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Electronic supplementary information (ESI)

Synthesis and *in vitro* assessment of a bifunctional closomer probe for fluorine (¹⁹F) magnetic resonance and optical bimodal cellular imaging

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Experimental procedures

General. Common reagents and chromatographic solvents were obtained from commercial suppliers (Sigma-Aldrich, Fisher Scientific) and used without any further purification. Lipophilic Sephadex LH-20 was obtained from GE Healthcare. NMR spectra were recorded on Bruker Avance 400 or 500 MHz spectrometers. The high-resolution mass spectrometry analysis was performed using Applied Biosystems Mariner ESI-TOF. Fluorescence spectra were recorder on a Varian Cary Eclipse fluorescence spectrophotometer.



Synthesis of tert-butyl (1-(3,5-bis(trifluoromethyl)phenyl)-1-oxo-5,8,11-trioxa-2-azatridecan-13yl)carbamate (**S1**):

Compound **S1** was prepared according to the literature reported procedure.¹

Synthesis of tert-butyl (1-(3,5-bis(trifluoromethyl)phenyl)-1-oxo-5,8,11-trioxa-2-azatridecan-13-yl)carbamate (**S2**):

In a 100 mL round bottom flask (RBF), amine **S1** (1.50 g, 5.13 mmol), 3,5-Bis(trifluoromethyl) benzoic acid (1.32 g, 5.13 mmol), *N*-Ethyl-*N'*-(3-dimethylaminopropyl) carbodiimide hydrochloride (1.28 g, 6.67 mmol) and 4-dimethylamino pyridine (0.81 g, 6.67 mmol) were dissolved in 30 mL dry dichloromethane (DCM). The resultant mixture was stirred for 12 h at room temperature (RT) under argon (Ar) atmosphere. Reaction mixture was then diluted with dichloromethane (100 ml) and washed with 5% HCl/water (100 mL) and then with aq. NaHCO₃ (100 mL). Organic layer was separated, dried over sodium sulfate (Na₂SO₄) and concentrated. Product **S2** was purified by silica gel column chromatography using 1-3% methanol (MeOH)-DCM gradient as mobile phase. The pure product was obtained as viscous oil. Yield: 2.30 g (84%). ¹H NMR (400 MHz, CDCl₃): δ 8.31 (s, 2H), 7.95 (s, 1H), 7.51 (brs, 1H), 5.04 (brs, 1H), 3.70 - 3.56 (m, 10H), 3.51 - 3.47 (m, 2H), 3.22 (q, 2H, *J* = 5.3 & 10.6 Hz), 1.39 (m, 9H). ¹³C NMR

(100 MHz, CDCl₃): δ 165.43, 164.63, 156.76, 137.54, 133.10, 132.83, 132.49, 132.16, 130.56, 128.46, 125.44, 122.44, 119.73, 79.99, 71.43, 71.38, 71.34, 71.25, 71.13, 71.04, 70.99, 70.93, 70.85, 70.29, 69.69, 54.15, 41.06 and 29.06. ¹⁹F NMR (376 MHz, CDCl₃): δ -62.92. HRMS (m/z): calcd. for C₂₂H₃₀F₆N₂O₆ [M+Na]⁺ 555.1900. Found: 555.0855.

N-(2-(2-(2-(2-aminoethoxy)ethoxy)ethoxy)ethyl)-3,5-bis(trifluoromethyl)benzamide (S3):

Compound **S2** (2.30 g, 4.32 mmol) was stirred with 20% trifluroacetic acid (TFA)-DCM (15 mL) at 0 °C for 3 h under Ar atmosphere. Reaction mixture was then concentrated under vacuum. The residue was then dissolved in 100 mL DCM and washed with aq. NaHCO₃ solution (100 mL). Organic layer was then separated, dried over Na₂SO₄ and concentrated. The crude product was purified via column chromatography over alumina (IV) using 1-5% MeOH-DCM gradient as mobile phase. The pure product was obtained as colorless oil. Yield: 1.63 g (87%). ¹H NMR (400 MHz, CDCl₃): δ 8.84 (brs, 1H), 8.30 (s, 2H), 7.86 (s, 1H), 3.56 - 3.51 (m, 12H), 3.36 (m, 2H), 2.66 (m, 2H), 2.17-1.27 (brs, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 165.53, 137.72, 132.86, 132.52, 132.19, 131.85, 128.67, 127.87, 125.16, 125.10, 122.45, 119.74, 73.77, 71.21, 71.04, 70.95, 70.62, 42.09 and 40.99. ¹⁹F NMR (376 MHz, CDCl₃): δ -63.00. HRMS (m/z): calcd. for C₁₇H₂₂F₆N₂O₄ [M+H]⁺ 433.1557. Found: 433.1916.



2-(2-(2-(prop-2-yn-1-yloxy)ethoxy)ethoxy)ethanamine (S4):

Compound **S4** was prepared according to the literature reported procedure.²

2-(6-(diethylamino)-3-(diethyliminio)-3H-xanthen-9-yl)-5-(N-(2-(2-(2-(prop-2-yn-1-yloxy)ethoxy)ethoxy)ethyl)sulfamoyl)benzenesulfonate (**S5**):

In a 100 mL round-bottom flask, sulforhodamine B acid chloride (0.50 g, 0.87 mmol) was dissolved in 10 mL tetrahydrofuran (THF) and 2 mL *N*, *N*-dimethylformamide (DMF). This mixture was cooled to 0 °C, and added DIPEA (0.17 g, 1.30 mmol) followed by the slow addition of amine **S4** (0.21 g, 1.13 mmol) in 10 ml THF. The reaction mixture was gradually allowed warmed to room temperature and stirred for 12 h under an Ar atmosphere. The reaction mixture was then concentrated under vacuum, and the pure product was obtained as pink colored power after purification by column chromatography over alumina (IV) using 1-5 % MeOH/DCM gradient as the mobile phase. Yield: 0.40 g (63%). ¹H NMR (400 MHz, CDCl₃): δ 8.56 (s, 1H), 7.89 (d, 1H, *J* = 7.6 Hz), 7.15 (d, 1H, *J* = 7.8 Hz), 7.02 (d, 2H, *J* = 9.4 Hz), 6.69 (d, 2H, *J* = 9.4 Hz), 6.58 (s, 2H), 4.04 (d, 2H, *J* = 1.5 Hz), 3.55 - 3.43 (m, 18H), 3.07 (t, 2H, *J* = 5.0 & 10 Hz), 2.37 (s, 1H), 1.16 (t, 12H, *J* = 6.8 & 13.6 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 157.92, 157.58, 155.60, 146.87, 142.35, 133.74, 132.84, 130.32, 127.53, 126.89, 114.14, 113.63, 95.76, 79.40, 74.82, 70.42, 70.24, 70.16, 69.73, 68.93, 58.27, 45.83, 42.90 and 12.39. HRMS (m/z): calcd. for C₃₆H₄₄N₃O₉S₂ [M+H]⁺ 728.2675. Found: 727.9888. calcd. for C₃₆H₄₄N₃O₉S₂ [M+Na]⁺ 750.2495. Found: 749.9365. calcd. for C₃₆H₄₄N₃O₉S₂ [2M+Na]⁺ 1477.5092. Found: 1477.6736.



Synthesis of Closomer 2.

Closomer 2 was prepared according to the literature reported procedure.³

Synthesis of Closomer 3.

A mixture of Closomer **2** (0.10 g, 0.03 mmol) and amine **S3** (0.86 g, 2.00 mmol) in 15 mL acetonitrile (ACN) was stirred at RT for 3 days. Progress of the reaction was monitored via mass spectrometry and ¹¹B NMR analysis. After completion, the reaction mixture was concentrated in rotavap and dried under vacuum. The crude product was then purified by gel filtration chromatography over Lipophilic Sephadex LH-20 using MeOH as the mobile phase to obtain pure product as colorless viscous oil. Purified product was passed through an ion exchange resin column to exchange TBA⁺ with Na⁺ ion. Yield: 0.16 g (84%). ¹H NMR (400 MHz, CD₃CN): δ 8.35 (s, 22H), 8.11 (s, 11H), 7.84 (brs, 11H), 6.56 (brs, 11H), 3.99 (m, 2H), 3.62-3.46 (m, 184H), 3.10 (m, 22H). ¹³C NMR (100 MHz, CDCl₃): δ 165.09, 156.98, 137.77, 132.58, 132.17, 131.83, 131.49, 128.71, 125.51, 125.45, 122.74, 70.95 – 70.68 (multiple peaks-PEG-CH₂-), 70.29, 69.73, 51.32, 41.81 and 40.61. ¹¹B NMR (128 MHz, CD₃CN): δ -63.35. HRMS (m/z): calcd. for C₂₁₄H₂₆₃B₁₂F₆₆N₂₅O₇₄ [M]²⁻ 2876.5638. Found: 2876.6501.

Synthesis of ¹⁹F-B₁₂-FL

In a 50 mL round bottom flask, the azide containing Closomer **3** (0.37 g, 0.064 mmol), alkyne terminated sulforhodamine B, **S5**, (60.8 mg, 0.084 mmol), and copper (I) iodide (12.2 mg, 0.064 mmol) were dissolved in a 1:1 mixture of THF and ACN (10 mL). To this mixture DIPEA (13.0 mg, 0.10 mmol) was added and the reaction mixture was vigorously stirred at RT for 12 h under an Ar atmosphere. The progress of the reaction was monitored by mass spectrometry analysis. After completion, the reaction mixture was concentrated to dryness, dissolved in ethyl acetate and filtered through a celite pad. The filtrate was concentrated and purified via gel filtration column chromatography over Lipophilic Sephadex LH-20 using DCM as an eluent to afford the pure product as pink solid after drying. Yield: 380 mg (91%). ¹H NMR (400 MHz, CD₃OD): δ 8.68 (s, 1H), 8.44 (m, 23H), 8.13 (m, 13H), 7.55 (m, 1H), 7.14 (m, 2H), 7.00 (m, 2H), 6.98 (m, 2H), 4.64 (m, 4H), 3.90 (m, 8H), 3.65 (m, 206H), 3.33 (m, 34H), 1.30 (m, 12H). ¹³C NMR (100 MHz, CDCl₃+CD₃OD drops): δ 165.71, 158.63, 157.15, 156.30, 148.05, 143.08, 137.34, 134.40, 132.95, 132.62, 132.28, 131.95, 137.34, 128.59, 128.18, 127.85, 125.28, 125.14, 122.43, 119.72, 114.94, 114.31, 96.45, 71.08 – 70.20 (multiple peaks-PEG-CH₂-), 46.55, 43.70, 41.63, 40.74 and 13.15. ¹¹B NMR (128 MHz, CDCl₃+CD₃OD drops): δ -16.39 and -18.74. ¹⁹F NMR (376 MHz, CDCl₃+CD₃OD drops): δ -16.39 and -18.74.

CDCl₃ + CD₃OD drops): δ -63.05. HRMS (m/z): calcd. for C₂₅₀H₃₀₈B₁₂F₆₆N₂₈O₈₃S₂ [M]²⁻ 3240.0425. Found: 3241.0366 [M]²⁻+H.

Cell Culture and maintenance: A549, DLD1, T47D and EMT-6 (ATCC, Manassas, VA) cells were maintained in DMEM + 10% FBS, 5% CO₂ and humidified atmosphere. When the flasks reached 90% confluence, cells were harvested using TrypLE (GIBCO, Grand Island, NY) according to the manufacturer's protocol. Briefly, cells were rinsed using 1X PBS to remove FBS and incubated with TrypLE for 10 min at 37 °C. Cells were then centrifuged at 323g for 8 min in a Fisher Scientific Accuspin 3R centrifuge (Pittsburgh, PA). Cell were resuspended in DMEM + 10% FBS and counted in an automatic cell counter (Invitrogen, Grand Island, NY).

Time dependent cellular uptake of ¹⁹F-B¹²-FL in A549 cells: A549 cells at the density of 300000 cells/time point were incubated with 100 μ M of ¹⁹F-B₁₂-FL (dissolved in DMSO) for different time points (5 min, 30 min, 1 h and 3 h). Final DMSO concentration on the cells was less than 2%. Cells were harvested using TrypLE cell detachment solution and washed 3 times with 1X PBS at 323g for 8 mins. Pellets were frozen at -20 ^oC and lyophilized. The residue was extracted with DMSO (500 μ L each) and the filtrates were analyzed by ¹⁹F NMR and fluorescence spectroscopy (Figure-S1).

Dose dependent cellular uptake of ¹⁹F-B¹²-FL in A549 cells: A549 cells at the density of 300000 cells/dose were incubated with different doses of ¹⁹F-B₁₂-FL (0.1-80 μ M, dissolved in DMSO) for 1hr. Final DMSO concentration on the cells was less than 2%. Cells were harvested using TrypLE cell detachment solution and washed 3 times with 1X PBS at 323g for 8 min. Pellets were frozen at -20 ⁰C and lyophilized. The residue was extracted with DMSO (500 μ L each) and the filtrates were analyzed by ¹⁹F NMR and fluorescence spectroscopy.

Evaluation of toxicity of ¹⁹F-B₁₂-**FL on A549, DLD1, T47D and EMT-6 cells**: A549, DLD1, T47D and EMT-6 cells at the density of 10000/well were plated in 96 well plates. Cells were treated with different doses of ¹⁹**F-B**₁₂-**FL** ranging from 0.1-100 μ M for 72 h followed by addition of MTT reagent for 4 h. The reaction was stopped after 4 h with stop solution provided with MTT kit. Plates were read at 570 nm using microplate reader.

Intracellular localization of ¹⁹F-B₁₂-FL in T47D by confocal microscopy: T47D cells at the density of 100000 cells/chamber were plated in chamber slide overnight for adherence and then

incubated with green Lysotracker at 37 °C for 30 min followed by washing with 1X PBS for 3-4 times. Cells were then incubated with ¹⁹**F-B**₁₂-**FL** (10 μ M) for 3 h following which cells were washed 3-4 times with 1X PBS. Slides were mounted and observed under confocal and fluorescent microscope and pictures were taken.



Fig. S1 Time dependent uptake of ¹⁹**F-B**₁₂**-FL** by human A549 cells. Figure shows fluorescence intensity of cell lysates post incubation of 100 μ M of ¹⁹**F-B**₁₂**-FL** at various time-points

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Chemical Formula: C₂₂H₃₀F₆N₂O₆









.CF₃ H₂N² Η 3 **S**3 ĊF₃

Chemical Formula: $C_{17}H_{22}F_6N_2O_4$

















$$\begin{array}{c} & - & 8.686 \\ & - & 8.445 \\ & - & 8.133 \\ & - & 8.133 \\ & 7.146 \\ & 7.1126 \\ & 7.1126 \\ & 7.1126 \\ & 7.1126 \\ & 7.1126 \\ & 7.1126 \\ & 6.937$$





