

## Supplementary Information

### Enhancement of (Stereo)Selectivity in Dynamic Kinetic Resolution Using Core-Shell Nanozeolite@enzyme as Bi-functional Catalyst

Wanlu Wang, Xiang Li, Zhoujun Wang, Yi Tang, Yahong Zhang\*

Department of Chemistry and Shanghai Key Laboratory of Molecular Catalysis and Innovative Materials, Fudan University, Shanghai 200433 (P. R. China), [zhangyh@fudan.edu.cn](mailto:zhangyh@fudan.edu.cn)

#### Experimental process

The one-pot DKR of racemic 1-PE was carried out in a glass reactor with a magnetic stirring under microwave irradiation at a desired temperature. Typically, racemic 1-PE (0.10 - 0.4 mmol) and acyl donor (0.20 - 0.8 mmol) were firstly added into n-hexane (4.0 mL). And then, 20 mg of the core-shell bi-functional catalysts (H $\beta$ -PDDA@CALB)MSs were added into the above-mentioned reaction mixture. The reaction mixture was irradiated by microwave under magnetic stirring at 50 - 60 °C for a desired time. For mixed catalyst system, 0.2 mmol racemic 1-PE and 0.4 mmol acyl donor were firstly added into n-hexane (4.0 mL). And then, 20 mg of H $\beta$ -ZMSs and 10 mg of Novozym®435 were added into the above-mentioned reaction mixture. The reaction mixture was irradiated by microwave under magnetic stirring at 50 °C for a desired time. The DKR processes of 1-(4-bromophenyl)-ethanol and 1-(p-tolyl) ethanol catalyzed by (H $\beta$ -PDDA@CALB)MSs and mixed catalysts using VA as acyl donor are the same as those of 1-PE at 50 °C.

Yields and enantiomeric excess ( $ee$ ) of products ( $ee_p$ ) and  $ee$  of substrates ( $ee_s$ ) were monitored by gas chromatography (GC, Shimadzu GC 2010 Plus) on a CP-CHIRASIL-DEX CB chiral column (25 m  $\times$  0.25  $\mu$ m, CP7502) with FID detection and calculated with area normalization method. The  $ee_s$  and conversion of substrate, and the selectivity, yield and  $ee$  of the R-ester, as well as the enzyme activity (EA) of free and immobilized CALB were calculated by the following equations:

$$ee_s = \frac{[\text{Substrate}]_{(R,t)} - [\text{Substrate}]_{(S,t)}}{[\text{Substrate}]_{(R,t)} + [\text{Substrate}]_{(S,t)}} \times 100\% \quad (1)$$

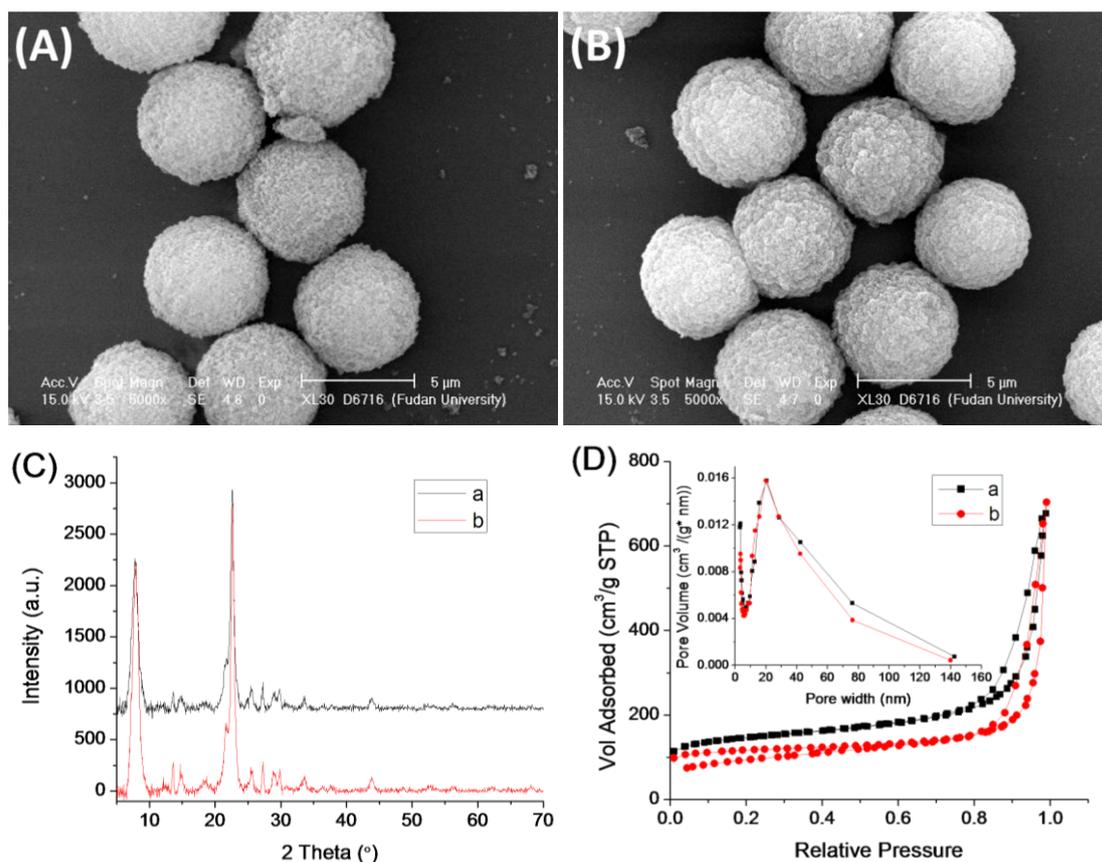
$$\text{Conv.} = \frac{[\text{Product}]_{(t)} + [\text{By-product}]_{(t)}}{[\text{Product}]_{(t)} + [\text{By-product}]_{(t)} + [\text{Substrate}]_{(t)}} \times 100\% \quad (2)$$

$$\text{Selec.} = \frac{[\text{Product}]_{(t)}}{[\text{Product}]_{(t)} + [\text{By-product}]_{(t)}} \times 100\% \quad (3)$$

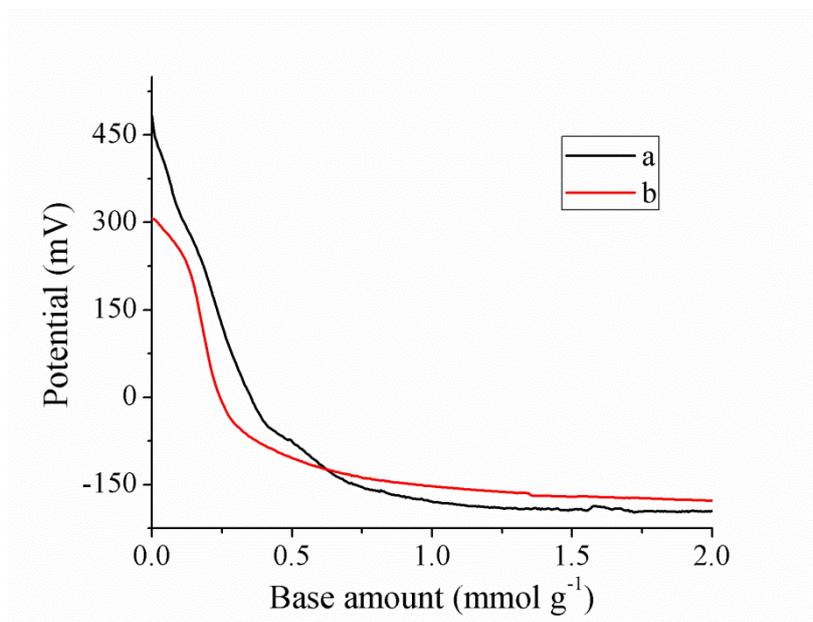
$$Y. = \text{Conv.} \times \text{Selec.} \quad (4)$$

$$ee_p = \frac{[\text{Product}]_{(R,t)} - [\text{Product}]_{(S,t)}}{[\text{Product}]_{(R,t)} + [\text{Product}]_{(S,t)}} \times 100\% \quad (5)$$

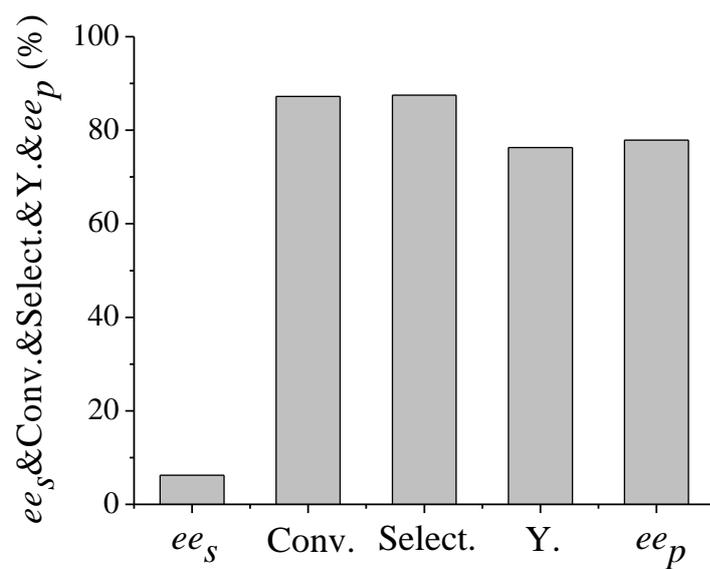
$$\text{EA} = \frac{[\text{Product}]_{(t)} \times V}{t \times m_{\text{CALB}}} \quad (6)$$



**Fig. S1** SEM images of (A) H $\beta$ -ZMSs and (B) (H $\beta$ -PDDA@CALB)MSs as well as XRD patterns (C) and N<sub>2</sub> adsorption/desorption isotherms and pore size distributions (D) of (a) H $\beta$ -ZMSs and (b) (H $\beta$ -PDDA@CALB)MSs.



**Fig. S2** Potentiometric titration curves of (a) H $\beta$ -ZMSs (b) (H $\beta$ -PDDA)MSs. Typically, the solid (0.10 g) was suspended in 25 mL of acetonitrile, and agitated for 3 h. Then, the suspension was titrated with 0.025 mol L<sup>-1</sup> n-butylamine in acetonitrile at a rate of 0.1 mL min<sup>-1</sup> on a potentiometric titration meter (ZDJ-5, Shanghai Leici instrument Factory). The electrode potential variation was measured with a manual continuous titration model using a double junction electrode. The addition continued until no further change of voltage was recorded. The initial electrode potential ( $E_i$ ) indicates the maximum acid strength and the value of meq amine/g solid at the beginning of the plateau indicates the total acid amount.



**Fig. S3** DKR result of 1-PE catalyzed by mixed (H $\beta$ -PDDA)MSs and Novozyme® 435 catalysts using VA as acyl donor at 50 °C. The concentrations of 1-PE and VA are 50 and 100 mmol L<sup>-1</sup>, respectively. The reaction time is 1 h.

**Table S1.** The physical and textural properties of H $\beta$ -ZMSs and (H $\beta$ -PDDA@CALB)MSs.

Catalyst	S <sub>BET</sub> <sup>a</sup> (m <sup>2</sup> g <sup>-1</sup> )	S <sub>micropore</sub> (m <sup>2</sup> g <sup>-1</sup> )	S <sub>external</sub> (m <sup>2</sup> g <sup>-1</sup> )	V <sub>micropore</sub> <sup>b</sup> (cm <sup>3</sup> g <sup>-1</sup> )	V <sub>mesopore</sub> <sup>b</sup> (cm <sup>3</sup> g <sup>-1</sup> )
H $\beta$ -ZMSs	605	459	146	0.23	0.63
(H $\beta$ -PDDA@CALB)MSs	397	285	111	0.14	0.43

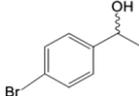
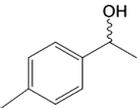
<sup>a</sup> Micropore area and external surface area were calculated by t-plot method. <sup>b</sup> Micropore and mesopore volumes were obtained by t-plot method and desorption data using BJH model between 1.7 and 300 nm width, respectively.

**Table S2.** The immobilized amount and EA of CALB on H $\beta$ -ZMSs and (H $\beta$ -PDDA)MSs as well as the EA of free CALB.

	Immobilized amount of CALB <sup>a</sup> (mg g <sup>-1</sup> )	EA of immobilized CALB <sup>b</sup> ( $\mu$ mol mg <sup>-1</sup> min <sup>-1</sup> )
Free CALB	--	0.17
H $\beta$ -ZMS@CALB	210	not detected
(H $\beta$ -PDDA@CALB)MS	83	0.80

<sup>a</sup> The immobilization of CALB achieved in the aqueous solution of 2 mL containing 6 mg of CALB and 20 mg of H $\beta$ -ZMSs or (H $\beta$ -PDDA)MSs at 35 °C. The immobilization amounts of CALB was obtained through measuring the UV absorbance of the protein solution at  $\lambda = 220$  nm before and after adsorption. To accurately calculate the adsorption amount, the curve was calibrated by using a series of CALB solutions with different concentrations before measurement. The immobilized amount of CALB was expressed as: mg enzyme (g carrier)<sup>-1</sup>, which is abbreviated as mg g<sup>-1</sup>. <sup>b</sup> The EA of CALB immobilized on the H $\beta$ -ZMSs and (H $\beta$ -PDDA)MSs and free CALB were identified by their catalytic performances in the trans-esterification of rac-1-PE. In the catalytic process, 0.20 mmol of rac-1-PE and 0.40 mmol of VA were added into n-hexane (4.0 mL), and then, the immobilized/free CALB were added into the reaction medium. The reaction mixture was irradiated by microwave under magnetic stirring at 50 °C for 15 min. The activity of CALB was obtained and normalized against the time and the amount of CALB. The activity unit was expressed as:  $\mu$ mol product (mg enzyme)<sup>-1</sup> (reaction min)<sup>-1</sup>, which is abbreviated as  $\mu$ mol mg<sup>-1</sup> min<sup>-1</sup>.

**Table S3.** DKR results of 1-(4-bromophenyl)-ethanol and 1-(p-tolyl) ethanol catalyzed by (H $\beta$ -PDDA@CALB)MSs and mixed catalysts using VA as acyl donor at 50 °C.<sup>a</sup>

Substrate	Catalyst	t (h)	Results of DKR				
			<i>ee<sub>s</sub></i> (%)	Conv. (%)	Select. (%)	Y. (%)	<i>ee<sub>p</sub></i> (%)
	(H $\beta$ -PDDA@CALB)MSs	1	66.4	47.4	93.6	44.4	97.9
		2	64.3	62.7	94.2	59.1	96.6
		3	69.0	69.8	93.5	65.2	95.5
	H $\beta$ -ZMSs + Novozyme®435	0.25	48.9	60.1	92.7	55.7	88.9
		0.5	12.5	83.6	91.8	76.7	79.4
		1	7.3	94.9	88.5	84.0	58.2
	(H $\beta$ -PDDA@CALB)MSs	1	11.4	44.6	84.7	37.8	77.1
		2	15.5	67.6	84.4	57.1	76.5
		3	15.8	77.6	84.8	65.8	76.0
	H $\beta$ -ZMSs + Novozyme®435 <sup>b</sup>	0.25	5.3	81.5	68.8	56.1	26.2
		0.5	18.6	97.7	58.8	57.4	2.3

<sup>a</sup> The concentrations of substrate and VA are 50 and 100 mmol L<sup>-1</sup>, respectively. <sup>b</sup> 10 mg H $\beta$ -ZMSs and 5 mg Novozyme®435 are used.