Supporting Information For:

Copper-Catalyzed 5-Endo-trig Cyclization of Ketoxime Carboxylates: A Facile Synthesis of 2-Arylpyrroles

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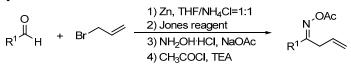
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1. General Information

¹H and ¹³C NMR spectra were recorded on Varian instrument (400 MHz) and (100 MHz), The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. Coupling constants, J were reported in Hertz unit (Hz). Preparative TLC was performed on TLC plate, Analytical thin layer chromatography was performed on 10-25um silica gel GF254, visualization was carried out with UV light. Flash column chromatography was performed with SiO₂ (Silicycle Silica Gel (200-300 mesh)). Unless otherwise stated, all reagents and solvents were purchased from commercial suppliers and used without further purification.

2. General Procedure for Preparation of Ketoxime Acetates

Procedure for Preparation of Ketoxime Acetates 1a-1m:



1) To a solution of the aldehyde (1.0 equiv) in anhydrous THF was added the allylbromide (2.0 equiv) and saturated aqueous NH₄Cl. Then, the zinc dust (2.0 equiv) was slowly added to the solution at 0 °C and the resulting suspension was stirred overnight at this temperature. After completion of the reaction, filtered and extracted with ethyl acetate for 3 times. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude allylic alcohol product was directly used in the next step without further purification.^[1]

2) Allylic alcohol (1.0 equiv) was dissolved in diethyl ether and the solution was kept at 0°C with stirring. The Jones reagent (2.0-4.0 equiv) was added dropwise at this temperature. Then, the resulting mixture was allowed to warm to room temperature and stirred for 1 h. After completion of the reaction, the diethyl ether layer was separated from the aqueous layer, which was extracted with diethyl ether for 3 times. The combined diethyl ether layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude ketone product was directly used in the next step without further purification.^[2] 3) To a solution of hydroxylamine hydrochloride (5.0 equiv) in water was stirred at room temperature. Sodium acetate (7.0 equiv) was dissolved in ethanol and added to the solution. The mixture was stirred at this temperature while the ketone (dissolved in ethanol) was added. The resulting suspension was stirred overnight and concentrated in vacuo. Then, the mixture was extracted with ethyl acetate 3 times and the combined extracts were washed with water and brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude material was purified by flash chromatography on silica gel to afford the corresponding ketoxime.^[3]

4) To a solution of the ketoxime (1 equiv) and TEA (Et₃N) (2 equiv) in anhydrous CH_2Cl_2 (20 mL) was stirred at 0°C while CH_3COCl (1.1 equiv) in CH_2Cl_2 (5 ml) was added dropwise. Then, the resulting mixture was stirred for 2h at the same temperature. After completion of the reaction, water was added and the mixture extracted with CH_2Cl_2 for 3 times. The combined organic extracts were washed with water, brine, dried over Na_2SO_4 , filtered and concentrated in vacuo. The crude material was purified by flash chromatography on silica gel to afford the corresponding ketoxime acetate with hexanes/ethyl acetate as the eluent.^[4]

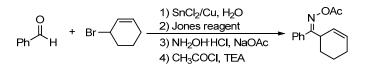
Procedure for Preparation of Ketoxime Acetates 1n and 1p:

$$\begin{array}{c} O \\ R^{1} \\ H \end{array} + Br \\ R^{2} \\ R^{2} \\ R^{2} \end{array} \begin{array}{c} 1) In, THF/H_{2}O=1:1 \\ 2) Jones reagent \\ 3) NH_{2}OH HCI, NaOAc \\ 4) CH_{3}COCI, TEA \\ R^{3} \\ R^{4} \end{array} \begin{array}{c} OAc \\ N \\ R^{2} \\ R^{1} \\ R^{3} \\ R^{4} \end{array}$$

1) To a solution of the 3-bromo-2-methylprop-1-ene (1.1 equiv) or its analogue and indium (1.1 equiv) in THF/H₂O (1:1) was stirred at room temperature while the aldehyde (1.0 equiv) was added to the solution. Then, the resulting mixture was stirred for 10-24 h at the same temperature. After completion of the reaction, saturated ammonium chloride was added at 0°C. The THF layer was separated from the aqueous layer, which was extracted with diethyl ether for 3 times. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude allylic alcohol product was directly used in the next step without further purification.^[5]

Steps 2), 3), and 4) are same as above-mentioned.

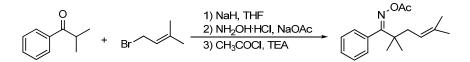
Procedure for Preparation of Ketoxime Acetate 1o:



1) Benzaldehyde (1.0 equiv) and 3-bromo-1-cyclohexene (1.8 equiv) were added in water and the solution was kept at room temperature with stirring while the $SnCl_2 \cdot H_2O$ (2.0 equiv) and copper powder (1.0 equiv) were added. Then, the resulting mixture was vigorously stirred at this temperature for 24 h. After completion of the reaction, the mixture was extracted with ethyl acetate (3×30 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered and evaporated. The crude material was purified by flash chromatography on silica gel to afford the cyclohex-2-en-1-yl(phenyl)methanol.^[6]

Steps 2), 3), and 4) are same as above-mentioned.

Procedure for Preparation of Ketoxime Acetate 1q:



1) To a solution of the isobutyrophenone (0.60 mL, 3.88 mmol) in anhydrous THF (15 mL) was added the NaH (186 mg, 4.65 mmol). The solution was stirred at room temperature for 0.5h. Then, the solution of 1-bromo-3-methylbut-2-ene (0.69 g, 4.66 mmol) in THF (5.0 mL) was slowly added to the solution at this temperature and the resulting suspension was stirred at reflux for 8 h. After completion of the reaction, the reaction mixture was cooled to room temperature. The reaction was quenched by adding saturated aqueous NH₄Cl, and the mixture was extracted with ethyl acetate for 3 times. The combined organic extracts were washed with water, brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude material was purified by flash chromatography on silica gel.^[7]

Steps 2), and 3) are same as above-mentioned.

Reference:

[1] Dam, J. H.; Fristrup, P.; Madsen, R. J. Org. Chem. 2008, 73, 3228.

[2] Jiang, D.; Peng, J.; Chen, Y. Org. Lett. 2008, 10, 1695.

[3] Garzan, A.; Jaganathan, A.; Marzijarani, N. S.; Yousefi, R.; Whitehead, D. C.; Jackson, J. E.; Borhan, B. *Chem. Eur. J.* 2013, **19**, 9015.

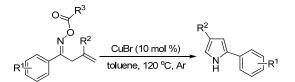
[4] Zaman, S.; Mitsuru, K.; Abell, A. D. Org. Lett. 2005, 7, 609.

[5] Zhu, M.-K.; Zhao, J.-F.; Loh, T.-P. J. Am. Chem. Soc. 2010, 132, 6284.

[6] Tan, X.-H.; Tao, C.-Z.; Hou, Y.-Q.; Luo, L.; Liu, L.; Guo, Q.-X. *Chin. J. Chem.* 2005, **23**, 237.

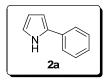
[7] Han, B.; Yang, X.-L.; Fang, R.; Yu, W.; Wang, C.; Duan, X.-Y.; Liu, S. Angew. Chem., Int. Ed. 2012, **51**, 8816.

3. General Procedure for Synthesis of 2-Arylpyrroles

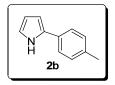


Ketoxime carboxylate (0.3 mmol), CuBr (10 mol%, 4.3 mg), and toluene (3 mL) was charged in a 25 mL round bottom flask. Then, the flask was evacuated and back-filled with Ar (3-times, balloon) and stirred at 120 °C. When the reaction was completed (detected by TLC), the mixture was cooled to room temperature. The reaction was quenched with H₂O (10 mL) and extracted with ethyl acetate (3×10 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and then evaporated in vacuo. The residue was purified by column chromatography on silica gel to afford the corresponding pyrroles with hexanes/ethyl acetate as the eluent.

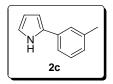
4. Spectroscopic Data for Products



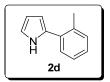
2a: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.28$ (s, 1 H), 7.62 (d, J = 7.6 Hz, 2 H), 7.34 (t, J = 7.6 Hz, 2 H), 7.14 (t, J = 7.2 Hz, 1 H), 6.84 (s, 1 H), 6.51 (s, 1 H), 6.11 (s, 1 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 133.0$, 131.1, 128.7, 125.4, 123.3, 119.3, 109.1, 105.6. HRMS Calcd (ESI) m/z for C₁₀H₁₀N: [M+H]⁺ 144.0808, found: 144.0814.



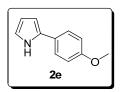
2b: ¹H NMR (CDCl₃, 400 MHz): $\delta = 8.38$ (s, 1 H), 7.36 (d, J = 7.6 Hz, 2 H), 7.17 (d, J = 7.2 Hz, 2 H), 6.83 (s, 1 H), 6.48 (s, 1 H), 6.29 (s, 1 H), 2.35 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 135.8$, 132.1, 129.9, 129.5, 123.7, 118.4, 109.8, 105.3, 21.1. HRMS Calcd (ESI) m/z for C₁₁H₁₂N: [M+H]⁺ 158.0964, found: 158.0967.



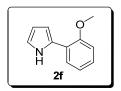
2c: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.25$ (s, 1 H), 7.46-7.41 (m, 2 H), 7.22 (t, J = 7.2 Hz, 1 H), 6.96 (d, J = 6.8 Hz, 1 H), 6.83 (s, 1 H), 6.49 (s, 1 H), 6.11 (s, 1 H), 2.32 (s, 3 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 137.7$, 133.0, 131.2, 128.6, 126.1, 124.0, 120.6, 119.2, 109.0, 105.5, 21.2. HRMS Calcd (ESI) m/z for C₁₁H₁₀N: [M-H]⁺ 156.0819, found: 156.0823.



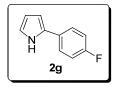
2d: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 10.98$ (s, 1 H), 7.35 (d, J = 7.2 Hz, 1 H), 7.21-7.15 (m, 2 H), 7.11-7.08 (m, 1 H), 6.80 (s, 1 H), 6.19 (s, 1 H), 6.10 (s, 1 H), 2.36 (s, 3 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 134.1$, 133.0, 130.8, 130.3, 127.7, 126.0, 125.8, 118.4, 108.5, 108.4, 21.3. HRMS Calcd (ESI) m/z for C₁₁H₁₂N: [M+H]⁺ 158.0964, found: 158.0960.



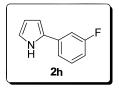
2e: ¹H NMR (CDCl₃, 400 MHz): $\delta = 8.33$ (s, 1 H), 7.40 (d, J = 8.0 Hz, 2 H), 6.91 (d, J = 8.4 Hz, 2 H), 6.82 (s, 1 H), 6.41 (s, 1 H), 6.28 (s, 1 H), 3.82 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 158.1$, 132.0, 125.8, 125.2, 118.2, 114.3, 109.8, 104.8, 55.2. HRMS Calcd (ESI) m/z for C₁₁H₁₂NO: [M+H]⁺ 174.0913, found: 174.0920.



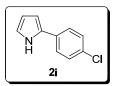
2f: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 10.94$ (s, 1 H), 7.62 (d, J = 7.2 Hz, 1 H), 7.15 (t, J = 7.6 Hz, 1 H), 7.04 (d, J = 8.0 Hz, 1 H), 6.96 (t, J = 7.6 Hz, 1 H), 6.83 (s, 1 H), 6.60 (s, 1 H), 6.12 (s, 1 H), 3.87 (s, 3 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 154.7$, 127.9, 126.5, 126.0, 121.4, 120.7, 118.6, 111.8, 108.2, 107.8, 55.3. HRMS Calcd (ESI) m/z for C₁₁H₁₂NO: [M+H]⁺ 174.0913, found: 174.0920.



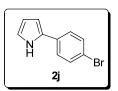
2g: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.28$ (s, 1 H), 7.63 (d, J = 5.2 Hz, 2 H), 7.18 (t, J = 7.2 Hz, 2 H), 6.84 (s, 1 H), 6.47 (s, 1 H), 6.11 (s, 1 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 160.4$ (d, $J_{CF} = 240.6$ Hz), 130.2, 129.7, 125.1 (d, $J_{CF} = 7.8$ Hz), 119.3, 115.5 (d, $J_{CF} = 21.3$ Hz), 109.1, 105.5. HRMS Calcd (ESI) m/z for C₁₀H₇FN: [M-H]⁺ 160.0568, found: 160.0570.



2h: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.37$ (s, 1 H), 7.46 (d, J = 8.0 Hz, 2 H), 7.38-7.33 (m, 1 H), 6.94 (t, J = 6.8 Hz, 1 H), 6.88 (s, 1 H), 6.60 (s, 1 H), 6.13 (s, 1 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 162.8$ (d, $J_{CF} = 240.6$ Hz), 135.4 (d, $J_{CF} = 8.6$ Hz), 130.6 (d, $J_{CF} = 8.8$ Hz), 129.9, 120.0, 119.3, 111.8 (d, $J_{CF} = 21.1$ Hz), 109.7 (d, $J_{CF} = 22.7$ Hz), 109.3, 106.8. HRMS Calcd (ESI) m/z for C₂₀H₁₇F₂N₂: [2M+H]⁺ 323.1354, found: 323.1339.

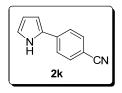


2i: ¹H NMR (CDCl₃, 400 MHz): $\delta = 8.41$ (s, 1 H), 7.39 (d, J = 8.4 Hz, 2 H), 7.32 (d, J = 8.4 Hz, 2 H), 6.87 (s, 1 H), 6.51 (s, 1 H), 6.30 (s, 1 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 131.6$, 131.1, 130.9, 128.9, 124.9, 119.2, 110.2, 106.3. HRMS Calcd (ESI) m/z for C₁₀H₇ClN: [M-H]⁺ 176.0273, found: 176.0275.

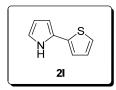


2j: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.37$ (s, 1 H), 7.58 (d, J = 8.4 Hz, 2 H), 7.51

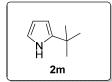
(d, J = 8.0 Hz, 2 H), 6.88 (s, 1 H), 6.55 (s, 1 H), 6.13 (s, 1 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 132.2$, 131.5, 129.9, 125.2, 119.9, 117.9, 109.3, 106.3. HRMS Calcd (ESI) m/z for C₁₀H₇BrN: [M-H]⁺219.9767, found: 219.9778.



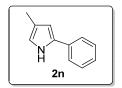
2k: ¹H NMR (DMSO- d^6 , 400 MHz): $\delta = 11.55$ (s, 1 H), 7.74 (s, 4 H), 6.95 (s, 1 H), 6.71 (s, 1 H), 6.15 (s, 1 H); ¹³C NMR (DMSO- d^6 , 100 MHz): $\delta = 137.1$, 132.7, 129.4, 123.5, 121.7, 119.4, 110.0, 108.7, 106.9. HRMS Calcd (ESI) m/z for C₁₁H₈N₂Na: [M+Na]⁺ 191.0580, found: 191.0579.



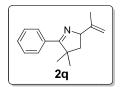
21: ¹H NMR (CDCl₃, 400 MHz): $\delta = 8.29$ (s, 1 H), 7.14 (d, J = 4.0 Hz, 1 H), 7.02 (s, 1 H), 7.01 (s, 1 H), 6.80 (s, 1 H), 6.41 (s, 1 H), 6.25 (s, 1 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 136.2$, 127.6, 126.6, 122.6, 120.8, 118.5, 109.9, 106.6. HRMS Calcd (ESI) m/z for C₈H₈NS: [M+H]⁺ 150.0372, found: 150.0374.



2m: ¹H NMR (CDCl₃, 400 MHz): $\delta = 7.92$ (s, 1 H), 6.59 (s, 1 H), 6.05 (s, 1 H), 5.87 (s, 1 H), 1.22 (s, 9 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 141.8$, 115.9, 107.7, 102.3, 31.3, 30.6.



2n: ¹H NMR (CDCl₃, 400 MHz): $\delta = 8.08$ (s, 1 H), 7.39 (d, J = 7.2 Hz, 2 H), 7.31 (t, J = 7.2 Hz, 2 H), 7.16 (t, J = 7.2 Hz, 1 H), 6.55 (s, 1 H), 6.36 (s, 1 H), 2.14 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 132.8$, 131.9, 128.8, 125.9, 123.6, 120.5, 116.8, 107.4, 11.9. HRMS Calcd (ESI) m/z for C₁₁H₁₂N: [M+H]⁺ 158.0964, found: 158.0970.



2q: ¹H NMR (CDCl₃, 400 MHz): δ = 7.75 (d, *J* = 3.6 Hz, 2 H), 7.38 (s, 3 H), 5.03 (s, 1 H), 4.86 (s, 1 H), 4.51 (t, *J* = 7.6 Hz, 1 H), 2.15-2.10 (m, 1 H), 1.81 (s, 3 H), 1.74 (t, *J* = 9.6 Hz, 1 H), 1.39 (s, 3 H), 1.34 (s, 3 H); ¹³C NMR (CDCl₃, 100 MHz): δ = 179.6, 146.9, 134.6, 129.4, 128.0, 127.8, 110.0, 72.2, 50.3, 47.2, 27.0, 25.8, 19.8. HRMS Calcd (ESI) m/z for C₁₅H₂₀N: [M+H]⁺214.1590, found: 214.1597.

5. Copies of ¹H and ¹³C NMR Spectra

