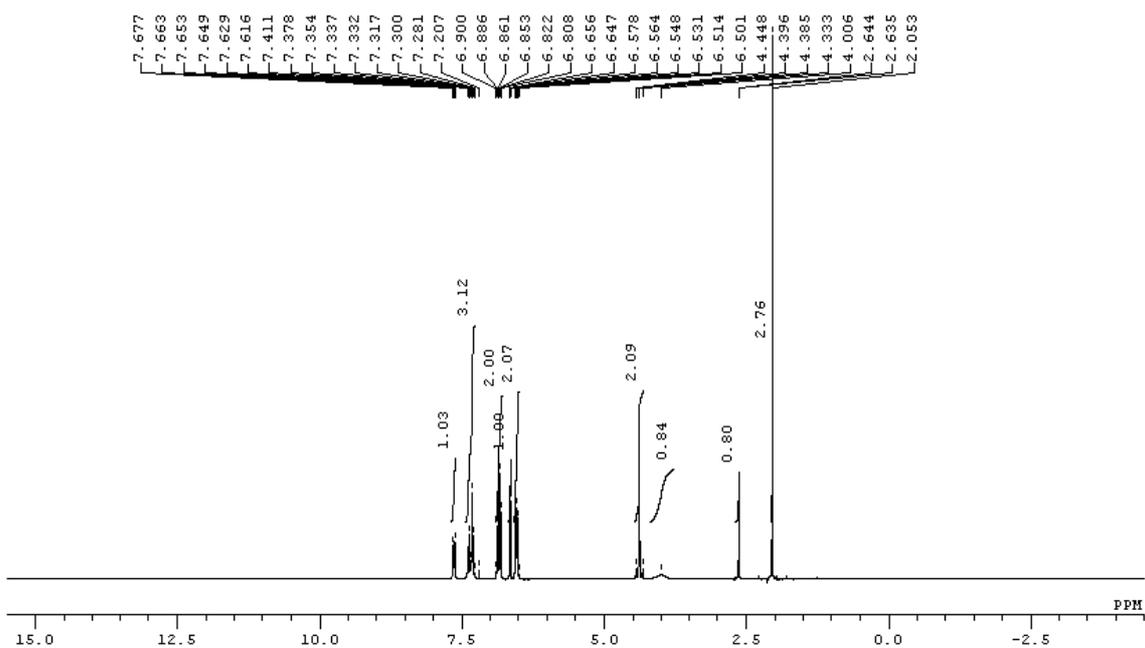


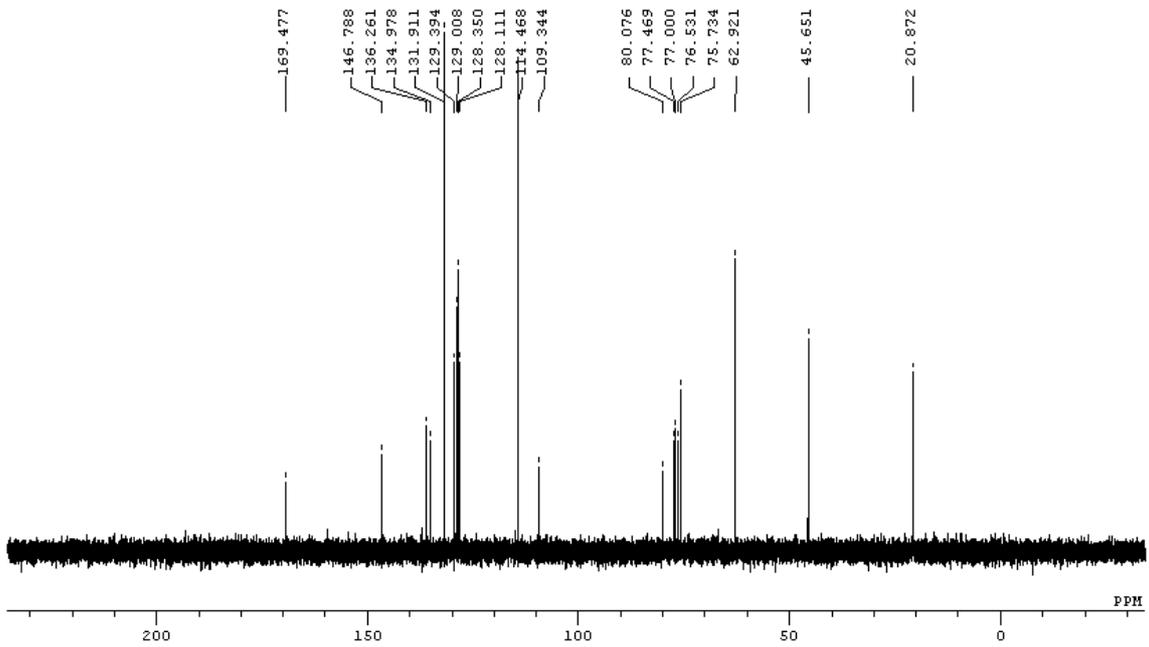
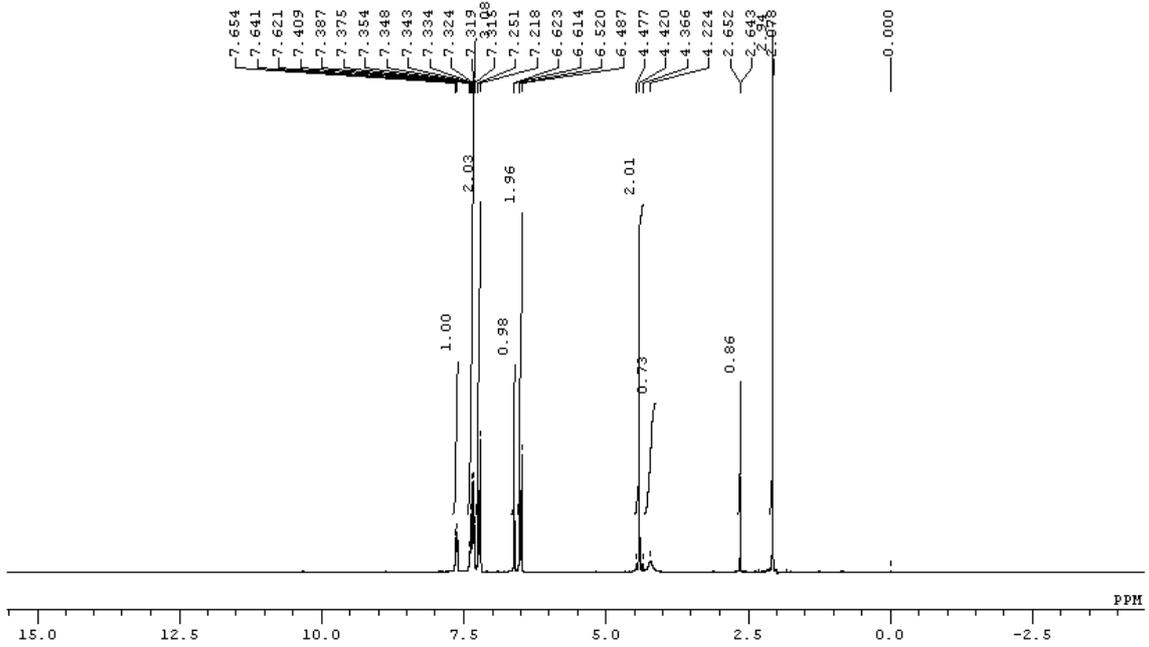
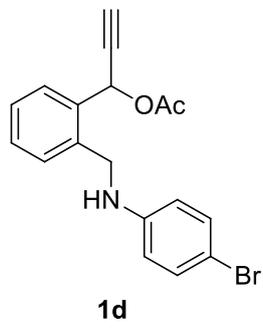
**1b**

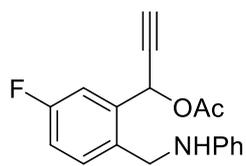




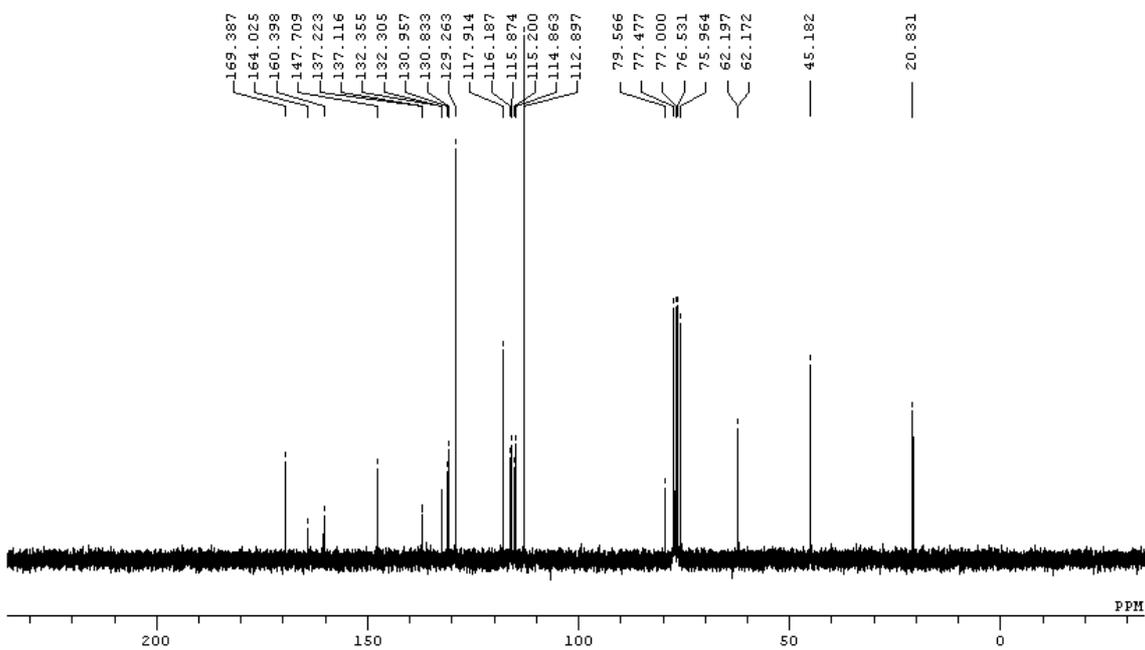
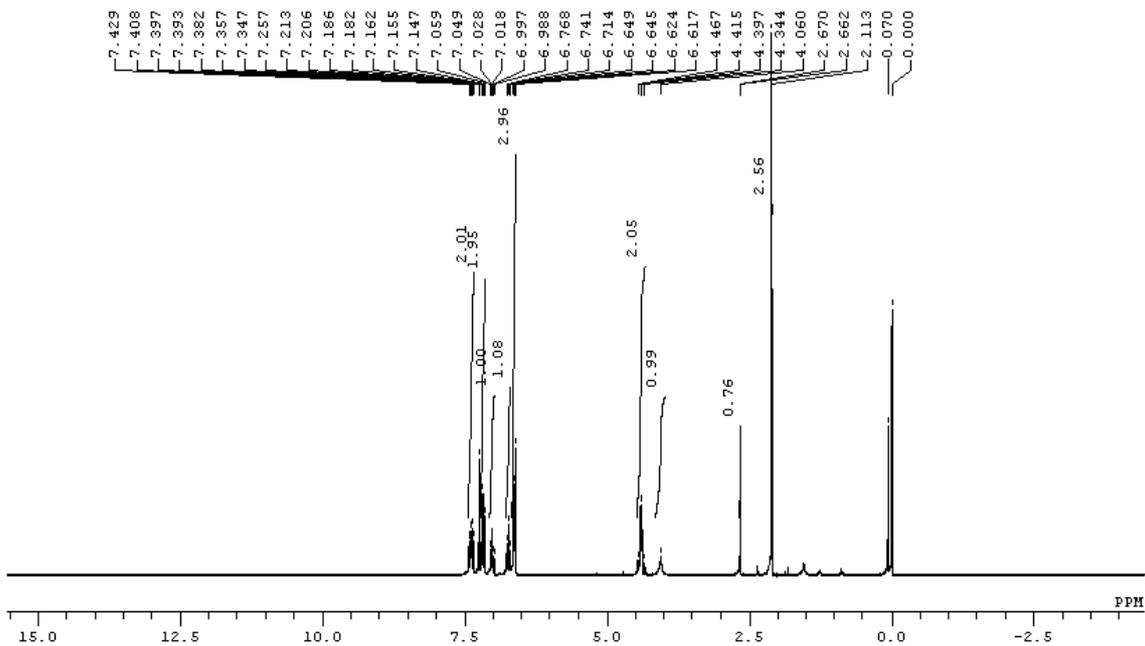
**1c**

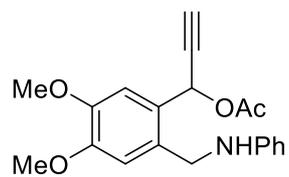




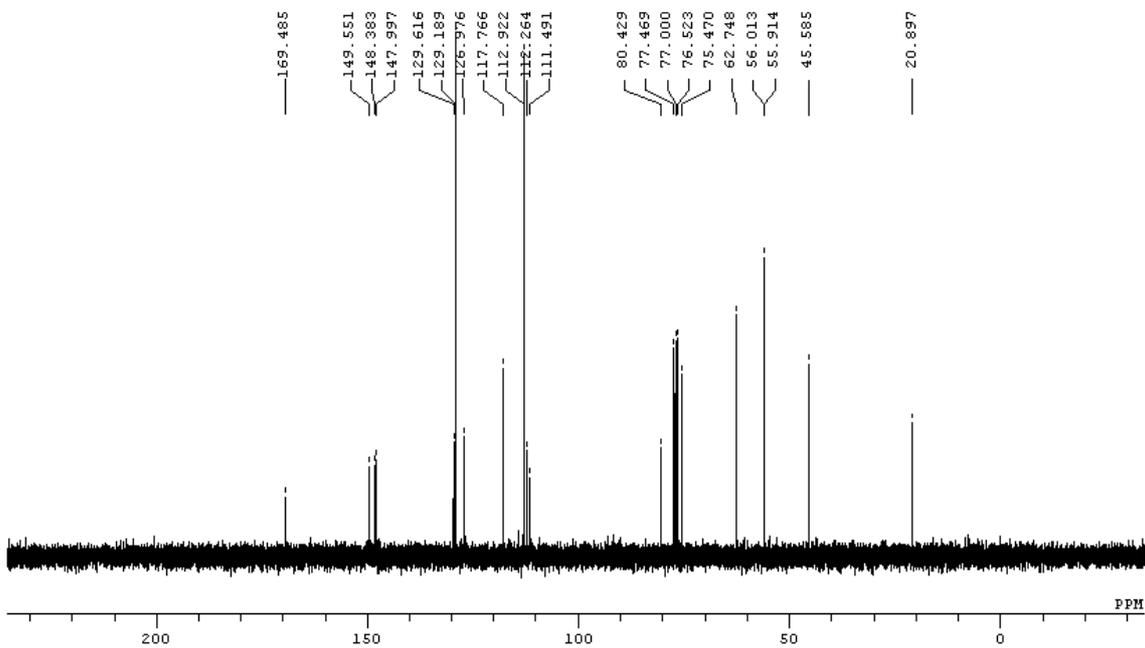
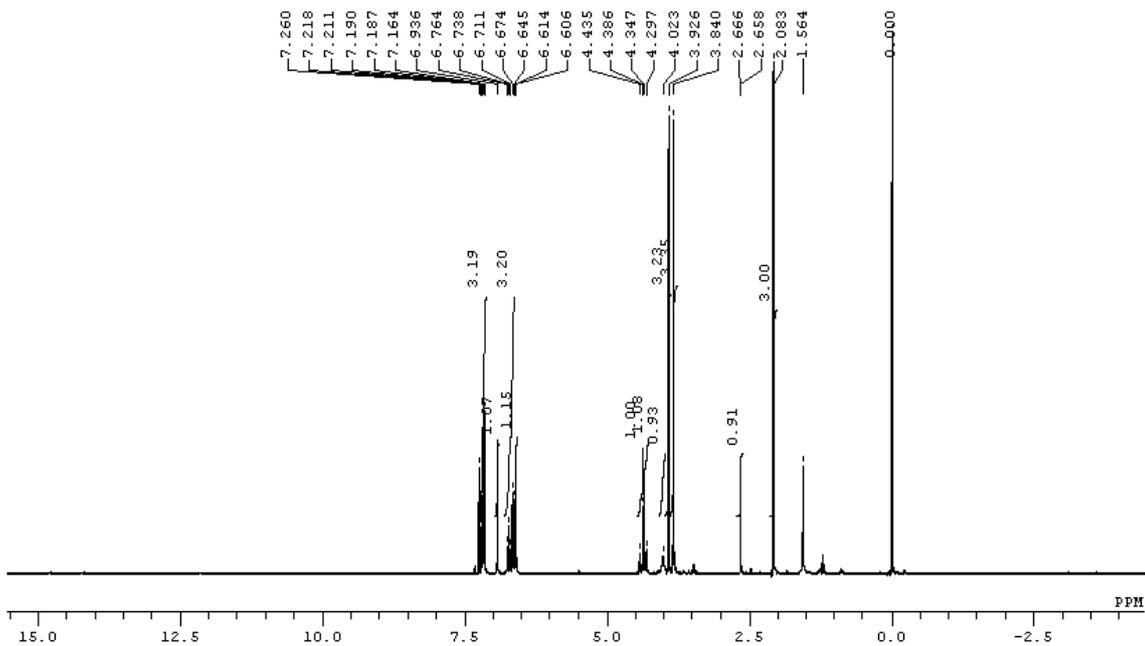


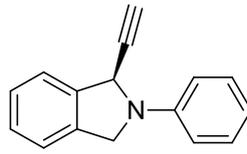
**1e**



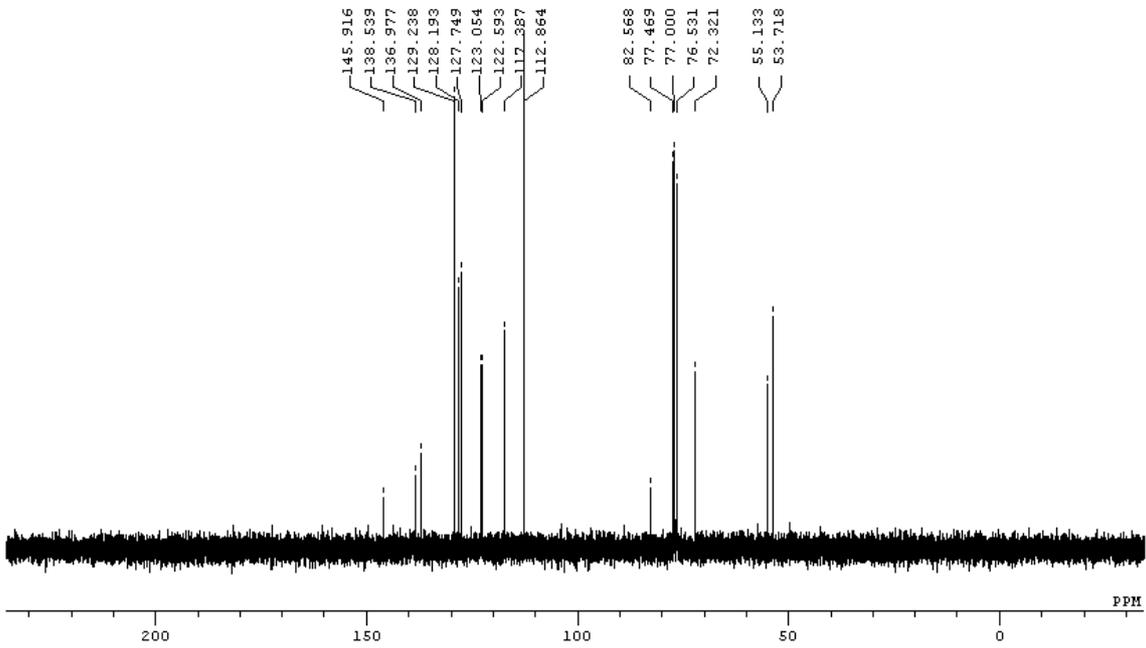
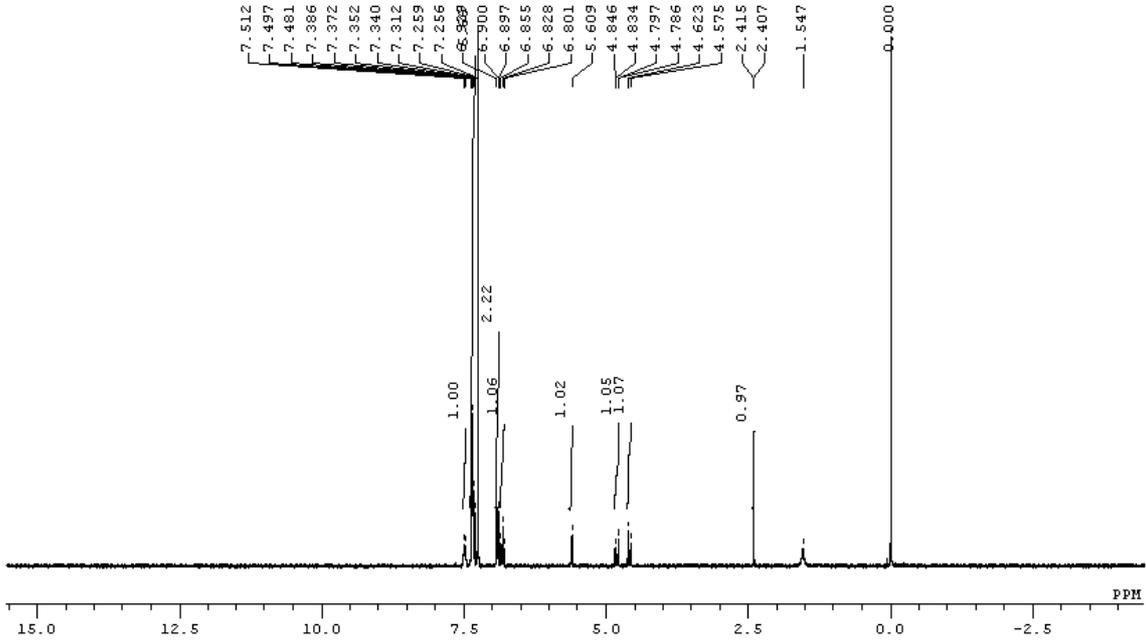


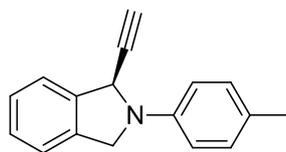
**1f**



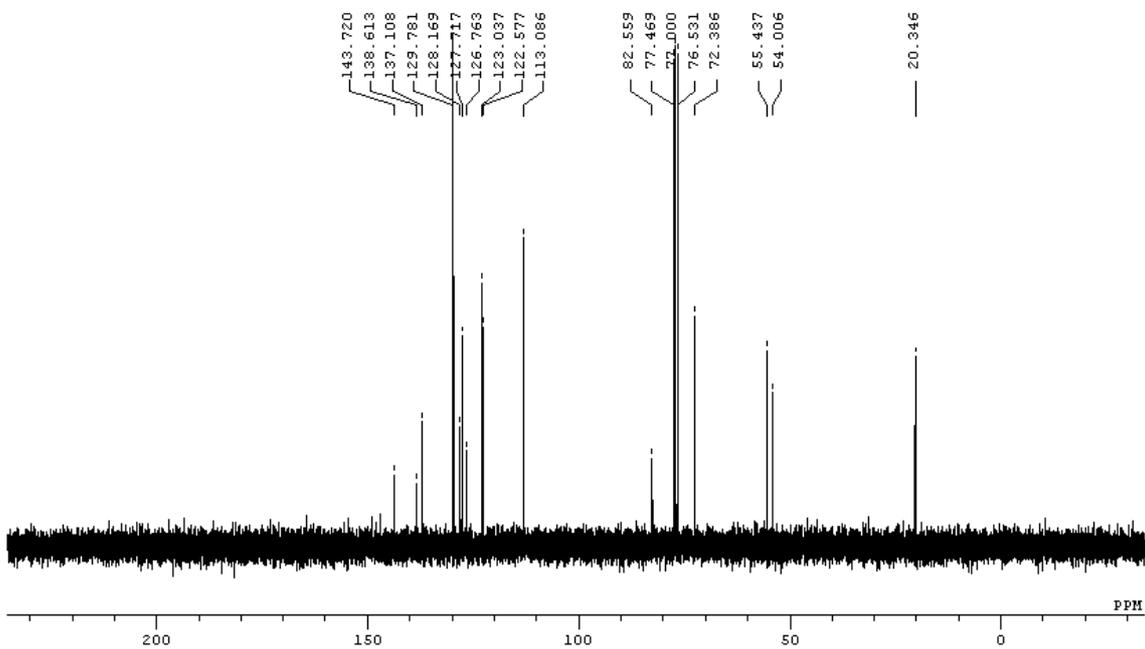
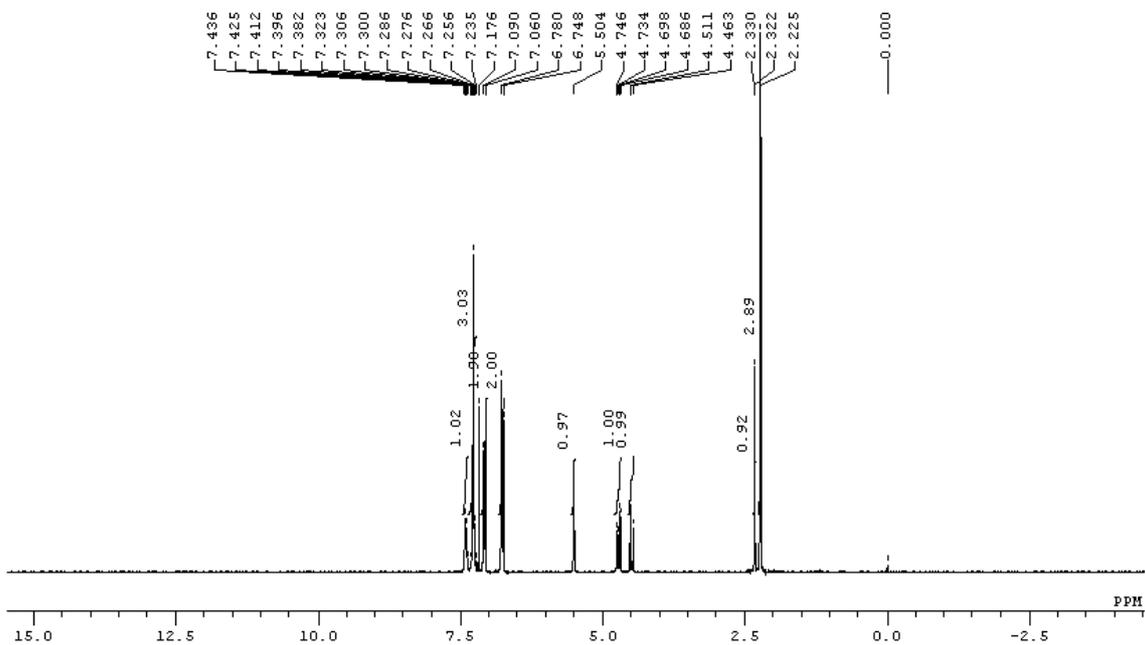


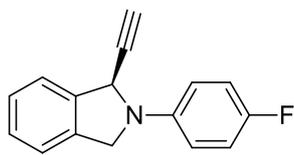
2a



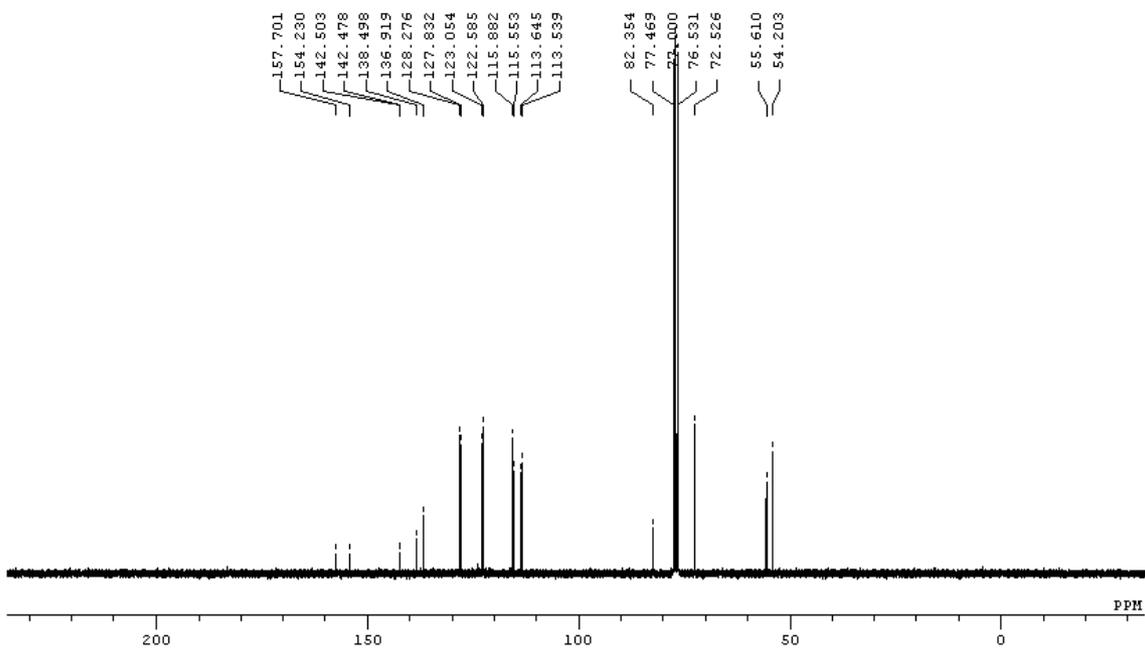
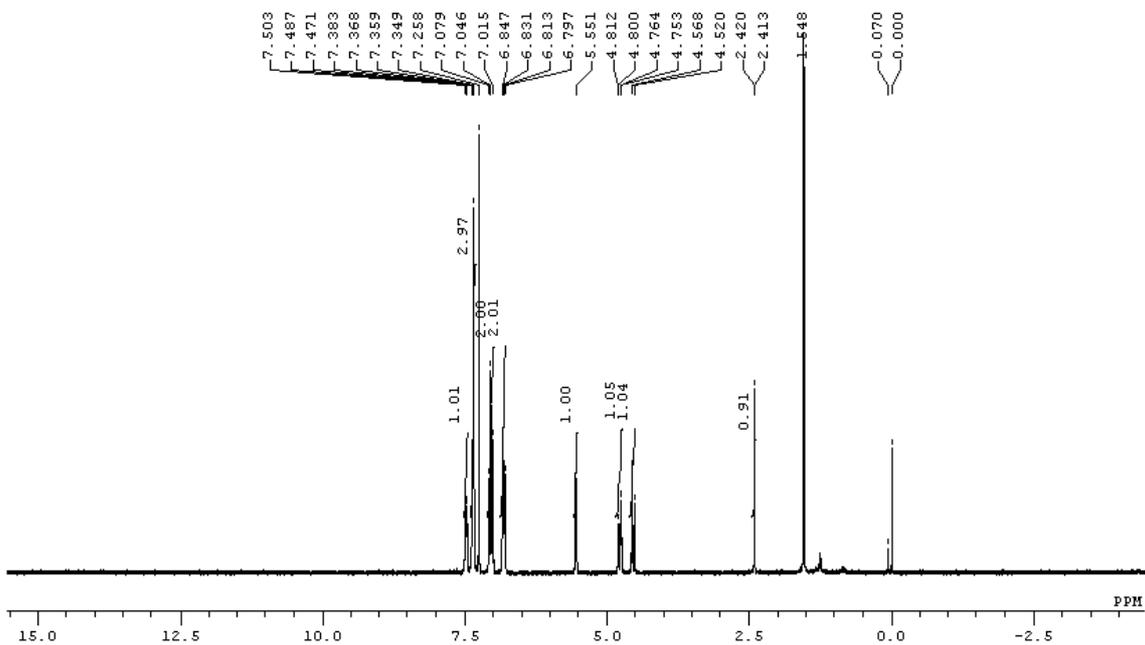


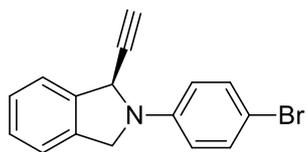
2b



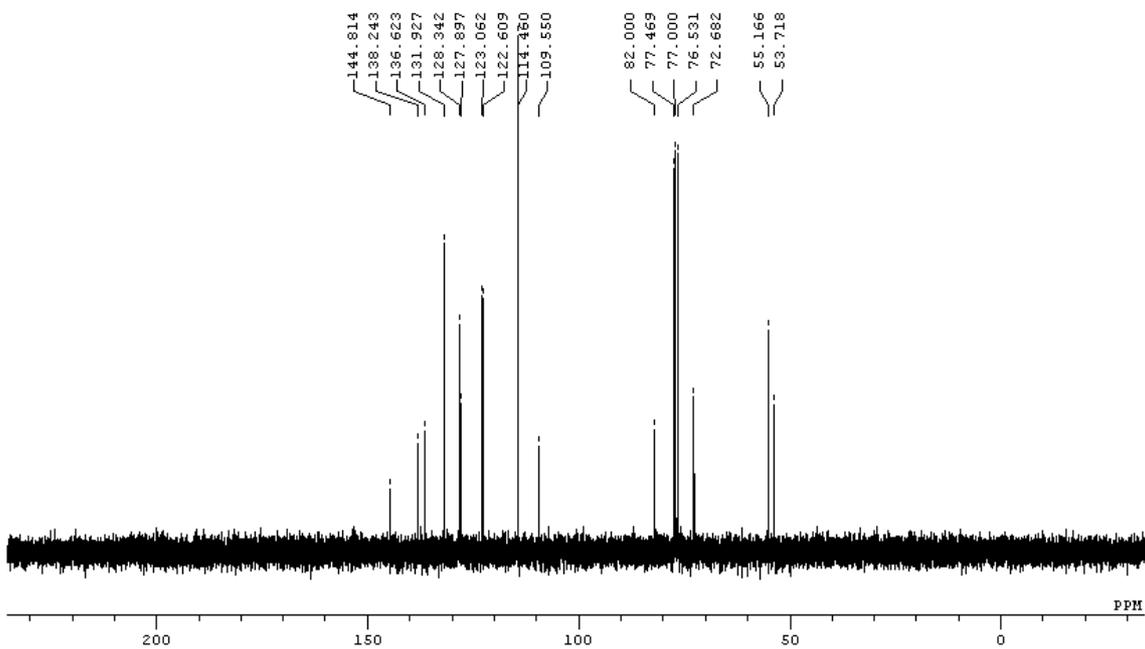
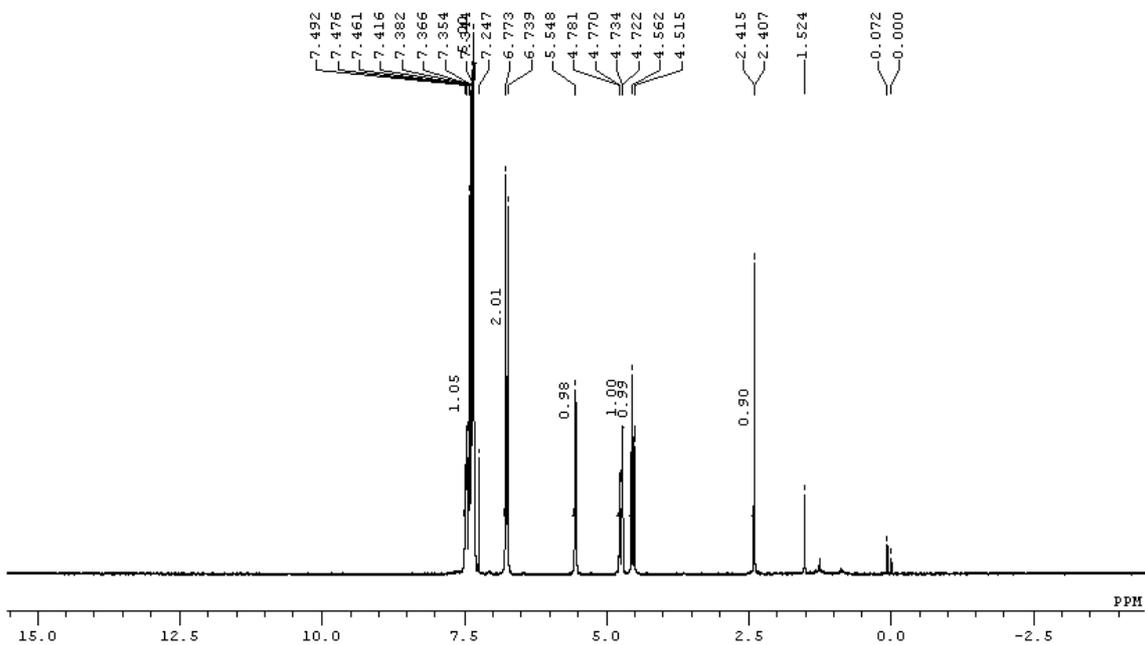


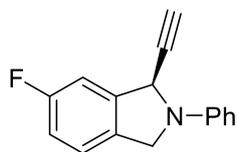
**2c**



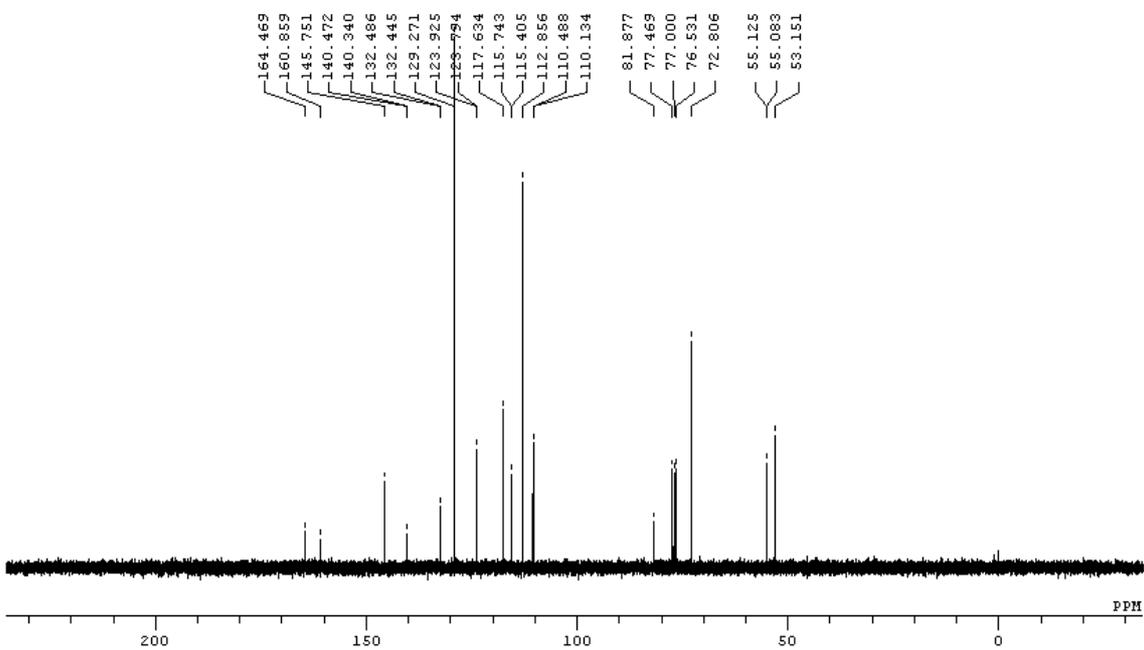
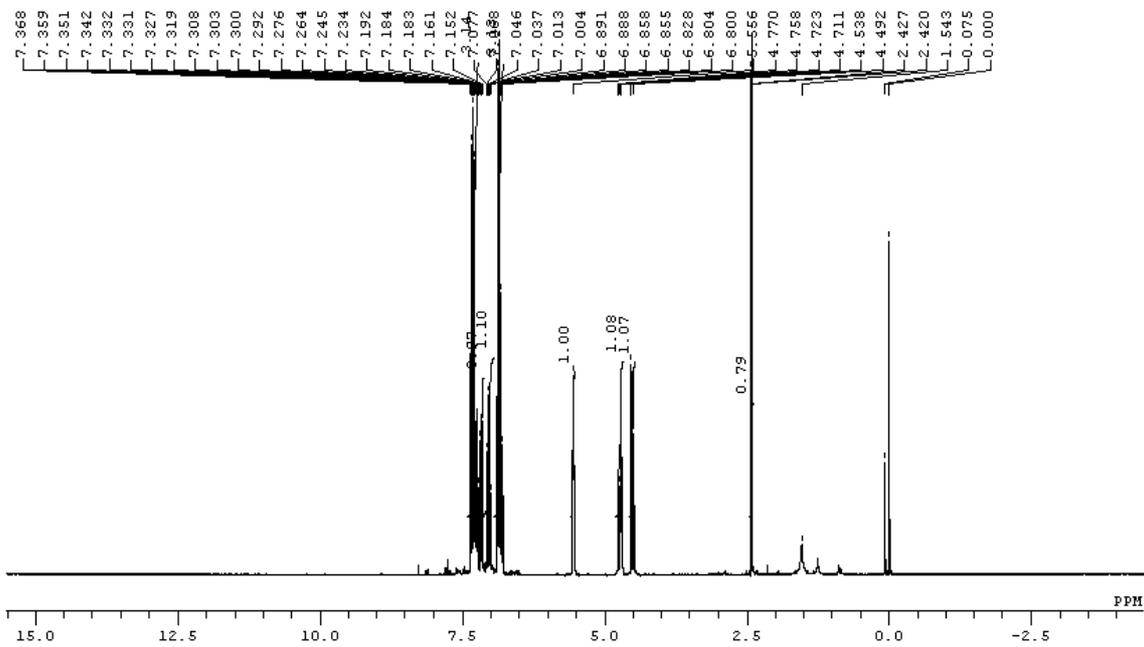


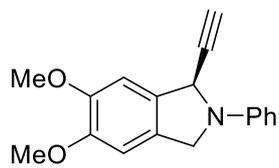
2d



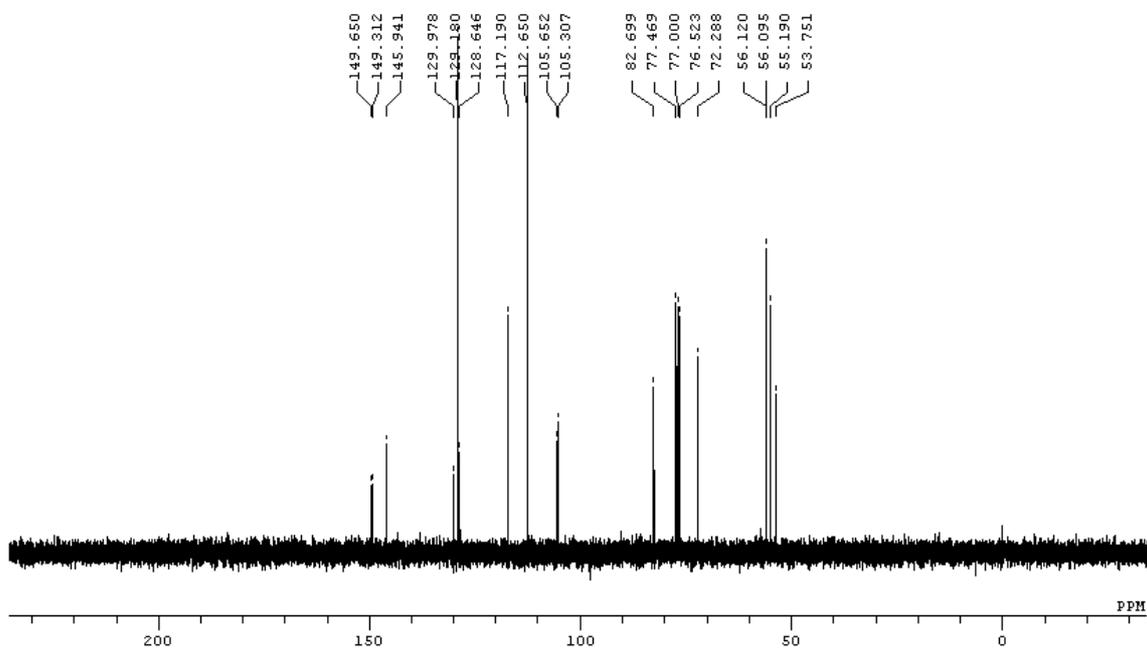
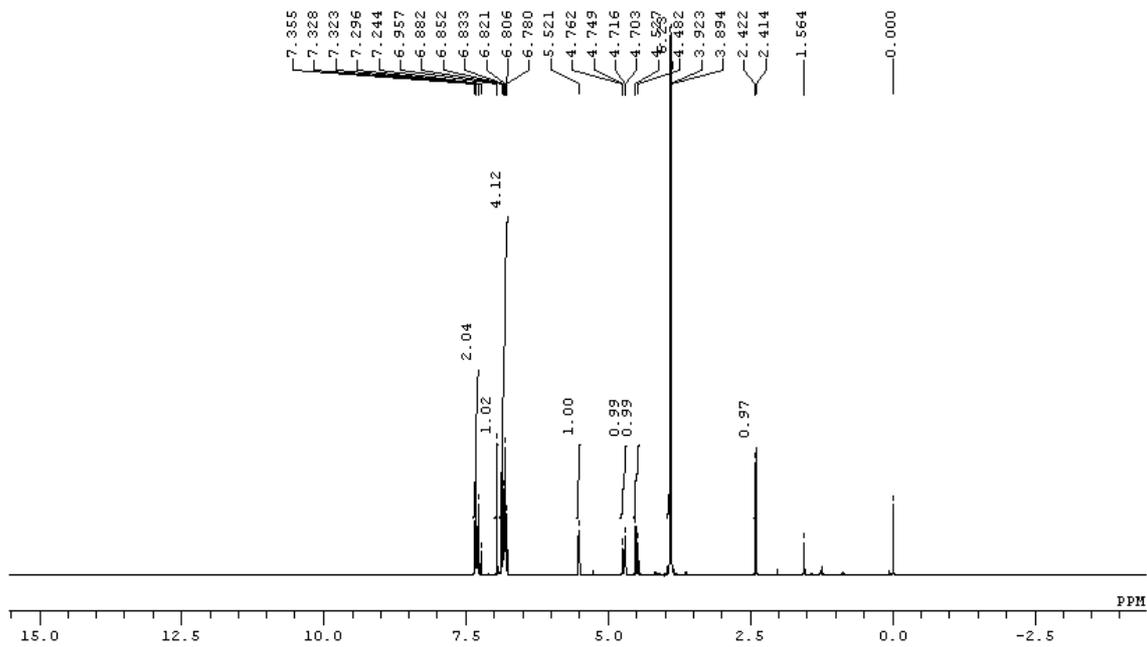


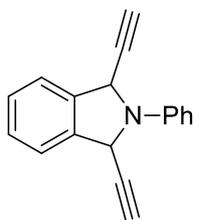
2e



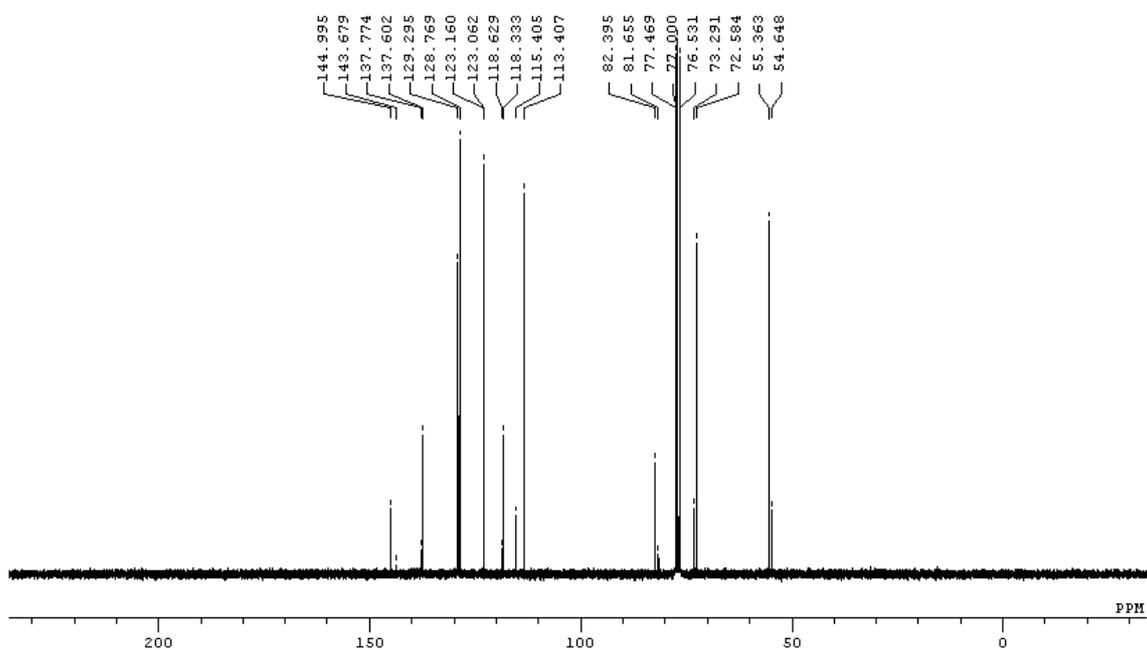
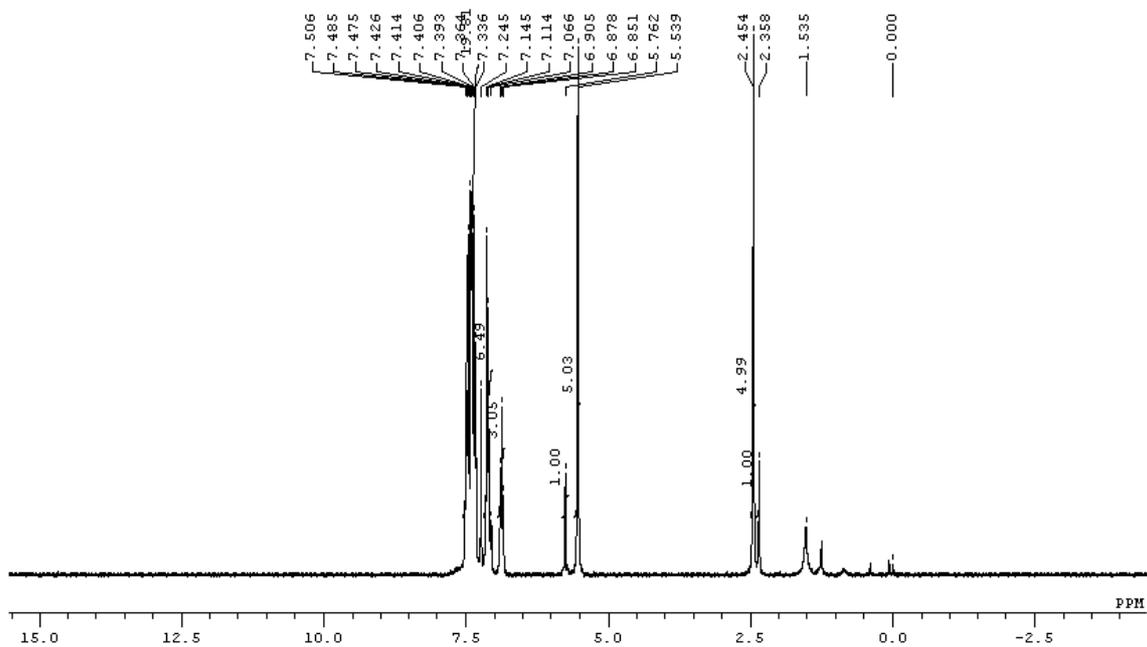


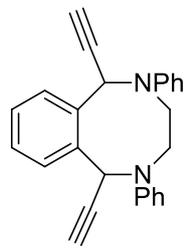
2f



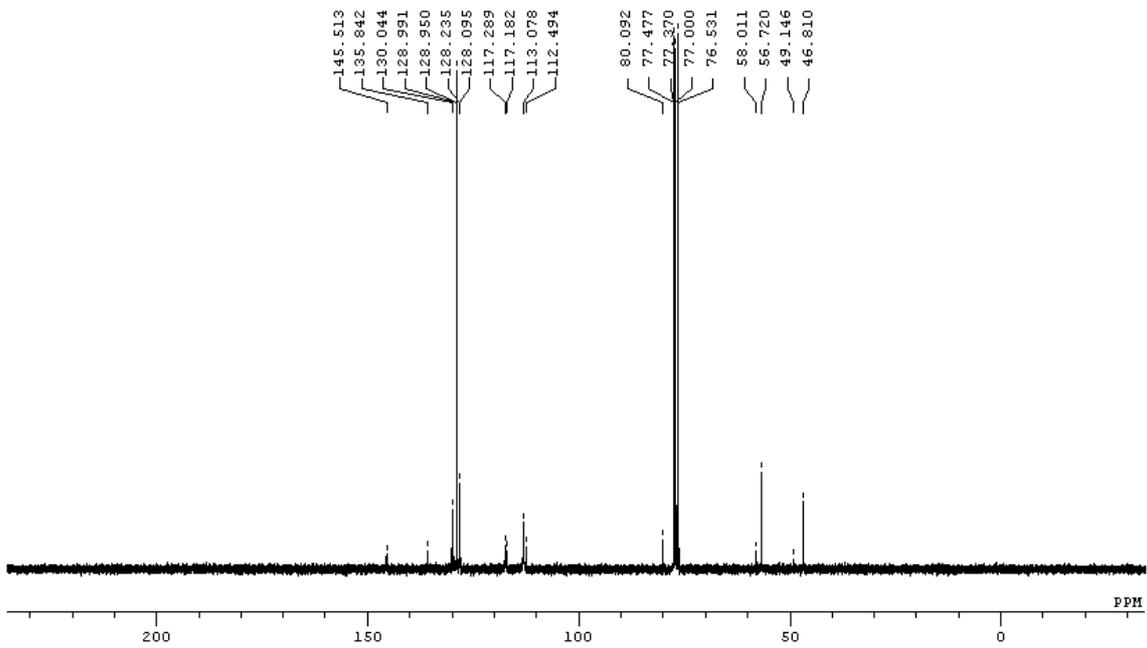
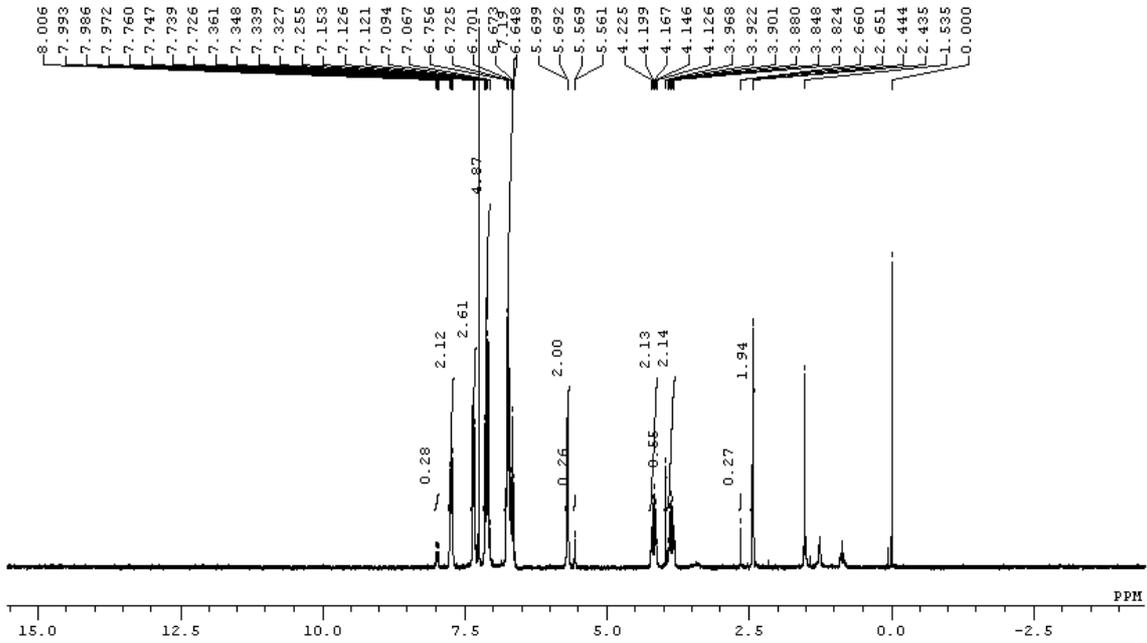


4a

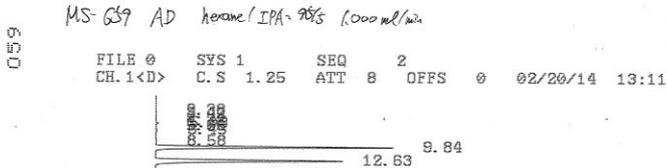




4b



**Charts of Propargylic Aminated Products by HPLC Analysis  
2a(rac)**



D-7500 INTEGRATOR REPORT

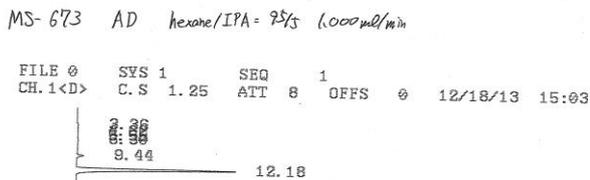
ANALYZED: 02/20/14 13:11 REPORTED: 02/20/14 13:26

SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 2

FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
8	9.84	2072754	50.194	BB
9	12.63	2056724	49.806	BB
TOTAL		4129478	100.000	
PEAK REJ :		50000		

2a



D-7500 INTEGRATOR REPORT

ANALYZED: 12/18/13 15:03 REPORTED: 12/18/13 15:20

SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 1

FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
5	9.44	60380	3.370	VB
8	12.18	1731319	96.630	BB
TOTAL		1791699	100.000	
PEAK REJ :		50000		

2b(rac)

MS-523 IA hexane/IPA = 95/5 1.000 mL/min

FILE # SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 8 OFFS @ 07/18/13 20:19

6.86	9.93
13.87	

D-7500 INTEGRATOR REPORT

ANALYZED: 07/18/13 20:19 REPORTED: 07/18/13 20:35  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 2  
FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
7	6.86	1738923	50.048	BV
10	9.93	1735572	49.952	BB
TOTAL		3474495	100.000	

PEAK REJ : 50000

2b

MS-51 IA hexane/IPA = 95/5 1.000 mL/min

FILE # SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 8 OFFS @ 07/19/13 20:11

7.23	11.49
18.77	

D-7500 INTEGRATOR REPORT

ANALYZED: 07/19/13 20:11 REPORTED: 07/19/13 20:26  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 1  
FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
5	7.23	92457	3.805	BB
8	11.49	2337592	96.195	VB
TOTAL		2430049	100.000	

PEAK REJ : 50000

2c(rac)

MS-536 IA hexane/IPA = 95/5 (1000 mL/min)

FILE # SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 8 OFFS 0 07/24/13 20:00

7.38	7.38
13.10	13.10
17.38	

D-7500 INTEGRATOR REPORT

ANALYZED: 07/24/13 20:00 REPORTED: 07/24/13 20:21  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 1

FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
6	7.38	2603234	50.388	BB
9	13.10	2563137	49.612	BB
TOTAL		5166371	100.000	
PEAK REJ :		50000		

2c

MS-685 IA hexane/IPA = 95/5 (1000 mL/min)

FILE # SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 8 OFFS 0 01/17/14 14:00

13.68	13.68
10.44	

D-7500 INTEGRATOR REPORT

ANALYZED: 01/17/14 14:00 REPORTED: 01/17/14 14:21  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 1

FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

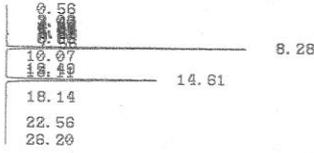
NO.	RT	AREA	CONC	BC
6	7.00	48387	2.673	BB
8	13.68	1689114	97.327	BB
TOTAL		1735511	100.000	
PEAK REJ :		10000		

113

2d(rac)

MS-491 IA hexane/IPA = 95/5 1.000 ml/min

FILE 0 SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 8 OFFS 0 06/06/13 14:44



D-7500 INTEGRATOR REPORT

ANALYZED: 06/06/13 14:44 REPORTED: 06/06/13 15:15  
 SYSTEM : 1 OPERATOR:  
 METHOD : SEQ : 1  
 CHANNEL : 1 <DIGITAL>  
 FILE : 0 MODULE T-PROG : DETECTOR= 1  
 CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
11	8.28	1239415	50.013	BB
15	14.61	1238754	49.987	BB
TOTAL		2478169	100.000	
PEAK REJ :		50000		

2d

MS-507 IA n/I = 95/5 1.000 ml/min

FILE 0 SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 7 OFFS 0 07/03/13 12:45



D-7500 INTEGRATOR REPORT

ANALYZED: 07/03/13 12:45 REPORTED: 07/03/13 13:05  
 SYSTEM : 1 OPERATOR:  
 METHOD : SEQ : 2  
 CHANNEL : 1 <DIGITAL>  
 FILE : 0 MODULE T-PROG : DETECTOR= 1  
 CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
5	8.44	22067	2.060	BB
6	15.71	1046952	97.940	BB
TOTAL		1071019	100.000	
PEAK REJ :		20000		

2e(rac)

FILE 0 SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 5 OFFS 0 05/02/14 15:31

3.05  
9.74  
12.04

D-7500 INTEGRATOR REPORT

ANALYZED: 05/02/14 15:31 REPORTED: 05/02/14 15:47

SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 2

FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
4	9.74	23048	50.471	BB
5	12.04	22618	49.529	BB
TOTAL		45666	100.000	
PEAK REJ :		20000		

2014. 5/2

PN-265 IA hex/IPA = 97/3 1.000 ml/min.

2e

FILE 0 SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 6 OFFS 0 05/06/14 17:19

4.18  
9.48  
11.56 9.48

D-7500 INTEGRATOR REPORT

ANALYZED: 05/06/14 17:19 REPORTED: 05/06/14 17:34

SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 2

FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
2	4.18	5794	0.964	BB
4	9.48	589843	98.088	BB
5	11.56	5706	0.949	BB
TOTAL		601343	100.000	
PEAK REJ :		5000		

2014. 5/6

PN-269 IA hex/IPA = 97/3, 1.000 ml/min

2f(rac)

FILE 0 SYS 1 SEQ 1  
CH. 1<D> C. S 1.25 ATT 6 OFFS 0 05/08/14 14:03

3.38  
9.04  
21.56  
23.43  
28.38  
30.69

D-7500 INTEGRATOR REPORT

ANALYZED: 05/08/14 14:03 REPORTED: 05/08/14 14:36  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 1  
FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
6	28.38	170956	50.413	BB
7	30.69	168152	49.587	BB
TOTAL		339108	100.000	

PEAK REJ : 30000

PN-268

IA, hex/IPA = 95/5, 1.000 ml/min.

2f

FILE 0 SYS 1 SEQ 2  
CH. 1<D> C. S 1.25 ATT 6 OFFS 0 05/07/14 21:59

3.37  
27.00  
29.26

D-7500 INTEGRATOR REPORT

ANALYZED: 05/07/14 21:59 REPORTED: 05/07/14 22:33  
SYSTEM : 1  
METHOD : OPERATOR:  
CHANNEL : 1 <DIGITAL> SEQ : 2  
FILE : 0 MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : 0

NO.	RT	AREA	CONC	BC
2	27.00	435741	97.918	BB
3	29.26	9267	2.082	BB
TOTAL		445008	100.000	

PEAK REJ : 9000

2014.5/7

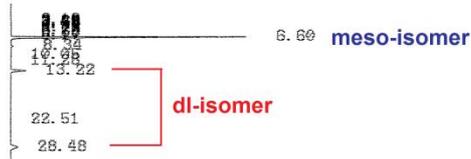
PN-272

IA, hex/IPA = 95/5, 1.000 ml/min

4a(rac)

MS 429 IA hexane/IPA = 95/5 1000 uL/min

FILE # SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 9 OFFS @ 02/21/14 11:48



D-7500 INTEGRATOR REPORT

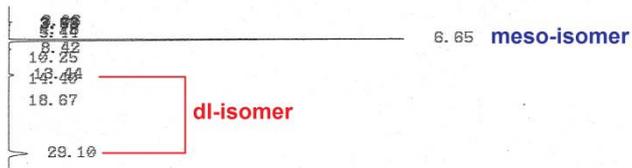
ANALYZED: 02/21/14 11:48 REPORTED: 02/21/14 12:20  
SYSTEM : 1 OPERATOR:  
METHOD : SEQ : 1  
CHANNEL : 1 <DIGITAL>  
FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
10	6.60	2130246	80.970	BV
14	13.22	258415	9.822	VB
16	28.48	242247	9.208	VB
TOTAL		2630908	100.000	
PEAK REJ :		200000		

4a

MS 660 IA hexane/IPA = 95/5 1000 uL/min

FILE # SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 9 OFFS @ 02/22/14 15:53



D-7500 INTEGRATOR REPORT

ANALYZED: 02/22/14 15:53 REPORTED: 02/22/14 16:27  
SYSTEM : 1 OPERATOR:  
METHOD : SEQ : 2  
CHANNEL : 1 <DIGITAL>  
FILE : @ MODULE T-PROG : DETECTOR= 1  
CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

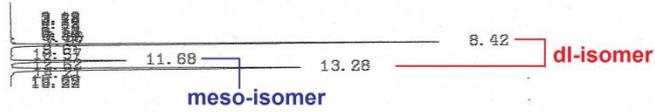
NO.	RT	AREA	CONC	BC
7	6.65	3595306	84.748	BB
10	13.44	80786	1.904	BV
13	29.10	566278	13.348	BB
TOTAL		4242370	100.000	
PEAK REJ :		30000		

4b(rac)

05

MS-730-B IA  $k_{av}/I^2A = 95\%$  1.000ml/min

FILE @ SYS 1 SEQ 1  
CH.1<D> C.S 1.25 ATT 1@ OFFS @ 02/21/14 21:38



D-7500 INTEGRATOR REPORT

ANALYZED: 02/21/14 21:38 REPORTED: 02/21/14 21:56  
 SYSTEM : 1 OPERATOR:  
 METHOD : CHANNEL : 1 <DIGITAL> SEQ : 1  
 FILE : @ MODULE T-PROG : DETECTOR= 1  
 CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
10	8.42	9934813	42.169	VV
13	11.68	3530165	14.984	VV
15	13.28	10094476	42.847	VV
TOTAL		23559454	100.000	
PEAK REJ :		100000		

4b

MS-486 IA  $k/I = 96\%$  1.000ml/min

FILE @ SYS 1 SEQ 2  
CH.1<D> C.S 1.25 ATT 6 OFFS @ 06/07/13 17:49



D-7500 INTEGRATOR REPORT

ANALYZED: 06/07/13 17:49 REPORTED: 06/07/13 18:22  
 SYSTEM : 1 OPERATOR:  
 METHOD : CHANNEL : 1 <DIGITAL> SEQ : 2  
 FILE : @ MODULE T-PROG : DETECTOR= 1  
 CALC-METHOD: AR/HI% <AREA> COMPONENT TBL : @

NO.	RT	AREA	CONC	BC
2	3.38	5352	0.371	BB
5	6.78	34524	2.396	BB
7	8.02	8846	0.614	VB
8	10.98	1218714	84.585	BV
9	12.40	171270	11.887	VB
10	15.31	2111	0.147	BB
TOTAL		1440817	100.000	
PEAK REJ :		2000		

05