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Supporting Information

Biomass Derived Furfural-Based Facile Synthesis of Protected (2S)-phenyl-3-piperidone, a Common Intermediate for Many Drugs

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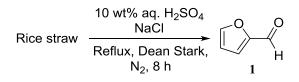
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1. General Methods

All reagents were commercially purchased and were used as received for the reactions. All reactions were carried out in oven-dried glassware while THF was freshly distilled from Na/Benzophenone ketyl and DCM was freshly distilled from Calcium Hydride. Rice straw used was collected from a rice straw farm in Jiangxi Province Fengcheng Area, China after being sun-dried. Upon receiving the rice straw, it was further dried in a vacuum oven at 80 °C for 6 h and stored in an air-tight container. The rice straw is cut into small pieces of about 2-3 cm in length before being used for reaction. Thin-layer chromatography (TLC) was conducted with Merck 60 F254 precoated silica gel plate (0.2 mm thickness) and visualized under UV, by potassium permanganate or ceric ammonium molybdate stain. Flash chromatography was performed using Merck silica gel 60 with distilled solvents. ¹H-NMR spectra were performed on a Bruker Avance 300, Bruker Avance 400 or Bruker Avance 500 NMR spectrometer and are reported in ppm downfield from SiMe₄ (δ 0.0), relative to the signal of chloroform-d ($\delta = 7.26$, singlet) or methanol-d₄ ($\delta = 3.31$, quintet). Data reported as: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, b = broad; coupling constant(s) in Hz; integration. Proton-decoupled ¹³C-NMR spectra were recorded on Bruker Avance 300 (75 MHz) or 400 (100MHz) or 500 (125 MHz) spectrometer and are reported in ppm using solvent as an internal standard (CDCl₃ at 77.16 ppm, CD₃OD at 49.15 ppm). IR spectra were recorded using nujol mull technique on NaCl plates on a Shimadzu IRPrestige-21 FT-IR Spectrophotometer or under attenuated total reflection (ATR) conditions on a PerkinElmer Spectrum 100 FT-IR Spectrometer and were reported in frequency of absorption (cm⁻¹). High-resolution mass spectral analysis (HRMS) was performed on Q-Tof Premier mass spectrometer (Waters Corporation).

2. Synthesis and characterization of compounds

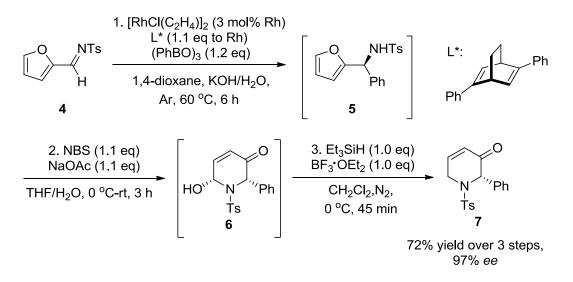


Furan-2-carbaldehyde (1)

To a 250 mL round-bottom flask equipped with a stir bar was added rice straw (10.5 g, 2-3 cm length), NaCl (14 g) and 10 wt% aqueous H₂SO₄ (70 mL). The round-bottom flask was then fitted with a Dean Stark trap with a stopcock at the bottom of the trap and then fitted with water condenser. DCM (10 mL) was added into the Dean Stark trap initially and the reaction mixture was heated to reflux at 150 °C. After 1 h, the DCM in the Dean Stark trap was collected *via* the stopcock and fresh DCM (10 mL) was added into the Dean Stark trap through the top of the water condenser. This process was repeated hourly for a total of 8 h and the combined organic layers were washed with saturated aqueous NaHCO₃ solution (40 mL) and brine (10 mL) before drying over anhydrous magnesium sulphate, filtered and concentrated under reduced pressure to give **1** as a pale yellow oil (851 mg, 8.86 mmol, 8.1 wt%) ¹H NMR (500 MHz, CDCl₃) δ (ppm): 9.68 (s, 1H), 7.70 (t, *J* = 0.8 Hz, 1H), 7.26 (dd, *J* = 3.6 Hz, 0.5 Hz 1H), 6.61 (dd, *J* = 3.6 Hz, 1.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 177.9, 152.9, 148.1, 121.2, 112.6; Other characterization data are similar to the authentic sample.

N-(furan-2-ylmethylene)-4-methylbenzenesulfonamide (4)

1 (14.0 g, 146 mmol, 1.0 eq), 4-methylbenzenesulfonamide (18.8 g, 110 mmol, 0.75 eq), boron trifluoride etherate (1.0 mL, 1.15 g, 8.1 mmol, 5.5 mol%) and toluene (150 mL) were added into a round-bottom flask fitted with a Dean Stark trap. The mixture was heated at reflux for 2 days and activated charcoal was added and stirred for 1 h. The mixture was filtered and the filtrate concentrated under reduced pressure to give a brown solid. Recrystallization from benzene gave *N*-(furan-2-ylmethylene)-4-methylbenzenesulfonamide **4** as brown crystals (20.5 g, 82.5 mmol, 75%). mp = 100-101 °C (lit¹ 100-101 °C); TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.32$; ¹H NMR (300 MHz, CDCl₃) δ (ppm): 8.81 (s, 1H), 7.87 (d, *J* = 8.3 Hz, 2H), 7.74 (d, *J* = 1.6 Hz, 1H), 7.34-7.31 (m, 3H), 6.65 (dd, *J* = 3.6 Hz, 1.7 Hz, 1H), 2.43 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 155.7, 149.8, 149.1, 144.6, 135.2, 129.8, 128.1, 124.7, 113.7, 21.7; FTIR (Nujol, NaCl, cm⁻¹): 1607, 1315, 1155, 932; HRMS (ESI) *m/z* Calcd for C₁₂H₁₂NO₃S [M+H]⁺: 250.0538; found: 250.0541.



(S)-N-(furan-2-yl(phenyl)methyl)-4-methylbenzenesulfonamide (5)

To the solution of $[RhCl(C_2H_4)_2)]_2$ (5.8 mg, 0.015 mmol, 3 mol% Rh) and (*R*,*R*)-Ph-bod* (8.5 mg, 0.033 mmol, 1.1 eq to Rh) in anhydrous 1,4-dioxane (2.5 mL) was added aqueous KOH (65.0 µL, 3.1 M, 20 mol% KOH, H₂O: 1 eq to boron) at room temperature and stirred for 15 min. This solution containing the catalyst was added to the solution of imine **4** (249 mg, 1.0 mmol, 1.0 eq) and 2,4,6-triphenylboroxine (374 mg, 1.2 mmol, 1.2 eq) in anhydrous 1,4-dioxane (4.0 mL) at the same temperature. After 6 h stirring at 60 °C, the mixture was passed through a short silica gel column (pre-treated with methanol, eluent: ethyl acetate) to give **5** as the crude product and was immediately subjected to the next step without any further purification.

Data for **5** after purification using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 5:1) to give (*S*)-N-(furan-2-yl(phenyl)methyl)-4-methylbenzenesulfonamide **5** as a pale yellow solid in 97% yield, *ee* = 99%. The *ee* was determined on Chiralcel OD-H column with hexane/2-propanol = 90:10, flow = 0.5 mL/min, wavelength = 220 nm. Retention times: 20.5 min [(*S*)-enantiomer], 22.0 min [(*R*)-enantiomer]. mp = 129-130 °C; TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.53$; $[\alpha]^{22}_{D} = -18.1$ (c = 1.03, CHCl₃) for 99% *ee* (Lit²: $[\alpha]^{20}_{D} = -21.6$ (c = 1.03, CHCl₃) for 99% ee.); ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.58 (d, *J* = 8.3 Hz, 2H), 7.26-7.22 (m, 4H), 7.19-7.14 (m, 4H), 6.19 (dd, *J* = 3.2 Hz, 1.9 Hz, 1H), 5.99 (d, *J* = 3.2 Hz, 1H), 5.61 (d, *J* = 7.7 Hz, 1H), 5.23 (d, *J* = 7.6 Hz, 1H), 2.38 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 152.3, 143.3, 142.7, 138.4, 137.5, 129.5, 128.7, 128.1, 127.3, 127.2, 110.3, 108.5, 55.6, 21.6; FTIR (Nujol, NaCl, cm⁻¹): 3265, 1597, 1319, 1159, 928; HRMS (ESI) *m/z* Calcd for C₁₈H₁₈NO₃S [M+H]⁺: 328.1007; found: 328.0992.

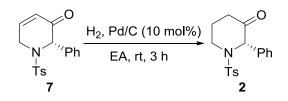
Crude **5** from the previous step was dissolved in a mixture of THF (10.0 mL) and H₂O (3.3 mL) at 0 °C. Sodium acetate (90 mg, 1.1 mmol, 1.1 eq) was added before *N*-bromosuccinimide (196 mg, 1.1 mmol, 1.1 eq) was slowly added in portions at 0 °C over 15 min. The mixture was stirred for 3 h at room temperature after the addition of *N*-bromosuccinimide before solid Na₂S₂O₃ and brine (7 mL) was added. The mixture was extracted with ethyl acetate (3 x 20 mL) and the combined organic layers were washed with

brine (25 mL), dried over anhydrous magnesium sulphate, filtered and concentrated under reduced pressure to give 6 as the crude product and 6 was immediately subjected to the next step without any further purification.

Data for *rac*-6 (recrystallized in CH₂Cl₂ from crude product as pale yellow crystals): mp = 122-123 °C (Decomposed); TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.26$; ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.64 (d, J = 8.3 Hz, 2H), 7.56-7.53 (m, 2H), 7.33-7.24 (m, 5H), 6.86 (dd, J = 10.4 Hz, 4.1 Hz, 1H), 6.11 (dd, J = 10.4 Hz, 1.4 Hz, 1H), 5.99-5.96 (m, 1H), 5.47 (s, 1H), 3.48-3.46 (m, 1H), 2.40 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 191.2, 144.6, 143.7, 136.8, 136.3, 130.3, 128.8, 128.4, 127.8, 127.6, 127.0, 73.6, 64.2, 21.7; FTIR (Nujol, NaCl, cm⁻¹): 3480, 1682, 1649, 1597, 1321, 1155; HRMS (ESI) *m/z* Calcd for C₁₈H₁₈NO₄S [M+H]⁺: 344.0957; found: 344.0970.

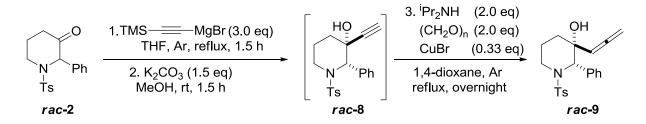
(S)-2-phenyl-1-tosyl-1,6-dihydropyridin-3(2H)-one (7)

The crude rearrangement product 6 was dissolved in anhydrous CH₂Cl₂ (9.0 mL) and cooled down to 0 °C under N₂ protection. Triethylsilane (159 µL, 116 mg, 1.0 mmol, 1.0 eq) was added followed by boron trifluoride etherate (123 µL, 142 mg, 1.0 mmol, 1.0 eq) and the mixture was allowed to stirred at 0 °C for 45 min before H₂O (10 mL) was added to quench the reaction. The mixture was extracted with CH₂Cl₂ (3 x 20 mL). The combined organic layers were washed with brine (25 mL), dried over anhydrous magnesium sulphate and filtered. Concentration under reduced pressure and purification using silica gel chromatography (Eluent: Hexane/Ethyl acetate = 4:1) gave (S)-2-phenyl-1-tosyl-1,6dihydropyridin-3(2H)-one 7 as a brown solid (234 mg, 0.72 mmol, 72% over 3 steps), ee = 97%. The *ee* was determined on Chiralcel OD-H column with hexane/2-propanol = 90:10, flow = 0.5 mL/min, wavelength = 220 nm. Retention times: 29.7 min [(R)-enantiomer], 34.0 min [(S)-enantiomer]. mp = 131-132 °C (lit³ = yellow oil); TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.42$; $[\alpha]_{D}^{21} = +123$ (c = 1.32, CH₂Cl₂) for 97% *ee*. (Lit³: $[\alpha]_{D}^{20} = -145$ (c = 0.3, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ (ppm): 7.62 (d, J = 8.3 Hz, 2H), 7.35-7.29 (m, 5H), 7.25 (d, J = 8.0 Hz, 2H), 6.69 (ddd, J = 10.4 Hz, 4.9 Hz, 1.9 Hz, 1H), 5.94 (ddd, J = 10.4 Hz, 2.4 Hz, 1.6 Hz, 1H), 5.62 (s, 1H), 4.46 (ddd, J = 20.9 Hz, 4.8 Hz, 1.4 Hz, 1H), 3.84 (dt, J = 20.9 Hz, 2.4 Hz, 1.6 Hz, 1H), 2.40 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 192.3, 144.6, 144.2, 136.5, 133.2, 130.1, 129.1, 128.7, 127.8, 127.2, 127.0, 64.1, 41.8, 21.7; FTIR (Nujol, NaCl, cm⁻¹): 1688, 1628, 1597, 1341, 1159; HRMS (ESI) *m/z* Calcd for C₁₈H₁₈NO₃S [M+H]⁺: 328.1007; found: 328.1020.



(S)-2-phenyl-1-tosylpiperidin-3-one (2)

(S)-2-phenyl-1-tosyl-1,6-dihydropyridin-3(2H)-one 7 (151 mg, 0.462 mmol, 1.0 eq) was dissolved in ethyl acetate (10 mL) and palladium on activated charcoal (10 wt%, 49 mg, 0.046 mmol, 10 mol%) was added. The round bottom flask was evacuated and refilled with H₂ thrice using a H₂ balloon. The reaction was stirred for 3 h at room temperature before being filtered through a pad of celite. Concentration under reduced pressure gave (S)-2phenyl-1-tosylpiperidin-3-one 2 as a pale yellow solid (152 mg, 0.462 mmol, 100%), ee = 97%. The *ee* was determined on Chiralcel OD-H column with hexane/2-propanol = 90:10, flow = 0.5 mL/min, wavelength = 220 nm. Retention times: 19.7 min [(R)-enantiomer], 23.0 min [(S)-enantiomer]. mp = 152-153 °C (lit³ = deliquescent solid, lit⁴ = 152-154 °C); TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.50$; $[\alpha]_{D}^{23} = -10.0$ (c = 1.01, CH₂Cl₂) for 97% *ee* (Lit⁴: $[\alpha]_{D}^{20} = +5$ (c = 0.2, CH₂Cl₂)); ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.71 (d, J = 8.3 Hz, 2H), 7.36-7.26 (m, 7H), 5.57 (s, 1H), 3.86 (dt, J = 14.0 Hz, 5.0 Hz, 1H), 3.44 (ddd, J = 14.0 Hz, 9.6 Hz, 4.3 Hz, 1H), 2.43 (s, 3H), 2.40-2.31 (m, 1H), 2.17 (dt, J = 16.0 Hz, 5.3 Hz, 1H), 1.79-1.61 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 204.6, 144.0, 137.3, 134.0, 130.1, 129.3, 128.3, 127.2, 125.9, 66.9, 41.3, 36.9, 23.6, 21.7; FTIR (Nujol, NaCl, cm⁻¹): 1721, 1595, 1342, 1157; HRMS (ESI) *m/z* Calcd for C₁₈H₂₀NO₃S [M+H] ⁺: 330.1164; found: 330.1166.



rac-3-ethynyl-2-phenyl-1-tosylpiperidin-3-ol (rac-8)

An oven-dried 50 mL two-neck round bottom flask equipped with a stir bar and a reflux condenser was cooled under vacuum and back-filled with Ar thrice before being charged with methylmagnesium bromide (2.0 mL of a 3.0 M solution in ether, 6.0 mmol, 3.0 eq), anhydrous THF (2.7 mL) and trimethylsilylacetylene (1.3 mL, 882 mg, 9.0 mmol, 4.5 eq). The mixture was heated at reflux for 1.5 h before **rac-2** (658 mg, 2.0 mmol, 1.0 eq) in anhydrous THF (16.0 mL) was added and the mixture was allowed to stir at reflux for 1.5 h before saturated aqueous NH₄Cl solution (10 mL) was added to quench the reaction. The layers were separated and the aqueous phase extracted with ethyl acetate (3 x 15 mL). The combined organic extracts were washed with saturated aqueous NaHCO₃ solution (30 mL),

brine (30 mL), dried over anhydrous magnesium sulphate and filtered. Concentration under reduced pressure gave the crude product which was used immediately in the next step without further purification.

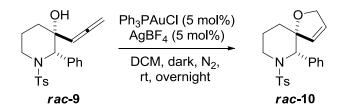
The crude product from the Grignard reaction was dissolved in MeOH (20 mL) and solid K_2CO_3 (414 mg, 3.0 mmol, 1.5 eq) was added and the mixture was allowed to stir at room temperature for 1.5 h. The solvent was removed under reduced pressure and saturated aqueous NH₄Cl solution (10 mL) and EA (20 mL) were added and the layers were separated. The aqueous phase was extracted with ethyl acetate (3 x 10 mL). The combined organic extracts were washed with saturated aqueous NaHCO₃ solution (20 mL), brine (20 mL), dried over anhydrous magnesium sulphate and filtered. Concentration under reduced pressure gave *rac*-8 as the crude product which was used immediately in the next step without further purification.

Data for *rac*-8 after purification using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 2:1) to give *rac*-3-ethynyl-2-phenyl-1-tosylpiperidin-3-ol *rac*-8 as a white solid. mp = 169-170 °C; TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.34$; ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.41 (d, *J* = 8.3 Hz, 2H), 7.34-7.32 (m, 2H), 7.26-7.22 (m, 3H), 7.06 (d, *J* = 8.1 Hz, 2H), 5.31 (s, 1H), 3.82 (dd, *J* = 13.8 Hz, 4.1 Hz, 1H), 3.39-3.29 (m, 1H), 2.53 (s, 1H), 2.34 (s, 3H), 2.02-1.98 (m, 2H), 1.94 (s, 1H), 1.87-1.83 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ (ppm): 143.0, 136.9, 136.2, 129.7, 129.2, 128.4, 128.3, 127.4, 86.1, 73.6, 69.0, 65.6, 41.2, 32.5, 22.0, 21.6; FTIR (ATR, cm⁻¹): 3469, 3297, 3067, 3032, 1597, 1324, 1161; HRMS (ESI) *m/z* Calcd for C₂₀H₂₂NO₃S [M+H]⁺: 356.1320; found: 356.1311.

rac-2-phenyl-3-(propa-1,2-dien-1-yl)-1-tosylpiperidin-3-ol (rac-9)

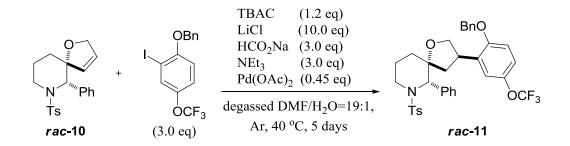
Isopropylamine (560 μ L, 404 mg, 4.0 mmol, 2.0 eq) was added to a suspension of crude *rac*-**8**, paraformaldehyde (120 mg, 4.0 mmol, 2.0 eq) and CuBr (95 mg, 0.67 mg, 33 mol%) in anhydrous 1,4-dioxane (16.0 mL). The reaction was refluxed overnight and then cooled to room temperature before being filtered through a pad of celite. Concentration under reduced pressure and purification using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 5:1) gave *rac*-2-phenyl-3-(propa-1,2-dien-1-yl)-1-tosylpiperidin-3-ol *rac*-**9** as a yellow solid (510 mg, 1.38 mmol, 69%) over 3 steps.

Melting point = 96-97 °C; TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.38$; ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.30 (d, J = 8.2 Hz, 2H), 7.25-7.16 (m, 5H), 7.02 (d, J = 8.0 Hz, 2H), 5.52 (t, J = 6.7 Hz, 1H), 5.01 (s, 1H), 4.99-4.92 (m, 2H), 3.80 (dd, J = 13.1 Hz, 4.5 Hz, 1H), 3.38 (td, J = 12.1 Hz, 4.3 Hz, 1H), 2.32 (s, 3H), 1.97-1.91 (m, 2H), 1.82-1.79 (m, 1H), 1.73-1.67 (m, 1H), 1.63 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 206.7, 142.8, 136.8, 129.7, 129.2, 128.2 x 2, 128.0, 127.2, 98.1, 80.1, 71.3, 65.6, 41.2, 32.1, 21.5 x 2; FTIR (Nujol, NaCl, cm⁻¹): 3532, 1960, 1597, 1329, 1153; HRMS (ESI) *m*/*z* Calcd for C₂₁H₂₄NO₃S [M+H] +: 370.1477; found: 370.1461.



rac-6-phenyl-7-tosyl-1-oxa-7-azaspiro[4.5]dec-3-ene (rac-10)

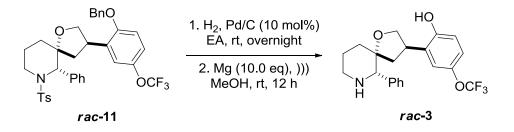
α-allenic alcohol *rac*-9 (200 mg, 0.54 mmol, 1.0 eq), chloro(triphenylphosphine)gold (I) (13 mg, 0.027 mmol, 5 mol%) and silver tetrafluoroborate (5 mg, 0.027 mmol, 5 mol%) were dissolved in anhydrous CH₂Cl₂ (2.0 mL) under N₂ and in the dark. The mixture was allowed to stir at room temperature in the dark overnight. The mixture was then filtered through a pad of celite and concentrated under reduced pressure. Purification using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 7:1) gave *rac*-6-phenyl-7-tosyl-1-oxa-7-azaspiro[4.5]dec-3-ene *rac*-10 as a pale yellow solid (170 mg, 0.46 mmol, 85%). mp = 105-106 °C; TLC (Hexane/Ethyl Acetate = 2:1): R_f = 0.60; ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.39 (d, *J* = 8.3 Hz, 2H), 7.26-7.24 (m, 2H), 7.17-7.13 (m, 3H), 7.07 (d, *J* = 8.1 Hz, 2H), 6.07 (dt, *J* = 6.7 Hz, 2.5 Hz, 1H), 5.87 (d, *J* = 6.2 Hz, 1H), 4.96 (s, 1H), 4.55 (dt, *J* = 13.2 Hz, 2.0 Hz, 1H), 4.29 (dd, *J* = 13.1 Hz, 2.0 Hz, 1H), 3.87-3.81 (m, 1H), 3.35 (ddd, *J* = 13.2 Hz, 11.1 Hz, 5.1 Hz, 1H), 2.35 (s, 3H), 1.95-1.82 (m, 3H), 1.70-1.65 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 142.8, 137.8, 137.5, 132.1, 129.3, 129.3, 127.7, 127.6, 127.2 x 2, 90.4, 75.4, 64.7, 41.3, 30.9, 22.5, 21.6; FTIR (Nujol, NaCl, cm⁻¹): 1651, 1599, 1331, 1171; HRMS (ESI) *m*/z Calcd for C₂₁H₂₄NO₃S [M+H]⁺: 370.1477; found: 370.1480.



rac-3-(2-(benzyloxy)-5-(trifluoromethoxy)phenyl)-6-phenyl-7-tosyl-1-oxa-7-azaspiro[4.5]decane (*rac*-11)

An oven-dried 10 mL Schlenk tube equipped with a stir bar was charged with spirocycle *rac*-**10** (243 mg, 0.66 mmol, 1.0 eq), 1-(benzyloxy)-2-iodo-4-(trifluoromethoxy)benzene (779 mg, 1.98 mmol, 1.98 mmol, 3.0 eq), tetrabutylammonium chloride (220 mg, 0.79 mmol, 1.2 eq), lithium chloride (279 mg, 6.6 mmol, 10.0 eq), sodium formate (134 mg, 1.98 mmol, 3.0 eq), triethylamine (275 μ L, 200 mg, 1.98 mmol, 3.0 eq) and a solution of DMF/H₂O = 19:1 (2.5 mL) under Ar. The mixture was degassed in liquid N₂, allowed to warm to room temperature and backfilled with Ar. The degassing procedure was repeated thrice before palladium (II) acetate (66 mg, 0.30 mmol, 0.45 eq) was added and the mixture was degassed

again before being heated to 40 °C and stirred for 5 days under Ar. The mixture was filtered through a pad of celite, concentrated under reduced pressure and purified using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 10:1 to 6:1) to give *rac*-3-(2-(benzyloxy)-5-(trifluoromethoxy)phenyl)-6-phenyl-7-tosyl-1-oxa-7-azaspiro[4.5]decane *rac*-11 as a pale brown oil (235 mg, 0.37 mmol, 56%). TLC (Hexane/Ethyl Acetate = 2:1): $R_f = 0.68$; ¹H NMR (300 MHz, CDCl₃) δ (ppm): 7.45-7.35 (m, 5H), 7.28-7.21 (m, 4H), 7.18-7.12 (m, 4H), 7.05 (d, *J* = 9.2 Hz, 1H), 6.97 (d, *J* = 8.1 Hz, 2H), 6.89 (d, *J* = 8.9 Hz, 1H), 5.13 (d, *J* = 11.8 Hz, 1H), 5.08 (d, *J* = 11.9 Hz, 1H), 5.02 (s, 1H), 4.20 (t, *J* = 7.7 Hz, 1H), 3.98-3.87 (m, 1H), 3.81-3.76 (m, 2H), 3.18 (dt, *J* = 12.2 Hz, 4.5 Hz, 1H), 2.74 (dd, *J* = 12.6 Hz, 7.7 Hz, 1H), 2.29 (s, 3H), 2.07-2.00 (m, 2H), 1.84-1.81 (m, 2H), 1.60-1.55 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 155.2, 142.9, 142.7, 137.7, 136.7, 136.6, 131.9, 129.5, 129.1, 128.9, 128.3, 128.0, 127.5, 127.4, 127.1, 121.0, 120.7 (q, *J* = 254.6 Hz), 120.2, 112.6, 83.6, 72.4, 70.8, 64.2, 43.0, 41.5, 39.1, 31.6, 23.4, 21.5; FTIR (ATR, cm⁻¹): 3064, 3033, 1599, 1334, 1152; HRMS (ESI) *m/z* Calcd for C₃₅H₃₅F₃NO₅S [M+H]⁺: 638.2188; found: 638.2167.



rac-2-(6-phenyl-1-oxa-7-azaspiro[4.5]decan-3-yl)-4-(trifluoromethoxy)phenol (rac-3)

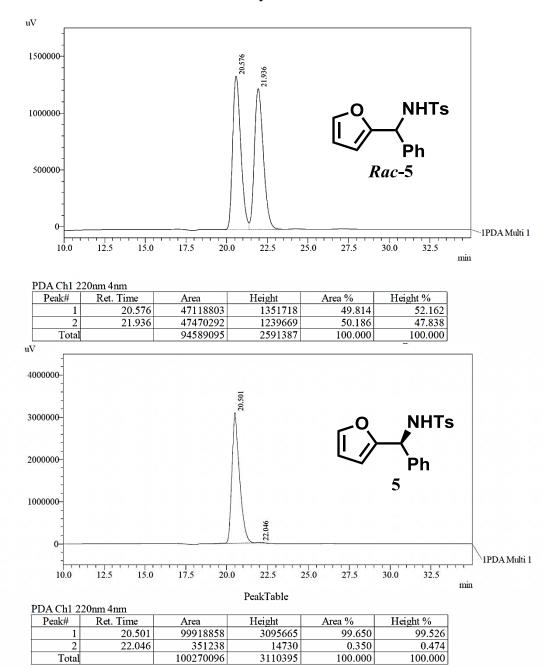
The reductive Heck reaction product *rac*-11 (126 mg, 0.20 mmol, 1.0 eq) was dissolved in ethyl acetate (2.0 mL) before palladium on activated charcoal (10 wt%, 21 mg, 0.02 mmol, 10 mol%) was added. The round bottom flask was evacuated and refilled with H_2 thrice using a H_2 balloon. The reaction was stirred overnight at room temperature before being filtered through a pad of celite. Concentration under reduced pressure gave the crude product which was used in the next step without further purification.

The crude hydrogenation product was dissolved in anhydrous MeOH (3 mL) and magnesium powder (48 mg, 2.0 mmol, 10.0 eq) was added. The suspension was sonicated overnight and 15% aqueous HCl solution (1 mL) was added and the mixture allowed to stir for an additional 15 min before saturated aqueous NaHCO₃ solution was added to neutralise the mixture. Ethyl acetate (30 mL) was added and the layers separated. The aqueous phase was extracted with ethyl acetate (3 x 30 mL) and the combined organic extracts were washed with brine (50 mL), dried over anhydrous magnesium sulphate and filtered. Concentration under reduced pressure and purification using silica gel chromatography (Eluent: Hexane/Ethyl Acetate = 2:1 to Ethyl Acetate/Methanol = 2:1) to give *rac*-2-(6-phenyl-1-oxa-7-azaspiro[4.5]decan-3-yl)-4-(trifluoromethoxy)phenol *rac*-3 (40 mg, 0.102 mmol, 51%) as a pale yellow solid. mp = 203-204 °C (decomposed) (lit⁵ = yellow oil); TLC (Ethyl Acetate/ Methanol = 2:1): R_f = 0.32; ¹H NMR (300 MHz, CD₃OD) δ (ppm): 7.48 (dd, *J* = 7.8 Hz, 1.6 Hz, 2H), 7.38-7.31 (m,

3H), 6.84 (dd, J = 8.7 Hz, 2.0 Hz, 1H), 6.75 (d, J = 2.6 Hz, 1H), 6.64 (d, J = 8.7 Hz, 1H), 3.96 (t, J = 7.5 Hz, 1H), 3.65 (dd, J = 9.9 Hz, 8.0 Hz, 1H), 3.58 (s, 1H), 3.14-3.09 (m, 1H), 2.73 (dd, J = 12.7 Hz, 2.9 Hz, 1H), 2.28-2.14 (m, 2H), 2.05-1.97 (m, 2H), 1.80-1.70 (m, 2H), 1.60 (d, J = 11.9 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ (ppm): 156.0, 142.9, 141.8, 130.4, 129.8, 129.2, 128.8, 122.2 (q, J = 252.7 Hz), 121.7, 121.1, 116.6, 83.6, 72.9, 70.1, 47.7, 42.8, 40.4, 38.7, 24.6; FTIR (ATR, cm⁻¹): 3290, 3062, 3032, 1607, 1510, 1494; HRMS (ESI) *m/z* Calcd for C₂₁H₂₃F₃NO₃ [M+H] ⁺: 394.1630; found: 394.1611.

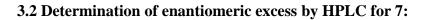
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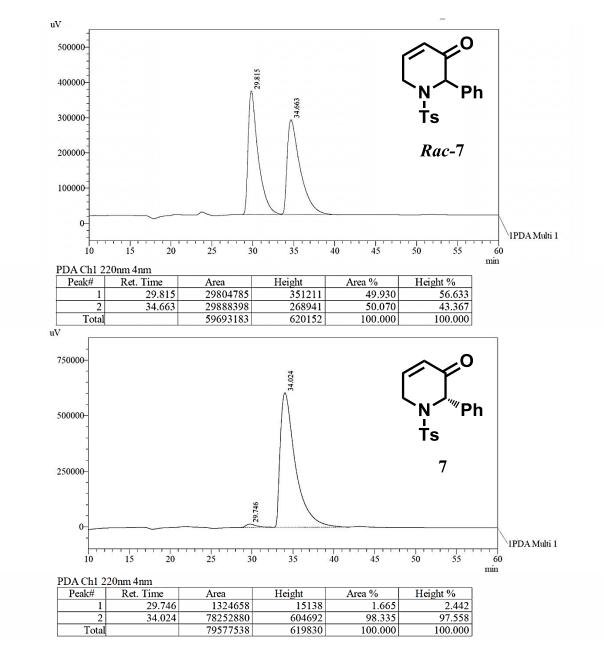
- 1. J. M. Harris and A. Padwa, Org. Lett., 2002, 4, 2029-2031.
- 2. N. Tokunaga, Y. Otomaru, K. Okamoto, K. Ueyama, R. Shintani and T. Hayashi, J. Am. Chem. Soc., 2004, 126, 13584-13585.
- 3. X. Gaucher, M. Jida and J. Ollivier, Synlett, 2009, 3320-3322.
- 4. C. G. Kokotos and V. K. Aggarwal, Chem. Commun., 2006, 2156-2158.
- 5. D. J. Wallace, J. M. Goodman, D. J. Kennedy, A. J. Davies, C. J. Cowden, M. S. Ashwood, I. F. Cottrell, U.-H. Dolling and P. J. Reider, *Org. Lett.*, 2001, **3**, 671-674.



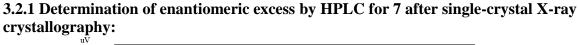
3.1 Determination of enantiomeric excess by HPLC for 5:

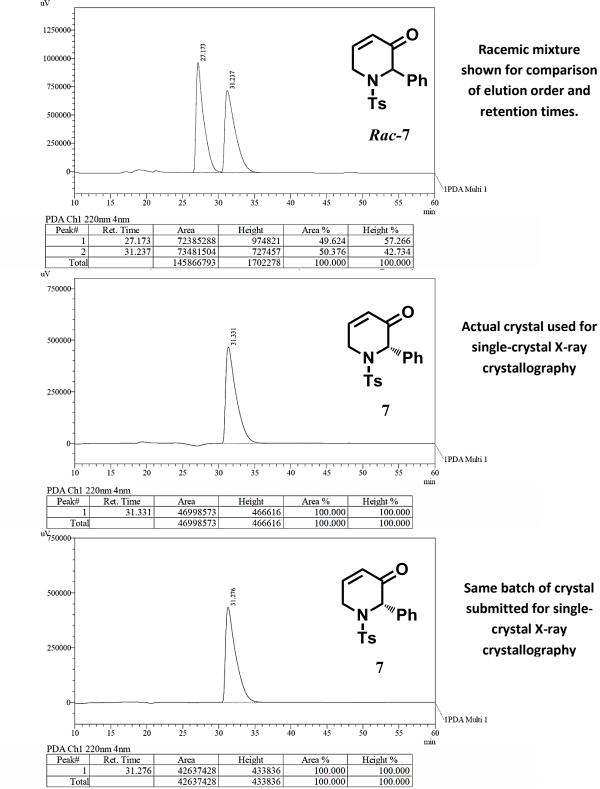
HPLC trace of **5** (Chiralpak OD-H, Hexanes:i-PrOH = 90:10, 0.5 mL/min, 220 nm)



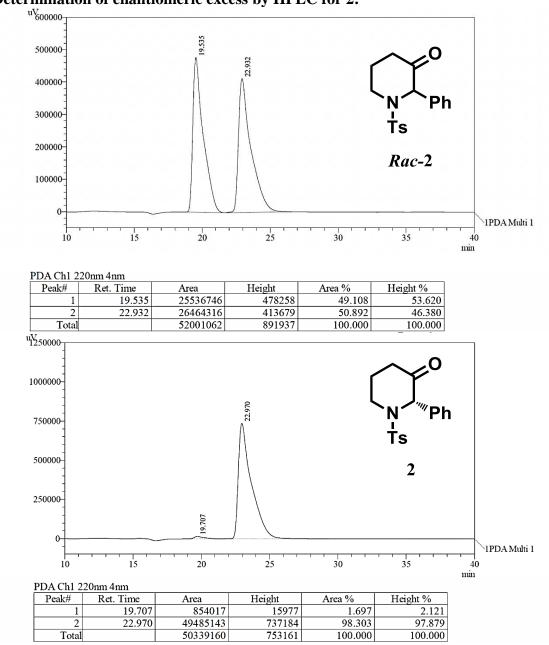


HPLC trace of 7 (Chiralpak OD-H, Hexanes:i-PrOH = 90:10, 0.5 mL/min, 220 nm)





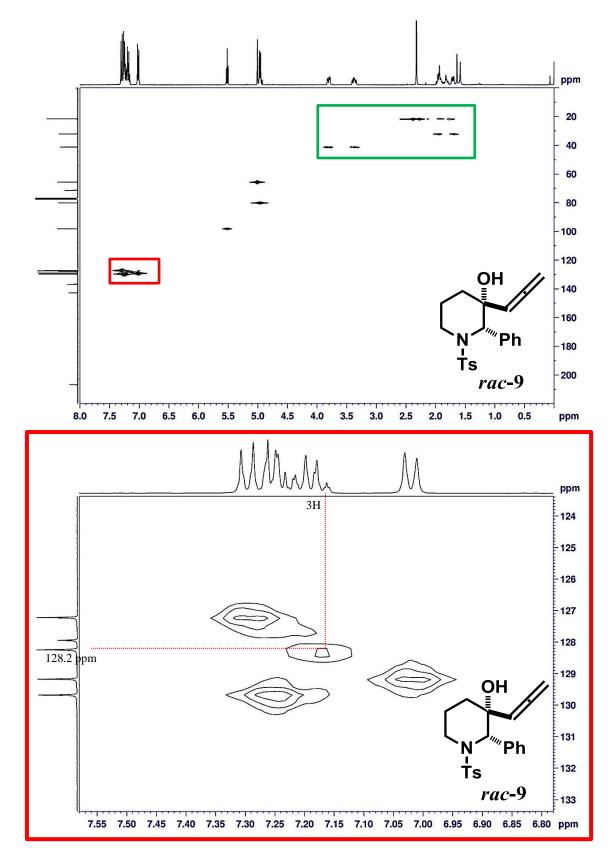
HPLC trace of **7** after X-ray (Chiralpak OD-H, Hexanes:i-PrOH = 90:10, 0.5 mL/min, 220 nm)

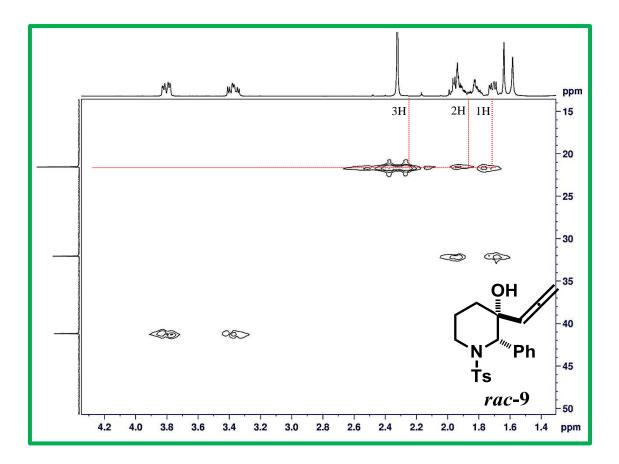


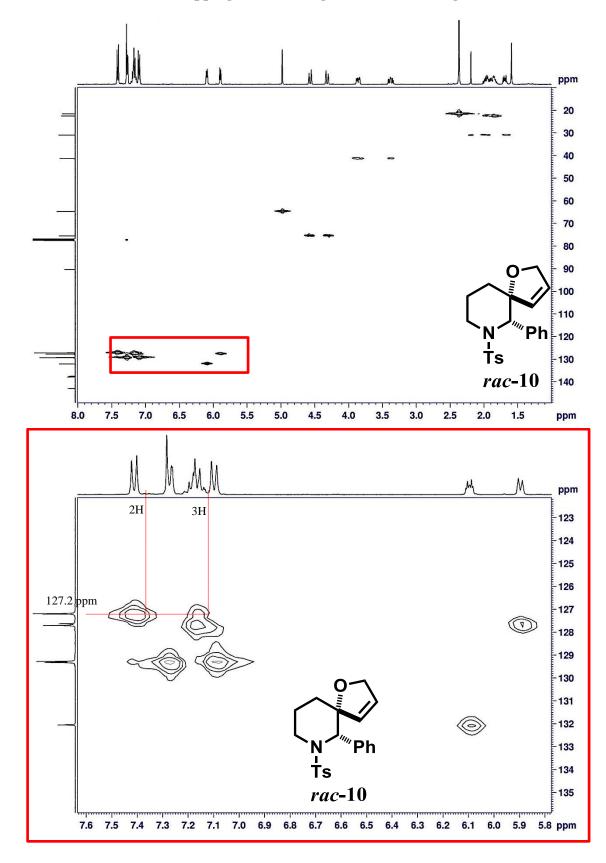
3.3 Determination of enantiomeric excess by HPLC for 2:

HPLC trace of 2 (Chiralpak OD-H, Hexanes:i-PrOH = 90:10, 0.5 mL/min, 220 nm)



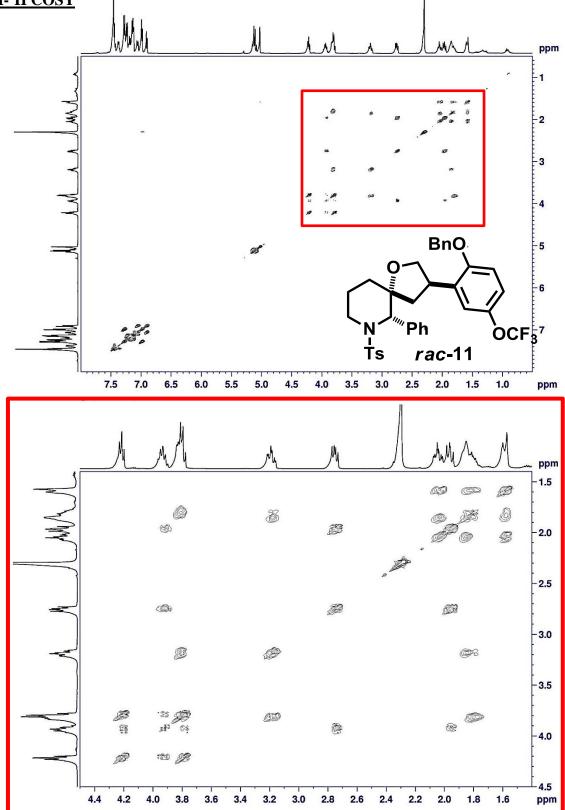


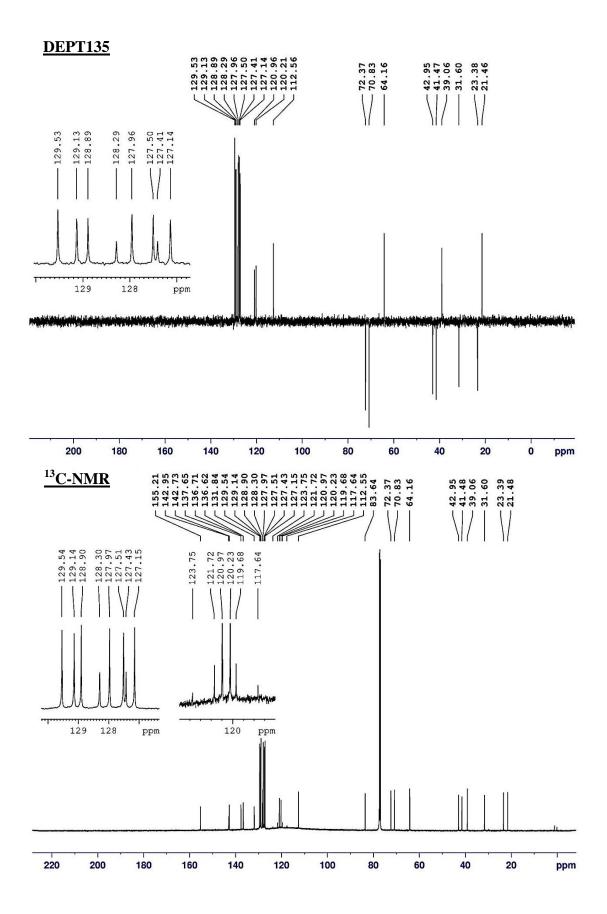




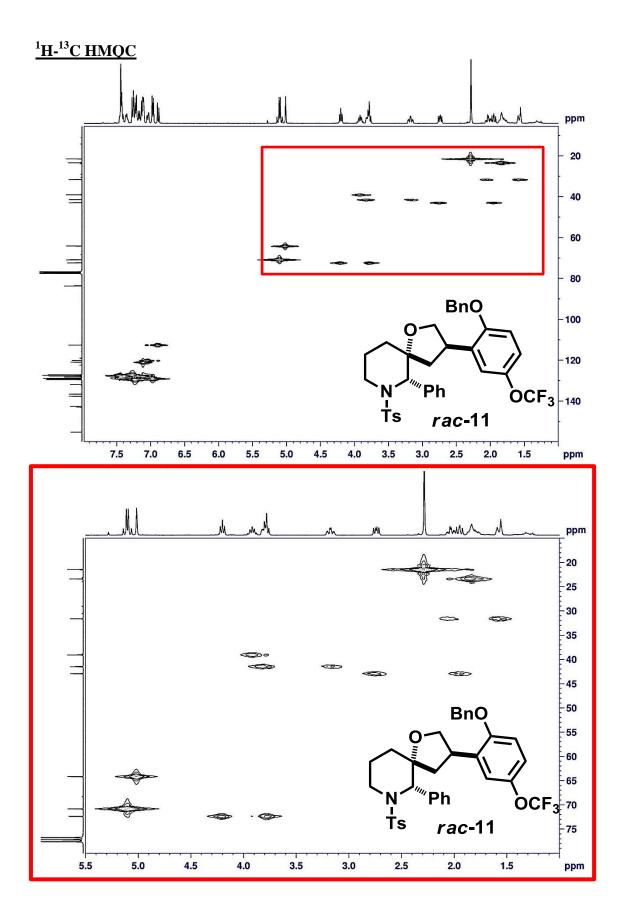
4.2 Determination of overlapping ¹³C NMR Signals in *rac*-10 using HMQC

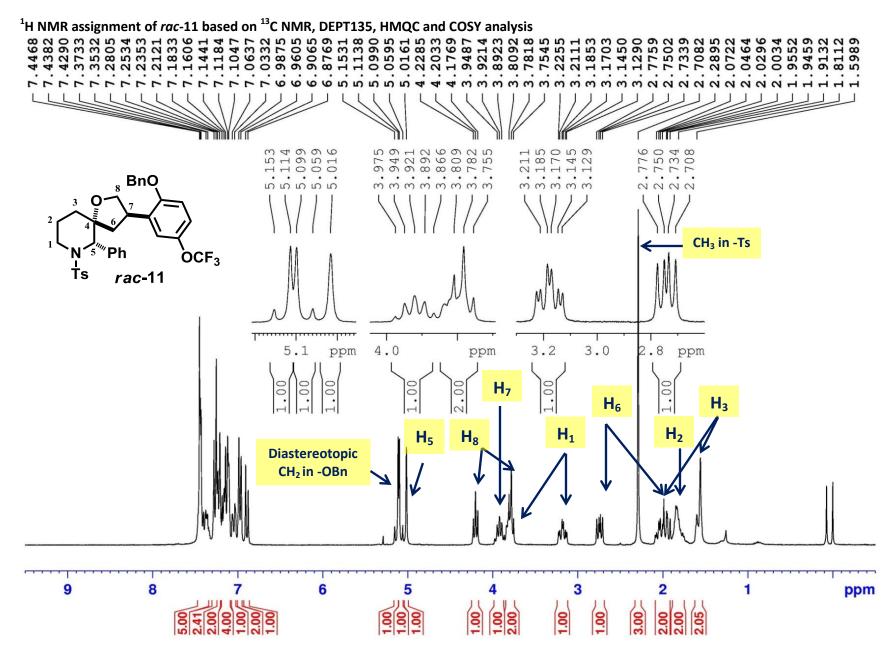
5. Determination of relative stereochemistry in *rac*-11 using NMR analysis
<u>¹H-¹H COSY</u>



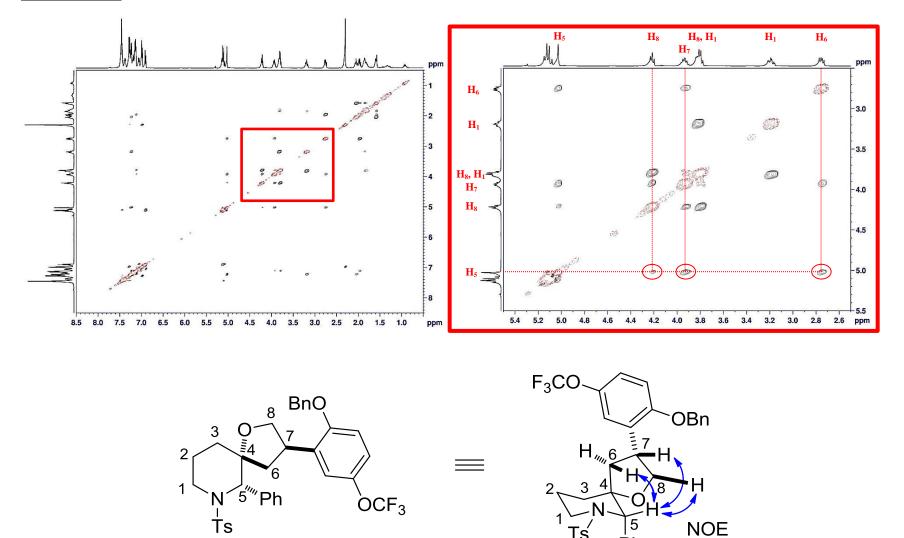


S19





¹H-¹H NOESY



rac-**11**

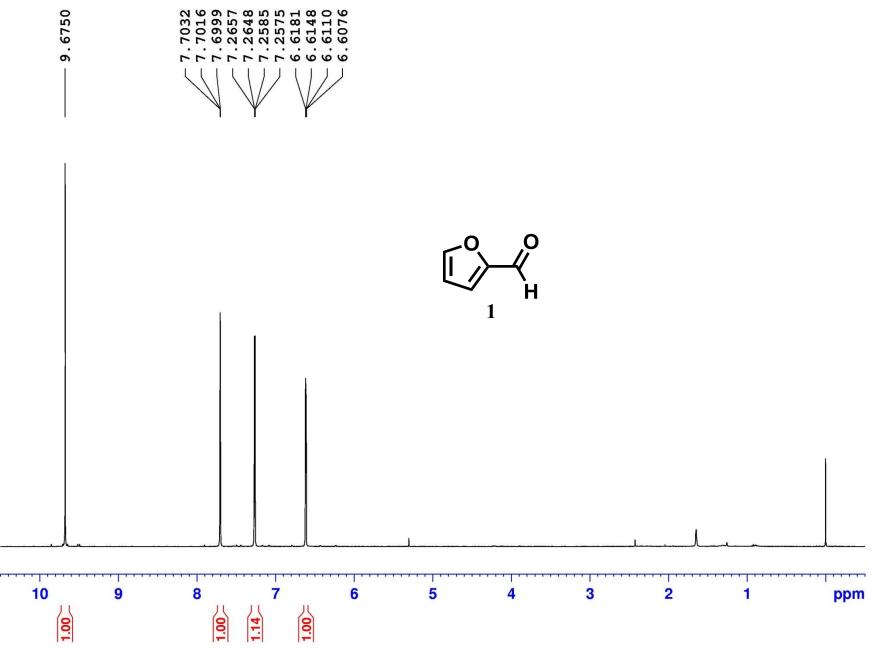
rac**-11**

^T5 Ph

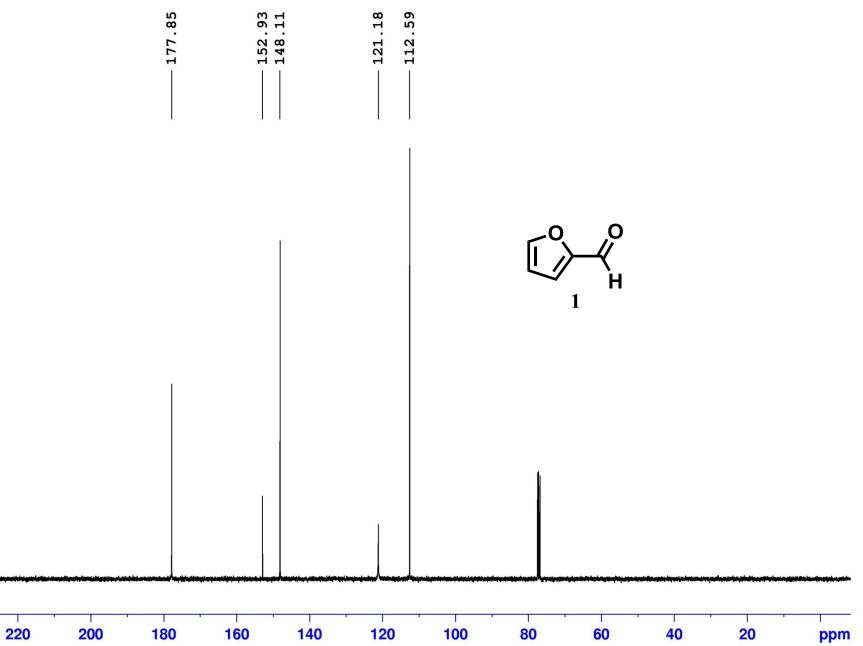
Тs

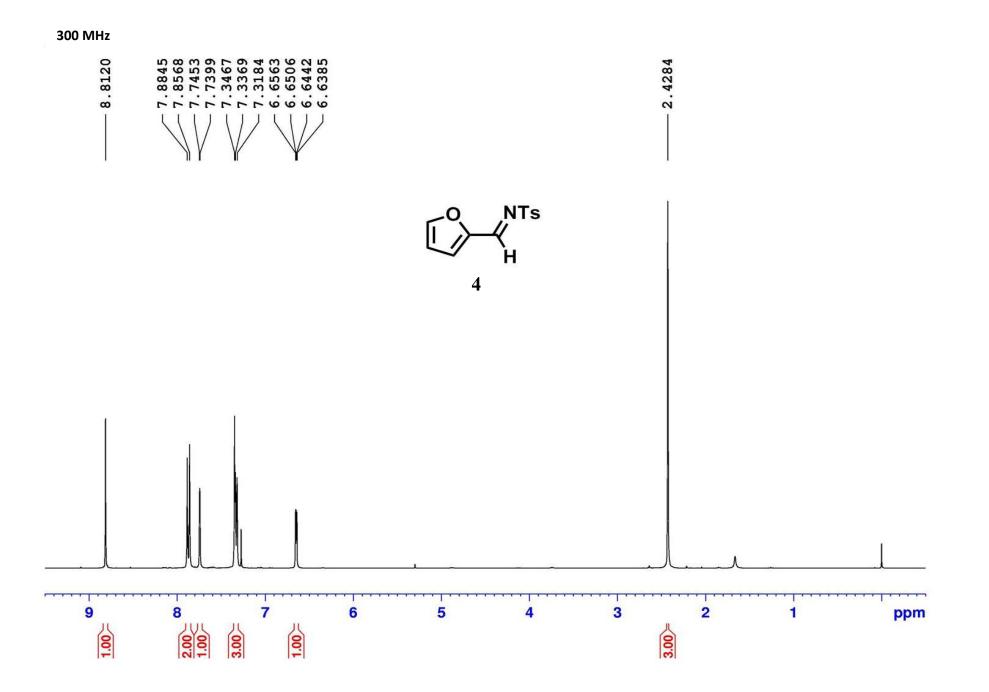
NOE

6. ¹H NMR and ¹³C NMR Spectra 500 MHz



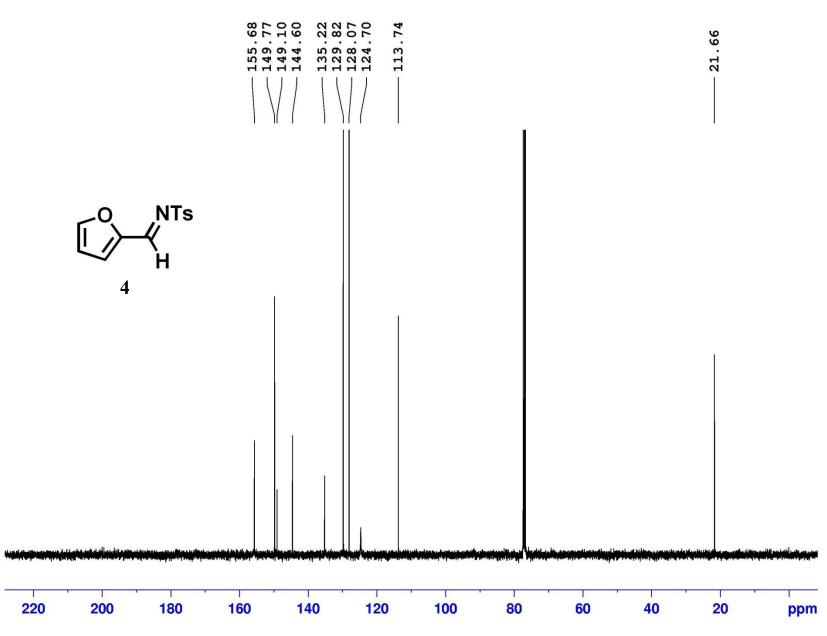




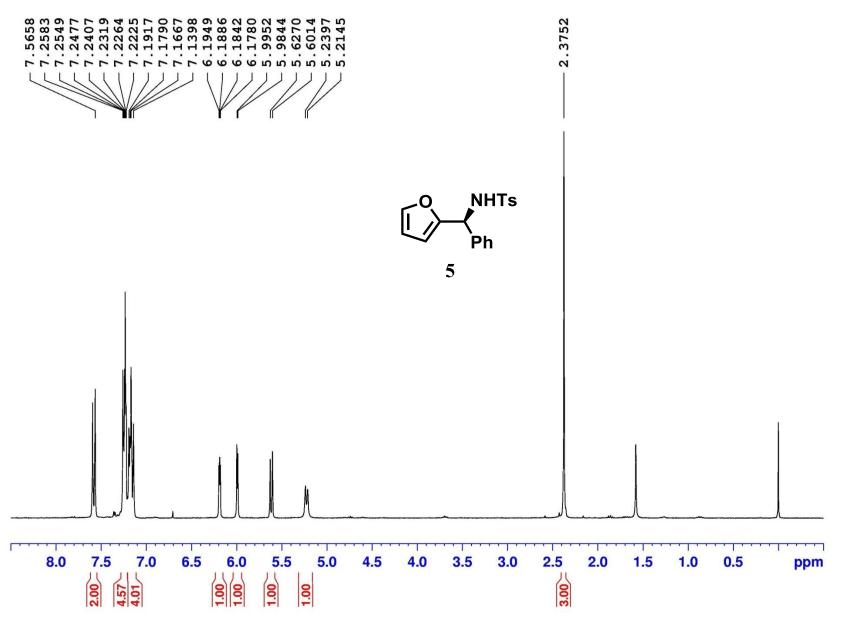


S25

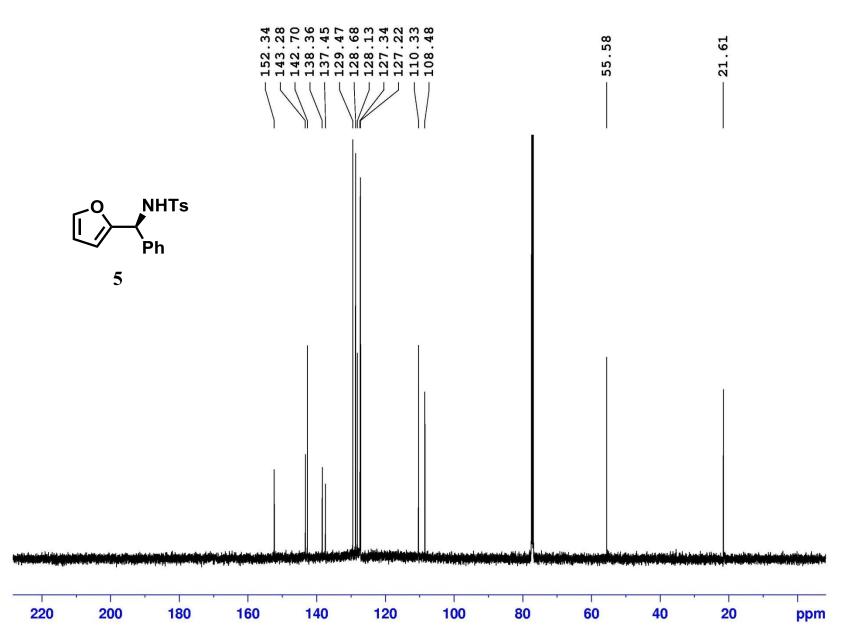




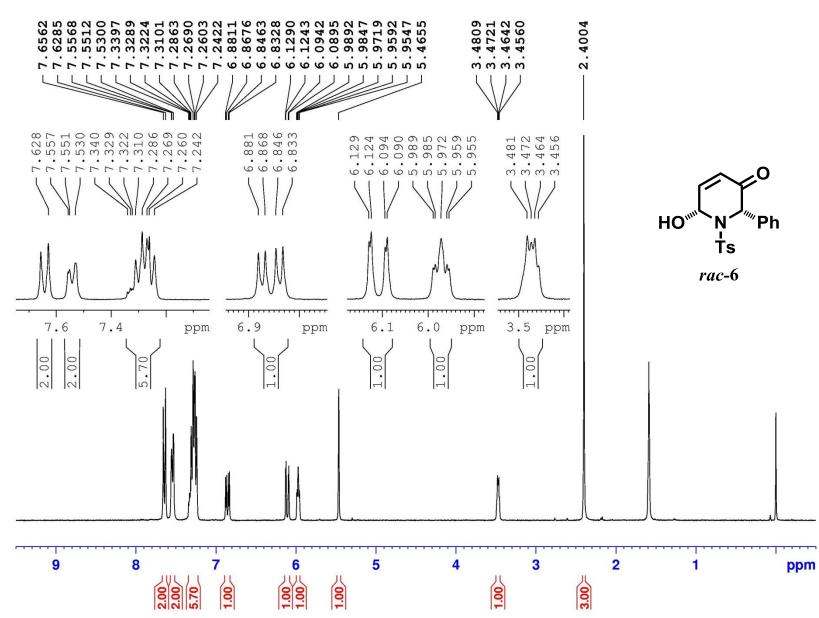




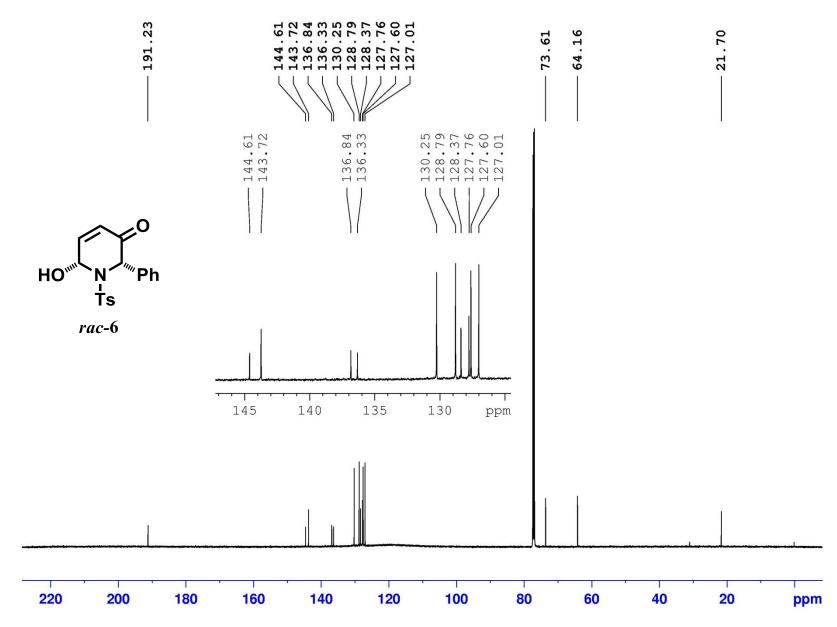
125 MHz



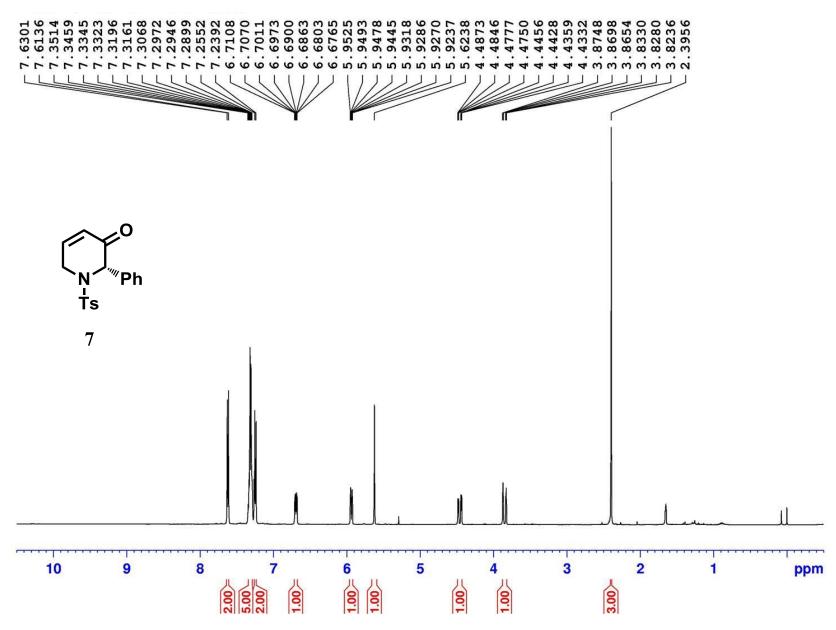


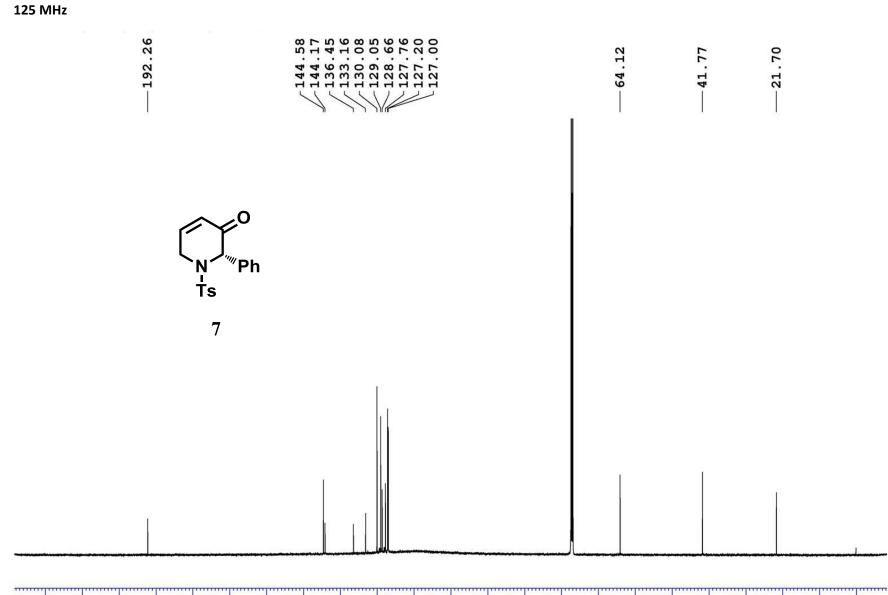




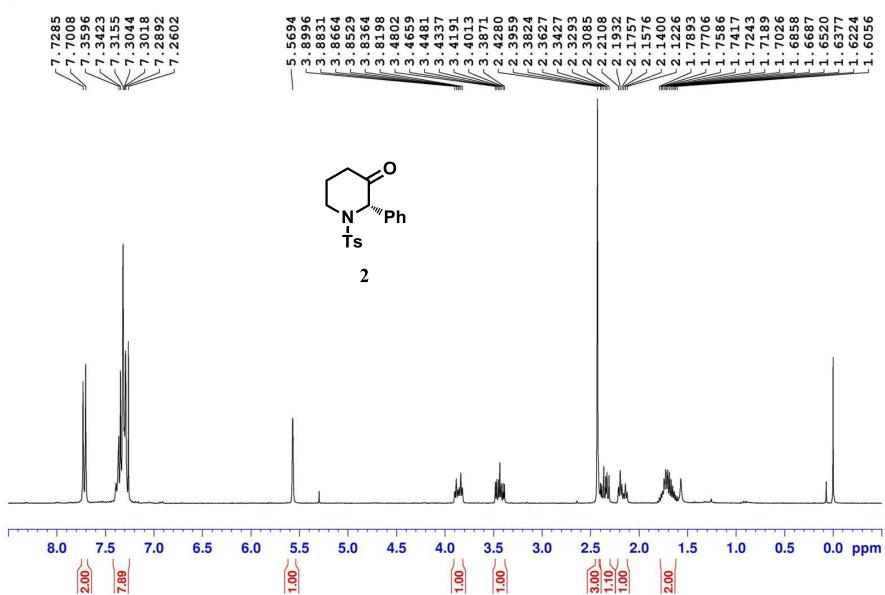


500 MHz

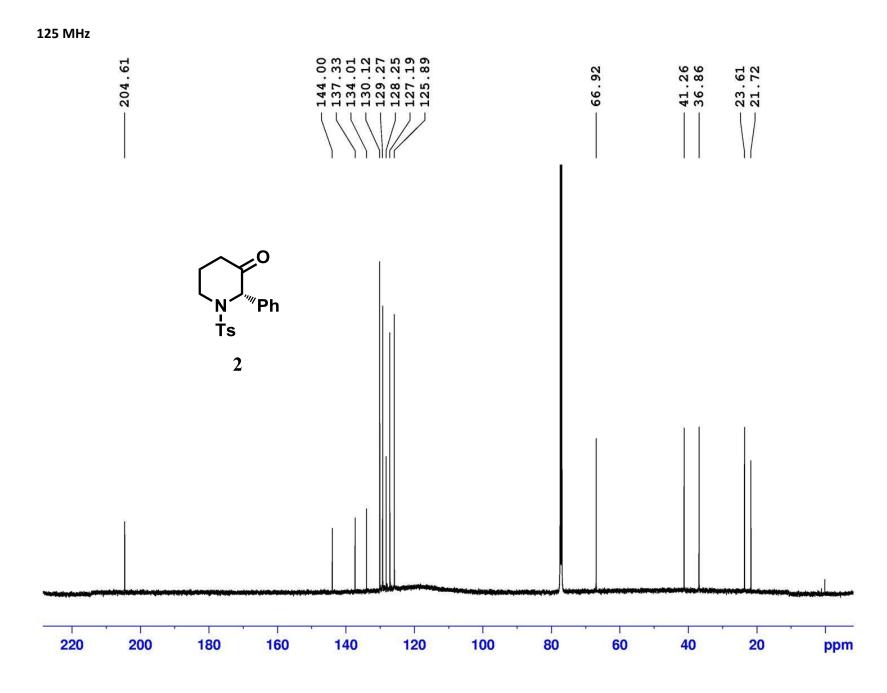




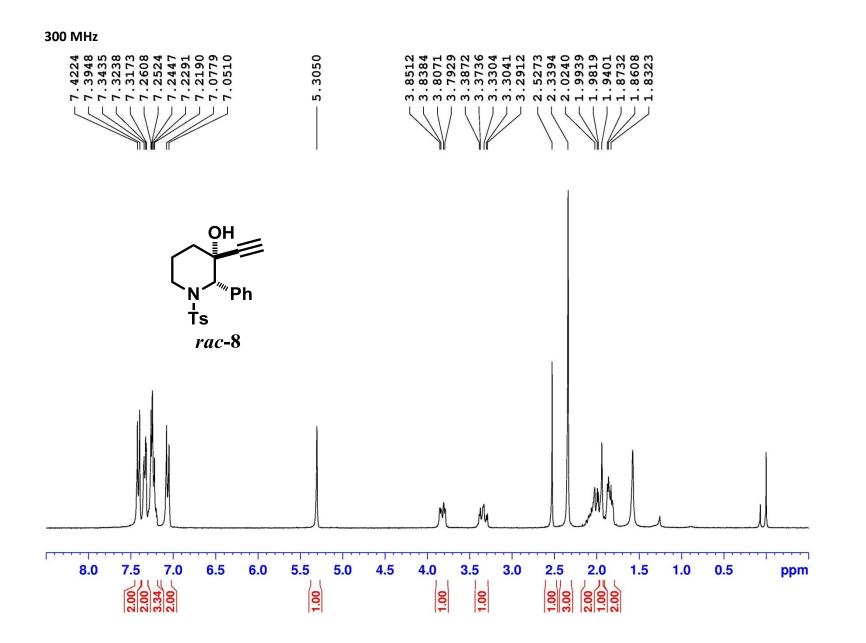
220	210	200	190	180	170	160	150	140	130	120	110	100	90	80	70	60	50	40	30	20	10	ppm



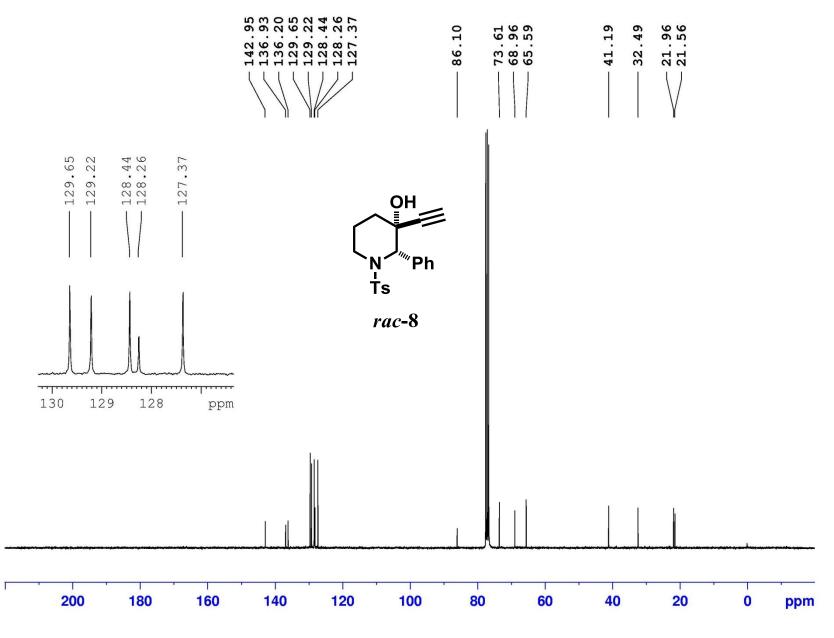
300 MHz



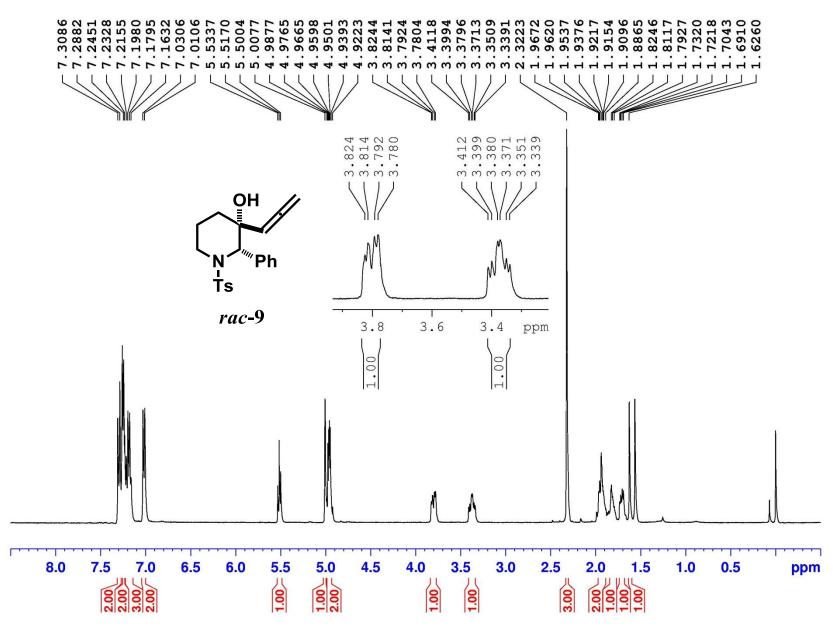
S34

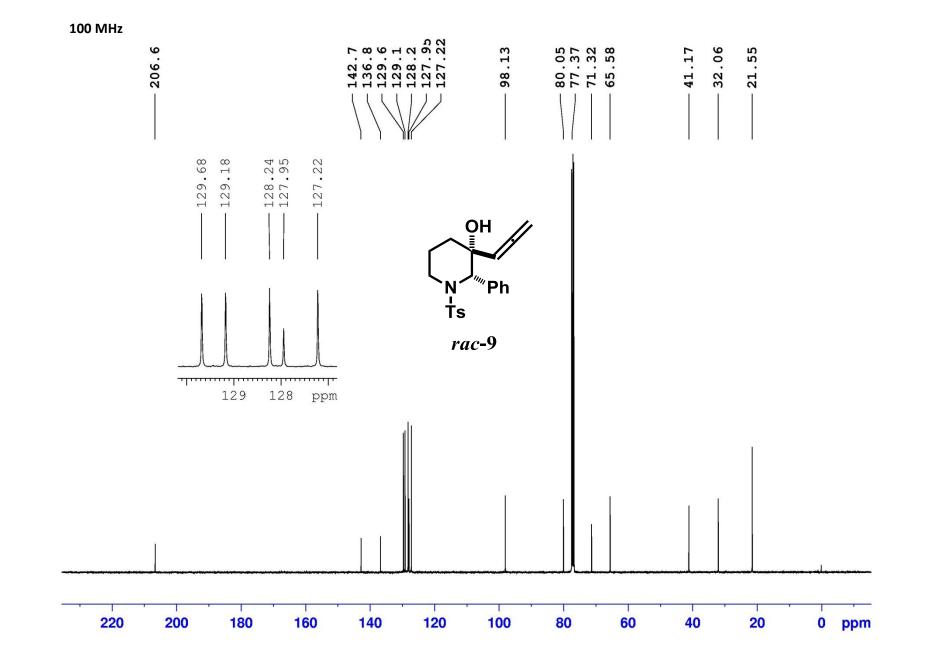


100 MHz



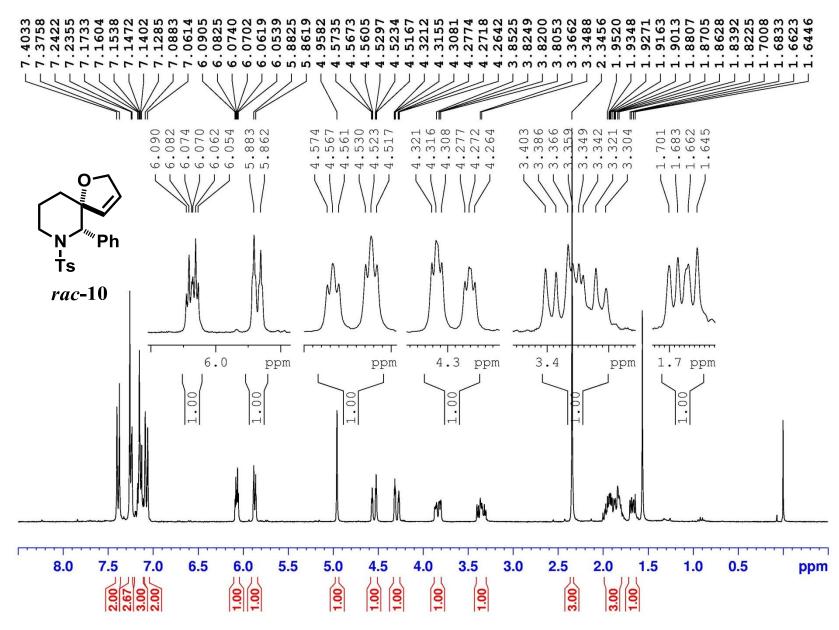




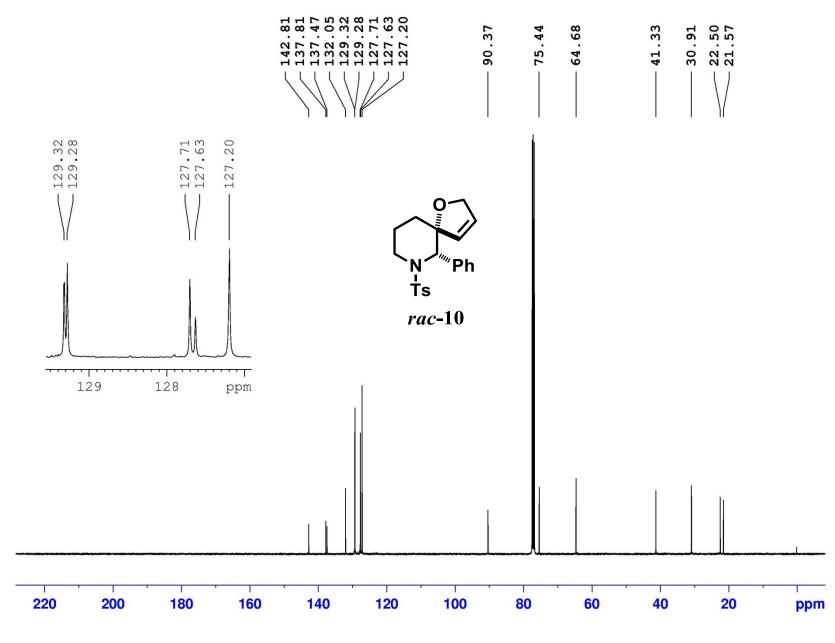


S38



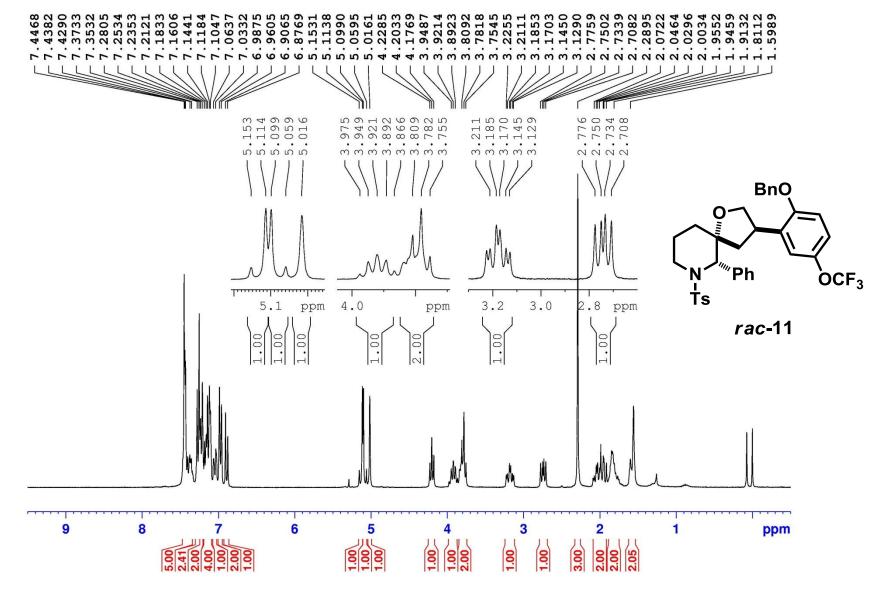


S39

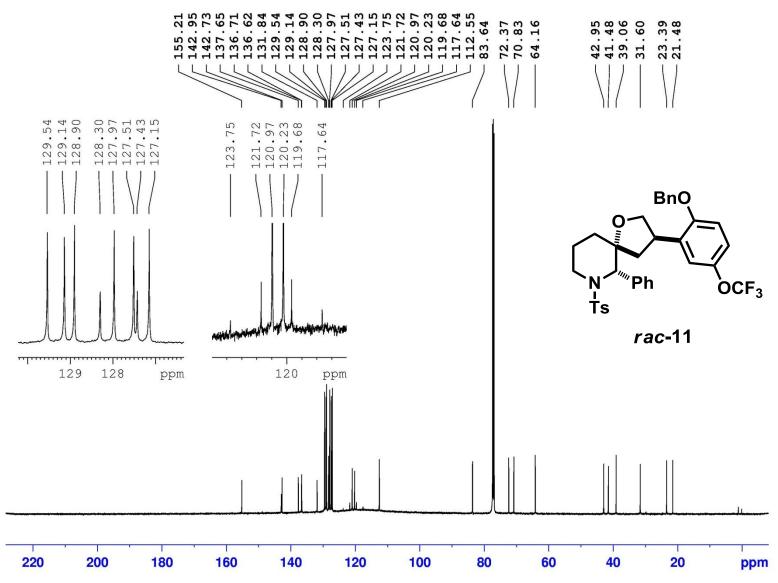


100 MHz

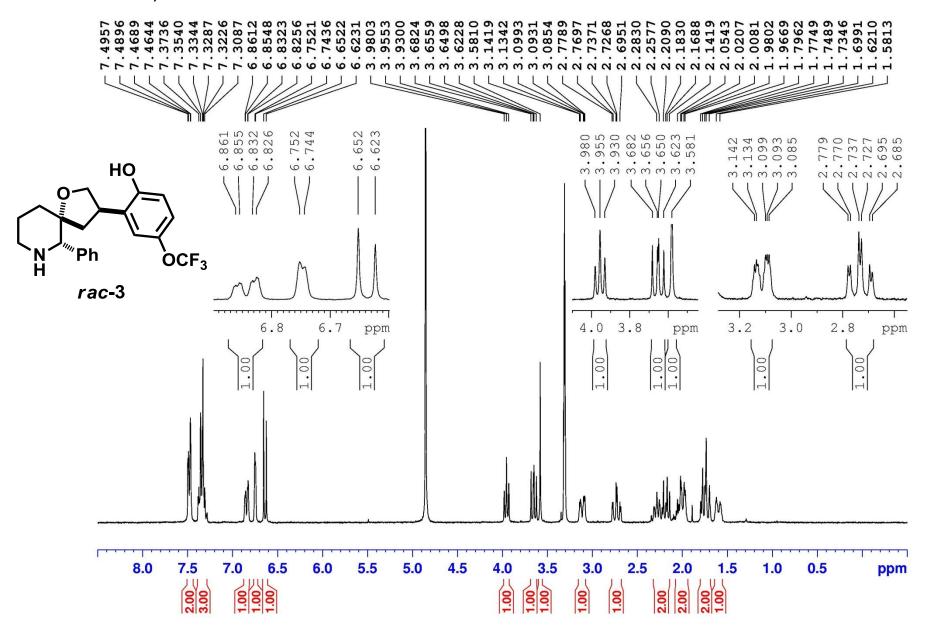


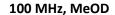


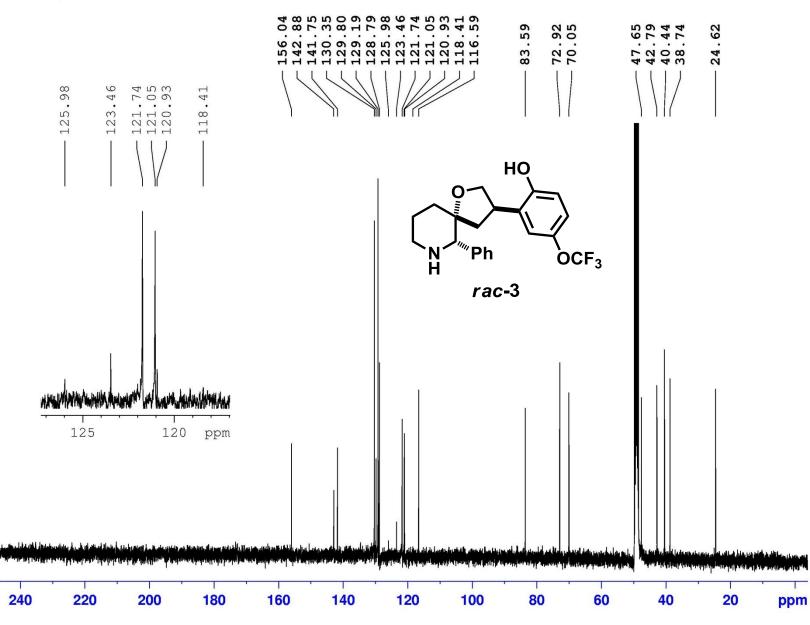




300 MHz, MeOD

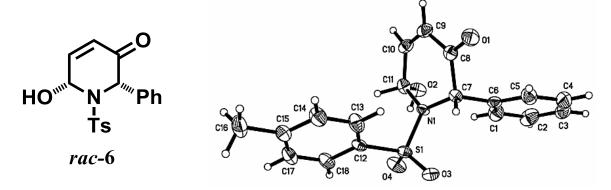






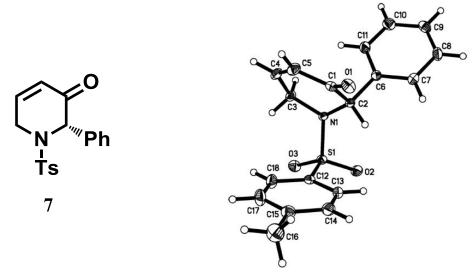
7.1 X-Ray Structure for Rac-6

Cambridge Crystallographic Data Centre Deposition Number: 917487

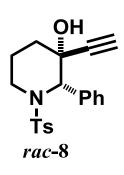


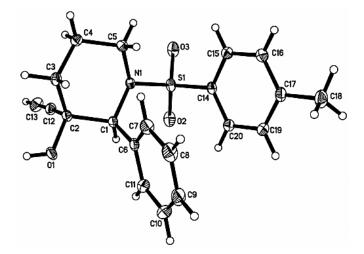
7.2 X-Ray Structure for 7

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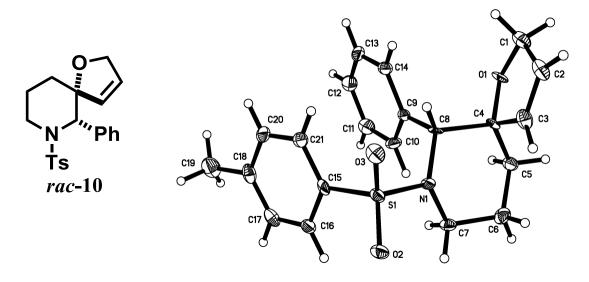






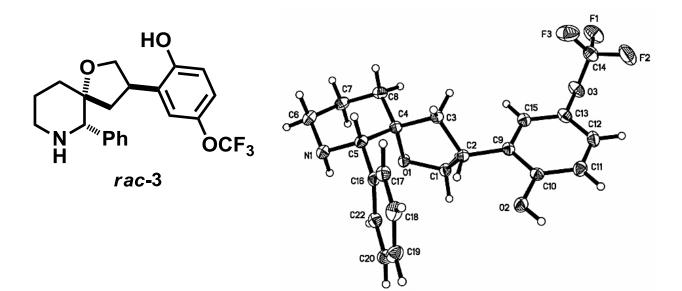
7.4 X-Ray Structure for *Rac*-10

Cambridge Crystallographic Data Centre Deposition Number: 917489



7.5 X-Ray Structure for *Rac-3*

Cambridge Crystallographic Data Centre Deposition Number: 917486



8. List of references found in reference 5 of main text

5 a) M. S. Kramer, N. Cutler, J. Feighner, R. Shrivastava, J. Carman, J. J. Sramek, S. A. Reines, G. Liu, D. Snavely, E. Wyatt-Knowles, J. J. Hale, S. G. Mills, M. MacCoss, C. J. Swain, T. Harrison, R. G. Hill, F. Hefti, E. M. Scolnick, M. A. Cascieri, G. G. Chicchi, S. Sadowski, A. R. Williams, L. Hewson, D. Smith, E. J. Carlson, R. J. Hargreaves and N. M. J. Rupniak, *Science*, 1998, **281**, 1640-1645.

b) For GR203040, see: P. Ward, D. R. Armour, D. E. Bays, B. Evans, G. M. P. Giblin, N. Heron, T. Hubbard, K. Liang and D. Middlemiss, *J. Med. Chem.*, 1995, **38**, 4985-4992.

c) For GR205171, see: C. J. Gardner, D. R. Armour, D. T. Beattie, J. D. Gale, A. B. Hawcock, G. J. Kilpatrick, D. J. Twissell and P. Ward, *Regul. Pept.*, 1996, **65**, 45-53.

d) For T-2328, see: Y. Watanabe, H. Asai, T. Ishii, S. Kiuchi, M. Okamoto, H. Taniguchi, M. Nagasaki and A. Saito, *J. Pharm. Sci.*, 2008, **106**, 121-127.

e) For CJ-17,493, see: Y. Shishido, H. Wakabayashi, H. Koike, N. Ueno, S. Nukui, T. Yamagishi, Y. Murata, F. Naganeo, M. Mizutani, K. Shimada, Y. Fujiwara, A. Sakakibara, O. Suga, R. Kusano, S. Ueda, Y. Kanai, M. Tsuchiya and K. Satake, *Bioorg. Med. Chem.*, 2008, **16**, 7193-7205.

f) For L-741,671, see: T. Ladduwahetty, R. Baker, M. A. Cascieri, M. S. Chambers, K. Haworth, L. E. Keown, D. E. MacIntyre, J. M. Metzger, S. Owen, W. Rycroft, S. Sadowski, E. M. Seward, S. L. Shepheard, C. J. Swain, F. D. Tattersall, A. P. Watt, D. W. Williamson and R. J. Hargreaves, *J. Med. Chem.*, 1996, **39**, 2907-2914.

g) For L-733,060, see: T. Harrison, B. J. Williams, C. J. Swain and R. G. Ball, *Bioorg. Med. Chem. Lett.*, 1994, **4**, 2545-2550.

h) For CP-99,994, see: M. C. Desai, S. L. Lefkowitz, P. F. Thadeio, K. P. Longo and R. M. Snider, *J. Med. Chem.*, 1992, **35**, 4911-4913.

i) For CP-122,721, see: T. J. Rosen, K. J. Coffman, S. McLean, R. T. Crawford, D. K. Bryce, Y. Gohda, M. Tsuchiya, A. Nagahisa, M. Nakane and J. A. Lowe III, *Bioorg. Med. Chem. Lett.*, 1998, **8**, 281-284.

9. Generally accepted mechanism for the aza-Achmatowicz Rearrangement

